

Supporting Information

Systematic Study of Fluorescein-Functionalized Macrophotoinitiators for Colorimetric Bioassays

Jungkyu K. Lee, Brandon. W. Heimer, and Hadley D. Sikes*

Department of Chemical Engineering, Massachusetts Institute of Technology, Cambridge,
MA 02139

* To whom correspondence should be addressed.

Tel: +1-617-253-5224

Fax: +1-617-253-2272

Email: sikes@mit.edu

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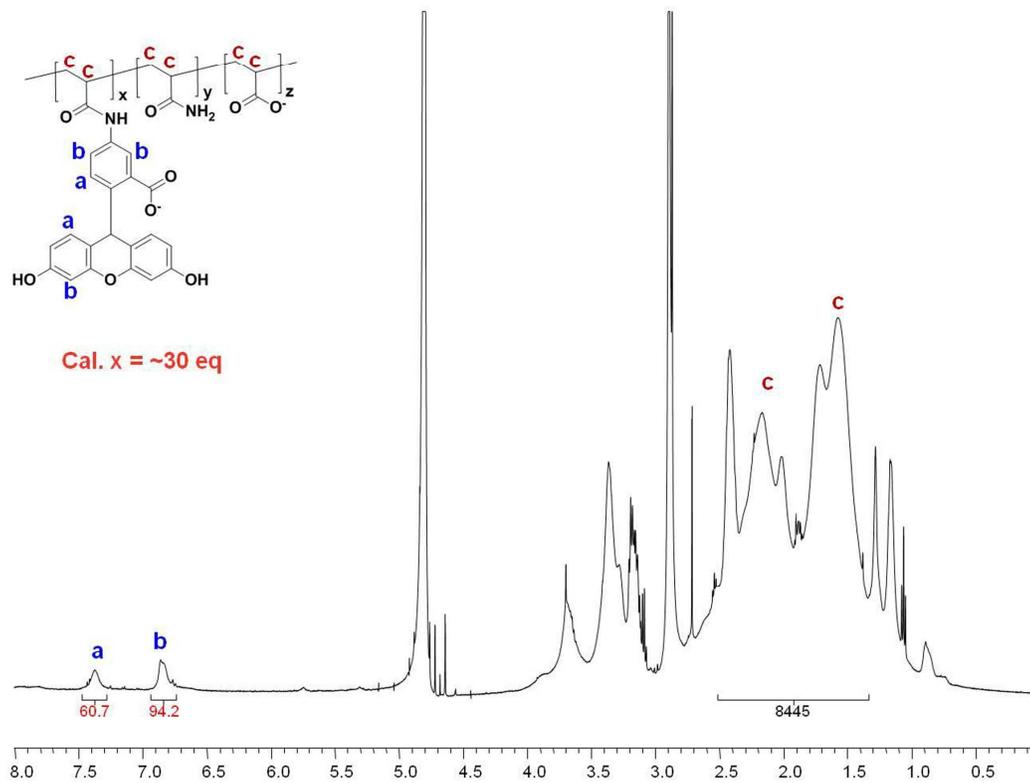
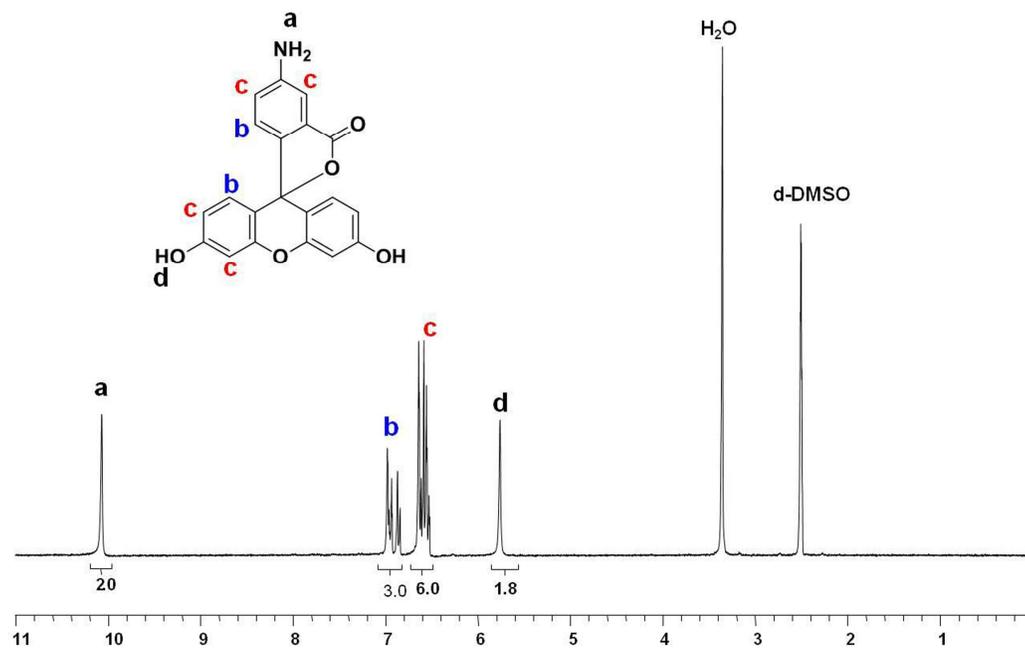
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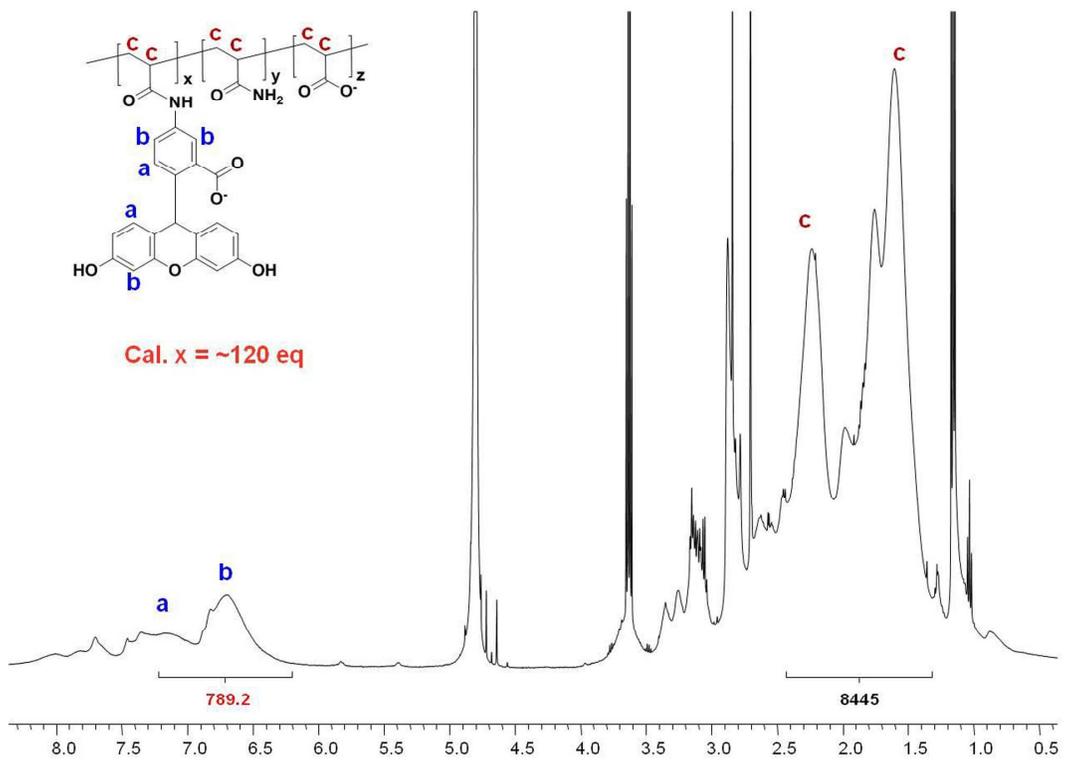
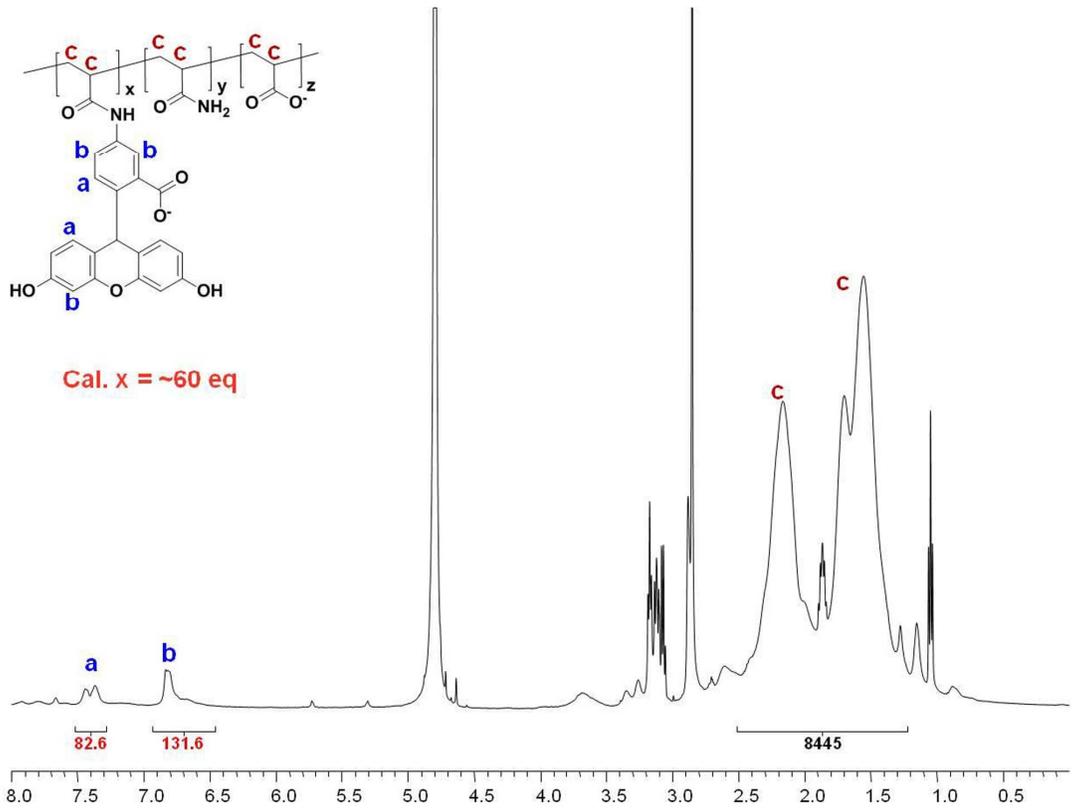
Figure S5. Stained chip images after photopolymerization using neutravidin/macrophotoinitiator **11** with varying irradiation time.

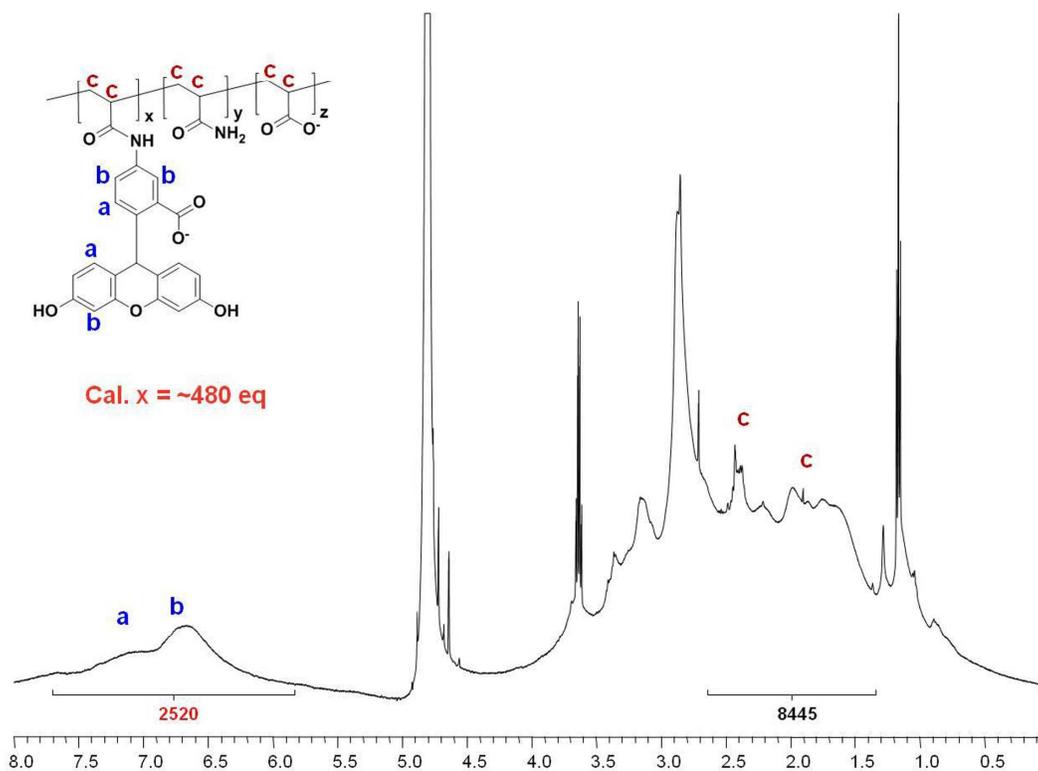
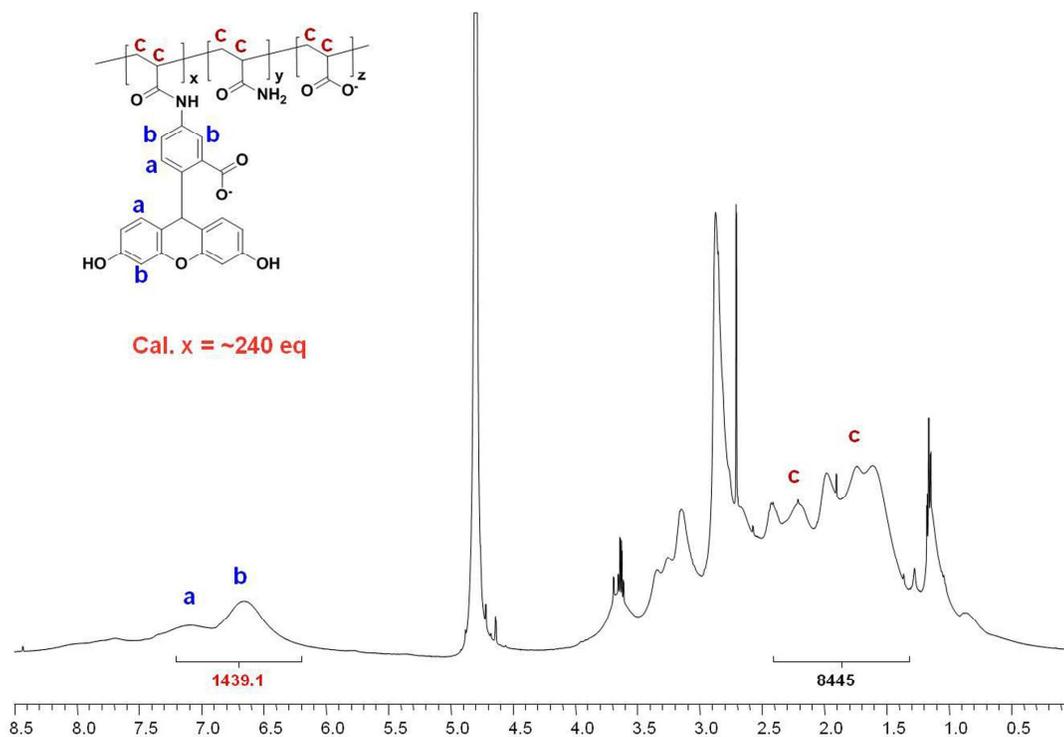
Figure S6. Analysis of colorimetric signal intensity.

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Figure S1. ^1H NMR spectra and peak assignment of fluoresceinamine and compound **2-6**.^a

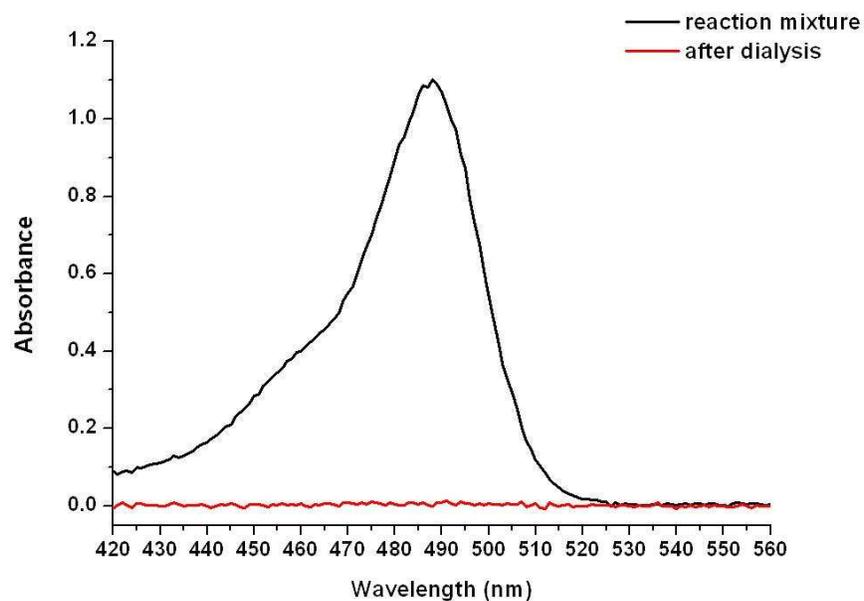






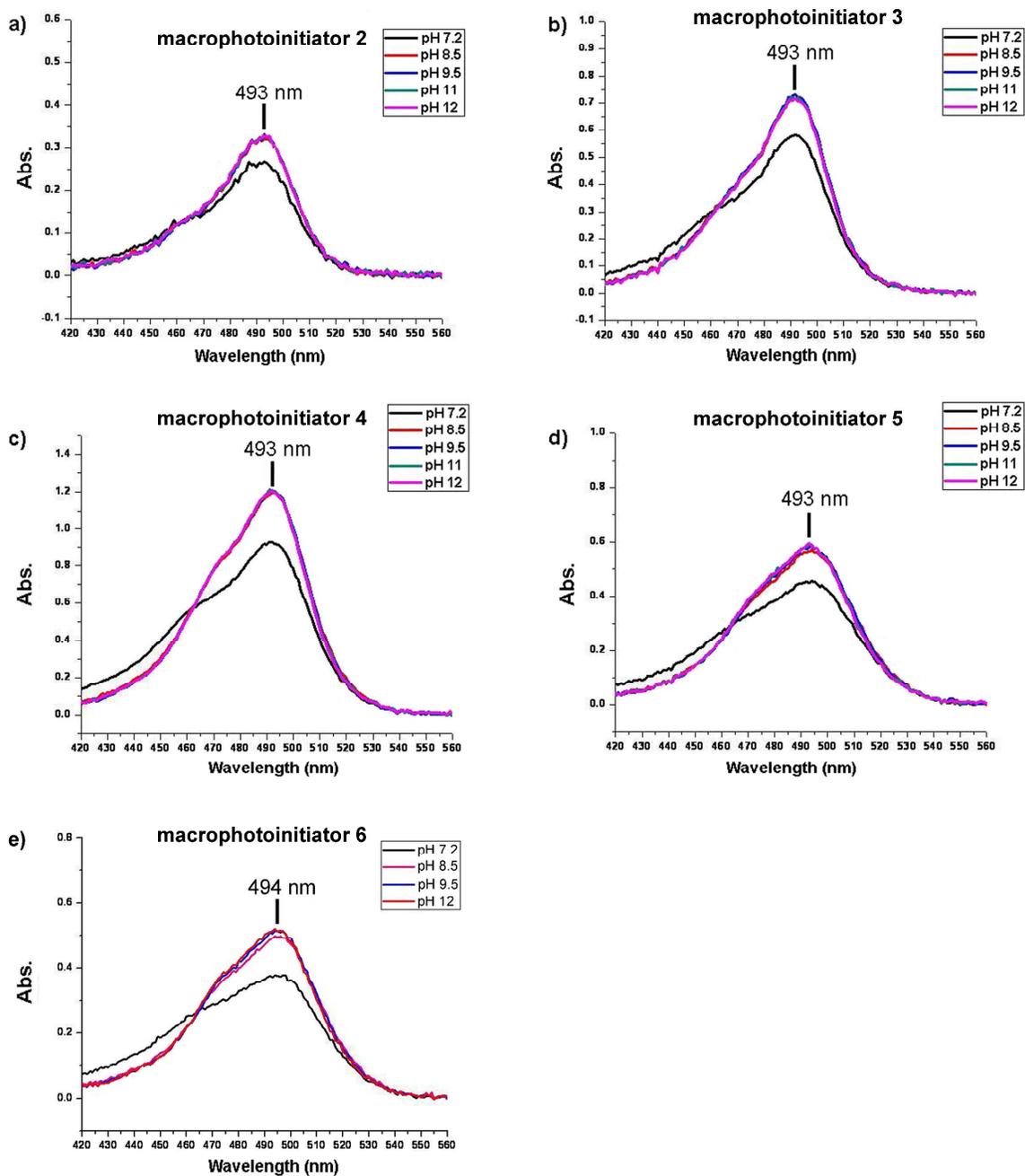
^a Due to the solubility, fluoresceinamine and macrophotoinitiators **2-6** were measured with DMSO-d₆ and D₂O, respectively.

Figure S2. UV/Vis absorption spectra of a physical blend of fluoresceinamine and copolymer **1** before and after dialysis.^a



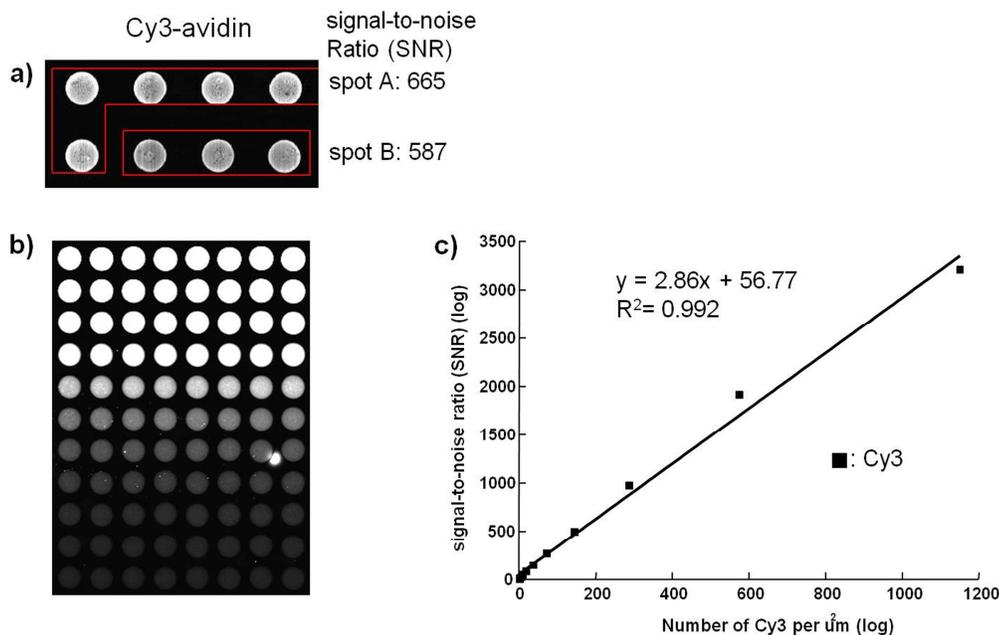
^aPhysical blend of copolymer (50 nmol, 10 mg) and fluoresceinamine (24 μ mol, 8.34 mg) was dissolved in a mixture of DMSO (0.5 mL) and pH 8.0 buffer solution (1.5 mL), and then was stirred at 35°C for 2 days. The dialysis was carried out as described in the Methods section. Before and after dialysis, the same volume concentration of the resulting mixtures was used to measure UV/Vis absorbance.

Figure S3. UV/Vis absorbance spectra of macrophotoinitiator **2-6** under varying pH conditions.^a



^aAs pH was increased, macrophotoinitiators **2-7** exhibited a hyperchromatic effect.

Figure S4. Determination of the density of biotin-DNA molecules on test surfaces by fluorescence intensity.



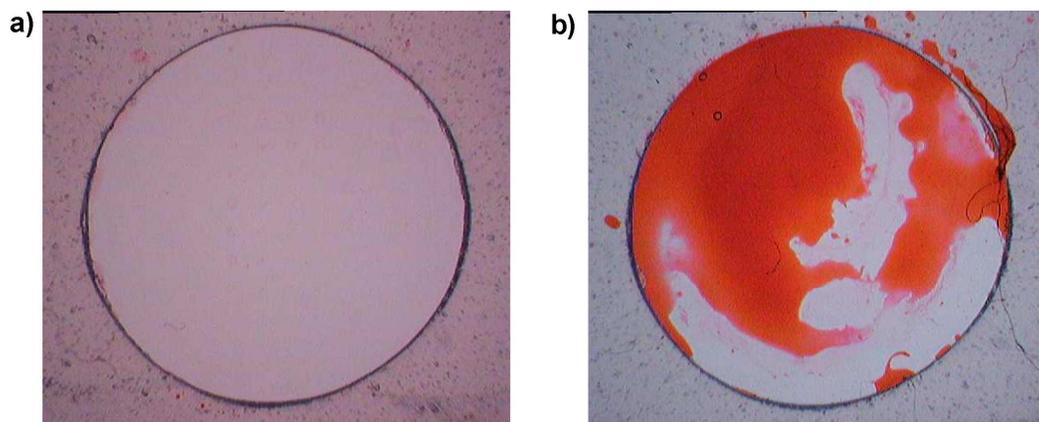
a) Fluorescence image of streptavidin-(Cy3)₂ bound on the test surfaces and average fluorescence intensity of each spot. b) Fluorescence image of a Cy3 calibration chip (Full Moon Biosystems), and c) its standard curve; black squares indicated the signal-to-noise ratio against 11 different numbers of Cy3 per square micrometer. The number of Cy3 on streptavidin was determined by following equation: $n_{\text{cy3}}/n_{\text{sa}} = (\text{Abs}_{\text{cy3},552} / \epsilon_{\text{cy3},552}) / [(\text{Abs}_{\text{sa},280} - \text{Abs}_{\text{cy3},280} - \text{Abs}_{1\% \text{BSA},280}) / \epsilon_{\text{sa},280}]$, and $\epsilon_{\text{cy3},552} = 150,000 \text{ M}^{-1}\text{cm}^{-1}$, $\epsilon_{\text{cy3},280} = 12,000 \text{ M}^{-1}\text{cm}^{-1}$, $\epsilon_{\text{BSA},280} = 43824 \text{ M}^{-1}\text{cm}^{-1}$, $\epsilon_{\text{sa},280} = 173,000 \text{ M}^{-1}\text{cm}^{-1}$.¹⁻³ $\text{SNR} = [(\text{Average of signal intensity}) - (\text{Average of background intensity})] / (\text{standard deviation of background intensity})$.

1. <http://www.nlv.ch/Molbiology/sites/Fluorescence1.htm>

2. <http://www.piercenet.com/files/TR0006-Extinction-coefficients.pdf>

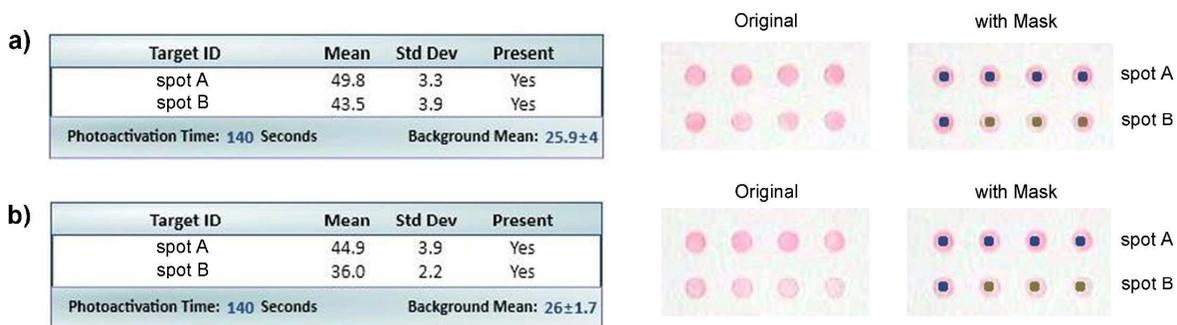
3. <http://medicine.yale.edu/labs/henegariu/www/tavi/FISHdyes.html>

Figure S5. Stained chip images after the photopolymerization using neutravidin/macrophotoinitiator **11** with varying irradiation time.^a



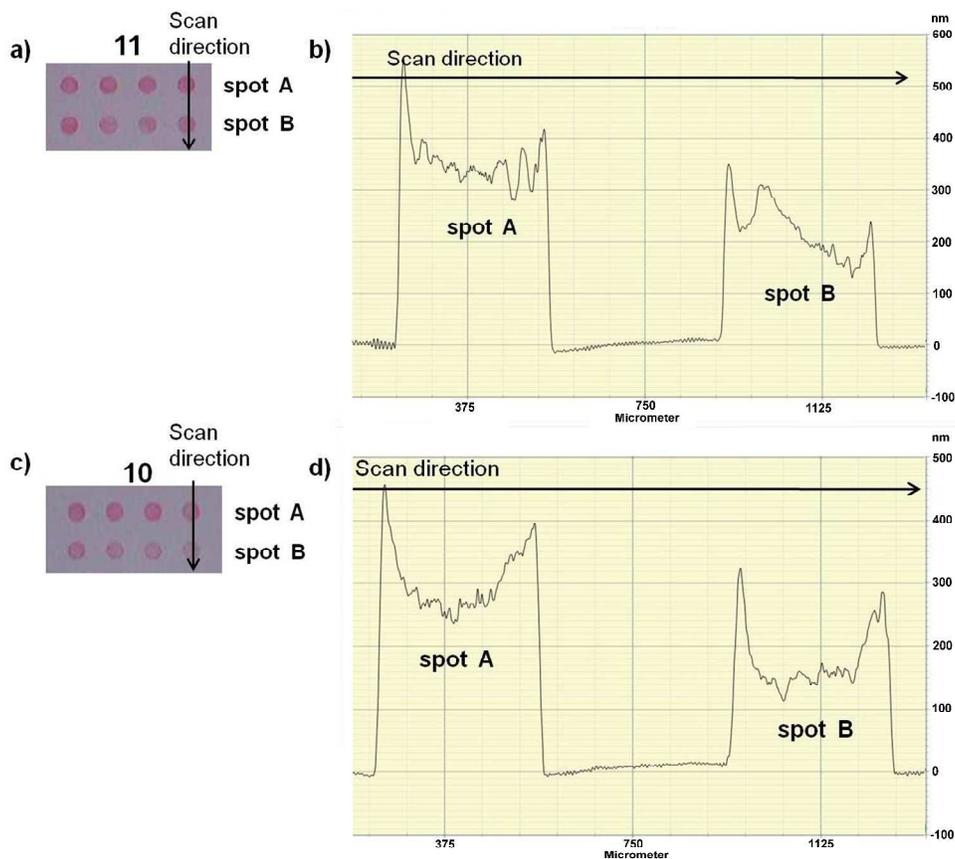
^aThe raw images of (a) 100 sec of irradiation; no polymerization detected, and (b) >190 sec of irradiation; hydrogel was excessively grown on the whole areas of the chip (bulk polymerization).

Figure S6. Analysis of color intensity after the photopolymerization of macrophotoinitiator **10** and **11** at 140 sec of irradiation, followed by the staining process.^a



^a Mean intensity for each row was obtained by the original image with mask, where the software calculated the mean intensity from the spots on each row; a) macrophotoinitiator **11** and b) macrophotoinitiator **10** was measured by three different areas on each row..

Figure S7. The images of surface profile after the photopolymerization using neutravidin/macrophotoinitiator **10** and **11** at 140 sec of irradiation.^a



^a The thickness of resulting polymer films were measured by Dektak 150 profilometer; a) image of a stained chip after photopolymerization with **11**, and b) the image of surface profile of a), c) image of a stained chip after photopolymerization with **10**, and d) the image of surface profile of c). The black arrow indicated a direction of the scanning target spots.