Origins of Enantioselectivity in Allylic Substitution Reactions Catalyzed by Metallacyclic Iridium Complexes

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General Procedures. All reactions were conducted in flame or oven-dried round-bottomed flasks fitted with rubber septa under a positive pressure of argon or nitrogen, or in 1-dram vials sealed with a screw cap fitted with PTFE silicon septum under an atmosphere of Argon, unless otherwise stated. Air and moisture-sensitive reagents were transferred via syringe, or were handled in an argon-filled or nitrogen-filled drybox (Innovative Technologies, Newburyport, Massachusetts) equipped with an oxygen sensor (working oxygen level <5 ppm) and low-temperature refrigeration unit (-35 °C). Organic solutions were concentrated by rotary evaporation at 23–35 °C. Flash-column chromatography was performed as described by Still et. al.¹ employing silica gel (40–63 μ m particle size) purchased from Silicycle. Analytical thin-layer chromatography (TLC) was performed using glass plates pre-coated with silica gel (0.25 mm). TLC plates were visualized by exposure to ultraviolet light (UV) or submersion in aqueous potassium permanganate solution (KMnO4), followed by brief heating by a heat gun (175 °C, 3–5 s).

Instrumentation. Proton nuclear magnetic resonance spectra (¹H NMR) were recorded at 500 MHz at 22 °C unless otherwise stated. Chemical shifts are expressed in parts per million (ppm, δ scale) downfield from tetramethylsilane and are referenced to residual protium in the NMR solvent (CHCl3, δ 7.26; THF, δ 1.73, 3.58; CH₂Cl₂, δ 5.32). Data are represented as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet and/or multiple resonances, br = broad, app = apparent), integration, coupling constant in Hertz. Proton-decoupled carbon nuclear magnetic resonance spectra (¹³C NMR) were recorded at 125 MHz at 22 °C. Chemical shifts are expressed in parts per million (ppm, δ scale) downfield from tetramethylsilane and are referenced to the carbon resonances of the solvent (CDCl3, δ 77.0; THF, δ 25.5, 67.7; CH₂Cl₂, δ 53.5) or to an external standard (CFCl₃ = 0 for ¹⁹F and 85% H₃PO₄ = 0 for 31P). Two-dimensional COSY (Correlation Spectroscopy) and Heteronuclear Multiple Quantum Coherence (HMQC) were recorded at 500 MHz at 22 °C. Gas chromatography was performed using an HP 6890 series gas chromatograph equipped with an HP-5 column (25 m, 0.2 mm I.D., 0.33 µm film). GC/MS analyses were performed on an Agilent 6890N GC equipped with a 5973 MS and an HP-5ms column (30 m x 0.25 mm ID x 0.25 µm film). HPLC analyses were carried out on a Waters chromatography system (1525 binary pump, 717+autosampler, 2487 dual wavelength detector). Elemental analyses were obtained at the University of Illinois Microanalysis Laboratory.

Materials. Commercial solvents and reagents were used as received with the following exceptions. Tetrahydrofuran, dichloromethane and benzene were deoxygenated and dried by sparging with argon and passing through columns of activated alumina and supported copper according to the method of Pangborn et al.² Aniline, *N*-methylaniline and octylamine were all dried over KOH, distilled under reduced pressure and freeze pump thawed before transferring into the drybox. Sodium dimethylmalonate was prepared by the reaction of 1 equiv. of the corresponding malonate with 1 equiv NaH in THF or THF-*d*8 directly before use. Lithium phenolate was prepared by reaction of 1 equiv. phenol with 1 equiv. of a 2.5 molar solution of n-butyllithium in hexane at 0 °C. Triethylamine was dried over CaSO₄ and distilled before use. All allylic carbonates (**4b**, **4b-D**-(*R***) and 6**) were synthesized by the reaction of the corresponding allylic alcohols with alkyl chloroformate in the presence of pyridine. Monodeuterated enantioenriched alcohols (±)-(*E*)-1-[²H₁]-3-Phenylprop-2-en-1-ol (corresponding to **4b-D**-(*R***)) and (***S***)-5-phenylpent-1-en-3-ol (corresponding to 5b-(***S*)) were prepared according to the method described previously.³⁻⁴ Ethylene complex **1a** was prepared according to an earlier report by our group.⁵ Cinnamyliridium complex **3e** containing the ligand **1b** was prepared according to the procedure reported previously by Helmchen and coworkers.⁶

Synthesis of allylic trifluoroacetates: Triethylamine (1.2 euiv.) was added to the solution of allylic alcohol (1 equiv) in THF (0.5 M). The resulting solution was cooled to 0° C, and acetic acid anhydride (1 equiv.) was added. The reaction mixture was allowed to stir for 4 hours while allowing the system to reach room temperature. The solvents were removed under reduced pressure, and the crude product was purified by column chromatography over silica gel with 8:2 hexane:EtOAc as the eluent.

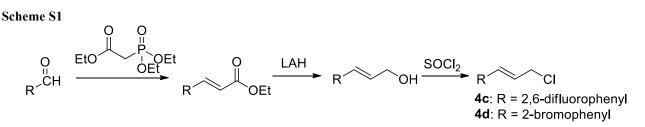
Cinnamyl trifluoromethylacetate, 4a: Colorless oil, 97% isolated yield. ¹H NMR (499 MHz, Benzene) δ 7.07 – 6.97 (m, 5H), 6.21 (d, *J* = 15.9 Hz, 1H), 5.74 (dt, *J* = 15.9, 6.8 Hz, 1H), 4.29 (d, *J* = 6.8 Hz, 1H). ¹³C NMR (126 MHz, Benzene) δ 157.2 (q, *J*_F = 42.0 Hz), 136.8, 135.6, 128.7, 128.6, 127.0, 120.1, 115.1 (q, *J*_F = 285.8 Hz), 68.4.

5-phenylpent-1-en-3-yl 2,2,2-trifluoroacetate, 5b: Colorless oil, 80% isolated yield. ¹H NMR (499 MHz, CDCl₃) δ 7.36 – 7.29 (m, 2H), 7.29 – 7.22 (m, 1H), 7.19 (dd, J = 7.8, 0.9 Hz, 2H), 5.91 – 5.81 (m, 1H), 5.45 – 5.32 (m, 3H), 2.81 – 2.61 (m, 2H), 2.22 – 2.11 (m, 1H), 2.10 – 2.00 (m, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 156.9 (q, J_F = 41.6 Hz), 140.4, 134.0, 128.7, 128.4, 126.5, 119.7, 114.7 (q, J_F = 287.0 Hz), 79.1, 35.5, 31.2.

5-phenylpent-2-en-1-yl 2,2,2-trifluoroacetate, 51: Colorless oil, 88% yield. ¹H NMR (499 MHz, CDCl₃) δ 7.40 – 7.12 (m, 5H), 6.01 – 5.92 (m, 1H), 5.71 – 5.59 (m, 1H), 4.79 (d, J = 6.7 Hz, 2H), 2.80 – 2.71 (m, 2H), 2.51 – 2.37 (m, 2H). ¹³C NMR (126 MHz, CDCl₃) ¹³C NMR (126 MHz, CDCl₃) δ 157.6 (q, $J_F = 42.2$ Hz), 141.4, 138.8, 128.6, 126.3, 122.3, 122.4, 114.8 (q, $J_F = 285.8$ Hz), 68.8, 35.3, 34.2.

Synthesis of allylic chlorides:

Allylic chlorides 4c and 4d were prepared starting from corresponding aldehydes by a Horner-Wadsworth-Emmons reaction, followed by reduction with lithium aluminum hydride to form the allylic alcohol. The allylic alcohol was subsequently converted to the allylic chloride with SOCl₂ to give 4c or 4d (Scheme S1).



(2,6-difluoro)cinnamylchloride, 4c: ¹H NMR (500 MHz, CDCl₃) δ 7.22 – 7.12 (m, 1H), 6.94 – 6.82 (m, 2H), 6.76 – 6.61 (m, 2H), 4.25 (d, *J* = 6.3 Hz, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 161.1 (dd, *J_F* = 252.2, 7.5 Hz), 131.8 (t, *J_F* = 8.2 Hz), 129.0 (t, *J_F* = 10.9 Hz), 120.5, 113.4, 111.7 (dd, *J_F* = 20.8, 5.4 Hz), 46.0.

(2-bromo)cinnamylchloride, 4d: ¹H NMR (500 MHz, CDCl₃) δ 7.62 – 7.47 (m, 2H), 7.32 – 7.23 (m, 1H), 7.17 – 7.08 (m, 1H), 7.01 (d, J = 15.6 Hz, 1H), 6.27 (dt, J = 15.5, 7.1 Hz, 1H), 4.27 (d, J = 7.1 Hz, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 135.8, 133.1, 132.7, 129.6, 127.9, 127.7, 127.3, 123.9, 45.1.

Synthesis of allyliridium complexes

Cinnamyliridium complex 2c: 100 mg of Ir(I) ethylene complex **1a**-(*R*,*R*,*R*) (0.115 mmol) was dissolved in 5 ml THF, and 53.0 mg of cinnamyl trfiluoroacetate 4a (0.230 mmol, method A) or 44.0 mg of methylcinnamyl carbonate 4b (0.230 mmol, method B) was added to the solution. The solution was allowed to stir for 5 min, after which time 22.4 mg AgBF₄ (0.115 mmol) was added. Large amounts of dark precipitate formed quickly upon addition of the AgBF₄. The precipitate was removed by syringe filtration through a 25 mm diameter and 0.2 µm pore size filter, and the filtrate was added dropwise to 10 ml of pentane while stirring vigorously. The precipitated product was collected by filtration and died under vacuum to give 90 mg (75%, Method A) or 96 mg (80%, Method B) of allyliridium complex 2c after drying. Crystals suitable for single-crystal structural analysis were obtained by dissolving 2c in a small amount THF and layering with benzene. Anal. Calc'd for C₅₉H₅₆BF₄IrNO₂P: C, 63.21; H, 5.03; N, 1.25. Found: C, 63.45; H, 5.21; N, 1.45. ¹H NMR (500 MHz, CD₂Cl₂) δ 8.32 (d, J = 8.9 Hz, 1H), 8.25 (d, J = 8.8 Hz, 1H), 8.14 - 8.07 (m, 2H), 7.84 (d, J = 8.8 Hz, 1H), 8.14 - 8.8 Hz, 1H), 8.14 (d, J = 8.8 H Hz, 1H), 7.71 (d, J = 8.8 Hz, 1H), 7.67 – 7.56 (m, 4H), 7.56 – 7.34 (m, 15H), 7.22 – 7.15 (m, 2H), 5.63 (t, J = 12.1 Hz, 1H), 5.26 - 5.14 (m, 1H), 4.93 - 4.78 (m, 1H), 4.24 - 4.14 (m, 1H), 4.02 - 3.88 (m, 2H), 3.56 - 3.47 (m, 1H), 3.02 - 3.48 (m, 2H), 3.56 - 3.47 (m, 1H), 3.02 - 3.48 (m, 2H), 3.56 - 3.47 (m, 2H), 3.02 - 3.48 (m, 2H), 3.56 - 3.47 (m, 2H), 3.02 - 3.48 (m, 2H), 3.56 - 3.47 (m, 2H), 3.02 - 3.48 (m, 2H), 3.56 - 3.47 (m, 2H), 3.02 - 3.48 (m, 2H), 3.56 - 3.47 (m, 2H), 3.02 - 3.48 (m, 2H), 3.56 - 3.47 (m, 2H), 3.02 - 3.48 (m, 2H), 3.56 - 3.47 (m, 2H), 3.02 - 3.48 (m, 2H), 3.56 - 3.47 (m, 2H), 3.02 - 3.48 (m, 2H), 3.56 - 3.47 (m, 2H), 3.02 - 3.48 (m, 2H), 3.56 - 3.47 (m, 2H), 3.02 - 3.48 (m, 2H), 3.56 - 3.47 (m, 2H), 3.02 - 3.48 (m 2.89 (m, 2H), 2.89 – 2.71 (m, 2H), 2.55 – 2.38 (m, 2H), 2.27 – 2.16 (m, 1H), 2.15 – 2.04 (m, 2H), 1.93 – 1.79 (m, 1H), 1.69 - 1.54 (m, 2H), 1.24 (t, J = 12.3 Hz, 1H), 0.61 (d, J = 7.4 Hz, 3H). ¹³C NMR (126 MHz, CD₂Cl₂) δ 148.83, 148.70, 147.50, 147.43, 142.42, 142.32, 140.22, 133.96, 133.91, 133.17, 132.92, 132.24, 132.19, 132.10, 131.95, 129.92, 129.68, 128.97, 128.87, 128.78, 128.67, 128.56, 128.54, 128.49, 128.40, 128.36, 127.61, 127.57, 127.38, 127.17, 126.67, 126.54, 126.24, 124.49, 122.80, 121.93, 121.19, 121.16, 120.97, 101.48, 97.22, 92.84, 91.02, 86.55, 84.15, 66.12, 65.87, 60.62, 39.95, 35.08, 33.32, 28.13, 26.85, 18.66, 17.47. ³¹P NMR (202 MHz, THF) δ 118.5.

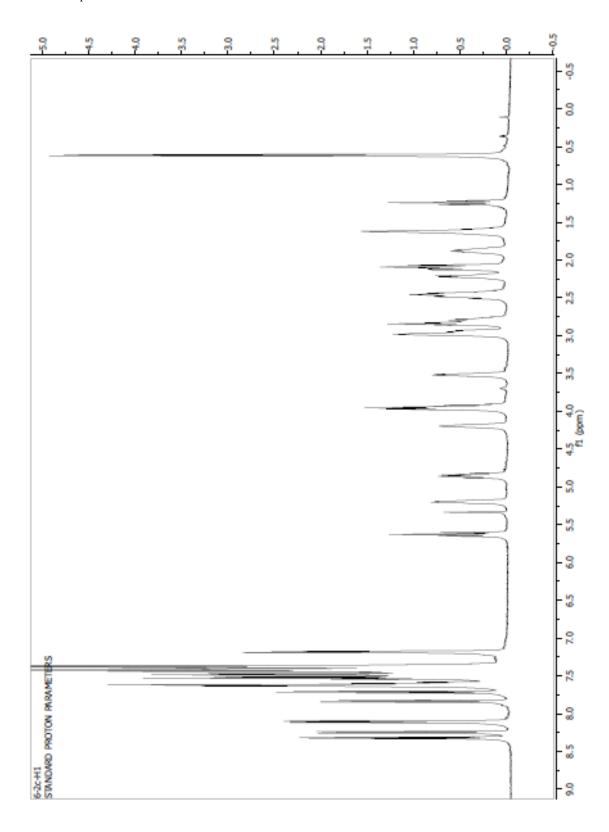
(2,6-difluoro)cinnamyliridium complex 2d: 100.0 mg of Ir(I) ethylene complex 1a-(R,R,R) (0.115 mmol) was dissolved in 5 ml benzene, and 23.9 mg (2,6-difluoro)cinnamylchloride 4c (0.127 mmol) was added. The resulting solution was added to a solution for 22.4 mg AgBF₄ (0.115 mmol) in 2 ml THF, and the resulting mixture was allowed to stir vigorously for 15 min. The precipitate was removed by syringe filtration through a 25 mm diameter and 0.2 µm pore size filter, and the filtrate was left in an argon-filled dry-box with the cap slightly unscrewed for 48 h. Complex 4c (86 mg, 65% yield) crystallized and was collected after drying under vacuum. Anal. Calc. for C₅₉H₅₄BF₆IrNO₂P: 61.24; H, 4.70; N, 1.21. Found: C, 61.32; H, 4.81; N, 1.33. ¹H NMR (500 MHz, THF) δ 8.30 (d, J = 8.9 Hz, 1H), 8.22 (d, J = 8.8 Hz, 1H), 8.08 (dd, J = 8.2, 2.8 Hz, 2H), 7.81 (d, J = 8.8 Hz, 1H), 7.68 (d, J = 8.8 Hz, 1H), 7.64 - 7.54 (m, 2H), 7.51 -7.44 (m, 3H), 7.42 - 7.30 (m, 10H), 7.17 - 7.11 (m, 2H), 7.06 (dd, J = 10.4, 8.6 Hz, 2H), 5.58 (t, J = 12.3 Hz, 1H), 5.48-5.37 (m, 1H), 5.25 (dd, J = 14.6, 6.4 Hz, 1H), 4.44 - 4.25 (m, 1H), 4.01 - 3.81 (m, 2H), 3.49 (dd, J = 20.9, 14.2 Hz, 2H), 3.20 - 3.07 (m, 1H), 2.80 (t, J = 7.1 Hz, 1H), 2.65 - 2.48 (m, 2H), 2.47 - 2.33 (m, 2H), 2.24 (dd, J = 14.6, 7.6 Hz, 1H), 1.99 (d, J = 11.0 Hz, 1H), 1.95 – 1.85 (m, 1H), 1.61 (dddd, J = 18.5, 14.4, 12.0, 6.8 Hz, 2H), 1.17 (t, J = 12.1 Hz, 1H), 0.58 (d, J = 7.4 Hz, 3H). ¹³C NMR (126 MHz, CD₂Cl₂) δ 147.38, 141.96, 141.85, 140.02, 133.14, 132.90, 132.28, 132.20, 132.14, 132.02, 130.91, 129.63, 128.96, 128.89, 128.77, 128.75, 128.49, 128.35, 128.33, 127.69, 127.65, 127.39, 127.16, 126.76, 126.62, 126.17, 124.60, 120.97, 120.95, 120.89, 120.88, 113.66, 113.47, 110.00, 102.39, 93.98, 93.22, 85.34, 74.30, 74.06, 66.42, 66.18, 60.59, 60.55, 54.09, 53.87, 53.65, 53.44, 53.22, 40.48, 35.21, 35.11, 27.88, 27.64, 18.45. ³¹P NMR (202 MHz, THF) δ 115.6.

(2-bromo)cinnamyliridium complex 2e: Ir(I) ethylene complex 1a-(*R*,*R*,*R*) (50.0 mg, 0.058 mmol) was dissolved in 3 ml of either benzene, THF or CH₂Cl₂. (2-bromo)cinnamylchloride (26.8 mg, 0.115 mmol) (4d) was added to the solution. The resulting solution was added to 11.3 mg of AgBF₄ (0.058 mmol) dissolved in 1 ml of THF and allowed to stir vigorously for 15 min. The resulting precipitate was removed by filtration through a 25 mm diameter, 0.2 µm pore size filter, and the filtrate was added dropwise to 10 ml of pentane while stirring vigorously. The precipitated product was collected on a glass frit and dried under vacuum. Yield: 40 mg (60%) from the reaction in benzene, 31 mg (50%) from the reaction in THF, and 36 mg (55%) from the reaction in CH₂Cl₂. Crystals suitable for solid-state structural analysis were obtained by dissolving the complex in a small amount of THF and layering the resulting solution with benzene. Anal. Calc. for C₅₉H₅₅BBrF₄IrNO₂P: 59.05; H, 4.62; N, 1.17. Found: C, 59.17; H, 4.76; N, 1.39. ¹H NMR (500 MHz, CD₂Cl₂) δ 8.33 (d, *J* = 8.9 Hz, 1H), 8.25 (d, *J* = 8.8 Hz, 1H), 8.11 (dd, *J* = 8.2, 2.6 Hz, 2H), 7.85 (d, *J* = 8.8 Hz, 1H), 7.76 (dd, *J* = 7.8, 1.3 Hz, 1H), 7.71 (d, *J* = 8.8 Hz, 1H), 7.66 – 7.57 (m, 3H), 7.55 (t, *J* = 7.6 Hz, 2H), 7.50 – 7.31 (m, 12H), 7.24 – 7.13 (m, 2H), 5.90 (t, *J* = 12.0 Hz, 1H), 5.40 – 5.27 (m, 1H), 5.02 – 4.87 (m, 1H), 4.35 (d, *J* = 5.3 Hz, 1H), 4.07 – 3.86 (m, 2H), 3.64 – 3.56 (m, 1H), 3.23 (b, *J* = 9.3 Hz, 2H), 2.75 (t, *J* = 6.8 Hz, 1H), 2.56 – 2.39 (m, 3H), 2.35 (dd, *J* = 12.0, 4.5 Hz, 1H), 2.24 (dd, *J* = 14.5, 7.8 Hz, 1H), 2.02 (d, *J* = 10.6 Hz, 1H), 1.95 (b, *J* = 19.2 Hz, 1H), 1.74

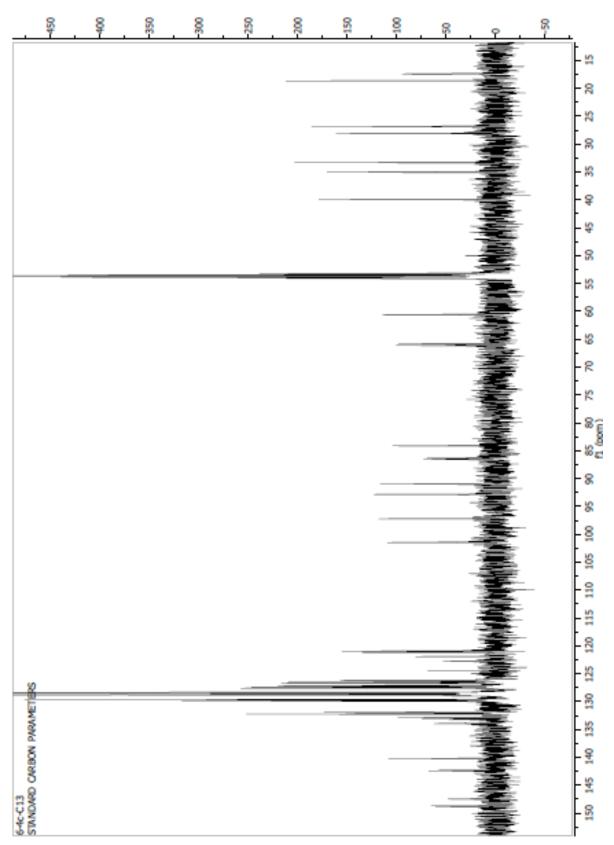
- 1.53 (m, 2H), 1.39 (t, J = 12.4 Hz, 1H), 0.61 (d, J = 7.4 Hz, 3H). ¹³C NMR (126 MHz, CD₂Cl₂) δ 148.74, 148.62, 147.47, 147.40, 142.12, 142.02, 139.99, 135.21, 134.09, 134.03, 133.17, 132.93, 132.28, 132.20, 132.15, 132.06, 130.48, 129.75, 129.12, 128.96, 128.89, 128.79, 128.70, 128.52, 128.42, 127.67, 127.63, 127.42, 127.16, 126.75, 126.61, 126.35, 122.76, 121.86, 121.08, 120.94, 103.13, 99.14, 94.38, 93.15, 86.13, 85.92, 85.66, 65.88, 65.63, 60.66, 38.22, 37.17, 35.07, 28.82, 27.80, 18.58, 16.19. ³¹P NMR (202 MHz, THF) δ 118.5. ³¹P NMR (202 MHz, THF) δ 115.4.

Phenethylallyliridium complex 2f-BF₄: 100.0 mg of [Ir(COD)Cl]₂ (0.149 mmol) and 160.7 mg phosphoramidite ligand L1a-(R,R,R) (0.298 mmol) were dissolved in 4 ml THF and 58.0 mg AgBF₄ (0.298 mmol) dissolved in 1 ml THF was added to the resulting solution followed by 132.0 mg of methyl (5-phenylpent-2-en-1-yl) carbonate. The reaction mixture was allowed to stir for 12 h. The large amount of white precipitate that forms over the course of the reaction was filtered out by syringe filtration through a 25 mm diameter, 0.2 µm pore size filter. The filtrate was added dropwise to 15 ml pentane while stirring vigorously. The precipitated product was collected over a glass frit and dried under vacuum. Yield: 271 mg (85%). ¹H NMR (500 MHz, CD_2Cl_2) δ 8.32 (d, J = 8.9 Hz, 1H), 8.23 (d, J = 8.8 Hz, 1H), 8.11 (dd, J =7.6, 6.4 Hz, 2H), 7.84 (d, J = 8.8 Hz, 1H), 7.67 (d, J = 8.8 Hz, 1H), 7.64 - 7.59 (m, 2H), 7.49 (t, J = 7.5 Hz, 2H), 7.46 -7.27 (m, 15H), 7.08-7.18 (m, 2H), 5.08 (dd, J = 13.9, 7.8 Hz, 1H), 4.45 (dd, J = 19.2, 10.9 Hz, 1H), 4.19 (dd, J = 8.3, 4.0 Hz, 1H), 3.99-3.79 (m, 2H), 3.66 (dd, *J* = 11.8, 5.6 Hz, 1H), 3.38 - 3.20 (m, 2H), 3.12 (dtd, *J* = 21.0, 13.9, 7.4 Hz, 2H), 3.00 (dd, J = 14.6, 8.2 Hz, 1H), 2.79 (t, J = 7.7 Hz, 1H), 2.76 - 2.66 (m, 1H), 2.55 - 2.40 (m, 2H), 2.39 - 2.26 (m, 1H), 2.55 - 2.40 (m, 2H), 2.39 - 2.26 (m, 2H), 2.39 - 2.262.20-2.07 (m, 2H), 1.92 (d, J = 11.3 Hz, 1H), 1.89 – 1.80 (m, 1H), 1.71 – 1.50 (m, 3H), 1.10 (t, J = 11.8 Hz, 1H), 0.58 (d, J = 7.4 Hz, 3H). ³¹P NMR (202 MHz, CDCl₃) δ 120.57. ¹³C NMR (126 MHz, CD₂Cl₂) δ 146.81, 145.44, 140.35, 140.26, 138.67, 138.29, 131.21, 130.94, 130.15, 129.89, 127.51, 127.06, 126.91, 126.70, 126.52, 126.28, 125.55, 125.39, 125.15, 124.75, 124.65, 124.50, 124.31, 120.81, 119.94, 119.13, 119.02, 103.71, 100.51, 90.51, 86.70, 80.47, 76.19, 75.94, 64.03, 63.76, 58.47, 42.09, 34.81, 34.62, 32.98, 30.55, 26.09, 23.92, 16.62, 12.47.

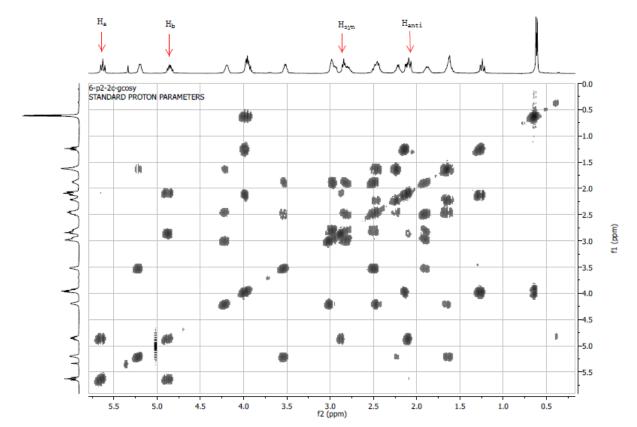
NMR spectra of allyliridium compexes. Cinnamyliridium complex 2c: ¹H-NMR spectrum:



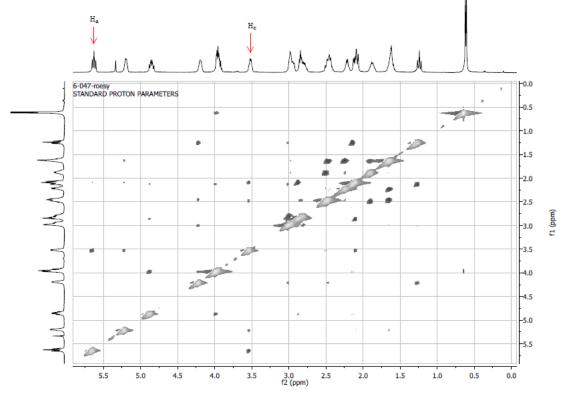
¹³C-NMR spectrum:

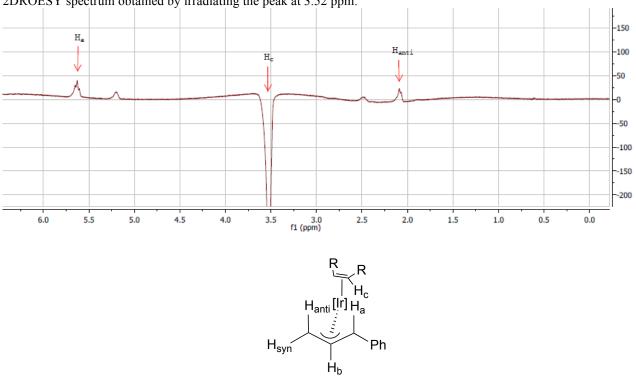


gCOSY spectrum:



2DROESY spectrum



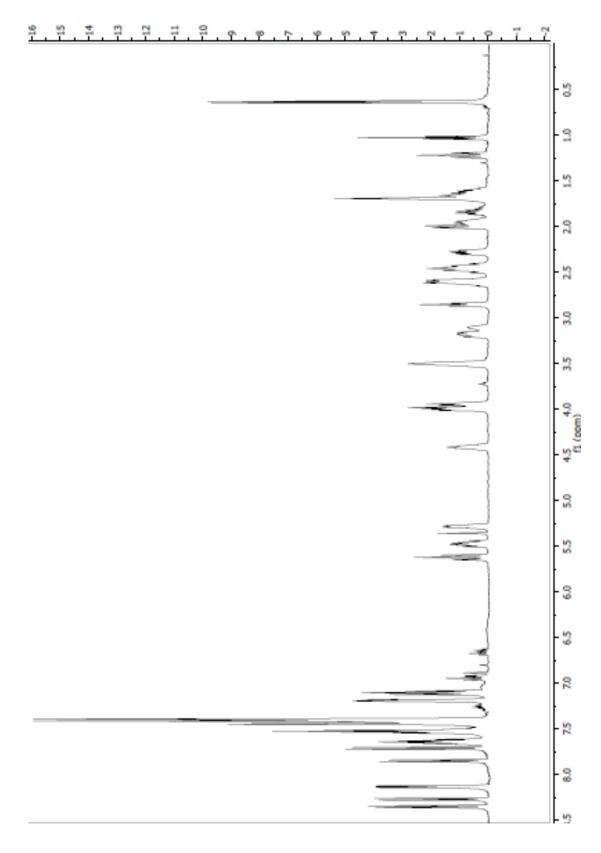


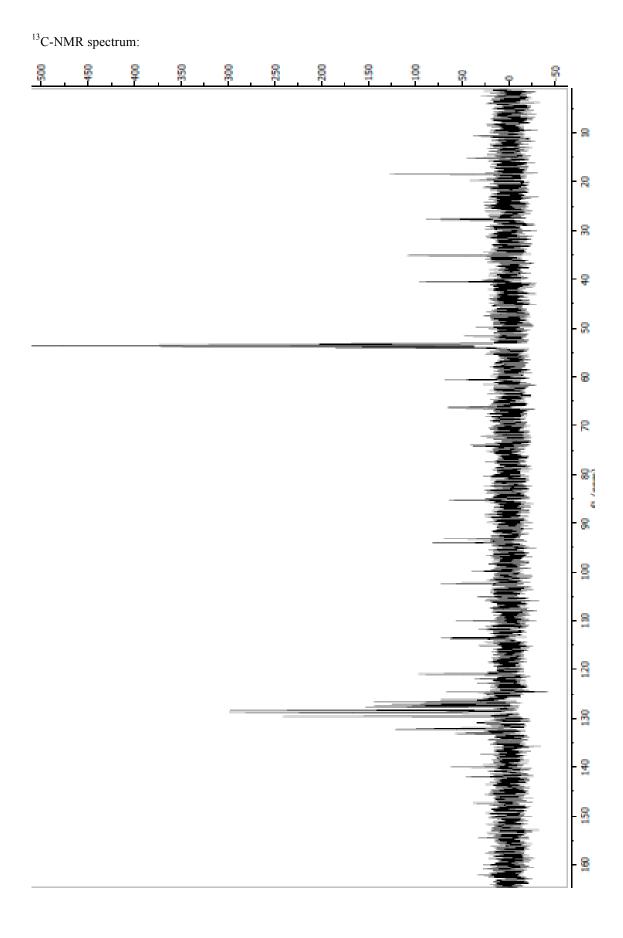
2DROESY spectrum obtained by irradiating the peak at 3.52 ppm.

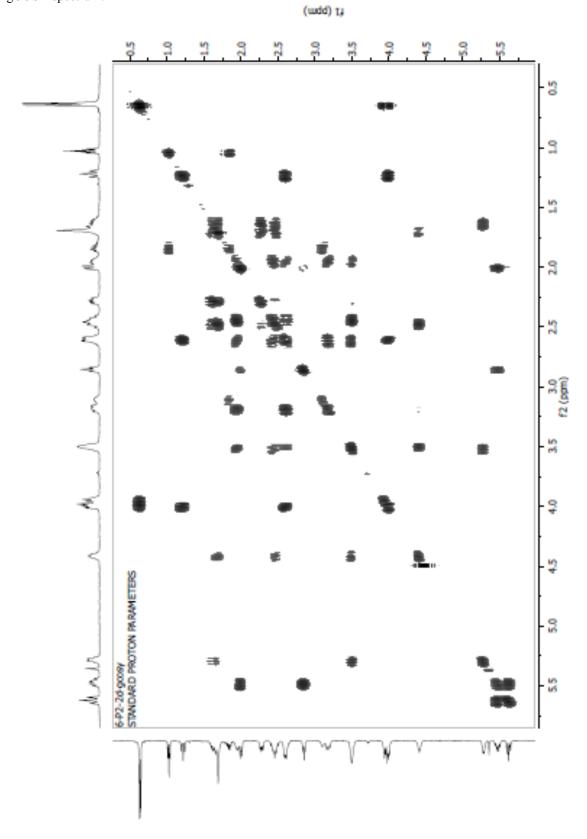
Assignment of the anti and syn allylic hydrogens:

The benzylic proton of the allyl (H_a as depicted in the figure above) should have the most downfield chemical shift among the allylic hydrogens and couple to only one other proton (H_b) in the gCOSY spectrum. Only the signal at 5.61 ppm meets these criteria; this peak was assigned to H_a . The proton coupled to H_a should be H_b and the signal at 4.85 ppm was assigned to H_b. H_b is coupled to two other protons which should be H_{anti} and H_{syn}. The signals at 2.83 and 2.05 are the remaining signals coupled to that of H_b. H_{anti} is located closer to the metal than H_{syn} (2.23 Å vs. 2.77 Å). Because of this structural feature Hanti will experience a stronger shielding from the metal than H_{syn}.⁷⁻⁸ The peak at 2.05 ppm was, therefore, assigned to Hanti and the peak at 2.83 ppm was assigned to Hsvn. This assignment of allylic protons was confirmed by 1D and 2D-ROESY NMR spectroscopy. The solid-state structure of the allyliridium complex shows that both H_a and H_{anti} should be close enough to the olefin proton (labeled as H_c) to observe an NOE (The H_a - H_c distance is 2.011 Å, and the H_{anti}-H_c distance is 2.383 Å), whereas H_{syn} is located too far (The H_{syn}-H_c distance is 4.001 Å) from H_c to observe an NOE. The 2D-ROESY NMR spectrum of the cinnamyliridium complex shows a strong NOE between H_a 5.61 ppm and the multiplet at 3.52 ppm. Thus, the multiplet at 3.52 ppm was assigned to H_c. The 2D-ROESY spectrum also shows that the peak at 3.52 ppm is close in space to the proton with a chemical shift close to 2.05 ppm. Because there are two protons at this chemical shift in ¹H-NMR spectrum, one corresponding to the allylic proton and the other to a proton from cyclooctadiene, we could not immediately assign the chemical shift at 2.05 ppm to H_{syn}. A 1D-ROESY spectrum was obtained with irradiation at the chemical shift of 3.52 ppm (H_c) to observe the peak shape of the coupled proton at 2.05 ppm. The peak shape of the coupled proton at 2.05 ppm is an apparent doublet, which is the peak shape observed in the ¹H-NMR spectrum for the allylic proton. No coupling was observed between H_c and protons with a chemical shift of 2.83. Thus the chemical shift at 2.05 ppm was assigned to H_{anti}, and the chemical shift at 2.83 ppm was assigned to H_{syn}. The outcome of the ROESY-NMR was in good agreement with our assignment of syn and anti allylic protons based on the ¹H NMR chemical shifts.

(2,6-difluoro)cinnamyliridium complex 2d: ¹H-NMR spectrum:

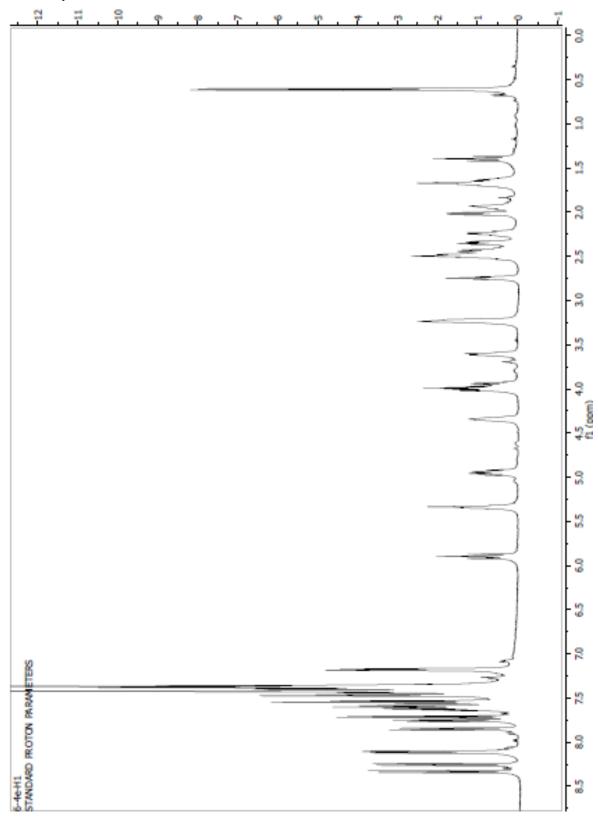


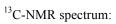


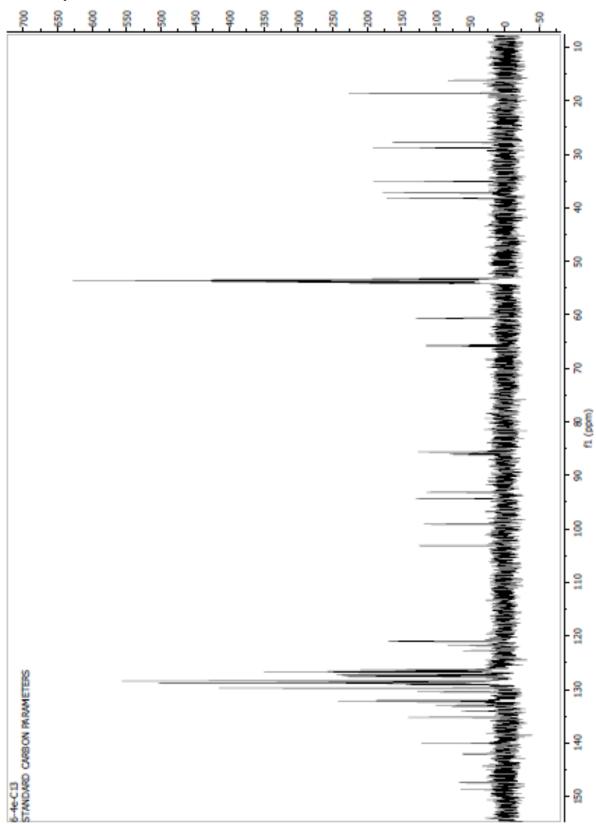


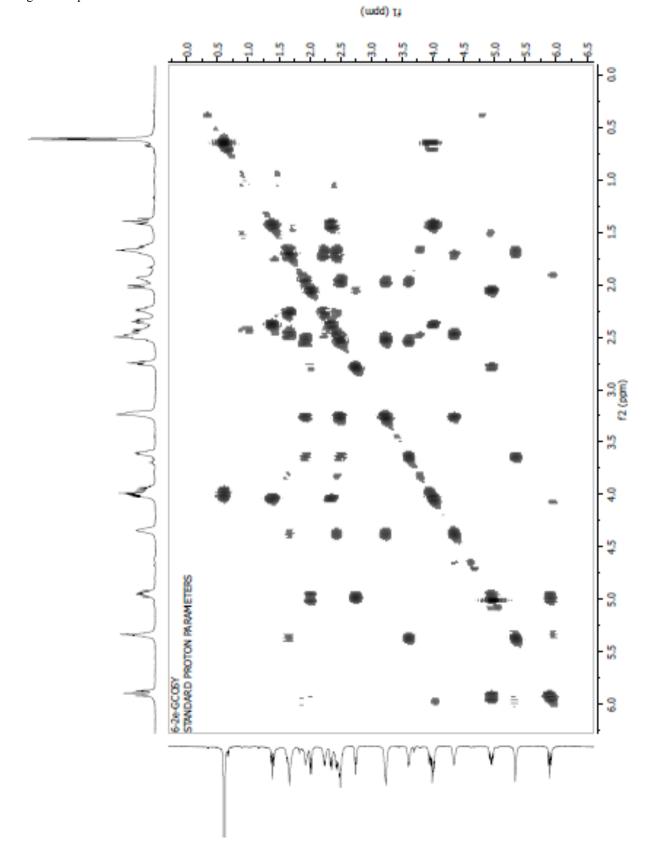
gCOSY spectrum:

(2-bromo)cinnamyliridium complex 2e: ¹H-NMR spectrum:



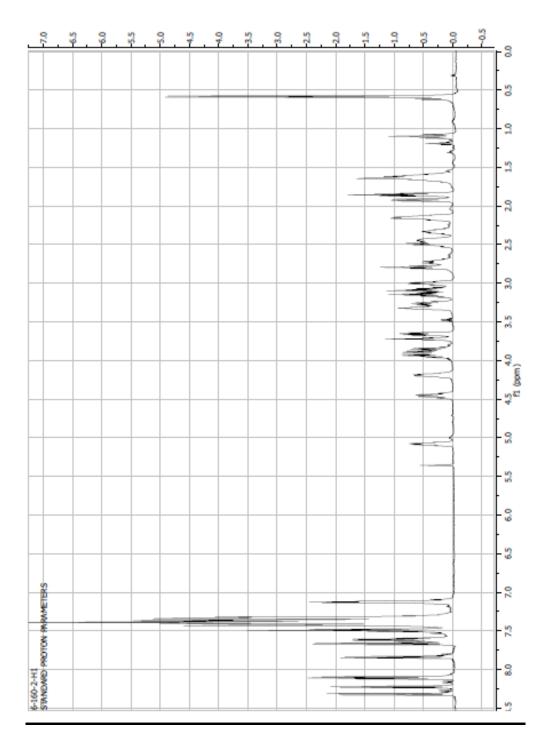




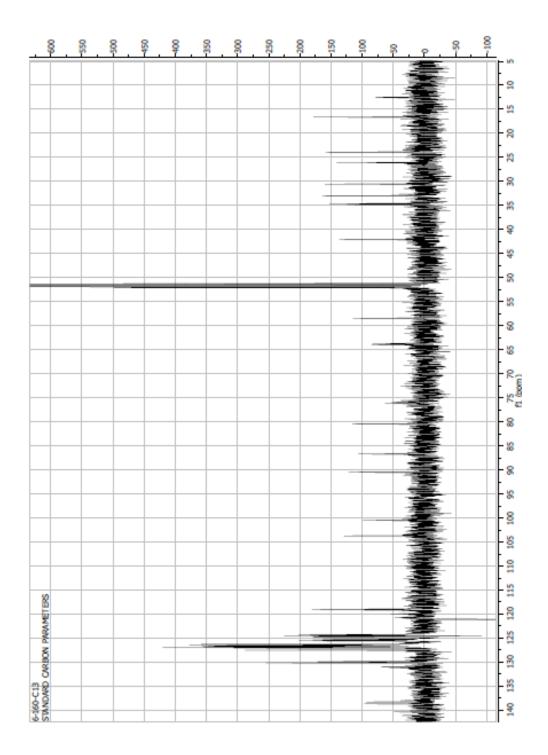


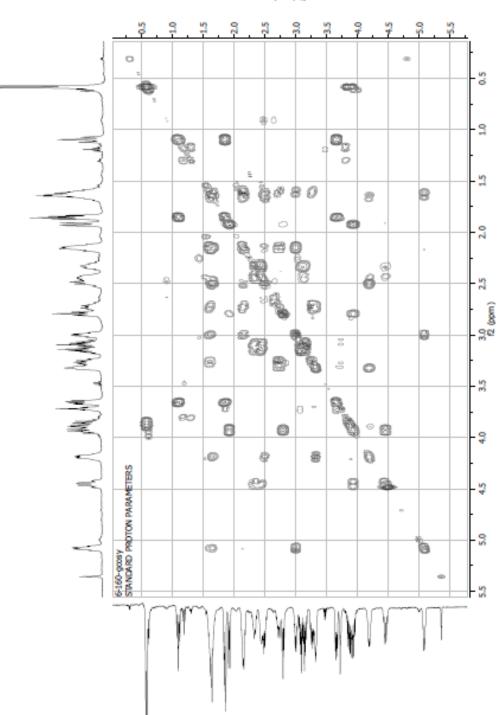
gCOSY spectrum:

Phenethylallyliridium complex 2f-BF4 ¹H-NMR spectrum:



¹³C-NMR spectrum:





(wdd) tj

Kinetic studies:

Rate of interconversion of allyliridium diastereomers:

General procedure for the measurement of the rate of interconversion:

1a-(*S,S,S)* (40 mg, 0.046 mmol) was dissolved in THF (1 ml for the reaction at -40 °C and 0.7 ml for the reaction at -30 °C) and transferred into a screw capped NMR tube with a sealed capillary tube containing ³¹P internal standard. The tube was placed in a dry-ice acetone bath and **5b-(***S***)** was added by syringe (62 μ l, 0.28 mmol). The tube was transferred to NMR-probe cooled to -30 or -40 °C and decay of **1a-(***S,S,S)* as well as formation and decay of **2f-TFA-(***S,S,S,R)* was monitored.

Reaction at -40 °C.

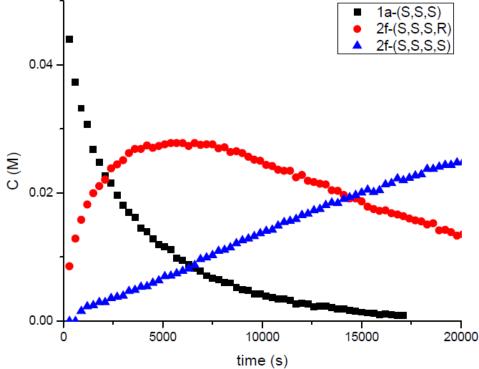


Figure S1. Plots of the change of concentration over time for 1a-(S,S,S), 2f-TFA-(S,S,S,R) and 2f-TFA-(S,S,S,S) at - 40 °C

The 1^{st} order decay of **1a-(***S***,***S***,***S***)** corresponds to oxidative addition of **5b-(***S***)** to **1a-(***S***,***S***,***S***)**. The rate constant for oxidative addition can be determined by plotting $\ln[1a-(S,S,S)]$ versus time.

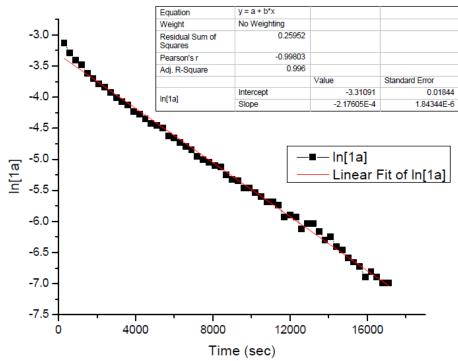


Figure S2. Plot $\ln[1a-(S,S,S)]$ versus time for the reaction of 1a-(S,S,S) with 5b-(S) at -40 °C.



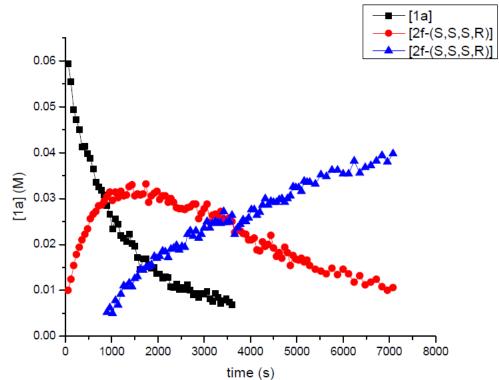


Figure S3. Plots of the change of concentration over time for **1a**-(*S*,*S*,*S*), **2f**-**TFA**-(*S*,*S*,*S*,*R*) and **2f**-**TFA**-(*S*,*S*,*S*,*S*) at - 30 ℃.

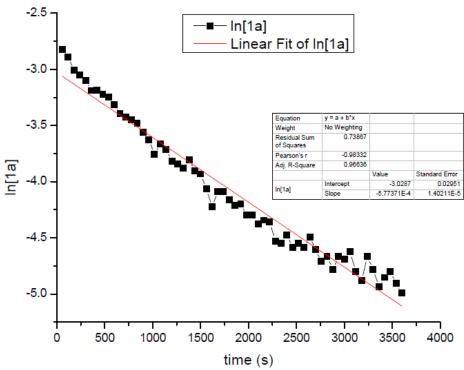


Figure S4. Plot ln[1a-(*S*,*S*,*S*)] versus time for the reaction of 1a-(*S*,*S*,*S*) with 5b-(*S*) at -30 °C.

Calculation of the rate of isomerization of 2f-TFA-(S,S,S,R).⁹⁻¹⁰

2f-TFA-minor is produced by **1a-(***S***,***S***,***S***)** and depleted by production of **2f-TFA-major**. The change in concentration of **2f-TFA-minor** is governed by the relationship:

 $d[2f - minor]/dt = k_1[1a - (S, S, S)] - k_2[2f - minor]$ (1)

Integrating gives the single transcendental equation:

 $[2\mathbf{f} - \mathbf{minor}] = \{ ([1\mathbf{a} - (\mathbf{S}, \mathbf{S}, \mathbf{S})]_0 k_1) / (k_2 - k_1) \} (e^{-k_1 t} - e^{-k_2 t}) (2) \}$

Equation 2 cannot be solved exactly but can be accurately approximated via the mathematical treatment of Emanuel.¹¹ Equation 2 can be simplified to

 $\beta_{max} = \kappa [\kappa/(1-\kappa)]$ (3) (this equation corresponds to the equation 2 in the paper)

where $\beta_{max} = ([\mathbf{2f} - \mathbf{minor}]_{max} / [\mathbf{1a} - (S, S, S)]_0)$ and $\kappa = k_2 / k_1$

The value of βmax was obtained directly from Figure 5 (for -40 °C) or Figure S3 (for -30 °C), and κ can be calculated iteratively so that equation 3 is satisfied. The value of k_2 is calculated from the product of κ and k_1 .

Measurements of the rates of nucleophilic attack

The rate of nucleophlic attack on allyliridium complexes was measured by ³¹P NMR spectroscopy on an Unity Inova 400 MHz instrument at -30, -40 or -60 °C. The rate constants for nucleophilic attack on allyliridium complexes **2c**, **2d**, **2f**(*S*,*S*,*S*,*S*) and **3c** were determined by conducting reactions of the complexes with 10 equiv of tetrabutyl ammonium acetate or a combination of 25 equiv of primary or a secondary amine nucleophile and 25 equiv of triethylamine as proton acceptor. The rate constants for nucleophilic attack were determined by plotting the natural logarithms of concentration vs. time.

Reaction of 2c with aniline at -30 °C in CH₂Cl₂ (table 2, entry 1)

Complex 2c (20.0 mg, 0.0178 mmol) was dissolved in 0.6 ml CH₂Cl₂ and transferred into a screw capped NMR tube and cooled in a dry-ice acetone bath. 41 μ l of aniline (0.45 mmol) and 62 μ l of triethylamine (0.45 mmol) were added to the NMR tube by syrings. The NMR tube was placed into a probe cooled to -30 °C and ³¹P-NMR spectra were recorded every 3 min.

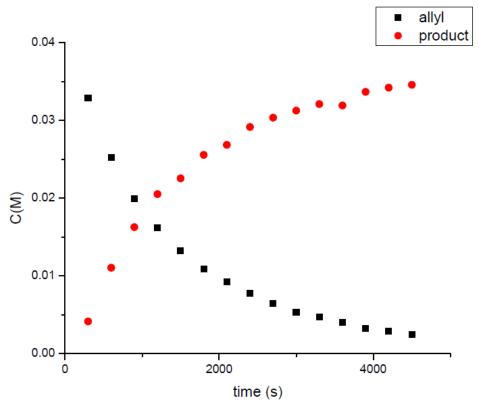
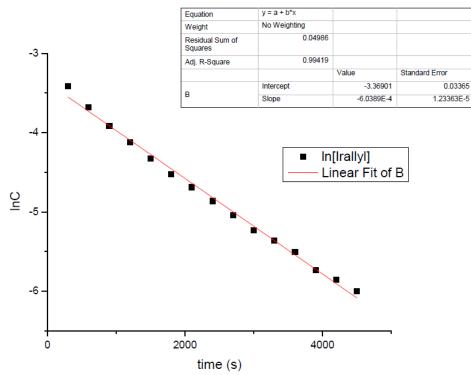
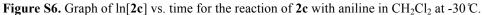


Figure S5. Plot of the change of concentration over time for the reaction of 2c with aniline in CH₂Cl₂ at -30 °C.





Reaction of 2c with aniline at -40 °C in THF (table 2, entry 2)

2c (30 mg, 0.026 mmol) was dissolved in 0.6 ml THF and transferred into a screw capped NMR tube and cooled in a dry-ice acetone bath. 61 μ l aniline (0.65 mmol) and 84 μ l triethylamine (0.445 mmol) were injected into the NMR tube. The NMR tube was placed into a probe cooled to -40 °C and ³¹P-NMR spectra were recorded every min.

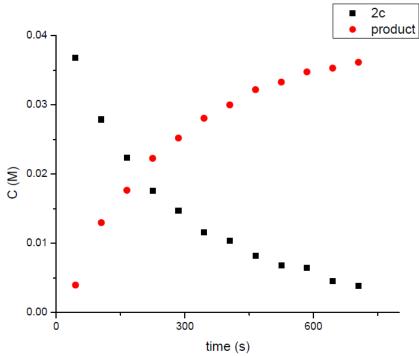


Figure S7. Plot of the change of concentration over time for the reaction of 2c with aniline in THF at -40 °C.

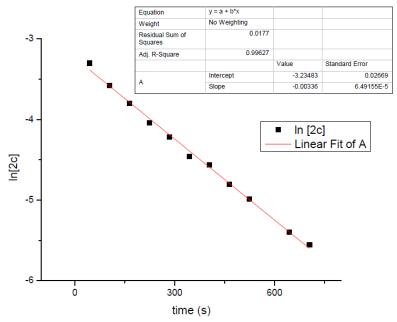


Figure S8. Graph of ln[2c] vs. time for the reaction of 2c with aniline in THF at -40 °C.

Reaction of 2d with aniline at -30 °C in CH₂Cl₂ (table 2, entry 4)

2d (30 mg, 0.026 mmol) was dissolved in 0.6 ml CH₂Cl₂ and transferred into a screw capped NMR tube and cooled in a dry-ice acetone bath. 61 μ l aniline (0.445 mmol) and 84 μ l triethylamine (0.445 mmol) were injected into the NMR tube. The NMR tube was placed into a probe cooled to -30 °C and ³¹P-NMR spectra were recorded every 4 min.

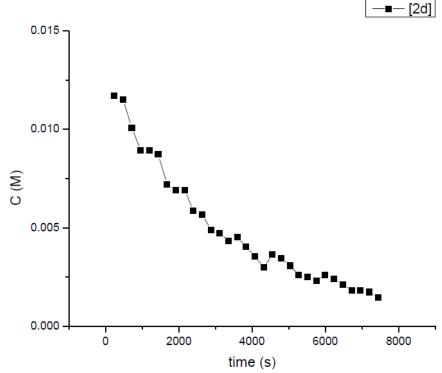


Figure S9. Plot of the change of concentration over time for the reaction of 2d with aniline in CH₂Cl₂ at -30 °C.

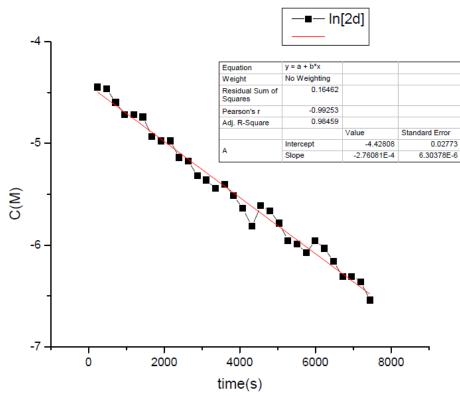


Figure S10. Graph of $\ln[2d]$ vs. time for the reaction of 2d with aniline in CH₂Cl₂ at -30 °C.

Reaction of 2d with N-methyl aniline at -30 °C in CH₂Cl₂ (table 2, entry 5)

2d (40 mg, 0.037 mmol) was dissolved in 0.6 ml CH₂Cl₂ and transferred into a screw-capped NMR tube and cooled in a dry-ice acetone bath. 113 μ l of *N*-methyl aniline (1.05 mmol) and 56 μ l of triethylamine (1.05 mmol) were injected into the NMR tube. The NMR tube was placed into a probe cooled to -30 °C, and ³¹P-NMR spectra were recorded every 4 min.

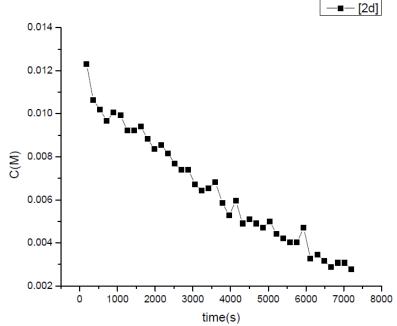


Figure S11. Plot of the change of concentration over time for the reaction of **2d** with N-methyl aniline in CH_2Cl_2 at - 30 °C.

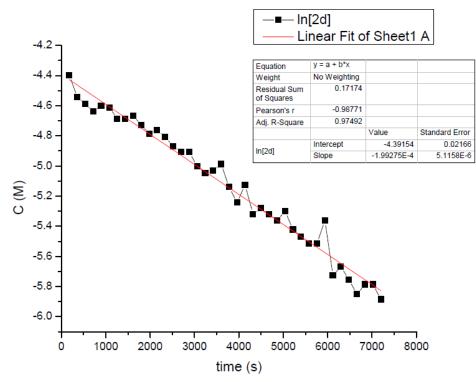


Figure S12. Graph of ln[2d] vs. time for the reaction of 2d with *N*-methyl aniline in CH₂Cl₂ at -30 °C.

Reaction of 3c with aniline at -40 °C in THF (table 2, entry 6)

3c (20 mg, 0.018 mmol) was dissolved in 0.6 ml THF and transferred into a screw capped NMR tube and cooled in a dry-ice acetone bath. 41 μ l aniline (0.45 mmol) and 62 μ l triethylamine (0.45 mmol) were injected into the NMR tube. The NMR tube was placed into a probe cooled to -40 °C and ³¹P-NMR spectra were recorded every 4 min.

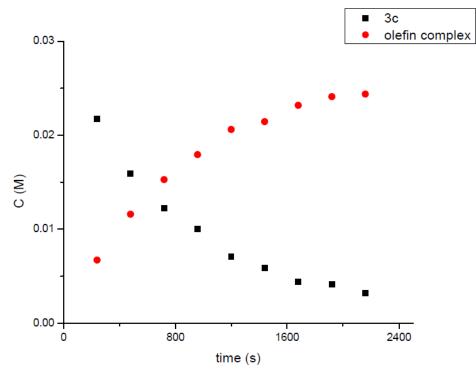


Figure S13. Plot of the change of concentration over time for the reaction of 3c with aniline in THF at -40 °C.

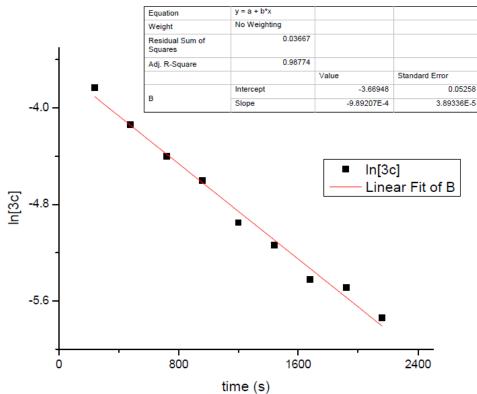


Figure S14. Graph of ln[3c] vs. time for the reaction of 3c with aniline in THF at -40°C.

Reaction of 2f(S,S,S,S) with aniline at -40 °C in THF (table 2, entry 8)

1a(*R*,*R*,*R*) (40 mg, 0.046 mmol) was dissolved in 1 ml THF and 5-phenylpent-2-en-1-yl 2,2,2-trifluoroacetate, **5l** (12 μ l, 51 mmol) was added to the solution and allowed to stir for 15 min. The reaction mixture was transferred into a screw capped NMR tube and cooled in a dry-ice acetone bath. 93 μ l aniline (1 mmol) and 139 μ l triethylamine (1 mmol) were injected into the NMR tube. The NMR tube was placed into a probe cooled to -40 °C and ³¹P-NMR spectra were recorded every min.

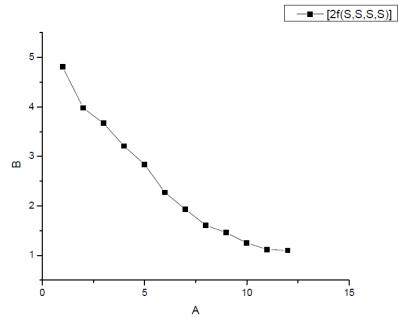


Figure S15. Plot of the change of concentration over time for the reaction of 2f(S,S,S,S) with aniline in THF at -40 °C.

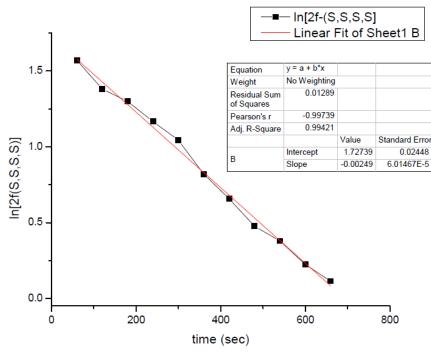


Figure S16. Graph of ln[2f(S,S,S,S)] vs. time for the reaction of 2f(S,S,S,S) with aniline in THF at -40 °C.

Measurement of activation parameters of nucleophilic attack

Reaction of 2f(*R*,*R*,*R*,*R*)-BF₄ with aniline at -40,-30,-20 and -10 °C in THF:

2f(*R*,*R*,*R*)-**BF**₄ (40 mg, 0.037 mmol) was dissolved in 0.5 ml THF and transferred into a screw capped NMR tube and cooled in a dry-ice acetone bath. 86 μ l aniline (0.94 mmol) and 130 μ l triethylamine (0.94 mmol) dissolved in 0.3 ml THF were injected into the NMR tube, making the overall volume of the solution 1 ml. The NMR tube was placed into a probe cooled to -40, -30, -20 or -10 °C and ³¹P-NMR spectra were recorded every 3 min.

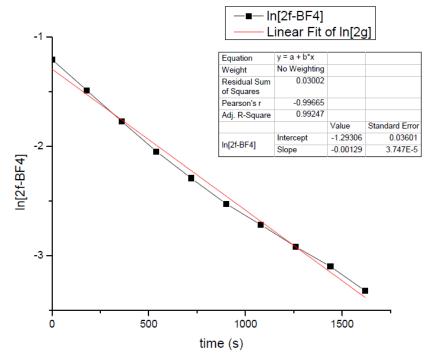


Figure S17. Graph of ln[2f(S,S,S,S)-BF₄] vs. time for the reaction of 2f(S,S,S,S)-BF₄ with aniline in THF at -10 °C.

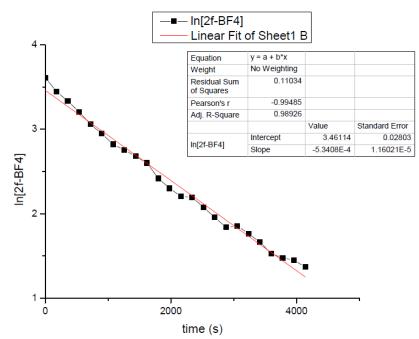


Figure S18. Graph of ln[2f(S,S,S,S)-BF₄] vs. time for the reaction of 2f(S,S,S,S)-BF₄ with aniline in THF at -20 °C.

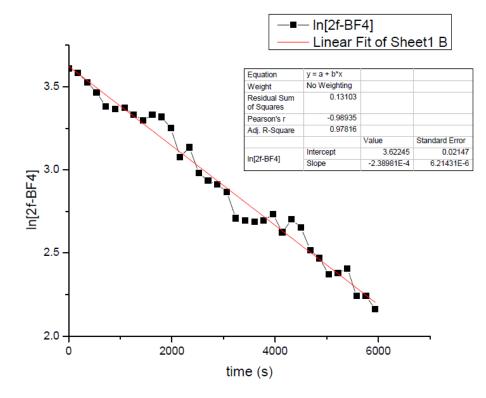


Figure S19. Graph of ln[2f(*S*,*S*,*S*,*S*)-BF₄] vs. time for the reaction of 2f(*S*,*S*,*S*,*S*)-BF₄ with aniline in THF at -30 °C.

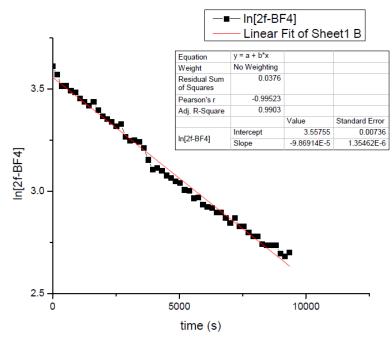


Figure S20. Graph of ln[2f(*S*,*S*,*S*,*S*)-BF₄] vs. time for the reaction of 2f(*S*,*S*,*S*,*S*)-BF₄ with aniline in THF at -40 °C.

Eyring plot for the reaction of 2f-BF₄ with aniline:

The natural logarithms of the rate constants divided by temperature $(\ln(k/T))$ for the reaction of **2f-BF**₄ with aniline measured at -10, -20, -30 and -40 °C were plotted against the reciprocal of the temperature (1/T) to determine the activation parameters for the nucleophilic attack.

	T (°C)	k_{Nu}	T (K)	1/T	ln(k/T)
1	-40	9.9 x 10 ⁻⁵	233	0.004292	-14.671
2	-30	2.4 x 10 ⁻⁴	243	0.004115	-13.827
3	-20	5.3 x 10 ⁻⁴	253	0.003953	-13.076
4	-10	1.3 x 10 ⁻³	263	0.03802	-12.225

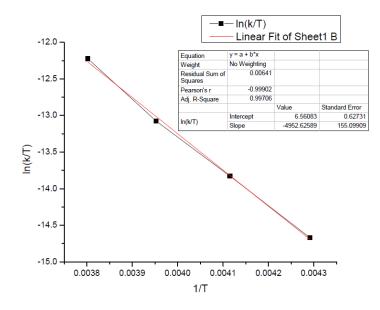


Figure S21. Eyring plot for the reaction of 2f-BF₄ with aniline.

From Figure S21:

Slope = $-4952 = -\Delta H/R$; $\Delta H^{\ddagger}_{nuc} = 4952 \times 8.314 = 41.2 \text{ kJ/mol} = 9.8 \text{ kcal/mol}$

Intercept = $6.6 = \ln(k_B/h) + \Delta S/R$; $\Delta S^{\dagger}_{nuc} = [6.6 - \ln(1.38 \times 10^{-23}/6.27 \times 10^{-34})] \times 8.314 = (6.6-23.8) \times 8.314 = -143 \text{ J/mol} \cdot \text{K} = -34.2 \text{ eu}$

Thus, the nucleophilic attack on a representative allyliridium complex has a typical entropy of activation for a bimolecular reaction.

For the reaction of aniline with 2f-TFA

At -40 °C $\Delta G^{\ddagger}_{nuc} = 75.7 \text{ kJ/mol}$

Estimation of the value of $\Delta\Delta H^{\dagger}$ for nucleophilic attack versus isomerization at -40 °C from $\Delta\Delta G^{\dagger}$ and $\Delta\Delta S^{\dagger}$.

 $\Delta \Delta G^{\dagger} = \Delta \Delta H^{\dagger} - T \bullet \Delta \Delta S^{\dagger} ;$

Assuming that $\Delta S^{\ddagger}_{nuc}$ for **2f-TFA** is similar to that for **2f-BF**₄ and that $\Delta S^{\ddagger}_{epi}$ is close to zero because it is unimolecular reaction,

 $\Delta\Delta S^{\dagger} = -143 \text{ J/mol} \cdot \text{K}$

At T = 233 K;

 $\Delta \Delta H^{\dagger} = \Delta \Delta G^{\dagger} + T \bullet \Delta \Delta S^{\dagger}$

 $\Delta\Delta G^{\dagger}$ = -7.6 kJ/mol from the ratio of rate constants for nucleophilic attack and epimerization of **2f-TFA**.

 $T \cdot \Delta \Delta S^{\dagger} = 143 \text{ x } 233 = -33.3 \text{ kJ/mol}$

And $\Delta\Delta H^{\dagger} = -40.9 \text{ kJ/mol};$

The temperature dependence of the ratio of rate constants of nucleophilic attack and epimerization can be derived as follows:

$$k_{Nu} = \frac{k_B T}{h} e^{\Delta H^{\dagger}/RT + \Delta S^{\dagger}/R} \quad (3)$$

$$k_{epi} = \frac{k_B T}{h} e^{\Delta H^{\dagger}/RT + \Delta S^{\dagger}/R} \quad (4)$$

$$\frac{k_{Nu}}{k_{epi}} = e^{\Delta \Delta H^{\dagger}/RT} \times e^{\Delta \Delta S^{\dagger}/R} \quad (5)$$

$$\frac{\left(\frac{k_{Nu}}{k_{epi}}\right)_{T_1}}{\left(\frac{k_{Nu}}{k_{epi}}\right)_{T_2}} = \frac{e^{\Delta \Delta H^{\dagger}/RT_1} \times e^{\Delta \Delta S^{\dagger}/R}}{e^{\Delta \Delta H^{\dagger}/RT_2} \times e^{\Delta \Delta S^{\dagger}/R}} \quad (6)$$

$$\frac{\left(\frac{k_{Nu}}{k_{epi}}\right)_{T_1}}{\left(\frac{k_{Nu}}{k_{epi}}\right)_{T_2}} = e^{(\Delta \Delta H^{\dagger}/RT_1) - (\Delta \Delta H^{\dagger}/RT_2)} \quad (7)$$

To determine the change in relative rate constants with temperature, the two temperatures and the value of $\Delta\Delta H^{\dagger}$ are used:

For the two temperatures $T_1 = -40$ °C; $T_2 = 25$ °C

 $(k_{Nu}/k_{epi})_{T2}/(k_{Nu}/k_{epi})_{T1} = e^{(-40900/(8.314 \text{ x } 298))} - (-40900/(8.314 \text{ x } 233))] = 100.$

For the two temperatures $T_1 = -40$ °C; $T_2 = 50$ °C

 $(k_{Nu}/k_{epi})_{T2}/(k_{Nu}/k_{epi})_{T1} = e^{(-40900/(8.314 x 323)) - (-40900/(8.314 x 233))]} = 360.$

Thus, a temperature increase from -40 $^{\circ}$ C to 25 $^{\circ}$ C and to 50 $^{\circ}$ C will result in 100 and 360 times increase respectively in the ratio of rate constants for epimerization versus nucleophilic attack.

Catalytic reactions of 2-bromocinnamyl carbonate 5: Ethyl 3-(2-bromophenyl)-2-propen-1-yl carbonate **5** (0.50 mmol, 143 mg) was added to the solution of **1a** (0.02 mmol 17.4 mg) in 1 ml THF followed by nucleophile (0.3 mmol). The solution was allowed to stir for the indicated time at indicated temperature. After removing the solvent yields were determined by ¹H-NMR with mesitylene as internal standart. The products were purified by flash column chromatography through silica using with Hex:EtOAc eluent.

N-(1-(2-bromophenyl)allyl)aniline, 7a: ¹H NMR (500 MHz, CDCl₃) δ 7.58 (dd, J = 8.0, 1.1 Hz, 1H), 7.44 (dd, J = 7.8, 1.7 Hz, 1H), 7.30 – 7.24 (m, 1H), 7.16 – 7.09 (m, 3H), 6.74 – 6.63 (m, 1H), 6.53 (dd, J = 8.6, 0.9 Hz, 2H), 6.04 (ddd, J = 17.1, 10.3, 5.6 Hz, 1H), 5.36 (t, J = 5.4 Hz, 1H), 5.26 (dt, J = 10.3, 1.2 Hz, 1H), 5.24 – 5.19 (m, 1H), 4.13 (b, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 146.89, 140.57, 137.53, 133.33, 129.39, 129.10, 128.58, 128.11, 124.02, 118.04, 117.22, 113.61, 59.48. HRMS-ESI (m/z): [MH]⁺ calcd. for C₁₅H₁₅BrN, 289.1903; found, 289.1901. HPLC conditions: Daicel CHIRALCEL OJ-H (0.46 cm x 25 cm); hexane/2-propanol =99.5/0.5; flow rate = 1 mL/min; detection wave length = 254 nm; TR = 12.1 (*R*), 14.3 (*S*) min.

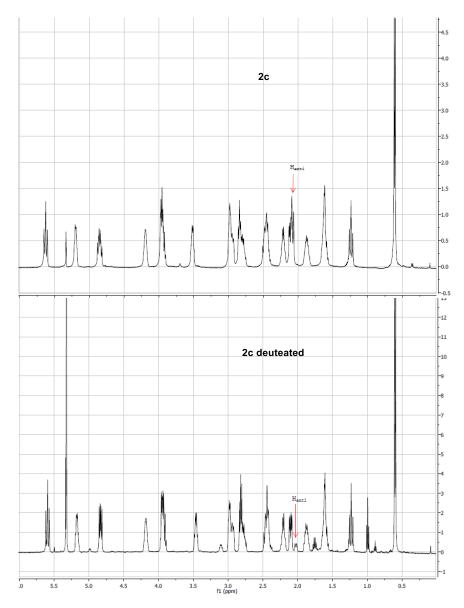
dimethyl 2-(1-(2-bromophenyl)allyl)malonate, 7b: ¹H NMR (499 MHz, CDCl₃) δ 7.58 (dd, J = 8.0, 1.2 Hz, 1H), 7.30 – 7.25 (m, 1H), 7.23 (dd, J = 7.8, 1.7 Hz, 1H), 7.12 – 7.06 (m, 1H), 5.97 (ddd, J = 17.1, 10.2, 8.0 Hz, 1H), 5.15 (dd, J = 15.0, 14.0 Hz, 2H), 4.70 (dd, J = 10.6, 8.1 Hz, 1H), 4.02 (d, J = 10.6 Hz, 1H), 3.75 (s, 3H), 3.56 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 168.20, 167.81, 139.38, 136.43, 133.68, 128.77, 128.66, 127.86, 124.99, 117.92, 56.43, 52.85, 52.77, 48.14. HRMS-ESI (m/z): [MH]⁺ calcd. for C₁₅H₁₅BrN, 328.1784; found, 328.1781. HPLC conditions: Daicel

CHIRALCEL OJ-H (0.46 cm x 25 cm); hexane/2-propanol =95/5; flow rate = 1 mL/min; detection wave length = 254 nm; $t_R = 32.1$ (*S*), 35.3 (*R*) min.

1-bromo-2-(1-phenoxyallyl)benzene, 7c: ¹H NMR (499 MHz, CDCl₃) δ 7.60 (dd, J = 19.8, 7.9 Hz, 2H), 7.40 – 7.23 (m, 3H), 7.23 – 7.09 (m, 1H), 7.01 – 6.82 (m, 3H), 6.25 – 6.03 (m, 2H), 5.48 (d, J = 16.5 Hz, 1H), 5.34 (d, J = 9.9 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 157.51, 139.27, 136.06, 132.87, 129.54, 129.38, 128.36, 128.17, 122.63, 121.17, 117.07, 115.85, 78.67. HRMS-ESI (m/z): [MH]⁺ calcd. for C₁₅H₁₅BrN, 290.1751; found, 290.1750. HPLC conditions: Daicel CHIRALCEL OJ-H (0.46 cm x 25 cm); hexane/2-propanol =99.5/0.5; flow rate = 1 mL/min; detection wave length = 254 nm; t_R = 29.3 (*S*), 31.5 (*R*) min.

Stereochemical route of the reaction: Oxidative addition:

Reaction between 1a and monodeuterated cinnamyl trifluoroacetate, 4a-D: 1a-(R,R,R) (30 mg, 0.035 mmol) was dissolved in 1 ml C₆H₆, and **4a-D** (10.2 mg, 0.042 mmol) was added to it. AgBF₄ (6.8 mg, 0.035 mmol) was dissolved in 0.5 ml THF and added to the first solution. The mixture was allowed to stir for 5 min. Precipitated AgCl was removed by filtration, and the solution was left in the drybox overnight with the cap slightly unscrewed. Crystals formed overnight and were washed with C₆H₆ and dried under vacuum. The obtained crystals were dissolved in CD₂Cl₂, and the ratio between the *syn* and *anti* deuterated complexes was determined by ¹H-NMR spectroscopy.



NMR spectra of fully protiated 2c and 2c obtained after reaction with 4a-D.

Stereochemistry of nucleophilic attack.

Cinnamyliridium complex **2c** monodeutrated at anti allylic hydrogen (40 mg 0.038 mmol) was dissolved in 0.5 ml CH₂Cl₂. Aniline (7 μ l, 0.076 mmol) and triethylamine (11 μ l 0.076 mmol) was added to the solution. The reaction mixture was allowed to stir for 15 min. Triphenylphosphine (20 mg, 0.076 mmol) was added to the solution to free the olefin. The product was purified by preparatory TLC plate chromatography over silicagel with 9:1 Hexane : Ethylacetate eluent. The ratio between the cis and trans deuterated product was determined through ¹H-NMR of the CDCl₃ solution of the product. Ee was determined by chiral HLPC. HPLC conditions: Daicel CHIRALCEL OJ-H (0.46 cm x 25 cm); hexane/2-propanol =99/1; flow rate= 1 mL/min; detection wave length = 254 nm; TR = 29 (R), 31 (S) min.

Catalytic reactions to determine the stereochemical route.

Reactions of 4b-D-(R) with aniline catalyzed by 1a-(R,R,R), 1a-(S,S,S) or 1b-(S,S,S):

In a small, vial linear allylic carbonate **4b-D**-(*R*) (39 mg, 0.2 mmol) was combined with the catalyst (either one of **1a**-(*R*,*R*,*R*) 7 mg, 0.08 mmol; **1a**-(*S*,*S*,*S*) 7mg, 0.08 mmol or **1b**-(*S*,*S*,*S*) 7.5 mg, 0.08 mmol) in 1ml THF. Aniline (22 μ l, 0.24 mmol) was added to each of the vials. The reaction mixture was allowed to stir for 1h. Solvents were removed under reduced pressure. Yields and branched to linear ratios were determined by ¹H-NMR with mesitylene standard. The crude reaction product was purified by flash column chromatography with 9:1 hexane:ethylacetate eluent phase. Ees were determined by chiral HLPC. HPLC conditions: Daicel CHIRALCEL OJ-H (0.46 cm x 25 cm); hexane/2-propanol =99/1; flow rate= 1 mL/min; detection wave length = 254 nm; TR = 29 (R), 31 (S) min.

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