Supporting Information

Hydration of Terminal Alkynes Catalyzed by Water-Soluble Cobalt Porphyrin Complexes

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1. General Comments

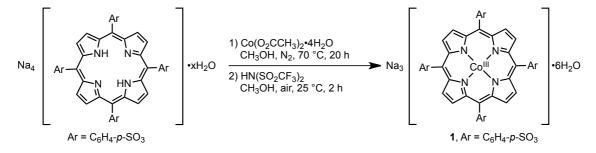
¹H and ¹³C NMR spectra were recorded on a JEOL ECA-600 (600 MHz for ¹H, 150 MHz for ¹³C), a JEOL ECA-500 (500 MHz for ¹H, 125 MHz for ¹³C), or a JEOL GSX (270 MHz for ¹H) at 27 °C. Chemical shifts are reported as δ in ppm and are internally referenced to tetramethylsilane (TMS, 0.0 ppm for ¹H) or CDCl₃ (77.06 ppm for ¹³C) and calculated relative to TMS. The following abbreviations are used: s = singlet, d = doublet, t = triplet, bs = broad singlet, dt = double triplet, ddd = double double double triplet, and m = multiplet). High-resolution mass spectra (HRMS) were obtained from JEOL JMS (EI or FAB). Low-resolution mass spectra (LRMS) were obtained from Agilent 5973 MSD (EI). IR spectra were recorded on a FT-IR6100 (JASCO) at 25 °C. UV spectra were recorded on a UV–2450 (SHIMADZU).

2. Materials

1-Decyne 1,8-nonadiyne (**2b**), 4-*t*-butylphenylacetylene (2a),(2i), 4-(4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)ethynylbenzene (21) and 5-hexyne-1-ol were purchased from Wako Chemicals. 10-Undecyne-1-ol, 10-undecynoic acid, and N,N-dimethyl-4-aminopyridine (DMAP) were purchased from TCI. Phenylacetylene (2h), 4-ethynyl benzonitrile 1-ethynyl-4-nitrobenzene (2m),(2n), and 5,10,15,20-tetraphenyl-21H,23H- porphine-p,p',p'',p'''-tetrasulfonic acid, tetrasodium salt [Na₄(H₂TPPS)], imidazole, trityl chloride (TrCl), boron trifluoride diethyl etherate, L-phenylalanine methyl hydrochloride, and ester *N*-(3-Dimethylaminopropyl)-*N*'-ethylcarbodiimide hydrochloride (EDAC) were purchased from Aldrich. Anhydrous methanol (water content: <50 ppm), anhydrous diethyl ether (water content: <50 ppm), anhydrous dichloromethane (water content: <10 ppm), anhydrous tetrahydrofuran (water content: <10 ppm), anhydrous toluene (water content: <10 ppm), cobalt(II) dichloride hexahydrate, cobalt(II) acetate tetrahydrate, trifluoro-N-[(trifluoromethyl)sulfonyl]methanesulfonamide $(HNTf_2),$ tert-butyldimethylsilyl chloride (TBDMSCl), sodium hydride (60% dispersion in mineral oil), and allylbromide were purchased from Kanto Chemicals. *tert*-Butyldiphenyl(hex-5-yn-1-yloxy)silane (2c)¹, hex-5-ynyl benzoate (2f)², *N*-(4-ethynylphenyl)acetamide $(2j)^3$, and 1-iodo-4-ethynylbenzene $(2k)^4$ were prepared according to the literature procedures.¹⁻⁴

3. Synthesis of 1 and 2

Tetrakis(p-sulfonatophenyl)porphyrin cobalt(III) trisodium salt hexahydrate (1)



To a 500 mL Schlenk flask equipped with a stirring bar, 5,10,15,20-tetraphenyl-21*H*,23*H*-porphine-p,p',p''-tetrasulfonic acid, tetrasodium

¹ Baldwin, J. E.; Romeril, S. P.; Lee, V.; Claridge, T. D. W.; *Org. Lett.* **2001**, *3*, 1145–1148.

² Chan, W.-K.; Ho, C.-M.; Wong, M.-K.; Che, C.-M. J. Am. Chem. Soc. **2006**, 128, 14796–14797.

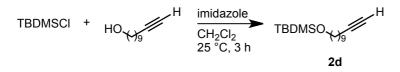
³ Tiecco, M.; Testaferri, L.; Temperini, A.; Bagnoli, L.; Marini, F.; Santi, C.; Terlizzi, R. *Eur. J. Org. Chem.* **2004**, 3447–3458.

⁴ Goeb, S.; Ziessel, R. Org. Lett. **2007**, *9*, 737–740.

salt hydrate [550.1 mg, 0.54 mmol (calculated as anhydrate)], cobalt acetate tetrahydrate (141.5 mg, 0.57 mmol), and dry CH₃OH (150 mL) were added under a N₂ atmosphere. The mixture was stirred at 70 °C for 20 h, cooled to 25 °C, and concentrated to around 30 mL under reduced pressure. Dry Et₂O (400 mL) was added to the mixture under a N₂ atmosphere. After an orange precipitate was generated, the supernatant was removed. The dissolution-reprecipitation process using dry CH₃OH and dry Et₂O was repeated two more times to afford tetrakis(p-sulfonatophenyl)porphyrin cobalt(II) tetrasodium salt as a orange solid. After this crude product was placed in a 300 mL recovery flask equipped with a stirring bar, dry CH₃OH (160 mL) and HNTf₂ (811.7 mg, 2.9 mmol) were added to the flask. The mixture was stirred under air at 25°C for 2 h, and was concentrated to around 20 mL under reduced pressure. Addition of dry Et₂O (200 mL) induced a formation of a purple precipitate, and the supernatant was removed The dissolution-reprecipitation process was repeated two more after centrifugation. The purple precipitate was dried under reduced pressure, and was dissolved in times. pure water (60 mL). The solution was dialyzed for 2 days (SPECTRUM CE, MWCO: 500-1000). The solution was concentrated to around 5 mL under reduced pressure, and passed through ion exchange resin (Dowex 50W×2 200–400, in Na⁺ form). The solution was concentrated, and dried under reduced pressure (0.08 mmHg) at 80 °C for 9 h to afford 1 (374.2 mg, 60% in two steps, hexahydrate) as a purple solid: mp >400 °C; ¹H NMR (500 MHz, CD₃OD) δ 8.26–8.48 (m, 16H), 9.34–9.37 (m, 8H); $^{13}C{^{1}H}$ NMR (125 MHz, CD₃OD) δ 120.8, 125.9, 135.2, 136.4, 144.1, 145.5, 146.6; UV-vis (CH₃OH) λ_{max} , nm (ϵ , M⁻¹cm⁻¹) 426.5 (1.92×10⁵), 539.0 (1.37×10⁴); elemental analysis calcd for [C₄₄H₃₆CoN₄Na₃O₁₈S₄]: C, 45.36; H, 3.11; N, 4.81, found: C, 46.33;

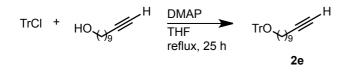
H, 3.33; N, 4.83.

tert-Butyldimethyl(undec-10-yn-1-yloxy)silane (2d)



To a solution of *tert*-butyldimethylsilyl chloride (4.52 g, 30.0 mmol) and imidazole (4.08 g, 59.9 mmol) in dry CH₂Cl₂ (30 mL) was added 10-undecyne-1-ol (3.0 mL, 15.6 mmol) at 25 °C under a N₂ atmosphere. The mixture was stirred at 25 °C for 3 h. The reaction was quenched with water (50 mL). After the mixture was extracted with CH₂Cl₂ (3 × 50 mL), the combined organic phases were washed with brine (30 mL), dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (2% ethyl acetate in *n*-hexane) to afford **2d** (4.24 g, 96%) as a colorless oil: IR (neat) 3314, 2930, 2857, 1097 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 0.04 (s, 6H), 0.89 (s, 9H), 1.27–1.41 (m, 10H), 1.49–1.60 (m, 4H), 1.93 (t, *J* = 2.8 Hz, 1H), 2.17 (dt, *J* = 2.8, 7.6 Hz, 2H), 3.59 (t, *J* = 6.8 Hz, 2H); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ –5.3, 18.4, 25.8, 26.0, 28.5, 28.7, 29.0, 29.4, 29.5, 32.9, 63.3, 68.0, 84.8; HRMS (FAB) calcd for [C₁₇H₃₅OSi⁺] ([MH⁺]) 283.2457; found 283.2463.

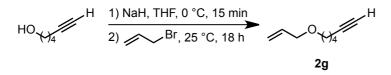




To a solution of trityl chloride (6.70 g, 24 mmol) and 4-(dimethylamino)pyridine (3.67

g, 30 mmol) in dry THF (50 mL) was added 10-undecyne-1-ol (3.80 mL, 20 mmol) at 25 °C under a N₂ atmosphere. The mixture was refluxed for 25 h, and was cooled to 25 °C. The reaction was quenched with water (50 mL). After the mixture was extracted with ethyl acetate (3 × 60 mL), the combined organic phases were washed with brine (30 mL), dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (2% ethyl acetate in *n*-hexane) to afford **2e** (6.69 g, 82%) as a colorless oil: IR (neat) 3306, 2930, 2856, 1071 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 1.20–1.42 (m, 10H), 1.45–1.54 (m, 2H), 1.58–1.67 (m, 2H), 1.86 (t, *J* = 2.8 Hz, 1H), 2.12 (dt, *J* = 2.8, 6.9 Hz, 2H), 3.06 (t, *J* = 6.9 Hz, 2H), 7.12–7.30 (m, 9H), 7.41–7.52 (m, 6H); ¹³C{¹H} NMR (150 MHz, CDCl₃) δ 18.4, 26.2, 28.5, 28.7, 29.0, 29.4(2C), 30.0, 63.6, 68.0, 84.7, 86.3, 126.8, 127.6, 128.7 , 144.5; HRMS (ESI) calcd for [C₃₀H₃₄NaO⁺] ([MNa⁺]) 433.2507; found 433.2516.

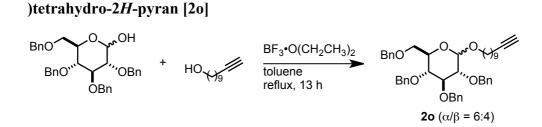
1-Allyalkoxy-5-hexyne (2g)



To a stirring suspension of NaH (144 mg, 6.0 mmol) in THF (30 mL) at 0 °C was slowly added 5-hexyl-1-ol (552 μ L, 5.0 mmol). After the mixture was stirred for 15 min at 0 °C, 3-bromo-1-propene (418 μ L, 4.8 mmol) was slowly added to the mixture, and the mixture was stirred for 18 h at 25 °C. The reaction was quenched with sat. aq. NH₄Cl, the layers were separated, and the aqueous phase was extracted with diethyl ether. The combined organic extracts were washed with brine, dried over Na₂SO₄, and concentrated under reduced pressure. The products was purified by SiO₂ column

chromatography (*n*-hexane only) to give **2g** (300 mg, 43%) as a colorless oil: IR (neat) 3300, 2116, 1647, 1107, 996, 915 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 1.70–1.60 (m, 4H), 1.94 (s, 1H), 2.19 (dt, J = 2.0, 6.8 Hz, 2H), 3.44 (t, J = 6.2 Hz, 2H), 3.95 (ddd, J = 1.2, 1.6, 5.6 Hz, 2H), 5.11 (ddt, J = 1.2, 1.7, 10.4 Hz, 1H), 5.22 (ddt, J = 1.6, 1.7, 17.2 Hz, 1H), 5.86 (ddt, J = 5.6, 10.4, 17.2 Hz, 1H); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 18.1, 25.1, 28.6, 68.3, 69.6, 71.7, 84.1, 116.6, 134.9; HRMS (EI) calcd for [C₉H₁₄O⁺] ([M⁺]) 138.1045; found 138.1049. These data are consistent with the literature values.⁵





In a 100 mL two-necked flask equipped with a stirring bar and a condenser, tetrabenzyl D-glucose (2.44 g, 4.5 mmol), 10-undecyne-1-ol (5.0 mL, 26.1 mmol), and BF₃·OEt₂ (170 μ L, 1.38 mmol) were dissolved in toluene (30 mL). The mixture was refluxed for 13 h, and cooled to 25 °C. The mixture was evaporated, and the residue was purified by silica gel column chromatography (11% ethyl acetate in *n*-hexane) to afford **20** (2.50 g, 80%, a mixture of isomers, $\alpha/\beta = 6:4^6$) as a colorless oil.

The α,β -mixture was purified by MPLC (Isolera One, Biotage, Ltd.; silica gel

⁵ Granados-Focil, S.; Woudenberg, R. C.; Yavuzcetin, O.; Tuominen, M. T.; Coughlin, E. B. *Macromolecules* **2007**, *40*, 8708–8713.

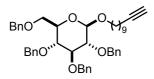
⁶ Vankaylapati, H.; Singh, G.; Tranoy, I. *Tetrahedron: Asymmetry* **2001**, *12*, 1373–1381.

column, Biotage[®] SNAP Cartridge KP-Sil 50 g; eluent, diethyl ether/hexane 1:16–1:1 in 20 minute; flow rate, 50 mL/min.) to afford each isomer of **20** in α ($\alpha/\beta = 99:1$) and β $(\alpha/\beta = 7:93)$ forms.

20α

Colorless oil; IR (ATR) 3294, 2924, 2858, 1065 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 1.23–1.41 (m, 10H), 1.47–1.54 (m, 2H), 1.57–1.67 (m, 2H), 1.93 (t, J = 2.8 Hz, 1H), 2.16 (dt, J = 2.8, 7.6 Hz, 2H), 3.38–3.44 (m, 1H), 3.55 (dd, J = 3.4, 9.6 Hz, 1H), 3.60– 3.66 (m, 3H), 3.72 (dd, J = 3.4, 10.3 Hz, 1H), 3.76–3.80 (m, 1H), 3.99 (t, J = 9.7 Hz, 1H), 4.47 (d, J = 11.7 Hz, 1H), 4.47 (d, J = 11.7 Hz, 1H), 4.60 (d, J = 12.4 Hz, 1H), 4.65 (d, J = 11.7 Hz, 1H), 4.74–4.85 (m, 4H), 4.99 (d, J = 10.3 Hz, 1H), 7.11–7.15 (m, 2H), 7.22–7.37 (m, 18H); ${}^{13}C{}^{1}H{}$ NMR (150 MHz, CDCl₃) δ 18.3, 26.1, 28.4, 28.7, 29.0, 29.3(2C), 68.1(2C), 68.5, 73.0, 73.4, 75.0, 75.6, 77.7, 80.1, 82.1, 84.7, 96.8, 127.4, 127.5, 127.6, 127.7, 127.8, 127.9, 128.0, 128.2, 128.3, 137.9, 138.2, 138.3, 138.9; HRMS (ESI) calcd for $[C_{45}H_{54}NaO_6^+]$ ([MNa⁺]) 713.3813; found 713.3818.

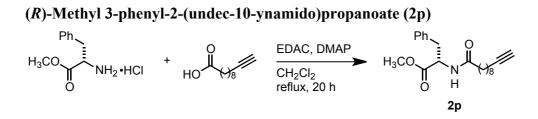
20β



Colorless oil; IR (ATR) 3294, 2927, 2858, 1068 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 1.24–1.44 (m, 10H), 1.47–1.53 (m, 2H), 1.60–1.69 (m, 2H), 1.93 (t, J = 2.8 Hz, 1H), 2.16 (dt, J = 2.8, 6.8 Hz, 2H), 3.42–3.49 (m, 2H), 3.50–3.59 (m, 2H), 3.61–3.69 (m, 2H), 3.74 (d, J = 11.0 Hz, 1H), 3.93-3.99 (m, 1H), 4.38 (d, J = 8.3 Hz, 1H), 4.51-4.57 (m, \mathbf{S}

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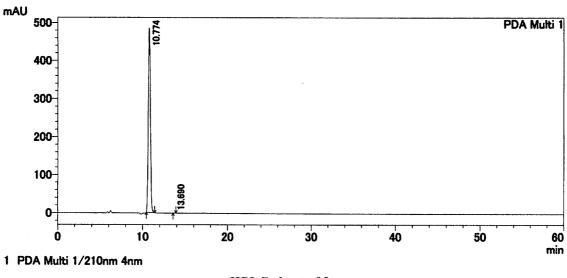
2H), 4.61 (d, J = 12.4 Hz, 1H), 4.71 (d, J = 11.0 Hz, 1H), 4.78 (d, J = 11.0 Hz, 1H), 4.81 (d, J = 11.0 Hz, 1H), 4.92 (d, J = 11.0 Hz, 1H), 4.95 (d, J = 11.0 Hz, 1H), 7.14– 7.17 (m, 2H), 7.24–7.36 (m, 18H); ¹³C{¹H} NMR (150 MHz, CDCl₃) δ 18.4, 26.2, 28.5, 28.7, 29.0, 29.4(2C),29.8, 68.1, 69.0, 70.1, 73.5, 74.8, 74.9, 75.0, 75.7, 78.0, 82.3, 84.7, 84.8, 103.6, 127.6, 127.6, 127.8, 127.9, 128.0, 128.1, 128.3, 128.4, 138.1, 138.2, 138.5, 138.6; HRMS (ESI) calcd for [C₄₅H₅₄NaO₆⁺] ([MNa⁺]) 713.3813; found 713.3814.



In a 50 mL two-necked flask equipped with a stirring bar and a condenser, undecynoic acid (397 mg, 2.2 mmol), L-phenylalanine methyl ester hydrochloride (474 mg, 2.2 mmol), EDAC (441 mg, 2.3 mmol), and DMAP (489 mg, 4.0 mmol) were dissolved in dry CH₂Cl₂ (8 mL). The mixture was refluxed for 20 h, and cooled to 25 °C. The reaction was quenched with water (20 mL). After the mixture was extracted with CH₂Cl₂ (3 × 20 mL), the combined organic phases were washed with sat. aq NaHCO₃ (10 mL) and brine (10 mL), dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (25% ethyl acetate in *n*-hexane) to afford **2p** (614.7 mg, 86%) as a colorless oil; IR (neat) 3292, 2931, 1747, 1650 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 1.24–1.40 (m, 8H), 1.48–1.60 (m, 4H), 1.93 (t, *J* = 2.8 Hz, 1H), 2.14–2.19 (m, 4H), 3.07 (dd, *J* = 6.2, 13.8 Hz, 1H), 3.72 (s, 3H), 4.89 (ddd, *J* = 6.2, 6.2, 7.6 Hz, 1H), 5.96 (d, *J* = 7.6 Hz, 1H), 7.09 (d, *J* = 6.9 Hz, 2H), 7.22–7.30 (m, 3H); ¹³C {¹H} NMR (150

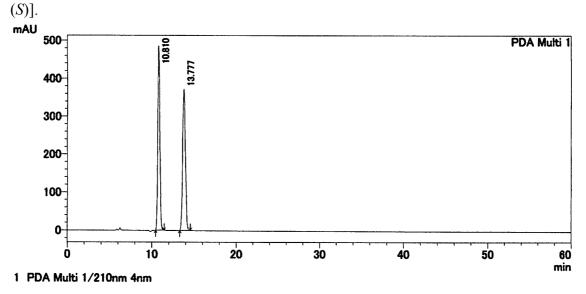
MHz, CDCl₃) δ 18.3, 25.4, 28.3, 28.5, 28.8, 29.0, 29.1, 36.4, 37.8, 52.2, 52.8, 68.0, 84.6, 127.0, 128.4, 129.1, 135.8, 172.1, 172.5; HRMS (FAB) calcd for [C₂₁H₃₀NO₃⁺] ([MH⁺]) 344.2220; found 344.2231. HPLC (Daicel Chiralcel OD-H, hexane/2-propanol 80:20,

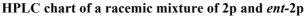
0.50 mL/min) $t_R 10.8 \text{ min}$.



HPLC chart of 2p

Enantiomer of **2p** (*ent*-**2p**) was prepared similarly using D-phenylalanine methyl ester hydrochloride. A racemic mixture of **2p** and *ent*-**2p** was analyzed by HPLC [Daicel Chiralcel OD-H, hexane/2-propanol 80:20, 0.50 mL/min, $t_R = 10.8 \text{ min } (R)$, 13.8 min





4. Hydration of Terminal Alkynes 2 to Methyl Ketones 3

A Representative Procedure for Hydration of Terminal Alkynes: Decan-2-one (3a, Table 1, Entry 15)

A mixture of alkyne **2a** (5.0 mmol, 691.0 mg), H₂O (22 mmol, 0.40 mL), **1** in CH₃OH (15 μ mol, 8 mM, 1.88 mL), HNTf₂ in CH₃OH (15 μ mol, 12 mM, 1.25 mL), and CH₃OH (1.88 mL) was heated at 80 °C for 12 h under air in a closed J. Young tube. The progress of the reaction was checked using TLC and GC. After the tube was cooled to 25 °C, *n*-hexane and water was added to the mixture. The layers were separated, and the aqueous phase was extracted with *n*-hexane. The combined organic extracts were washed with brine, dried over Na₂SO₄, and concentrated under reduced pressure. The crude product was purified by distillation (3.75 mmHg, 140 °C) to give **3a** as a colorless oil (749.0 mg, 96%): IR (neat) 1719 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 0.88 (t, *J* = 6.9 Hz, 3H), 1.20–1.38 (m, 10H), 1.50–1.66 (m, 2H), 2.14 (s, 3H), 2.42 (t, *J* = 7.5 Hz, 2H); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 14.0, 22.6, 23.8, 29.1, 29.2, 29.3, 29.8, 31.8, 43.8, 209.3; LRMS (EI) *m/z*: 156 (M⁺).

2,8-Nonanedione (3b) 0 0

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White solid, purified by recrystalization (from *n*-hexane); IR (neat) 1713, 1701 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 1.22–1.28 (m, 2H), 1.53–1.60 (m, 4H), 2.13 (s, 6H), 2.44 (t, J = 7.5 Hz, 4H); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 23.3, 28.4, 29.8, 43.3, 208.8; HRMS (FAB) calcd for [C₉H₁₆CsO₂⁺] ([MCs⁺]) 289.0199; found 289.0212.

6-tert-Butyldiphenylsilyloxy-2-hexanone (3c)

Colorless oil, purified by column chromatography on silica gel (5% ethyl acetate in *n*-hexane); ¹H NMR (500 MHz, CDCl₃) δ 1.07 (s, 9H), 1.50–1.68 (m, 4H), 2.11 (s, 3H), 2.41 (t, *J* = 7.5 Hz, 2H), 3.68 (t, *J* = 6.3 Hz, 2H), 7.35–7.50 (m, 6H), 7.60–7.68 (m, 4H); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 19.3, 20.4, 27.0, 29.9, 32.0, 43.5, 63.6, 127.7, 129.7, 134.1. 135.7, 209.1. LRMS (EI) *m/z*: 239 [Si(*t*-C₄H₉)(C₆H₅)₂⁺].

11-[(tert-Butyldimethylsilyl)oxy]undecan-2-one (3d)



Colorless oil, purified by column chromatography on silica gel (5% ethyl acetate in *n*-hexane); IR (neat) 2929, 2856, 1719 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 0.04 (s, 6H), 0.89 (s, 9H), 1.25–1.34 (m, 10H), 1.48–1.60 (m, 4H), 2.12 (s, 3H), 2.41 (t, *J* = 7.6 Hz, 2H), 3.59 (t, *J* = 6.2 Hz, 2H); ¹³C{¹H} NMR (150 MHz, CDCl₃) δ –5.5, 18.2, 23.7, 25.6, 25.8, 29.0, 29.2(2C), 29.3, 29.6, 32.7, 43.6, 63.0, 208.6; HRMS (ESI) calcd for [C₁₇H₃₇O₂Si⁺] ([MH⁺]) 301.2563; found 301.2583.

11-(Trityloxy)undecan-2-one (3e) TrO (y_9)

Colorless oil, purified by column chromatography on silica gel (5% ethyl acetate in *n*-hexane); IR (neat) 2929, 2854, 1717 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 1.16–1.43 (m, 10H), 1.45–1.65 (m, 4H), 2.00 (s, 3H), 2.31 (t, *J* = 7.8 Hz, 2H), 3.06 (t, *J* = 6.4 Hz, 2H), 7.06–7.28 (m, 9H), 7.44 (t, *J* = 7.8 Hz, 6H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 23.5, 26.0, 28.9, 29.1(2C), 29.2, 29.5, 29.8, 43.4, 63.3, 86.0, 126.5, 127.4, 128.4, S 12

144.0, 208.5; HRMS (ESI) calcd for $[C_{30}H_{36}NaO_2^+]$ ([MNa⁺]) 451.2613; found 451.2626.

5-Oxohexyl benzoate (3f)

Colorless oil; IR (neat) 1718, 1274 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 1.65–1.80 (m, 4H), 2.14 (s, 3H), 2.44 (t, *J* = 6.9 Hz, 2H), 2.52 (t, *J* = 6.8 Hz, 2H), 4.32 (t, *J* = 6.2 Hz 2H), 7.35–7.60 (m, 3H), 8.02–8.04 (m, 2H); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 20.1, 28.1, 29.8, 42.9, 64.4, 128.3, 129.4, 130.2, 132.8, 166.5, 208.3; HRMS (FAB) calcd for [C₁₃H₁₇O₂⁺] ([MH⁺]) 221.1178; found 221.1184.

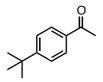
1-(1-Propen-2-oxy)-5-heptanone (3g)

Colorless oil, purified by distillation (3.75 mmHg, 130 °C); IR (neat) 1718, 1106 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 1.60–1.70 (m, 4H) 2.13 (s, 3H), 2.46 (t, *J* = 6.8 Hz, 2H), 3.43 (t, *J* = 6.2 Hz, 2H), 3.95 (ddd, *J* = 1.3, 1.6, 5.7 Hz, 2H), 5.14 (ddt, *J* = 1.3, 1.7, 10.4 Hz, 1H), 5.23 (ddt, *J* = 1.6, 1.6, 17.2 Hz, 1H), 5.88 (ddt, *J* = 5.7, 10.4, 17.2 Hz, 1H); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 20.5, 29.1, 29.8, 43.4, 69.9, 71.7, 116.7, 134.9, 208.8; HRMS (FAB) calcd for [C₉H₁₇O₂⁺] ([MH⁺]) 157.1229; found 157.1229.

Acetophenone (3h)

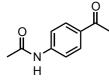
Colorless oil, purified by distillation (3.75 mmHg, 140 °C); IR (neat) 1684 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 2.60 (s, 3H), 7.40–7.59 (m, 3H), 7.92–8.01 (m, 2H); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 26.5, 128.2, 128.5, 133.0, 137.0, 198.0; LRMS (EI) *m/z*: 120 (M⁺).

4-t-Butylacetophenone (3i)



Colorless oil; IR (neat) 1678 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 1.34 (s, 9H), 2.58 (s, 3H), 7.40–7.50 (m, 2H), 7.90–8.00 (m, 2H); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 26.5, 31.0, 35.0, 125.4, 128.2, 134.5, 156.7, 197.7; HRMS (FAB) calcd for [C₁₂H₁₆NaO⁺] ([MNa⁺]) 199.1093; found 199.1091.

N-Acetyl-4-acetoanilide (3j)



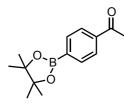
White solid; IR (neat) 3277, 1671 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 2.22 (s, 3H), 2.58 (s, 3H), 7.62–7.66 (m, 2H), 7.90–8.30 (m, 3H); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 24.8, 26.5, 118.8, 129.8, 132.8, 142.3, 168.6, 197.1; HRMS (EI) calcd for [C₁₀H₁₁NO₂] ([M]⁺) 177.0790; found 177.0786.

4-Iodoacetophenone (3k)



White solid; IR (neat) 1666 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 2.58 (s, 3H), 7.65–7.84 (m, 4H); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 26.7, 101.3, 129.9, 136.5, 138.0, 197.4; HRMS (EI) calcd for [C₈H₇IO] ([M]⁺) 245.9542; found 245.9534.

1-[4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)phenyl]ethanone (3l)



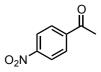
White solid; IR (neat) 2990, 1682 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 1.36 (s, 12H), 2.61 (s, 3H), 7.91 (m, 4H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 24.8, 26.6, 84.1, 127.2, 127.5, 164.8, 138.9, 198.3; HRMS (FAB) calcd for [C₁₄H₁₉BO₃⁺] ([MH⁺]) 247.1506; found 247.1522.

4-Acetylbenzonitrile (3m)



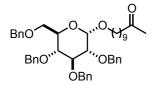
White solid; IR (neat) 1692 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 2.66 (s, 3H), 7.77–7.81 (m, 2H), 8.04–8.08 (m, 2H); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 26.7, 116.3, 117.9, 128.6, 132.5, 139.8, 196.5; HRMS (FAB) calcd for [C₉H₇CsNO⁺] ([MCs⁺]) 277.9577; found 277.9585.

4-Nitroacetophenone (3n)



White solid, purified by column chromatography on silica gel (50% CH₂Cl₂ in *n*-hexane); IR (neat) 1684, 1604, 1355, 831 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 2.70 (s, 3H), 8.11–8.14 (m, 2H), 8.30–8.34 (m, 2H); ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 26.9, 123.8, 129.2, 141.3, 150.3, 196.3; HRMS (FAB) calcd for [C₈H₇CsNO₃⁺] ([MCs⁺]) 297.9475; found 297.9476.

11-[[(2*S*,3*R*,4*S*,5*R*,6*R*)-3,4,5-tris(benzyloxy)-6-[(benzyloxy)methyl]tetrahydro-2*H*-p yran-2-yl]oxy]undecan-2-one [$3o(\alpha/\beta = 99:1)$]

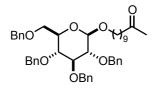


Pale yellow oil; IR (ATR) 2924, 2858, 1712, 1065 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 1.22–1.40 (m, 10H), 1.50–1.69 (m, 4H), 2.11 (s, 3H), 2.40 (t, *J* = 7.8 Hz, 2H), 3.38– 3.45 (m, 1H), 3.52–3.68 (m, 4H), 3.68–3.82 (m, 2H), 3.99 (t, *J* = 9.6 Hz, 1H), 4.47 (d, *J* = 11.9 Hz, 1H), 4.57–4.68 (m, 2H), 4.72–4.87 (m, 4H), 4.99 (d, *J* = 11.0 Hz, 1H), 7.10– 7.17 (m, 2H), 7.19–7.40 (m, 18H); ¹³C{¹H} NMR (150 MHz, CDCl₃) δ 23.8, 26.1, 29.2, 29.4, 29.8, 43.8, 68.2, 68.5, 70.1, 73.1, 73.5, 75.1, 75.7, 77.8, 80.1, 82.1, 96.9, 127.5, 127.6, 127.7, 127.8, 127.9, 128.0, 128.3, 128.4, 138.0, 138.3, 138.4, 138.9, 209.3; HRMS (ESI) calcd for [C₄₅H₅₆NaO₇⁺] ([MNa⁺]) 731.3918; found 731.3926.

The β isomer (**3o** β) was obtained and characterized by the hydration of **2o**($\alpha/\beta = 6:4$) followed by chromatographic separation using MPLC (Isolera One, Biotage, Ltd.; silica gel column, Biotage[®] SNAP Ultra 25 g; eluent, diethyl ether/hexane 1:16–1:1 in 27

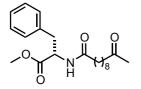
minute; flow rate, 25 mL/min.).

30β



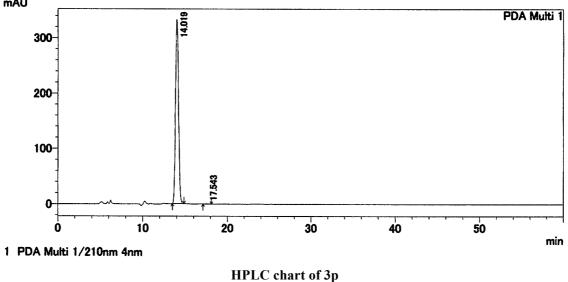
Colorless oil; IR (ATR) 2924, 2858, 1712, 1068 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 1.22–1.43 (m, 10H), 1.51–1.69 (m, 4H), 2.12 (s, 3H), 2.40 (t, J = 6.8 Hz, 2H), 3.42–3.49 (m, 2H), 3.49–3.60 (m, 2H), 3.60–3.70 (m, 2H), 3.74 (d, J = 11.0 Hz, 1H), 3.92–3.99 (m, 1H), 4.39 (d, J = 8.3 Hz, 1H), 4.51–4.63 (m, 3H), 4.71 (d, J = 11.0 Hz, 1H), 4.74–4.85 (m, 2H), 4.89–4.97 (m, 2H), 7.14–7.17 (m, 2H), 7.23–7.37 (m, 18H); ¹³C{¹H} NMR (150 MHz, CDCl₃) δ 23.9, 26.2, 29.2, 29.4, 29.5, 29.8, 29.9, 43.9, 69.1, 70.2, 73.5, 74.8, 74.9, 75.1, 75.7, 78.0, 82.4, 84.8, 103.7, 127.6, 127.7, 127.8, 127.9, 128.0, 128.2, 128.4, 138.2, 138.3, 138.6, 138.7; HRMS (ESI) calcd for [C₄₅H₅₆NaO₇⁺] ([MNa⁺]) 731.3918; found 731.3906.

(*R*)-Methyl 2-(10-oxoundecanamido)-3-phenylpropanoate (3p)



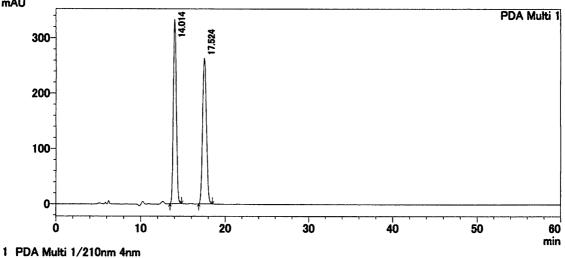
White solid, purified by recrystallization (from *n*-hexane); IR (neat) 3326, 2925, 1741, 1644 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 1.20–1.39 (m, 8H), 1.50–1.66 (m, 4H), 2.07–2.23 (m, 5H), 2.41 (t, *J* = 7.8 Hz, 2H), 3.08 (dd, *J* = 6.0, 16.0 Hz, 1H), 3.14 (dd, *J* = 6.0, 16.0 Hz, 1H), 3.72 (s, 3H), 4.86–4.93 (ddd, *J* = 6.0, 6.0, 7.4 Hz, 1H), 5.93 (d, *J* = 7.4 Hz, 1H), 7.03–7.35 (m, 5H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 23.7, 25.4, 29.0, 29.1, 29.8, 36.4, 37.8, 43.7, 52.2, 52.8, 127.0, 128.5, 129.2, 135.9, 172.1, 172.5, 209.2;

HRMS (FAB) calcd for [C₂₁H₃₂NO₄⁺] ([MH⁺]) 362.2331; found 362.2329. HPLC



(Daicel Chiralcel OD-H, hexane/2-propanol 80:20, 0.50 mL/min) $t_R = 14.0$ min. **mAU**

Enantiomer of **3p** was prepared by the hydration of the enantiomer of **2p**. A racemic mixture of **3p** and *ent*-**3p** was analyzed by HPLC [Daicel Chiralcel OD-H, hexane/2-propanol 80:20, 0.50 mL/min, $t_R = 14.0 \min(R)$, 17.5 min (*S*)].



HPLC chart of a racemic mixture of 3p and ent-3p

Gram-Scale Hydration of 2b (Scheme 3)

A mixture of **2b** (5.05 g, 6.32 mL, 42.0 mmol), **1** (0.251 mmol, 292.6 mg), HNTf₂ (0.126 mmol, 35.3 mg), H₂O (6.7 mL), and CH₃OH (42 mL) was heated at 80 °C for 12 h under air in a glass autoclave. After the autoclave was cooled to 25 °C, ethyl acetate and water was added to the mixture. The layers were separated, and the aqueous phase was extracted with ethyl acetate. The combined organic extracts were washed several times with brine, dried over Na₂SO₄, and concentrated under reduced pressure. **3b** was obtained in 92% yield (6.03 g) as a pale yellow solid after recrystallization with *n*-hexane.

5. Tables

	+ 2 CH ₃ OH 2h	5, HCl CHCl ₃ 50 °C, 12 h, air 3h	H ₃ CO OCH ₃ + CH ₃ 4h
		Ar Ar Ar Co Ar Ar Ar	
		5	
entry	Ar		yield of 4h (%) ^b
entry 1	Ar C ₆ H ₄ -p-CO ₂ CH ₃	5	yield of 4h (%) ^b 90
		5 yield of 3h (%) ^b	
1	C ₆ H ₄ - <i>p</i> -CO ₂ CH ₃	5 yield of 3h (%) ^b 4	90
1 2	C ₆ H ₄ -p-CO ₂ CH ₃ C ₆ F ₅	5 yield of 3h (%) ^b 4 2	90 85

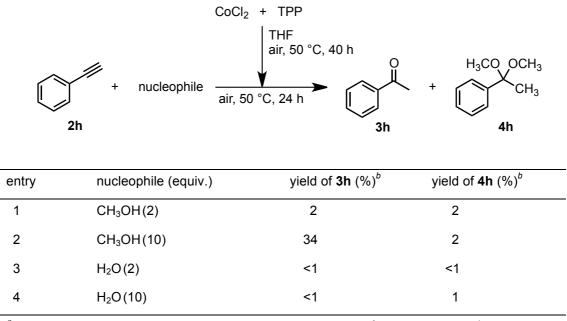
 Table S1.
 Substituent Effect on the Aromatic Ring of Tetraaryl Porphyrins^a

^{*a*}Molar ratio: **5**:HCl:**2h**: CH₃OH = 1:2:2000:22300. [**2h**] $_0$ = 1.82 M. ^{*b*}Determined by ¹H NMR using 1,1,2,2-tetrachloroethane as an internal standard.

Procedure for Table S1

A mixture of **2h** (2.0 mmol, 0.22 mL), 1,1,2,2-tetrachloroethane (50 μ L), solution of HCl in CH₃OH (2.0 μ mol, 10 mM, 0.2 mL), CH₃OH (22.3 mmol, 0.7 mL), and solution of **5** in CHCl₃ (1.0 μ mol, 5.0 mM, 0.2 mL) was heated at 50 °C for 12 h under air in a closed J. Young tube. After the mixture was cooled to room temperature, an aliquot of the mixture was directly injected into a NMR tube, then yields of the products were determined by ¹H NMR.

Table S2.Addition of Water or Methanol to $2h^a$



^{*a*}Molar ratio: CoCl₂:TPP:**2h** = 1:1:400. [**2h**]₀ = 1.0 M. ^{*b*}Determined by ¹H NMR using 1,1,2,2-tetrachloroethane as an internal standard. TPP = 5,10,15,20-tetraphenyl-21*H*,23*H*-porphyrin. THF = tetrahydrofuran.

Procedure for Table S2

A mixture of TPP (10 μ mol, 6.1 mg), THF (2 mL), and solution of CoCl₂ in THF (10 μ mol, 5.0 mM, 2 mL) was heated at 50 °C for 40 h under air in a closed J. Young tube. After the catalyst solution was cooled to room temperature, **2h** (4.0 mmol, 0.44 mL) and nucleophile was added and heated at 50 °C for 24 h under air. After the reaction, 1,1,2,2-tetrachloroethane was added to the mixture. An aliquot of the mixture was directly injected into a NMR tube, then yields of the products were determined by ¹H NMR.

6. General Procedure for Kinetic Measurement Using Calorimeter

Reactions were performed using an Ominical SuperCRC reaction calorimeter. The instrument contains a differential scanning calorimeter (DSC), which compares the heat released or consumed in a sample vessel to a reference vessel. The reaction vessels were 16 mL borosilicate screw-thread vials fit with open-top black phenolic screw caps and white PTFE septa charged with PTFE stir bars. Sample volume did not exceed 3.5 mL. In a calorimetric experiment, **2h** and CoCl₂ was added to the reaction vessel with dry CH₃OH. The vessel containing the same amount of **2h** and CH₃OH was prepared as a reference vessel. The reaction vessel and reference vessel ware placed in the calorimeter and stirred until thermal equilibrium was reached. Two syringes containing 5 mM TPP solution in CHCl₃ and CHCl₃ were placed in the injection port of the reaction vessel and the reference vessel, respectively. These two syringes were allowed to thermally equilibrate (ca. 40 min). The reaction was initiated by injecting the solution to the reaction vessel and the reference vessel at the same time in order to remove the thermal difference. The temperature of DSC was monitored using internal thermometer in calorimeter and kept on 50-52 °C. A raw data curve was produced by measuring the heat flow from the sample vessel every ten seconds during the reaction. After the reaction, the yields of the products were determined by ¹H NMR using 1,1,2,2-tetrachloroethane as an internal standard. A data curve was transformed into a square wave allowing for the response time of the instrument to be calculated using the WinCRC turbo software. The reaction rate, which is directly proportional to the heat flow, fraction conversion, and instantaneous concentration of reactants/products can all

be calculated from the data curve. Details of kinetic measurement using calorimeter are described in the literature.⁷

7. Figures

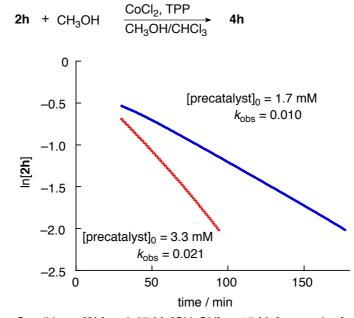


Figure S1. Determination of Reaction Order in [2h] and [precatalyst]

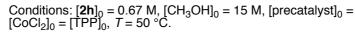
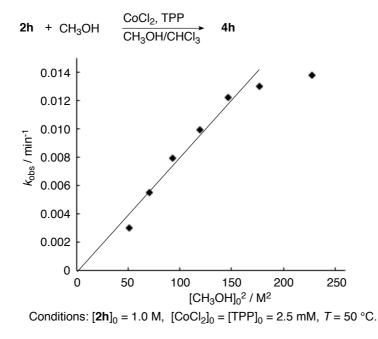
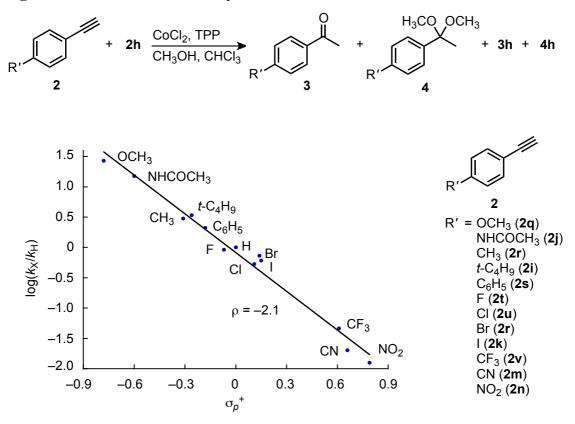


Figure S2. Determination of Reaction Order in [CH₃OH]



⁷ Blackmond, D. G. Angew. Chem. Int. Ed. 2005, 44, 4302 – 4320.

Figure S3. Hammett Plot Analysis



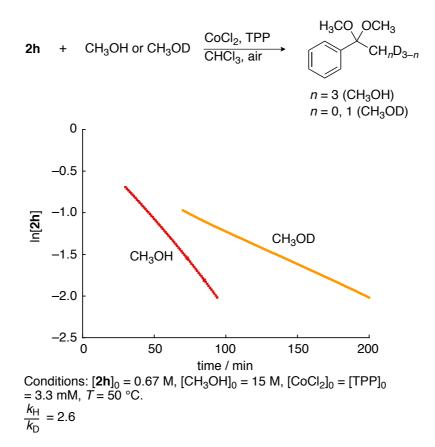
Conditions: $[2]_0 = [2h]_0 = 0.17 \text{ mM}$, $[CH_3OH]_0 = 15 \text{ M}$, $[catalyst]_0 = [CoCl_2]_0 = [TPP]_0$, $T = 50 \degree C$. Unless otherwise noted, reaction rate was measured using 2h as an internal reference. Reaction rate for 2v, 2n, and 2m were calibrated using 2r, 2v, and 2m, respectively.

Procedure for Figure S3

A mixture of **2h** (0.22 mL, 2.0 mmol), **2** (0.5 mmol), CH₃OH (0.78 mL), CoCl₂ (5.0 mM in CH₃OH, 1mL, 5.0 µmol) and TPP (5.0 mM in CHCl₃, 1.0 mL, 5.0 µmol) was heated at 50 °C under air in the closed glass tube. The yields of **4h**, **4q**, **4j**, **4r**, **4i**, **4s**, **4t**, **4u**, **4r**, **4k**, **4v**, **4m**, and **4n** were determined by ¹H NMR using 1,1,2,2-tetrachloroethane as an internal standard based on the signals at 1.54 ppm (s, 3H), 1.53 ppm (s, 3H), 1.52 ppm (s, 3H), 1.59 ppm (s, 3H), 1.59 ppm (s, 3H), 1.59 ppm (s, 3H), 1.55 ppm (s, 3H), 1.52 ppm (s, 3H), respectively. The yields of **3h**, **3q**, **3j**, **3r**, **3i**, **3s**, **3t**, **3u**, **3r**, **3k**, **3v**, **3m**, and **3n** were determined based on the signals at

2.60 ppm (s, 3H), 2.57 ppm (s, 3H), 2.42 ppm (s, 3H), 2.59 ppm (s, 3H), 2.67 ppm (s, 3H), 2.67 ppm (s, 3H), 2.61 ppm (s, 3H), 2.61 ppm (s, 3H), 2.60 ppm (s, 3H), 2.60 ppm (s, 3H), 2.67 ppm (s, 3H), 2.67 ppm (s, 3H), and 2.71 ppm (s, 3H), respectively.

Figure S4. Kinetic Isotope Effect



Procedure for Figure S4

According to the general procedure on kinetic measurement using calorimeter, **1a** (0.22 mL, 2.0 mmol) and CH₃OD (0.78 mL) were charged both in the reaction vessel and the reference vessel. CoCl₂ (10 mM in CH₃OD, 1mL, 10 μ mol) and CH₃OD (1.0 mL) were added to the reaction vessel and the reference vessel respectively, and the reaction was started by adding TPP (10 mM in CHCl₃, 1.0 mL, 10 μ mol) and CHCl₃ (1.0 mL) to the reaction vessel and the reference vessel, respectively. After the kinetic measurement, ¹H

NMR and GCMS analysis of the reaction mixture indicated consumption of **1a** (100 %), and the presence of (2,2-D₂)**2a**, (2,2,2-D₃)**2a**, (2,2-D₂)**3a**, and (2,2,2-D₃)**3a**.