

SUPPORTING INFORMATION FOR:

A Chemoenzymatic Total Synthesis of (+)-Clividine

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X-ray Crystallographic Studies

X-ray Crystallographic Data

Oxalate salt of C11b-epi-ent-1. $C_{17}H_{20}NO_5^+ \cdot C_2HO_4^-$, $M = 407.38$, $T = 200$ K, monoclinic, space group $P2_1$, $Z = 2$, $a = 7.8532(4)$, $b = 7.8328(4)$, $c = 15.1100(8)$ Å, $\beta = 93.534(3)^\circ$, $V = 927.69(8)$ Å³, $D_x = 1.458$ g cm⁻³, 1761 unique data ($\theta = 2.7$ – 25°), $R = 0.035$ [for 1629 with $I > 2.0\sigma(I)$]; $R_w = 0.086$ (all data), $S = 1.03$.

Picrate salt of ent-1. $2(C_{17}H_{20}NO_5^+) \cdot 2(C_6H_2N_3O_7^-) \cdot 7.87H_2O \cdot 0.315CH_3CN$, $M = 1247.6$, $T = 200$ K, monoclinic, space group $P2_1$, $Z = 2$, $a = 8.6590(2)$, $b = 35.4747(9)$, $c = 9.0925(2)$ Å, $\beta = 101.1071(12)^\circ$, $V = 2740.68(11)$ Å³, $D_x = 1.512$ g cm⁻³, 4933 unique data ($\theta = 2.6$ – 25.1°), $R = 0.050$ [for 3901 with $I > 2.0\sigma(I)$]; $R_w = 0.110$ (all data), $S = 1.02$.

Compound 6. $C_{11}H_{14}BrNO_3$, $M = 288.14$, $T = 200$ K, monoclinic, space group $P2_1$, $Z = 2$, $a = 10.0349(3)$, $b = 5.8790(1)$, $c = 11.4499(3)$ Å, $\beta = 113.6267(15)^\circ$, $V = 618.87(3)$ Å³, $D_x = 1.546$ g cm⁻³, 2718 unique data ($\theta = 2.6$ – 27.5°), $R = 0.025$ [for 2619 with $I > 2.0\sigma(I)$]; $R_w = 0.065$ (all data), $S = 1.01$, Flack parameter = $-0.015(7)$.

Compound 10. $C_{20}H_{21}NO_6$, $M = 371.39$, $T = 200$ K, orthorhombic, space group $P2_12_12_1$, $Z = 4$, $a = 9.7236(2)$, $b = 13.2103(2)$, $c = 14.0965(2)$ Å, $V = 1810.72(6)$ Å³, $D_x = 1.362$ g cm⁻³, 2977 unique data ($\theta = 2.6$ – 30°), $R = 0.034$ [for 2703 with $I > 2.0\sigma(I)$]; $R_w = 0.087$, $S = 1.00$.

Compound 15. $C_{17}H_{19}NO_5$, $M = 317.34$, $T = 200$ K, monoclinic, space group $P2_1$, $Z = 2$, $a = 8.6115(8)$, $b = 7.9108(8)$, $c = 11.2630(12)$ Å, $\beta = 105.651(6)^\circ$, $V = 738.83(13)$ Å³, $D_x = 1.426$ g cm⁻³, 1411 unique data ($\theta = 2.7$ – 25.1°), $R = 0.081$ [for 1103 with $I > 2.0\sigma(I)$]; $R_w = 0.236$ (all data), $S = 1.02$.

Structure Determination

Images were measured on a Nonius Kappa CCD diffractometer (MoK α , graphite monochromator, $\lambda = 0.71073$ Å) and data extracted using the DENZO package.¹ Structure solution was by direct methods (SIR92).² The structures of the abovementioned compounds were refined using the CRYSTALS program package.³ Atomic coordinates, bond lengths and angles, and displacement parameters have been deposited at the Cambridge Crystallographic Data Centre (CCDC nos. 813598–813602 for the oxalate salt of compound C11b-epi-ent-1, the picrate salt of compound ent-1 and compounds 6, 10 and 15, respectively). These data can be obtained free-of-charge via www.ccdc.cam.ac.uk/data_request/cif, by emailing data_request@ccdc.cam.ac.uk, or by contacting The Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033.

Anisotropic Displacement Ellipsoid Plots

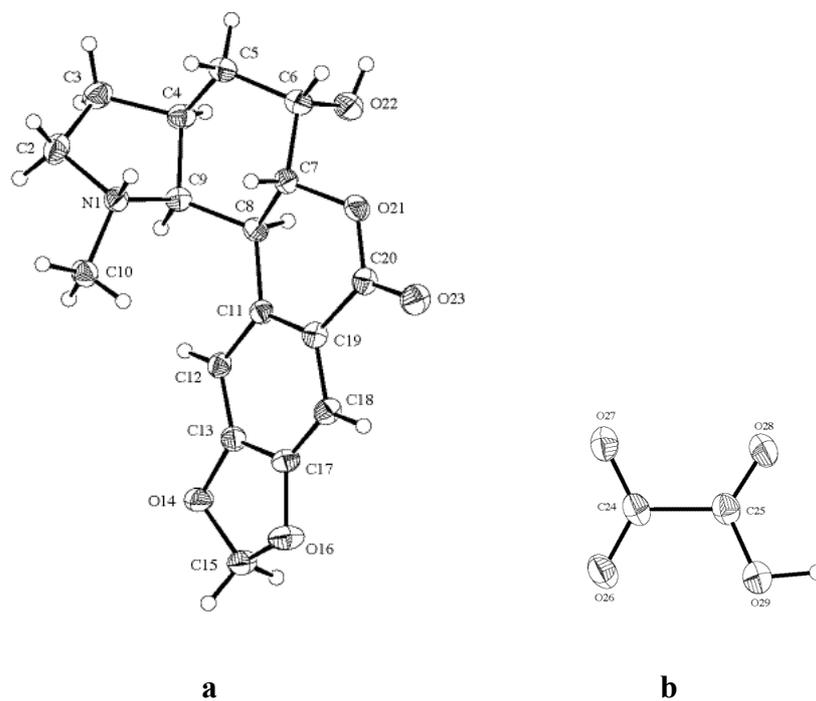


Figure S1. Structure of the oxalate salt of C11b-*epi-ent-1* (CCDC 813598, **a** = C₁₇H₂₀NO₅⁺ cation, **b** = C₂HO₄⁻ anion) with labelling of selected atoms. Anisotropic displacement ellipsoids display 30% probability levels. Hydrogen atoms are drawn as circles with small radii.

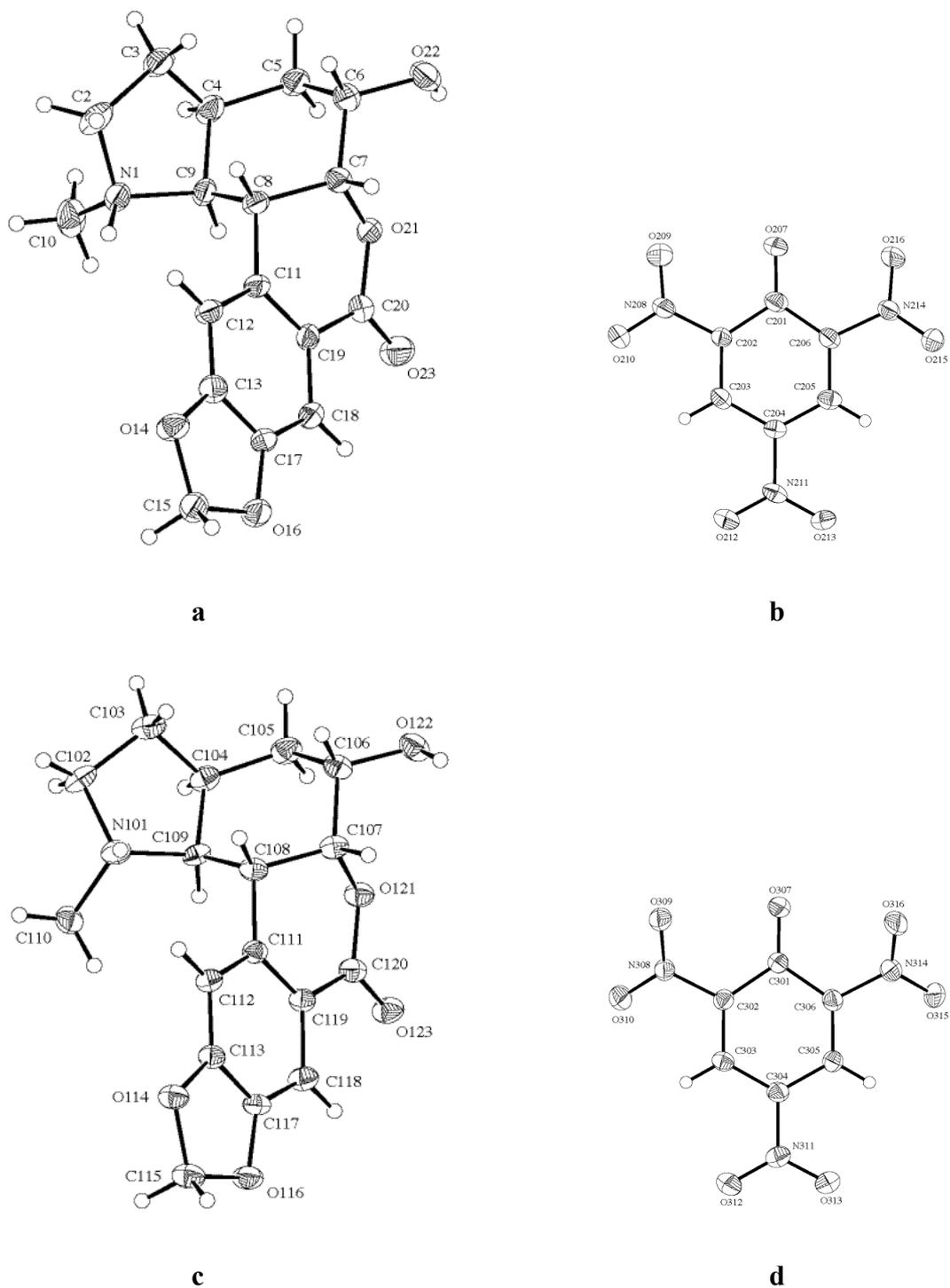


Figure S2. Structure of the picrate salt of *ent-1* (CCDC 813599, **a** = $C_{17}H_{20}NO_5^+$ cation one, **b** = $C_6H_2N_3O_7^-$ anion one, **c** = $C_{17}H_{20}NO_5^+$ cation two, **d** = $C_6H_2N_3O_7^-$ anion two) with labelling of selected atoms. Anisotropic displacement ellipsoids display 30% probability levels. Hydrogen atoms are drawn as circles with small radii.

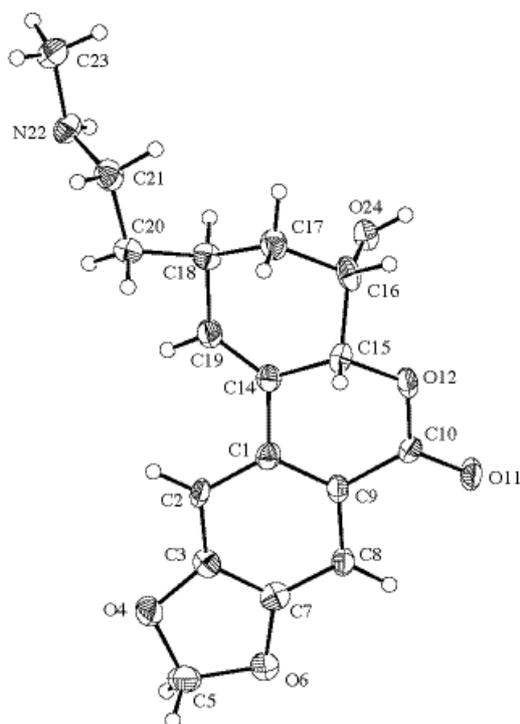
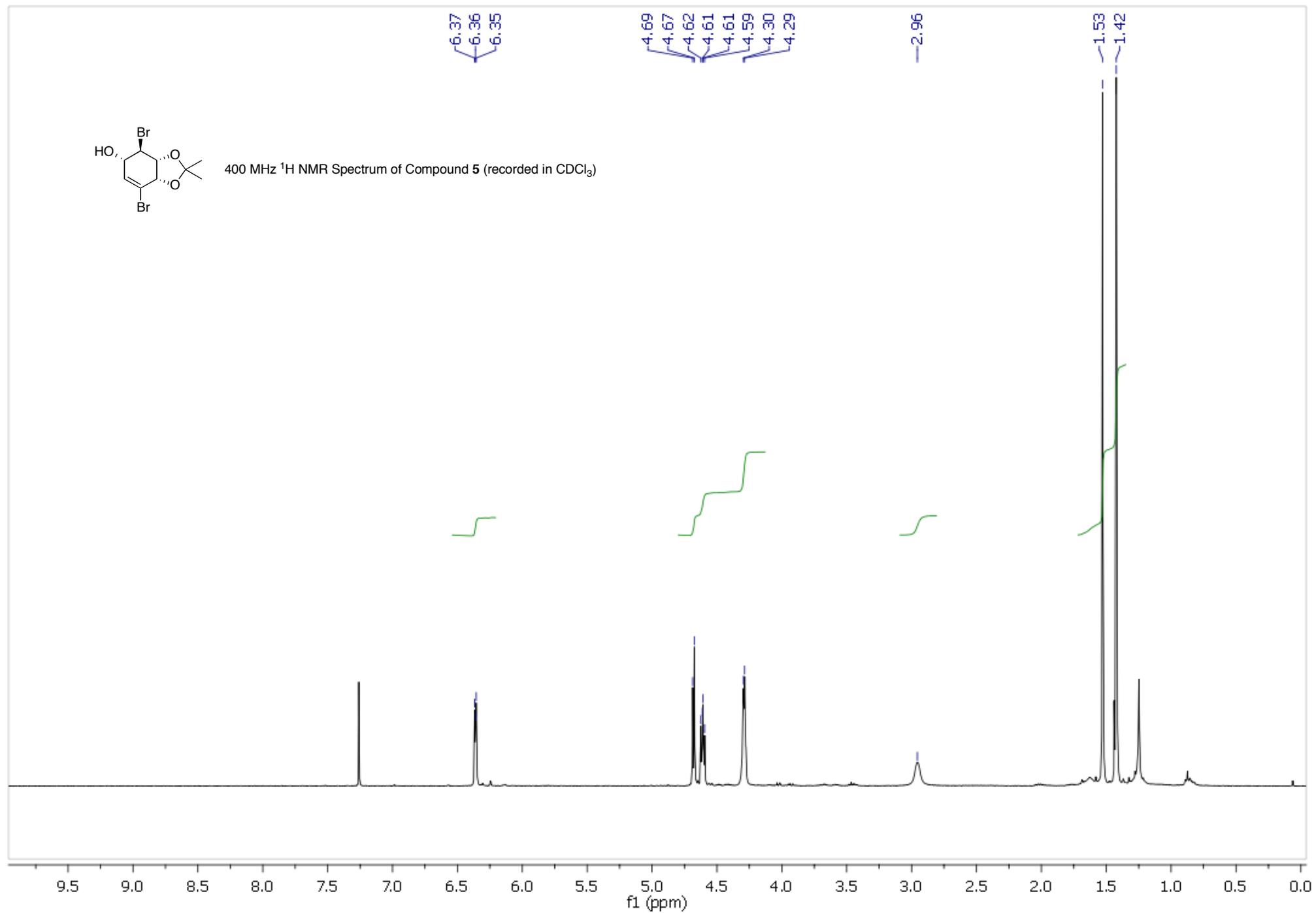
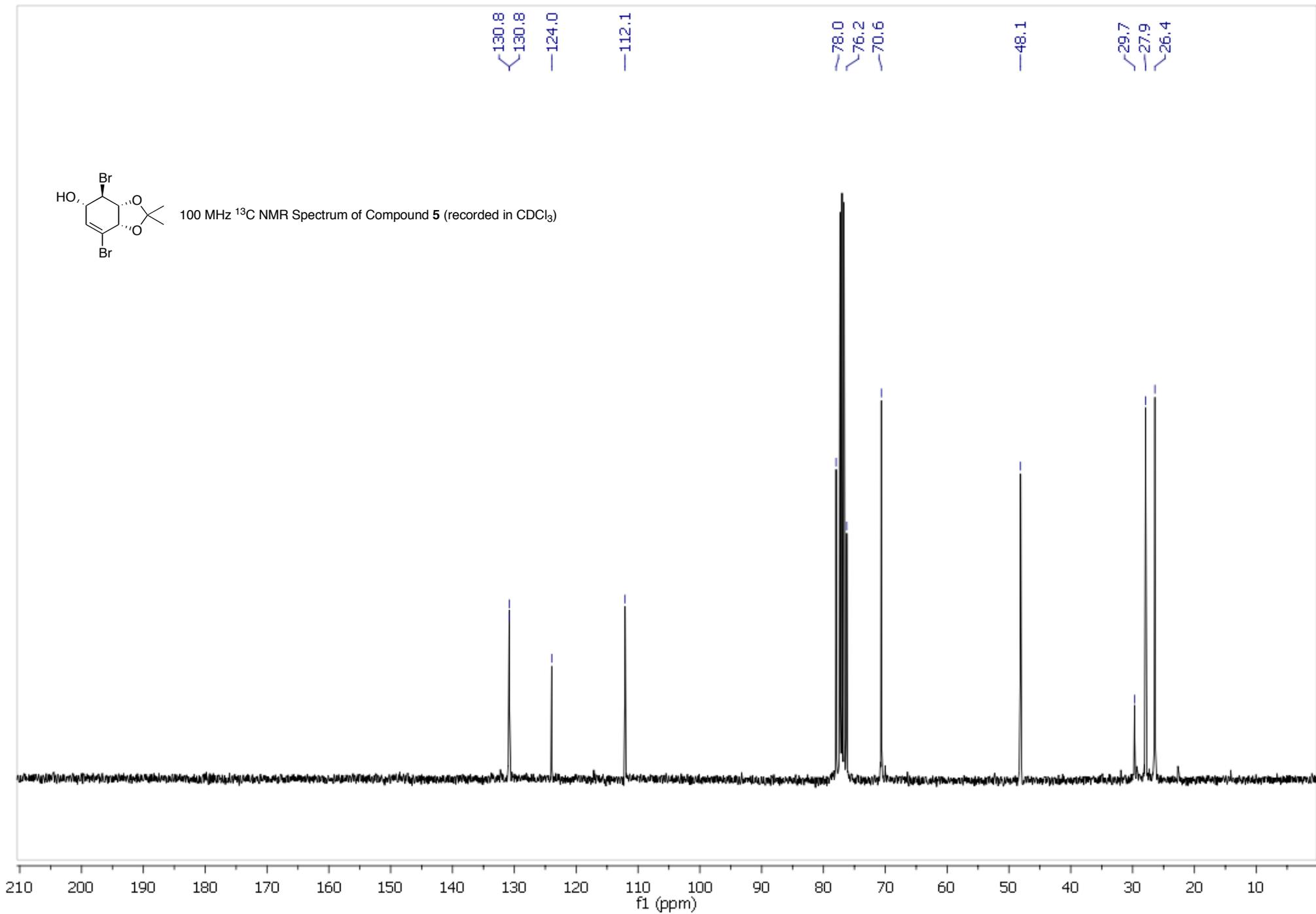


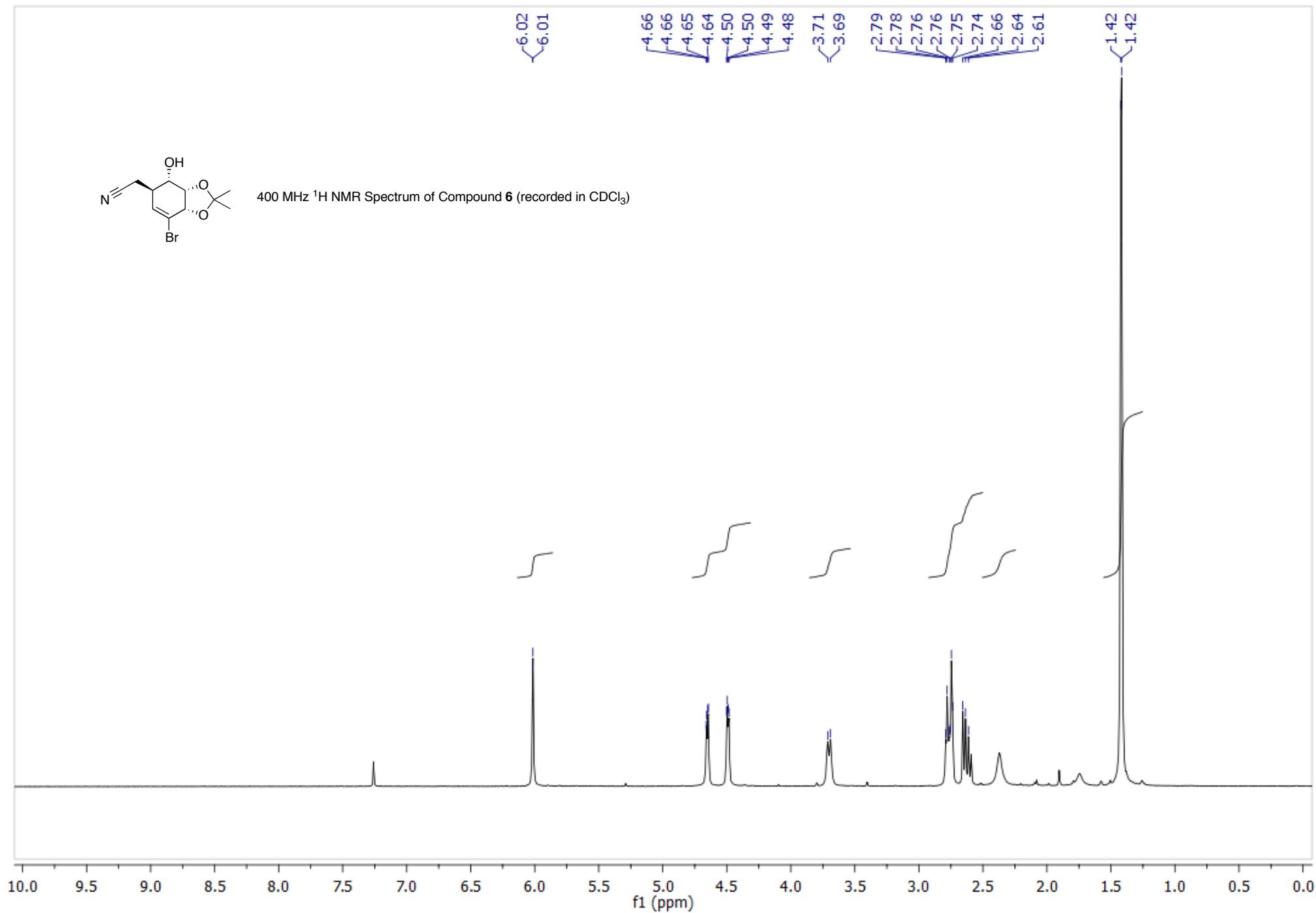
Figure S5. Structure of compound **15** (CCDC 813602, C₁₇H₁₉NO₅) with labelling of selected atoms. Anisotropic displacement ellipsoids display 30% probability levels. Hydrogen atoms are drawn as circles with small radii.

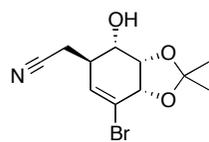
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- (1) DENZO–SMN. Otwinowski, Z.; Minor, W. Processing of X-ray diffraction data collected in oscillation mode. In *Methods in Enzymology, Volume 276: Macromolecular Crystallography, Part A*; Carter Jr., C. W.; Sweet, R. M., Eds.; Academic Press: New York, 1997; pp. 307–326.
- (2) SIR92. Altomare, A.; Cascarano, G.; Giacovazzo, C.; Guagliardi, A.; Burla, M. C.; Polidori, G.; Camalli, M. *J. Appl. Crystallogr.*, **1994**, *27*, 435.
- (3) Betteridge, P. W.; Carruthers, J. R.; Cooper, R. I.; Prout, K.; Watkin, D. J. *J. Appl. Crystallogr.*, **2003**, *36*, 1487.

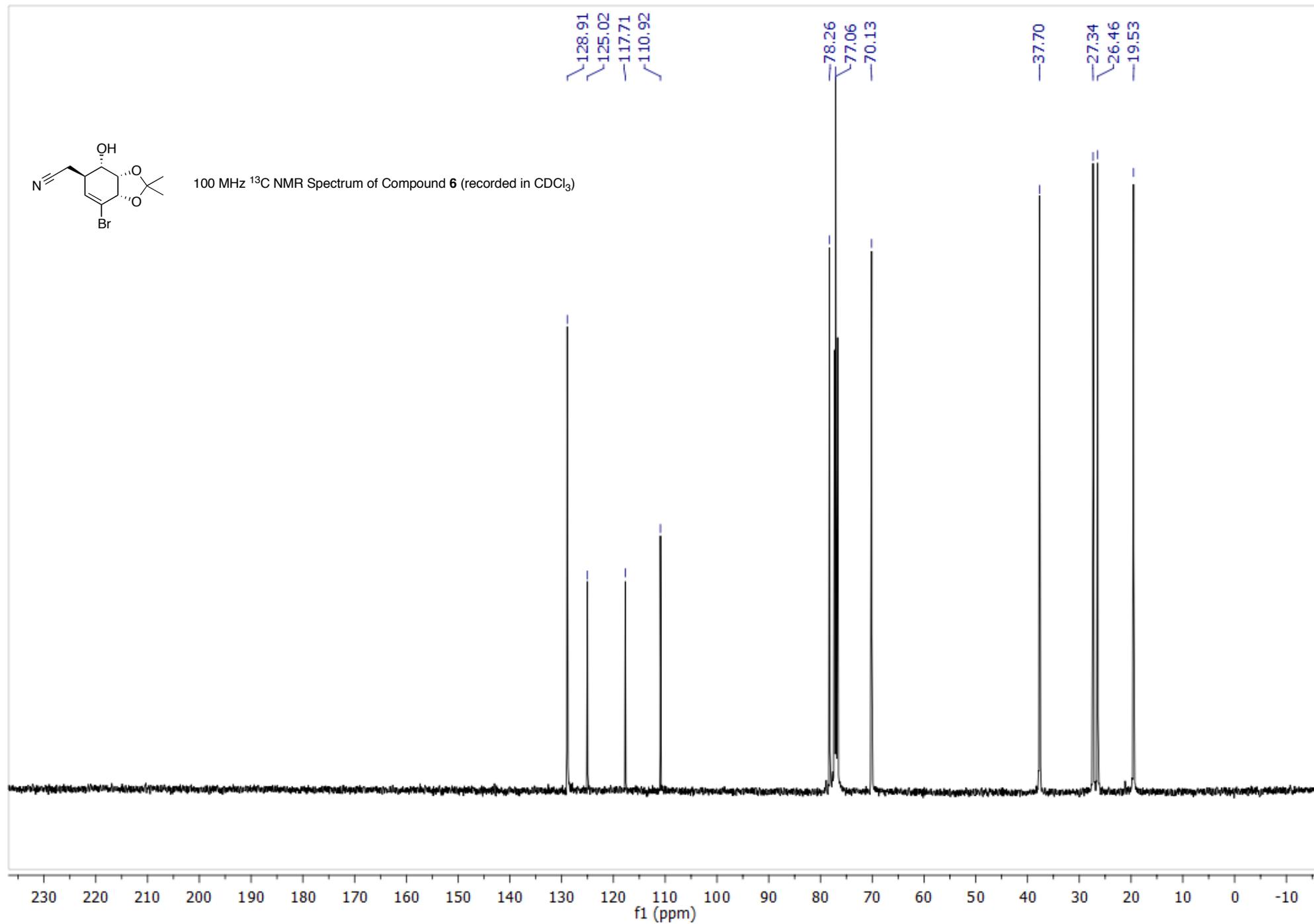


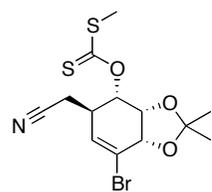




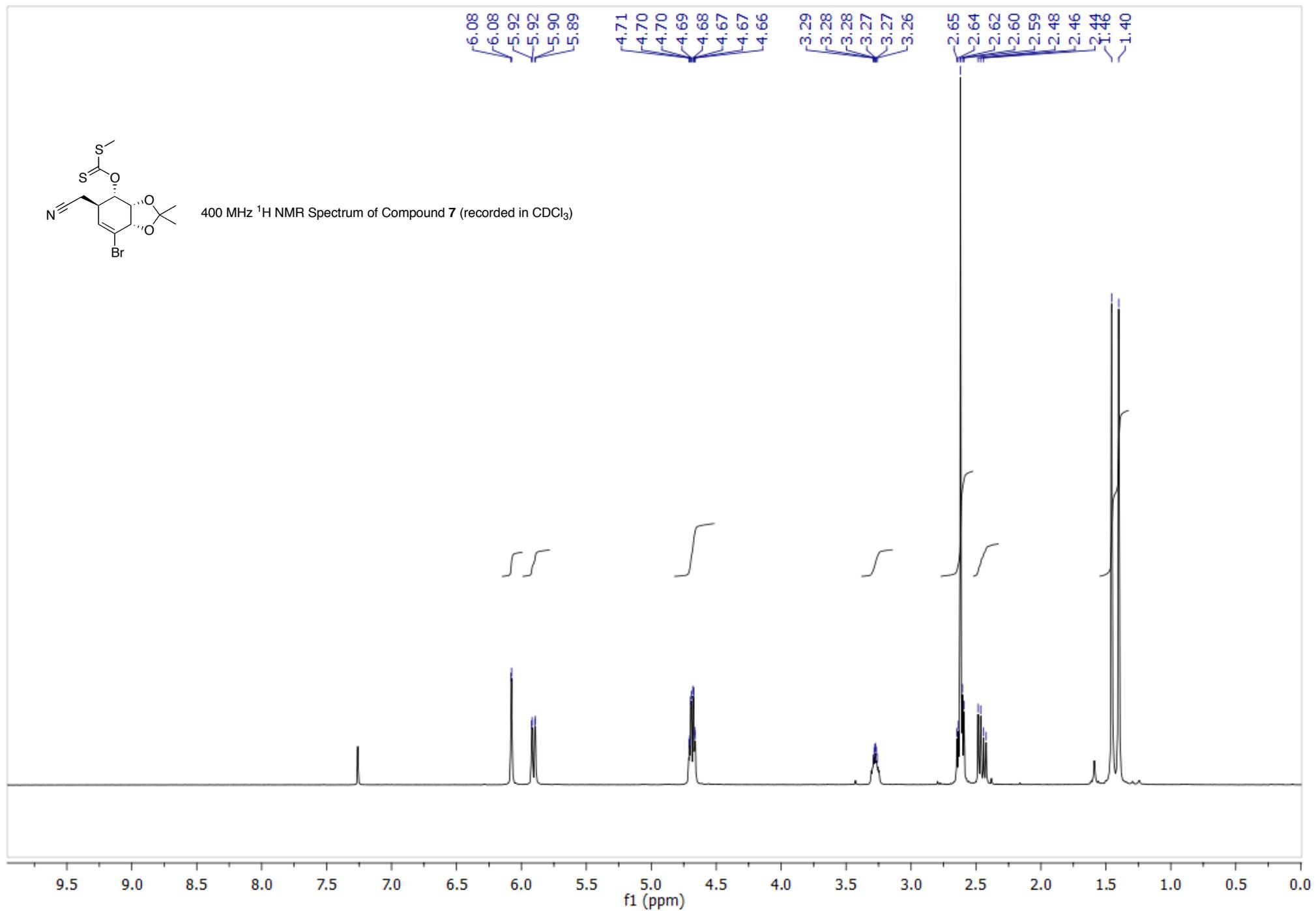


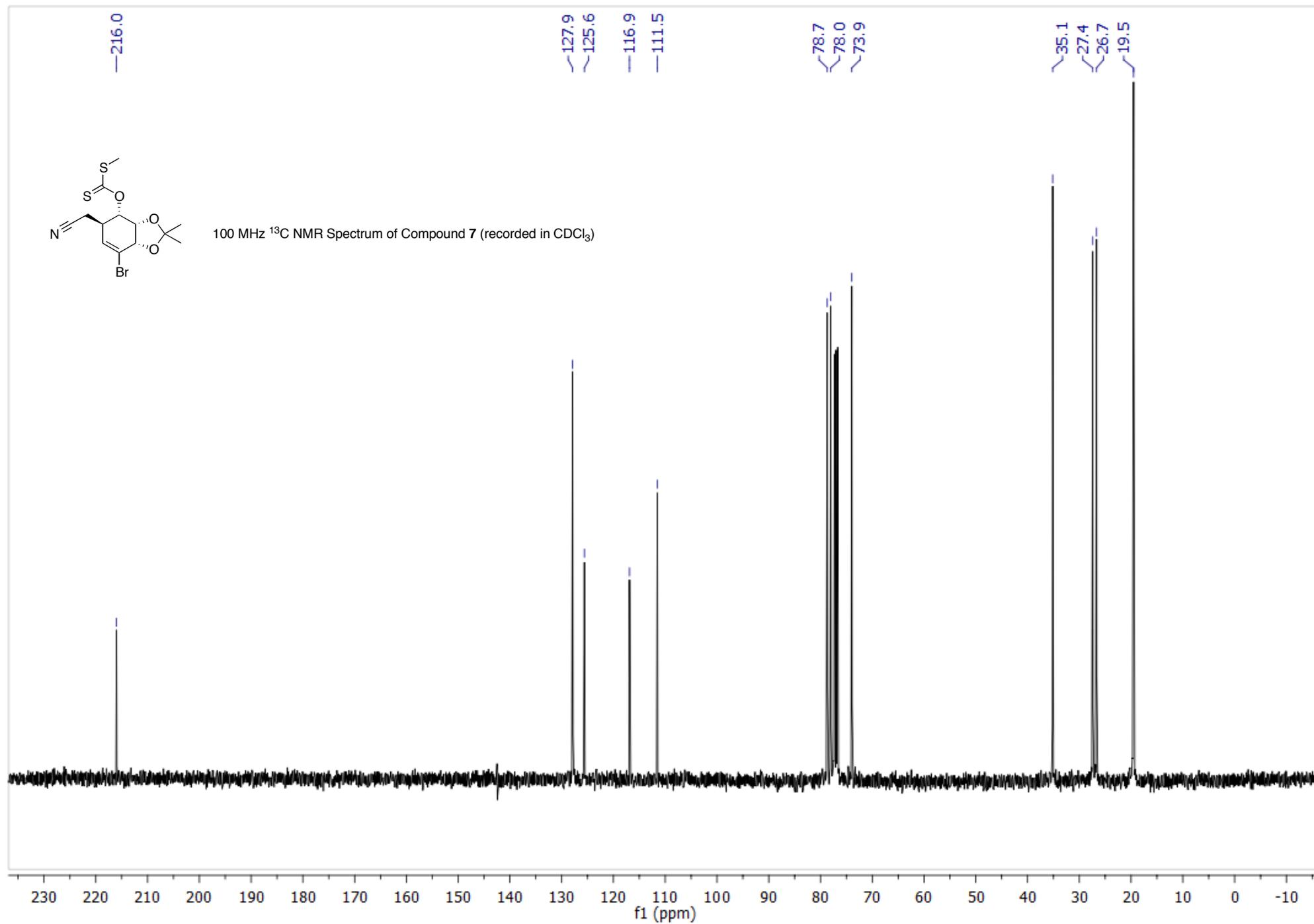
100 MHz ^{13}C NMR Spectrum of Compound 6 (recorded in CDCl_3)

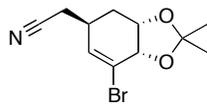




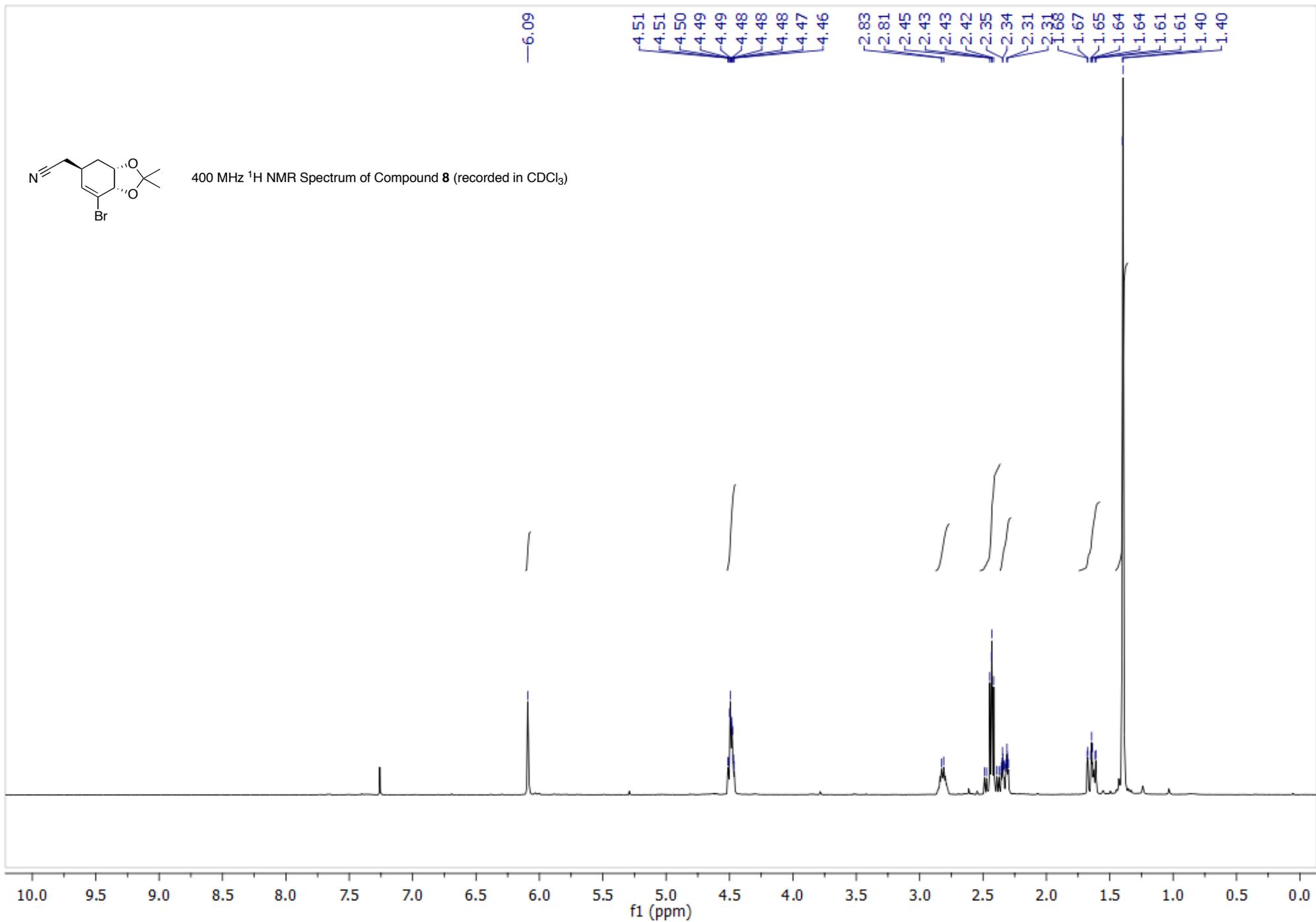
400 MHz ^1H NMR Spectrum of Compound 7 (recorded in CDCl_3)

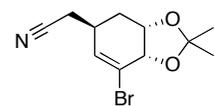




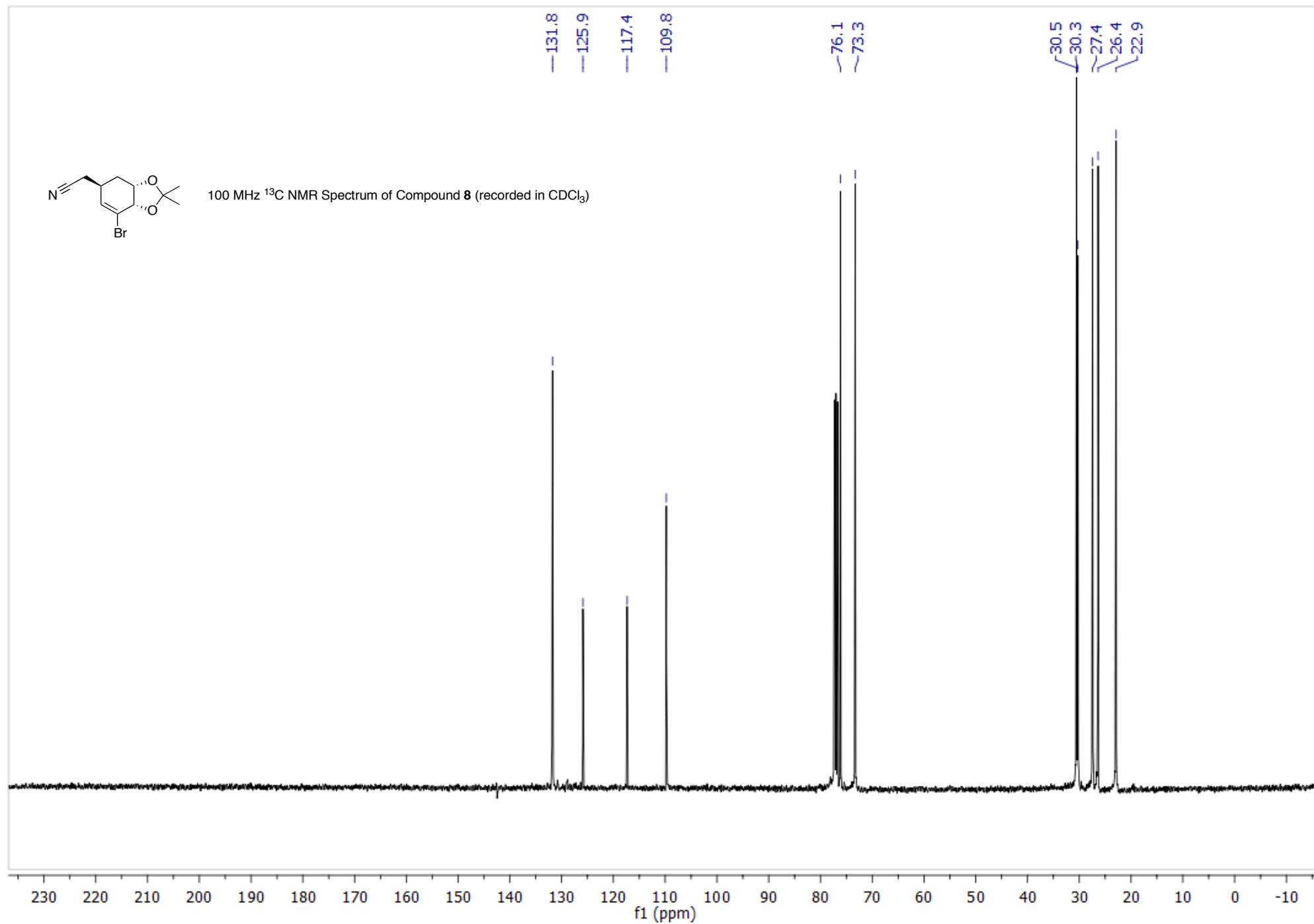


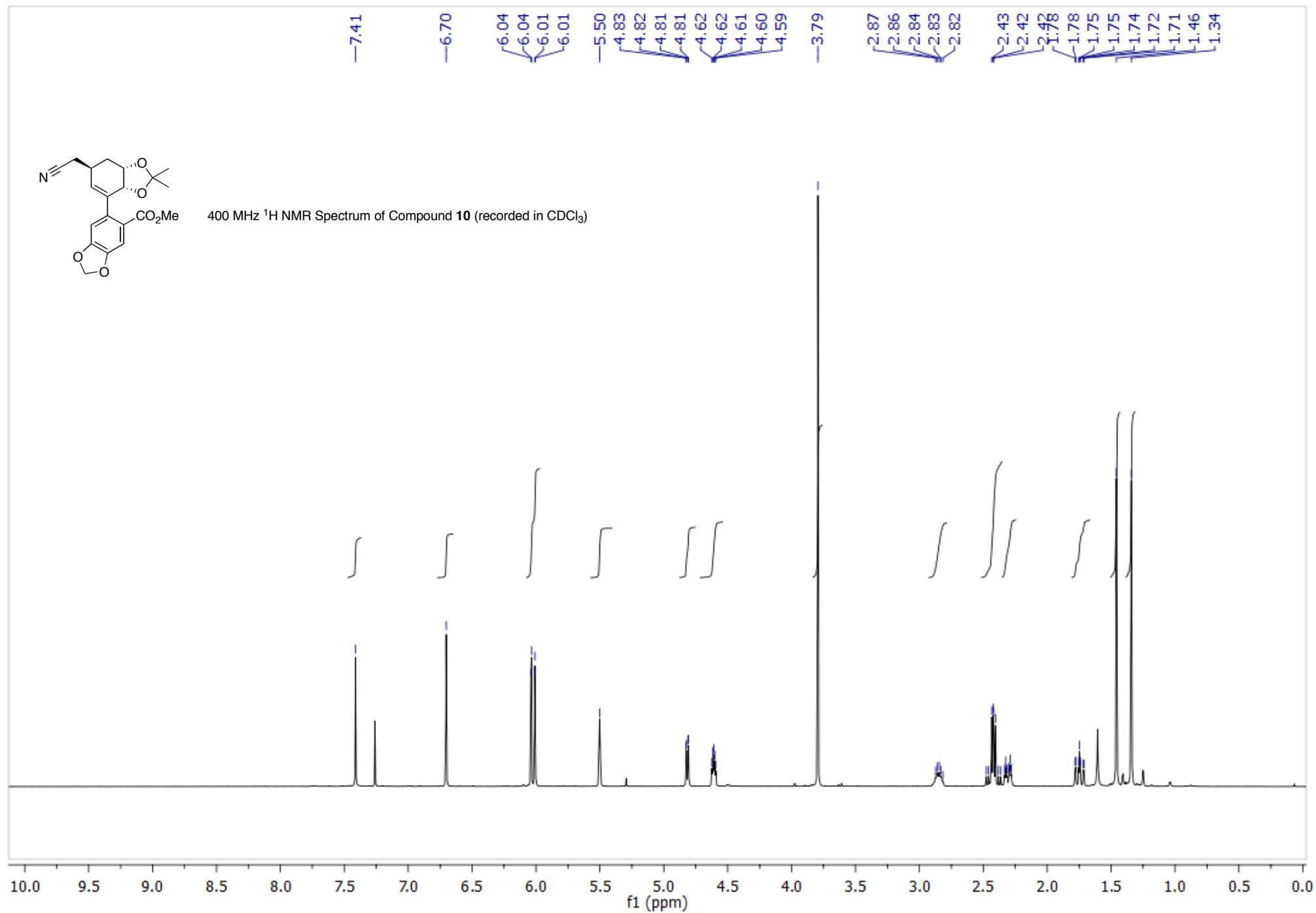
400 MHz ^1H NMR Spectrum of Compound **8** (recorded in CDCl_3)

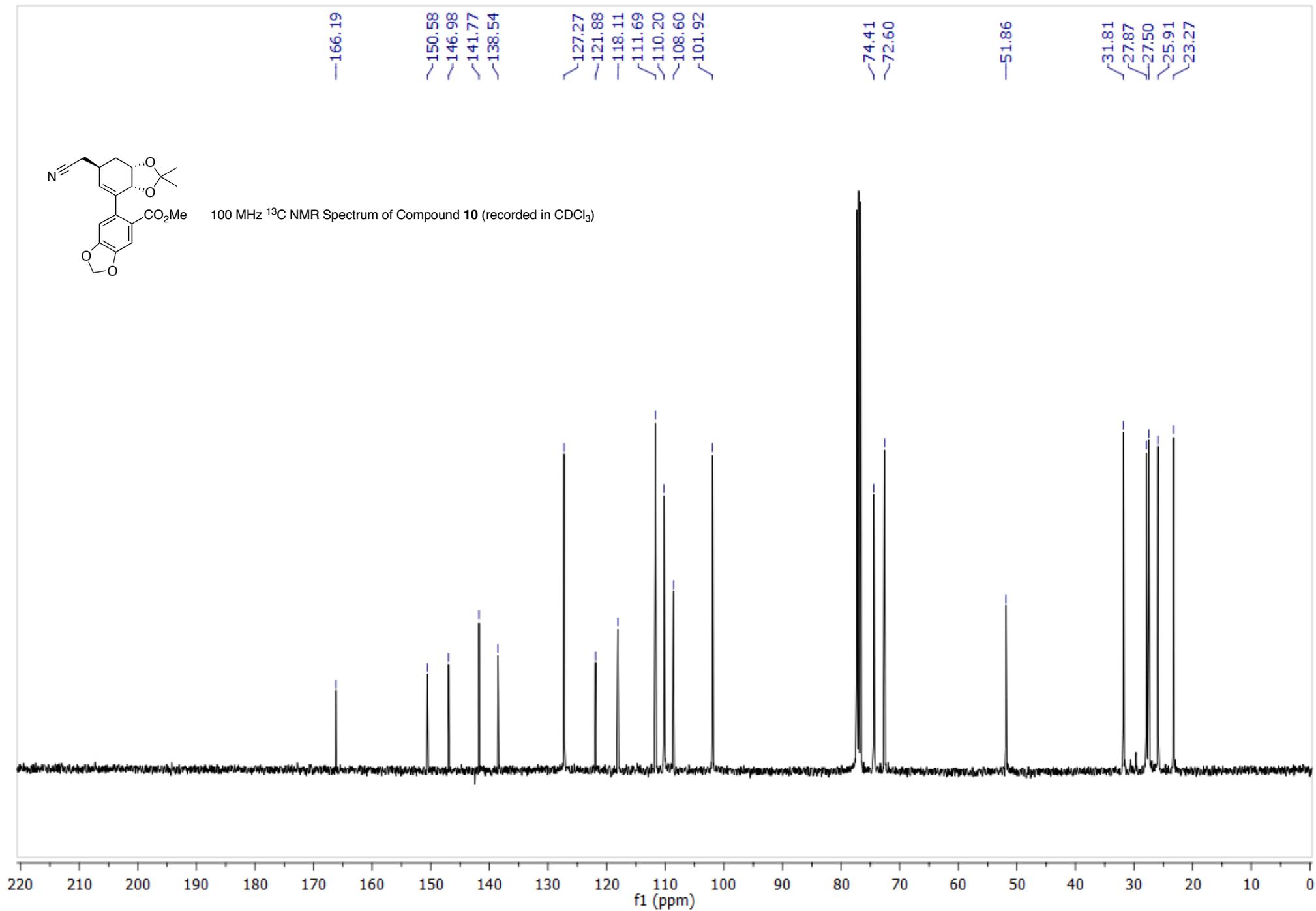


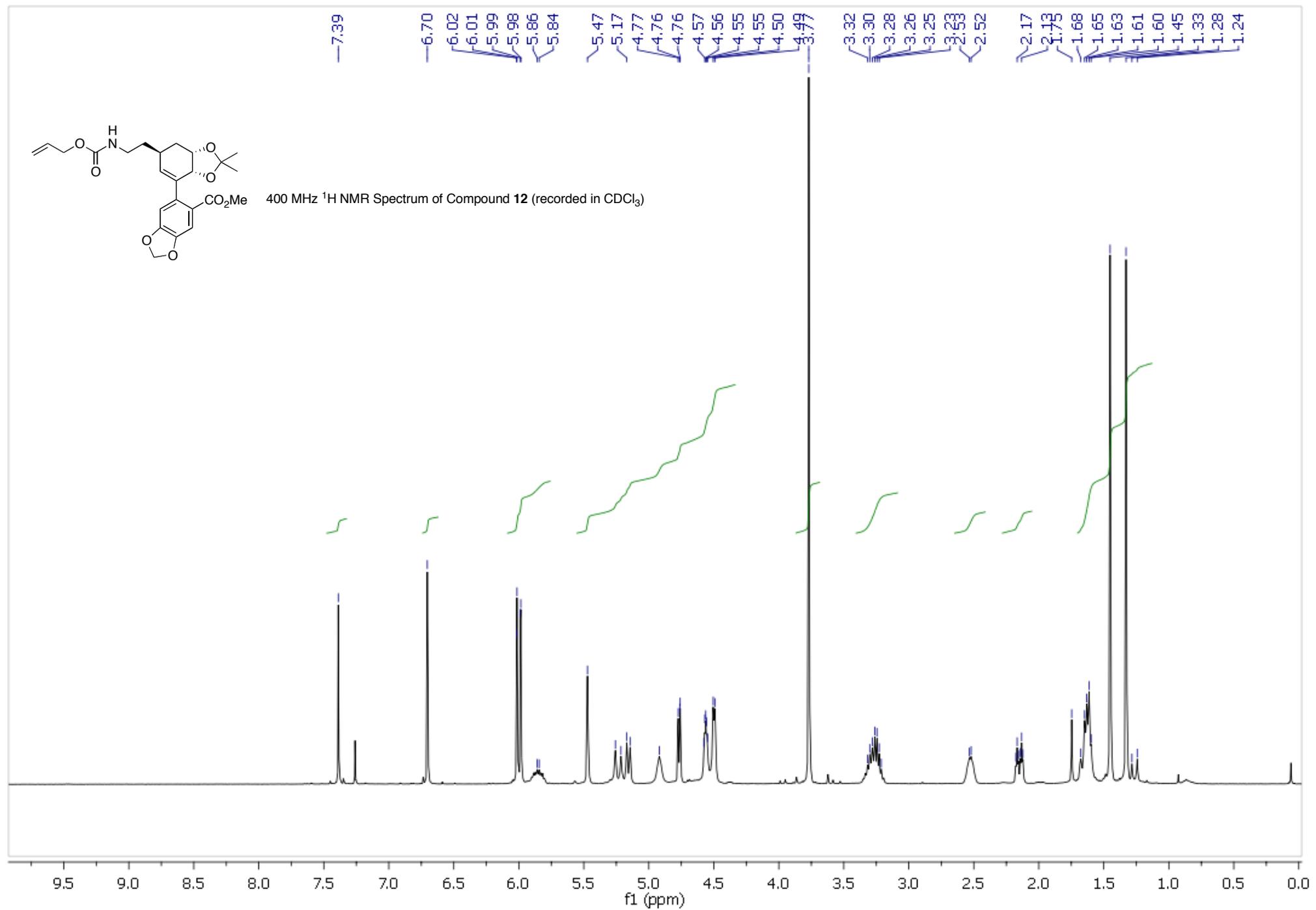


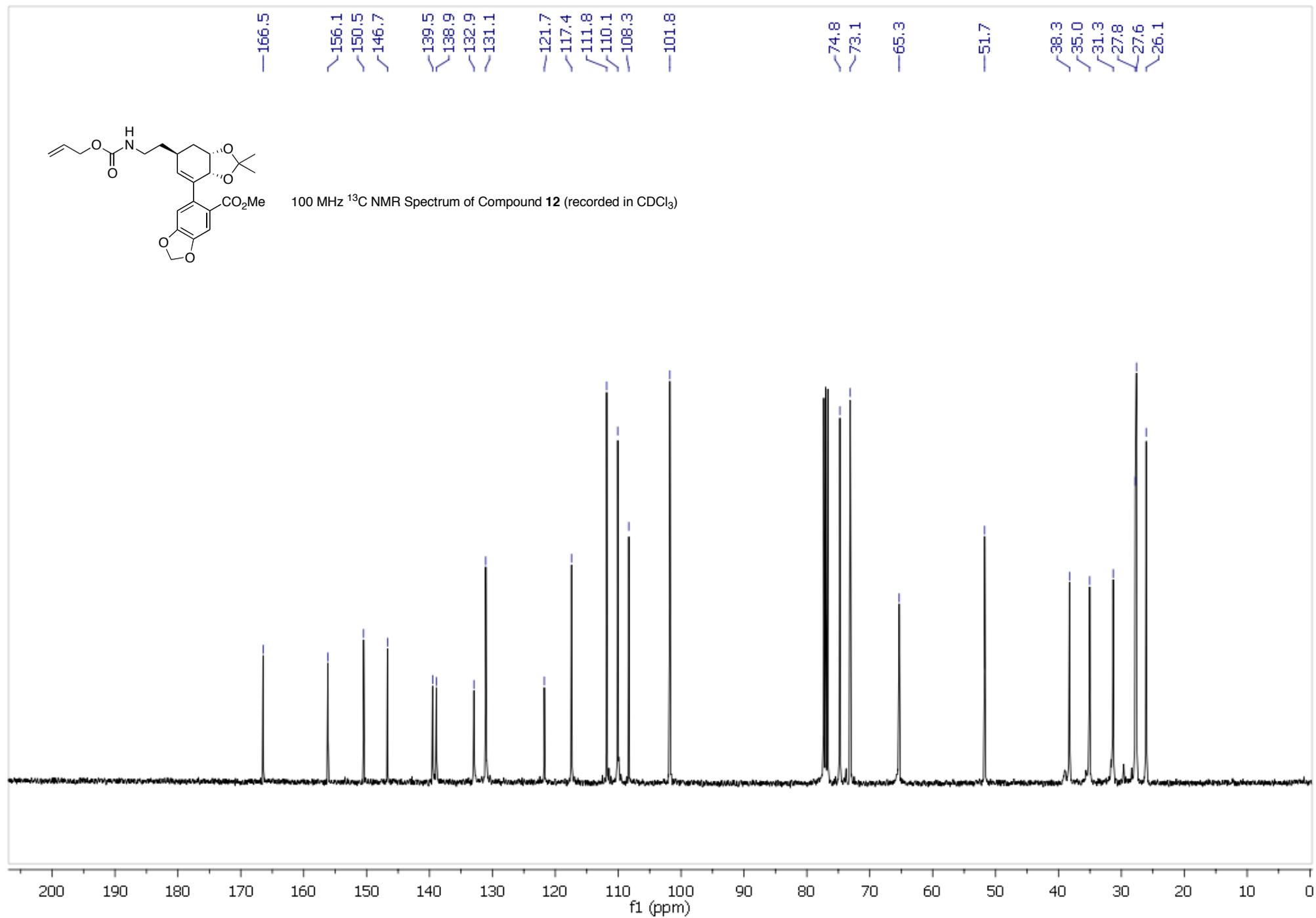
100 MHz ^{13}C NMR Spectrum of Compound **8** (recorded in CDCl_3)

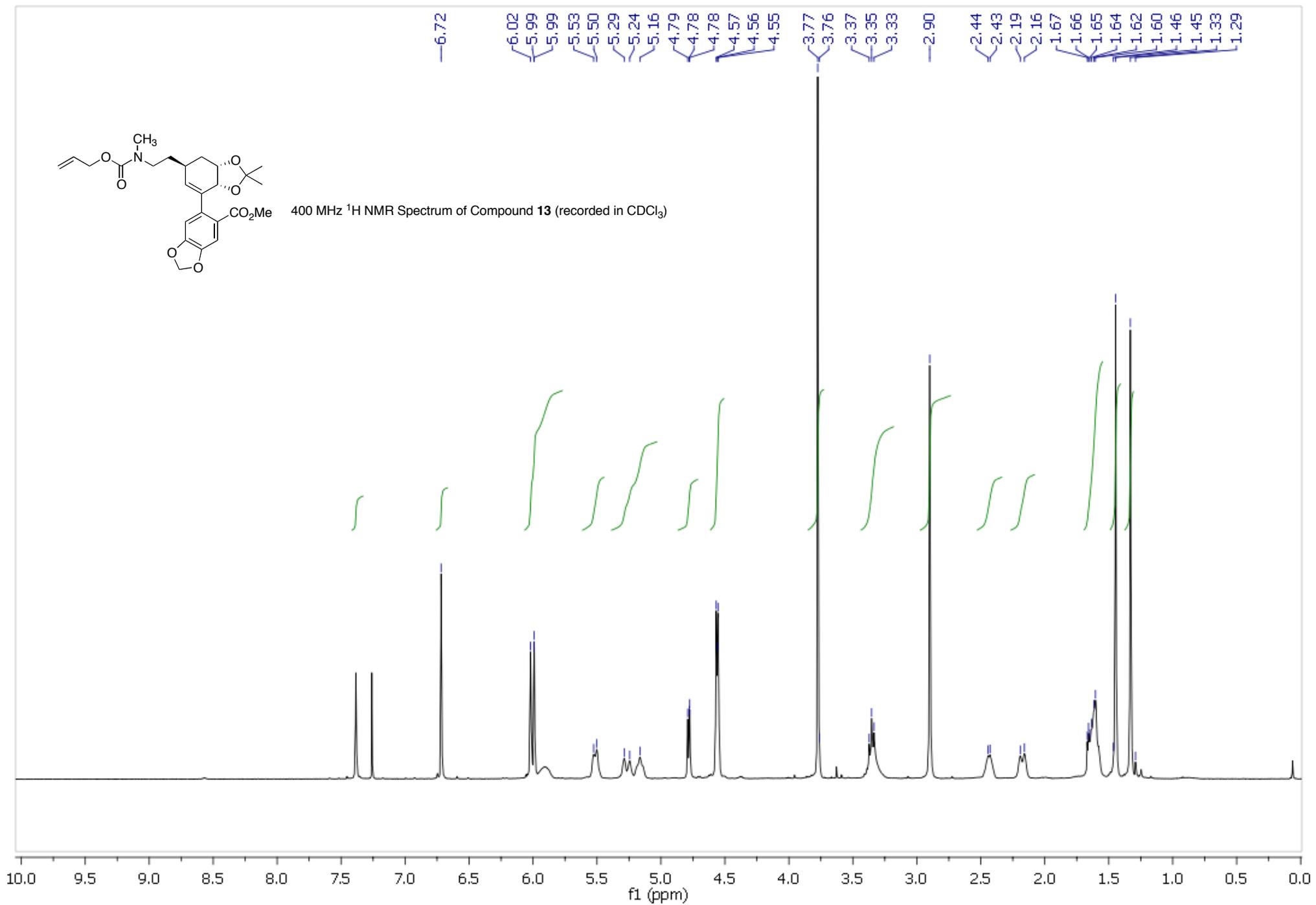


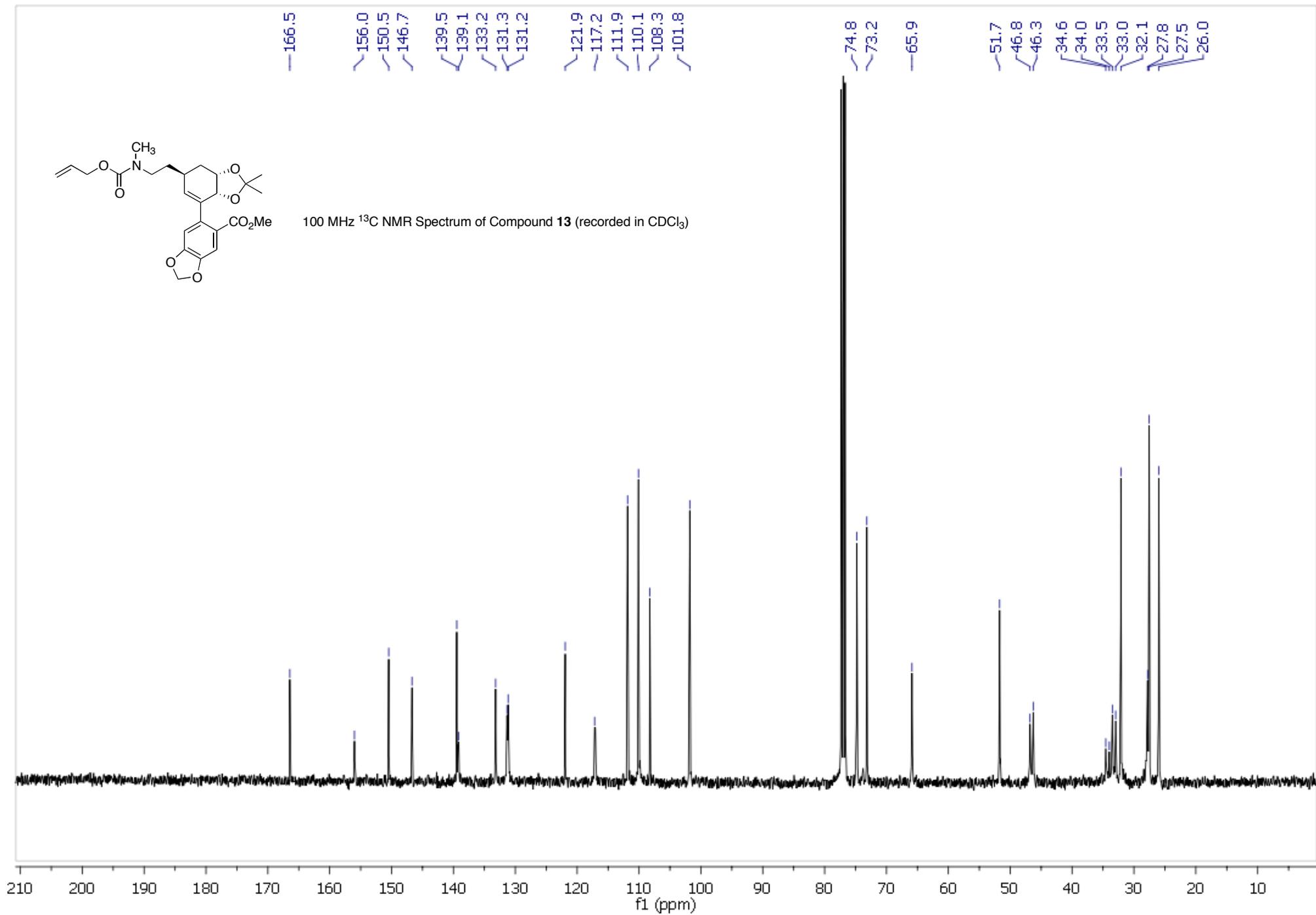


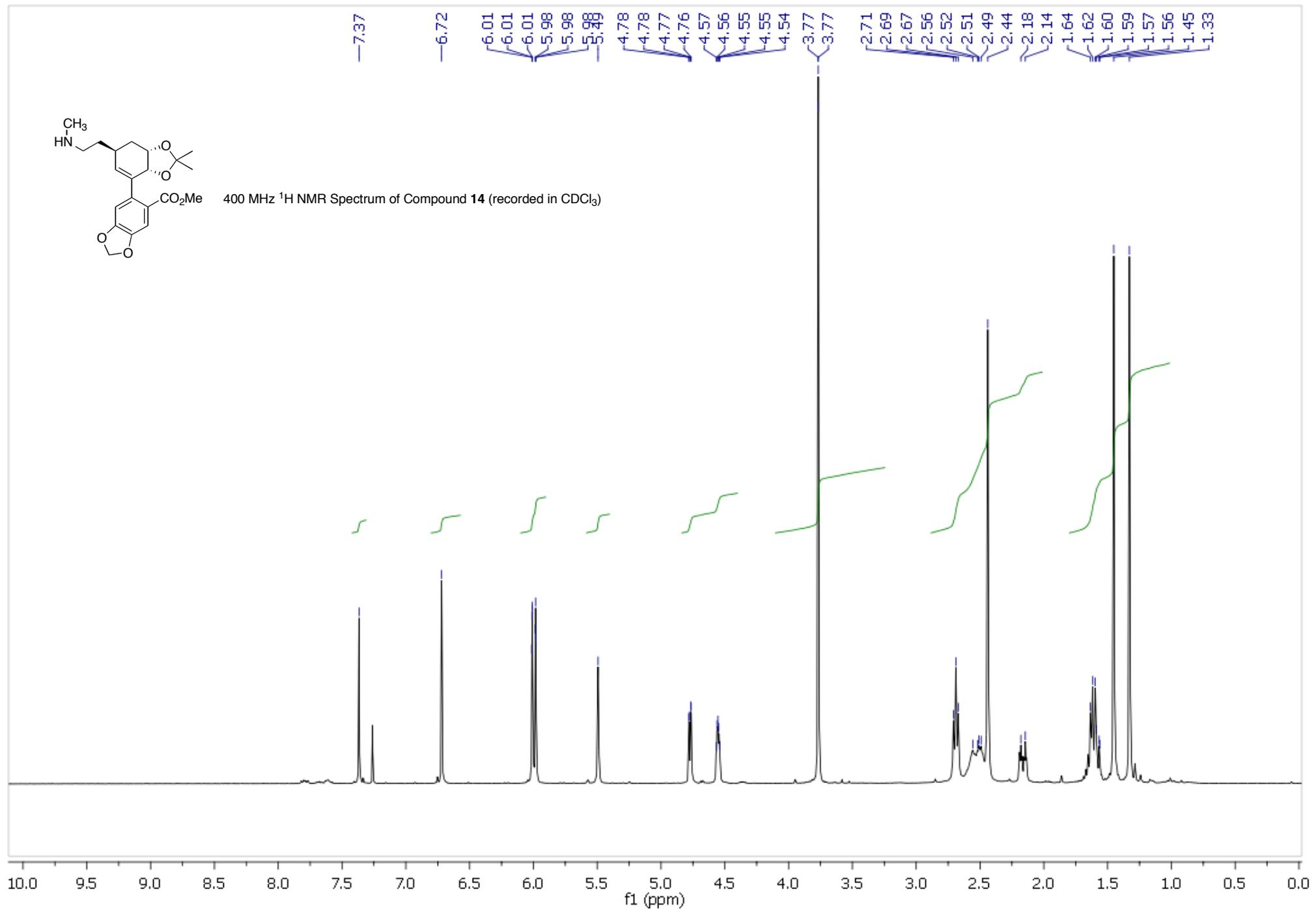


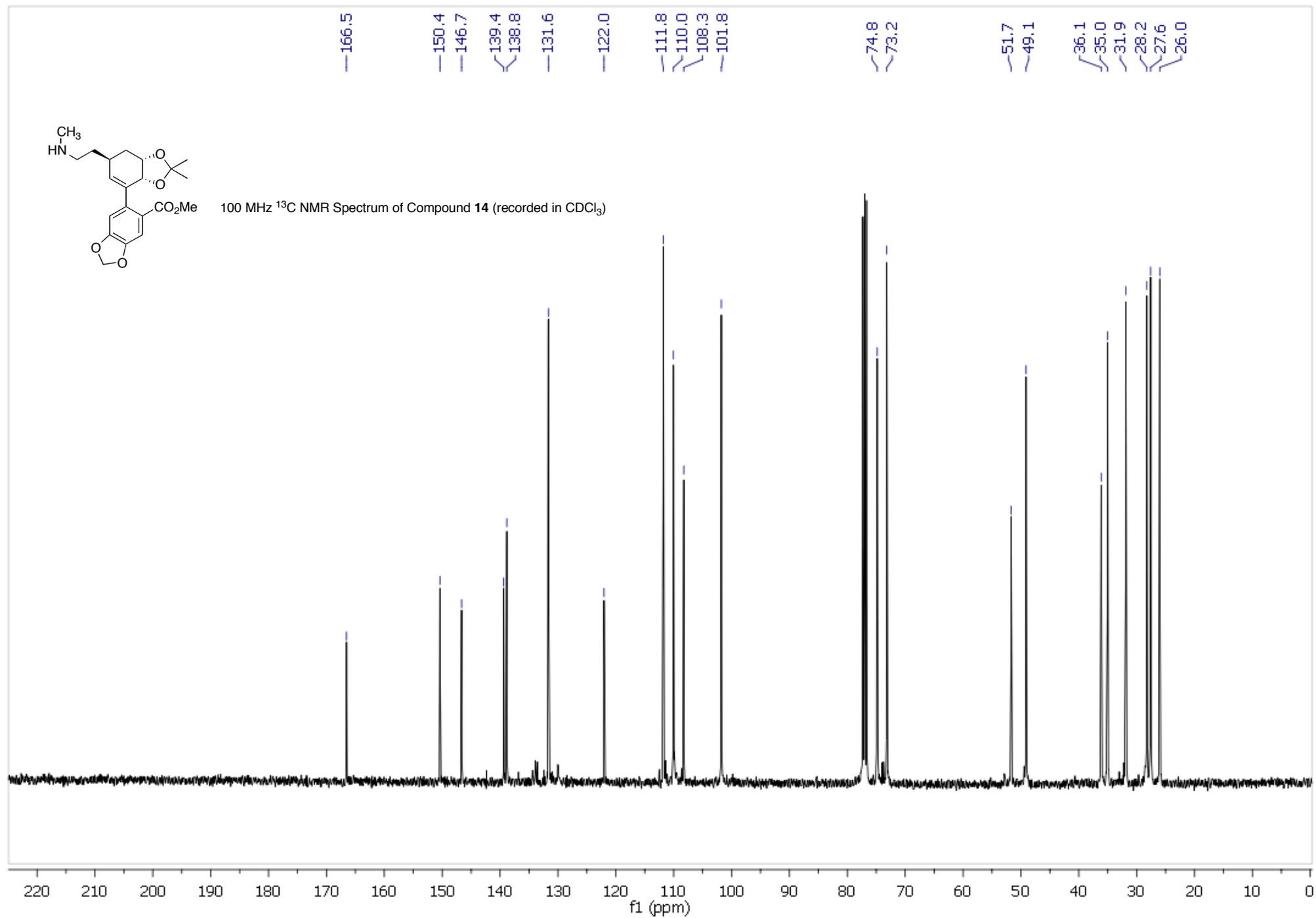


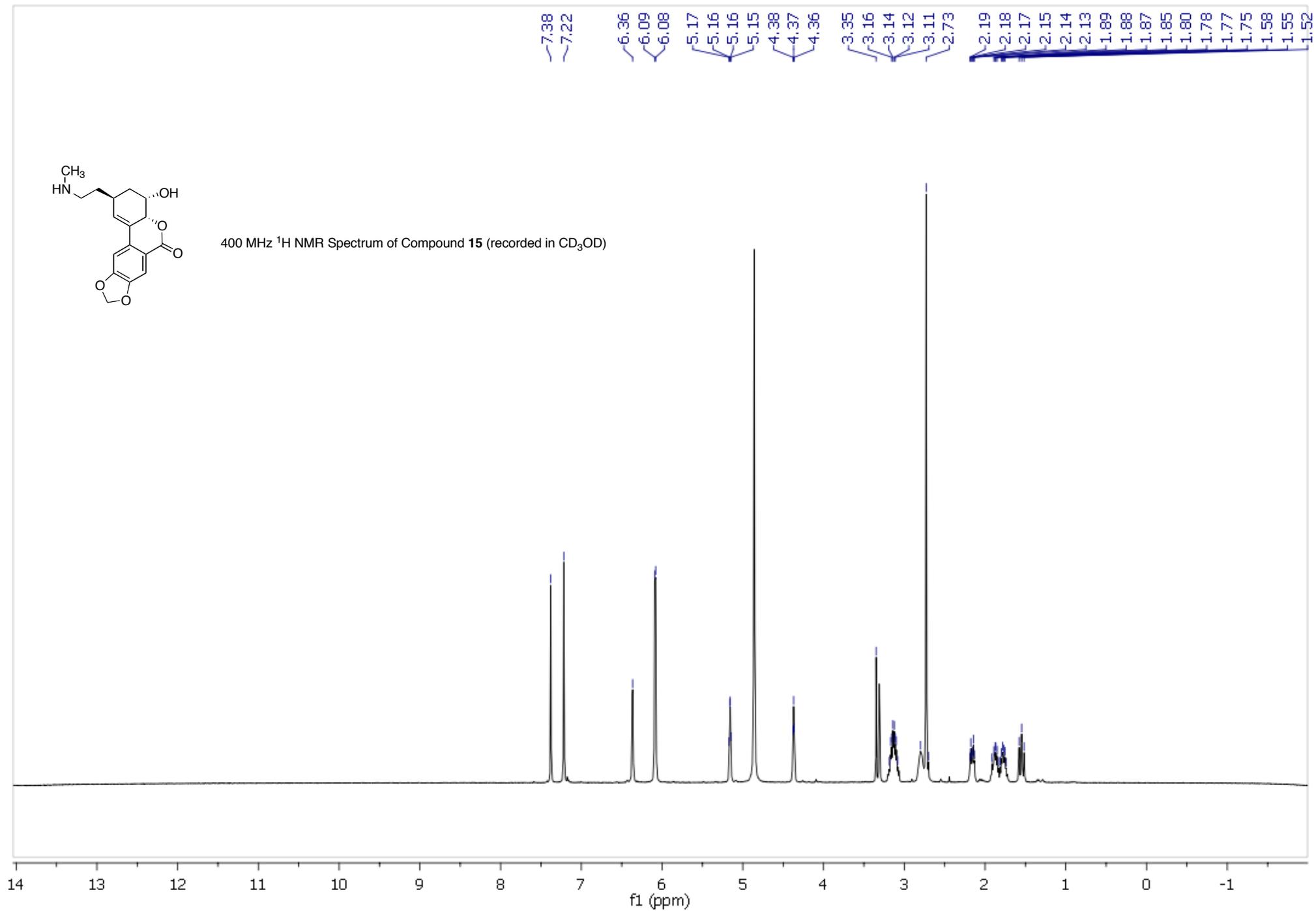


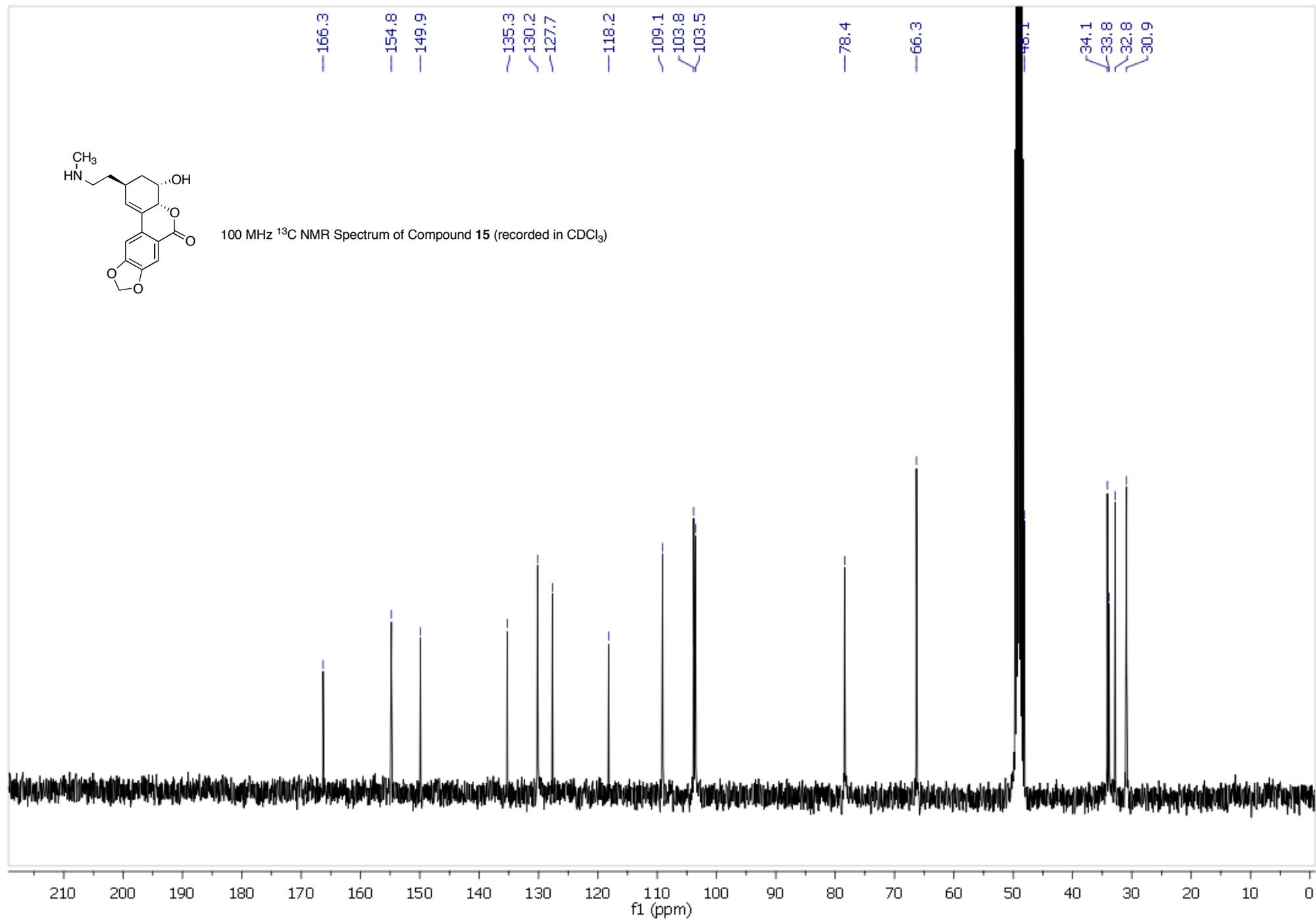


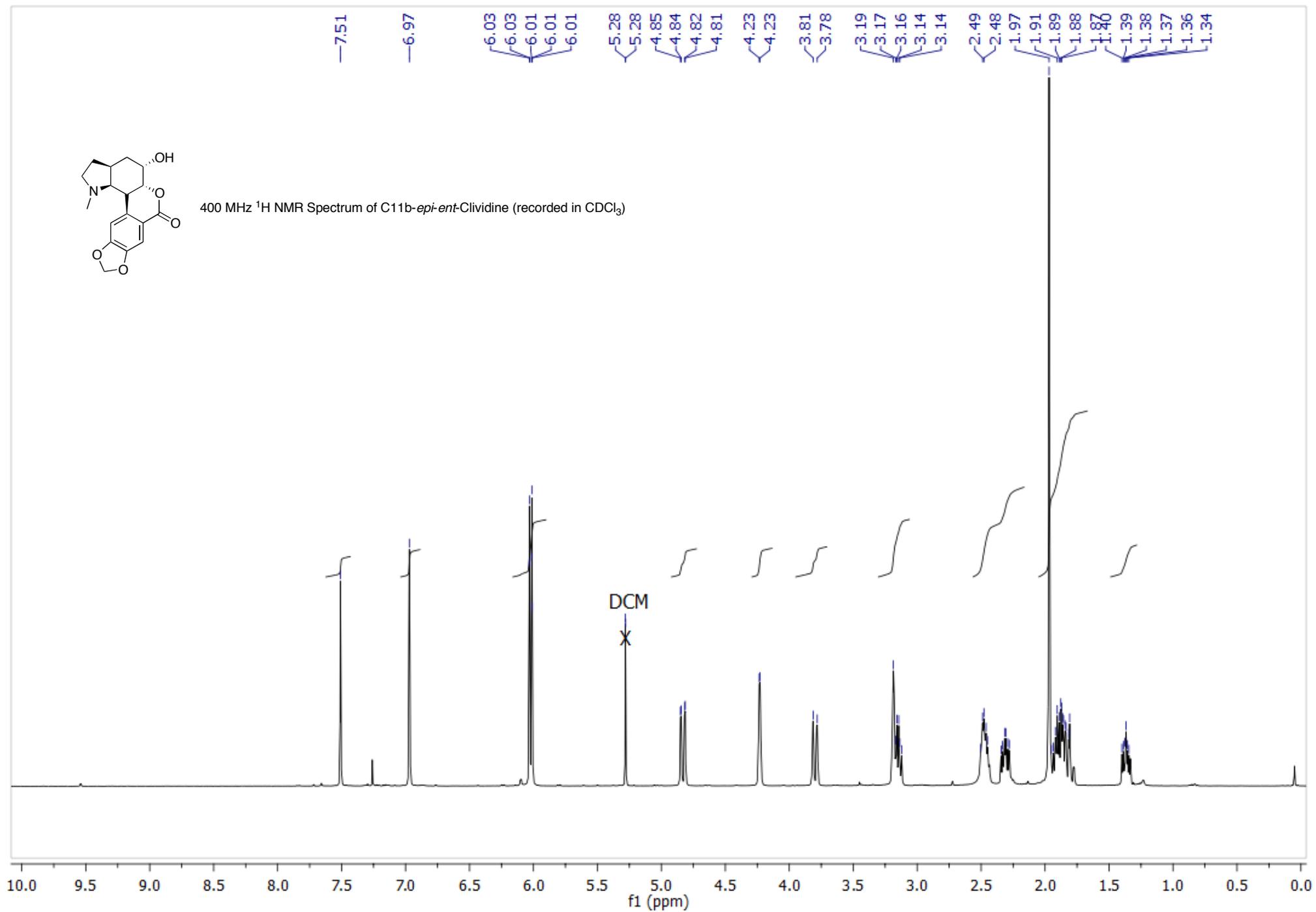


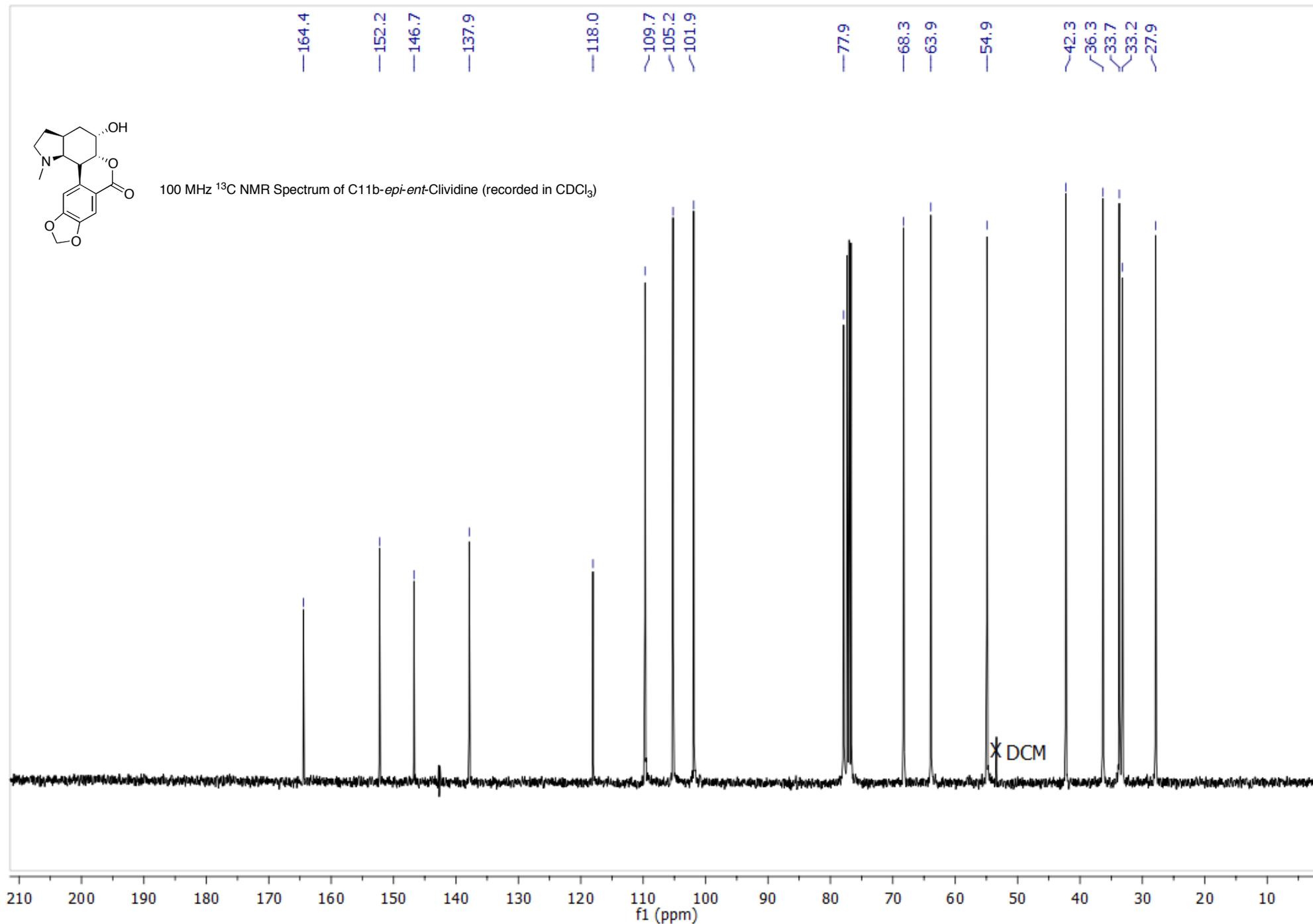


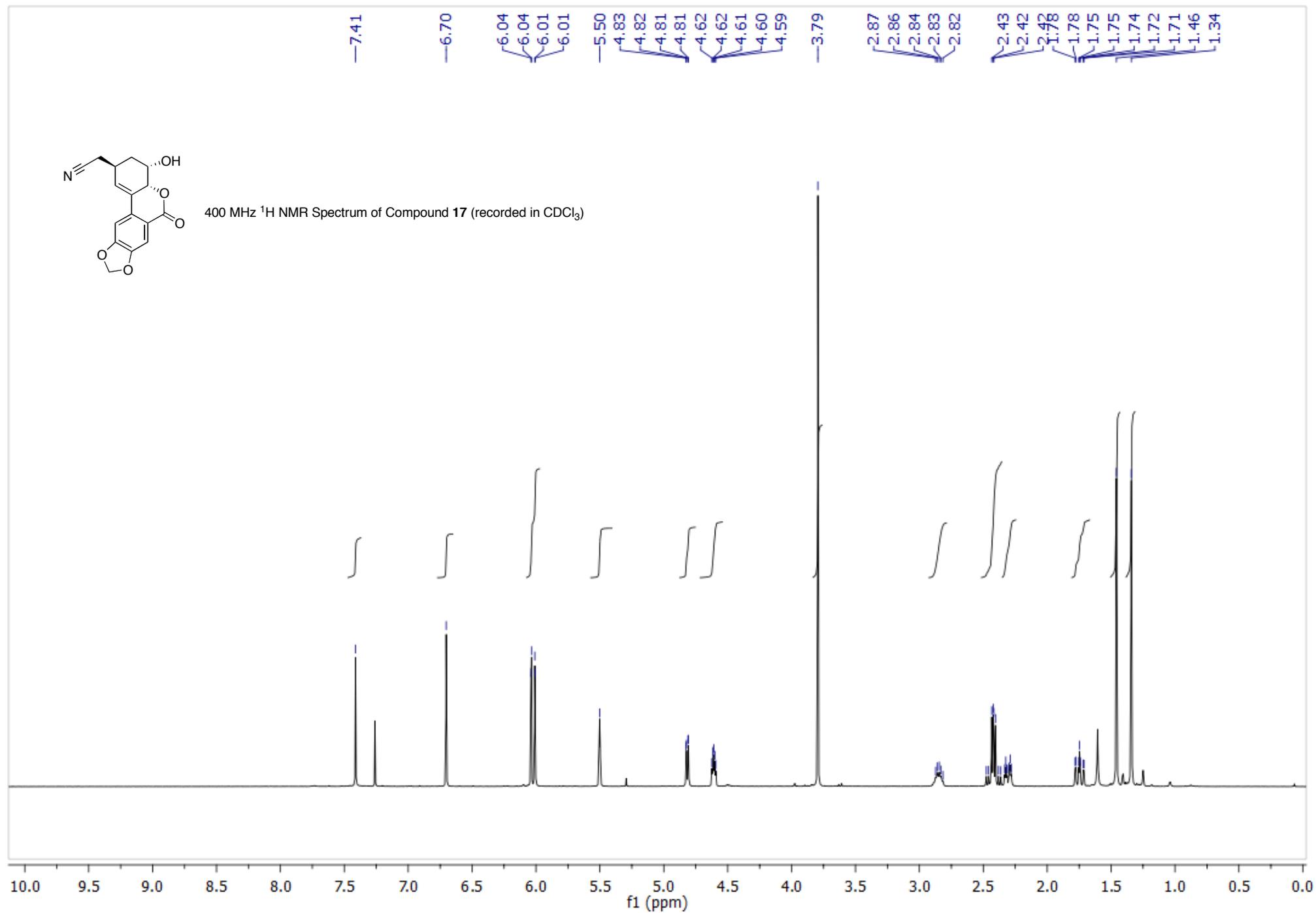


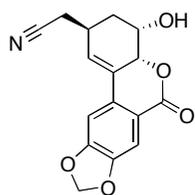












100 MHz ^{13}C NMR Spectrum of Compound 17 (recorded in d_6 -DMSO)

