Supporting Information

Disproportionation of Dipyrrolylquinoxaline Radical Anions via the Internal Protons of the Pyrrole Moieties

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Experimental Details

Chemicals. Acetonitrile was distilled over CaH₂. Tetra-*n*-butylammonium perchlorate (Bu₄NClO₄) was purchased from Sigma Chemical Co. or Fluka Chemika Co., recrystallized from ethanol and dried under vacuum at 40 °C for at least one week prior to use. Cobaltocene (CoCp₂), tetrabutylammonium hydroxide (Bu₄NOH, 37% in MeOH) and quinoxaline (6) were purchased commercially and used as received without further purification. 2,3-Dipyrrol-2'-yl-6,7-dinitroquinoxaline (1H₂),^{S1} 2,3-dipyrrol-2'-yl-6-nitroquinoxaline (2H₂)^{S1} and 2,3-dipyrrol-2'- ylquinoxaline (3H₂)^{S1}, 6,7-dinitroquinoxaline (4)^{S2} and 6-nitroquinoxaline (5)^{S2} were synthesized as according literature. CD₃CN and DMSO-*d*₆ were purchased from Cambridge Isotope Laboratories, Co.Ltd.

Absorption Spectral Measurements. UV–vis absorption spectra and spectral changes were recorded on a Hewlett Packard 8453 diode array spectrophotometer using 1 cm cuvette. ¹H NMR spectra of synthesized chemicals and reaction products were recorded on a JEOL AL300 (300 MHz) spectrometer. The chemical shifts are expressed in ppm relative to tetramethylsilane (TMS).

Electrochemistry. Cyclic voltammograms were obtained with an EG&G Princeton Applied Research (PAR) 173 potentiostat/galvanostat. A homemade three-electrode cell was used and consisted of a platinum button or glassy carbon working electrode, a platinum counter electrode and a homemade saturated calomel reference electrode (SCE). The SCE was separated from the bulk of the solution by a fritted glass bridge of low porosity which contained the solvent/supporting electrolyte mixture. UV-visible spectroelectrochemical experiments were performed with a home-built thin-layer cell that had a light transparent platinum net working electrode.^{S3} Potentials were applied and monitored with an EG&G PAR Model 173 potentiostat. Time-resolved UV-visible spectra were recorded with a Hewlett-Packard Model 8453 diode array spectrophotometer.

EPR Measurements. EPR spectra of $1H_2^-$ and 4^- were recorded on a JEOL JES-RE1XE spectrometer. The magnitude of modulation was chosen to optimize the resolution and signal-to-noise (S/N) ratio of the observed spectra under non-saturating microwave power conditions. The g values and hyperfine coupling constants were

calibrated using an Mn²⁺ marker.

Calculations. Theoretical calculations of the properties of molecules were performed using Density-Functional Theory (DFT) with the UB3LYP density-functional and the 6-31G(d) basis set.⁸⁴ All calculations were performed using Gaussian 09, revision A.02.⁸⁵ Graphical outputs of the computational results were generated with the Gauss View software program (ver. 3.09) developed by Semichem, Inc.⁸⁶

(S1) (a) Black, C. B.; Andrioletti, B.; Try, A. C.; Ruiperez, C.; Sessler, J. L. J. Am. Chem. Soc. 1999, 121, 10438. (b) Anzenbacher, P. Jr., Try, A. C.; Miyaji, H.; Jursíková, K.; Lynch, V. M.; Marquez, M.; Sessler, J. L. J. Am. Chem. Soc. 2000, 122, 10268.

(S2) Fisons Pest Control Ltd. *Quinoxaline fungicides*. **1963**, *26*, dpp. BE 631044 19631104.

(S3) Lin, X. Q.; Kadish. K. M. Anal. Chem. 1985, 57, 1498.

(S4) Becke, A. D. J. Chem. Phys. 1993, 98, 5648.

(S5) Gaussian 09, Revision A.02, Gaussian, Inc.: Wallingford CT, 2004.

(S6) Dennington II, R.; Keith, T.; Millam, J.; Eppinnett, K.; Hovell, W. L.; Gilliland, R.

Gaussview; Semichem, Inc.,: Shawnee Mission, KS,. 2003.

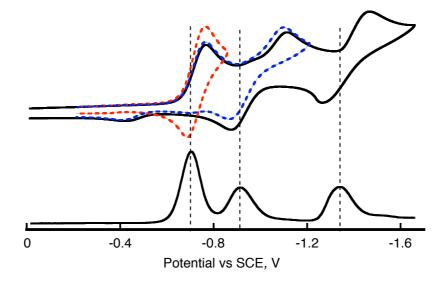


Figure S1. Cyclic voltammogram and differential pulse voltammogram of $1H_2$ in deaerated MeCN containing Bu_4NPF_6 (0.10 M) at 298 K; scan rate: 0.1 V s⁻¹.

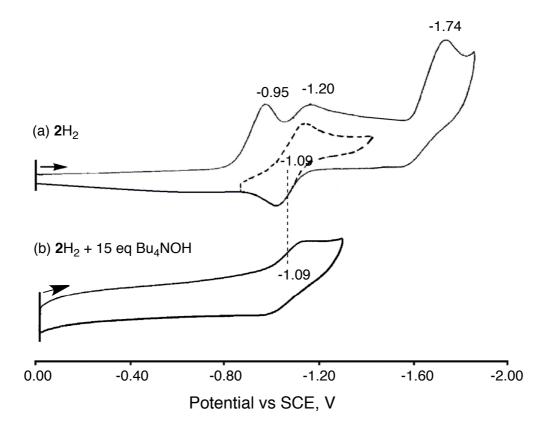
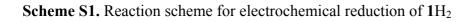
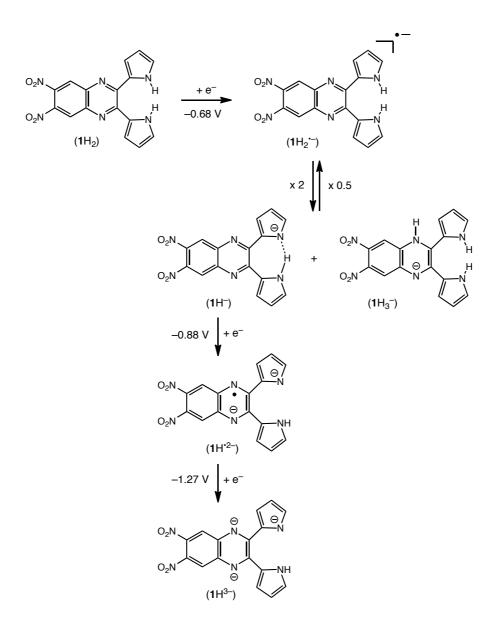


Figure S2. Cyclic voltammogram of (a) $2H_2$ and (b) $2H^-$ produced by the reaction of $2H_2$ with Bu₄NOH in MeCN containing Bu₄NClO₄ (0.10 M) at 298 K; scan rate: 100 mV s⁻¹.





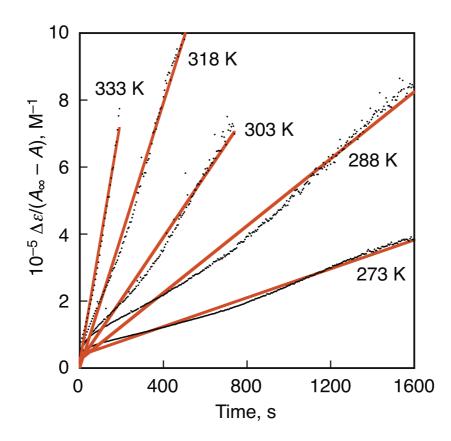


Figure S3. Second-order plots for disproportionation of $1H_2^{-}$ produced in electron transfer from CoCp₂ (2.5 × 10⁻⁵ M) to $1H_2$ (2.5 × 10⁻⁵ M) in deaerated MeCN at various temperatures (273-333 K).

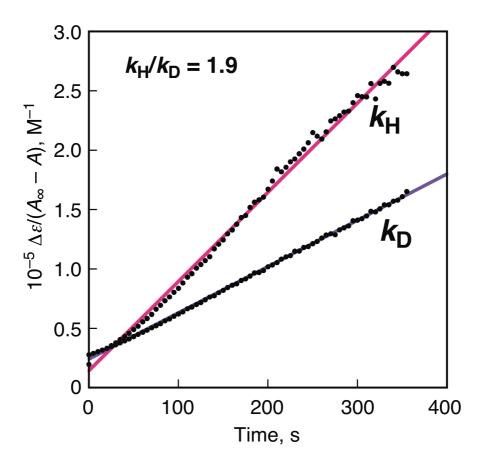


Figure S4. Second-order plots for the disproportionation of $1H_2^{-1}$ (5.0 x 10^{-5} M) in the presence of H₂O (5.0 x 10^{-4} M) and D₂O (5.0 x 10^{-4} M) in deaerated MeCN at 298 K.

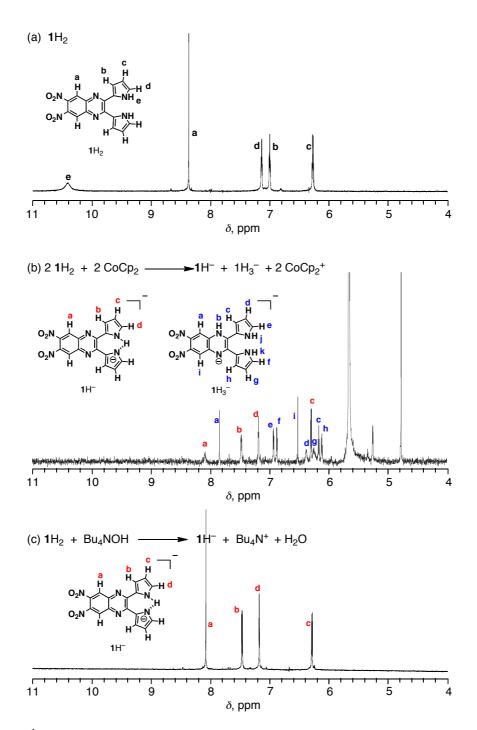


Figure S5. ¹H NMR spectra of (a) $1H_2$ (a-e in black), (b) $1H^-$ (a-d in red) and $1H_3^-$ (a-i in blue) produced by electron-transfer reduction of $1H_2$ (5.0 mM) with CoCp₂ (5.0 mM) and (c) $1H^-$ produced by deprotonation of $1H_2$ (5.0 mM) with Bu₄NOH (5.0 mM) in deaerated CD₃CN. The signals of b, k and j N-H protons of $1H_3^-$ are too broad to observe due to the small amout of H_2O present in CD₃CN. Asterisk (*) denote the signals of CoCp⁺ and decomposed products.

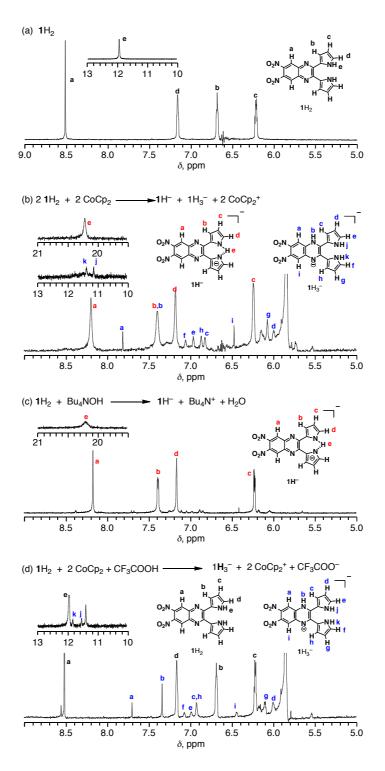


Figure S6. ¹H NMR spectra of (a) $1H_2$ (a-e in black), (b) $1H^-$ (a-e in red) and $1H_3^-$ (a-k in blue) produced by the electron-transfer reduction of $1H_2$ (5.0 mM) with CoCp₂ (5.0 mM) and (c) $1H^-$ (a-e in red) produced by deprotonation of $1H_2$ (5.0 mM) with Bu₄NOH (5.0 mM) (d) $1H_3^-$ (a-k in blue) produced by the electron-transfer reduction of $1H_2$ (5.0 mM) with CoCp₂ (4.0 mM) in the presence of trifluoroacetic acid (2.0 mM) in deaerated DMSO- d_6 . The signals of $1H_2$ as a starting material are shown with black labels. Asterisk (*) denote the signals of CoCp₂⁺ and decomposed product.

¹H NMR Assignments

a) In CD₃CN

1H₂: ¹H NMR (300 MHz, CD₃CN) δ 6.24 – 6.29 (2H, m, pyrrole H), 6.97 – 7.03 (2H, m, pyrrole H), 7.10 – 7.16 (2H, m, pyrrole H), 8.36 (2H, s, quinoxaline H), 10.40 (2H, br, s, pyrrole NH)

1H⁻: ¹H NMR (300 MHz, CD₃CN) δ 6.28 – 6.32 (2H, dd, J = 3.7, 2.2 Hz, pyrrole H), 7.15 – 7.20 (2H, dd, J = 2.2, 1.5 Hz, pyrrole H), 7.44 – 7.50 (2H, dd, J = 3.7, 1.5 Hz, pyrrole H), 8.05 (2H, br, s, quinoxaline H)

1H₃⁻: ¹H NMR (300 MHz, CD₃CN) δ 6.08 – 6.13 (1H, t, J = 2.4 Hz, pyrrole H), 6.15 – 6.19 (1H, t, J = 2.4 Hz, pyrrole H), 6.20 – 6.28 (1H, br, m, pyrrole H), 6.35 – 6.40 (1H, br, m, pyrrole H), 6.51 (1H, s, quinoxaline H), 6.86 – 6.90 (1H, dd, J = 2.4, 1.8 Hz, pyrrole H), 6.91 – 6.95 (1H, dd, J = 2.4, 1.8 Hz, pyrrole H), 7.83 (1H, s, quinoxaline H)

b) In DMSO-d₆

1H₂: ¹H NMR (300 MHz, DMSO- d_6) δ 6.20 – 6.28 (2H, m, pyrrole H), 6.68 – 6.77 (2H, m, pyrrole H), 7.15 – 7.22 (2H, m, pyrrole H), 8.50 (2H, s, quinoxaline H), 11.94 (2H, br, s, pyrrole NH)

1H⁻: ¹H NMR (300 MHz, DMSO- d_6) δ 6.22 (2H, m, pyrrole H), 7.17 (2H, m, pyrrole H), 7.38 (2H, m, pyrrole H), 8.17 (2H, s, quinoxaline H), 20.20 (1H, br, s, NH)

1H₃⁻: ¹H NMR (300 MHz, DMSO-*d*₆) δ 5.99 (1H, m, pyrrole H), 6.07 (1H, m, pyrrole H), 6.47 (1H, s, quinoxaline H), 6.82 (2H, m, pyrrole H), 6.96 (1H, m, pyrrole H), 7.06 (1H, m, pyrrole H), 7.4 (1H, br, s, quinoxaline NH), 7.81 (1H, s, quinoxaline H), 11.12 (1H, br, s, pyrrole NH), 11.36 (1H, br, s, pyrrole NH).

The NMR spectra of $1H_2$, $1H^-$ and $1H_3^-$ in DMSO- d_6 are virtually the same as those in CD₃CN although small shifts are observed due to the difference in the solvent and the hydrogen-bonded NH proton is only observed in DMSO- d_6 . The NMR spectra of $1H_3^-$ produced by the electron-transfer reduction of $1H_2$ with CoCp₂ in the presence of trifluoroacetic acid (Figure S6d) are also vertually the same as those of $1H_3^-$ produced by disproportionation of $1H_2^-$ (Figure S6b), although small shifts are observed due to the presence of the trifluoroacetate anion.

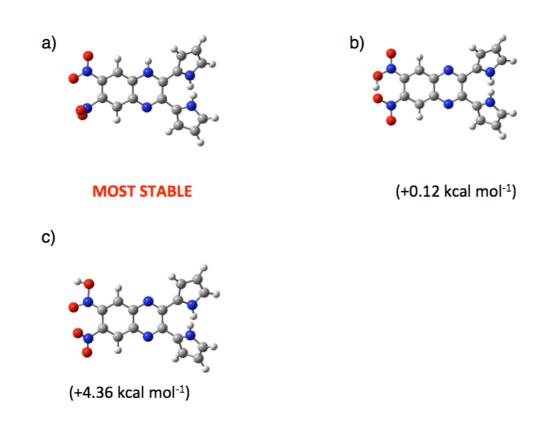


Figure S7. Optimized structures of $1H_3^-$ isomers as determined via DFT calculations carried out at the B3LYP/6-31G(d) level. Relative energies are given in parentheses.

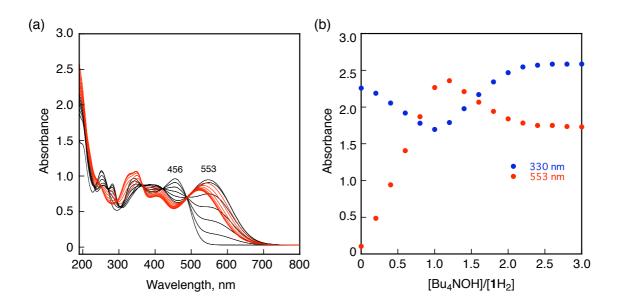


Figure S8. (a) Titration of $1H_2$ (0.50 mM) with Bu₄NOH in MeCN at 298 K. The black line shows the titration up to 1 equiv of Bu₄NOH to produce $1H^-$ and red line shows the titration from 1 equiv to 2 equiv of Bu₄NOH to produce 1^{2-} . (b) Plots of absorbance at 330 and 553 nm vs [Bu₄NOH]/[1H₂].

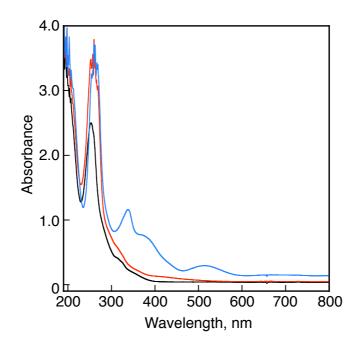


Figure S9. UV-vis absorption spectra of **4** (5.0×10^{-5} M, black), and **4**^{•-} (blue) produced by electron-transfer reduction of **4** (5.0×10^{-5} M) with CoCp₂ (5.0×10^{-5} M), and **4**^{•-} with CF₃COOH (5.0×10^{-5} M) (red) in deaerated MeCN at 298 K.

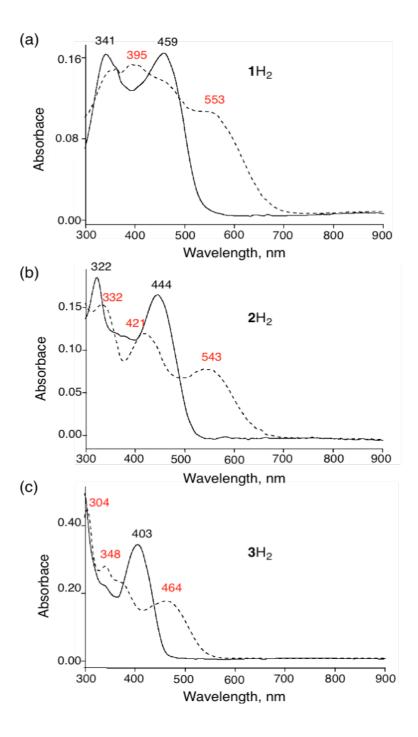


Figure S10. UV-vis absorption spectral changes in electrolysis of (a) $1H_2$ (3.3×10^{-4} M), (b) $2H_2$ (3.1×10^{-4} M) and (c) $3H_2$ (9.2×10^{-4} M) in the presence of Bu₄NClO₄ (0.20 M) in MeCN at applied potentials of -0.80 V (vs SCE) for 347 s, -0.96 V for 310 s and -1.70 V for 180 s, respectively.

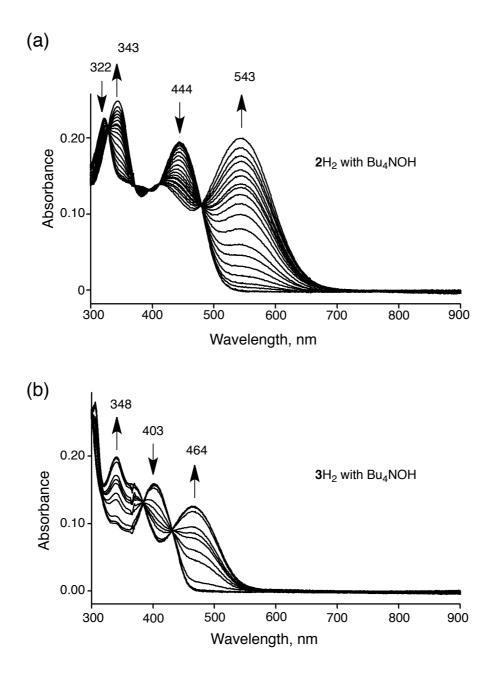


Figure S11. UV-vis absorption spectral change of (a) $2H_2$ (1.7×10^{-5} M) and (b) $3H_2$ (2.0×10^{-5} M) in MeCN titrated with Bu₄NOH (1.0×10^{-2} M).

<i>Т</i> , К	[1 H ₂ •-], mM	Κ
243	3.7×10^{-1}	0.48
258	3.2×10^{-1}	0.78
273	2.0×10^{-1}	2.8
288	1.0×10^{-1}	13
303	4.5×10^{-2}	87
318	1.9×10^{-2}	5.4×10^2

Table S1. Equilibrium Constants between $1H_2^-$ and $1H^-/1H_3^-$ in MeCN at Various Temperatures

K values were determined by EPR spectra calibrated by DPPH.

Table S2. Rate Constants for the Disproportionation of 1H₂⁻⁻ in MeCN at Various Temperatures

<i>Т</i> , К	$k, M^{-1} s^{-1}$
273	1.1 x 10 ²
288	$2.5 \ge 10^2$
303	$4.6 \ge 10^2$
318	$1.0 \ge 10^3$
333	$1.8 \ge 10^3$

k values were determined by using $\Delta \varepsilon_{553} = 1.8 \times 10^4 \text{ M}^{-1} \text{ cm}^{-1}$.