

Supporting Information

Design of Chiral Bis-Phosphoric Acid Catalyst Derived from (R)-3,3'-Di(2-hydroxy-3-arylphenyl)binaphthol: Catalytic Enantioselective Diels-Alder Reaction of α,β -Unsaturated Aldehydes with Amidodienes

Norie Momiyama,* Tohru Konno, Yuichi Furiya, Takeaki Iwamoto, and Masahiro Terada*

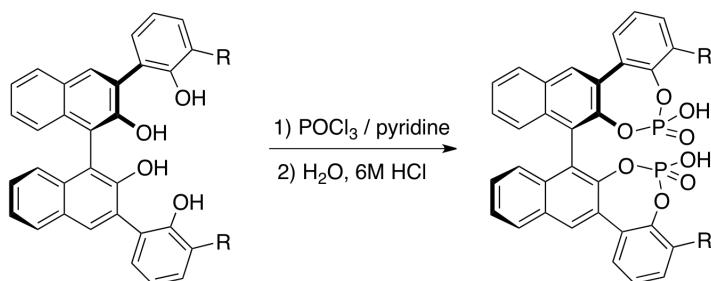
*Department of Chemistry, Graduate School of Science, Tohoku University,
6-3 Aoba Aramakiaza Aoba-ku, Sendai 980-8578, Japan*

Experimental Section.

General. All reactions were carried out under an atmosphere of standard grade nitrogen gas (oxygen <10 ppm) in flame-dried glassware with magnetic stirring. Toluene was used as anhydrous in solvent line system KANTO. α,β -Unsaturated aldehydes were distilled over calcium hydride before used. Purification of reaction products was carried out by flash column chromatography on silica gel 60 (spherical, neutral, 100–210 μm ; KANTO). Analytical thin layer chromatography (TLC) was performed on E. Merck precoated (0.25 mm) silica gel 60-F₂₅₄ plates. Visualization was accomplished with UV light and phosphomolybdic acid solution in ethanol by heating.

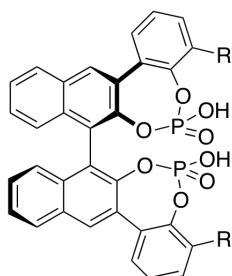
Infrared (IR) spectra were recorded on a Shimazu FTIR-8600PC spectrometer. ^1H NMR spectra were recorded on a JEOL JNM-A-500 (500 MHz) spectrometer at ambient temperature. NMR solvents were purchased from ACROS (CDCl_3), CIL, Inc. (DMSO-d_6), and used as received. Data are reported as follows: chemical shifts are reported in ppm from tetramethylsilane on the δ scale, with solvent resonance employed as internal standard (CDCl_3 7.26 ppm, DMSO-d_6 2.49 ppm), multiplicity (b = broad, s = singlet, d = doublet, t = triplet, q = quartet, and m = multiplet), integration, coupling constant (Hz) and assignment. ^{13}C NMR spectra were recorded on a JEOL JNM-A-500 (125.7 MHz) spectrometer at ambient temperature. Chemical shifts are reported in ppm from tetramethylsilane on the δ scale, with solvent resonance employed as internal standard (CDCl_3 77.0 ppm, DMSO-d_6 39.50 ppm). ^{31}P NMR spectra were recorded on a JEOL JNM-A-300 (121.5 MHz) spectrometer at ambient temperature. Chemical shifts are reported in ppm from 85% H_3PO_4 on the δ scale, with solvent resonance employed as internal standard (DMSO-d_6 0.4 ppm). High-performance liquid chromatography (HPLC) was performed on a Jasco equipped with a variable wavelength detector using Chiralcel OD-H column (0.46 cm x 25 cm), AD-3 column (0.46 cm x 25 cm), and IA column (0.46 cm x 25 cm) from Daicel. Optical rotations were measured on a Jasco P-1020 digital polarimeter with a sodium lamp and reported as follows; $[\alpha]_D^{T\ ^\circ\text{C}}$ ($c = \text{g}/100 \text{ mL}$, solvent). Mass spectra were obtained on Bruker Daltonics APEX III FT-ICR-MS spectrometer and HITACHI M-2500 spectrometer in Instrumental Analysis Center for Department of Chemistry, Graduate School of Science, Tohoku University.

General Procedure for Synthesis of Chiral Bis-Phosphoric Acid 1



Corresponding tetraphenol was synthesized based on reported procedure.¹ A flame-dried 20mL two-necked round bottom flask equipped with a magnetic stir bar was charged with tetraphenol (0.84 mmol, 1 equiv), and pyridine (8.0 mL) was added. To the resulting solution was added phosphorus oxychloride (4.2 mmol, 5 equiv) at room temperature, then stirred at 70 °C for 24 h. After cooling to room temperature, H₂O (8.0 mL) was added and the resulting suspension was further stirred at 70 °C for 12 h. The resulting mixture was diluted with CH₂Cl₂ (30 mL), followed by washed with 6N HCl (3×20 mL), and combined organic layers were concentrated under reduced pressure. The resultant solids were dissolved in MeOH (10 mL). To the resulting solution was added conc. HCl (5 mL) at room temperature, and stirred further at this temperature for 1 h. The mixture was extracted with CH₂Cl₂ (3×20 mL), and then, the organic layers were dried over Na₂SO₄ and concentrated under reduced pressure after filtration. The residual crude product was purified by column chromatography on silica gel (Silica gel 60 extra pure for column chromatography: Catalogue No. 107754 Merck KgaA) with elution by dichloromethane:methanol (100:1) to give metal- or HCl-free chiral phosphoric acid as white solid.*

***IMPORTANCE:** To exclude the contamination of metals and HCl, short pass column chromatography was conducted on Silica gel 60 extra pure (Catalogue No. 107754 Merck KgaA). The present purification procedure is valid to produce “metal- and/or HCl-free chiral phosphoric acid”. Details were reported in our recent publication “Metal-Free Chiral Phosphoric Acid or Chiral Metal Phosphate as Active Catalyst in the Activation of N-Acyl Aldimines” (*Synlett* **2011**, 1255-1258.)²



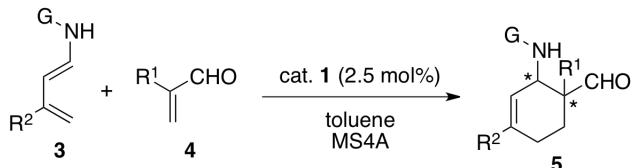
1e: R = 2,4,6-*i*-Pr₃C₆H₂

Chiral Bis-Phosphoric Acid 1e

White solid; TLC R_f = 0.48 (10:1 dichloromethane:methanol); IR (KBr) 3432(br), 2961, 2869, 1605, 1448, 1423, 1211, 1194, 1095, 996, 895 cm⁻¹; ¹H NMR (DMSO, 500 MHz) δ 8.43 (s, 1H, Ar-H), 8.14 (d, 1H, J = 8.1 Hz, Ar-H), 7.90 (d, 1H, J = 7.7 Hz, Ar-H), 7.52 (t, 1H, J = 7.5 Hz, Ar-H), 7.44 (t, 1H, J = 7.7 Hz, Ar-H), 7.38 (t, 1H, J = 7.7 Hz, Ar-H), 7.22

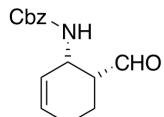
(d, 1H, $J = 7.7$ Hz, Ar-H), 7.16 (d, 1H, $J = 8.5$ Hz, Ar-H), 7.01 (s, 1H, Ar-H), 6.95 (s, 1H, Ar-H), 3.77 (bs, OH), 2.86 (m, 1H, CH), 2.69 (m, 1H, CH), 2.29 (m, 1H, CH), 1.21 (d, 6H, $J = 6.8$ Hz, CH_3), 1.06 (d, 6H, $J = 6.8$ Hz, CH_3), 0.98 (d, 3H, $J = 6.8$ Hz, CH_3), 0.82 (d, 3H, $J = 6.8$ Hz, CH_3); ^{13}C NMR ($CDCl_3$, 125.65 MHz) δ 147.1, 146.9, 146.5 (d, $J_{P,C} = 7.6$ Hz), 146.2, 144.8 (d, $J_{P,C} = 9.4$ Hz), 133.0, 132.6, 132.3, 132.0, 130.8, 130.5, 129.6, 129.3, 128.6, 127.1, 125.5, 125.4, 124.8, 122.6, 122.5, 120.7, 119.9, 33.4, 30.5, 30.1, 25.9, 24.6, 24.0, 23.9, 23.4, 23.0; ^{31}P NMR (DMSO, 121.5 MHz) δ -3.34; Anal. HRMS (ESI) Exact Mass Calcd. for $C_{62}H_{64}O_8P_2$ ([M-H] $^-$) 997.4004. Found: 997.4001.

General Procedure for Reactions of α,β -Unsaturated Aldehydes with Amidodiienes.

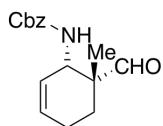


To the solution of catalyst (2.5 mol%, 0.005 mmol) and MS4A (150 mg) in toluene was added α,β -unsaturated aldehyde **4** (0.3 mmol) at room temperature, and the resulting solution was stirred at this temperature for 30 min. Then, amidodiene **3** (0.2 mmol) in toluene (0.5 mL) was added at -80 °C, and the resulting solution was stirred at this temperature for 48 h. The reaction mixture was quenched by saturated $NaHCO_3$ aq. (10 mL) and the aqueous layer was extracted with EtOAc (20 mLx3). The combined organic layers were washed with brine, dried over Na_2SO_4 and concentrated under reduced pressure after filtration. The residual crude product was chromatographed on silica gel using a mixture of ethyl acetate and hexane as the eluant to give the amido aldehyde **5**.

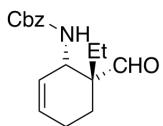
Benzyl (1*S*, 6*R*)-6-formylcyclohex-2-enylcarbamate (5aa)



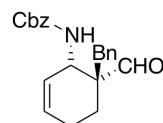
Purification by silica gel column chromatography with elution by hexane:ethyl acetate (12:1) provided as colorless oil. TLC $R_f = 0.40$ (4:1 hexane:ethyl acetate); $[\alpha]_D^{24} 111.1^\circ$ ($c = 1.11$, $CHCl_3$); IR (ATR) 3319, 3030, 2935, 2837, 1715, 1518, 1236, 1062 cm $^{-1}$; 1H NMR ($CDCl_3$, 500 MHz) δ (ppm): 9.81 (s, 1H, CHO), 7.37-7.30 (m, 5H, Ar-H), 5.86-5.84 (m, 1H, CH), 5.71-5.68 (m, 1H, CH), 5.11-5.03 (m, 3H, NH, CH_2), 4.74 (bs, 1H, CH), 2.79-2.77 (m, 1H, CH_2), 2.09-2.05 (m, 2H, CH_2), 2.00-1.96 (m, 1H, CH_2), 1.77-1.74 (m, 1H, CH_2); ^{13}C NMR ($CDCl_3$, 125.6 MHz) δ 202.5, 155.7, 136.1, 130.6, 128.4 (2C), 128.1, 128.0 (2C), 126.2, 66.9, 50.9, 45.5, 23.3, 18.5; Anal. HRMS (ESI) Exact Mass Calcd. for $C_{15}H_{17}NO_3$ ([M+Na] $^+$): 282.1106. Found: 282.1101. Enantiomeric excess was determined by HPLC analysis of the corresponding benzoylether with a Chiralcel OD-H column (95:5 hexane:ethanol), 0.6 mL/min; minor enantiomer $t_r = 24.1$ min, major enantiomer $t_r = 35.5$ min.

Benzyl (1*S*, 6*R*)-6-formyl-6-methylcyclohex-2-enylcarbamate (5ab)

Purification by silica gel column chromatography with elution by hexane:ethyl acetate (12:1) provided as colorless oil. TLC $R_f = 0.28$ (4:1 hexane:ethyl acetate); $[\alpha]_D^{24} 60.1^\circ$ ($c = 0.84$, CHCl_3); IR (ATR) 3324, 3031, 2931, 1720, 1500, 1455, 1325, 1234, 1132 cm^{-1} ; ^1H NMR (CDCl_3 , 500 MHz) δ 9.47 (s, 1H, CHO), 7.38-7.29 (m, 5H, Ar-H), 5.75-5.72 (m, 1H, CH), 5.62-5.59 (m, 1H, CH), 5.40 (d, 1H, $J = 9.5$ Hz, NH), 5.12 (d, 1H, $J = 12.0$ Hz, CH_2), 5.07 (d, 1H, $J = 12.0$ Hz, CH_2), 4.30 (d, 1H, $J = 10.0$ Hz, CH), 2.14-2.07 (m, 1H, CH_2), 2.01-1.89 (m, 2H, CH_2), 1.70-1.66 (m, 1H, CH_2), 1.17 (s, 3H, CH_3); ^{13}C NMR (CDCl_3 , 125.6 MHz) δ 204.2, 156.3, 136.4, 128.5, 128.1, 128.0, 127.4, 66.9, 51.8, 49.3, 31.6, 27.8, 22.6, 22.2, 18.8, 14.1; Anal. HRMS (ESI) Exact Mass Calcd. for $\text{C}_{16}\text{H}_{19}\text{NO}_3$ ([M+Na] $^+$): 296.1263. Found: 296.1257. Enantiomeric excess was determined by HPLC analysis of the corresponding benzoylether with a Chiralcel OD-H column (97:3 hexane:ethanol), 1.0 mL/min; minor enantiomer $t_r = 12.9$ min, major enantiomer $t_r = 23.8$ min.

Benzyl (1*S*, 6*R*)-6-ethyl-6-formylcyclohex-2-enylcarbamate (5ac)

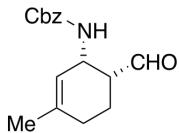
Purification by silica gel column chromatography with elution by hexane:ethyl acetate (12:1) provided as colorless oil. TLC $R_f = 0.30$ (4:1 hexane:ethyl acetate); $[\alpha]_D^{24} 101.8^\circ$ ($c = 1.00$, CHCl_3); IR (ATR) 3319, 2965, 2926, 1721, 1499, 1455, 1326, 1234, 1050 cm^{-1} ; ^1H NMR (CDCl_3 , 500 MHz) δ 9.52 (s, 1H, CHO), 7.37-7.28 (m, 5H, Ar-H), 5.77-5.75 (m, 1H, CH), 5.63-5.59 (m, 1H, CH), 5.26 (d, 1H, $J = 9.4$ Hz, NH), 5.11 (d, 1H, $J = 12.2$ Hz, CH_2), 5.05 (d, 1H, $J = 12.2$ Hz, CH_2), 4.35 (d, 1H, $J = 9.8$ Hz, CH), 2.01 (m, 2H, CH_2), 1.81-1.74 (m, 2H, CH_2), 1.72-1.62 (m, 1H, CH_2), 1.65-1.57 (m, 1H, CH_2), 0.91 (t, 3H, CH_3); ^{13}C NMR (CDCl_3 , 125.6 MHz) δ 205.0, 156.1, 136.3, 129.0, 128.5 (2C), 128.1, 128.0 (2C), 127.0, 66.9, 52.6, 50.0, 24.3, 22.6, 21.9, 8.2; Anal. HRMS (ESI) Exact Mass Calcd. for $\text{C}_{17}\text{H}_{21}\text{NO}_3$ ([M+Na] $^+$): 310.1419. Found: 310.1413. Enantiomeric excess was determined by HPLC analysis of the corresponding benzoylether with a Chiralcel OD-H column (97:3 hexane:ethanol), 1.0 mL/min; minor enantiomer $t_r = 12.1$ min, major enantiomer $t_r = 21.7$ min.

Benzyl (1*S*, 6*R*)-6-benzyl-6-formylcyclohex-2-enylcarbamate (5ad)

Purification by silica gel column chromatography with elution by hexane:ethyl acetate (12:1) provided as colorless oil. TLC $R_f = 0.35$ (4:1 hexane:ethyl acetate); $[\alpha]_D^{24} 25.7^\circ$ ($c = 1.29$, CHCl_3); IR (ATR) 3423, 3321, 3029, 2941, 2839,

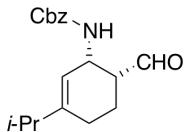
1718, 1496, 1455, 1323, 1055, 1026 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 9.54 (s, 1H, CHO), 7.37-7.15 (m, 10H, Ar-H), 5.72-5.70 (m, 1H, CH), 5.63-5.60 (m, 2H, CH, NH), 5.14 (d, 1H, *J* = 12.0 Hz, CH₂), 5.10 (d, 1H, *J* = 12.0 Hz, CH₂), 4.42 (d, 1H, *J* = 9.8 Hz, CH), 3.01 (d, 1H, *J* = 14.1 Hz, CH₂), 2.84 (d, 1H, *J* = 13.7 Hz, CH₂), 2.14-2.10 (m, 1H, CH₂), 1.96-1.92 (m, 2H, CH₂), 1.55-1.49 (m, 1H, CH₂); ¹³C NMR (CDCl₃, 125.6 MHz) δ 205.1, 156.2, 136.4, 135.1, 130.4 (2C), 128.5 (2C), 128.4 (2C), 128.1, 128.0 (2C), 127.8, 126.9, 66.9, 52.6, 50.5, 38.5, 25.0, 22.0(2C); Anal. HRMS (ESI) Exact Mass Calcd. for C₂₂H₂₃NO₃ ([M+Na]⁺): 372.1576. Found: 372.1568. Enantiomeric excess was determined by HPLC analysis of the corresponding benzoylether with a Chiralcel OD-H column (97:3 hexane:ethanol), 1.0 mL/min; minor enantiomer t_r = 15.3 min, major enantiomer t_r = 18.6 min

Benzyl (1*S*, 6*R*)-6-formyl-3-methylcyclohex-2-en-1-ylcarbamate (**5ba**)



Purification by silica gel column chromatography with elution by hexane:ethyl acetate (12:1) provided as colorless oil. TLC R_f = 0.24 (4:1 hexane:ethyl acetate); [α]_D²⁴ 100.0° (c = 1.20, CHCl₃); IR (ATR) 3318, 2931, 1694, 1517, 1454, 1376, 1224, 1067, 966 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 9.82 (s, 1H, CHO), 7.32-7.30 (m, 5H, Ar-H), 5.45-5.42 (m, 1H, CH), 5.10 (d, 1H, *J* = 8.2 Hz, CH₂), 5.04 (d, 1H, *J* = 8.2 Hz, CH₂), 4.96 (d, 1H, *J* = 8.1 Hz, CH), 4.72 (bs, 1H, NH), 2.69 (m, 1H, CH), 1.97-1.91 (m, 3H, CH₂), 1.85-1.72 (m, 1H, CH₂), 1.68 (s, 3H, CH₃); ¹³C NMR (CDCl₃, 125.6 MHz) δ 202.8, 155.8, 138.9, 136.3, 128.5 (2C), 128.1, 128.0 (2C), 120.6, 66.9, 50.9, 46.0, 28.4, 23.3, 18.6; Anal. HRMS (ESI) Exact Mass Calcd. for C₁₆H₁₉NO₃ ([M+Na]⁺): 296.1257. Found: 296.1256. Enantiomeric excess was determined by HPLC analysis of the corresponding benzoylether with a Chiralcel OD-H column (97:3 hexane:ethanol), 1.0 mL/min; minor enantiomer t_r = 14.8 min, major enantiomer t_r = 25.7 min.

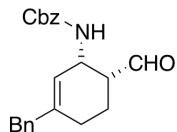
Benzyl (1*S*, 6*R*)-6-formyl-3-isopropylcyclohex-2-enylcarbamate (**5ca**)



Purification by silica gel column chromatography with elution by hexane:ethyl acetate (12:1) provided as colorless oil. TLC R_f = 0.34 (4:1 hexane:ethyl acetate); [α]_D²⁴ 95.6° (c = 1.28, CHCl₃); IR (ATR) 3324, 2959, 2871, 1715, 1517, 1455, 1330, 1227, 1064 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 9.82 (s, 1H, CHO), 7.37-7.26 (m, 5H, Ar-H), 5.43 (d, 1H, *J* = 4.25 Hz, CH), 5.11 (d, 1H, *J* = 12.4 Hz, CH₂), 5.04 (d, 1H, *J* = 12.4 Hz, CH₂), 4.95 (d, 1H, *J* = 8.5 Hz, NH), 4.75-4.72 (m, 1H, CH₂), 2.70 (d, 1H, *J* = 11.1 Hz, CH₂), 2.21-2.16 (m, 1H, CH), 2.05-1.93 (m, 3H, CH₂), 1.71-1.67 (m, 1H, CH₂), 0.99 (d, 6H, *J* = 6.8 Hz, CH₃); ¹³C NMR (CDCl₃, 125.6 MHz) δ 202.8, 155.7, 148.3, 136.2, 128.5(2C), 128.2, 128.1(2C), 118.0, 66.9, 51.3, 45.9, 34.8, 24.5, 21.3, 21.0, 18.7; Anal. HRMS (ESI) Exact Mass Calcd. for C₁₈H₂₃NO₃ ([M+Na]⁺): 324.1570. Found: 324.1570. Enantiomeric excess was determined by HPLC analysis of the corresponding benzoylether with a Chiralcel OD-H column (97:3 hexane:ethanol), 1.0 mL/min; minor enantiomer t_r = 20.4 min, major

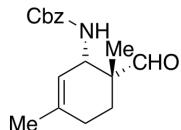
enantiomer $t_r = 30.9$ min.

Benzyl (1*S*, 6*R*)-3-benzyl-6-formylcyclohex-2-enylcarbamate (5da)



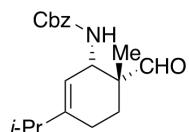
Purification by silica gel column chromatography with elution by hexane:ethyl acetate (12:1) provided as white solid. TLC $R_f = 0.26$ (4:1 hexane:ethyl acetate); $[\alpha]_D^{24} 88.6^\circ$ ($c = 0.90$, CHCl_3); IR (ATR) 3325, 3029, 2900, 2834, 1715, 1517, 1496, 1454, 1230, 1064 cm^{-1} ; ^1H NMR (CDCl_3 , 500 MHz) δ 9.79 (s, 1H, CHO), 7.39-7.26 (m, 7H, Ar-H), 7.23-7.19 (m, 1H, Ar-H), 7.11 (d, 2H, $J = 6.80$ Hz, Ar-H), 5.49 (d, 1H, $J = 4.3$ Hz, CH), 5.10 (d, 1H, $J = 12.0$ Hz, CH_2), 5.05-5.02 (m, 2H, CH_2NH), 4.76 (bs, 1H, CH_2), 3.27 (s, 2H, CH_2), 2.72 (d, 1H, $J = 4.7$ Hz, CH_2), 1.95-1.92 (m, 3H, CH_2), 1.74-1.69 (m, 1H, CH_2); ^{13}C NMR (CDCl_3 , 125.6 MHz) δ 202.6, 155.8, 141.8, 138.7, 136.2, 128.8(2C), 128.5(2C), 128.4(2C), 128.2, 128.1(2C), 126.4, 122.1, 67.0, 50.9, 46.1, 43.8, 26.4, 18.8; Anal. HRMS (ESI) Exact Mass Calcd. for $\text{C}_{22}\text{H}_{23}\text{NO}_3$ ($[\text{M}+\text{Na}]^+$): 372.1570. Found: 372.1569. Enantiomeric excess was determined by HPLC analysis of the corresponding benzoylether with a Chiralcel OD-H column (97:3 hexane:ethanol), 1.0 mL/min; minor enantiomer $t_r = 24.3$ min, major enantiomer $t_r = 42.5$ min.

Benzyl (1*S*, 6*R*)-6-formyl-3,6-dimethylcyclohex-2-enylcarbamate (5bb)



Purification by silica gel column chromatography with elution by hexane:ethyl acetate (12:1) provided as colorless oil. TLC $R_f = 0.30$ (4:1 hexane:ethyl acetate); $[\alpha]_D^{24} 72.1^\circ$ ($c = 1.57$, CHCl_3); IR (ATR) 3324, 2965, 2930, 2728, 1721, 1499, 1455, 1231, 1041 cm^{-1} ; ^1H NMR (CDCl_3 , 500 MHz) δ 9.49 (s, 1H, CHO), 7.37-7.30 (m, 5H, Ar-H), 5.33 (d, 1H, $J = 1.3$ Hz, NH), 5.28 (d, 1H, $J = 8.5$ Hz, CH), 5.11 (d, 1H, $J = 12.2$ Hz, CH_2), 5.06 (d, 1H, $J = 12.2$ Hz, CH_2), 4.26 (d, 1H, $J = 9.8$ Hz, CH), 1.99-1.87 (m, 3H, CH_2), 1.68-1.65 (m, 4H, CH_3CH_2), 1.15 (s, 3H, CH_3); ^{13}C NMR (CDCl_3 , 125.6 MHz) δ 204.5, 156.6, 136.5, 136.4, 128.5(2C), 128.1, 128.0(2C), 121.4, 66.8, 52.1, 49.0, 27.6, 27.0, 23.0, 18.5; Anal. HRMS (ESI) Exact Mass Calcd. for $\text{C}_{17}\text{H}_{21}\text{NO}_3$ ($[\text{M}+\text{Na}]^+$): 310.1414. Found: 310.1414. Enantiomeric excess was determined by HPLC analysis of the corresponding benzoylether with a Chiralcel OD-H column (97:3 hexane:ethanol), 1.0 mL/min; minor enantiomer $t_r = 14.7$ min, major enantiomer $t_r = 22.8$ min.

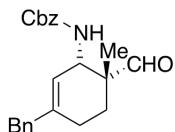
Benzyl (1*S*, 6*R*)-6-formyl-3-isopropyl-6-methylcyclohex-2-enylcarbamate (5cb)



Purification by silica gel column chromatography with elution by hexane:ethyl acetate (12:1) provided as colorless

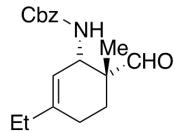
oil. TLC $R_f = 0.30$ (4:1 hexane:ethyl acetate); $[\alpha]_D^{24} 75.0^\circ$ ($c = 1.50$, CHCl₃); IR (ATR) 3439, 3327, 2960, 2930, 1719, 1498, 1455, 1229, 1037 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 9.49 (s, 1H, CHO), 7.38-7.26 (m, 5H, Ar-H), 5.33-5.29 (m, 2H, CH, NH), 5.12 (d, 1H, J = 12.2 Hz, CH₂), 5.06 (d, 1H, J = 12.2 Hz, CH₂), 4.28 (d, 1H, J = 9.0 Hz, CH), 2.20-2.14 (m, 1H, CH), 2.04-1.88 (m, 3H, CH₂), 1.68-1.63 (m, 1H, CH₂), 1.14 (s, 3H, CH₃), 0.97 (d, 6H, J = 6.8 Hz, CH₃); ¹³C NMR (CDCl₃, 125.6 MHz) δ 204.4, 156.3, 145.9, 136.4, 128.5 (2C), 128.1, 128.0 (2C), 118.9, 66.8, 52.2, 49.2, 34.5, 27.7, 22.9, 21.2, 21.1, 18.6; Anal. HRMS (ESI) Exact Mass Calcd for C₁₉H₂₅NO₃(M+Na)⁺: 338.1727. Found: 338.1726. Enantiomeric excess was determined by HPLC analysis of the corresponding benzoylether with a Chiralcel OD-H column (97:3 hexane:ethanol), 1.0 mL/min; minor enantiomer $t_r = 7.3$ min, major enantiomer $t_r = 11.2$ min.

Benzyl (1*S*, 6*R*)-3-benzyl-6-formyl-6-methylcyclohex-2-enylcarbamate (5db)



Purification by silica gel column chromatography with elution by hexane:ethyl acetate (12:1) provided as colorless oil. TLC $R_f = 0.27$ (4:1 hexane:ethyl acetate); $[\alpha]_D^{24} 54.8^\circ$ ($c = 1.73$, CHCl₃); IR (ATR) 3324, 3027, 2927, 2834, 2721, 1718, 1495, 1454, 1226, 1028 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 9.44 (s, 1H, CHO), 7.38-7.27 (m, 7H, Ar-H), 7.20 (t, 1H, J = 7.3 Hz, Ar-H), 7.11 (d, 2H, J = 7.3 Hz, Ar-H), 5.43 (s, 1H, NH), 5.40 (d, 1H, J = 9.8 Hz, CH), 5.12 (d, 1H, J = 12.4 Hz, CH₂), 5.07 (d, 1H, J = 12.4 Hz, CH₂), 4.33 (d, 1H, J = 9.4 Hz, CH), 3.26 (d, 1H, J = 15.2 Hz, CH₂), 3.22 (d, 1H, J = 15.2 Hz, CH₂), 1.93-1.80 (m, 3H, CH₂), 1.66-1.60 (m, 1H, CH₂), 1.15 (s, 3H, CH₃); ¹³C NMR (CDCl₃, 125.6 MHz) δ 204.3, 156.3, 139.3, 138.9, 136.4, 128.7 (2C), 128.5 (2C), 128.4 (2C), 128.1, 128.0 (2C), 126.3, 123.3, 66.9, 52.3, 49.2, 43.6, 28.0, 25.0, 18.6; Anal. HRMS (ESI) Exact Mass Calcd for C₂₃H₂₅NO₃(M+Na)⁺: 386.1727. Found: 386.1726. Enantiomeric excess was determined by HPLC analysis of the corresponding benzoylether with a Chiralcel OD-H column (97:3 hexane:ethanol), 1.0 mL/min; minor enantiomer $t_r = 16.5$ min, major enantiomer $t_r = 35.4$ min.

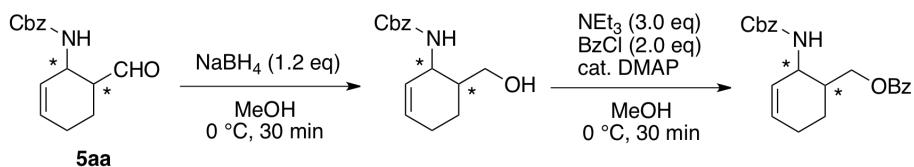
Benzyl (1*S*, 6*R*)-3-ethyl-6-formyl-6-methylcyclohex-2-enylcarbamate (5eb)



Purification by silica gel column chromatography with elution by hexane:ethyl acetate (12:1) provided as colorless oil. TLC $R_f = 0.28$ (4:1 hexane:ethyl acetate); $[\alpha]_D^{24} 78.4^\circ$ ($c = 1.57$, CHCl₃); IR (ATR) 3324, 2964, 2932, 1719, 1499, 1456, 1229, 1037 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 9.50 (s, 1H, CHO), 7.38-7.27 (m, 5H, Ar-H), 5.32-5.28 (m, 2H, CH, NH), 5.12 (d, 1H, J = 12.2 Hz, CH₂), 5.06 (d, 1H, J = 12.2 Hz, CH₂), 4.28 (d, 1H, J = 9.8 Hz, CH), 1.95 (m, 5H, CH₂), 1.69-1.63 (m, 1H, CH₂), 1.15 (s, 3H, CH₃), 0.98 (t, 3H, J = 7.5 Hz, CH₃); ¹³C NMR (CDCl₃, 125.6 MHz) δ 204.5, 156.3, 141.8, 136.4, 128.5 (2C), 128.1, 128.0 (2C), 119.7, 66.8, 52.2, 49.1, 29.6, 27.7, 25.3, 18.6, 12.0; Anal. HRMS (ESI) Exact Mass Calcd for C₁₈H₂₃NO₃(M+Na)⁺: 324.1570. Found: 324.1570. Enantiomeric excess was determined by HPLC analysis of the corresponding benzoylether with a Chiralcel OD-H column (97:3 hexane:ethanol), 1.0

mL/min; minor enantiomer $t_r = 7.9$ min, major enantiomer $t_r = 12.8$ min.

Representative Procedure for Derivatization of Amido Aldehyde to Benzoylether.



To the solution of **5aa** (22.5 mg, 0.10 mmol) in MeOH (2.0 mL) was added NaBH₄ (45.0 mg, 0.12 mmol) at 0 °C. The resulting mixture was stirred at this temperature for 30 min. The reaction mixture was quenched by water, then the aqueous layer was extracted with EtOAc (10 mLx3). The combined organic extracts were washed with brine, dried over Na₂SO₄ and concentrated under reduced pressure after filtration, used in next step without purification.

To the solution of amido alcohol and 4-dimethylaminopyridine (1.0 mg) in CH₂Cl₂ (2.0 mL) was added triethylamine (42 μL, 0.30 mmol) and benzoyl chloride (23.0 μL, 0.20 mmol) at room temperature. The resulting solution was stirred at room temperature for 3 h. The reaction mixture was diluted with CH₂Cl₂ (5.0 mL) and quenched by water. The aqueous layer was extracted with CH₂Cl₂ (10 mLx3), the combined organic layers were washed with brine, dried over Na₂SO₄ and concentrated under reduced pressure after filtration. The residual crude product was chromatographed on silica gel using a mixture of ethyl acetate and hexane as the eluant to give the product as a white solid.

Determination of Absolute Configuration.

Benzyl (1*R*, 6*S*)-6-formylcyclohex-2-enylcarbamate **5aa** was synthesized based on reported procedure.³ Enantiomeric excess of **5aa** was determined by HPLC analysis after derivatization to the corresponding benzoylether based on above procedure. Absolute configuration was determined by comparison with HPLC analysis of reported benzyl (1*R*, 6*S*)-6-formylcyclohex-2-enylcarbamate **5aa**. Absolute configuration of other products was determined by analogy.

Reference

- (1) Ishihara, K.; Kurihara, H.; Matsumoto, M.; Yamamoto, H. *J. Am. Chem. Soc.* **1998**, *120*, 6920-6930.
- (2) Terada, M.; Kanomata, K. *Synlett* **2011**, 1255-1258.
- (3) Wipf, P.; Wang, X. *Tetrahedron Lett.* **2000**, *41*, 8747-8751.

DPFLE bis-phosphoric acid TriPals

Fri Sep 24 22:13:59 2010

1H

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500.00 MHz

0.00 KHz

16241.00 Hz

16334

6997.90 Hz

32

2.3413 sec

ACQTIME

PD

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1H

6.75 sec

IRNUC

1H

26.2 c

CTEMP

DMSO

POINT

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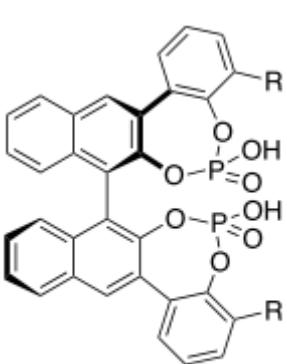
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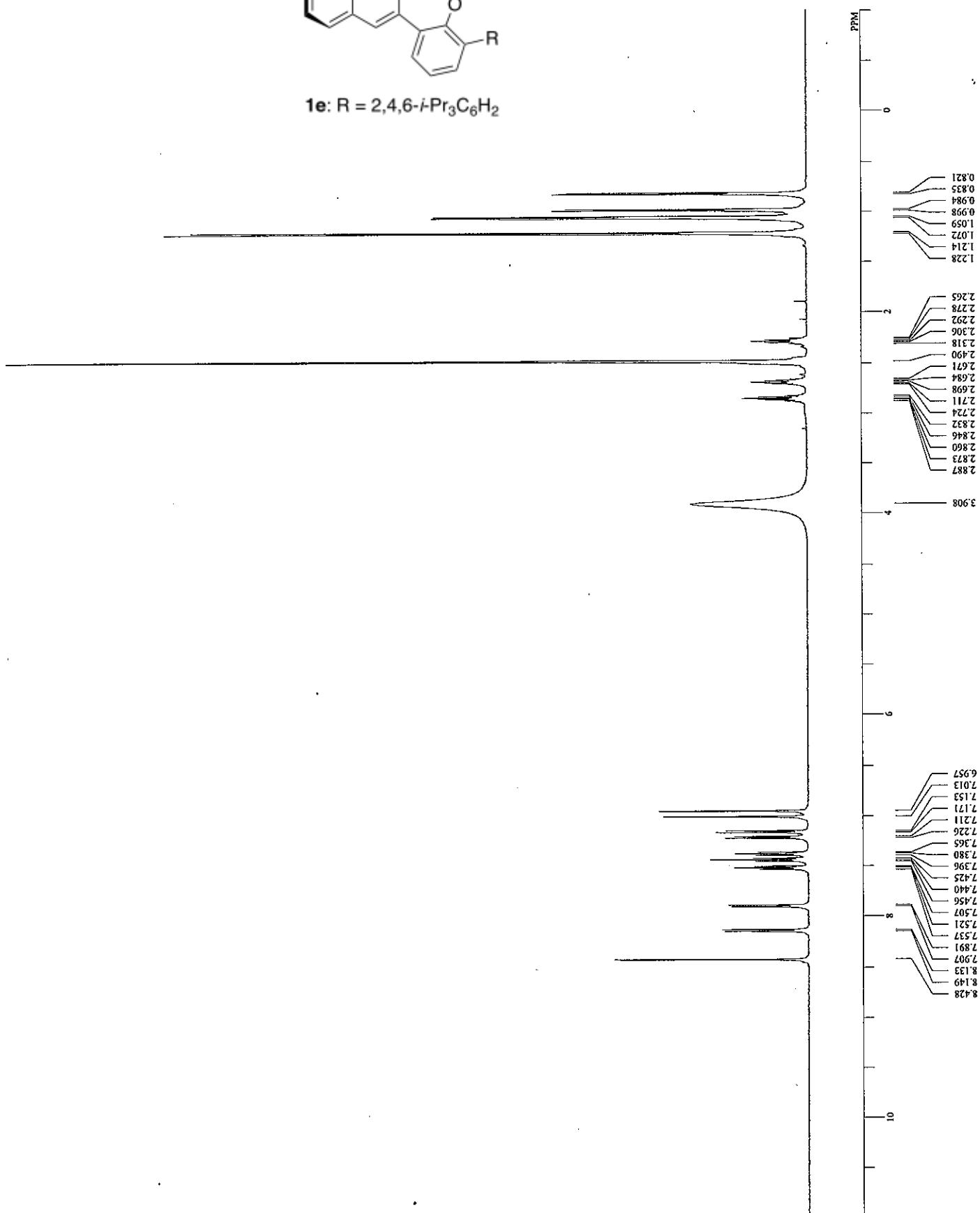
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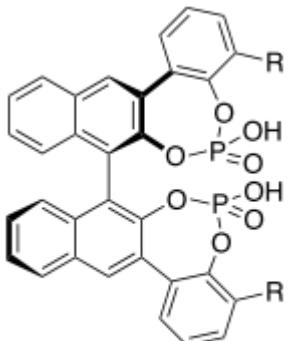
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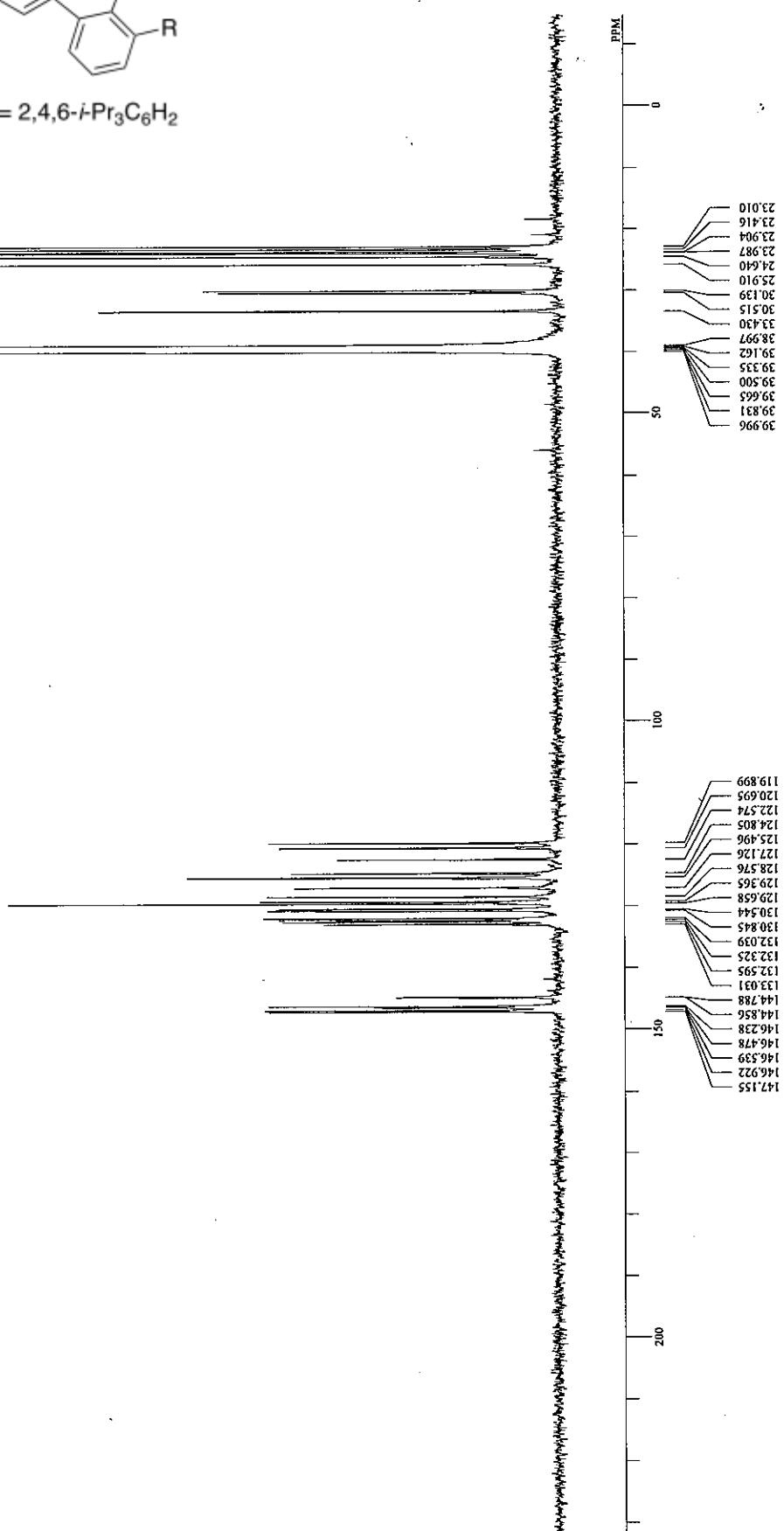
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DATIM



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DPFLE bis-phosphoric acid TriPals

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0.00 KHz

16241.00 Hz

16334

6997.90 Hz

32

2.3413 sec

ACQTIME

PD

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1H

6.75 sec

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1H

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CTEMP

DMSO

POINT

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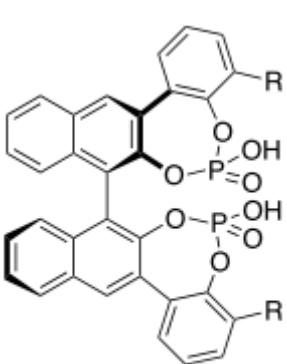
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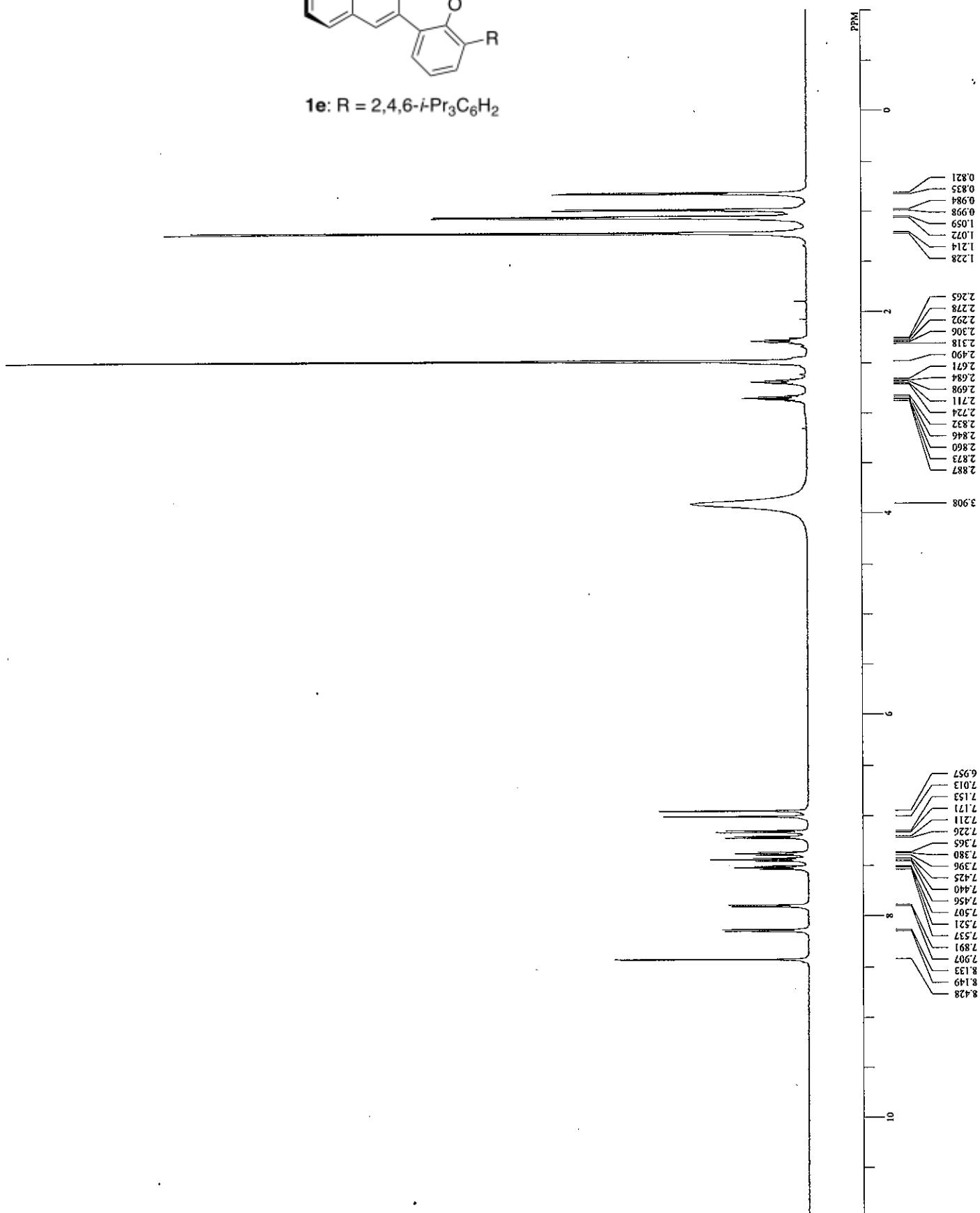
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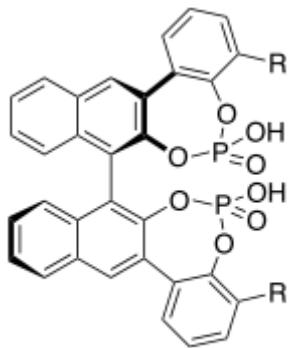


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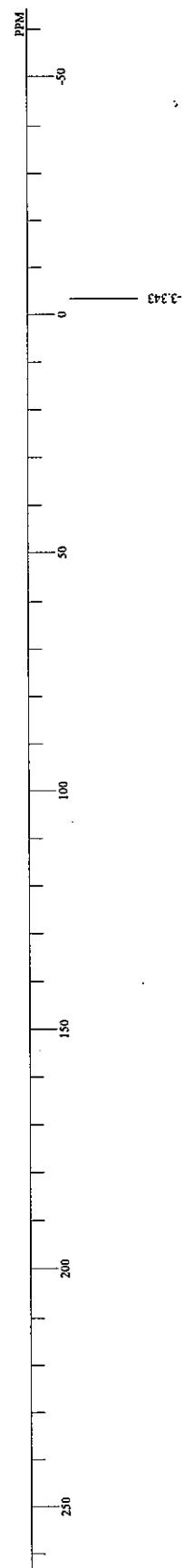


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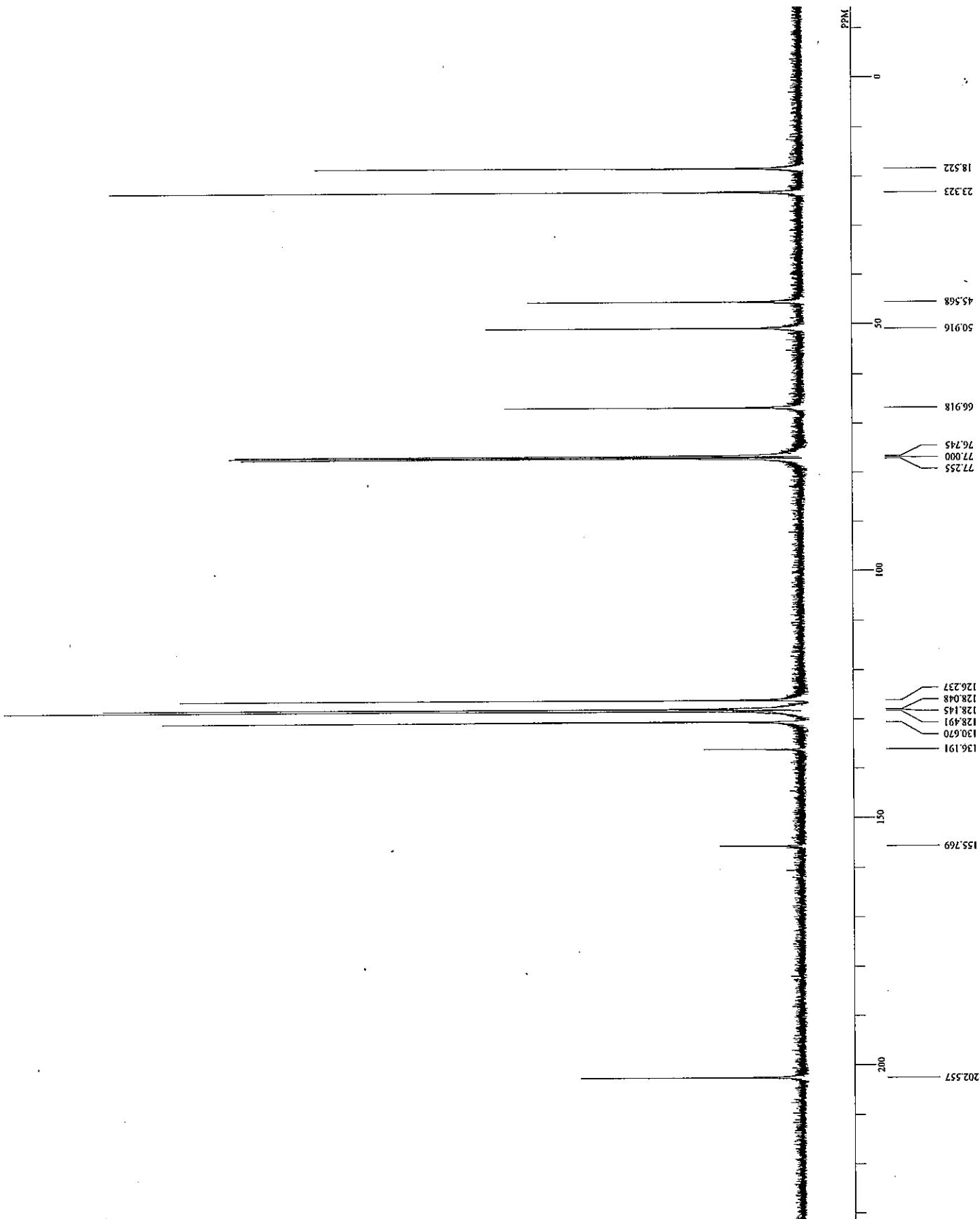
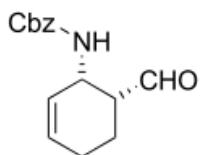
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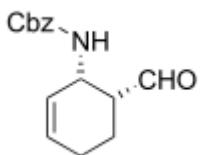
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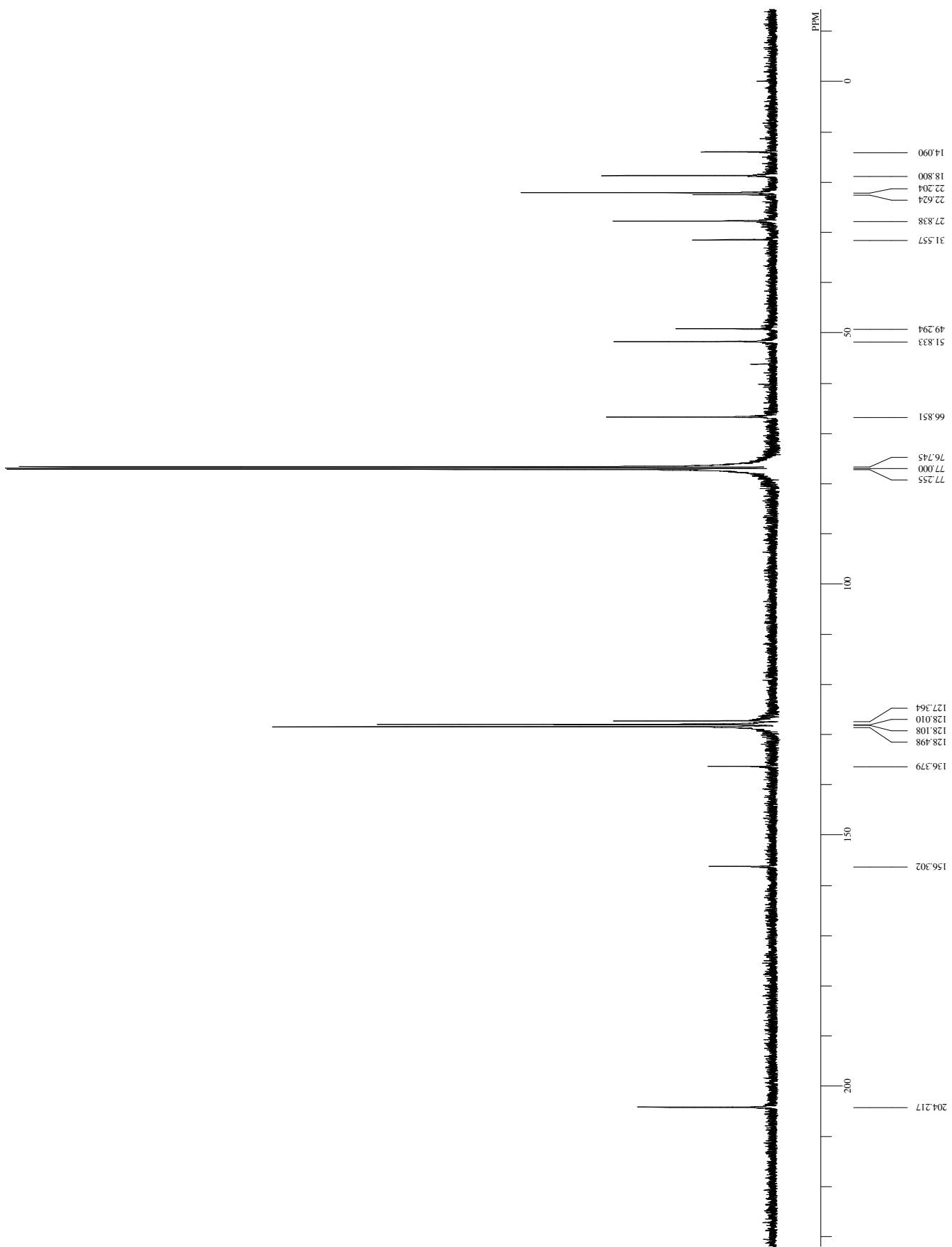
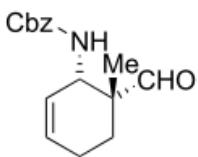
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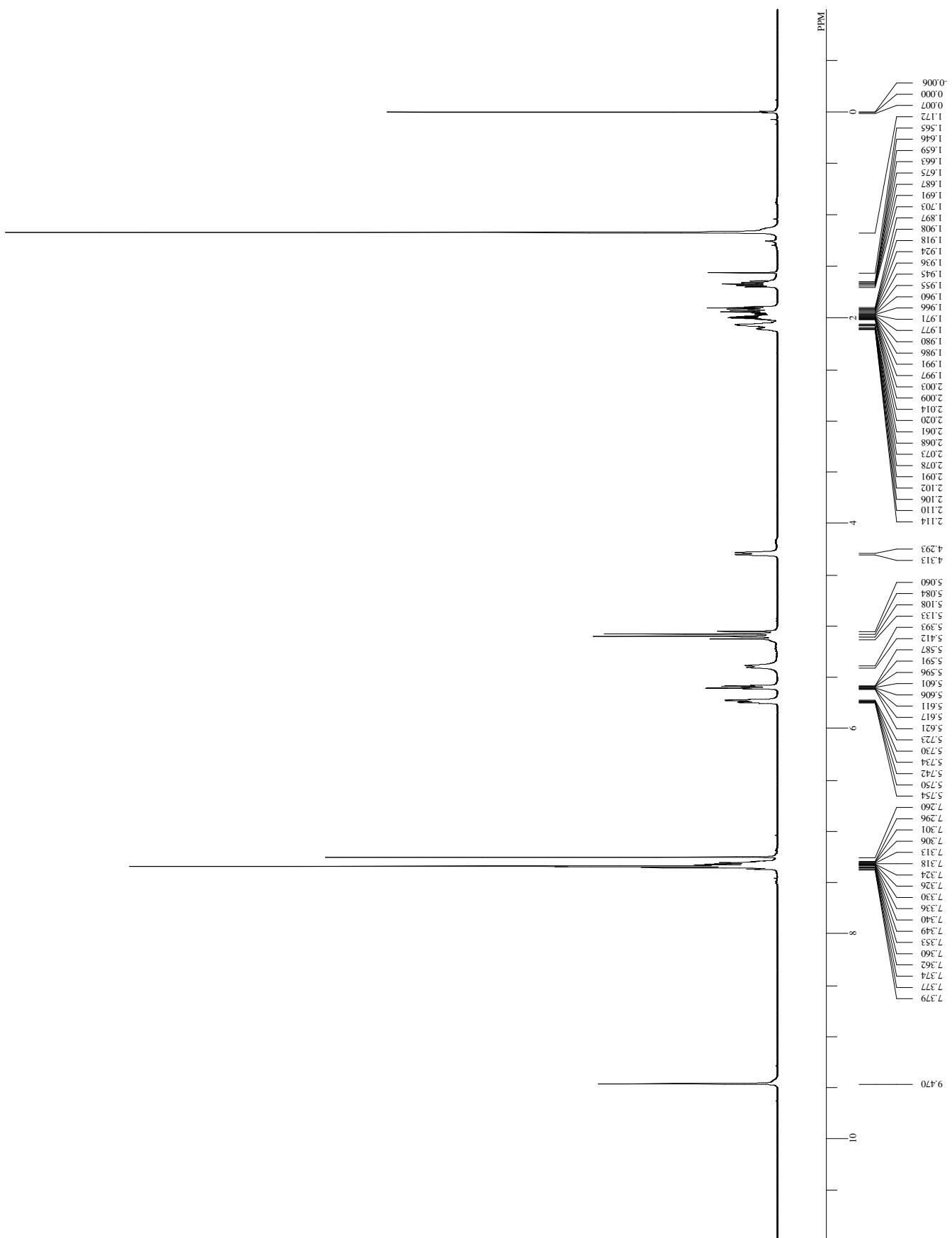
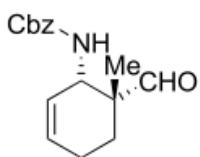
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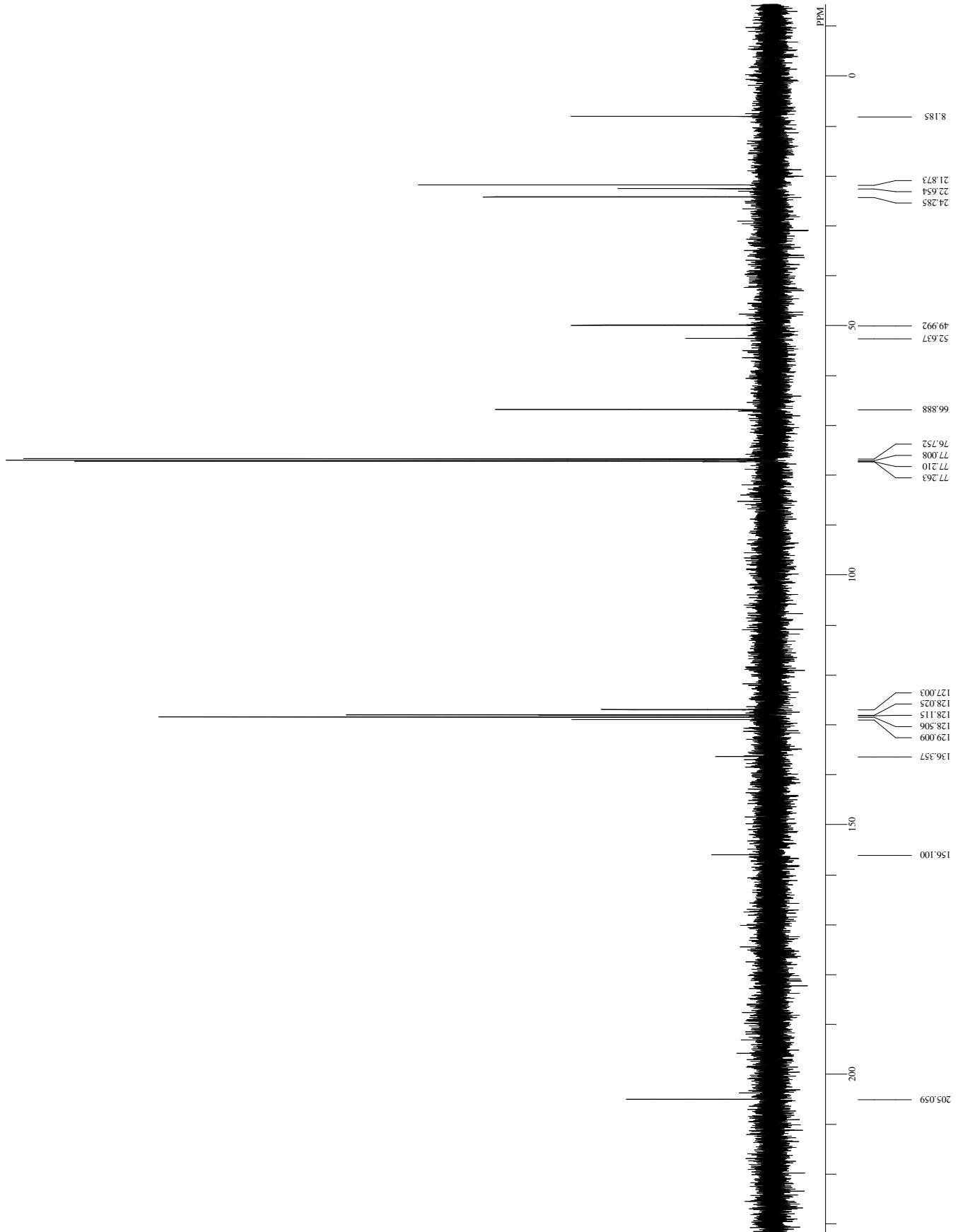
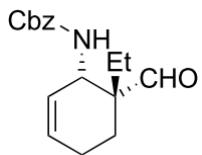
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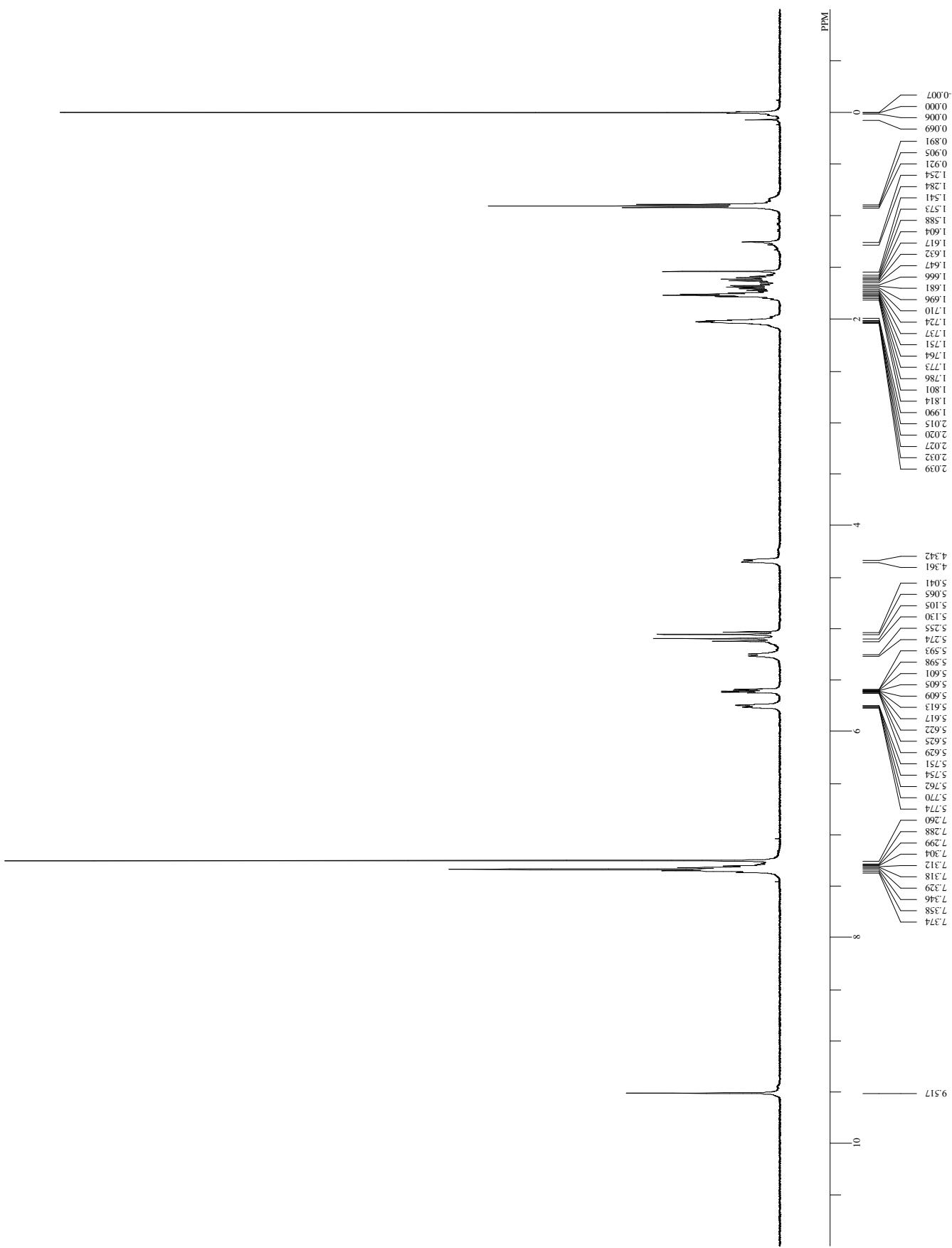
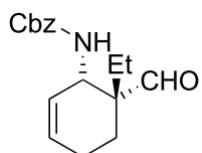
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EXMOD
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PD
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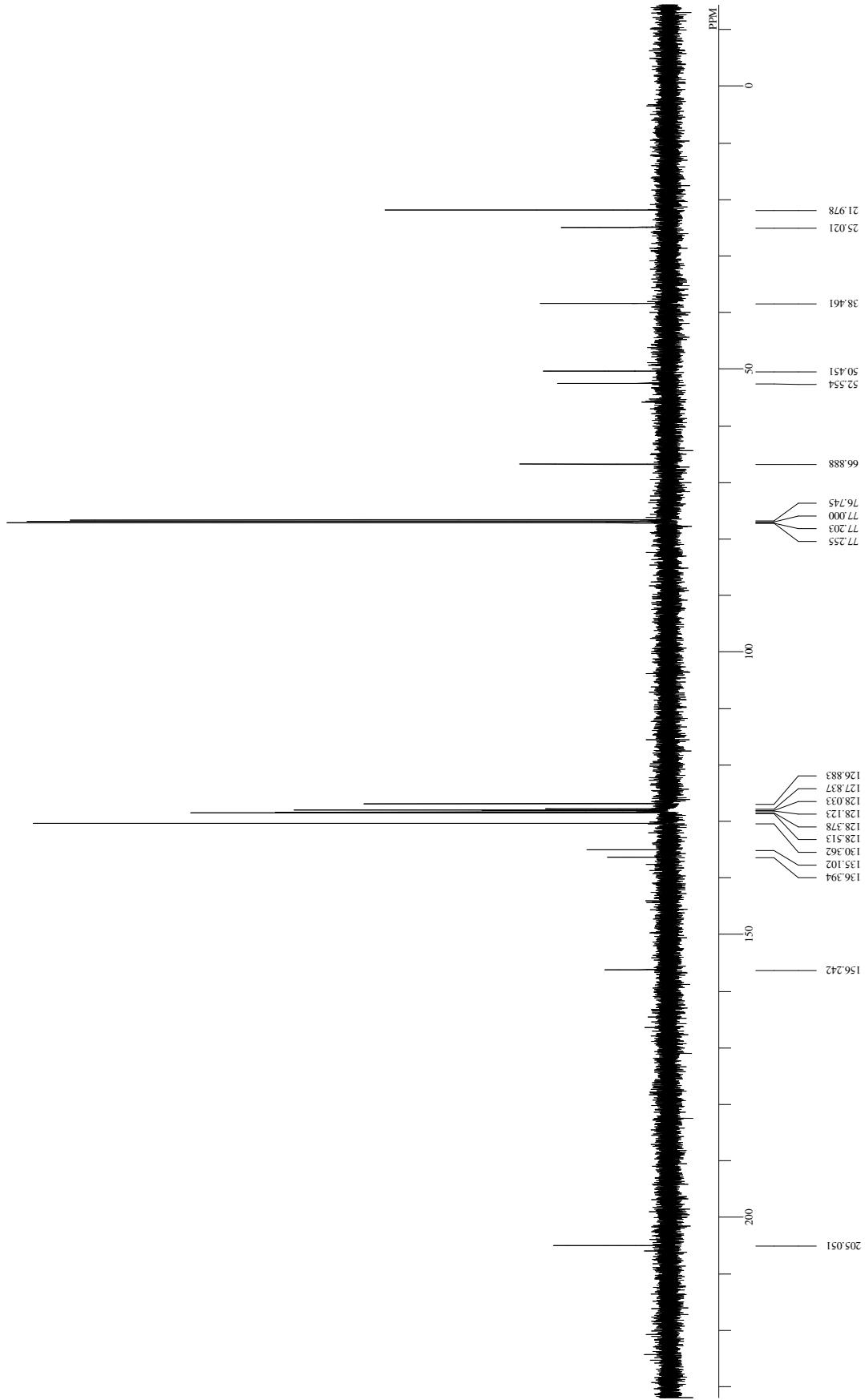
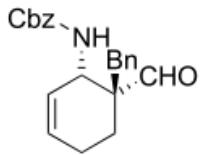
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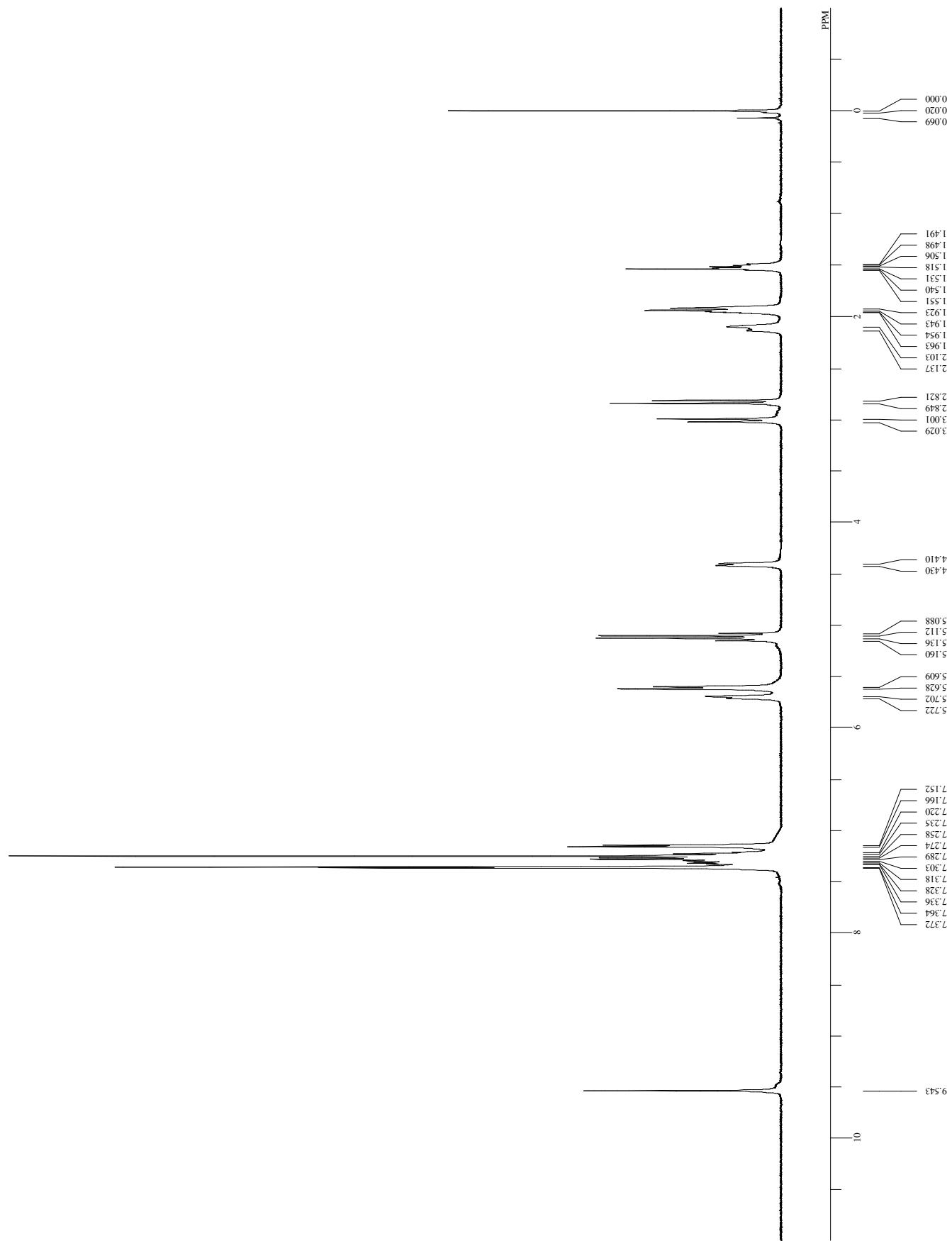
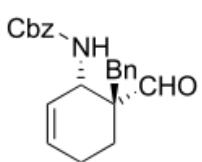
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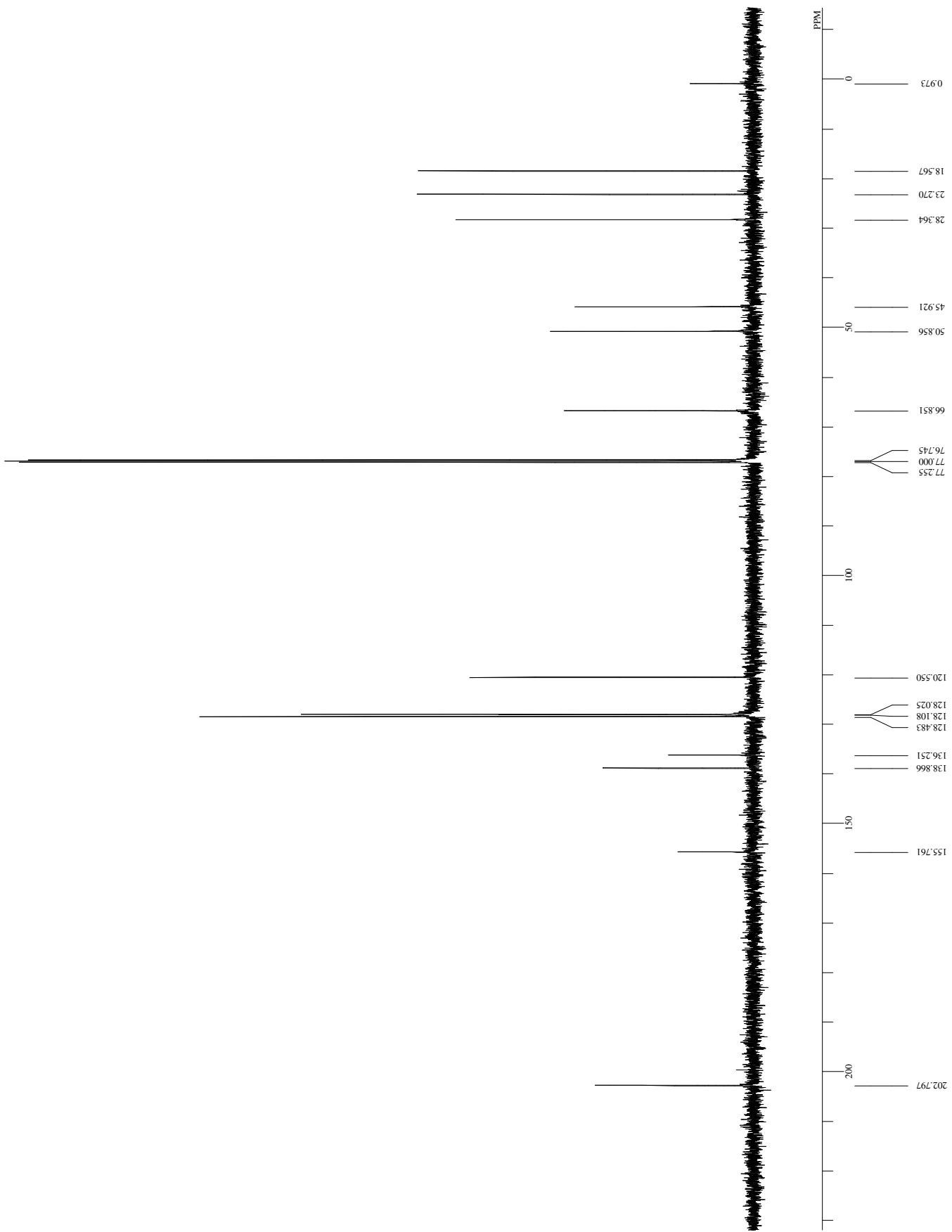
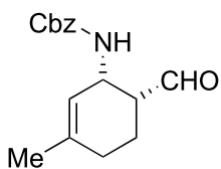
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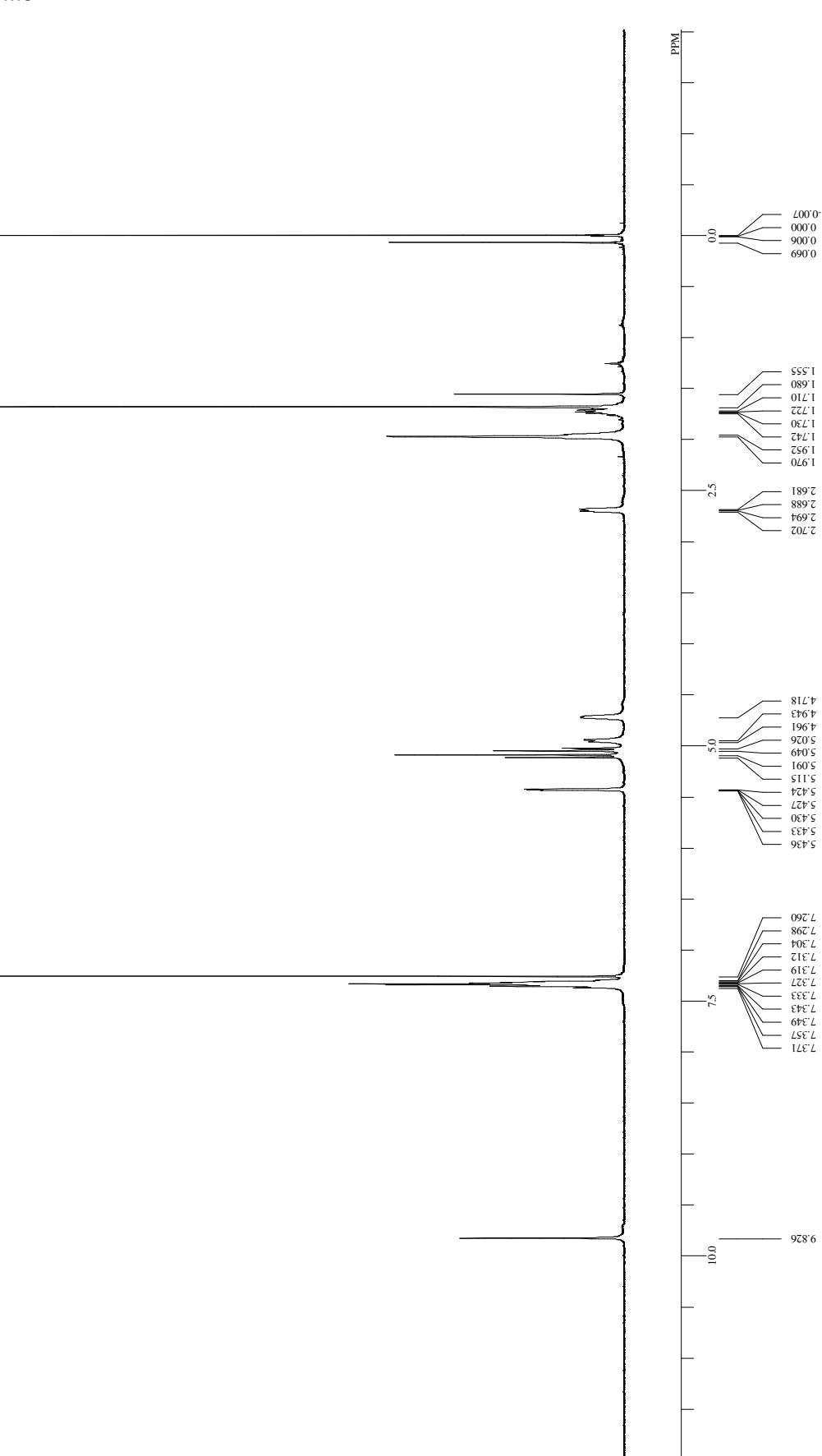
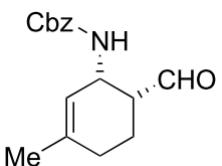
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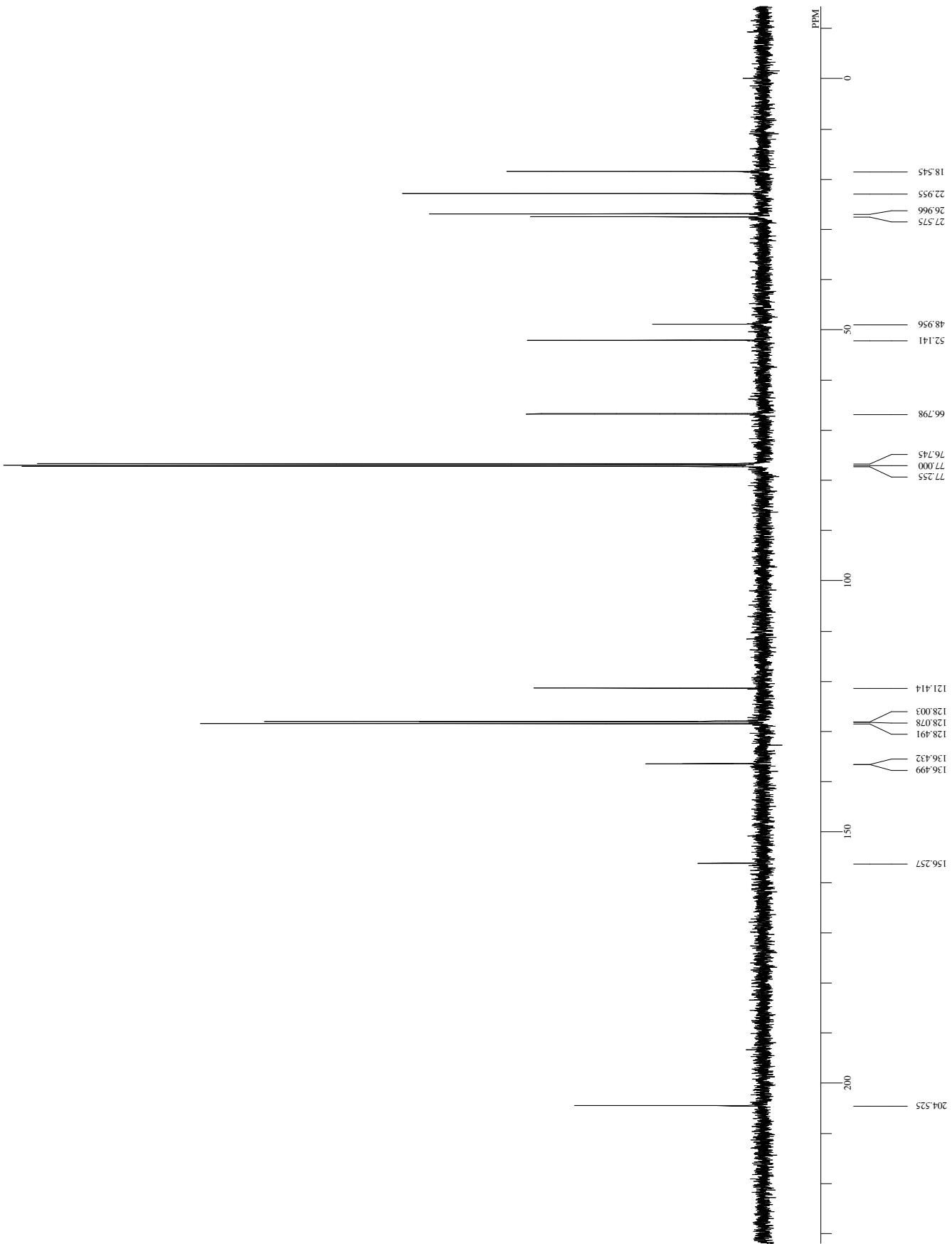
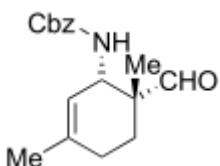
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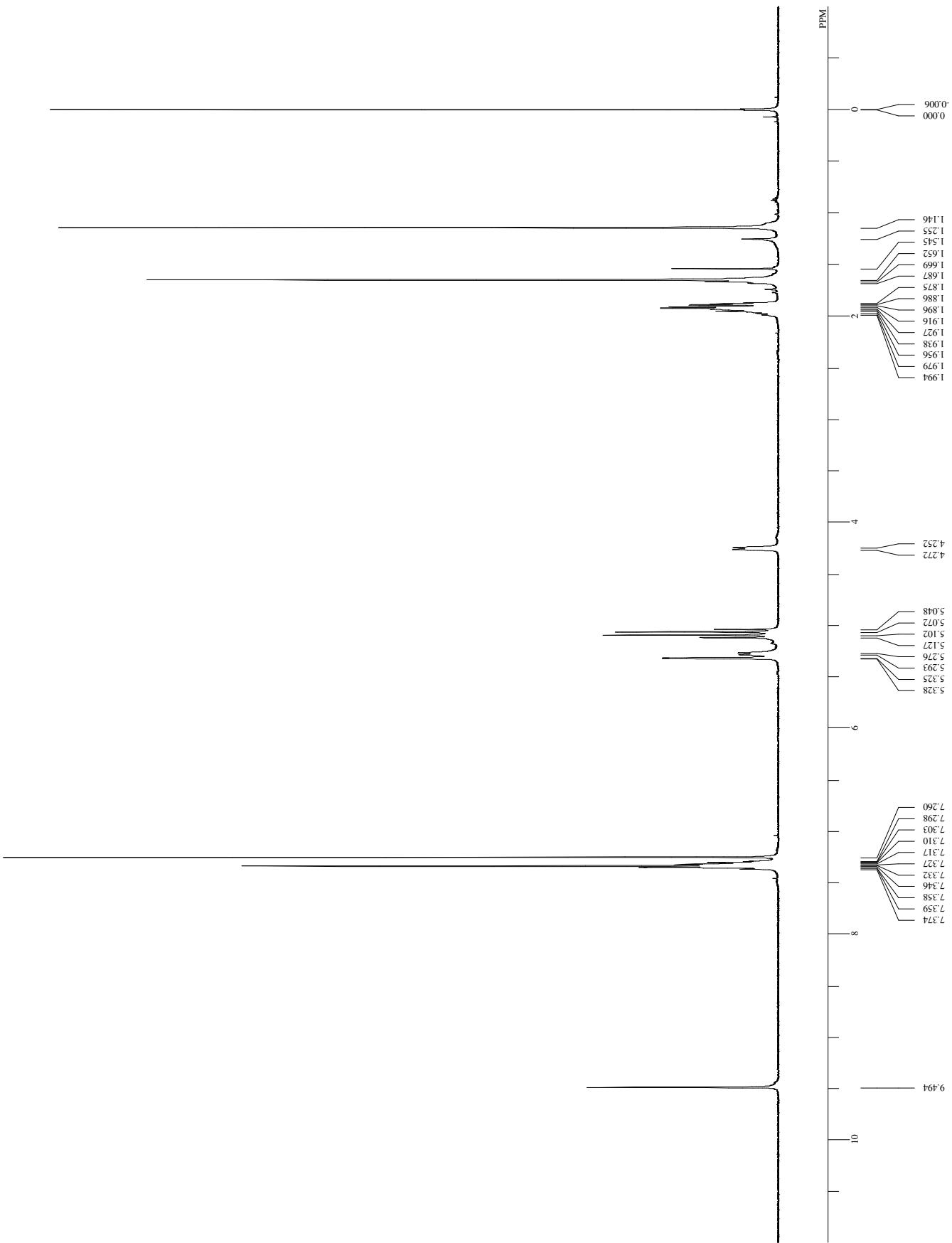
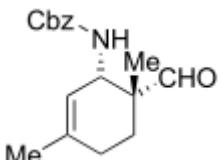
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RGAIN 28



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BF 22
RGAIN



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Fri Feb 18 15:14:20 2011

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0.00 kHz

OBIN
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POINT
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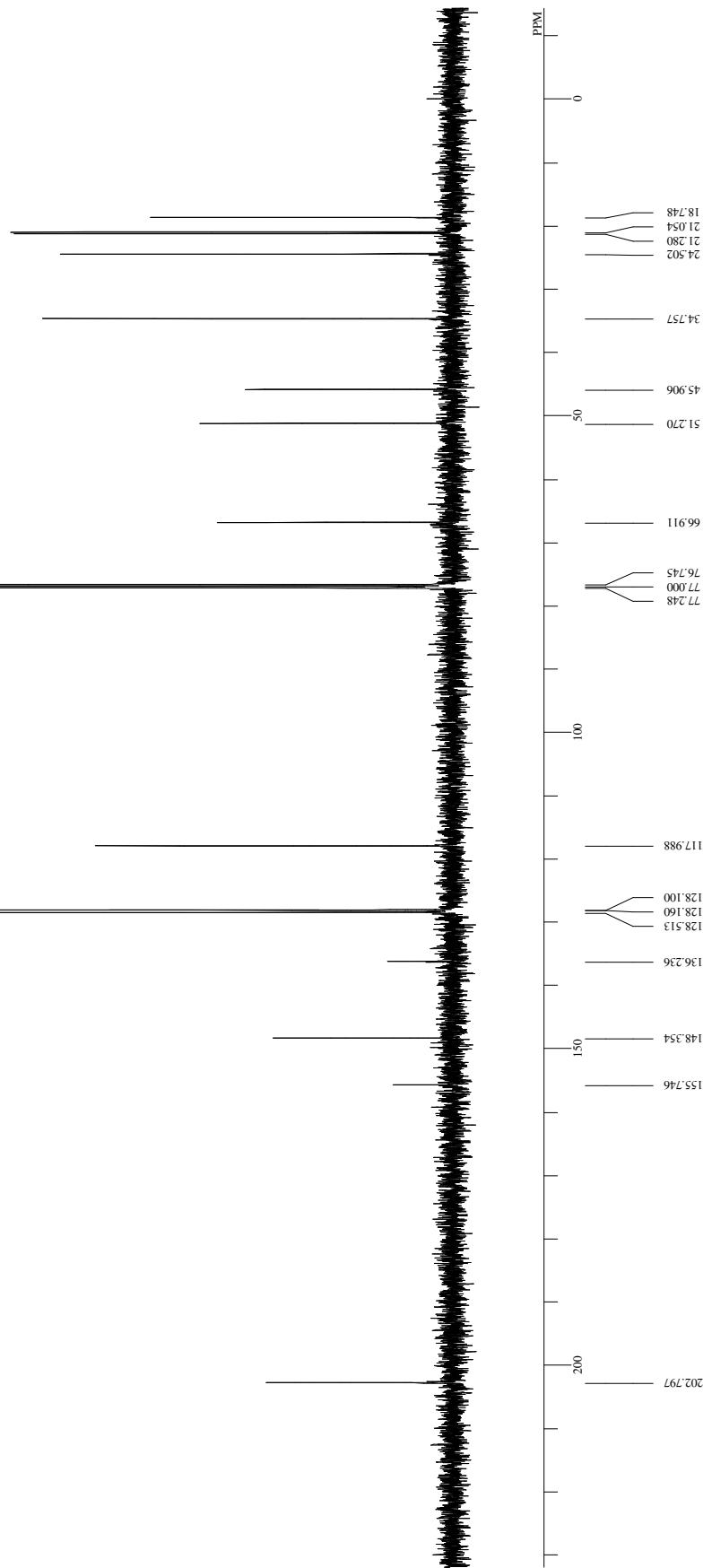
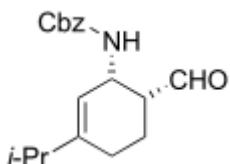
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BF
2.00 Hz

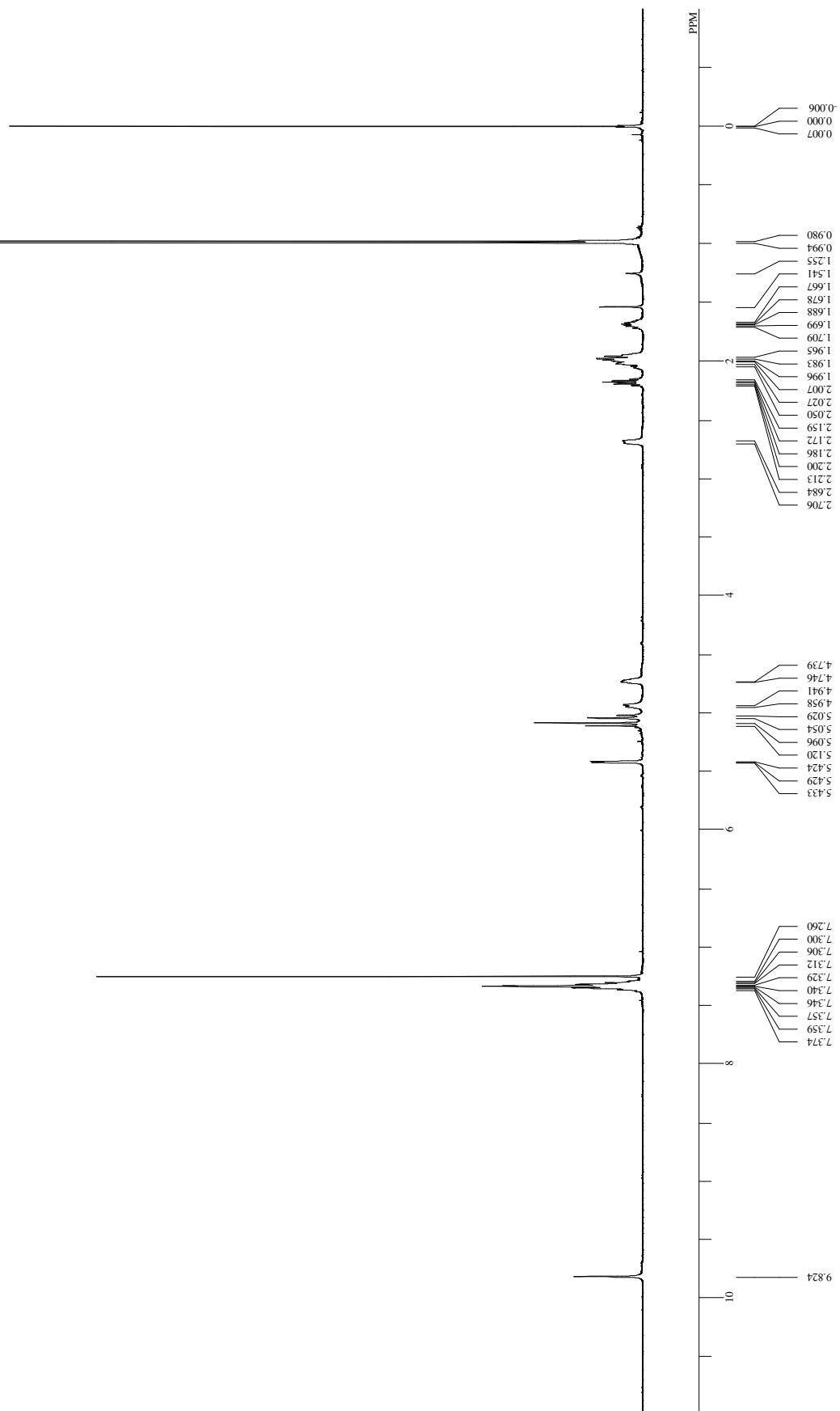
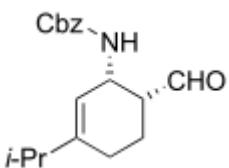
RGAIN
28



E:\Group C\1-HNMR\101021\hex5911-Pr\data.xls

DFILE
COMNT
DATIM
OBNUC
IH
EXMOD
OBRQ
OFFSET
OBIN
POINT
FREQU
SCANS
ACQTM
PD
PW1
IRNUC
CTEMP
SLVNT
EXREF
BF
RGAIN

500.00 MHz
0.00 kHz
16240.00 Hz
1.6384
6997.90 Hz
16
2,3413 sec
1.6587 sec
6.75 ussec
26.1 c
 CDCl_3
0.00 ppm
0.10 Hz
23

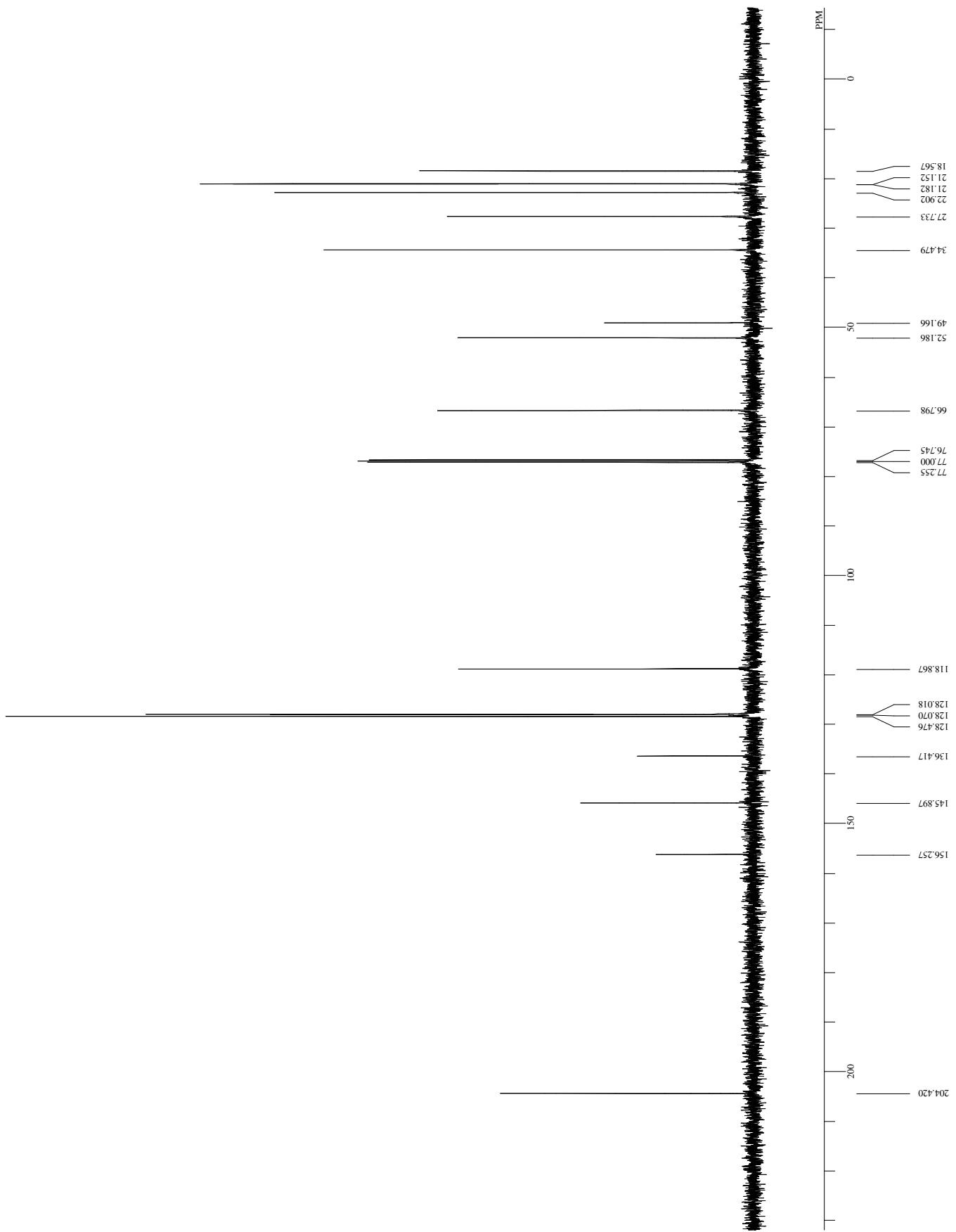
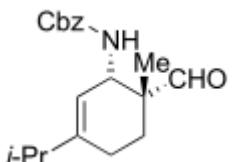


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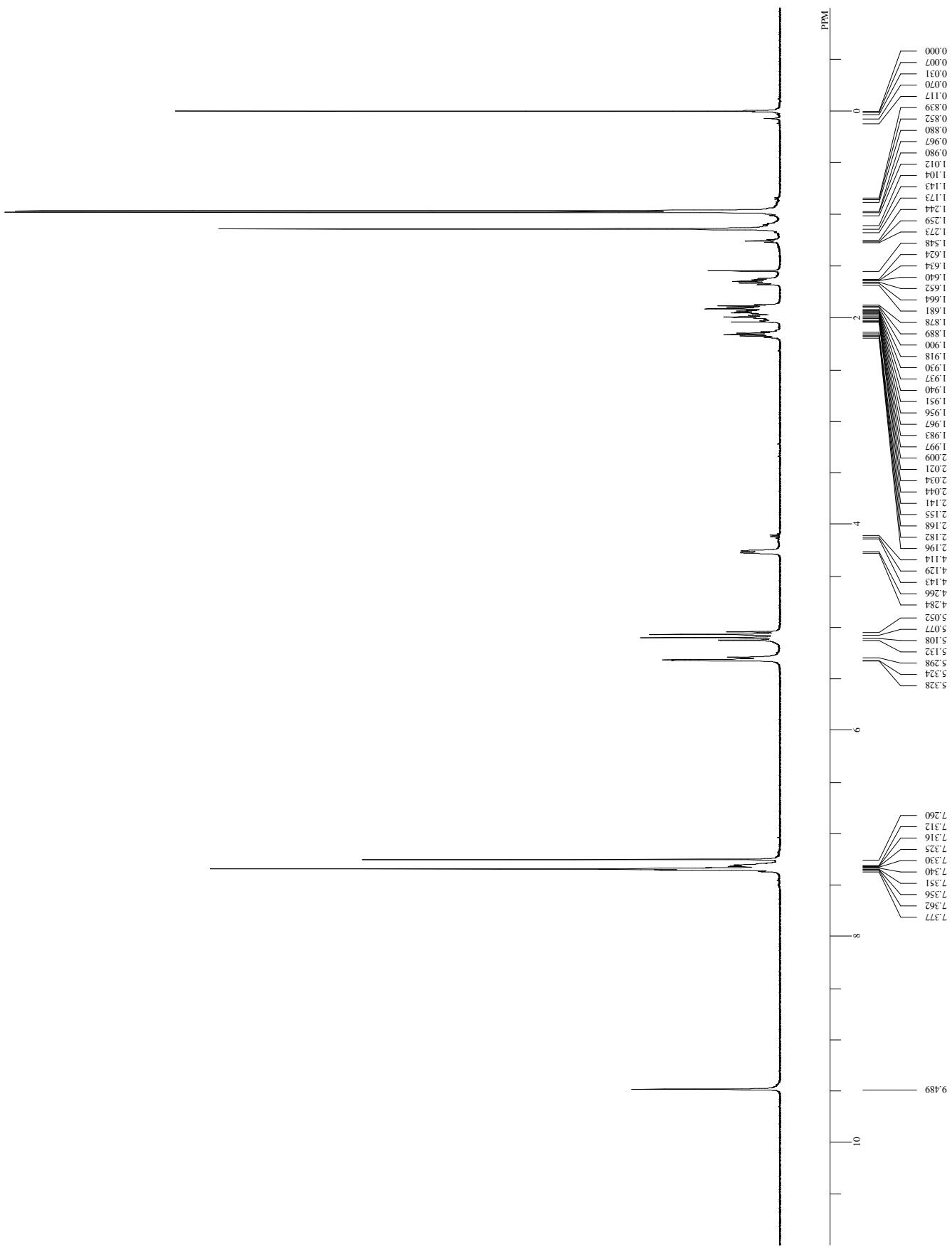
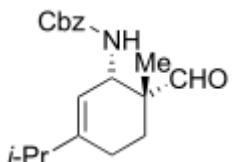
E:\Group\CV-13NMR\carbone57-1.prfals
E\Group CV-13NMR\carbone57-1.prfals

WED Jan 26 21:04:01 2011
SINGL           13C
EXMOD          OBERQ
OBIN           OBETI
POINT          POINT
FREQU          FREQU
SCANS          SCANS
ACQTM          ACQTM
PD              PD
PWL             PWL
IRNUC          IRNUC
CTEMP          CTEMP
SLVNT          SLVNT
EXREF          EXREF
BF              BF
RGAIN          RGAIN

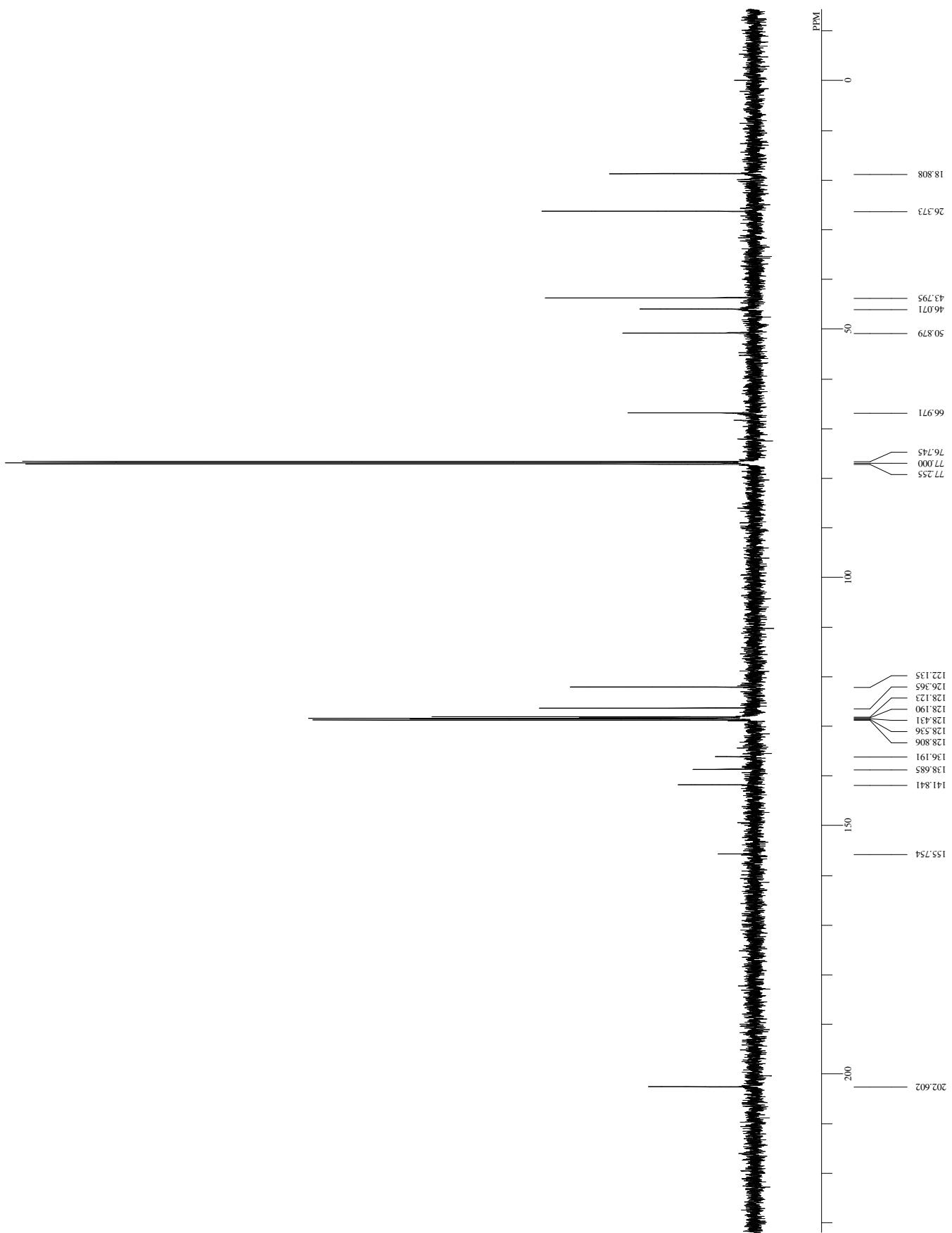
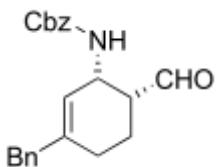
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DFILE E:\Group C\i-NMR\101021-\ex587 i-PrP.als



E:\Group C\13NMR\carbon\Cbz(Bn)-acroleinals
 DFILE COMINT
 DATIM Fri Feb 18 15:30:48 2011
 OBNUC 13C
 EXMOD SINGL
 OBRQ 125.65 MHz
 OFFSET 0.00 kHz
 OBFIN 12900.90 Hz
 POINT 32768
 FREQU 30959.75 Hz
 SCANS 200
 ACQTM 1.0584 sec
 PD 1.9016 sec
 PW1 5.00 uscc
 IRNUC 1H
 CTMP 24.8 c
 SLVNT CDCL₃
 EXREF 77.00 ppm
 BF 200 Hz
 RGAIN 28

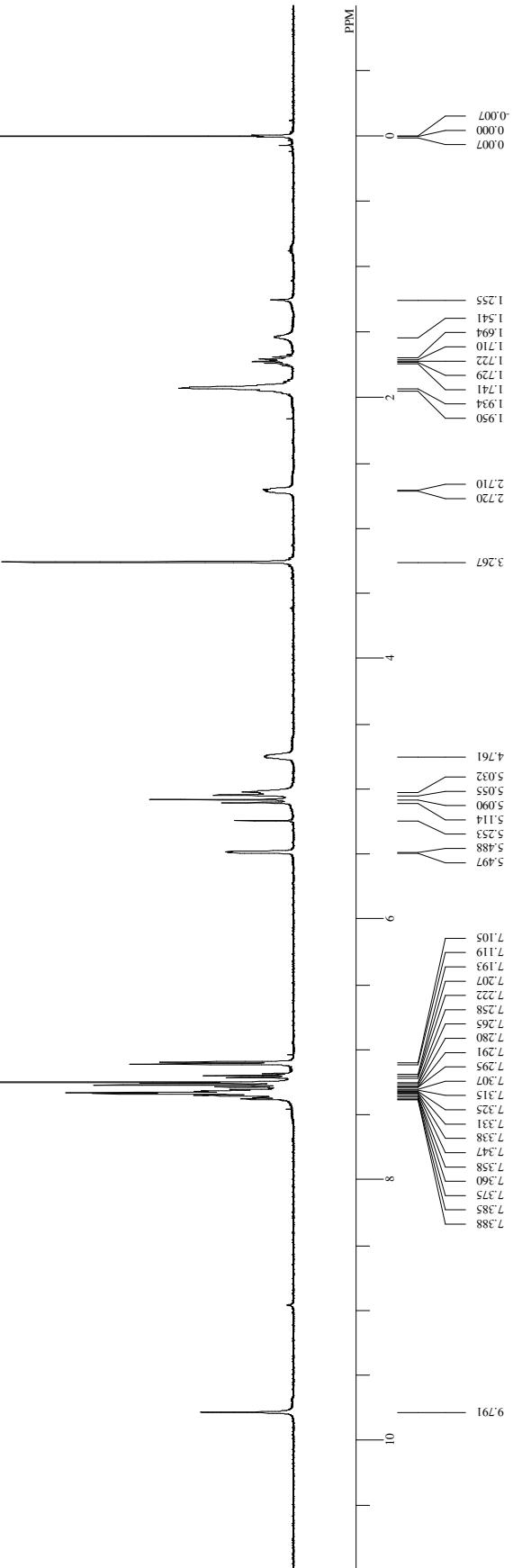
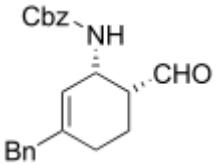


E:\Group C\j-i\NMR\101021-\ex591 Bnals

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    Thu Feb 17 21:24:21 2011
  ORBNIC 1H
  EXAMOD SINGL
  500.000 MHz
  0.00 kHz
  1624.000 Hz
  16384
  699.90 Hz
  16
  SCANS 2.3413 sec
  ACQTM 1.6587 sec
  OFFSET 6.75 ussec
  OBPN 1H
  POINT CDL3.5
  26.8 e
  IRNUC
  EXREFR
  SELVNT
  BFBF
  REGAIN

```



DFILE E:\Group C\1H-NMR\carbonec578.Bruks

COMNT Wed Jan 26 20:53:24 2011

DATIM 13C

OBNUC SINGL

EXMOD OBRQ

125.65 MHz

OFFSET 0.00 kHz

OBFIN 12900.90 Hz

POINT 32768

FREQU 30959.75 Hz

SCANS 206

ACQTM 1.0584 sec

PD 1.9016 sec

PWL 5.00 usec

IRNUC 1H

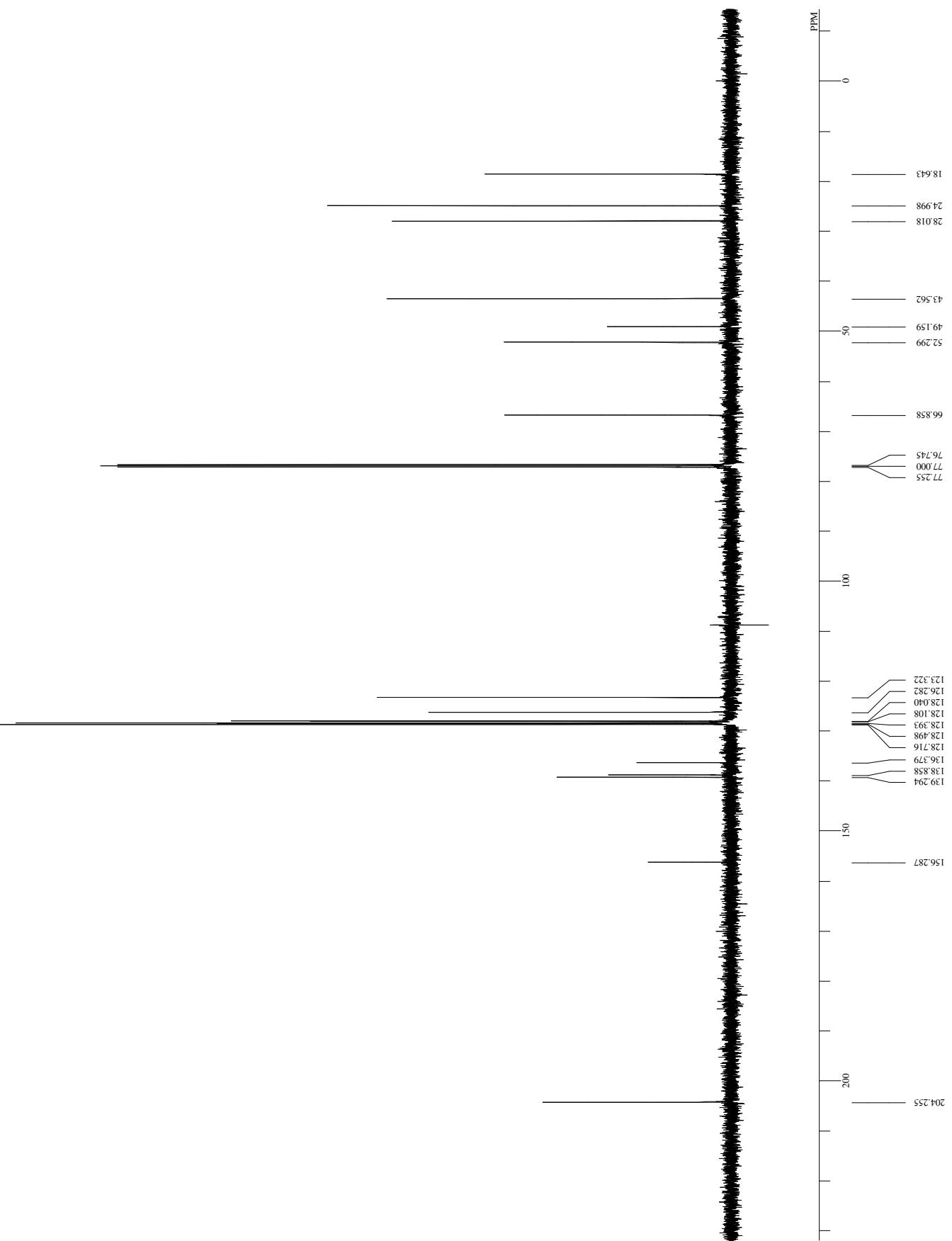
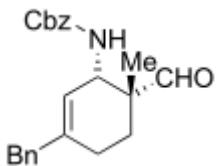
CTEMP 27.1 c

SLVNT CDCL₃

EXREF 77.00 ppm

BF 100 Hz

RGAIN 28



DFILE E:\Group C\JNMR101021\ex578.Bn.P.xls

COMNT Wed Jan 26 10:31:28 2011

DATIM IH

OBNUC SINGL

OBRQ 500.00 MHz

OBST 0.00 kHz

OBIN 16240.00 Hz

POINT 16384

FREQU 6997.90 Hz

SCANS 8

ACQTM 2.3413 sec

PD 1.687 sec

PWL 6.75 usec

IRNUC IH

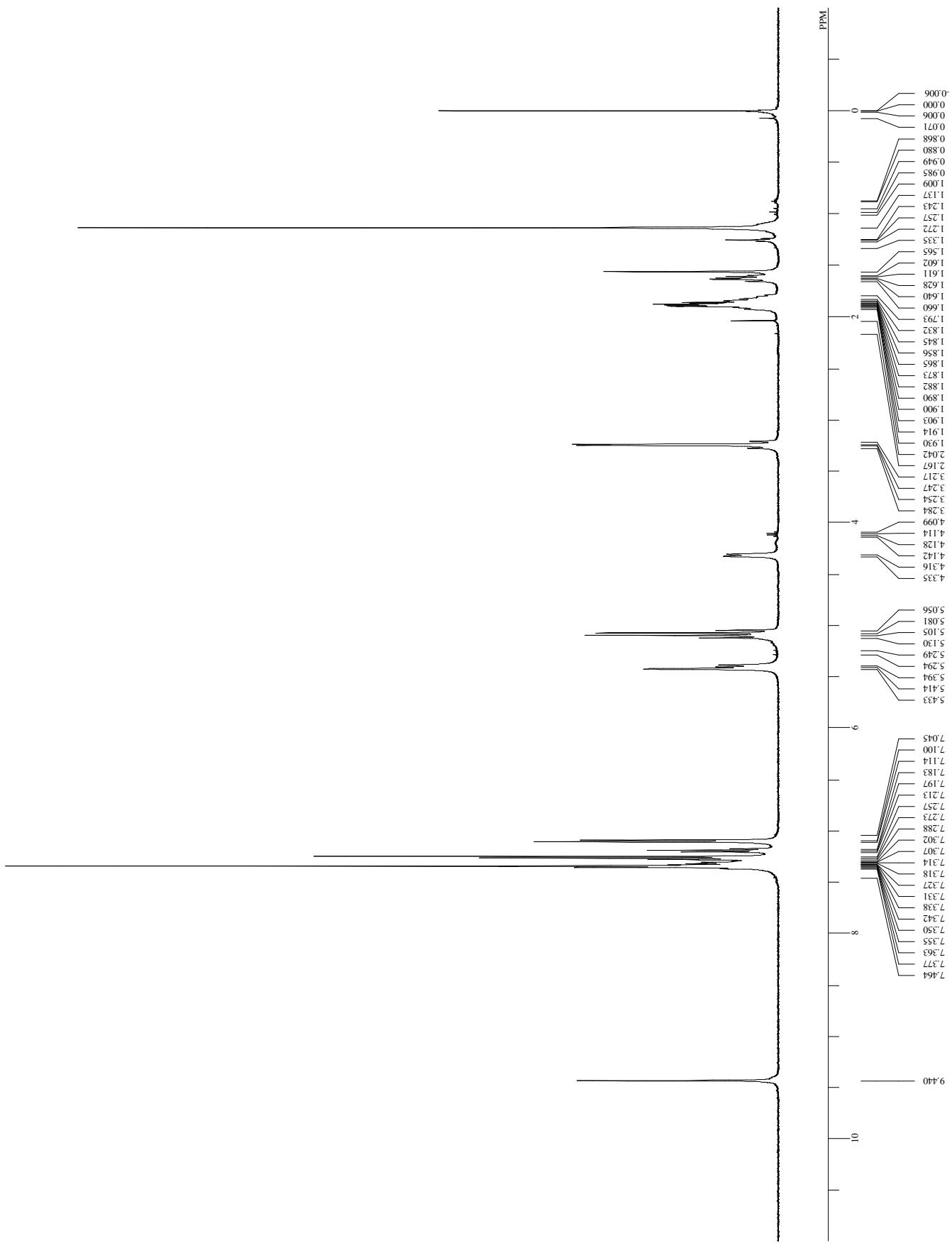
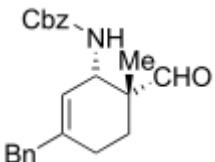
CTEMP 25.9 c

SLVNT CDCL₃

EXREF 0.00 ppm

BF 0.10 Hz

RGAIN 20



DFILE E:\Group C\1H-NMR\carbonec578.Etabs

COMNT Wed Jan 26 21:19:07 2011

DATIM

OBNUC

EXMOD

OBRQ

OBST

OBIN

POINT

FREQU

SCANS

ACQTM

PD

PWL

IRNUC

CTEMP

SLVNT

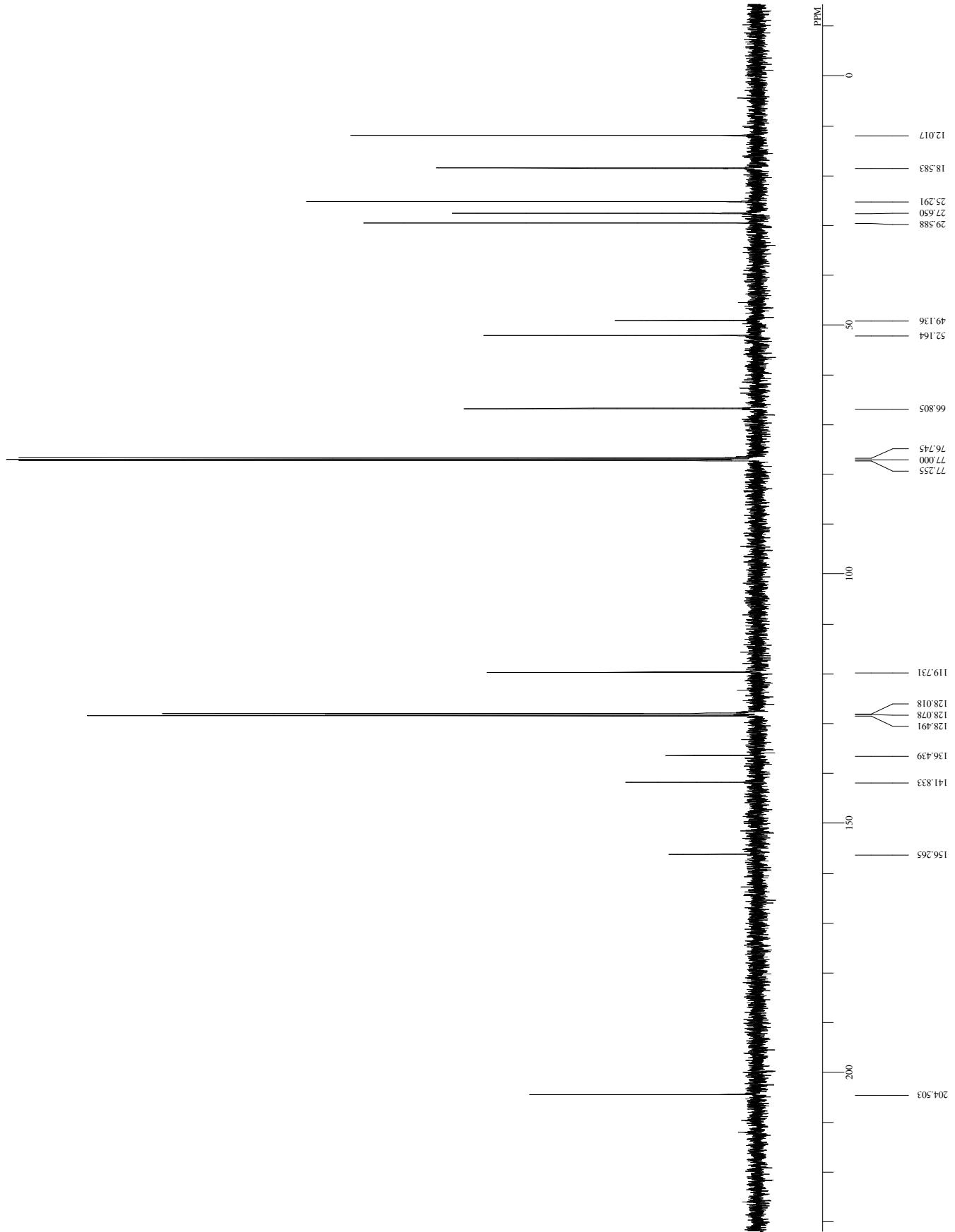
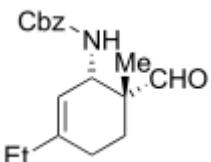
CDCL₃

EXREF

BF

RGAIN

28



E:\Group C\1-HNMR\101021\ex578.E1.Pals

COMNT
DATIM

Tue Jan 25 16:47:42 2011

OBNUC

IH

EXMOD

SINGL

OBRQ

500.00 MHz

OFFSET

0.00 kHz

OBPN

16240.00 Hz

POINT

16384

FREQU

6997.90 Hz

SCANS

8

ACQTM

2.3413 sec

PD

1.687 sec

PWL

6.75 usec

IRNUC

IH

CTEMP

25.5 c

SLVNT

CDCL₃

EXREF

0.00 ppm

BF

0.10 Hz

RGAIN

19

