

1 **Response of Anaerobic Granular Sludge to a Shock Load of Zinc Oxide**
2 **Nanoparticles during Biological Wastewater Treatment**

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12 **Synthetic Wastewater.** The synthetic wastewater contained carbon source (glucose and sodium acetate (pH
13 6.9 ± 0.1) with a chemical oxygen demand (COD) ratio of 1:1) and inorganic nutrients. The inorganic
14 nutrients consisted of (mg/L of tap water) 1000 NH₄Cl, 500 KH₂PO₄, 200 CaCl₂, 200 MgCl₂ ·6H₂O, 50 FeCl₃,
15 0.5 H₃BO₃, 0.5 (NH₄)₆Mo₇O₂₄ ·4H₂O, 0.5 ZnSO₄ ·7H₂O, 0.5 CuSO₄ ·5H₂O, 0.5 CoCl₂ ·6H₂O, 0.5 AlCl₃ ·6H₂O,
16 4 EDTA, 1 MnCl₂ ·4H₂O, 1 NiCl₂ 6H₂O, and 34.8 SDBS (as dispersing reagent). The calculated ionic
17 strength of the synthetic wastewater is 32.4 mM.

18 **Detailed Operation of Parent UASB Reactor.** In the initial 15 d, the UASB reactor was operated at an
19 organic loading rate (OLR) of 2.0 kg-COD/m³/day and a substrate COD concentration of 2500 mg/L with a
20 hydraulic retention time (HRT) of 30 h. Then the influent OLR was gradually increased to 5, 8, 12 and 16
21 kg-COD/m³/day, and the corresponding influent COD was 2500, 3200, 4000 and 4000 mg/L, respectively,
22 which resulted in the HRT decreased to 12, 9.6, 8 and 6 h. In each stage the acclimation was not finished
23 until the COD removal efficiency reached relatively stable.

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Table S1. Excitation and Emission Wavelengths for Dyes and Associated Targets.

Dye	Excitation (nm)	Emission (nm)	Targets
FITC	488	500-550	Proteins
Concanavalin A	561	570-590	α -D-glucopyranose polysaccharides
Calcofluor white	400	410-480	β -D-glucopyranose polysaccharides
Nile red	514	625-700	Lipids
SYTO 63	633	650-670	Total cells
SYTOX Blue	458	460-480	Dead cells

Table S2. The Statistical Analysis Results of Different Dosages of ZnO NPs and Their Released Zn²⁺ Affecting Individual Compositions of EPS (Compared with the Control).

			Proteins	Polysaccharides	DNA	Lipids	Humic acids
ZnO NPs (mg/g-TSS)	F _{observed}	2.66	1.00	1.95	4.59	2.09	
10	F _{significance}	7.71	7.71	7.71	7.71	7.71	
	P _(0.05)	0.18	0.37	0.24	0.10	0.22	
	F _{observed}	3.99	5.54	0.48	6.97	0.97	
50	F _{significance}	7.71	7.71	7.71	7.71	7.71	
	P _(0.05)	0.12	0.08	0.53	0.06	0.38	
	F _{observed}	685.60	7.61	73.65	33.83	92.03	
100	F _{significance}	7.71	7.71	7.71	7.71	7.71	
	P _(0.05)	1.26×10 ⁻⁵	0.05	0.00	4.35×10 ⁻³	6.60×10 ⁻⁴	
	F _{observed}	981.92	6.42	259.88	189.05	163.05	
200	F _{significance}	7.71	7.71	7.71	7.71	7.71	
	P _(0.05)	6.18×10 ⁻⁶	0.06	8.66×10 ⁻⁵	1.62×10 ⁻⁴	2.17×10 ⁻⁴	
Zn ²⁺ (mg/L)	F _{observed}	0.00	0.76	0.25	0.44	1.05	
7.8	F _{significance}	7.71	7.71	7.71	7.71	7.71	
	P _(0.05)	0.99	0.43	0.65	0.54	0.36	
	F _{observed}	1.27	2.85	0.74	0.12	0.77	
19.2	F _{significance}	7.71	7.71	7.71	7.71	7.71	
	P _(0.05)	0.32	0.17	0.44	0.75	0.43	
	F _{observed}	409.94	3.19	44.16	34.20	35.67	
30.5	F _{significance}	7.71	7.71	7.71	7.71	7.71	
	P _(0.05)	3.51×10 ⁻⁵	0.15	2.66×10 ⁻³	4.26×10 ⁻³	3.95×10 ⁻⁴	
	F _{observed}	959.48	5.20	113.81	83.55	83.03	
40.9	F _{significance}	7.71	7.71	7.71	7.71	7.71	
	P _(0.05)	6.47×10 ⁻⁶	0.08	4.37×10 ⁻⁴	7.95×10 ⁻⁴	8.05×10 ⁻⁴	

Table S3. Main Functional Groups Observed from FTIR Spectra of EPS Studied.^{1,2}

Wave number (cm ⁻¹)	Vibration type	Functional type
3200–3400	Stretching vibration of OH	OH into polymeric compounds
2960–2930	Asymmetric stretching vibration of CH ₂	
2860	Symmetric stretching vibration of CH ₂	
1740	Stretching vibration of C=O	Carboxylic acids
1635	Stretching vibration of C=O and C–N (Amide I)	Proteins (peptidic bond)
1540	Stretching vibration of C–N and deformation vibration of N–H (Amide II)	Proteins (peptidic bond)
1455	Deformation vibration of CH ₂	
1400	Stretching vibration of C=O	Carboxylates
1240	Deformation vibration of C=O	Carboxylic acids
1100	Stretching vibration C–O–C	Polysaccharides
1055	Stretching vibration of O–H	

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Table S4. The Statistical Analysis Results of Different Dosages of ZnO NPs and Their Released Zn²⁺ Affecting Methane Production (Compared with the Control).

	ZnO NPs (mg/g-TSS)				Zn ²⁺ (mg/L)			
	10	50	100	200	7.8	19.2	30.5	40.9
F _{observed}	0.02	7.05	327.75	771.53	1.01	0.32	104.63	438.23
F _{significance}	7.71	7.71	7.71	7.71	7.71	7.71	7.71	7.71
P _(0.05)	0.90	0.06	15.47×10 ⁻⁵	9.99×10 ⁻⁶	0.37	0.60	5.15×10 ⁻⁴	3.08×10 ⁻⁵

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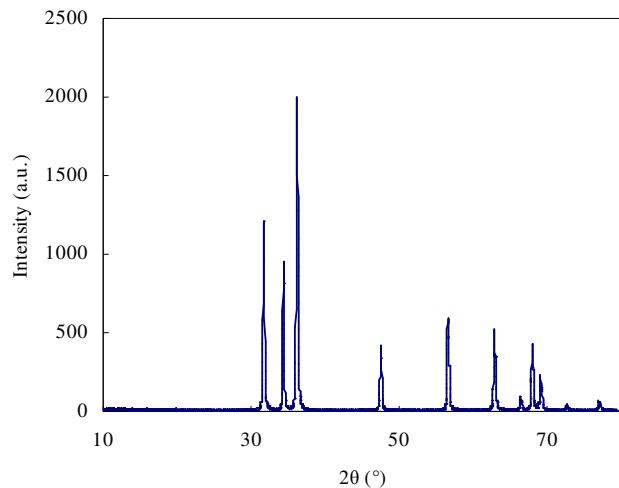


Figure S1. X-ray diffraction (XRD) pattern of ZnO NPs

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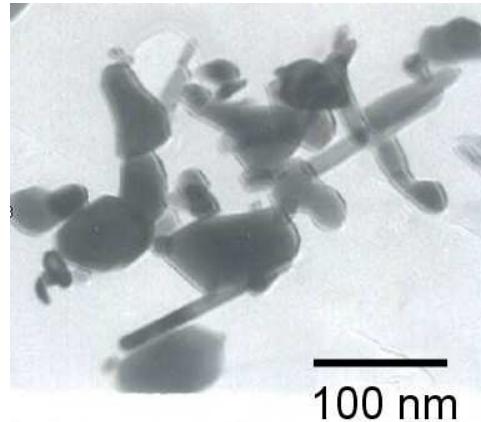
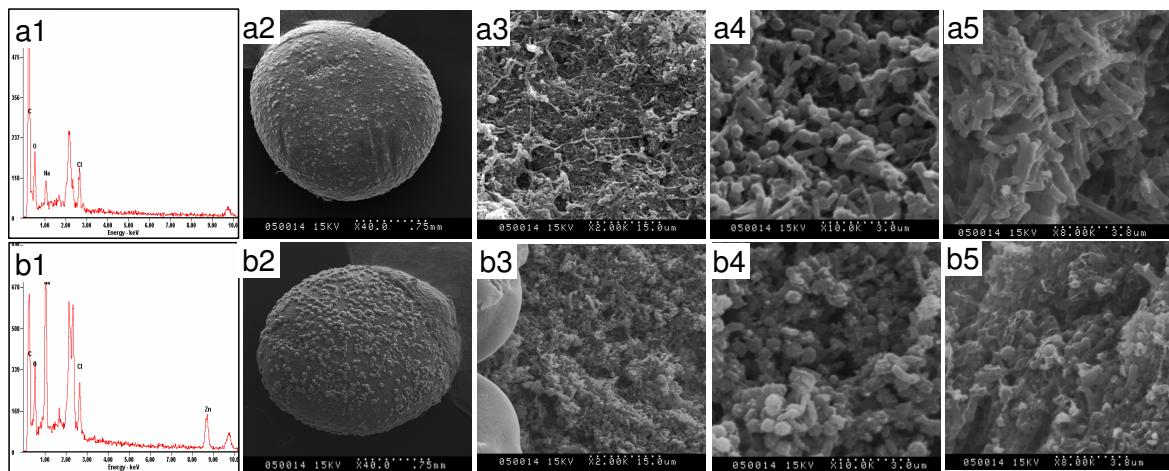
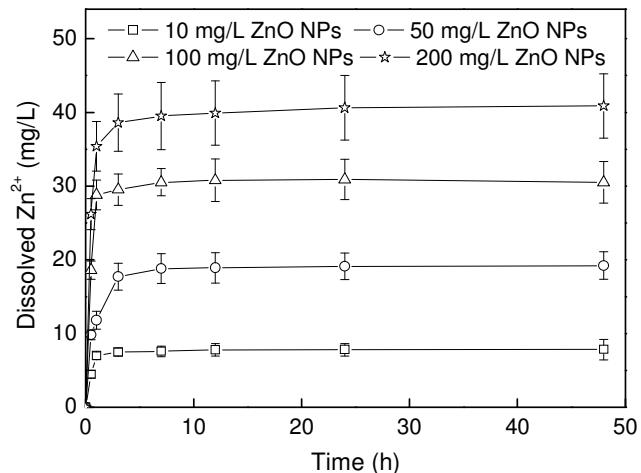


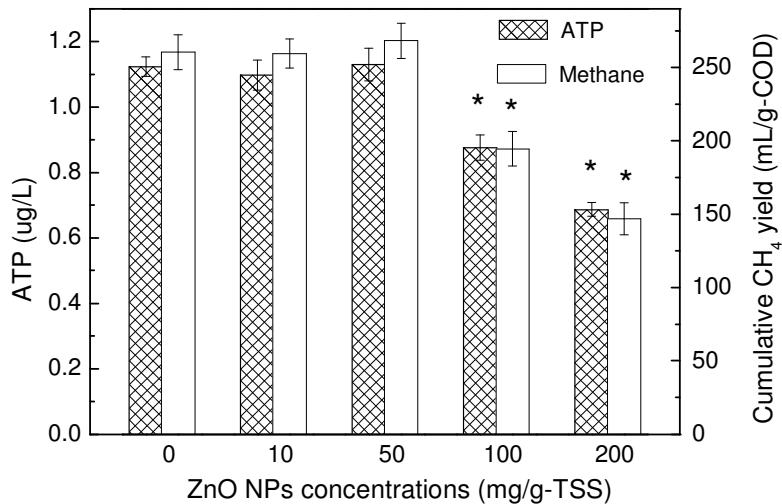
Figure S2. Transmission electron microscopy (TEM) micrograph of ZnO NPs suspension.



35 **Figure S3.** The EDX (a1 and b1) and SEM (a2-a5 and b2-b5) analyses of AGS in the absence (a1-a5)
36 and presence (b1-b5) of 200 mg/g-TSS of ZnO NPs. a2 and b2, a3 and b3, a4 and b4, and a5 and b5
37 represent the morphology, surface at magnification of 2000 and 10000, and interior of AGS, respectively.



39 **Figure S4.** Dissolution dynamic curves of ZnO NPs in synthetic wastewater containing 0.1 mM SDBS
40 (pH=6.90 ± 0.10). Error bars represent standard deviations of triplicate tests.



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42 **Figure S5.** Effect of ZnO NPs on ATP content of AGS. For direct comparison, the total methane
 43 productions in the presence of different dosages of ZnO NPs are also included. Asterisks indicate
 44 statistical differences ($p < 0.05$) from the control. Error bars represent standard deviations of duplicate
 45 tests.

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