

“Diversity Oriented Synthesis of a Vinblastine-Templated Library of 7-Aryl Octahydroazonino[5,4-*b*]indoles via a Three-Component Reaction”

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Supporting Information

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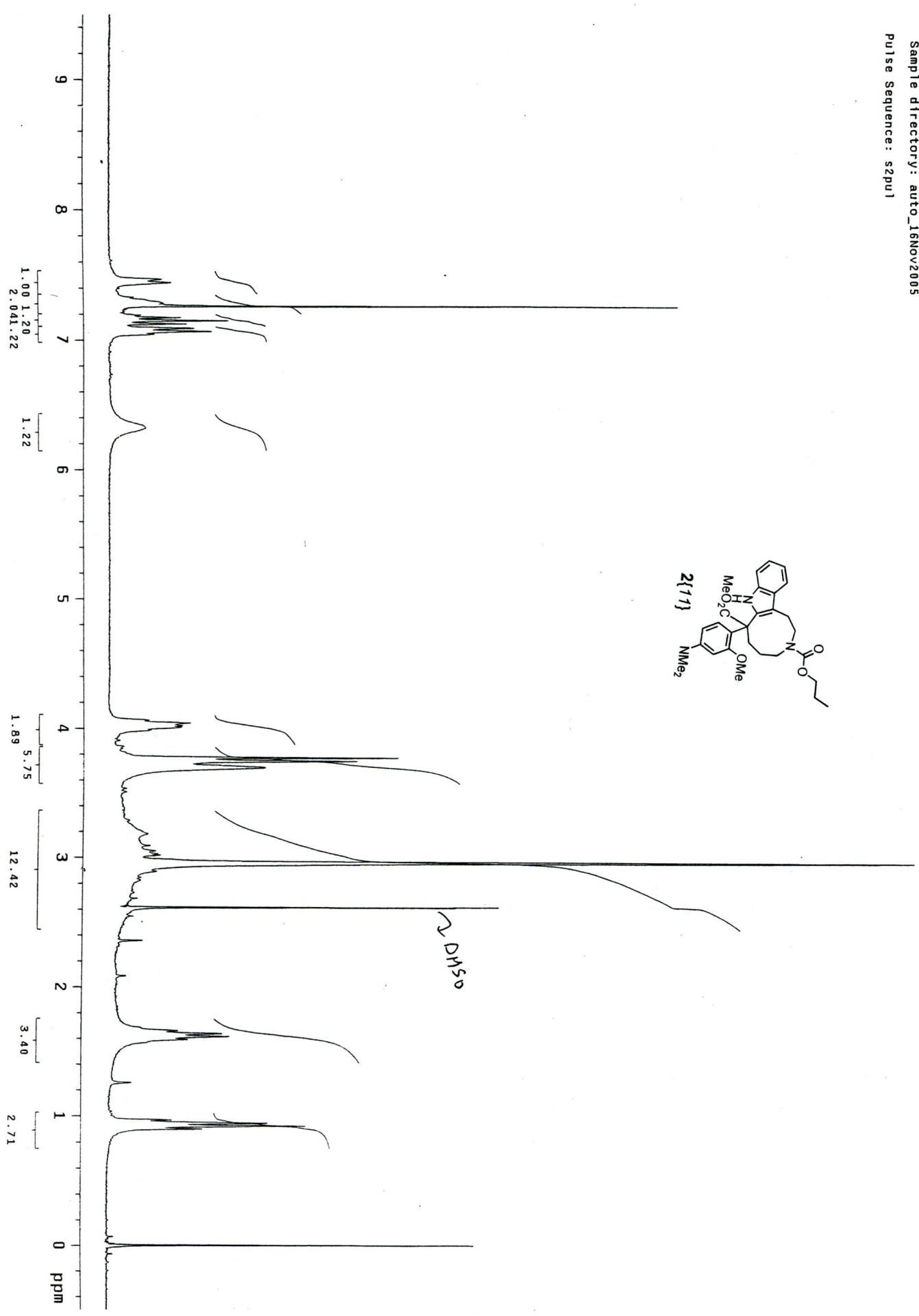
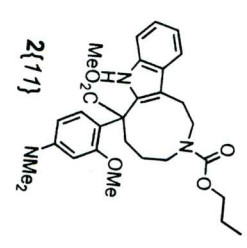
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General Methods. All reagents and solvents were purchased and used without further purification. Reactions were performed into two-drum round bottom vials in 24-well plates. All reactions were monitored by reverse phase HPLC on a Shimadzu LC system. Routine LC/MS was run in an integrated LC/MS system with a Shimadzu LC and a Micromass DMZ MS instrument. Solvent was removed from reaction vials utilizing a Savant or Genevac evaporation system. ^1H NMR and ^{13}C NMR spectra were recorded on a Varian Mercury spectrometer. Low-resolution mass spectra were recorded in ES^+ mode.

Purification Conditions. The crude products were dissolved in 500 μL of DMSO and then purified by reverse phase prep-HPLC with mass triggered fraction collection. An Xterra Prep MS-C18 column (19 $\times$ 50 mm) was utilized with water and acetonitrile as gradients (80 mL/min) and NH_3 (10 mM) as modifier. Samples were collected in 24-vial bar-coded purification blocks followed by solvent evaporation. The dried samples were then weighed using a balance controlled by an automation system to calculate the recovery, and then diluted in DMSO to a 30 mM concentration. An aliquot of 20 μL was pulled from each vial and then diluted into 500 μL of DMSO for characterization. The samples were analyzed by a 2.5-minute reverse-phase HPLC/ $\text{UV}_{214\text{nm}}$ /ELSD/MS method to confirm the MS of the products and to determine their purity. The samples were finally reformatted in a 96-well plate for biological screening.

Archive directory: /export/home/auto/vnmr-sys/data
Sample directory: auto_16Nov2005
Pulse Sequence: s2pu1



VIN-185-C13
13C OBSERVE

pad=2 run with findz0 before acquisition

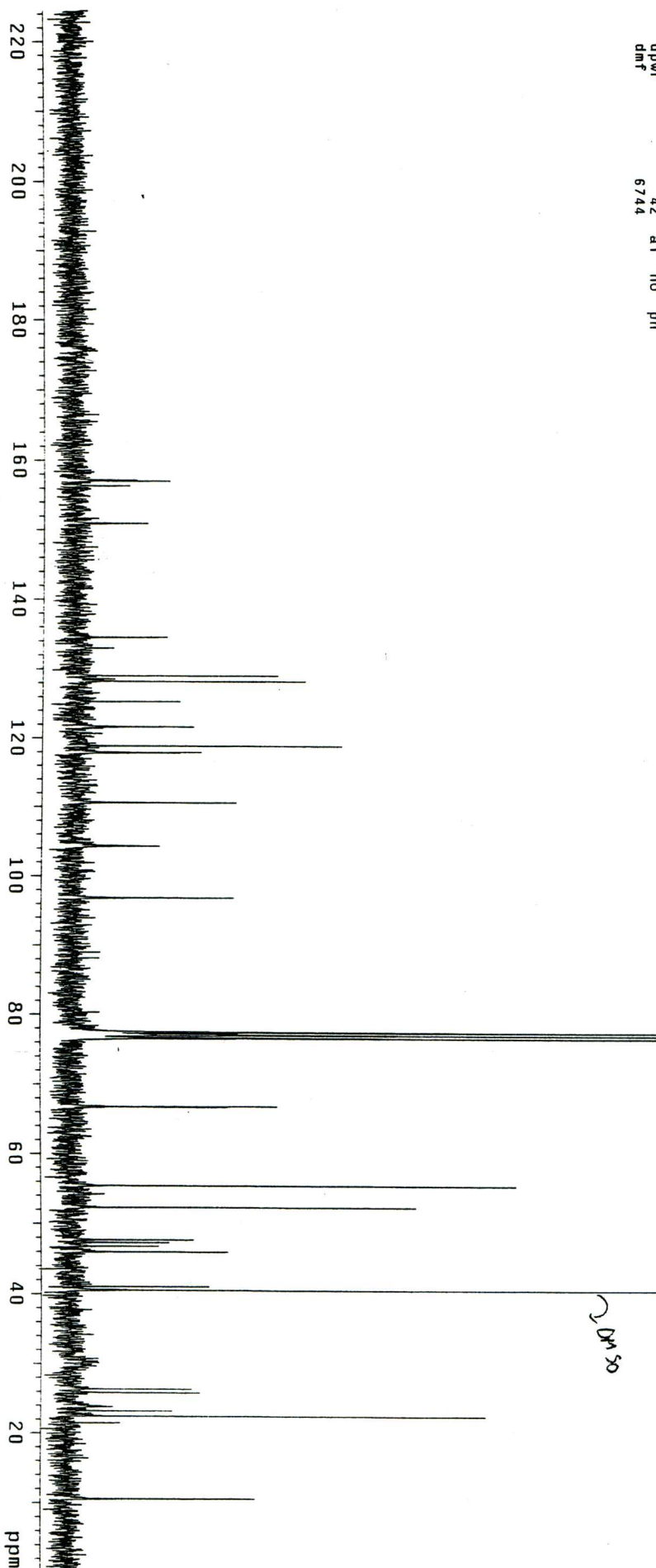
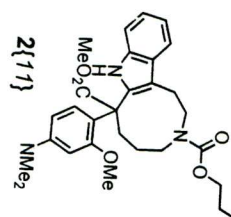
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o_17.10.05/VIN-185~		pw90	13.500
-C1301.fid	alfa		20.000

ACQUISITION		FLAGS	
sw	18832.4	11	n
at	1.815	in	n
np	68362	dp	y
fb	10400	hs	nm
bs	64		
dl	2.000	lb	1.00
nt	5000	fn	not used
ct	5000	DISPLAY	

TRANSMITTER		SP		WD	
tn	C13	sp	-37.7	wd	16960.8
sfreq	75.390	rf1	6946.8		
tof	740.3	rfp	5804.4		
tpwr	58	rp	-78.4		
pw	6.750	tp	-301.2		

DECOUPLER		H1		WC	
dn	H1	sc	250	wc	0
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dm	yy	th	50		
dmm	w	at			
dpwr	42	no			
dmf	6744	ph			

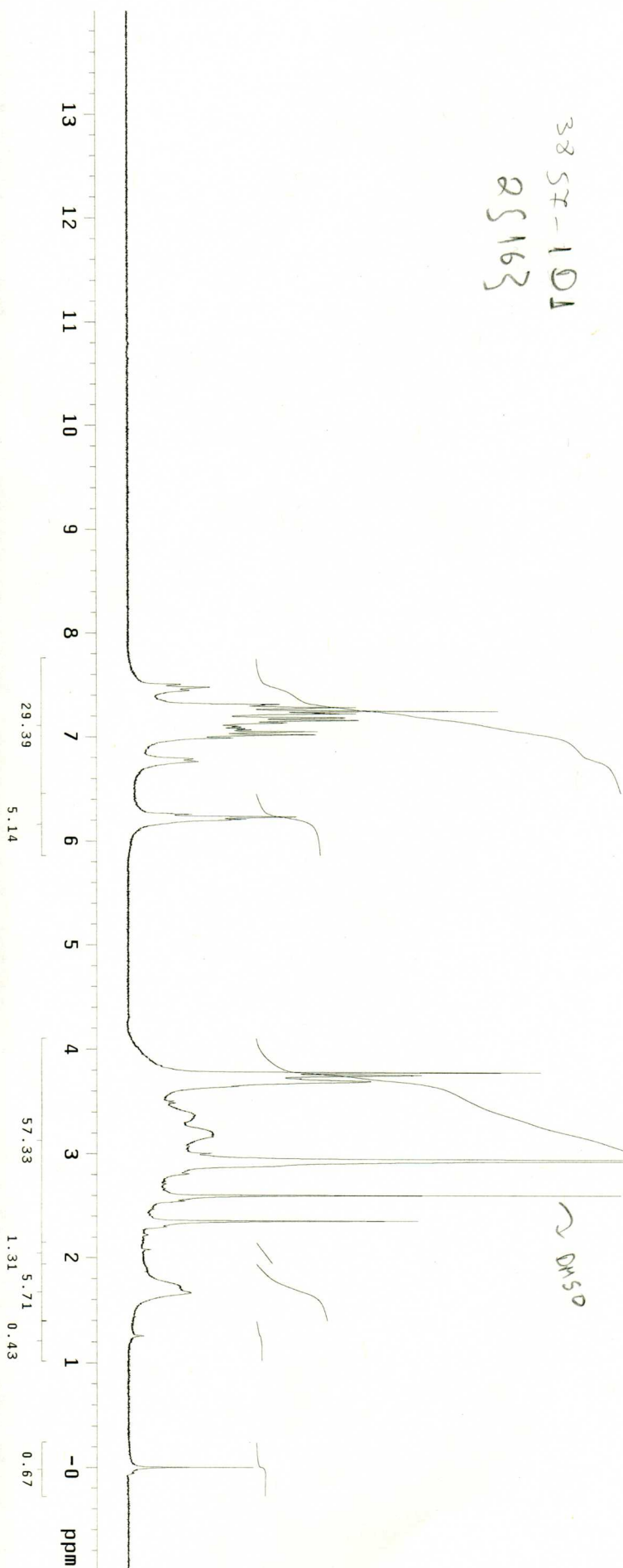
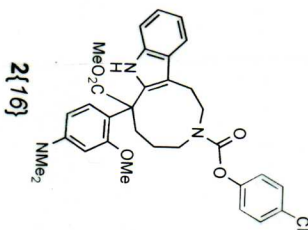


1B6
STANDARD 1H OBSERVE

Automation directory: /export/home/auto/walk_auto/auto_12.10.05
File : 1B602

Pulse Sequence: s2pul
Solvent: cdcl3
Ambient temperature
Sample #20
File: 1B601
Mercury-3000B "horror"

Relax. delay 1.000 sec
Pulse 45.0 degrees
Acq. time 1.998 sec
Width 4796.2 Hz
32 repetitions
OBSERVE H1, 299.788555 MHz
DATA PROCESSING
FT size 32768
Total time 1 min, 39 sec



IC1
STANDARD 1H OBSERVE

Automation directory: /export/home/auto/walk_auto/auto_12.10.05
File: IC102

Pulse Sequence: s2pul

Solvent: cdcl3

Ambient temperature

Sample #21

File: IC101

Mercury-300BB "horror"

Relax. delay 1.000 sec

Pulse 45.0 degrees

Acq. time 1.998 sec

Width 4796.2 Hz

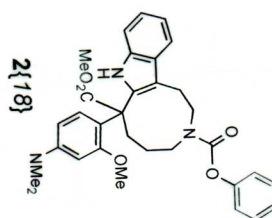
32 repetitions

OBSERVE H1, 299.785529 MHz

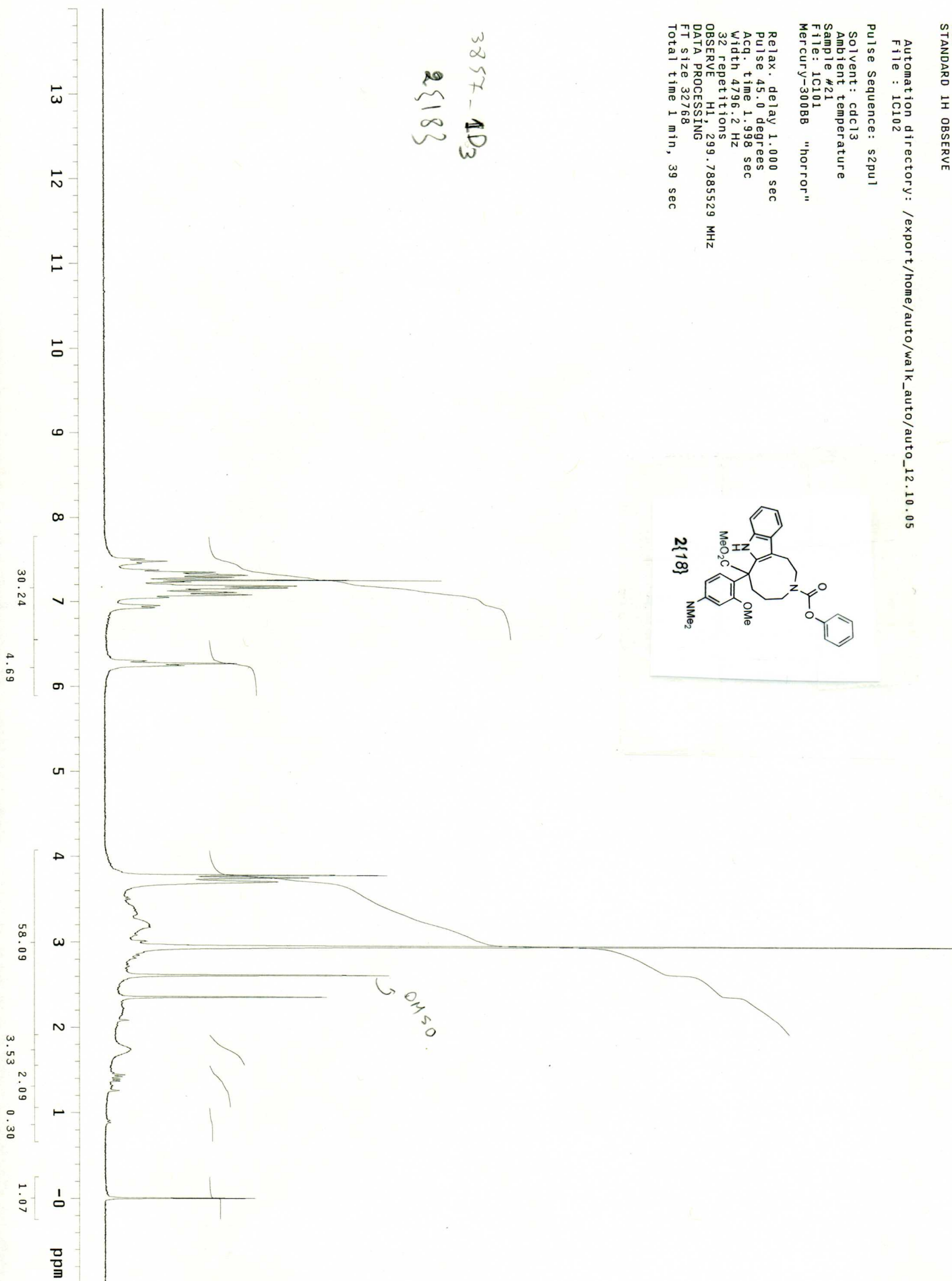
DATA PROCESSING

FT size 32768

Total time 1 min, 39 sec



3857-1D3
2{183}



vin-2B4-c13
STANDARD 1H OBSERVE

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File: vin-2B4-c1302

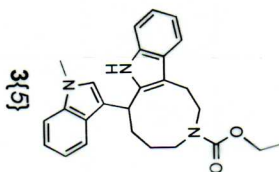
Pulse Sequence: s2pul

Solvent: cdcl3
Ambient temperature
Sample #4
File: vin-2B4-c1301
Mercury-300BB "horror"

Relax. delay 1.000 sec
Pulse 45.0 degrees
Acq. time 1.998 sec
Width 4796.2 Hz
32 repetitions
OBSERVE H1, 299.785540 MHz
DATA PROCESSING
F1 size 32768
Total time 1 min, 39 sec

3854.45

3{5}



13 12 11 10 9 8 7 6 5 4 3 2 1 -0 ppm

0.09

34.46

2.32

47.43

14.13

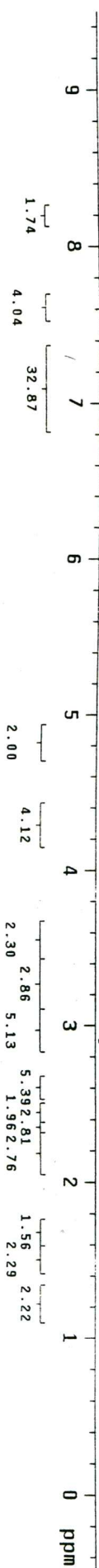
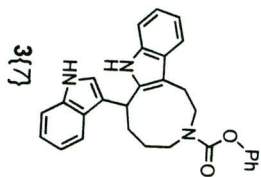
1.57

expt s2pu1

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solvent cdc13 gain not used
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auto/walk-auto/aut~ hst 0.008
_11.10.05/28501.f~ pw90 17.250
id atfa 20.000

ACQUISITION FLAGS
sw 4796.2 11 n
at 1.998 11 y
np 19166 11 y
fb 2500 11 y
bs 16 11 y
dl 1.000 11 y
nt 32 11 y
ct 32 11 y
tn TRANSMITTER H1 f1
sfrq 299.790 rf1
tof 315.5 rf1
tpwr 60 tp
pw 8.625 PLOT
DECOUPLER wc 250
dn C13 SC 0
dof 0 VS 962
dm nm th 2
dmm c at cdc ph
dpwr 44
dmf 13900

20M50



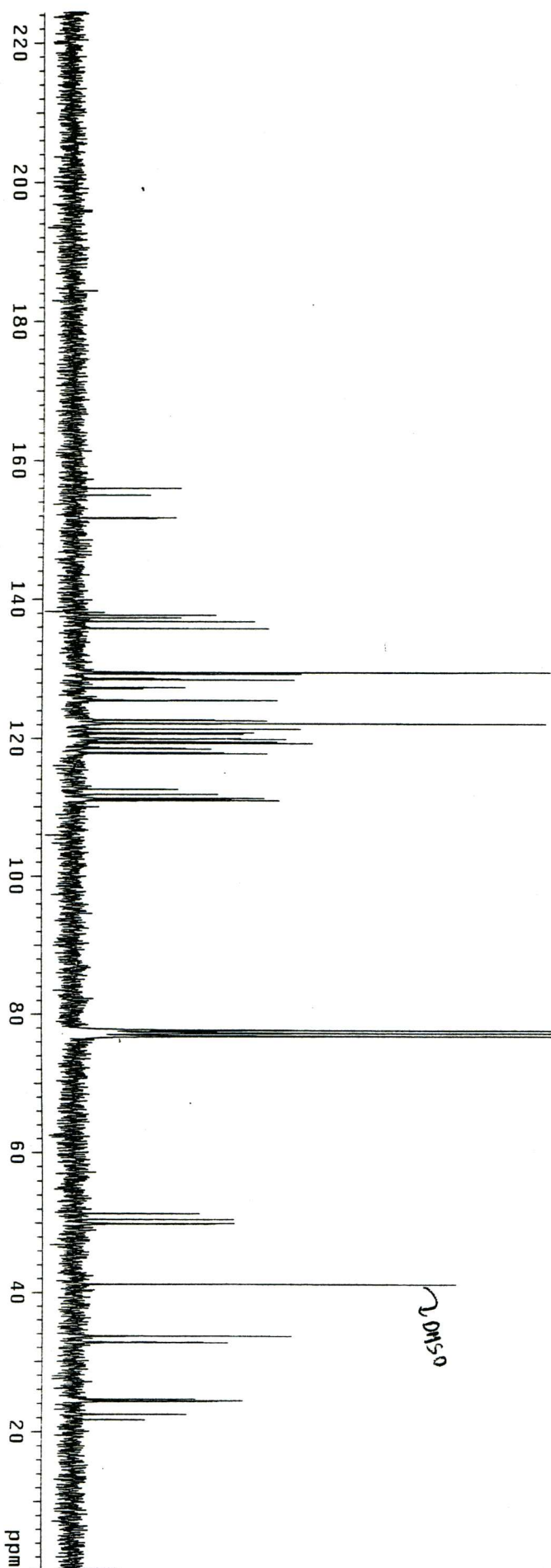
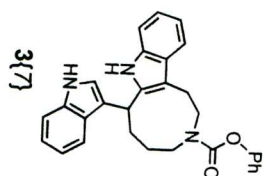
vin-285-c13
expt s2pu1

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date	Oct 23 2005	temp	not used
solvent	cdcl3	gain	not used
file	/export/home/~	spin	20
auto/walk	auto/aut~	hst	0.008
o_20.10.05/vin-185~		pw90	13.500
-c1301.f1d		alpha	20.000

ACQUISITION		PROCESSING	
sw	18832.4	fl	n
at	1.815	in	n
np	68362	dp	y
fb	10400	hs	nn
bs	64		
dl	2.000	lb	1.00
nt	5000	fn	not used
ct	5000		

TRANSMITTER		DISPLAY	
tn	C13	sp	-37.9
stfq	75.390	wp	16960.8
tof	740.3	rf1	1124.2
tpwr	58	rfp	0
pw	6.750	lp	-97.0
			-264.3

DECOUPLER		PLOT	
dn	H1	wc	250
dof	0	sc	0
dm	yyv	vs	3123
dmm	w	th	10
dpvr	42	ai	no
dmf	6744	ph	



expi szpul

SAMPLE SPECIAL
date Oct 11 2005 temp not used
solvent cdc13 gain not used
file /export/home/~ spin 20
auto/walk_auto/aut~ hst 0.008
o_11.10.05/28601.f~ pw90 17.250
id a1fa 20.000

ACQUISITION
sw 4796.2 11
at 1.998 1n
np 19166 1n
fb 2600 16
bs 16
di 1.000 1n
nt 32 32
ct 32 32

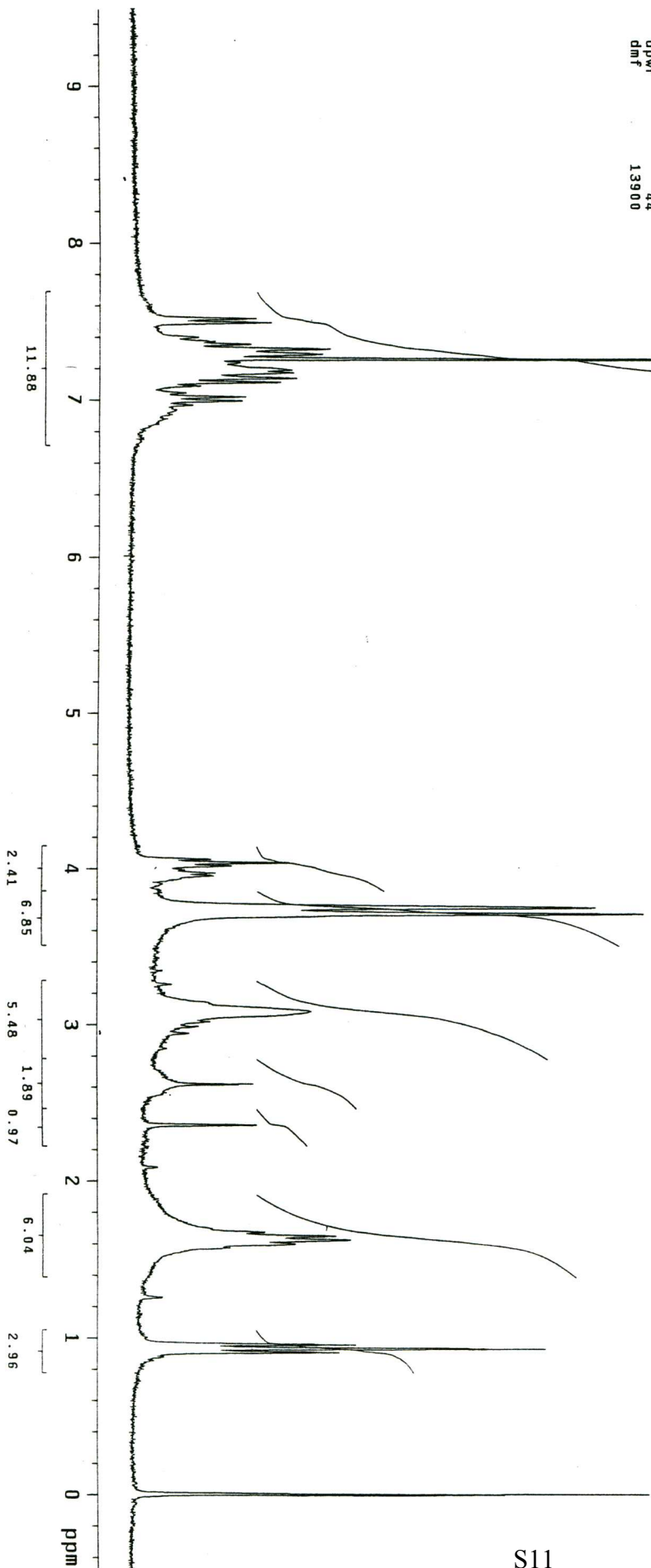
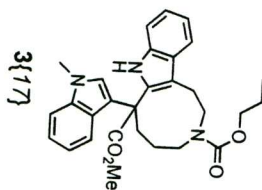
TRANSMITTER
tn H1
sfreq 299.790 rfp
tof 315.5 rfp
tpwr 60 1p
pw 8.625 1p

DECOUPLER
dn C13
dof 0 vs
dm nnn th
dmm C at
dpwr 44 cdc
dmf 13900 ph

PROCESSING
not used
display not used

FLAGS
n
n
y
nn

PLOT
250
1148
10



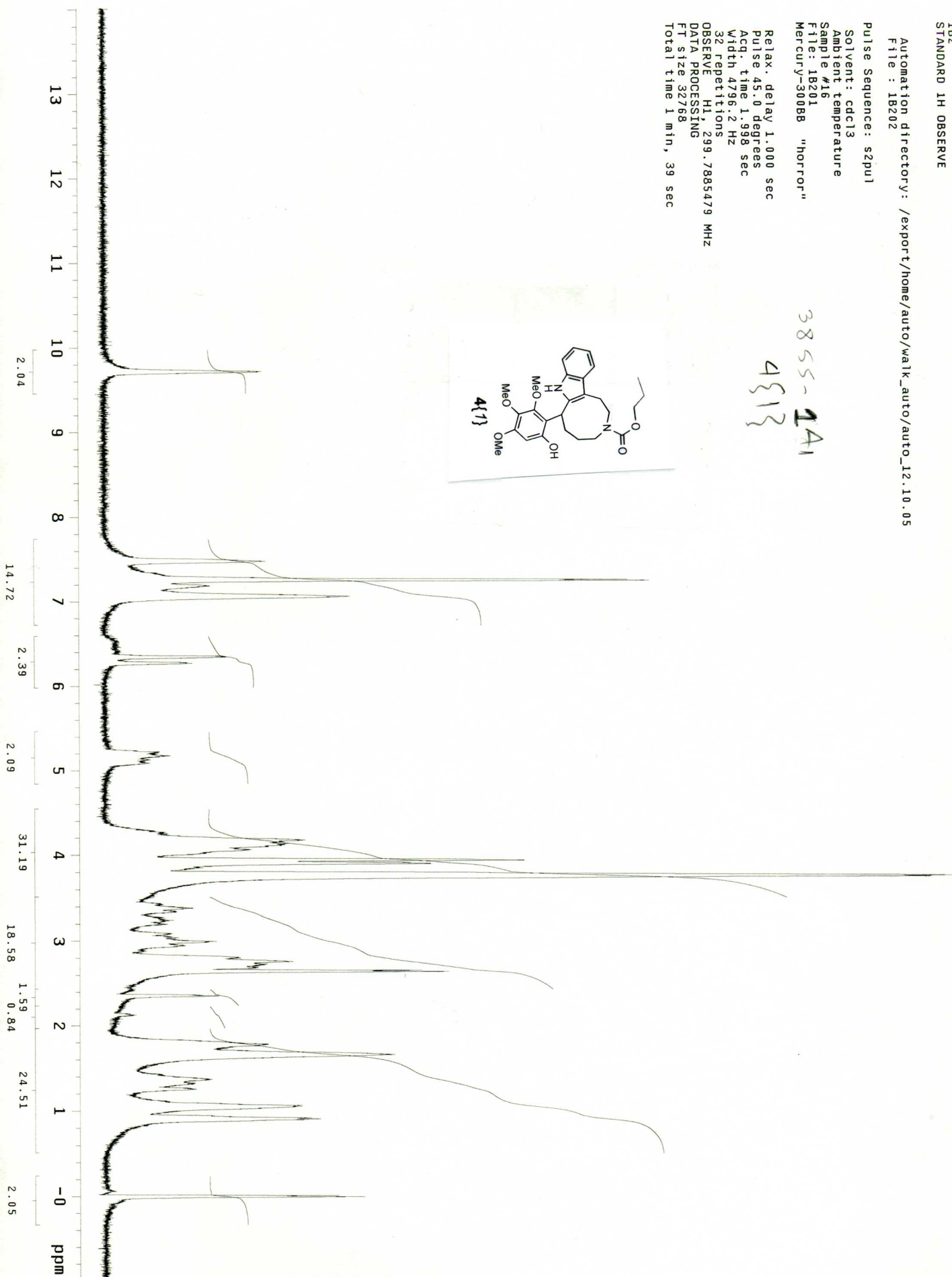
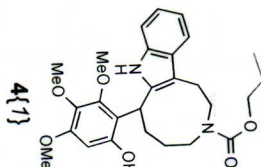
1B2
STANDARD 1H OBSERVE

Automation directory: /export/home/auto/auto_walk_auto_12.10.05
File : 1B202

Pulse Sequence: szpul
Solvent: cdcl3
Ambient temperature
Sample #16
File: 1B201
Mercury-300BB "horror"

Relax. delay 1.000 sec
Pulse 45.0 degrees
Acq. time 1.998 sec
Width 4796.2 Hz
32 repetitions
OBSERVE H1, 299.7865479 MHz
DATA PROCESSING
FT size 32768
Total time 1 min, 39 sec

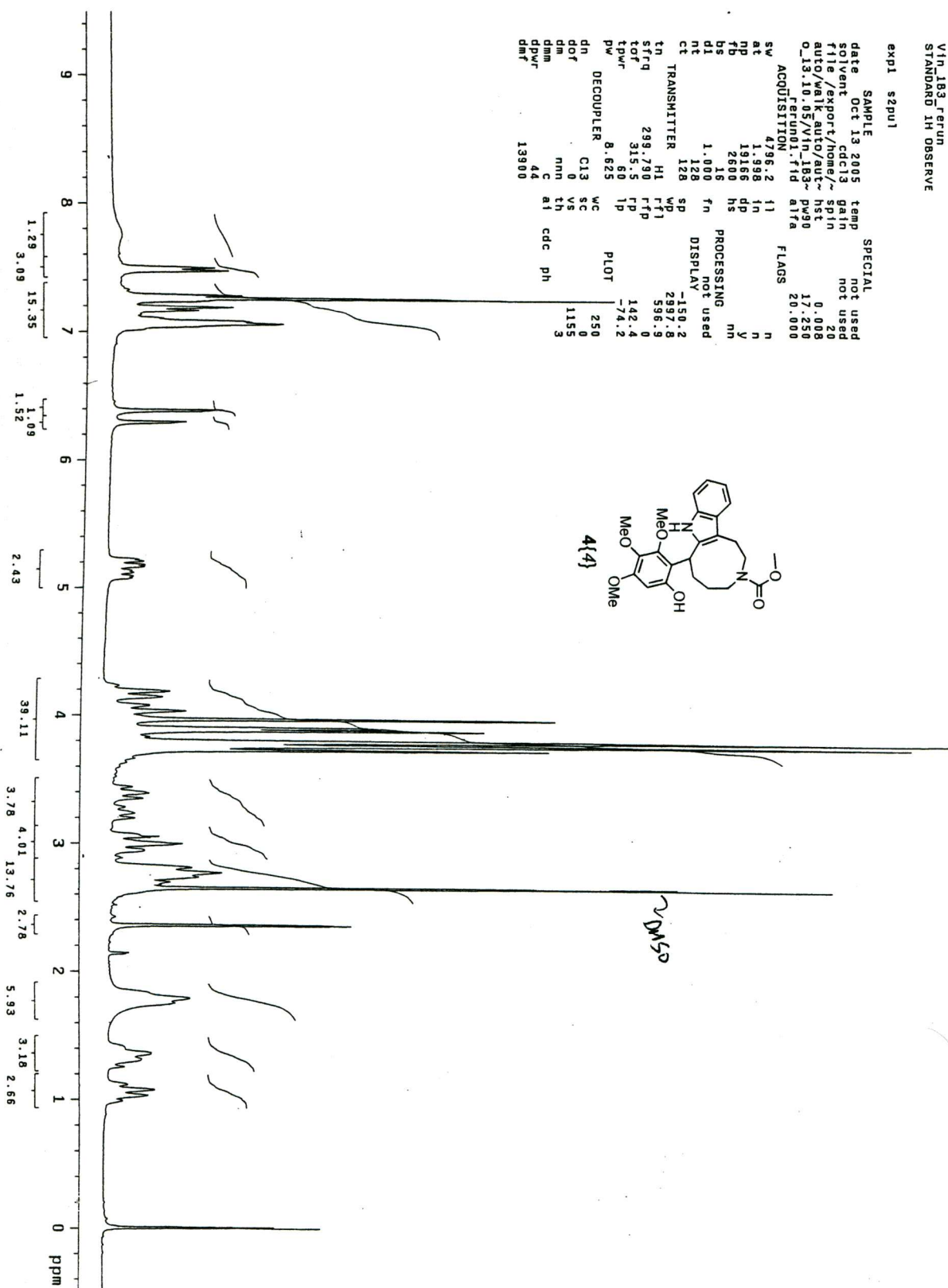
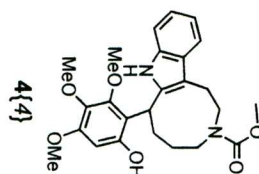
3855-1A1
4{1}



VIN_183_ferun
STANDARD 1H OBSERVE

expt1 s2pu1

SAMPLE		SPECIAL	
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solvent	cdcl3	gain	not used
file	/export/home/~	spin	20
auto	auto/home/~	hsc	0.008
o-13	10.85/VIN_183-	pw90	17.250
ferun01	titd	alra	20.000
ACQUISITION		FLAOS	n
sv	4795.2	fn	n
at	1199.2	dp	y
mp	1915	hs	m
fb	2600	hs	m
bs	16	hs	m
dl	1.000	fn	not used
nt	128	fn	not used
ct	128	sp	-150.2
tn	TRANSMITTER	wp	2997.6
stfrq	299.790	rfl	596.9
tof	315.5	rfl	142.4
tpwr	60	lp	-74.2
pw	8.625	lp	-74.2
DECOUPLER		WC	250
dn	CD3	sc	0
dof	0	vs	1155
dm	mn	th	3
dmm	c	at	cdc
dpwr	44	ph	
dmf	13900		



VIN-183-C13
13C OBSERVE

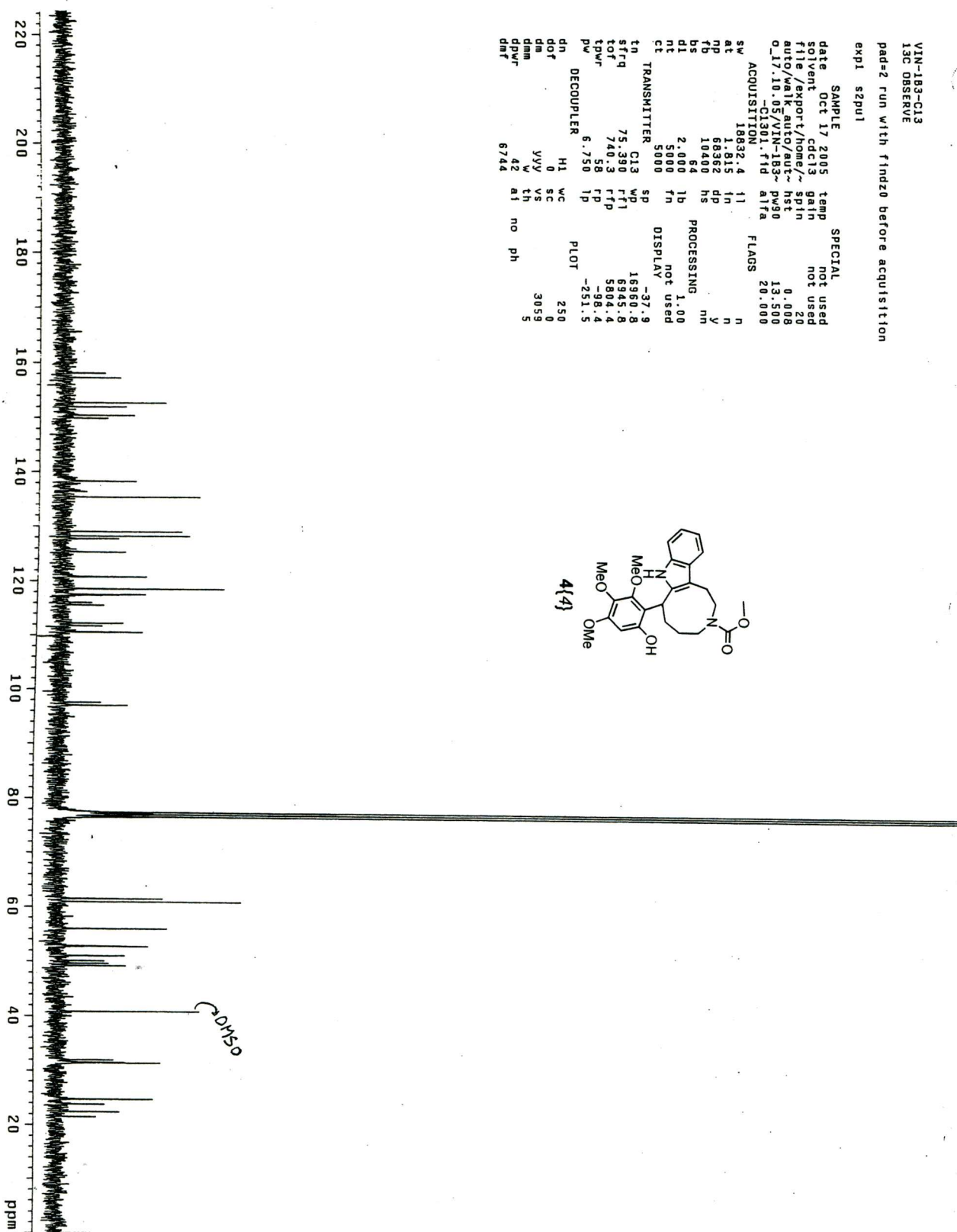
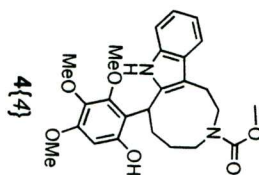
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expt szpul

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solvent CDCl3 gain not used
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auto/waltz16/aut/ PR80 13.500
O-17.10.05/VIN-183- PR80 20.000
C1301.fid a1fa 20.000

ACQUISITION FLADS
sw 100.625 11 n
at 1.815 1n y
np 68362 1n y
fb 10400 1n y
bs 64 1n y
d1 2.000 1n y
nt 5000 1n y
ct 5000 1n y

TRANSMITTER SP DISPLAY
tn C13 WD -37.9
sfrq 75.390 rfi 16960.8
tof 740.3 rfp 6945.8
tpw 58 rfp 5804.4
pw 6.750 1p -98.4
DECOUPLER H1 WC PLOT -251.5
dn 0 SC 250
dof 0 SC 0
dm 0 SC 0
dmm yvy vs 3059
dpr 42 ai no ph 5
dmf 6744
```



1B4
STANDARD 1H OBSERVE

Automation directory: /export/home/auto/walk_auto/auto_12.10.05
File : 1B402

Pulse Sequence: s2pul

Solvent: cdcl3
Ambient temperature

Sample #18

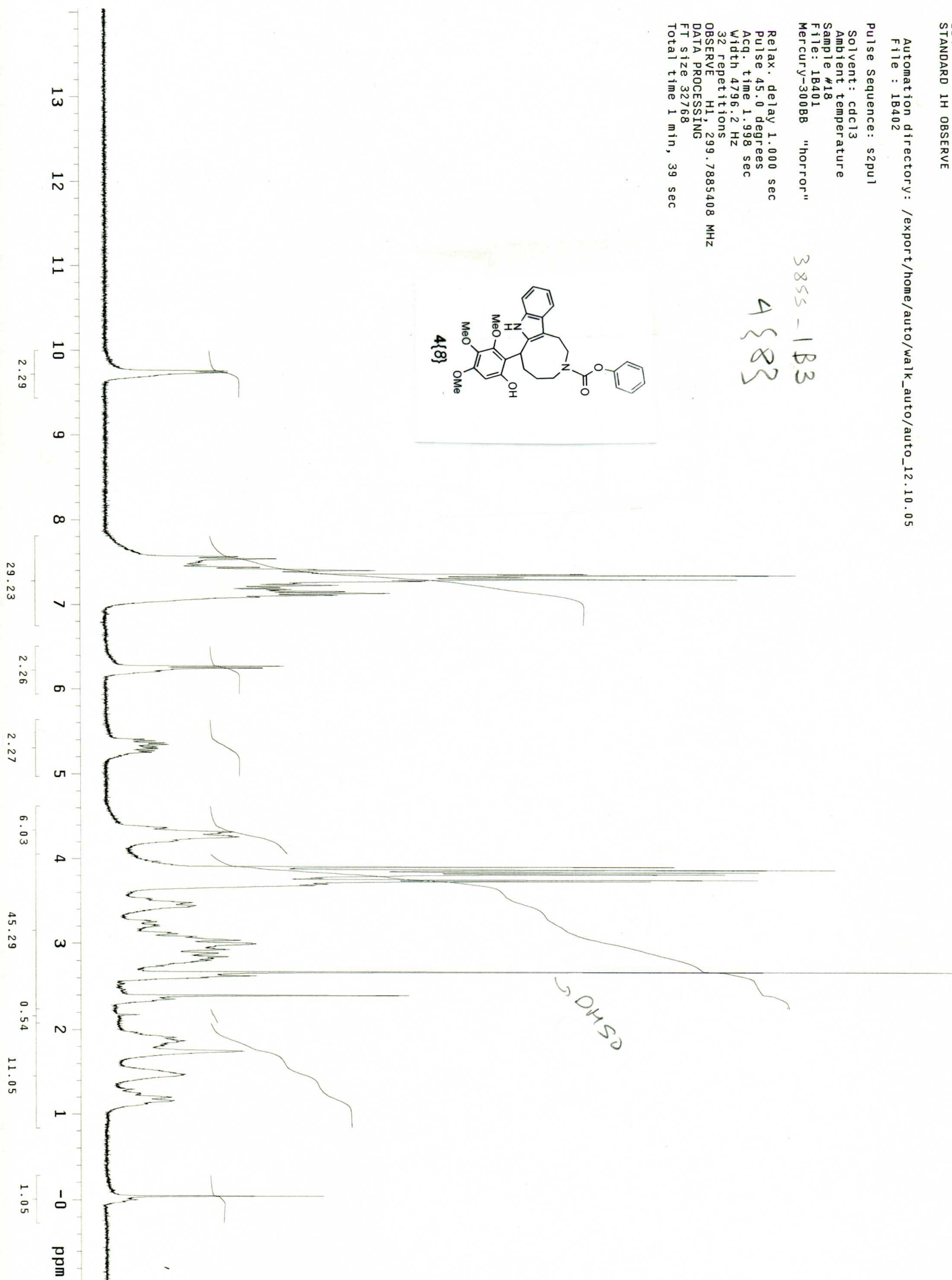
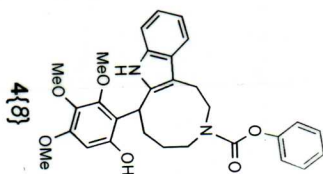
File: 1B401

Mercury-300BB "horror"

3855-1B3

4{82}

Relax. delay 1.000 sec
Pulse 45.0 degrees
Acq. time 1.998 sec
Width 4796.2 Hz
32 repetitions
OBSERVE H1 299.785408 MHz
DATA PROCESSING
FT size 32768
Total time 1 min, 39 sec

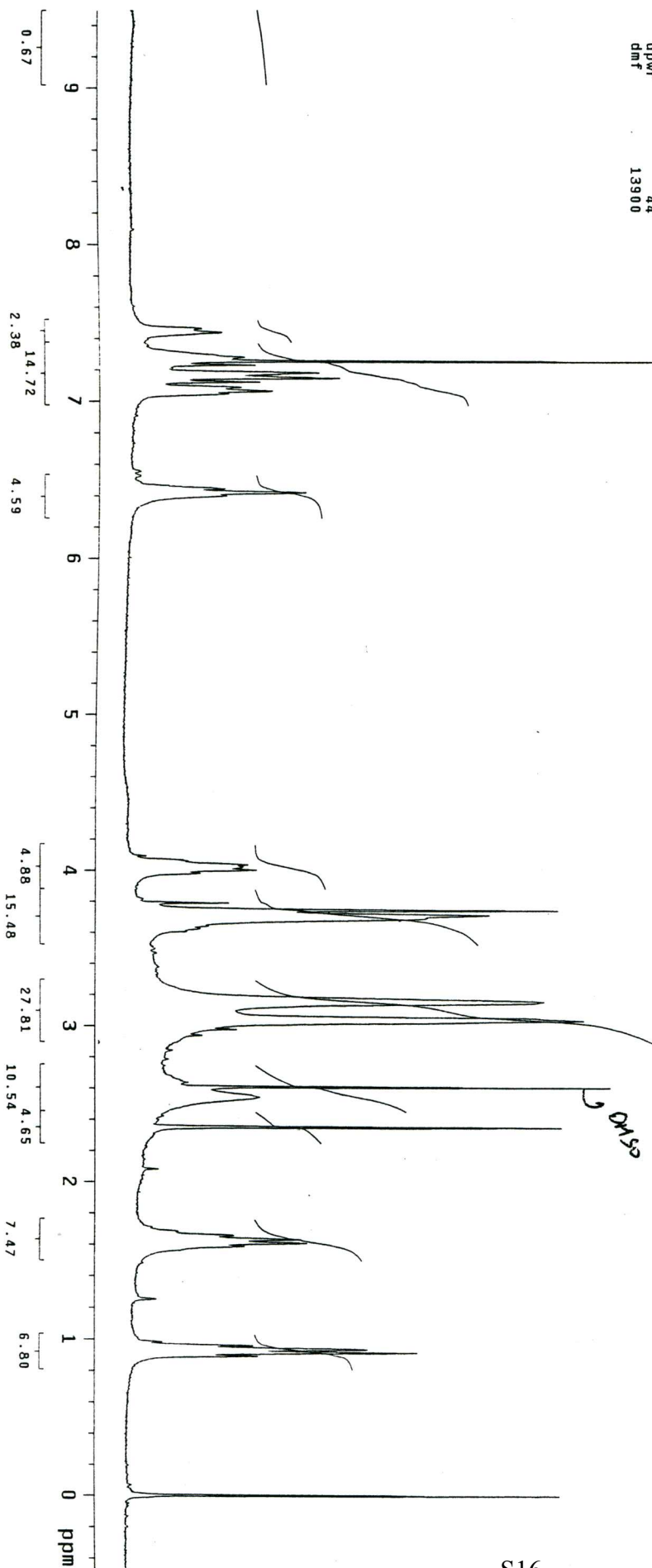
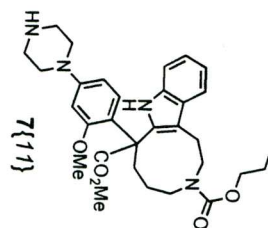


expt s2pu1

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solvent	cdcl3	gain	not used
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auto/walk	auto/aut~	hst	0.008
o_11.10.05/2A501.f~	pw90	17.250	
	id	20.000	
	alfa		

ACQUISITION		PROCESSING	
sw	4796.2	fn	not used
at	1.998	sp	-150.2
np	19166	wp	2997.8
fb	2600	h1	597.5
bs	16	rfl	0
dl	1.000	rff	139.3
nt	32	tp	-65.9
ct	32	ip	
tn	299.790	pl	
stfrq	315.5		
tofr	60		
tpwr	8.625		

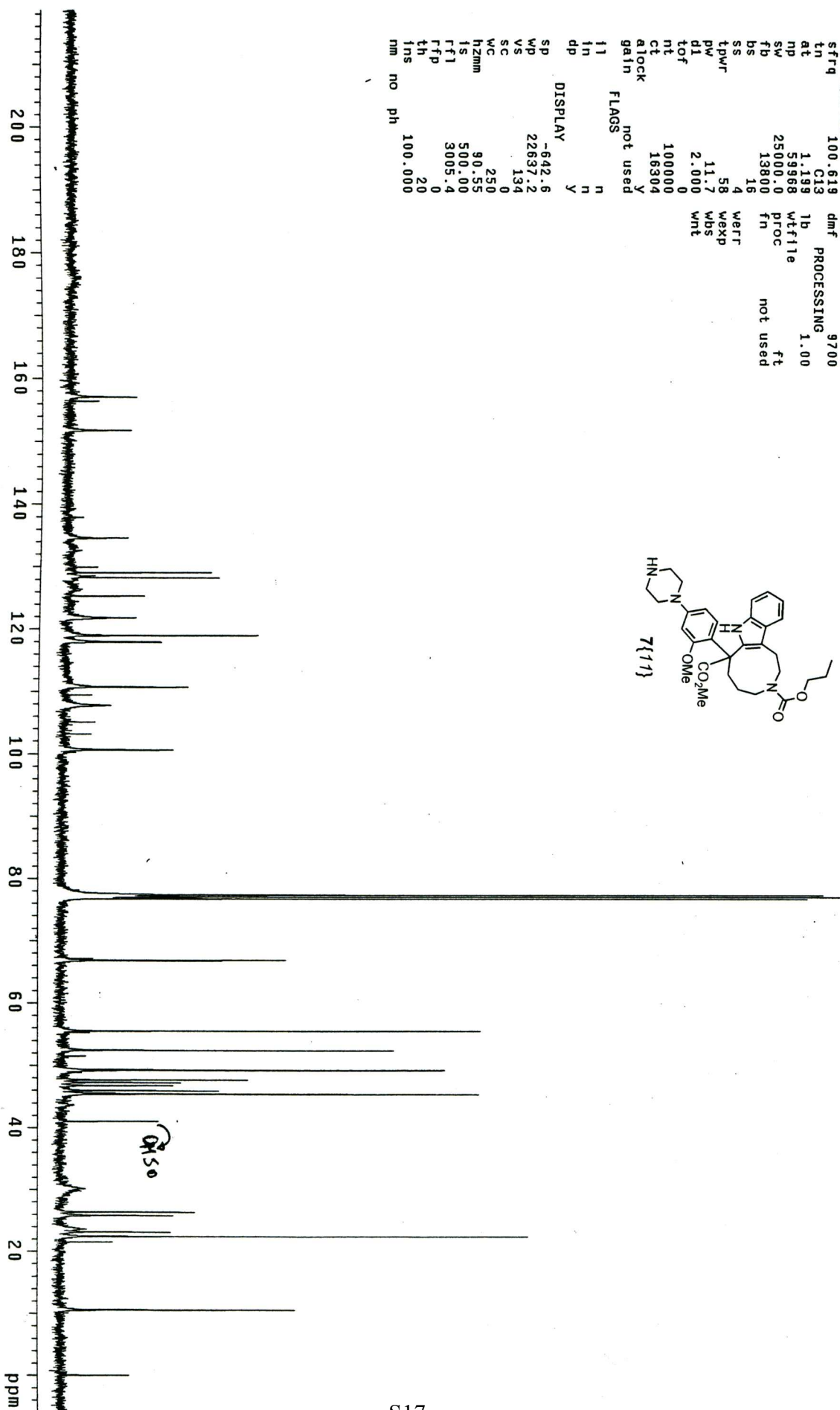
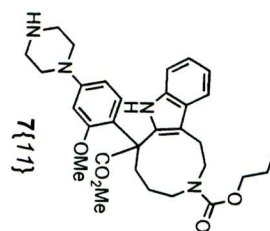
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dot	0	vs	1791
dm	nnn	th	16
dmm	C	at	cdc
dpwr	44	ph	
dmt	13900		



vin-2A5-c13
exptl std13c

SAMPLE DEC. & VT
date Oct 24 2005 dfrq 400.120
solvent CDC13 dn H1
file /export/home/~ dpwr 37
mkaselj/vin-2A5-C1~ dof 0
3 yyy
v

ACQUISITION
sfrq 100.619 dmm 9700
tn C13 dmf
at 1.19 lb PROCESSING 1.00
np 59968 wtfll
sw 2500.0 proc ft
fb 13800 fn not used
bs 16
ss 4 weff
tpwr 58 wexp
pw 11.7 wbs
d1 2.000 wnt
tof 0
nt 100000
ct 16304
atlock y
gain not used
FLAGS
f1 n
in n
dp y
DISPLAY
sp -642.6
wp 22637.2
vs 134
sc 0
wc 250
hzmm 90.35
is 500.00
rf1 3005.4
rfp 0
th 20
ins 100.000
nm no ph



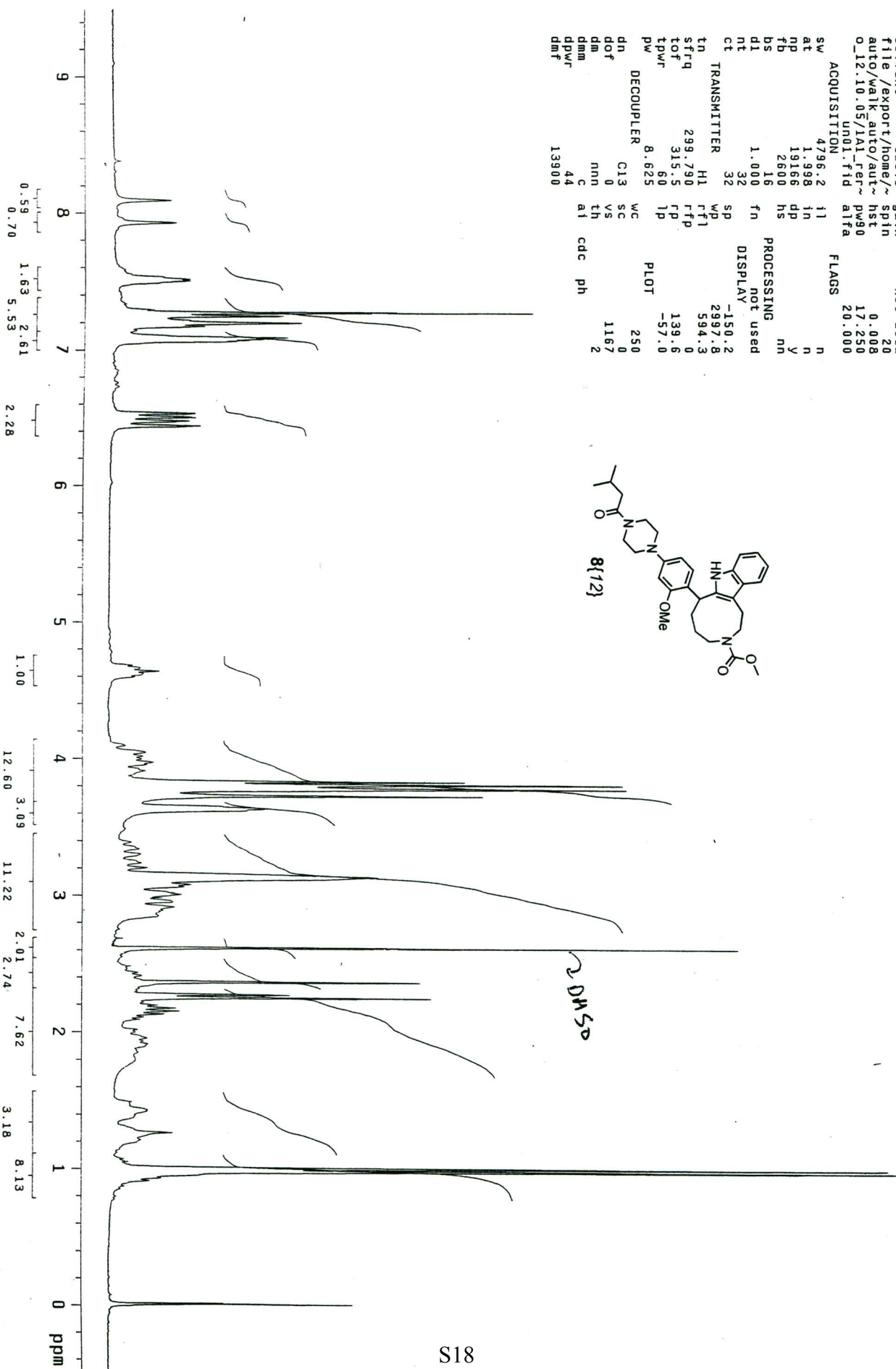
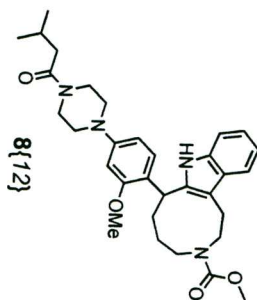
expl s2pul

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auto/walk	auto/aut~	hst	0.008
o_12.10.05/1A1_ref~	pw90		17.250
	un01.fid	at1a	20.000

ACQUISITION		FLAGS	
sw	4796.2	11	n
at	1.998	in	n
np	19166	dp	y
fb	2600	hs	nn
bs	16		
dl	1.000	fn	not used
nt	32		
ct	32		

TRANSMITTER		PROCESSING	
tn	H1	sp	-150.2
sfreq	299.790	wp	2997.8
tofr	315.5	rfl	594.3
tpwr	60	fp	139.6
	8.625	tp	-57.0

DECOUPLER		PLOT	
dn	C13	wc	250
dof	0	sc	0
dm	nmn	th	1167
dmm	c		2
dpwr	44		
dmf	13900		



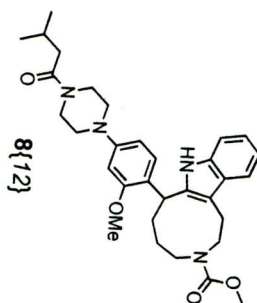
pad=2 run with findz0 before acquisition
expi szpu1

SAMPLE		SPECIAL	
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solvent	cdcl3	gain	not used
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auto/walk	auto/aut~	hst	0.008
o_13.10.05/VIN-1A1~		pw90	13.500
-C1301.fid		alfa	20.000

ACQUISITION		FLAGS	
sw	18832.4	11	n
at	1.815	in	n
np	68362	dp	y
fb	10400	hs	nn
bs	64		
di	2.000	1b	not used
nt	5000	fn	1.00
ct	5000	DISPLAY	not used

TRANSMITTER		DISPLAY	
tn	C13	sp	-37.9
sfrq	75.390	wp	16960.8
tof	740.3	rf1	6946.7
tpwr	58	rfp	5804.4
pw	6.750	ip	-93.9
			-269.0

DECOUPLER		PLOT	
dn	H1	WC	250
dof	0	SC	0
dm	yyv	VS	3049
dmm	w	th	11
dpwr	42	at	no
dmf	6744	ph	



1A2
STANDARD 1H OBSERVE

Automation directory: /export/home/auto/walk_auto/auto_12.10.05
File: 1A202

Pulse Sequence: s2pu1

Solvent: cdcl3
Ambient temperature

Sample #10

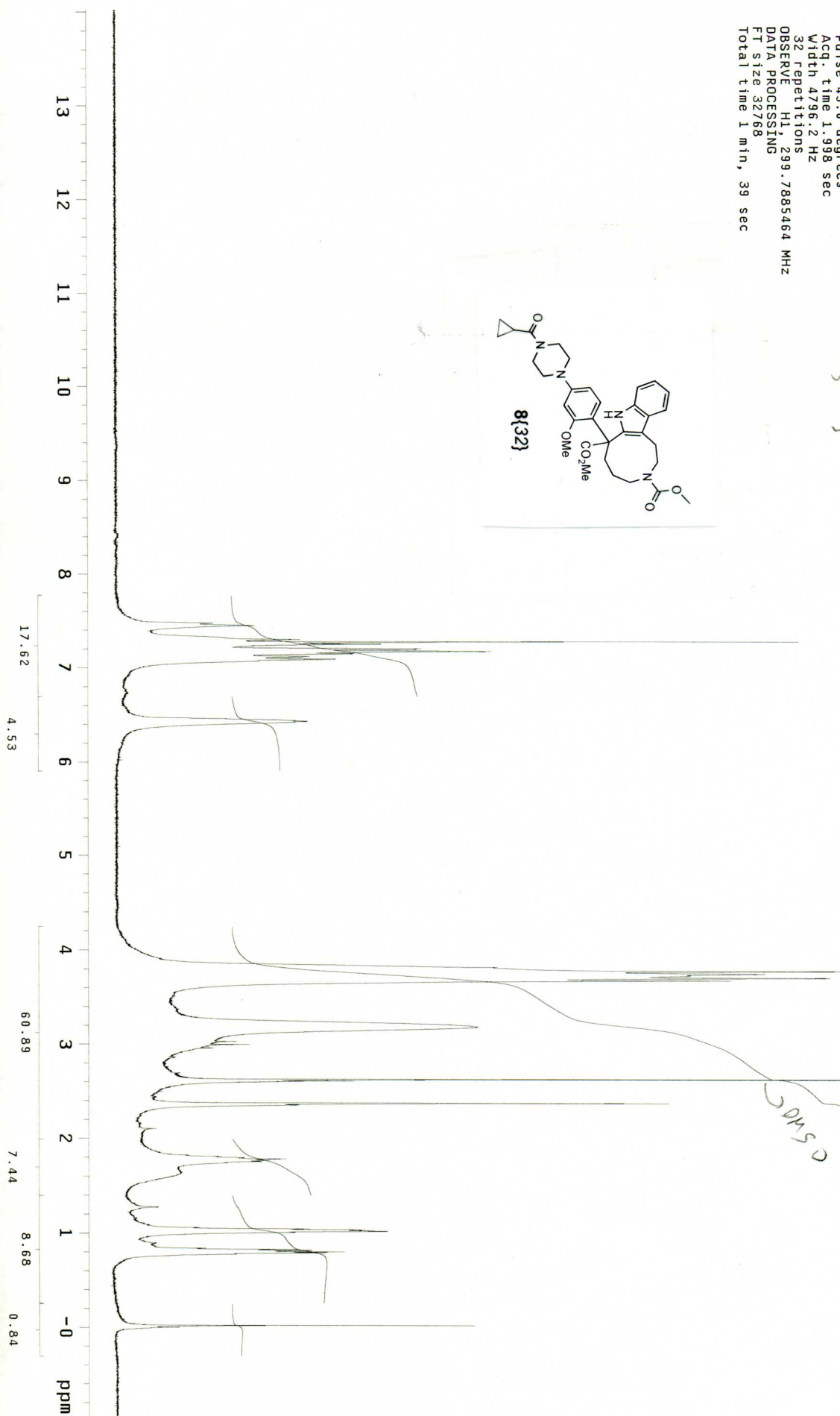
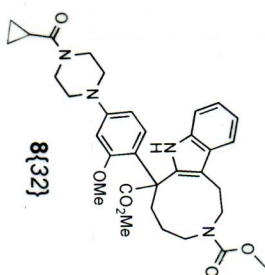
File: 1A201

Mercury-300BB "horror"

3839-245

8{32}

Relax. delay 1.000 sec
Pulse 45.0 degrees
Acq. time 1.998 sec
Width 4796.2 Hz
32 repetitions
OBSERVE H1, 299.785464 MHz
DATA PROCESSING
FT size 32768
Total time 1 min, 39 sec



1A3
STANDARD 1H OBSERVE

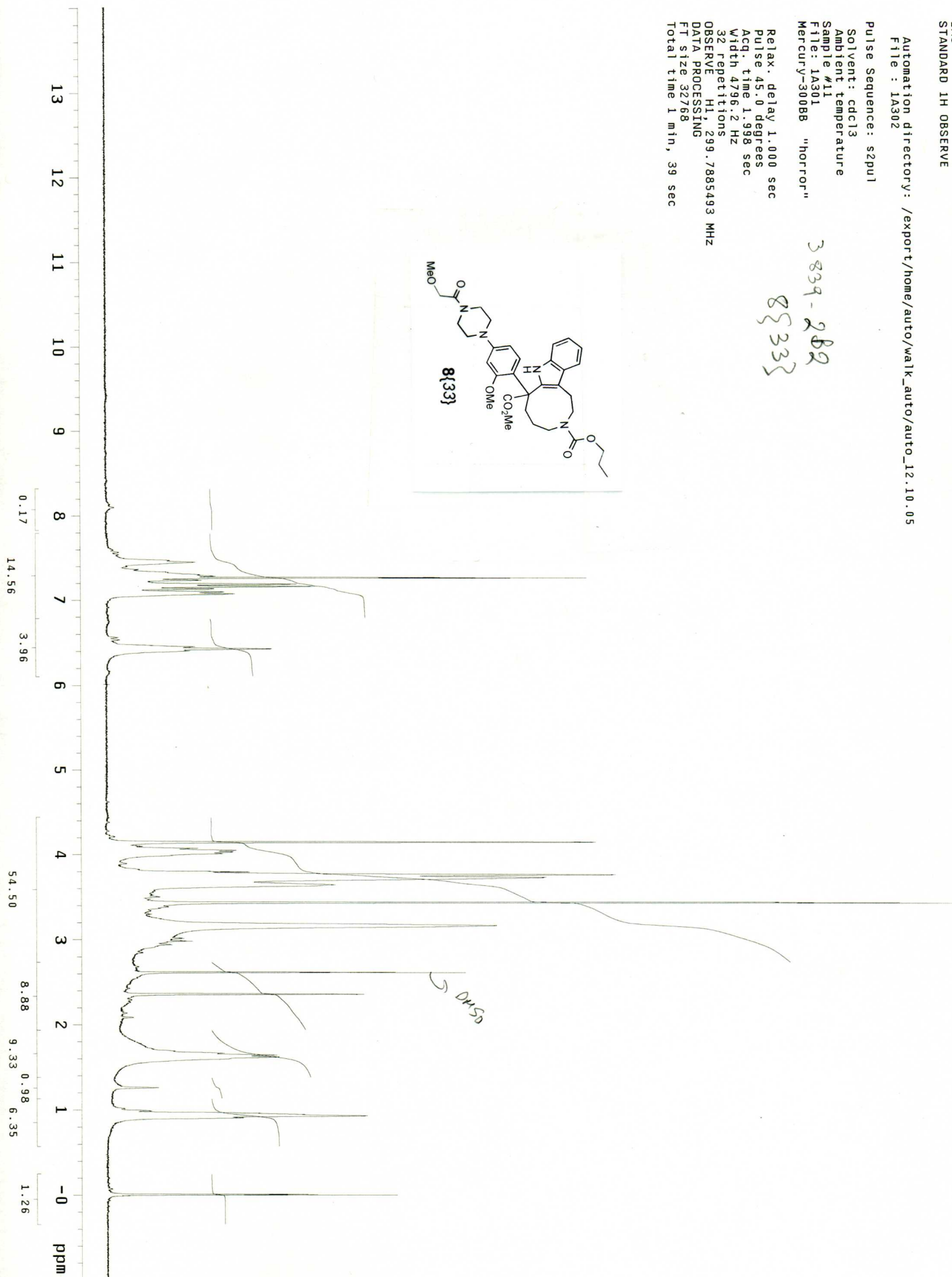
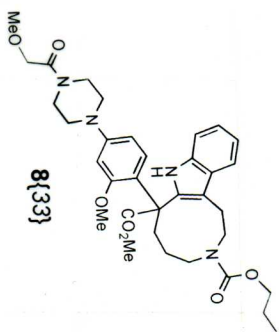
Automation directory: /export/home/auto/walk_auto/auto_12.10.05
File : 1A302

Pulse Sequence: s2pu1

Solvent: cdcl3
Ambient temperature
Sample #11
File: 1A301
Mercury-300BB "horror"

3839-282
8{333}

Relax. delay 1.000 sec
Pulse 45.0 degrees
Acq. time 1.998 sec
Width 4796.2 Hz
32 repetitions
OBSERVE H1, 299.7885493 MHz
DATA PROCESSING
FT size 32768
Total time 1 min, 39 sec



1A4 STANDARD	1H OBSERVE
1	1
2	2
3	3
4	4
5	5
6	6
7	7
8	8
9	9
10	10
11	11
12	12
13	13
14	14
15	15
16	16
17	17
18	18
19	19
20	20
21	21
22	22
23	23
24	24
25	25
26	26
27	27
28	28
29	29
30	30
31	31
32	32
33	33
34	34
35	35
36	36
37	37
38	38
39	39
40	40
41	41
42	42
43	43
44	44
45	45
46	46
47	47
48	48
49	49
50	50
51	51
52	52
53	53
54	54
55	55
56	56
57	57
58	58
59	59
60	60
61	61
62	62
63	63
64	64
65	65
66	66
67	67
68	68
69	69
70	70
71	71
72	72
73	73
74	74
75	75
76	76
77	77
78	78
79	79
80	80
81	81
82	82
83	83
84	84
85	85
86	86
87	87
88	88
89	89
90	90
91	91
92	92
93	93
94	94
95	95
96	96
97	97
98	98
99	99
100	100

```
Automation directory: /export/home/auto/walk_auto/auto_12.10.05
File : 1A402
```

Pulse Sequence: s2pu1

Solvent: cdcl3
Ambient temperature

Sample #12

Mercury-300BB "horror"

Relax. delay 1.000 sec
Pulse 45.0 degrees
Aer + time 1.000 sec

Acq. time 1.998 sec

32 repetitions

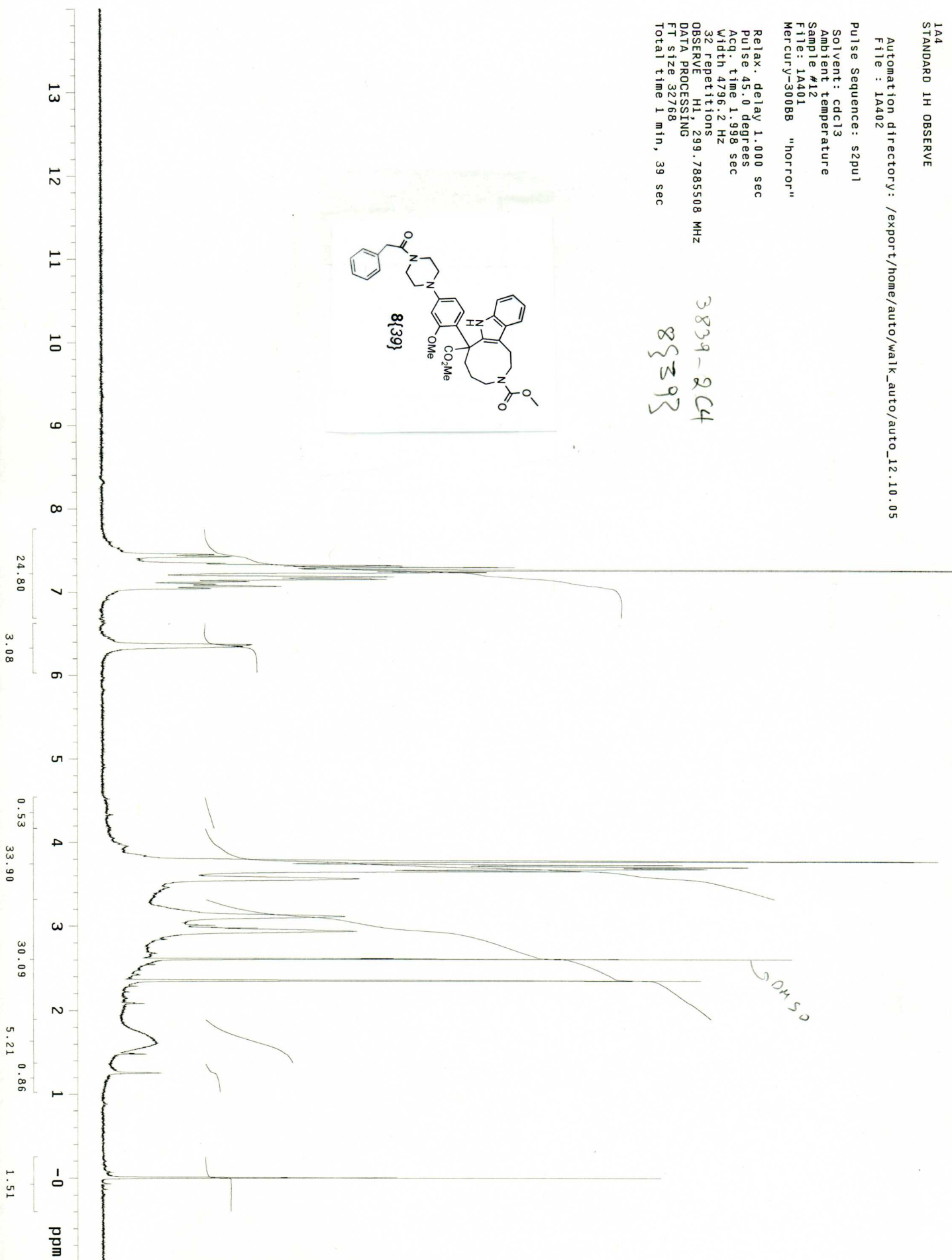
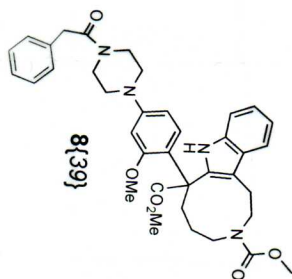
OBSERVE H1, 299.7885508 MHZ

DATA PROCESSING
FT size 32768

Total time 1 min, 39 sec

3839-2C4

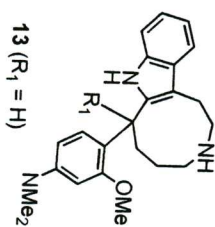
8
W
W



17-75-A-NH-Proton

Archive directory: /export/home/auto/vnmr/sys/data
Sample directory: auto_18Nov2005

Pulse Sequence: s2pul



3.77 5.36
6.11 6.79

3.68

10.18

48.14

4.27

4.24

7.46

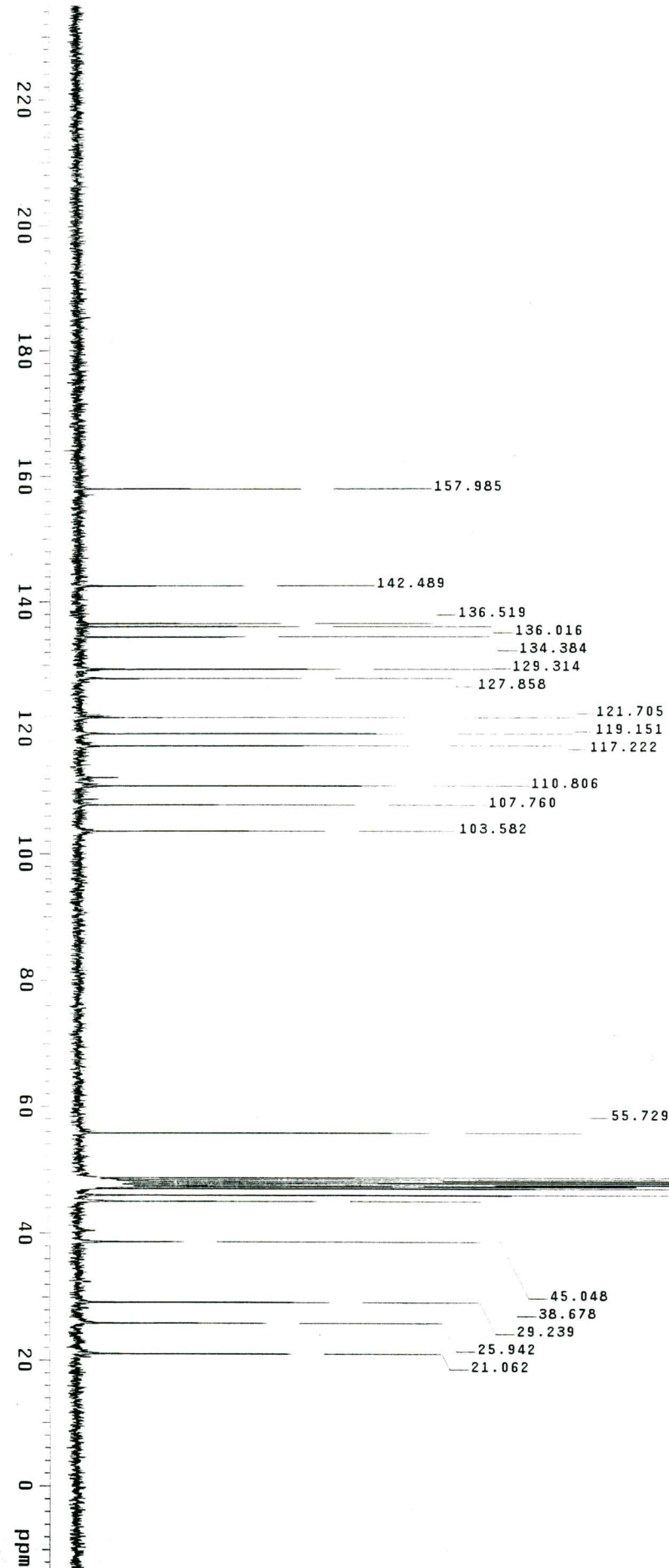
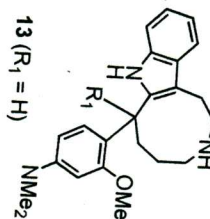
9 8 7 6 5 4 3 2 1 0 ppm

17-75-A-NH-Carbon
13C OBSERVE

pad=2 run with findz0 before acquisition

Archive directory: /export/home/auto/vnmr/sys/data
Sample directory: auto_19Nov2005

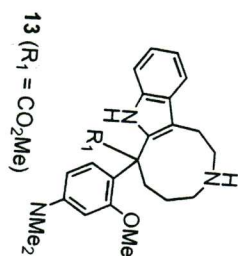
Pulse Sequence: s2pu1



17-75-B-NH-Proton

Archive directory: /export/home/auto/vnmr/sys/data
Sample directory: auto_18Nov2005

Pulse Sequence: s2pu1



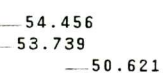
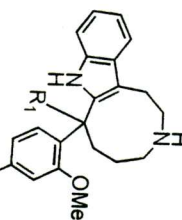
0.79
2.52 1.82
1.00 0.53

2.77
2.74
14.38
1.04

1.80

Archive directory: /export/home/auto/vnmrsys/data
Sample directory: auto_19Nov2005

13 ($R_1 = \text{CO}_2\text{Me}$)



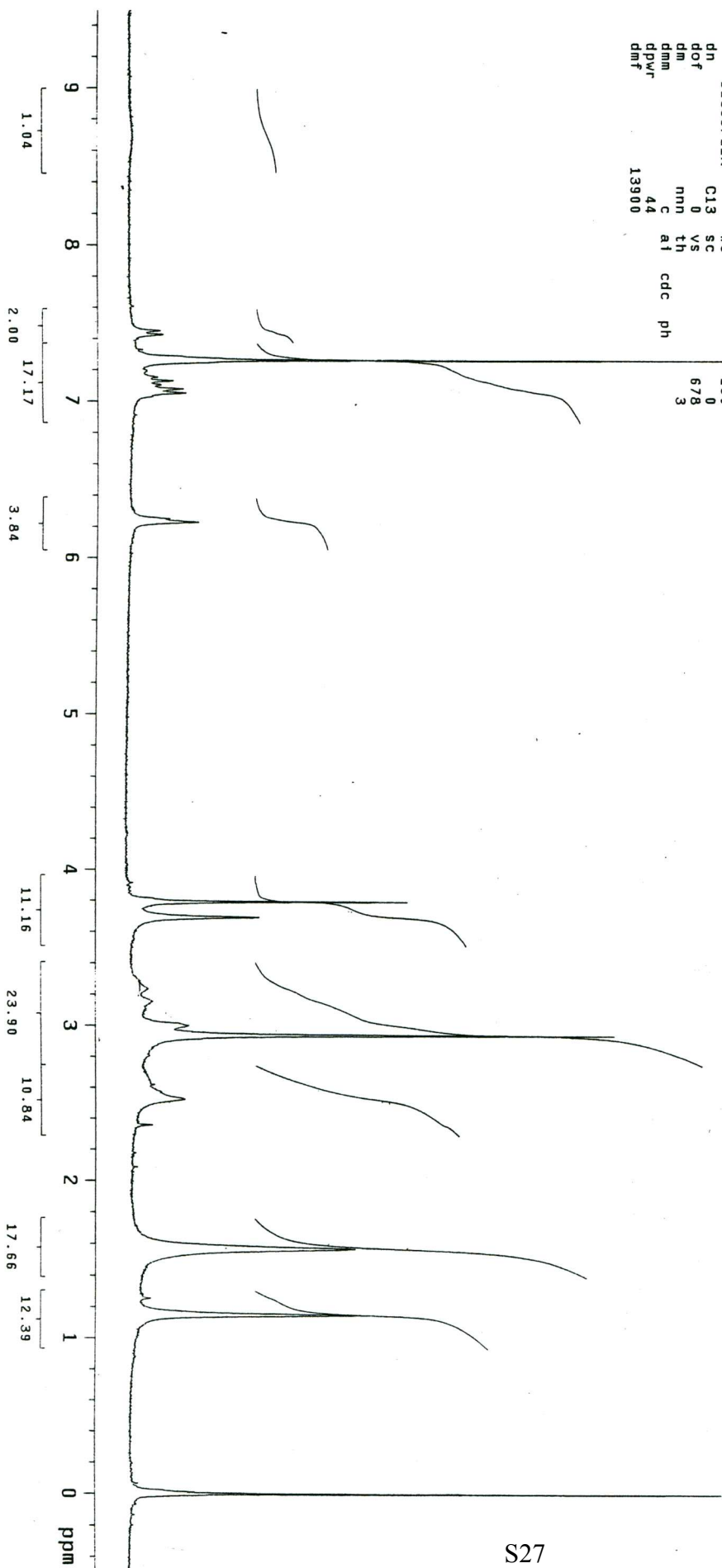
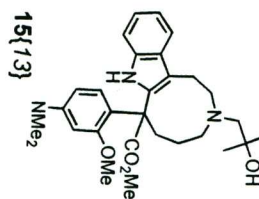
—47.236
—46.950
—46.668
—46.382
~~—46.096~~
—45.814
—45.528
—44.381

exp1 s2pu1

SAMPLE		SPECIAL	
date	Oct 13 2005	temp	not used
solvent	cdcl3	gain	not used
file	/export/home/~	spin	20
auto/walk	auto/aut~	hst	0.008
0_13.10.05/Vin02.f~	pw90		17.250
			20.000

ACQUISITION		FLAGS	
sw	4796.2	f1	n
at	1.998	in	y
np	1916	dp	
fb	2600	hs	nm
bs	16		
d1	1.000	fn	
nt	128	not used	
ct	128	DISPLAY	
tn	TRANSMITTER	WD	-150.2
stfq	H1	rfl	2997.8
tof	299.790	rtp	590.8
tpwr	315.5	tp	126.0
pw	8.625	ip	-61.7

DECOUPLER		PLOT	
dn	C13	WC	250
dof	0	VS	0
dm	nnn	th	678
dmm	c	at	3
dpwr	44	cdc	
dmf	13900	ph	



Vin 1B1 re-run
STANDARD 1H OBSERVE

Automation directory: /export/home/auto/walk_auto/auto_13.10.05
File : Vin04

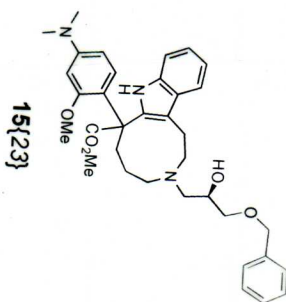
Pulse Sequence: s2pu1

Solvent: cdc13
Ambient temperature
Sample #15
File: Vin03
Mercury-300BB "horror"

Relax. delay 1.000 sec
Pulse 45.0 degrees
Acq. time 1.998 sec

Width 4796.2 Hz
128 repetitions
OBSERVE H1, 299.785508 MHz
DATA PROCESSING
FT size 32768
Total time 6 min, 38 sec

3912-104
15{23}



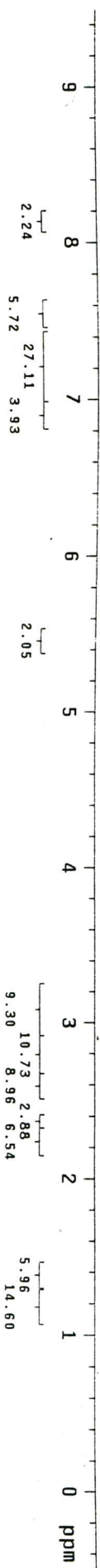
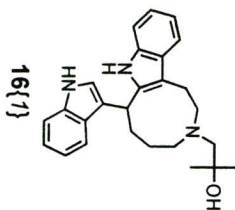
expt s2pu1

SAMPLE		SPECIAL	
date	Oct 12 2005	temp	not used
solvent	cdcl3	gain	not used
file	/export/home/~	spin	20
auto/walk	auto/aut~	hst	0.008
id	0_12.10.05/1C501.f~	pw90	17.250
		alpha	20.000

ACQUISITION		PROCESSING	
sw	4796.2	fn	not used
at	1.998	dp	nm
np	19166	hs	nm
fb	2600		
bs	16		
d1	1.000		
nt	32		
ct	32		

TRANSMITTER		DISPLAY	
tn	H1	rf1	-150.2
strq	299.790	rfp	2997.8
tof	315.5	fp	597.8
tpwr	60	tp	128.18
	8.625		158.18

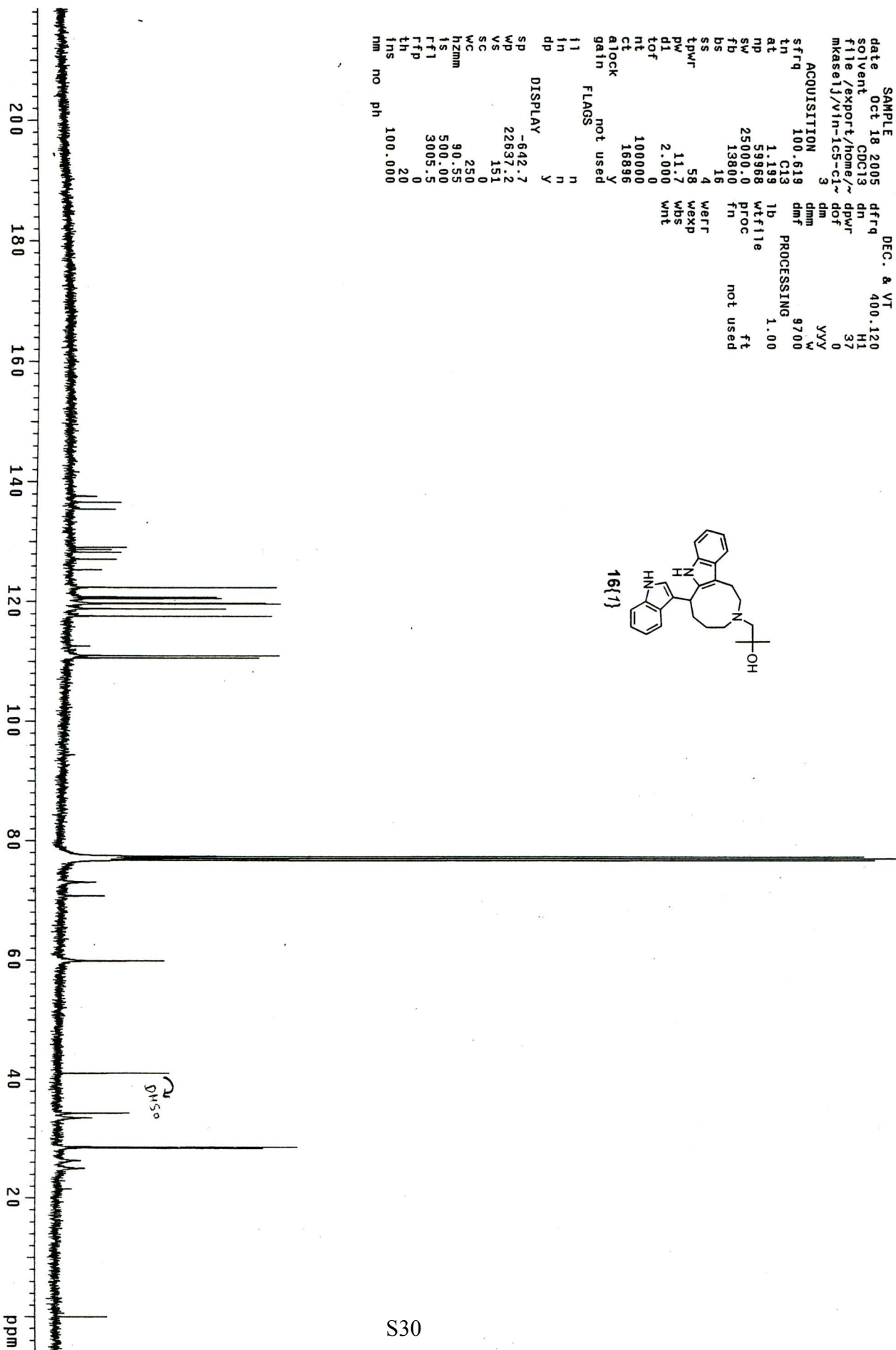
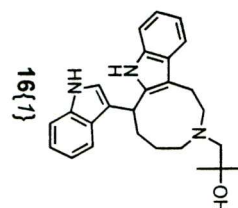
DECOUPLER		PLOT	
dn	C13	wc	250
dof	0	sc	0
dm	nmn	th	781
dmm	0		2
dpwr	44		
dmf	13900		



Vin-1C5-C13

expl std13c

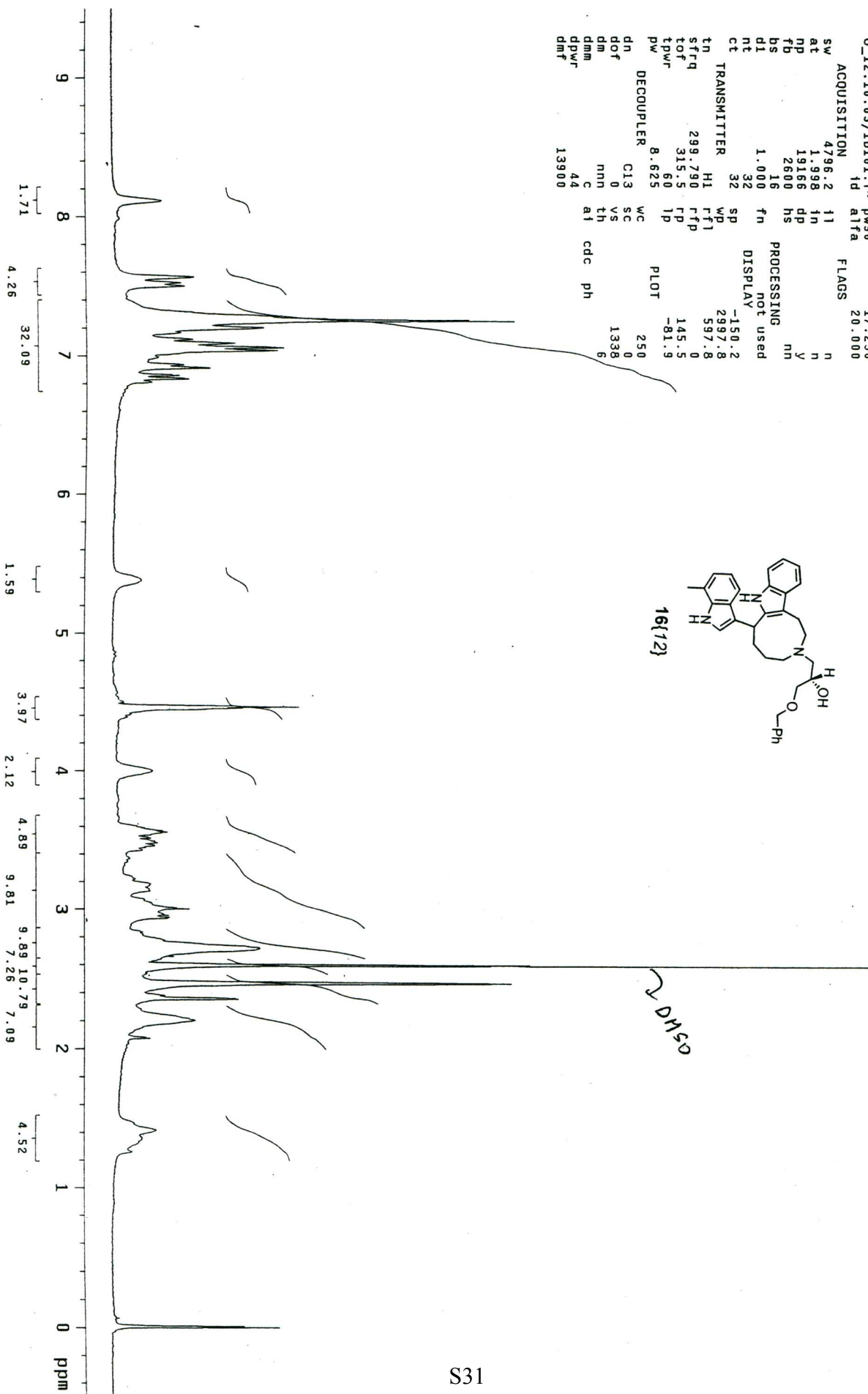
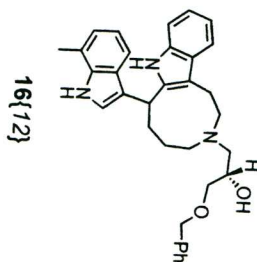
SAMPLE		DEC. & VT	
date	Oct 18 2005	dfrq	400.120
solvent	CDC13	dn	H1
file	/export/home/~	dpwr	37
mkaselj	Vin-1C5-C13	dof	0
ACQUISITION		dm	yyy
sfrq	100.619	dmm	w
in	C13	dmf	9700
at	1.199	lb	1.00
np	59968	wtfile	
sw	25000.0	proc	ft
fb	13800	fn	not used
bs	16		
ss	4	werf	
tpwr	58	wexp	
dl	11.7	wbs	
tof	2.000	wnt	
nt	100000		
ct	16856		
atlock	Y		
gain	not used		
FLAGS			
fl	n		
in	n		
dp	Y		
DISPLAY			
sp	-642.7		
wp	22637.2		
vs	151		
sc	0		
wc	250		
h2nm	90.55		
is	500.00		
rfl	3005.5		
rfp	0		
th	20		
ins	100.000		
nm	no		



exp1 s2pu1

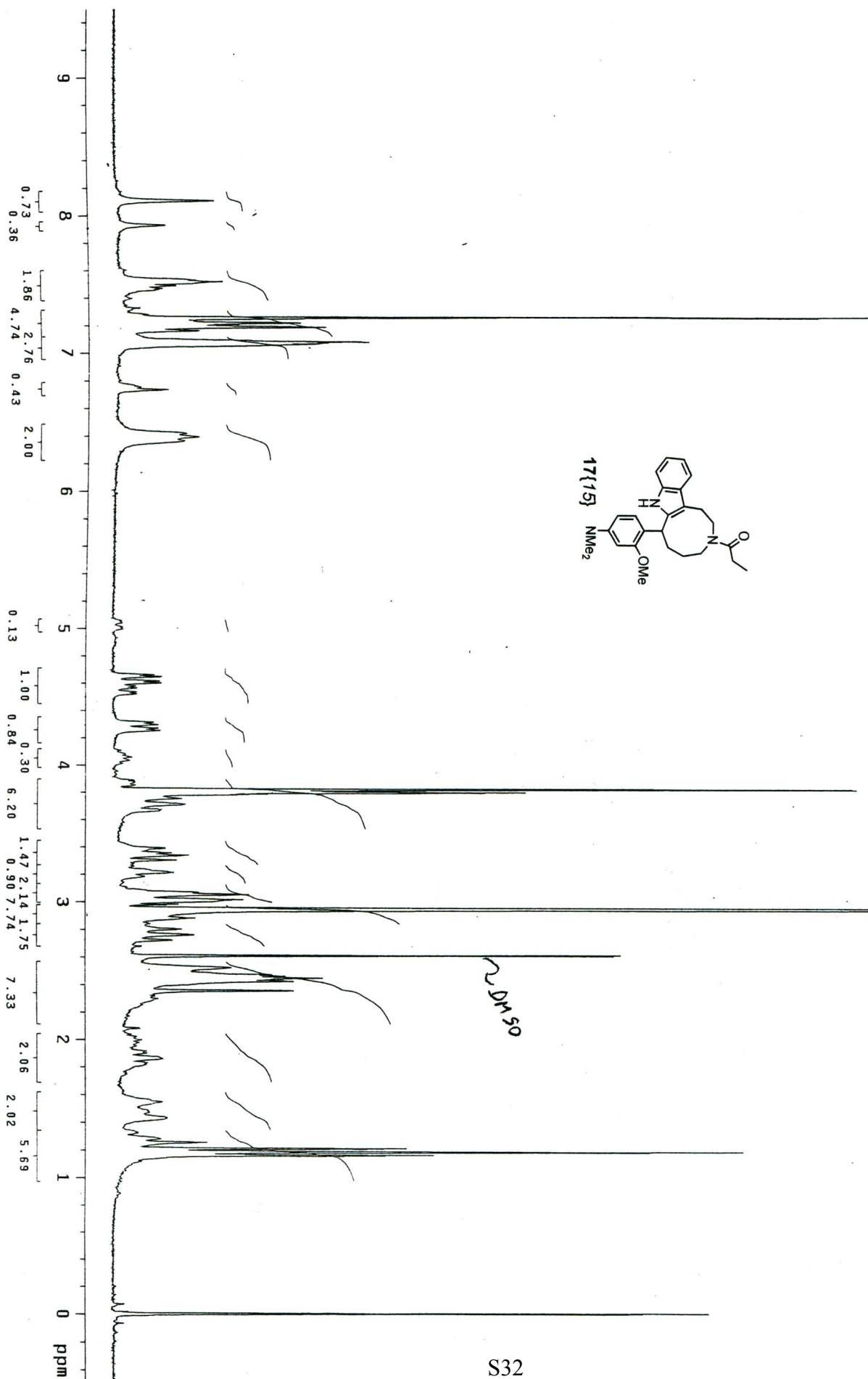
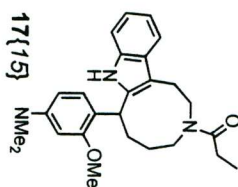
SAMPLE SPECIAL
date Oct 12 2005 temp not used
solvent cdcl3 gain not used
file /export/home/~ spin 20
auto/walk-auto/aut~ hst 0.008
O_12.10.05/10101.f~ pw90 17.250
id alfa 20.000

ACQUISITION
sw 4796.2 11 n
at 1.938 1n n
np 19166 dp y
fb 2600 hs nn
bs 16
dl 1.000 fn not used
nt 32
ct 32 sp
TRANSMITTER
tn 32 wp -150.2
sfreq 299.790 rf1 2997.8
tof 315.5 rfp 597.8
tpwr 60 lp 145.5
pw 8.625 PLOT -81.9
DECOUPLER wc 250
dn C13 SC 0
dof 0 VS 1338
dm nnn th 6
dmm c at cdc ph
dpwr 44
dmf 13900



1D3
STANDARD 1H OBSERVE

Archive directory: /export/home/auto/vnmr/sys/data
Sample directory: auto_16Nov2005
Pulse Sequence: s2pu1



exp1 s2pu1

SAMPLE

date Oct 12 2005 temp not used
solvent cdc13 gain 20
title /export/home/~ spin 0.008
auto/walk-auto/aut~ hst 17.250
o_12.10.05/1C201.f~ pw90 20.000

SPECIAL

ACQUISITION

FLAGS

sw 4796.2 f1 n
at 1.998 f2 y
np 1916 dp n
fb 2600 hs n
bs 16
d1 1.000 fn not used
nt 32
ct 32

PROCESSING

not used

DISPLAY

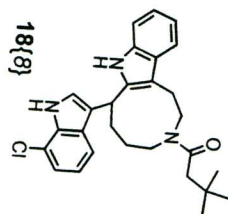
-150.2
2997.8
597.5

TRANSMITTER

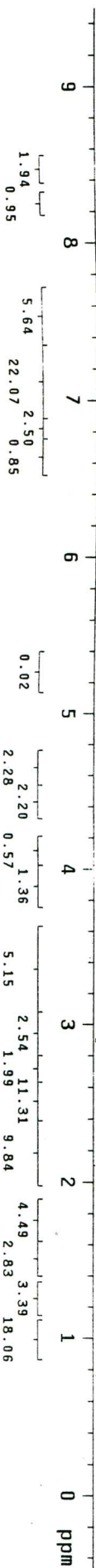
h1 rft
sfrq 299.790 rfp
tof 315.5 rp
tdwr 60 lp
pw 8.625

DECOUPLER

wc 250
dn c13 sc 789
dot 0 vs 2
dm nnn th
dmm c at cdc ph
dpwr 44
dmf 13900



20050



2B1
STANDARD 1H OBSERVE

Automation directory: /export/home/auto/walk_auto/auto_11.10.05
File : 2B102

Pulse Sequence: s2pul

Solvent: cdc13

Ambient temperature

Sample #14

File: 2B101

Mercury-300BB "horror"

Relax. delay 1.000 sec

Pulse 45.0 degrees

Acq. time 1.998 sec

Width 4796.2 Hz

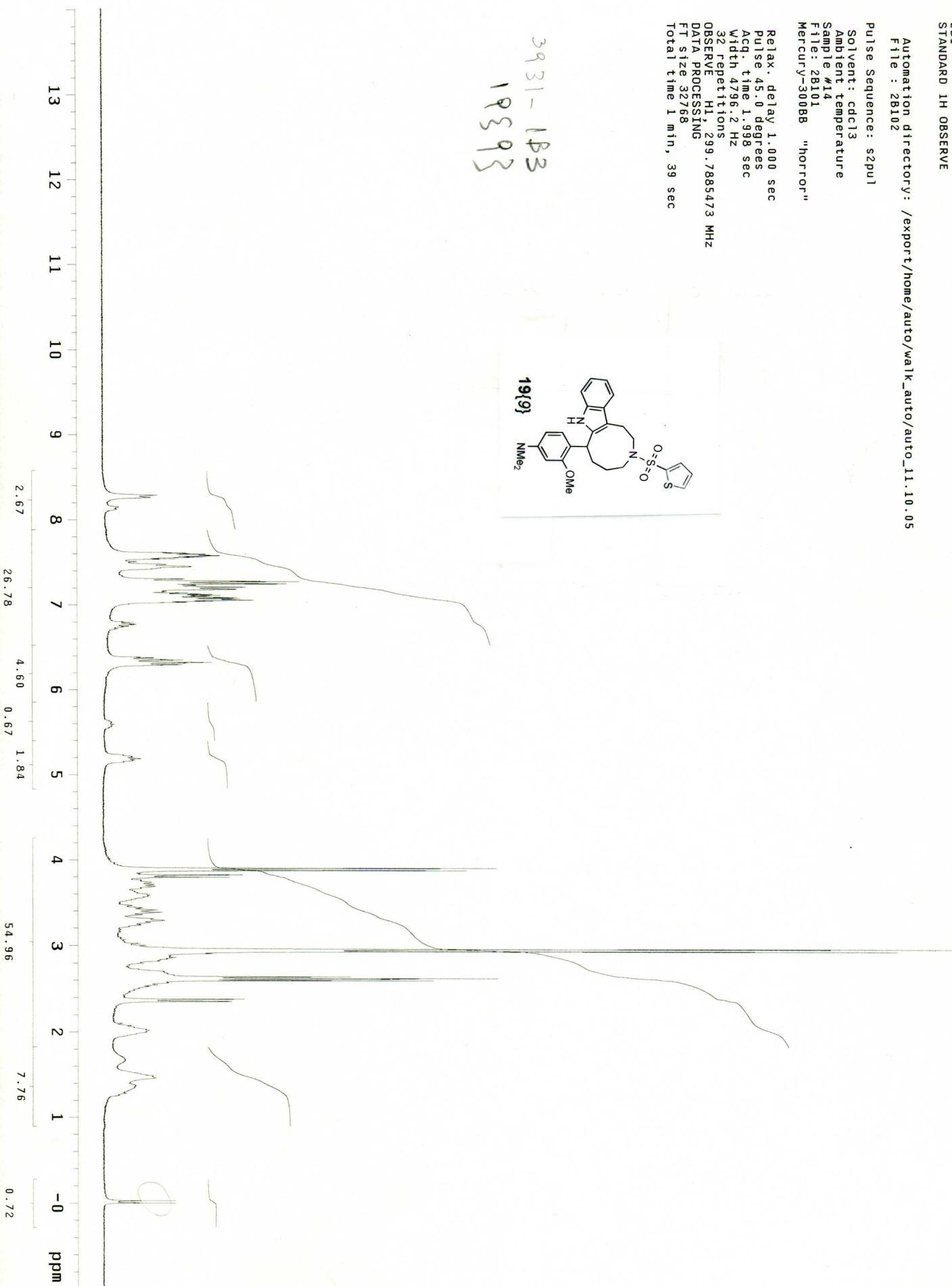
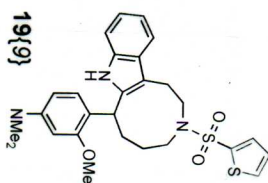
32 repetitions

OBSERVE H1 299.785473 MHz

DATA PROCESSING

FT size 32768

Total time 1 min, 39 sec



2B2
STANDARD 1H OBSERVE

Automation directory: /export/home/auto/walk_auto/auto_11.10.05
File : 2B202

Pulse Sequence: s2pul

Solvent: cdc13
Ambient temperature

Sample #15

File: 2B201
Mercury-300BB "horror"

Relax. delay 1.000 sec
Pulse 45.0 degrees

Acq. time 1.998 sec

Width 4796.2 Hz

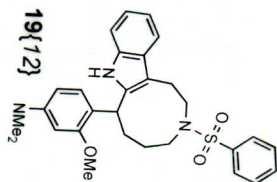
32 repetitions

OBSERVE H1 299.785529 MHz

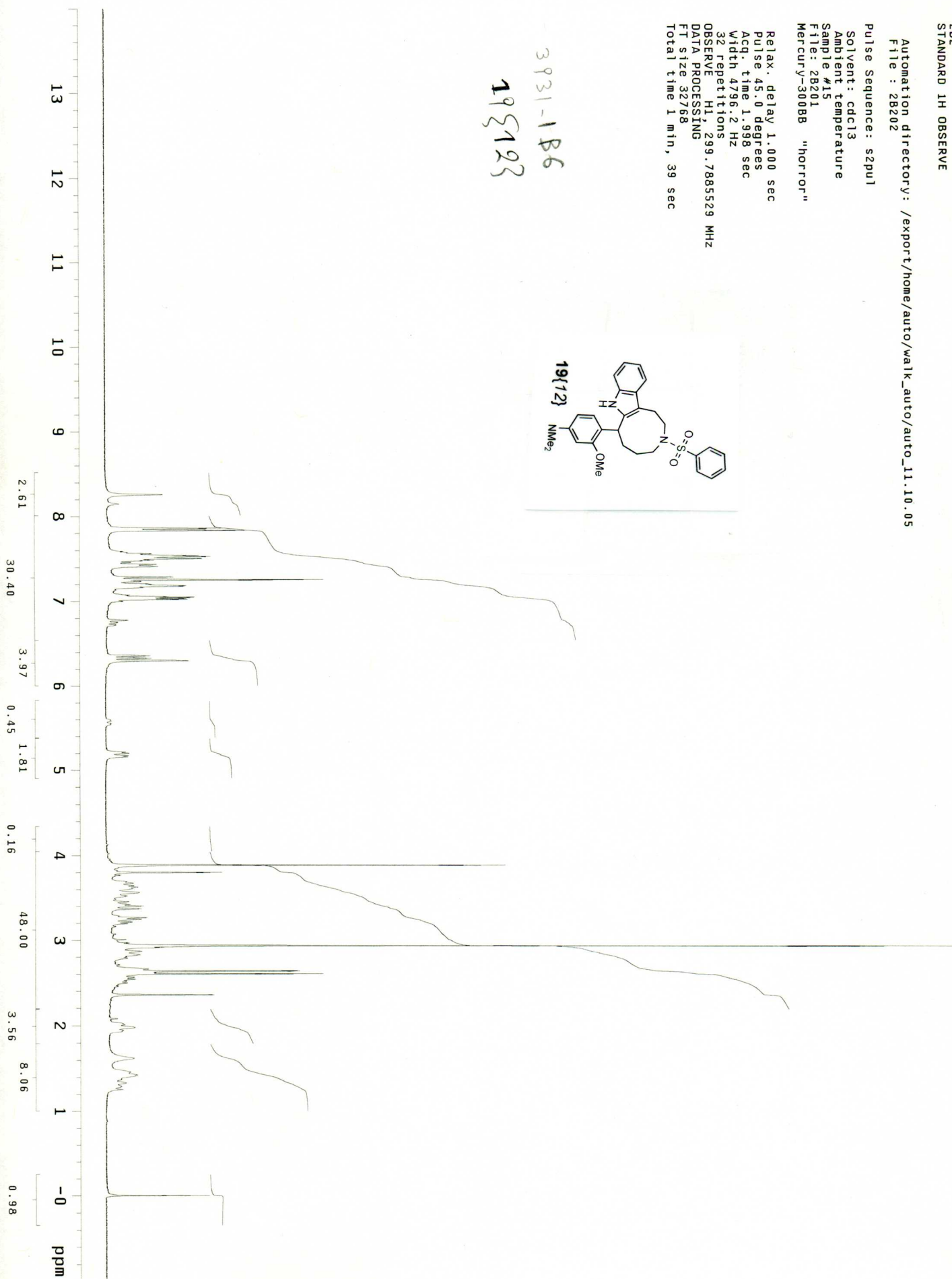
DATA PROCESSING

FT size 32768

Total time 1 min, 39 sec



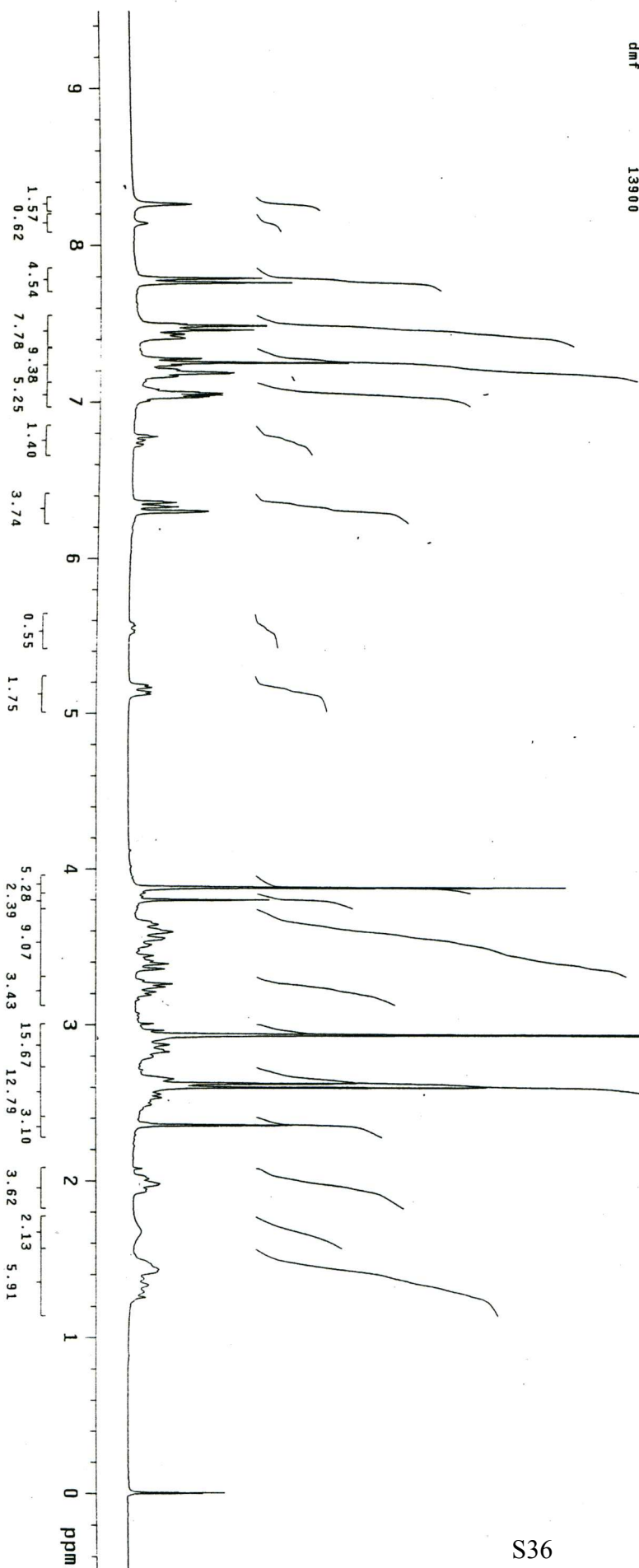
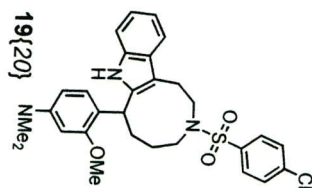
3931-1B6
195123



expl s2pu1

SAMPLE		SPECIAL	
date	Oct 11 2005	temp	not used
solvent	cdcl3	gain	not used
file	/export/home/~	spn	20
auto/walk	auto/aut~	hst	0.008
o_11.10.05/28301.f~	pw90	17.250	
	id	alpha	20.000

ACQUISITION		PROCESSING	
sw	4796.2	fn	not used
at	1.998	in	n
np	19166	dp	y
fb	2600	hs	nn
bs	16		
dl	1.000	fn	not used
nt	32	sp	-150.2
ct	32	wp	2997.8
tn	299.790	rf1	599.6
sfreq	315.5	rffp	143.5
tof	60	ip	-77.8
tpwr	8.625		
pw		PLOT	250
DECOUPLER	C13	WC	0
dn	0	VS	635
dof	nnn	th	6
dm	c	at	cdc ph
dmm	44		
dpwr	13900		
dmt			



Tables describing the synthesis layout of chemsets 2-4, 7-10, 15-20^a

Table 1. Array layout of chemset 2

Table 2. Array layout of chemset 3

Table 3. Array layout of chemset 4

Table 4. Array layout of chemset 7

Table 5. Array layout of chemset 8

Table 6. Array layout of chemset 9

Table 7. Array layout of chemset 10

Table 8. Array layout of chemset 15

Table 9. Array layout of chemset 16

Table 10. Array layout of chemset 17

Table 11. Array layout of chemset 18

Table 12. Array layout of chemset 19

Table 13. Array layout of chemset 20

^aNumbers in parentheses represent the isolated yield (% yield or μmol quantity), the % purity by $\text{UV}_{214\text{nm}}$ and ELSD (evaporative light scattering detector), respectively, of each individual compound prepared on 100 μmol scale and recovered after mass-triggered high throughput reverse phase HPLC purification.

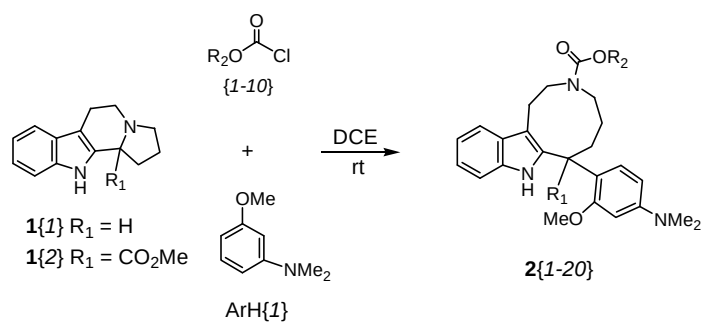
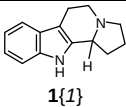
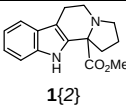
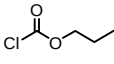
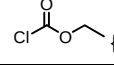
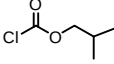
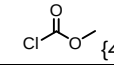
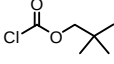
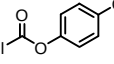
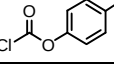
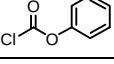
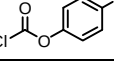
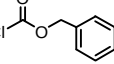


Table 1. Array layout of chemset $\text{2}\{1-20\}$

Library Scaffolds		 $\text{1}\{1\}$	 $\text{1}\{2\}$
Chloroformates $\{1-10\}$	$\text{ArH}\{1\}$	Product	Product
 $\{1\}$		$\text{2}\{1\}$ (47.5, 89, 100)	$\text{2}\{11\}$ (27.6, 100, 100)
 $\{2\}$		$\text{2}\{2\}$ (49.5, 90, 100)	$\text{2}\{12\}$ Not isolated
 $\{3\}$		$\text{2}\{3\}$ (59.9, 90, 100)	$\text{2}\{13\}$ (42.0, 100, 100)
 $\{4\}$		$\text{2}\{4\}$ (52.4, 89, 100)	$\text{2}\{14\}$ (30.9, 96, 100)
 $\{5\}$		$\text{2}\{5\}$ (58.9, 90, 100)	$\text{2}\{15\}$ (41.3, 100, 100)
 $\{6\}$		$\text{2}\{6\}$ (53, 89, 100)	$\text{2}\{16\}$ (63, 100, 100)
 $\{7\}$		$\text{2}\{7\}$ (55.3, 90, 100)	$\text{2}\{17\}$ (61.2, 100, 100)
 $\{8\}$		$\text{2}\{8\}$ (26.3, 74, 100)	$\text{2}\{18\}$ (64.3, 100, 100)
 $\{9\}$		$\text{2}\{9\}$ (57.5, 88, 100)	$\text{2}\{19\}$ (65, 100, 100)
 $\{10\}$		$\text{2}\{10\}$ (13.9, 60, 100)	$\text{2}\{20\}$ (25.1, 100, 100)

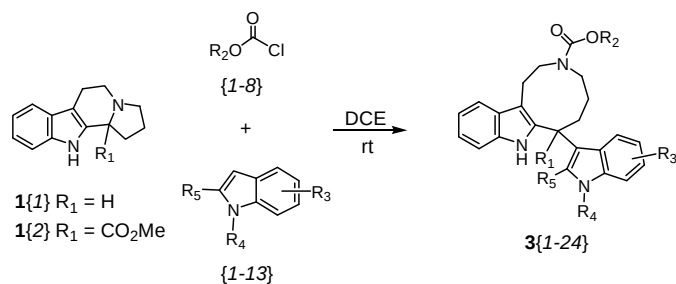
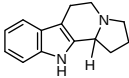
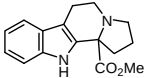
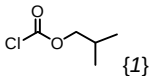
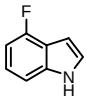
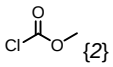
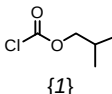
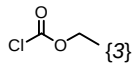
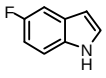
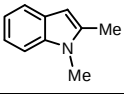
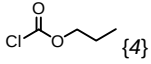
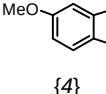
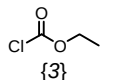
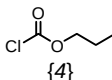
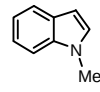
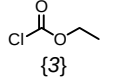
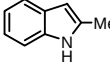
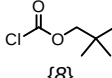
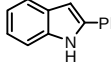
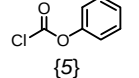
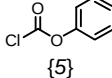
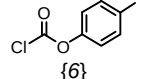
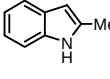
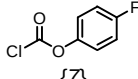
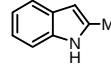
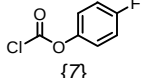
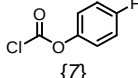
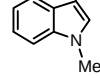
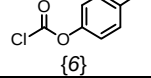
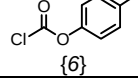
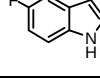
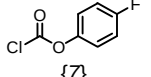
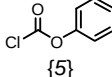
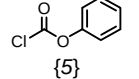
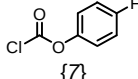
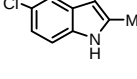


Table 2. Array layout of chemset 3{1-24}

Library Scaffolds		 1{1}	Library Scaffolds		 1{2}
Chloroformates {1-8}	Indoles{1-13}	Product	Chloroformates {1-8}	Indoles{1-13}	Product
 {1}	 {1}	3{1} (42.7, 95, 100)	 {2}	 {9}	3{13} Not isolated
 {2}	 {2}	3{2} Not isolated	 {1}	 {1}	3{14} (12.8, 96, 100)
 {3}	 {3}	3{3} (28.7, 100, 100)	 {3}	 {10}	3{15} (27.2, 100, 100)
 {4}	 {4}	3{4} (29.7, 94, 100)	 {3}	 {3}	3{16} (11.6, 100, 100)
 {3}	 {5}	3{5} (60.6, 100, 100)	 {4}	 {5}	3{17} (16.7, 100, 100)
 {3}	 {6}	3{6} (46.1, 93, 100)	 {8}	 {11}	3{18} (29.0, 100, 100)
 {5}	 {1}	3{7} (41.9, 100, 100)	 {5}	 {1}	3{19} Not isolated
 {6}	 {6}	3{8} Not isolated	 {7}	 {6}	3{20} Not isolated
 {7}	 {5}	3{9} (60.5, 100, 100)	 {7}	 {5}	3{21} (27.4, 100, 100)
 {6}	 {3}	3{10} (63.8, 100, 100)	 {6}	 {3}	3{22} Not isolated
 {7}	 {7}	3{11} (58.7, 100, 100)	 {5}	 {12}	3{23} (14.6, 85, 100)
 {5}	 {8}	3{12} (46, 85, 100)	 {7}	 {13}	3{24} Not isolated

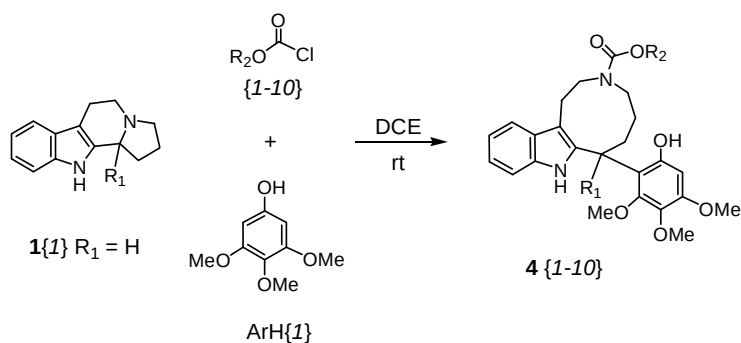


Table 3. Array layout of chemset **4**{1-10}

Library Scaffolds		 1 {1}
Chloroformates {1-10}	ArH{1}	Product
 1	3,4,5-trimethoxyphenol	4 {1} (28.0, 100, 100)
 2	3,4,5-trimethoxyphenol	4 {2} (33.8, 96, 100)
 3	3,4,5-trimethoxyphenol	4 {3} Not isolated
 4	3,4,5-trimethoxyphenol	4 {4} (43.7, 100, 100)
 5	3,4,5-trimethoxyphenol	4 {5} (10.2, 71, 76)
 6	3,4,5-trimethoxyphenol	4 {6} Not isolated
 7	3,4,5-trimethoxyphenol	4 {7} (25.1, 100, 100)
 8	3,4,5-trimethoxyphenol	4 {8} (43.4, 100, 100)
 9	3,4,5-trimethoxyphenol	4 {9} (47.3, 100, 100)
 10	3,4,5-trimethoxyphenol	4 {10} Not isolated

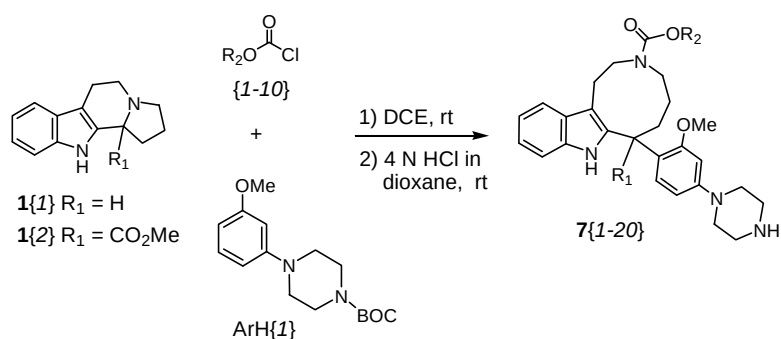


Table 4. Array layout of chemset 7{1-20}

Library Scaffolds			
Chloroformates {1-10}	ArH{1}	Product	Product
{1}		7{1} (27.2, 96, 100)	7{11} (28.8, 100, 100)
{2}		7{2} (26.1, 100, 100)	7{12} (32.3, 100, 100)
{3}		7{3} (28.1, 93, 100)	7{13} (30.5, 94, 100)
{4}		7{4} (28.8, 96, 100)	7{14} (34.2, 97, 100)
{5}		7{5} (23.1, 88, 94)	7{15} (27.7, 77, 88)
{6}		7{6} (18.0, 82, 92)	7{16} (32.3, 73, 87)
{7}		7{7} (24.5, 82, 90)	7{17} (33.5, 69, 81)
{8}		7{8} Not isolated	7{18} (33.4, 79, 93)
{9}		7{9} (25.2, 81, 92)	7{19} (37.2, 78, 91)
{10}		7{10} (25.6, 83, 97)	7{20} (29.1, 91, 100)

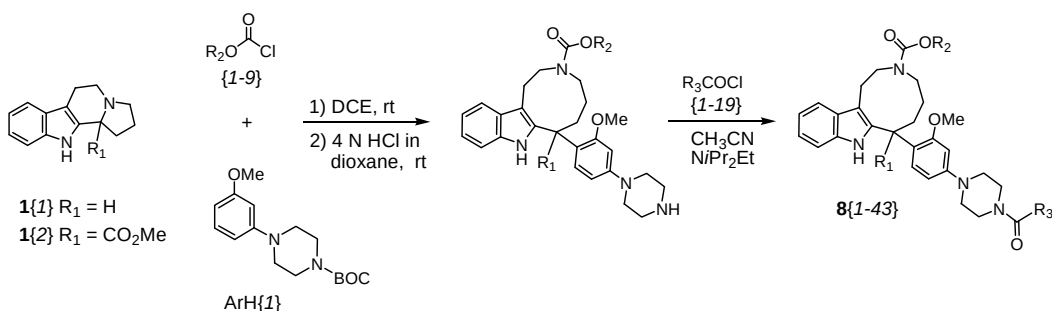
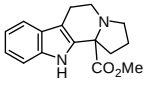
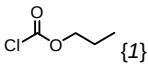
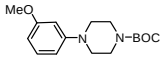
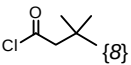
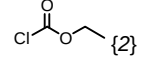
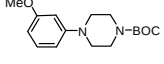
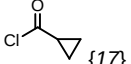
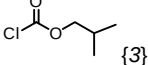
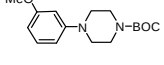
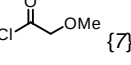
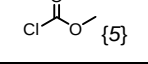
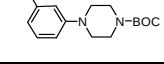
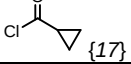
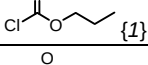
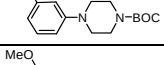
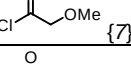
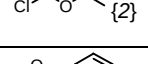
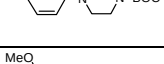
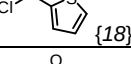
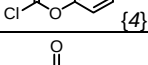
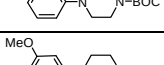
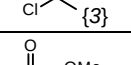
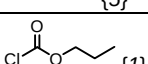
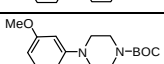
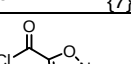
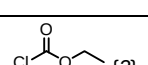
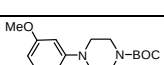
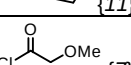
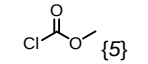
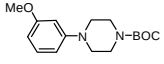
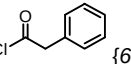
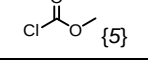
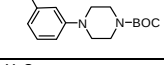
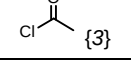
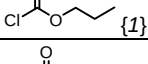
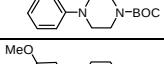
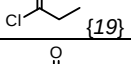
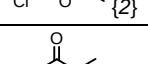
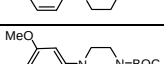
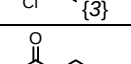
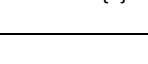
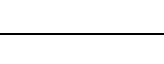
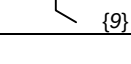


Table 5. Array layout of chemset **8{1-43}**

Library Scaffolds				Library Scaffolds			
Chloroformates {1-9}	ArH{1}	Acid Chlorides {1-19}	Product	Chloroformates {1-9}	ArH{1}	Acid Chlorides {1-19}	Product
{1}	{1}	{1}	8{1} (22.2, 81, 95)	{3}	{1}	{11}	8{15} Not isolated
{2}	{1}	{2}	8{2} (30.7, 100, 100)	{6}	{1}	{11}	8{16} Not isolated
{2}	{1}	{3}	8{3} (33.2, 78, 100)	{7}	{1}	{12}	8{17} Not isolated
{3}	{1}	{4}	8{4} (6.0, 100, 100)	{5}	{1}	{13}	8{18} (26.9, 100, 100)
{4}	{1}	{5}	8{5} (18.8, 100, 93)	{2}	{1}	{6}	8{19} (9.0, 66, 78)
{5}	{1}	{6}	8{6} (20.1, 100, 100)	{2}	{1}	{14}	8{20} (15.8, 100, 100)
{1}	{1}	{3}	8{7} (52.9, 84, 100)	{3}	{1}	{12}	8{21} (14.9, 100, 69)
{2}	{1}	{7}	8{8} (27.1, 80, 92)	{4}	{1}	{15}	8{22} (25.7, 100, 100)
{3}	{1}	{8}	8{9} (22.3, 100, 100)	{8}	{1}	{7}	8{23} (34.2, 88, 97)
{6}	{1}	{9}	8{10} (6.0, 100, 100)	{5}	{1}	{7}	8{24} (39.1, 85, 100)
{7}	{1}	{7}	8{11} Not isolated	{5}	{1}	{11}	8{25} Not isolated
{5}	{1}	{10}	8{12} (31.4, 100, 100)	{5}	{1}	{3}	8{26} (40.3, 95, 100)
{1}	{1}	{4}	8{13} (18.9, 72, 79)	{9}	{1}	{7}	8{27} (28.9, 78, 86)
{2}	{1}	{11}	8{14} Not isolated	{9}	{1}	{16}	8{28} (22.7, 100, 100)

Table 5 (contd)

Library Scaffolds			 1{2}
Chloroformates {1-9}	ArH{1}	Acid Chlorides {1-19}	Product
 {1}		 {8}	8{29} (16.4, 100, 100)
 {2}		 {17}	8{30} (3.9, 49, 81)
 {3}		 {7}	8{31} (3.3, 44, 83)
 {5}		 {17}	8{32} (36.8, 100, 100)
 {1}		 {7}	8{33} (26.8, 100, 100)
 {2}		 {18}	8{34} (3.7, 38, 40)
 {4}		 {3}	8{35} (36.1, 80, 93)
 {5}		 {7}	8{36} (47.9, 58, 86)
 {1}		 {11}	8{37} Not isolated
 {2}		 {7}	8{38} (3.0, 32, 38)
 {5}		 {6}	8{39} (24.5, 100, 100)
 {5}		 {3}	8{40} (52.0, 55, 81)
 {1}		 {19}	8{41} (3.4, 44, 71)
 {2}		 {3}	8{42} (3.7, 61, 87)
 {5}		 {9}	8{43} (20.8, 100, 100)

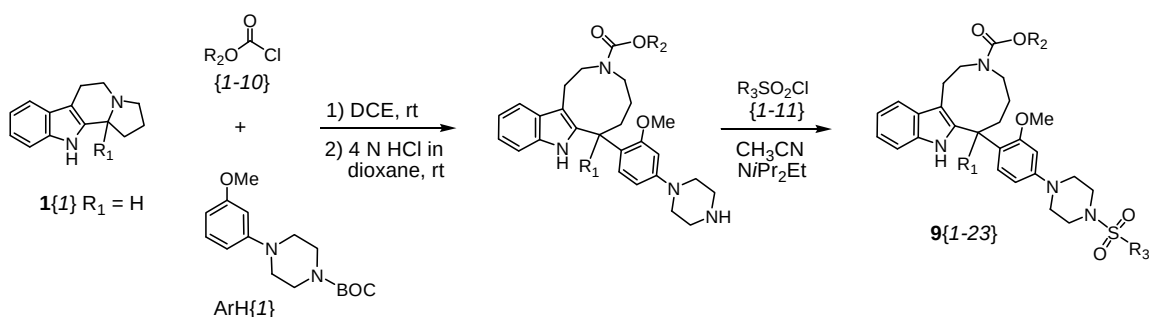
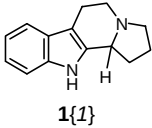
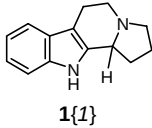
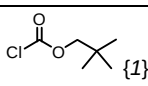
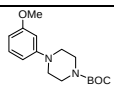
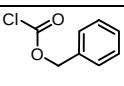
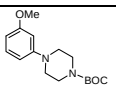
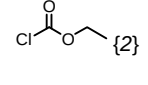
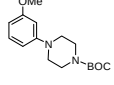
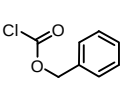
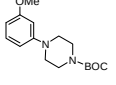
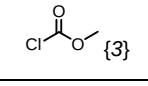
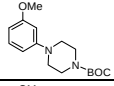
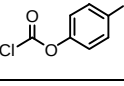
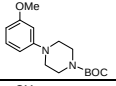
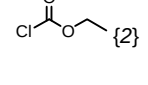
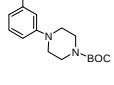
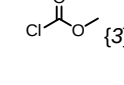
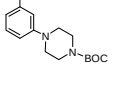
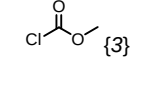
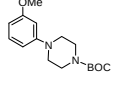
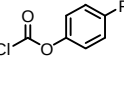
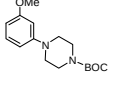
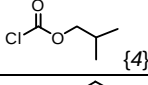
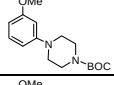
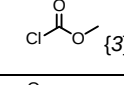
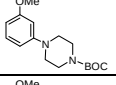
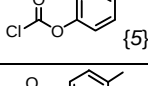
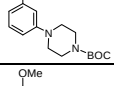
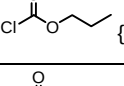
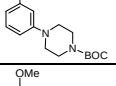
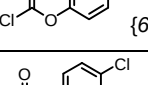
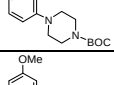
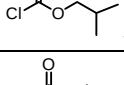
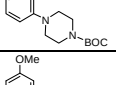
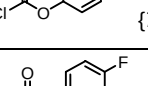
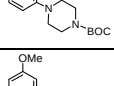
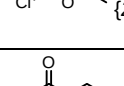
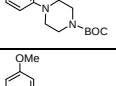
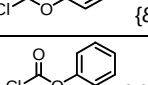
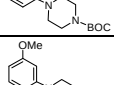
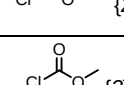
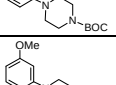
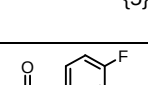
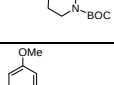
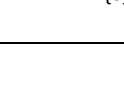
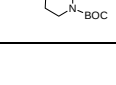
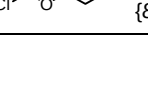
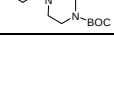


Table 6. Array layout of chemset **9**{1-23}

Library Scaffolds			 1 {1}	Library Scaffolds			 1 {1}
Chloroformates {1-10}	ArH{1}	Sulfonyl Chlorides {1-11}	Product	Chloroformates {1-10}	ArH{1}	Sulfonyl Chlorides {1-11}	Product
 {1}		 {1}	9 {1} Not isolated	 {9}		 {1}	9 {13} (7.7, 69, 86)
 {2}		 {2}	9 {2} (5.9, 100, 100)	 {9}		 {2}	9 {14} (4.5, 95, 100)
 {3}		 {3}	9 {3} (13.8, 97, 100)	 {6}		 {8}	9 {15} (5.5, 100, 100)
 {2}		 {4}	9 {4} (17.3, 100, 100)	 {3}		 {7}	9 {16} (11.6, 90, 100)
 {3}		 {5}	9 {5} (13.7, 100, 100)	 {8}		 {8}	9 {17} (9.3, 100, 100)
 {4}		 {6}	9 {6} (5.2, 100, 100)	 {3}		 {1}	9 {18} (7.5, 68, 91)
 {5}		 {3}	9 {7} (5.3, 80, 100)	 {10}		 {3}	9 {19} (5.9, 100, 100)
 {6}		 {2}	9 {8} (3.9, 77, 100)	 {4}		 {9}	9 {20} (9.7, 94, 100)
 {7}		 {1}	9 {9} (6.2, 65, 87)	 {2}		 {10}	9 {21} (13.3, 100, 100)
 {8}		 {2}	9 {10} (4.8, 74, 86)	 {2}		 {11}	9 {22} (4.7, 28, 12)
 {5}		 {5}	9 {11} (11.6, 88, 100)	 {3}		 {4}	9 {23} (17.3, 91, 100)
 {8}		 {7}	9 {12} (5.4, 80, 95)				

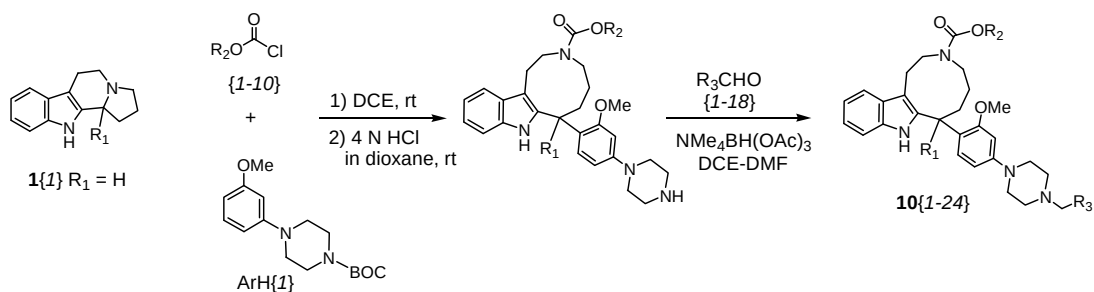
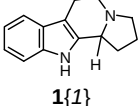
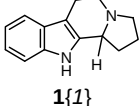
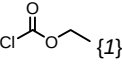
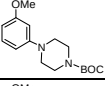
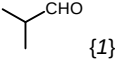
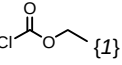
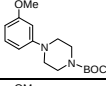
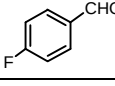
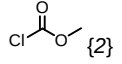
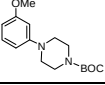
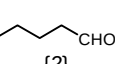
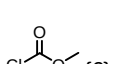
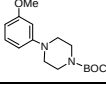
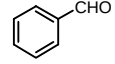
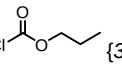
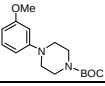
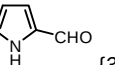
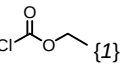
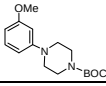
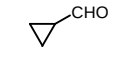
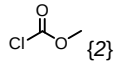
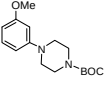
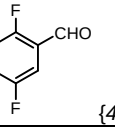
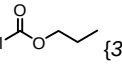
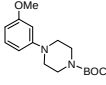
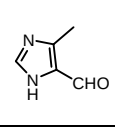
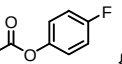
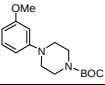
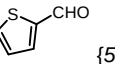
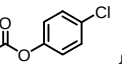
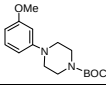
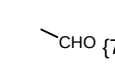
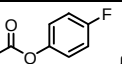
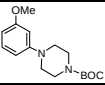
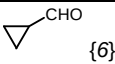
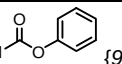
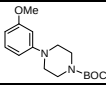
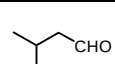
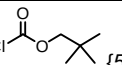
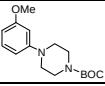
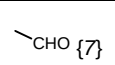
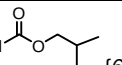
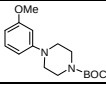
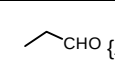
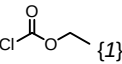
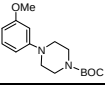
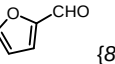
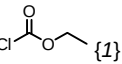
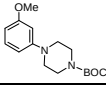
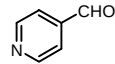
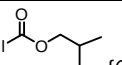
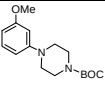
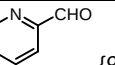
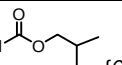
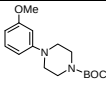
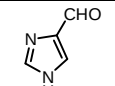
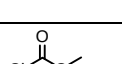
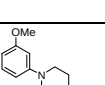
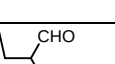
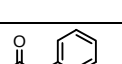
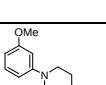
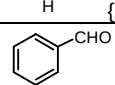
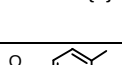
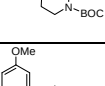
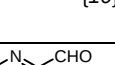
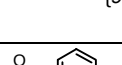
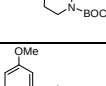
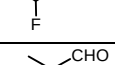
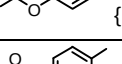
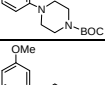
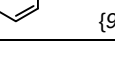
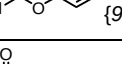
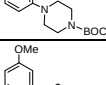
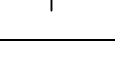


Table 7. Array layout of chemset **10**{1-24}

Library Scaffolds			 1 {1}	Library Scaffolds			 1 {1}
Chloroformates {1-10}	ArH{1}	Aldehydes {1-18}	Product	Chloroformates {1-10}	ArH{1}	Aldehydes {1-18}	Product
 {1}		 {1}	10 {1} (9.1, 100, 100)	 {1}		 {12}	10 {13} (8.9, 100, 100)
 {2}		 {2}	10 {2} (12.8, 100, 100)	 {2}		 {13}	10 {14} (7.1, 96, 100)
 {3}		 {3}	10 {3} (8.4, 100, 100)	 {1}		 {6}	10 {15} (7.4, 92, 100)
 {2}		 {4}	10 {4} (5.9, 74, 100)	 {3}		 {14}	10 {16} (12.4, 100, 100)
 {4}		 {5}	10 {5} Not isolated	 {8}		 {7}	10 {17} (12.2, 100, 100)
 {4}		 {6}	10 {6} (9.2, 100, 100)	 {9}		 {15}	10 {18} (8.9, 94, 100)
 {5}		 {7}	10 {7} (6.1, 100, 100)	 {6}		 {11}	10 {19} (6.4, 100, 100)
 {1}		 {8}	10 {8} (6.6, 95, 100)	 {1}		 {16}	10 {20} Not isolated
 {6}		 {9}	10 {9} (8.0, 96, 100)	 {6}		 {17}	10 {21} Not isolated
 {2}		 {10}	10 {10} (7.7, 100, 100)	 {9}		 {18}	10 {22} Not isolated
 {7}		 {9}	10 {11} (7.8, 100, 100)	 {9}		 {1}	10 {23} (9.4, 92, 100)
 {7}		 {11}	10 {12} (9.6, 100, 100)	 {10}		 {7}	10 {24} (10.4, 96, 100)

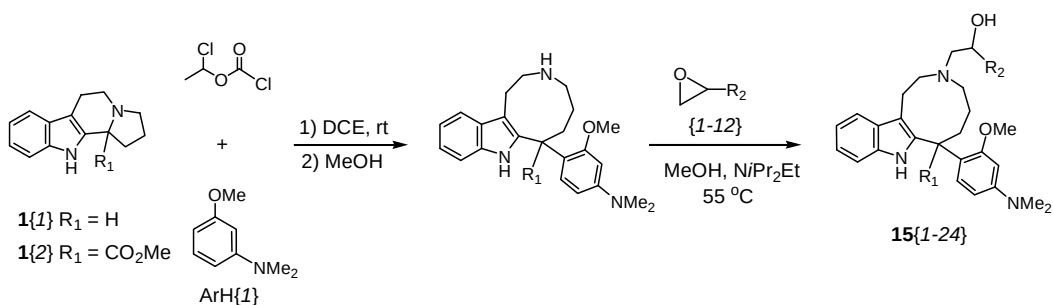


Table 8. Array layout of chemset **15{1-24}**

Library Scaffolds		 1{1}	 1{2}
ArH{1}	Epoxides{1-12}	Product	Product
	{1}	15{1} (31.7, 85, 100)	15{13} (27.1, 100, 100)
	{2}	15{2} (48.7, 88, 100)	15{14} (33.9, 100, 100)
	{3}	15{3} Not isolated	15{15} (5.8, 79, 100)
	{4}	15{4} (52.7, 90, 100)	15{16} Not isolated
	{5}	15{5} (35.4, 94, 100)	15{17} (41.3, 100, 100)
	{6}	15{6} (24.0, 100, 100)	15{18} (21.1, 56, 83)
	{7}	15{7} (24.8, 100, 100)	15{19} (30.3, 76, 100)
	{8}	15{8} Not isolated	15{20} (25.7, 100, 100)
	{9}	15{9} (31.7, 100, 100)	15{21} (28.9, 100, 100)
	{10}	15{10} (45.2, 100, 100)	15{22} (56.5, 88, 100)
	{11}	15{11} (42.7, 92, 100)	15{23} (46.3, 100, 100)
	{12}	15{12} (27.6, 85, 100)	15{24} (44.4, 75, 100)

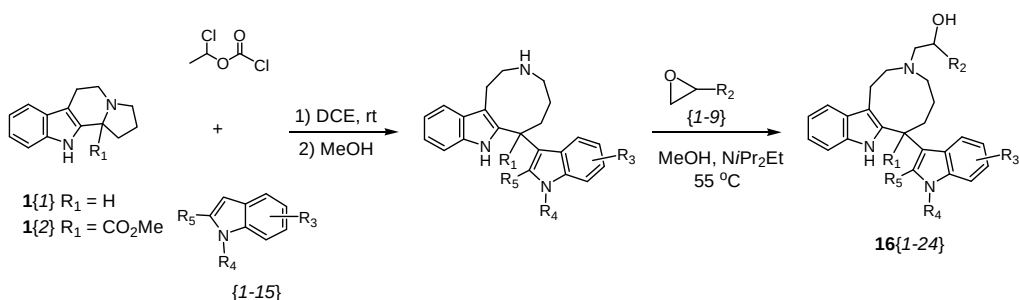


Table 9. Array layout of chemset **16{1-24}**

Library Scaffolds		 1{1}	Library Scaffolds		 1{2}
Indoles{1-15}	Epoxides{1-9}	Product	Indoles{1-15}	Epoxides{1-9}	Product
{1}	{1}	16{1} (15.6, 97, 100)	{12}	{1}	16{13} (8.4, 65, 89)
{2}	{2}	16{2} (41.9, 76, 94)	{1}	{4}	16{14} (11.7, 26, 16)
{3}	{3}	16{3} (44.6, 50, 100)	{7}	{6}	16{15} (10.5, 73, 93)
{4}	{4}	16{4} (21.7, 72, 88)	{3}	{4}	16{16} Not isolated
{5}	{1}	16{5} (24.9, 91, 100)	{5}	{1}	16{17} (3.0, 69, 97)
{6}	{4}	16{6} (38.9, 61, 79)	{8}	{3}	16{18} (16.3, 67, 100)
{1}	{5}	16{7} (12.5, 97, 100)	{1}	{8}	16{19} (8.8, 93, 100)
{7}	{1}	16{8} (29.7, 86, 100)	{6}	{9}	16{20} (13.2, 67, 100)
{8}	{6}	16{9} (32.1, 100, 100)	{13}	{3}	16{21} (27.0, 50, 100)
{9}	{3}	16{10} (36.0, 95, 100)	{11}	{7}	16{22} (8.9, 33, 45)
{10}	{7}	16{11} (26.3, 94, 100)	{14}	{1}	16{23} Not isolated
{11}	{8}	16{12} (29.6, 97, 100)	{15}	{4}	16{24} (14.2, 71, 96)

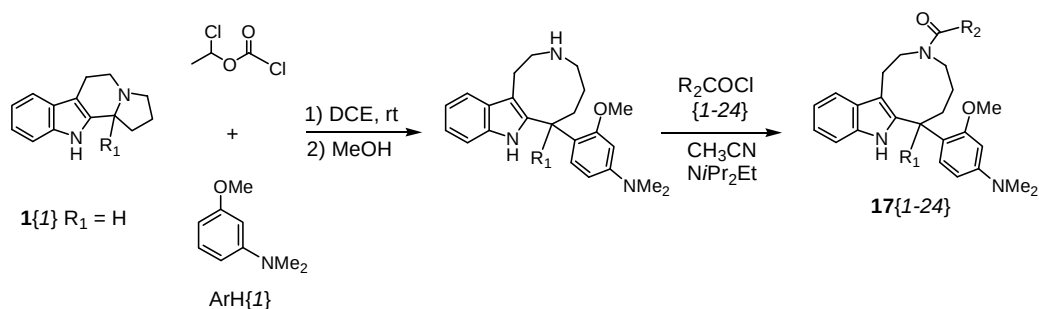
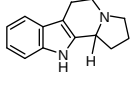
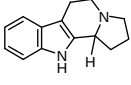
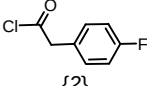
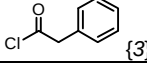
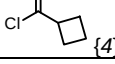
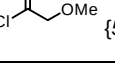
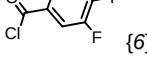
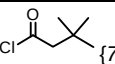
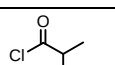
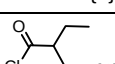
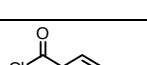
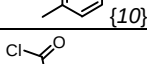
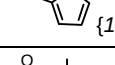


Table 10. Array layout of chemset $17\{1-24\}$

Library Scaffolds		 $1\{1\}$	Library Scaffolds		 $1\{1\}$
ArH{1}	Acid Chlorides {1-24}	Product	ArH{1}	Acid Chlorides {1-24}	Product
<i>m</i> -dimethylamino anisole	 $\{1\}$	$17\{1\}$ (33.5, 0, 100)	<i>m</i> -dimethylamino anisole	 $\{13\}$	$17\{13\}$ (30.5, 97, 100)
<i>m</i> -dimethylamino anisole	 $\{2\}$	$17\{2\}$ (24.8, 74, 100)	<i>m</i> -dimethylamino anisole	 $\{14\}$	$17\{14\}$ Not isolated
<i>m</i> -dimethylamino anisole	 $\{3\}$	$17\{3\}$ (29.5, 67, 100)	<i>m</i> -dimethylamino anisole	 $\{15\}$	$17\{15\}$ (37.6, 100, 100)
<i>m</i> -dimethylamino anisole	 $\{4\}$	$17\{4\}$ (42.3, 89, 100)	<i>m</i> -dimethylamino anisole	 $\{16\}$	$17\{16\}$ (47.4, 95, 100)
<i>m</i> -dimethylamino anisole	 $\{5\}$	$17\{5\}$ (34.1, 100, 100)	<i>m</i> -dimethylamino anisole	 $\{17\}$	$17\{17\}$ (36.6, 80, 100)
<i>m</i> -dimethylamino anisole	 $\{6\}$	$17\{6\}$ (34.8, 97, 100)	<i>m</i> -dimethylamino anisole	 $\{18\}$	$17\{18\}$ (16.9, 88, 100)
<i>m</i> -dimethylamino anisole	 $\{7\}$	$17\{7\}$ (30.8, 100, 100)	<i>m</i> -dimethylamino anisole	 $\{19\}$	$17\{19\}$ (29.9, 100, 100)
<i>m</i> -dimethylamino anisole	 $\{8\}$	$17\{8\}$ (44.6, 94, 100)	<i>m</i> -dimethylamino anisole	 $\{20\}$	$17\{20\}$ (38.8, 83, 100)
<i>m</i> -dimethylamino anisole	 $\{9\}$	$17\{9\}$ (40.6, 83, 100)	<i>m</i> -dimethylamino anisole	 $\{21\}$	$17\{21\}$ (40.1, 87, 100)
<i>m</i> -dimethylamino anisole	 $\{10\}$	$17\{10\}$ (39.3, 85, 100)	<i>m</i> -dimethylamino anisole	 $\{22\}$	$17\{22\}$ (45.1, 94, 100)
<i>m</i> -dimethylamino anisole	 $\{11\}$	$17\{11\}$ (22.6, 65, 100)	<i>m</i> -dimethylamino anisole	 $\{23\}$	$17\{23\}$ (17.3, 87, 100)
<i>m</i> -dimethylamino anisole	 $\{12\}$	$17\{12\}$ (42.8, 87, 100)	<i>m</i> -dimethylamino anisole	 $\{24\}$	$17\{24\}$ (35.9, 90, 100)

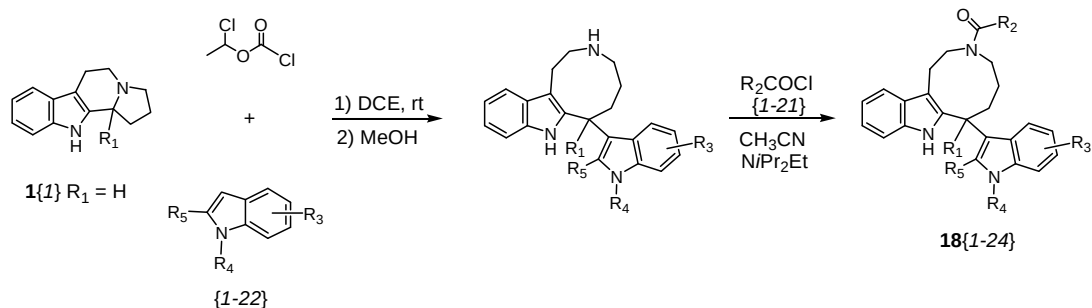
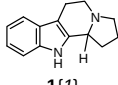
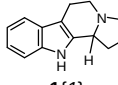
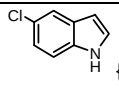
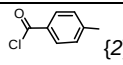
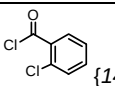
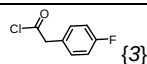
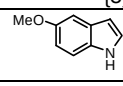
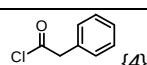
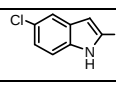
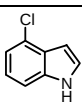
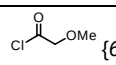
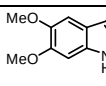
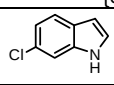
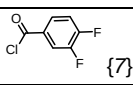
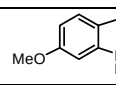
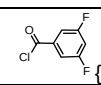
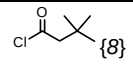
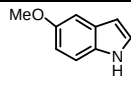
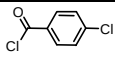
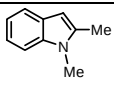
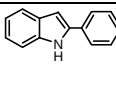
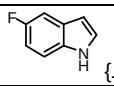
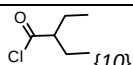
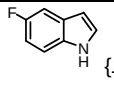
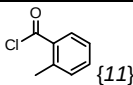
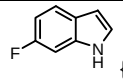
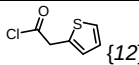
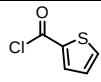


Table 11. Array layout of chemset **18**{1-24}

Library Scaffolds		 1{1}	Library Scaffolds		 1{1}
Indoles {1-22}	Acid Chlorides {1-21}	Product	Indoles {1-22}	Acid Chlorides {1-21}	Product
 {1}	 {1}	18 {1} Not isolated	 {12}	 {13}	18 {13} (38.6, 59, 100)
 {2}	 {2}	18 {2} Not isolated	 {13}	 {14}	18 {14} Not isolated
 {3}	 {3}	18 {3} Not isolated	 {14}	 {15}	18 {15} Not isolated
 {4}	 {4}	18 {4} Not isolated	 {15}	 {16}	18 {16} (25.3, 98, 100)
 {5}	 {5}	18 {5} Not isolated	 {16}	 {17}	18 {17} Not isolated
 {6}	 {6}	18 {6} (18.5, 59, 60)	 {17}	 {18}	18 {18} Not isolated
 {7}	 {7}	18 {7} Not isolated	 {18}	 {19}	18 {19} Not isolated
 {8}	 {8}	18 {8} (46.9, 90, 100)	 {19}	 {20}	18 {20} Not isolated
 {9}	 {9}	18 {9} (36.2, 28, 100)	 {20}	 {21}	18 {21} (13.5, 83, 100)
 {10}	 {10}	18 {10} (33.5, 77, 100)	 {21}	 {16}	18 {22} (24.8, 89, 100)
 {10}	 {11}	18 {11} (30.2, 67, 96)	 {22}	 {6}	18 {23} Not isolated
 {11}	 {12}	18 {12} Not isolated	 {1}	 {18}	18 {24} Not isolated

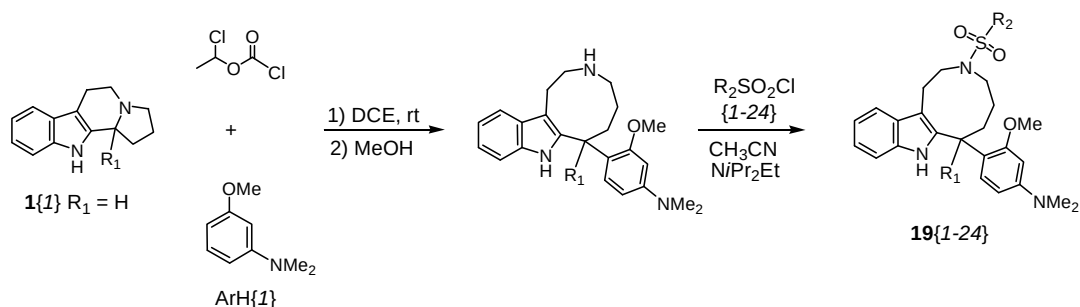
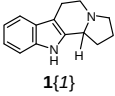
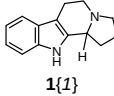
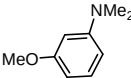
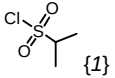
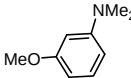
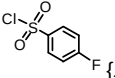
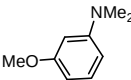
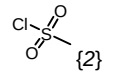
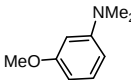
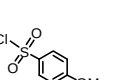
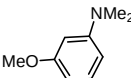
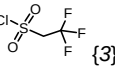
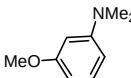
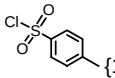
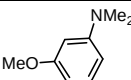
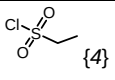
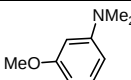
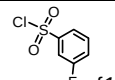
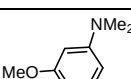
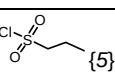
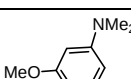
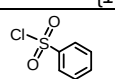
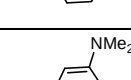
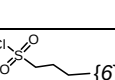
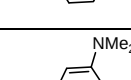
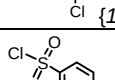
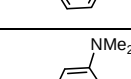
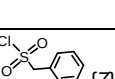
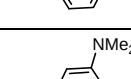
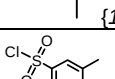
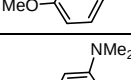
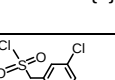
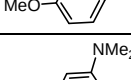
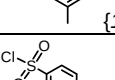
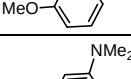
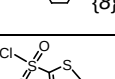
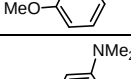
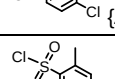
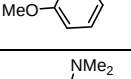
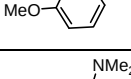
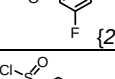
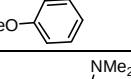
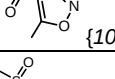
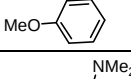
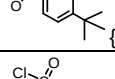
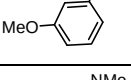
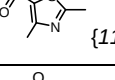
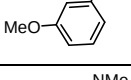
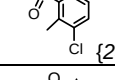


Table 12. Array layout of chemset **19**{1-24}

Library Scaffolds		 1 {1}	Library Scaffolds		 1 {1}
ArH{1}	Sulfonyl Chlorides{1-24}	Product	ArH{1}	Sulfonyl Chlorides{1-24}	Product
	 {1}	19 {1} Not isolated		 {13}	19 {13} (32.8, 100, 100)
	 {2}	19 {2} (18.9, 100, 100)		 {14}	19 {14} (30.7, 100, 100)
	 {3}	19 {3} Not isolated		 {15}	19 {15} (31.1, 100, 100)
	 {4}	19 {4} (16.3, 68, 93)		 {16}	19 {16} (30.7, 100, 100)
	 {5}	19 {5} (10.1, 60, 88)		 {17}	19 {17} (29.4, 100, 100)
	 {6}	19 {6} Not isolated		 {18}	19 {18} (30.3, 100, 100)
	 {7}	19 {7} (10.0, 100, 100)		 {19}	19 {19} (32.0, 100, 100)
	 {8}	19 {8} (5.6, 100, 100)		 {20}	19 {20} (30.3, 100, 100)
	 {9}	19 {9} (33.8, 100, 100)		 {21}	19 {21} (29.7, 100, 100)
	 {10}	19 {10} (29.3, 94, 100)		 {22}	19 {22} (27.3, 97, 100)
	 {11}	19 {11} (30.1, 97, 100)		 {23}	19 {23} (27.0, 100, 100)
	 {12}	19 {12} (29.6, 100, 100)		 {24}	19 {24} (29.9, 100, 100)

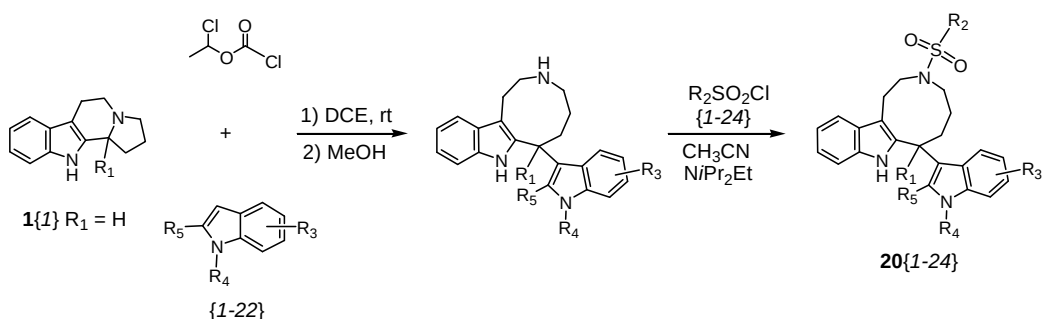


Table 13. Array layout of chemset **20**{1-24}

Library Scaffolds			Library Scaffolds	
Indoles {1-22}	Sulfonyl Chlorides{1-24}	Product	Indoles {1-22}	Product
		20 {1} (4.2, 31, 25)		20 {13} (12.5, 93, 100)
		20 {2} (15.0, 88, 100)		20 {14} Not isolated
		20 {3} Not isolated		20 {15} Not isolated
		20 {4} Not isolated		20 {16} (5.4, 0, 100)
		20 {5} Not isolated		20 {17} Not isolated
		20 {6} (5.2, 100, 100)		20 {18} Not isolated
		20 {7} (6.5, 45, 98)		20 {19} (8.7, 84, 100)
		20 {8} Not isolated		20 {20} (7.3, 0, 100)
		20 {9} Not isolated		20 {21} (20.1, 100, 100)
		20 {10} (22.1, 88, 100)		20 {22} Not isolated
		20 {11} (112.6, 100, 100)		20 {23} Not isolated
		20 {12} (15.0, 87, 100)		20 {24} (17.6, 100, 100)