# Asymmetric lodolactonization Utilizing Chiral Squaramides 

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## General Information

All commercially available reagents and solvents were used in the form they were supplied without any further purification. The stated yields are based on isolated material. The melting points are uncorrected. Thin layer chromatography was performed on silica gel $60 \mathrm{~F}_{254}$ aluminum-backed plates fabricated by Merck. Flash column chromatography was performed on silica gel $60(40-63 \mu \mathrm{~m})$ fabricated by Merck. NMR spectra were recorded on a Bruker AVII- 400 or a Bruker DPX- 300 spectrometer at 400 MHz or 300 MHz respectively for ${ }^{1} \mathrm{H}$ NMR and at 100 MHz or 75 MHz respectively for ${ }^{13} \mathrm{C}$ NMR. Coupling constants $(J)$ are reported in hertz and chemical shifts are reported in parts per million ( $\delta$ ) relative to the central residual protium solvent resonance in ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right.$ $=\delta 7.27$, DMSO- $d_{6}=\delta 2.50$ and TFA $\left.-d=\delta 11.50\right)$ and the central carbon solvent resonance in ${ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right.$ $=\delta 77.00 \mathrm{ppm}$ and DMSO- $d_{6}=\delta 39.43$ ). Mass spectra were recorded at 70 eV on Waters Prospec Q spectrometer using EI as the method of ionization. High resolution mass spectra were recorded on Waters Prospec Q spectrometer using EI as the method of ionization. Optical rotations were measured using a 1 mL cell with a 1.0 dm path length on a Perkin Elmer 341 polarimeter. Determination of enantiomeric excess was performed by HPLC on an Agilent Technologies 1200 Series instrument with diode array detector set at 254 nm and equipped with a chiral stationary phase (Chiralpak AD-H $5 \mu \mathrm{~m} 4.6 \times 250 \mathrm{~mm}$ ), applying the conditions stated. Alternatively, determination of enantiomeric excess was performed by GLC on a Varian 3380 instrument with split (1:30) injection, FID detector and equipped with a chiral stationary phase (Chiraldex G-TA $0.12 \mu \mathrm{~m} 0.25$ $\mathrm{mm} \times 30 \mathrm{~m}$ ), applying the conditions stated.

## Preparation of Squaramide Catalysts




11a, R =

11b,


Scheme S-1 Synthetic route to chiral squaramides 5a, 5b, 6a and 6b.
General procedure for the preparation of squaramide catalysts (Scheme S-1): Step 1. 3,4-Dimethoxycyclobut-3-ene-1,2-dione (9) (1.0 equiv.) was suspended/dissolved in either MeOH or $\mathrm{CH}_{2} \mathrm{Cl}_{2}(0.07 \mathrm{M})$ and the amine $\mathbf{1 0 a}, \mathbf{b}$ or 11a, $\mathbf{b}$ (1.1 equiv.) was added. The resulting mixture was stirred at ambient temperature for 48 hours. The mixture was then filtered; the collected solid residue was washed with ice-cold MeOH and dried in vacuo to afford the corresponding squaramate $\mathbf{1 2 a}, \mathbf{b}^{1}$ or 13a, $\mathbf{b}^{2,3}$. Step 2. 3-(Benzylamino)- or 3-(arylamino)-4-methoxycyclobut-3-ene-1,2-dione 12a, b or 13a, b ( 1.0 equiv.) was suspended in $\mathrm{MeOH}(0.07 \mathrm{M})$ and $(1 R, 2 R)$ $N^{1}, N^{1}$-dipentylcyclohexane-1,2-diamine (14) (1.0 equiv.) was added. The resulting mixture was stirred at ambient temperature for 24 hours. The mixture was then filtered; the collected solid residue was washed with ice-cold MeOH and dried in vacuo to afford the corresponding squaramide $\mathbf{5 a}, \mathbf{b}$ or $\mathbf{6 a}, \mathbf{b}$.

Notice! ( $1 R, 2 R$ )- $N^{1}, N^{1}$-dipentylcyclohexane-1,2-diamine (14) was prepared according to literature procedure ${ }^{4}$ from commercially available ( $1 R, 2 R$ )-cyclohexane-1,2-diamine (Aldrich; $98 \%$, optical purity $e e$ : $99 \%$ by GLC). The optical antipode, $(1 S, 2 S)$ - $N^{1}, N^{1}$-dipentylcyclohexane-1,2-diamine (ent-14), was prepared in the same manner from commercially available ( $1 S, 2 S$ )-cyclohexane-1,2-diamine D-tartrate (Aldrich; 99\%).

3-(((1R,2R)-2-(Dipentylamino)cyclohexyl)amino)-4-((4-(trifluoromethyl)benzyl)amino)cyclobut-3-ene-1,2dione (5a).


Prepared according to the general procedure by reacting 3-methoxy-4-((4-(trifluoromethyl)benzyl)amino)-cyclobut-3-ene-1,2-dione (12a) ( $0.250 \mathrm{~g}, 0.88 \mathrm{mmol}$ ) with ( $1 R, 2 R$ )- $N^{1}, N^{1}$-dipentylcyclohexane-1,2-diamine (14) $(0.223 \mathrm{~g}, 0.88 \mathrm{mmol})$ to afford the title compound $\mathbf{5 a}$ after recrystallization from pentane. Yield: $0.211 \mathrm{~g}(48 \%)$, colourless solid; M.p.: $204-205{ }^{\circ} \mathrm{C}$, decomp.; TLC (hexanes/EtOAc 1:1): $R_{\mathrm{f}}=0.06 ;[\alpha]_{D}^{20}=-634.9(\mathrm{c}=0.04$, $\mathrm{CHCl}_{3}$ ); ${ }^{1} \mathrm{H}$ NMR ( 400 MHz, DMSO- $d_{6}$ ) $\delta 7.85(\mathrm{bs}, 1 \mathrm{H}), 7.70(\mathrm{~d}, J=8.0 \mathrm{~Hz}, 2 \mathrm{H}), 7.53(\mathrm{~d}, J=8.0 \mathrm{~Hz}, 2 \mathrm{H}), 7.16$ (bs, 1H), $4.80(\mathrm{~s}, 2 \mathrm{H}), 4.00-3.57(\mathrm{~m}, 1 \mathrm{H}), 2.43-2.28(\mathrm{~m}, 3 \mathrm{H}), 2.28-2.16(\mathrm{~m}, 2 \mathrm{H}), 2.05-1.90(\mathrm{~m}, 1 \mathrm{H}), 1.84-$ $1.73(\mathrm{~m}, 1 \mathrm{H}), 1.73-1.53(\mathrm{~m}, 2 \mathrm{H}), 1.39-0.97(\mathrm{~m}, 16 \mathrm{H}), 0.76(\mathrm{t}, J=6.7 \mathrm{~Hz}, 6 \mathrm{H}){ }^{13}{ }^{3} \mathrm{C}$ NMR ( 101 MHz , DMSO$\left.d_{6}\right) \delta 182.6,181.9,168.1,166.9,144.0,128.2(2 \mathrm{C}), 124.4-126.4(\mathrm{~m}, 3 \mathrm{C}), 124.2\left(\mathrm{q},{ }^{1} J_{\mathrm{CF}}=272.0 \mathrm{~Hz}, 1 \mathrm{C}\right), 63.8$, 54.1, 49.4 (2C), 46.1, 34.6, 29.0 (2C), 28.5 (2C), 25.0, 24.6, 24.0, 22.1 (2C), 13.9 (2C); HRMS (EI): Exact mass calculated for $\mathrm{C}_{28} \mathrm{H}_{40} \mathrm{~F}_{3} \mathrm{~N}_{3} \mathrm{O}_{2}[M]^{+}: 507.3073$, found 507.3066.

3-((3,5-Bis(trifluoromethyl)benzyl)amino)-4-(((1R,2R)-2-(dipentylamino)cyclohexyl)amino)cyclobut-3-ene-1,2-dione (5b).


Prepared according to the general procedure by reacting 3 -((3,5-bis(trifluoromethyl)benzyl)amino)-4-methoxycyclobut-3-ene-1,2-dione (12b) $(1.050 \mathrm{~g}, 2.97 \mathrm{mmol})$ with ( $1 R, 2 R$ )- $N^{1}, N^{1}$-dipentylcyclohexane-1,2diamine (14) ( $0.756 \mathrm{~g}, 2.97 \mathrm{mmol})$ to afford the title compound $\mathbf{5 b}$. Yield: $1.045 \mathrm{~g}(85 \%)$, colourless solid; M.p.: $216-218{ }^{\circ} \mathrm{C}$, decomp.; TLC (hexanes/EtOAc 1:1): $R_{\mathrm{f}}=0.06 ;[\alpha]_{D}^{20}=-86.0\left(\mathrm{c}=0.05, \mathrm{CHCl}_{3}\right) ;{ }^{1} \mathrm{H}$ NMR (300 MHz, DMSO- $d_{6}$ ) $\delta 8.02(\mathrm{~s}, 3 \mathrm{H}), 7.87(\mathrm{bs}, 1 \mathrm{H}), 7.24(\mathrm{bs}, 1 \mathrm{H}), 5.08-4.67(\mathrm{~m}, 2 \mathrm{H}), 4.05-3.57(\mathrm{~m}, 1 \mathrm{H}), 2.41-$ $2.26(\mathrm{~m}, 3 \mathrm{H}), 2.26-2.12(\mathrm{~m}, 2 \mathrm{H}), 2.04-1.85(\mathrm{~m}, 1 \mathrm{H}), 1.83-1.72(\mathrm{~m}, 1 \mathrm{H}), 1.72-1.54(\mathrm{~m}, 2 \mathrm{H}), 1.41-0.87(\mathrm{~m}$, $16 \mathrm{H}), 0.70(\mathrm{t}, J=6.4 \mathrm{~Hz}, 6 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR ( 101 MHz, DMSO- $\left.d_{6}+\mathrm{DCl}\right) \delta$ 183.0, 182.4, 169.3, 167.9, 142.8, 130.9 $\left(\mathrm{q},{ }^{2} J_{\mathrm{CF}}=32.9 \mathrm{~Hz}, 2 \mathrm{C}\right), 129.0(2 \mathrm{C}), 123.8\left(\mathrm{q},{ }^{1} J_{\mathrm{CF}}=272.9 \mathrm{~Hz}, 2 \mathrm{C}\right), 121.6,64.1,52.4,52.0,51.3,46.3,34.4,28.8$, 28.7, 24.7, 24.3 (2C), 24.1, 24.0, 22.1, 21.9, 14.2, 14.0; HRMS (EI): Exact mass calculated for $\mathrm{C}_{29} \mathrm{H}_{39} \mathrm{~F}_{6} \mathrm{~N}_{3} \mathrm{O}_{2}$ $[M]^{+}: 575.2946$, found 575.2937.

3-(((1R,2R)-2-(Dipentylamino)cyclohexyl)amino)-4-((4-(trifluoromethyl)phenyl)amino)cyclobut-3-ene-1,2dione (6a).


Prepared according to the general procedure by reacting 3-methoxy-4-((4-(trifluoromethyl)phenyl)amino)-cyclobut-3-ene-1,2-dione (13a) ( $0.763 \mathrm{~g}, 2.81 \mathrm{mmol}$ ) with ( $1 R, 2 R$ )- $N^{1}, N^{1}$-dipentylcyclohexane-1,2-diamine (14) $(0.715 \mathrm{~g}, 2.81 \mathrm{mmol})$ to afford the title compound $\mathbf{6 a}$. Yield: $0.905 \mathrm{~g}(66 \%)$, colourless solid; M.p.: $200-201{ }^{\circ} \mathrm{C}$, decomp.; TLC (hexanes/EtOAc 1:1): $R_{\mathrm{f}}=0.10 ;[\alpha]_{D}^{20}=-141.7\left(\mathrm{c}=0.13, \mathrm{CHCl}_{3}\right) ;{ }^{1} \mathrm{H}$ NMR ( 300 MHz, TFA) $\delta$
7.59 (d, $J=8.2 \mathrm{~Hz}, 2 \mathrm{H}), 7.36$ (d, $J=8.2 \mathrm{~Hz}, 2 \mathrm{H}), 4.75-4.52(\mathrm{~m}, 1 \mathrm{H}), 3.76-3.45(\mathrm{~m}, 2 \mathrm{H}), 3.33$ (td, $J=12.6,4.3$ $\mathrm{Hz}, 1 \mathrm{H}), 3.14(\mathrm{td}, J=12.3,5.2 \mathrm{~Hz}, 1 \mathrm{H}), 2.89(\mathrm{td}, J=12.2,4.6 \mathrm{~Hz}, 1 \mathrm{H}), 2.36-2.21(\mathrm{~m}, 1 \mathrm{H}), 2.21-2.06(\mathrm{~m}, 1 \mathrm{H})$, $2.04-1.50(\mathrm{~m}, 8 \mathrm{H}), 1.50-1.35(\mathrm{~m}, 2 \mathrm{H}), 1.35-1.14(\mathrm{~m}, 8 \mathrm{H}), 0.94-0.68(\mathrm{~m}, 6 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR ( 101 MHz , DMSO$\left.d_{6}\right) \delta 184.5,179.6,169.7,162.0,142.8,126.6(2 \mathrm{C}), 124.5\left(\mathrm{q},{ }^{1} J_{\mathrm{CF}}=271.0 \mathrm{~Hz}\right), 122.1\left(\mathrm{q},{ }^{2} J_{\mathrm{CF}}=32.1 \mathrm{~Hz}\right), 117.6$ (2C), 64.1 (2C), 54.6, 49.2, 34.2, 29.2 (2C), 28.5 (2C), 24.9, 24.6, 23.6, 22.2 (2C), 14.0 (2C); HRMS (EI): Exact mass calculated for $\mathrm{C}_{27} \mathrm{H}_{38} \mathrm{~F}_{3} \mathrm{~N}_{3} \mathrm{O}_{2}[M]^{+}: 493.2916$, found 493.2921.

3-((3,5-Bis(trifluoromethyl)phenyl)amino)-4-(((1R,2R)-2-(dipentylamino)cyclohexyl)amino)cyclobut-3-ene-1,2-dione (6b).


Prepared according to the general procedure by reacting 3-((3,5-bis(trifluoromethyl)phenyl)amino)-4-methoxycyclobut-3-ene-1,2-dione (13b) ( $1.000 \mathrm{~g}, 2.94 \mathrm{mmol}$ ) with ( $1 R, 2 R$ )- $N^{1}, N^{1}$-dipentylcyclohexane-1,2diamine (14) ( $0.748 \mathrm{~g}, 2.94 \mathrm{mmol})$ to afford the title compound $\mathbf{6 b}$. Yield: $1.050 \mathrm{~g}(64 \%)$, colourless solid; M.p.: $195-196{ }^{\circ} \mathrm{C}$, decomp.; TLC (hexanes/EtOAc 1:1): $R_{\mathrm{f}}=0.12 ;[\alpha]_{D}^{20}=-107.6\left(\mathrm{c}=0.10, \mathrm{CHCl}_{3}\right) ;{ }^{1} \mathrm{H}$ NMR (300 $\mathrm{MHz}, \mathrm{TFA}) \delta 7.87(\mathrm{~s}, 2 \mathrm{H}), 7.79(\mathrm{~s}, 1 \mathrm{H}), 4.74(\mathrm{td}, J=11.2,4.2 \mathrm{~Hz}, 1 \mathrm{H}), 3.89-3.54(\mathrm{~m}, 2 \mathrm{H}), 3.44(\mathrm{td}, J=12.5$, $6.3 \mathrm{~Hz}, 1 \mathrm{H}), 3.25(\mathrm{td}, J=12.4,5.2 \mathrm{~Hz}, 1 \mathrm{H}), 2.99(\mathrm{td}, J=12.2,4.5 \mathrm{~Hz}, 1 \mathrm{H}), 2.47-2.32(\mathrm{~m}, 1 \mathrm{H}), 2.32-2.17(\mathrm{~m}$, $1 \mathrm{H}), 2.16-1.46(\mathrm{~m}, 10 \mathrm{H}), 1.46-1.26(\mathrm{~m}, 8 \mathrm{H}), 1.02-0.81(\mathrm{~m}, 6 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(101 \mathrm{MHz}\right.$, DMSO- $\left.d_{6}\right) \delta 184.5$, $179.7,170.0,161.4,141.3,131.4\left(\mathrm{q},{ }^{2} J_{\mathrm{CF}}=32.9,31.6 \mathrm{~Hz}, 2 \mathrm{C}\right), 123.1\left(\mathrm{q},{ }^{1} J_{\mathrm{CF}}=273.0 \mathrm{~Hz}, 2 \mathrm{C}\right), 117.5(2 \mathrm{C}), 114.3$, 64.2 (2C), 54.7, 49.2, 34.1, 29.2 (2C), 28.5 (2C), 24.8, 24.6, 23.7, 22.2 (2C), 13.9 (2C); HRMS (EI): Exact mass calculated for $\mathrm{C}_{28} \mathrm{H}_{37} \mathrm{~F}_{6} \mathrm{~N}_{3} \mathrm{O}_{2}[M]^{+}: 561.2790$, found 561.2799.

3-((3,5-Bis(trifluoromethyl)phenyl)amino)-4-(((1S,2S)-2-(dipentylamino)cyclohexyl)amino)cyclobut-3-ene-1,2-dione (ent-6b).


Prepared according to the general procedure by reacting 3-((3,5-bis(trifluoromethyl)phenyl)amino)-4-methoxycyclobut-3-ene-1,2-dione (13b) ( $0.240 \mathrm{~g}, 0.71 \mathrm{mmol}$ ) with ( $1 R, 2 R$ )- $N^{1}, N^{1}$-dipentylcyclohexane-1,2diamine (ent-14) $(0.180 \mathrm{~g}, 0.71 \mathrm{mmol})$ to afford the title compound ent-6b. Yield: $0.164 \mathrm{~g}(41 \%)$, colourless solid; TLC (hexanes/EtOAc 1:1): $R_{\mathrm{f}}=0.12 ;[\alpha]_{D}^{20}=110.6\left(\mathrm{c}=0.11, \mathrm{CHCl}_{3}\right)$.


Figure S-1 ${ }^{1} \mathrm{H}$-NMR spectrum of compound $\mathbf{5 a}$.


| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | ```M:/ Jørn/ NMR/ AVI400/ JET-176-017-35/ 11/ fid``` |
| 2 Title | JET-176-017-35 |
| 3 Comment | C13CPD DMSO \{D: $\backslash$ uio $\backslash$ AVII $400-05\}$ jornet 12 |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | spect |
| 8 Author |  |
| 9 Solvent | DMSO |
| 10 Temperature | 295.7 |
| 11 Pulse Sequence | zgpg30 |
| 12 Experiment | 1D |
| 13 Number of Scans | 2048 |
| 14 Receiver Gain | 1620 |
| 15 Relaxation Delay | 2.0000 |
| 16 Pulse Width | 7.0000 |
| 17 Acquisition Time | 1.2977 |
| 18 Acquisition Date | 2012-05-05T18:45:00 |
| 19 Modification Date | 2012-05-05T18:45:59 |
| $20 \begin{aligned} & \text { Spectrometer } \\ & \text { Frequency }\end{aligned}$ | 100.64 |
| 21 Spectral | 25252.5 |
| 22 Lowest Frequency | -1102.0 |
| 23 Nucleus | 13C |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |

Figure S-2 ${ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{5 a}$.


Figure $\mathbf{S}-\mathbf{3}^{1} \mathrm{H}$-NMR spectrum of compound $\mathbf{5 b}$.


Figure $\mathbf{S}-4{ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{5 b}$.


Figure $\mathbf{S - 5}{ }^{1} \mathrm{H}-\mathrm{NMR}$ spectrum of compound $\mathbf{6 a}$.


Figure S-6 ${ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{6 a}$.


Figure $S-7{ }^{1} \mathrm{H}-\mathrm{NMR}$ spectrum of compound $\mathbf{6 b}$.


| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | ```M:/ Jørn/ NMR/ AVI400/ JET-176-091-183/ 11/ fid``` |
| 2 Title | JET-176-091-183 |
| 3 Comment | C13CPD DMSO \{D: $\backslash$ uio $\backslash$ AVII400-05\} jornet 14 |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | spect |
| 8 Author |  |
| 9 Solvent | DMSO |
| 10 Temperature | 295.7 |
| 11 Pulse Sequence | zgpg30 |
| 12 Experiment | 1D |
| 13 Number of Scans | 2048 |
| 14 Receiver Gain | 2050 |
| 15 Relaxation Delay | 2.0000 |
| 16 Pulse Width | 7.0000 |
| 17 Acquisition Time | 1.2977 |
| 18 Acquisition Date | 2012-05-05T22:51:00 |
| 19 Modification Date | 2012-05-05T22:51:49 |
| 20 Spectrometer Frequency | 100.64 |
| 21 Spectral Width | 25252.5 |
| 22 Lowest Frequency | -1101.2 |
| 23 Nucleus | 13 C |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |

Figure S-8 ${ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{6 b}$.


Figure $\mathbf{S - 9}$ MS spectrum of compound $\mathbf{5 a}$.


Figure S-10 HRMS spectrum of compound 5a.


Figure S-11 MS spectrum of compound $\mathbf{5 b}$.


Figure S-12 HRMS spectrum of compound $\mathbf{5 b}$.


Figure S-13 MS spectrum of compound 6a.


Figure S-14 HRMS spectrum of compound $\mathbf{6 a}$.


Figure S-15 MS spectrum of compound $\mathbf{6 b}$.


Figure S-16 HRMS spectrum of compound $\mathbf{6 b}$.

## Preparation of Starting Materials

General procedure for the preparation of $\gamma$ - and $\delta$-unsaturated acids: Step 1. 4-Aryl-4-oxobutanoic acid (1.0 equiv.) or 5-aryl-5-oxopentanoic acid ( 1.0 equiv.) was suspended in MeOH ( 0.25 M ), the suspension was cooled to $0{ }^{\circ} \mathrm{C}$ and acetyl chloride ( 1.2 equiv.) was added. Cooling was discontinued and the resulting homogeneous mixture was stirred overnight at ambient temperature. The mixture was diluted with $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ and washed in succession with satd. aq. $\mathrm{NaHCO}_{3}$, brine and water. The organic phase was dried over $\mathrm{MgSO}_{4}$, filtered and the solvent was evaporated in vacuo to afford the corresponding 4-aryl-4-oxobutanoic acid methyl ester or 5-aryl-5oxopentanoic acid methyl ester. Step 2. Triphenylphosphonium bromide ( 1.3 equiv.) was suspended in THF ( 0.75 M) and cooled to $0{ }^{\circ} \mathrm{C}$, whereupon KHMDS ( 0.5 M in toluene, 1.3 equiv.) was added in one go. The resulting mixture was stirred for 1 hour then cooled to $-78^{\circ} \mathrm{C}$ and a solution of the 4 -aryl-4-oxobutanoic acid methyl ester or 5-aryl-5-oxopentanoic acid methyl ester ( 1.0 equiv.) in THF ( $\sim 0.25 \mathrm{M}$ ) was added in a dropwise manner. Cooling was discontinued and the resulting mixture was stirred at ambient temperature until TLC indicated full conversion of the starting material. The mixture was treated with satd. aq. $\mathrm{NH}_{4} \mathrm{Cl}$ and extracted with EtOAc. The combined organic extracts were dried over $\mathrm{MgSO}_{4}$, filtered and the solvent was evaporated in vacuo. The residue was purified by flash chromatography on silica (hexanes, followed by hexanes/EtOAc in various proportions) to afford the corresponding 4-aryl-4-oxobutanoic acid methyl ester or 5-arylhex-5-enoic acid methyl ester. Step 3. 4-Arylpent-4-enoic methyl ester or 5-arylhex-5-enoic acid methyl ester ( 1.0 equiv.) was dissolved in THF and water was added ( $8: 1 \mathrm{THF} /$ water, $\sim 0.10 \mathrm{M}$ ), followed by $\mathrm{LiOH}_{2} \mathrm{O}$ ( 1.5 equiv.). The resulting biphasic mixture was stirred vigorously overnight at ambient temperature, whereupon the solvent was evaporated in vacuo. The residue was dissolved in 1 M aq. NaOH and washed with $\mathrm{CH}_{2} \mathrm{Cl}_{2}$. The pH was adjusted to approx. 2 and the aq. phase was extracted twice with $\mathrm{CH}_{2} \mathrm{Cl}_{2}$. The combined organic extracts were dried over $\mathrm{MgSO}_{4}$, filtered and evaporated in vacuo to afford the corresponding 4-arylpent-4-enoic acid or 5-arylhex-5-enoic acid, respectively.

Alternatively, some of the $\gamma$ - and $\delta$-unsaturated acids were prepared directly from the corresponding 4-aryl-4oxobutanoic acids or 5-aryl-5-oxopentanoic acids according to the procedure of Takemiya et al. ${ }^{5}$

## 5-Phenylhex-5-enoic acid (1a). ${ }^{4}$



Prepared from commercially available 5-oxo-5-phenylpentanoic acid (15a) by a Wittig reaction according to the procedure of Takemiya et al. ${ }^{5}$ All physical data were in full agreement with those reported in the literature. ${ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 11.82(\mathrm{~s}, 1 \mathrm{H}), 7.15-7.63(\mathrm{~m}, 5 \mathrm{H}), 5.34(\mathrm{~s}, 1 \mathrm{H}), 5.11(\mathrm{~s}, 1 \mathrm{H}), 2.61(\mathrm{t}, J=7.5 \mathrm{~Hz}, 2 \mathrm{H})$, $2.41(\mathrm{t}, J=7.4 \mathrm{~Hz}, 2 \mathrm{H}), 1.83(\mathrm{p}, J=7.4 \mathrm{~Hz}, 2 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR ( $101 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 180.2,147.3,140.7,128.3$ (2C), 127.5, 126.1 (2C), 113.1, 34.4, 33.3, 23.0.

5-(Naphthalen-2-yl)hex-5-enoic acid (1b). ${ }^{4}$


Prepared according to the general procedure using 5-(naphthalen-2-yl)-5-oxopentanoic acid (15b). All physical data were in full agreement with those reported in the literature. ${ }^{1} \mathrm{H}$ NMR $\left(300 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 11.23(\mathrm{bs}, 1 \mathrm{H}), 7.77-$ $7.96(\mathrm{~m}, 4 \mathrm{H}), 7.61(\mathrm{dd}, J=8.6,1.8 \mathrm{~Hz}, 1 \mathrm{H}), 7.42-7.56(\mathrm{~m}, 2 \mathrm{H}), 5.51(\mathrm{~d}, J=1.3 \mathrm{~Hz}, 1 \mathrm{H}), 5.23(\mathrm{~d}, J=1.5 \mathrm{~Hz}, 1 \mathrm{H}), 2.73(\mathrm{t}, J$ $=7.5 \mathrm{~Hz}, 2 \mathrm{H}), 2.45(\mathrm{t}, J=7.4 \mathrm{~Hz}, 2 \mathrm{H}), 1.89(\mathrm{p}, J=7.4 \mathrm{~Hz}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C} \operatorname{NMR}\left(75 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 180.1,147.1,137.9,133.4$, $132.8,128.1,127.9,127.5,126.1,125.8,124.7,124.5,113.6,34.4,33.3,23.1$.


Prepared according to the general procedure using 5-oxo-5-( $p$-tolyl)pentanoic acid (15c). All physical data were in full agreement with those reported in the literature. ${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}\right.$, DMSO- $\left.d_{6}\right) \delta 7.34(\mathrm{~d}, J=8.0 \mathrm{~Hz}, 2 \mathrm{H})$, $7.15(\mathrm{~d}, J=7.9 \mathrm{~Hz}, 2 \mathrm{H}), 5.29(\mathrm{~d}, J=2.0 \mathrm{~Hz}, 1 \mathrm{H}), 5.01(\mathrm{~d}, J=1.9 \mathrm{~Hz}, 1 \mathrm{H}), 2.48(\mathrm{t}, J=7.4 \mathrm{~Hz}, 2 \mathrm{H}), 2.29(\mathrm{~s}, 3 \mathrm{H})$, $2.22(\mathrm{t}, J=7.4 \mathrm{~Hz}, 2 \mathrm{H}), 1.60(\mathrm{p}, J=7.4 \mathrm{~Hz}, 2 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR ( 101 MHz, DMSO- $d_{6}$ ) $\delta 174.3,147.0,137.2,136.8$, 129.0 (2C), 125.7 (2C), 111.8, 33.8, 33.0, 23.2, 20.7.

5-(4-Methoxyphenyl)hex-5-enoic acid (1d). ${ }^{4}$


Prepared according to the general procedure using 5-(4-methoxyphenyl)-5-oxopentanoic acid (15d). All physical data were in full agreement with those reported in the literature. ${ }^{1} \mathrm{H} \mathrm{NMR}\left(300 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 7.31-7.41(\mathrm{~m}, 2 \mathrm{H})$, $6.84-6.91(\mathrm{~m}, 2 \mathrm{H}), 5.25(\mathrm{~d}, J=1.4 \mathrm{~Hz}, 1 \mathrm{H}), 5.01(\mathrm{~d}, J=1.4 \mathrm{~Hz}, 1 \mathrm{H}), 3.82(\mathrm{~s}, 3 \mathrm{H}), 2.56(\mathrm{t}, J=7.4 \mathrm{~Hz}, 2 \mathrm{H}), 2.39(\mathrm{t}, J=7.4$ $\mathrm{Hz}, 2 \mathrm{H}), 1.81(\mathrm{p}, J=7.5 \mathrm{~Hz}, 2 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR ( $75 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 179.5,159.1,146.5,133.1,127.2$ (2C), 113.7 (2C), 111.5, 55.3, 34.5, 33.2, 23.1.

## 5-(4-Fluorophenyl)hex-5-enoic acid (1e). ${ }^{4}$



Prepared according to the general procedure using 5-(4-fluorophenyl)-5-oxopentanoic acid (15e). All physical data were in full agreement with those reported in the literature. ${ }^{1} \mathrm{H}$ NMR $\left(300 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 11.35(\mathrm{bs}, 1 \mathrm{H})$, $7.32-7.45(\mathrm{~m}, 2 \mathrm{H}), 6.97-7.08(\mathrm{~m}, 2 \mathrm{H}), 5.24-5.30(\mathrm{~m}, 1 \mathrm{H}), 5.04-5.11(\mathrm{~m}, 1 \mathrm{H}), 2.56(\mathrm{t}, J=7.5 \mathrm{~Hz}, 1 \mathrm{H}), 2.39$ $(\mathrm{t}, J=7.5 \mathrm{~Hz}, 1 \mathrm{H}), 1.80(\mathrm{p}, J=7.5 \mathrm{~Hz}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(75 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 180.0,162.3\left(\mathrm{~d},{ }^{1} J_{\mathrm{CF}}=246 \mathrm{~Hz}\right), 146.3$, $136.7\left(\mathrm{~d},{ }^{4} J_{\mathrm{CF}}=3.3 \mathrm{~Hz}\right), 127.7\left(\mathrm{~d},{ }^{3} J_{\mathrm{CF}}=7.9 \mathrm{~Hz}, 2 \mathrm{C}\right), 115.2\left(\mathrm{~d},{ }^{2} J_{\mathrm{CF}}=21.3 \mathrm{~Hz}, 2 \mathrm{C}\right), 113.0,34.5,33.2,22.9$.

## 5-(4-Chlorophenyl)hex-5-enoic acid (1f). ${ }^{4}$



Prepared from 5-(4-chlorophenyl)-5-oxopentanoic acid (15f) by a Wittig reaction according to the procedure of Takemiya et al. ${ }^{5}$ All physical data were in full agreement with those reported in the literature. ${ }^{1} \mathrm{H}$ NMR ( 400 $\left.\mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 9.76(\mathrm{bs}, 1 \mathrm{H}), 7.28-7.38(\mathrm{~m}, 4 \mathrm{H}), 5.31(\mathrm{~d}, J=1.3 \mathrm{~Hz}, 1 \mathrm{H}), 5.11(\mathrm{~d}, J=2.0 \mathrm{~Hz}, 1 \mathrm{H}), 2.55(\mathrm{t}, J=$ $7.5 \mathrm{~Hz}, 2 \mathrm{H}), 2.39(\mathrm{t}, J=7.4 \mathrm{~Hz}, 2 \mathrm{H}), 1.79(\mathrm{p}, J=7.4 \mathrm{~Hz}, 2 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR ( $101 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 179.6,146.2$, 139.1, 133.3, 128.5 (2C), 127.4 (2C), 113.6, 34.3, 33.2, 22.9.

## 5-(4-Bromophenyl)hex-5-enoic acid (1g). ${ }^{4}$



Prepared according to the general procedure using 5-(4-bromophenyl)-5-oxopentanoic acid (15g). All physical data were in full agreement with those reported in the literature. ${ }^{1} \mathrm{H}$ NMR $\left(300 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 11.44(\mathrm{bs}, 1 \mathrm{H})$, $7.42-7.52(\mathrm{~m}, 2 \mathrm{H}), 7.24-7.33(\mathrm{~m}, 2 \mathrm{H}), 5.33(\mathrm{~s}, 1 \mathrm{H}), 5.12(\mathrm{~s}, 1 \mathrm{H}), 2.56(\mathrm{t}, J=7.5 \mathrm{~Hz}, 2 \mathrm{H}), 2.40(\mathrm{t}, J=7.4 \mathrm{~Hz}$, 2 H ), 1.80 (p, $J=7.4 \mathrm{~Hz}, 2 \mathrm{H}$ ); 13C NMR ( $75 \mathrm{MHz}, \mathrm{CDCl} 3$ ) $\delta 180.0,146.2,139.5,131.4$ (2C), 127.7 (2C), 121.4, 113.6, 34.2, 33.2, 22.9.

6-Methyl-5-methyleneheptanoic acid (1h).4,5


Prepared from commercially available 6-methyl-5-methyleneheptanoic acid (15h) by a Wittig reaction according to the procedure of Takemiya et al. ${ }^{5}$ All physical data were in full agreement with those reported in the literature. ${ }^{1} \mathrm{H}$ NMR ( $300 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 11.09(\mathrm{bs}, 1 \mathrm{H}), 4.76-4.83(\mathrm{~m}, 1 \mathrm{H}), 4.67-4.74(\mathrm{~m}, 1 \mathrm{H}), 2.38(\mathrm{t}, J=7.5 \mathrm{~Hz}, 2 \mathrm{H})$, 2.24 (hept, $J=7.0 \mathrm{~Hz}, 1 \mathrm{H}), 2.09(\mathrm{t}, J=7.5 \mathrm{~Hz}, 2 \mathrm{H}), 1.80(\mathrm{p}, J=7.5 \mathrm{~Hz}, 2 \mathrm{H}), 1.03(\mathrm{~d}, J=7.0 \mathrm{~Hz}, 6 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(75 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 180.3,154.6,107.1,33.6$ (3C), 23.0, 21.8 (2C).

## 4-Phenylpent-4-enoic acid (3a). ${ }^{4}$



Prepared according to the general procedure using 4-oxo-4-phenylbutanoic acid (16a). All physical data were in full agreement with those reported in the literature. ${ }^{1} \mathrm{H}$ NMR $\left(300 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 10.56(\mathrm{bs}, 1 \mathrm{H}), 7.23-7.48$ (m, $5 \mathrm{H}), 5.35(\mathrm{~s}, 1 \mathrm{H}), 5.14(\mathrm{~s}, 1 \mathrm{H}), 2.88(\mathrm{t}, J=7.7 \mathrm{~Hz}, 2 \mathrm{H}), 2.56(\mathrm{t}, J=7.7 \mathrm{~Hz}, 2 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR ( $\left.75 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta$ $179.5,146.5,140.4,128.4$ (2C), 127.7, 126.1 (2C), 112.9, 33.0, 30.1.

## 4-(4-Chlorophenyl)pent-4-enoic acid (3b). ${ }^{6}$



Prepared according to the general procedure using 4-(4-chlorophenyl)-4-oxobutanoic acid (16b). All physical data were in full agreement with those reported in the literature. ${ }^{1} \mathrm{H}$ NMR ( $300 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 10.60(\mathrm{bs}, 1 \mathrm{H})$, $7.28-7.38(\mathrm{~m}, 4 \mathrm{H}), 5.32(\mathrm{~s}, 1 \mathrm{H}), 5.14(\mathrm{~s}, 1 \mathrm{H}), 2.83(\mathrm{t}, J=7.7 \mathrm{~Hz}, 1 \mathrm{H}), 2.54(\mathrm{t}, J=7.7 \mathrm{~Hz}, 2 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR ( 75 $\mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 179.3,145.4,138.8,133.5,128.6$ (2C), 127.4 (2C), 113.5, 32.8, 30.0.


Figure $\mathbf{S - 1 7}{ }^{1} \mathrm{H}-\mathrm{NMR}$ spectrum of compound $\mathbf{1 a}$.


| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | M:/ Jørn/ NMR/ AVI400/ JET-176-016-33/ 11/ fid |
| 2 Title | JET-176-016-33 |
| 3 Comment | C13CPD CDC13 \{D: $\backslash$ uio $\backslash$ AVII400-05\} jornet 15 |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | spect |
| 8 Author |  |
| 9 Solvent | CDC13 |
| 10 Temperature | 295.8 |
| 11 Pulse Sequence | zgpg30 |
| 12 Experiment | 1D |
| 13 Number of Scans | 1024 |
| 14 Receiver Gain | 812 |
| 15 Relaxation Delay | 2.0000 |
| 16 Pulse Width | 7.0000 |
| 17 Acquisition Time | 1.2977 |
| 18 Acquisition Date | 2012-05-05T23:58:00 |
| 19 Modification Date | 2012-05-05T23:58:21 |
| 20 Spectrometer Frequency | 100.64 |
| 21 Spectral | 25252.5 |
| 22 Lowest Frequency | -1060.9 |
| 23 Nucleus | 13 C |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |

Figure S-18 ${ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{1 a}$.


Figure S-19 ${ }^{1} \mathrm{H}$-NMR spectrum of compound $\mathbf{1 b}$.


| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | M:/ NMR/ JNPD-142/ <br> 11/fid |
| 2 Title | JNPD-142/ 11 |
| 3 Comment | ```5-(Napht-2-yl)hex-5- enoic acid C13CPD CDCI3 {D:\ uio\ DPX300-09} jmnolsoe 43``` |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | DPX300 |
| 8 Author |  |
| 9 Solvent | CDCI3 |
| 10 Temperature | 298.2 |
| 11 Pulse Sequence | zgpg30 |
| 12 Experiment | 1D |
| 13 Number of Scans | 1024 |
| 14 Receiver Gain | 10321 |
| 15 Relaxation Delay | 2.0000 |
| 16 Pulse Width | 5.8000 |
| 17 Acquisition Time | 1.8219 |
| 18 Acquisition Date | 2012-05-13T21:53:49 |
| 19 Modification Date | 2012-05-13T21:53:53 |
| 20 Spectrometer Frequency | 75.48 |
| 21 Spectral Width | 17985.6 |
| 22 Lowest Frequency | -1452.0 |
| 23 Nucleus | 13 C |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |

Figure $\mathbf{S - 2 0}{ }^{13} \mathbf{C}$-NMR spectrum of compound $\mathbf{1 b}$.


| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | M:/ Jørn/ NMR/ <br> AVI400/ <br> JET-222-107-25/ 10/ <br> fid |
| 2 Title | JET-222-107-25 |
| 3 Comment | PROTON CDCI3 \{D: $\backslash$ uio \( |
| ) AVII400-05\} jornet 18 |  |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | spect |
| 8 Author |  |
| 9 Solvent | CDCI3 |
| 10 Temperature | 294.8 |
| 11 Pulse Sequence | zg30 |
| 12 Experiment | 1D |
| 13 Number of Scans | 16 |
| 14 Receiver Gain | 81 |
| 15 Relaxation Delay | 1.0000 |
| 16 Pulse Width | 12.1500 |
| 17 Acquisition Time | 3.9846 |
| 18 Acquisition Date | 2012-05-06T04:11:00 |
| 19 Modification Date | 2012-05-06T04:11:39 |
| 20 Spectrometer Frequency | 400.18 |
| 21 Spectral Width | 8223.7 |
| 22 Lowest Frequency | -3541.8 |
| 23 Nucleus | 1H |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |

Figure S-21 ${ }^{1} \mathrm{H}$-NMR spectrum of compound $\mathbf{1 c}$.


| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | M:/ Jørn/ NMR/ <br> AVI400/ <br> JET-222-107-25/ 11/ <br> fid |
| 2 Title | JET-222-107-25 |
| 3 Comment | C13CPD CDC13 \{D: $\backslash$ uio AVII400-05\} jornet 18 |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | spect |
| 8 Author |  |
| 9 Solvent | CDC13 |
| 10 Temperature | 295.7 |
| 11 Pulse Sequence | zgpg30 |
| 12 Experiment | 1D |
| 13 Number of Scans | 2048 |
| 14 Receiver Gain | 724 |
| 15 Relaxation Delay | 2.0000 |
| 16 Pulse Width | 7.0000 |
| 17 Acquisition Time | 1.2977 |
| 18 Acquisition Date | 2012-05-06T06:08:00 |
| 19 Modification Date | 2012-05-06T06:08:29 |
| 20 Spectrometer Frequency | 100.64 |
| 21 Spectral Width | 25252.5 |
| 22 Lowest Frequency | -1576.6 |
| 23 Nucleus | 13 C |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |

Figure $\mathbf{S}-\mathbf{2 2}{ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{1 c}$.


Figure S-23 ${ }^{1} \mathrm{H}$-NMR spectrum of compound 1 d .


| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | M:/ NMR/ JNPD-185/ 21/fid |
| 2 Title | JNPD-185/ 21 |
| 3 Comment | Hydrolysis of $p$ MeOPh d-unsaturated ester. 2 h C13CPD CDCI3 \{D: $\backslash$ uio DPX300-09\} jmnolsoe 57 |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | DPX300 |
| 8 Author |  |
| 9 Solvent | CDCI3 |
| 10 Temperature | 298.2 |
| 11 Pulse Sequence | zgpg30 |
| 12 Experiment | 1D |
| 13 Number of Scans | 1024 |
| 14 Receiver Gain | 13004 |
| 15 Relaxation Delay | 2.0000 |
| 16 Pulse Width | 5.8000 |
| 17 Acquisition Time | 1.8219 |
| 18 Acquisition Date | 2012-09-01T21:38:30 |
| 19 Modification Date | 2012-09-01T21:38:33 |
| 20 Spectrometer Frequency | 75.48 |
| 21 Spectral Width | 17985.6 |
| 22 Lowest Frequency | -1448.2 |
| 23 Nucleus | 13C |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |

Figure $\mathbf{S}-\mathbf{2 4}{ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{1 d}$.


Figure S-25 ${ }^{1} \mathrm{H}$-NMR spectrum of compound $\mathbf{1 e}$.


Figure S-26 ${ }^{13}$ C-NMR spectrum of compound $\mathbf{1 e}$.

| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | M:/ NMR/ JNPD-140/ 10 fid |
| 2 Title | JNPD-140/ 10 |
| 3 Comment | F-Ketoacid PROTON CDCI3 \{D: $\backslash$ uio DPX300-09\} jmnolsoe 53 |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | DPX300 |
| 8 Author |  |
| 9 Solvent | CDC13 |
| 10 Temperature | 298.2 |
| 11 Pulse Sequence | zg30 |
| 12 Experiment | 1D |
| 13 Number of Scans | 16 |
| 14 Receiver Gain | 181 |
| 15 Relaxation Delay | 1.0000 |
| 16 Pulse Width | 10.0000 |
| 17 Acquisition Time | 5.3084 |
| 18 Acquisition Date | 2012-05-10T11:33:59 |
| 19 Modification Date | 2012-05-10T11:34:03 |
| 20 Spectrometer Frequency | 300.13 |
| 21 Spectral Width | 6172.8 |
| 22 Lowest Frequency | -1236.0 |
| 23 Nucleus | 1 H |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |


| Parameter | Value |
| :---: | :---: |
| $\begin{array}{ll} 1 & \begin{array}{l} \text { Data File } \\ \text { Name } \end{array} \end{array}$ | M:/ NMR/ JNPD-140/ <br> 11/fid |
| 2 Title | JNPD-140/ 11 |
| 3 Comment | F-Ketoacid C13CPD CDCl3 \{D: $\backslash$ uio $\backslash$ DPX300-09\} jmnolsoe 53 |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | DPX300 |
| 8 Author |  |
| 9 Solvent | CDCl3 |
| 10 Temperature | 298.2 |
| 11 Pulse Sequence | zgpg30 |
| 12 Experiment | 1D |
| 13 Number of Scans | 1024 |
| 14 Receiver Gain | 11585 |
| 15 Relaxation Delay | 2.0000 |
| 16 Pulse Width | 5.8000 |
| 17 Acquisition Time | 1.8219 |
| 18 Acquisition Date | 2012-05-10T12:41:22 |
| 19 Modification Date | 2012-05-10T12:41:25 |
| 20 Spectrometer Frequency | 75.48 |
| 21 Spectral | 17985.6 |
| 22 Lowest Frequency | -1448.2 |
| 23 Nucleus | 13C |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |



Figure S-27 ${ }^{1} \mathrm{H}$-NMR spectrum of compound $\mathbf{1 f}$.


| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | M:/ Jørn/ NMR/ <br> AVI400/ <br> JET-222-104-19/ 11/ <br> fid |
| 2 Title | JET-222-104-19 |
| 3 Comment | C13CPD CDCI3 \{D: $\backslash$ uio \( |
| ) AVII400-05\} jornet 16 |  |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | spect |
| 8 Author |  |
| 9 Solvent | CDC13 |
| 10 Temperature | 295.7 |
| 11 Pulse Sequence | zgpg30 |
| 12 Experiment | 1D |
| 13 Number of Scans | 2048 |
| 14 Receiver Gain | 1620 |
| 15 Relaxation Delay | 2.0000 |
| 16 Pulse Width | 7.0000 |
| 17 Acquisition Time | 1.2977 |
| 18 Acquisition Date | 2012-05-06T02:02:00 |
| 19 Modification Date | 2012-05-06T02:02:12 |
| 20 Spectrometer Frequency | 100.64 |
| 21 Spectral | 25252.5 |
| 22 Lowest Frequency | -1057.2 |
| 23 Nucleus | 13C |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |

Figure S-28 ${ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{1 f}$.


Figure S-29 ${ }^{1} \mathrm{H}-\mathrm{NMR}$ spectrum of compound $\mathbf{1 g}$.


Figure $\mathbf{S}-\mathbf{3 0}{ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{1 g}$.


Figure S-31 ${ }^{1} \mathrm{H}-\mathrm{NMR}$ spectrum of compound $\mathbf{1 h}$.


| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | M:/ NMR/ JNPD-154/ 11/fid |
| 2 Title | JNPD-154/ 11 |
| 3 Comment | i-Pr d-lactone pre. Acid C13CPD CDCI3 \{D: $\backslash$ uio $\mathrm{DPX} 300-09\}$ jmnolsoe 38 |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | DPX300 |
| 8 Author |  |
| 9 Solvent | CDC13 |
| 10 Temperature | 298.2 |
| 11 Pulse Sequence | zgpg30 |
| 12 Experiment | 1D |
| 13 Number of Scans | 1024 |
| 14 Receiver Gain | 10321 |
| 15 Relaxation Delay | 2.0000 |
| 16 Pulse Width | 5.8000 |
| 17 Acquisition Time | 1.8219 |
| 18 Acquisition Date | 2012-06-06T15:50:10 |
| 19 Modification Date | 2012-06-06T15:50:14 |
| 20 Spectrometer Frequency | 75.48 |
| 21 Spectral Width | 17985.6 |
| 22 Lowest Frequency | -1446.8 |
| 23 Nucleus | 13 C |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |

Figure $\mathbf{S - 3 2}{ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{1 h}$.


Figure S-33 ${ }^{1} \mathrm{H}-\mathrm{NMR}$ spectrum of compound $\mathbf{3 a}$.


Figure $\mathbf{S - 3 4}{ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{3 a}$.


Figure S-35 ${ }^{1} \mathrm{H}$-NMR spectrum of compound $\mathbf{3 b}$.


Figure $\mathbf{S}-\mathbf{3 6}{ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{3 b}$.

## Enantioselective Iodolactonization Utilizing Chiral Squaramides

General procedure for asymmetric iodolactonization with chiral squaramides: Iodine ( 0.15 equiv.) and $N$ iodosuccinimide ( 1.0 equiv.) was dissolved in a combination of acetone/ $\mathrm{CH}_{2} \mathrm{Cl}_{2}(1: 1,0.20 \mathrm{M})$. Subsequently, the squaramide 5-8 ( 0.15 equiv.) was added and the resulting mixture was cooled to $-78^{\circ} \mathrm{C}$. A solution of unsaturated acid 1 or $\mathbf{3}$ ( 1.0 equiv.) in a combination of acetone/ $\mathrm{CH}_{2} \mathrm{Cl}_{2}(1: 1,0.20 \mathrm{M})$ was added and the resulting mixture was stirred at $-78^{\circ} \mathrm{C}$ for 24 hours. The reaction mixture was treated with satd. aq. $\mathrm{Na}_{2} \mathrm{~S}_{2} \mathrm{O}_{3}(10 \mathrm{ml})$ while still in the cold, allowed to equilibrate to ambient temperature and then $\mathrm{EtOAc}(30 \mathrm{ml})$ was added. The phases were separated and the organic phase was washed with aq. $\mathrm{NaOH}(2 \times 20 \mathrm{ml}, 1.0 \mathrm{M})$ and brine ( 20 ml ). The organic phase was dried over $\mathrm{MgSO}_{4}$, filtered and evaporated in vacuo. The residue was purified by column chromatography on silica (hexanes/EtOAc 4:1) to afford the corresponding iodolactone 2 or 4.

Notice! The iodolactones were observed to be very labile in the condensed state under vacuum. Thus, to avoid decomposition, great care had to be taken when evaporating the solvent in vacuo after isolation by flash chromatography. Once all visible traces of solvent had been removed it was of paramount importance to equilibrate back to ambient pressure immediately. Without this precaution, the isolated iodolactone would turn black spontaneously. Due to the instability of the iodolactones they were stored under argon and refrigerated.

## (S)-6-(Iodomethyl)-6-phenyltetrahydro-2H-pyran-2-one (2a). ${ }^{4}$



Prepared according to the general procedure using 5-phenylhex-5-enoic acid (1a) ( $40 \mathrm{mg}, 210 \mu \mathrm{~mol}$ ), squaramide $\mathbf{6 b}(17 \mathrm{mg}, 31 \mu \mathrm{~mol})$, iodine ( $8 \mathrm{mg}, 31 \mu \mathrm{~mol}$ ) and $N$-iodosuccinimide ( $47 \mathrm{mg}, 210 \mu \mathrm{~mol}$ ). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes $/$ i $\operatorname{PrOH} 98: 2,1 \mathrm{~mL} / \mathrm{min}): t_{r}\left(e_{1}\right.$, major) $=24.04 \mathrm{~min}$ and $t_{r}\left(e_{2}\right.$, minor $)=27.20$ min. Yield: 55 mg ( $83 \%$ ) of colourless oil; ee: $87 \%$; TLC (hexanes/EtOAc 1:1): $R_{\mathrm{f}}=0.48$, visualized with anisaldehyde; $[\alpha]_{D}^{20}=27.2\left(\mathrm{c}=0.06, \mathrm{CHCl}_{3}\right) ;{ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 7.30-7.54(\mathrm{~m}, 5 \mathrm{H}), 3.57(\mathrm{~d}, J=1.4$ $\mathrm{Hz}, 2 \mathrm{H}), 2.28-2.59(\mathrm{~m}, 4 \mathrm{H}), 1.69-1.98(\mathrm{~m}, 1 \mathrm{H}), 1.45-1.69(\mathrm{~m}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR ( $\left.101 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 170.4$, 140.1, 128.9 (2C), 128.3, 125.1 (2C), 84.3, 32.0, 28.9, 17.7, 16.5; HRMS (EI): Exact mass calculated for $\mathrm{C}_{12} \mathrm{H}_{13} \mathrm{IO}_{2}[M]^{+}: 315.9960$, found 315.9972.

## (R)-6-(Iodomethyl)-6-phenyltetrahydro-2H-pyran-2-one (ent-2a). ${ }^{4}$



Prepared according to the general procedure using 5-phenylhex-5-enoic acid (1a) ( $40 \mathrm{mg}, 210 \mu \mathrm{~mol}$ ), squaramide ent- $\mathbf{6 b}(17 \mathrm{mg}, 31 \mu \mathrm{~mol}$ ), iodine ( $8 \mathrm{mg}, 31 \mu \mathrm{~mol}$ ) and $N$-iodosuccinimide ( $47 \mathrm{mg}, 210 \mu \mathrm{~mol}$ ). Purified by column chromatography on silica (hexanes/EtOAc $4: 1$ ). The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes $/$ PrOH 98:2, $1 \mathrm{~mL} / \mathrm{min}$ ): $t_{r}\left(e_{1}\right.$, minor $)=27.00 \mathrm{~min}$ and $t_{r}\left(e_{2}\right.$, major) $=31.00$ min . Yield: 52 mg ( $79 \%$ ) of colourless oil; ee: $82 \%$; TLC (hexanes/EtOAc 1:1): $R_{\mathrm{f}}=0.48$, visualized with anisaldehyde; $[\alpha]_{D}^{20}=-26.0\left(\mathrm{c}=0.07, \mathrm{CHCl}_{3}\right) ;{ }^{1} \mathrm{H} \operatorname{NMR}\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 7.30-7.54(\mathrm{~m}, 5 \mathrm{H}), 3.57(\mathrm{~d}, J=$ $1.4 \mathrm{~Hz}, 2 \mathrm{H}), 2.28-2.59(\mathrm{~m}, 4 \mathrm{H}), 1.69-1.98(\mathrm{~m}, 1 \mathrm{H}), 1.45-1.69(\mathrm{~m}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C} \operatorname{NMR}\left(101 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 170.4$, 140.1, 128.9 (2C), 128.3, 125.1 (2C), 84.3, 32.0, 28.9, 17.7, 16.5.
(S)-6-(Iodomethyl)-6-(naphthalen-2-yl)tetrahydro-2H-pyran-2-one (2b). ${ }^{4}$


Prepared according to the general procedure using 5-(naphthalen-2-yl)hex-5-enoic acid (1b) ( $40 \mathrm{mg}, 166 \mu \mathrm{~mol}$ ), squaramide $\mathbf{6 b}(14 \mathrm{mg}, 25 \mu \mathrm{~mol})$, iodine ( $7 \mathrm{mg}, 25 \mu \mathrm{~mol}$ ) and $N$-iodosuccinimide ( $37 \mathrm{mg}, 166 \mu \mathrm{~mol}$ ). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes $\left./{ }^{i} \operatorname{PrOH} 99: 1,1 \mathrm{~mL} / \mathrm{min}\right): t_{r}\left(e_{1}\right.$, major $)=66.14 \mathrm{~min}$ and $t_{r}\left(e_{2}\right.$, minor $)=82.74$ $\min$. Yield: $55 \mathrm{mg}(91 \%)$ of colourless oil; ee: $92 \%$; TLC (hexanes/EtOAc 1:1): $R_{\mathrm{f}}=0.56$, visualized with anisaldehyde; $[\alpha]_{D}^{20}=22.0\left(\mathrm{c}=0.13, \mathrm{CHCl}_{3}\right) ;{ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 7.81-7.97(\mathrm{~m}, 4 \mathrm{H}), 7.48-7.62(\mathrm{~m}$, $2 \mathrm{H}), 7.41$ (dd, $J=8.6,2.0 \mathrm{~Hz}, 1 \mathrm{H}), 3.67$ (d, $J=2.8 \mathrm{~Hz}, 2 \mathrm{H}$ ), $2.29-2.77$ (m, 4H), 1.78-1.93 (m, 1H), $1.52-1.69$ $(\mathrm{m}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(101 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 170.5,137.4,133.0,132.8,129.0,128.3,127.5,126.8$ (2C), 125.0, 122.3, 84.6, 32.1, 29.0, 17.4, 16.6; HRMS (EI): Exact mass calculated for $\mathrm{C}_{16} \mathrm{H}_{15} \mathrm{IO}_{2}[M]^{+}: 366.0117$, found 366.0109.

## (S)-6-(Iodomethyl)-6-(p-tolyl)tetrahydro-2H-pyran-2-one (2c). ${ }^{4}$



Prepared according to the general procedure using 5 -( $p$-tolyl)hex-5-enoic acid (1c) ( $40 \mathrm{mg}, 196 \mu \mathrm{~mol}$ ), squaramide $\mathbf{6 b}(17 \mathrm{mg}, 29 \mu \mathrm{~mol})$, iodine ( $7 \mathrm{mg}, 29 \mu \mathrm{~mol}$ ) and $N$-iodosuccinimide ( $44 \mathrm{mg}, 196 \mu \mathrm{~mol}$ ). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes $/ i \operatorname{PrOH} 98: 2,1 \mathrm{~mL} / \mathrm{min}): t_{r}\left(e_{1}\right.$, major $)=25.32 \mathrm{~min}$ and $t_{r}\left(e_{2}\right.$, minor $)=34.16$ min . Yield: 51 mg ( $80 \%$ ) of colourless oil; ee: $86 \%$; TLC (hexanes/EtOAc 1:1): $R_{\mathrm{f}}=0.56$, visualized with anisaldehyde; $[\alpha]_{D}^{20}=24.6\left(\mathrm{c}=0.10, \mathrm{CHCl}_{3}\right) ;{ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 7.25(\mathrm{~d}, J=8.4 \mathrm{~Hz}, 2 \mathrm{H}), 7.20(\mathrm{~d}, J=$ $8.2 \mathrm{~Hz}, 2 \mathrm{H}), 3.55(\mathrm{~s}, 2 \mathrm{H}), 2.40-2.54(\mathrm{~m}, 2 \mathrm{H}), 2.35(\mathrm{~s}, 3 \mathrm{H}), 2.27-2.40(\mathrm{~m}, 2 \mathrm{H}), 1.74-1.89(\mathrm{~m}, 1 \mathrm{H}), 1.66-1.49$ $(\mathrm{m}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR ( $101 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 170.5,138.2,137.1,129.6$ (2C), 125.1 (2C), 84.3, 31.9, 28.9, 21.0, 17.9, 16.5; HRMS (EI): Exact mass calculated for $\mathrm{C}_{13} \mathrm{H}_{15} \mathrm{IO}_{2}[M]^{+}: 330.0117$, found 330.0108 .

## (S)-6-(Iodomethyl)-6-(4-methoxyphenyl)tetrahydro-2H-pyran-2-one (2d). ${ }^{4}$



Prepared according to the general procedure using 5-(4-methoxyphenyl)hex-5-enoic acid (1d) ( $48 \mathrm{mg}, 218 \mu \mathrm{~mol}$ ), squaramide $\mathbf{6 b}(18 \mathrm{mg}, 33 \mu \mathrm{~mol})$, iodine ( $8 \mathrm{mg}, 33 \mu \mathrm{~mol}$ ) and $N$-iodosuccinimide ( $49 \mathrm{mg}, 218 \mu \mathrm{~mol}$ ). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes $/$ PrOH 90:10, $1 \mathrm{~mL} / \mathrm{min}$ ): $t_{r}\left(e_{1}\right.$, major) $=12.91$ min and $t_{r}\left(e_{2}\right.$, minor) $=15.56$ min . Yield: 65 mg ( $87 \%$ ) of colourless oil; ee: $12 \%$; TLC (hexanes/EtOAc 1:1): $R_{\mathrm{f}}=0.53$, visualized with anisaldehyde; $[\alpha]_{D}^{20}=1.6\left(\mathrm{c}=0.06, \mathrm{CHCl}_{3}\right) ;{ }^{1} \mathrm{H}$ NMR $\left(300 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 7.24-7.33(\mathrm{~m}, 2 \mathrm{H}), 6.86-6.95(\mathrm{~m}$, $2 \mathrm{H}), 3.81(\mathrm{~s}, 3 \mathrm{H}), 3.50-3.58(\mathrm{~m}, 2 \mathrm{H}), 2.39-2.55(\mathrm{~m}, 2 \mathrm{H}), 2.25-2.39(\mathrm{~m}, 2 \mathrm{H}), 1.75-1.90(\mathrm{~m}, 1 \mathrm{H}), 1.48-1.70$
(m, 1H); ${ }^{13} \mathrm{C}$ NMR ( $75 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 170.5,159.4,132.0,126.5$ (2C), 114.2 (2C), 84.2, 55.3, 31.8, 28.9, 18.1, 16.5; HRMS (EI): Exact mass calculated for $\mathrm{C}_{13} \mathrm{H}_{15} \mathrm{IO}_{3}[M]^{+}: 346.0066$, found 346.0064 .
(S)-6-(4-Fluorophenyl)-6-(iodomethyl)tetrahydro-2H-pyran-2-one (2e). ${ }^{4}$


Prepared according to the general procedure using 5-(4-fluorophenyl)hex-5-enoic acid (1e) ( $42 \mathrm{mg}, 200 \mu \mathrm{~mol}$ ), squaramide $\mathbf{6 b}$ ( $17 \mathrm{mg}, 30 \mu \mathrm{~mol}$ ), iodine ( $8 \mathrm{mg}, 30 \mu \mathrm{~mol}$ ) and $N$-iodosuccinimide ( $45 \mathrm{mg}, 200 \mu \mathrm{~mol}$ ). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes $/ 2 \operatorname{PrOH} 90: 10,1 \mathrm{~mL} / \mathrm{min}): t_{r}\left(e_{1}\right.$, major $)=10.70 \mathrm{~min}$ and $t_{r}\left(e_{2}\right.$, minor $)=12.17$ min . Yield: $55 \mathrm{mg}(83 \%)$ of colourless oil; ee: $90 \%$; TLC (hexanes/EtOAc 1:1): $R_{\mathrm{f}}=0.49$, visualized with anisaldehyde; $[\alpha]_{D}^{20}=26.0\left(\mathrm{c}=0.10, \mathrm{CHCl}_{3}\right) ;{ }^{1} \mathrm{H}$ NMR $\left(300 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 7.29-7.47(\mathrm{~m}, 2 \mathrm{H}), 7.02-7.17(\mathrm{~m}$, $2 \mathrm{H}), 3.54(\mathrm{~s}, 1 \mathrm{H}), 2.41-2.58(\mathrm{~m}, 2 \mathrm{H}), 2.41-2.26(\mathrm{~m}, 2 \mathrm{H}), 1.76-1.93(\mathrm{~m}, 1 \mathrm{H}), 1.48-1.71(\mathrm{~m}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(101 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 170.1,162.4\left(\mathrm{~d},{ }^{1} J_{\mathrm{CF}}=248 \mathrm{~Hz}\right), 136.0\left(\mathrm{~d},{ }^{4} J_{\mathrm{CF}}=3.2 \mathrm{~Hz}\right), 127.1\left(\mathrm{~d},{ }^{3} J_{\mathrm{CF}}=8.3 \mathrm{~Hz}, 2 \mathrm{C}\right), 115.9$ $\left(\mathrm{d},{ }^{2} J_{\mathrm{CF}}=21.5 \mathrm{~Hz}, 2 \mathrm{C}\right), 84.1,31.9,28.9,17.5,16.5$; HRMS (EI): Exact mass calculated for $\mathrm{C}_{12} \mathrm{H}_{12} \mathrm{FIO} \mathrm{O}_{2}[M]^{+}$: 333.9866, found 333.9867 .
(S)-6-(4-Chlorophenyl)-6-(iodomethyl)tetrahydro-2H-pyran-2-one (2f). ${ }^{4}$


Prepared according to the general procedure using 5-(4-chlorophenyl)hex-5-enoic acid (1f) ( $40 \mathrm{mg}, 178 \mu \mathrm{~mol}$ ), squaramide $\mathbf{6 b}(15 \mathrm{mg}, 27 \mu \mathrm{~mol})$, iodine ( $7 \mathrm{mg}, 27 \mu \mathrm{~mol}$ ) and $N$-iodosuccinimide ( $40 \mathrm{mg}, 178 \mu \mathrm{~mol}$ ). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes $\left./{ }^{i} \operatorname{PrOH} 98: 2,1 \mathrm{~mL} / \mathrm{min}\right): t_{r}\left(e_{1}\right.$, major $)=31.58 \mathrm{~min}$ and $t_{r}\left(e_{2}\right.$, minor $)=46.09$ $\min$. Yield: $49 \mathrm{mg}(78 \%)$ of colourless oil; ee: $96 \%$; TLC (hexanes/EtOAc 1:1): $R_{\mathrm{f}}=0.46$, visualized with anisaldehyde; $[\alpha]_{D}^{20}=27.2\left(\mathrm{c}=0.20, \mathrm{CHCl}_{3}\right) ;{ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 7.38(\mathrm{~d}, J=8.8 \mathrm{~Hz}, 2 \mathrm{H}), 7.32(\mathrm{~d}, J=$ $8.8 \mathrm{~Hz}, 2 \mathrm{H}), 2.42-2.59(\mathrm{~m}, 2 \mathrm{H}), 2.27-2.41(\mathrm{~m}, 2 \mathrm{H}), 1.78-1.93(\mathrm{~m}, 1 \mathrm{H}), 1.50-1.66(\mathrm{~m}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR (101 $\mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 170.0,138.8,134.5,129.1$ (2C), 126.7 (2C), 84.1, 32.0, 29.0, 17.1, 16.5; HRMS (EI): Exact mass calculated for $\mathrm{C}_{12} \mathrm{H}_{12} \mathrm{ClIO}_{2}[M]^{+}: 349.9571$, found 349.9577.

## (S)-6-(4-Bromophenyl)-6-(iodomethyl)tetrahydro-2H-pyran-2-one (2g). ${ }^{4}$



Prepared according to the general procedure using 5-(4-bromophenyl)hex-5-enoic acid ( $\mathbf{1 g}$ ) ( $22 \mathrm{mg}, 80 \mu \mathrm{~mol}$ ), squaramide $\mathbf{6 b}(7 \mathrm{mg}, 12 \mu \mathrm{~mol})$, iodine ( $3 \mathrm{mg}, 27 \mu \mathrm{~mol}$ ) and $N$-iodosuccinimide ( $18 \mathrm{mg}, 178 \mu \mathrm{~mol}$ ). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes $/ \operatorname{PrOH} 90: 10,1 \mathrm{~mL} / \mathrm{min}): t_{r}\left(e_{1}\right.$, major $)=11.83 \mathrm{~min}$ and $t_{r}\left(e_{2}\right.$, minor $)=15.15$
min. Yield: 23 mg (73\%) of colourless oil; ee: 91\%; TLC (hexanes/EtOAc 1:1): $R_{\mathrm{f}}=0.50$, visualized with anisaldehyde; $[\alpha]_{D}^{20}=23.3\left(\mathrm{c}=0.15, \mathrm{CHCl}_{3}\right) ;{ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 7.54(\mathrm{~d}, J=8.6 \mathrm{~Hz}, 2 \mathrm{H}), 7.26(\mathrm{~d}, J=$ $8.7 \mathrm{~Hz}, 2 \mathrm{H}), 3.54(\mathrm{~s}, 2 \mathrm{H}), 2.42-2.57(\mathrm{~m}, 2 \mathrm{H}), 2.28-2.42(\mathrm{~m}, 2 \mathrm{H}), 1.78-1.92(\mathrm{~m}, 1 \mathrm{H}), 1.50-1.70(\mathrm{~m}, 1 \mathrm{H}),{ }^{13} \mathrm{C}$ NMR ( $101 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 170.0,139.4,132.1$ (2C), 127.0 (2C), 122.6, 84.1, 31.9, 29.0, 16.9, 16.5; HRMS (EI): Exact mass calculated for $\mathrm{C}_{12} \mathrm{H}_{12} \mathrm{BrIO}_{2}[M]^{+}: 393.9065$, found 393.9070 .

## (S)-6-(Iodomethyl)-6-isopropyltetrahydro-2H-pyran-2-one (2h). ${ }^{4,}$



Prepared according to the general procedure using 6-methyl-5-methyleneheptanoic acid (1h) ( $29 \mathrm{mg}, 186 \mu \mathrm{~mol}$ ), squaramide $\mathbf{6 b}(16 \mathrm{mg}, 28 \mu \mathrm{~mol})$, iodine ( $7 \mathrm{mg}, 28 \mu \mathrm{~mol}$ ) and $N$-iodosuccinimide ( $42 \mathrm{mg}, 186 \mu \mathrm{~mol}$ ). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral GLC analysis (Chiraldex G-TA, $170{ }^{\circ} \mathrm{C}$ isothermal): $t_{r}\left(e_{1}\right.$, major) $=22.56 \mathrm{~min}$ and $t_{r}\left(e_{2}\right.$, minor $)=22.91 \mathrm{~min}$. Yield: 40 $\mathrm{mg}(77 \%)$ of colourless oil; $e e$ : $16 \%$; TLC (hexanes/EtOAc 1:1): $R_{\mathrm{f}}=0.57$, visualized with anisaldehyde; $[\alpha]_{D}^{20}=$ $1.50\left(\mathrm{c}=0.07, \mathrm{CHCl}_{3}\right) ;{ }^{1} \mathrm{H} \operatorname{NMR}\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 3.42(\mathrm{~s}, 2 \mathrm{H}), 2.45-2.59(\mathrm{~m}, 1 \mathrm{H}), 2.31-2.45(\mathrm{~m}, 1 \mathrm{H}), 2.09$ $-2.25(\mathrm{~m}, 1 \mathrm{H}), 1.95-2.06(\mathrm{~m}, 1 \mathrm{H}), 1.76-1.95(\mathrm{~m}, 3 \mathrm{H}), 0.97(\mathrm{~d}, J=7.0 \mathrm{~Hz}, 6 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR (101 MHz, CDCl3) $\delta 170.4,84.8,35.2,29.5,26.7,16.7,16.6,16.4,13.0$; HRMS (EI): Exact mass calculated for $\mathrm{C}_{9} \mathrm{H}_{15} \mathrm{IO}{ }_{2}[M]^{+}$: 282.0119, found 282.0124.

## (R)-5-(Iodomethyl)-5-phenyldihydrofuran-2(3H)-one (4a). ${ }^{4,6}$



Prepared according to the general procedure using 4-phenylpent-4-enoic acid (3a) ( $40 \mathrm{mg}, 227 \mu \mathrm{~mol}$ ), squaramide $\mathbf{6 b}(19 \mathrm{mg}, 34 \mu \mathrm{~mol})$, iodine ( $9 \mathrm{mg}, 34 \mu \mathrm{~mol}$ ) and $N$-iodosuccinimide ( $51 \mathrm{mg}, 227 \mu \mathrm{~mol}$ ). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes $\left./{ }^{i} \operatorname{PrOH} 98: 2,1 \mathrm{~mL} / \mathrm{min}\right): t_{r}\left(e_{1}\right.$, minor $)=20.69 \mathrm{~min}$ and $t_{r}\left(e_{2}\right.$, major $)=23.69$ min . Yield: $59 \mathrm{mg}(86 \%)$ of colourless oil; ee: 7\%; TLC (hexanes/EtOAc 1:1): $R_{\mathrm{f}}=0.55$, visualized with anisaldehyde; $[\alpha]_{D}^{20}=5.70\left(\mathrm{c}=0.02, \mathrm{CHCl}_{3}\right) ;{ }^{1} \mathrm{H} \mathrm{NMR}\left(300 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 7.31-7.49(\mathrm{~m}, 5 \mathrm{H}), 3.64(\mathrm{~d}, J=0.9$ $\mathrm{Hz}, 2 \mathrm{H}$ ), $2.41-2.88(\mathrm{~m}, 4 \mathrm{H}){ }^{13} \mathrm{C}$ NMR ( $101 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 175.3,140.5,128.8$ (2C), 128.5, 124.8 (2C), 86.0, 33.9, 29.2, 16.3; HRMS (EI): Exact mass calculated for $\mathrm{C}_{11} \mathrm{H}_{11} \mathrm{IO}_{2}[M]^{+}: 301.9804$, found 301.9793.
(S)-5-(4-Chlorophenyl)-5-(iodomethyl)dihydrofuran-2(3H)-one (4b). ${ }^{7}$


Prepared according to the general procedure using 4-(4-chlorophenyl)pent-4-enoic acid ( $\mathbf{3 b}$ ) ( $42 \mathrm{mg}, 200 \mu \mathrm{~mol}$ ), squaramide $\mathbf{6 b}$ ( $17 \mathrm{mg}, 30 \mu \mathrm{~mol}$ ), iodine ( $8 \mathrm{mg}, 30 \mu \mathrm{~mol}$ ) and $N$-iodosuccinimide ( $45 \mathrm{mg}, 200 \mu \mathrm{~mol}$ ). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC
analysis (Chiralpak AD-H, hexanes $\left./{ }^{i} \operatorname{PrOH} 98: 2,1 \mathrm{~mL} / \mathrm{min}\right): t_{r}\left(e_{1}\right.$, major $)=25.97 \mathrm{~min}$ and $t_{r}\left(e_{2}\right.$, minor $)=27.98$ min . Yield: 57 mg ( $85 \%$ ) of colourless oil; ee: $14 \%$; TLC (hexanes/EtOAc 1:1): $R_{\mathrm{f}}=0.52$, visualized with anisaldehyde; $[\alpha]_{D}=0.0\left(\mathrm{c}=0.21, \mathrm{CHCl}_{3}\right) ;{ }^{1} \mathrm{H}$ NMR $\left(300 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta 7.30-7.43(\mathrm{~m}, 4 \mathrm{H}), 3.59(\mathrm{~s}, 2 \mathrm{H}), 2.65$ $-2.88(\mathrm{~m}, 2 \mathrm{H}), 2.43-2.65(\mathrm{~m}, 2 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR ( $101 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta 174.9,139.1,134.6,129.0$ (2C), 126.3 (2C), 85.6, 33.8, 29.1, 15.7; HRMS (EI): Exact mass calculated for $\mathrm{C}_{11} \mathrm{H}_{10} \mathrm{ClIO}_{2}[M]^{+}: 335.9414$, found 335.9407.


Figure S-37 ${ }^{1} \mathrm{H}$-NMR spectrum of compound $\mathbf{2 a}$.

Figure S-38 ${ }^{13} \mathrm{C}$-NMR spectrum of compound 2a.


| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | M:/ Jørn/ NMR/ AVI400/ <br> JET-222-118-47/ 10/ fid |
| 2 Title | JET-222-118-47 |
| 3 Comment | PROTON CDCI3 \{D: $\}$ <br> uio $\backslash$ AVII400-05\} <br> jornet 28 |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | spect |
| 8 Author |  |
| 9 Solvent | CDC13 |
| 10 Temperature | 295.2 |
| 11 Pulse Sequence | zg30 |
| 12 Experiment | 1D |
| 13 Number of Scans | 16 |
| 14 Receiver Gain | 36 |
| 15 Relaxation Delay | 1.0000 |
| 16 Pulse Width | 12.1500 |
| 17 Acquisition Time | 3.9846 |
| 18 Acquisition Date | 2012-06-12T10:27:00 |
| 19 Modification Date | 2012-06-12T10:28:05 |
| 20 Spectrometer Frequency | 400.18 |
| 21 Spectral Width | 8223.7 |
| 22 Lowest Frequency | -1645.0 |
| 23 Nucleus | 1 H |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |

Figure $\mathbf{S - 3 9}{ }^{1} \mathrm{H}-\mathrm{NMR}$ spectrum of compound $\mathbf{2 b}$.


Figure S-40 ${ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{2 b}$.


| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | M:/ Jørn/ NMR/ AVI400/ <br> JET-222-109-29/ 10/ fid |
| 2 Title | JET-222-109-29 |
| 3 Comment | PROTON CDCI3 \{D: $\backslash$ uio AVII400-05\} jornet 82 |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | spect |
| 8 Author |  |
| 9 Solvent | CDC13 |
| 10 Temperature | 294.6 |
| 11 Pulse Sequence | zg30 |
| 12 Experiment | 1 D |
| 13 Number of Scans | 16 |
| 14 Receiver Gain | 90 |
| 15 Relaxation Delay | 1.0000 |
| 16 Pulse Width | 12.1500 |
| 17 Acquisition Time | 3.9846 |
| 18 Acquisition | 2012-05-10T01:04:00 |
| 19 Modification Date | 2012-05-10T01:04:58 |
| 20 Spectrometer Frequency | 400.18 |
| 21 Spectral Width | 8223.7 |
| 22 Lowest Frequency | -1644.6 |
| 23 Nucleus | 1 H |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |

Figure $\mathrm{S}-\mathbf{4 1}{ }^{1} \mathrm{H}-\mathrm{NMR}$ spectrum of compound $\mathbf{2 c}$.


| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | M:/ Jørn/ NMR/ <br> AVI400/ <br> JET-222-109-29/ 11/ <br> fid |
| 2 Title | JET-222-109-29 |
| 3 Comment | C13CPD CDCl3 \{D: 1 uio AVII400-05\} jornet 82 |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | spect |
| 8 Author |  |
| 9 Solvent | CDCI3 |
| 10 Temperature | 295.6 |
| 11 Pulse Sequence | zgpg30 |
| 12 Experiment | 1D |
| 13 Number of Scans | 1024 |
| 14 Receiver Gain | 2050 |
| 15 Relaxation Delay | 2.0000 |
| 16 Pulse Width | 7.0000 |
| 17 Acquisition Time | 1.2977 |
| 18 Acquisition Date | 2012-05-10T02:04:00 |
| 19 Modification Date | 2012-05-10T02:04:23 |
| 20 Spectrometer Frequency | 100.64 |
| 21 Spectral Width | 25252.5 |
| 22 Lowest Frequency | -1061.8 |
| 23 Nucleus | 13 C |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |

Figure $\mathbf{S}-\mathbf{4 2}{ }^{13} \mathrm{C}-\mathrm{NMR}$ spectrum of compound $\mathbf{2 c}$.


Figure $\mathbf{S}-\mathbf{4 3}{ }^{1} \mathrm{H}-\mathrm{NMR}$ spectrum of compound $\mathbf{2 d}$.


Figure $\mathbf{S}-\mathbf{4 4}{ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{2 d}$.


| Parameter | Value |
| :---: | :---: |
| Data File | M:/ Jørn/ NMR/ |
| Name | JEt-222-112-35/ 10/ fid |
| Title | JEt-222-112-35/ 10 |
| Comment | PROTON CDCI3 \{D: uio \DPX300-09\} jornet 2 |
| 4 Origin | Bruker BioSpin GmbH |
| Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | DPX300 |
| 8 Author |  |
| Solvent | CDC13 |
| 10 Temperature | 298.2 |
| 11 Pulse Sequence | zg30 |
| 12 Experiment | 1D |
| 13 Number of Scans | 16 |
| 14 Receiver Gain | 228 |
| 15 Relaxation Delay | 1.0000 |
| 16 Pulse Width | 10.0000 |
| 17 Acquisition | 5.3084 |
| 18 Acquisition Date | 2012-05-16T18:11:38 |
| 19 Modification Date | 2012-05-16T18:11:43 |
| 20 Spectrometer Frequency | 300.13 |
| 21 Spectral | 6172.8 |
| 22 Lowest Frequency | -1236.3 |
| 23 Nucleus | 1H |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |

Figure $\mathbf{S}-\mathbf{4 5}{ }^{1} \mathrm{H}$-NMR spectrum of compound 2e.


Figure S-46 ${ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{2 e}$.


Figure $\mathrm{S}-\mathbf{4 7}{ }^{1} \mathrm{H}$-NMR spectrum of compound $\mathbf{2 f}$.



Figure $\mathbf{S}-48{ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{2 f}$.


| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | M:/ Jørn/ NMR/ AVI400/ JET-222-123-56/ 10/ fid |
| 2 Title | JET-222-123-56 |
| 3 Comment | PROTON CDCI3 \{D: $\backslash$ uio AVII400-05\} jornet 10 |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | spect |
| 8 Author |  |
| 9 Solvent | CDCI3 |
| 10 Temperature | 295.1 |
| 11 Pulse Sequence | zg30 |
| 12 Experiment | 1 D |
| 13 Number of Scans | 16 |
| 14 Receiver Gain | 40 |
| 15 Relaxation Delay | 1.0000 |
| 16 Pulse Width | 12.1500 |
| 17 Acquisition | 3.9846 |
| 18 Acquisition Date | 2012-06-25T20:19:00 |
| 19 Modification Date | 2012-06-25T20:19:14 |
| 20 Spectrometer Frequency | 400.18 |
| 21 Spectral | 8223.7 |
| 22 Lowest Frequency | -1644.2 |
| 23 Nucleus | 1 H |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |

Figure $\mathbf{S}-\mathbf{4 9}{ }^{1} \mathrm{H}-\mathrm{NMR}$ spectrum of compound $\mathbf{2 g}$.


Figure S-50 ${ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{2 g}$.


| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | M:/ Jørn/ NMR/ <br> AVI400/ <br> JET-222-121-53/ 10/ <br> fid |
| 2 Title | JET-222-121-53 |
| 3 Comment | PROTON CDCI3 \{D: $\backslash$ uio $\backslash$ AVII400-05\} jornet 26 |
| 4 Origin | Bruker BioSpin GmbH |
| Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | spect |
| 8 Author |  |
| Solvent | CDC13 |
| 10 Temperature | 295.2 |
| 11 Pulse Sequence | zg30 |
| 12 Experiment | 1D |
| 13 Number of Scans | 16 |
| 14 Receiver Gain | 25 |
| $\begin{aligned} & 15 \text { Relaxation } \\ & \text { Delay } \end{aligned}$ | 1.0000 |
| 16 Pulse Width | 12.1500 |
| 17 Acquisition Time | 3.9846 |
| 18 Acquisition Date | 2012-06-12T09:14:00 |
| 19 Modification Date | 2012-06-12T09:14:34 |
| 20 Spectrometer Frequency | 400.18 |
| 21 Spectral | 8223.7 |
| 22 Lowest Frequency | -1645.0 |
| 23 Nucleus | 1H |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |

Figure $\mathbf{S - 5 1}{ }^{1} \mathrm{H}$-NMR spectrum of compound $\mathbf{2 h}$.



Figure S-52 ${ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{2 h}$.


Figure S-53 ${ }^{1} \mathrm{H}-\mathrm{NMR}$ spectrum of compound $\mathbf{4 a}$.


Figure S-54 ${ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{4 a}$.

| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | M:/ Jørn/ NMR/ <br> JET-222-122-55/ 10/ fid |
| 2 Title | JET-222-122-55/ 10 |
| 3 Comment | PROTON CDCI3 \{D: $\backslash$ uio DPX300-09\} jornet 2 |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | DPX300 |
| 8 Author |  |
| 9 Solvent | CDC13 |
| 10 Temperature | 298.2 |
| 11 Pulse Sequence | zg30 |
| 12 Experiment | 1D |
| 13 Number of Scans | 16 |
| 14 Receiver Gain | 287 |
| 15 Relaxation Delay | 1.0000 |
| 16 Pulse Width | 10.0000 |
| 17 Acquisition Time | 5.3084 |
| 18 Acquisition Date | 2012-06-14T17:08:16 |
| 19 Modification Date | 2012-06-14T17:08:21 |
| 20 Spectrometer Frequency | 300.13 |
| 21 Spectral Width | 6172.8 |
| 22 Lowest Frequency | -1236.3 |
| 23 Nucleus | 1H |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |


| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | $\begin{aligned} & \text { M:/ Jørn/ NMR/ } \\ & \text { AVI400/ } \\ & \text { JET-222-122-55/ 10/ } \\ & \text { fid } \end{aligned}$ |
| 2 Title | JET-222-122-55 |
| 3 Comment | C13CPD CDCI3 \{D: $\backslash$ uio $\backslash$ AVII $400-05\}$ jornet 5 |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | spect |
| 8 Author |  |
| 9 Solvent | CDCI3 |
| 10 Temperature | 296.1 |
| 11 Pulse Sequence | zgpg30 |
| 12 Experiment | 1D |
| 13 Number of Scans | 1024 |
| 14 Receiver Gain | 1620 |
| 15 Relaxation Delay | 2.0000 |
| 16 Pulse Width | 7.0000 |
| 17 Acquisition Time | 1.2977 |
| 18 Acquisition Date | 2012-06-18T15:25:00 |
| 19 Modification Date | 2012-06-18T15:25:58 |
| 20 Spectrometer Frequency | 100.64 |
| 21 Spectral | 25252.5 |
| 22 Lowest Frequency | -1062.3 |
| 23 Nucleus | 13 C |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |



Figure S-55 ${ }^{1} \mathrm{H}$-NMR spectrum of compound $\mathbf{4 b}$.

| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | M:/ Jørn/ NMR/ JET-222-120-51/ 30/ fid |
| 2 Title | JET-222-120-51/ 30 |
| 3 Comment | PROTON CDCI3 \{D: 1 uio \ DPX300-09\} jornet 40 |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | DPX300 |
| 8 Author |  |
| 9 Solvent | CDC13 |
| 10 Temperature | 298.2 |
| 11 Pulse Sequence | zg30 |
| 12 Experiment | 1 D |
| 13 Number of Scans | 16 |
| 14 Receiver Gain | 256 |
| $\begin{aligned} & 15 \text { Relaxation } \\ & \text { Delay } \end{aligned}$ | 1.0000 |
| 16 Pulse Width | 10.0000 |
| 17 Acquisition Time | 5.3084 |
| 18 Acquisition Date | 2012-06-14T12:37:40 |
| 19 Modification Date | 2012-06-14T12:37:45 |
| 20 Spectrometer Frequency | 300.13 |
| 21 Spectral Width | 6172.8 |
| 22 Lowest Frequency | -1236.3 |
| 23 Nucleus | 1 H |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |


| Parameter | Value |
| :---: | :---: |
| 1 Data File Name | M:/ Jørn/ NMR/ AVI400/ <br> JET-222-120-51/ 10/ fid |
| 2 Title | JET-222-120-51 |
| 3 Comment | C13CPD CDC13 \{D: $\backslash$ uio AVII400-05\} jornet 98 |
| 4 Origin | Bruker BioSpin GmbH |
| 5 Owner | tnmr |
| 6 Site |  |
| 7 Spectrometer | spect |
| 8 Author |  |
| 9 Solvent | CDCI3 |
| 10 Temperature | 296.2 |
| 11 Pulse Sequence | zgpg30 |
| 12 Experiment | 1D |
| 13 Number of Scans | 1024 |
| 14 Receiver Gain | 1820 |
| 15 Relaxation Delay | 2.0000 |
| 16 Pulse Width | 7.0000 |
| 17 Acquisition Time | 1.2977 |
| 18 Acquisition Date | 2012-06-14T16:24:00 |
| 19 Modification Date | 2012-06-14T16:24:17 |
| 20 Spectrometer Frequency | 100.64 |
| 21 Spectral Width | 25252.5 |
| 22 Lowest Frequency | -1059.9 |
| 23 Nucleus | 13 C |
| 24 Acquired Size | 32768 |
| 25 Spectral Size | 65536 |

Figure $\mathbf{S - 5 6}{ }^{13} \mathrm{C}$-NMR spectrum of compound $\mathbf{4 b}$.


Figure S-57 MS spectrum of compound 2a.


Figure S-58 HRMS spectrum of compound 2a.


Figure S-59 MS spectrum of compound $\mathbf{2 b}$.


Figure S-60 HRMS spectrum of compound $\mathbf{2 b}$.


Figure S-61 MS spectrum of compound 2c.


Figure S-62 HRMS spectrum of compound 2c.


Figure S-63 MS spectrum of compound 2d.


Figure S-64 HRMS spectrum of compound 2d.


Figure S-65 MS spectrum of compound $\mathbf{2 e}$.


Figure S-66 HRMS spectrum of compound $\mathbf{2 e}$.


Figure S-67 MS spectrum of compound 2 f .


Figure S-68 HRMS spectrum of compound $\mathbf{2 f}$.


Figure S-69 MS spectrum of compound $\mathbf{2 g}$.


Figure S-70 HRMS spectrum of compound 2g.


Figure S-71 MS spectrum of compound $\mathbf{2 h}$.


Figure S-72 HRMS spectrum of compound $\mathbf{2 h}$.


Figure S-73 MS spectrum of compound 4a.


Figure S-74 HRMS spectrum of compound 4a.


Figure S-75 MS spectrum of compound $\mathbf{4 b}$.


Figure S-76 HRMS spectrum of compound $\mathbf{4 b}$.


Use Multiplier \& Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=254 nm

| $\begin{gathered} \text { Peak } \\ \# \end{gathered}$ | $\begin{aligned} & \text { RetTime } \\ & {[\text { min }]} \end{aligned}$ | Type | Width <br> [min] | Area |  | Heicht |  | Area |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  | mAU | * 3 | [mAU | ] |  |
| 1 | 5.356 | VB | 0.1622 |  | 4.70993 | 4.318 | $34 \mathrm{e}-1$ | 0.4359 |
| 2 | 7.875 | BV | 0.1490 |  | 2.14533 |  | 28923 | 2.9749 |
| 3 | 24.044 | BB | 0.4905 | 974 | 4.59747 | 30. | . 63984 | 90.1937 |
| 4 | 27.196 | B | 0.4987 |  | 10795 |  | 2539 | 6.3956 |

Figure S-77 HPLC chromatogram of chiral compound 2a.


Figure S-78 HPLC chromatogram of racemic compound 2a.


Figure S-79 HPLC chromatogram of chiral compound ent-2a.



| Sorted By | : | Siqnal |  |  |
| :---: | :---: | :---: | :---: | :---: |
| Multiplier | : | 1.0000 |  |  |
| Dilution | : | 1.0000 |  |  |
| Sample Amount | : | 1.00000 | [ $\mathrm{ng} / \mathrm{ul}$ ] | ( not used |
| Use Multiplier \& | lution | Factor with | ISTDs |  |
| Signal 1: VWD1 A, Wavelength=254 nm |  |  |  |  |
| $\begin{aligned} & \text { Peak RetTime Type } \\ & \# \text { [min] } \end{aligned}$ | Width | Area | Height | $\begin{gathered} \text { Area } \\ \text { \% } \end{gathered}$ |
|  | [min] | mAU *s | [mAU ] |  |
| 13.739 VB | 0.2032 | 14.80722 | 1.04493 | 1.2256 |
| $2 \quad 7.720 \mathrm{VB}$ | 0.1454 | 15.28908 | 1. 61476 | 1.2655 |
| 322.810 VB | 0.4551 | 587.88745 | 19.91185 | 48.6604 |
| 425.865 BB | 0.5126 | 590.16034 | 17.77557 | 48.8485 |

Figure S-80 HPLC chromatogram of racemic compound $\mathbf{2 a}$.


Figure S-81 HPLC chromatogram of chiral compound $\mathbf{2 b}$.


Figure S-82 HPLC chromatogram of racemic compound $\mathbf{2 b}$.


Figure S-83 HPLC chromatogram of chiral compound $\mathbf{2 c}$.



Figure S-84 HPLC chromatogram of racemic compound $2 \mathbf{2 c}$.



Figure S-85 HPLC chromatogram of chiral compound 2d.


Figure S-86 HPLC chromatogram of racemic compound $\mathbf{2 d}$.


| $\begin{gathered} \text { Peak } \\ \# \end{gathered}$ | $\begin{aligned} & \text { RetTime } \\ & {[\text { min }]} \end{aligned}$ | Type | Width <br> [min] | $\operatorname{mAU}^{\text {Area }}$ | $\begin{aligned} & \text { Height } \\ & {[\mathrm{mAU}]} \end{aligned}$ | $\begin{gathered} \text { Area } \\ 8 \end{gathered}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | 3.068 | BV | 0.0941 | 6.46438 | 1.08766 | 0.7673 |
| 2 | 4.236 | V | 0.1514 | 5.44301 | $5.25429 \mathrm{e}-1$ | 0.6461 |
| 3 | 5.368 | VB | 0.1207 | 15.27190 | 1.85640 | 1.8128 |
| 4 | 10.696 | VB | 0.2249 | 772.95050 | 52.73806 | 91.7508 |
| 5 | 12.170 | BB | 0.2499 | 42.31588 | 2.57544 | 5.0230 |

Figure S-87 HPLC chromatogram of chiral compound 2e.


Figure S-88 HPLC chromatogram of racemic compound $\mathbf{2 e}$.


| Area Percent Report |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
| Sorted By | : | Siqnal |  |  |
| Multiplier | : | 1.0000 |  |  |
| Dilution | : | 1.0000 |  |  |
| Sample Amount | : | 1.00000 | [ $\mathrm{ng} / \mathrm{ul}$ ] | ( not |

Use Multiplier \& Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=254 nm


Figure S-89 HPLC chromatogram of chiral compound $\mathbf{2 f}$.



| Sorted By | $:$ | Siqnal |  |
| :--- | :---: | :---: | :--- |
| Multiplier | $:$ | 1.0000 |  |
| Dilution | $:$ | 1.0000 |  |
| Sample Amount | $:$ | 1.00000 | [ng/ul] (not used in calc.) |
| Use Multiplier \& | Dilution Factor with |  |  |

Signal 1: VWD1 A, Wavelength=254 nm

| $\begin{gathered} \text { Peak } \\ \# \end{gathered}$ | $\begin{aligned} & \text { RetTime } \\ & \text { [min] } \end{aligned}$ | Type | Width [min] | ${ }_{\mathrm{mAU}}{ }^{2}$ | $\begin{aligned} & \text { Area } \\ & { }^{\prime}{ }^{2} \end{aligned}$ | $\begin{array}{r} \mathrm{He} \\ {[\mathrm{~m} \mathbf{A U}} \end{array}$ | $\begin{aligned} & \text { eight } \\ & \text { Uir } \end{aligned}$ | $\begin{gathered} \text { Area } \\ 8 \end{gathered}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | 7.165 | BV | 0.1352 |  | 9.76059 |  | 2.23410 | 0.6601 |
| 2 | 8.083 | VB | 0.1659 |  | 5.50255 |  | 3.23448 | 1.1859 |
| 3 | 31.346 | BB | 0.6430 | 1468 | 8.44702 |  | 5.05751 | 49.0527 |
| 4 | 45.712 | BB | 1.0581 | 1469 | 9.89929 |  | 1.38708 | 49.1012 |

Figure S-90 HPLC chromatogram of racemic compound $\mathbf{2 f}$.



| Sorted By | $:$ | Siqnal |  |
| :--- | :---: | :---: | :--- |
| Multiplier | $\vdots$ | 1.0000 |  |
| Dilution | $\vdots$ | 1.0000 |  |
| Sample Amount | $\vdots$ | 1.00000 | [nq/ul] (not used in calc.) |
| Use Multiplier \& Dilution Factor with ISTDs |  |  |  |

Signal 1: VWD1 A, Wavelength=254 nm

| $\begin{gathered} \text { Peak } \\ \# \end{gathered}$ | $\begin{aligned} & \text { RetTime } \\ & {[\text { min }]} \end{aligned}$ | Type | Width <br> [min] | $\mathrm{mAX}^{2}$ | $\begin{gathered} \text { Area } \\ { }^{*} \text { s } \end{gathered}$ | $\begin{aligned} & \text { Height } \\ & {[\mathrm{mAU} \quad]} \end{aligned}$ | $\begin{gathered} \text { Area } \\ 8 \end{gathered}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | 3.141 | VV | 0.1089 |  | 4.83858 | 1.83550 | 1.0290 |
| 2 | 5.453 | VV | 0.1161 |  | 9.32522 | 5.10360 | 2.7271 |
| 3 | 11.826 | VV | 0.2480 | 1324 | 4.15820 | 82.04984 | 91.8253 |
| 4 | 15.151 | VV | 0.3620 |  | 3.71920 | 2.65011 | 4.4187 |
| Totals : |  |  |  | 1442 | 2.04120 | 91.63906 |  |

Figure S-91 HPLC chromatogram of chiral compound $\mathbf{2 g}$.


Figure S-92 HPLC chromatogram of racemic compound $\mathbf{2 g}$.


Figure S-93 GLC chromatogram of chiral compound $\mathbf{2 h}$.


Figure S-94 GLC chromatogram of racemic compound $\mathbf{2 h}$.


Figure S-95 HPLC chromatogram of chiral compound 4a.



Use Multiplier \& Dilution Factor with ISTDs

Signal 1: VWD1 A, wavelength=254 nm

| $\begin{gathered} \text { Peak } \\ \# \end{gathered}$ | $\begin{aligned} & \text { RetTime } \\ & {[\mathrm{min}]} \end{aligned}$ | Type | Width <br> [min] | Area |  | Height |  | rea |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  | maU | *s | [mAU | ] |  |
| 1 | 19.757 | BB | 0.3764 | 445 | 2344 | 18. | 735 | 49.9590 |
| 2 | 22.817 | BB | 0.4345 | 445 | 75348 |  | 7343 | 50.041 |

Figure S-96 HPLC chromatogram of racemic compound 4a.



Figure S-97 HPLC chromatogram of chiral compound 4b.


Figure S-98 HPLC chromatogram of racemic compound $\mathbf{4 b}$.

## References

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