

Supplemental Information

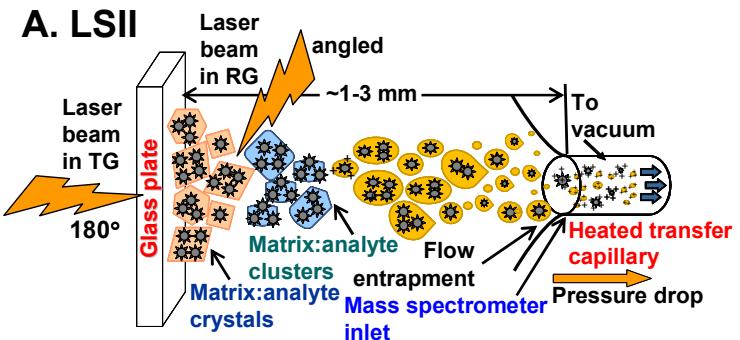
Laserspray Ionization Imaging of Multiply Charged Ions using a Commercial Vacuum MALDI Ion Source

Ellen D. Inutan¹, James Wager-Miller², Ken Mackie², Sarah Trimpin^{1*}

¹*Department of Chemistry, Wayne State University, Detroit, MI 48202 USA*

²*Gill Center for Biomolecular Science, Indiana University, Bloomington, IN 47405*

USA



B. MAII

Tapping
Glass plate
To vacuum
Heated transfer capillary
Pressure drop
Matrix:analyte crystals

Scheme S1. Cluster model representation at atmospheric pressure of (A) LSII in transmission geometry and reflection geometry laser ablation and (B) MAII with no laser of the matrix/analyte crystals forming charged matrix/analyte clusters. Multiply charged ions are produced in the presence of a pressure drop region from atmospheric pressure to vacuum and desolvation of the matrix from the clusters in the heated transfer capillary.

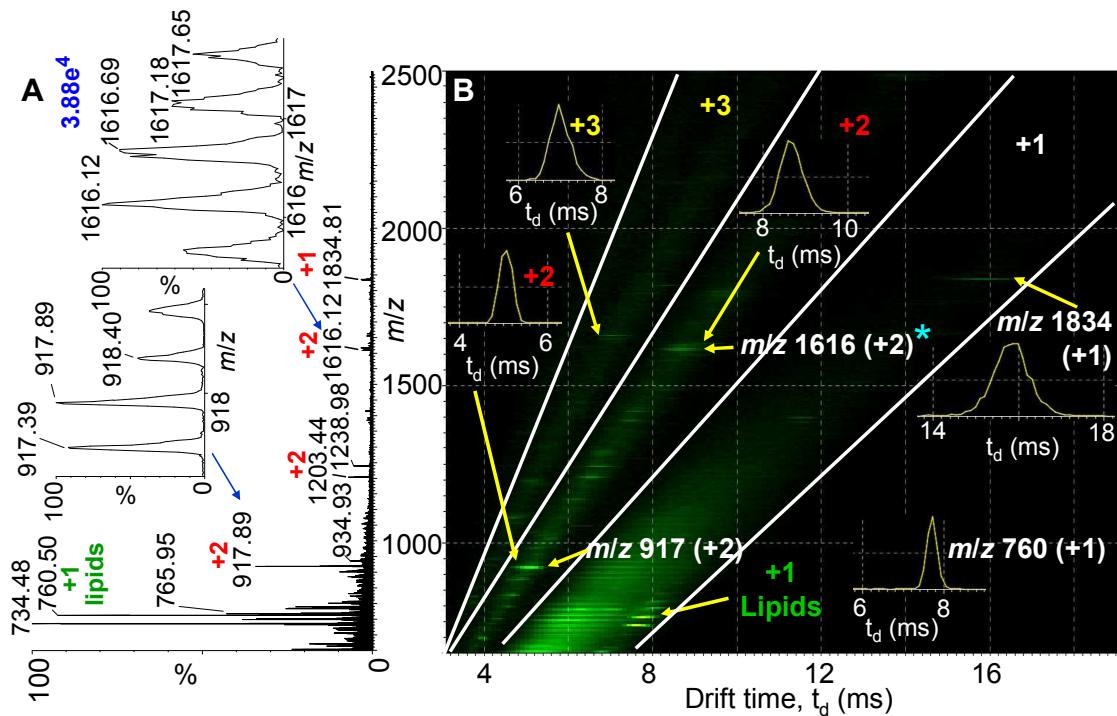


Figure S1. LSIV-IMS-MS (**A**) total mass spectrum and (**B**) two-dimensional plot of drift time vs. m/z of delipidified mouse brain tissue spotted with 2,5-DHAP matrix and acquired using ‘LSI settings’^{39,42} on a commercial intermediate pressure MALDI source of the SYNAPT G2 mass spectrometer. Insets show (**A**) isotopic distribution of +2 ions of the identified *N*-acetylated MBP peptide (m/z 917.39) and the highest MW detected, ~3.2 kDa (m/z 1616.12) (see *), and (**B**) drift times of +3, +2, and +1 ions.

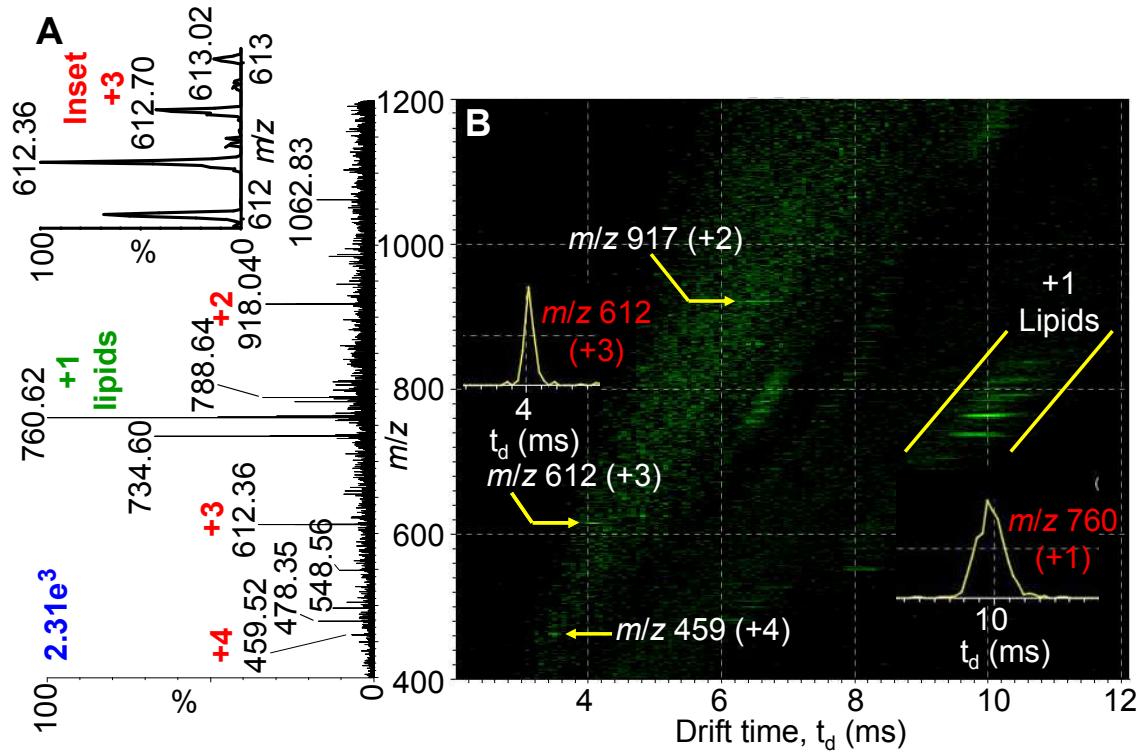


Figure S2. LSII-IMS-MS (A) total mass spectrum and (B) two-dimensional plot of drift time vs. m/z of non delipidified mouse brain tissue spotted with 2,5-DHAP matrix and acquired using the nanoESI source of the SYNAPT G2 mass spectrometer with a homebuilt skimmer cone to perform LSII in transmission geometry. Insets show in (A) isotopic distribution and (B) drift times of +3 charge state ion (m/z 612.36) of the identified *N*-acetylated MBP peptide and +1 drift time of a lipid (m/z 760.62).

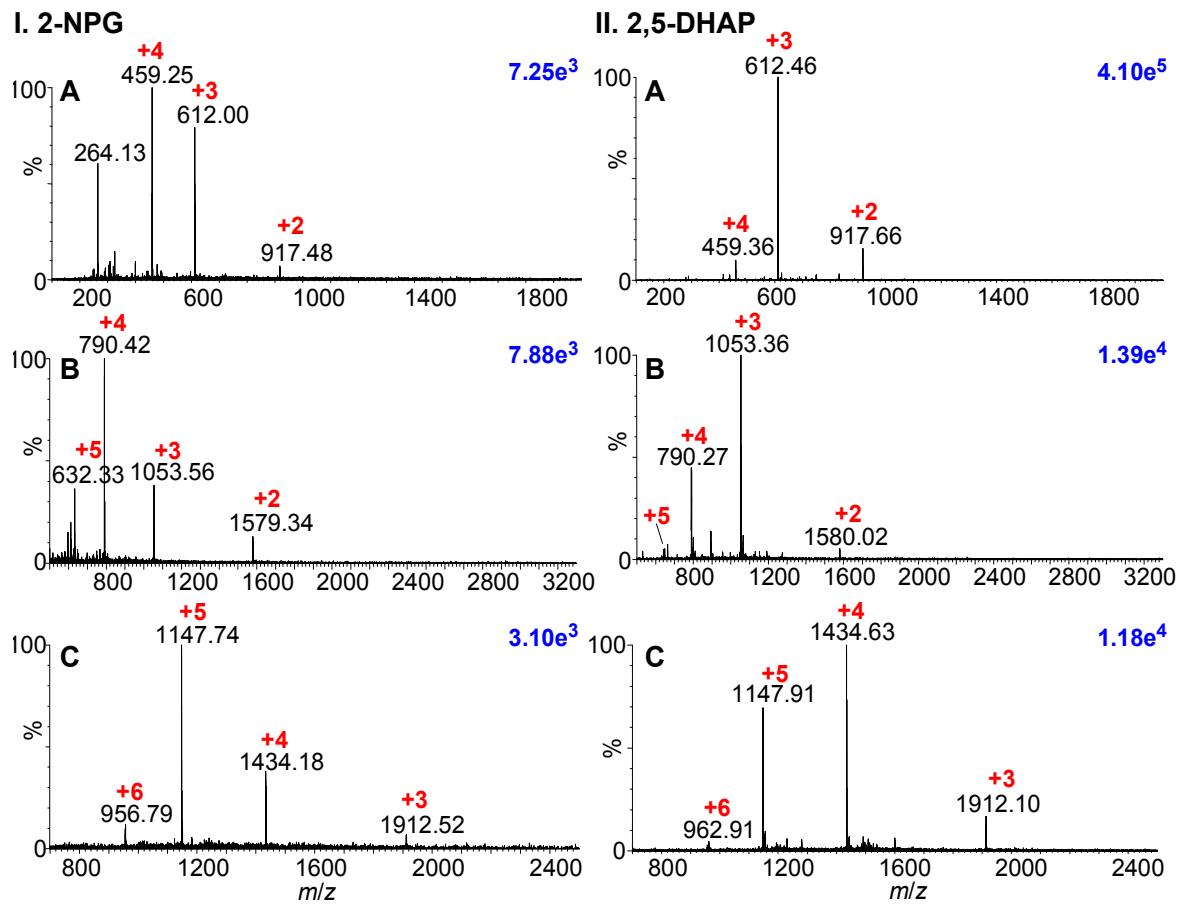


Figure S3. LSIV-MS using (I) 2-NPG matrix at low laser power (4.2 J cm^{-2}) and (II) 2,5-DHAP matrix at higher laser power (7.3 J cm^{-2}) of (A) *N*-acetylated MBP peptide (MW 1833 Da), (B) galanin (MW 3150 Da), and (C) bovine insulin (MW 5731 Da) acquired using ‘LSI settings’^{39,42} on a commercial intermediate pressure MALDI source of the SYNAPT G2 mass spectrometer.

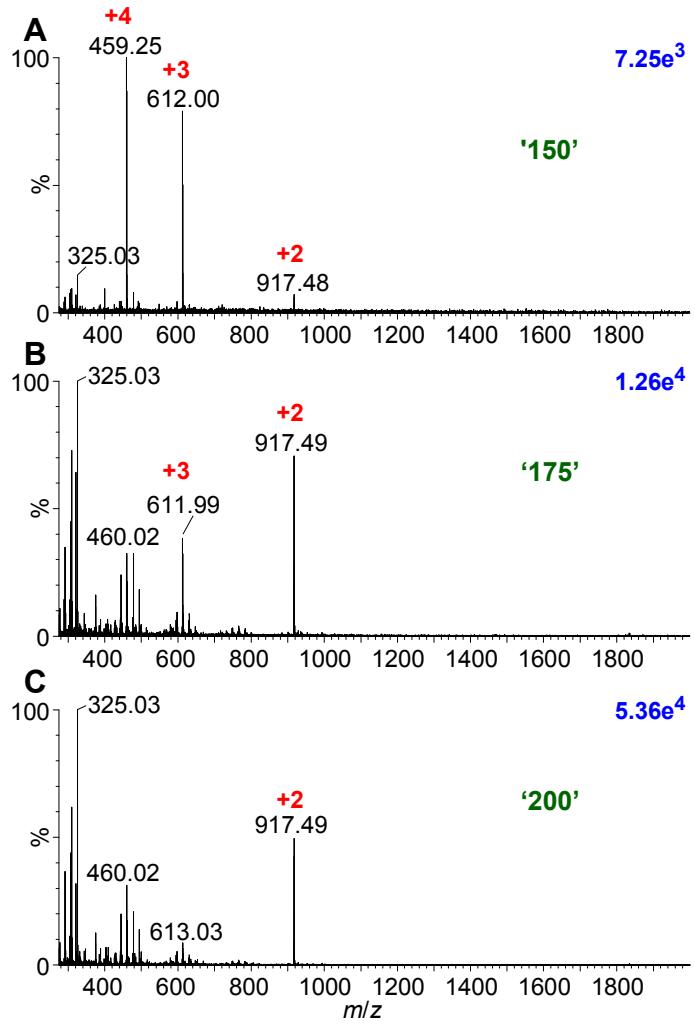


Figure S4. LSIV-MS of *N*-acetylated MBP peptide (MW 1833 Da) with 2-NPG matrix using different laser power: (A) 4.2, (B) 5.7, and (C) 7.3 J cm^{-2} acquired using 'LSI settings'^{39,42} on a commercial intermediate pressure MALDI source of the SYNAPT G2 mass spectrometer.

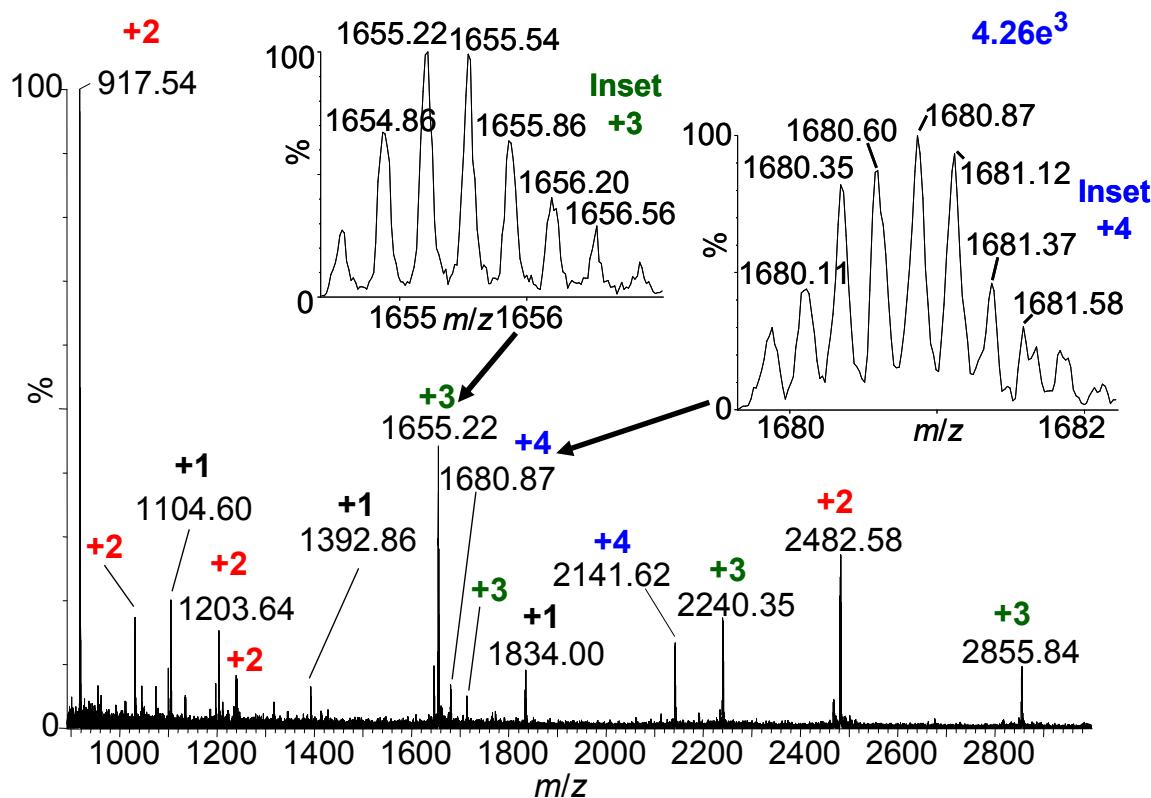


Figure S5. LSIV-IMS-MS of delipidified mouse brain tissue spotted with 2-NPG matrix shown in **Figure 2A**. Shown here is the total mass spectrum of the multiply charged peptide and protein ions with the large molecular weight detected, an ~8.6 kDa protein (m/z 2855.84) with insets of the isotopic distributions of higher charge states: +3 (m/z 1655.22) and +4 (m/z 1680.87) ions. Data acquired using ‘LSI settings’^{39,42} on a commercial intermediate pressure MALDI source of the SYNAPT G2 mass spectrometer.

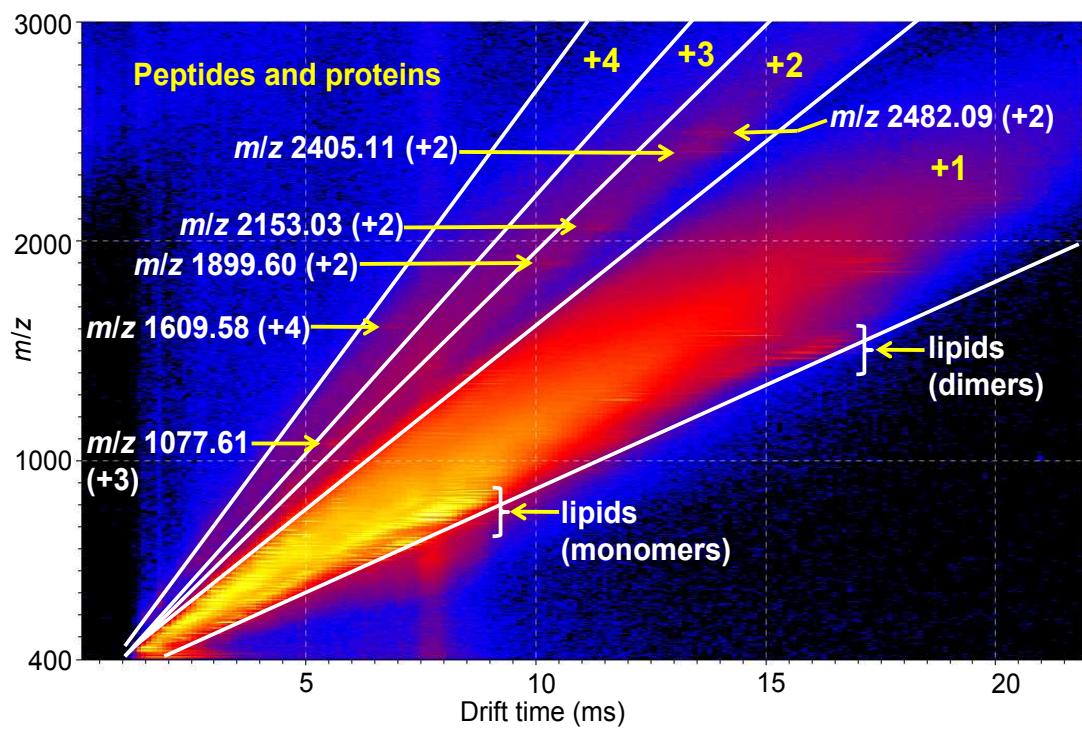


Figure S6. LSIV-IMS-MS two-dimensional plot of drift time vs. m/z of delipidified mouse brain tissue mounted on CHCA precoated glass plate spray coated with binary matrix mixture of 90% 2,5-DHAP and 10% 2-NPG and acquired using ‘LSI settings’^{39,42} on a commercial intermediate pressure MALDI source of SYNAPT G2 mass spectrometer.