

Supporting Information

Synthesis of Enaminones by Rhodium-Catalyzed Denitrogenative Rearrangement of 1-(*N*-Sulfonyl-1,2,3-triazol-4-yl)alkanols

Tomoya Miura, Yuuta Funakoshi, Masao Morimoto, Tsuneaki Biyajima, and Masahiro Murakami

Department of Synthetic Chemistry and Biological Chemistry, Kyoto University,
Katsura, Kyoto 615-8510, Japan

Table of Contents:

S2	General Methods and Materials		
	Spectroscopic Data (1a , 1b)		
S3	Spectroscopic Data (1c , 1d , 1e , 1f , 1g)		
S4	Spectroscopic Data (3a , 3d , 3e , 3f , 3g)		
S5	Spectroscopic Data (3h , 3i , <i>cis</i> - 3j , <i>trans</i> - 3j)		
S6	Typical Procedure for the Denitrogenative Rearrangement Reaction of 1-(<i>N</i> -Tosyl-1,2,3-triazol-4-yl)alkanols (Table 1, entry 1).		
	Spectroscopic Data (2a , 2d , 2e , 2e')		
S7	Spectroscopic Data (2f' , 2g , 2g' , 2h)		
S8	Spectroscopic Data (4a , 4b , 4c , 4d)		
S9	Spectroscopic Data (4e , 4f , 4g , 4h)		
S10	Spectroscopic Data (4i , 4j , 4j')		
	Typical Procedure for the One-pot Synthesis of Enaminones from Propargylic Alcohols (Eq 3).		
S11	Spectroscopic Data (4k , 6 , 7)		
S12	Spectroscopic Data (8 , 9)		
S13	Spectroscopic Data (10)		
S14–15	¹ H and ¹³ C NMR Spectra of 1a	S65–66	¹ H and ¹³ C NMR Spectra of 4c
S16–17	¹ H and ¹³ C NMR Spectra of 1b	S67–68	¹ H and ¹³ C NMR Spectra of 4d
S18–19	¹ H and ¹³ C NMR Spectra of 1c	S69–70	¹ H and ¹³ C NMR Spectra of 4e
S20–21	¹ H and ¹³ C NMR Spectra of 1d	S71–72	¹ H and ¹³ C NMR Spectra of 4f
S22–23	¹ H and ¹³ C NMR Spectra of 1e	S73–74	¹ H and ¹³ C NMR Spectra of 4g
S24–25	¹ H and ¹³ C NMR Spectra of 1f	S75–76	¹ H and ¹³ C NMR Spectra of 4h
S26–27	¹ H and ¹³ C NMR Spectra of 1g	S77–78	¹ H and ¹³ C NMR Spectra of 4i
S28–29	¹ H and ¹³ C NMR Spectra of 3a	S79–80	¹ H and ¹³ C NMR Spectra of 4j
S30–31	¹ H and ¹³ C NMR Spectra of 3d	S81–82	¹ H and ¹³ C NMR Spectra of 4j'
S32–33	¹ H and ¹³ C NMR Spectra of 3e	S83–84	¹ H and ¹³ C NMR Spectra of 4k
S34–35	¹ H and ¹³ C NMR Spectra of 3f	S85–86	¹ H and ¹³ C NMR Spectra of 6
S36–37	¹ H and ¹³ C NMR Spectra of 3g	S87–88	¹ H and ¹³ C NMR Spectra of 7
S38–39	¹ H and ¹³ C NMR Spectra of 3h	S89–90	¹ H and ¹³ C NMR Spectra of 8
S40–41	¹ H and ¹³ C NMR Spectra of 3i	S91–92	¹ H and ¹³ C NMR Spectra of 9
S42–43	¹ H and ¹³ C NMR Spectra of <i>cis</i> - 3j	S93–94	¹ H and ¹³ C NMR Spectra of 10
S44–45	¹ H and ¹³ C NMR Spectra of <i>trans</i> - 3j	S95–96	¹ H and ¹³ C NMR Spectra of 11
S46–47	¹ H and ¹³ C NMR Spectra of 2a		
S48–49	¹ H and ¹³ C NMR Spectra of 2d		
S50–51	¹ H and ¹³ C NMR Spectra of 2e		
S52–53	¹ H and ¹³ C NMR Spectra of 2e'		
S54	¹ H of 2f'		
S55–56	¹ H and ¹³ C NMR Spectra of 2g		
S57–58	¹ H and ¹³ C NMR Spectra of 2g'		
S59–60	¹ H and ¹³ C NMR Spectra of 2h		
S61–62	¹ H and ¹³ C NMR Spectra of 4a		
S63–64	¹ H and ¹³ C NMR Spectra of 4b		

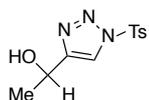
General Methods.

Rhodium(II)-catalyzed reactions were carried out with a Biotage Initiator 2.5 microwave synthesizer. IR measurements were performed on a FTIR SHIMADZU DR-8000 spectrometer fitted with a Pike Technologies MIRacle Single Reflection ATR adapter. ^1H and ^{13}C NMR spectra were recorded on a Varian Mercury-vx400 (^1H at 400.44 MHz and ^{13}C at 100.69 MHz) spectrometer. NMR data were obtained in CDCl_3 . Proton chemical shifts were referenced to the residual proton signal of the solvent at 7.26 ppm (CHCl_3). Carbon chemical shifts were referenced to the carbon signal of the solvent at 77.0 ppm (CDCl_3). High-resolution mass spectra were recorded on a Thermo Scientific Exactive (ESI and APCI) spectrometer. Flash column chromatography was performed with silica gel 60N (Kanto) and diol-silica gel DIOL MB 100–40/75 (Fuji Silysia Chemical Ltd.). Preparative thin-layer chromatography (PTLC) was performed on silica gel plates with PF254 indicator (Merck). Recycling preparative HPLC was carried out on COSMOSIL SSL-II (Nacalai) with a Japan Analytical Industry LC-9110 NEXT. Gel permeation chromatography (GPC) was carried out with a Japan Analytical Industry LC-908.

Materials.

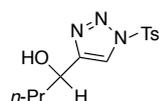
Chloroform (Wako, dehydrated, amylene as stabilizer) was distilled from phosphorus oxide (Wako). Toluene (Nacalai) was used as received from the commercial sources. $\text{Rh}_2(\text{Oct})_4$ (Aldrich), $\text{Cu}(\text{OAc})_2 \cdot \text{H}_2\text{O}$ (Wako), and *o*-aminophenol (nacalai) were used as received from the commercial sources. 3-Butyn-2-ol (**5a**, Aldrich), 1-ethynyl-1-cyclohexanol (**5c**, TCI), mestranol (**5k**, TCI) were used as received from the commercial sources. 1-(*N*-Sulfonyl-1,2,3-triazol-4-yl)alkanols **1a–h** and 1-(*N*-sulfonyl-1,2,3-triazol-4-yl)cycloalkanols **3a–j** were prepared from the corresponding propargylic alcohols according to the literature procedures.^{1,2} The analytical data of compounds **1h**,² **2b**,³ **2c**,³ **2f**,⁴ **3b**,¹ and **3c**¹ have already reported.

1a:



IR (ATR): 3315, 2978, 1595, 1394, 1192, 1178, 1113, 1009 cm^{-1} ; ^1H NMR: δ = 1.57 (d, J = 6.8 Hz, 3H), 2.12–2.36 (br, 1H), 2.44 (s, 3H), 5.06 (q, J = 6.4 Hz, 1H), 7.38 (d, J = 8.0 Hz, 2H), 7.98 (d, J = 8.4 Hz, 2H), 8.05 (s, 1H); ^{13}C NMR: δ = 21.7, 22.8, 62.5, 120.3, 128.5, 130.4, 132.7, 147.3, 152.1; HRMS (ESI⁺): Calcd for $\text{C}_{11}\text{H}_{14}\text{N}_3\text{O}_3\text{S}$, $\text{M}+\text{H}^+$ 268.0750. Found m/z 268.0743.

1b:



IR (ATR): 3342, 3267, 3153, 2955, 2870, 1595, 1387, 1171, 1018, 980 cm^{-1} ; ^1H NMR: δ = 0.94 (t, J = 7.6 Hz, 3H), 1.31–1.55 (m, 2H), 1.74–1.91 (m, 2H), 2.36–2.68 (br, 1H), 2.45 (s, 3H), 4.89 (dd, J = 7.2, 5.6 Hz, 1H), 7.38 (d, J = 8.0 Hz, 2H), 7.98 (d, J = 8.4 Hz, 2H), 8.04 (s, 1H); ^{13}C NMR: δ = 13.6, 18.3, 21.7, 39.0, 66.3, 120.5, 128.5, 130.3, 132.8, 147.3, 151.4; HRMS (ESI⁺): Calcd for $\text{C}_{13}\text{H}_{18}\text{N}_3\text{O}_3\text{S}$, $\text{M}+\text{H}^+$ 296.1063. Found m/z 296.1055.

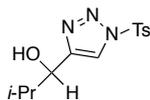
1 Raushel, J.; Fokin, V. V. *Org. Lett.* **2010**, *12*, 4952.

2 Liu, Y.; Wang, X.; Xu, J.; Zhang, Q.; Zhao, Y.; Hu, Y. *Tetrahedron* **2011**, *67*, 6294.

3 Liu, P.; Shan, G.; Chen, S.; Rao, Y. *Tetrahedron Lett.* **2012**, *53*, 936.

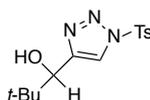
4 Xiao, F.; Wang, J. *J. Org. Chem.* **2006**, *71*, 5789

1c:



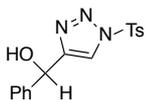
IR (ATR): 3298, 3101, 2968, 1593, 1393, 1379, 1194, 1177, 1024, 988 cm^{-1} ; ^1H NMR: δ = 0.81–0.98 (m, 6H), 2.06–2.18 (m, 1H), 2.45 (s, 3H), 2.66–3.46 (br, 1H), 4.67 (d, J = 5.6 Hz, 1H), 7.38 (d, J = 8.0 Hz, 2H), 7.98 (d, J = 8.0 Hz, 2H), 8.04 (s, 1H); ^{13}C NMR: δ = 16.9, 18.2, 21.6, 33.7, 71.6, 121.1, 128.4, 130.3, 132.7, 147.2, 150.2; HRMS (ESI⁺): Calcd for $\text{C}_{13}\text{H}_{18}\text{N}_3\text{O}_3\text{S}$, $\text{M}+\text{H}^+$ 296.1063. Found m/z 296.1055.

1d:



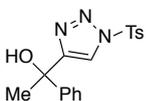
IR (ATR): 3263, 3103, 2968, 1389, 1196, 1177, 1057, 1024, 1016, 982 cm^{-1} ; ^1H NMR: δ = 0.92 (s, 9H), 2.02–2.48 (br, 1H), 2.45 (s, 3H), 4.57 (s, 1H), 7.38 (d, J = 8.0 Hz, 2H), 7.98 (d, J = 8.4 Hz, 2H), 8.02 (s, 1H); ^{13}C NMR: δ = 21.7, 25.3, 35.3, 75.0, 121.3, 128.5, 130.4, 132.9, 147.3, 149.0; HRMS (ESI⁺): Calcd for $\text{C}_{14}\text{H}_{20}\text{N}_3\text{O}_3\text{S}$, $\text{M}+\text{H}^+$ 310.1220. Found m/z 310.1211.

1e:



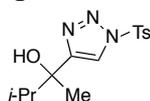
IR (ATR): 3336, 3155, 1593, 1456, 1387, 1217, 1194, 1177, 1043, 1011, 966 cm^{-1} ; ^1H NMR: δ = 2.44 (s, 3H), 2.44–3.16 (br, 1H), 5.98 (s, 1H), 7.29–7.43 (m, 7H), 7.87 (s, 1H), 7.96 (d, J = 8.8 Hz, 2H); ^{13}C NMR: δ = 21.7, 68.6, 121.2, 126.3, 128.2, 128.6, 130.3, 132.6, 140.8, 147.3, 150.9; HRMS (ESI⁺): Calcd for $\text{C}_{16}\text{H}_{16}\text{N}_3\text{O}_3\text{S}$, $\text{M}+\text{H}^+$ 330.0907. Found m/z 330.0897.

1f:

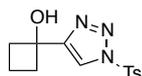


IR (ATR): 3422, 3162, 1391, 1196, 1177, 1138, 1113, 1005, 986 cm^{-1} ; ^1H NMR: δ = 1.96 (s, 3H), 2.45 (s, 3H), 2.75–3.05 (br, 1H), 7.24–7.30 (m, 1H), 7.30–7.37 (m, 2H), 7.38 (d, J = 8.0 Hz, 2H), 7.43–7.48 (m, 2H), 7.92 (s, 1H), 7.98 (d, J = 8.4 Hz, 2H); ^{13}C NMR: δ = 21.7, 30.3, 71.9, 120.4, 125.0, 127.4, 128.2, 128.6, 130.4, 132.7, 145.3, 147.3, 154.4; HRMS (ESI⁺): Calcd for $\text{C}_{17}\text{H}_{18}\text{N}_3\text{O}_3\text{S}$, $\text{M}+\text{H}^+$ 344.1063. Found m/z 344.1053.

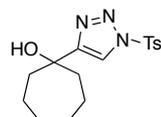
1g:



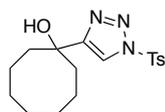
IR (ATR): 3422, 3123, 2966, 1593, 1385, 1192, 1178, 1092, 999 cm^{-1} ; ^1H NMR: δ = 0.81–0.89 (m, 6H), 1.53 (s, 3H), 2.08–2.34 (br, 1H), 2.10 (sept, J = 6.8 Hz, 1H), 2.45 (s, 3H), 7.39 (d, J = 8.0 Hz, 2H), 7.991 (d, J = 7.6 Hz, 2H), 7.995 (s, 1H); ^{13}C NMR: δ = 16.8, 17.0, 21.7, 24.8, 37.8, 73.4, 120.3, 128.5, 130.3, 132.9, 147.2, 154.0; HRMS (ESI⁺): Calcd for $\text{C}_{14}\text{H}_{20}\text{N}_3\text{O}_3\text{S}$, $\text{M}+\text{H}^+$ 310.1220. Found m/z 310.1211.

3a:

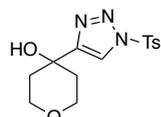
IR (ATR): 3287, 3113, 1593, 1396, 1196, 1177, 1015 cm^{-1} ; ^1H NMR: δ = 1.74–1.91 (m, 1H), 1.87–2.02 (m, 1H), 2.30–2.44 (m, 2H), 2.44 (s, 3H), 2.47–2.59 (m, 2H), 2.80–2.98 (br, 1H), 7.38 (d, J = 8.0 Hz, 2H), 7.99 (d, J = 8.4 Hz, 2H), 8.07 (s, 1H); ^{13}C NMR: δ = 12.5, 21.8, 37.1, 71.8, 119.5, 128.7, 130.4, 132.9, 147.3, 153.2; HRMS (ESI⁺): Calcd for $\text{C}_{13}\text{H}_{16}\text{N}_3\text{O}_3\text{S}$, $\text{M}+\text{H}^+$ 294.0907. Found m/z 294.0902.

3d:

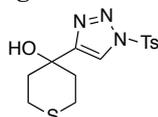
IR (ATR): 3385, 3148, 2920, 1385, 1192, 1177 cm^{-1} ; ^1H NMR: δ = 1.48–1.77 (m, 8H), 1.96 (dd, J = 14.4, 8.4 Hz, 2H), 2.10 (dd, J = 14.8, 10.0 Hz, 2H), 2.28–2.44 (br, 1H), 2.44 (s, 3H), 7.38 (d, J = 8.0 Hz, 2H), 7.98 (d, J = 8.4 Hz, 2H), 8.00 (s, 1H); ^{13}C NMR: δ = 21.77, 21.81, 29.2, 41.8, 73.4, 119.1, 128.7, 130.4, 133.0, 147.2, 156.3; HRMS (ESI⁺): Calcd for $\text{C}_{16}\text{H}_{22}\text{N}_3\text{O}_3\text{S}$, $\text{M}+\text{H}^+$ 336.1376. Found m/z 336.1371.

3e:

IR (ATR): 3487, 3130, 2895, 2843, 1593, 1389, 1194, 1178, 1013, 999 cm^{-1} ; ^1H NMR: δ = 1.42–1.62 (m, 5H), 1.56–1.76 (m, 5H), 2.03–2.11 (m, 4H), 2.23–2.27 (br, 1H), 2.45 (s, 3H), 7.38 (d, J = 8.0 Hz, 2H), 7.99 (d, J = 8.0 Hz, 2H), 8.00 (s, 1H); ^{13}C NMR: δ = 21.6, 21.8, 24.5, 28.0, 36.5, 73.1, 119.6, 128.7, 130.4, 133.0, 147.3, 155.0; HRMS (ESI⁺): Calcd for $\text{C}_{17}\text{H}_{24}\text{N}_3\text{O}_3\text{S}$, $\text{M}+\text{H}^+$ 350.1533. Found m/z 350.1526.

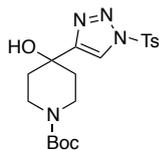
3f:

IR (ATR): 3402, 3125, 2860, 1595, 1389, 1196, 1180, 1020 cm^{-1} ; ^1H NMR: δ = 1.83 (d, J = 13.2 Hz, 2H), 2.17 (td, J = 12.4, 4.8 Hz, 2H), 2.46 (s, 3H), 3.79 (d, J = 11.6 Hz, 2H), 3.89 (t, J = 10.8 Hz, 2H), 7.40 (d, J = 8.0 Hz, 2H), 8.00 (d, J = 8.4 Hz, 2H), 8.03 (s, 1H); ^{13}C NMR: δ = 21.8, 37.8, 63.3, 66.9, 119.5, 128.7, 130.5, 132.8, 147.5, 154.1; HRMS (ESI⁺): Calcd for $\text{C}_{14}\text{H}_{18}\text{N}_3\text{O}_4\text{S}$, $\text{M}+\text{H}^+$ 324.1013. Found m/z 324.1007.

3g:

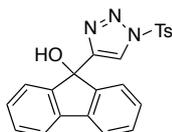
IR (ATR): 3400, 3153, 2980, 1591, 1391, 1194, 1184, 1020 cm^{-1} ; ^1H NMR: δ = 2.13–2.25 (m, 4H), 2.34–2.48 (br, 1H), 2.46 (s, 3H), 2.44–2.53 (m, 2H), 3.05–3.15 (m, 2H), 7.40 (d, J = 8.8 Hz, 2H), 7.995 (d, J = 8.0 Hz, 2H), 8.003 (s, 1H); ^{13}C NMR: δ = 21.8, 23.7, 38.5, 68.1, 119.3, 128.8, 130.5, 132.8, 147.5, 154.6; HRMS (ESI⁺): Calcd for $\text{C}_{14}\text{H}_{18}\text{N}_3\text{O}_3\text{S}_2$, $\text{M}+\text{H}^+$ 340.0784. Found m/z 340.0780.

3h:



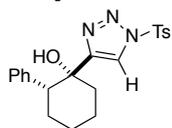
IR (ATR): 3427, 3161, 1666, 1591, 1427, 1387, 1173, 1146, 1076, 989 cm^{-1} ; ^1H NMR: δ = 1.46 (s, 9H), 1.85 (d, J = 12.4 Hz, 2H), 2.01 (td, J = 12.0, 4.8 Hz, 2H), 2.46 (s, 3H), 3.29 (t, J = 10.8 Hz, 2H), 3.87 (br, 2H), 7.40 (d, J = 8.0 Hz, 2H), 8.00 (d, J = 8.0 Hz, 2H), 8.01 (s, 1H); ^{13}C NMR: (-60 °C) δ = 22.0, 28.2, 36.1, 36.3, 38.1, 39.1, 67.1, 79.8, 119.5, 128.6, 130.5, 131.6, 147.8, 154.2, 154.4; HRMS (ESI $^+$): Calcd for $\text{C}_{19}\text{H}_{27}\text{N}_4\text{O}_5\text{S}$, $\text{M}+\text{H}^+$ 423.1697. Found m/z 423.1688.

3i:



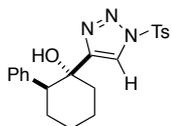
IR (ATR): 3256, 3169, 2980, 1591, 1452, 1389, 1192, 1178, 989 cm^{-1} ; ^1H NMR: δ = 2.45 (s, 3H), 2.99–3.10 (br, 1H), 7.32 (td, J = 7.2, 0.8 Hz, 2H), 7.37 (d, J = 8.4 Hz, 2H), 7.41 (td, J = 7.6, 0.8 Hz, 2H), 7.61 (d, J = 7.6 Hz, 2H), 7.66 (d, J = 7.6 Hz, 2H), 7.84 (s, 1H), 7.96 (d, J = 8.4 Hz, 2H); ^{13}C NMR: δ = 21.8, 78.3, 120.3, 120.5, 124.8, 128.5, 128.8, 129.8, 130.4, 132.7, 139.5, 146.8, 147.4, 149.9; HRMS (ESI $^+$): Calcd for $\text{C}_{22}\text{H}_{18}\text{N}_3\text{O}_3\text{S}$, $\text{M}+\text{H}^+$ 404.1063. Found m/z 404.1059.

cis-**3j**:



IR (ATR): 3362, 1389, 1192, 1177, 1005 cm^{-1} ; ^1H NMR: δ = 1.48–1.62 (m, 1H), 1.68–1.97 (m, 5H), 2.07–2.32 (m, 3H), 2.47 (s, 3H), 3.15 (dd, J = 13.2, 3.6 Hz, 1H), 6.83 (d, J = 7.2 Hz, 2H), 6.96 (t, J = 7.6 Hz, 2H), 7.06 (tt, J = 7.6, 1.6 Hz, 1H), 7.34 (d, J = 8.0 Hz, 2H), 7.48 (s, 1H), 7.80 (d, J = 8.4 Hz, 2H); ^{13}C NMR: δ = 21.0, 21.7, 25.8, 27.2, 38.4, 51.8, 72.5, 120.6, 126.4, 127.7, 128.3, 128.6, 130.2, 133.2, 140.9, 146.8, 155.0; HRMS (ESI $^+$): Calcd for $\text{C}_{21}\text{H}_{24}\text{N}_3\text{O}_3\text{S}$, $\text{M}+\text{H}^+$ 398.1533. Found m/z 398.1521.

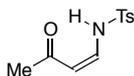
trans-**3j**:



IR (ATR): 3362, 1387, 1194, 1177, 1053, 1005 cm^{-1} ; ^1H NMR: δ = 1.43–1.58 (m, 1H), 1.72–1.85 (m, 2H), 1.86 (dd, J = 13.2, 4.4 Hz, 1H), 1.94–2.04 (m, 1H), 2.11–2.54 (m, 4H), 2.48 (s, 3H), 2.89 (dd, J = 13.2, 3.6 Hz, 1H), 6.77 (d, J = 6.8 Hz, 2H), 7.02 (t, J = 7.6 Hz, 2H), 7.12 (t, J = 7.6 Hz, 1H), 7.38 (d, J = 8.0 Hz, 2H), 7.57 (s, 1H), 7.89 (d, J = 8.8 Hz, 2H); ^{13}C NMR: δ = 21.7, 22.7, 26.1, 28.3, 40.3, 55.2, 73.3, 122.1, 126.7, 127.7, 128.4, 128.9, 130.3, 133.2, 140.2, 147.0, 152.5; HRMS (ESI $^+$): Calcd for $\text{C}_{21}\text{H}_{24}\text{N}_3\text{O}_3\text{S}$, $\text{M}+\text{H}^+$ 398.1533. Found m/z 398.1521.

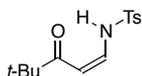
Typical Procedure for the Denitrogenative Rearrangement Reaction of 1-(*N*-Tosyl-1,2,3-triazol-4-yl)-alkanols (Table 1, entry 1). A 2-5 mL Biotage[®] microwave vial was charged with Rh₂(Oct)₄ (0.8 mg, 1 μmol), freshly prepared **1a** (53.5 mg, 0.20 mmol), and CHCl₃ (4 mL). The vial was capped with a Teflon pressure cap. The reaction mixture was heated at 140 °C for 15 min under microwave irradiation. After the reaction mixture was cooled, the solvent was removed under reduced pressure. The residue was purified by recycling preparative HPLC (CH₂Cl₂) to give the product **2a** (42.8 mg, 0.18 mmol, 89%).

2a:



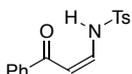
Purified by recycling preparative HPLC (CH₂Cl₂); IR (ATR): 3109, 1680, 1657, 1574, 1354, 1167, 1151, 1090, 966 cm⁻¹; ¹H NMR: δ = 2.13 (s, 3H), 2.42 (s, 3H), 5.45 (d, *J* = 8.4 Hz, 1H), 6.95 (t, *J* = 8.8 Hz, 1H), 7.31 (d, *J* = 8.0 Hz, 2H), 7.73 (d, *J* = 8.4 Hz, 2H), 11.45 (d, *J* = 8.0 Hz, 1H); ¹³C NMR: δ = 21.5, 30.1, 103.2, 126.6, 130.0, 136.8, 139.7, 144.5, 200.5; HRMS (ESI⁺): Calcd for C₁₁H₁₄NO₃S, M+H⁺ 240.0689. Found *m/z* 240.0683.

2d:



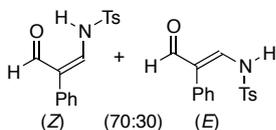
Purified by recycling preparative HPLC (CH₂Cl₂); IR (ATR): 3117, 2968, 1674, 1578, 1560, 1356, 1242, 1231, 1163, 1080, 924 cm⁻¹; ¹H NMR: δ = 1.11 (s, 9H), 2.42 (s, 3H), 5.65 (d, *J* = 8.4 Hz, 1H), 7.05 (dd, *J* = 10.4, 8.4 Hz, 1H), 7.32 (d, *J* = 8.0 Hz, 2H), 7.74 (d, *J* = 8.4 Hz, 2H), 11.57 (d, *J* = 10.4 Hz, 1H); ¹³C NMR: δ = 21.6, 26.6, 42.9, 98.7, 126.8, 130.0, 137.1, 140.7, 144.4, 208.5; HRMS (ESI⁺): Calcd for C₁₄H₂₀NO₃S, M+H⁺ 282.1158. Found *m/z* 282.1150.

2e:

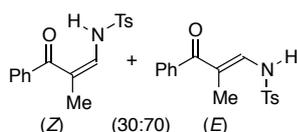


Purified by recycling preparative HPLC (CH₂Cl₂/ethyl acetate = 100:1); IR (ATR): 3115, 1638, 1558, 1456, 1354, 1232, 1159, 1015 cm⁻¹; ¹H NMR: δ = 2.42 (s, 3H), 6.19 (d, *J* = 8.8 Hz, 1H), 7.26 (t, *J* = 9.4 Hz, 1H), 7.33 (d, *J* = 8.4 Hz, 2H), 7.41–7.48 (m, 2H), 7.50–7.57 (m, 1H), 7.78 (d, *J* = 8.4 Hz, 2H), 7.87 (d, *J* = 7.2 Hz, 2H), 11.94 (d, *J* = 10.0 Hz, 1H); ¹³C NMR: δ = 21.5, 99.2, 126.7, 127.7, 128.6, 130.0, 132.8, 136.9, 137.5, 141.9, 144.5, 192.0; HRMS (ESI⁺): Calcd for C₁₆H₁₆NO₃S, M+H⁺ 302.0845. Found *m/z* 302.0837.

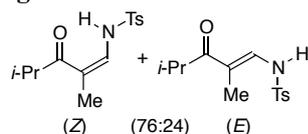
2e':



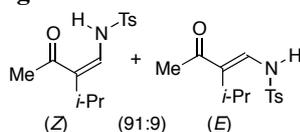
Purified by recycling preparative HPLC (CH₂Cl₂/ethyl acetate=100:1); IR (ATR): 3236, 1684, 1628, 1595, 1541, 1331, 1248, 1159, 1084 cm⁻¹; ¹H NMR: (*Z*) δ = 2.44 (s, 3H), 7.26-7.46 (m, 8H), 7.78 (d, *J* = 8.0 Hz, 2H), 9.68 (d, *J* = 3.6 Hz, 1H), 11.63 (d, *J* = 10.8 Hz, 1H); (*E*) δ = 2.47 (s, 3H), 7.11 (d, *J* = 8.0, 2H), 7.26-7.46 (m, 7H), 7.77 (d, *J* = 8.4 Hz, 2H), 9.42 (s, 1H); ¹³C NMR: (*Z* and *E*) δ = 21.61, 21.65, 117.2, 124.5, 126.8, 126.9, 126.9, 128.7, 129.0, 129.1, 129.3, 129.7, 130.2, 130.3, 135.1, 136.0, 136.6, 139.0, 143.4, 144.9, 145.2, 189.3, 193.6; HRMS (ESI⁺): Calcd for C₁₆H₁₆NO₃S, M+H⁺ 302.0845. Found *m/z* 302.0837.

2f':

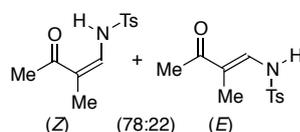
It is difficult to get a large amount of **2f'** due to the minor products. Therefore, only ^1H NMR was shown here. Purified by recycling preparative HPLC ($\text{CH}_2\text{Cl}_2/\text{ethyl acetate}=100:1$); ^1H NMR: (Z) $\delta = 1.93$ (d, $J = 1.2$ Hz, 3H); 2.44 (s, 3H), 7.04 (dq, $J = 10.8, 1.2$ Hz, 1H), 7.30-7.55 (m, 7H), 7.79 (d, $J = 8.8$ Hz, 2H), 11.44 (d, $J = 10.4$ Hz, 1H); (E) $\delta = 1.83$ (d, $J = 1.2$ Hz, 3H); 2.46 (s, 3H), 6.83 (d, $J = 12.0$ Hz, 1H), 7.10 (dq, $J = 12.0, 1.2$ Hz, 1H), 7.30-7.55 (m, 7H), 7.69 (d, $J = 8.4$ Hz, 2H).

2g:

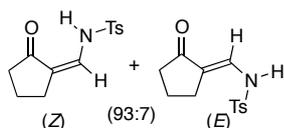
Purified by recycling preparative HPLC ($\text{CH}_2\text{Cl}_2/\text{ethyl acetate} = 100:1$); IR (ATR): 3354, 3260, 3192, 2970, 2932, 2872, 1715, 1607, 1597, 1342, 1157, 1088, 1047 cm^{-1} ; ^1H NMR: (Z) $\delta = 1.02$ (d, $J = 6.8$ Hz, 6H), 1.92 (d, $J = 1.2$ Hz, 3H), 2.40 (s, 3H), 2.86 (septet, $J = 6.8$ Hz, 1H), 6.82 (dq, $J = 10.4, 1.2$ Hz, 1H), 7.29 (d, $J = 8.0$ Hz, 2H), 7.72 (d, $J = 8.4$ Hz, 2H), 11.56 (d, $J = 10.4$ Hz, 1H); (E) $\delta = 1.06$ (d, $J = 6.8$ Hz, 6H), 1.64 (d, $J = 1.2$ Hz, 3H), 2.42 (s, 3H), 3.14 (septet, $J = 6.8$ Hz, 1H), 7.33 (d, $J = 8.0$ Hz, 2H), 7.41 (dd, $J = 12.0, 1.2$ Hz, 1H), 7.76 (d, $J = 8.4$ Hz, 2H), (N-H missing); ^{13}C NMR: (Z) $\delta = 16.9, 18.3, 21.5, 36.5, 109.2, 126.6, 129.9, 137.3, 137.5, 144.1, 208.4$; (E) $\delta = 9.5, 19.7, 21.6, 33.7, 116.5, 130.1, 133.6, 136.8, 144.7, 203.2$; HRMS (ESI $^+$): Calcd for $\text{C}_{14}\text{H}_{20}\text{NO}_3\text{S}$, $\text{M}+\text{H}^+$ 282.1158. Found m/z 282.1150.

2g':

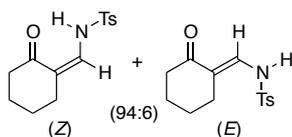
Purified by recycling preparative HPLC ($\text{CH}_2\text{Cl}_2/\text{ethyl acetate} = 100:1$); IR (ATR): 3204, 3051, 2963, 1651, 1574, 1433, 1360, 1263, 1155, 1090 cm^{-1} ; ^1H NMR: (Z) $\delta = 1.10$ (d, $J = 6.4$ Hz, 6H), 2.22 (s, 3H), 2.41 (s, 3H), 2.74 (quint, $J = 6.8$ Hz, 1H), 6.84 (d, $J = 10.8$ Hz, 1H), 7.30 (d, $J = 8.4$ Hz, 2H), 7.71 (d, $J = 8.4$ Hz, 2H), 11.64 (d, $J = 10.4$ Hz, 1H); ^{13}C NMR: (Z) $\delta = 21.5, 23.3, 27.9, 28.3, 121.7, 126.5, 129.9, 135.3, 137.4, 144.1, 202.4$; HRMS (ESI $^+$): Calcd for $\text{C}_{14}\text{H}_{20}\text{NO}_3\text{S}$, $\text{M}+\text{H}^+$ 282.1158. Found m/z 282.1150.

2h:

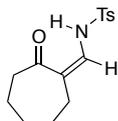
Purified by recycling preparative HPLC ($\text{CH}_2\text{Cl}_2/\text{ethyl acetate} = 100:1$); IR (ATR): 3269, 2930, 1732, 1639, 1593, 1408, 1337, 1269, 1157, 1086 cm^{-1} ; ^1H NMR: (Z) $\delta = 1.89$ (d, $J = 1.2$ Hz, 3H), 2.16 (s, 3H), 2.42 (s, 3H), 6.78 (dq, $J = 10.4, 1.2$ Hz, 1H), 7.30 (d, $J = 8.0$ Hz, 2H), 7.73 (d, $J = 8.4$ Hz, 2H), 11.41 (d, $J = 10.4$ Hz, 1H); (E) $\delta = 1.64$ (d, $J = 1.2$ Hz, 3H), 2.26 (s, 3H), 2.44 (s, 3H), 6.96 (d, $J = 12.0$ Hz, 1H), 7.35 (d, $J = 8.0$ Hz, 2H), 7.35-7.40 (m, 1H), 7.77 (d, $J = 8.4$ Hz, 2H); ^{13}C NMR: (Z) $\delta = 17.5, 21.5, 28.8, 110.3, 126.6, 129.9, 136.5, 137.4, 144.2, 202.4$; (E) $\delta = 9.2, 21.6, 25.0, 118.3, 126.7, 130.2, 134.6, 136.7, 144.8, 196.5$; HRMS (ESI $^+$): Calcd for $\text{C}_{12}\text{H}_{16}\text{NO}_3\text{S}$, $\text{M}+\text{H}^+$ 254.0845. Found m/z 254.0841.

4a:

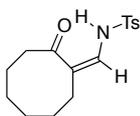
Purified by recycling preparative HPLC (CH₂Cl₂); IR (ATR): 3113, 1709, 1597, 1350, 1339, 1207, 1157, 1088, 1007 cm⁻¹; ¹H NMR: (Z) δ = 1.91 (quint, *J* = 7.6 Hz, 2H), 2.31 (t, *J* = 7.6 Hz, 2H), 2.41 (s, 3H), 2.55 (td, *J* = 7.2, 2.0 Hz, 2H), 6.79 (s, 1H), 7.31 (d, *J* = 8.4 Hz, 2H), 7.73 (d, *J* = 8.4 Hz, 2H), 10.71 (s, 1H); ¹³C NMR: (Z) δ = 21.2, 21.5, 27.2, 39.3, 114.5, 126.7, 130.0, 131.7, 137.2, 144.3, 209.5; HRMS (ESI⁺): Calcd for C₁₃H₁₆NO₃S, M+H⁺ 266.0845. Found m/z 266.0841.

4b:

Purified by recycling preparative HPLC (CH₂Cl₂); IR (ATR): 3138, 2947, 1670, 1553, 1335, 1219, 1157, 1084 cm⁻¹; ¹H NMR: (Z) δ = 1.62–1.71 (m, 2H), 1.70–1.80 (m, 2H), 2.34 (t, *J* = 6.8 Hz, 2H), 2.38 (td, *J* = 6.8, 1.2 Hz, 2H), 2.41 (s, 3H), 6.81 (dt, *J* = 10.0, 1.2 Hz, 1H), 7.30 (d, *J* = 8.0 Hz, 2H), 7.73 (d, *J* = 8.4 Hz, 2H), 11.58 (d, *J* = 10.0 Hz, 1H); ¹³C NMR: (Z) δ = 21.5, 22.1, 23.2, 28.3, 38.6, 111.8, 126.6, 129.9, 137.2, 137.4, 144.2, 202.4; HRMS (ESI⁺): Calcd for C₁₄H₁₈NO₃S, M+H⁺ 280.1002. Found m/z 280.1002.

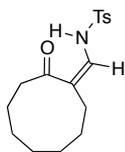
4c:

Purified by recycling preparative HPLC (CH₂Cl₂); IR (ATR): 3274, 3115, 2922, 1651, 1566, 1346, 1258, 1167, 1142, 1092 cm⁻¹; ¹H NMR: δ = 1.56–1.74 (m, 6H), 2.27–2.33 (m, 2H), 2.41 (s, 3H), 2.48–2.54 (m, 2H), 6.84 (d, *J* = 10.4 Hz, 1H), 7.30 (d, *J* = 8.0 Hz, 2H), 7.72 (d, *J* = 8.4 Hz, 2H), 11.46 (d, *J* = 10.4 Hz, 1H); ¹³C NMR: δ = 21.5, 24.8, 30.4, 31.2, 32.4, 44.6, 116.6, 126.6, 129.9, 136.6, 137.3, 144.1, 206.6; HRMS (ESI⁺): Calcd for C₁₅H₂₀NO₃S, M+H⁺ 294.1158. Found m/z 294.1153.

4d:

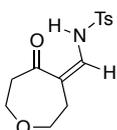
Purified by recycling preparative HPLC (CH₂Cl₂); IR (ATR): 3175, 3112, 2924, 1645, 1564, 1354, 1261, 1163, 1086 cm⁻¹; ¹H NMR: δ = 1.37–1.60 (m, 6H), 1.65–1.74 (m, 2H), 2.38 (t, *J* = 6.0 Hz, 2H), 2.41 (s, 3H), 2.52 (t, *J* = 6.4 Hz, 2H), 6.83 (d, *J* = 10.0 Hz, 1H), 7.30 (d, *J* = 8.4 Hz, 2H), 7.72 (d, *J* = 8.4 Hz, 2H), 11.68 (d, *J* = 10.0 Hz, 1H); ¹³C NMR: δ = 21.6, 25.6, 26.0, 28.9, 29.9, 32.7, 39.6, 115.8, 126.6, 129.9, 137.2, 137.3, 144.1, 207.4; HRMS (ESI⁺): Calcd for C₁₆H₂₂NO₃S, M+H⁺ 308.1315. Found m/z 308.1313.

4e:



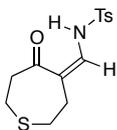
Purified by recycling preparative HPLC (CH₂Cl₂); IR (ATR): 3210, 2924, 1643, 1558, 1350, 1256, 1157, 1088 cm⁻¹; ¹H NMR: δ = 1.37–1.56 (m, 6H), 1.51–1.68 (m, 2H), 1.64–1.82 (m, 2H), 2.32–2.45 (m, 2H), 2.40 (s, 3H), 2.51 (t, *J* = 6.4 Hz, 2H), 6.84 (d, *J* = 10.4 Hz, 1H), 7.29 (d, *J* = 8.0 Hz, 2H), 7.72 (d, *J* = 8.0 Hz, 2H), 11.72 (d, *J* = 10.0 Hz, 1H); ¹³C NMR: δ = 21.5, 24.3, 24.6, 26.1, 27.6, 29.9, 31.0, 39.2, 117.2, 126.6, 129.9, 137.3, 138.2, 144.1, 207.7; HRMS (ESI⁺): Calcd for C₁₇H₂₄NO₃S, M+H⁺ 322.1471. Found m/z 322.1465.

4f:



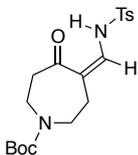
Purified by recycling preparative HPLC (CH₂Cl₂/ethyl acetate=100:1); IR (ATR): 3308, 1682, 1651, 1595, 1566, 1346, 1263, 1159, 1146 cm⁻¹; ¹H NMR: δ = 2.41 (s, 3H), 2.46–2.51 (m, 2H), 2.71–2.75 (m, 2H), 3.68–3.76 (m, 4H), 6.89 (d, *J* = 10.4 Hz, 1H), 7.31 (d, *J* = 8.0 Hz, 2H), 7.73 (d, *J* = 8.4 Hz, 2H), 11.42 (d, *J* = 10.4 Hz, 1H); ¹³C NMR: δ = 21.5, 35.0, 48.3, 66.0, 72.3, 114.5, 126.6, 130.0, 137.1, 137.8, 144.4, 204.4; HRMS (ESI⁺): Calcd for C₁₄H₁₈NO₄S, M+H⁺ 296.0951. Found m/z 296.0940.

4g:



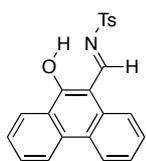
Purified by recycling preparative HPLC (CH₂Cl₂/ethyl acetate=100:1); IR (ATR): 3179, 3028, 2897, 1647, 1560, 1354, 1263, 1155, 1146, 1082 cm⁻¹; ¹H NMR: δ = 2.42 (s, 3H), 2.64–2.80 (m, 6H), 2.92–2.95 (m, 2H), 6.92 (d, *J* = 10.4 Hz, 1H), 7.31 (d, *J* = 8.4 Hz, 2H), 7.73 (d, *J* = 8.4 Hz, 2H), 11.48 (d, *J* = 10.8 Hz, 1H); ¹³C NMR: δ = 21.5, 25.6, 32.4, 35.2, 47.5, 114.5, 126.6, 130.0, 137.0, 138.5, 144.4, 204.1; HRMS (ESI⁺): Calcd for C₁₄H₁₈NO₃S₂, M+H⁺ 312.0723. Found m/z 312.0717.

4h:



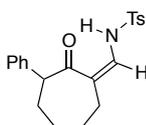
Purified by recycling preparative HPLC (CH₂Cl₂/ethyl acetate=100:1); IR (ATR): 3179, 2974, 2930, 1688, 1651, 1574, 1418, 1362, 1244, 1161, 1088 cm⁻¹; ¹H NMR: δ = 1.44 (s, 9H), 2.37–2.46 (m, 2H), 2.42 (s, 3H), 2.60–2.66 (m, 2H), 3.46–3.54 (m, 4H), 6.91 (d, *J* = 10.4 Hz, 1H), 7.31 (d, *J* = 8.4 Hz, 2H), 7.73 (d, *J* = 8.0 Hz, 2H), 11.47 (d, *J* = 10.0 Hz, 1H); ¹³C NMR: δ = 21.5, 28.3, 33.0–33.8 (br), 41.7–42.6 (br), 46.1, 47.6–48.9 (br), 80.2, 114.1, 126.6, 129.9, 137.0, 138.5, 144.4, 154.6, 204.2; HRMS (ESI⁺): Calcd for C₁₉H₂₇N₂O₅S, M+H⁺ 395.1635. Found m/z 395.1630.

4i:



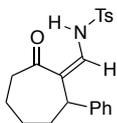
Purified by recycling preparative HPLC (CH₂Cl₂); IR (ATR): 3065, 1589, 1541, 1487, 1321, 1294, 1153, 1088 cm⁻¹; ¹H NMR: δ = 2.44 (s, 3H), 7.37 (d, *J* = 8.0 Hz, 2H), 7.55 (t, *J* = 6.8 Hz, 1H), 7.64 (t, *J* = 6.8 Hz, 2H), 7.81 (t, *J* = 8.4 Hz, 1H), 7.93 (d, *J* = 8.4 Hz, 2H), 8.19 (d, *J* = 8.4 Hz, 1H), 8.50 (d, *J* = 8.0 Hz, 1H), 8.55 (dd, *J* = 8.4, 5.6 Hz, 2H), 9.93 (s, 1H), (O-H missing); ¹³C NMR: δ = 21.7, 105.2, 120.3, 122.8, 123.5, 124.8, 125.4, 125.7, 125.8, 127.3, 127.7, 128.4, 130.0, 132.0, 135.1, 135.9, 144.7, 165.6, 166.0; HRMS (ESI⁺): Calcd for C₂₂H₁₈NO₃S, M+H⁺ 376.1002. Found m/z 376.0995.

4j:



Purified by recycling preparative HPLC (CH₂Cl₂); IR (ATR): 3192, 2924, 2853, 1651, 1574, 1352, 1250, 1167, 1072, 1055 cm⁻¹; ¹H NMR: δ = 1.32–1.46 (m, 1H), 1.66–1.80 (m, 1H), 1.90–2.13 (m, 4H), 2.35 (dd, *J* = 15.2, 6.0 Hz, 1H), 2.42 (s, 3H), 2.55–2.66 (m, 1H), 3.89 (d, *J* = 10.4 Hz, 1H), 6.95 (d, *J* = 10.4 Hz, 1H), 7.10 (d, *J* = 8.0 Hz, 2H), 7.24–7.37 (m, 5H), 7.70 (d, *J* = 8.4 Hz, 2H), 11.40 (d, *J* = 10.4 Hz, 1H); ¹³C NMR: δ = 21.5, 30.1, 30.6, 32.1, 33.0, 57.5, 115.7, 126.7, 126.8, 128.1, 128.4, 129.8, 137.2, 137.4, 140.8, 144.1, 204.7; HRMS (ESI⁺): Calcd for C₂₁H₂₄NO₃S, M+H⁺ 370.1471. Found m/z 370.1461.

4j':

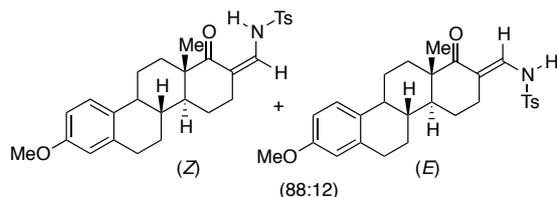


Purified by preparative thin-layer chromatography (CHCl₃/ethyl acetate = 25:1) and recycling preparative HPLC (Hexane/CH₂Cl₂/ethyl acetate=70:15:15); IR (ATR): 3179, 2926, 2856, 1645, 1568, 1360, 1259, 1167, 1150, 1086 cm⁻¹; ¹H NMR: δ = 1.51–1.65 (m, 1H), 1.63–1.82 (m, 2H), 1.83–1.96 (m, 1H), 2.06–2.23 (m, 2H), 2.36–2.48 (m, 1H), 2.40 (s, 3H), 2.58–2.69 (m, 1H), 3.72–3.80 (m, 1H), 6.39 (d, *J* = 10.8 Hz, 1H), 7.12 (d, *J* = 7.2 Hz, 2H), 7.23–7.30 (m, 3H), 7.35 (t, *J* = 7.2 Hz, 2H), 7.55 (d, *J* = 8.4 Hz, 2H), 11.57 (d, *J* = 10.8 Hz, 1H); ¹³C NMR: δ = 21.6, 24.7, 28.5, 35.3, 44.1, 45.9, 119.9, 126.6, 126.7, 127.8, 128.8, 129.8, 137.2, 139.0, 142.9, 144.1, 206.3; HRMS (ESI⁺): Calcd for C₂₁H₂₄NO₃S, M+H⁺ 370.1471. Found m/z 370.1462.

Typical Procedure for the One-pot Synthesis of Enaminones from Propargylic Alcohols (equation 3).

A 2-5 mL Biotage[®] microwave vial was charged with 2-aminophenol (1.23 g, 11.3 μmol), Cu(OAc)₂·H₂O (3.9 mg, 19.5 μmol), tosyl azide (38.4 mg, 0.19 mmol), but-3-yn-2-ol (**5a**, 14.4 mg, 0.21 mmol), and CHCl₃ (1 mL). The vial was capped with a Teflon pressure cap. The reaction mixture was stirred at room temperature for 24 h. To the resulting green solution were added Rh₂(Oct)₄ (1.57 mg, 2 μmol) and CHCl₃ (3 mL). Then, the reaction mixture was heated at 140 °C for 15 min under microwave irradiation. After being cooled to room temperature, the resulting mixture was passed through a pad of diol silica and eluted with ethyl acetate (50 mL). The filtrate was concentrated under reduced pressure. The residue was purified by recycling preparative HPLC (CH₂Cl₂) to give the product **2a** (33.8 mg, 0.14 mmol, 69%).

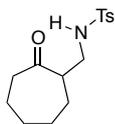
4k:



Purified by recycling preparative HPLC (CH_2Cl_2); IR (ATR): 3244, 2926, 1651, 1574, 1499, 1352, 1254, 1161, 1088 cm^{-1} ; ^1H NMR: (Z) δ = 0.98 (s, 3H), 1.22–1.56 (m, 6H), 1.98–2.14 (m, 2H), 2.14–2.26 (m, 2H), 2.32–2.48 (m, 2H), 2.42 (s, 3H), 2.52–2.60 (m, 1H), 2.81–2.89 (m, 2H), 3.77 (s, 3H), 6.63 (d, J = 2.4 Hz, 1H), 6.72 (dd, J = 8.4, 2.4 Hz, 1H), 6.82 (d, J = 10.4 Hz, 1H), 7.21 (d, J = 8.8 Hz, 1H), 7.31 (d, J = 8.0 Hz, 2H), 7.74 (d, J = 8.4 Hz, 2H), 11.38 (d, J = 10.4 Hz, 1H); ^{13}C NMR: (Z) δ = 17.5, 21.2, 21.5, 25.8, 26.3, 26.5, 30.0, 33.1, 39.3, 42.8, 44.8, 46.1, 55.1, 109.7, 111.6, 113.4, 126.2, 126.6, 129.9, 132.2, 136.9, 137.3, 137.6, 144.1, 157.5, 207.8; HRMS (ESI⁺): Calcd for $\text{C}_{28}\text{H}_{34}\text{NO}_4\text{S}$, $\text{M}+\text{H}^+$ 480.2203. Found m/z 480.2192.

Procedure for the Hydrogenation Reaction of Enaminone 4c Catalyzed by Pd/C (Scheme 2). A side-arm tube equipped with a stirrer bar was charged with enaminone **4c** (57.7 mg, 0.20 mmol) and Pd/C (6.9 mg, 12 wt%), and ethyl acetate (3 mL). The tube was connected to a hydrogen balloon and immersed in a dry ice/acetone bath. After ten vacuum/ H_2 -filling cycles, the cooling bath was removed. The reaction mixture was stirred for 24 h at 40 °C, and then, cooled to room temperature. The resulting mixture was passed through a pad of Celite and eluted with ethyl acetate. The filtrate was concentrated under reduced pressure. The residue was purified by preparative thin-layer chromatography (chloroform/ethyl acetate = 25:1) to give the product **6** (50.3 mg, 0.17 mmol, 86%).

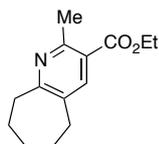
6:



IR (ATR): 3279, 2926, 1693, 1325, 1155, 1092 cm^{-1} ; ^1H NMR: δ = 1.19–1.36 (m, 2H), 1.44–1.95 (m, 6H), 2.30–2.52 (m, 2H), 2.42 (s, 3H), 2.78–2.88 (m, 1H), 3.00 (t, J = 6.8 Hz, 2H), 5.06 (t, J = 6.8 Hz, 1H), 7.30 (d, J = 8.0 Hz, 2H), 7.72 (d, J = 8.4 Hz, 2H); ^{13}C NMR: δ = 21.5, 23.3, 29.0, 29.1, 29.2, 43.5, 44.6, 51.3, 126.9, 129.7, 137.1, 143.3, 215.5; HRMS (ESI⁺): Calcd for $\text{C}_{15}\text{H}_{22}\text{NO}_3\text{S}$, $\text{M}+\text{H}^+$ 296.1315. Found m/z 296.1308.

Procedure for the Reaction of Enaminone 4c with Ethyl Acetoacetate (Scheme 2). A side-arm tube equipped with a stirrer bar and reflux condenser was charged with enaminone **4c** (61.8 mg, 0.21 mmol) and ammonium acetate (23.0 mg, 0.30 mmol). The tube was evacuated and refilled with argon three times, and ethyl acetoacetate (34.0 mg, 0.26 mmol) and AcOH (2 mL) were added. After being heated at 140 °C for 12 h, the reaction mixture was cooled to room temperature and neutralized with 1 M NaOH aq. The aqueous layer was extracted with ethyl acetate (2 mL x 4). The combined organic extracts were dried over Na_2SO_4 and concentrated under reduced pressure. The residue was purified by preparative thin-layer chromatography (hexane/ethyl acetate = 4:1) to give the product **7** (24.5 mg, 0.11 mmol, 50%).

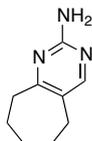
7:



IR (ATR): 2922, 1720, 1597, 1556, 1443 cm^{-1} ; ^1H NMR: δ = 1.39 (t, J = 7.2 Hz, 3H), 1.54–1.76 (m, 4H), 1.84–1.92 (m, 2H), 2.74–2.82 (m, 2H), 2.77 (s, 3H), 3.01–3.08 (m, 2H), 4.36 (q, J = 7.2 Hz, 2H), 7.88 (s, 1H); ^{13}C NMR: δ = 14.3, 24.2, 26.3, 27.9, 32.4, 34.5, 39.4, 60.9, 122.8, 135.2, 138.6, 156.2, 166.0, 166.9; HRMS (ESI⁺): Calcd for $\text{C}_{14}\text{H}_{20}\text{NO}_2$, $[\text{M}+\text{H}]^+$ 234.1489. Found m/z 234.1486.

Procedure for the Reaction of Enaminone 4c with Guanidine (Scheme 2). To a side-arm tube equipped with a stirrer bar and reflux condenser was charged with enaminone **4c** (76.2 mg, 0.26 mmol), guanidine hydrochloride (30.5 mg, 0.32 mmol) and NaOH (14.7 mg, 0.37 mmol). The tube was evacuated and refilled with argon three times, and *i*-PrOH (5 mL) was added. After being refluxed at 110 °C for 24 h, the reaction mixture was cooled to room temperature and concentrated under reduced pressure. The residue was purified by preparative thin-layer chromatography (hexane/ethyl acetate = 1:4) to give the product **8** (26.2 mg, 0.16 mmol, 62%).

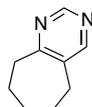
8:



IR (ATR): 3314, 3159, 2914, 1655, 1591, 1556, 1483, 1437 cm^{-1} ; ^1H NMR: δ = 1.52–1.69 (m, 4H), 1.76–1.86 (m, 2H), 2.53–2.60 (m, 2H), 2.72–2.78 (m, 2H), 5.18 (br s, 2H), 7.92 (s, 1H); ^{13}C NMR: δ = 25.9, 28.3, 31.0, 32.3, 38.9, 124.9, 156.7, 161.5, 172.5; HRMS (ESI⁺): Calcd for $\text{C}_9\text{H}_{14}\text{N}_3$, $[\text{M}+\text{H}]^+$ 164.1182. Found m/z 164.1183.

Procedure for the Reaction of Enaminone 4c with Formamidine (Scheme 2). To a side-arm tube equipped with a stirrer bar and reflux condenser was charged with enaminone **4c** (61.1 mg, 0.21 mmol) and formamidine hydrochloride (84.5 mg, 1.1 mmol). The tube was evacuated and refilled with argon three times, and pyridine (1 mL) was added. After being refluxed at 130 °C for 20 h, the reaction mixture was cooled to room temperature and concentrated under reduced pressure. The residue was purified by preparative thin-layer chromatography (1st.: hexane/ethyl acetate = 1:1, 2nd.: chloroform/ethyl acetate = 100:1) to give the product **9** (19.4 mg, 0.13 mmol, 63%).

9:⁵

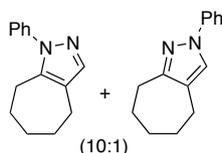


IR (ATR): 2922, 2853, 1572, 1551, 1456, 1447, 1396 cm^{-1} ; ^1H NMR: δ = 1.62–1.75 (m, 4H), 1.84–1.95 (m, 2H), 2.71–2.79 (m, 2H), 2.94–3.02 (m, 2H), 8.37 (s, 1H), 8.89 (s, 1H); ^{13}C NMR: δ = 25.7, 27.4, 31.9, 32.3, 39.0, 135.5, 155.5, 156.3, 171.3; HRMS (ESI⁺): Calcd for $\text{C}_9\text{H}_{13}\text{N}_2$, $[\text{M}+\text{H}]^+$ 149.1073. Found m/z 149.1074.

⁵ Boger, D. L.; Schumacher, J.; Mullican, M. D.; Patel, M.; Panek, J. S. *J. Org. Chem.* **1982**, *47*, 2673.

Procedure for the Reaction of Enaminone 4c with Phenylhydrazine (Scheme 2). To a side-arm tube equipped with a stirrer bar and reflux condenser was charged with enaminone **4c** (69.7 mg, 0.24 mmol). The tube was evacuated and refilled with argon three times, and phenyl hydrazine (28.9 mg, 0.27 mmol) and EtOH (4 mL) were added. After being refluxed at 100 °C for 12 h, the reaction mixture was cooled to room temperature and concentrated under reduced pressure. The residue was purified by preparative thin-layer chromatography (hexane/ethyl acetate = 5:1) to give the product **9** (42.9 mg, 0.20 mmol, 85% yield, 10:1 r.r.).

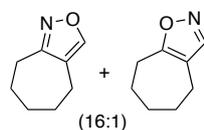
10:⁶



IR (ATR): 1501, 1398 cm^{-1} ; ^1H NMR: δ = 1.60–1.76 (m, 4H), 1.81–1.90 (m, 2H), 2.60–2.67 (m, 2H), 2.74–2.82 (m, 2H), 7.33–7.42 (m, 4H), 7.42–7.49 (m, 2H); ^{13}C NMR: δ = 25.6, 27.09, 27.15, 28.5, 31.7, 121.9, 125.4, 127.4, 128.8, 139.6, 139.9, 142.1; HRMS (ESI⁺): Calcd for $\text{C}_{14}\text{H}_{17}\text{N}_2$, $[\text{M}+\text{H}]^+$ 213.1386. Found m/z 213.1384.

Procedure for the Reaction of Enaminone 4c with Hydroxylamine (Scheme 2). To a side-arm tube equipped with a stirrer bar was charged with enaminone **4c** (178.3 mg, 0.61 mmol) and hydroxylamine hydrochloride (218.9 mg, 3.2 mmol). The tube was evacuated and refilled with argon three times, and MeOH (3 mL) was added. After being heated at 70 °C for 4 h, the reaction mixture was cooled to room temperature and neutralized with NaHCO_3 aq. The aqueous layer was extracted with Et_2O (4 mL x 4). The combined organic extracts were dried over Na_2SO_4 and concentrated under reduced pressure. The residue was purified by preparative thin-layer chromatography (hexane/ethyl acetate = 5:1) to give the product **10** (54.3 mg, 0.40 mmol, 66%, 16:1 r.r.).

11:⁷

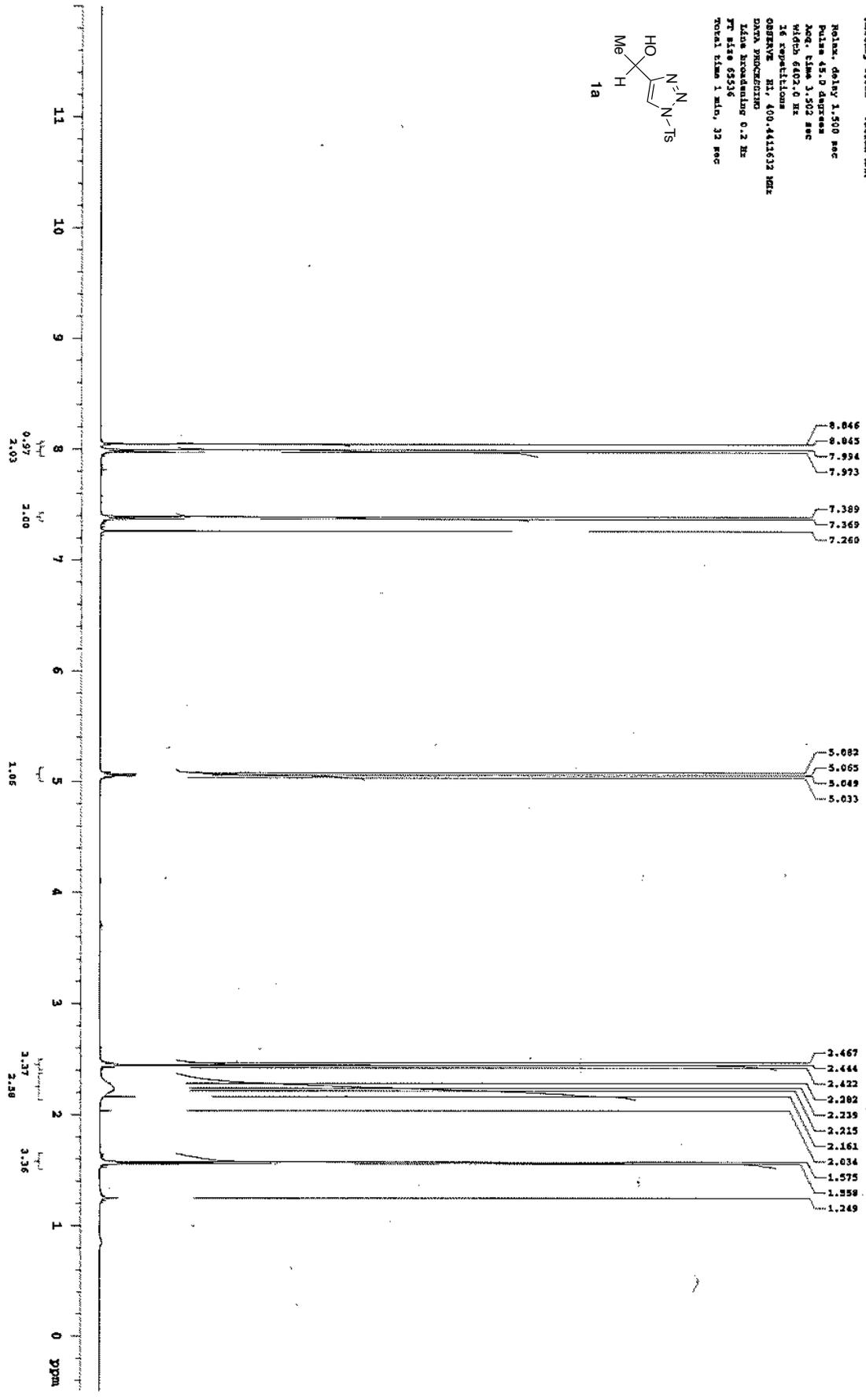
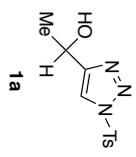


IR (ATR): 2922, 2851, 1614, 1443, 1414 cm^{-1} ; ^1H NMR: δ = 1.60–1.74 (m, 4H), 1.78–1.86 (m, 2H), 2.50–2.56 (m, 2H), 2.76–2.84 (m, 2H), 8.02 (s, 1H); ^{13}C NMR: δ = 23.2, 27.0, 27.2, 29.0, 31.9, 120.2, 153.6, 164.9; HRMS (APCI): Calcd for $\text{C}_8\text{H}_{12}\text{NO}$, $[\text{M}+\text{H}]^+$ 138.0913. Found m/z 138.0914.

6 Cho, C. S.; Patel, D. B. *Tetrahedron* **2006**, 62, 6388.

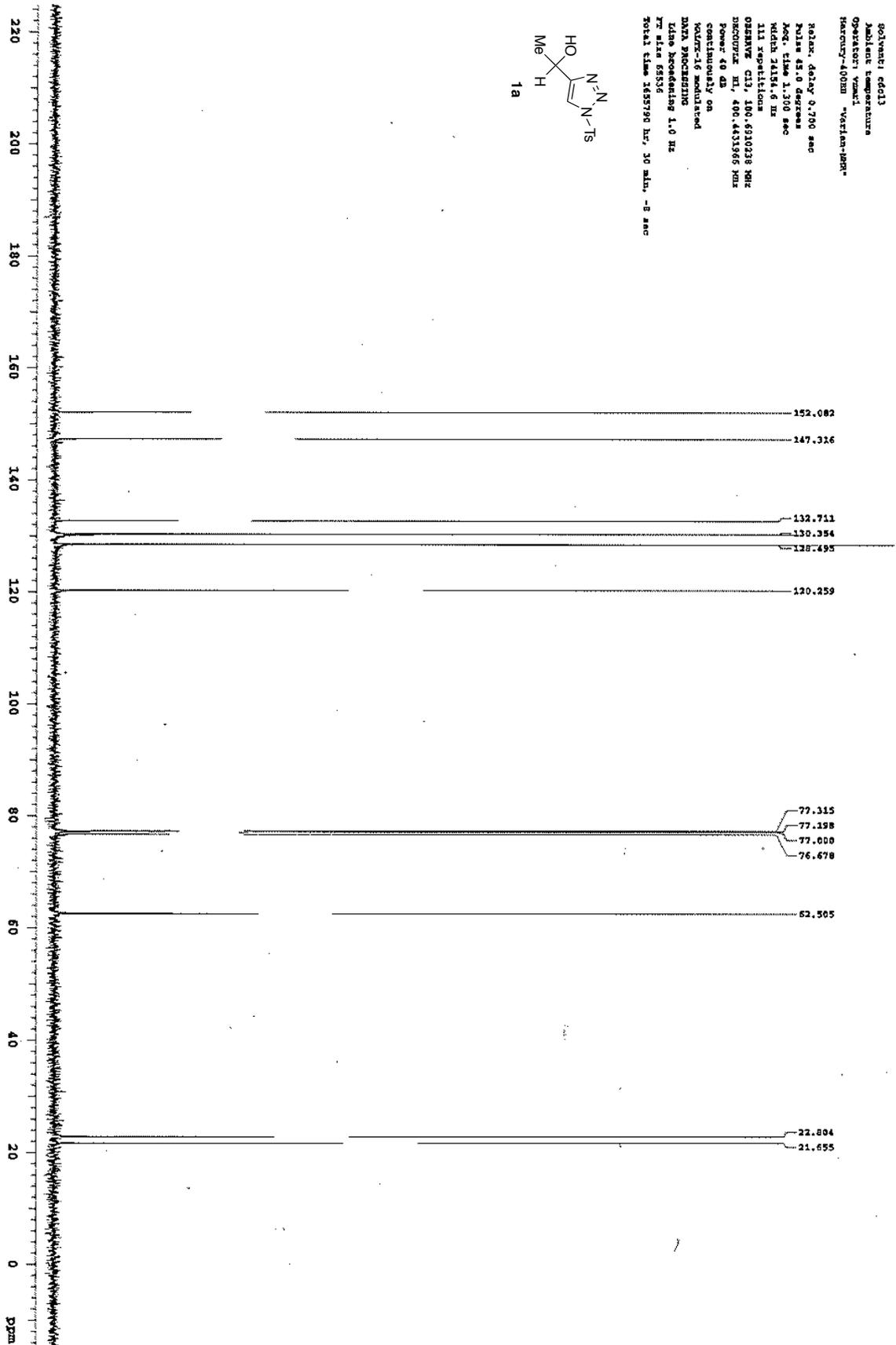
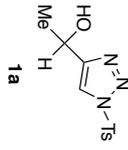
7 Ichino, T.; Arimoto, H.; Uemura, D. *Chem. Commun.* **2006**, 1742.

SOLVENT: d6-DMSO
 Ambient Temperature
 Operator: vsmw1
 Program: 400M1 "Varian-PMN"
 Relax. delay 1.500 sec
 Pulse 15.0 degrees
 Acq. time 3.302 sec
 Width 6402.0 Hz
 16 repetitions
 OBSERVE F1, 400.441612 MHz
 DATA PROCESSING
 F2 Acq. Bandwidth 0.2 Hz
 FT size 65536
 Total time 1 min, 32 sec

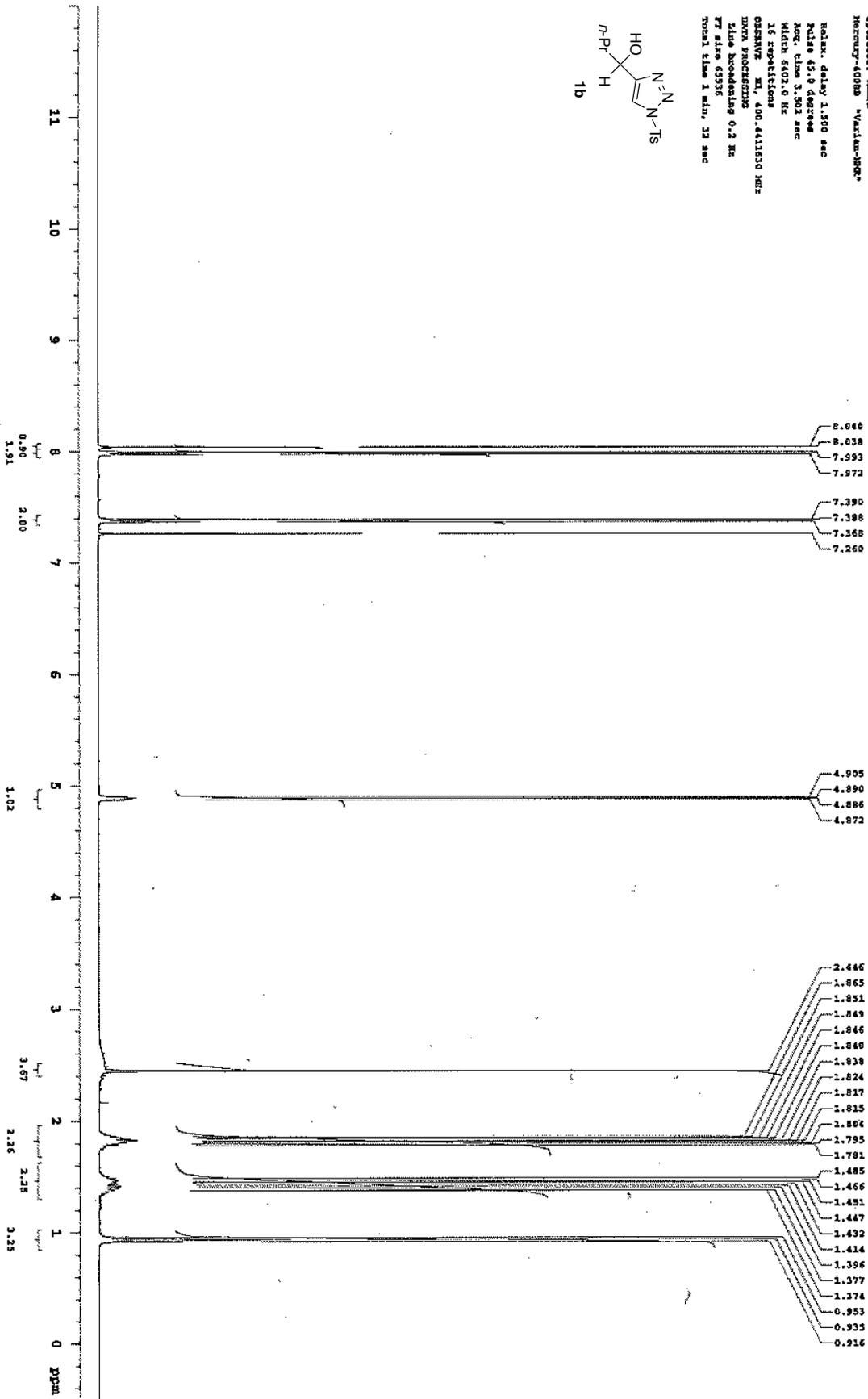
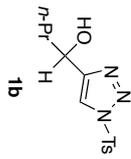


Solvent: cdcl3
Molalnt temperature
Operator: VAM:1
Nucleus: 13C NMR Varian-125P

Relax. delay 0.700 sec
Pulse 45.0 degrees
Acq. time 1.300 sec
Width 24154.6 Hz
111 repetitions
OBSERVE: C13 100.630238 MHz
INSTRUM: NI 400.4431966 MHz
Power 40 dB
SOLVENT: cdcl3
CONTINUOUSLY ON
DATA PROCESSING
Line broadening 1.0 Hz
FM file 68156
Total time 165708 hr, 30 min, -8 sec



Solvent: cdcl3
 Ambient temperature
 Operator: ymwt
 Mercury-6000 Varian-JNM
 Pulse delay: 1.500 sec
 Pulse: 45.0 degrees
 Acq. time: 3.502 sec
 Width: 6602.0 Hz
 IS: 100000000
 OBSERVE: H1, 400.441630 MHz
 DATA PROCESSING
 Zero: 0.2 Hz
 FT size: 63336
 Total time: 1 min, 33 sec



Pulse Sequence: zgpg30

Solvent: CDCl3

Substrate Temperature

Operator: ymzr1

Hexony-40183 *YALIAS-HM-

Relax. delay: 0.700 sec

Pulse: zgpg30

Acq. time: 1.100 sec

Width: 24194.6 Hz

450 experiments

DIRNAME: C:\100_631022

DIRGLOB: *.1, 400_443186

Power: 40 dB

continuously on

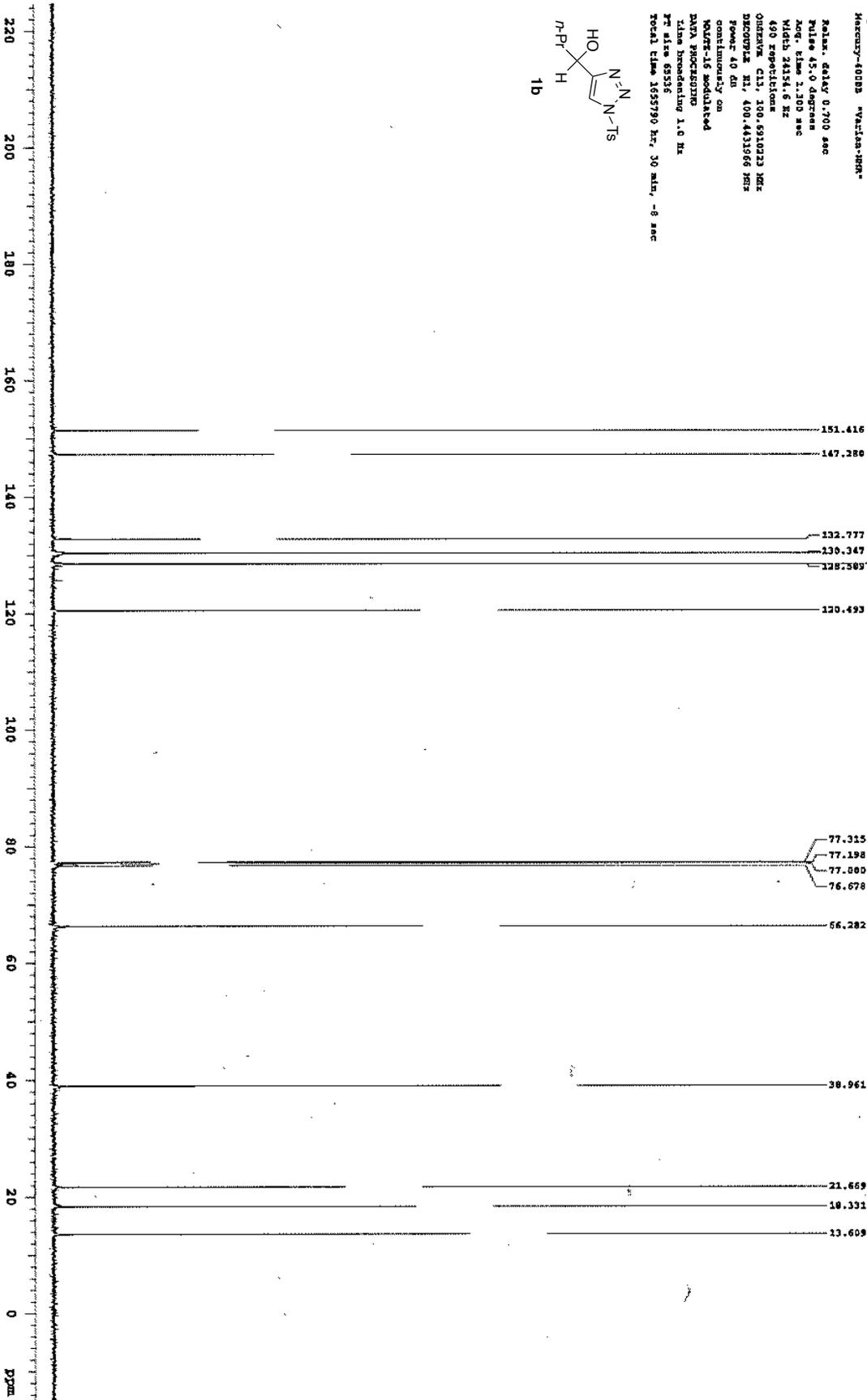
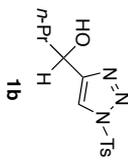
NAME: 1S

NAME: PROCRS310

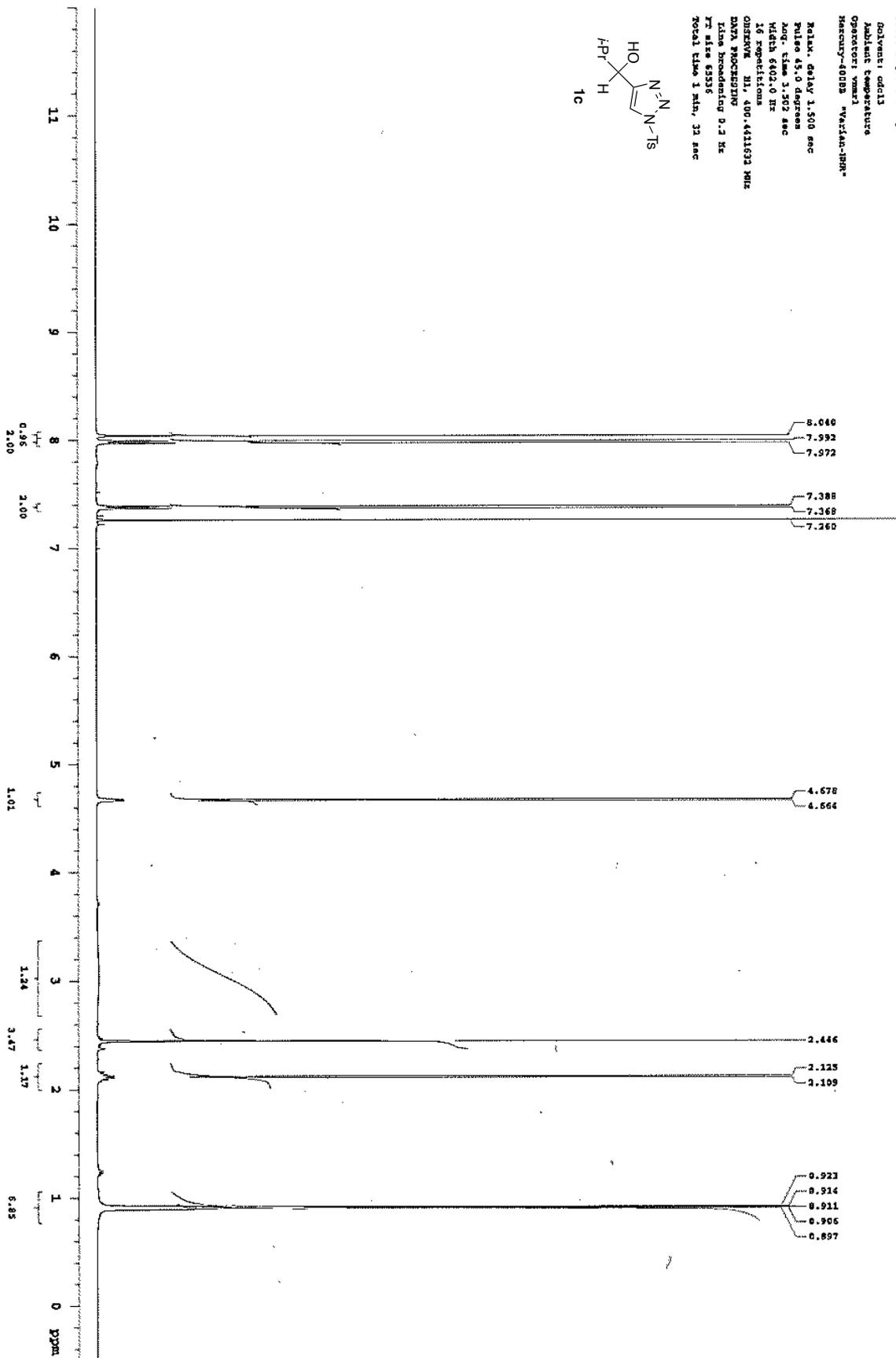
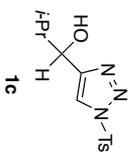
Line broadening: 1.0 Hz

F1 file: 63535

Total time: 165790 hr, 30 min, -8 sec



Pulse Sequence: zgpg30
 Solvent: dcd13
 Acquire Temperature: 300.2
 Operator Name: wvlsjg-jm
 Frequency: 400MHz ¹³C-NMR
 Relax. delay: 1.500 sec
 Pulse: 45.0 degree
 Acq. time: 1.502 sec
 Width: 6402.0 Hz
 16 repetitions
 OBSERVE: H1, 400.441633 MHz
 DATA PROCESSING
 Line broadening: 0.2 Hz
 FT: size 65336
 Total time: 1 min, 32 sec



Pulse sequence: zgpg1

Solvent: cdcl3

Acquire temperature

Operator: vsmc1

Hardware: 400MR Varian-800*

Relax. delay: 0.700 sec

Puls. pr. 0.00000000

Acq. time: 1.300 sec

NUC1: 131P4.8 Hz

31 experiments

OSBANK: C13, 100.631067 MHz

PROCPAR: NL, 400.451986 MHz

Power: 4.0 dB

acquire: on

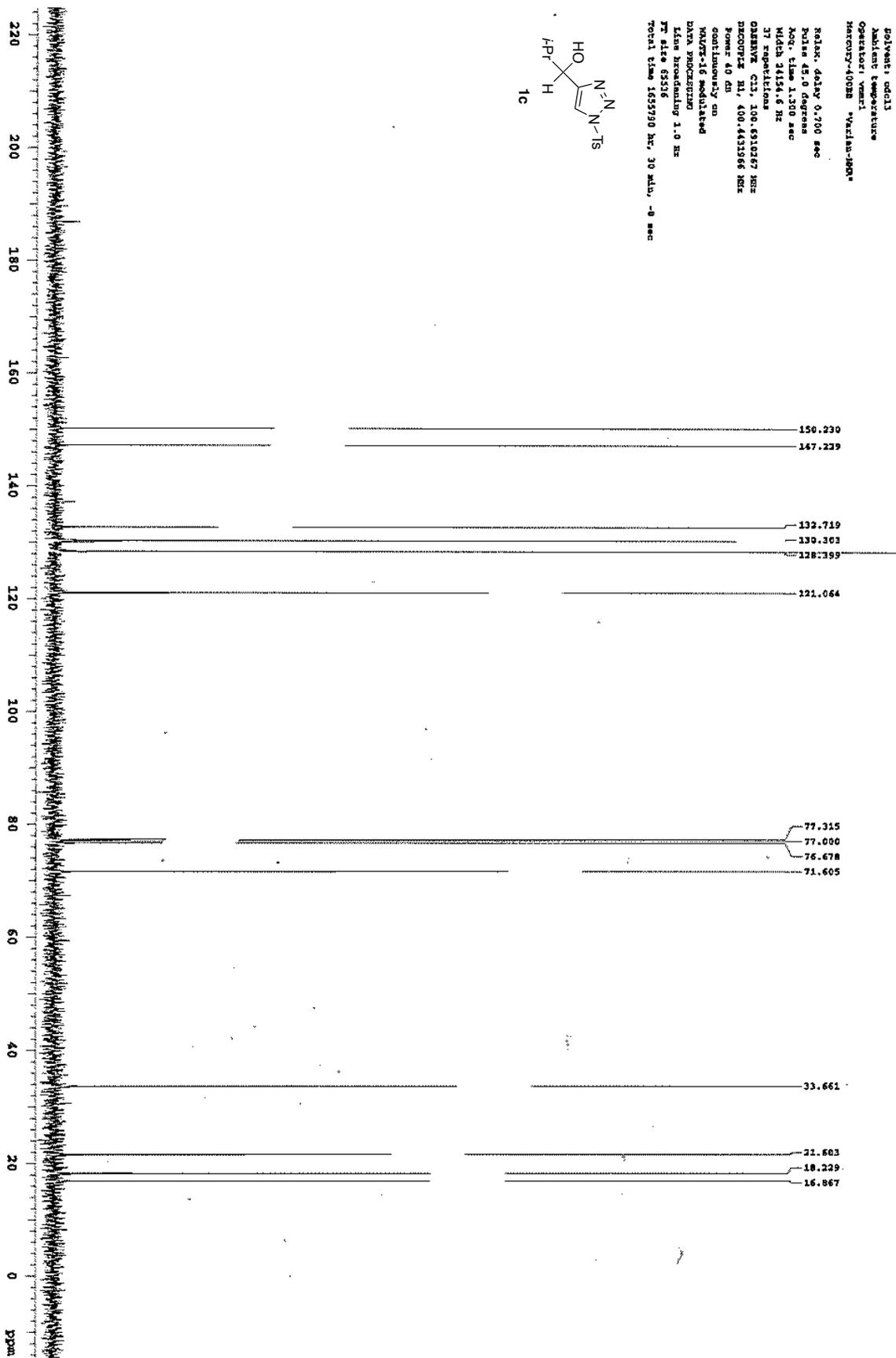
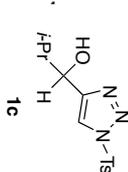
NAME: 13

DATA PROCESSING

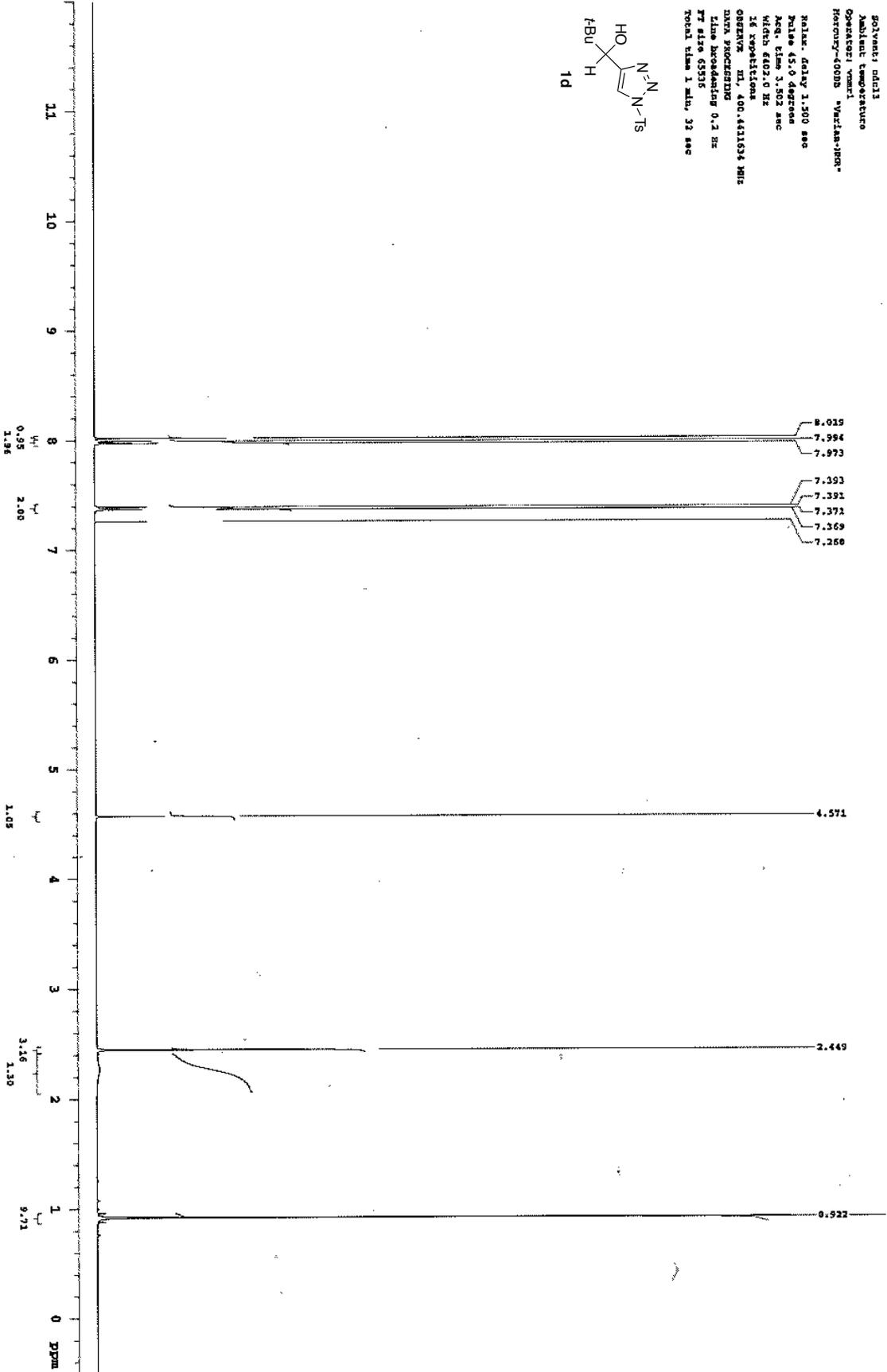
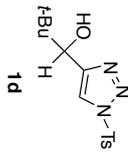
Line broadening: 1.0 Hz

IR file: 03136

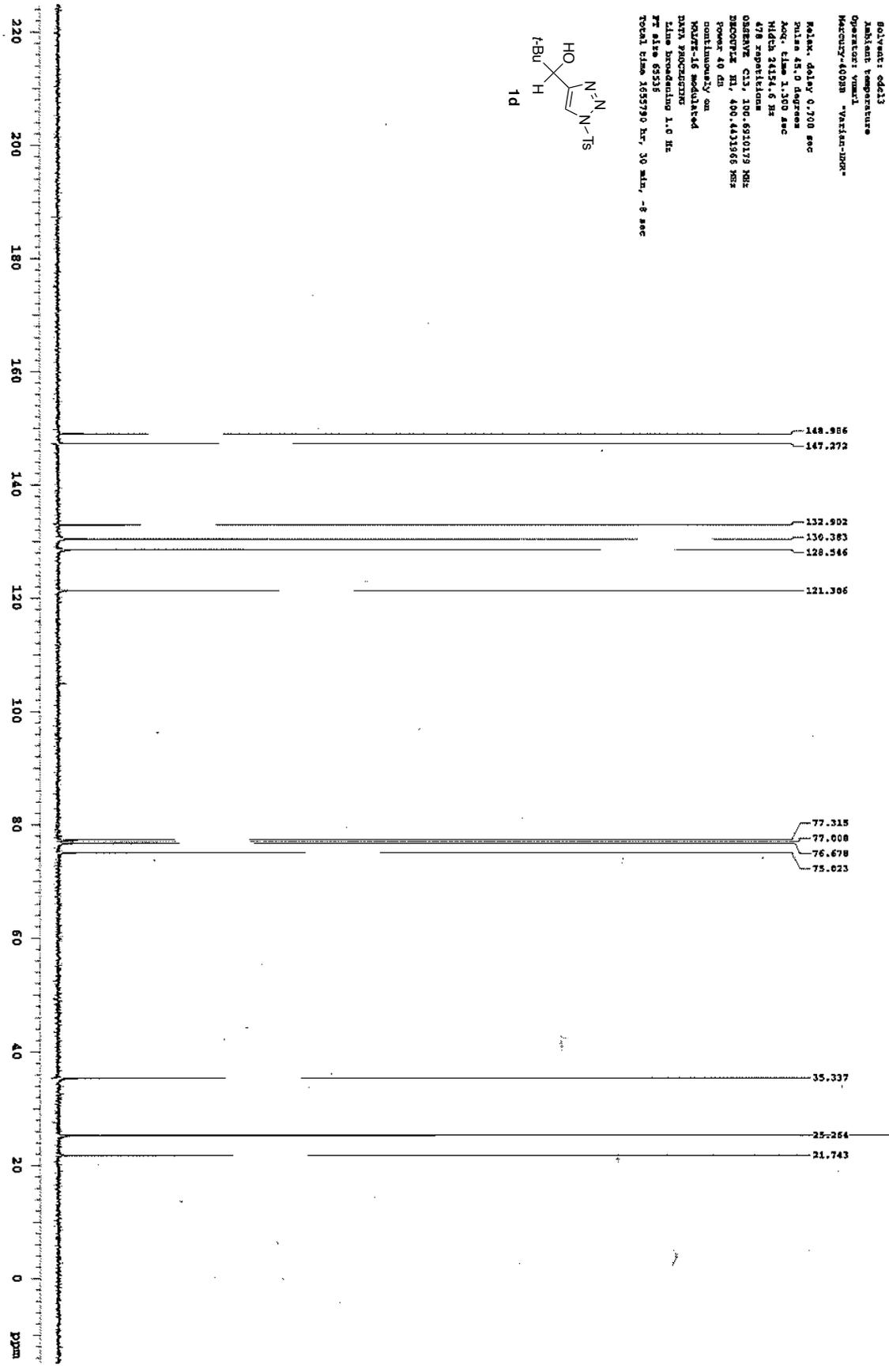
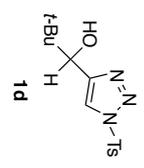
Total time: 1655790 hr, 30 min, -8 sec



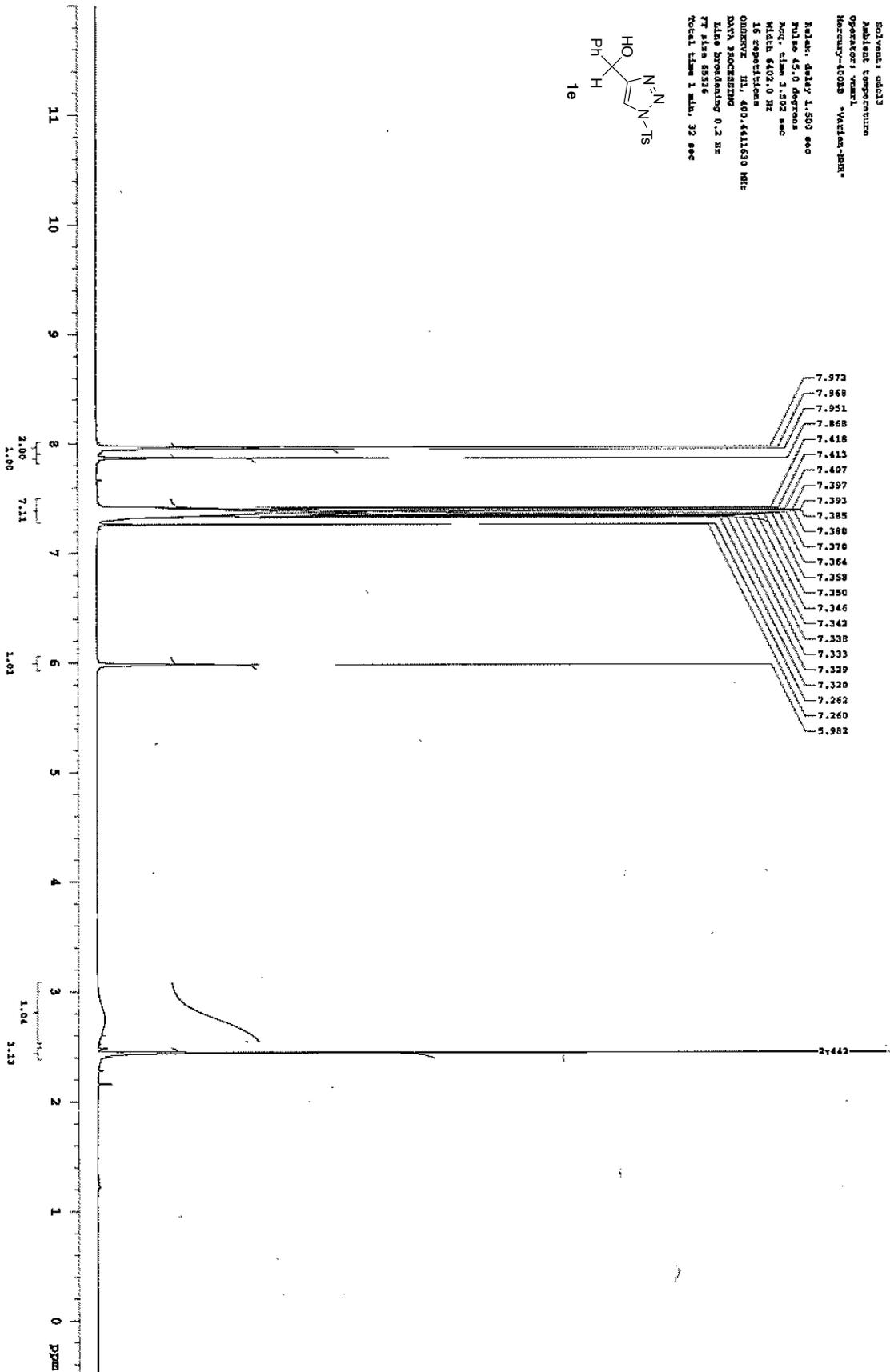
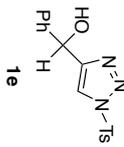
Solvent: dmf-d₂
 Ambient temperature
 Operator: vsmel
 Mercury-60000 "Varian-IBM"
 Relax. delay 1.500 sec
 Pulse 45.0 degrees
 Acq. time 3.502 sec
 Width 602.0 Hz
 IS type: 13C
 OBSERVE: H1, 400.481534 MHz
 DATA PROCESSING
 Line broadening 0.2 Hz
 FT size 65536
 Total time 1 min, 32 sec

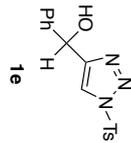
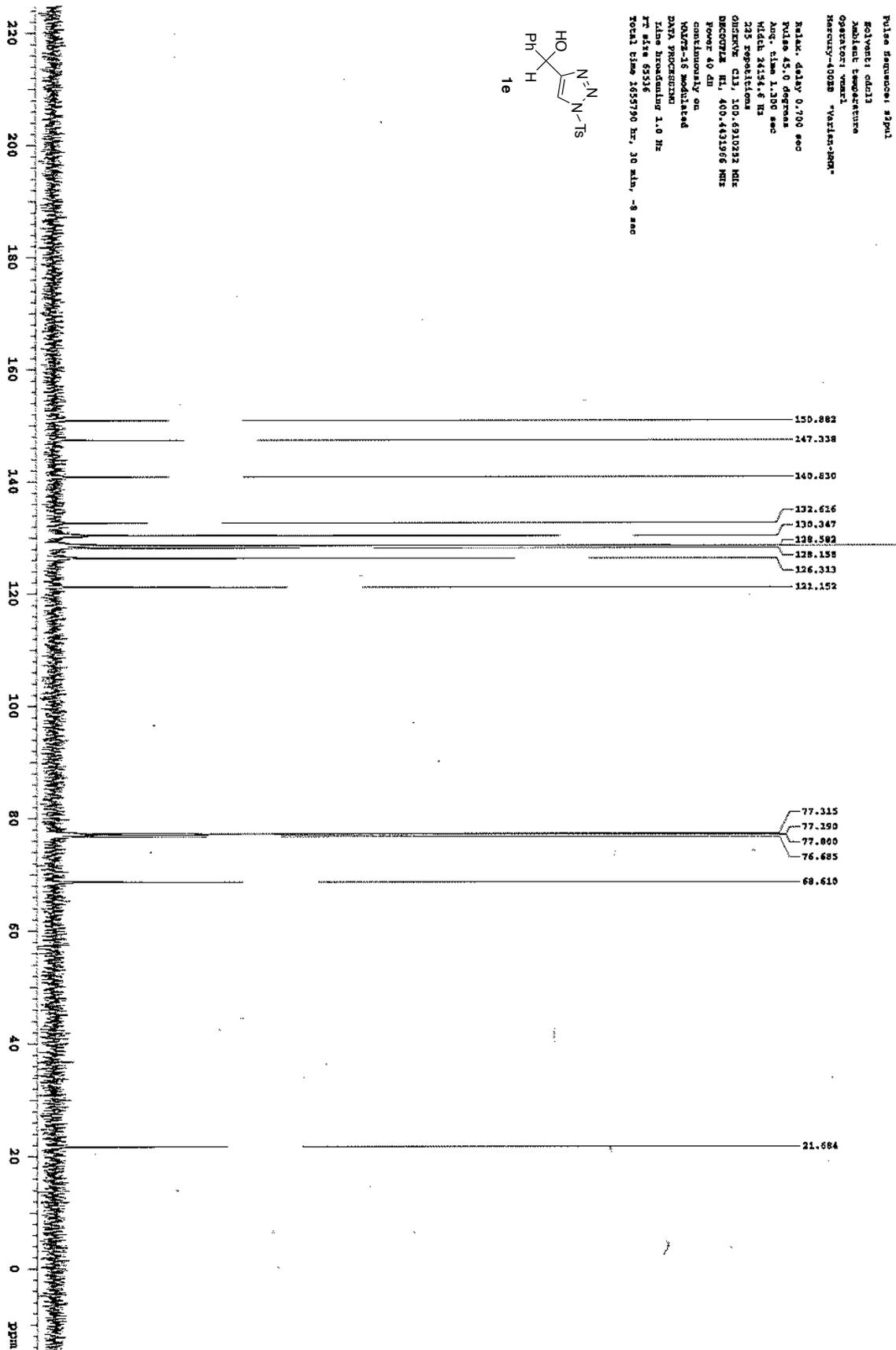


Solvent: cdcl3
 Ambient Temperature
 Operator: yuma1
 Mercury-400MHz -Varian-DMR-
 Pulse delay 0.700 sec
 Relax 45.0 degrees
 Acq. time 1.300 sec
 High 3434.6 Hz
 478 repetitions
 OBSERVE C13 100.6310179 MHz
 PROBEHZ H1 400.431966 MHz
 Power 40 dB
 continuously on
 WATER-16 modulation
 D1M1 300000000
 Line resolution 1.0 Hz
 FT file 023105
 Total time 1033790 hr, 30 min, -8 sec



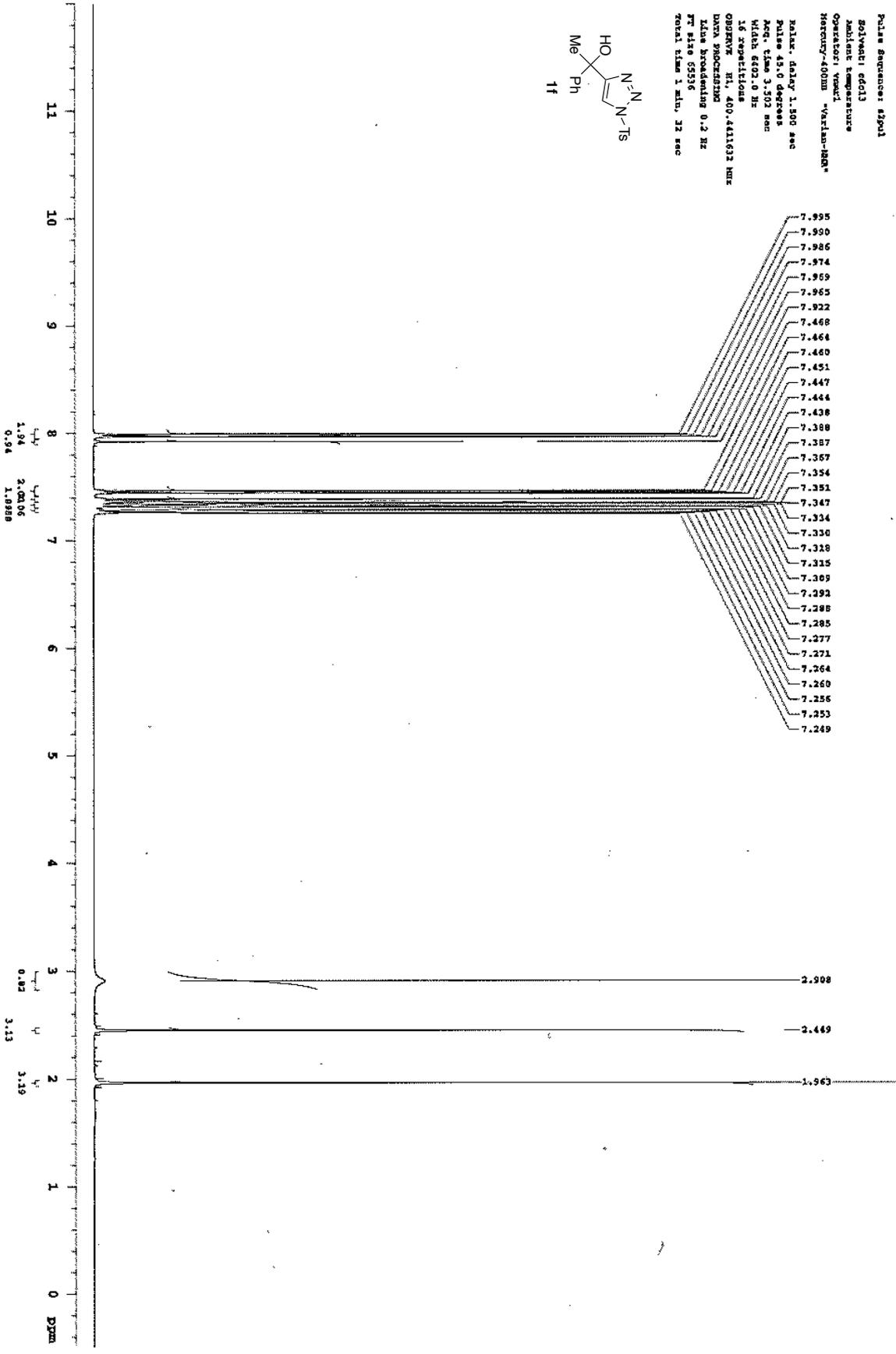
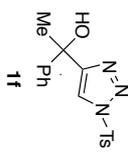
Solvent: dms-d6
 Ambient temperature
 Operator: nmr1
 Name: 4-028 "VARLEN-201"
 Date: 02/02/88
 Pulse delay 1.300 sec
 Pulse 45.0 degrees
 Acq. time 3.502 sec
 MHz 6402.0 Hz
 16 repetitions
 OHSRVE HL 400.441830 MHz
 DATA PROCESSING
 Also broadening 0.2 Hz
 FT also 65315
 Total time 1 min, 32 sec





Pulse program: zgpg30
 Solvent: cdcl3
 Solvent temperature: 25.00
 Operator: vsm
 Mercury-4000 "Varian-90M"
 Pulse delay: 0.100 sec
 Pulse: 15.0 degrees
 Acq. time: 1.200 sec
 Nucleus: 13C
 233 experiments
 OMS: 100.631032 MHz
 DECOUPL: 100.631032 MHz
 Power: 40 dB
 continuously on
 VOLTAGE: 15 modulation
 DATA PROCESSING:
 Line broadening: 1.0 Hz
 FT size: 65536
 Total time: 1655790 hr, 30 min, -8 sec

Pulse sequence: zgpg30
 Solvent: d6d13
 Ambient temperature
 Operator: yowat1
 Mercury-400mhz "Varian-400"
 Relax. delay 1.500 sec
 Pulse 45.0 degrees
 Acq. time 3.502 min
 Width 6402.0 Hz
 16 repetitions
 OBSERVE: H1, 400.441632 MHz
 DATA PROCESSING
 Line broadening 0.2 Hz
 FT file 65336
 Total time 1 min, 32 sec



Pulse sequence: zgpg31

Solvent: cdcl3

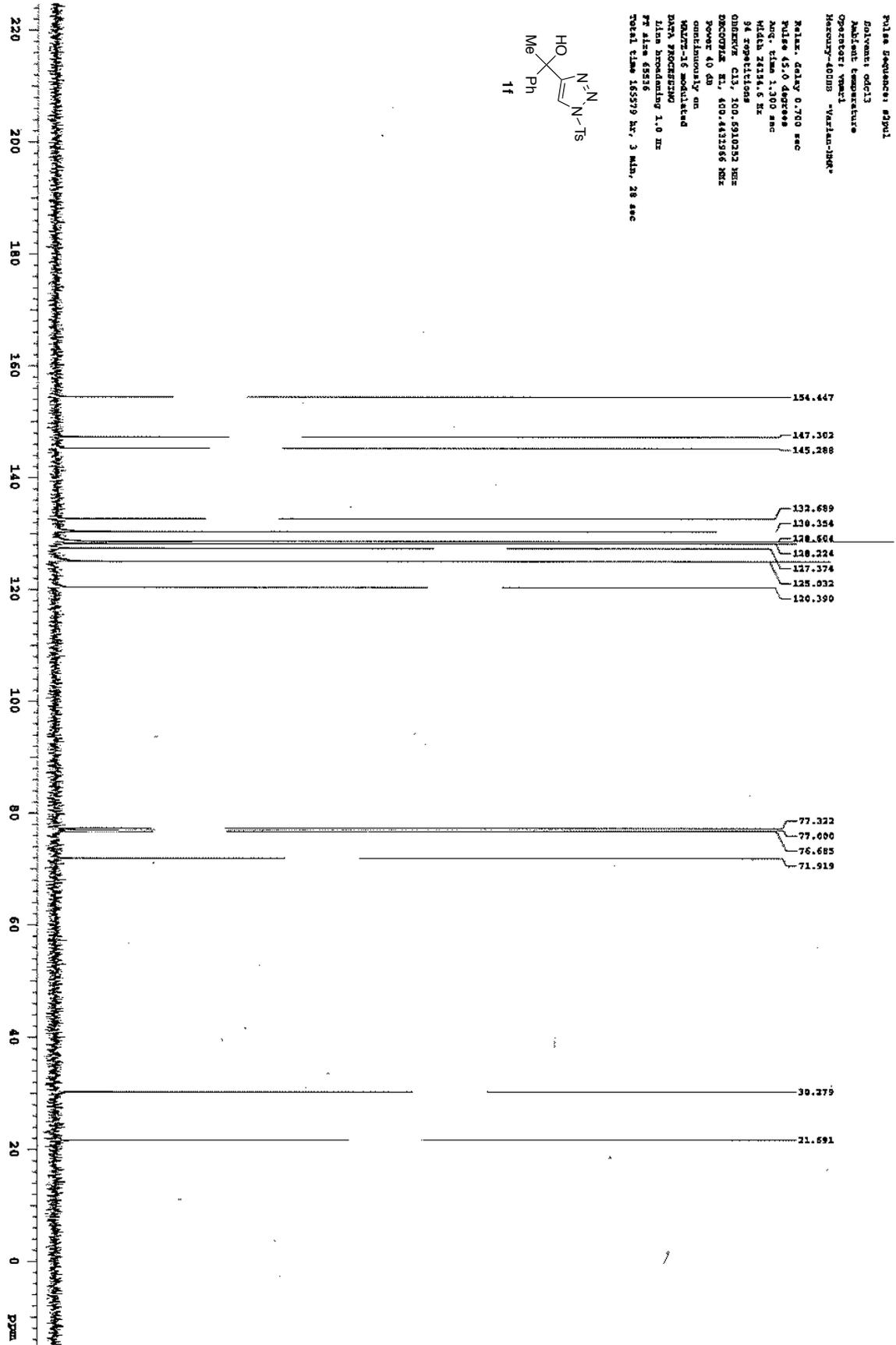
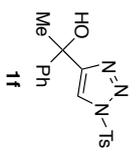
Ambient temperature

Operator: yemw1

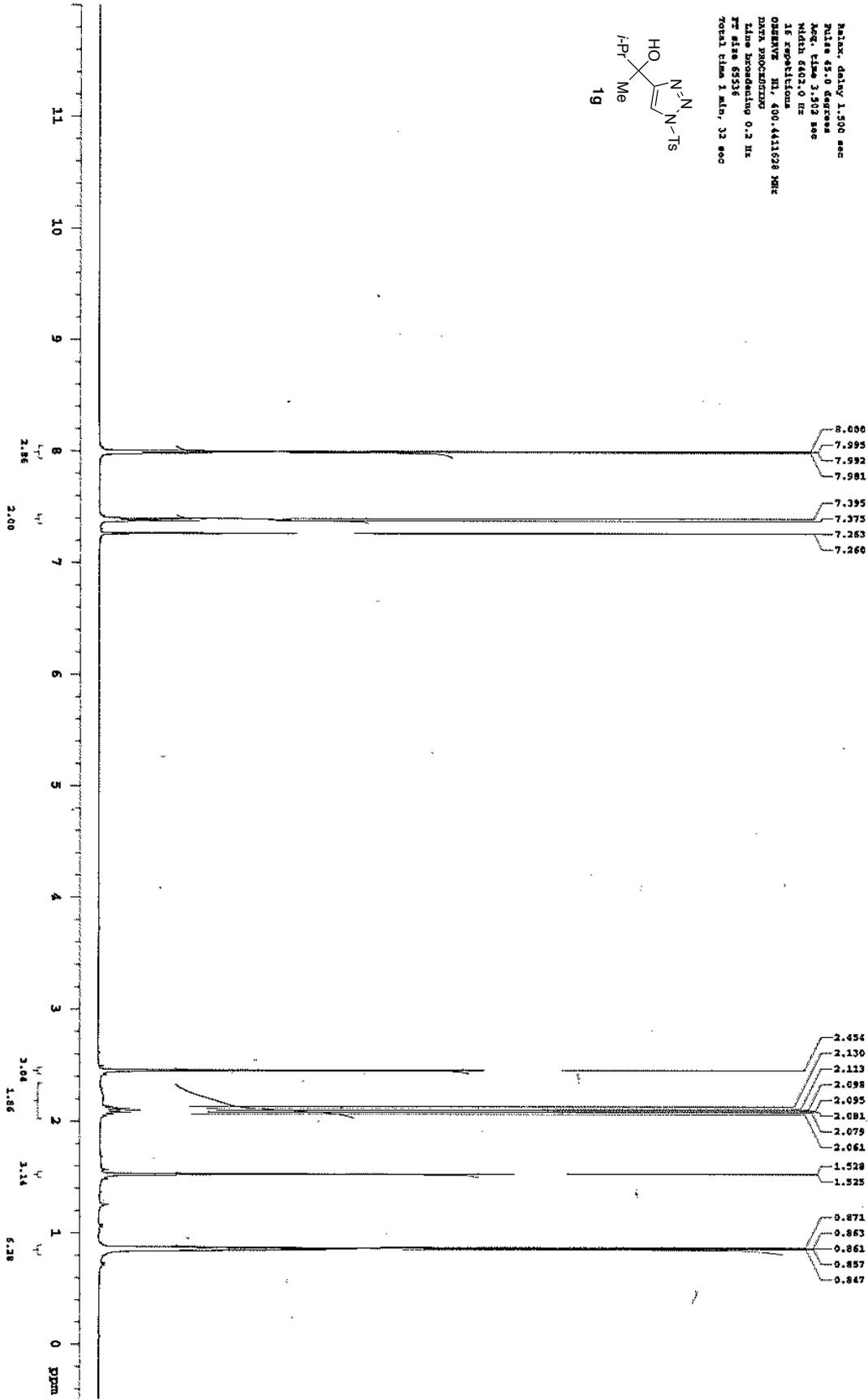
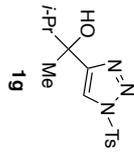
Acquire Date: 11/18/03

Acquire Time: 11:00:00

Acquire Date: 11/18/03

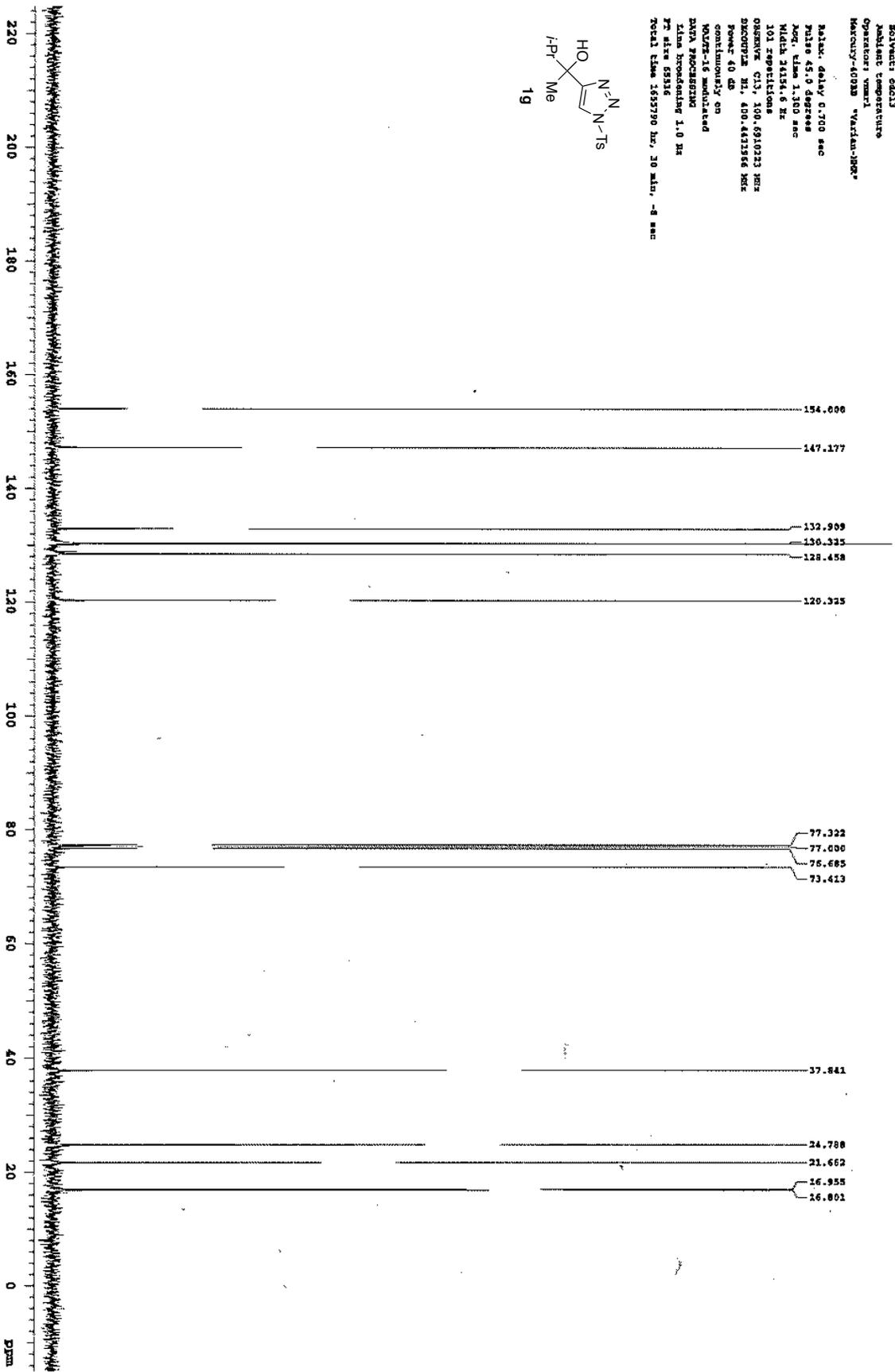
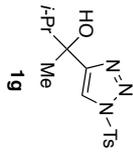


Solvent: ndcl3
 Ambient temperature
 Operator: vmas1
 Name: 400MHz-Varian-2828
 Relax. delay 1.500 sec
 Pulse 45.0 degree
 Acq. time 3.503 sec
 Width 6402.0 Hz
 IS repetitions
 OBSERVE: H1, 400.441528 MHz
 DATA PROCESSING
 Line broadening 0.2 Hz
 FT size 65536
 Total time 1 min, 32 sec

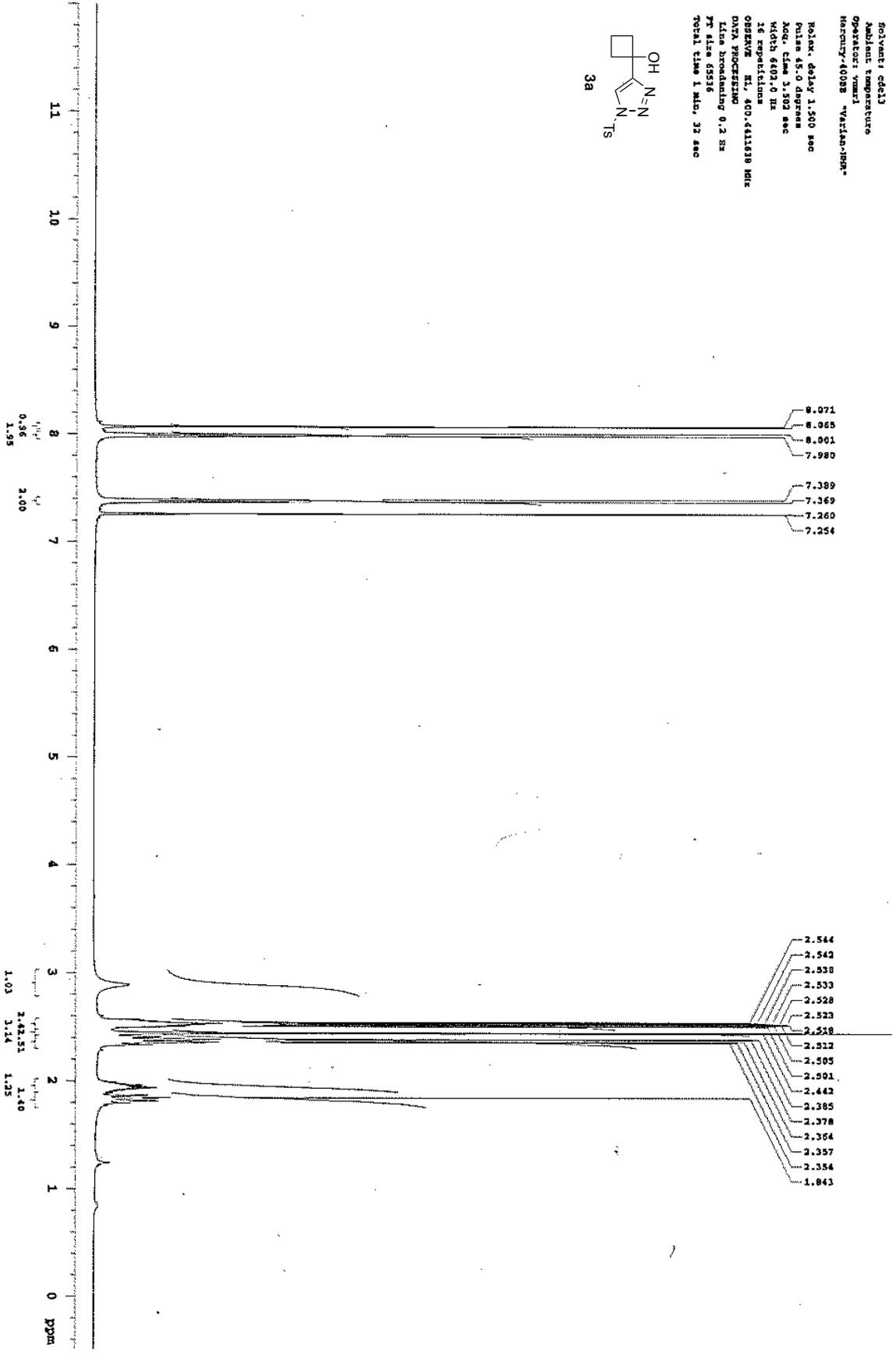
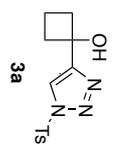


Solvent: cdcl3
 Ambient temperature
 Operator: vmm:1
 Mercury-4020 "Varian-80"

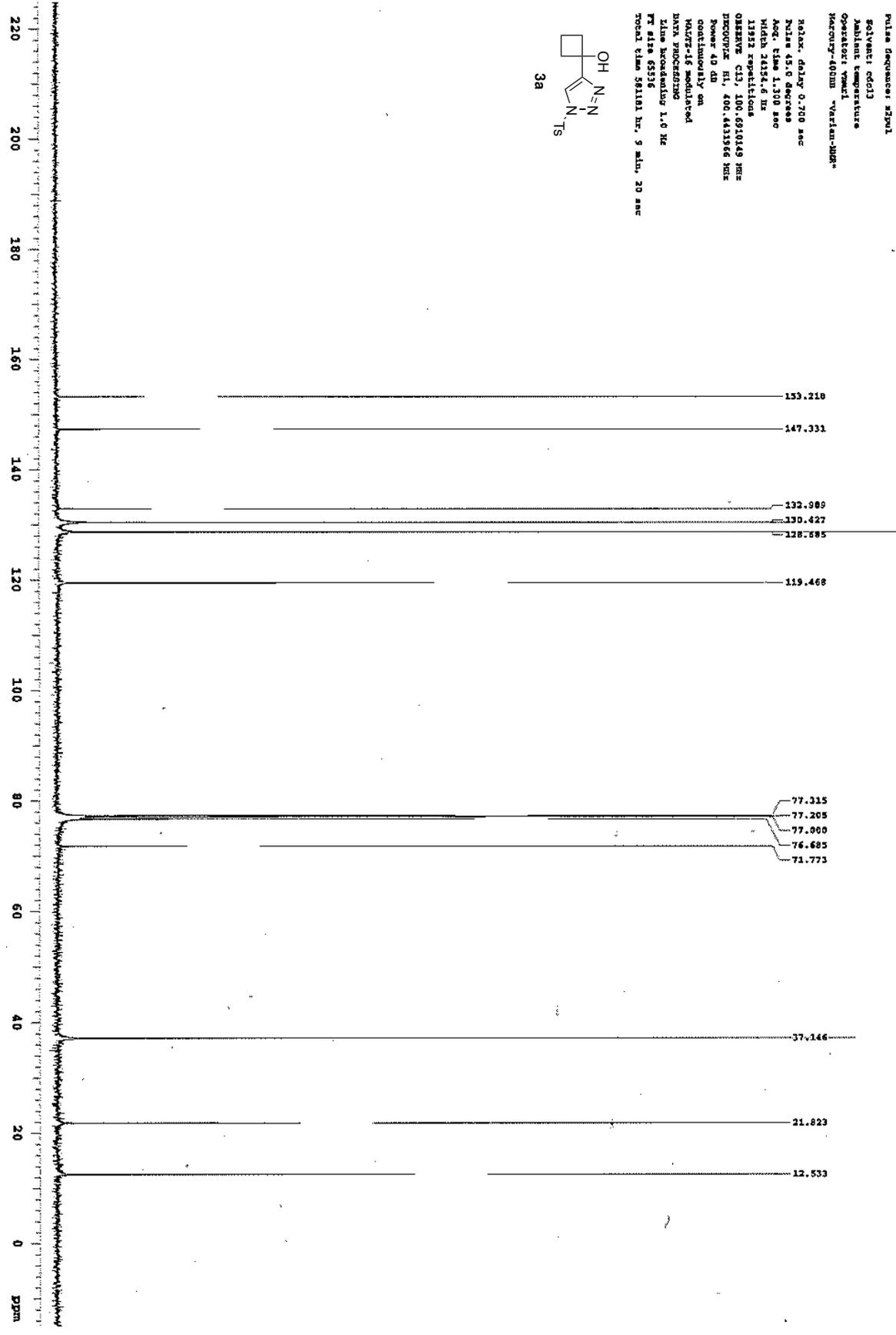
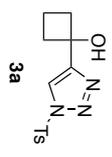
 Pulse delay 0.700 sec
 Pulse 45.0 degrees
 Acq. time 1.300 sec
 Width 34154.6 Hz
 101 repetitions
 OBSERVE C13, 100.631022 MHz
 ZNOFFSET H1, 400.441156 MHz
 Power 40 db
 continuously on
 VOLTAGE-15 modulated
 DATA PROCESSING
 Alpha broadening 1.0 Hz
 FT size 65516
 Total time 1651790 Hz, 30 min, -8 min



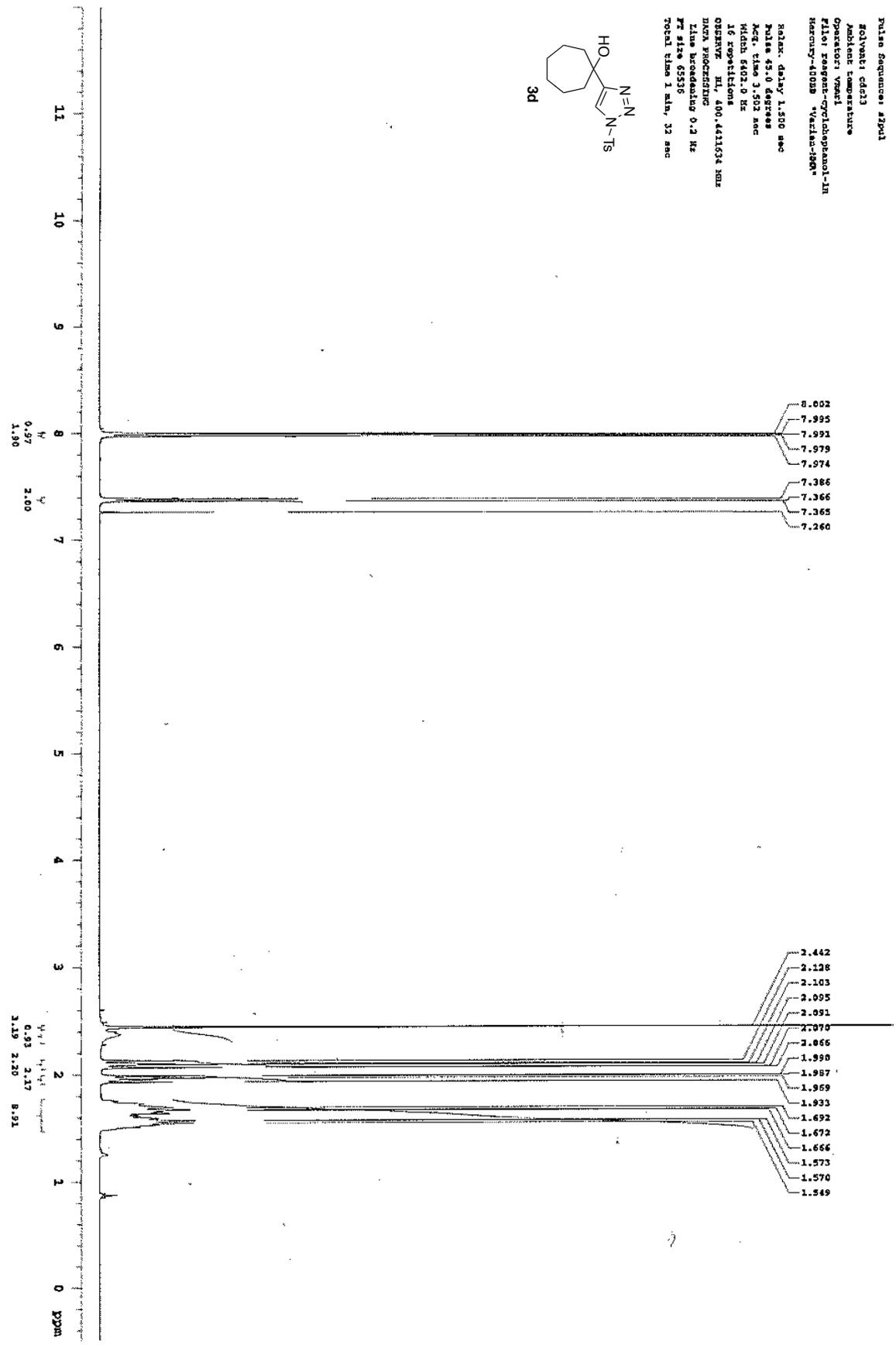
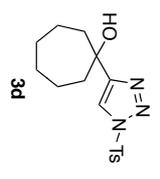
Solvent: cdcl3
 Ambient temperature
 Operator: ymaw1
 Frequency: 400MHz "Varian-JNM-
 Pulse delay 1.500 sec
 Pulse 45.0 degrees
 Acq. time 3.503 sec
 Width 6402.0 Hz
 16 repetitions
 OBSERVE H1, 400.441818 MHz
 DATA PROCESSING
 Line broadening 0.2 Hz
 FT file 65316
 Total time 1 min, 32 sec



Pulse sequence: zgpg30
 Solvent: cdcl3
 Ambient temperature
 Operator: ymml
 Mercury-400mhz "Varian-MR"
 Relax delay: 0.700 sec
 Pulse 45.0 degrees
 Acq. time 1.310 sec
 Width 24224.6 Hz
 13121 repetitions
 OBSERVE C13, 100, 6910149 MHz
 PROCPROG H1, 400, 641356 MHz
 Power 40 dB
 continuously on
 HALFT-16 modulated
 DATA PROCESSING
 Line broadening 1.0 Hz
 FT size 65536
 Total time 58191 hr, 9 min, 20 sec

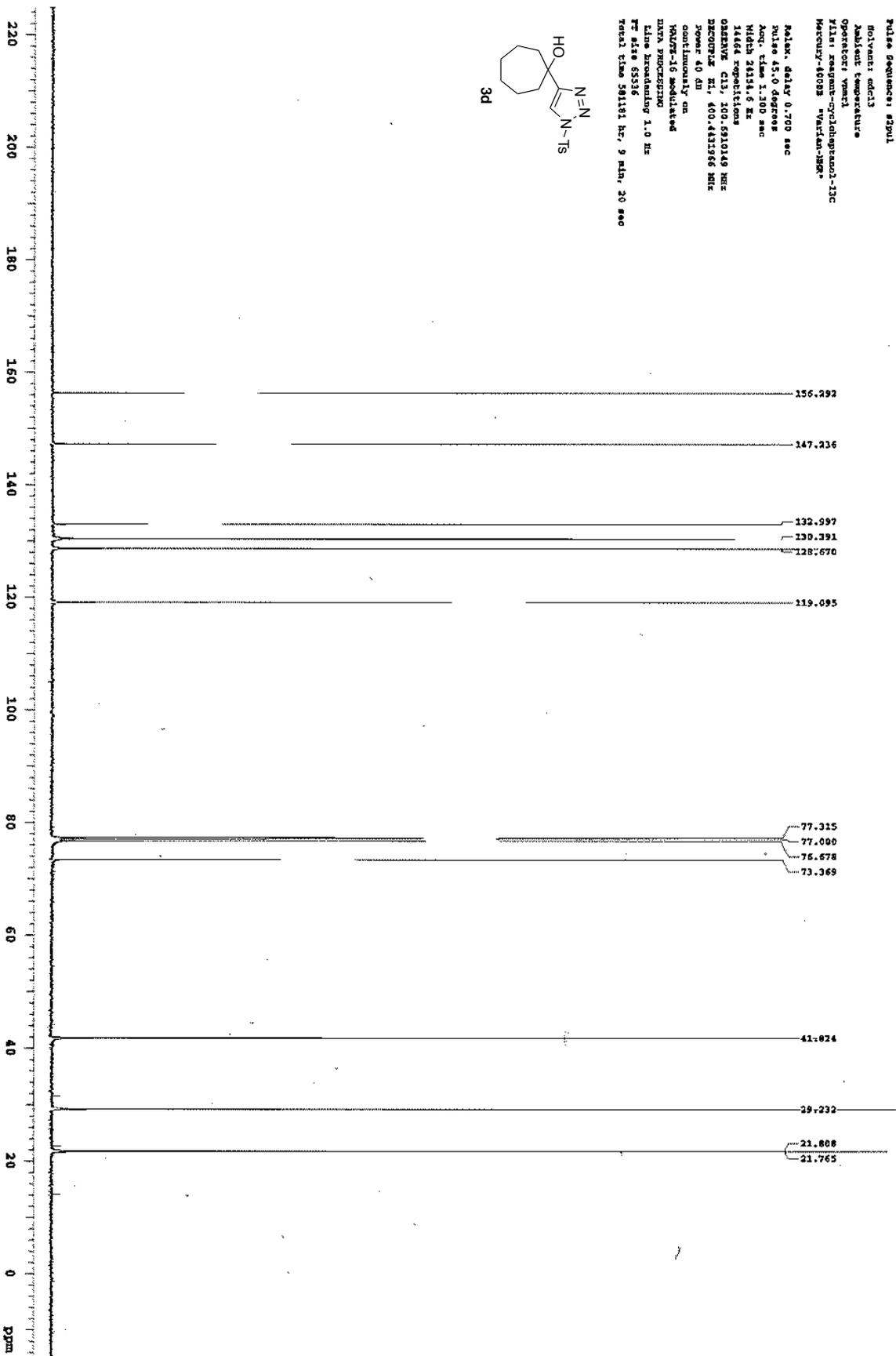
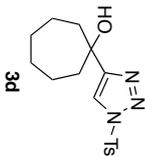


Pulse Sequence: zgpg30
 Solvent: cdcl3
 Solvent Temperature:
 Operator: vmac1
 File: reagent-cyclohexanol-in
 Name: 4028 "Verlaan-180"
 Relax. delay: 1.500 sec
 Pulse: 45.0 degrees
 Acq. time: 3.503 sec
 Width: 6402.0 Hz
 16 repetitions
 OBSERVE: H1, 400.4411534 MHz
 DATA PROCESSING:
 Line broadening: 0.2 Hz
 FT size: 65536
 Total time: 1 min, 33 sec



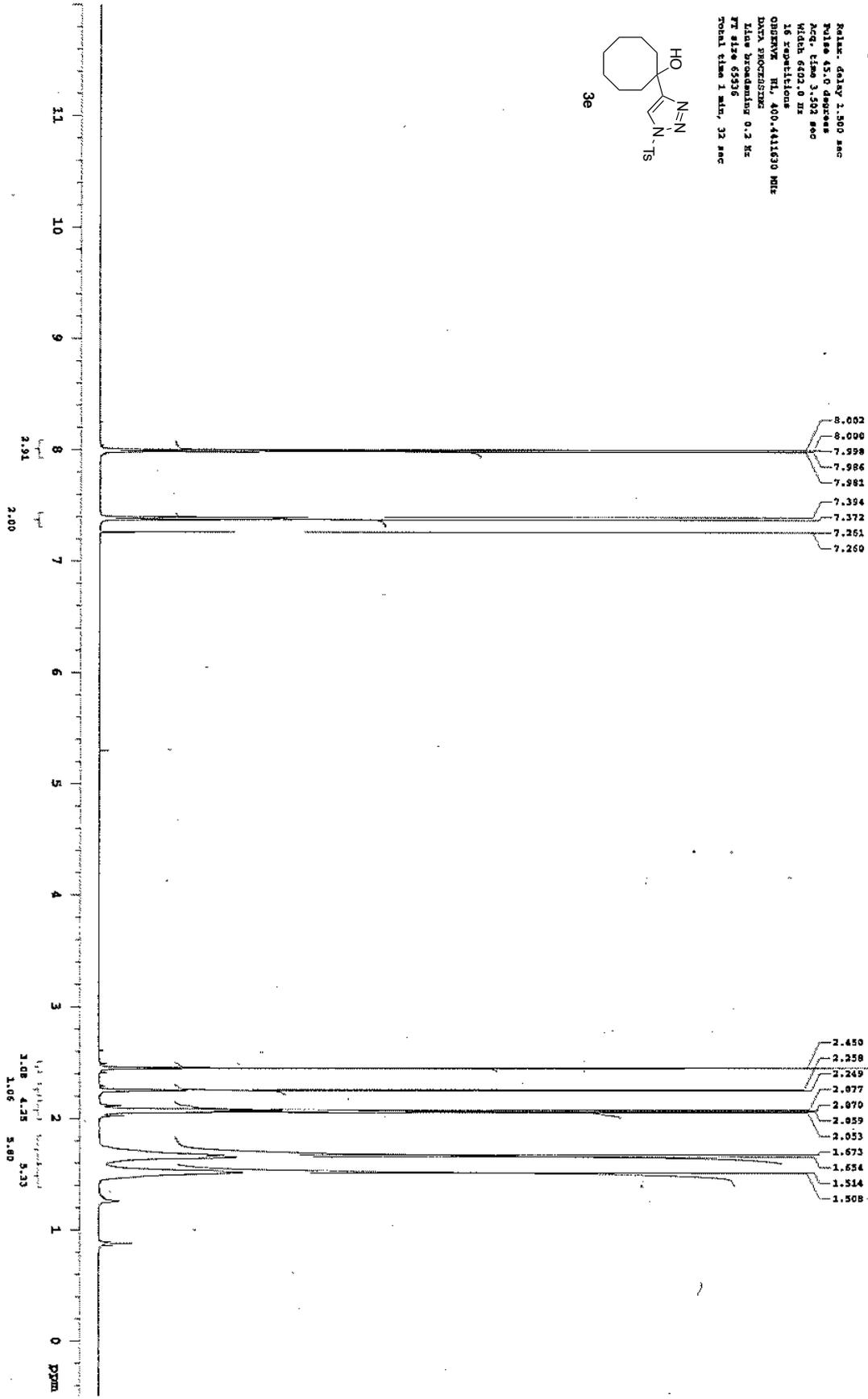
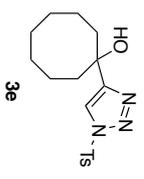
Pulse sequence: zgpg30

Solvent: cdcl3
Acquisition temperature:
Operator: yamyl
Pulse program: zgpg30
Magnetic field: Varian-1H
Pulse delay: 0.700 sec
Pulse: 45.0 degree
Acq. time: 1.300 sec
High: 2134.6 Hz
1464 repetitions
Date_01: 10/28/10
DECODE: 11, 400.413186 MHz
Power: 10.00 W
Sensitivity: 0.00
NOISE: 16.000000
DATA PROCESSING
Line broadening: 1.0 Hz
F2: 414.55356
F1: 101.625181 MHz, 9 min, 30 sec



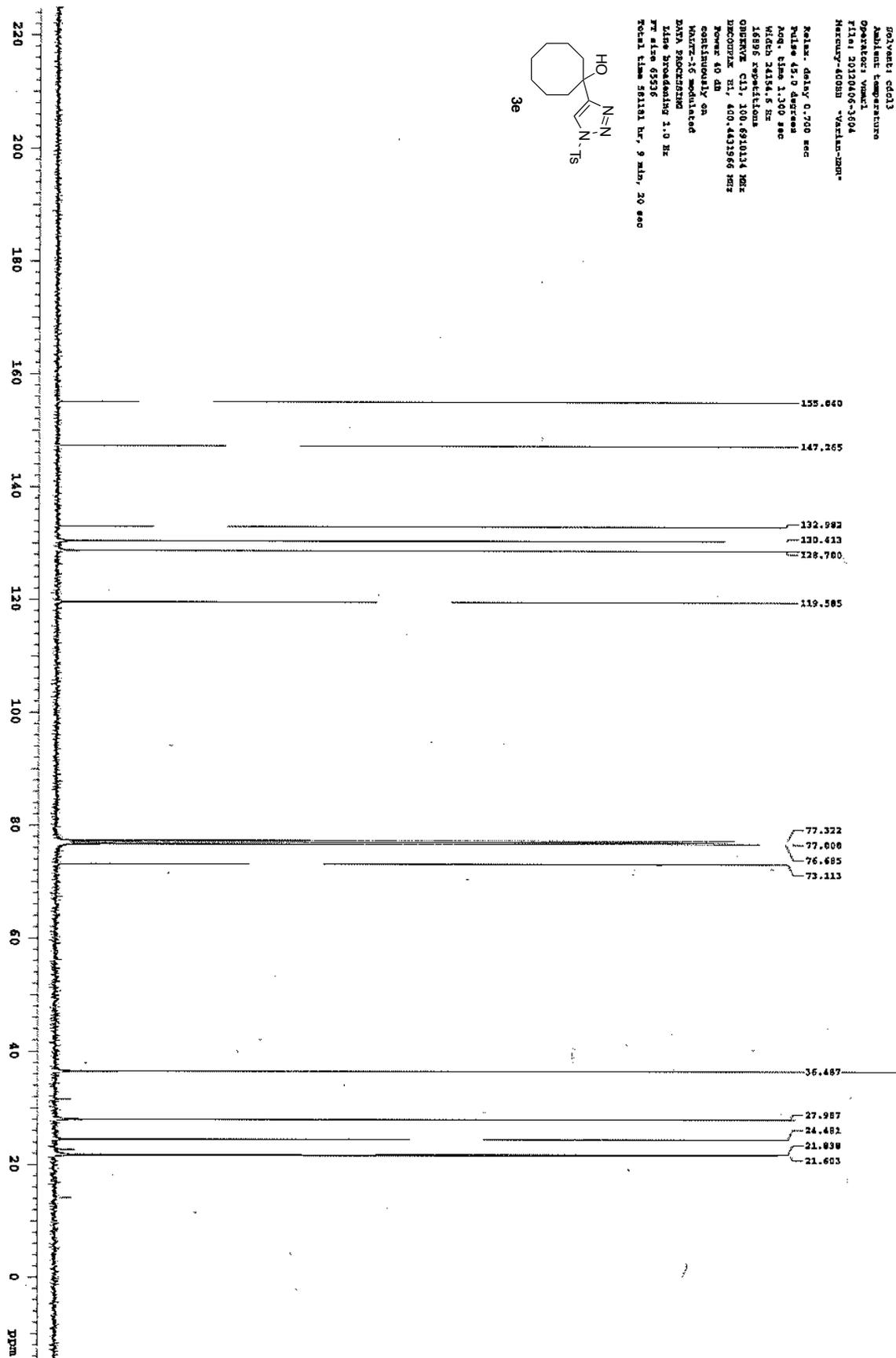
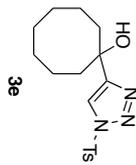
Pulse sequence: zgpg30
 Solvent: cdcl3
 Ambient temperature
 Operator: vmar1
 Mercury-400DB "Varian-DMC"

Pulse delay: 1.500 sec
 Pulse: 45.0 degrees
 Acq. time: 3.592 sec
 Width: 6402.0 Hz
 16 repetitions
 OBSERVE: HL, 400.441630 MHz
 DATA PROCESSING:
 Line broadening: 0.2 Hz
 FT size: 65536
 Total time: 1 min, 32 sec



Solvent: cdcl3
Ambient temperature
Operator: Yumari
File: 20120406-1604
Mercury-400MH "Varian-Vnmr"

Pulse delay: 0.700 sec
Pulse: 42.0 degrees
Acq. time: 1.300 sec
Width: 24184.5 Hz
16395 repetitions
OBSERVE: C13, 100, 6910134 MHz
PNOUSE: H1, 400, 431966 MHz
Power: 40 dB
continuously on
MAGNET-16 modulated
DATA ACQUISITION
1/400: 1.0 Hz
PT: 4.000 sec
Total time: 511.01 hr, 9 min, 20 sec



Pulse sequence: zgpg1

Solvent: dms-d6

Substant: compound

Operator: ymari

Frequency: 400MHz ¹H NMR

Pulse delay: 1.500 sec

Pulse: 45.0 degrees

Acq time: 3.592 sec

Width: 6102.0 Hz

16 experiments

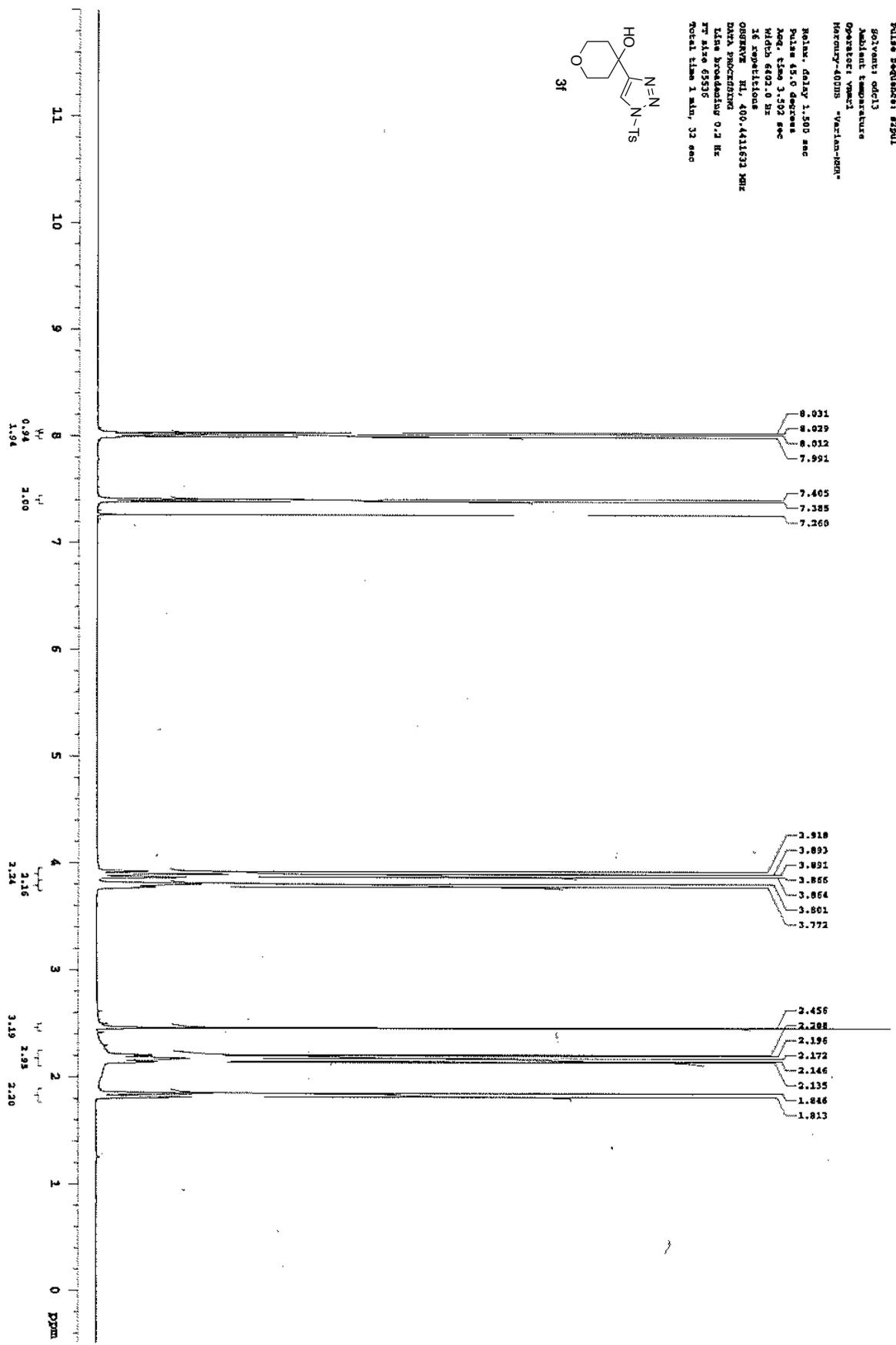
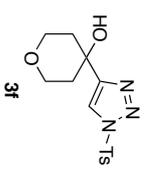
ORIGIN: HJ, 400.411533 MHz

DATA PROCESSING

Line broadening: 0.2 Hz

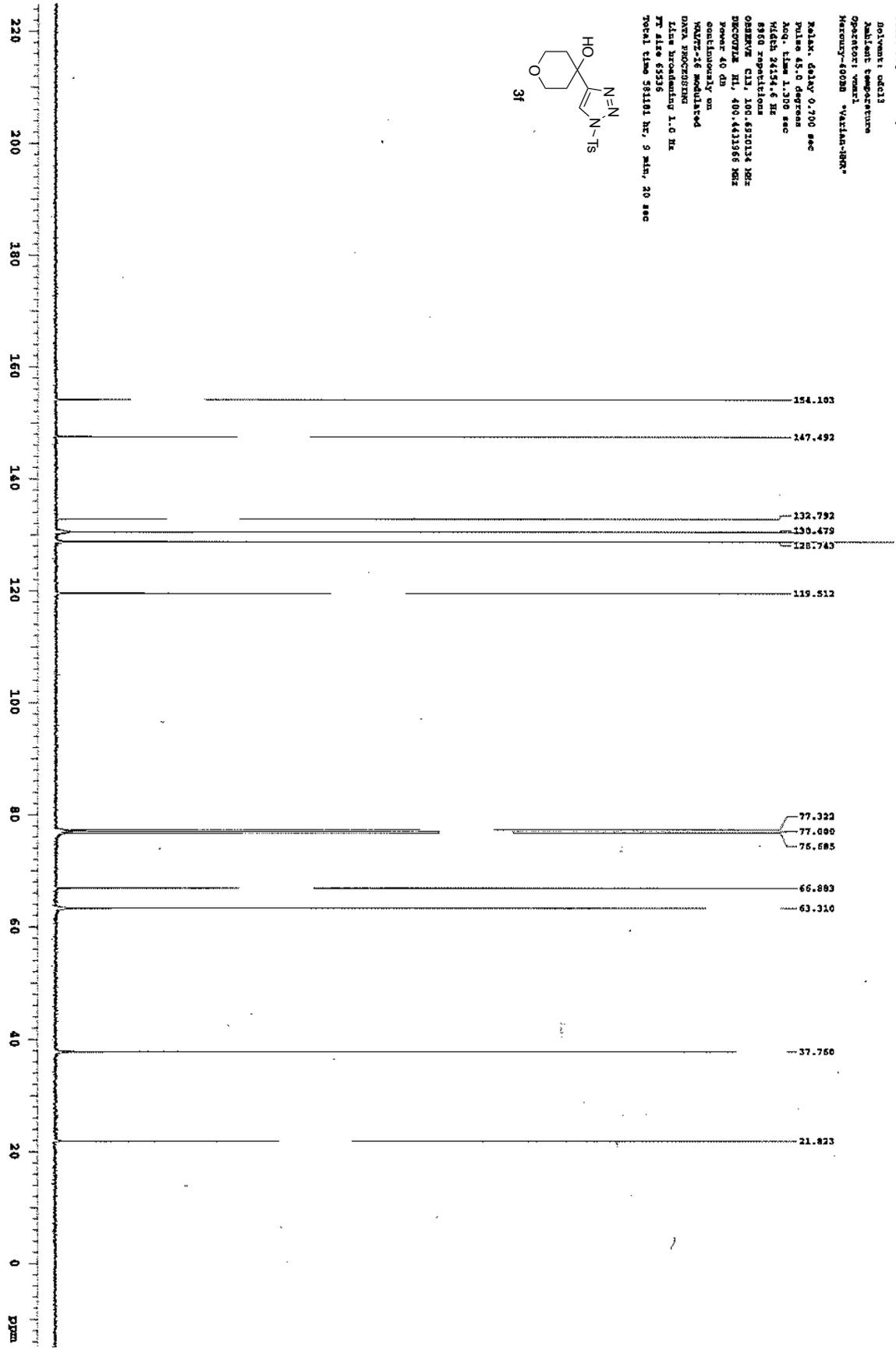
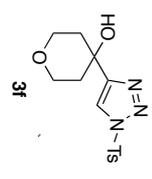
FT Aise: 65510

Relax time: 1 min, 32 sec



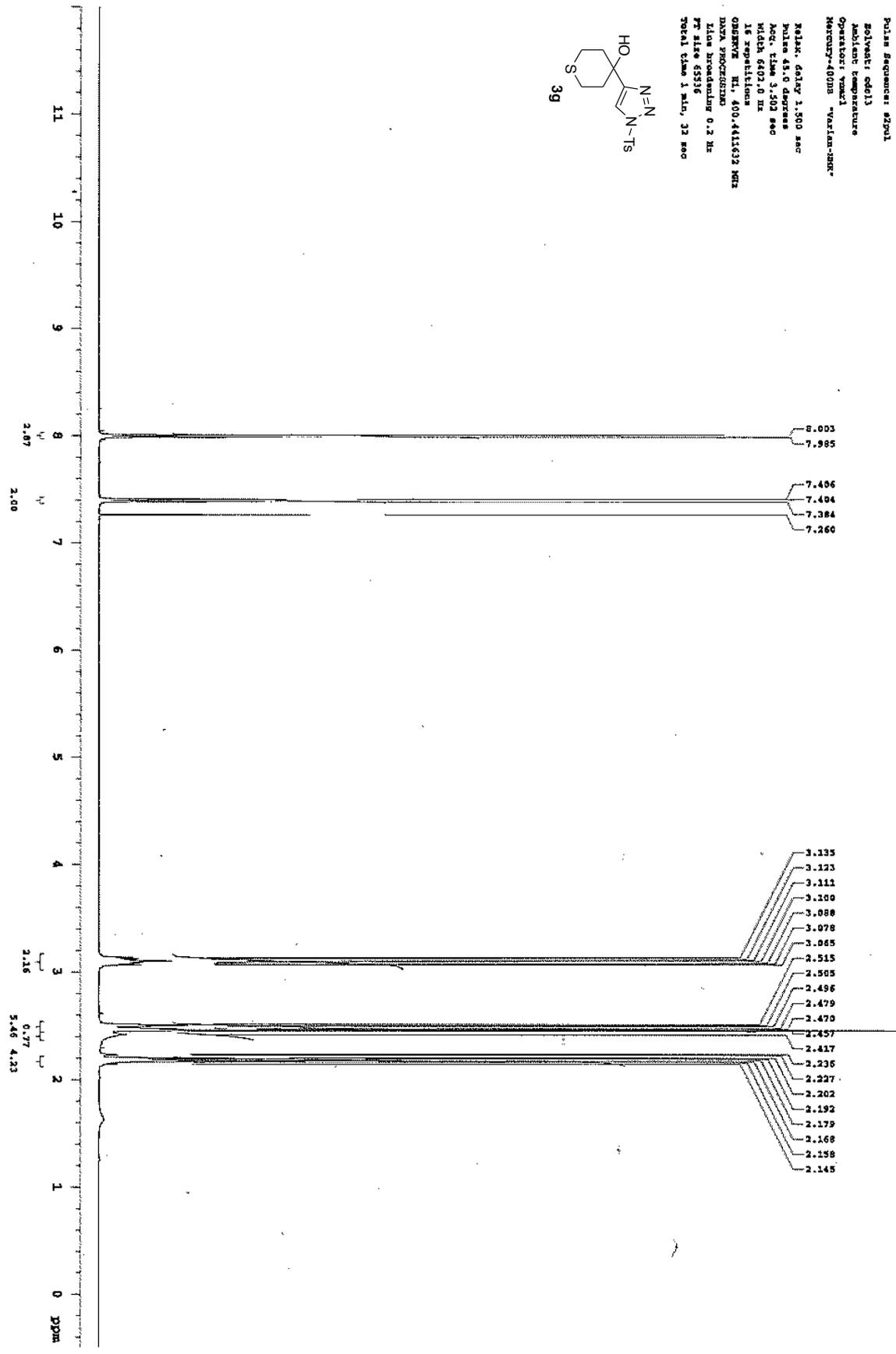
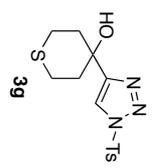
Solvent: dcd13
Sample: 3f
Operator: ymari
Mercury-400MHz Varian-380*

Relax. delay 0.300 sec
Pulse 45.0 degrees
Acq. time 1.300 sec
Width 2434.6 Hz
8160 repetitions
Observer: CIL 100-632013 JMR-
DECEMBER 01, 2004 03:06 PM
F2 F1 40 40
Date_ Acquired: 04
Date_ Processed: 04
Date_ Printed: 1.0 Hz
F2 File: 63205
Total time 58.104 hr, 9 min, 20 sec

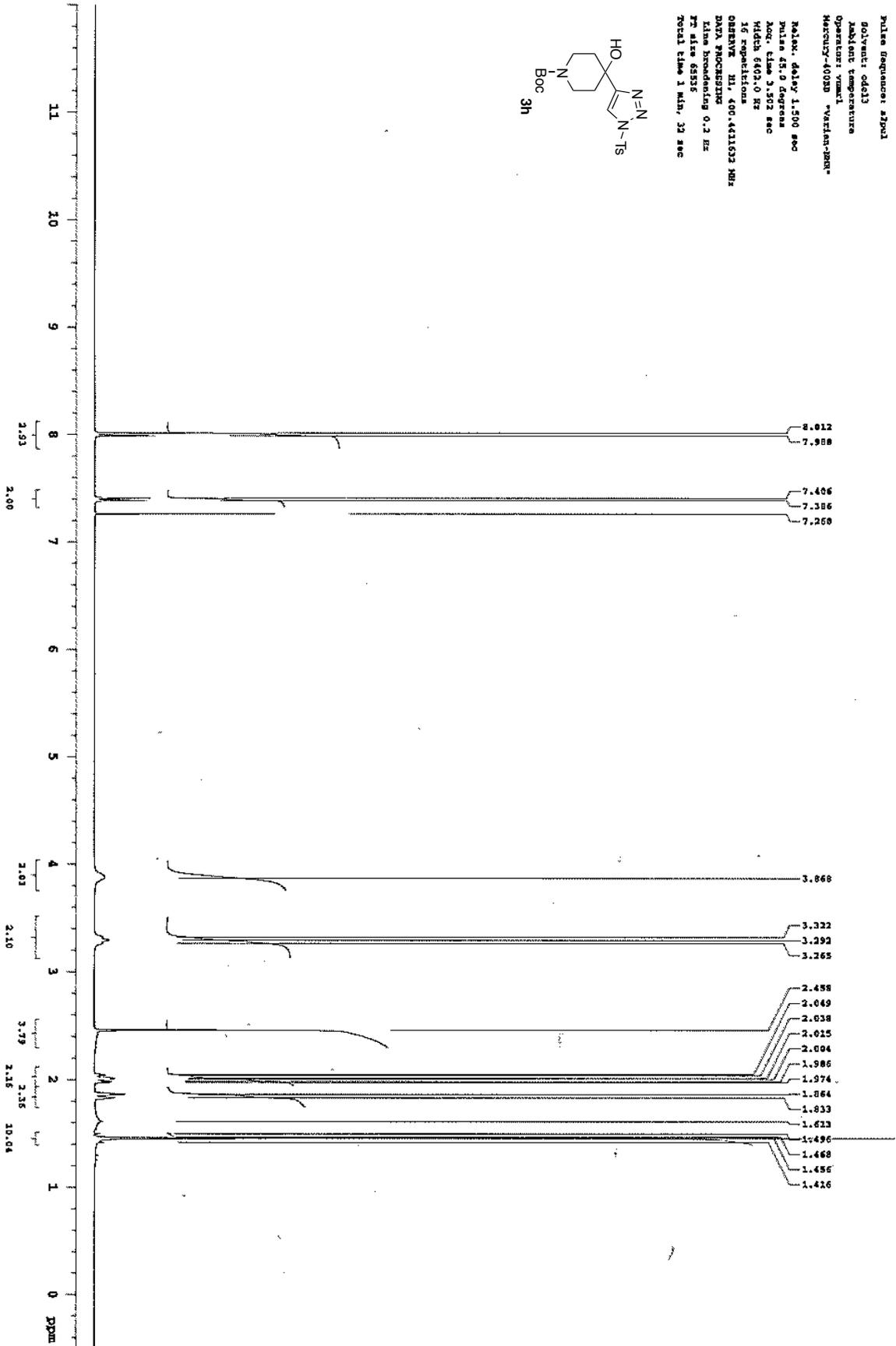
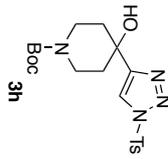


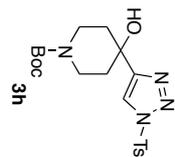
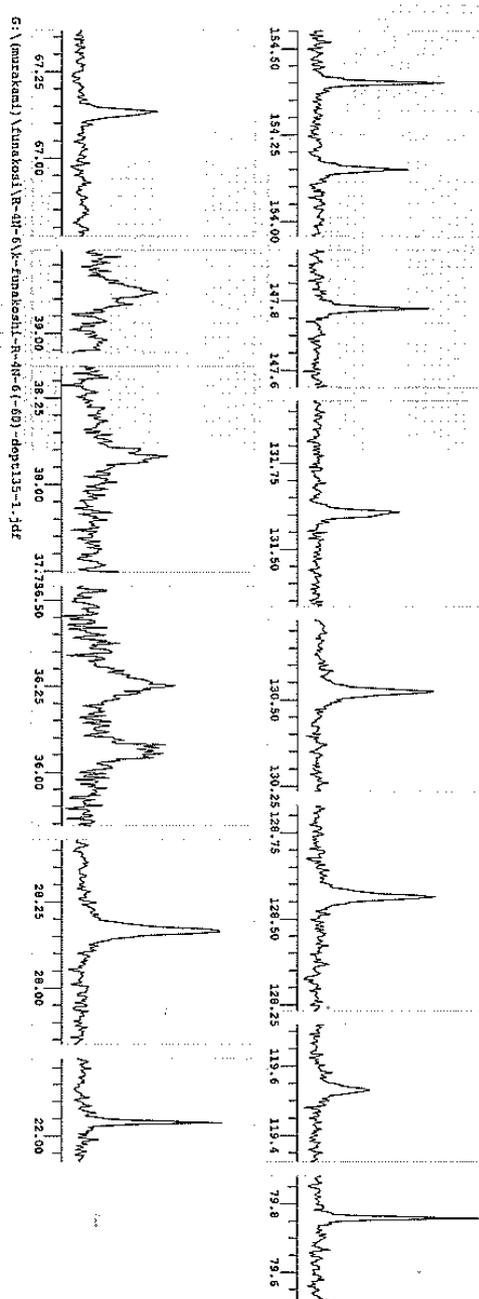
Pulse Sequence: zgpg30
 Solvent: cdcl3
 Molten: temperature
 Operator: vaw1
 Name: 400M-400M "Varian-MR"

 Relax. delay 1.500 sec
 Pulse 45.0 degrees
 Acq. time 3.500 sec
 WALTZ 6402.0 Hz
 16 repetitions
 OBSERVE H1, 400.4411532 MHz
 DATA PROCESSING
 Line broadening 0.2 Hz
 FT size 45336
 Total time 1 min, 32 sec

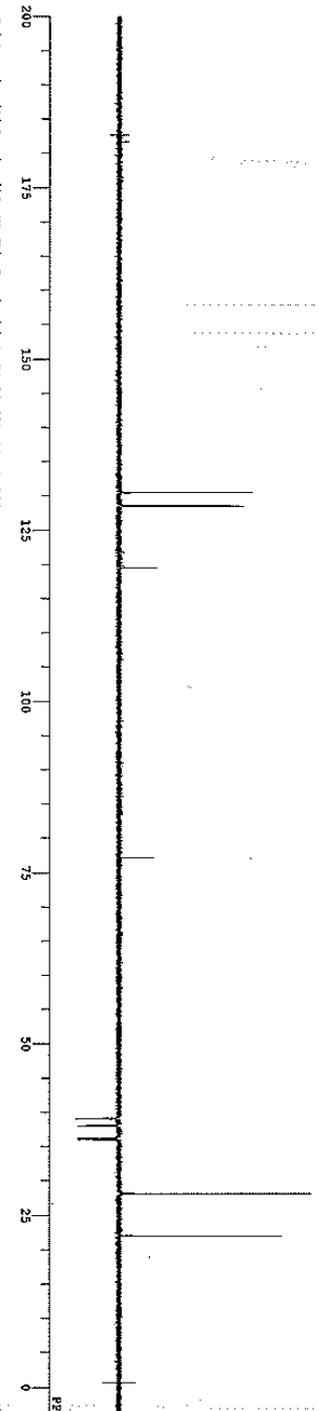


Pulse frequency: 490.1
 Solvent: cdcl3
 Solvent temperature:
 Operator: ymaki
 Machine: 400MHz Varian-DMX
 Relax delay: 1.500 sec
 pulse: 45.0 degrees
 Acq. time: 3.502 sec
 Width: 6400.0 Hz
 16 repetitions
 OBSERVE: H1, 400.641532 MHz
 DATA PROCESSING
 Line broadening: 0.2 Hz
 F2 alias: 85315
 Total time: 1 Min, 32 sec





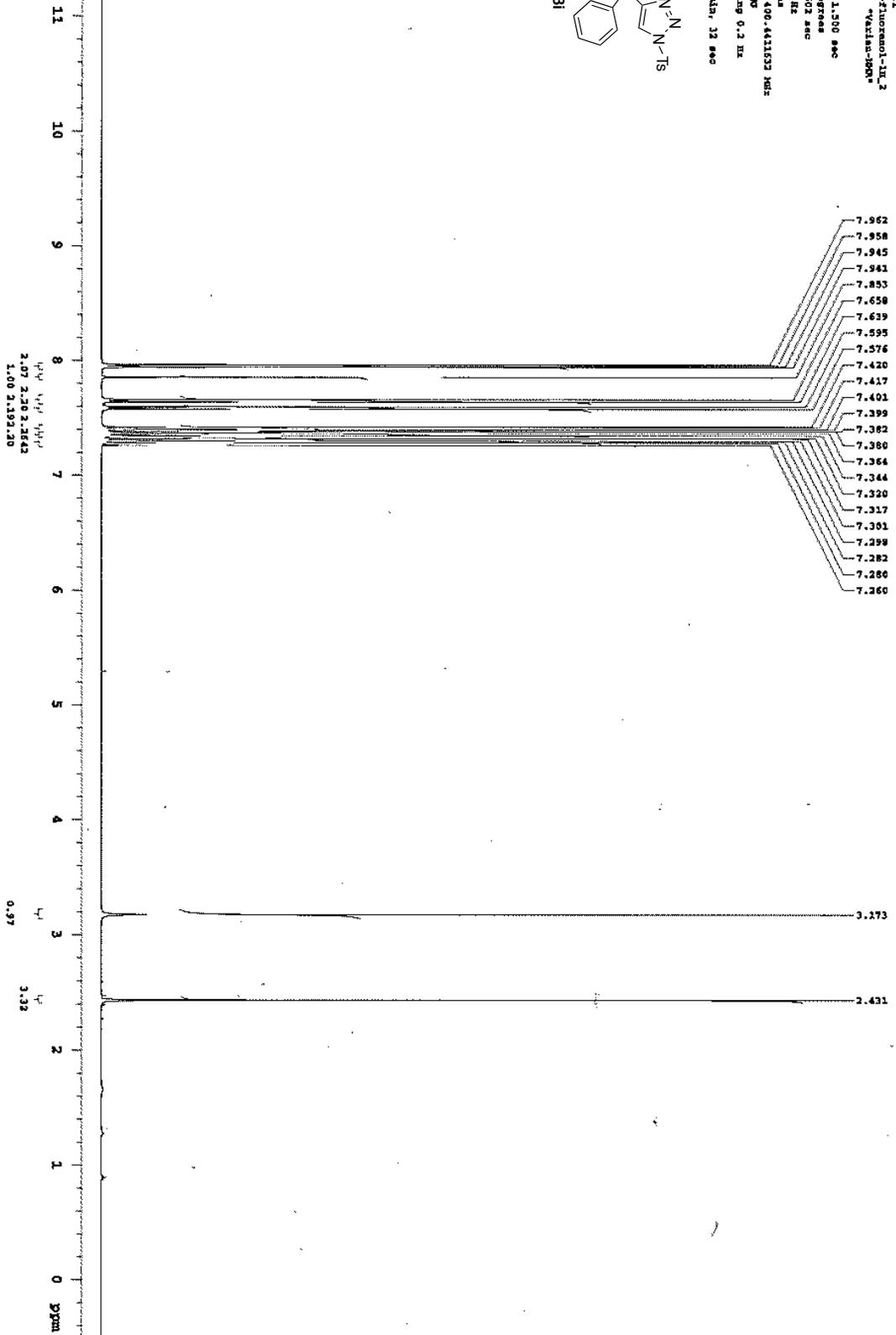
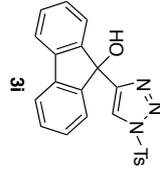
930375



DEPT135 13C
 COUNT 3897
 DATE_1 19-07-2012 12:16:54
 DEPT135
 EXH000 100.53 MHz
 OBFREQ 5.166 Hz
 PULPROG zgpg30
 PROCNO 2
 XCPDPR 2.3598 sec
 PD 2.0000 sec
 F1 6.00 usec
 IRRUCD 1H
 SLEWHT 77.00 ppm
 EXH000 0.40 Hz
 RECALC 50

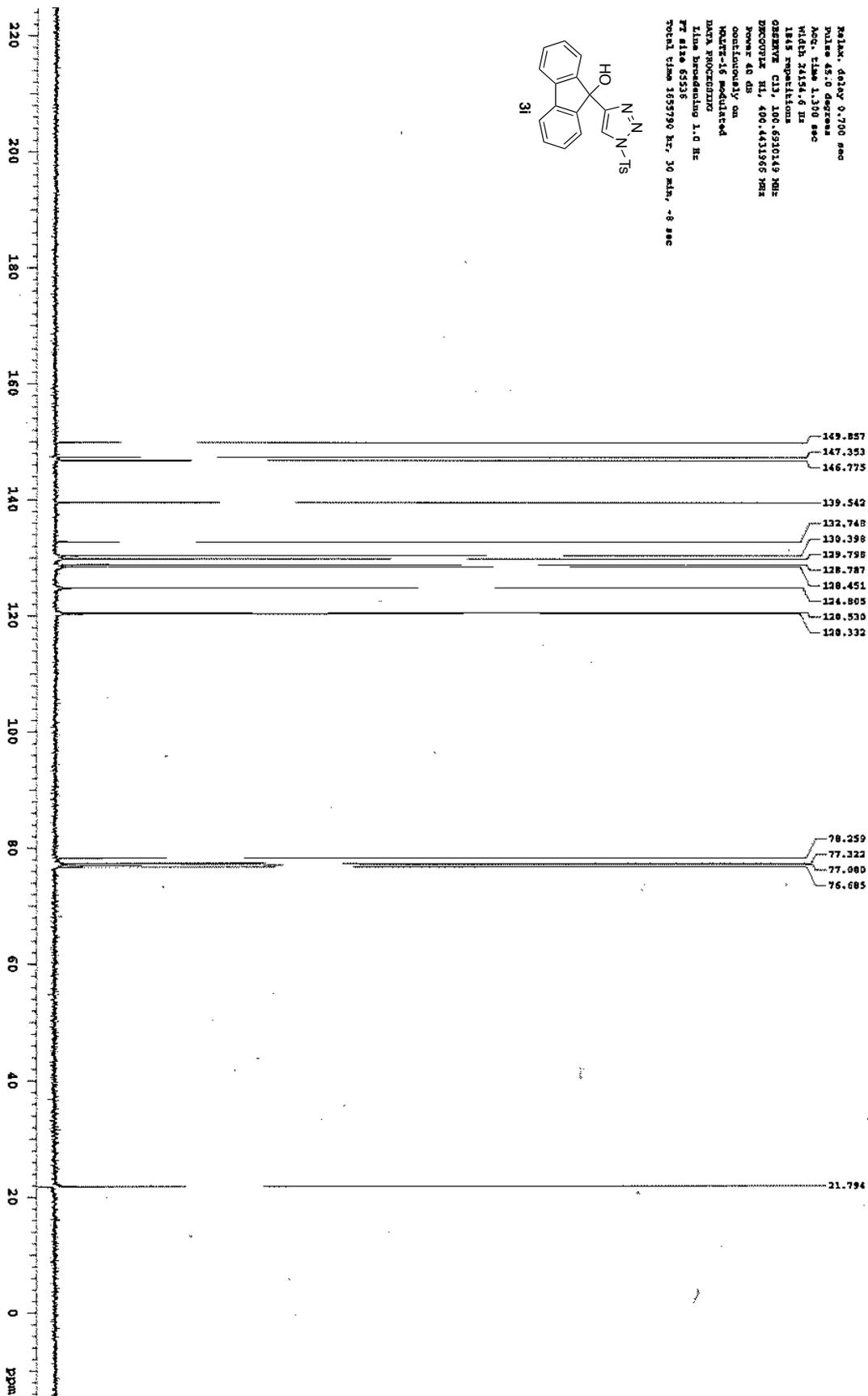
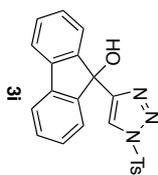
DEPT135 13C
 COUNT 3897
 DATE_1 19-07-2012 12:16:54
 DEPT135
 EXH000 100.53 MHz
 OBFREQ 5.166 Hz
 PULPROG zgpg30
 PROCNO 2
 XCPDPR 2.3598 sec
 PD 2.0000 sec
 F1 6.00 usec
 IRRUCD 1H
 SLEWHT 77.00 ppm
 EXH000 0.40 Hz
 RECALC 50

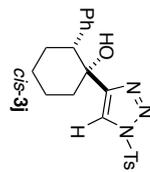
Solvent: cdcl3
 Ambient temperature
 Operator: vmmwl
 File: rmgdnt-fluoreno1-111_2
 Mercury-403B *Varian-189P*
 Pulse delay 1.500 sec
 Pulse 45.0 degrees
 Acq. time 3.502 sec
 Width 6002.0 Hz
 16 repetitions
 OBSERVE HI, 400.441573 MHz
 DATA PROCESSING
 Line broadening 0.2 Hz
 FT size 65536
 Total time 1 min, 12 sec



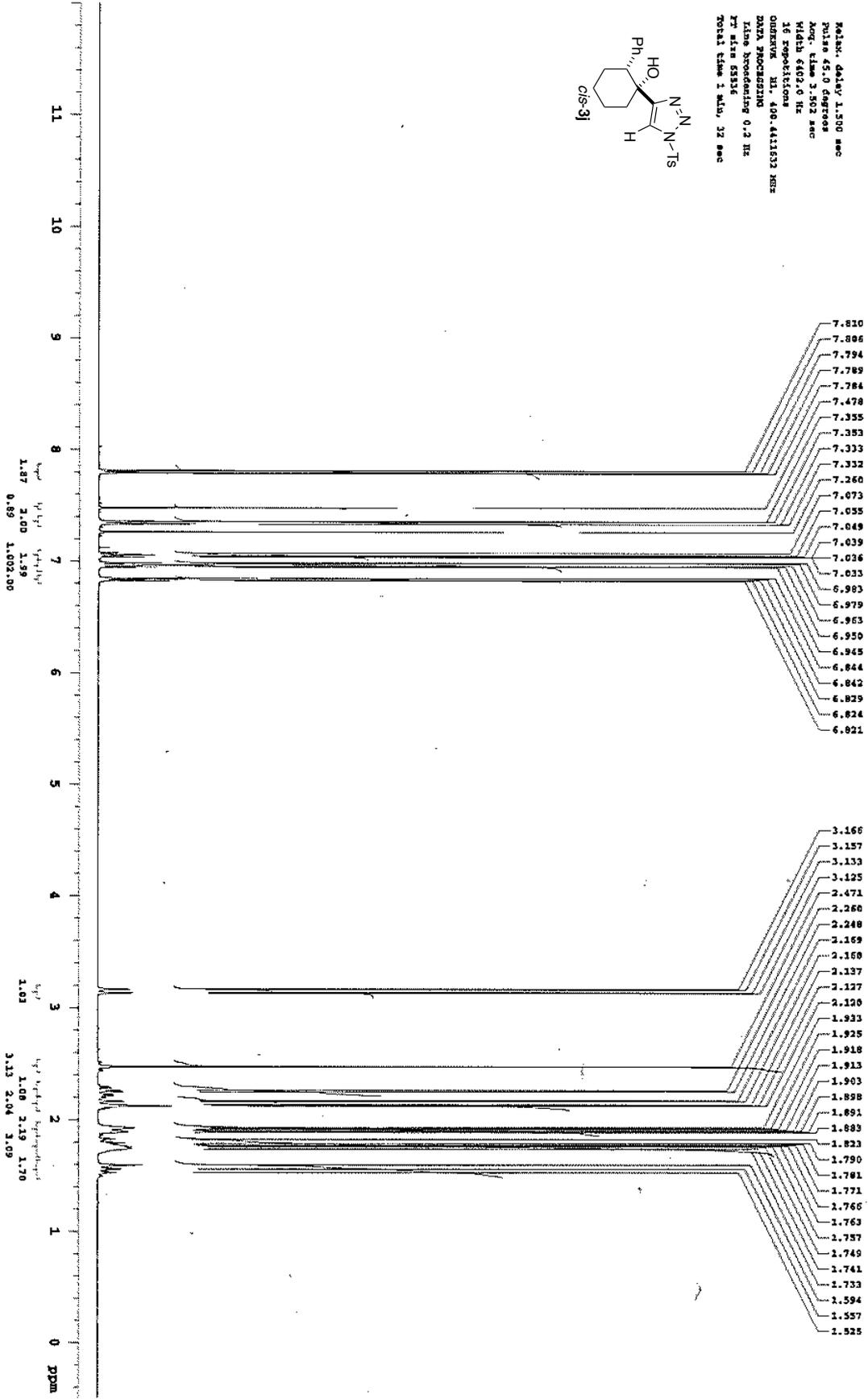
Mercury-400MHz Varian-SM-

Peak: 4242 0.100 sec
Pulse: 4570 degrees
Acq. Time: 1.170 sec
Width: 14154.0 Hz
1845 repetitions
OBSERVE CH: 100.630149 MHz
PROBHD: H1, 400.441365 MHz
PROB: 40 mm
continuously on
PULPROG: zgpg30
DATA PROCESSING
Time Resolving: 1.0 Hz
FT file: 65535
Total time: 165790 Hz, 30 min, -8 sec





Relax. delay 1.500 sec
 Pulse 45.0 degrees
 Acq. time 3.502 sec
 Width 6402.0 Hz
 16 repetitions
 CHANNEL M1 400.441833 MHz
 DATA PROCESSING
 Name broadening 0.2 Hz
 FT file 55315
 Total time 1 min, 32 sec



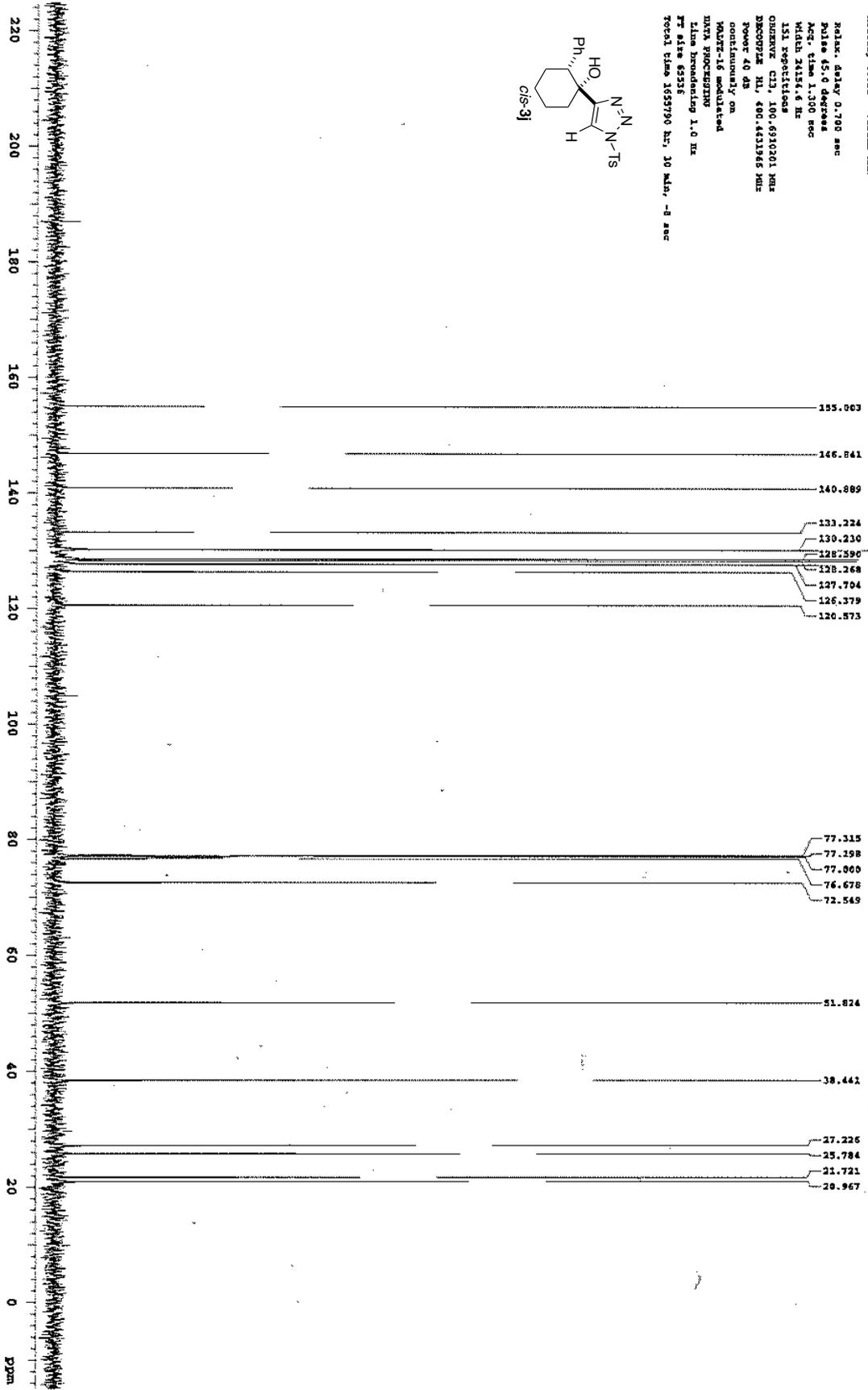
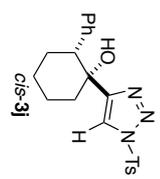
=====

Solvent: dcdcl3
Pulsed Temperature
Operator: ymwt
Hexony-400MHz Varian-EMR-

Phase delay 0.795 sec
Pulse 45.0 degrees
Acq time 1.310 sec
Width 2418.4 Hz

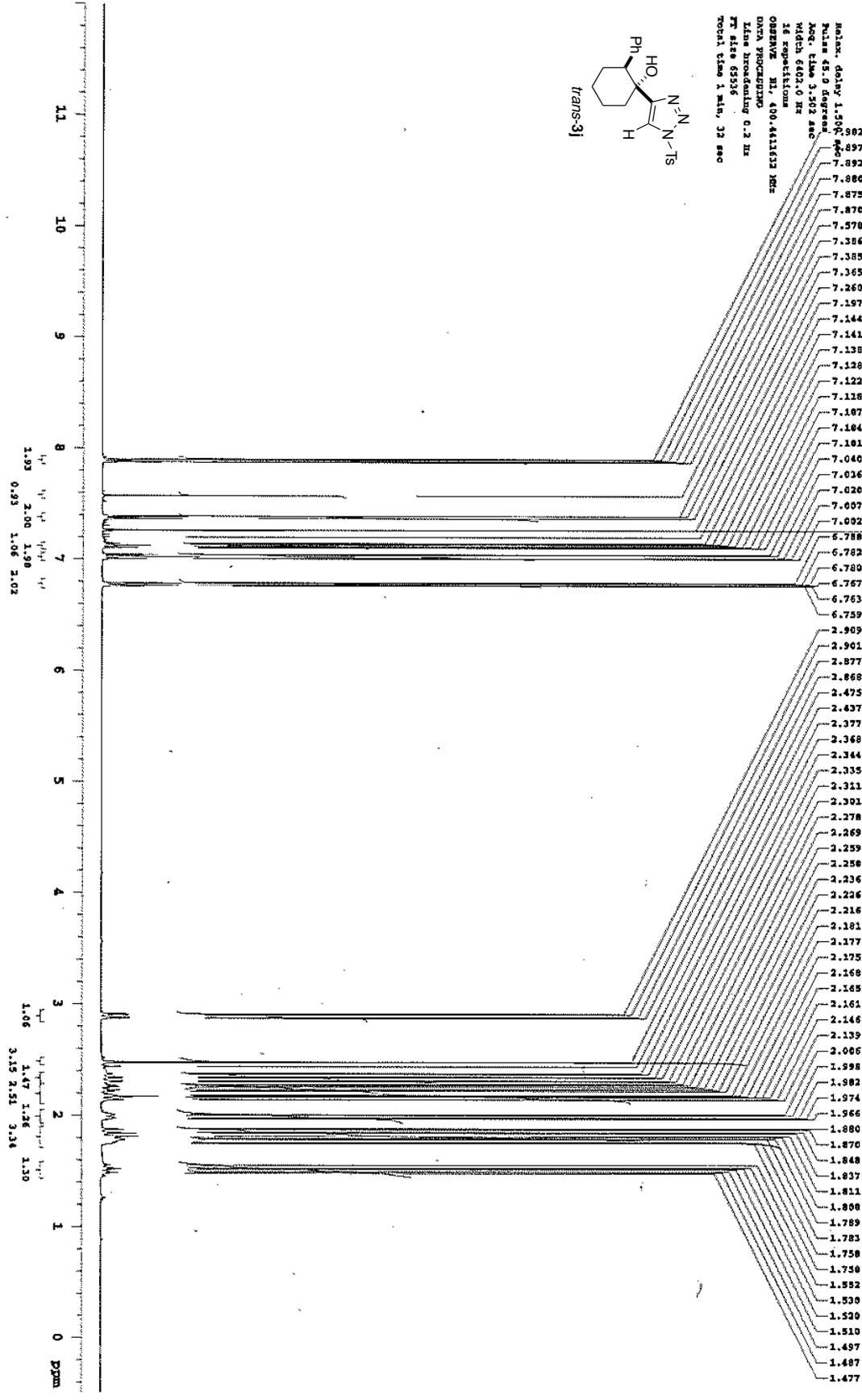
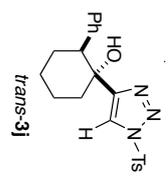
131 Spectral
Chemical C12, 100.6310261 MHz
Decouple N1, 800.421565 MHz
Power 4.0 dB
Continuously on

DATA PROCESSING
Line broadening 1.0 Hz
F2 file 63235
Total time 1833790 Hz, 30 Min, -8 sec



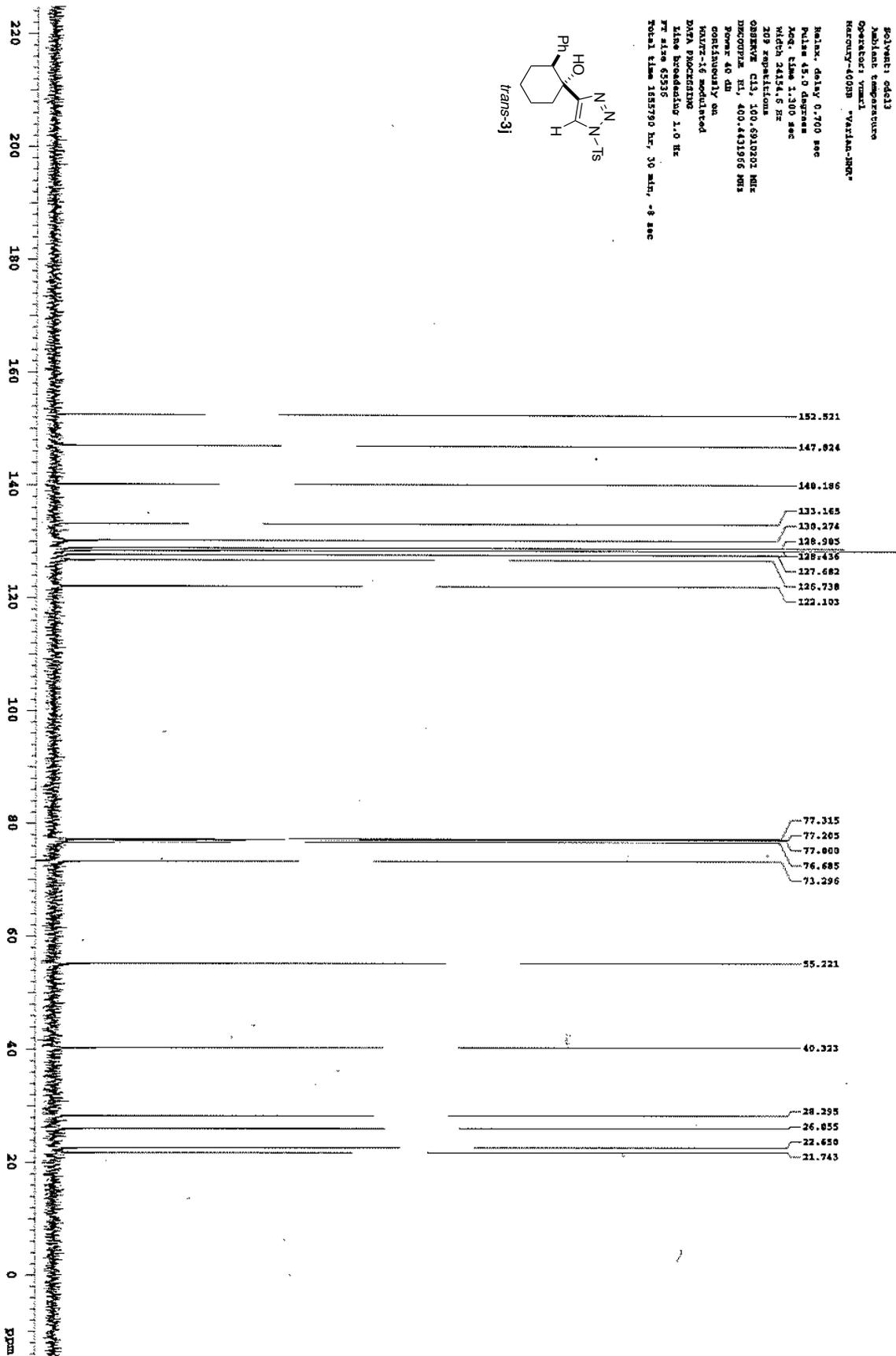
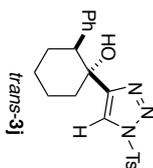
Pulse Sequence: zgpg31
 Solvent: cdcl3
 Ambient temperature
 Operator: vmm1
 Mercury-400WB "Varian-100"

Relax. delay 1.500 sec
 Pulse 45.0 degrees
 Acq. time 3.503 sec
 Width 6403.0 Hz
 16 repetitions
 OBSERVE HI, 400.441633 MHz
 DATA PROCESSING
 Line broadening 0.2 Hz
 FT file 65336
 Total time 1 min, 32 sec

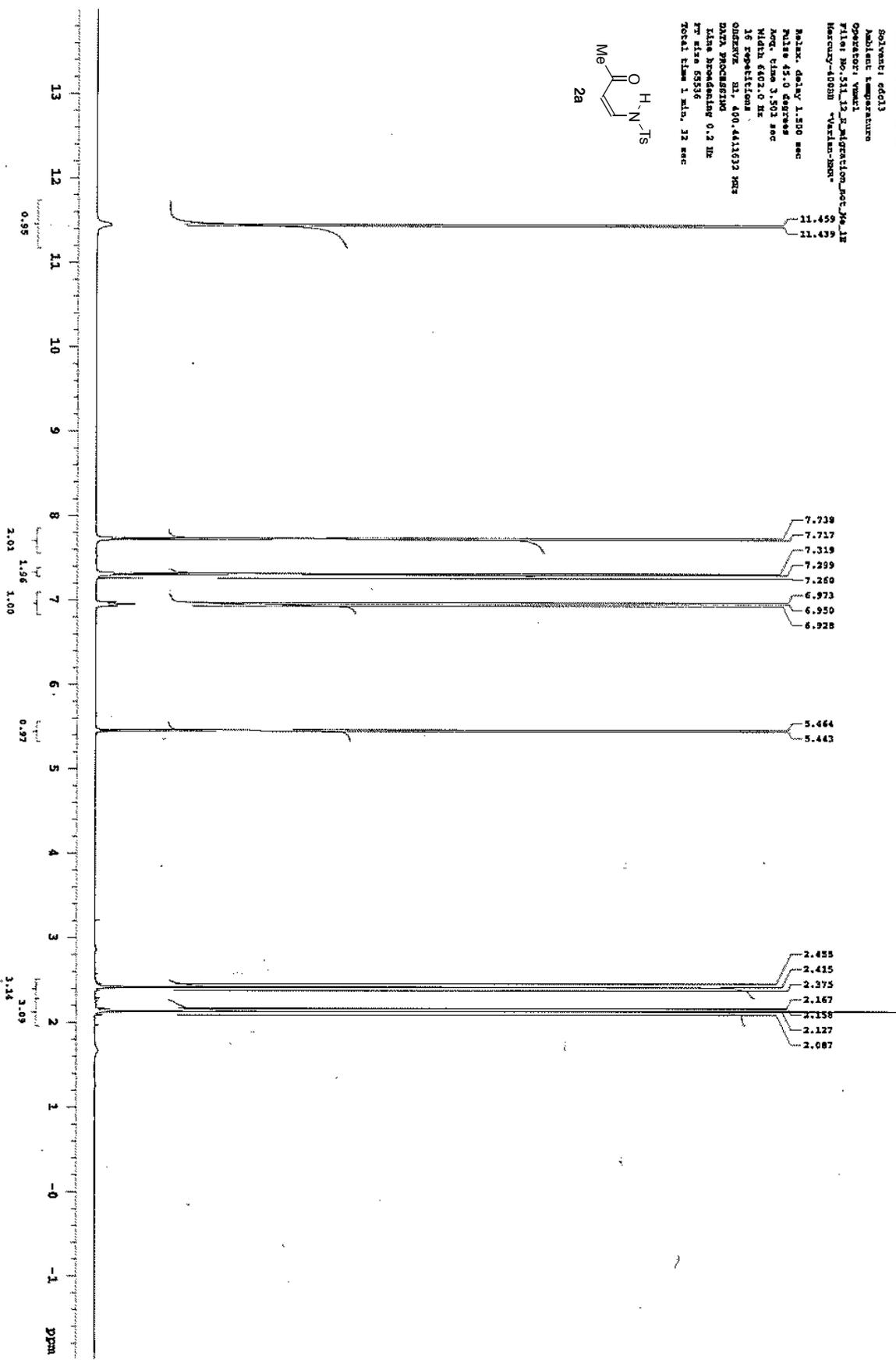
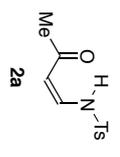


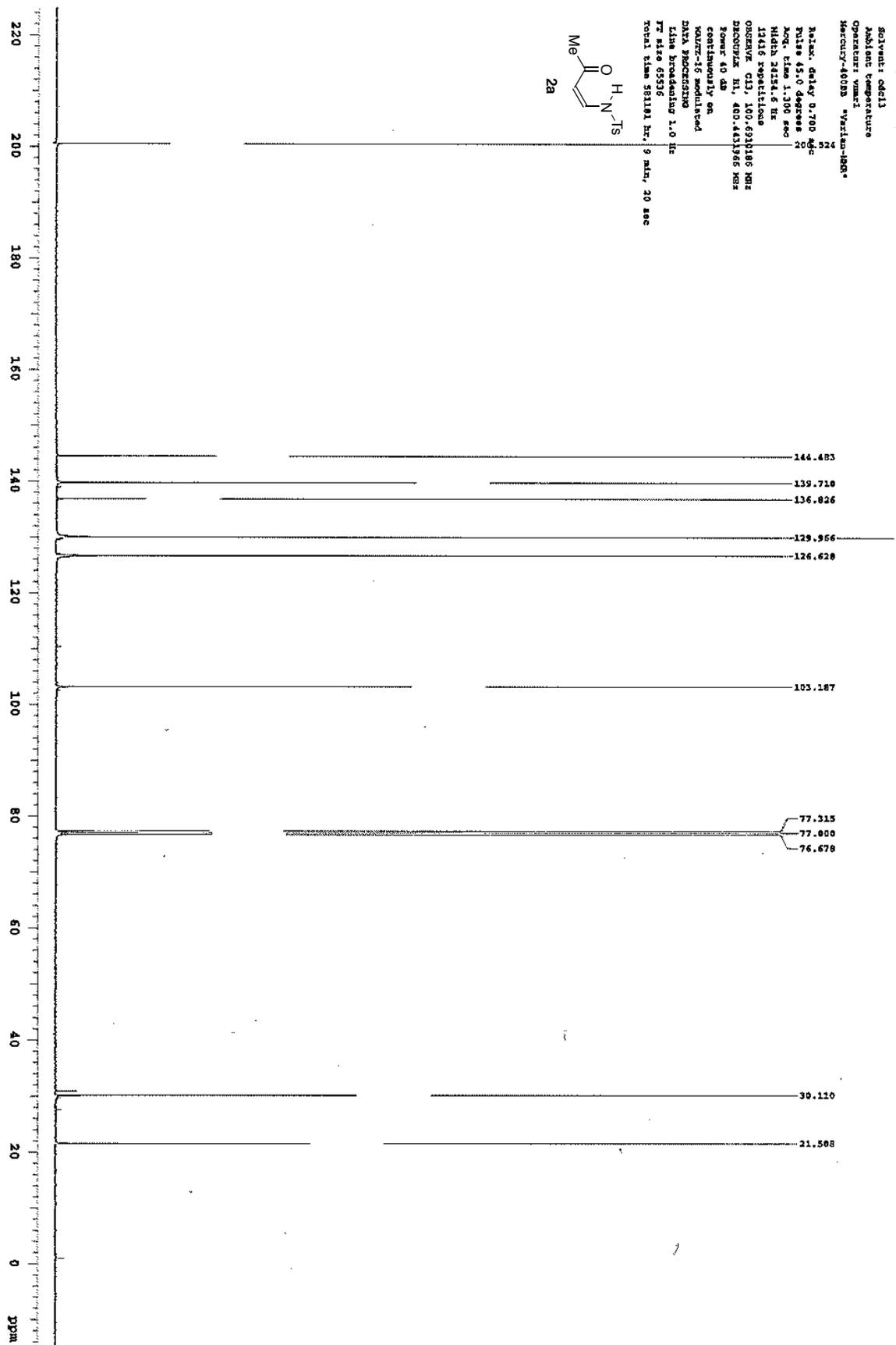
Pulse Sequence: zgpg30
Solvent: cdcl3
Ambient temperature
Operator: ymuel
Nucleus: 13C NMR

Relax. delay: 0.700 sec
Pulse: 45.0 degree
Acq. Elm: 1.300 sec
Width: 24154.6 Hz
109 experiments
OBSERVE: C13, 100.621020 MHz
INSTRUM: ELI, 400.4431956 MHz
Power: 40 dB
continuously on
NUC1: 13C modulated
DATA PROCESSING
Line broadening: 1.0 Hz
FT size: 65536
Total time: 185770 hr, 30 min, -8 sec



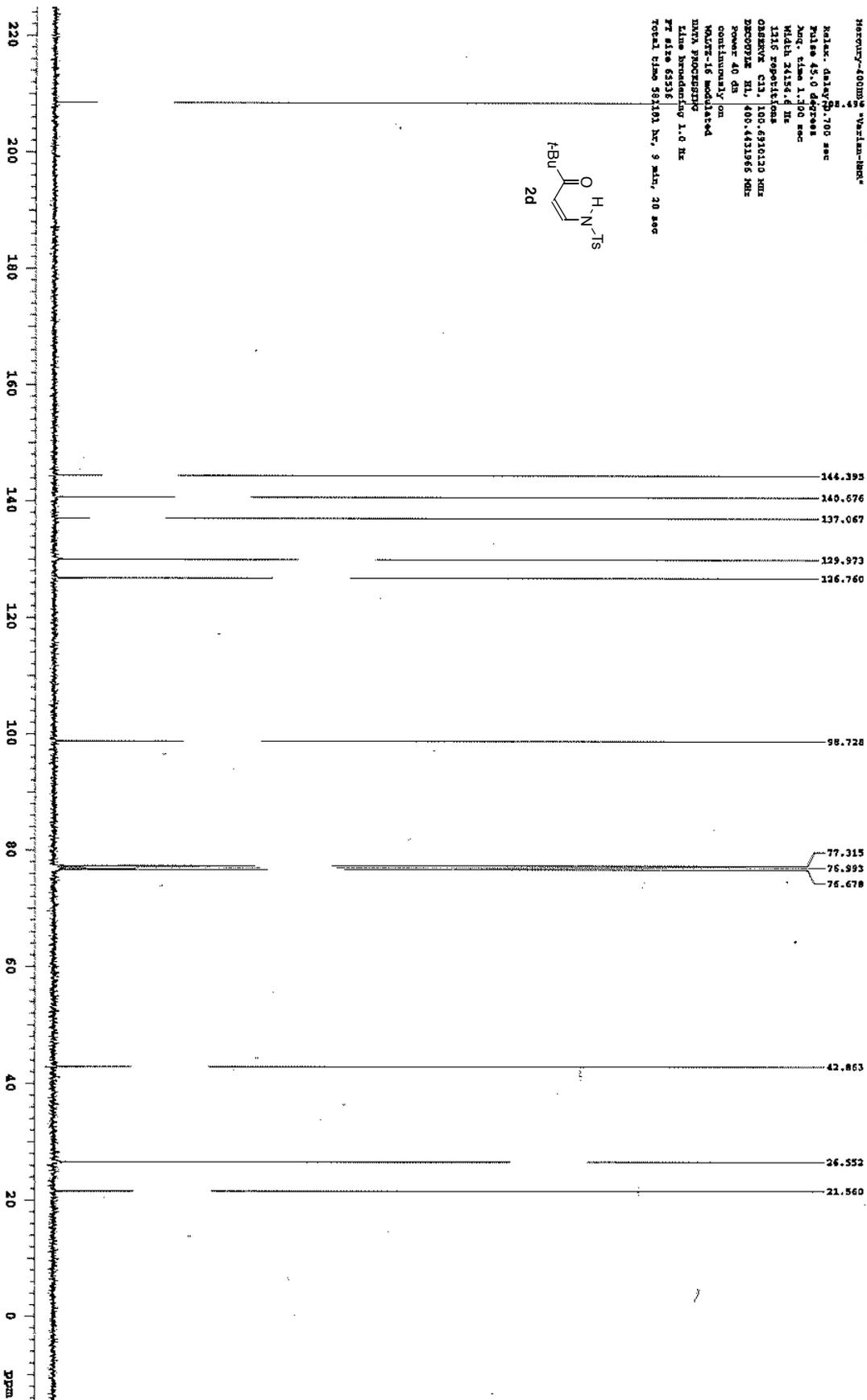
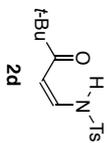
Pulse sequence: zgpg30
 Solvent: dcd13
 Ambient temperature:
 Operator: ymaci
 File: No. 511_13_13_Migration_acq_16_13
 Mercury-400HD "Varian-800"
 Relax. delay: 1.500 sec
 Pulse: 45.0 degrees
 Acq. time: 3.501 sec
 Width: 6402.0 Hz
 16 repetitions
 OMRNAME: 11_400_441613_16Z
 DATA PROCESSING
 Line broadening: 0.2 Hz
 FT file: 65836
 Total time: 1 min, 32 sec



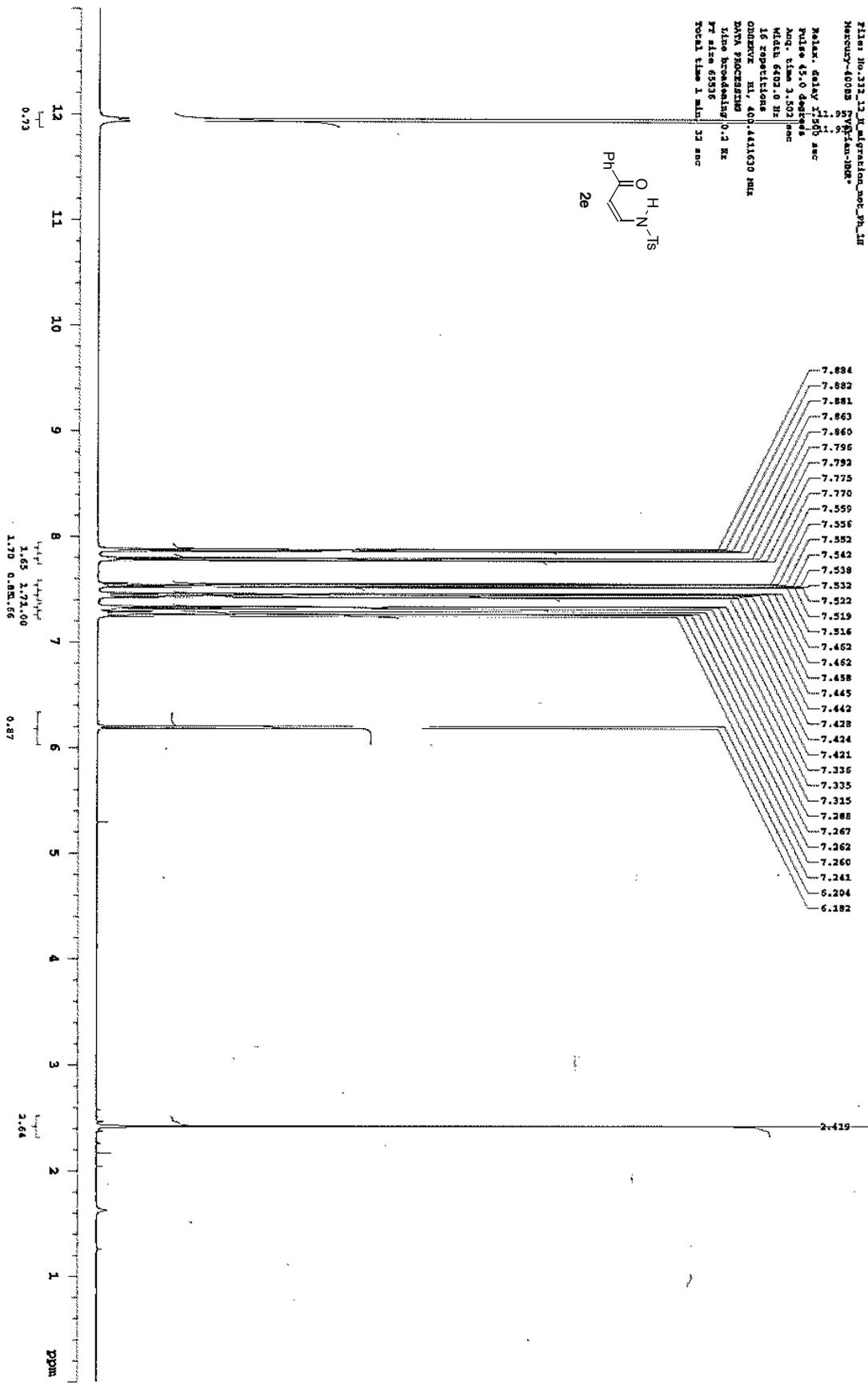
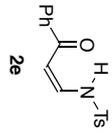


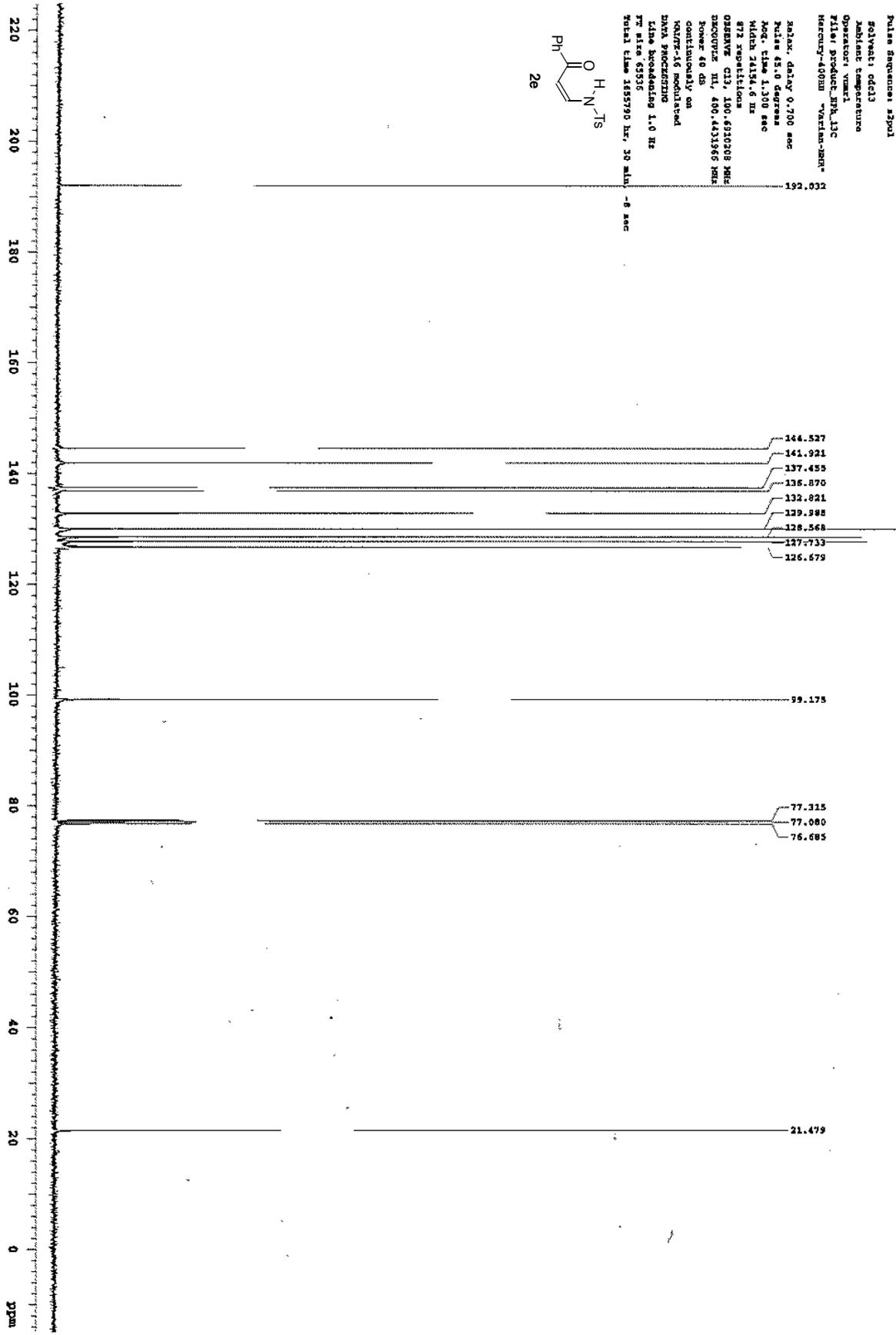
Operator: Yvonne
File: 80-122-13-Substitution_job_cmu_13c
Memory: 400MB, 400MHz, 400MHz

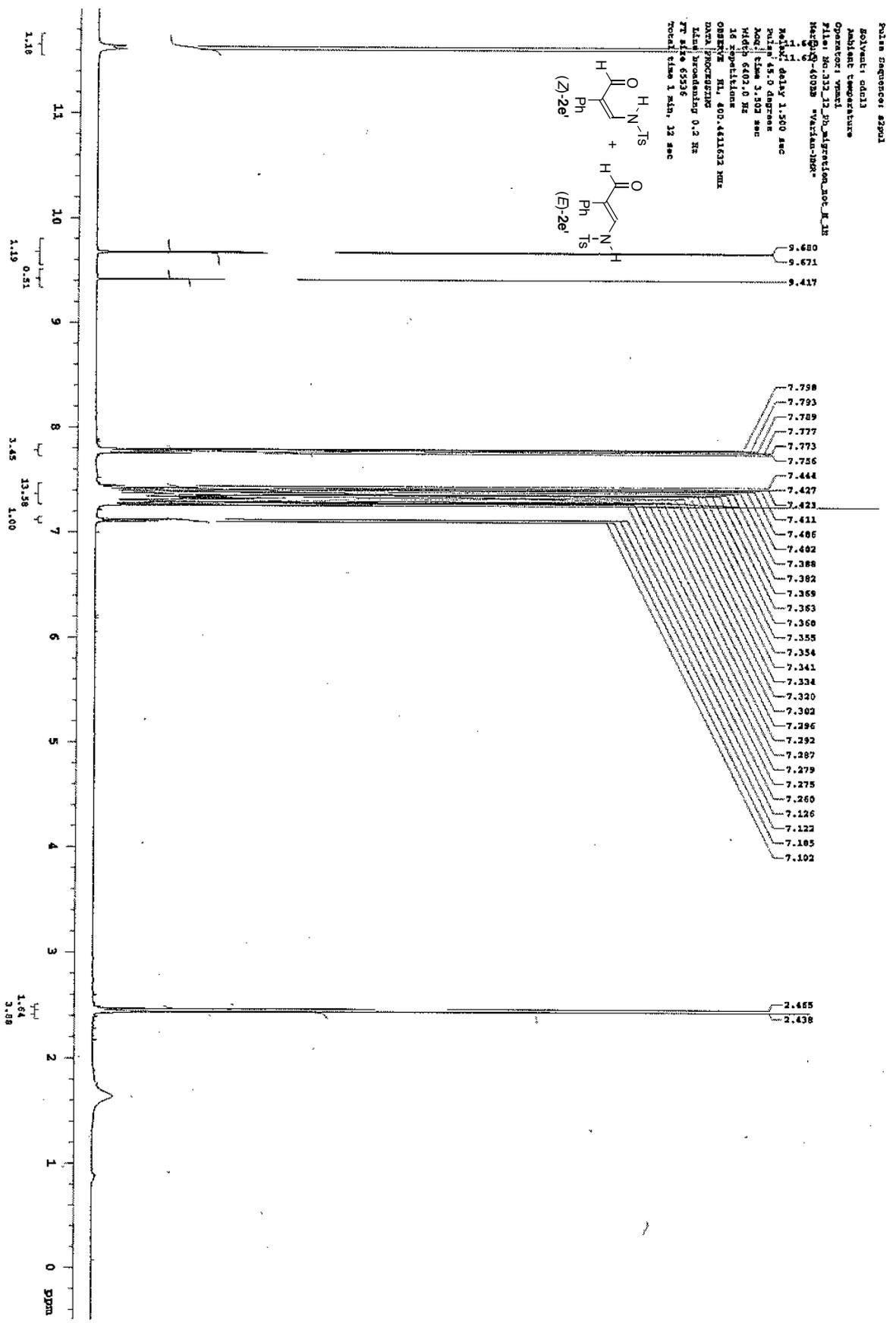
Relax. delay: 7.00 sec
Pulse: 45.0 degrees
Acq. time: 1.100 sec
Width: 24156.6 Hz
13115 repetitions
OBSERVE CH1, 100.6310120 MHz
DECOUPLE CH1, 400.413865 MHz
Power: 40 dB
continuously on
NMR-15 modulated
DATA PROCESSING
Line broadening: 1.0 Hz
TF file: 63236
Total time: 581101 hr, 9 min, 30 sec

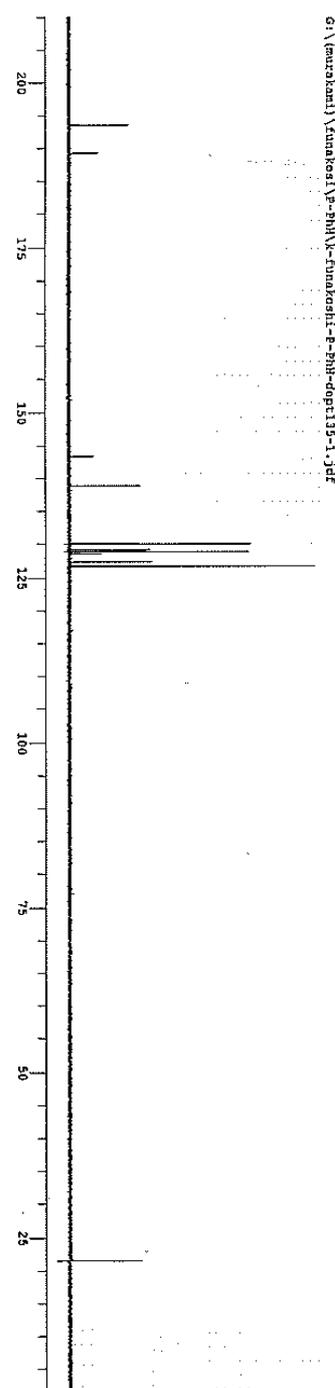
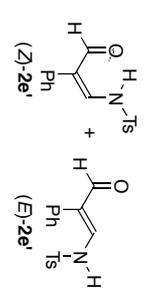
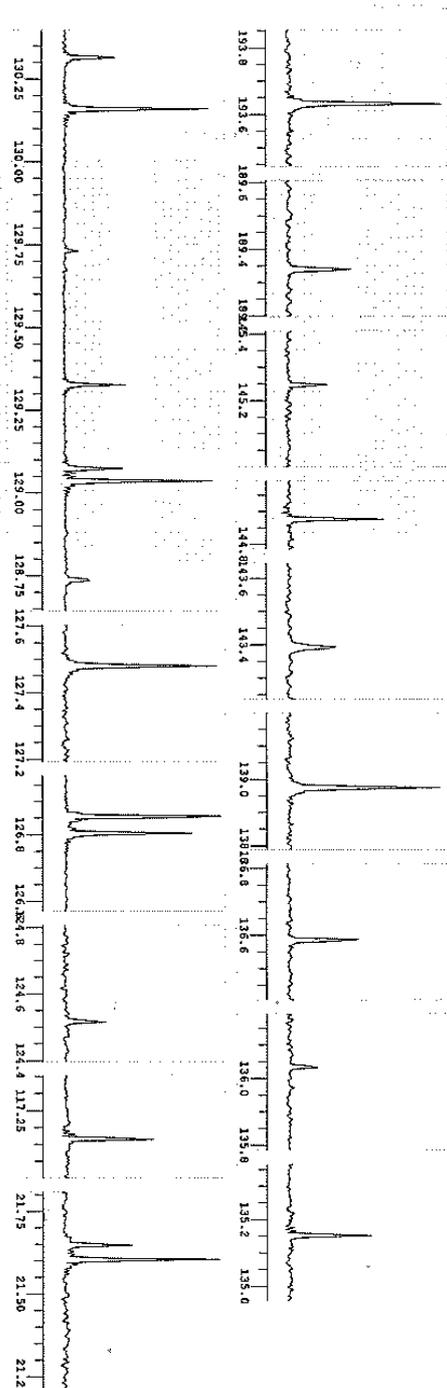


F101: No. 312.12, H. migration, not. Ph. 1H
 Masscu-400MS C₁₃H₁₁N₂O₂S
 Relax. delay 1.500 sec
 Pulse 45.0 degrees
 Acq. time 3.501 sec
 Width 6402.0 Hz
 16 repetitions
 ODMREV XL, 403.441630 MHz
 DATA PROCESSING
 Line broadening 0.2 Hz
 FT size 65536
 Total time 1 min. 33 sec



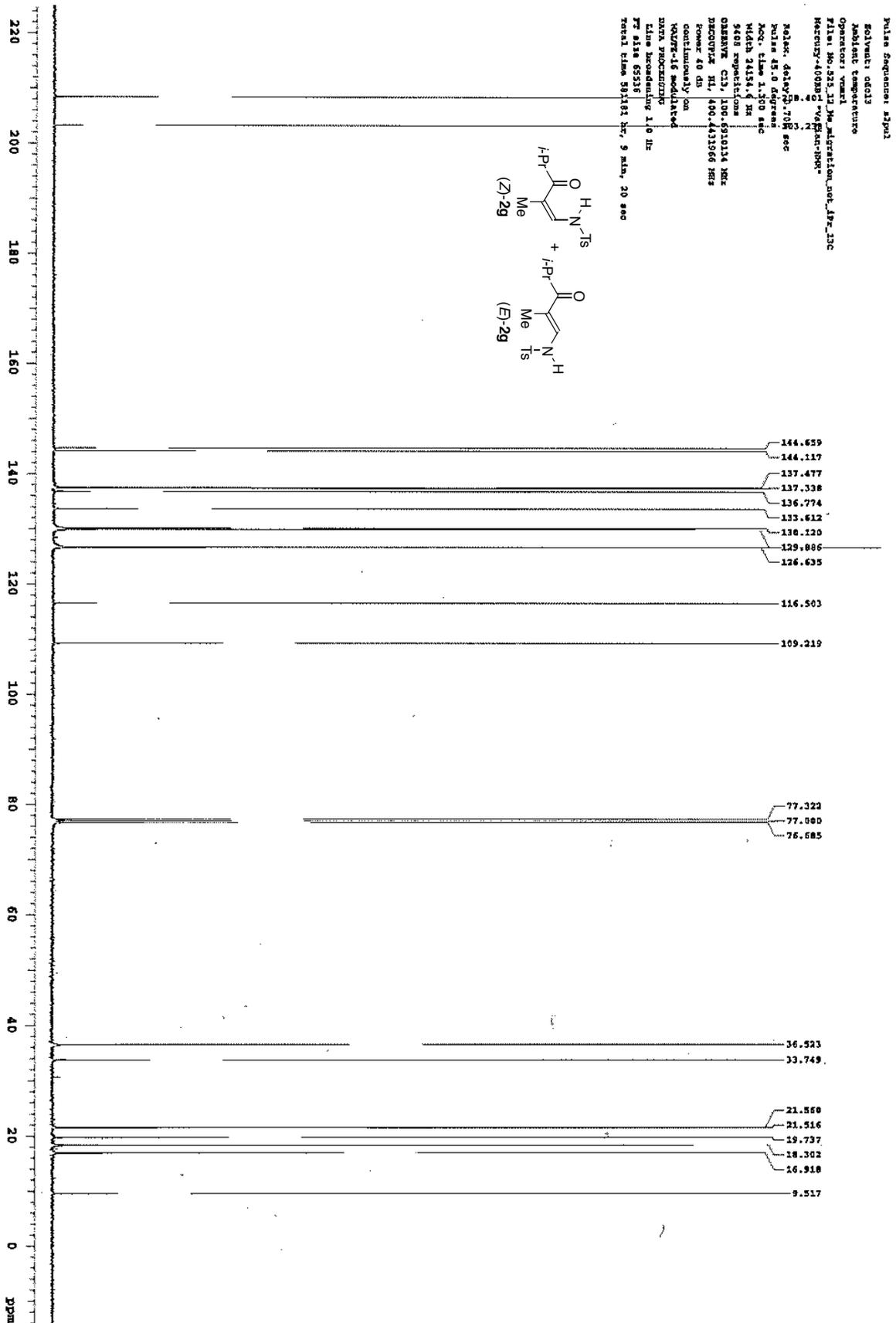


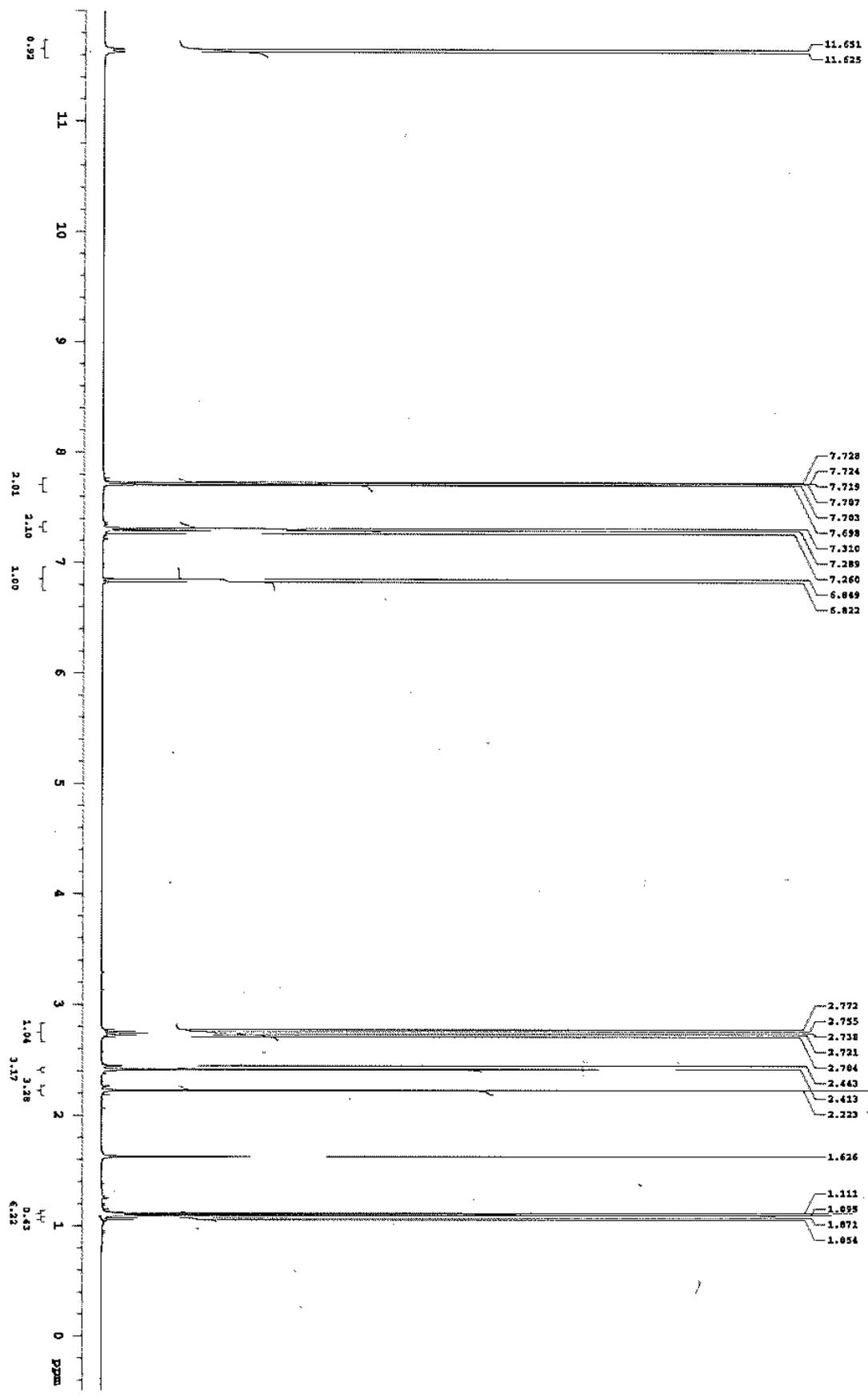
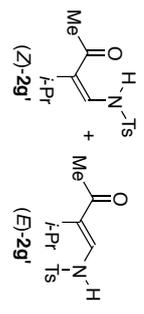


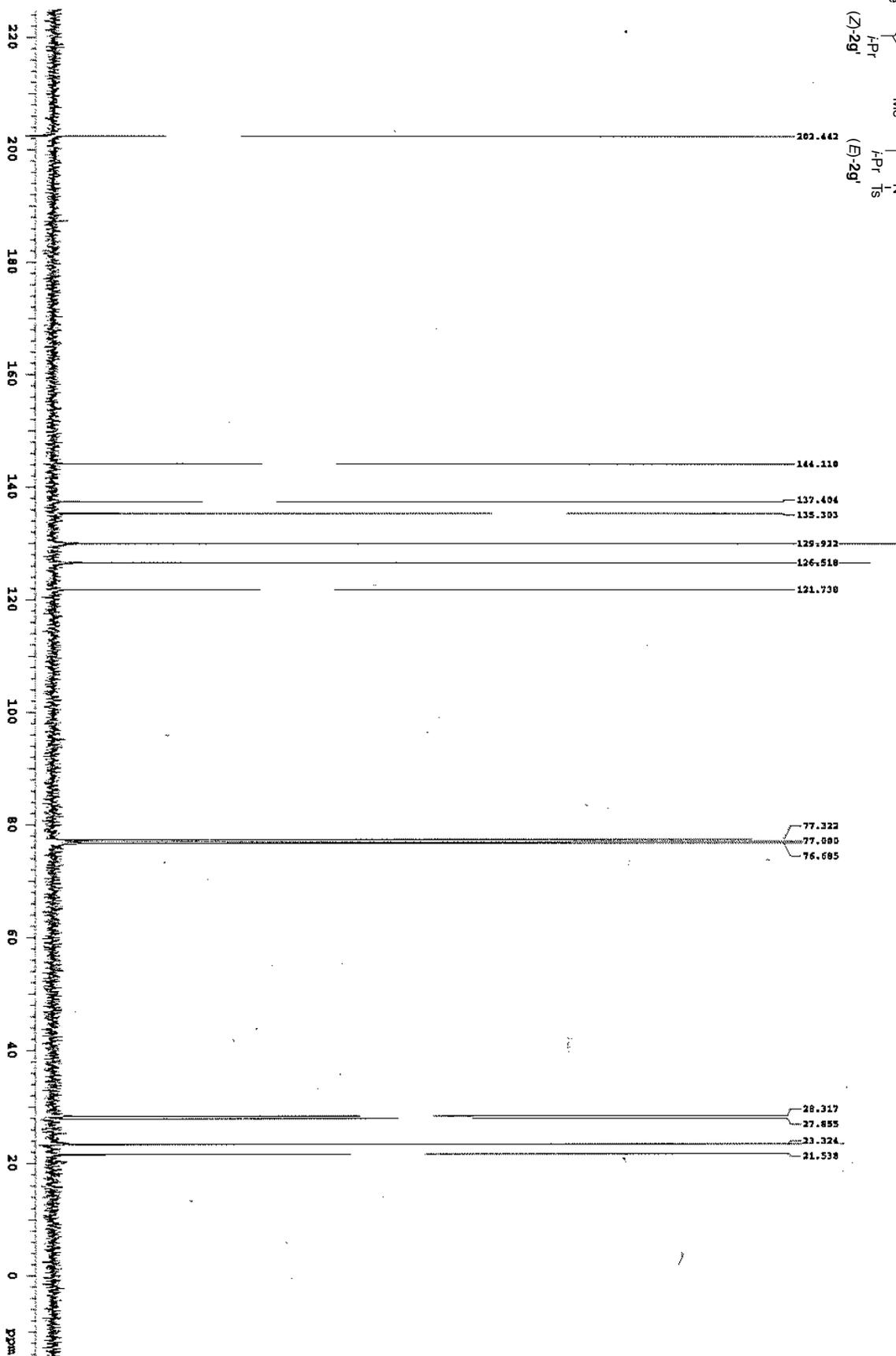
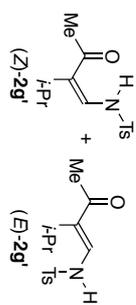


DEPT135 k-funakoshi-p-rnh-dept135-1.jdf
 DEPT135 DEPT135 decoupling
 DATE_ 01-08-2012 10:13:09
 CONTC single pulse decoupled gated NMR
 13C
 EXPOD single pulse dec
 OBPRC 109.53 MHz
 OBF10 5.86 Hz
 POINT0 65536
 FREQ0 31407.03 Hz
 SCANS 526
 ACQTR 1.0433 sec
 PD 1.7000 sec
 2.97 sec
 IANUC 1H
 CTMRP 22.7 c
 SLVTR CDCL3 77.00 PPM
 EXREF BF 0.24 Hz
 ROBIN 60

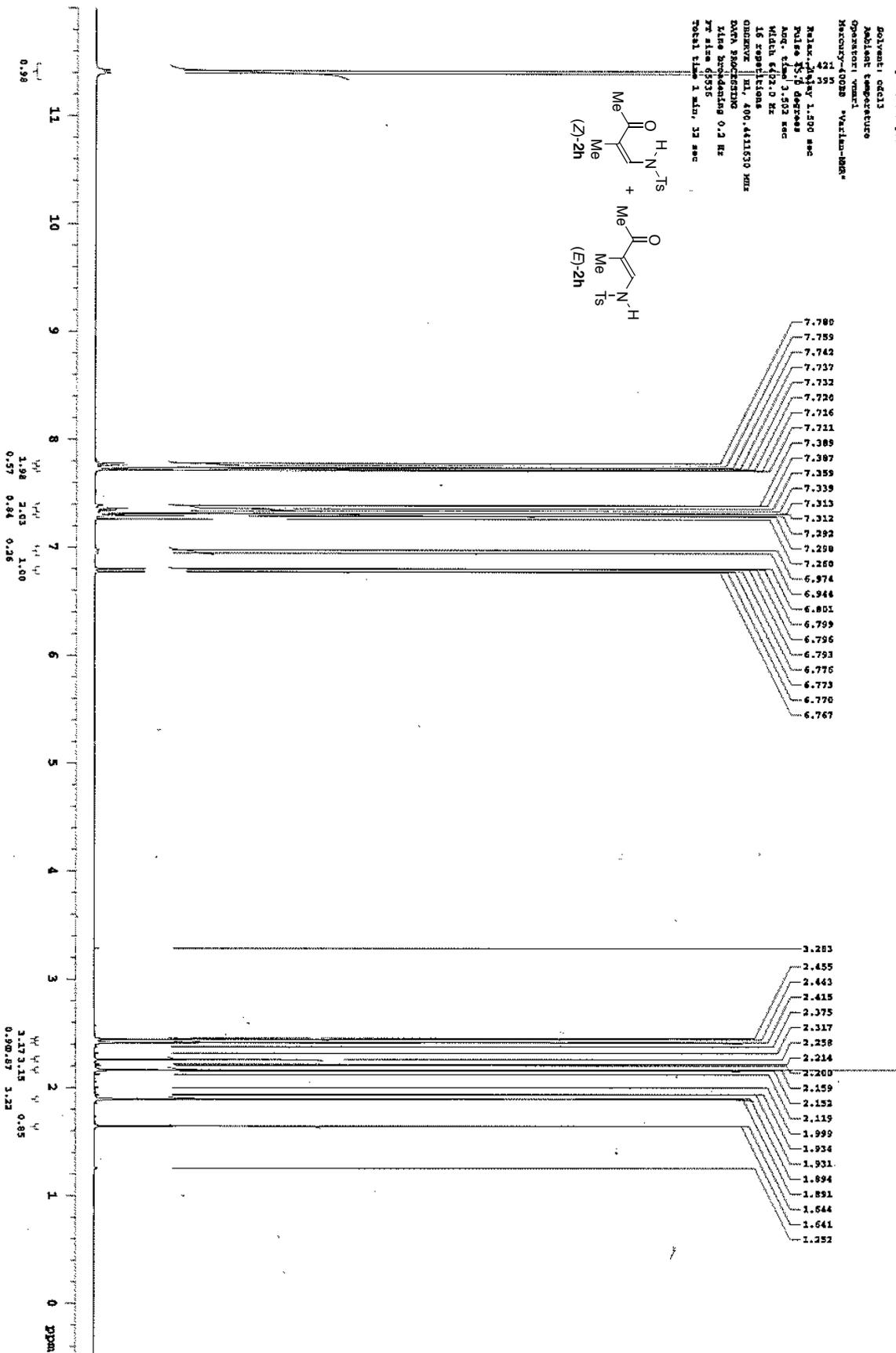
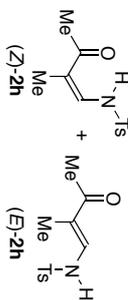
DEPT135 k-funakoshi-p-rnh-dept135-1.jdf
 DEPT135 DEPT135 decoupling
 DATE_ 01-08-2012 10:13:09
 CONTC single pulse decoupled gated NMR
 13C
 EXPOD single pulse dec
 OBPRC 109.53 MHz
 OBF10 5.86 Hz
 POINT0 65536
 FREQ0 31407.03 Hz
 SCANS 526
 ACQTR 1.0433 sec
 PD 1.7000 sec
 2.97 sec
 IANUC 1H
 CTMRP 22.5 c
 SLVTR CDCL3 77.00 PPM
 EXREF BF 0.24 Hz
 ROBIN 60

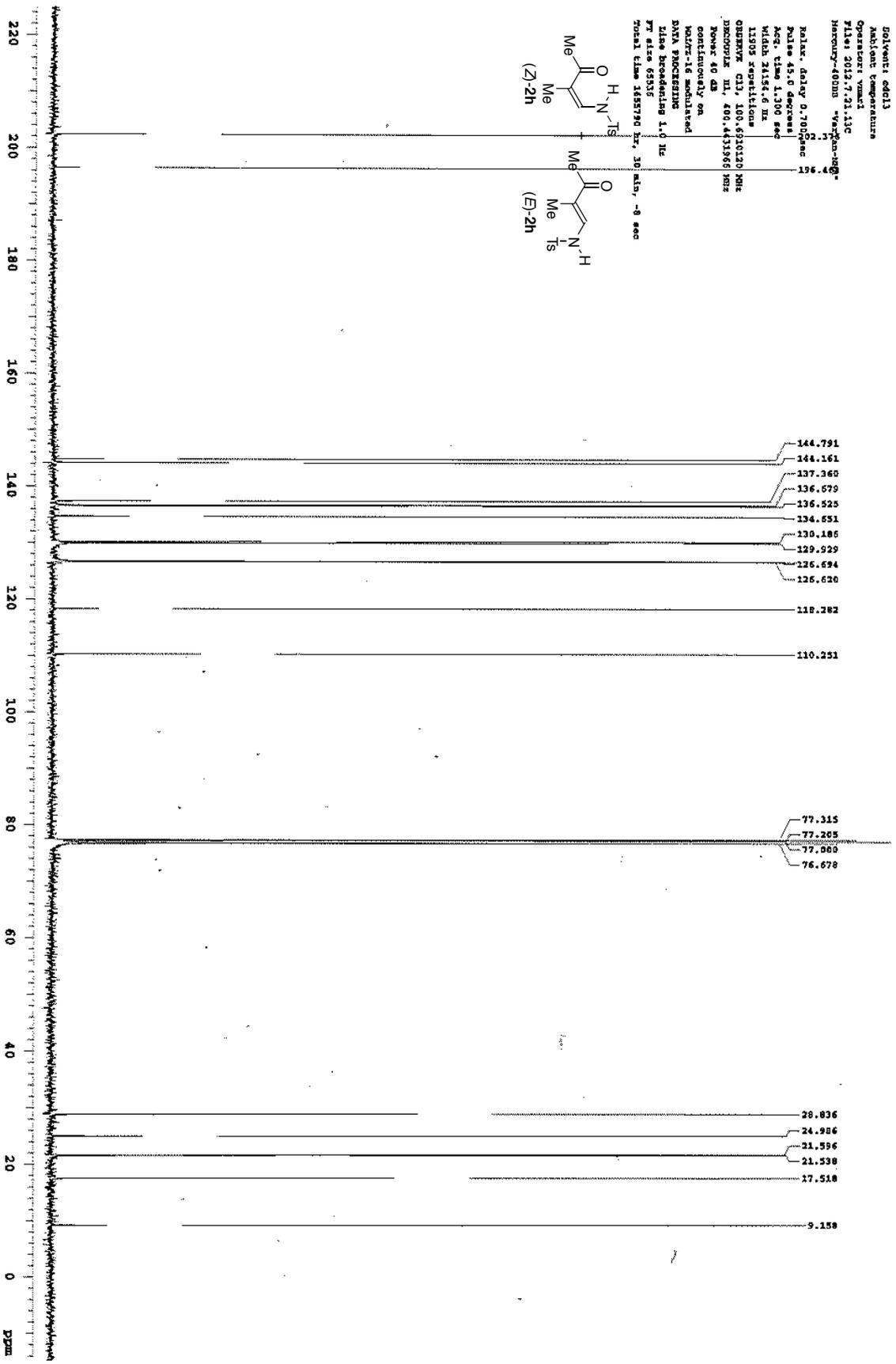






Pulse frequency: 83001
 Solvent: cdcl3
 Ambient temperature
 Operator: vsmc
 Machinery: JNM-PMX 100
 Relax. delay: 1.500 sec
 Pulse: 15.0 degrees
 Acq. time: 1.503 sec
 Width: 6321.0 Hz
 16 repetitions
 GEMREX: HI, 400.441630 MHz
 DMA SPOCKETING
 Line broadening: 0.1 Hz
 XT also 65535
 Total time: 1 min, 33 sec





Pulse Programme: sfp01

Solvent: cdcl3

Operator: vrmx1
File: j01170_01clopopy1a_product_13c_2f1d
Format: 400Mg -Varian-DMR

Pulse: delay 0.700 sec

Pulse 45.0 degrees

Acq. time 1.100 sec

Width 24154.6 Hz

15360 repetitions

ORIGIN: C11, 100.6910137 MHz

PROCYLE III, 400.441966 MHz

Power 40 dB

continuously on

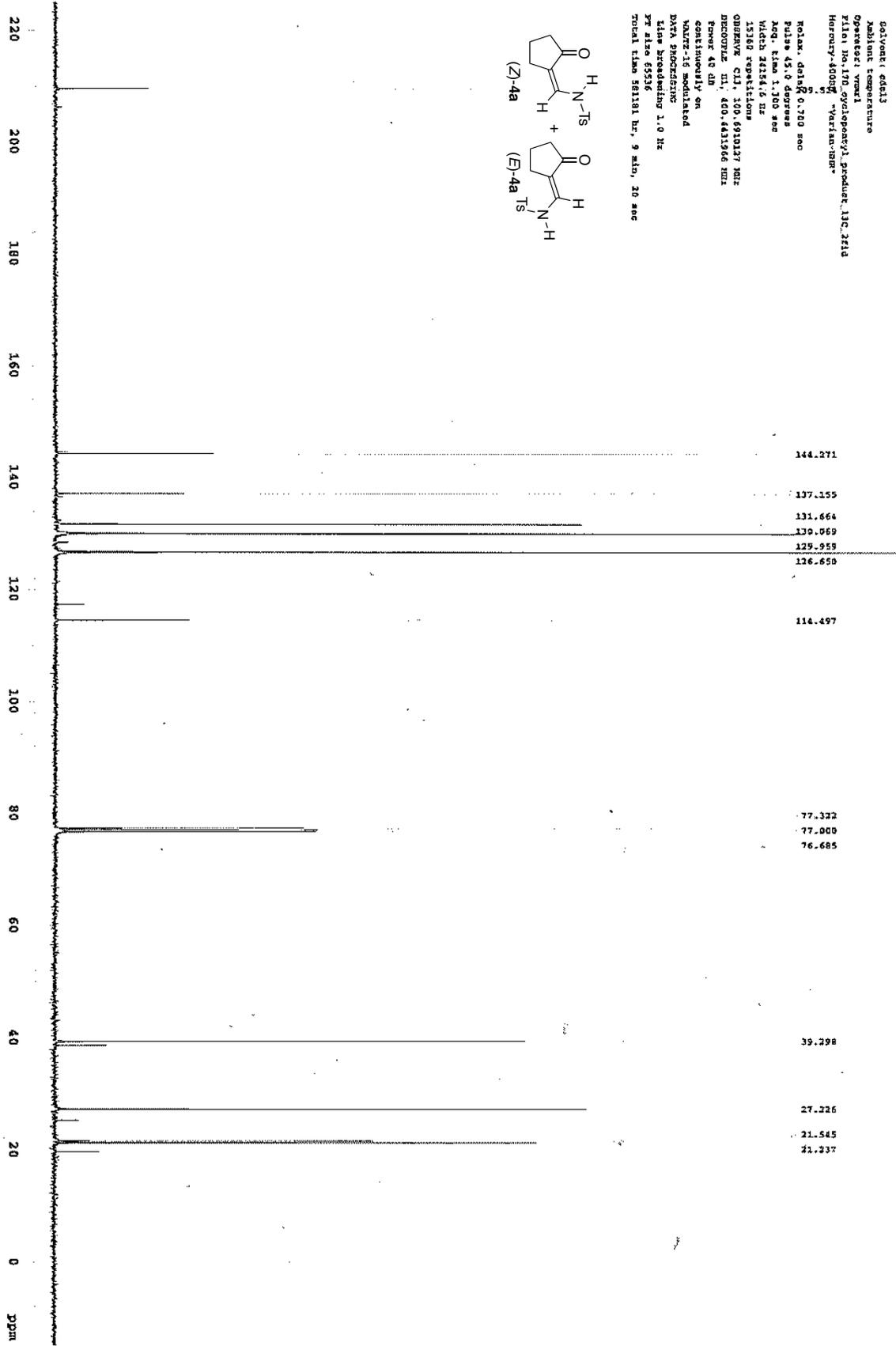
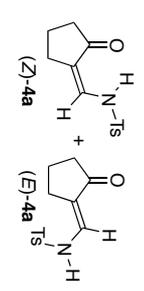
WALTZ-16 modulation

DVPA synchronization

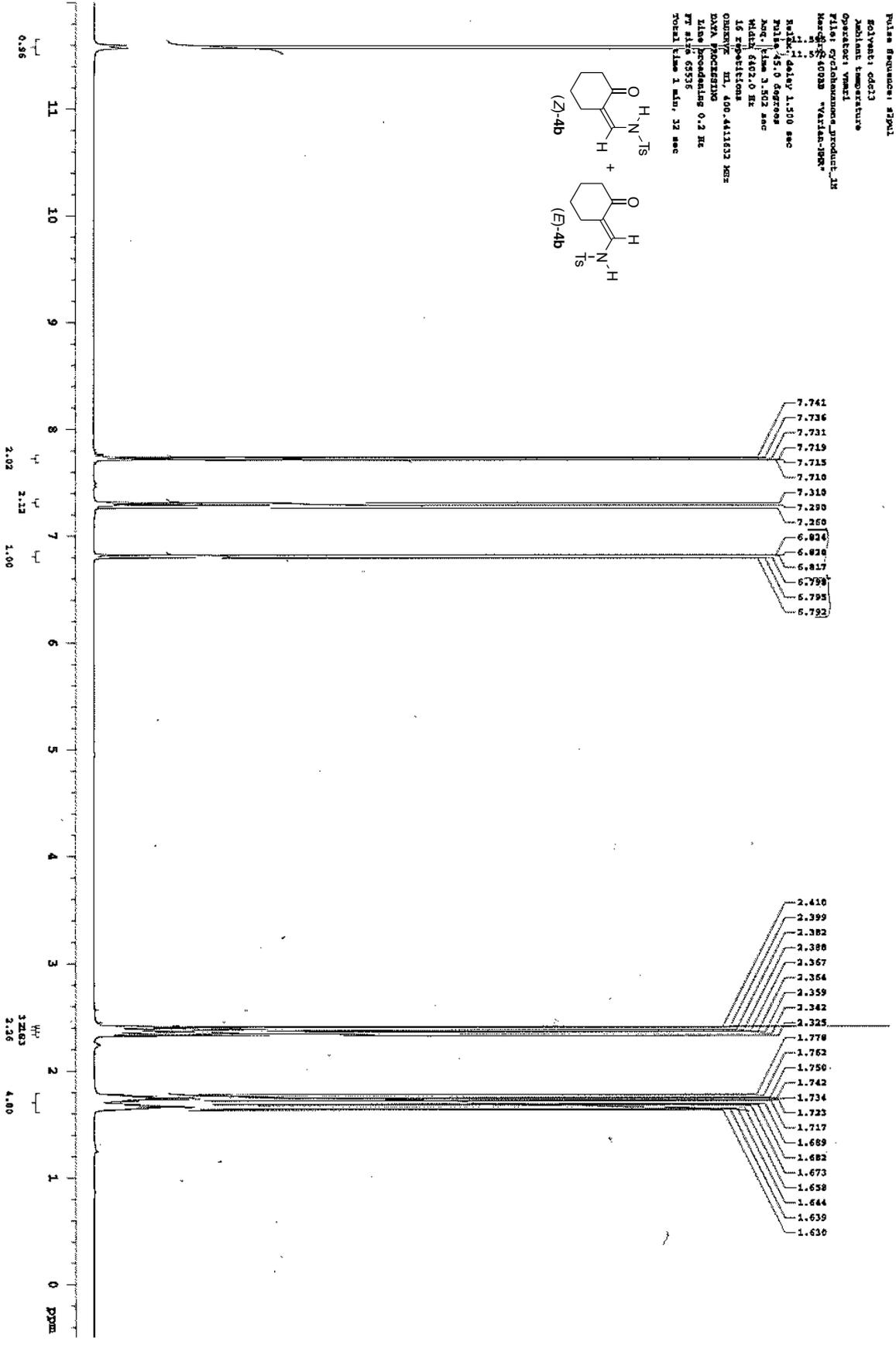
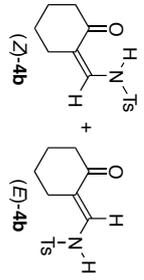
Class broadening 1.0 Hz

PT also 65936

Total time 581581 hr, 9 min, 10 sec



Pulse Sequence: zgpg30
 Solvent: cdcl3
 Solvent temperature
 Operator: ymari
 File: C:\chemstation\data\product_11
 Name: 4-1023 "Varian-100"
 No. 1
 Date: 11/11/91
 Time: 11:11
 Ratio: delay 1.500 sec
 Pulse: 43.0 degrees
 Acq. time 3.502 sec
 Width: 6402.0 Hz
 16 repetitions
 CHANNEL F1, 400.441832 MHz
 DATA PROCESSING
 Line Processing: 0.2 Hz
 FT size: 65536
 Total time: 1 min, 32 sec



Pulse sequence: zgpg30

Solvent: cdcl3

Sample temperature: 300 K

Operator: ymami

Acquisition date: 2008-08-20

File name: 08280802

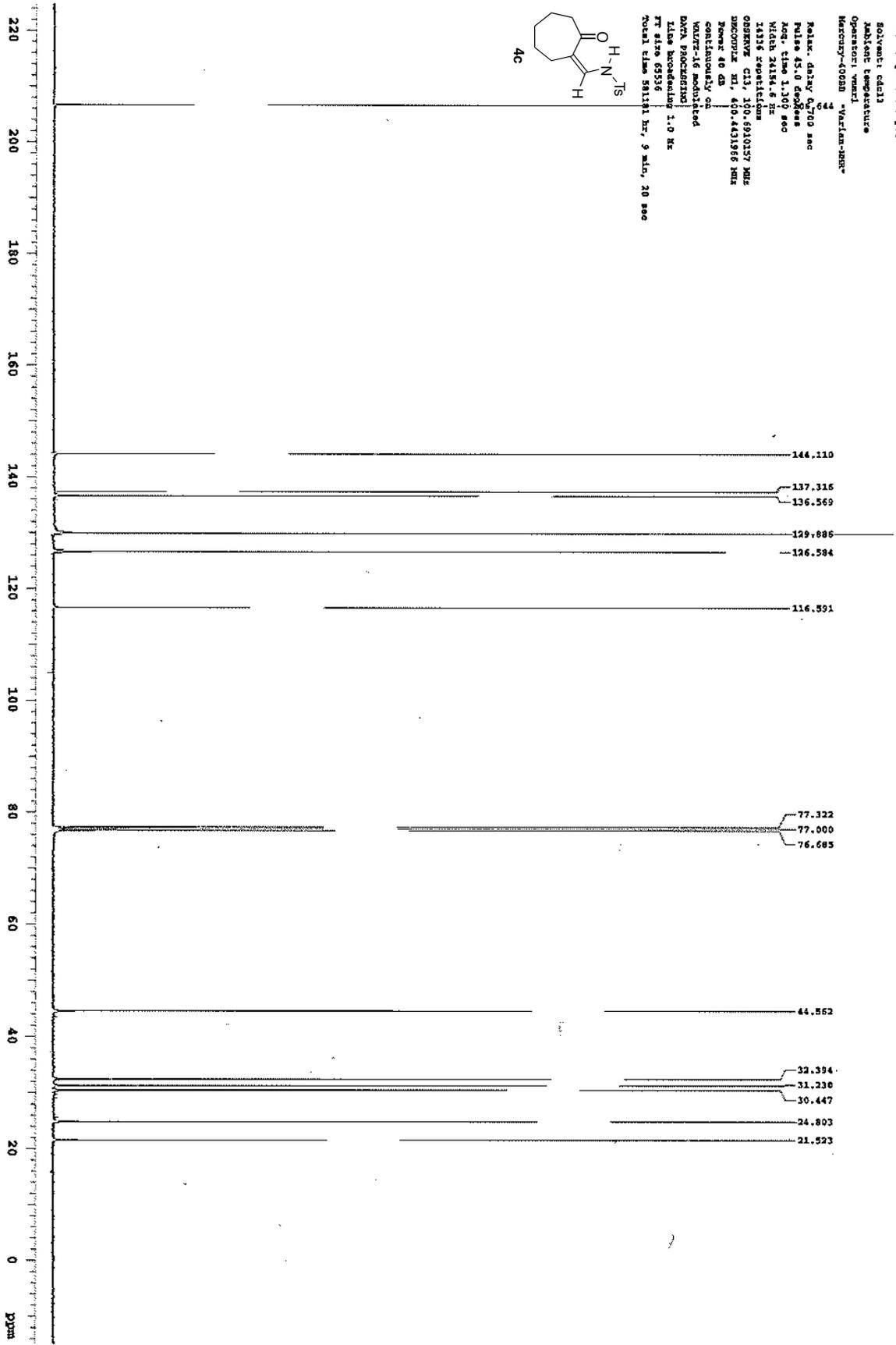
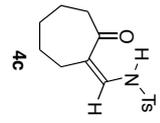
Sample name: 4c

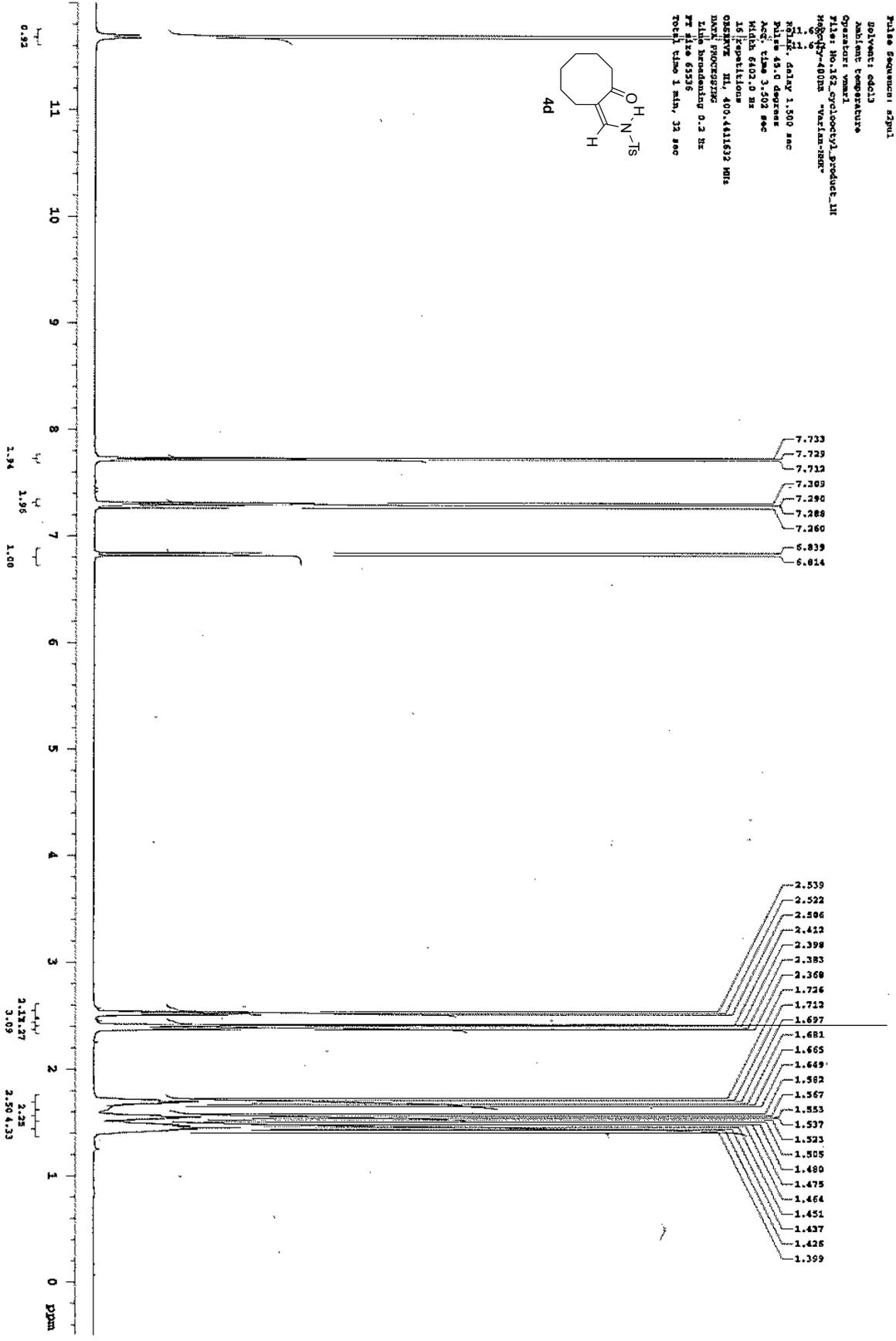
Weight: 2.1845 g

Volume: 10.00 mL

Concentration: 0.21845 g/mL

Label: 13C





Pulse sequence: zgpg30

Solvent: cdcl3

Ambient temperature

Operator: ymwt

Mercury-400DB "Varian-900"

Pulse delay: 1.100 sec

Pulse 45.0 degrees

Acq. time: 1.070 sec

NUC1: 13C13

NUC2: 13C13

NAME: zgpg30

ORIGIN: C:\PROG\DATA\13C

INSTRUM: HPL 400-431966 MHz

PROBHD: 5 mm QNP 1H/13

PROBHD: 5 mm QNP 1H/13

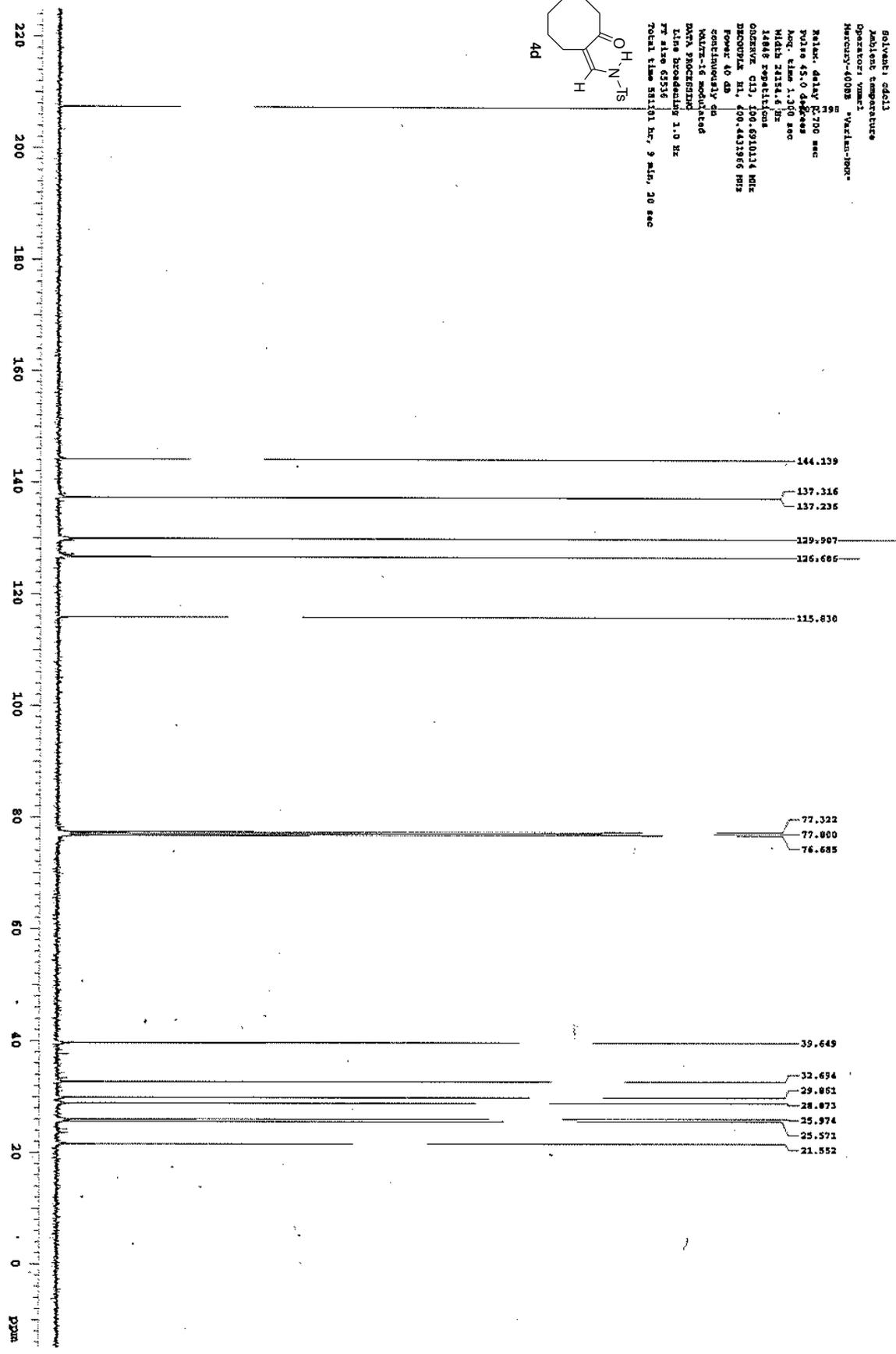
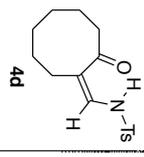
CONTAINER: zgpg30

DATA PROCESSING

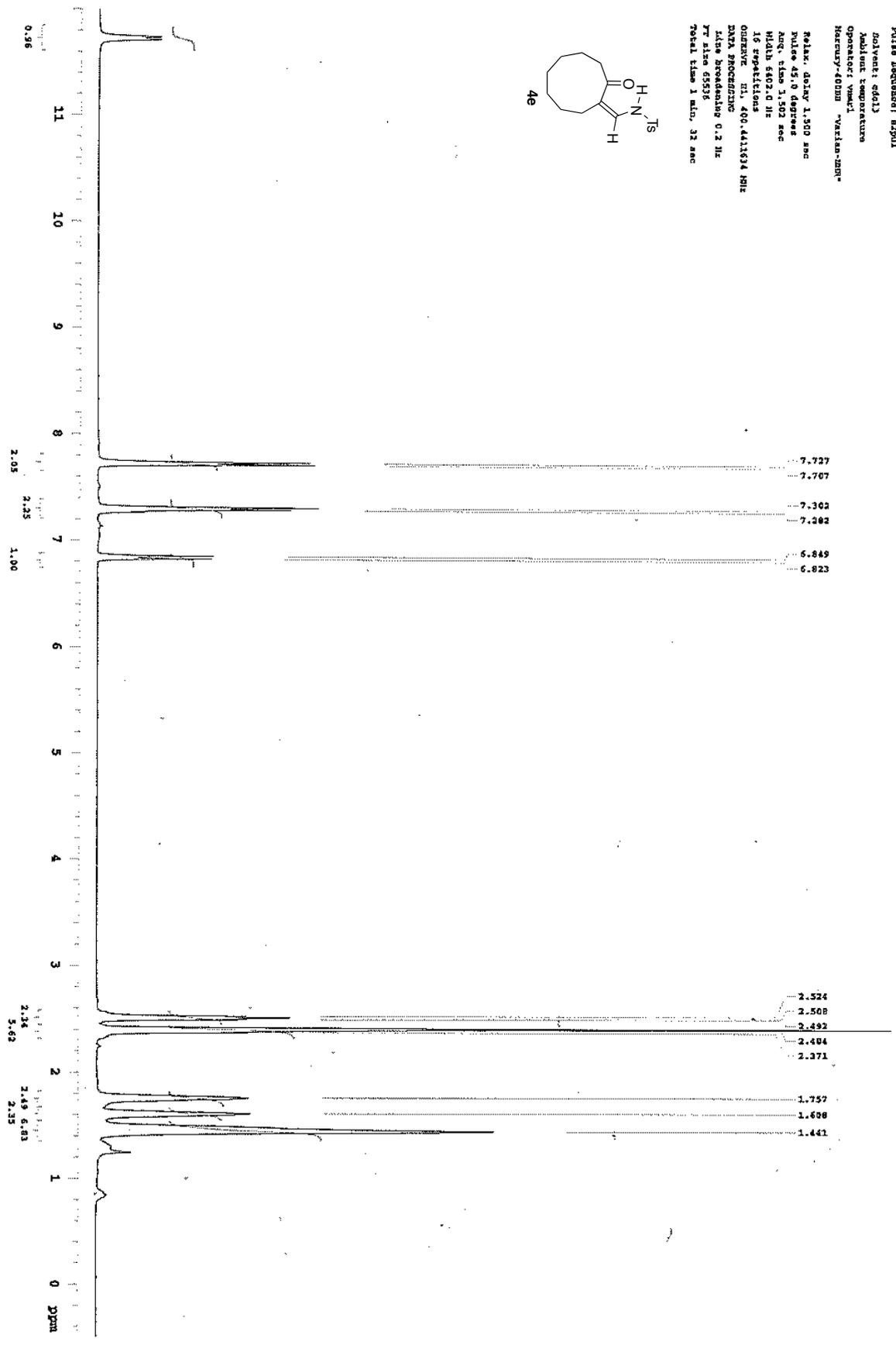
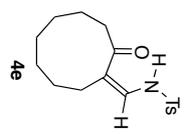
LINE BROADENING 1.0 Hz

PR size 65536

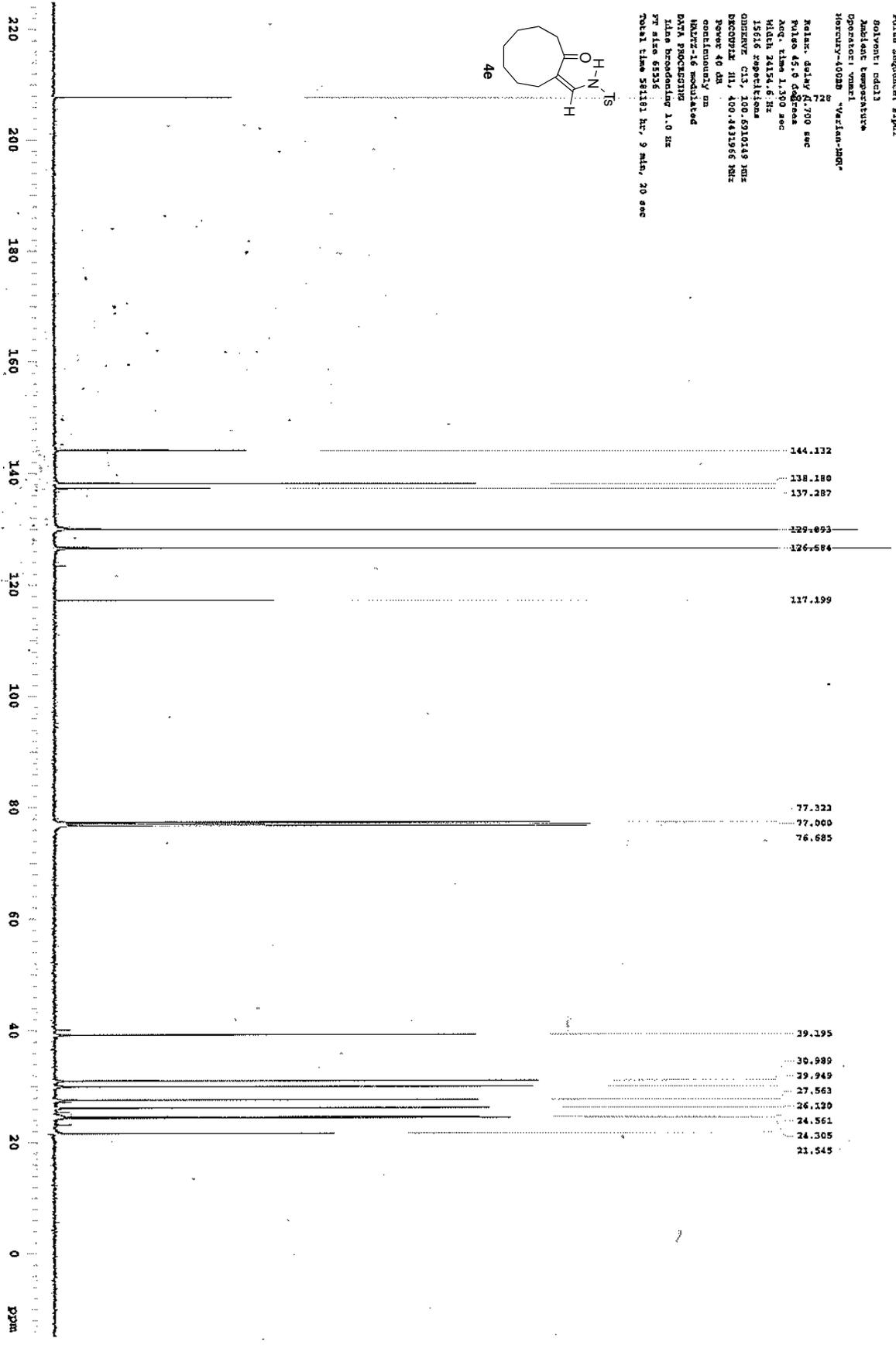
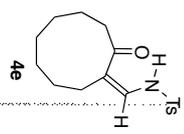
Total time 51:01 hr, 9 min, 20 sec

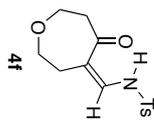


Pulse Program: h2pm1
 Solvent: cdcl3
 Ambient Temperature
 Operator: vma1
 Mercury-400m "Varian-imp"
 Relax delay 1.500 sec
 Pulse 45.0 degrees
 Acq. time 1.502 sec
 MHz 502.0 Hz
 16 repetitions
 QUREX: H1, 400.441834 MHz
 DATA PROCESSING
 Line broadening 0.2 Hz
 FT scan 65536
 Total time 1 min, 32 sec

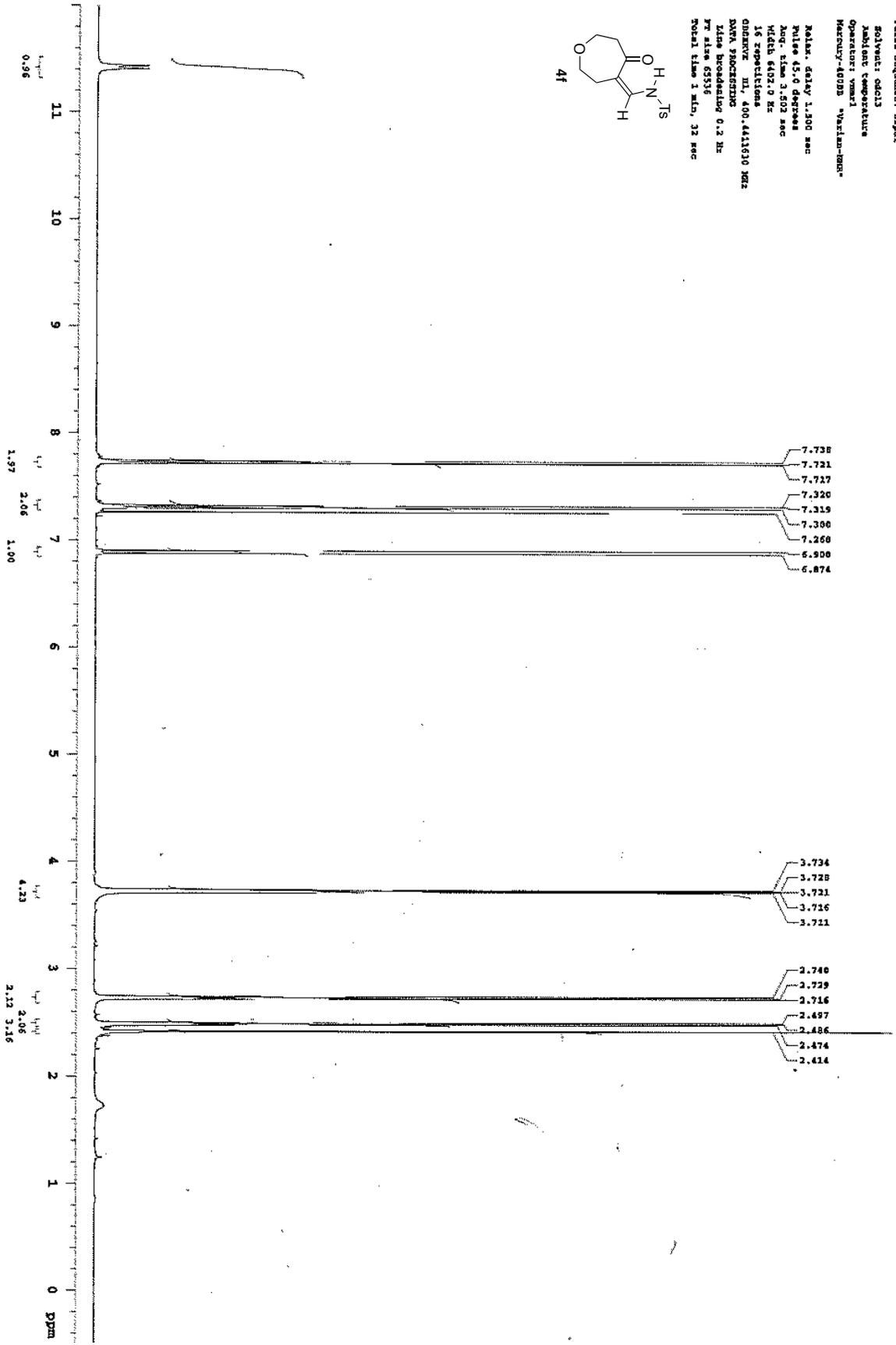


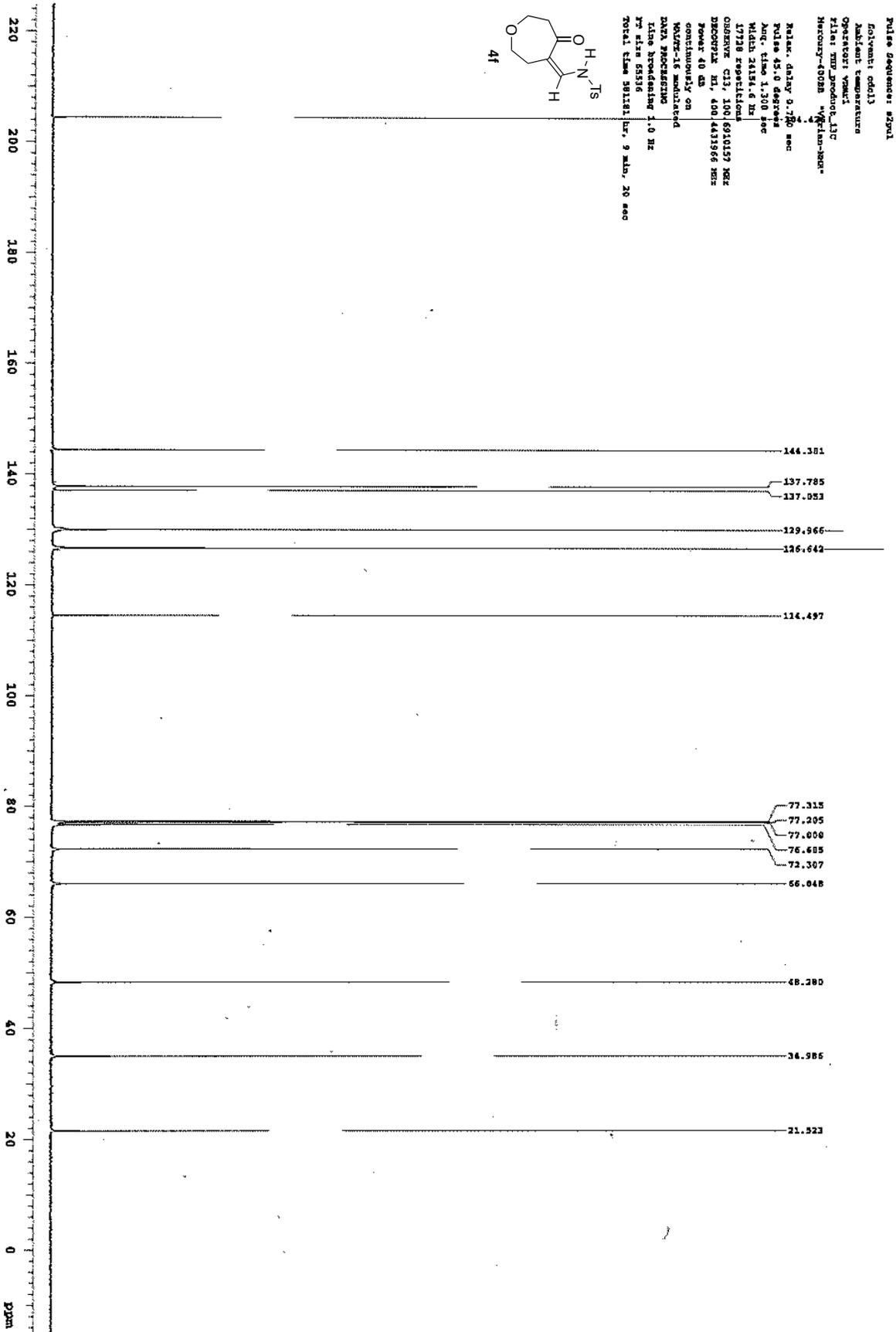
Pulse Sequence: zgpg30
 Solvent: cdcl3
 Ambient Temperature
 Operator: ymami Varian-200
 Frequency: 100.625 MHz
 Nucleus: 13C
 Relax. delay: 6.700 sec
 Pulse: 45.0 degrees
 Acq. time: 1.350 sec
 Width: 24154.6 Hz
 15616 repetitions
 OBSERVE: C13, 100.6310149 MHz
 DECOUPLE: H1, 100.431965 MHz
 Power: 40 dB
 continuously on
 INSTR: 16-modulated
 DATA PROCESSOR
 Data Processing: 1.0 Hz
 FT date: 851316
 Total time: 301.81 hr, 9 min, 20 sec

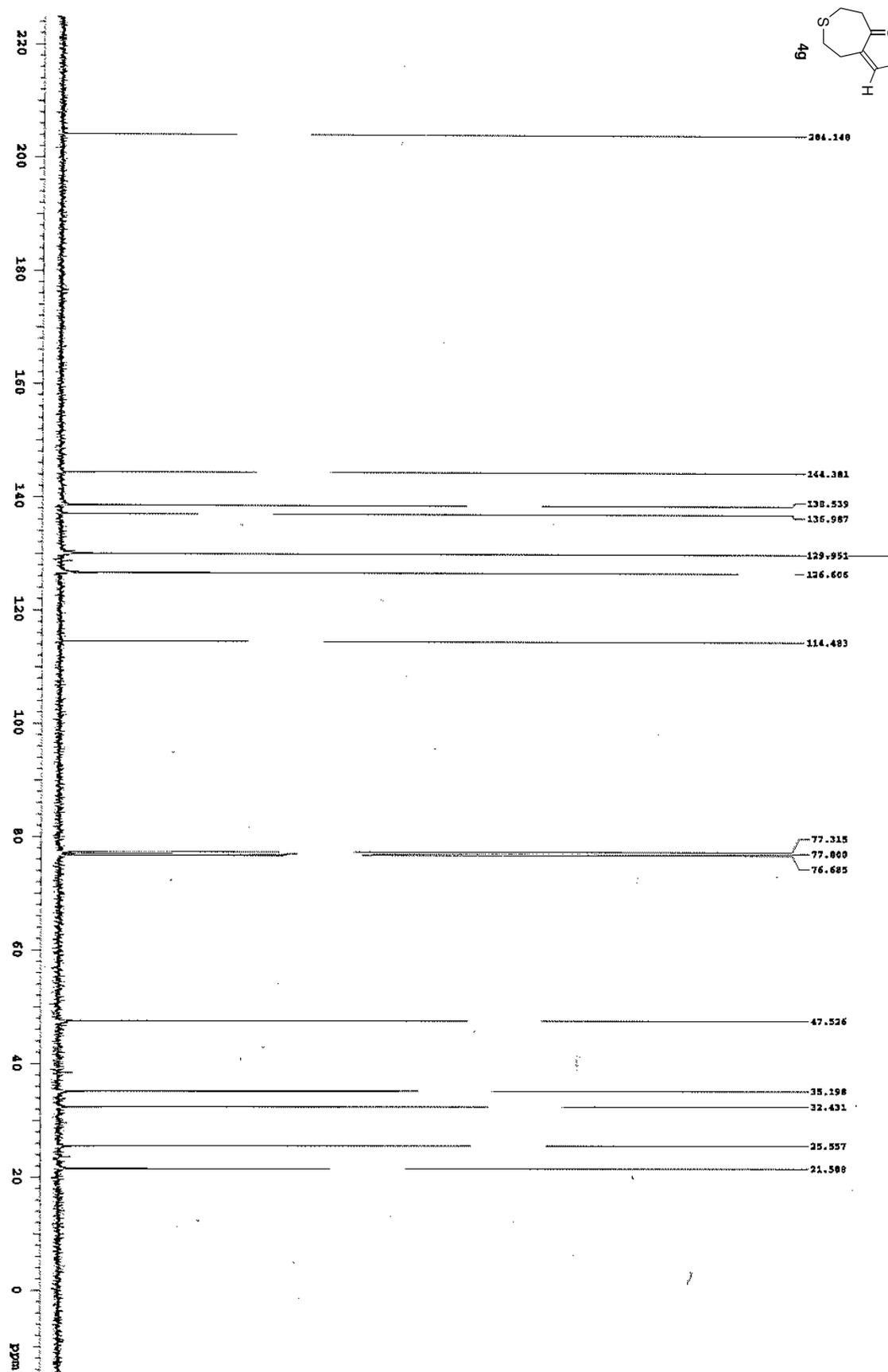
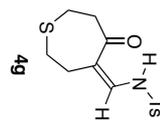




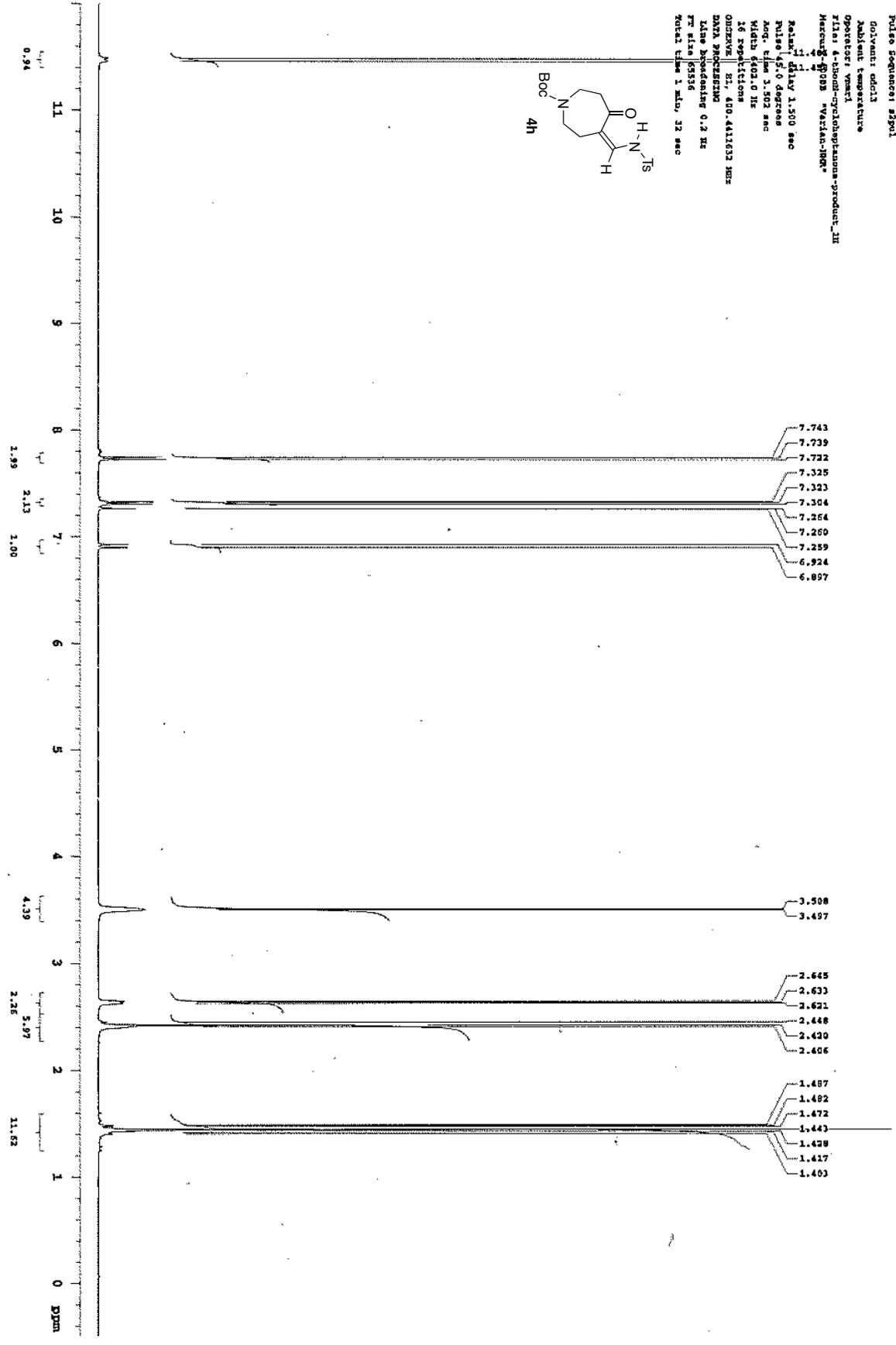
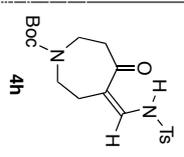
Pulse sequence: zgpg1
 Solvent: cdcl3
 Ambient temperature
 Operator: ymas1
 Mercury-400DB "Varian-800"
 Relax. delay: 1.500 sec
 Pulse: 45.0 degrees
 Acq. time: 3.502 sec
 Width: 6402.0 Hz
 16 repetitions
 Channel: H1, 400.441610 MHz
 DATA PROCESSING
 Line broadening: 0.2 Hz
 FT size: 65536
 Total time: 1 min, 32 sec

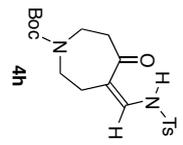
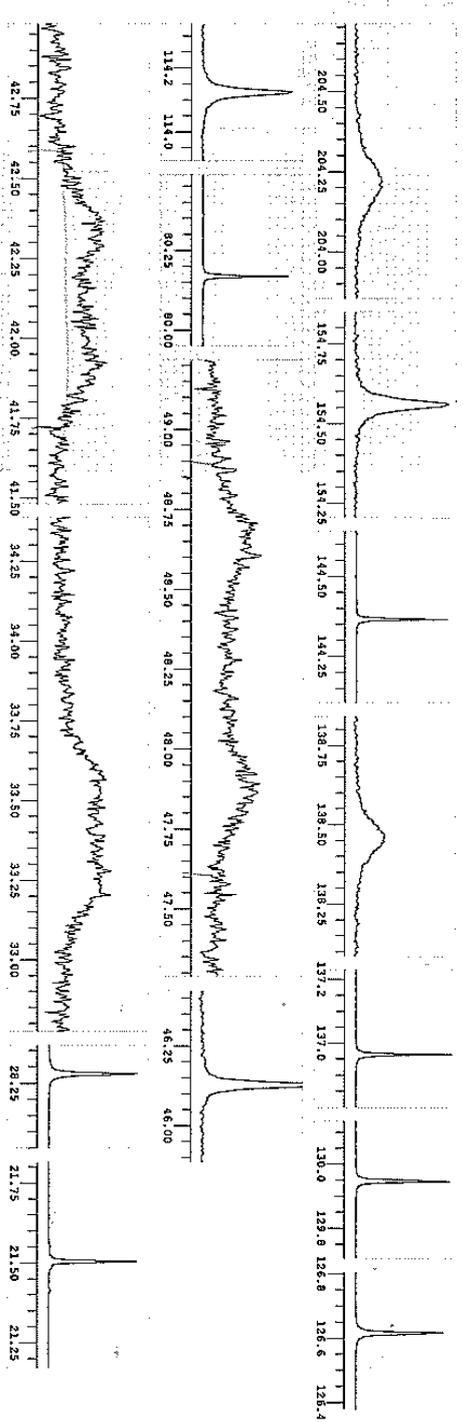




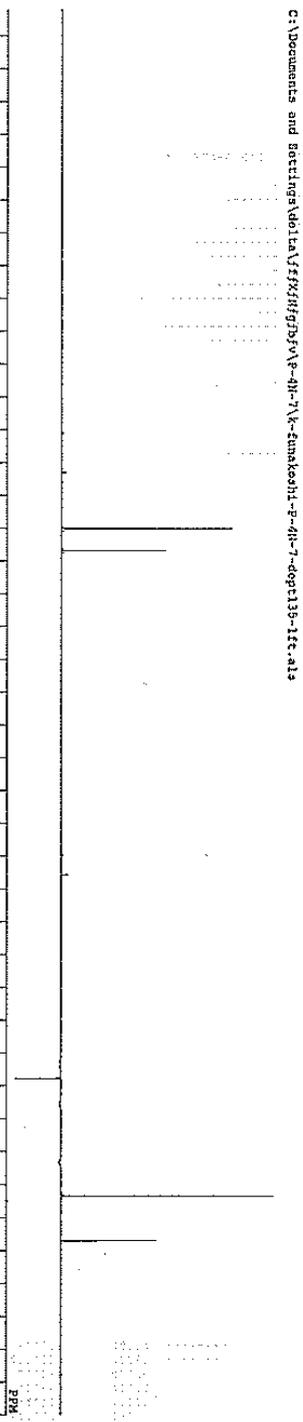


Pulse Sequence: zgpg31
 Solvent: cdcl3
 Subst: temp: rt
 Operator: smat
 Title: 4-(tert-butyl)-piperidin-2-one-2-ylidene-1H-imidazole-5-carboxamide
 Name: zgpg31
 Name: VASIA-MNR
 Relax: delay 1.500 sec
 Pulse: 9.0 degree
 Acq: time 3.502 sec
 Width: 6400.0 Hz
 16 repetitions
 CHANNEL: SI, 400.441633 MHz
 DATA PROCESSING
 Lame: boosting 0.3 Hz
 FT: file: 05336
 Total time: 1 min, 32 sec





930874



DEPT135 k-funakoeh1-p-4h-7-dept135-1tc.a
 DEPT135 DEPT135 decoupling
 DATE_ 19-07-2012 08:13:23
 EXPROD dept-ox2
 OBPRQ 100.53 MHz
 OBPTH 5.86 Hz
 POINT 52428
 FREQD 22123.56 Hz
 SCANS 1200
 SCAH 2.1200 sec
 DS 21.0028 sec
 PD 8.00 usec
 INRG 1H
 CTRF 22.6 c
 SINT CDCL3 77.00 ppm
 EXRF 0.21 Hz
 ROIN 60

DEPT135 k-funakoeh1-p-4h-7-ine-1tc.a
 DEPT135 DEPT135 decoupled gated HO
 DATE_ 19-07-2012 06:50:48
 EXPROD sing1g pulg.dac
 OBPRQ 100.53 MHz
 OBPTH 5.86 Hz
 POINT 52428
 FREQD 22123.56 Hz
 SCANS 6800
 SCAH 2.3698 sec
 DS 5.1000 sec
 PD 2.67 usec
 INRG 1H
 CTRF 22.4 c
 SINT CDCL3 77.00 ppm
 EXRF 0.21 Hz
 ROIN 60

Pulse sequence: zgpg30

Solvent: d6d13

Acquire temperature

Operator: vsm1

P1: 0.166 sec

Acq: 400 MHz

Mar: 400 MHz

Relax. delay: 1.500 sec

Pulse: 45.0 degrees

Acq. time: 1.502 sec

Width: 6402.0 Hz

16 repetitions

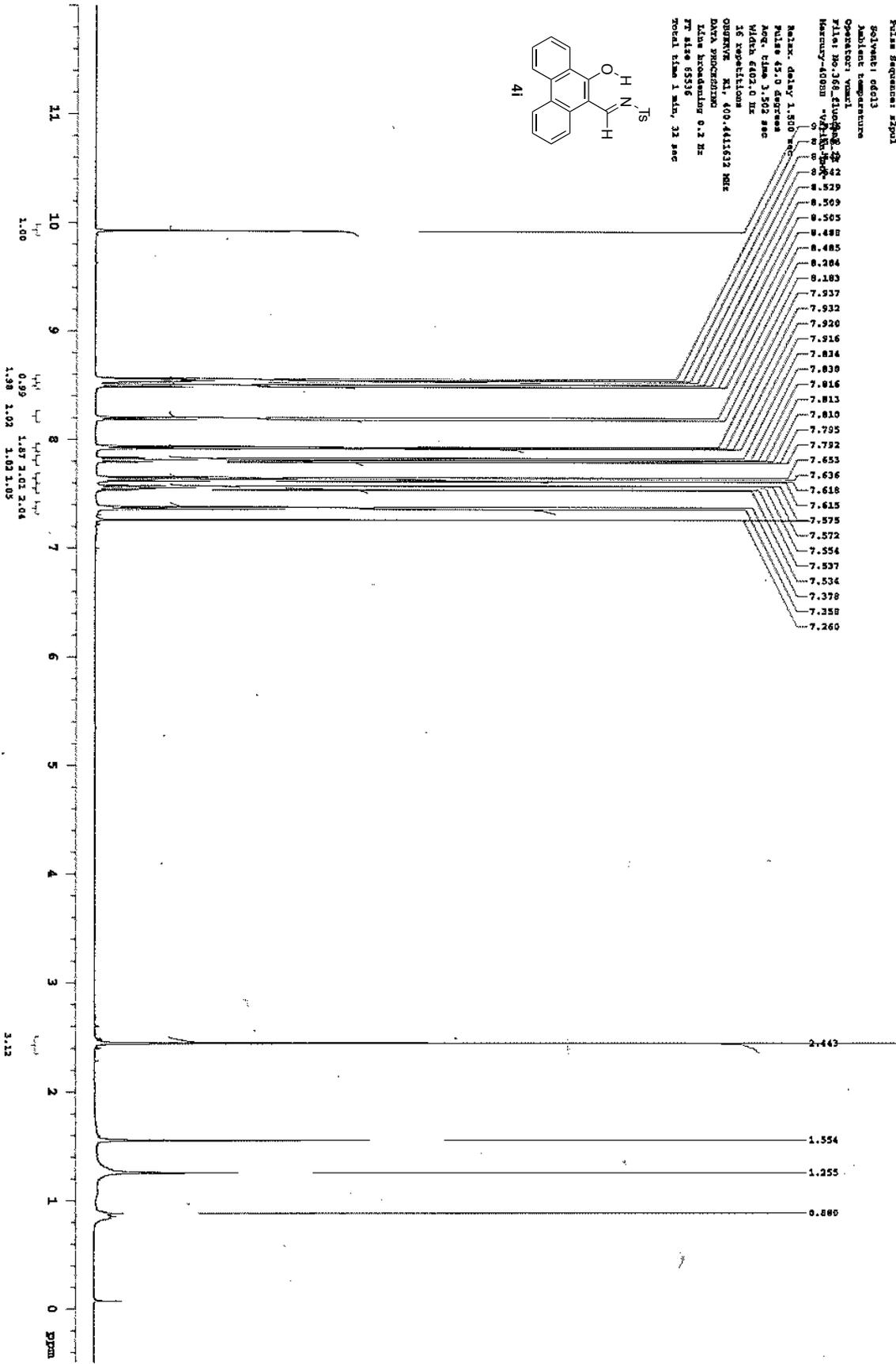
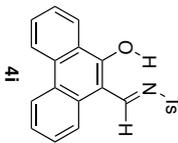
ORGANIC: KI, 400-441832 MHz

DATA PROCESSING

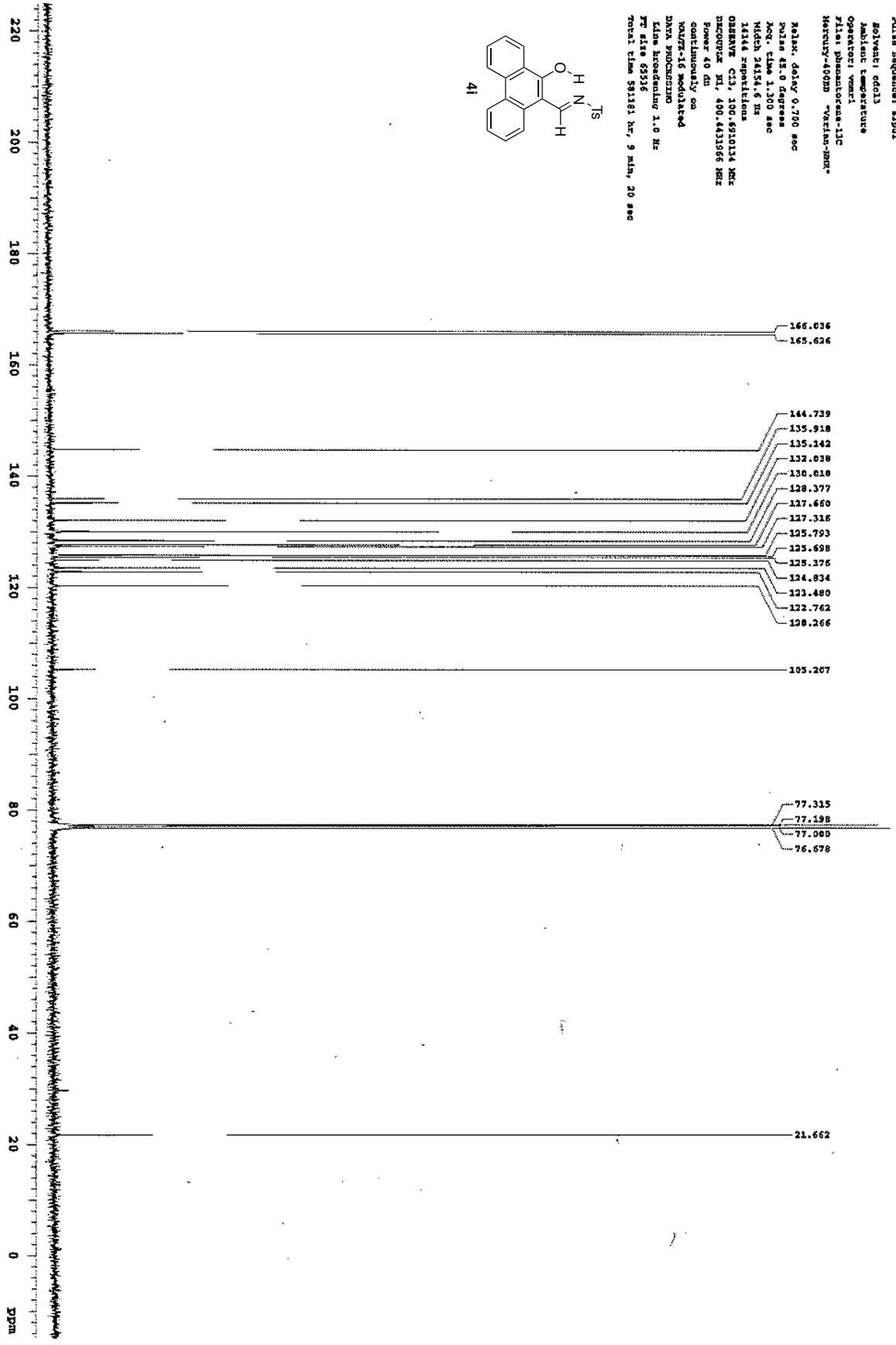
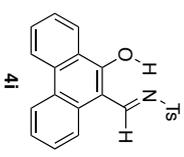
Line broadening: 0.2 Hz

FT size: 65536

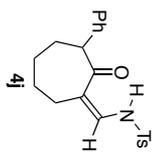
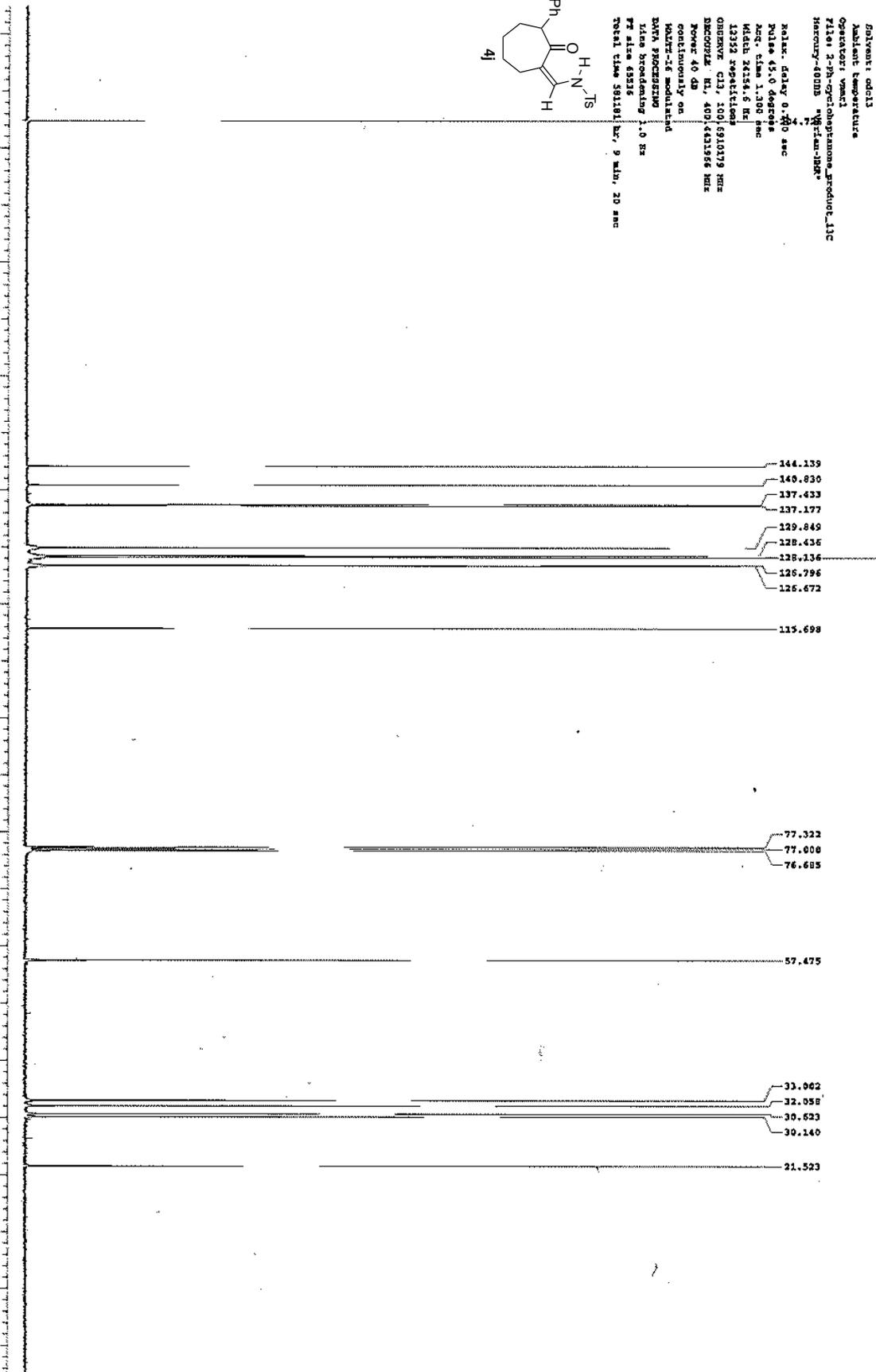
Total time: 1 min, 31 sec



Run name: 06013
 Solvent: Toluene
 Name: 06013
 Operator: vsm
 File: 06013-13C
 Method: 400M - 13C
 Pulse delay: 0.700 sec
 Pulse: 45.0 degrees
 Acq. time: 1.300 sec
 Width: 24554.6 Hz
 14344 repetitions
 OBSERVE: C13 100.631014 MHz
 INVERT: N1 400.431066 MHz
 Power: 40 dB
 Continuously on
 WALTZ-16 modulation
 Data processing:
 F2: 100.631014 MHz
 F1: 400.431066 MHz
 Total time: 59.181 hr, 9 min, 20 sec



220 200 180 160 140 120 100 80 60 40 20 0 ppm



Pulse Sequence: zgpg31
Solvent: cdcl3
Acquisition Temperature: 300.2 K
Operator: ymwt
File: 3-Ph-cyclohexanone_product_11c
Name: 3-Ph-cyclohexanone_product_11c
Pulse Delay: 0.100 sec
Pulse: 45.0 degrees
Acq. time: 1.100 sec
Date_ Time: 11/14/05 11:53
13332 400MHz
CHMURV: C13, 100/6910179 MHz
INSTRUM: H1, 400/441196 MHz
Power: 40 dB
Continuously on
PULPROG: zgpg31
DATA PROCESSING: None
Name: 3-Ph-cyclohexanone_product_11c
PT: 1.100 sec
Total time: 59.181 hr, 9 min, 29 sec

Pulse Sequence: rzpml

Solvent: cdcl3

Ambient Temperature

Operator: ymwt1

File: 20120714013

Sample: 49028 Pyridin-4HCl

Acquisition

Pulse delay 0.200 sec

Pulse 45.0 degrees

Acq. time 1.207 sec

Wd. 2418.6 Hz

16384 repetitions

Channel: C1, 100.6210112 MHz

Decouple: H1, 400.4831268 MHz

Power: 40 dB

Conditionally on

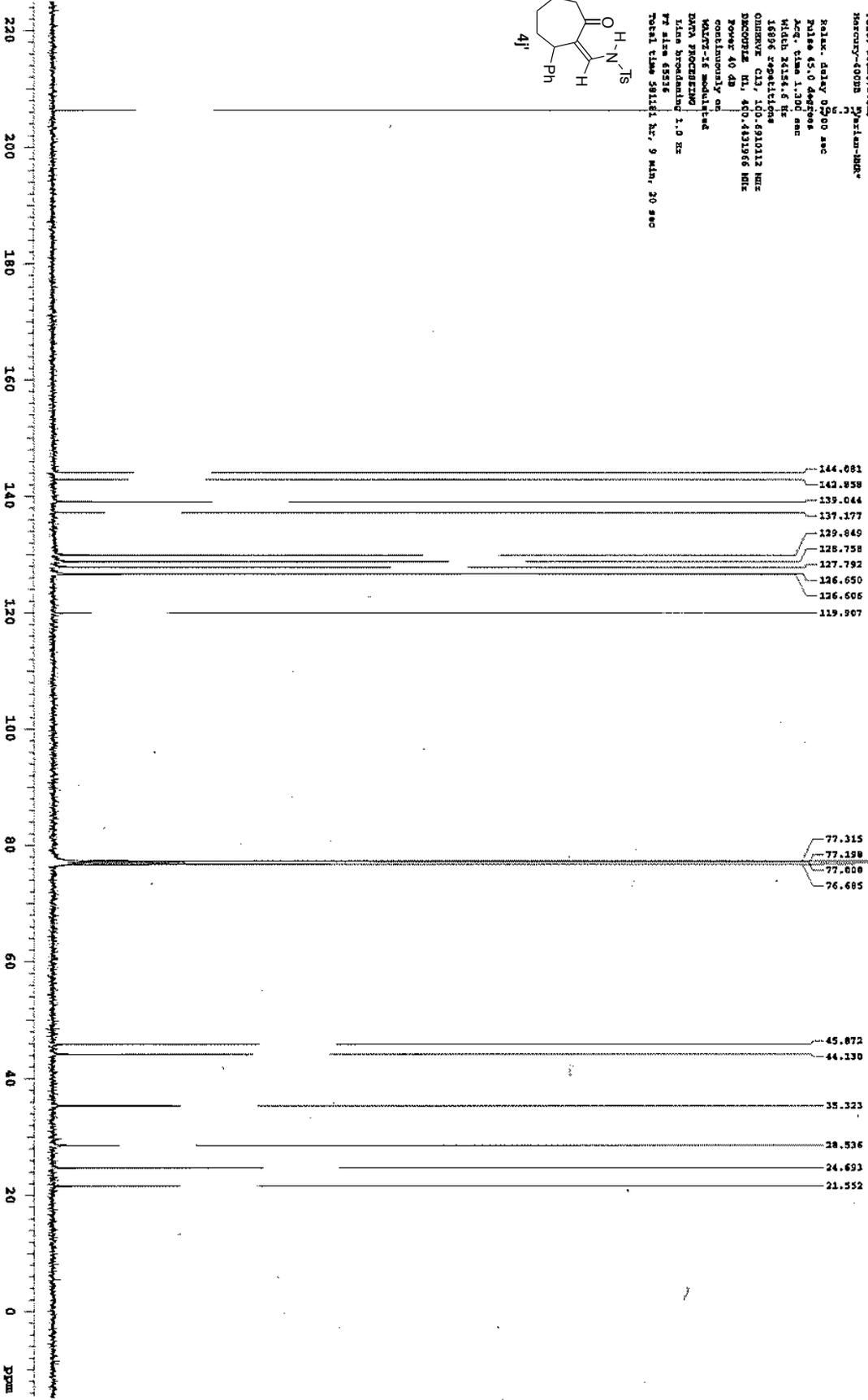
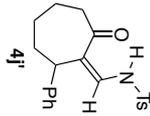
WATER-18 model: r2

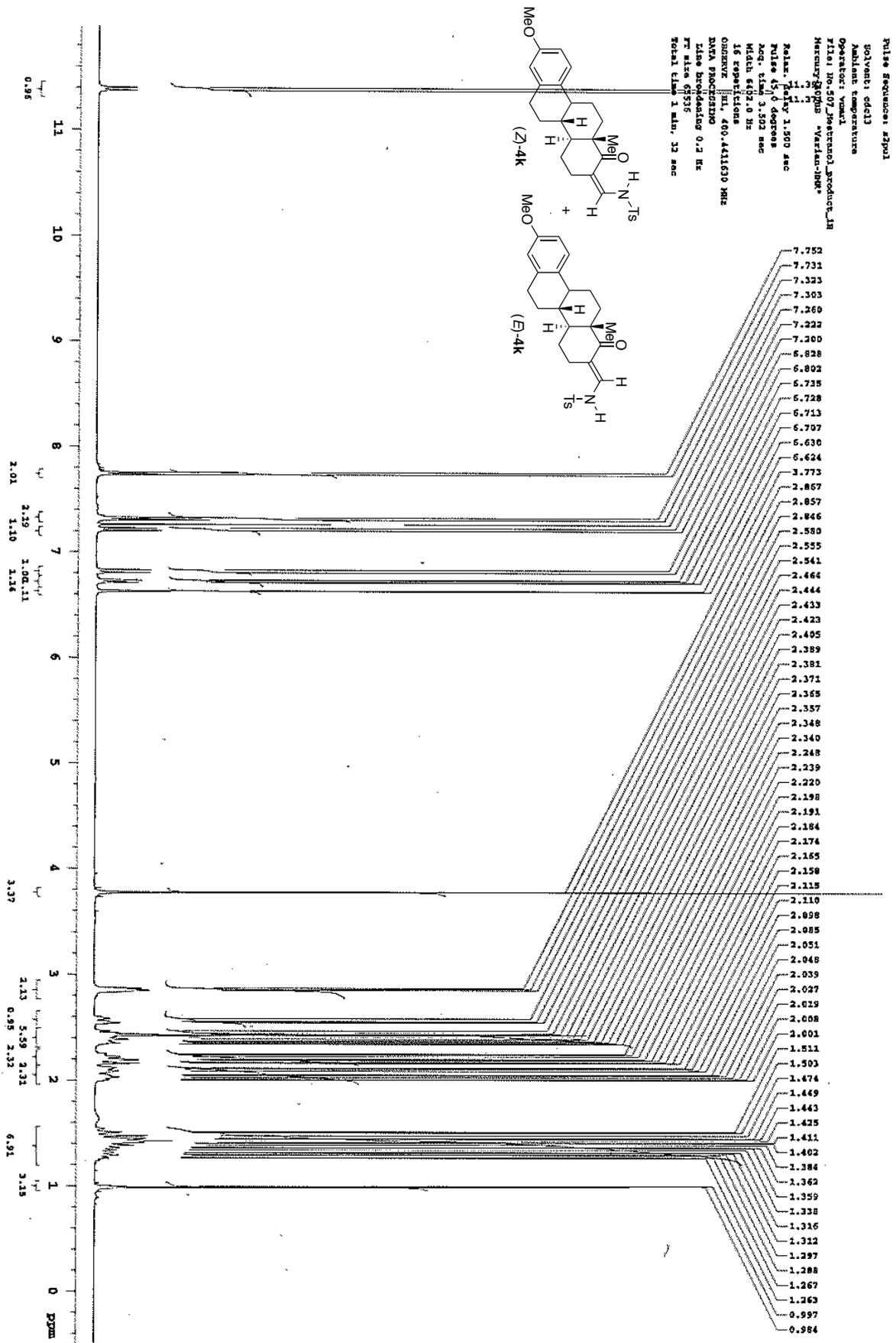
DATA PROCESSING

Line broadening 1.0 Hz

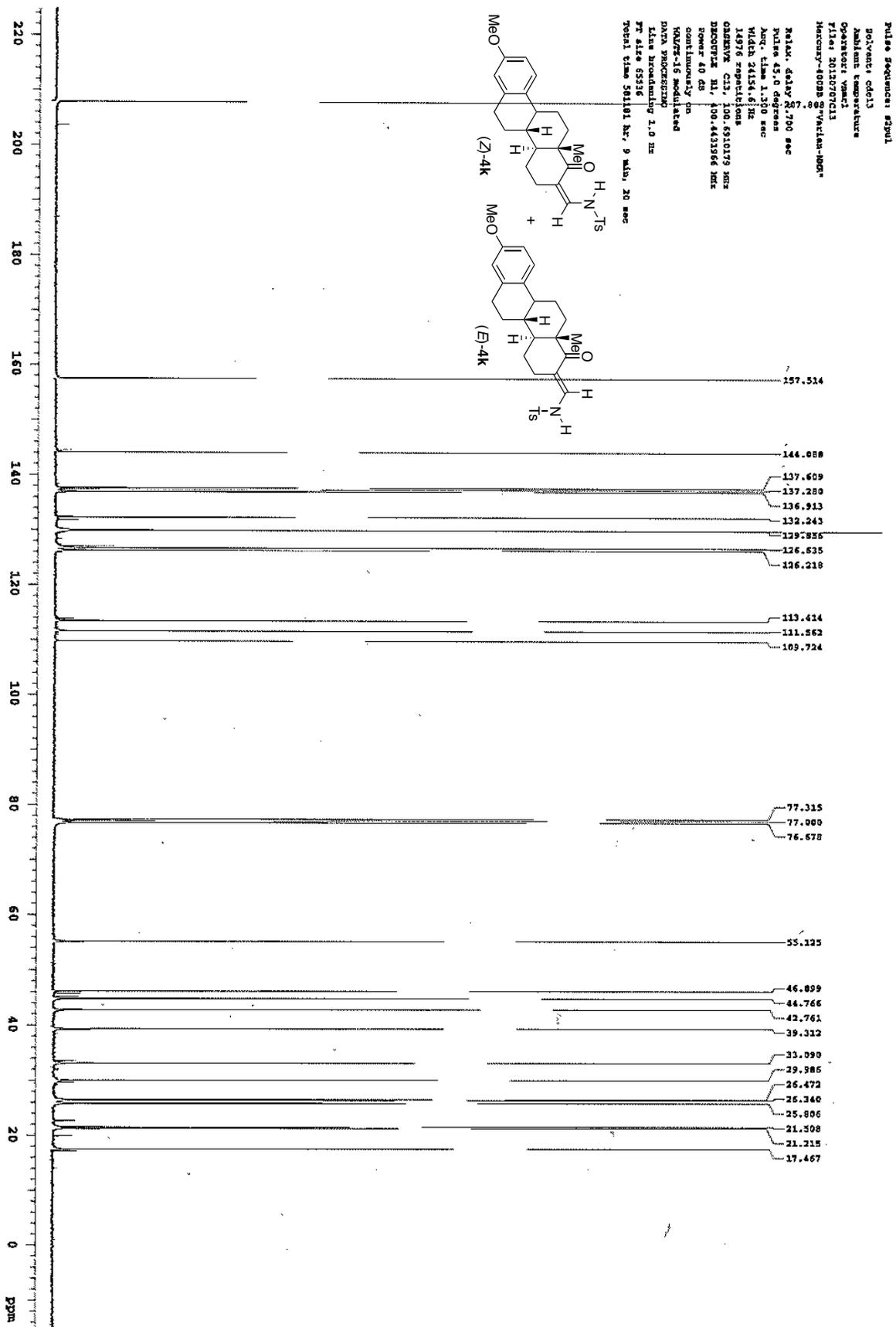
FT scan 45316

Total time 301.811 hr, 9 min, 20 sec

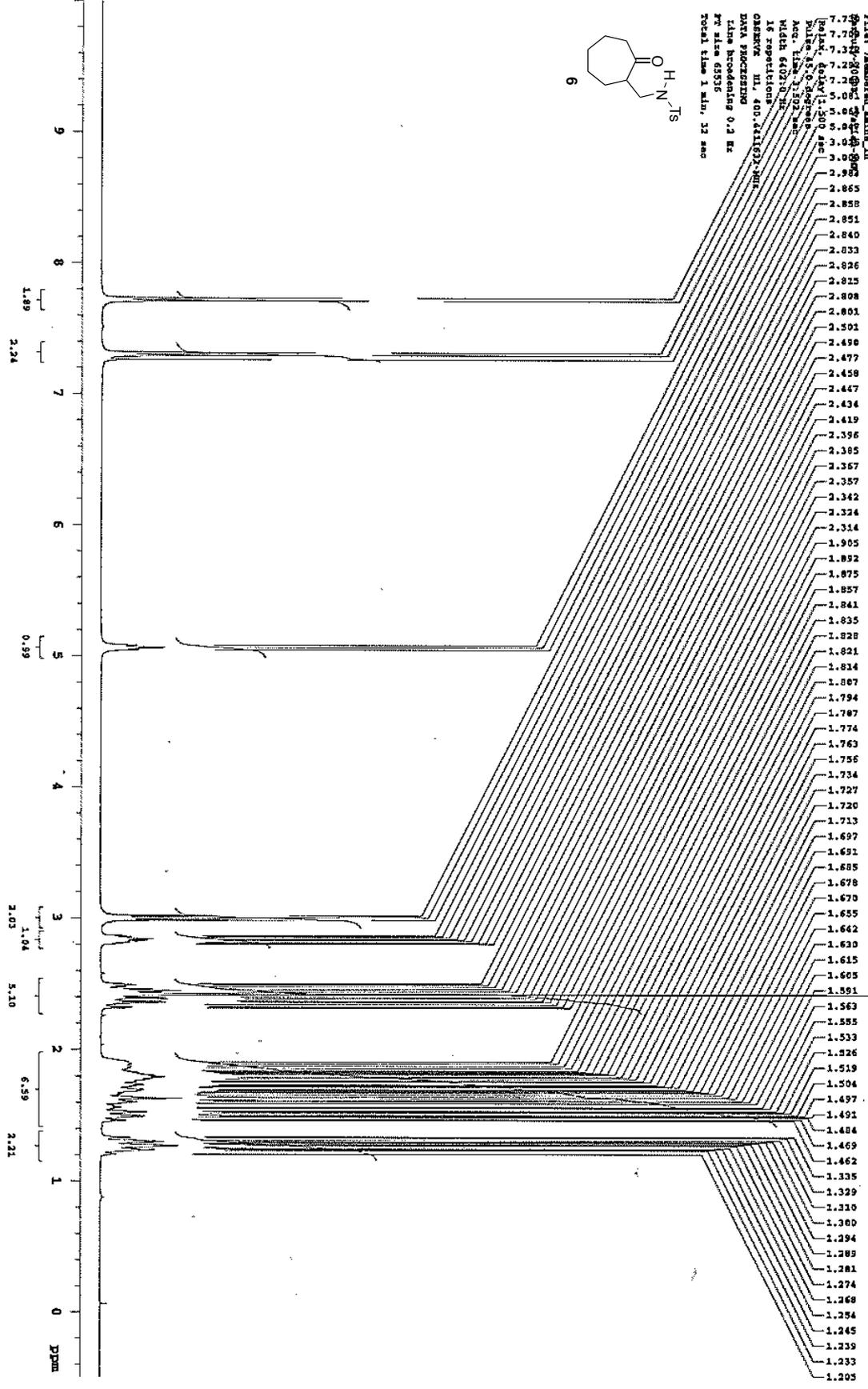
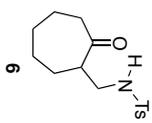




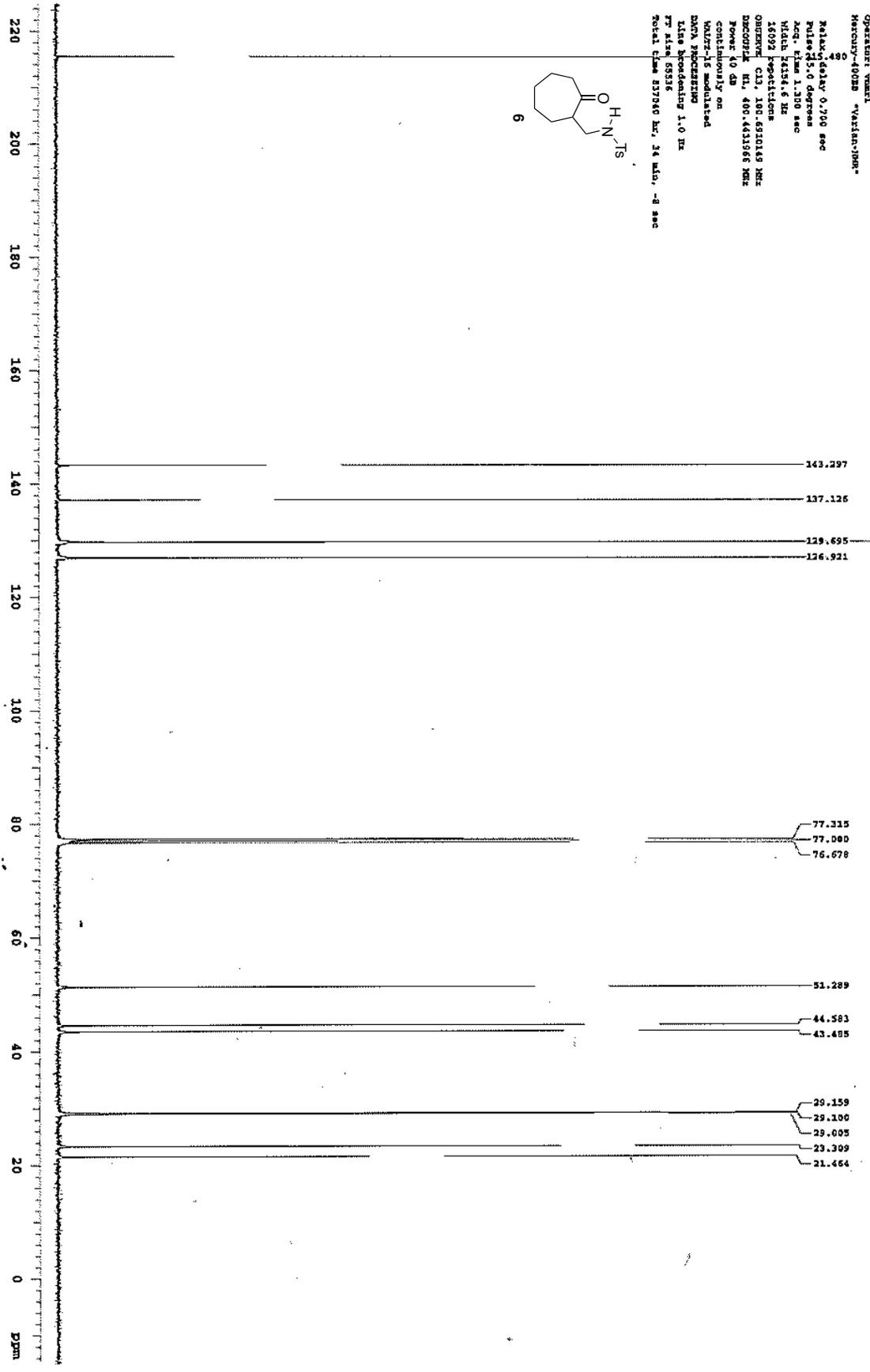
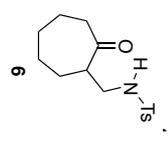
Pulse sequence: zgpg1
 Solvent: cdcl3
 Solvent temperature: 300.2
 Operator: ymml
 P1: 10.507
 P2: 1.000
 P3: 1.000
 P4: 1.000
 P5: 1.000
 P6: 1.000
 P7: 1.000
 P8: 1.000
 P9: 1.000
 P10: 1.000
 P11: 1.000
 P12: 1.000
 P13: 1.000
 P14: 1.000
 P15: 1.000
 P16: 1.000
 P17: 1.000
 P18: 1.000
 P19: 1.000
 P20: 1.000
 P21: 1.000
 P22: 1.000
 P23: 1.000
 P24: 1.000
 P25: 1.000
 P26: 1.000
 P27: 1.000
 P28: 1.000
 P29: 1.000
 P30: 1.000
 P31: 1.000
 P32: 1.000
 P33: 1.000
 P34: 1.000
 P35: 1.000
 P36: 1.000
 P37: 1.000
 P38: 1.000
 P39: 1.000
 P40: 1.000
 P41: 1.000
 P42: 1.000
 P43: 1.000
 P44: 1.000
 P45: 1.000
 P46: 1.000
 P47: 1.000
 P48: 1.000
 P49: 1.000
 P50: 1.000
 P51: 1.000
 P52: 1.000
 P53: 1.000
 P54: 1.000
 P55: 1.000
 P56: 1.000
 P57: 1.000
 P58: 1.000
 P59: 1.000
 P60: 1.000
 P61: 1.000
 P62: 1.000
 P63: 1.000
 P64: 1.000
 P65: 1.000
 P66: 1.000
 P67: 1.000
 P68: 1.000
 P69: 1.000
 P70: 1.000
 P71: 1.000
 P72: 1.000
 P73: 1.000
 P74: 1.000
 P75: 1.000
 P76: 1.000
 P77: 1.000
 P78: 1.000
 P79: 1.000
 P80: 1.000
 P81: 1.000
 P82: 1.000
 P83: 1.000
 P84: 1.000
 P85: 1.000
 P86: 1.000
 P87: 1.000
 P88: 1.000
 P89: 1.000
 P90: 1.000
 P91: 1.000
 P92: 1.000
 P93: 1.000
 P94: 1.000
 P95: 1.000
 P96: 1.000
 P97: 1.000
 P98: 1.000
 P99: 1.000
 P100: 1.000



Pulse sequence: zgpg30
 Solvent: cdcl3
 Ambient temperature
 Operator: smwt
 File: Zamboni_alkn_1H
 Date_Time: 20070717 11:46:08
 Name: 6
 Pulse program: zgpg30
 Acq. time: 2:52.266
 Width: 6402.8 Hz
 16 repetitions
 OBSERVE: 1H, 400, 411(32)-NMR
 DATA PROCESSING
 Line broadening: 0.2 Hz
 FT waltz 65316
 Total time: 1 min, 32 sec

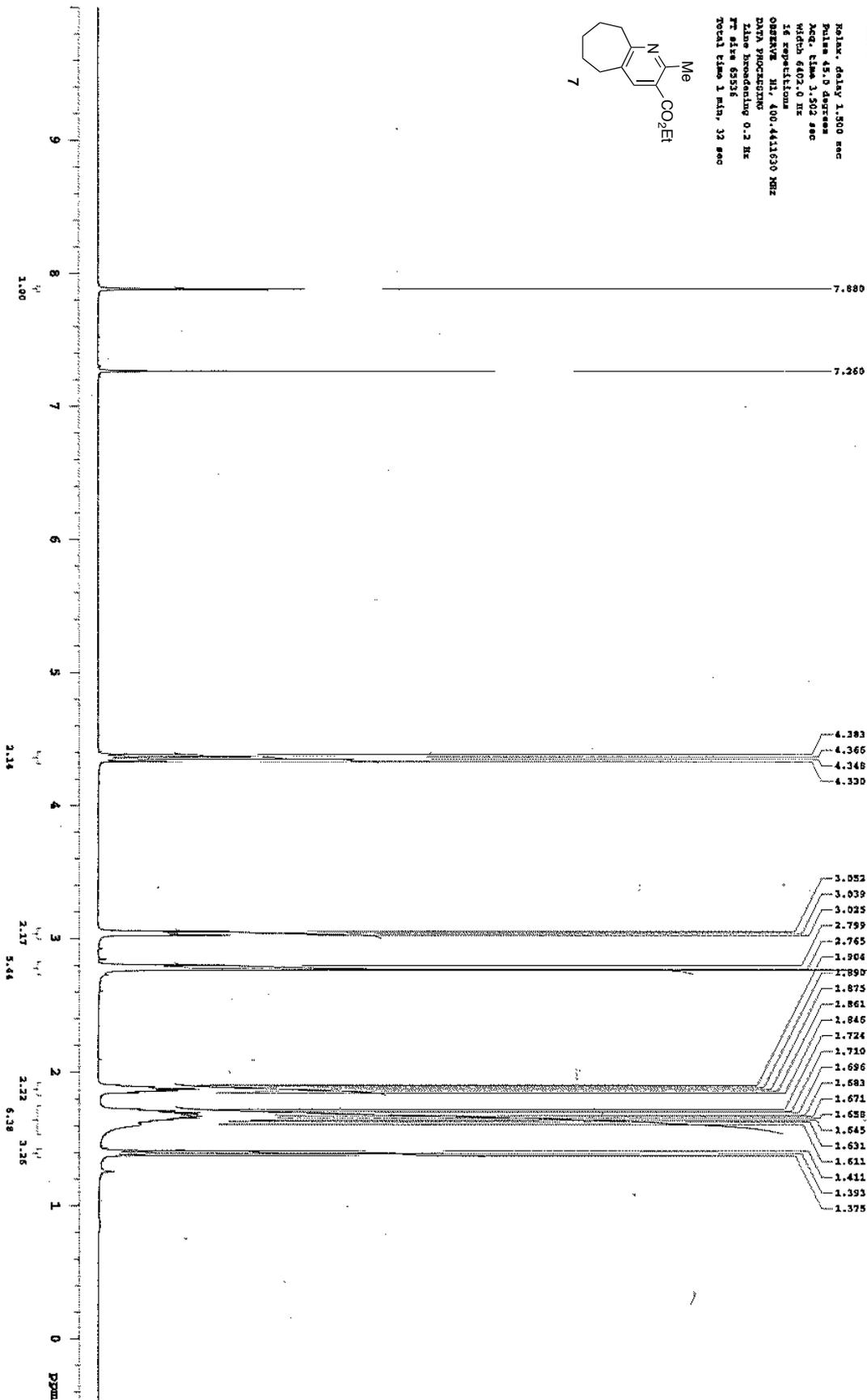
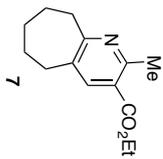


Solvent: cdcl3
 Ambient temperature
 Operator: nmr1
 Mercury-400HD "Varian-jms"
 0
 Relax delay 0.700 sec
 Pulse 5.0 degrees
 Acq. time 1.100 sec
 Width 2156.6 Hz
 16092 points
 OBSERVE C13, 100.6310149 MHz
 DECOUPLE H1, 400.431966 MHz
 Power 40 dB
 continuously on
 WALTZ-16 modulated
 DUMA PROCESSION
 Line broadening 1.0 Hz
 FT Aiac 65316
 Total time 237940 Hz, 34 min., -8 sec

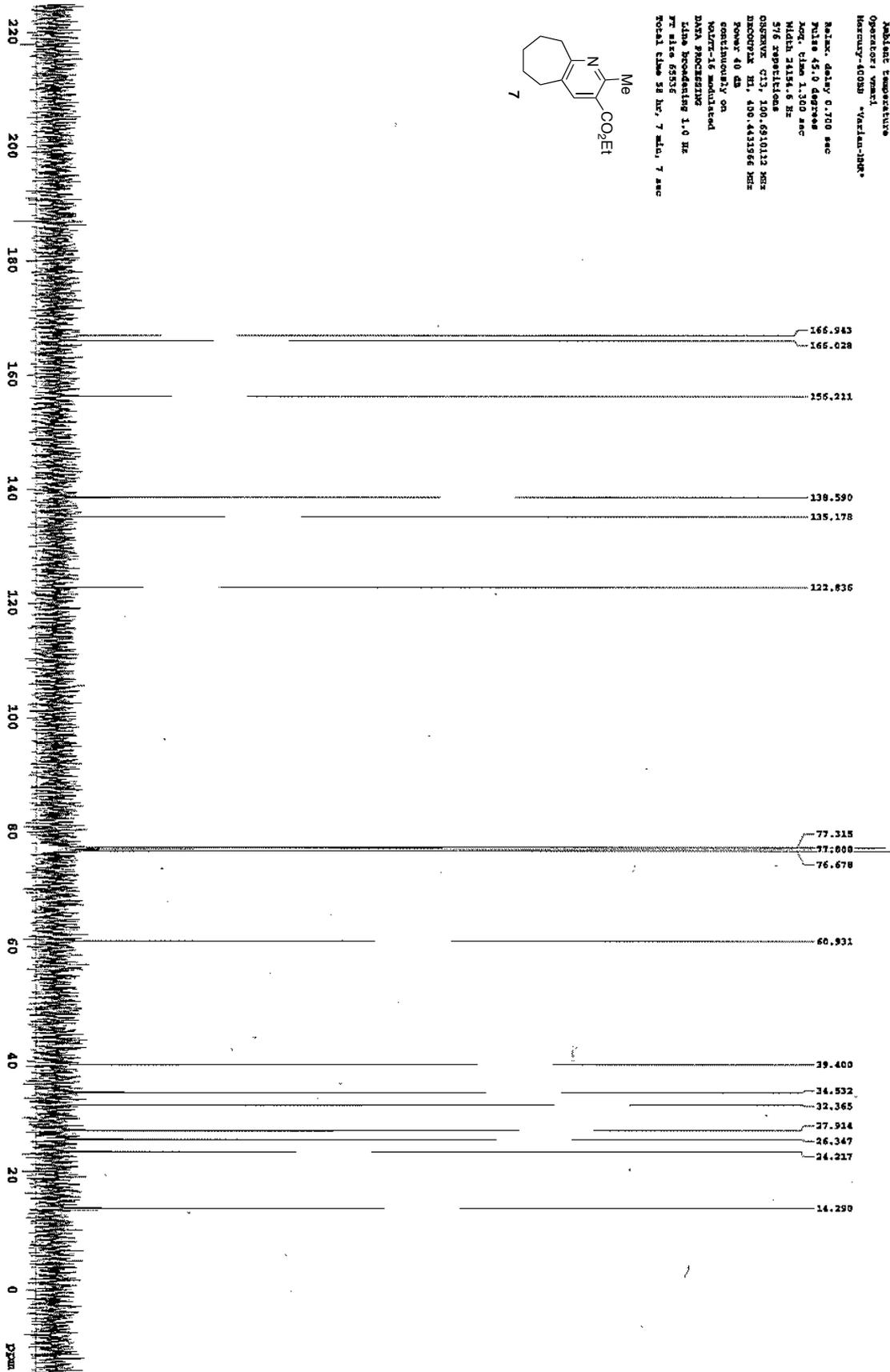
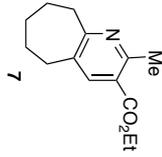


Pulse sequence: s2p01
 Solvent: cdcl3
 Ambient temperature
 Operator: ymml
 Mercury-400MHz "Varian-100"

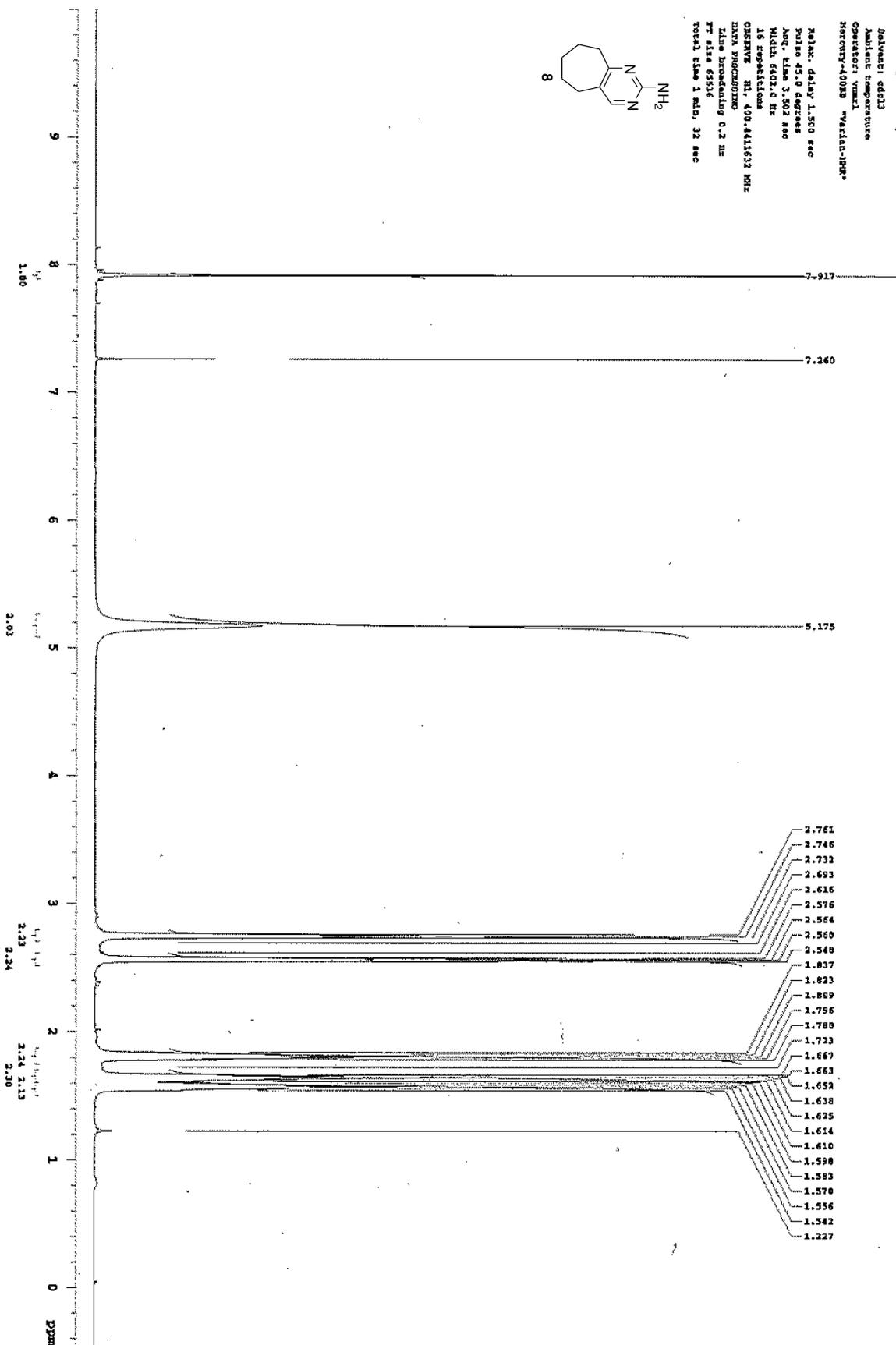
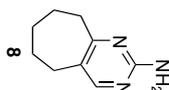
Relax delay 1.500 sec
 Pulse 45.0 degrees
 Acq. time 3.302 sec
 Width 6402.0 Hz
 IS repetitions
 OBSERVE HI, 400.441630 MHz
 DATA PROCESSING
 Xline broadening 0.2 Hz
 FT size 65536
 Total time 1 min, 32 sec



Solvent: cdcl3
 Substrate: Emameprazine
 Operator: j. frank
 Instrument: Varian-100M
 Name: 45154-6
 Pulse delay: 0.700 sec
 Pulse: 45.0 degrees
 Acq. time: 1.300 sec
 Width: 24154.6 Hz
 576 repetitions
 Observed: C13, 100.631013 MHz
 INCREMENT: 0.1, 400.443196 MHz
 Power: 40 dB
 continuously on
 NMR-16 multiscan
 DATA PROCESSING
 Line broadening: 1.0 Hz
 FT size: 65535
 Total time: 58 hr, 7 min, 7 sec



Pulse sequence: zgpg30
 Solvent: dcd3
 Ambient temperature
 Operator: ymml
 Frequency: 400MHZ Varian-DMR
 Relax. delay: 1.500 sec
 Pulse 45.0 degree
 Acq. time: 3.502 sec
 Width: 8402.0 Hz
 16 repetitions
 OBSERVE HI, 400.411632 MHz
 INVA PROGRAM
 Line broadening: 0.2 Hz
 FT size: 65536
 Total time: 1 min, 32 sec



=====

Pulse sequence: zgpg1

Solvent: cdcl3

Acquisition temperature

Operator: YMAKI

Hardware: 400MHz Varian-DMX

Relax. delay: 0.700 sec

Pulses: 45.0 degrees

Acq. time: 1.300 sec

Width: 24344.6 Hz

384 repetitions

ORIGIN: C13, 100.630157 MHz

DECODE: H1, 400.4431865 MHz

Power: 40 dB

continuously on

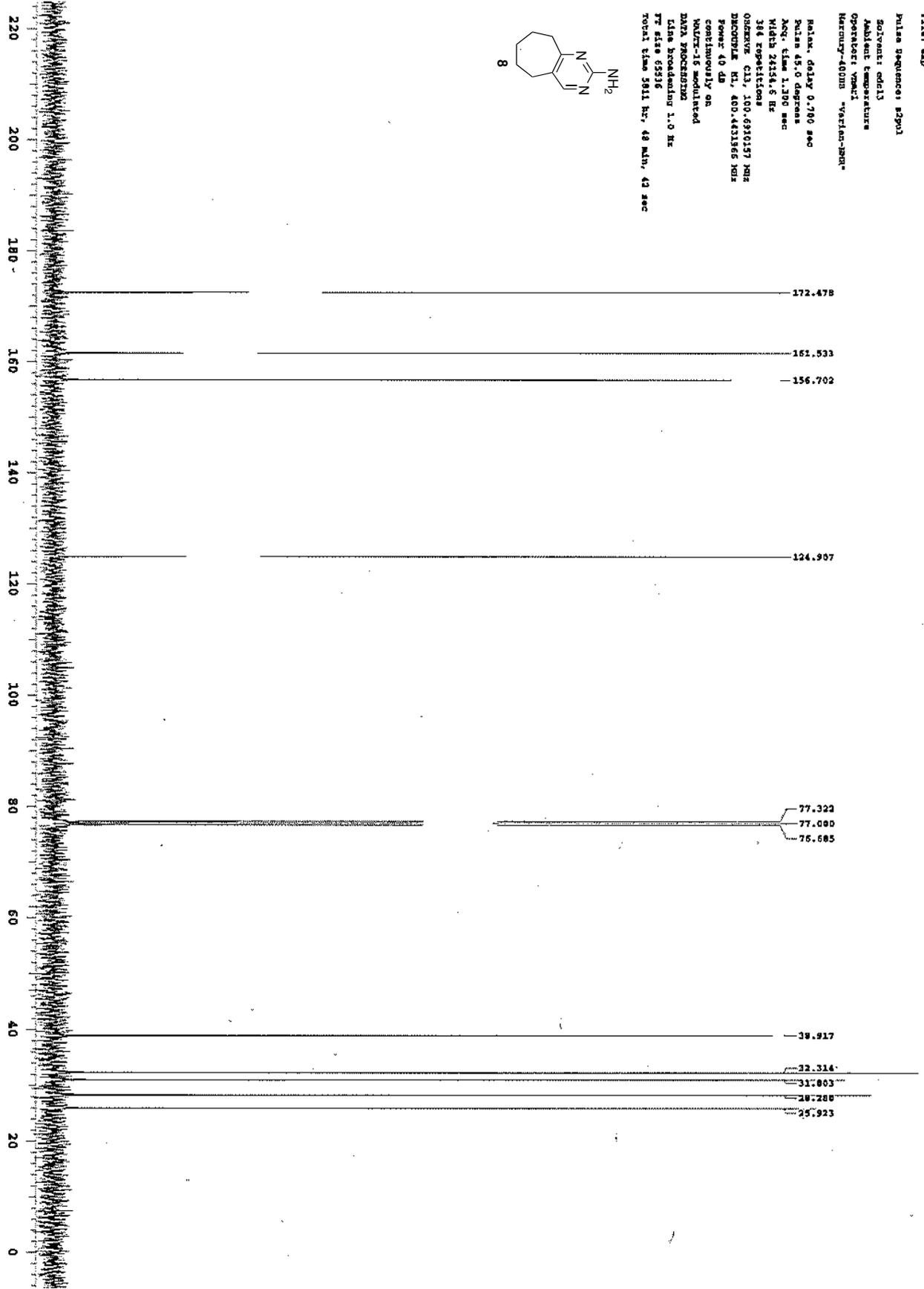
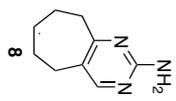
NUC1: 13 modulation

DATA PROCESSING:

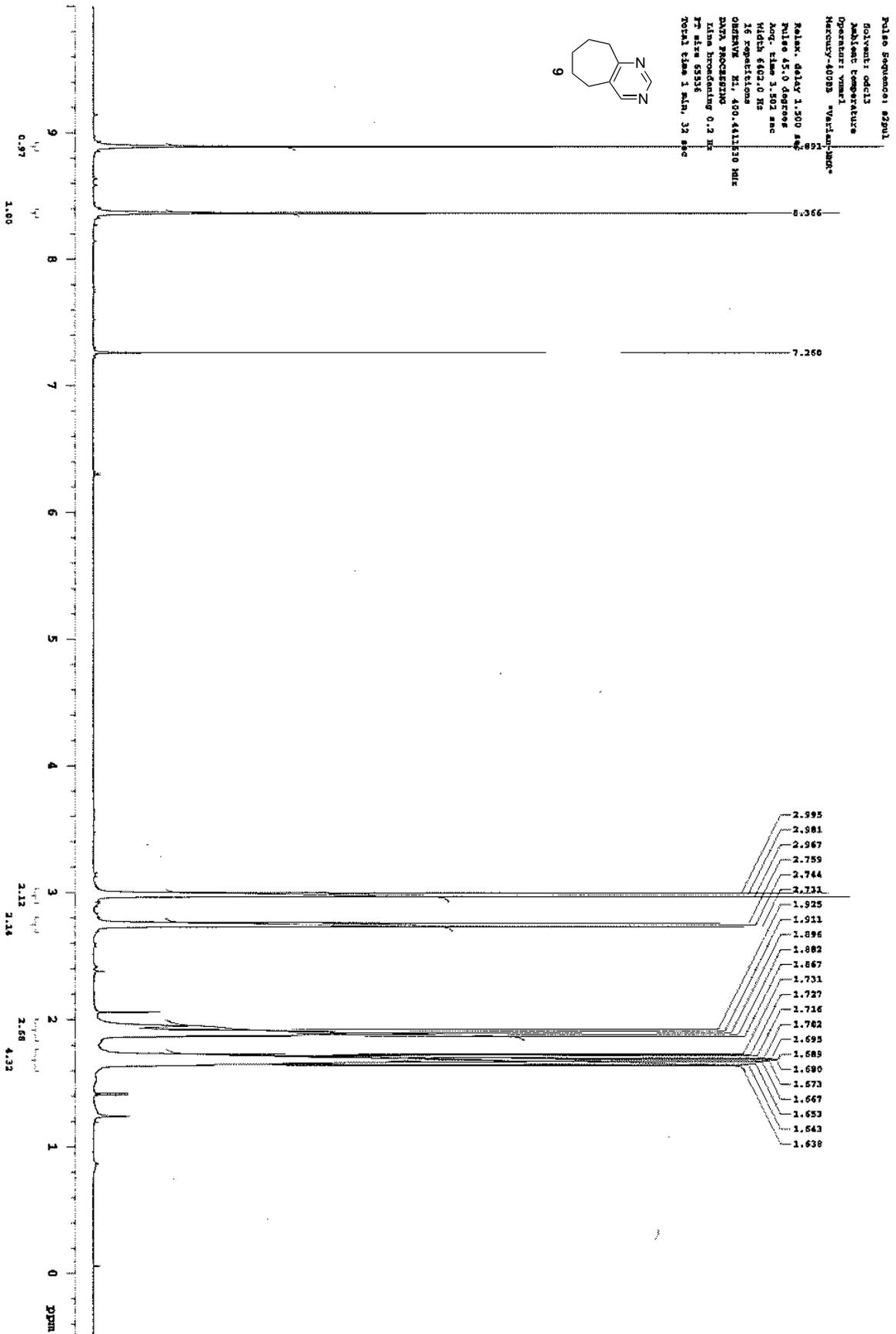
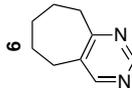
Line broadening: 1.0 Hz

TF size: 65536

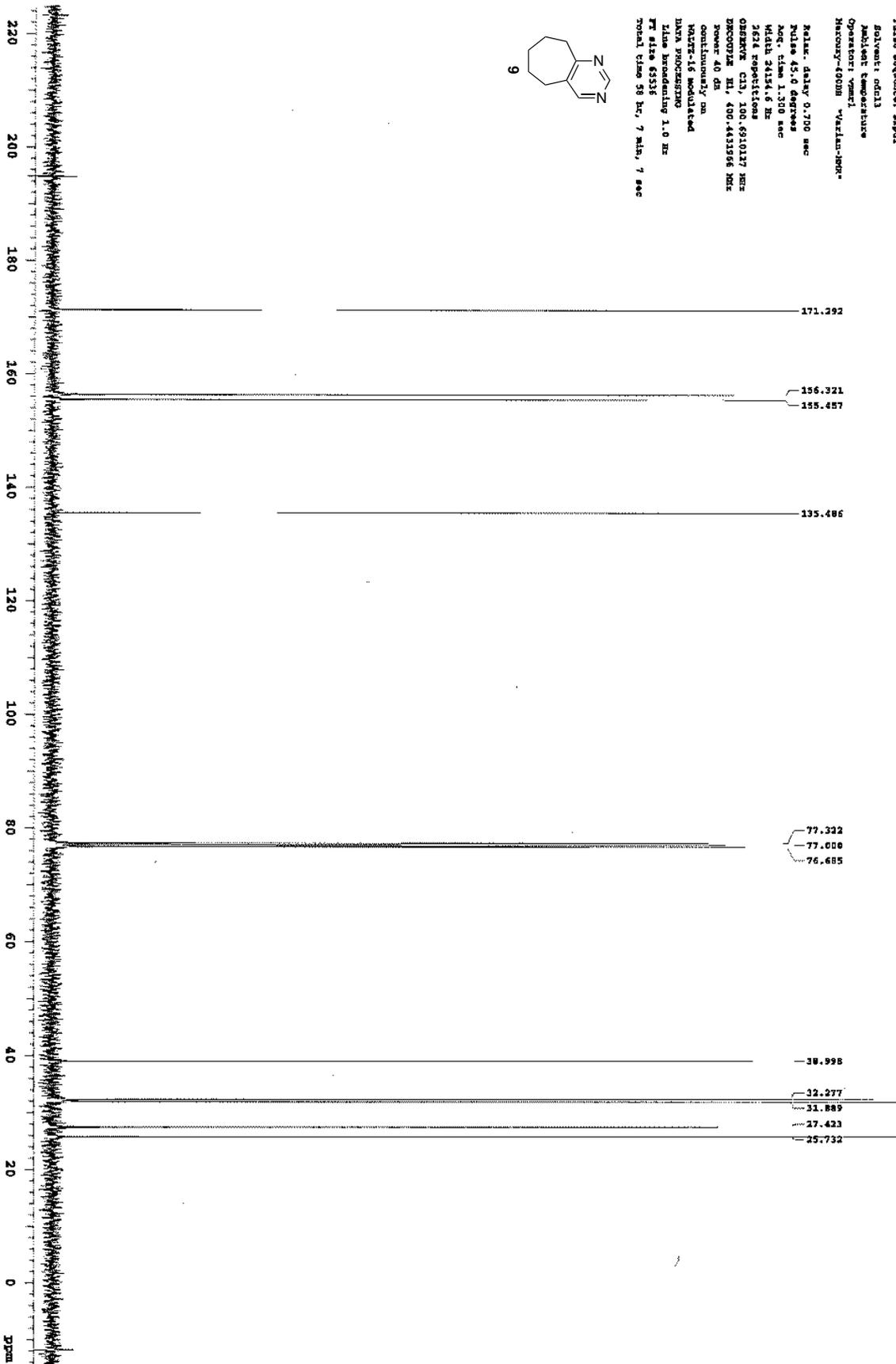
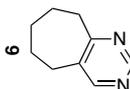
Total time: 3811 Hz, 48 min, 43 sec



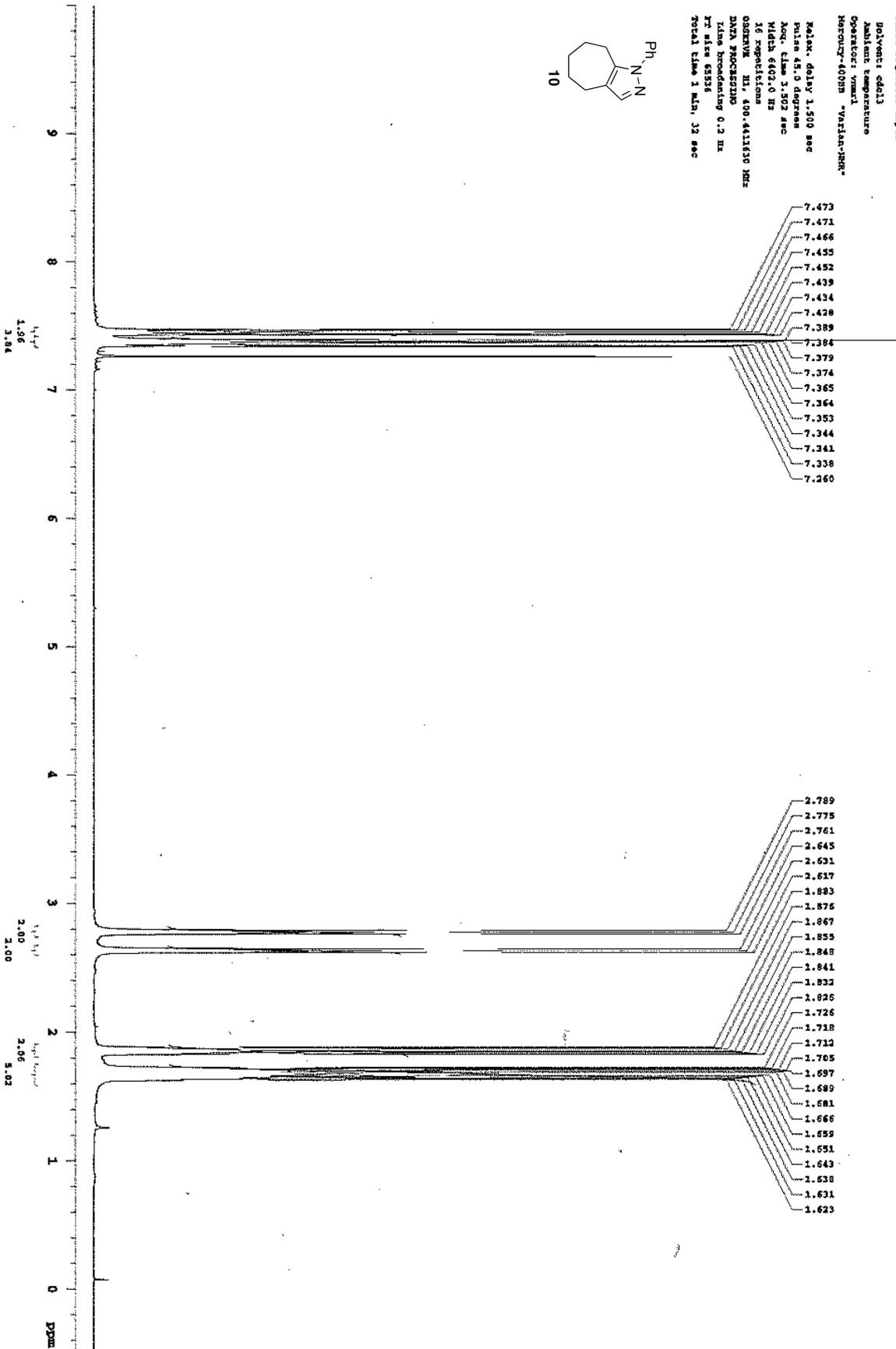
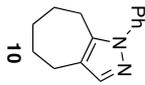
Pulse Sequence: zgpg30
 Solvent: cdcl3
 Ambient Temperature
 Operator: ymas1
 Mercury-400MHz "Varian-DMX"
 Relax. delay 1.500 sec
 Pulse 45.0 degrees
 Acq. time 3.501 sec
 Width 6402.0 Hz
 16 repetitions
 OBSERVE: H1, 400.441130 MHz
 DATA PROCESSING
 Gain broadening 0.2 Hz
 FT size 65316
 Total time 1 min, 32 sec



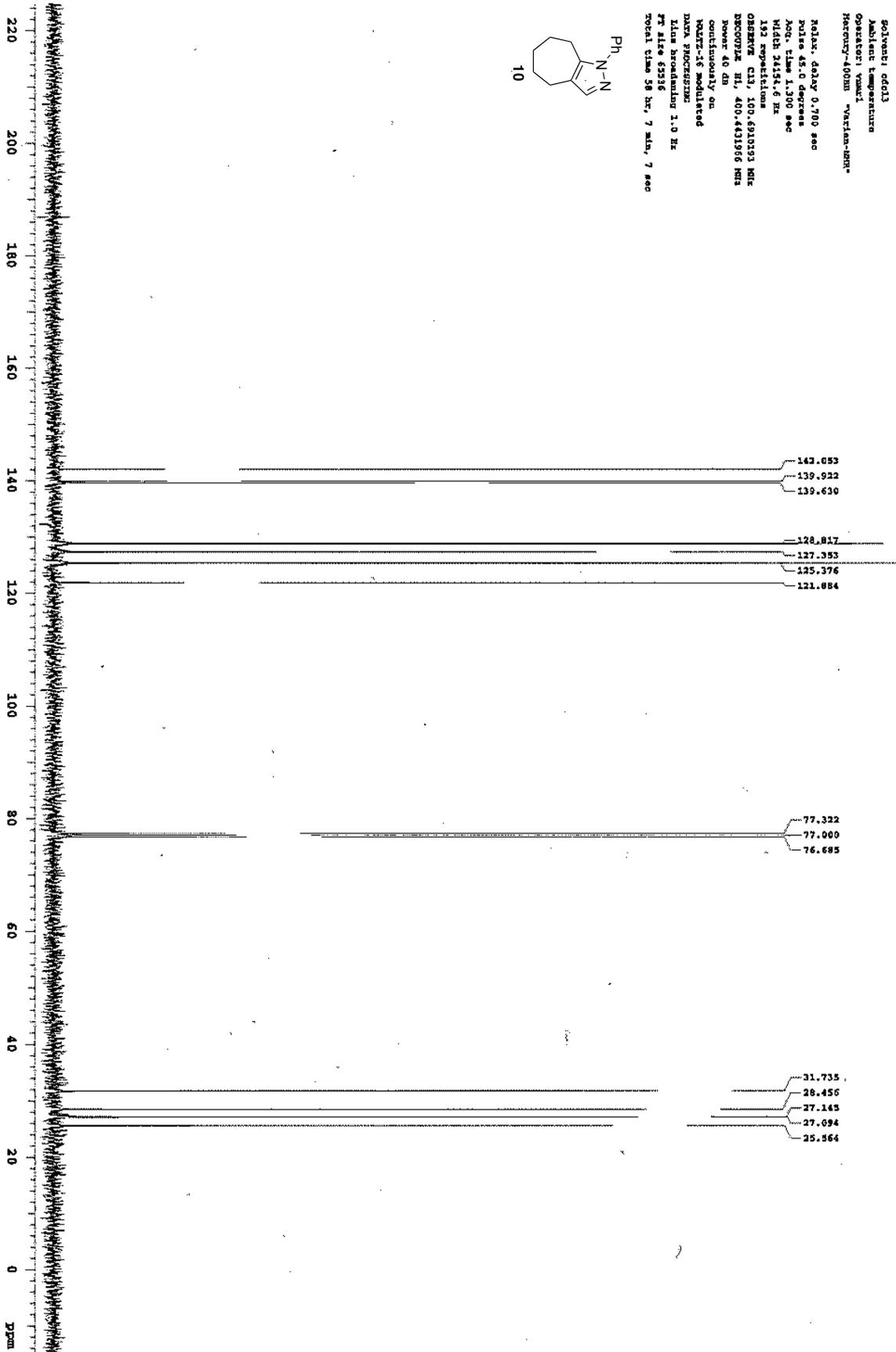
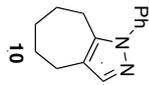
Pulse sequence: zgpg30
 Solvent: cdcl3
 Solvent temperature
 Operator: vamsi
 Macro: zgpg30 "Varian-90M"
 Relax. delay 0.700 sec
 Pulse 45.0 degrees
 Acq. time 1.300 sec
 Width 24124.8 Hz
 2564 repetitions
 OBSERVE CH1, 100.6310117 MHz
 EXCITE CH1, 400.4431966 MHz
 Power 40 dB
 continuously on
 VAPOR-16 Modulated
 DATA PROCESSING
 Line broadening 1.0 Hz
 FT size 65536
 total time 58 hr, 7 min, 7 sec



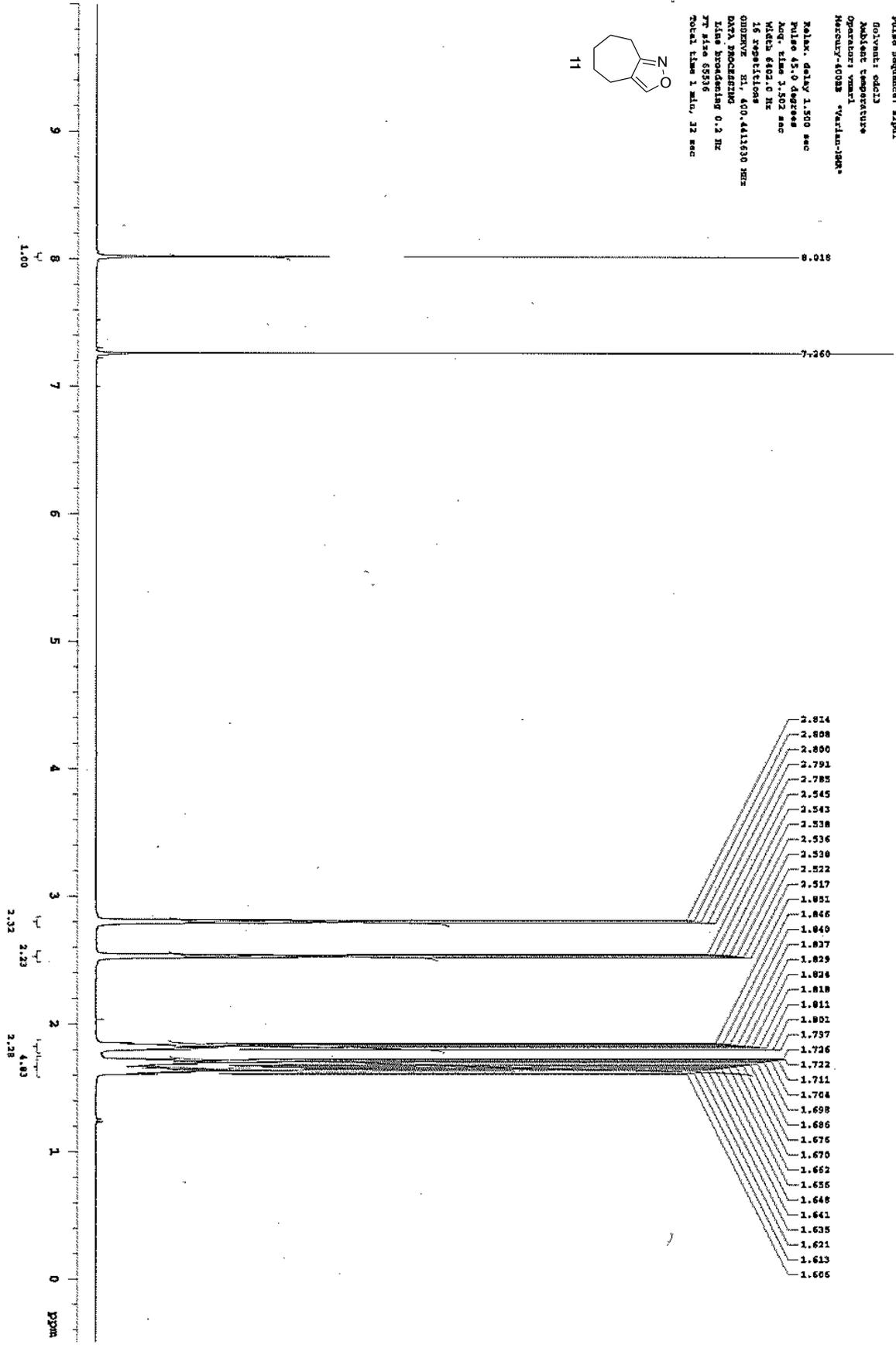
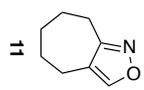
Pulse Sequence: zgpg30
 Solvent: cdcl3
 Ambient temperature
 Operator: vmm1
 Name: HPC-008H - Varian-PPM-
 Pulse delay 1.500 sec
 Relax 45.0 degrees
 Acq. time 1.502 sec
 Width 6602.0 Hz
 16 Repetitions
 OBSERVE H1, 400.441830 MHz
 DATA PROCESSING
 F2 value 65255
 IR size 65255
 Total time 1 min, 32 sec



Pulse Sequence: zgpg31
 Solvent: cdcl3
 Acquisition Temperature
 Operator: vsmc
 Frequency: 100MHz
 Relax Delay: 0.700 sec
 Pulse: 45.0 degrees
 Acq. Time: 1.300 sec
 Width: 24154.6 Hz
 133 repetitions
 OBSERVE CH: 100.6210193 MHz
 DECOUPLE CH: 400.431966 MHz
 Power: 40 dB
 continuously on
 HOLTZ-16 modulated
 MAX PROGS: 2
 Scan Broadening: 1.0 Hz
 FT Aft: 65336
 Total time: 58 hr, 7 min, 7 sec



Pulse Sequence: zgpg31
 Solvent: cdcl3
 Substance: ynamid
 Operator: ynamid
 Mercury-400BS "Varian-100"
 Relax. delay: 1.500 sec
 Pulse: 45.0 degrees
 Acq. time: 1.502 sec
 Width: 6402.0 Hz
 16 repetitions
 OHSERVE: SI, 400, 4411630 MHz
 DATA PROCESSING
 Line broadening: 0.2 Hz
 FT: also 65336
 Total time: 1 min, 12 sec



Pulse Sequence: zgpg30
Solvent: cdcl3
Ambient temperature
Operator: ymml
Acronym: 400MH "Varian-100"

Pulse delay: 0.700 sec
Pulse: 45.0 degrees
Acq. time: 1.100 sec
Width: 24154.6 Hz
4116 repetitions
OBSERVE CH: 100.6210127 MHz
PROBHD: H1, 400.4431956 MHz
Power: 40 dB
continuously on
NUC1: 15 modulated
DATA STRUCTING
SOLV: H2O
ILAS: H2O
IT size: 65536
Total time: 581 hr, 10 min, 52 sec

