

Supporting Information

**The 3-Deoxy Analogue of α -GalCer: Disclosing the
Role of the 4-Hydroxyl Group for CD1d-Mediated
NKT Cell Activation**

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Won-Jea Cho, Chang-Yuil Kang, and Sanghee Kim**

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A. Figures

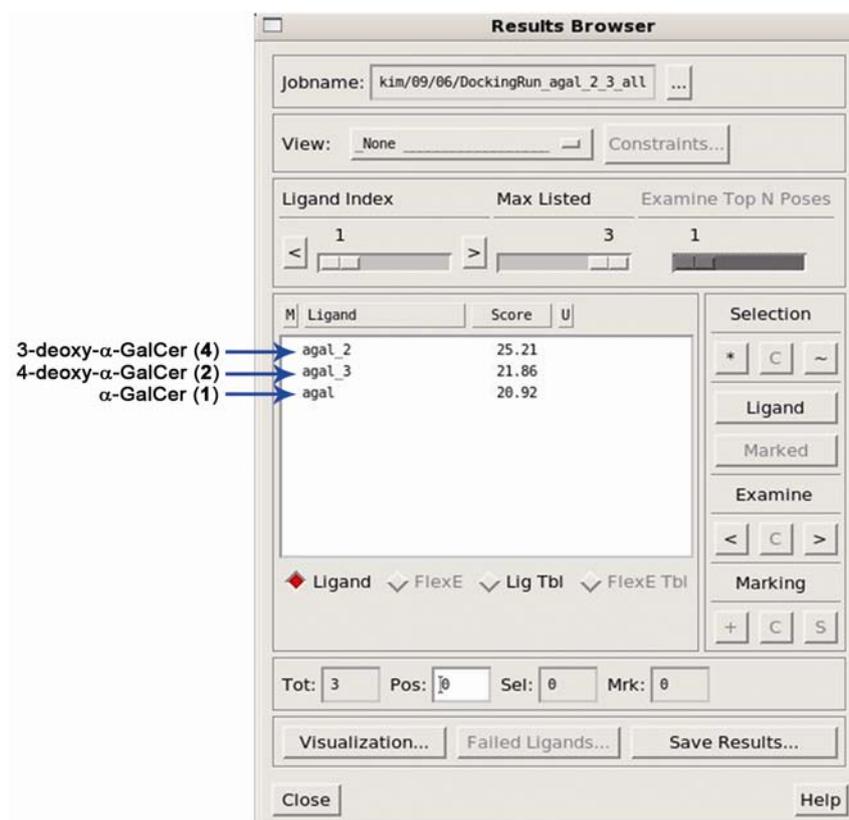


Figure S1. Surflex-Dock docking scores of compounds 1, 2, and 4.

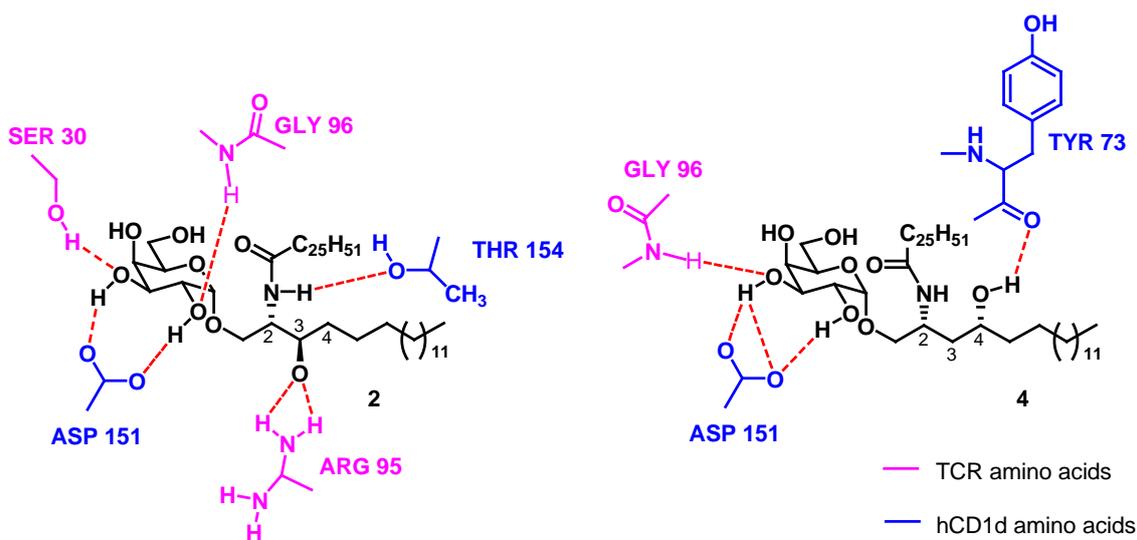


Figure S2. The hydrogen bonding modes of the docked compounds 2 and 4.

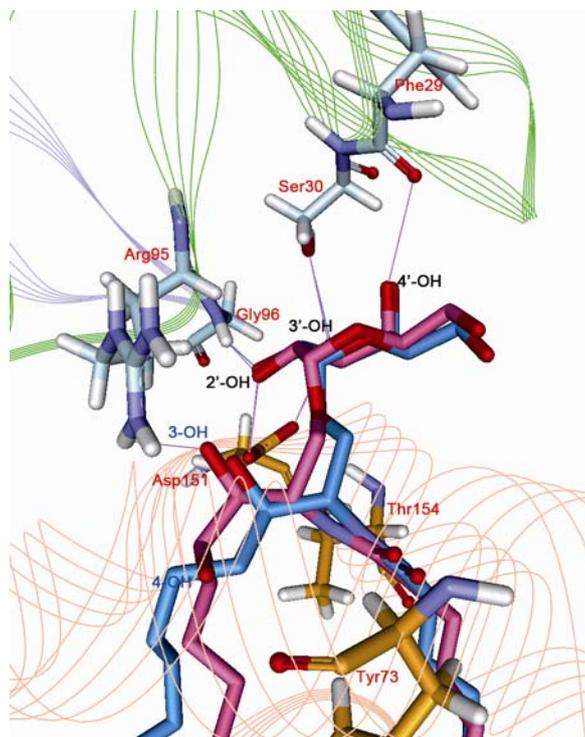


Figure S3. Comparison of the docking model of 4-deoxy- α -GalCer **2** (blue backbone) with the crystal structure of α -GalCer **1** (pink backbone, Protein Data Bank code 2PO6). The key amino acid residues are shown in light blue (TCR residues) and orange (CD1d residues).

B. Experimental Methods

(a) Biological methods

Glycolipids were dissolved in a 0.5% solution of Tween 20 in saline as a vehicle at a concentration of 200 $\mu\text{g}/\text{mL}$, and diluted with sterile PBS immediately prior to use.

Determination of the stimulating activity for NKT hybridoma cells^{1,2}

Mouse CD1d-transfected rat basophilic leukemia (RBL) cells were loaded with 8 ng/mL α -GalCer (**1**), 4-deoxy- α -GalCer (**2**) or 3-deoxy- α -GalCer (**4**). After 4 h, free glycolipids were removed by washing three times with PBS 3, and the RBL cells were incubated with DN32.D3 NKT hybridoma cells for 16 h. The level of IL-2 secreted into the supernatant was determined by an Enzyme-Linked Immunosorbent Assay (ELISA).

Evaluation of the cytokine levels produced by primary splenocytes³

Splenocytes from naïve C57BL/6 mice were cultured in the presence of 0.5 ng/mL α -GalCer (**1**), 4-deoxy- α -GalCer (**2**)⁴ or 3-deoxy- α -GalCer (**4**) for 72 h. The levels of IFN- γ and IL-4 secreted into the supernatant were determined by ELISA.

(b) Molecular modeling methods

In order to perform docking studies of α -GalCer analogues with hCD1d of NKT, the Surflex-Dock program in Sybyl version 8.1.1 (Tripos Associates) was operated on an IBM computer (Intel Pentium 4, 2.8GHz CPU, 1GB memory) running the Red Hat Linux 4.0 operating system. The X-ray crystallographic structure of hCD1d complex (PDB code 2PO6) was used for docking analysis. The structures of the α -GalCer

analogues were drawn into the Sybyl package with standard bond lengths and angles and minimized using the conjugate gradient method. The Gasteiger-Hückel charge, with a distance-dependent dielectric function, was applied for the minimization process.

Surflex-Dock docks the molecules into a receptor-binding site that is defined to a protomol. The protomol is a unique and important factor of the docking algorithm and is a computational representation of assumed ligands that interact with the binding site. Surflex-Dock's scoring function contains hydrophobic, polar, repulsive, entropic, and solvation terms. After running Surflex-Dock on the α -GalCer derivatives, the scores of 20 docked conformers were ranked in a molecular spreadsheet, and the best total score conformer was chosen as the binding mode of the ligand to the active site.

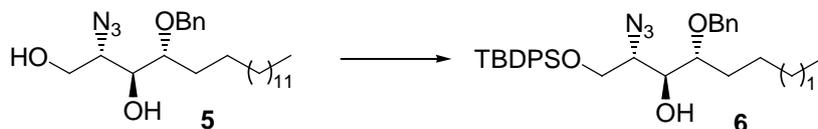
(c) Synthetic procedures and compound characterization

General methods

All chemicals were reagent grade and used as purchased. All reactions were performed under an inert atmosphere of dry argon or nitrogen and using dry distilled solvents. Reactions were monitored by TLC analysis using silica gel 60 F-254 thin layer plates. Flash column chromatography was carried out on silica gel (230–400 mesh). Optical rotations were measured using a polarimeter set for sodium D light (589.3 nm). ^1H NMR (500, 400 or 300 MHz) and ^{13}C NMR (100 or 75 MHz) spectra were recorded in δ units relative to the non-deuterated solvent as an internal reference. High-resolution mass spectra (HRMS) were recorded using fast atom bombardment (FAB). The purities of the products were >95% based on proton NMR spectra.

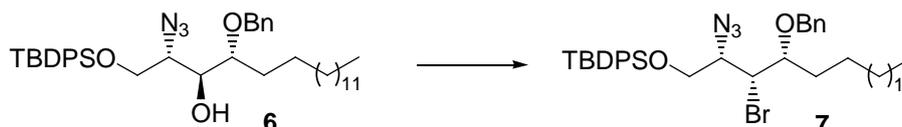
Compound synthesis

(2*S*,3*S*,4*R*)-2-Azido-4-(benzyloxy)-1-(*tert*-butyldiphenylsilyloxy)octadecan-3-ol (6)



Triethylamine (0.2 mL, 1.40 mmol) and TBDPS-Cl (0.29 mL, 1.13 mmol) were added to a cooled (0 °C) solution of **5** (405 mg, 0.93 mmol) in CH₂Cl₂ (10 mL). The reaction mixture was stirred at room temperature for 12 h and quenched with saturated aqueous NaHCO₃ solution. The mixture was then extracted with EtOAc. The organic extracts were washed with brine, dried over Na₂SO₄, and then concentrated under reduced pressure. The residue was purified using silica gel column chromatography (hexane/EtOAc, 20:1) to give **6** (622 mg, 99%) as a yellow liquid: ¹H NMR (400 MHz, CDCl₃) δ 0.87 (t, *J* = 6.4 Hz, 3H), 1.07 (s, 9H), 1.13–1.32 (m, 24H), 1.39–1.64 (m, 2H), 2.36 (d, *J* = 3.7 Hz, 1-OH), 3.42–3.53 (m, 2H), 3.82–3.90 (m, 2H), 4.02 (dd, *J* = 2.7, 10.8 Hz, 1H), 4.45 (dd, *J* = 11.3, 18.0 Hz, 2H), 7.22–7.30 (m, 4H), 7.37–7.45 (m, 6H), 7.66–7.70 (m, 5H); ¹³C NMR (100 MHz, CDCl₃) δ 14.1, 19.1, 22.7, 25.2, 26.7, 28.3, 29.3, 29.57, 29.61, 29.65, 29.7, 31.9, 63.4, 64.9, 70.3, 71.8, 79.5, 127.77, 127.83, 128.4, 129.8, 132.80, 132.84, 135.6, 138.1; HRMS (FAB) calcd for C₄₁H₆₂O₃N₃Si 672.4560 ([M+H]⁺), found 672.4537.

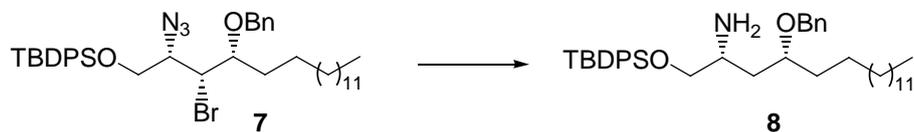
((2*S*,3*R*,4*R*)-2-Azido-4-(benzyloxy)-3-bromooctadecyloxy)(*tert*-butyl)diphenylsilane (7)



Triphenylphosphine (498 mg, 1.92 mmol) and carbon tetrabromide (630 mg, 1.92 mmol) were added to a solution of **6** (318 mg, 0.48 mmol) in CH₂Cl₂ (10 mL). The

reaction mixture was stirred at room temperature for 1 day. After the addition of EtOAc, the solid was filtered. The filtrate was evaporated and the crude product was purified using silica gel column chromatography (hexane/EtOAc, 20:1) to give **7** (250 mg, 72%) as a yellow liquid; ^1H NMR (400 MHz, CDCl_3) δ 0.81 (t, $J = 6.7$ Hz, 3H), 1.02 (s, 9H), 1.12–1.25 (m, 24H), 1.32–1.76 (m, 2H), 3.47–3.51 (m, 1H), 3.68–3.82 (m, 3H), 4.20–4.23 (m, 1H), 4.47 (dd, $J = 11.2, 19.6$ Hz, 2H), 7.16–7.20 (m, 4H), 7.26–7.40 (m, 6H), 7.57–7.62 (m, 5H); ^{13}C NMR (100 MHz, CDCl_3) δ 14.1, 19.1, 22.7, 25.5, 26.8, 26.82, 26.84, 29.36, 29.43, 29.51, 29.55, 29.6, 29.65, 29.69, 31.9, 55.6, 63.9, 65.0, 72.1, 79.9, 127.7, 127.76, 127.81, 127.84, 127.9, 128.4, 130.0, 135.57, 135.61, 135.63, 135.7, 137.9; HRMS (FAB) calcd for $\text{C}_{41}\text{H}_{61}\text{BrO}_2\text{N}_3\text{Si}$ 734.3716 ($[\text{M}+\text{H}]^+$), found 734.3742.

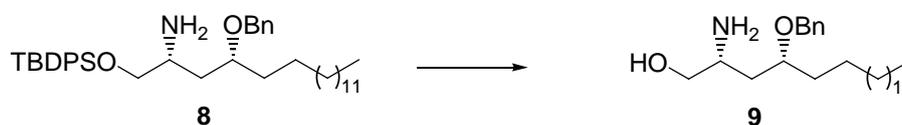
(2R,4R)-4-(Benzyloxy)-1-(tert-butylidiphenylsilyloxy)octadecan-2-amine (8)



Nikel(II) chloride (79 mg, 0.61 mmol) and sodium borohydride (46 mg, 1.22 mmol) were added to a solution of **7** (225 mg, 0.31 mmol) in EtOH (6 mL) at room temperature. The reaction mixture was stirred for 12 h and the solid was removed through a pad of Celite. The filtrate was extracted with EtOAc and water. The organic layer was washed with brine, dried over Na_2SO_4 , and then concentrated under reduced pressure. The residue was purified using silica gel column chromatography ($\text{CH}_2\text{Cl}_2/\text{MeOH}$, 10:1) to give **8** (164 mg, 85%) as a white solid: ^1H NMR (300 MHz, CDCl_3) δ 0.83 (t, $J = 6.7$ Hz, 3H), 1.01 (s, 9H), 1.12–1.27 (m, 24H), 1.32–1.60 (m, 2H), 2.00–2.18 (m, 2H), 2.97–3.05 (m, 1H), 3.38–3.44 (m, 2H), 3.56–3.64 (dd, $J = 4.1, 9.9$ Hz, 1H), 4.45 (d, $J = 11.6$

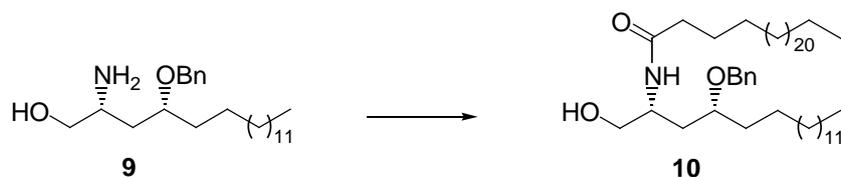
Hz, 1H), 4.31 (d, $J = 11.3$ Hz, 1H), 7.16–7.26 (m, 4H), 7.28–7.40 (m, 6H), 7.58–7.63 (m, 5H); ^{13}C NMR (100 MHz, CDCl_3) δ 14.1, 19.3, 22.6, 25.0, 25.1, 26.9, 29.3, 29.5, 29.7, 29.8, 31.9, 33.8, 38.0, 51.0, 68.6, 70.5, 77.5, 127.4, 127.56, 127.59, 127.67, 127.70, 127.73, 128.3, 129.7, 133.5, 135.55, 135.58, 135.7, 138.7; HRMS (FAB) calcd for $\text{C}_{41}\text{H}_{64}\text{O}_2\text{NSi}$ 630.4706 ($[\text{M}+\text{H}]^+$), found 630.4706.

(2*R*,4*R*)-2-Amino-4-(benzyloxy)octadecan-1-ol (9)



TBAF (0.36 mL, 1.0 M solution in THF, 0.36 mmol) was added to a solution of **8** (112 mg, 0.178 mmol) in THF (3 mL) at room temperature. The reaction mixture was stirred for 12 h and then diluted with water and extracted with CH_2Cl_2 . The organic layer was washed with brine, dried over Na_2SO_4 , and then concentrated under reduced pressure. The crude product was purified using silica gel column chromatography ($\text{CH}_2\text{Cl}_2/\text{MeOH}$, 10:1 to 5:1) to give **9** (56 mg, 80%) as a white solid: ^1H NMR (400 MHz, CDCl_3) δ 0.85 (t, $J = 6.7$ Hz, 3H), 1.15–1.31 (m, 24H), 1.33–1.49 (m, 3H), 1.57–1.76 (m, 2H), 1.82–1.90 (m, 1H), 3.25–3.30 (m, 1H), 3.40–3.43 (m, 1H), 3.57–3.64 (m, 2H), 3.73 (dd, $J = 2.8, 12.0$ Hz, 1H), 4.43 (d, $J = 11.5$ Hz, 1H), 4.55 (d, $J = 11.4$ Hz, 1H), 7.20–7.31 (m, 5H); ^{13}C NMR (100 MHz, CDCl_3) δ 14.1, 19.7, 22.7, 24.0, 24.6, 29.3, 29.56, 29.65, 29.7, 29.8, 31.9, 33.1, 33.6, 52.9, 58.9, 62.0, 70.3, 127.9, 128.2, 128.7, 137.7; HRMS (FAB) calcd for $\text{C}_{25}\text{H}_{46}\text{O}_2\text{N}$ 392.3529 ($[\text{M}+\text{H}]^+$), found 392.3530.

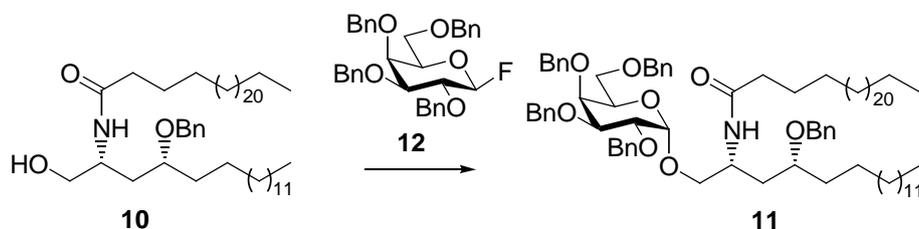
***N*-((2*R*,4*R*)-4-(Benzyloxy)-1-hydroxyoctadecan-2-yl)hexacosanamide (10)**



EDCI (197 mg, 0.26 mmol), DMAP (2 mg, 0.013 mmol), and hexacosanoic acid (49 mg, 0.26 mmol) were added to a stirred solution of **9** (50 mg, 0.13 mmol) in CH₂Cl₂ (2 mL). The reaction mixture was stirred under a nitrogen atmosphere for 6 h. Solvent was removed *in vacuo* and the resulting residue was purified using silica gel column chromatography (hexane/EtOAc, 1:1) to afford the desired products **10** (77 mg, 78%) as a white solid; ¹H NMR (300 MHz, CDCl₃) δ 0.86 (t, *J* = 6.7 Hz, 6H), 1.11–1.32 (m, 68H), 1.41–1.52 (m, 2H), 1.58–1.81 (m, 4H), 1.88–1.93 (m, 2H), 3.50–3.58 (m, 3H), 3.89–3.98 (m, 1H), 4.32 (d, *J* = 11.4 Hz, 1H), 4.63 (d, *J* = 11.2 Hz, 1H), 6.33 (d, *J* = 5.3 Hz, 1-NH), 7.27–7.35 (m, 5H); ¹³C NMR (100 MHz, CDCl₃) δ 14.1, 22.7, 24.6, 25.7, 29.2, 29.29, 29.35, 29.5, 29.60, 29.64, 29.7, 29.8, 31.9, 33.2, 35.2, 36.6, 52.3, 66.9, 70.5, 77.9, 127.7, 127.8, 128.5, 138.0, 174.5; HRMS (FAB) calcd for C₅₁H₉₆O₃N 770.7390 ([M+H]⁺), found 770.7375.

***N*-((2*R*,4*R*)-4-(Benzyloxy)-1-((2*S*,3*R*,4*S*,5*S*,6*R*)-3,4,5-tris(benzyloxy)-6-(benzyloxymethyl)tetrahydro-2*H*-pyran-2-yloxy)octadecan-2-yl)hexacosanamide**

(11)



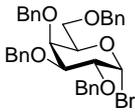
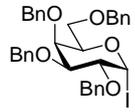
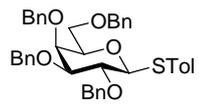
Freshly ground molecular sieves (4 Å, 50 mg), silver perchlorate (42 mg, 0.198 mmol), and tin(II) chloride (51 mg, 0.198 mmol) were added to a solution of compound **10** (51

mg, 0.066 mmol) in dry THF (2 mL) and stirred for 30 min. The mixture was cooled to $-10\text{ }^{\circ}\text{C}$, and a solution of β -tetrabenzylgalactosyl fluoride **12** (54 mg, 0.099 mmol) in dry THF (2 mL) was added dropwise. After 10 min, the mixture was warmed to room temperature and stirred for 2 h. The mixture was filtered through a pad of Celite and the filter cake was washed with EtOAc. The filtrate was washed with brine, dried over Na_2SO_4 , and concentrated under reduced pressure. The residue was purified using silica gel column chromatography (hexane/EtOAc, 6:1) to give **11** as a white solid (8 mg, 10%): ^1H NMR (500 MHz, CDCl_3) δ 0.86 (t, $J = 6.9$ Hz, 6H), 1.15–1.37 (m, 68H), 1.46–1.55 (m, 4H), 1.72–1.81 (m, 2H), 1.88–1.98 (m, 2H), 3.37 (dd, $J = 5.7, 9.4$ Hz, 1H), 3.40–3.46 (m, 1H), 3.51 (dd, $J = 6.8, 9.3$ Hz, 1H), 3.56 (dd, $J = 3.6, 11.0$ Hz, 1H), 3.73 (dd, $J = 3.6, 11.1$ Hz, 1H), 3.84–3.89 (m, 2H), 3.92 (t, $J = 6.3$ Hz, 1H), 4.01 (dd, $J = 3.7, 9.9$ Hz, 1H), 4.10–4.13 (m, 1H), 4.36 (dd, $J = 7.5, 11.6$ Hz, 2H), 4.46 (dd, $J = 8.8, 11.6$ Hz, 2H), 4.55 (d, $J = 11.6$ Hz, 1H), 4.62 (d, $J = 11.8$ Hz, 1H), 4.70 (d, $J = 11.7$ Hz, 1H), 4.75–4.77 (m, 2H), 4.80 (d, $J = 11.7$ Hz, 1H), 4.90 (d, $J = 11.6$ Hz, 1H), 6.17 (d, $J = 8.1$ Hz, 1-NH), 7.21–7.36 (m, 25H); ^{13}C NMR (100 MHz, CDCl_3) δ 14.2, 19.7, 22.7, 24.9, 25.7, 27.1, 29.4, 29.6, 29.7, 29.9, 30.0 (2C), 30.1, 31.9, 32.8, 33.5, 36.7, 37.1, 53.5, 59.0, 60.1, 70.4, 73.2 (2C), 73.5, 74.5, 74.9, 75.1, 79.0, 95.8, 98.1, 120.0, 127.5, 127.6, 127.7 (2C), 127.9 (2C), 128.3 (2C), 128.4 (2C), 138.2, 138.4, 138.6, 138.7, 138.8, 138.9, 173.9; HRMS (FAB) calcd for $\text{C}_{85}\text{H}_{130}\text{O}_8\text{N}$ 1292.9796 ($[\text{M}+\text{H}]^+$), found 1292.9763.

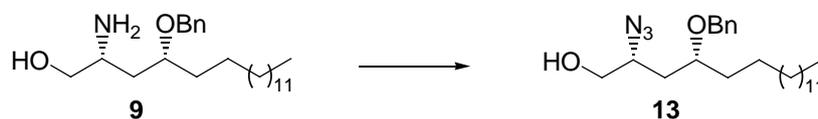
Attempts to increase the yield of **11** by varying the glycosyl donor and reaction conditions were not successful (Table S1, not optimized).

Table S1.

entry	donor	conditions	yield (%)	ref.
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1		Bu ₄ NBr, TMU, CH ₂ Cl ₂ , rt	< 10	5
2		<i>n</i> -Hex ₄ NBr, 4Å MS, CH ₂ Cl ₂ , rt	< 10	
3		Bu ₄ NI, DIPEA, CH ₂ Cl ₂ , rt	0	6
4		1-Benzenesulfinylpiperidine, Tf ₂ O, CH ₂ Cl ₂ , -78 °C to 0 °C	< 5	7

(2*R*,4*R*)-2-Azido-4-(benzyloxy)octadecan-1-ol (13)

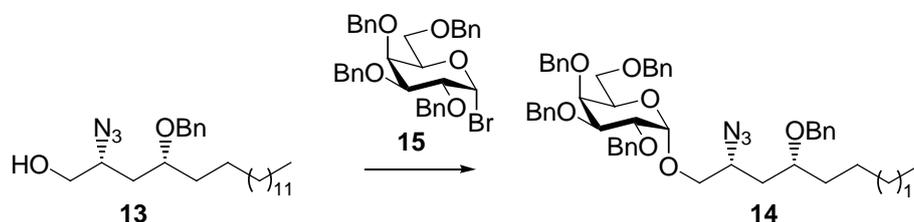


Synthesis of TfN₃ solution: A solution of NaN₃ (167 mg, 2.58 mmol) in H₂O (2 mL) was cooled in an ice bath and treated with 4 mL of CH₂Cl₂. The resulting biphasic mixture was stirred vigorously and treated with Tf₂O (0.09 mL, 0.52 mmol). After stirring for 2 h at 0 °C, the organic phase was separated and the aqueous phase was extracted twice with CH₂Cl₂. The combined organic layers were washed with saturated Na₂CO₃ solution and used without further purification.

Compound **9** (84 mg, 0.21 mmol) was dissolved in H₂O (2 mL) and treated with K₂CO₃ (44 mg, 0.32 mmol) and CuSO₄ hydrate (4 mg, 0.02 mmol). MeOH (4 mL) and the TfN₃ solution were added to the reaction mixture, followed by addition of more MeOH until homogeneity was observed. The reaction mixture was stirred for 12 h and the solvent was removed under reduced pressure. The crude product was purified using silica gel column chromatography (hexane/EtOAc, 6:1) to give **13** (71 mg, 79%) as a white solid: ¹H NMR (300 MHz, CDCl₃) δ 0.86 (t, *J* = 6.7 Hz, 3H), 1.59–1.30 (m, 24H), 1.48–1.65 (m, 2H), 1.70–1.89 (m, 2H), 2.15–2.19 (m, 1H), 3.50–3.68 (m, 4H), 4.43 (d,

$J = 11.5$ Hz, 1H), 4.55 (d, $J = 11.4$ Hz, 1H), 7.25–7.36 (m, 5H); ^{13}C NMR (100 MHz, CDCl_3) δ 14.1, 22.7, 25.0, 29.3, 29.55, 29.63, 29.66, 29.70, 31.9, 33.3, 34.6, 60.8, 64.6, 70.8, 75.7, 127.8, 127.9, 128.4, 138.1; HRMS (FAB) calcd for $\text{C}_{25}\text{H}_{44}\text{O}_2\text{N}_3$ 418.3434 ($[\text{M}+\text{H}]^+$), found 418.3427.

(2*S*,3*R*,4*S*,5*S*,6*R*)-2-((2*R*,4*R*)-2-Azido-4-(benzyloxy)octadecyloxy)-3,4,5-tris(benzyloxy)-6-(benzyloxymethyl)tetrahydro-2*H*-pyran (14**)⁸**

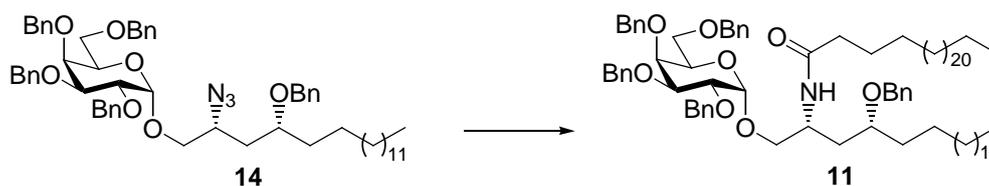


Triphenylphosphine (226 mg, 0.86 mmol) and carbon tetrabromide (285 mg, 0.86 mmol) were added to a solution of tetrabenzyl galactose (155 mg, 0.29 mmol) in dry CH_2Cl_2 (4 mL) and the resulting mixture was stirred at room temperature for 3 h. (*in situ* generation of compound **15**). *N,N*-tetramethylurea (0.06 mL, 0.48 mmol), Bu_4NBr (93 mg, 0.29 mmol) and acceptor **13** (40 mg, 0.10 mmol) dissolved in dry CH_2Cl_2 (3 ml) were then added by syringe. The reaction was stirred at room temperature for 8 days until complete consumption of the starting material observed by TLC. The reaction mixture was filtered through a pad of Celite and the filter cake was washed with CH_2Cl_2 . The organic mixture was washed with saturated aqueous NaHCO_3 solution and brine, dried over Na_2SO_4 , and concentrated under reduced pressure. The residue was purified using silica gel column chromatography (hexane/EtOAc, 8:1) to give **14** (75 mg, 83%) as a yellow liquid: ^1H NMR (300 MHz, CDCl_3) δ 0.85 (t, $J = 6.5$ Hz, 3H), 1.17–1.32 (m, 24H), 1.46–1.54 (m, 2H), 1.64–1.85 (m, 2H), 3.44–3.47 (m, 4H), 3.55–3.63 (m, 1H), 3.74 (dd, $J = 3.1, 10.4$ Hz, 1H), 3.90–3.93 (m, 3H), 4.01 (dd, $J = 3.3, 9.5$ Hz, 1H), 4.33–

4.47 (m, 4H), 4.53 (d, $J = 15.4$ Hz, 1H), 4.61–4.66 (m, 1H), 4.72 (d, $J = 12.5$ Hz, 1H), 4.78–4.81 (m, 3H), 4.90 (d, $J = 11.4$ Hz, 1H), 7.22–7.32 (m, 25H); ^{13}C NMR (100 MHz, CDCl_3) δ 14.1, 21.1, 22.7, 24.9, 29.4, 29.7, 29.8, 31.9, 35.6, 34.7, 58.6, 60.4, 69.0 (4C), 69.4, 70.7, 73.0, 73.2 (2C), 73.4, 74.7, 75.0, 75.1, 79.2, 96.0, 98.4, 127.4, 127.5, 127.6, 127.7 (2C), 127.8 (2C), 128.2 (2C), 128.3, 128.4, 137.4, 138.0, 138.5, 138.6, 138.7, 138.8, 138.9; HRMS (FAB) calcd for $\text{C}_{59}\text{H}_{78}\text{O}_7\text{N}_3$ 940.5840 ($[\text{M}+\text{H}]^+$), found 940.5822.

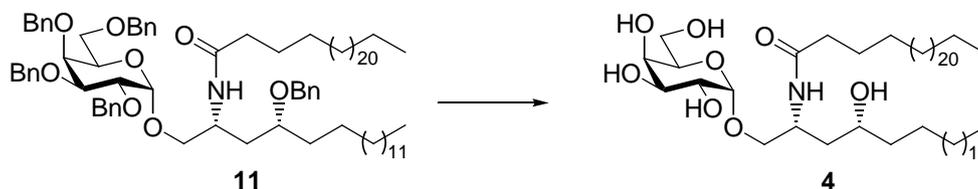
***N*-((2*R*,4*R*)-4-(Benzyloxy)-1-((2*S*,3*R*,4*S*,5*S*,6*R*)-3,4,5-tris(benzyloxy)-6-(benzyloxymethyl)tetrahydro-2*H*-pyran-2-yloxy)octadecan-2-yl)hexacosanamide**

(11)



Triphenylphosphine (24 mg, 0.12 mmol) was added to a solution of **14** (56 mg, 0.06 mmol) in a mixture of benzene (3 mL) and water (30 μL). The mixture was stirred overnight at 60 $^\circ\text{C}$ and concentrated under reduced pressure. The residue was dissolved in CH_2Cl_2 (3 mL) followed by addition of EDCI (27 mg, 0.12 mmol), DMAP (1 mg, 10 mol%) and hexacosanoic acid (44 mg, 0.12 mmol). The mixture was stirred overnight at room temperature and then concentrated under reduced pressure. The residue was purified using silica gel column chromatography (hexane/EtOAc, 3:1) to give **11** (42 mg, 54%) as a white solid: All of the spectral data for compound **11** were in agreement for the products of the two synthetic routes (synthesized via intermediates **10** and **12**, respectively).

***N*-((2*R*,4*R*)-4-Hydroxy-1-((2*S*,3*R*,4*S*,5*R*,6*R*)-3,4,5-trihydroxy-6-(hydroxymethyl)tetrahydro-2*H*-pyran-2-yloxy)octadecan-2-yl)hexacosanamide (**4**)**



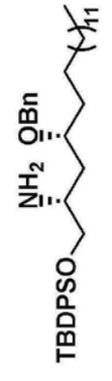
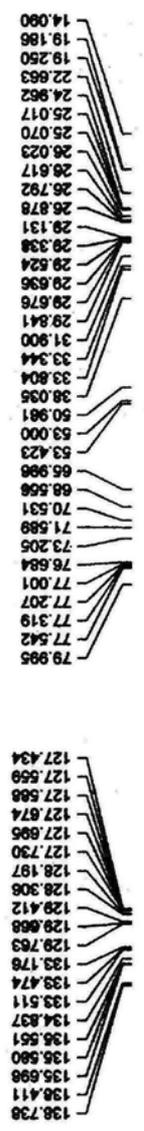
The compound **11** (15 mg, 0.01 mmol) and Pd(OH)₂ (7 mg, 50 wt%) were suspended in a solution of EtOH/CH₂Cl₂ (3:1, 2 mL). The reaction mixture was hydrogenated (1 atm) at room temperature. After 2 days, the palladium catalyst was removed by filtration through a syringe filter (PTFE, 0.50 μm, hydrophobic) and rinsed with EtOH/CH₂Cl₂ (9:1). The filtrate was concentrated *in vacuo*. The residual solid was triturated with hexane/EtOAc (1:1) to give **4** (7 mg, 72%) as a white solid: $[\alpha]_D^{25} +43.6$ (*c* 1.0, pyridine); ¹H NMR (300 MHz, C₅D₅N) δ 0.85 (t, *J* = 6.0 Hz, 6H), 1.16–1.48 (m, 68H), 1.57–1.88 (m, 4H), 2.12–2.28 (m, 2H), 2.45 (t, *J* = 7.3 Hz, 2H), 4.34–4.42 (ddd, *J* = 5.0, 10.4, 44.7 Hz, 2H), 4.32–4.62 (m, 6H), 4.62–4.73 (m, 1H), 4.78–4.86 (m, 1H), 5.35–5.45 (m, 1H), 5.42 (d, *J* = 3.8 Hz, 1H), 8.65 (d, *J* = 8.8 Hz, 1-NH); ¹³C NMR (100 MHz, C₅D₅N) δ 14.8, 21.7, 23.4, 26.2, 26.9, 30.1, 30.2, 30.3, 30.5, 32.6, 34.7, 37.2, 37.3, 47.4, 63.3, 71.0, 71.5, 72.1, 72.3, 72.8, 73.6, 102.4, 171.2, 173.6; HRMS (FAB) calcd for C₅₀H₁₀₀O₈N 842.7449 ([M+H]⁺), found 842.7462.

C. References of Supporting Information

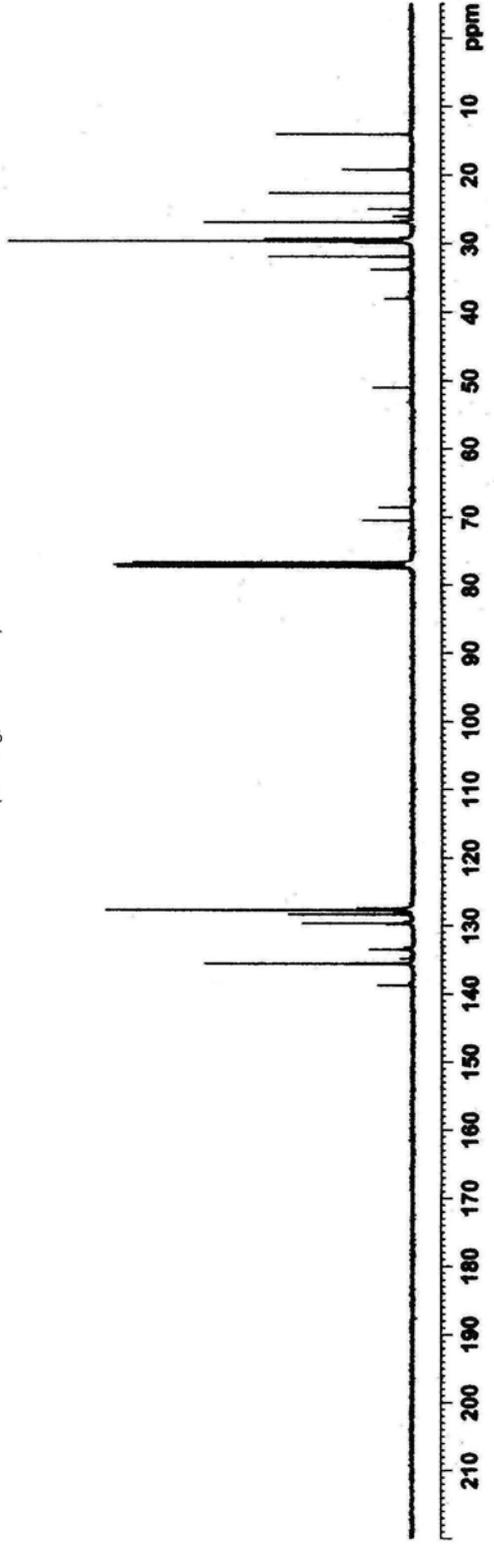
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Tsuji, M.; Wong, C.-H. Quantitative microarray analysis of intact glycolipid-CD1d interaction and correlation with cell-based cytokine production. *J. Am. Chem. Soc.* **2008**, *130*, 12348–12354.

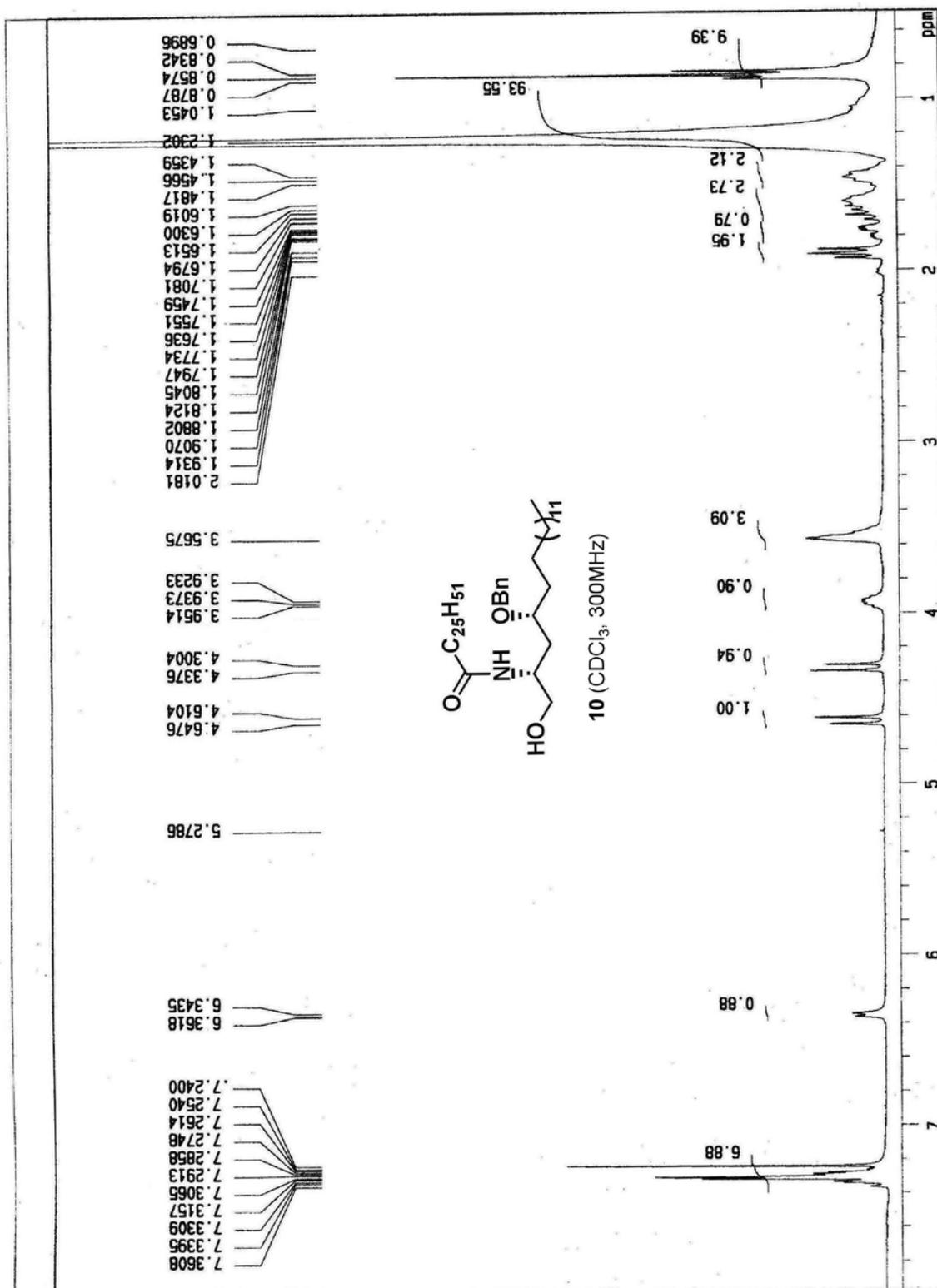
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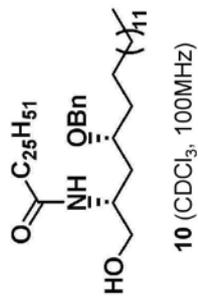


8 (CDCl₃, 100MHz)

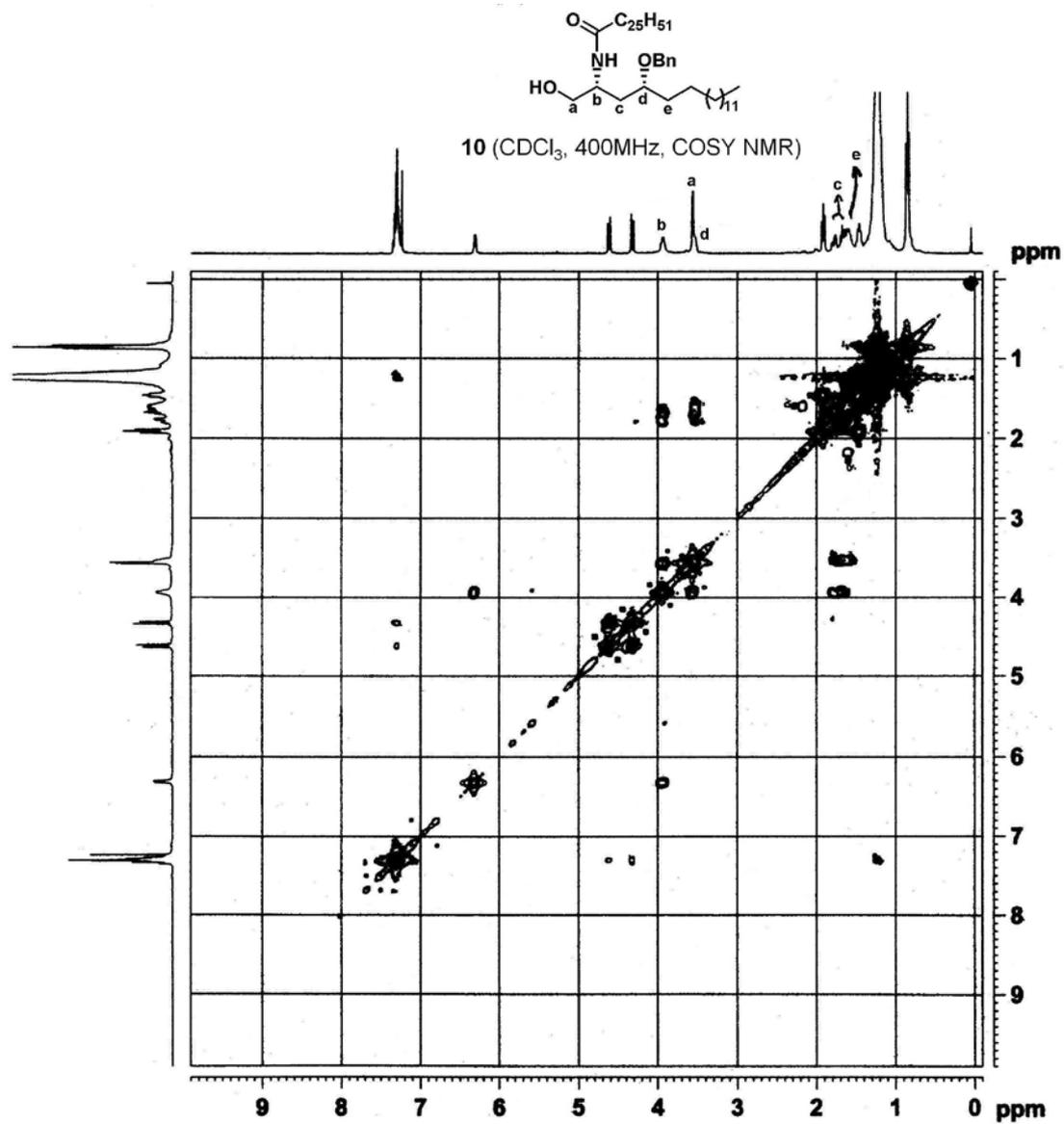


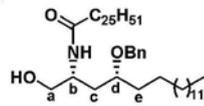
(b) ^1H NMR and ^{13}C NMR spectra of compound 10



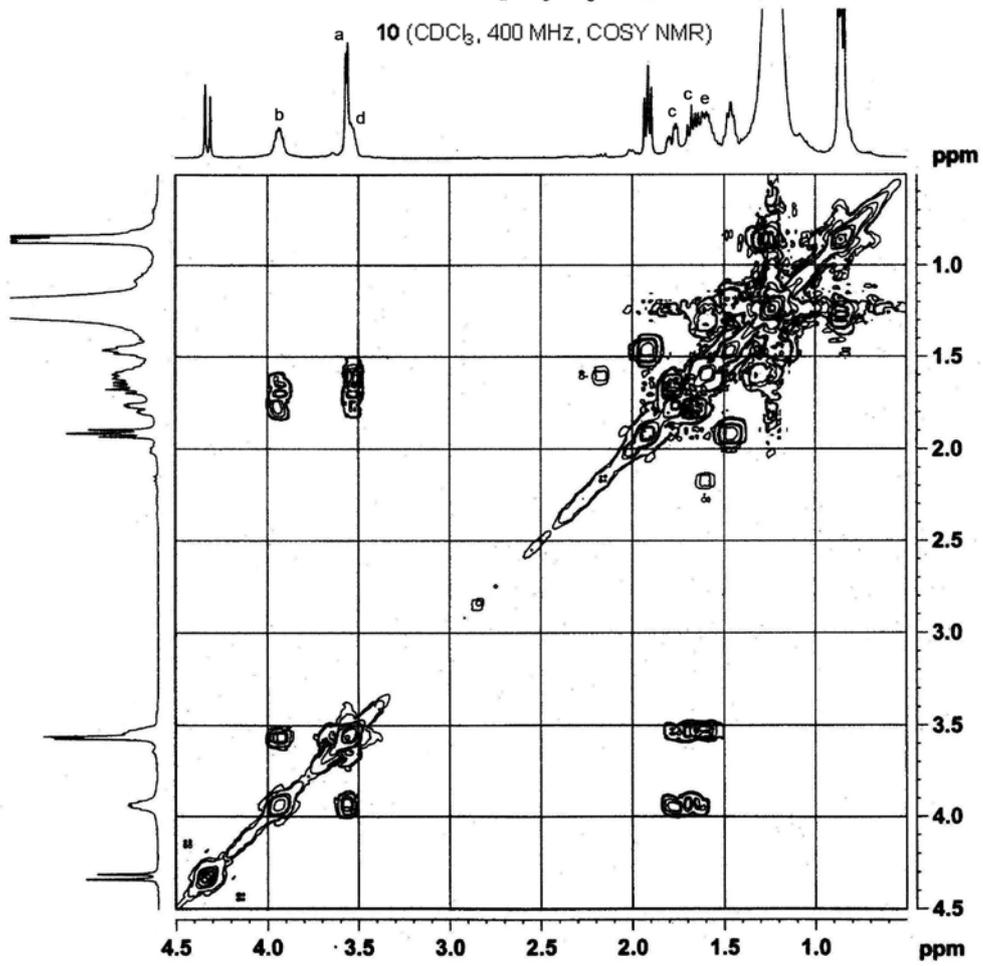


(c) COSY NMR spectra of compound 10

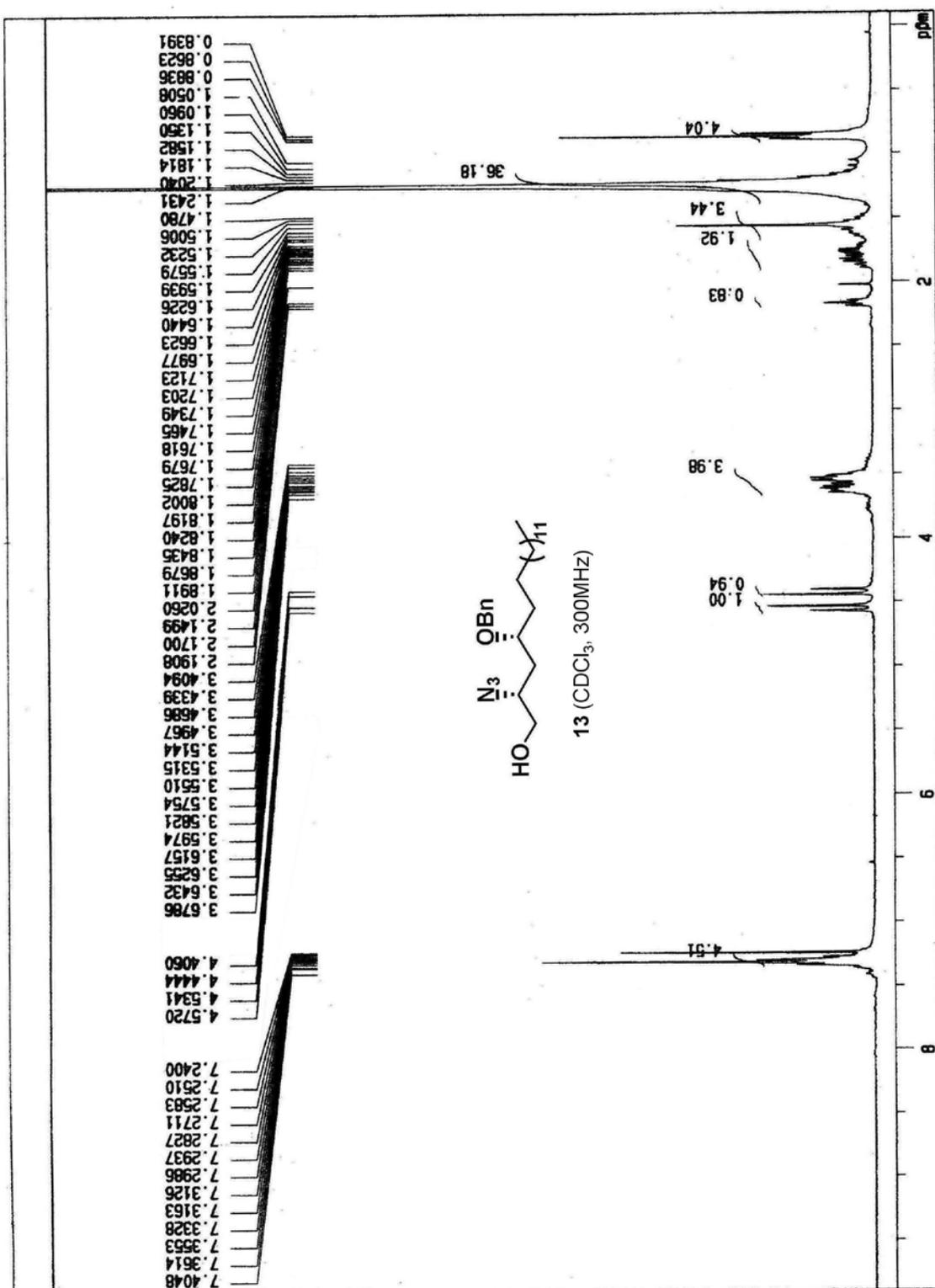




10 (CDCl₃, 400 MHz, COSY NMR)



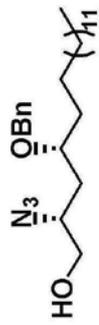
(d) ^1H NMR and ^{13}C NMR spectra of compound 13



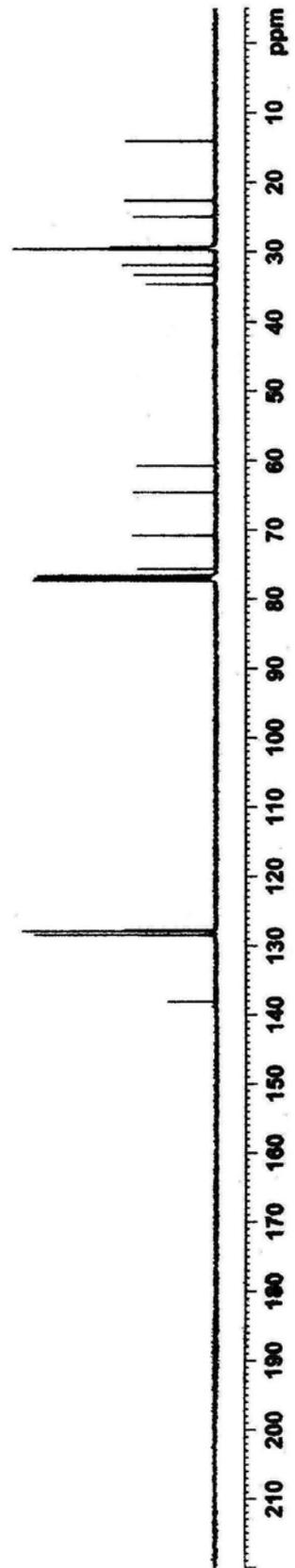
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70.813
75.680
76.685
77.002
77.320

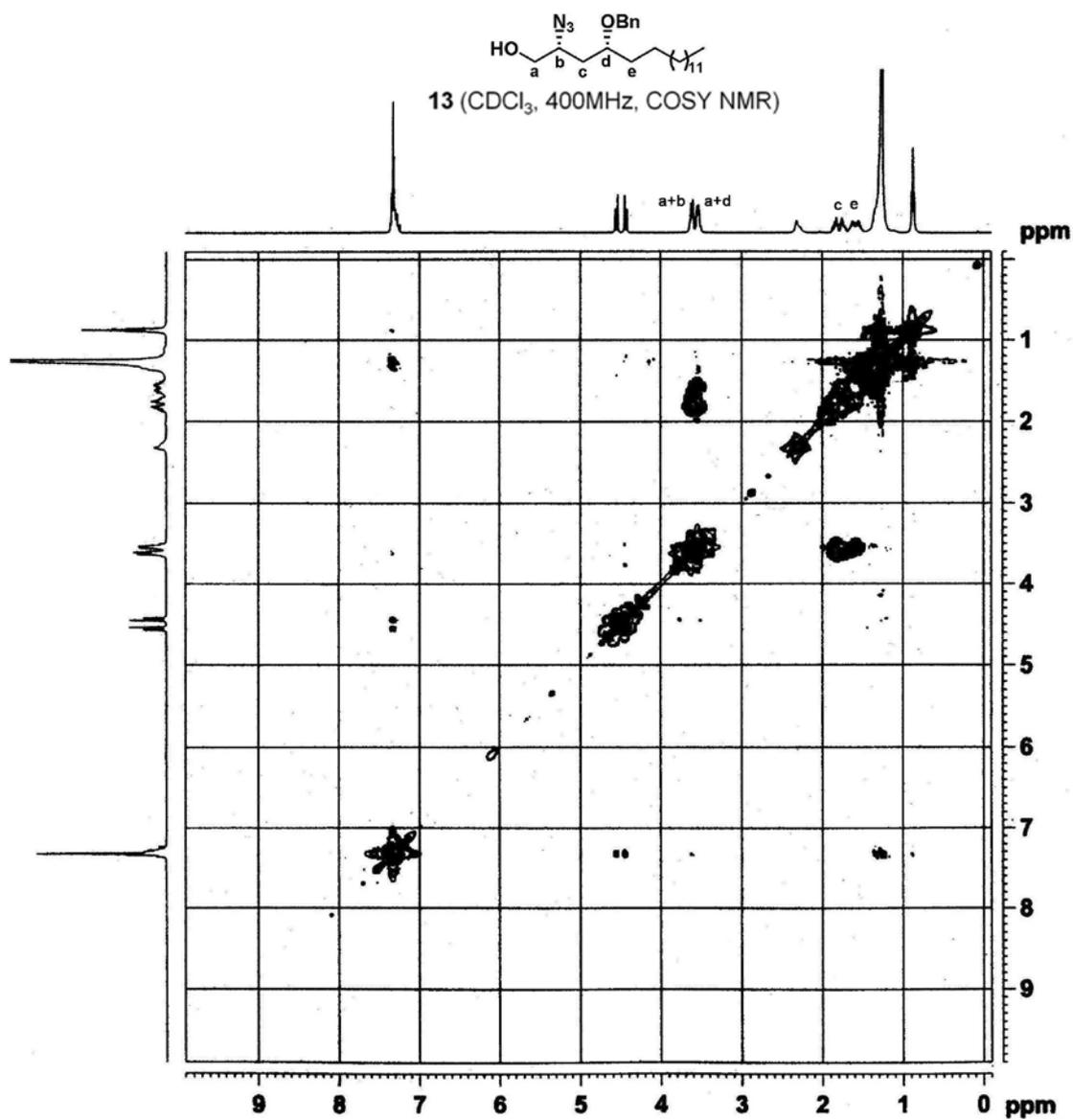
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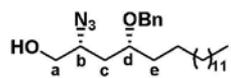


13 (CDCl₃, 100MHz)

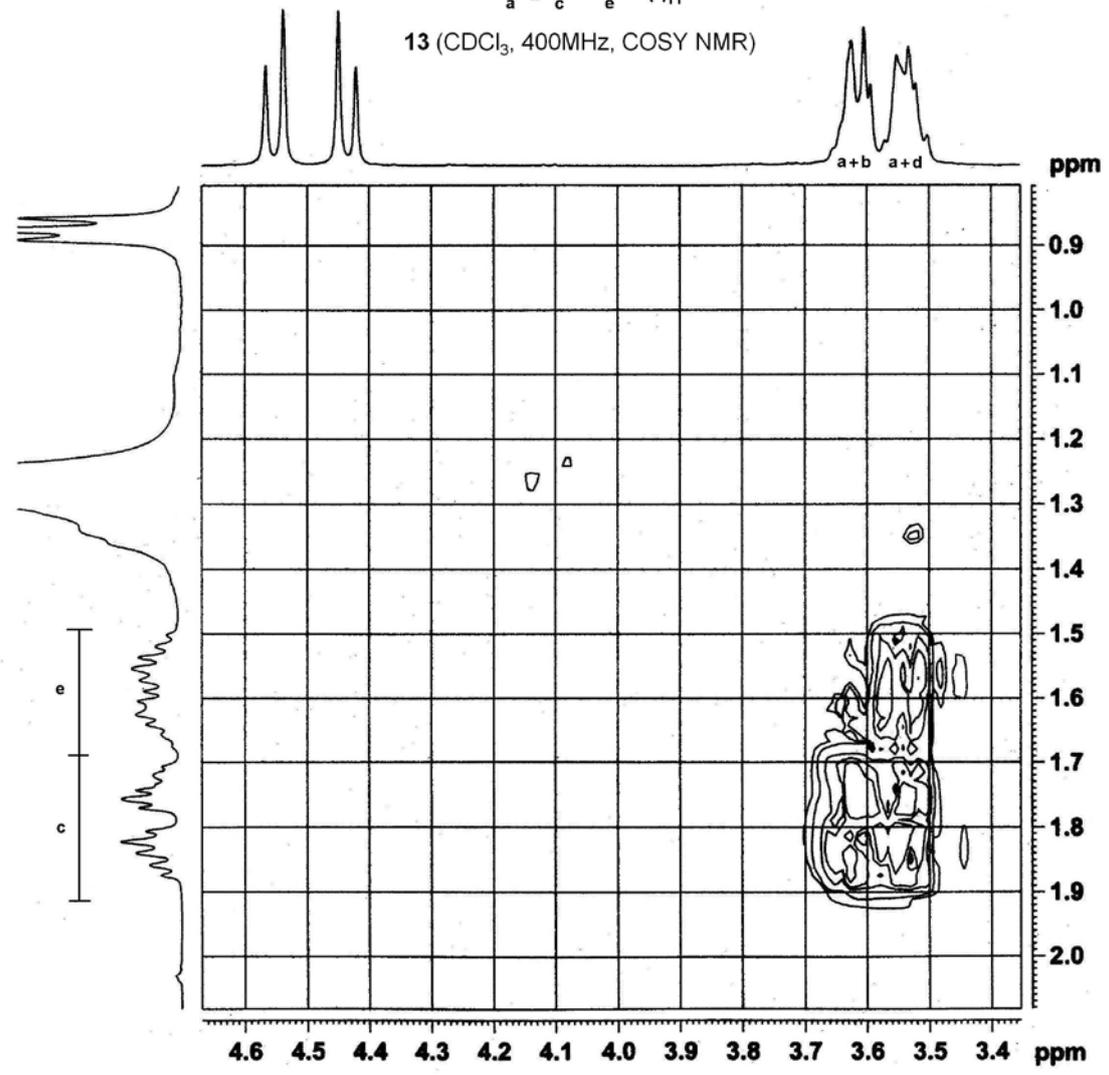


(e) COSY NMR spectra of compound 13





13 (CDCl₃, 400MHz, COSY NMR)





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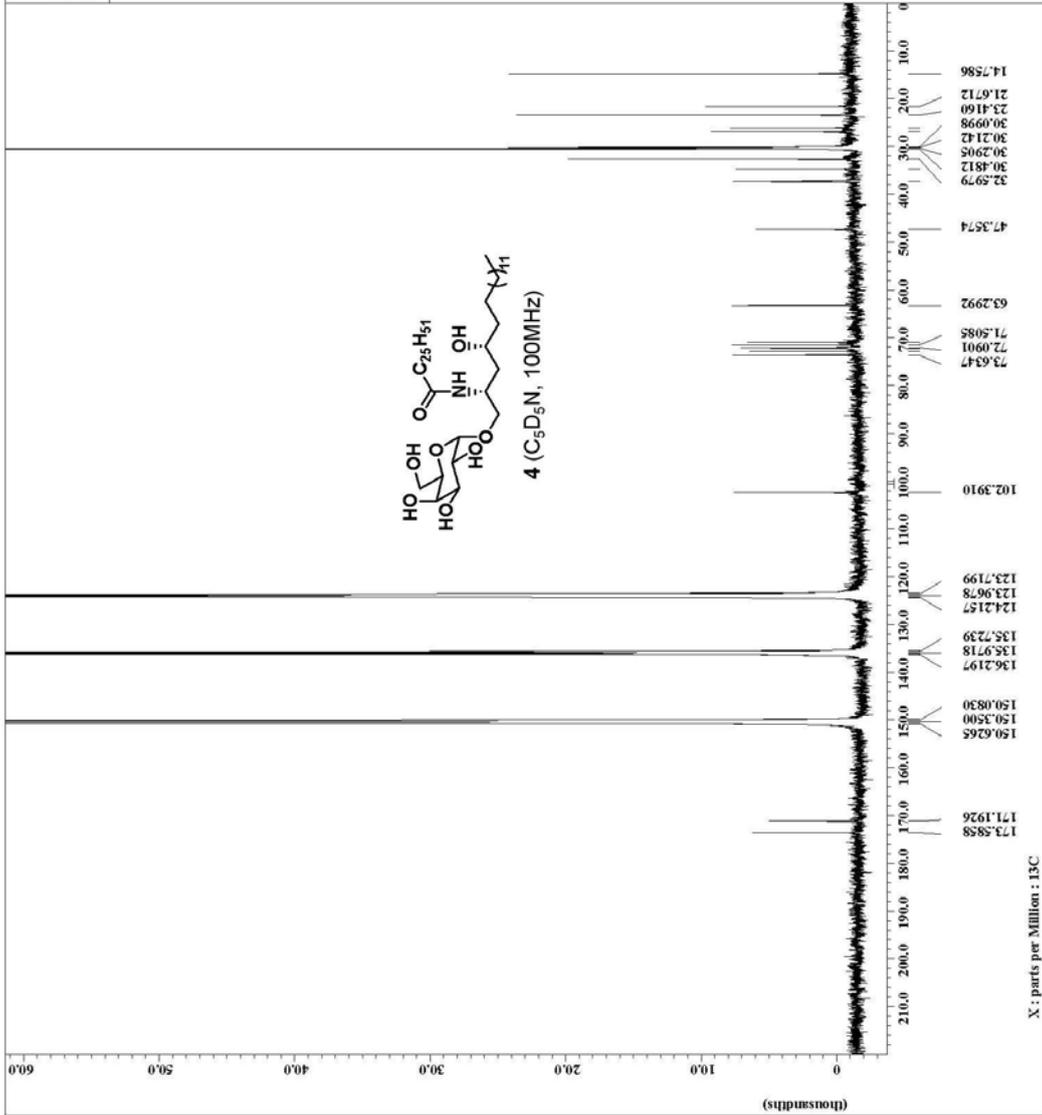
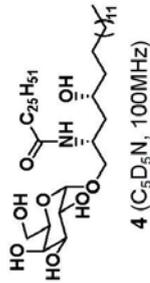
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Dim Unit    = 13C
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X: parts per Million : 13C

(g) ^1H NMR and ^{13}C NMR spectra of compound 2

