

Functionalization of Planar Chiral Fused Arene Ruthenium Complexes: Synthesis, X-ray Structures and Spectroscopic Characterization of Monodentate Triarylphosphines

Philipp Buchgraber, Audrey Mercier, Wee Chuan Yeo, Céline Besnard and E. Peter Kündig*

*Department of Organic Chemistry, University of Geneva
Quai Ernest Ansermet, 1211, Geneva 4, Switzerland
peter.kundig@unige.ch*

CONTENTS:

- 1. Crystallography**
- 2. NMR Tables**
- 3. NMR Spectra**
- 4. References**

1. Crystallography

The structures of **11b** and **12b** are of low quality due to a strong disorder observed for the counter-ion, the Cp* ring, and the solvent molecules. For both compounds, the Cp* and the PF₆ anion were modeled using two molecules. Distances and angles were restrained to ideal values. Restraints were also applied on anisotropic displacement parameters. Solvent molecules were taken into account using the bypass/squeeze method¹ in the program platon.² For **12b**, two voids were located, each containing 43 electrons, which were attributed to two CH₂Cl₂ molecules. For **11b**, four voids were located, each containing 57 electrons, which were attributed to four CH₂Cl₂ molecules. The attribution of CH₂Cl₂ as solvent molecule was straightforward for **11b**, for which the Fourier difference map revealed disordered CH₂Cl₂ molecules that could be identified (but a model using two disordered molecules per void, each with an occupancy of 0.5 was not good enough to model the disorder and hence squeeze was preferred to a disordered model of the solvent). For **12b**, the density in the Fourier difference map was much more diffuse and therefore the solvent molecule could not be attributed to CH₂Cl₂ or hexane (both have a number of electrons compatible with the one found by squeeze). The attribution of the nature of the solvent molecule was based on the elemental analysis. Besides a reasonable fit to the data, reflected in the low R values obtained at the end of the refinement, these disordered models are of lower quality. However, the anisotropic displacement parameters are reasonable in the relevant region of the structure. The phenyl substituents on the phosphine showed elongated anisotropic parameters which are compatible with a pendulum movement with center C13 and axis C13-C16 (Figure S1).

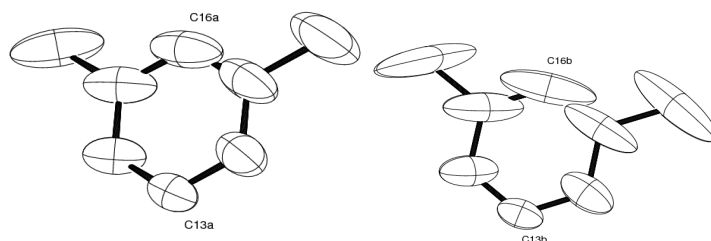
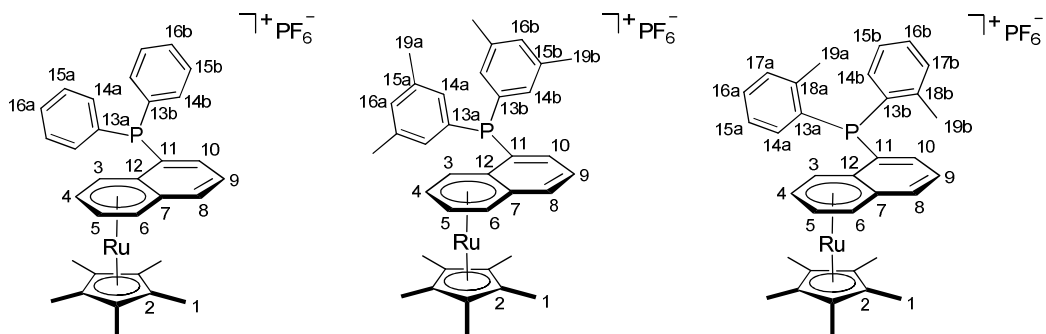


Figure S1. Anisotropic displacement parameters for the phosphine substituents in **11b**.

2. NMR Tables

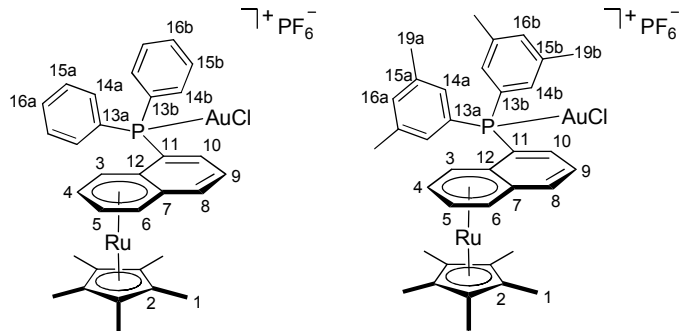
Table S1. ^{13}C NMR Spectroscopic Data for Phosphine Complexes 10b-12b^[a]



	10b		11b		12b	
Position	$\delta_{\text{C}}^{[\text{b}]}$	multiplicity ^[c]	$\delta_{\text{C}}^{[\text{b}]}$	multiplicity ^[c]	$\delta_{\text{C}}^{[\text{b}]}$	multiplicity ^[c]
1	10.0	d, $J = 1.9$ Hz	10.0	d, $J = 2.0$ Hz	10.1	d, $J = 1.1$ Hz
2	94.9	s	94.7	s	95.0	s
3	83.5	d, $J = 26.3$ Hz	83.6	d, $J = 26.0$ Hz	83.8	d, $J = 23.3$ Hz
4	88.0	d, $J = 1.6$ Hz	87.0	d, $J = 1.5$ Hz	87.8	d, $J = 1.4$ Hz
5	88.8	d, $J = 1.2$ Hz	88.6	d, $J = 1.3$ Hz	88.7	d, $J < 1.0$ Hz
6	86.1	d, $J = 1.2$ Hz	86.0	d, $J = 1.4$ Hz	85.7	d, $J < 1.0$ Hz
7	97.2	d, $J = 3.2$ Hz	97.3	d, $J = 3.1$ Hz	96.9	d, $J = 2.7$ Hz
8	128.7	s	128.3	s	128.4	d, $J = 1.4$ Hz
9	130.6	s	130.7	s	130.8	s
10	136.2	d, $J = 1.3$ Hz	136.1	d, $J = 1.5$ Hz	136.3	d, $J = 3.5$ Hz
11	137.7	d, $J = 20.3$ Hz	138.4	d, $J = 20.7$ Hz	136.8	d, $J = 21.0$ Hz
12	98.8	d, $J = 20.8$ Hz	98.8	d, $J = 20.6$ Hz	100.0	d, $J = 19.4$ Hz
13a	133.8	d, $J = 7.0$ Hz	133.6	d, $J = 6.2$ Hz	142.9	d, $J = 28.2$ Hz
13b	133.3	d, $J = 8.5$ Hz	133.0	d, $J = 7.9$ Hz	144.3	d, $J = 28.5$ Hz
14a	134.5	d, $J = 20.6$ Hz	132.7	d, $J = 21.2$ Hz	131.2	d, $J = 5.5$ Hz
15a	129.5	d, $J = 7.9$ Hz	139.3	d, $J = 8.4$ Hz	130.1	s
14b	135.1	d, $J = 20.7$ Hz	132.1	d, $J = 20.9$ Hz	130.9	d, $J = 5.9$ Hz
15b	129.7	d, $J = 7.8$ Hz	139.1	d, $J = 8.4$ Hz	130.8	s
16a	130.4	s	132.3	s	126.7	s
16b	130.6	s	132.0	s	127.5	d, $J = 1.4$ Hz
17a	--	--	--	--	134.0	s
17b	--	--	--	--	135.2	s
18a	--	--	--	--	133.5	d, $J = 6.8$ Hz
18b	--	--	--	--	130.9	d, $J = 8.0$ Hz
19a	--	--	21.4	s	21.3	d, $J = 22.9$ Hz
19b	--	--	21.3	s	22.0	d, $J = 22.0$ Hz

[a] Spectra recorded in CD_2Cl_2 at 125 MHz. [b] δ_{C} in ppm. [c] Coupling constants in Hertz (Hz).

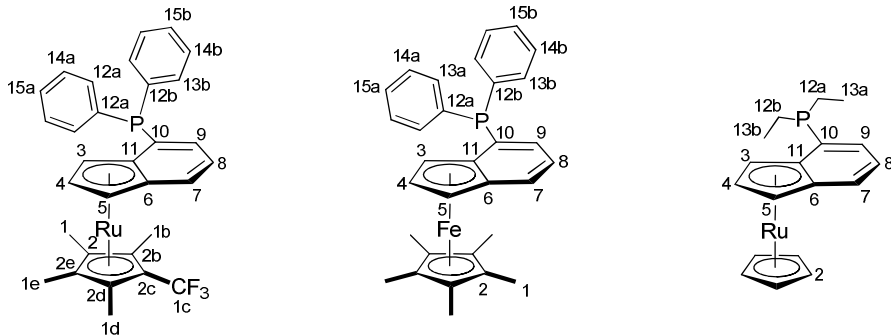
Table S2. ^{13}C NMR Spectroscopic Data for Gold(I) Complexes 13b and 14b^[a]



	13b		14b	
Position	$\delta_{\text{C}}^{[\text{b}]}$	multiplicity ^[c]	$\delta_{\text{C}}^{[\text{b}]}$	multiplicity ^[c]
1	10.7	s	10.7	s
2	96.2	s	96.1	s
3	83.2	d, $J = 14.4$ Hz	83.3	d, $J = 14.0$ Hz
4	87.6	s	87.5	s
5	89.0	s	88.8	s
6	86.3	d, $J < 1.0$ Hz	86.2	s
7	95.4	d, $J = 6.1$ Hz	95.2	d, $J = 6.0$ Hz
8	134.7	d, $J = 2.4$ Hz	134.4	d, $J = 2.8$ Hz
9	129.2	d, $J = 10.6$ Hz	129.3	d, $J = 10.7$ Hz
10	140.1	d, $J = 5.9$ Hz	140.0	d, $J = 6.0$ Hz
11	126.4	d, $J = 19.5$ Hz	126.9	d, $J = 55.8$ Hz
12	99.0	d, $J = 11.9$ Hz	99.3	d, $J = 11.7$ Hz
13a	126.1	d, $J = 14.3$ Hz	125.9	d, $J = 19.6$ Hz
13b	125.7	d, $J = 22.8$ Hz	125.4	d, $J = 15.9$ Hz
14a	135.4	d, $J = 14.4$ Hz	132.8	d, $J = 14.4$ Hz
15a	135.3	d, $J = 14.5$ Hz	132.5	d, $J = 14.5$ Hz
14b	130.4	d, $J = 12.4$ Hz	140.5	d, $J = 13.0$ Hz
15b	130.2	d, $J = 12.4$ Hz	140.2	d, $J = 13.1$ Hz
16a	133.6	d, $J = 2.7$ Hz	135.3	d, $J = 2.8$ Hz
16b	133.6	d, $J = 2.6$ Hz	135.3	d, $J = 2.5$ Hz
19a	--	--	21.5	s
19b	--	--	21.4	s

[a] Spectra recorded in CD_2Cl_2 at 125 MHz. [b] δ_{C} in ppm. [c] Coupling constants in Hertz (Hz)

Table S3. ^{13}C NMR Spectroscopic Data for Representative Indenyl Phosphines^[a]

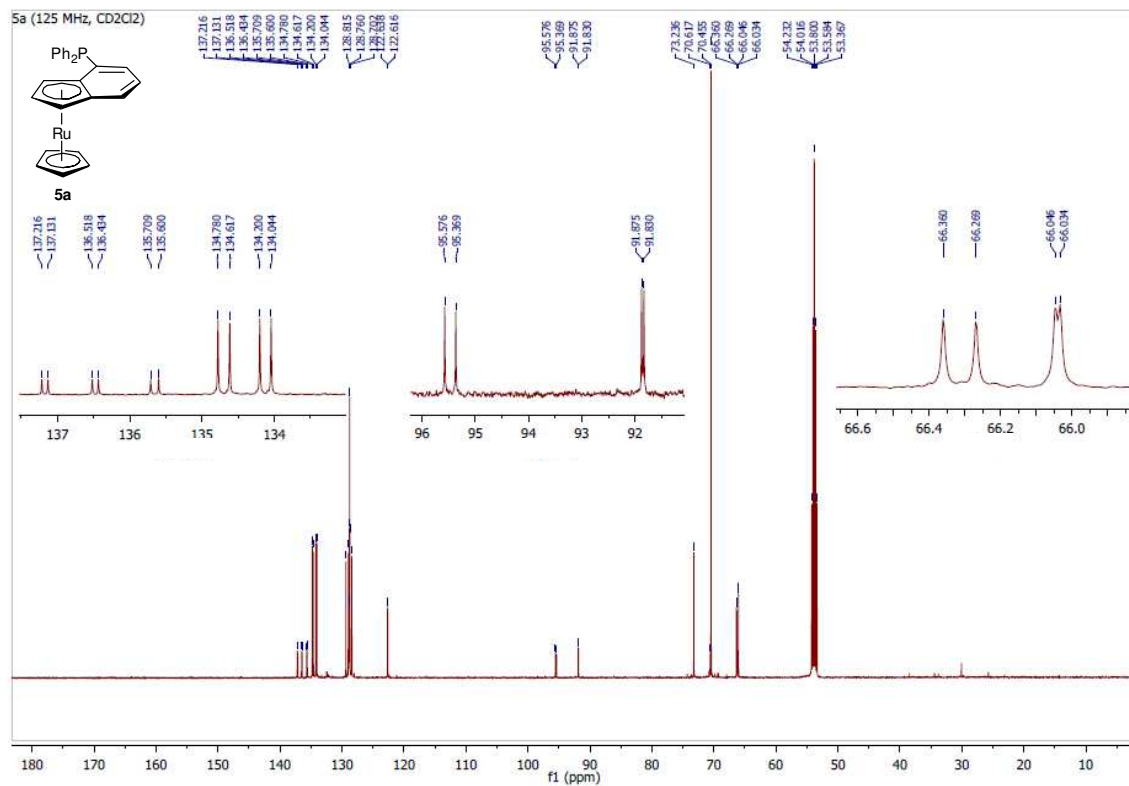
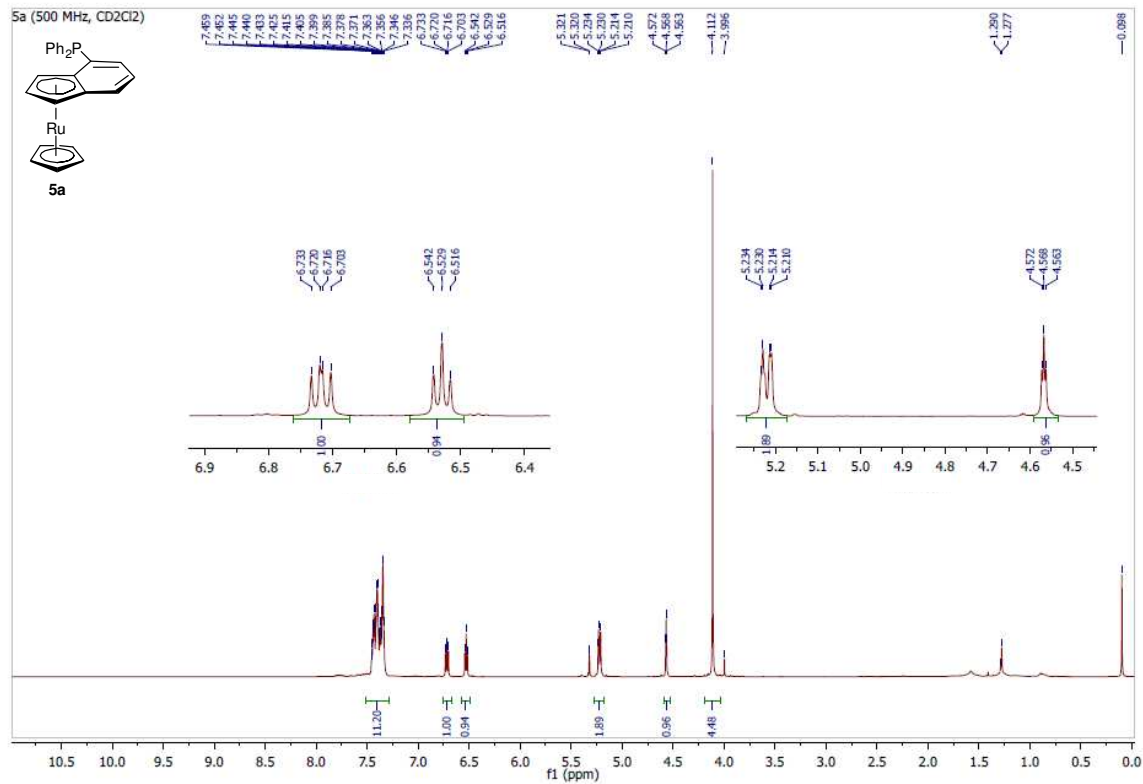


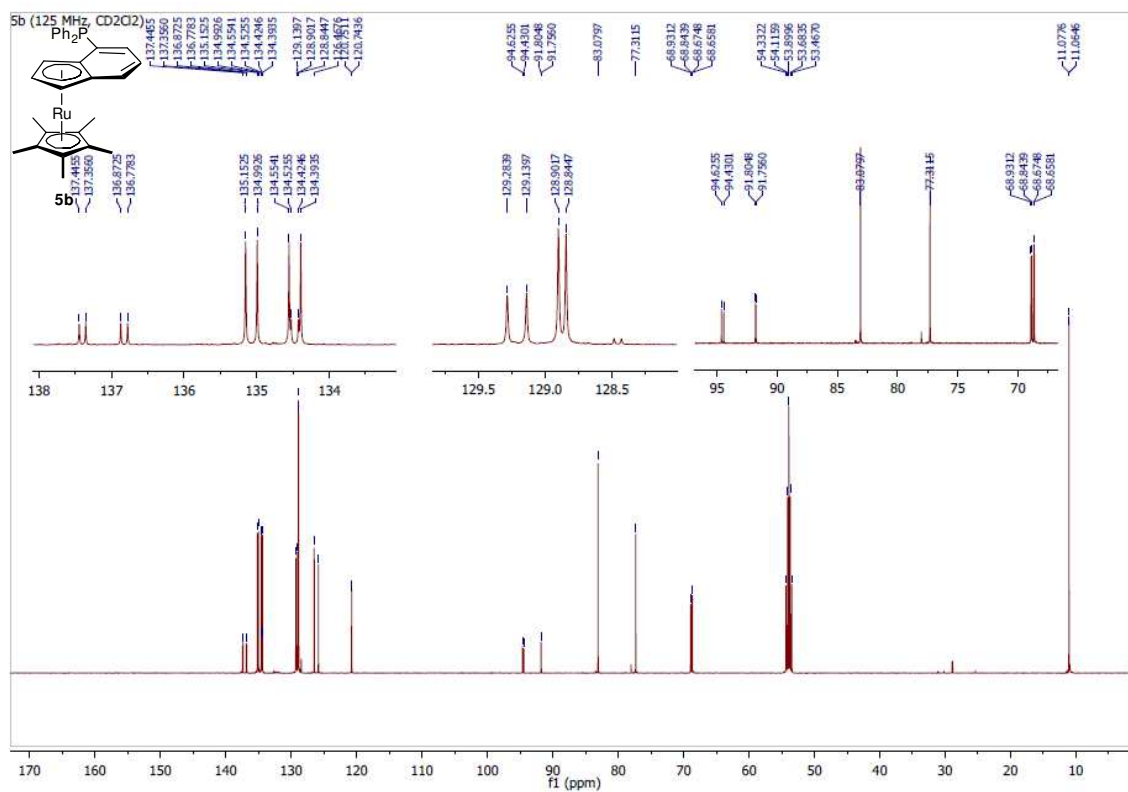
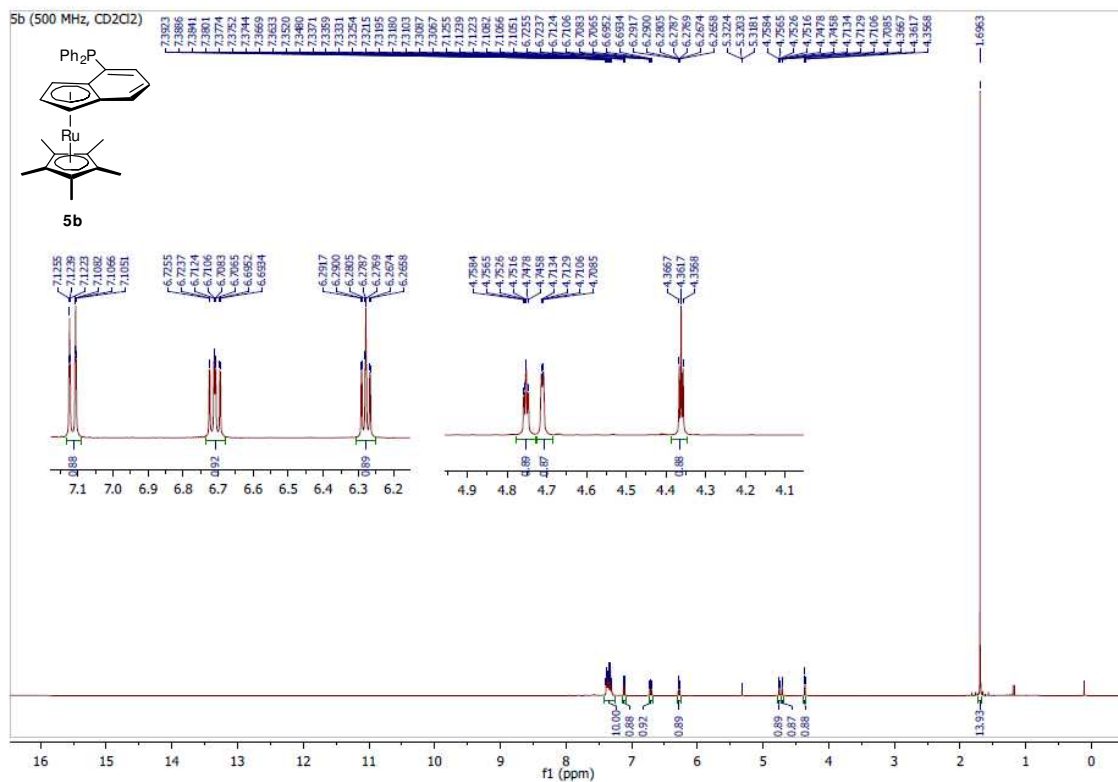
Position	5c			8b^[d]		7a	
	$\delta_{\text{C}}^{[\text{b}]}$	multiplicity ^[c]		$\delta_{\text{C}}^{[\text{b}]}$	multiplicity ^[c]	$\delta_{\text{C}}^{[\text{b}]}$	multiplicity ^[c]
1	9.7	d, $J = 3.0\text{ Hz}$		10.4	d, $J = 2.5\text{ Hz}$	--	--
2	85.7	s		78.0	s	70.4	s
2b	81.8	q, $J_{(\text{C},\text{F})} = 1.4\text{ Hz}$		--	--	--	--
1b	10.5	q, $J_{(\text{C},\text{F})} = 1.8\text{ Hz}$		--	--	--	--
2c	76.2	q, $J_{(\text{C},\text{F})} = 35.8\text{ Hz}$		--	--	--	--
1c	128.6	q, $J_{(\text{C},\text{F})} = 270\text{ Hz}$		--	--	--	--
2d	82.1	q, $J_{(\text{C},\text{F})} = 1.4\text{ Hz}$		--	--	--	--
1d	11.7	pent, $J \approx 2.0\text{ Hz}$		--	--	--	--
2e	86.0	s		--	--	--	--
1e	10.4	d, $J = 2.7\text{ Hz}$		--	--	--	--
3	69.8	d, $J = 11.0\text{ Hz}$		66.7	d, $J = 10.4\text{ Hz}$	66.1	d, $J = 9.6\text{ Hz}$
4	77.7	s		76.8	s	73.2	s
5	69.2	d, $J = 1.9\text{ Hz}$		66.3	d, $J = 1.9\text{ Hz}$	66.0	d, overlap
6	92.8	d, $J = 6.0\text{ Hz}$		88.2	d, $J = 6.0\text{ Hz}$	92.0	d, $J = 4.6\text{ Hz}$
7	126.2	s		128.8	s	127.6	s
8	122.2	s		122.2	s	122.7	s
9	127.1	s		127.4	s	126.0	s
10	134.8	d, $J = 14.0\text{ Hz}$		137.2	d, $J = 13.7\text{ Hz}$	136.4	d, $J = 19.2\text{ Hz}$
11	95.6	d, $J = 24.3\text{ Hz}$		91.2	d, $J = 23.6\text{ Hz}$	95.9	d, $J = 21.8\text{ Hz}$
12a	136.9	d, $J = 11.0\text{ Hz}$		137.0	d, $J = 10.9\text{ Hz}$	18.6	d, $J = 11.7\text{ Hz}$
12b	136.2	d, $J = 11.7\text{ Hz}$		137.0	d, $J = 12.0\text{ Hz}$	18.1	d, $J = 11.2\text{ Hz}$
13a	134.9	d, $J = 20.2\text{ Hz}$		135.2	d, $J = 20.2\text{ Hz}$	10.4	d, $J = 15.0\text{ Hz}$
13b	134.5	d, $J = 20.3\text{ Hz}$		134.4	d, $J = 20.1\text{ Hz}$	10.1	d, $J = 12.2\text{ Hz}$
14a	128.9	d, $J = 7.3\text{ Hz}$		128.8	d, $J = 7.3\text{ Hz}$	--	--
14b	128.9	d, $J = 7.3\text{ Hz}$		128.7	d, $J = 7.1\text{ Hz}$	--	--
15a	129.4	s		129.2	s	--	--
15b	129.2	s		128.8	s	--	--

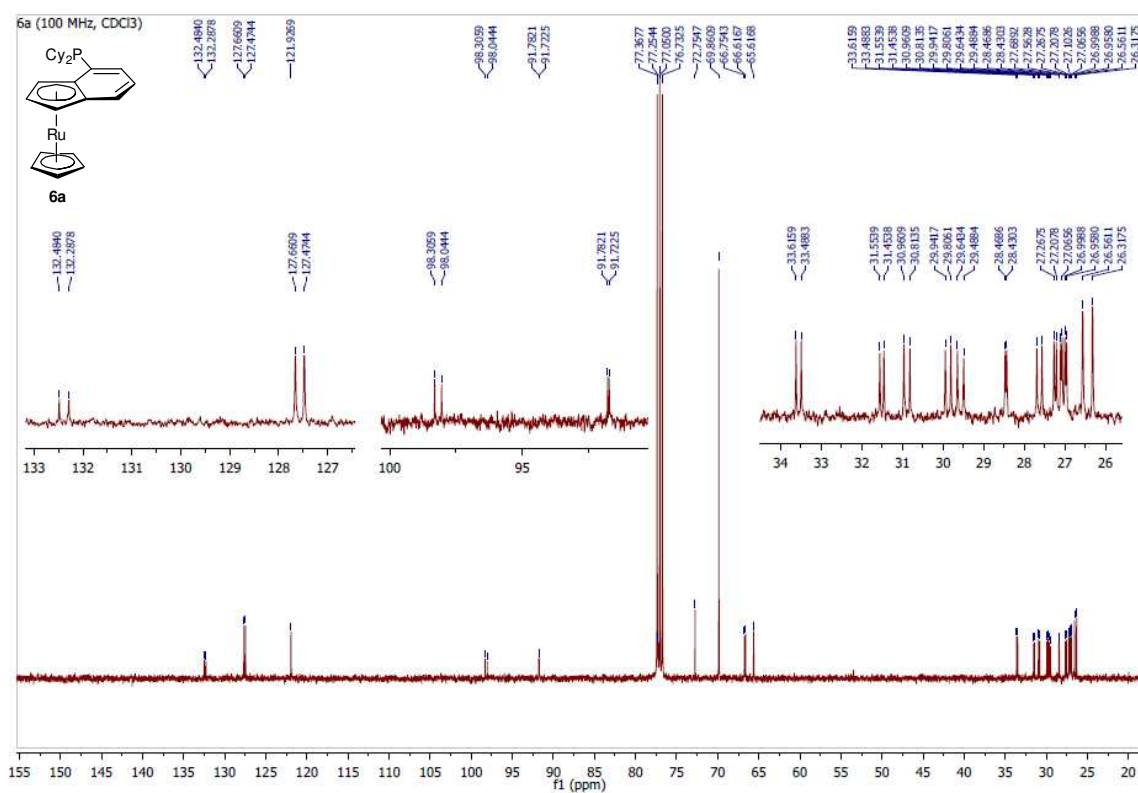
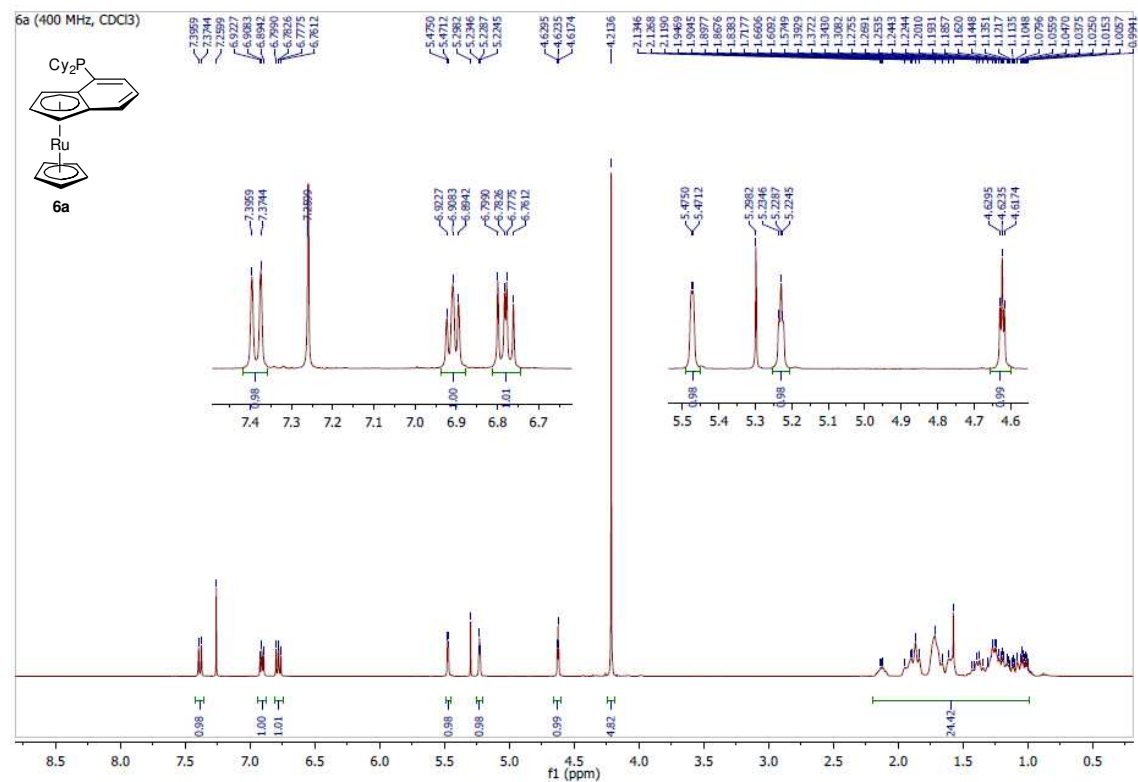
[a] Spectra recorded in CD_2Cl_2 at 125 MHz. [b] δ_{C} in ppm. [c] Coupling constants in Hertz (Hz).

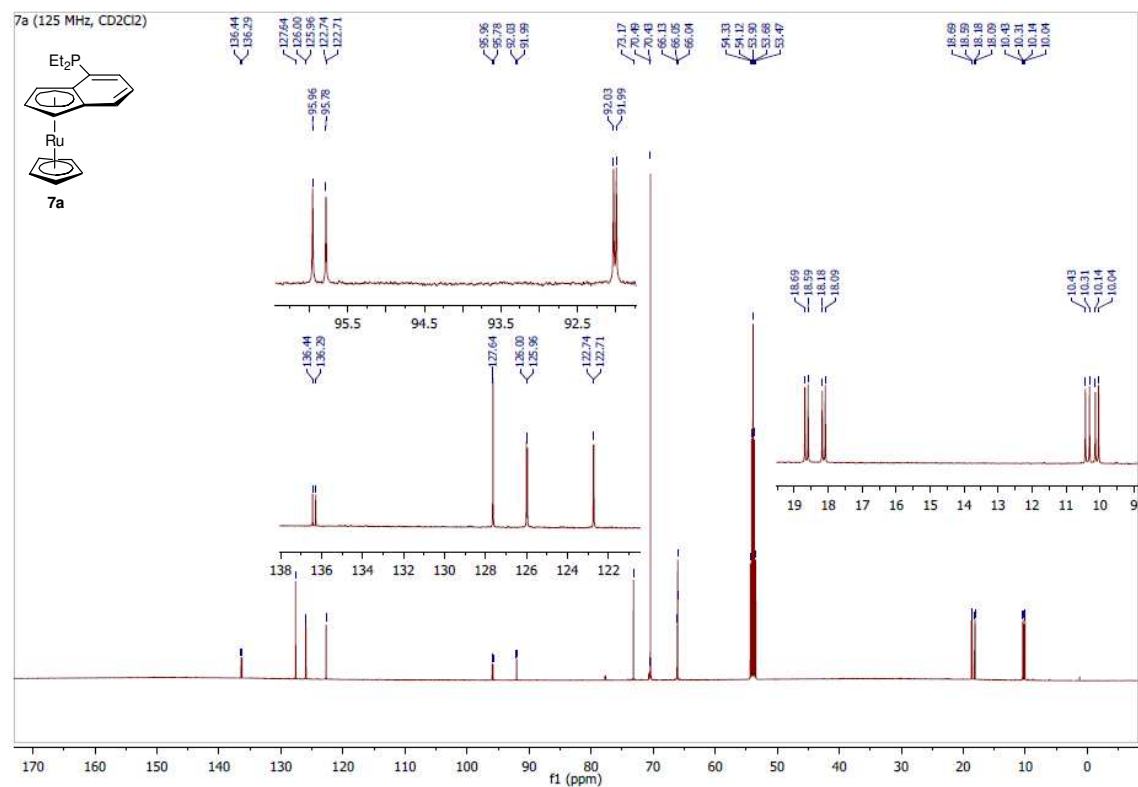
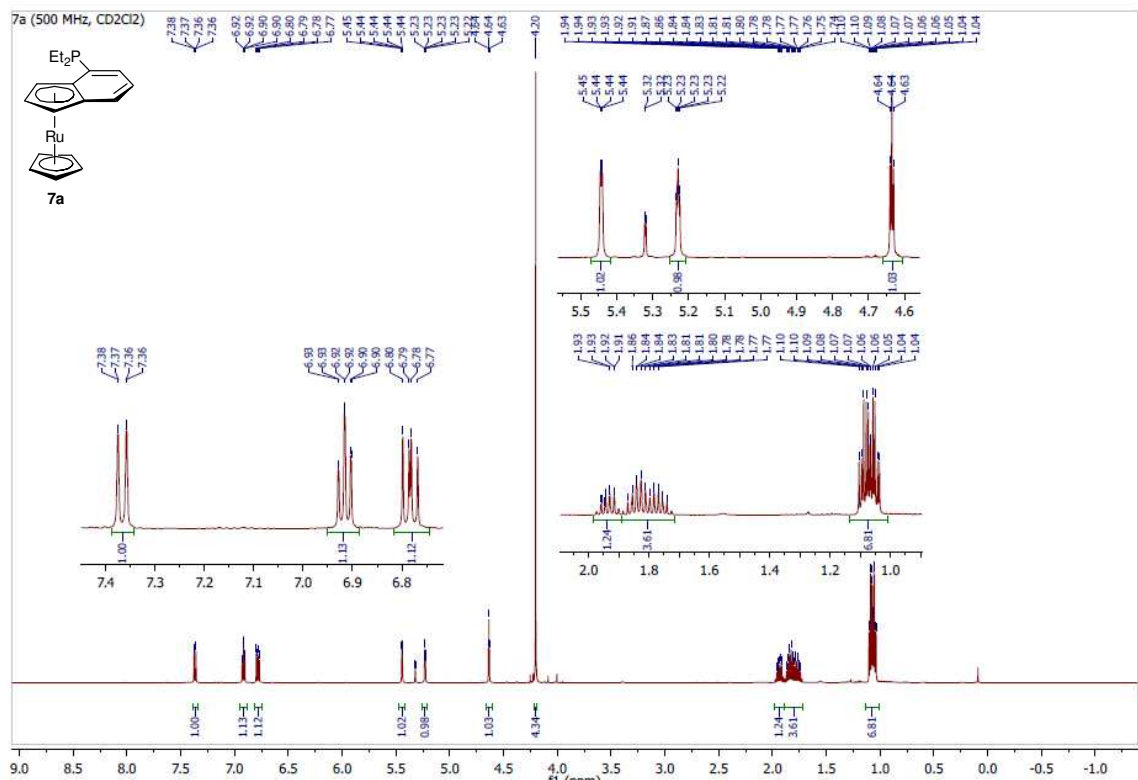
[d] Spectrum recorded in C_6D_6 at 125 MHz.

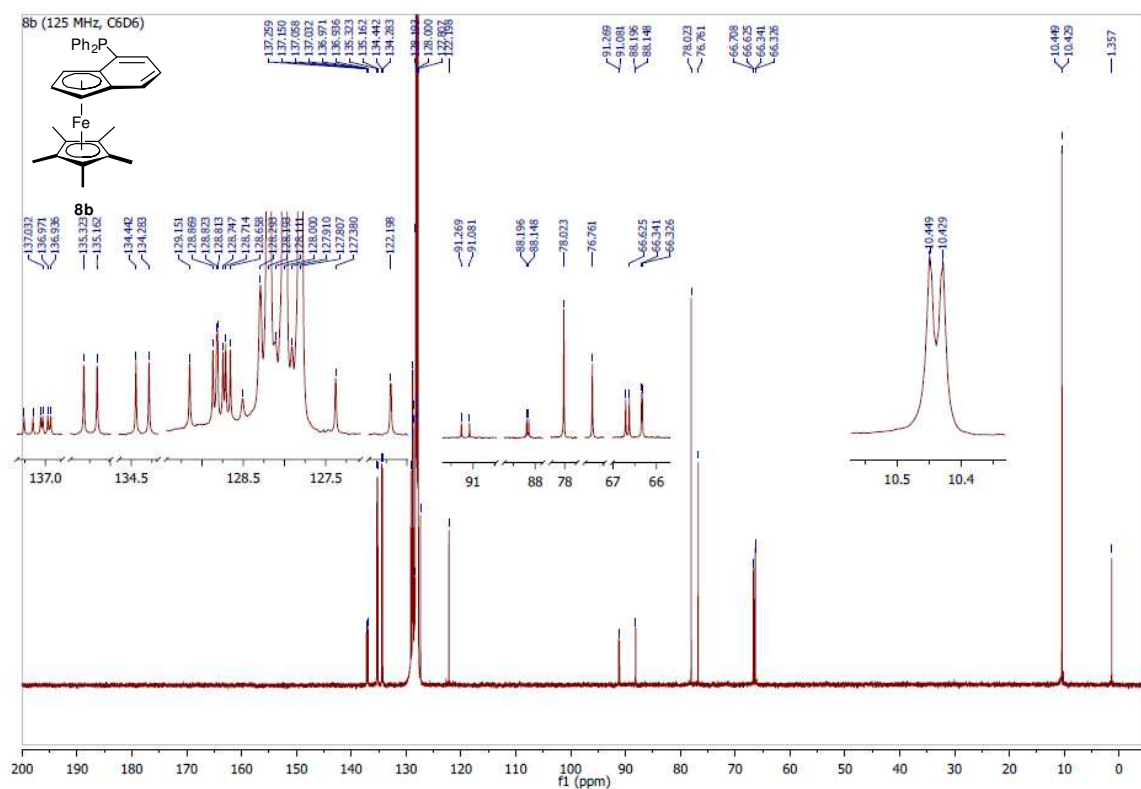
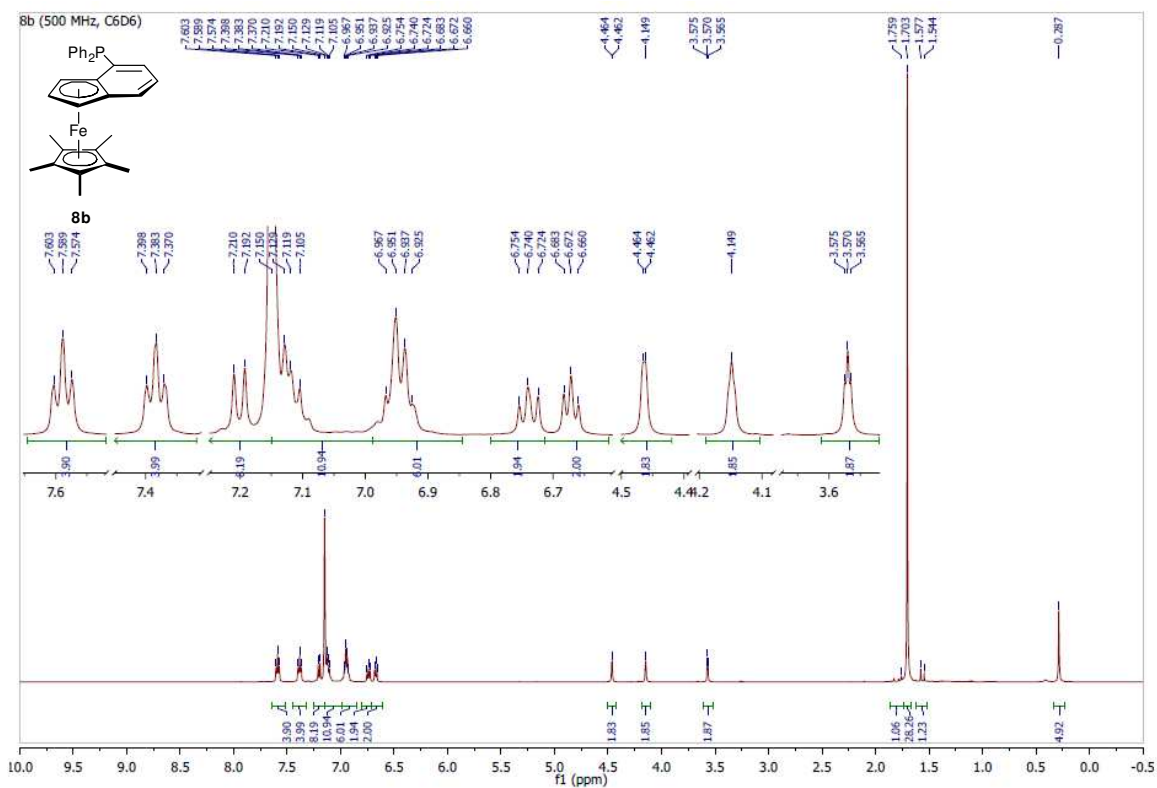
3. NMR Spectra

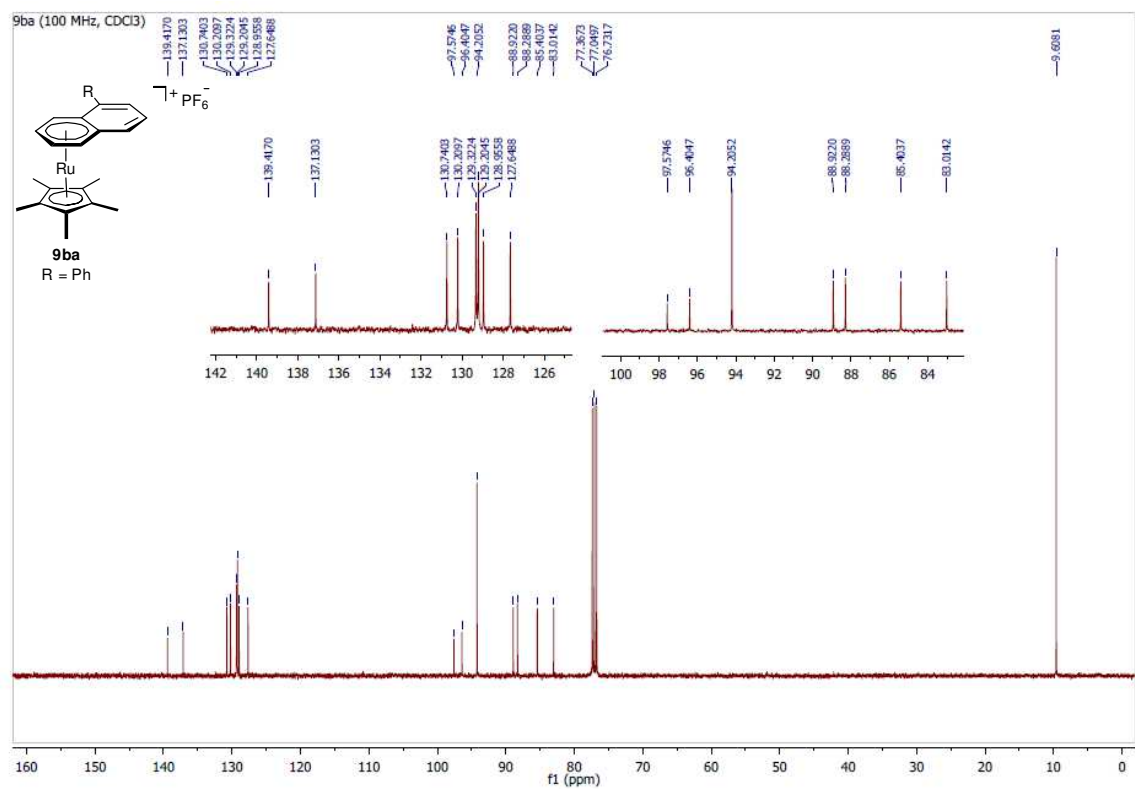
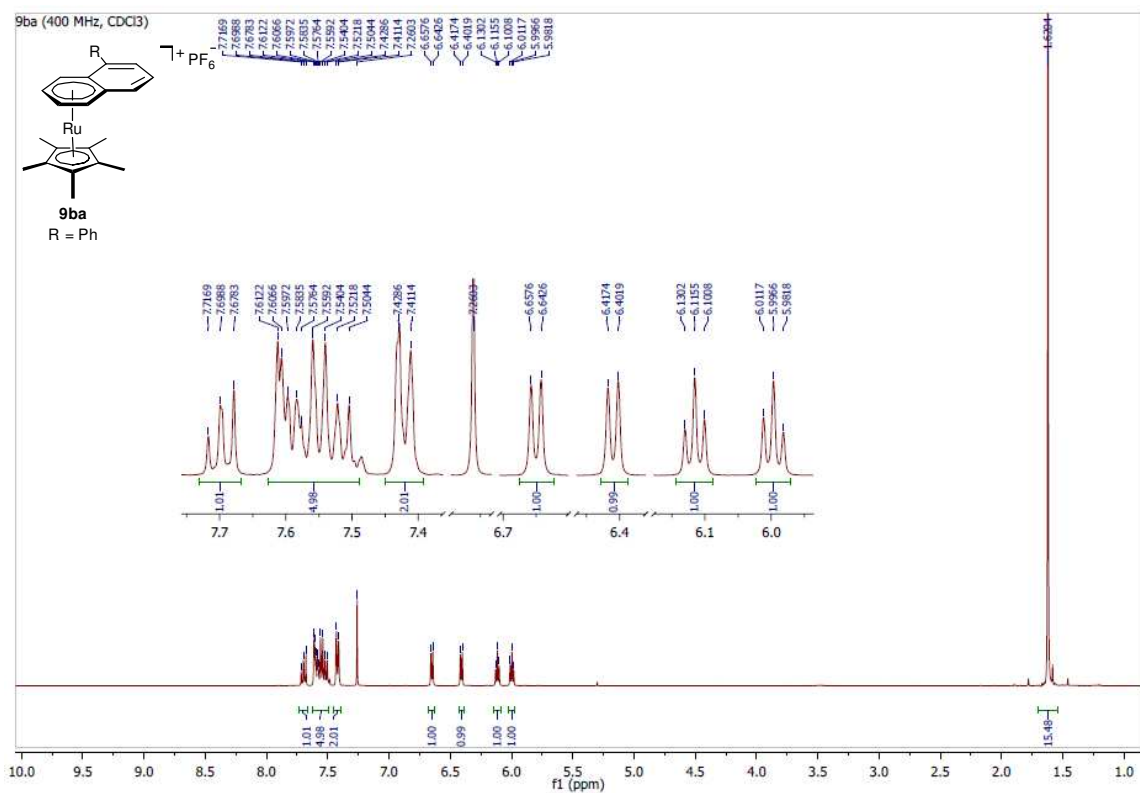


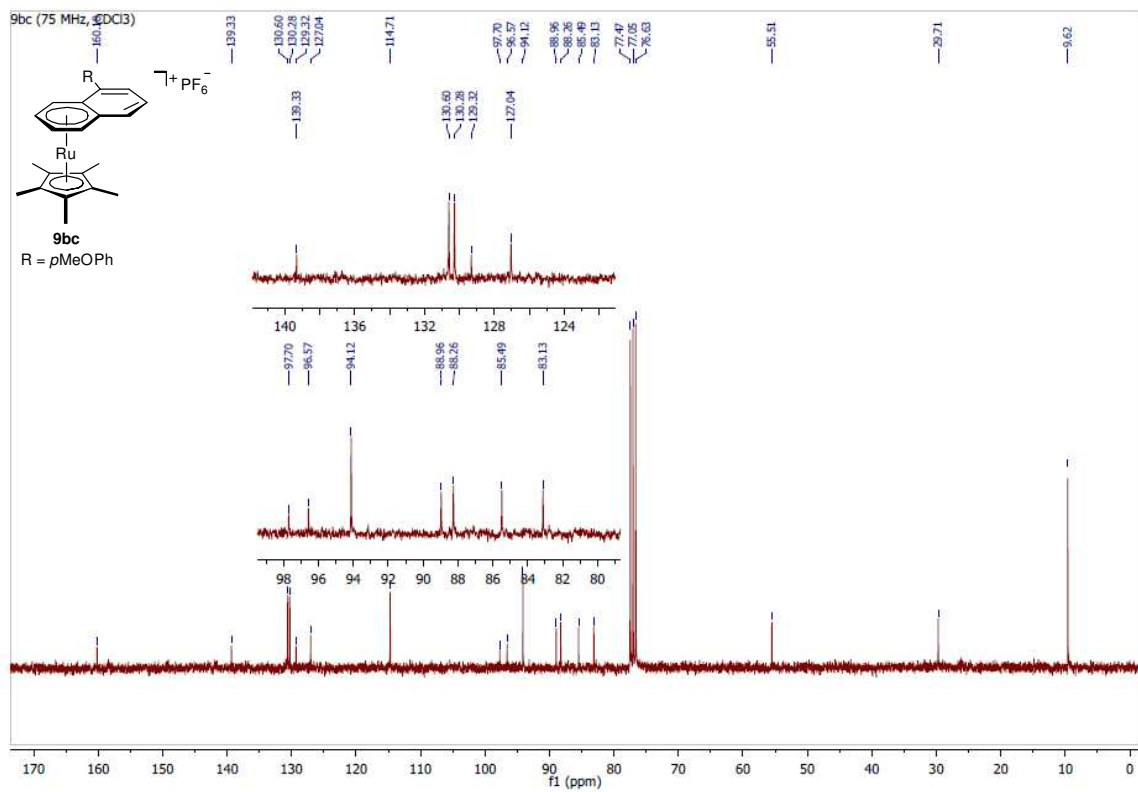
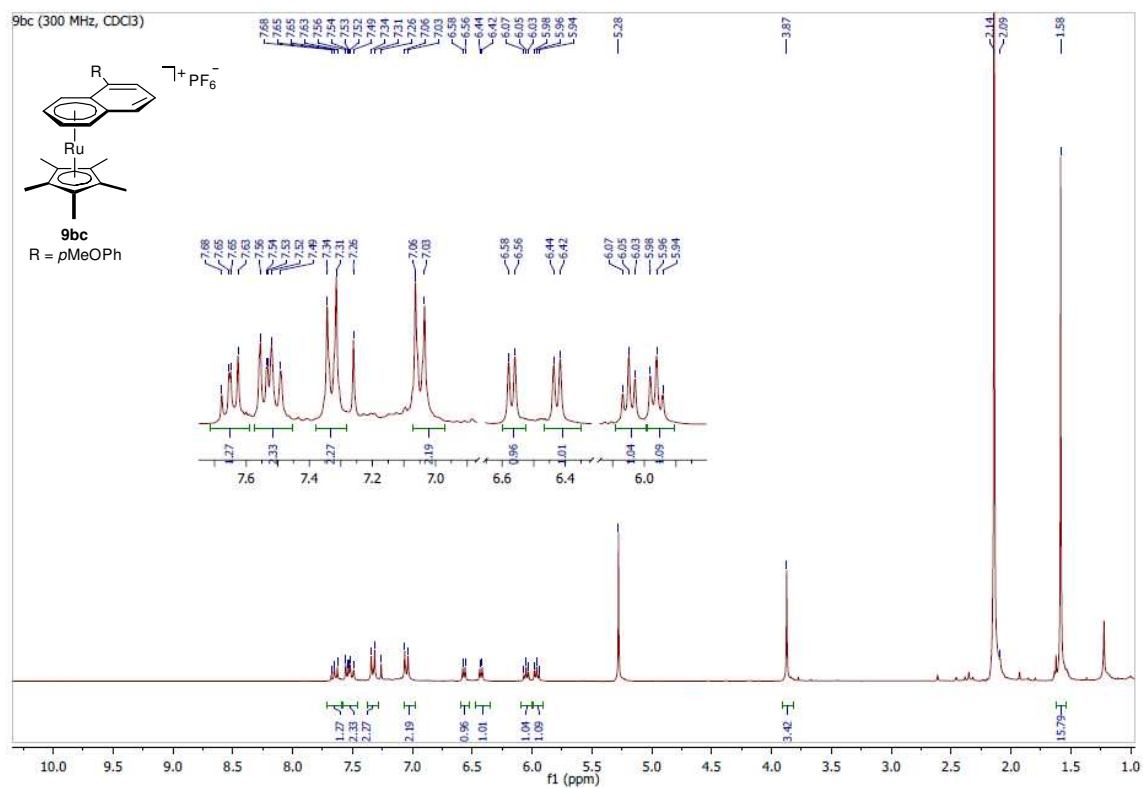


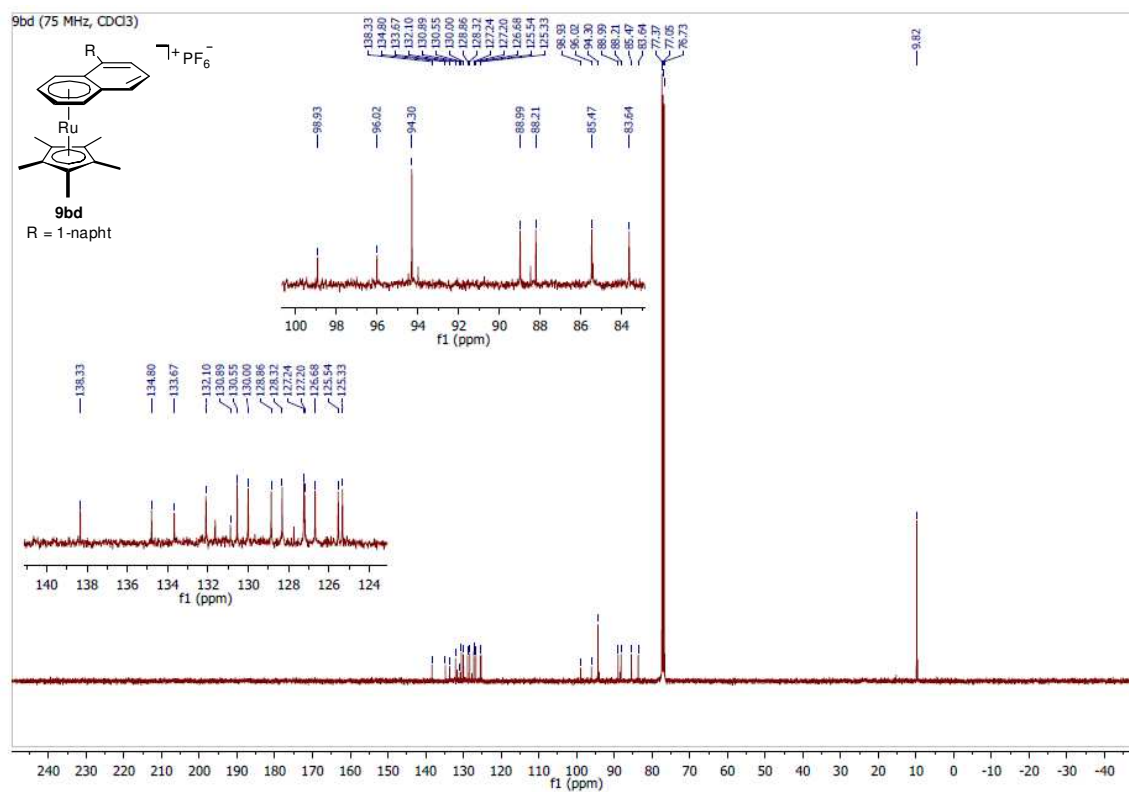
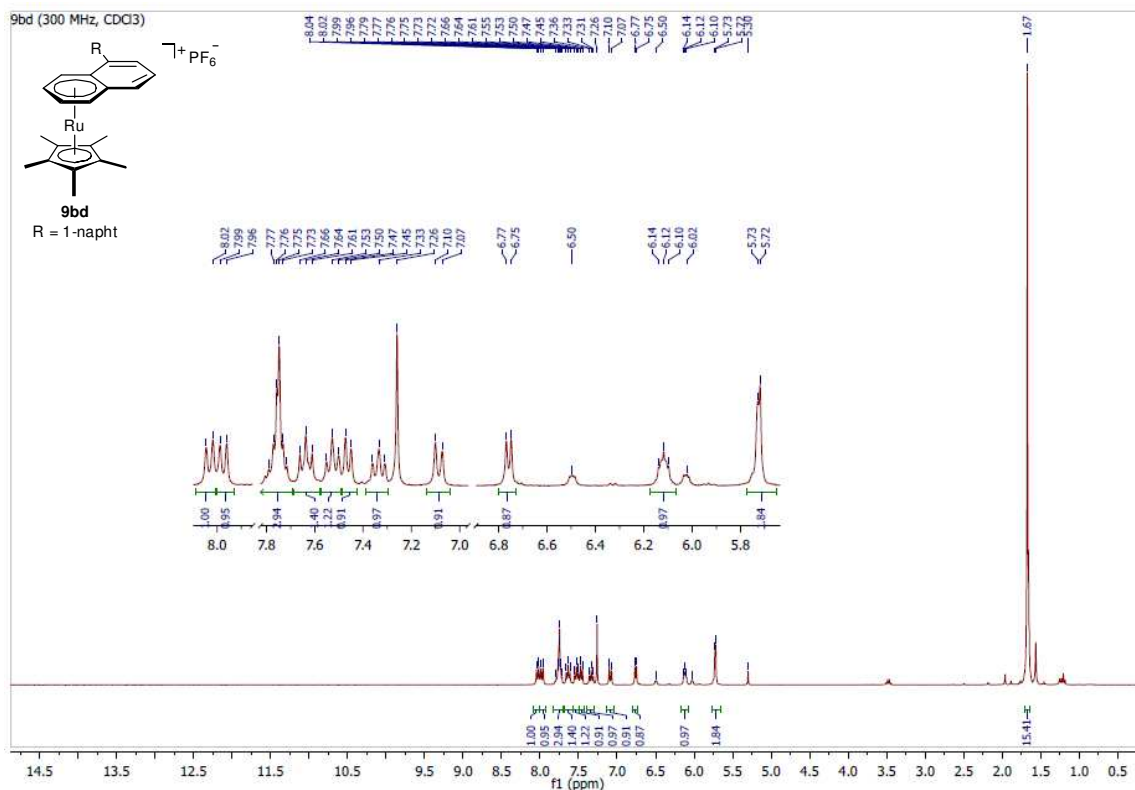


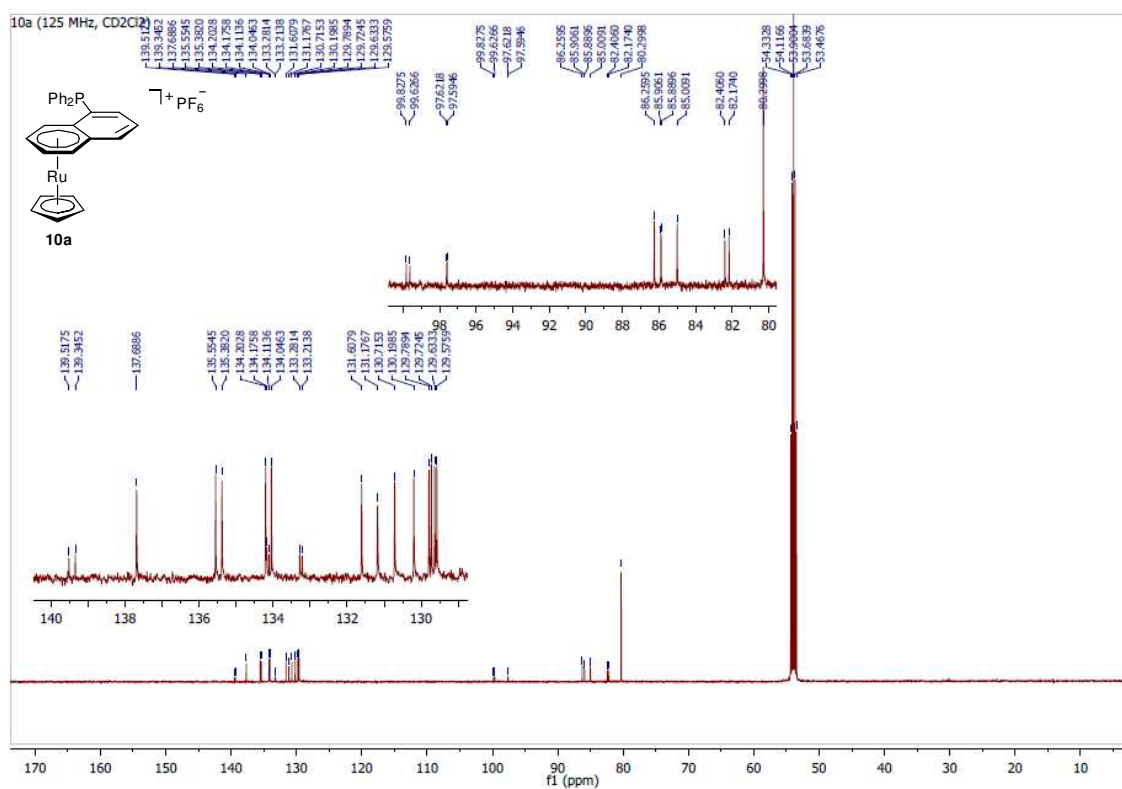
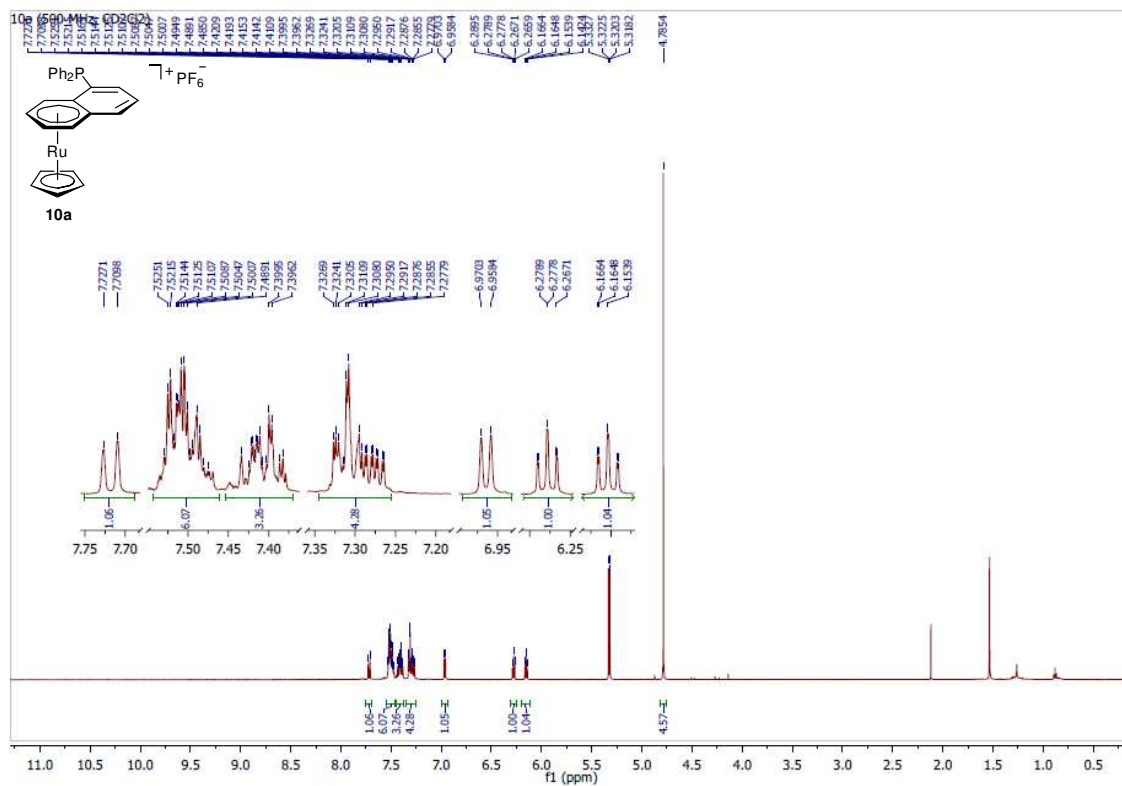


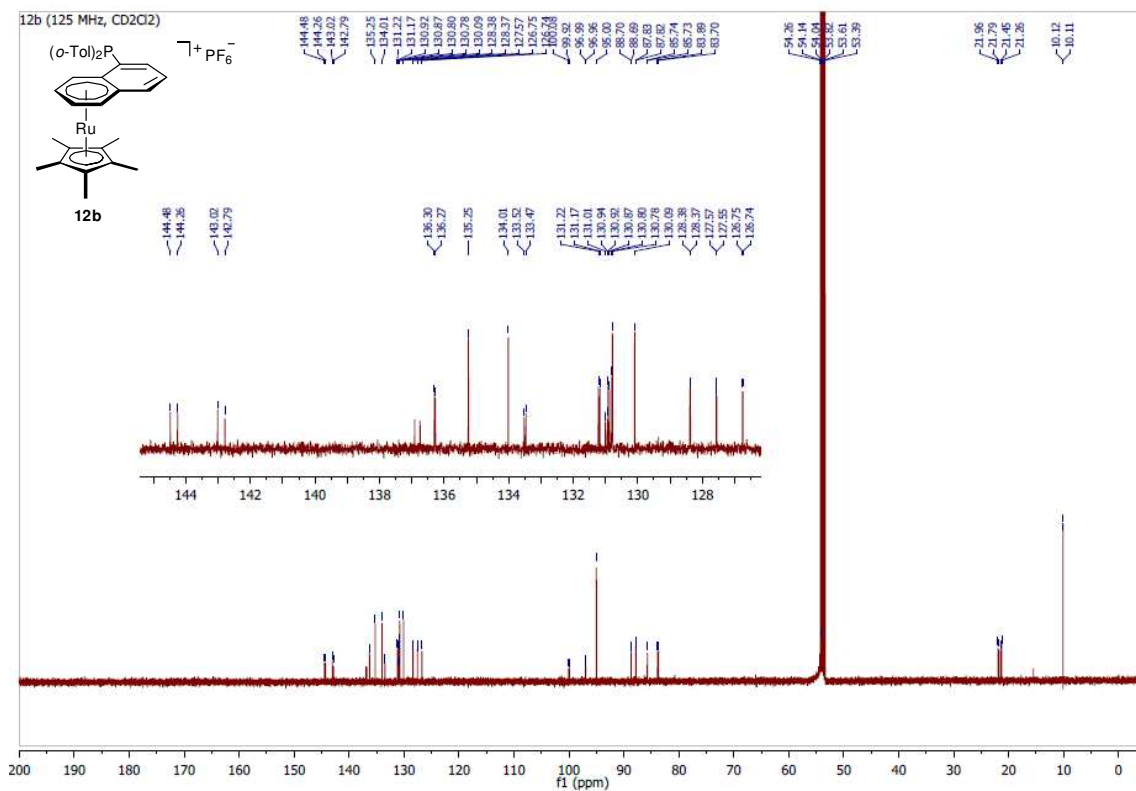
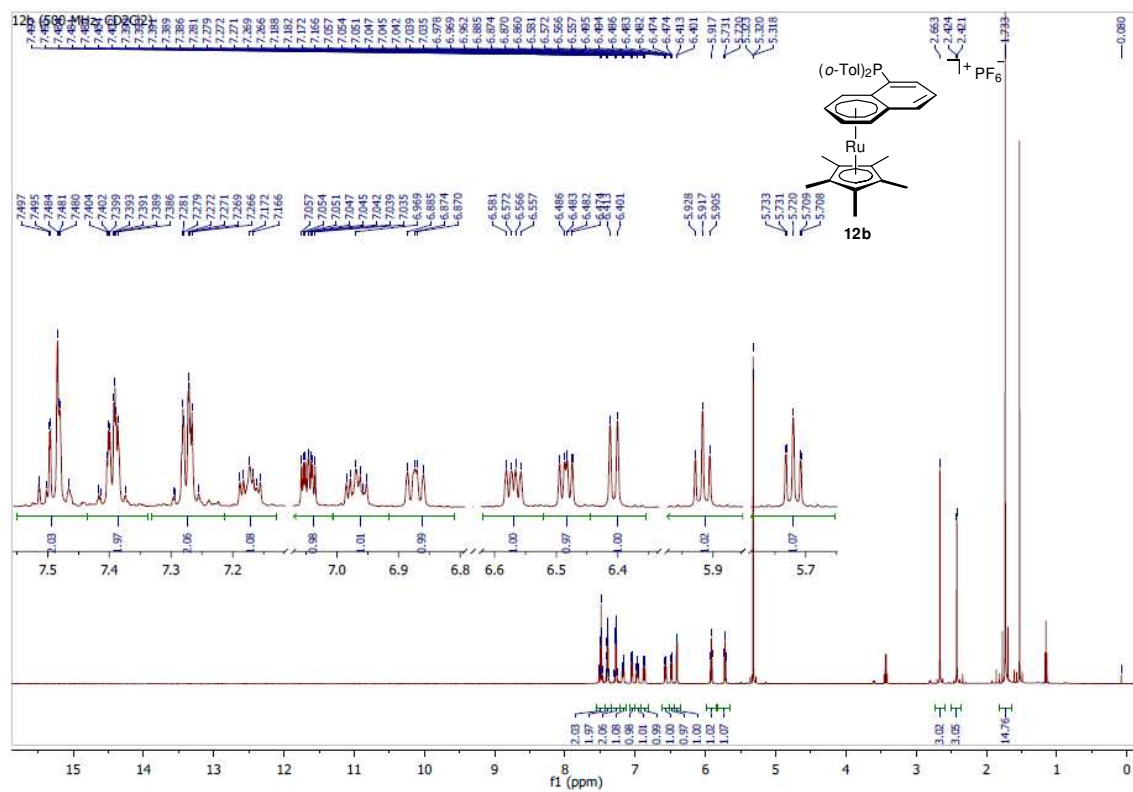


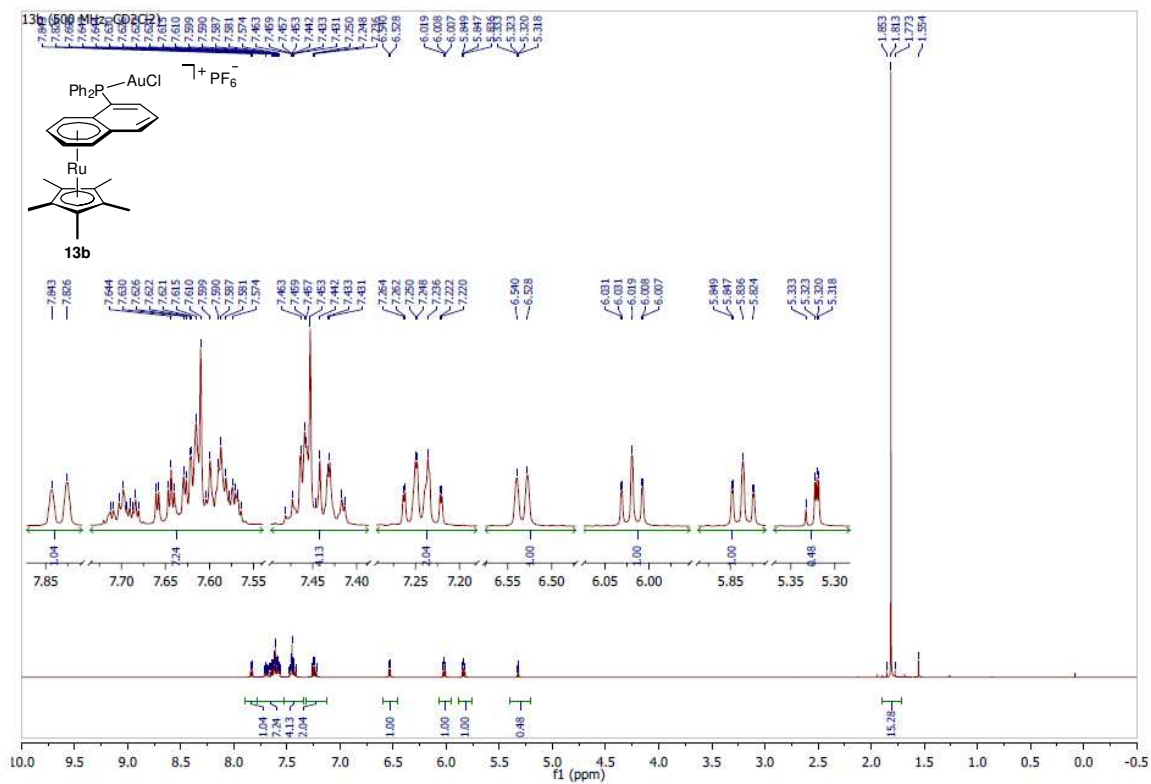












4. References

¹ Van der Sluis, P.; Spek, A. L. *Acta Cryst. A* **1990**, *46*, 194–201.

² Spek, A. L. *Acta Cryst. A* **1990**, *46*, C34.