Supporting Information for

[2Fe-2S] proteins in chlorosomes: CsmI and CsmJ participate in light-dependent control of energy transfer in chlorosomes of *Chlorobaculum tepidum*

Hui Li[†]*, Niels-Ulrik Frigaard^{*}[§], and Donald A. Bryant^{+ ||}*

[†]Department of Biochemistry and Molecular Biology, The Pennsylvania State University, University Park, PA 16802 USA

^{*}Present Address: Department of Genome Sciences, University of Washington, Seattle WA 98195 USA

[§]Present address: Section for Marine Biology, Department of Biology, University of Copenhagen, Strandpromenaden 5, 3000 Helsingør, Denmark

¹¹Department of Chemistry and Biochemistry, Montana State University, Bozeman, MT 59717 USA

* To whom correspondence should be addressed. 108 Althouse Laboratory, The Pennsylvania State University, University Park, PA 16802 USA; phone: (814) 865-1992; fax: (814) 863-7024; e-mail: dab14@psu.edu

Materials and Methods Addenda:

Cell viability testing. Cells in late exponential phase (OD_{600 nm} ~1.5) were diluted 100-fold in anoxic or oxic K-phosphate buffer, pH 7.0. Tubes containing anoxic cell suspensions were wrapped in foil (to maintain dark conditions) and were incubated in an anoxic chamber. Oxic cell suspensions were placed in darkness or under ambient (room) light (~50 µmol photons m⁻² s⁻¹) with their caps attached loosely. After 3 or 6 days, the cell suspensions were diluted 1×10^4 -fold, and an aliquot (30 µL) of cells was plated on CL plates solidified with 1.5% (w/v) phytagel. The plates were incubated in the light at ~40°C for 6 days in an anoxic chamber, and the resulting colonies were counted.

Pigment and quinone analyses. BChls, carotenoids, and quinones were extracted with acetonemethanol (150 µL, 7:2, v/v) from an aliquot of chlorosomes (1 µL). After mixing and filtration, ammonium acetate (0.1 volume of 1.0 M) was added prior to loading the extracts onto a 4.5 mm × 25 cm Discovery C₁₈ reversed-phase, high-performance liquid chromatography (HPLC) column (Supelco, Bellefonte, PA). The Agilent 1100 series HPLC system (Agilent Technologies, Palo Alto, CA) was controlled with Agilent ChemStation software. HPLC analyses were performed as described (*1, 2*). Pigment concentrations were determined by absorption spectroscopy in methanol using the following absorption coefficients: BChl *c*, 20 L· g⁻¹ cm⁻¹ at 635 nm; BChl *a*, 60 L · g⁻¹ cm⁻¹ at 770 nm; carotenoids, 265 L · g⁻¹ cm⁻¹ at 491 nm; chlorobiumquinones (1'-oxomenaquinone-7 and 1'-hydroxymenaquinone-7), 17 L · g⁻¹ cm⁻¹ at 270 nm; menaquinone-7, 26 L· g⁻¹ cm⁻¹ at 270 nm.

References for Supplement:

- Frigaard, N.-U., Takaichi, S.,Hirota, M., Shimada, K., Matsuura, K. 1997. Quinones in chlorosomes of green sulfur bacteria and their role in the redox-dependent fluorescence studied in chlorosome-like bacteriochlorophyll *c* aggregates. *Arch. Microbiol.* 167, 343-349.
- Frigaard, N.-U., Li, H., Milks, K. J., Bryant, D. A. 2004. Nine mutants of *Chlorobium tepidum* each unable to synthesize a different chlorosome protein still assemble functional chlorosomes. *J. Bacteriol.* 186, 646-53.
- Li, H., Jubilerer, S., Garcia Costas, A. M., Frigaard, N.-U., and Bryant D. A. (2009) Multiple antioxidant proteins protect *Chlorobaculum tepidum* against oxygen and reactive oxygen species. *Arch. Microbiol. 191*, 853-867.

Name	Size	Description	Antibiotic
	(kbp)		Resistance ^a
pHP45Ω	4.4	Contains a 2.1-kb <i>aadA</i> streptomycin and	Ap ^R Sm ^R Sp ^R
		spectinomycin cassette	
pMS255	3.7	Contains a 1.1-kb <i>aacC1</i> gentamicin cassette	$Ap^{R} Gm^{R}$
pRL409	5.4	^b Contains a 2.8-kb <i>cat-ermC</i> cassette	Ap ^R Cm ^R
			Em ^R
pET3d::csmI::aadA	7.3	Constructed by insertion of <i>aadA</i> cassette from	Ap ^R Sm ^R Sp ^R
		pHP45 Ω into the <i>Xcm</i> I site of <i>csmI</i> in	
		pET3d::csmI	
pET32a::csmX::aadA	8.6	Constructed by insertion of <i>aadA</i> cassette from	Ap ^R Sm ^R Sp ^R
		pHP45 Ω into the <i>Mfe</i> I site of <i>csmX</i> in	
		pET32a::csmX	
pET3d::csmI::cat-ermC	8.1	Constructed by deletion of <i>aadA</i> marker from	Ap ^R Cm ^R Em ^R
		pET3d:: <i>csmI</i> :: <i>aadA</i> , and then inserting <i>cat-ermC</i>	
		marker at the same XcmI site	
°pET3d::csmJ::aacC1	6.4	Constructed by insertion of <i>aacC1</i> from pMS255	Ap ^R Gm ^R
		into the ApoI site of <i>csmJ</i> in pET3d:: <i>csmJ</i>	

^aAp^R, ampicillin resistance; Em^R, erythromycin resistance; Gm^R, gentamicin resistance; Sm^R, streptomycin resistance; Sp^R, spectinomycin resistance.

^bThe Cm^R marker is functional in *E. coli,* and the Em^R marker is functional in *Cba. tepidum*.

^cThe EcoRI site within pET3d::*csmJ* was previously deleted.

	BChl a	Carotenoids	Chlorobium-	Mena-	Total
			quinones	quinone-7	Quinones
wild type	11 ± 1	67 ± 7	54 ± 4	10 ± 2	64 ± 6
csmI	12 ± 1	70 ± 3	46 ± 7	17 ± 5	63 ± 12
csmJ	15 ± 4	64 ± 2	62 ± 9	14 ± 2	76 ± 11
csmX	12 ± 3	67 ± 2	51 ± 11	12 ± 1	63 ± 12
csmI csmX	11 ± 1	56 ± 6	60 ± 11	13 ± 5	73 ± 16
csmJ csmX	13 ± 1	57 ± 2	66 ± 10	14 ± 4	80 ± 14
csmI csmJ	14 ± 2	52 ± 5	36 ± 8	25 ± 5	61 ± 13
csmI csmJ csmX	14 ± 1	53 ± 3	38 ± 8	28 ± 7	66 ± 15

Table S2. Pigment and quinone contents of chlorosomes from WT and mutant strains^a

^a Values are reported as mg per g of BChl c. Contents were determined by HPLC analysis as previously described (2). All values are the averages and standard deviations of at least two measurements for two separate chlorosome preparations from different cell cultures.

	low irradiance	moderate irradiance	high irradiance
	8 μ mol photons m ⁻² s ⁻¹	$30 \ \mu mol \ photons \ m^{-2} \ s^{-1}$	150 μ mol photons m ⁻² s ⁻¹
WT	0.047 ± 0.004	0.203 ± 0.007	0.277 ± 0.008
csmI	0.039 ± 0.005	0.203 ± 0.005	0.263 ± 0.010
csmJ	0.039 ± 0.002	0.170 ± 0.005	0.236 ± 0.005
csmX	0.044 ± 0.004	0.193 ± 0.006	0.275 ± 0.004
csmI csmX	0.041 ± 0.003	0.174 ± 0.012	0.248 ± 0.007
csmJ csmX	0.043 ± 0.003	0.168 ± 0.007	0.229 ± 0.006
csmI csmJ ^b	0.040 ± 0.004	0.168 ± 0.005	0.163 ± 0.007
csmI csmJ csmX	0.042 ± 0.005	0.174 ± 0.004	0.254 ± 0.009

Table S3. Growth rates of WT and mutant strains at three light intensities.^a

^a Specific growth rates (μ, h^{-1}) are reported for 47°C. The values are averages and standard errors for at least two independent cultures.

^b Three independent *csmI csmJ* mutants were isolated and characterized. One *csmI csmJ* mutant failed to grow at high irradiance and grew much more slowly at moderate irradiance. This strain was also shown to be temperature-sensitive. Two other strains of the *csmI csmJ* mutant grew a little more slowly than wild type at moderate irradiance but grew more slowly (~60%) than other mutants and wild type at high irradiance. The values reported here are the average values for these other two strains.

Strain/conditions ^a	72 h/ 3 d		144 h/ 6 d	
	Colonies ^b	Viability (%) ^c	Colonies ^b	Viability (%) ^c
WT (dark, anoxic)	86 ± 0	100	85 ± 5	99
WT (dark, oxic)	50 ± 4	58	24 ± 2	28
WT (light, oxic)	27 ± 3	31	0	≤1
csmI csmJ csmX (dark, anoxic)	90 ± 3	100	62 ± 3	69
csmI csmJ csmX (dark, oxic)	44 ± 8	49	12 ± 1	13
csmI csmJ csmX (light, oxic)	14 ± 4	16	0	≤1

Table S4. Cell viability after 3 and 6 days under anoxic and oxic conditions (± room light).

^a Strains and conditions: Wild type (WT) and *csmI csmJ csmX* mutant cells were incubated in the dark or under ambient room light (~50 μ mol photons m⁻² s⁻¹) for 3 d or 6 d (72 and 144 h, respectively) under oxic or anoxic conditions as specified (see supplemental Materials and Methods for additional details).

^b Each number is the average of the colonies that appeared on two plates after an identical dilution was performed (see supplemental Materials and Methods for additional details). ^c Viability was calculated as the percentage of colony-forming units relative to the control culture maintained under dark anoxic conditions for 72 h. Preliminary studies showed that no loss of viability occurred during this time (also see Li et al. (*3*)).

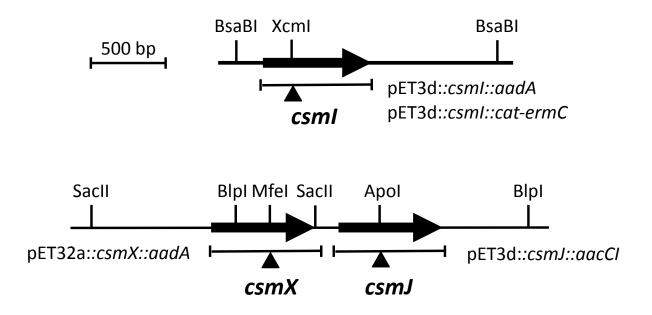


Figure S1. Restriction maps showing the physical maps of constructions used for inactivation of the *csmI*, *csmJ*, and *csmX* genes (see Table S1 and (2)).