Asymmetric lodolactonization Utilizing Chiral Squaramides

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Supporting Information

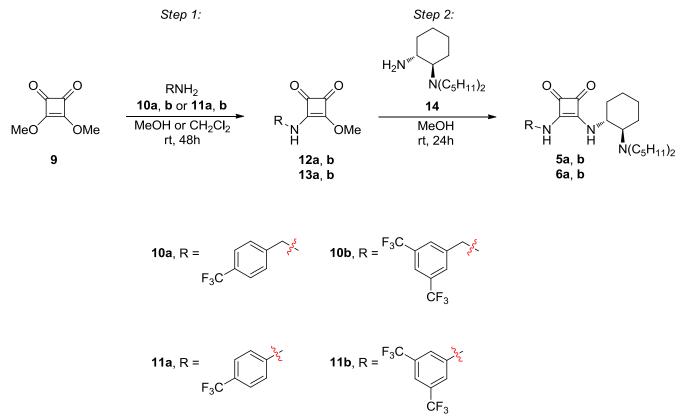
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General Information

All commercially available reagents and solvents were used in the form they were supplied without any further purification. The stated yields are based on isolated material. The melting points are uncorrected. Thin layer chromatography was performed on silica gel 60 F₂₅₄ aluminum-backed plates fabricated by Merck. Flash column chromatography was performed on silica gel 60 (40-63 µm) fabricated by Merck. NMR spectra were recorded on a Bruker AVII-400 or a Bruker DPX-300 spectrometer at 400 MHz or 300 MHz respectively for ¹H NMR and at 100 MHz or 75 MHz respectively for ¹³C NMR. Coupling constants (J) are reported in hertz and chemical shifts are reported in parts per million (δ) relative to the central residual protium solvent resonance in ¹H NMR (CDCl₃) = δ 7.27, DMSO- $d_6 = \delta$ 2.50 and TFA- $d = \delta$ 11.50) and the central carbon solvent resonance in ¹³C NMR (CDCl₃) = δ 77.00 ppm and DMSO- d_6 = δ 39.43). Mass spectra were recorded at 70 eV on Waters Prospec Q spectrometer using EI as the method of ionization. High resolution mass spectra were recorded on Waters Prospec Q spectrometer using EI as the method of ionization. Optical rotations were measured using a 1 mL cell with a 1.0 dm path length on a Perkin Elmer 341 polarimeter. Determination of enantiomeric excess was performed by HPLC on an Agilent Technologies 1200 Series instrument with diode array detector set at 254 nm and equipped with a chiral stationary phase (Chiralpak AD-H 5 µm 4.6 x 250 mm), applying the conditions stated. Alternatively, determination of enantiomeric excess was performed by GLC on a Varian 3380 instrument with split (1:30) injection, FID detector and equipped with a chiral stationary phase (Chiraldex G-TA 0.12 µm 0.25 mm x 30 m), applying the conditions stated.

Preparation of Squaramide Catalysts

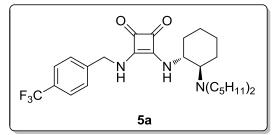


Scheme S-1 Synthetic route to chiral squaramides 5a, 5b, 6a and 6b.

General procedure for the preparation of squaramide catalysts (Scheme S-1): Step 1. 3,4-Dimethoxycyclobut-3ene-1,2-dione (9) (1.0 equiv.) was suspended/dissolved in either MeOH or CH_2Cl_2 (0.07 M) and the amine **10a**, **b** or **11a**, **b** (1.1 equiv.) was added. The resulting mixture was stirred at ambient temperature for 48 hours. The mixture was then filtered; the collected solid residue was washed with ice-cold MeOH and dried *in vacuo* to afford the corresponding squaramate **12a**, **b**¹ or **13a**, **b**^{2. 3}. Step 2. 3-(Benzylamino)- or 3-(arylamino)-4methoxycyclobut-3-ene-1,2-dione **12a**, **b** or **13a**, **b** (1.0 equiv.) was suspended in MeOH (0.07 M) and (*IR*,2*R*)- N^1 , N^1 -dipentylcyclohexane-1,2-diamine (**14**) (1.0 equiv.) was added. The resulting mixture was stirred at ambient temperature for 24 hours. The mixture was then filtered; the collected solid residue was washed with ice-cold MeOH and dried *in vacuo* to afford the corresponding squaramide **5a**, **b** or **6a**, **b**.

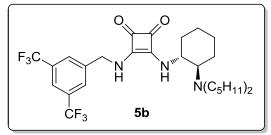
Notice! (1R,2R)- N^1 , N^1 -dipentylcyclohexane-1,2-diamine (14) was prepared according to literature procedure⁴ from commercially available (1R,2R)-cyclohexane-1,2-diamine (Aldrich; 98%, optical purity *ee*: 99% by GLC). The optical antipode, (1S,2S)- N^1 , N^1 -dipentylcyclohexane-1,2-diamine (*ent*-14), was prepared in the same manner from commercially available (1S,2S)-cyclohexane-1,2-diamine D-tartrate (Aldrich; 99%).

3-(((1*R*,2*R*)-2-(Dipentylamino)cyclohexyl)amino)-4-((4-(trifluoromethyl)benzyl)amino)cyclobut-3-ene-1,2-dione (5a).



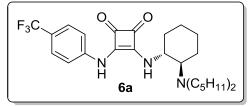
Prepared according to the general procedure by reacting 3-methoxy-4-((4-(trifluoromethyl)benzyl)amino)-cyclobut-3-ene-1,2-dione (**12a**) (0.250 g, 0.88 mmol) with $(1R,2R)-N^1,N^1$ -dipentylcyclohexane-1,2-diamine (**14**) (0.223 g, 0.88 mmol) to afford the title compound **5a** after recrystallization from pentane. Yield: 0.211 g (48%), colourless solid; M.p.: 204 – 205 °C, decomp.; TLC (hexanes/EtOAc 1:1): $R_f = 0.06$; $[\alpha]_D^{20} = -634.9$ (c = 0.04, CHCl₃); ¹H NMR (400 MHz, DMSO- d_6) δ 7.85 (bs, 1H), 7.70 (d, J = 8.0 Hz, 2H), 7.53 (d, J = 8.0 Hz, 2H), 7.16 (bs, 1H), 4.80 (s, 2H), 4.00 – 3.57 (m, 1H), 2.43 – 2.28 (m, 3H), 2.28 – 2.16 (m, 2H), 2.05 – 1.90 (m, 1H), 1.84 – 1.73 (m, 1H), 1.73 – 1.53 (m, 2H), 1.39 – 0.97 (m, 16H), 0.76 (t, J = 6.7 Hz, 6H); ¹³C NMR (101 MHz, DMSO- d_6) δ 182.6, 181.9, 168.1, 166.9, 144.0, 128.2 (2C), 124.4 – 126.4 (m, 3C), 124.2 (q, ¹ $J_{CF} = 272.0$ Hz, 1C), 63.8, 54.1, 49.4 (2C), 46.1, 34.6, 29.0 (2C), 28.5 (2C), 25.0, 24.6, 24.0, 22.1 (2C), 13.9 (2C); HRMS (EI): Exact mass calculated for C₂₈H₄₀F₃N₃O₂ [M]⁺: 507.3073, found 507.3066.

3-((3,5-*Bis*(trifluoromethyl)benzyl)amino)-4-(((1*R*,2*R*)-2-(dipentylamino)cyclohexyl)amino)cyclobut-3-ene-1,2-dione (5b).



Prepared according to the general procedure by reacting 3-((3,5-bis(trifluoromethyl)benzyl)amino)-4-methoxycyclobut-3-ene-1,2-dione (**12b**) (1.050 g, 2.97 mmol) with (1*R*,2*R*)- N^1 , N^1 -dipentylcyclohexane-1,2-diamine (**14**) (0.756 g, 2.97 mmol) to afford the title compound **5b**. Yield: 1.045 g (85%), colourless solid; M.p.: 216 – 218 °C, decomp.; TLC (hexanes/EtOAc 1:1): $R_f = 0.06$; $[\alpha]_D^{20} = -86.0$ (c = 0.05, CHCl₃); ¹H NMR (300 MHz, DMSO- d_6) δ 8.02 (s, 3H), 7.87 (bs, 1H), 7.24 (bs, 1H), 5.08 – 4.67 (m, 2H), 4.05 – 3.57 (m, 1H), 2.41 – 2.26 (m, 3H), 2.26 – 2.12 (m, 2H), 2.04 – 1.85 (m, 1H), 1.83 – 1.72 (m, 1H), 1.72 – 1.54 (m, 2H), 1.41 – 0.87 (m, 16H), 0.70 (t, J = 6.4 Hz, 6H); ¹³C NMR (101 MHz, DMSO- d_6 + DCl) δ 183.0, 182.4, 169.3, 167.9, 142.8, 130.9 (q, ${}^{2}J_{CF} = 32.9$ Hz, 2C), 129.0 (2C), 123.8 (q, ${}^{1}J_{CF} = 272.9$ Hz, 2C), 121.6, 64.1, 52.4, 52.0, 51.3, 46.3, 34.4, 28.8, 28.7, 24.7, 24.3 (2C), 24.1, 24.0, 22.1, 21.9, 14.2, 14.0; HRMS (EI): Exact mass calculated for C₂₉H₃₉F₆N₃O₂ [M]⁺: 575.2946, found 575.2937.

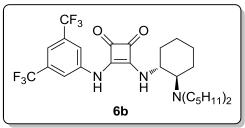
3-(((1*R*,2*R*)-2-(Dipentylamino)cyclohexyl)amino)-4-((4-(trifluoromethyl)phenyl)amino)cyclobut-3-ene-1,2-dione (6a).



Prepared according to the general procedure by reacting 3-methoxy-4-((4-(trifluoromethyl)phenyl)amino)-cyclobut-3-ene-1,2-dione (**13a**) (0.763 g, 2.81 mmol) with $(1R,2R)-N^1,N^1$ -dipentylcyclohexane-1,2-diamine (**14**) (0.715 g, 2.81 mmol) to afford the title compound **6a**. Yield: 0.905 g (66%), colourless solid; M.p.: 200 – 201 °C, decomp.; TLC (hexanes/EtOAc 1:1): $R_f = 0.10$; $[\alpha]_D^{20} = -141.7$ (c = 0.13, CHCl₃); ¹H NMR (300 MHz, TFA) δ

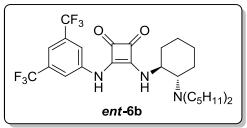
7.59 (d, J = 8.2 Hz, 2H), 7.36 (d, J = 8.2 Hz, 2H), 4.75 – 4.52 (m, 1H), 3.76 – 3.45 (m, 2H), 3.33 (td, J = 12.6, 4.3 Hz, 1H), 3.14 (td, J = 12.3, 5.2 Hz, 1H), 2.89 (td, J = 12.2, 4.6 Hz, 1H), 2.36 – 2.21 (m, 1H), 2.21 – 2.06 (m, 1H), 2.04 – 1.50 (m, 8H), 1.50 – 1.35 (m, 2H), 1.35 – 1.14 (m, 8H), 0.94 – 0.68 (m, 6H); ¹³C NMR (101 MHz, DMSO- d_6) δ 184.5, 179.6, 169.7, 162.0, 142.8, 126.6 (2C), 124.5 (q, ${}^{1}J_{CF} = 271.0$ Hz), 122.1 (q, ${}^{2}J_{CF} = 32.1$ Hz), 117.6 (2C), 64.1 (2C), 54.6, 49.2, 34.2, 29.2 (2C), 28.5 (2C), 24.9, 24.6, 23.6, 22.2 (2C), 14.0 (2C); HRMS (EI): Exact mass calculated for C₂₇H₃₈F₃N₃O₂ [M]⁺: 493.2916, found 493.2921.

3-((3,5-*Bis*(trifluoromethyl)phenyl)amino)-4-(((1*R*,2*R*)-2-(dipentylamino)cyclohexyl)amino)cyclobut-3-ene-1,2-dione (6b).

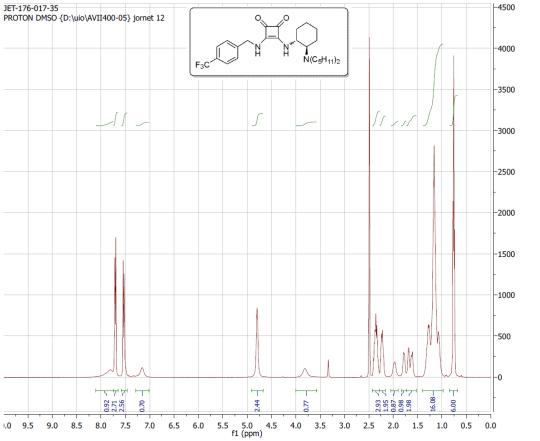


Prepared according to the general procedure by reacting 3-((3,5-bis(trifluoromethyl)phenyl)amino)-4-methoxycyclobut-3-ene-1,2-dione (**13b**) (1.000 g, 2.94 mmol) with (1*R*,2*R*)- N^1 , N^1 -dipentylcyclohexane-1,2-diamine (**14**) (0.748 g, 2.94 mmol) to afford the title compound **6b**. Yield: 1.050 g (64%), colourless solid; M.p.: 195 – 196 °C, decomp.; TLC (hexanes/EtOAc 1:1): $R_f = 0.12$; $[\alpha]_D^{20} = -107.6$ (c = 0.10, CHCl₃); ¹H NMR (300 MHz, TFA) δ 7.87 (s, 2H), 7.79 (s, 1H), 4.74 (td, J = 11.2, 4.2 Hz, 1H), 3.89 – 3.54 (m, 2H), 3.44 (td, J = 12.5, 6.3 Hz, 1H), 3.25 (td, J = 12.4, 5.2 Hz, 1H), 2.99 (td, J = 12.2, 4.5 Hz, 1H), 2.47 – 2.32 (m, 1H), 2.32 – 2.17 (m, 1H), 2.16 – 1.46 (m, 10H), 1.46 – 1.26 (m, 8H), 1.02 – 0.81 (m, 6H); ¹³C NMR (101 MHz, DMSO- d_6) δ 184.5, 179.7, 170.0, 161.4, 141.3, 131.4 (q, ² $_{JCF} = 32.9$, 31.6 Hz, 2C), 123.1 (q, ¹ $_{JCF} = 273.0$ Hz, 2C), 117.5 (2C), 114.3, 64.2 (2C), 54.7, 49.2, 34.1, 29.2 (2C), 28.5 (2C), 24.8, 24.6, 23.7, 22.2 (2C), 13.9 (2C); HRMS (EI): Exact mass calculated for C₂₈H₃₇F₆N₃O₂ [M]⁺: 561.2790, found 561.2799.

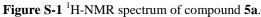
3-((3,5-*Bis*(trifluoromethyl)phenyl)amino)-4-(((1*S*,2*S*)-2-(dipentylamino)cyclohexyl)amino)cyclobut-3-ene-1,2-dione (*ent*-6b).

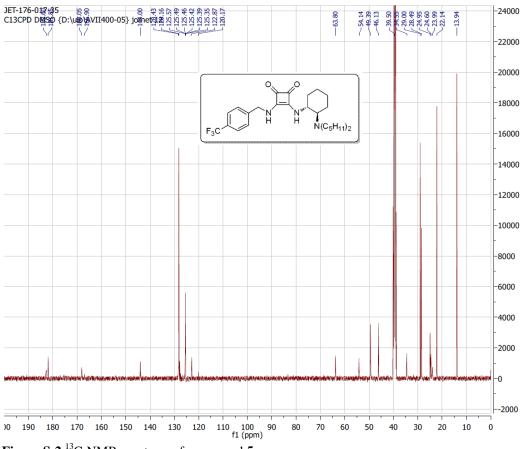


Prepared according to the general procedure by reacting 3-((3,5-bis(trifluoromethyl)phenyl)amino)-4-methoxycyclobut-3-ene-1,2-dione (**13b**) (0.240 g, 0.71 mmol) with (1*R*,2*R*)- N^1 , N^1 -dipentylcyclohexane-1,2-diamine (*ent*-14) (0.180 g, 0.71 mmol) to afford the title compound *ent*-6b. Yield: 0.164 g (41%), colourless solid; TLC (hexanes/EtOAc 1:1): $R_f = 0.12$; $[\alpha]_D^{20} = 110.6$ (c = 0.11, CHCl₃).



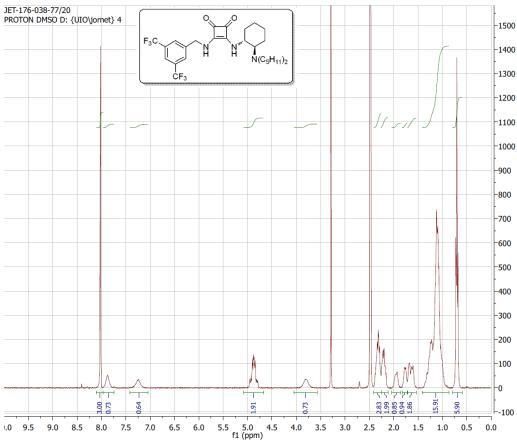
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5	Owner	tnmr
6	Site	
	Spectrometer	spect
8	Author	
9	Solvent	DMSO
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11	Pulse Sequence	zg30
12	Experiment	1D
13	Number of Scans	16
14	Receiver Gain	72
15	Relaxation Delay	1.0000
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22	Lowest Frequency	-1646.4
23	Nucleus	1H
24	Acquired Size	32768
25	Spectral Size	65536



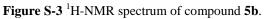


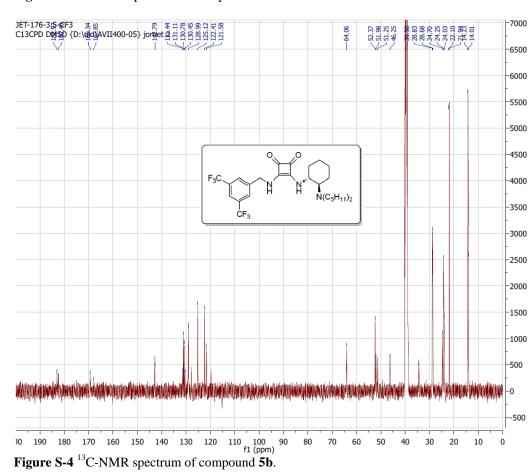
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5	Owner	tnmr
6	Site	
7	Spectrometer	spect
8	Author	
9	Solvent	DMSO
10	Temperature	295.7
11	Pulse Sequence	zgpg30
12	Experiment	1D
13	Number of Scans	2048
14	Receiver Gain	1620
15	Relaxation Delay	2.0000
16	Pulse Width	7.0000
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19	Modification Date	2012-05-05T18:45:59
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21	Spectral Width	25252.5
22	Lowest Frequency	-1102.0
23	Nucleus	13C
24	Acquired Size	32768
25	Spectral Size	65536

Figure S-2 ¹³C-NMR spectrum of compound 5a.

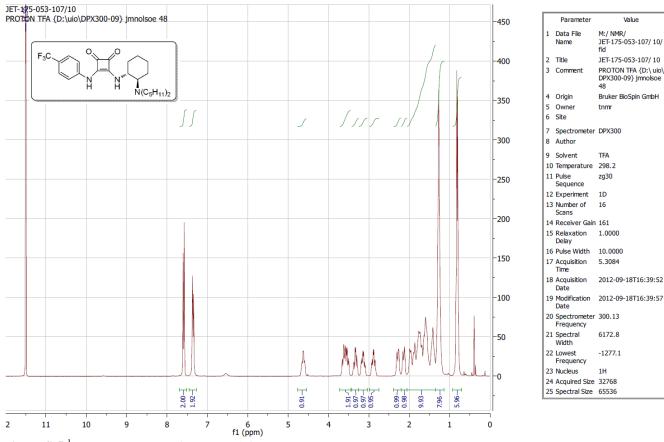


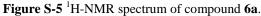
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4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
	Spectrometer Author	DPX300
9	Solvent	DMSO
10	Temperature	298.2
11	Pulse Sequence	zg30
12	Experiment	1D
13	Number of Scans	64
14	Receiver Gain	1149
15	Relaxation Delay	1.0000
16	Pulse Width	10.0000
17	Acquisition Time	5.3084
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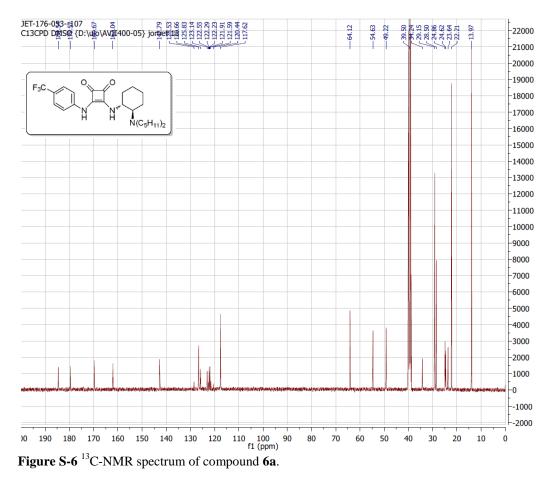




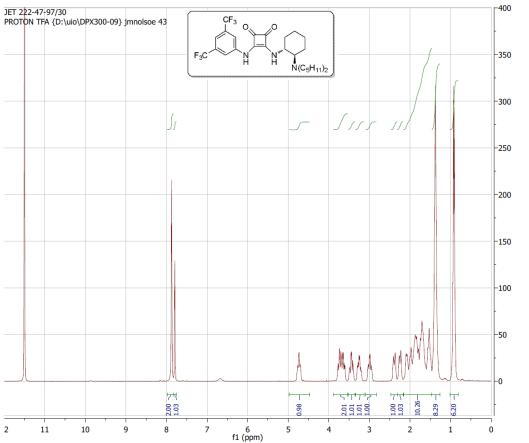
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4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
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7	Spectrometer	spect
	Author	
9	Solvent	DMSO
10	Temperature	300.0
11	Pulse Sequence	zgpg30
12	Experiment	1D
13	Number of Scans	1024
14	Receiver Gain	2050
15	Relaxation Delay	5.0000
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17	Acquisition Time	1.2977
18	Acquisition Date	2012-11-05T09:59:00
19	Modification Date	2012-11-05T09:59:39
20	Spectrometer Frequency	100.64
21	Spectral Width	25252.5
22	Lowest Frequency	-1038.7
23	Nucleus	13C
24	Acquired Size	32768
25	Spectral Size	65536



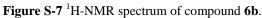


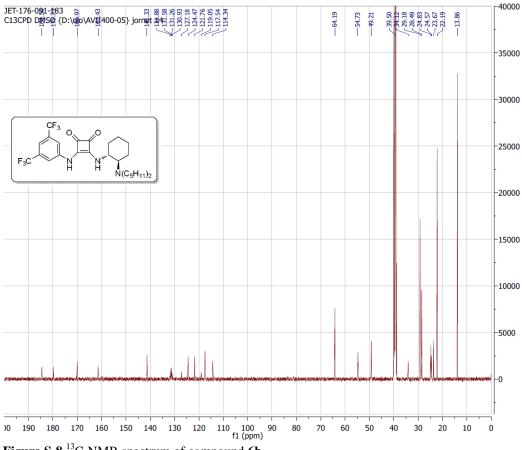


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9	Solvent	DMSO
10	Temperature	295.8
11	Pulse Sequence	zgpg30
12	Experiment	1D
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14	Receiver Gain	1150
15	Relaxation Delay	2.0000
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21	Spectral Width	25252.5
22	Lowest Frequency	-1101.7
23	Nucleus	13C
	Acquired Size	
25	Spectral Size	65536



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16	Pulse Width	10.0000
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20	Spectrometer Frequency	300.13
21	Spectral Width	6172.8
22	Lowest Frequency	-1275.9
23	Nucleus	1H
	Acquired Size	
25	Spectral Size	65536





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4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
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	Spectrometer Author	spect
9	Solvent	DMSO
10	Temperature	295.7
11	Pulse Sequence	zgpg30
12	Experiment	1D
13	Number of Scans	2048
14	Receiver Gain	2050
15	Relaxation Delay	2.0000
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Figure S-8¹³C-NMR spectrum of compound 6b.

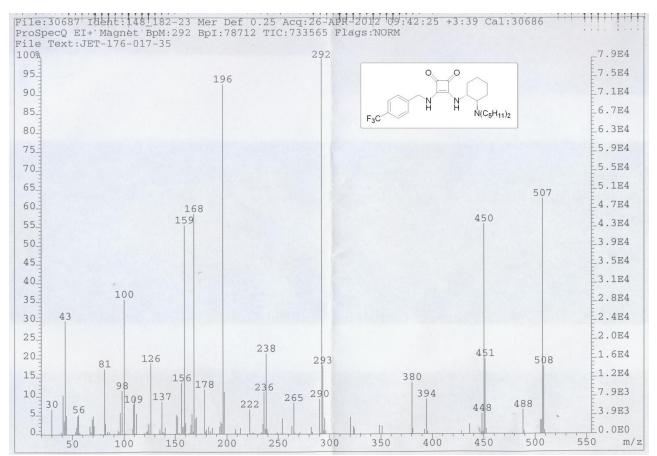


Figure S-9 MS spectrum of compound 5a.

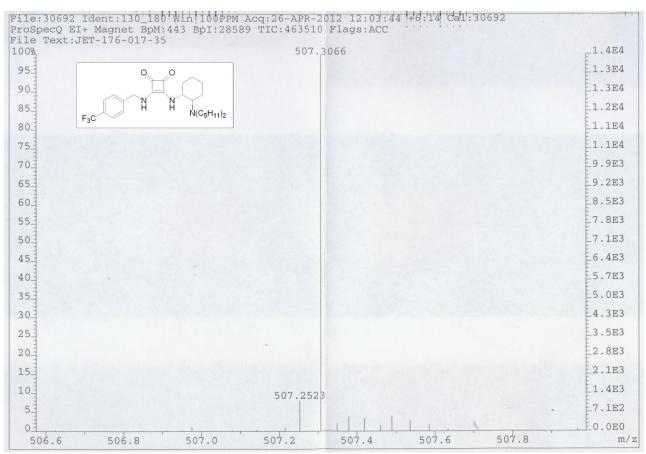


Figure S-10 HRMS spectrum of compound 5a.

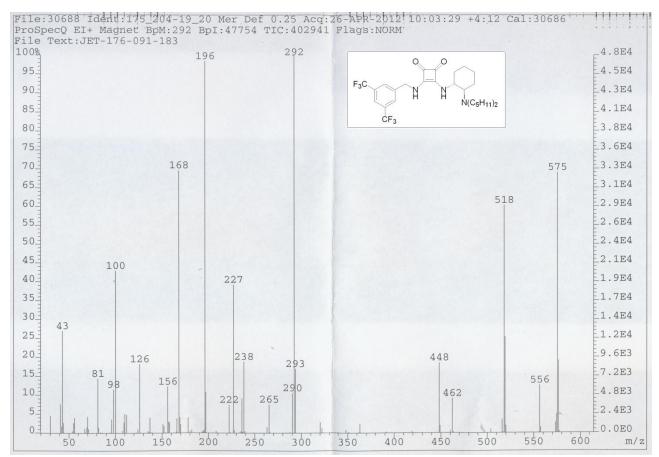


Figure S-11 MS spectrum of compound 5b.

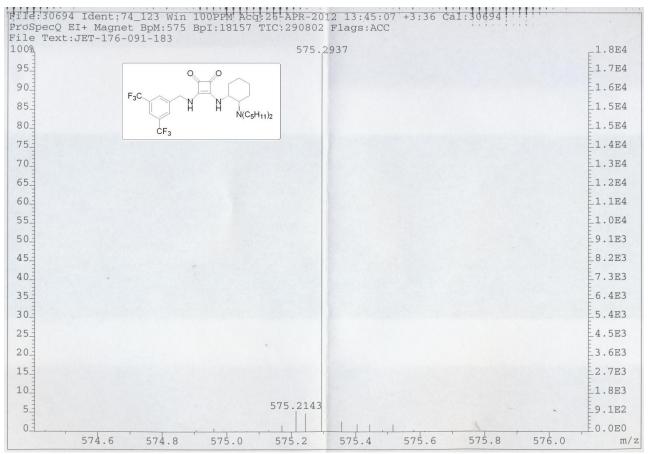
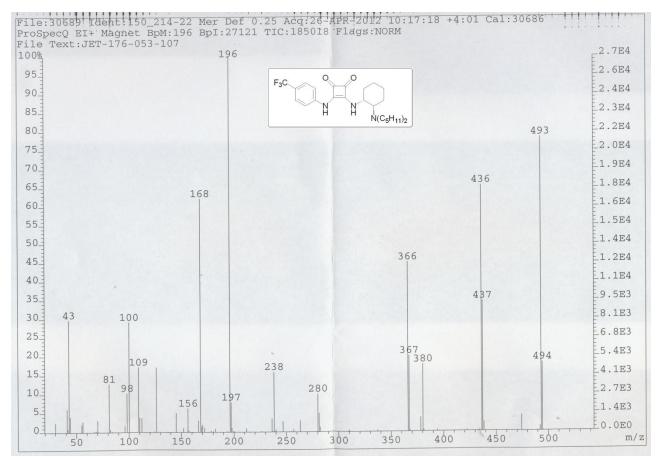
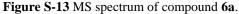


Figure S-12 HRMS spectrum of compound 5b.





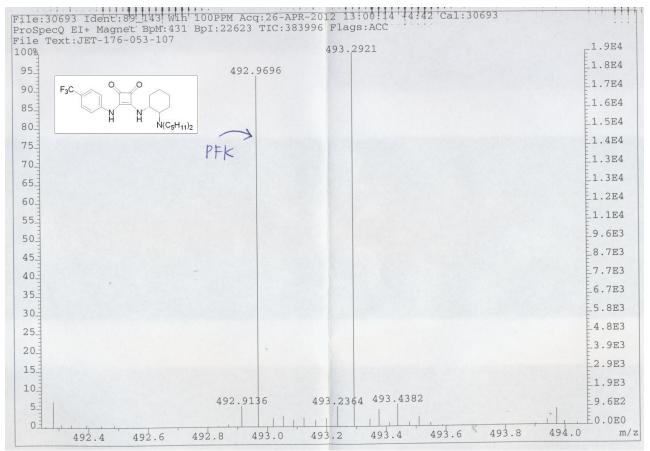
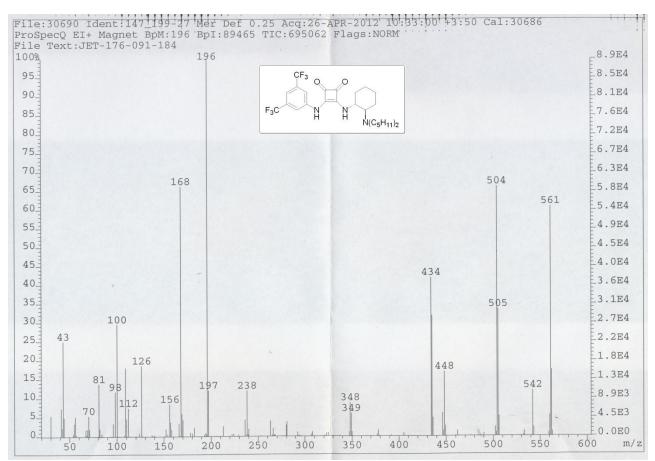
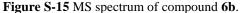


Figure S-14 HRMS spectrum of compound 6a.





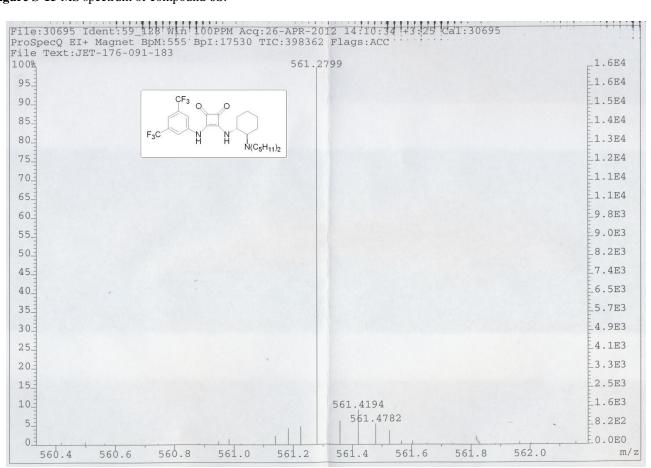


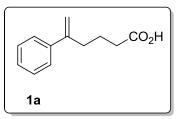
Figure S-16 HRMS spectrum of compound 6b.

Preparation of Starting Materials

General procedure for the preparation of γ - and δ -unsaturated acids: Step 1. 4-Aryl-4-oxobutanoic acid (1.0 equiv.) or 5-aryl-5-oxopentanoic acid (1.0 equiv.) was suspended in MeOH (0.25 M), the suspension was cooled to 0 °C and acetyl chloride (1.2 equiv.) was added. Cooling was discontinued and the resulting homogeneous mixture was stirred overnight at ambient temperature. The mixture was diluted with CH₂Cl₂ and washed in succession with satd. aq. NaHCO₃, brine and water. The organic phase was dried over MgSO₄, filtered and the solvent was evaporated in vacuo to afford the corresponding 4-aryl-4-oxobutanoic acid methyl ester or 5-aryl-5oxopentanoic acid methyl ester. Step 2. Triphenylphosphonium bromide (1.3 equiv.) was suspended in THF (0.75 M) and cooled to 0 °C, whereupon KHMDS (0.5 M in toluene, 1.3 equiv.) was added in one go. The resulting mixture was stirred for 1 hour then cooled to -78 °C and a solution of the 4-aryl-4-oxobutanoic acid methyl ester or 5-aryl-5-oxopentanoic acid methyl ester (1.0 equiv.) in THF (~0.25 M) was added in a dropwise manner. Cooling was discontinued and the resulting mixture was stirred at ambient temperature until TLC indicated full conversion of the starting material. The mixture was treated with satd. aq. NH_4Cl and extracted with EtOAc. The combined organic extracts were dried over MgSO₄, filtered and the solvent was evaporated in vacuo. The residue was purified by flash chromatography on silica (hexanes, followed by hexanes/EtOAc in various proportions) to afford the corresponding 4-aryl-4-oxobutanoic acid methyl ester or 5-arylhex-5-enoic acid methyl ester. Step 3. 4-Arylpent-4-enoic methyl ester or 5-arylhex-5-enoic acid methyl ester (1.0 equiv.) was dissolved in THF and water was added (8:1 THF/water, ~0.10 M), followed by LiOH H₂O (1.5 equiv.). The resulting biphasic mixture was stirred vigorously overnight at ambient temperature, whereupon the solvent was evaporated *in vacuo*. The residue was dissolved in 1 M aq. NaOH and washed with CH₂Cl₂. The pH was adjusted to approx. 2 and the aq. phase was extracted twice with CH₂Cl₂. The combined organic extracts were dried over MgSO₄, filtered and evaporated *in vacuo* to afford the corresponding 4-arylpent-4-enoic acid or 5-arylhex-5-enoic acid, respectively.

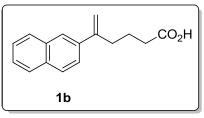
Alternatively, some of the γ - and δ -unsaturated acids were prepared directly from the corresponding 4-aryl-4oxobutanoic acids or 5-aryl-5-oxopentanoic acids according to the procedure of Takemiya *et al.*⁵

5-Phenylhex-5-enoic acid (1a).⁴

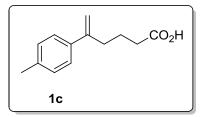


Prepared from commercially available 5-oxo-5-phenylpentanoic acid (**15a**) by a Wittig reaction according to the procedure of Takemiya *et al.*⁵ All physical data were in full agreement with those reported in the literature. ¹H NMR (400 MHz, CDCl₃) δ 11.82 (s, 1H), 7.15 – 7.63 (m, 5H), 5.34 (s, 1H), 5.11 (s, 1H), 2.61 (t, *J* = 7.5 Hz, 2H), 2.41 (t, *J* = 7.4 Hz, 2H), 1.83 (p, *J* = 7.4 Hz, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 180.2, 147.3, 140.7, 128.3 (2C), 127.5, 126.1 (2C), 113.1, 34.4, 33.3, 23.0.

5-(Naphthalen-2-yl)hex-5-enoic acid (1b).⁴

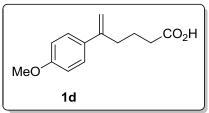


Prepared according to the general procedure using 5-(naphthalen-2-yl)-5-oxopentanoic acid (**15b**). All physical data were in full agreement with those reported in the literature. ¹H NMR (300 MHz, CDCl₃) δ 11.23 (bs, 1H), 7.77–7.96 (m, 4H), 7.61 (dd, *J* = 8.6, 1.8 Hz, 1H), 7.42 – 7.56 (m, 2H), 5.51 (d, *J* = 1.3 Hz, 1H), 5.23 (d, *J* = 1.5 Hz, 1H), 2.73 (t, *J* = 7.5 Hz, 2H), 2.45 (t, *J* = 7.4 Hz, 2H), 1.89 (p, *J* = 7.4 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 180.1, 147.1, 137.9, 133.4, 132.8, 128.1, 127.9, 127.5, 126.1, 125.8, 124.7, 124.5, 113.6, 34.4, 33.3, 23.1.



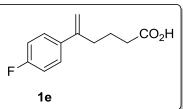
Prepared according to the general procedure using 5-oxo-5-(*p*-tolyl)pentanoic acid (**15c**). All physical data were in full agreement with those reported in the literature. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.34 (d, *J* = 8.0 Hz, 2H), 7.15 (d, *J* = 7.9 Hz, 2H), 5.29 (d, *J* = 2.0 Hz, 1H), 5.01 (d, *J* = 1.9 Hz, 1H), 2.48 (t, *J* = 7.4 Hz, 2H), 2.29 (s, 3H), 2.22 (t, *J* = 7.4 Hz, 2H), 1.60 (p, *J* = 7.4 Hz, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 174.3, 147.0, 137.2, 136.8, 129.0 (2C), 125.7 (2C), 111.8, 33.8, 33.0, 23.2, 20.7.

5-(4-Methoxyphenyl)hex-5-enoic acid (1d).⁴



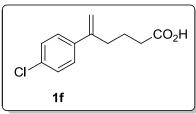
Prepared according to the general procedure using 5-(4-methoxyphenyl)-5-oxopentanoic acid (**15d**). All physical data were in full agreement with those reported in the literature. ¹H NMR (300 MHz, CDCl₃) δ 7.31 – 7.41 (m, 2H), 6.84 – 6.91 (m, 2H), 5.25 (d, *J* = 1.4 Hz, 1H), 5.01 (d, *J* = 1.4 Hz, 1H), 3.82 (s, 3H), 2.56 (t, *J* = 7.4 Hz, 2H), 2.39 (t, *J* = 7.4 Hz, 2H), 1.81 (p, *J* = 7.5 Hz, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 179.5, 159.1, 146.5, 133.1, 127.2 (2C), 113.7 (2C), 111.5, 55.3, 34.5, 33.2, 23.1.

5-(4-Fluorophenyl)hex-5-enoic acid (1e).⁴



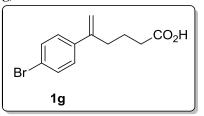
Prepared according to the general procedure using 5-(4-fluorophenyl)-5-oxopentanoic acid (**15e**). All physical data were in full agreement with those reported in the literature. ¹H NMR (300 MHz, CDCl₃) δ 11.35 (bs, 1H), 7.32 – 7.45 (m, 2H), 6.97 – 7.08 (m, 2H), 5.24 – 5.30 (m, 1H), 5.04 – 5.11 (m, 1H), 2.56 (t, *J* = 7.5 Hz, 1H), 2.39 (t, *J* = 7.5 Hz, 1H), 1.80 (p, *J* = 7.5 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 180.0, 162.3 (d, ¹*J*_{CF} = 246 Hz), 146.3, 136.7 (d, ⁴*J*_{CF} = 3.3 Hz), 127.7 (d, ³*J*_{CF} = 7.9 Hz, 2C), 115.2 (d, ²*J*_{CF} = 21.3 Hz, 2C), 113.0, 34.5, 33.2, 22.9.

5-(4-Chlorophenyl)hex-5-enoic acid (1f).⁴



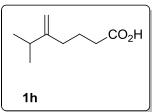
Prepared from 5-(4-chlorophenyl)-5-oxopentanoic acid (**15f**) by a Wittig reaction according to the procedure of Takemiya *et al.*⁵ All physical data were in full agreement with those reported in the literature. ¹H NMR (400 MHz, CDCl₃) δ 9.76 (bs, 1H), 7.28 – 7.38 (m, 4H), 5.31 (d, *J* = 1.3 Hz, 1H), 5.11 (d, *J* = 2.0 Hz, 1H), 2.55 (t, *J* = 7.5 Hz, 2H), 2.39 (t, *J* = 7.4 Hz, 2H), 1.79 (p, *J* = 7.4 Hz, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 179.6, 146.2, 139.1, 133.3, 128.5 (2C), 127.4 (2C), 113.6, 34.3, 33.2, 22.9.

5-(4-Bromophenyl)hex-5-enoic acid (1g).⁴



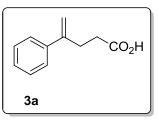
Prepared according to the general procedure using 5-(4-bromophenyl)-5-oxopentanoic acid (**15g**). All physical data were in full agreement with those reported in the literature. ¹H NMR (300 MHz, CDCl₃) δ 11.44 (bs, 1H), 7.42–7.52 (m, 2H), 7.24–7.33 (m, 2H), 5.33 (s, 1H), 5.12 (s, 1H), 2.56 (t, *J* = 7.5 Hz, 2H), 2.40 (t, *J* = 7.4 Hz, 2H), 1.80 (p, *J* = 7.4 Hz, 2H); 13C NMR (75 MHz, CDCl₃) δ 180.0, 146.2, 139.5, 131.4 (2C), 127.7 (2C), 121.4, 113.6, 34.2, 33.2, 22.9.

6-Methyl-5-methyleneheptanoic acid (1h).^{4, 5}



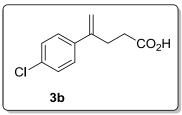
Prepared from commercially available 6-methyl-5-methyleneheptanoic acid (**15h**) by a Wittig reaction according to the procedure of Takemiya *et al.*⁵ All physical data were in full agreement with those reported in the literature. ¹H NMR (300 MHz, CDCl₃) δ 11.09 (bs, 1H), 4.76 – 4.83 (m, 1H), 4.67 – 4.74 (m, 1H), 2.38 (t, *J* = 7.5 Hz, 2H), 2.24 (hept, *J* = 7.0 Hz, 1H), 2.09 (t, *J* = 7.5 Hz, 2H), 1.80 (p, *J* = 7.5 Hz, 2H), 1.03 (d, *J* = 7.0 Hz, 6H); ¹³C NMR (75 MHz, CDCl₃) δ 180.3, 154.6, 107.1, 33.6 (3C), 23.0, 21.8 (2C).

4-Phenylpent-4-enoic acid (3a).⁴

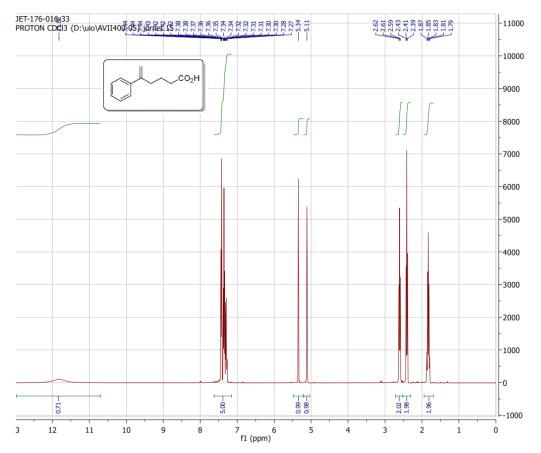


Prepared according to the general procedure using 4-oxo-4-phenylbutanoic acid (**16a**). All physical data were in full agreement with those reported in the literature. ¹H NMR (300 MHz, CDCl₃) δ 10.56 (bs, 1H), 7.23 – 7.48 (m, 5H), 5.35 (s, 1H), 5.14 (s, 1H), 2.88 (t, *J* = 7.7 Hz, 2H), 2.56 (t, *J* = 7.7 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 179.5, 146.5, 140.4, 128.4 (2C), 127.7, 126.1 (2C), 112.9, 33.0, 30.1.

4-(4-Chlorophenyl)pent-4-enoic acid (3b).⁶

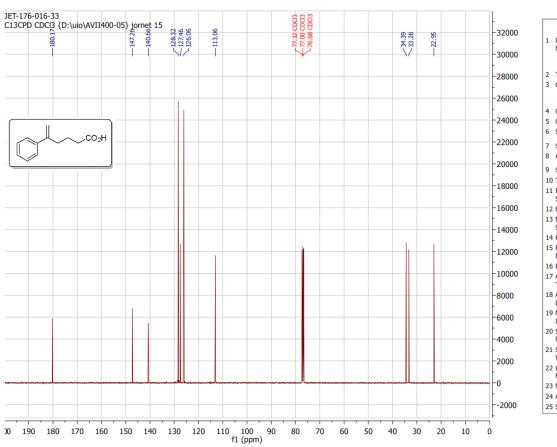


Prepared according to the general procedure using 4-(4-chlorophenyl)-4-oxobutanoic acid (**16b**). All physical data were in full agreement with those reported in the literature. ¹H NMR (300 MHz, CDCl₃) δ 10.60 (bs, 1H), 7.28 – 7.38 (m, 4H), 5.32 (s, 1H), 5.14 (s, 1H), 2.83 (t, *J* = 7.7 Hz, 1H), 2.54 (t, *J* = 7.7 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 179.3, 145.4, 138.8, 133.5, 128.6 (2C), 127.4 (2C), 113.5, 32.8, 30.0.



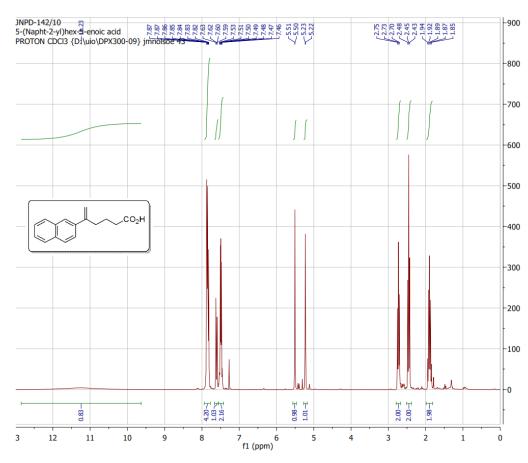
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4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
7	Spectrometer	spect
8	Author	
9	Solvent	CDCI3
10	Temperature	294.9
11	Pulse Sequence	zg30
12	Experiment	1D
13	Number of Scans	16
14	Receiver Gain	57
15	Relaxation Delay	1.0000
16	Pulse Width	12.1500
17	Acquisition Time	3.9846
18	Acquisition Date	2012-05-05T22:58:00
19	Modification Date	2012-05-05T22:58:50
20	Spectrometer Frequency	400.18
21	Spectral Width	8223.7
22	Lowest Frequency	-1644.6
23	Nucleus	1H
	Acquired Size	
25	Spectral Size	65536



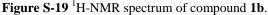


	Parameter	Value
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4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
7 8	Spectrometer Author	spect
9	Solvent	CDCl3
10	Temperature	295.8
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12	Experiment	1D
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22	Lowest Frequency	-1060.9
23	Nucleus	13C
24	Acquired Size	32768
25	Spectral Size	65536

Figure S-18¹³C-NMR spectrum of compound 1a.



ParameterValueParameterValue10/fdName10/fd2TitleJNPD-142/103CommentSecondant3CommentSecondant4OriginBruker BioSpin GmbH5Ownertmm6Site7SpectrometerDPX3008Author298.211Pulse2930Sequence1012Experiment1D13Number of16Scan1014Receiver Gain14415Relaxation1.000017Acquisition2012-05-13T20:46:3219Modification2012-05-13T20:46:3220Spectrometer30.13Frequencysitt41236.321Spectral41226.3Srequency14236.324Acquisition2012-05-13T20:46:3225Spectral526.3	-	_		
Name 10/ fid 2 Title JNPD-142/10 3 Comment 5-(Napht-2-y)hex-5-enoic acid PROTON CDCI3 (D:\ uio\DPX300-09) Jmolsoe 43 4 Origin Bruker BioSpin GmbH 5 5 Owner Imm 6 Site Filter 7 Spectrometer DPX300 8 Author 9 9 Solvent CDCI3 10 Temperature 298.2 11 Pulse 230 Sequence 213 12 Experiment 1D 13 Number of 16 Scans 14 Receiver Gain 14 Receiver Gain 144 15 Relaxation 1.0000 Delay 10.0000 17 7 Acquisition 2012-05-13T20:46:32 Date 20 Spectrameter 30.13 Frequency 2126.3 21 Spectrameter 30.13 Frequency 2126.	L		Parameter	Value
3 Comment 5-(Napht-2-yI)hex-5- enoic acid PROTON CDCI3 {D:1 uio\ DPX300-09} jmnolsoe 43 4 Origin Bruker BioSpin GmbH 5 Owner tnmr 6 Site - 7 Spectrometer DX300 8 Author - 9 Solvent CDCI3 10 Temperature 298.2 11 Pulse zg30 Sequence 12 Experiment 1D 13 Number of 16 Scans 14 Receiver Gain 144 15 Relaxation 1.0000 Delay 10.0000 17 Acquisition 2012-05-13T20:46:32 Date 20 Spectrometer 19 Modification 2012-05-13T20:46:32 Date 20 Spectrameter 10 Spectrameter 30.13 Frequency 123.6.3 21 Spectrameter 123.3 Width 1.236.3 Yeader 1236.3 23 Nucleus 1H	1	L		
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5 Owner tmm 6 Site - 7 Spectrometer DPX300 8 Author - 9 Solvent CDCI3 10 Temperature 298.2 11 Pulse 230 Sequence 10 - 12 Experiment 10 13 Number of 16 Scans - - 14 Receiver Gain 1.44 15 Relaxation 1.0000 Delay 10 0.03084 Time 30.13 - 17 Acquisition 2012-05-13T20:46:32 Date - 20 19 Modification 2012-05-13T20:46:32 Date - 20 20 Spectrameter 300.13 Frequency - 1236.3 21 Spectrameter - 23 Nucleus 1H 24	3	8	Comment	enoic acid PROTON CDCI3 {D:\ uio\ DPX300-09}
6 Site 7 Spectrometer DPX300 8 Author 9 9 Solvent CDCI3 10 Temperature 298.2 11 Pulse zg30 Sequence 10 10 12 Experiment 1D 13 Number of 16 Scans 1.0000 Delay 16 Pulse Width 10.0000 17 Acquisition 5.0304 Time 3004 10 18 Acquisition 2012-05-13T20:46:32 Date 20 Spectrometer 300.13 Frequency 30.13 Frequency 21 Spectral 172.8 Width -1236.3 Frequency 23 Nucleus 1H 24 Acquired Size 32768	4	ł	Origin	Bruker BioSpin GmbH
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Date 19 Modification 2012-05-13T20:46:32 Date 20 Spectrometer 300.13 Frequency 21 Spectral 6172.8 Width 22 Lowest -1236.3 Frequency 23 Nucleus 1H 24 Acquired Size 32768	1	17		5.3084
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Frequency 21 Spectral 6172.8 Width	1	9		2012-05-13T20:46:32
Width 22 Lowest -1236.3 Frequency 23 Nucleus 1H 24 Acquired Size 32768	2	20		300.13
Frequency 23 Nucleus 1H 24 Acquired Size 32768	2	21		6172.8
24 Acquired Size 32768	2	22		-1236.3
	2	23	Nucleus	1H
25 Spectral Size 65536	2	24	Acquired Size	32768
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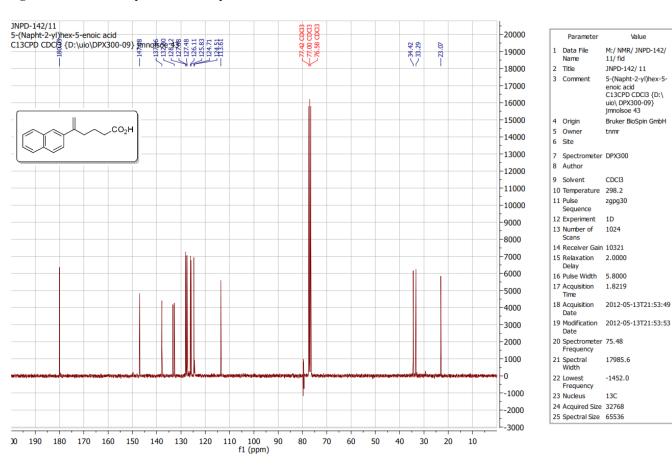
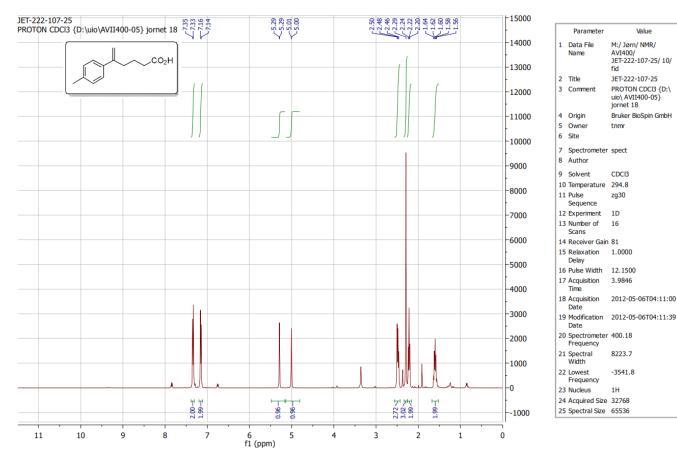
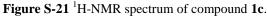
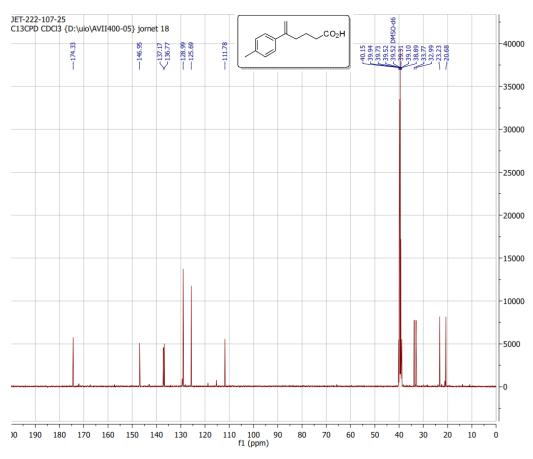


Figure S-20¹³C-NMR spectrum of compound 1b.

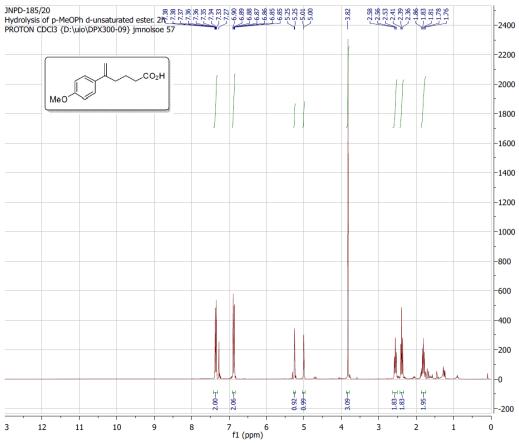






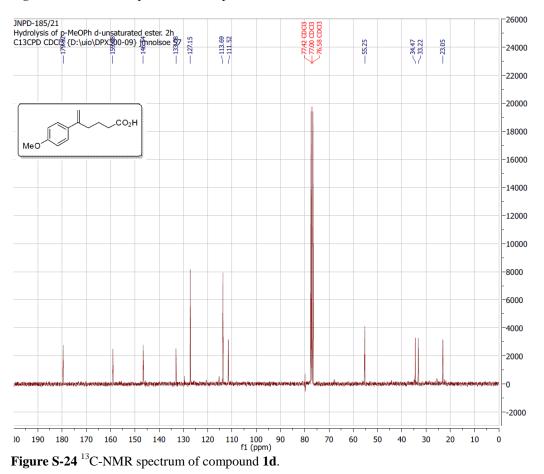
	Parameter	Value
1	Data File Name	M:/ Jørn/ NMR/ AVI400/ JET-222-107-25/ 11/ fid
2	Title	JET-222-107-25
3	Comment	C13CPD CDCl3 {D:\ uio\ AVII400-05} jornet 18
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
7	Spectrometer	spect
8	Author	
9	Solvent	CDCl3
10	Temperature	295.7
11	Pulse Sequence	zgpg30
12	Experiment	1D
13	Number of Scans	2048
14	Receiver Gain	724
15	Relaxation Delay	2.0000
16	Pulse Width	7.0000
17	Acquisition Time	1.2977
18	Acquisition Date	2012-05-06T06:08:00
19	Modification Date	2012-05-06T06:08:29
20	Spectrometer Frequency	100.64
21	Spectral Width	25252.5
22	Lowest Frequency	-1576.6
23	Nucleus	13C
24	Acquired Size	32768
25	Spectral Size	65536

Figure S-22 ¹³C-NMR spectrum of compound 1c.

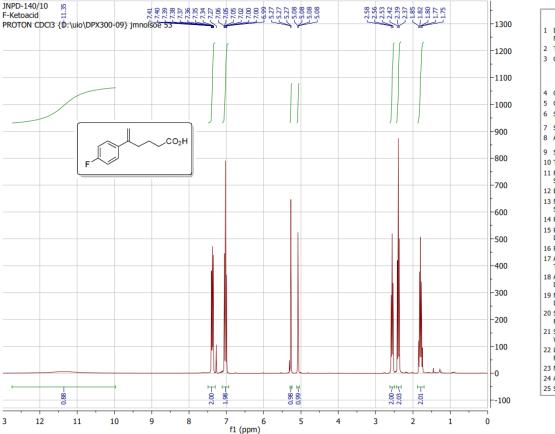


	Parameter	Value
1	Data File Name	M:/ NMR/ JNPD-185/ 20/ fid
2	Title	JNPD-185/ 20
3	Comment	Hydrolysis of p- MeOPh d-unsaturated ester. 2h PROTON CDCI3 {D:\ uio\ DPX300-09} jmnolsoe 57
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
7	Spectrometer	DPX300
	Author	
9	Solvent	CDCI3
-	Temperature	
	Pulse	zq30
	Sequence	-9
12	Experiment	1D
13	Number of Scans	16
14	Receiver Gain	228
15	Relaxation Delay	1.0000
16	Pulse Width	10.0000
17	Acquisition Time	5.3084
18	Acquisition Date	2012-09-01T20:12:35
19	Modification Date	2012-09-01T20:12:40
20	Spectrometer Frequency	300.13
21	Spectral Width	6172.8
22	Lowest Frequency	-1236.3
23	Nucleus	1H
	Acquired Size	
25	Spectral Size	65536



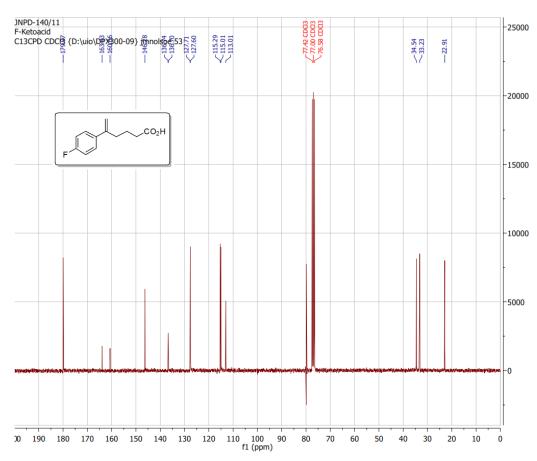


	Parameter	Value
1	Data File Name	M:/ NMR/ JNPD-185/ 21/ fid
2	Title	JNPD-185/ 21
3	Comment	Hydrolysis of p- MeOPh d-unsaturated ester. 2h C13CPD CDCl3 {D:\ uio\ DPX300-09} jmnolsoe 57
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
7	Spectrometer	DPX300
8	Author	
9	Solvent	CDCI3
10	Temperature	298.2
11	Pulse Sequence	zgpg30
12	Experiment	1D
13	Number of Scans	1024
14	Receiver Gain	13004
15	Relaxation Delay	2.0000
16	Pulse Width	5.8000
17	Acquisition Time	1.8219
18	Acquisition Date	2012-09-01T21:38:30
19	Modification Date	2012-09-01T21:38:33
20	Spectrometer Frequency	75.48
21	Spectral Width	17985.6
22	Lowest Frequency	-1448.2
23	Nucleus	13C
24	Acquired Size	32768
25	Spectral Size	65536



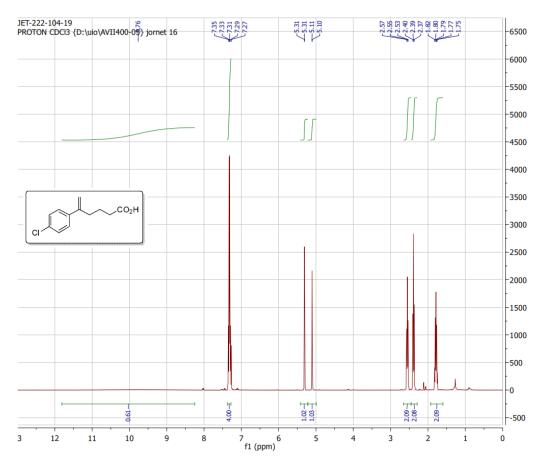
	Parameter	Value
1	Data File Name	M:/ NMR/ JNPD-140/ 10/ fid
2	Title	JNPD-140/ 10
3	Comment	F-Ketoacid PROTON CDCI3 {D:\ uio\ DPX300-09} jmnolsoe 53
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
	Spectrometer Author	DPX300
9	Solvent	CDCl3
10	Temperature	298.2
11	Pulse Sequence	zg30
12	Experiment	1D
13	Number of Scans	16
14	Receiver Gain	181
15	Relaxation Delay	1.0000
16	Pulse Width	10.0000
17	Acquisition Time	5.3084
18	Acquisition Date	2012-05-10T11:33:59
19	Modification Date	2012-05-10T11:34:03
20	Spectrometer Frequency	300.13
21	Spectral Width	6172.8
22	Lowest Frequency	-1236.0
23	Nucleus	1H
	Acquired Size	
25	Spectral Size	65536

Figure S-25 ¹H-NMR spectrum of compound 1e.



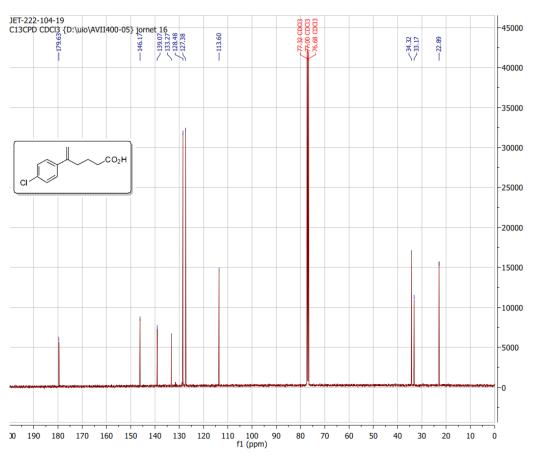
	Parameter	Value
1	Data File Name	M:/ NMR/ JNPD-140/ 11/ fid
2	Title	JNPD-140/ 11
3	Comment	F-Ketoacid C13CPD CDCl3 {D:\ uio\ DPX300-09} jmnolsoe 53
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
7	Spectrometer	DPX300
	Author	
9	Solvent	CDCI3
10	Temperature	298.2
11	Pulse Sequence	zgpg30
12	Experiment	1D
13	Number of Scans	1024
14	Receiver Gain	11585
15	Relaxation Delay	2.0000
16	Pulse Width	5.8000
17	Acquisition Time	1.8219
18	Acquisition Date	2012-05-10T12:41:22
19	Modification Date	2012-05-10T12:41:25
20	Spectrometer Frequency	75.48
21	Spectral Width	17985.6
22	Lowest Frequency	-1448.2
23	Nucleus	13C
24	Acquired Size	32768
25	Spectral Size	65536

Figure S-26¹³C-NMR spectrum of compound 1e.



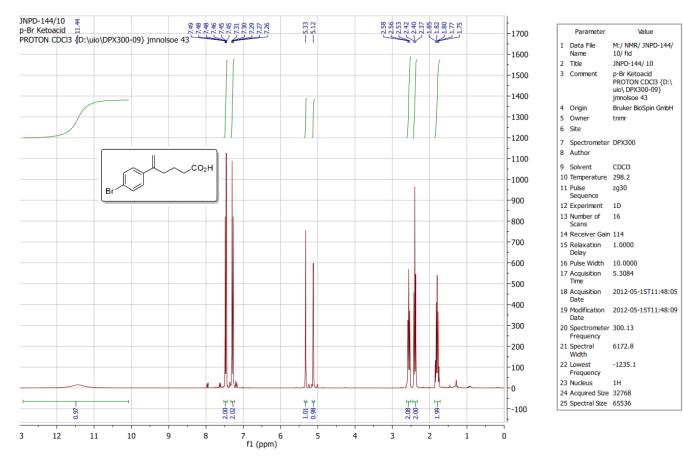
_		
	Parameter	Value
1	Data File Name	M:/ Jørn/ NMR/ AVI400/ JET-222-104-19/ 10/ fid
2	Title	JET-222-104-19
3	Comment	PROTON CDCl3 {D:\ uio\ AVII400-05} jornet 16
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
	Spectrometer Author	spect
9	Solvent	CDCl3
10	Temperature	294.9
11	Pulse Sequence	zg30
12	Experiment	1D
13	Number of Scans	16
14	Receiver Gain	72
15	Relaxation Delay	1.0000
16	Pulse Width	12.1500
17	Acquisition Time	3.9846
18	Acquisition Date	2012-05-06T00:05:00
19	Modification Date	2012-05-06T00:05:22
20	Spectrometer Frequency	400.18
21	Spectral Width	8223.7
22	Lowest Frequency	-1644.6
23	Nucleus	1H
24	Acquired Size	32768
25	Spectral Size	65536

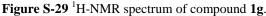


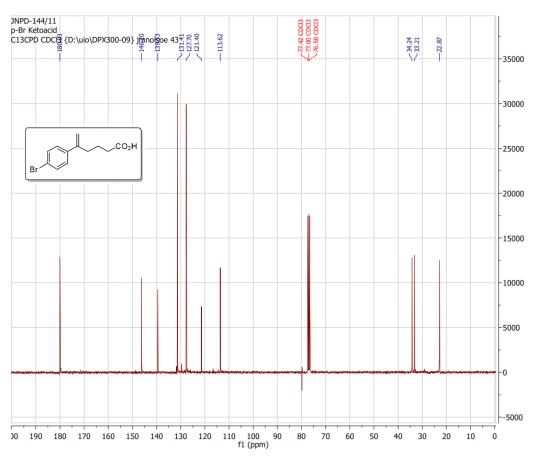


	Parameter	Value
1	Data File Name	M:/ Jørn/ NMR/ AVI400/ JET-222-104-19/ 11/ fid
2	Title	JET-222-104-19
3	Comment	C13CPD CDCl3 {D:\ uio\ AVII400-05} jornet 16
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
	Spectrometer Author	spect
9	Solvent	CDCl3
10	Temperature	295.7
11	Pulse Sequence	zgpg30
12	Experiment	1D
13	Number of Scans	2048
14	Receiver Gain	1620
15	Relaxation Delay	2.0000
16	Pulse Width	7.0000
17	Acquisition Time	1.2977
18	Acquisition Date	2012-05-06T02:02:00
19	Modification Date	2012-05-06T02:02:12
20	Spectrometer Frequency	100.64
21	Spectral Width	25252.5
22	Lowest Frequency	-1057.2
23	Nucleus	13C
24	Acquired Size	32768
25	Spectral Size	65536

Figure S-28 ¹³C-NMR spectrum of compound 1f.

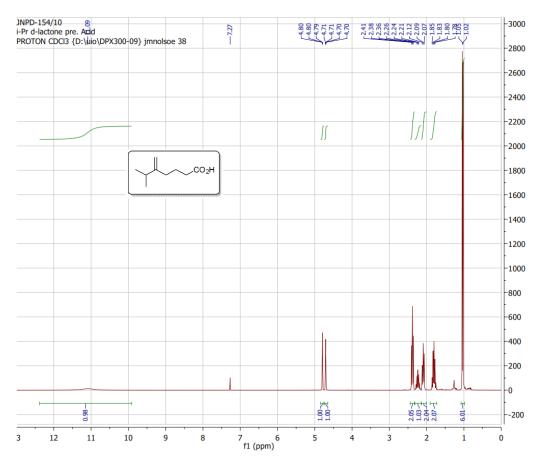




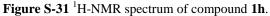


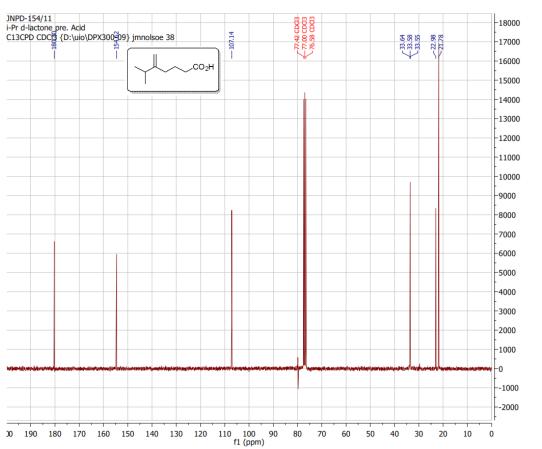
	Parameter	Value
1	Data File Name	M:/ NMR/ JNPD-144/ 11/ fid
2	Title	JNPD-144/ 11
3	Comment	p-Br Ketoacid C13CPD CDCl3 {D:\ uio\ DPX300-09} jmnolsoe 43
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
7	Spectrometer	DPX300
8	Author	
9	Solvent	CDCl3
10	Temperature	298.2
11	Pulse Sequence	zgpg30
12	Experiment	1D
13	Number of Scans	1024
14	Receiver Gain	11585
15	Relaxation Delay	2.0000
16	Pulse Width	5.8000
17	Acquisition Time	1.8219
18	Acquisition Date	2012-05-15T12:55:32
19	Modification Date	2012-05-15T12:55:35
20	Spectrometer Frequency	75.48
21	Spectral Width	17985.6
22	Lowest Frequency	-1451.5
23	Nucleus	13C
24	Acquired Size	32768
25	Spectral Size	65536

Figure S-30 ¹³C-NMR spectrum of compound 1g.



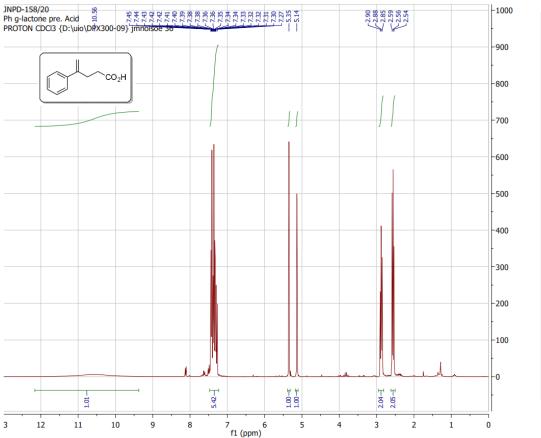
	Parameter	Value
1	Data File Name	M:/ NMR/ JNPD-154/ 10/ fid
2	Title	JNPD-154/ 10
3	Comment	i-Pr d-lactone pre. Acid PROTON CDCl3 {D: uio\ DPX300-09} jmnolsoe 38
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
L.	Spectrometer Author	DPX300
9	Solvent	CDCI3
10	Temperature	298.2
11	Pulse Sequence	zg30
12	Experiment	1D
13	Number of Scans	16
14	Receiver Gain	114
15	Relaxation Delay	1.0000
16	Pulse Width	10.0000
17	Acquisition Time	5.3084
18	Acquisition Date	2012-06-06T14:42:47
19	Modification Date	2012-06-06T14:42:52
20	Spectrometer Frequency	300.13
21	Spectral Width	6172.8
22	Lowest Frequency	-1236.3
23	Nucleus	1H
24	Acquired Size	32768
25	Spectral Size	65536



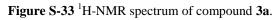


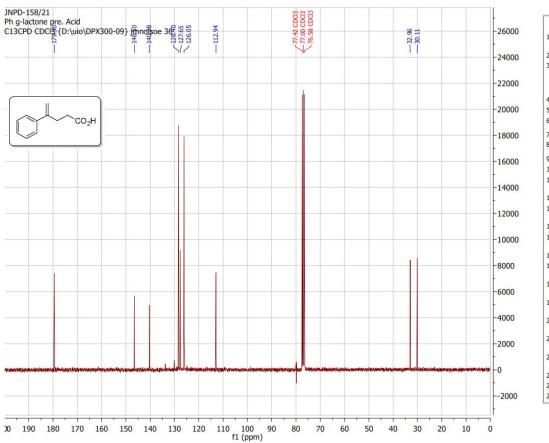
	Parameter	Value
1	Data File Name	M:/ NMR/ JNPD-154/ 11/ fid
2	Title	JNPD-154/ 11
3	Comment	i-Pr d-lactone pre. Acid C13CPD CDCl3 {D:\ uio\ DPX300-09} jmnolsoe 38
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
	Spectrometer Author	DPX300
9	Solvent	CDCl3
10	Temperature	298.2
11	Pulse Sequence	zgpg30
12	Experiment	1D
13	Number of Scans	1024
14	Receiver Gain	10321
15	Relaxation Delay	2.0000
16	Pulse Width	5.8000
17	Acquisition Time	1.8219
18	Acquisition Date	2012-06-06T15:50:10
19	Modification Date	2012-06-06T15:50:14
20	Spectrometer Frequency	75.48
21	Spectral Width	17985.6
22	Lowest Frequency	-1446.8
23	Nucleus	13C
24	Acquired Size	32768
25	Spectral Size	65536

Figure S-32 ¹³C-NMR spectrum of compound 1h.



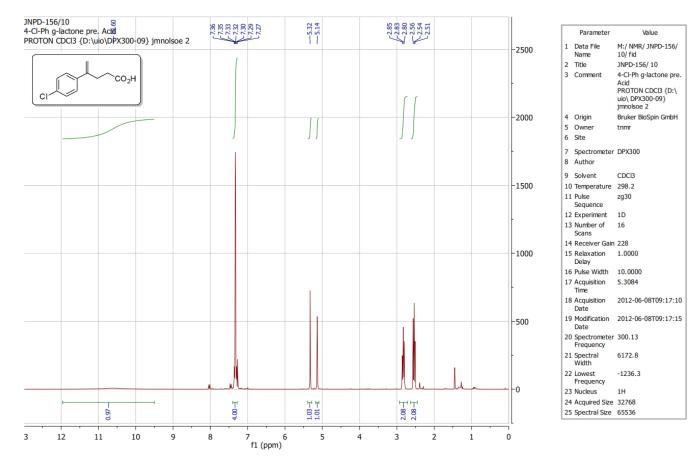
	Parameter	Value
1	Data File Name	M:/ NMR/ JNPD-158/ 20/ fid
2	Title	JNPD-158/ 20
3	Comment	Ph g-lactone pre. Acid PROTON CDCI3 {D:\ uio\ DPX300-09} jmnolsoe 36
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
7 8	Spectrometer Author	DPX300
9	Solvent	CDCl3
10	Temperature	298.2
11	Pulse Sequence	zg30
12	Experiment	1D
13	Number of Scans	16
14	Receiver Gain	203
15	Relaxation Delay	1.0000
16	Pulse Width	10.0000
17	Acquisition Time	5.3084
18	Acquisition Date	2012-06-11T09:07:17
19	Modification Date	2012-06-11T09:07:22
20	Spectrometer Frequency	300.13
21	Spectral Width	6172.8
22	Lowest Frequency	-1236.3
23	Nucleus	1H
	Acquired Size	
25	Spectral Size	65536

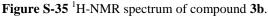


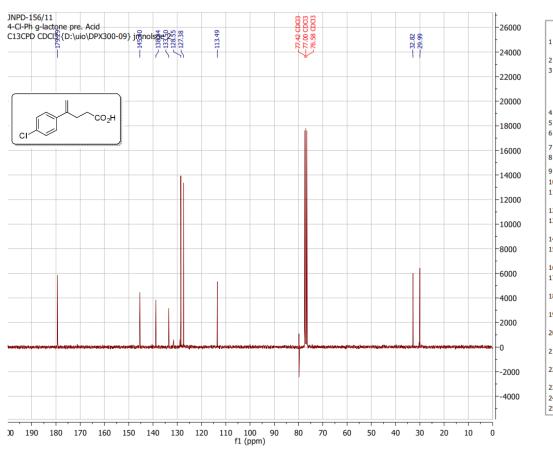


	Parameter	Value
	Data File Name	M:/ NMR/ JNPD-158/ 21/ fid
2	Title	JNPD-158/21
3	Comment	Ph g-lactone pre. Acid C13CPD CDCl3 {D: uio\ DPX300-09} jmnolsoe 36
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
7	Spectrometer	DPX300
8	Author	
9	Solvent	CDCI3
10	Temperature	298.2
11	Pulse Sequence	zgpg30
12	Experiment	1D
13	Number of Scans	
14	Receiver Gain	13004
15	Relaxation Delay	2.0000
16	Pulse Width	5.8000
17	Acquisition Time	1.8219
18	Acquisition Date	2012-06-11T10:14:40
19	Modification Date	2012-06-11T10:14:44
20	Spectrometer Frequency	75.48
21	Spectral Width	17985.6
22	Lowest Frequency	-1449.4
23	Nucleus	13C
24	Acquired Size	32768
25	Spectral Size	65536

Figure S-34 ¹³C-NMR spectrum of compound 3a.







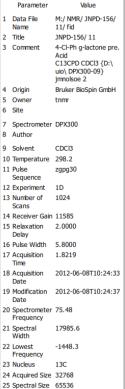


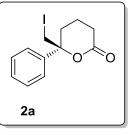
Figure S-36¹³C-NMR spectrum of compound 3b.

Enantioselective Iodolactonization Utilizing Chiral Squaramides

General procedure for asymmetric iodolactonization with chiral squaramides: Iodine (0.15 equiv.) and *N*-iodosuccinimide (1.0 equiv.) was dissolved in a combination of acetone/CH₂Cl₂ (1:1, 0.20 M). Subsequently, the squaramide **5-8** (0.15 equiv.) was added and the resulting mixture was cooled to -78 °C. A solution of unsaturated acid **1** or **3** (1.0 equiv.) in a combination of acetone/CH₂Cl₂ (1:1, 0.20 M) was added and the resulting mixture was treated with satd. aq. Na₂S₂O₃ (10 ml) while still in the cold, allowed to equilibrate to ambient temperature and then EtOAc (30 ml) was added. The phases were separated and the organic phase was washed with aq. NaOH (2 x 20 ml, 1.0 M) and brine (20 ml). The organic phase was dried over MgSO₄, filtered and evaporated *in vacuo*. The residue was purified by column chromatography on silica (hexanes/EtOAc 4:1) to afford the corresponding iodolactone **2** or **4**.

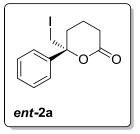
Notice! The iodolactones were observed to be very labile in the condensed state under vacuum. Thus, to avoid decomposition, great care had to be taken when evaporating the solvent *in vacuo* after isolation by flash chromatography. Once all visible traces of solvent had been removed it was of paramount importance to equilibrate back to ambient pressure immediately. Without this precaution, the isolated iodolactone would turn black spontaneously. Due to the instability of the iodolactones they were stored under argon and refrigerated.

(S)-6-(Iodomethyl)-6-phenyltetrahydro-2*H*-pyran-2-one (2a).⁴



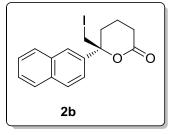
Prepared according to the general procedure using 5-phenylhex-5-enoic acid (**1a**) (40 mg, 210 µmol), squaramide **6b** (17 mg, 31 µmol), iodine (8 mg, 31 µmol) and *N*-iodosuccinimide (47 mg, 210 µmol). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes/ⁱPrOH 98:2, 1 mL/min): $t_r(e_1, \text{ major}) = 24.04 \text{ min and } t_r(e_2, \text{ minor}) = 27.20 \text{ min. Yield: 55 mg (83%) of colourless oil;$ *ee* $: 87%; TLC (hexanes/EtOAc 1:1): <math>R_f = 0.48$, visualized with anisaldehyde; $[\alpha]_D^{20} = 27.2$ (c = 0.06, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.30 – 7.54 (m, 5H), 3.57 (d, J = 1.4 Hz, 2H), 2.28 – 2.59 (m, 4H), 1.69 – 1.98 (m, 1H), 1.45 – 1.69 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 170.4, 140.1, 128.9 (2C), 128.3, 125.1 (2C), 84.3, 32.0, 28.9, 17.7, 16.5; HRMS (EI): Exact mass calculated for $C_{12}H_{13}IO_2[M]^+$: 315.9960, found 315.9972.

(R)-6-(Iodomethyl)-6-phenyltetrahydro-2H-pyran-2-one (ent-2a).⁴



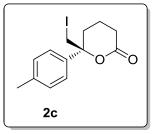
Prepared according to the general procedure using 5-phenylhex-5-enoic acid (**1a**) (40 mg, 210 µmol), squaramide *ent-6b* (17 mg, 31 µmol), iodine (8 mg, 31 µmol) and *N*-iodosuccinimide (47 mg, 210 µmol). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes/ⁱPrOH 98:2, 1 mL/min): $t_r(e_1, \text{minor}) = 27.00 \text{ min}$ and $t_r(e_2, \text{major}) = 31.00 \text{ min}$. Yield: 52 mg (79%) of colourless oil; *ee*: 82%; TLC (hexanes/EtOAc 1:1): $R_f = 0.48$, visualized with anisaldehyde; $[\alpha]_D^{20} = -26.0$ (c = 0.07, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.30 – 7.54 (m, 5H), 3.57 (d, J = 1.4 Hz, 2H), 2.28 – 2.59 (m, 4H), 1.69 – 1.98 (m, 1H), 1.45 – 1.69 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 170.4, 140.1, 128.9 (2C), 128.3, 125.1 (2C), 84.3, 32.0, 28.9, 17.7, 16.5.

(S)-6-(Iodomethyl)-6-(naphthalen-2-yl)tetrahydro-2H-pyran-2-one (2b).⁴



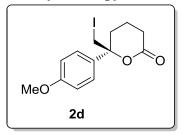
Prepared according to the general procedure using 5-(naphthalen-2-yl)hex-5-enoic acid (**1b**) (40 mg, 166 µmol), squaramide **6b** (14 mg, 25 µmol), iodine (7 mg, 25 µmol) and *N*-iodosuccinimide (37 mg, 166 µmol). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes/ⁱPrOH 99:1, 1 mL/min): $t_r(e_1, \text{ major}) = 66.14 \text{ min and } t_r(e_2, \text{ minor}) = 82.74 \text{ min. Yield: 55 mg (91%) of colourless oil;$ *ee* $: 92%; TLC (hexanes/EtOAc 1:1): <math>R_f = 0.56$, visualized with anisaldehyde; $[\alpha]_D^{20} = 22.0$ (c = 0.13, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.81 – 7.97 (m, 4H), 7.48 – 7.62 (m, 2H), 7.41 (dd, J = 8.6, 2.0 Hz, 1H), 3.67 (d, J = 2.8 Hz, 2H), 2.29 – 2.77 (m, 4H), 1.78 –1.93 (m, 1H), 1.52 –1.69 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 170.5, 137.4, 133.0, 132.8, 129.0, 128.3, 127.5, 126.8 (2C), 125.0, 122.3, 84.6, 32.1, 29.0, 17.4, 16.6; HRMS (EI): Exact mass calculated for C₁₆H₁₅IO₂ [*M*]⁺: 366.0117, found 366.0109.

(S)-6-(Iodomethyl)-6-(p-tolyl)tetrahydro-2H-pyran-2-one (2c).⁴



Prepared according to the general procedure using 5-(*p*-tolyl)hex-5-enoic acid (**1c**) (40 mg, 196 µmol), squaramide **6b** (17 mg, 29 µmol), iodine (7 mg, 29 µmol) and *N*-iodosuccinimide (44 mg, 196 µmol). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes/ⁱPrOH 98:2, 1 mL/min): $t_r(e_1, \text{ major}) = 25.32 \text{ min and } t_r(e_2, \text{ minor}) = 34.16 min. Yield: 51 mg (80%) of colourless oil;$ *ee* $: 86%; TLC (hexanes/EtOAc 1:1): <math>R_f = 0.56$, visualized with anisaldehyde; $[\alpha]_D^{20} = 24.6$ (c = 0.10, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.25 (d, *J* = 8.4 Hz, 2H), 7.20 (d, *J* = 8.2 Hz, 2H), 3.55 (s, 2H), 2.40 – 2.54 (m, 2H), 2.35 (s, 3H), 2.27 – 2.40 (m, 2H), 1.74 – 1.89 (m, 1H), 1.66 – 1.49 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 170.5, 138.2, 137.1, 129.6 (2C), 125.1 (2C), 84.3, 31.9, 28.9, 21.0, 17.9, 16.5; HRMS (EI): Exact mass calculated for C₁₃H₁₅IO₂ [*M*]⁺: 330.0117, found 330.0108.

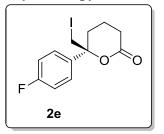
(S)-6-(Iodomethyl)-6-(4-methoxyphenyl)tetrahydro-2H-pyran-2-one (2d).⁴



Prepared according to the general procedure using 5-(4-methoxyphenyl)hex-5-enoic acid (**1d**) (48 mg, 218 µmol), squaramide **6b** (18 mg, 33 µmol), iodine (8 mg, 33 µmol) and *N*-iodosuccinimide (49 mg, 218 µmol). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes/^{*i*}PrOH 90:10, 1 mL/min): $t_r(e_1, \text{ major}) = 12.91 \text{ min and } t_r(e_2, \text{ minor}) = 15.56 \text{ min. Yield: 65 mg (87%) of colourless oil;$ *ee* $: 12%; TLC (hexanes/EtOAc 1:1): <math>R_f = 0.53$, visualized with anisaldehyde; $[\alpha]_D^{20} = 1.6$ (c = 0.06, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.24 – 7.33 (m, 2H), 6.86 – 6.95 (m, 2H), 3.81 (s, 3H), 3.50 – 3.58 (m, 2H), 2.39 – 2.55 (m, 2H), 2.25 – 2.39 (m, 2H), 1.75 – 1.90 (m, 1H), 1.48 – 1.70

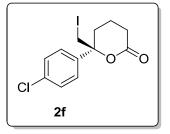
(m, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 170.5, 159.4, 132.0, 126.5 (2C), 114.2 (2C), 84.2, 55.3, 31.8, 28.9, 18.1, 16.5; HRMS (EI): Exact mass calculated for C₁₃H₁₅IO₃ [*M*]⁺: 346.0066, found 346.0064.

(S)-6-(4-Fluorophenyl)-6-(iodomethyl)tetrahydro-2H-pyran-2-one (2e).⁴



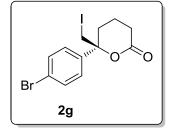
Prepared according to the general procedure using 5-(4-fluorophenyl)hex-5-enoic acid (**1e**) (42 mg, 200 µmol), squaramide **6b** (17 mg, 30 µmol), iodine (8 mg, 30 µmol) and *N*-iodosuccinimide (45 mg, 200 µmol). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes/¹PrOH 90:10, 1 mL/min): $t_r(e_1, \text{ major}) = 10.70 \text{ min and } t_r(e_2, \text{ minor}) = 12.17 min. Yield: 55 mg (83%) of colourless oil;$ *ee* $: 90%; TLC (hexanes/EtOAc 1:1): <math>R_f = 0.49$, visualized with anisaldehyde; $[\alpha]_D^{20} = 26.0 \text{ (c} = 0.10, \text{ CHCl}_3)$; ¹H NMR (300 MHz, CDCl₃) δ 7.29 – 7.47 (m, 2H), 7.02 – 7.17 (m, 2H), 3.54 (s, 1H), 2.41 – 2.58 (m, 2H), 2.41 – 2.26 (m, 2H), 1.76 – 1.93 (m, 1H), 1.48 – 1.71 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 170.1, 162.4 (d, ¹ $J_{CF} = 248 \text{ Hz}$), 136.0 (d, ⁴ $J_{CF} = 3.2 \text{ Hz}$), 127.1 (d, ³ $J_{CF} = 8.3 \text{ Hz}, 2C$), 115.9 (d, ² $J_{CF} = 21.5 \text{ Hz}, 2C$), 84.1, 31.9, 28.9, 17.5, 16.5; HRMS (EI): Exact mass calculated for C₁₂H₁₂FIO₂ [M]⁺: 333.9866, found 333.9867.

(S)-6-(4-Chlorophenyl)-6-(iodomethyl)tetrahydro-2*H*-pyran-2-one (2f).⁴



Prepared according to the general procedure using 5-(4-chlorophenyl)hex-5-enoic acid (**1f**) (40 mg, 178 µmol), squaramide **6b** (15 mg, 27 µmol), iodine (7 mg, 27 µmol) and *N*-iodosuccinimide (40 mg, 178 µmol). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes/ⁱPrOH 98:2, 1 mL/min): $t_r(e_1, \text{ major}) = 31.58 \text{ min and } t_r(e_2, \text{ minor}) = 46.09 \text{ min. Yield: 49 mg (78%) of colourless oil;$ *ee* $: 96%; TLC (hexanes/EtOAc 1:1): <math>R_f = 0.46$, visualized with anisaldehyde; $[\alpha]_D^{20} = 27.2$ (c = 0.20, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.38 (d, *J* = 8.8 Hz, 2H), 7.32 (d, *J* = 8.8 Hz, 2H), 2.42 – 2.59 (m, 2H), 2.27 –2.41 (m, 2H), 1.78 – 1.93 (m, 1H), 1.50 – 1.66 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 170.0, 138.8, 134.5, 129.1 (2C), 126.7 (2C), 84.1, 32.0, 29.0, 17.1, 16.5; HRMS (EI): Exact mass calculated for C₁₂H₁₂CIIO₂ [*M*]⁺: 349.9571, found 349.9577.

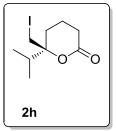
(S)-6-(4-Bromophenyl)-6-(iodomethyl)tetrahydro-2H-pyran-2-one (2g).⁴



Prepared according to the general procedure using 5-(4-bromophenyl)hex-5-enoic acid (**1g**) (22 mg, 80 µmol), squaramide **6b** (7 mg, 12 µmol), iodine (3 mg, 27 µmol) and *N*-iodosuccinimide (18 mg, 178 µmol). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes/^{*i*}PrOH 90:10, 1 mL/min): $t_r(e_1, major) = 11.83$ min and $t_r(e_2, minor) = 15.15$

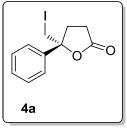
min. Yield: 23 mg (73%) of colourless oil; *ee*: 91%; TLC (hexanes/EtOAc 1:1): $R_f = 0.50$, visualized with anisaldehyde; $[\alpha]_D^{20} = 23.3$ (c = 0.15, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.54 (d, J = 8.6 Hz, 2H), 7.26 (d, J = 8.7 Hz, 2H), 3.54 (s, 2H), 2.42 – 2.57 (m, 2H), 2.28 – 2.42 (m, 2H), 1.78 – 1.92 (m, 1H), 1.50 – 1.70 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 170.0, 139.4, 132.1 (2C), 127.0 (2C), 122.6, 84.1, 31.9, 29.0, 16.9, 16.5; HRMS (EI): Exact mass calculated for C₁₂H₁₂BrIO₂ [M]⁺: 393.9065, found 393.9070.

(S)-6-(Iodomethyl)-6-isopropyltetrahydro-2*H*-pyran-2-one (2h).⁴,



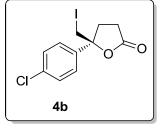
Prepared according to the general procedure using 6-methyl-5-methyleneheptanoic acid (**1h**) (29 mg, 186 µmol), squaramide **6b** (16 mg, 28 µmol), iodine (7 mg, 28 µmol) and *N*-iodosuccinimide (42 mg, 186 µmol). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral GLC analysis (Chiraldex G-TA, 170 °C isothermal): $t_r(e_1, \text{ major}) = 22.56 \text{ min and } t_r(e_2, \text{ minor}) = 22.91 \text{ min}$. Yield: 40 mg (77%) of colourless oil; *ee*: 16%; TLC (hexanes/EtOAc 1:1): $R_f = 0.57$, visualized with anisaldehyde; $[\alpha]_D^{20} = 1.50 \text{ (c} = 0.07, \text{CHCl}_3)$; ¹H NMR (400 MHz, CDCl₃) δ 3.42 (s, 2H), 2.45–2.59 (m, 1H), 2.31–2.45 (m, 1H), 2.09 – 2.25 (m, 1H), 1.95 – 2.06 (m, 1H), 1.76 – 1.95 (m, 3H), 0.97 (d, *J* = 7.0 Hz, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 170.4, 84.8, 35.2, 29.5, 26.7, 16.7, 16.6, 16.4, 13.0; HRMS (EI): Exact mass calculated for C₉H₁₅IO₂ [*M*]⁺: 282.0119, found 282.0124.

(*R*)-5-(Iodomethyl)-5-phenyldihydrofuran-2(3*H*)-one (4a).^{4, 6}



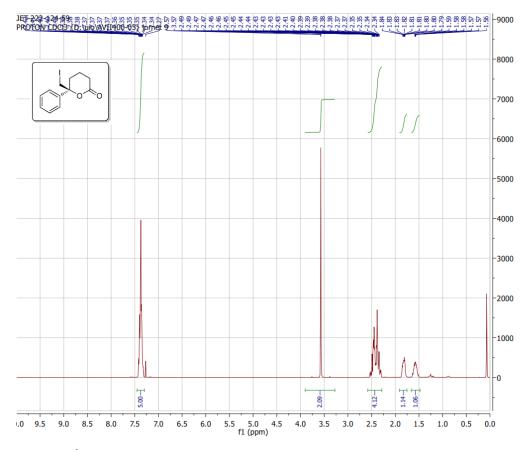
Prepared according to the general procedure using 4-phenylpent-4-enoic acid (**3a**) (40 mg, 227 µmol), squaramide **6b** (19 mg, 34 µmol), iodine (9 mg, 34 µmol) and *N*-iodosuccinimide (51 mg, 227 µmol). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC analysis (Chiralpak AD-H, hexanes/ⁱPrOH 98:2, 1 mL/min): $t_r(e_1, \text{ minor}) = 20.69 \text{ min}$ and $t_r(e_2, \text{ major}) = 23.69 \text{ min}$. Yield: 59 mg (86%) of colourless oil; *ee*: 7%; TLC (hexanes/EtOAc 1:1): $R_f = 0.55$, visualized with anisaldehyde; $[\alpha]_D^{20} = 5.70$ (c = 0.02, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.31 – 7.49 (m, 5H), 3.64 (d, *J* = 0.9 Hz, 2H), 2.41 – 2.88 (m, 4H); ¹³C NMR (101 MHz, CDCl₃) δ 175.3, 140.5, 128.8 (2C), 128.5, 124.8 (2C), 86.0, 33.9, 29.2, 16.3; HRMS (EI): Exact mass calculated for C₁₁H₁₁IO₂ [*M*]⁺: 301.9804, found 301.9793.

(S)-5-(4-Chlorophenyl)-5-(iodomethyl)dihydrofuran-2(3H)-one (4b).⁷

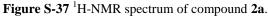


Prepared according to the general procedure using 4-(4-chlorophenyl)pent-4-enoic acid (**3b**) (42 mg, 200 μ mol), squaramide **6b** (17 mg, 30 μ mol), iodine (8 mg, 30 μ mol) and *N*-iodosuccinimide (45 mg, 200 μ mol). Purified by column chromatography on silica (hexanes/EtOAc 4:1). The enantiomeric excess was determined by chiral HPLC

analysis (Chiralpak AD-H, hexanes/ⁱPrOH 98:2, 1 mL/min): $t_r(e_1, \text{ major}) = 25.97 \text{ min and } t_r(e_2, \text{ minor}) = 27.98 \text{ min.}$ Yield: 57 mg (85%) of colourless oil; *ee*: 14%; TLC (hexanes/EtOAc 1:1): $R_f = 0.52$, visualized with anisaldehyde; $[\alpha]_D = 0.0 \text{ (c} = 0.21, \text{ CHCl}_3)$; ¹H NMR (300 MHz, CDCl₃) δ 7.30 – 7.43 (m, 4H), 3.59 (s, 2H), 2.65 – 2.88 (m, 2H), 2.43 – 2.65 (m, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 174.9, 139.1, 134.6, 129.0 (2C), 126.3 (2C), 85.6, 33.8, 29.1, 15.7; HRMS (EI): Exact mass calculated for C₁₁H₁₀ClIO₂ [*M*]⁺: 335.9414, found 335.9407.



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5	Owner	tnmr
6	Site	
7	Spectrometer	spect
8	Author	
9	Solvent	CDCI3
10	Temperature	295.0
11	Pulse Sequence	zg30
12	Experiment	1D
13	Number of Scans	16
14	Receiver Gain	32
15	Relaxation Delay	1.0000
16	Pulse Width	12.1500
17	Acquisition Time	3.9846
18	Acquisition Date	2012-06-18T17:26:00
19	Modification Date	2012-06-18T17:26:26
20	Spectrometer Frequency	400.18
21	Spectral Width	8223.7
22	Lowest Frequency	-1644.9
23	Nucleus	1H
24	Acquired Size	32768
25	Spectral Size	65536



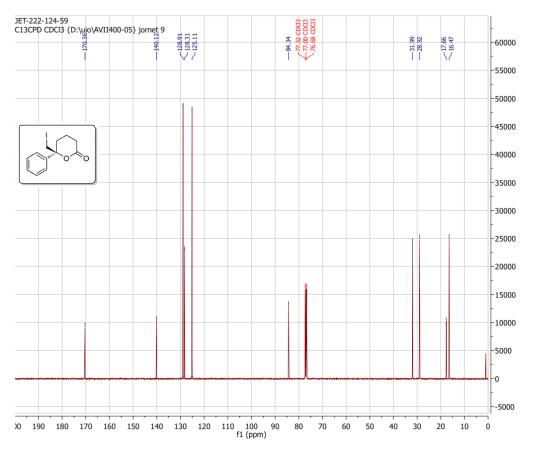
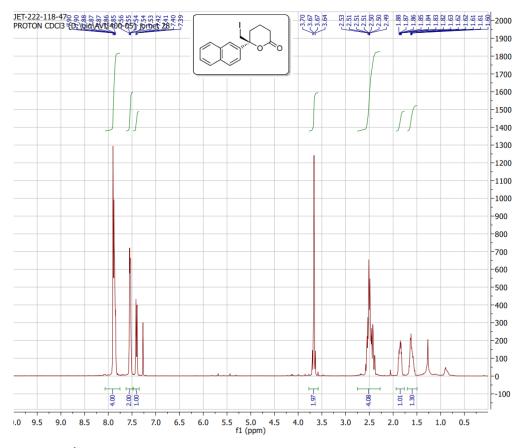
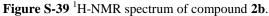


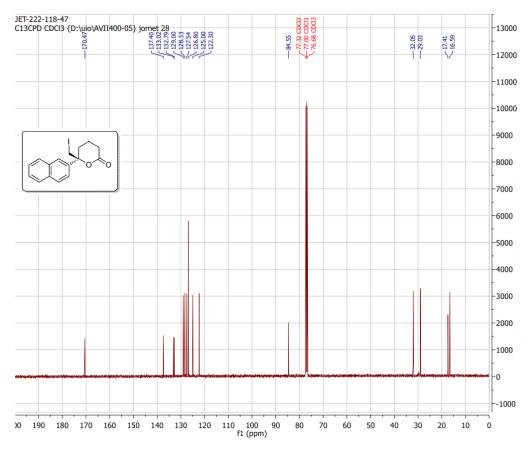


Figure S-38 ¹³C-NMR spectrum of compound 2a.



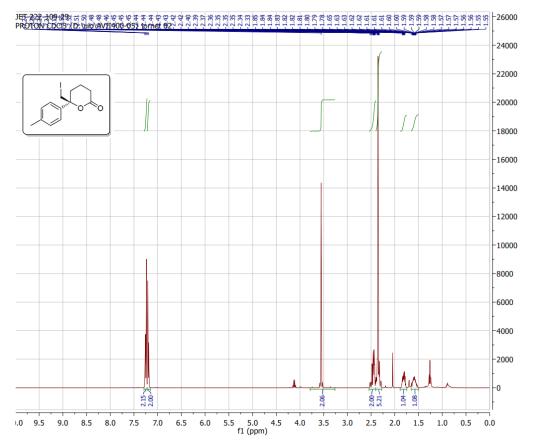
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4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
	Spectrometer Author	spect
9	Solvent	CDCI3
10	Temperature	295.2
	Pulse Sequence	zg30
12	Experiment	1D
13	Number of Scans	16
14	Receiver Gain	36
15	Relaxation Delay	1.0000
16	Pulse Width	12.1500
17	Acquisition Time	3.9846
18	Acquisition Date	2012-06-12T10:27:00
19	Modification Date	2012-06-12T10:28:05
20	Spectrometer Frequency	400.18
21	Spectral Width	8223.7
22	Lowest Frequency	-1645.0
23	Nucleus	1H
24	Acquired Size	32768
25	Spectral Size	65536



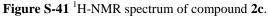


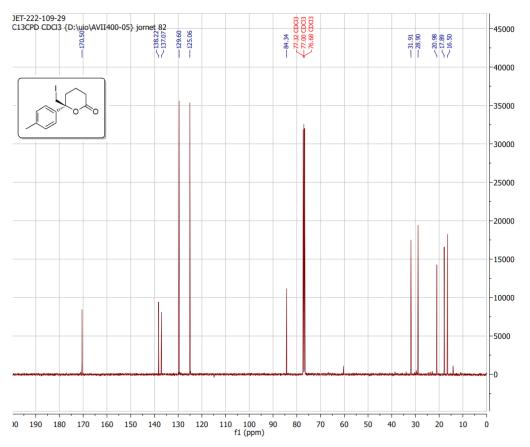
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4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
7	Spectrometer	spect
8	Author	
9	Solvent	CDCl3
10	Temperature	296.2
11	Pulse Sequence	zgpg30
12	Experiment	1D
13	Number of Scans	1024
14	Receiver Gain	724
15	Relaxation Delay	2.0000
16	Pulse Width	7.0000
17	Acquisition Time	1.2977
18	Acquisition Date	2012-06-12T11:27:0
19	Modification Date	2012-06-12T11:27:3
20	Spectrometer Frequency	100.64
21	Spectral Width	25252.5
22	Lowest Frequency	-1059.6
23	Nucleus	13C
24	Acquired Size	32768
25	Spectral Size	65536

Figure S-40 ¹³C-NMR spectrum of compound 2b.



	Parameter	Value
1	Data File Name	M:/ Jørn/ NMR/ AVI400/ JET-222-109-29/ 10/ fid
2	Title	JET-222-109-29
3		PROTON CDCl3 {D:\ uio\ AVII400-05} jornet 82
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
7	Spectrometer	spect
8	Author	
9	Solvent	CDCI3
10	Temperature	294.6
11	Pulse Sequence	zg30
12	Experiment	1D
13	Number of Scans	16
14	Receiver Gain	90
15	Relaxation Delay	1.0000
16	Pulse Width	12.1500
17	Acquisition Time	3.9846
18	Acquisition Date	2012-05-10T01:04:00
19	Modification Date	2012-05-10T01:04:58
20	Spectrometer Frequency	400.18
21	Spectral Width	8223.7
22	Lowest Frequency	-1644.6
23	Nucleus	1H
24	Acquired Size	32768
	Spectral Size	65526





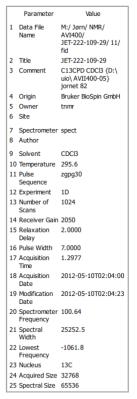
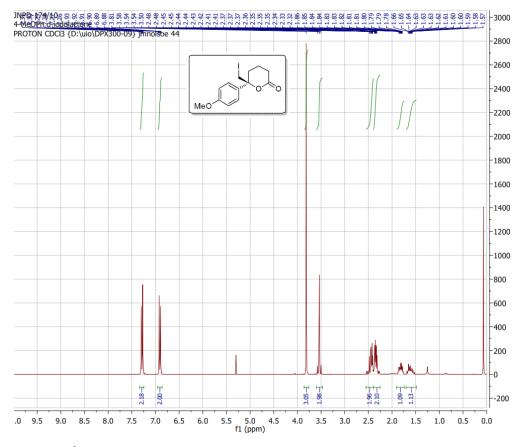
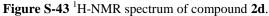
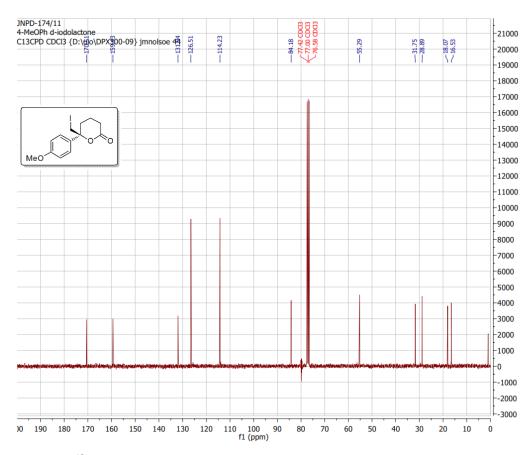


Figure S-42 ¹³C-NMR spectrum of compound **2c**.



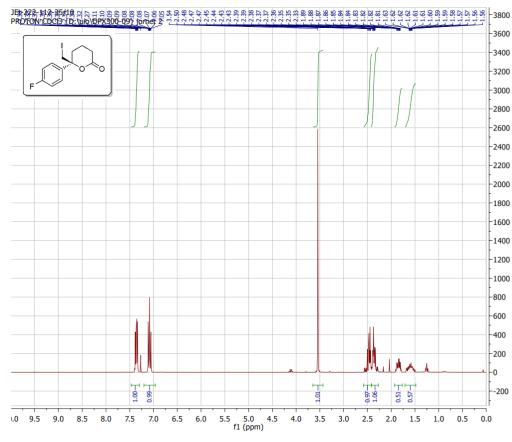
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4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
1.	Spectrometer Author	DPX300
9	Solvent	CDCI3
10	Temperature	298.2
	Pulse	zq30
	Sequence	
12	Experiment	1D
13	Number of Scans	16
14	Receiver Gain	228
15	Relaxation Delay	1.0000
16	Pulse Width	10.0000
17	Acquisition Time	5.3084
18	Acquisition Date	2012-07-15T18:10:16
19	Modification Date	2012-07-15T18:10:21
20	Spectrometer Frequency	300.13
21	Spectral Width	6172.8
22	Lowest Frequency	-1236.9
23	Nucleus	1H
24	Acquired Size	32768
25	Spectral Size	65536





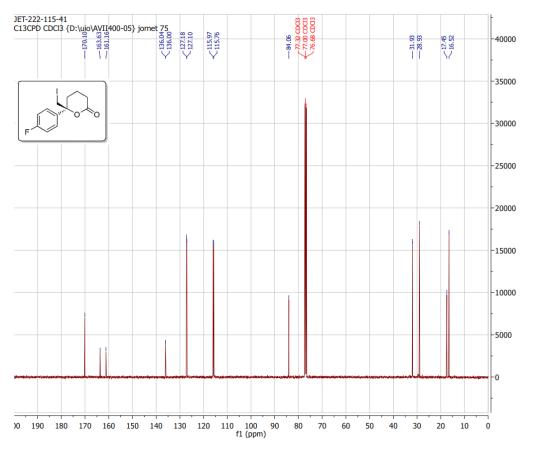
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4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
	Spectrometer Author	DPX300
-		
-		CDCI3
	Temperature	298.2
11	Pulse Sequence	zgpg30
12	Experiment	1D
13	Number of Scans	1024
14	Receiver Gain	11585
15	Relaxation Delay	2.0000
16	Pulse Width	5.8000
17	Acquisition Time	1.8219
18	Acquisition Date	2012-07-15T19:17:41
19	Modification Date	2012-07-15T19:17:45
20	Spectrometer Frequency	75.48
21	Spectral Width	17985.6
22	Lowest Frequency	-1450.2
23	Nucleus	13C
24	Acquired Size	32768
25	Spectral Size	65536

Figure S-44 ¹³C-NMR spectrum of compound 2d.



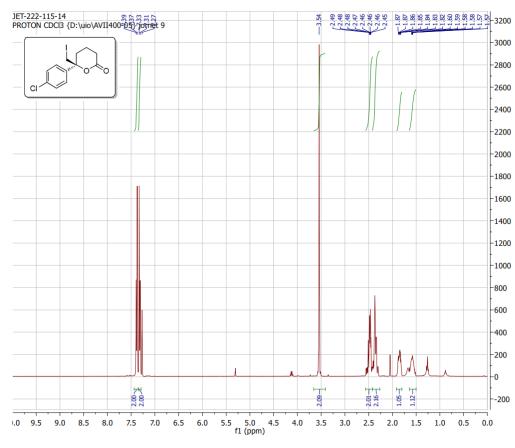
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5	Owner	tnmr
6	Site	
7	Spectrometer	DPX300
8	Author	
9	Solvent	CDCI3
10	Temperature	298.2
11	Pulse Sequence	zg30
12	Experiment	1D
13	Number of Scans	16
14	Receiver Gain	228
15	Relaxation Delay	1.0000
16	Pulse Width	10.0000
17	Acquisition Time	5.3084
18	Acquisition Date	2012-05-16T18:11:38
19	Modification Date	2012-05-16T18:11:43
20	Spectrometer Frequency	300.13
21	Spectral Width	6172.8
22	Lowest Frequency	-1236.3
23	Nucleus	1H
24	Acquired Size	32768
25	Spectral Size	65536

Figure S-45 ¹H-NMR spectrum of compound 2e.

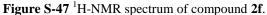


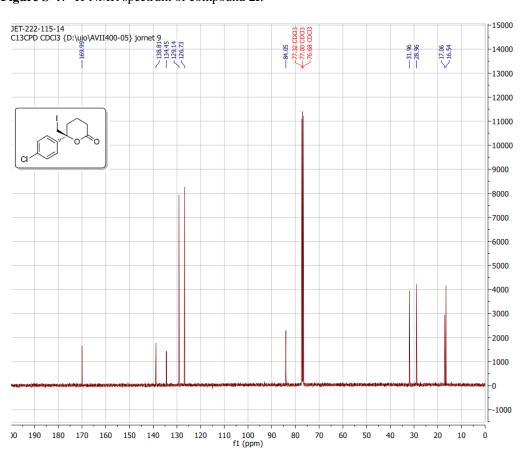
	D	M.L.
	Parameter	Value
1	Data File Name	M:/ Jørn/ NMR/ AVI400/ JET-222-116-43/ 10/
2	Title	fid JET-222-115-41
3		C13CPD CDCl3 {D:\ uio\ AVII400-05} jornet 75
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
	Spectrometer Author	spect
9	Solvent	CDCI3
10	Temperature	295.9
11	Pulse Sequence	zgpg30
12	Experiment	1D
13	Number of Scans	1024
14	Receiver Gain	2050
15	Relaxation Delay	2.0000
16	Pulse Width	7.0000
17	Acquisition Time	1.2977
18	Acquisition Date	2012-05-15T06:51:00
19	Modification Date	2012-05-15T06:51:18
20	Spectrometer Frequency	100.64
21	Spectral Width	25252.5
22	Lowest Frequency	-1060.8
23	Nucleus	13C
24	Acquired Size	32768
25	Spectral Size	65536

Figure S-46¹³C-NMR spectrum of compound 2e.



	Parameter	Value
1	Data File Name	M:/ Jørn/ NMR/ AVI400/ JET-222-115-41/ 10/ fid
2	Title	JET-222-115-14
3		PROTON CDCl3 {D:\ uio\ AVII400-05} jornet 9
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
7	Spectrometer	spect
8	Author	
9	Solvent	CDCl3
10	Temperature	295.2
11	Pulse Sequence	zg30
12	Experiment	1D
13	Number of Scans	16
14	Receiver Gain	45
15	Relaxation Delay	1.0000
16	Pulse Width	12.1500
17	Acquisition Time	3.9846
18	Acquisition Date	2012-06-25T19:12:00
19	Modification Date	2012-06-25T19:13:04
20	Spectrometer Frequency	400.18
21	Spectral Width	8223.7
22	Lowest Frequency	-1644.9
23	Nucleus	1H
24	Acquired Size	32768
25	Spectral Size	65536





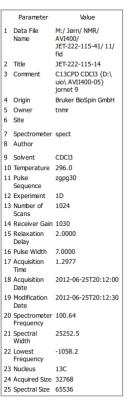
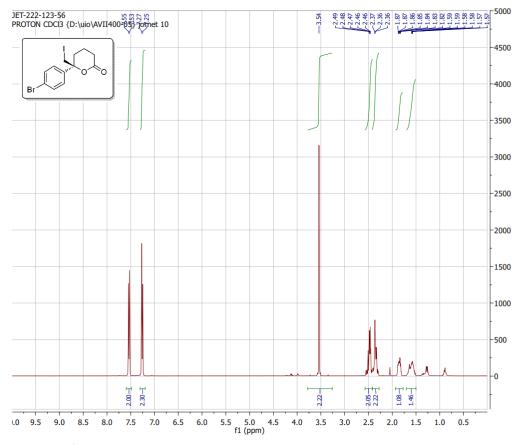
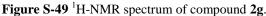
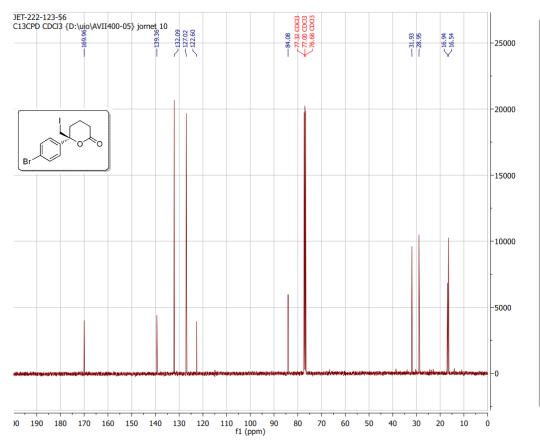


Figure S-48 ¹³C-NMR spectrum of compound 2f.



	Parameter	Value
1	Data File Name	M:/ Jørn/ NMR/ AVI400/ JET-222-123-56/ 10/ fid
2	Title	JET-222-123-56
3	Comment	PROTON CDCl3 {D:\ uio\ AVII400-05} jornet 10
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
7	Spectrometer	spect
8	Author	
9	Solvent	CDCI3
10	Temperature	295.1
11	Pulse Sequence	zg30
12	Experiment	1D
13	Number of Scans	16
14	Receiver Gain	40
15	Relaxation Delay	1.0000
16	Pulse Width	12.1500
17	Acquisition Time	3.9846
18	Acquisition Date	2012-06-25T20:19:00
19	Modification Date	2012-06-25T20:19:14
20	Spectrometer Frequency	400.18
21	Spectral Width	8223.7
22	Lowest Frequency	-1644.2
23	Nucleus	1H
24	Acquired Size	32768
25	Spectral Size	65536





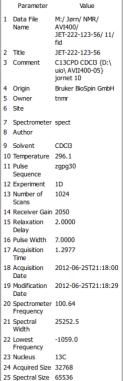
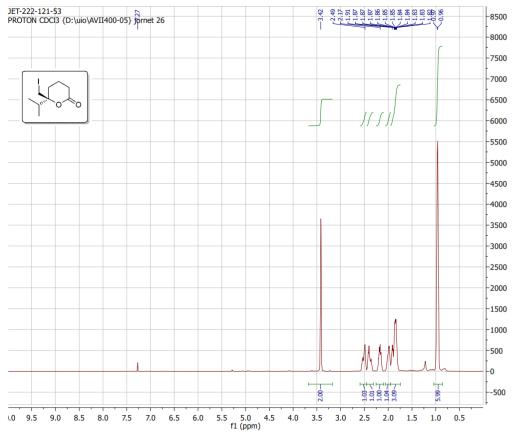
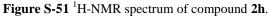
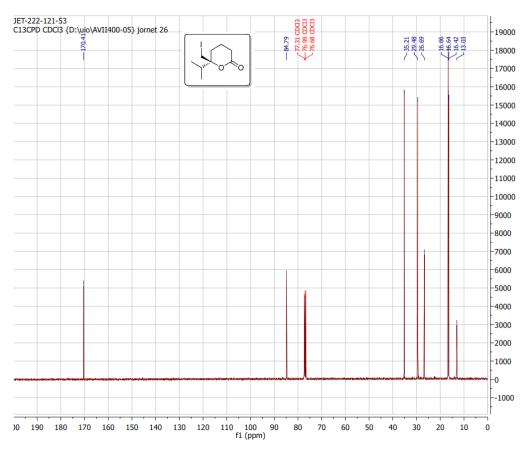


Figure S-50 ¹³C-NMR spectrum of compound 2g.



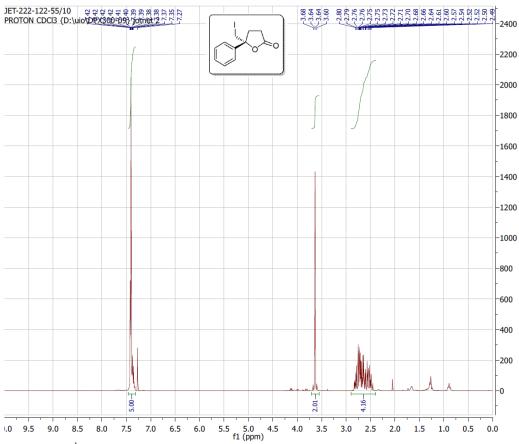
	Parameter	Value
1	Data File Name	M:/ Jørn/ NMR/ AVI400/ JET-222-121-53/ 10/ fid
2	Title	JET-222-121-53
3	Comment	PROTON CDCl3 {D:\ uio\ AVII400-05} jornet 26
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
7	Spectrometer	spect
8	Author	
9	Solvent	CDCI3
10	Temperature	295.2
	Pulse Sequence	zg30
12	Experiment	1D
13	Number of Scans	16
14	Receiver Gain	25
15	Relaxation Delay	1.0000
16	Pulse Width	12.1500
17	Acquisition Time	3.9846
18	Acquisition Date	2012-06-12T09:14:00
19	Modification Date	2012-06-12T09:14:34
20	Spectrometer Frequency	400.18
21	Spectral Width	8223.7
22	Lowest Frequency	-1645.0
23	Nucleus	1H
24	Acquired Size	32768
25	Spectral Size	65536





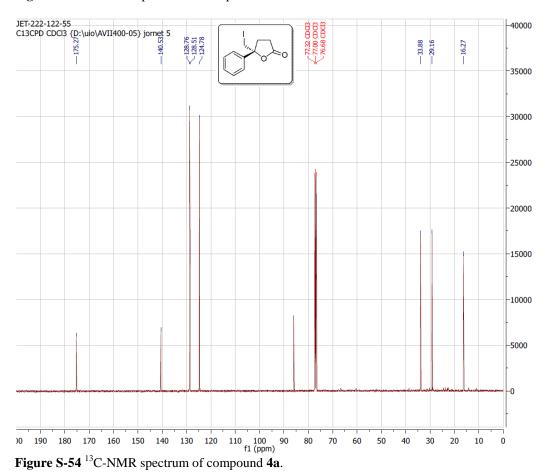
	Parameter	Value
1	Data File Name	M:/ Jørn/ NMR/ AVI400/ JET-222-121-53/ 11/ fid
2	Title	JET-222-121-53
3	Comment	C13CPD CDCl3 {D:\ uio\ AVII400-05} jornet 26
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
7	Spectrometer	spect
8	Author	
9	Solvent	CDCI3
10	Temperature	296.1
11	Pulse Sequence	zgpg30
12	Experiment	1D
13	Number of Scans	1024
14	Receiver Gain	812
15	Relaxation Delay	2.0000
16	Pulse Width	7.0000
17	Acquisition Time	1.2977
18	Acquisition Date	2012-06-12T10:13:00
19	Modification Date	2012-06-12T10:13:59
20	Spectrometer Frequency	100.64
21	Spectral Width	25252.5
22	Lowest Frequency	-1068.9
23	Nucleus	13C
	Acquired Size	
25	Spectral Size	65536

Figure S-52 ¹³C-NMR spectrum of compound 2h.



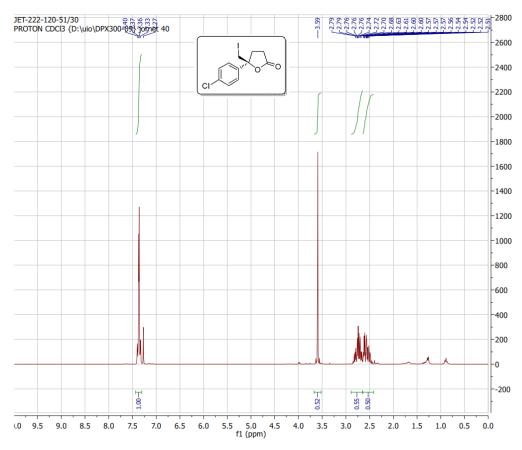
	Parameter	Value
1	Data File Name	M:/ Jørn/ NMR/ JET-222-122-55/ 10/ fid
2	Title	JET-222-122-55/ 10
3	Comment	PROTON CDCI3 {D:\ uio\ DPX300-09} jornet 2
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
7	Spectrometer	DPX300
	Author	
9	Solvent	CDCI3
10	Temperature	298.2
11	Pulse Sequence	zg30
12	Experiment	1D
13	Number of Scans	16
14	Receiver Gain	287
15	Relaxation Delay	1.0000
16	Pulse Width	10.0000
17	Acquisition Time	5.3084
18	Acquisition Date	2012-06-14T17:08:16
19	Modification Date	2012-06-14T17:08:21
20	Spectrometer Frequency	300.13
21	Spectral Width	6172.8
22	Lowest Frequency	-1236.3
	Nucleus	1H
	Acquired Size	
25	Spectral Size	65536



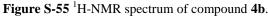


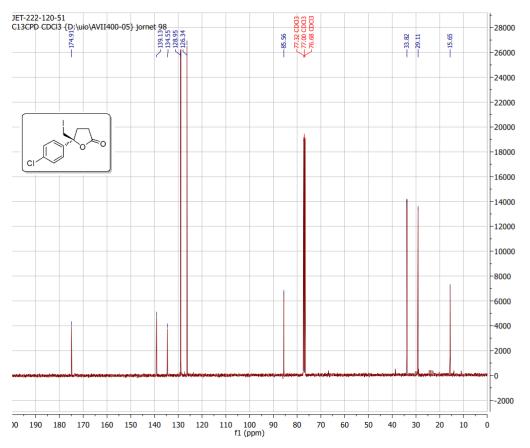
	Parameter	Value
1	Data File Name	M:/ Jørn/ NMR/ AVI400/ JET-222-122-55/ 10/ fid
2	Title	JET-222-122-55
3	Comment	C13CPD CDCl3 {D:\ uio\ AVII400-05} jornet 5
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
7	Spectrometer	spect
8	Author	
9	Solvent	CDCI3
10	Temperature	296.1
11	Pulse Sequence	zgpg30
12	Experiment	1D
13	Number of Scans	1024
14	Receiver Gain	1620
15	Relaxation Delay	2.0000
16	Pulse Width	7.0000
17	Acquisition Time	1.2977
18	Acquisition Date	2012-06-18T15:25:00
19	Modification Date	2012-06-18T15:25:58
20	Spectrometer Frequency	100.64
21	Spectral Width	25252.5
22	Lowest Frequency	-1062.3
23	Nucleus	13C
	Acquired Size	
25	Spectral Size	65536

S39



	Parameter	Value
1	Data File Name	M:/ Jørn/ NMR/ JET-222-120-51/ 30/ fid
2	Title	JET-222-120-51/ 30
3	Comment	PROTON CDCl3 {D:\ uio\ DPX300-09} jornet 40
4	Origin	Bruker BioSpin GmbH
5	Owner	tnmr
6	Site	
7	Spectrometer	DPX300
	Author	
9	Solvent	CDCI3
10	Temperature	298.2
11	Pulse Sequence	zg30
12	Experiment	1D
13	Number of Scans	16
14	Receiver Gain	256
15	Relaxation Delay	1.0000
16	Pulse Width	10.0000
17	Acquisition Time	5.3084
18	Acquisition Date	2012-06-14T12:37:40
19	Modification Date	2012-06-14T12:37:45
20	Spectrometer Frequency	300.13
21	Spectral Width	6172.8
22	Lowest Frequency	-1236.3
	Nucleus	1H
24	Acquired Size	32768
25	Spectral Size	65536





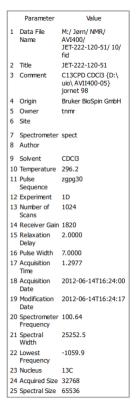
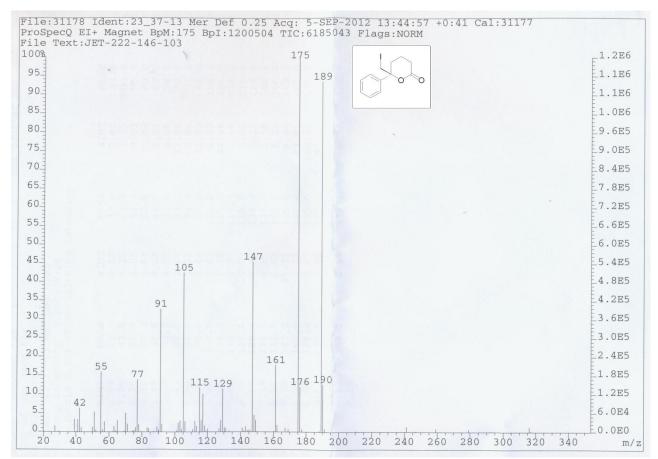
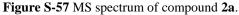


Figure S-56 ¹³C-NMR spectrum of compound 4b.





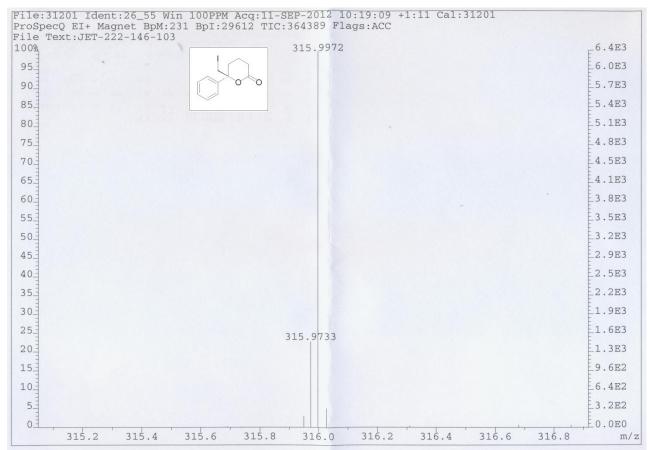


Figure S-58 HRMS spectrum of compound 2a.

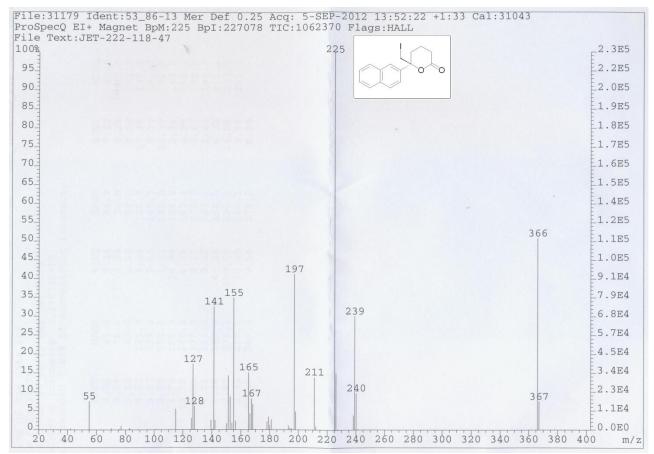


Figure S-59 MS spectrum of compound 2b.

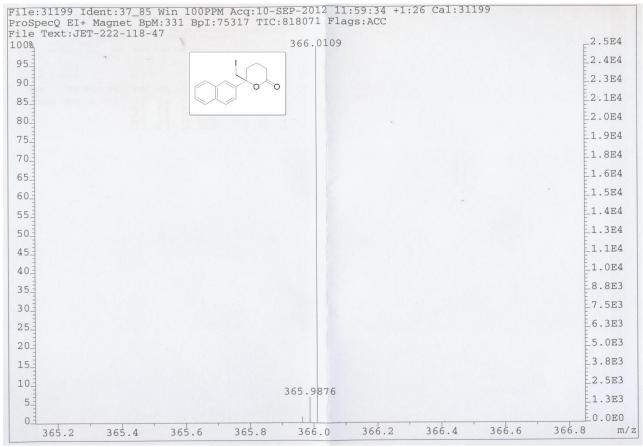


Figure S-60 HRMS spectrum of compound 2b.

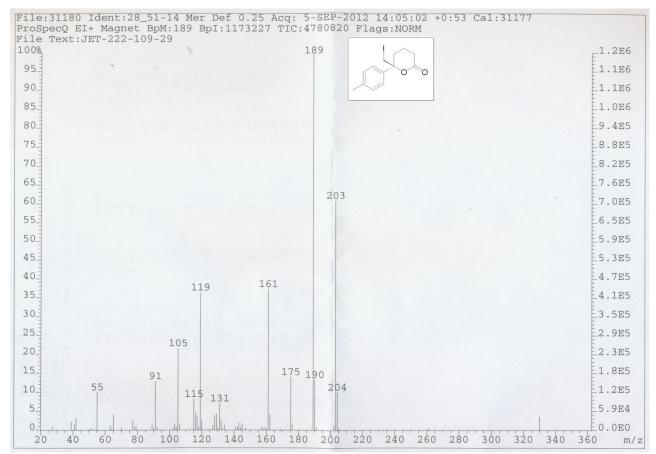


Figure S-61 MS spectrum of compound 2c.

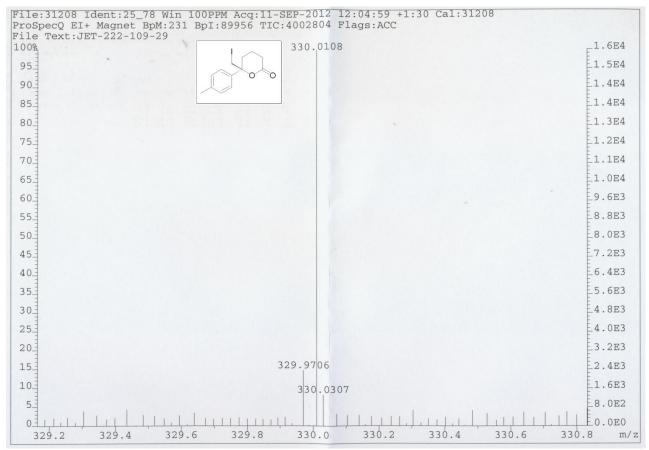


Figure S-62 HRMS spectrum of compound 2c.

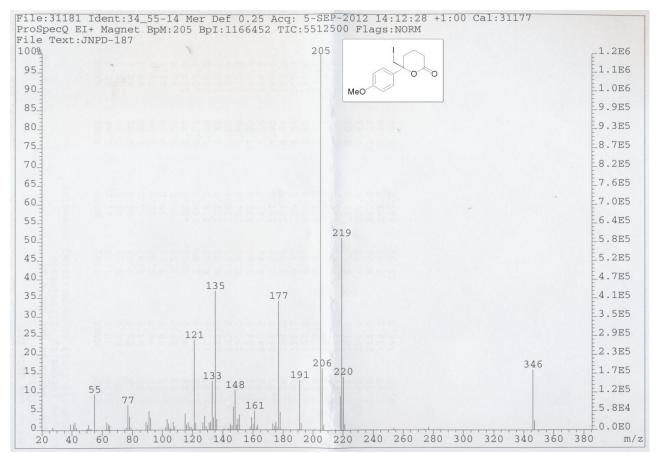


Figure S-63 MS spectrum of compound 2d.

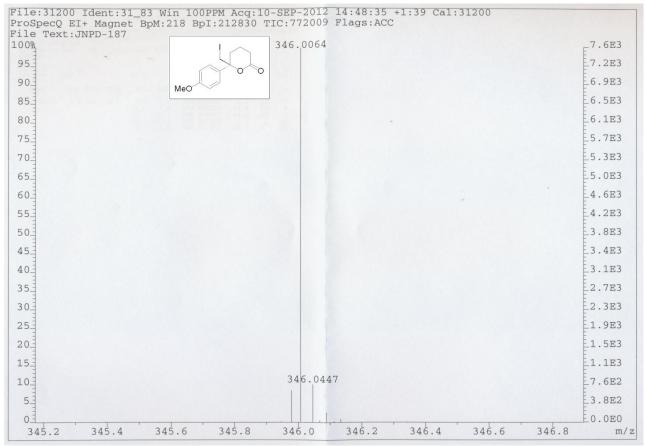


Figure S-64 HRMS spectrum of compound 2d.

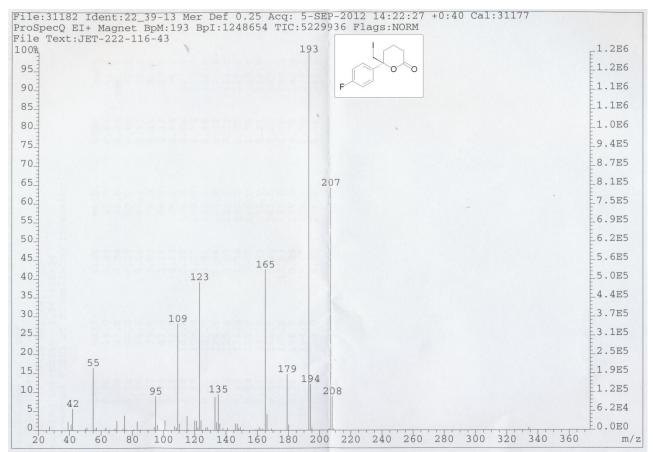


Figure S-65 MS spectrum of compound 2e.

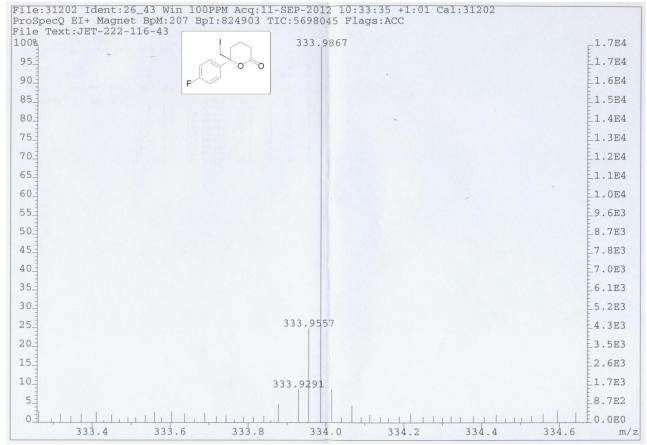


Figure S-66 HRMS spectrum of compound 2e.

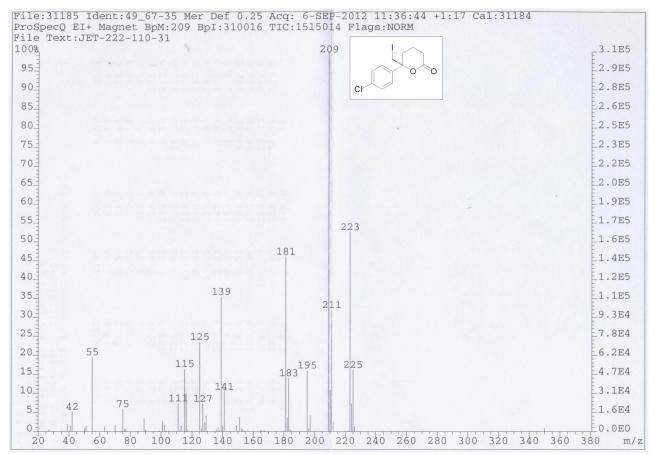


Figure S-67 MS spectrum of compound 2f.

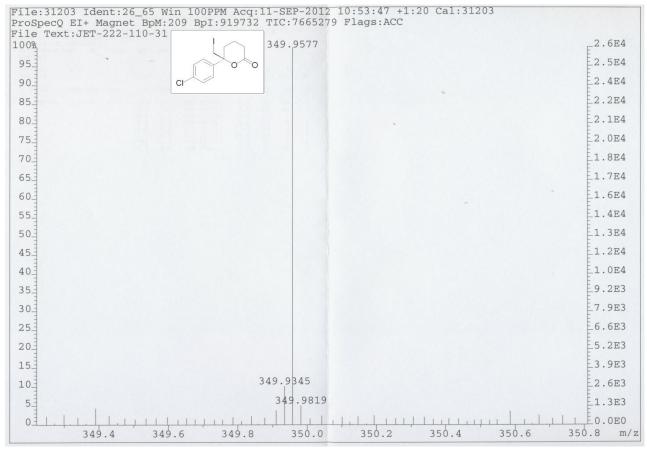


Figure S-68 HRMS spectrum of compound 2f.

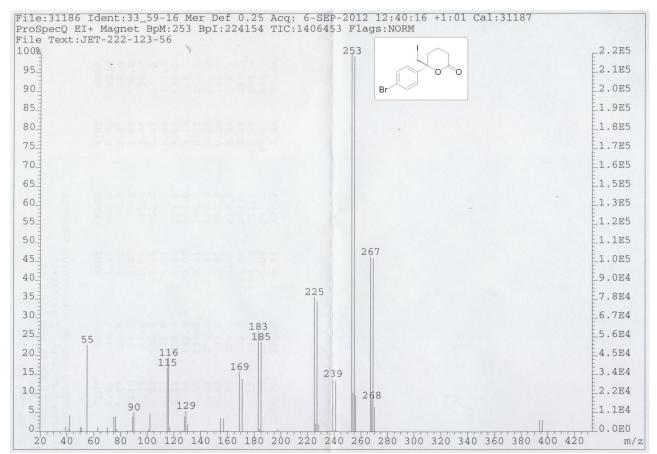


Figure S-69 MS spectrum of compound 2g.

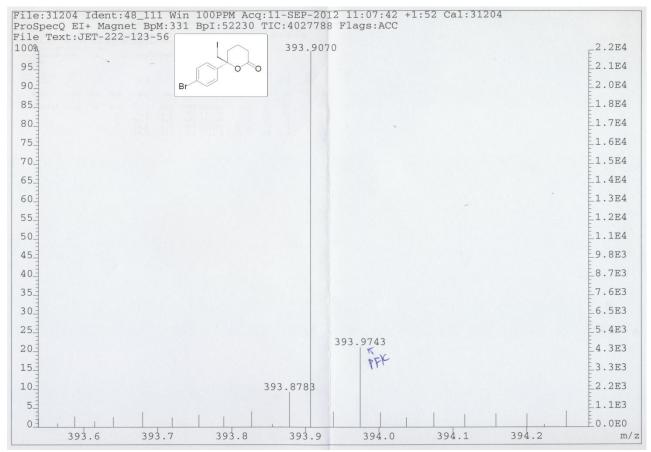
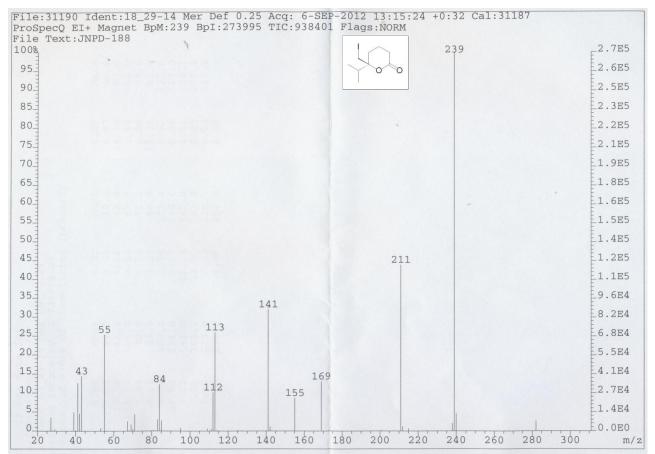
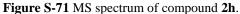


Figure S-70 HRMS spectrum of compound 2g.





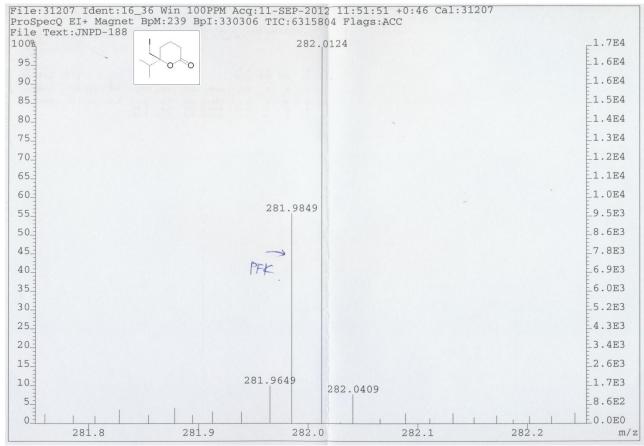


Figure S-72 HRMS spectrum of compound 2h.

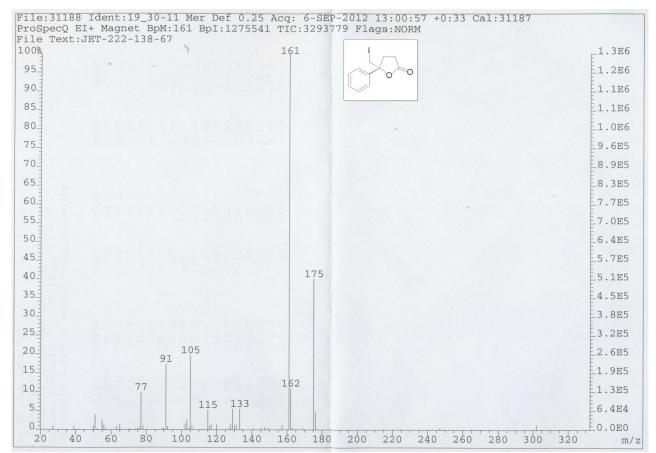


Figure S-73 MS spectrum of compound 4a.

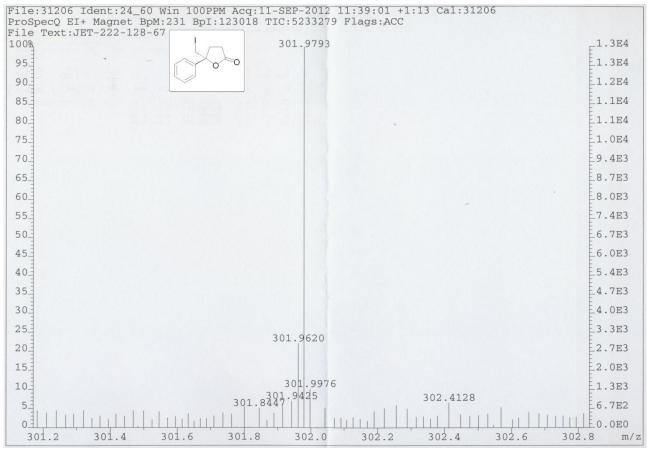


Figure S-74 HRMS spectrum of compound 4a.

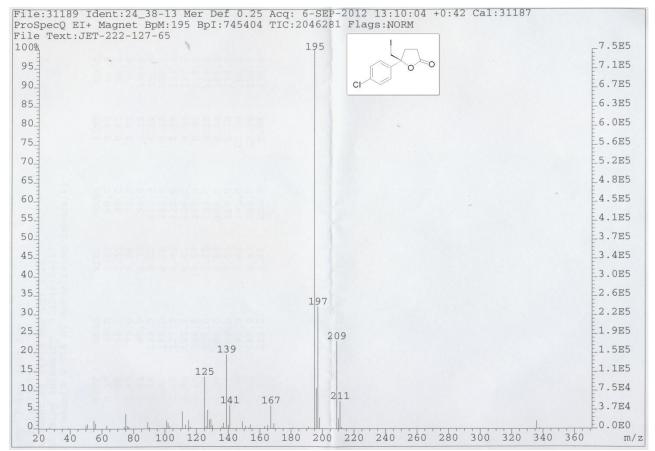


Figure S-75 MS spectrum of compound 4b.

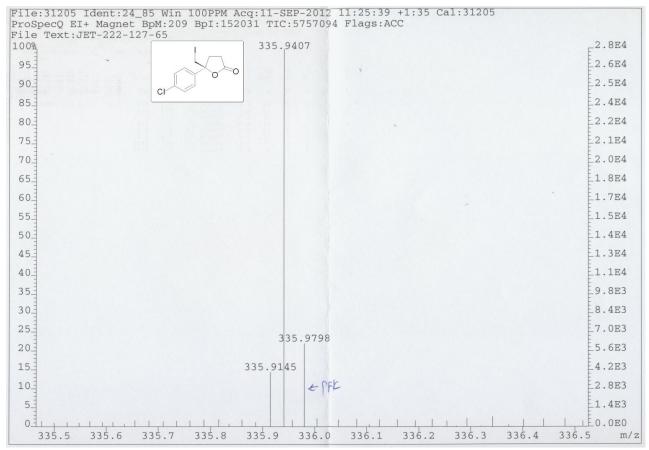


Figure S-76 HRMS spectrum of compound 4b.

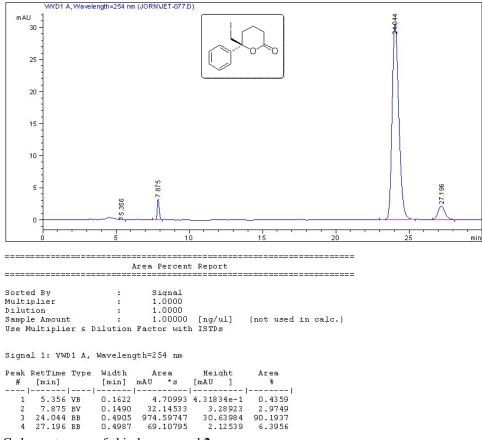


Figure S-77 HPLC chromatogram of chiral compound 2a.

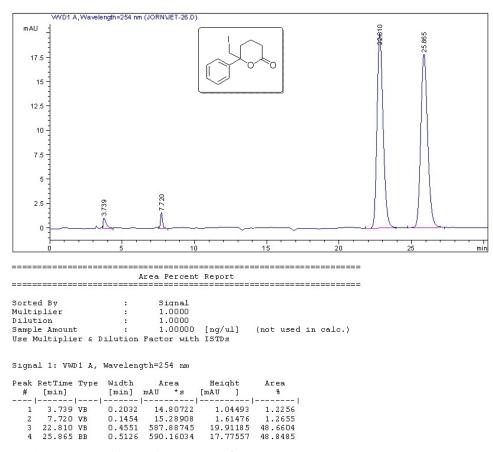


Figure S-78 HPLC chromatogram of racemic compound 2a.

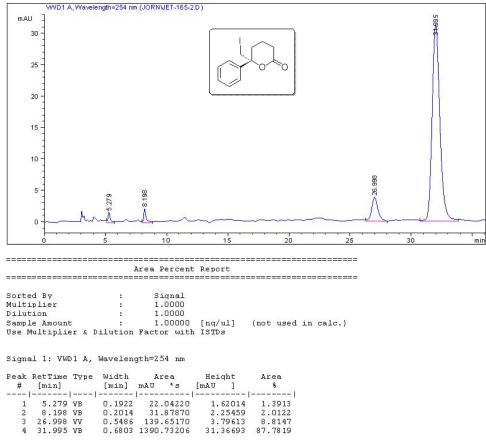


Figure S-79 HPLC chromatogram of chiral compound ent-2a.

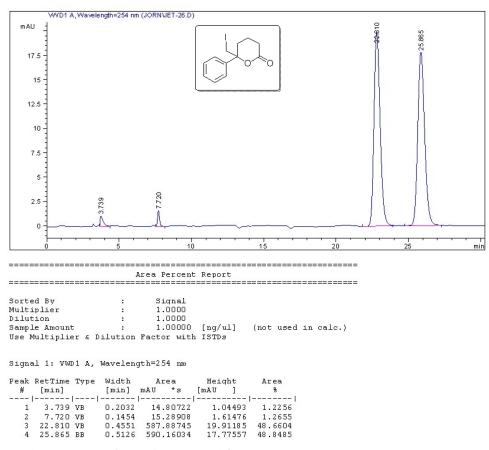


Figure S-80 HPLC chromatogram of racemic compound 2a.

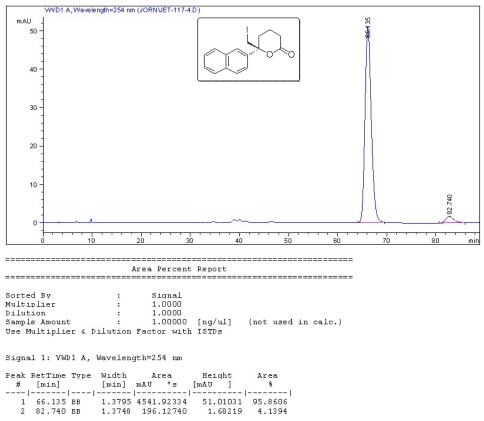


Figure S-81 HPLC chromatogram of chiral compound 2b.

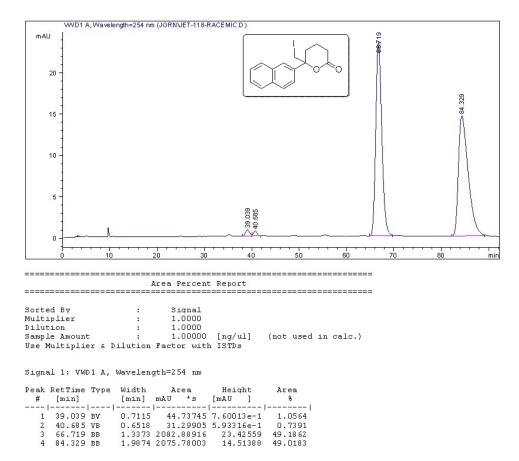


Figure S-82 HPLC chromatogram of racemic compound 2b.

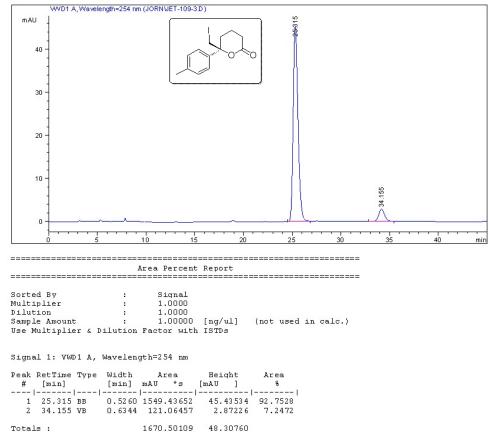


Figure S-83 HPLC chromatogram of chiral compound 2c.

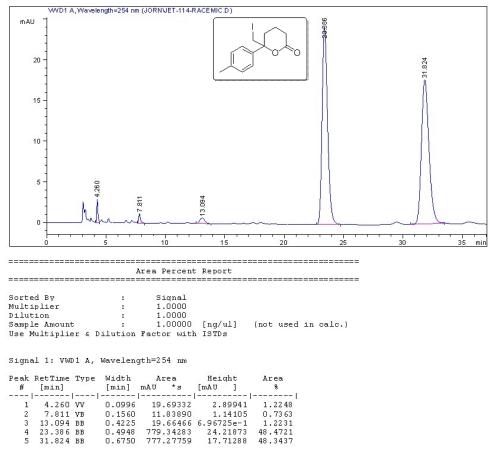


Figure S-84 HPLC chromatogram of racemic compound 2c.

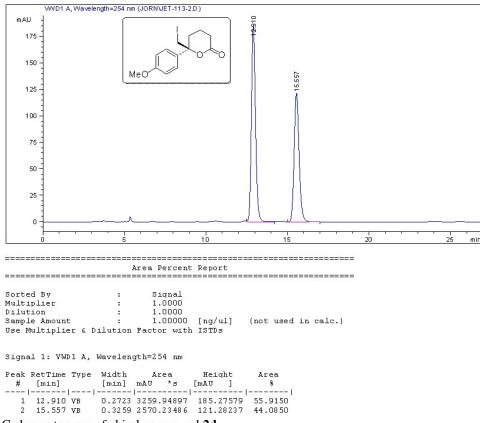


Figure S-85 HPLC chromatogram of chiral compound 2d.

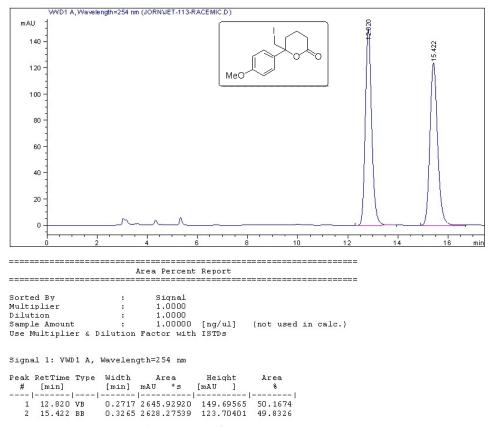
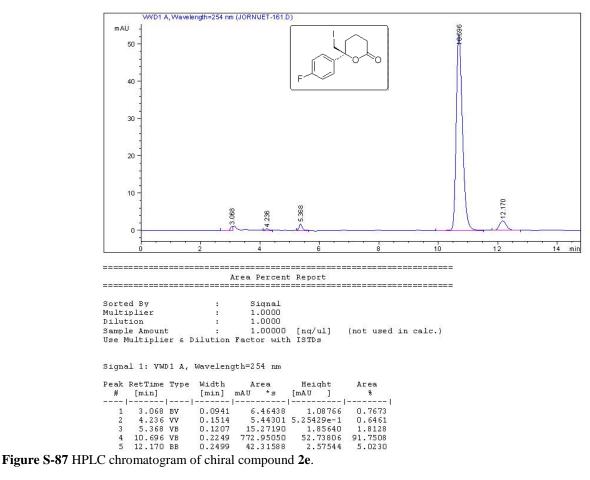


Figure S-86 HPLC chromatogram of racemic compound 2d.



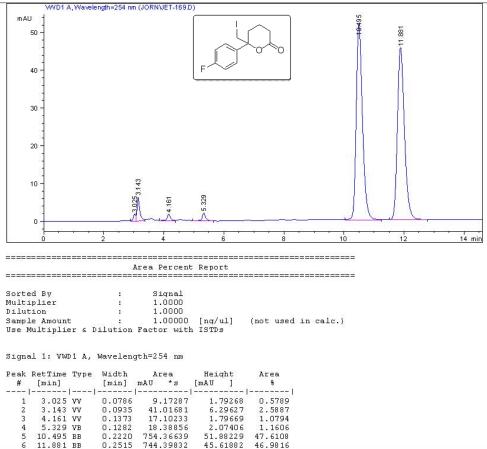
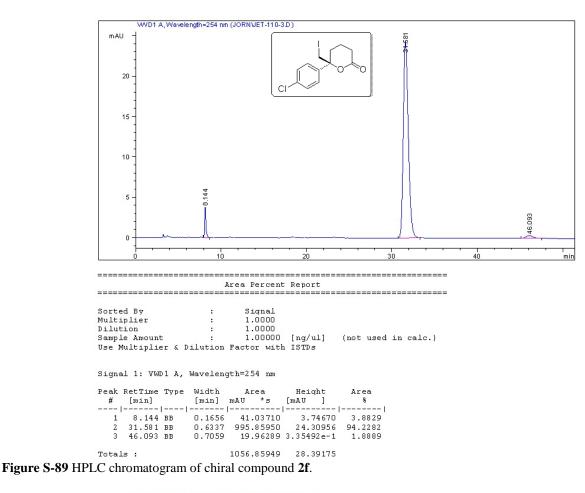


Figure S-88 HPLC chromatogram of racemic compound 2e.



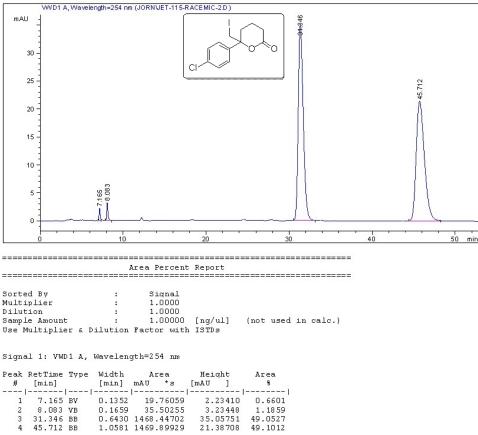
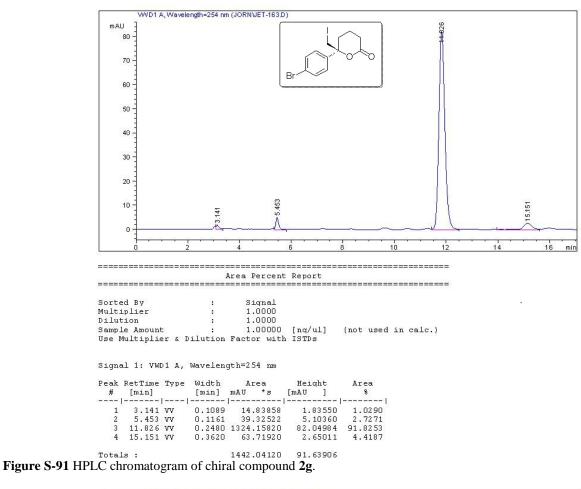


Figure S-90 HPLC chromatogram of racemic compound 2f.



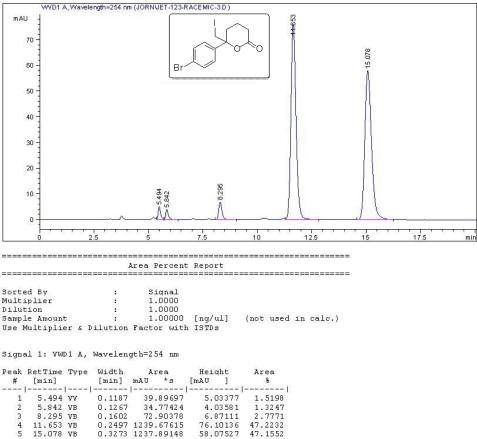


Figure S-92 HPLC chromatogram of racemic compound 2g.

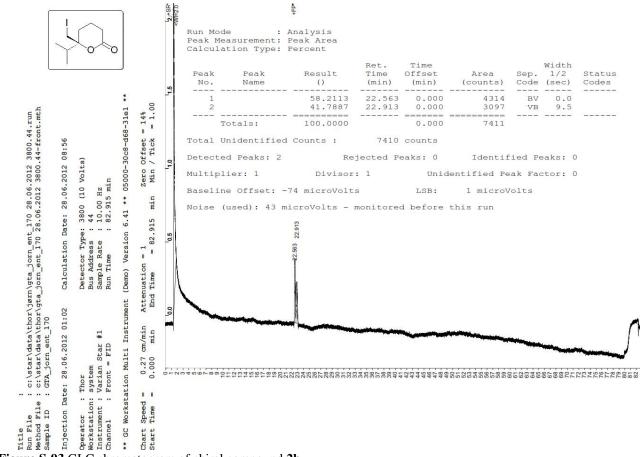
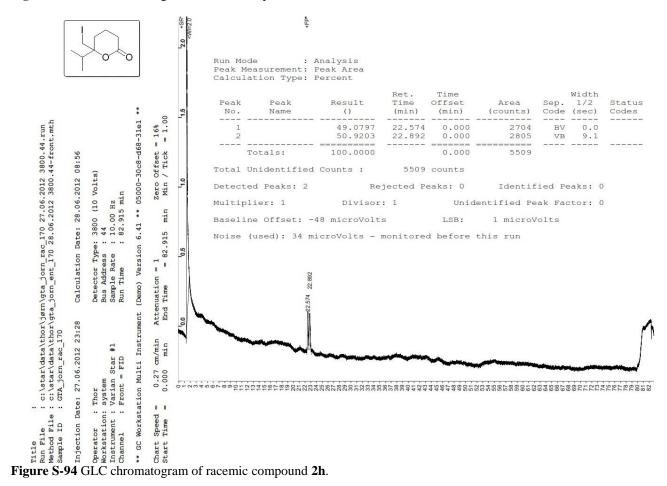


Figure S-93 GLC chromatogram of chiral compound 2h.



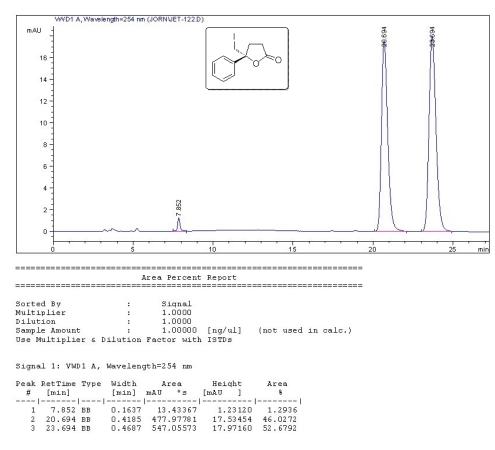


Figure S-95 HPLC chromatogram of chiral compound 4a.

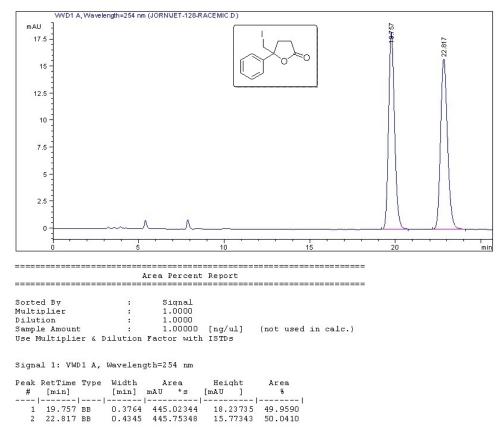
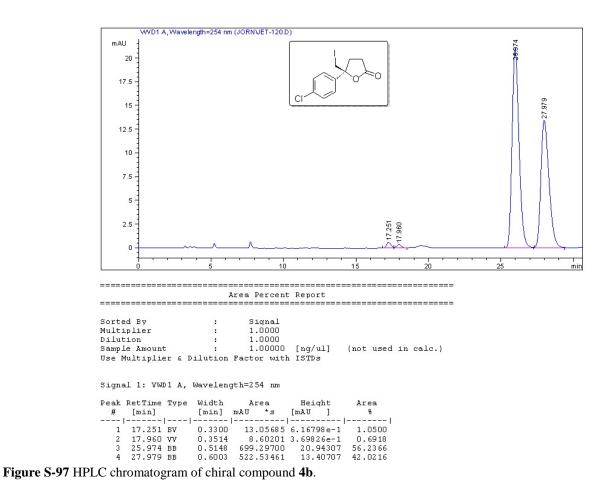


Figure S-96 HPLC chromatogram of racemic compound 4a.



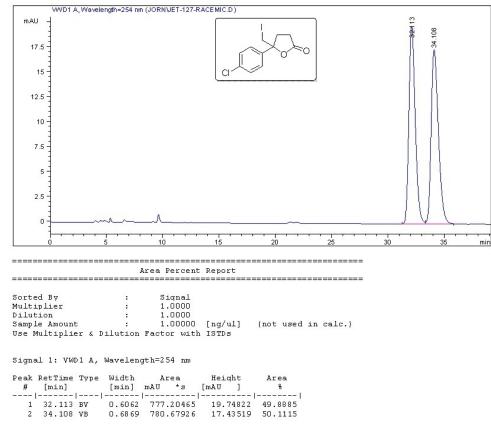


Figure S-98 HPLC chromatogram of racemic compound 4b.

References

- (1) Malerich, J. P.; Hagihara, K.; Rawal, V. H. J. Am. Chem. Soc. 2008, 130, 14416.
- (2) Konishi, H.; Lam, T. Y.; Malerich, J. P.; Rawal, V. H. Org. Lett. 2010, 12, 2028.
- (3) Zhu, Y.; Malerich, J. P.; Rawal, V. H. Angew. Chem. Int. Ed. 2010, 49, 153.
- (4) Veitch, G. E.; Jacobsen, E. N. Angew. Chem., Int. Ed. 2010, 49, 7332.
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- (6) Haas, J.; Piguel, S.; Wirth, T. Org. Lett. 2002, 4, 297.
- (7) For the antipodal (*R*)-enantiomer, see ref. 6.