Supporting Information

# Nardoaristolones A and B, Two Terpenoids with Unusual Skeletons from *Nardostachys chinensis* Batal

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## **General experimental procedures**

Optical rotations were measured on a Jasco P-1020 polarimeter with a 1 cm cell at room temperature. UV spectra were recorded on a JASCO V-550 UV/Vis spectrometer. IR spectra were obtained using a JASCO FT/IR-480 plus spectrometer. CD spectra were obtained on a Jasco J-810 spectropolarimeter at room temperature. HR-ESI-MS spectra were acquired using a Waters Synapt G2 mass spectrometer. The NMR spectra were measured with a Bruker AV-300/400/600 spectrometer at room temperature. Silica gel (200-300 mesh, Qingdao Marine Chemical Ltd., China), octadecylsilanized (ODS) silica gel (YMC Ltd., Japan), and Sephadex LH-20 (Amersham Pharmacia Biotech, Sweden) were used for open column chromatography (CC).

# Plant Material

The underground parts of *Nardostachys chinensis* were provided by Shijiazhuang Yiling Pharmaceutical Co., Ltd and identified by Dr. Qingcun Tian of the company.

### **Extraction and Isolation**

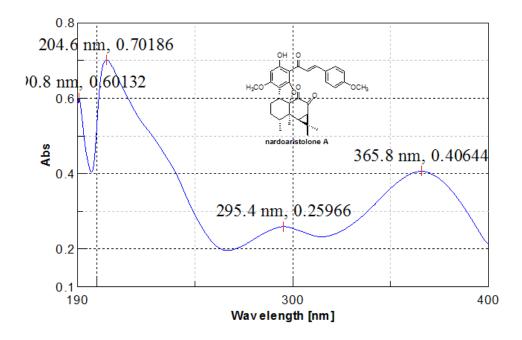
The air-dried medical materials (5 Kg) were refluxed twice with 60% EtOH for 2 hours each time. The crude extract (970.5g) was column chromatographed over a macroporous resin HP-20 eluted with EtOH-H<sub>2</sub>O in gradient. The 70% EtOH-H<sub>2</sub>O eluent (171.7 g) was fractionated by silica gel column chromatography eluted with CHCl<sub>3</sub>-CH<sub>3</sub>OH (100:0 $\rightarrow$ 0:100) to afford twelve fractions (Fr. 1-12). Fr. 3 (53.4 g, CHCl<sub>3</sub>/CH<sub>3</sub>OH 99:1) was then chromatographed on a silica gel column using petroleum ether-EtOAc gradient elution to yield 9 subfractions (Fr. 3.1-3.9). The subfraction Fr. 3.8 (9.1 g, P/E 100:10) was further subjected to ODS column chromatography eluted with MeOH-H<sub>2</sub>O (50:50 $\rightarrow$ 100:0) and purified by preparative HPLC on ODS column with 45% MeOH-H<sub>2</sub>O to yield compound **2** (89.2 mg). Another subfraction Fr. 3.9 (2.1 g, P/E 100:15) was also subjected to ODS column chromatography eluted with MeOH-H<sub>2</sub>O (50:50 $\rightarrow$ 100:0) and purified by Sephadex LH-20 column chromatography (CHCl<sub>3</sub>/CH<sub>3</sub>OH 7:3) to afford compound **1** (5.6 mg).

# Physico-chemical constants of 1-2

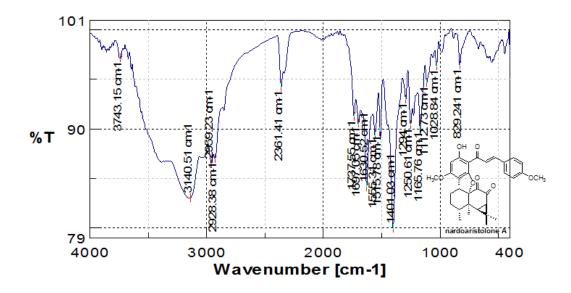
Nardoaristolone A (1): Yellow powder;  $[\alpha]_{D}^{26}$  -74.6 (*c* 0.50, CH<sub>3</sub>OH); UV (CH<sub>3</sub>OH)  $\lambda_{max}$  (log  $\varepsilon$ ) 204.6 (3.87), 295.4 (3.44), 365.8 (3.63) nm; IR (KBr)  $\lambda_{max}$  3140, 2361, 1630, 1515, 1401, 1250, 1165, 829 cm<sup>-1</sup>; CD (CH<sub>3</sub>OH)  $\lambda_{max}$  ( $\Delta\varepsilon$ ) 395 (+2.35), 318 (-2.61), 268 (-1.40), 235 (+0.41), 215 (-1.12) nm; HR-ESI-MS m/z 531.2380 ([M+H]<sup>+</sup>, calcd for C<sub>32</sub>H<sub>35</sub>O<sub>7</sub>, 531.2383).

Nardoaristolone B (**2**): Light yellow crystals;  $[\alpha]_{D}^{26}$  -19.6 (*c* 0.50, CH<sub>3</sub>OH); UV (CH<sub>3</sub>OH)  $\lambda_{max}$  (log  $\epsilon$ ) 254.2 (3.66) nm; IR (KBr)  $\lambda_{max}$  1704, 1671, 1454, 1370, 1214, 1033, 869 cm<sup>-1</sup>; CD (CH<sub>3</sub>OH)  $\lambda_{max}$  ( $\Delta\epsilon$ ) 398 (+1.11), 336 (-1.78), 269 (-2.54), 230 (+4.53), 214 (+4.10) nm; HR-ESI-MS m/z 219.1377 ([M+H]<sup>+</sup>, calcd for C<sub>14</sub>H<sub>19</sub>O<sub>2</sub>, 219.1385).

UV spectrum of nardoaristolone A (1) in CH<sub>3</sub>OH.

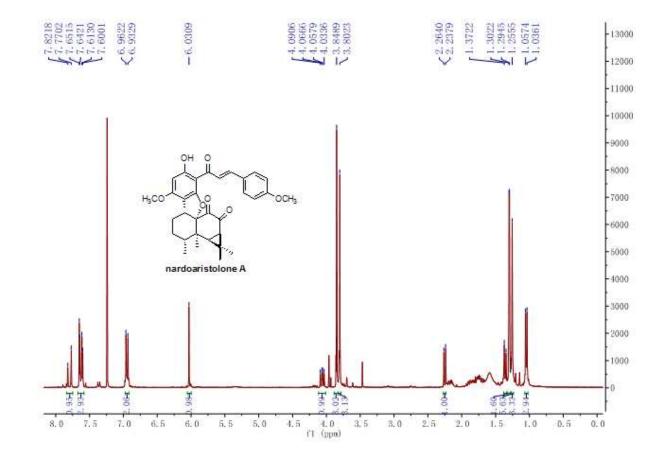


IR (KBr disc) spectrum of nardoaristolone A (1).

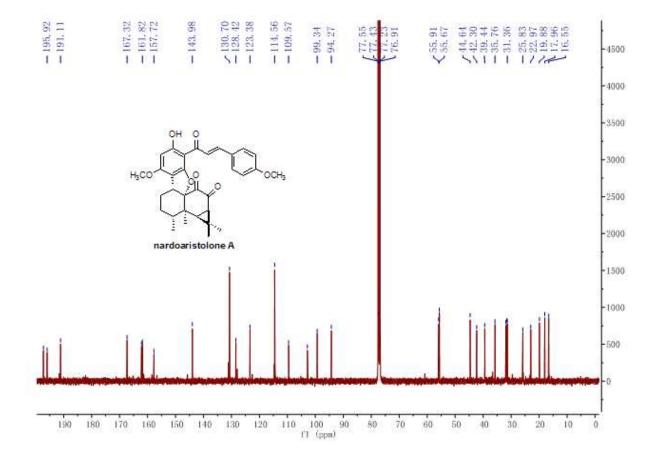


### HR-ESI-MS spectrum of nardoaristolone A (1).

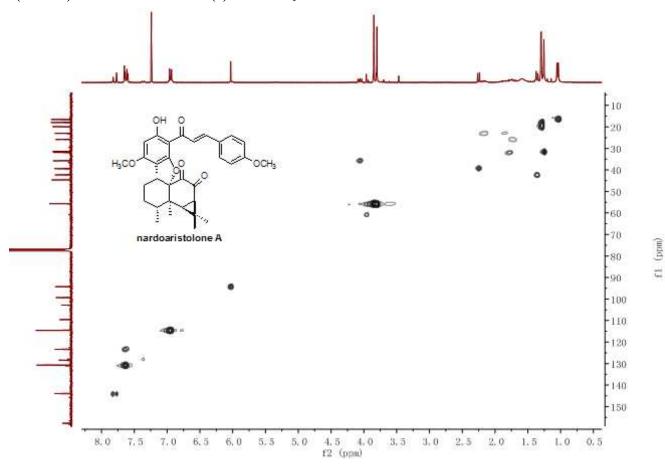
#### **Elemental Composition Report** Page 1 Single Mass Analysis Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3 Monoisotopic Mass, Even Electron Ions 146 formula(e) evaluated with 1 results within limits (up to 50 best isotopic matches for each mass) Elements Used: C: 0-200 H: 0-1000 O: 0-200 OH 0 E3-9-18-2 20120528-08 247 (1.993) Cm (247) 1: TOF MS ES+ 2.32e+005 OCH: H3CO 531.2380 100-16 532.2416 nardoaristolone A 553,2195 533.2439 554.2227 3.2439 535.6028<sup>541.2215</sup>542.7382 535.0 540.0 531.1752 547,8688 0 523.2419 526.3033 550.2088 559.1987 560.0 m/z 530.0 545.0 -525.0 550.0 555.0 Minimum: Maximum: -1.5 5.0 5, 0 Calc. Mass mita 531.2383 -0.3 PPM -0.6 DBE i-FIT Norm Conf(%) Formula 15.5 246.2 n/a n/a C32 H35 07 Mass 531.2380 -0.3



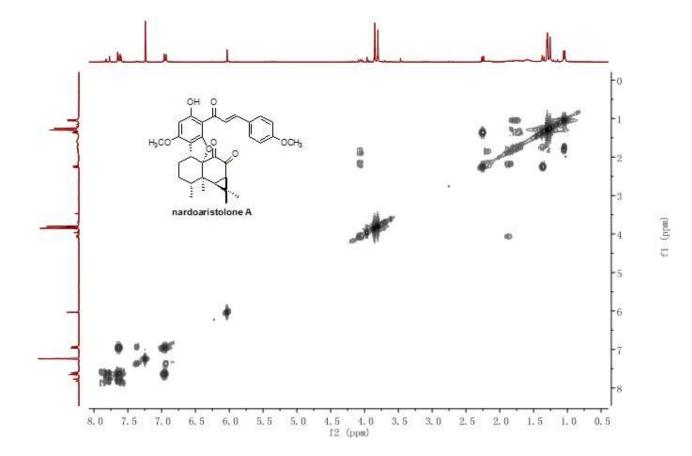
<sup>1</sup>H NMR (AV-300, 300 MHz) spectrum of nardoaristolone A (1) in CDCl<sub>3</sub>



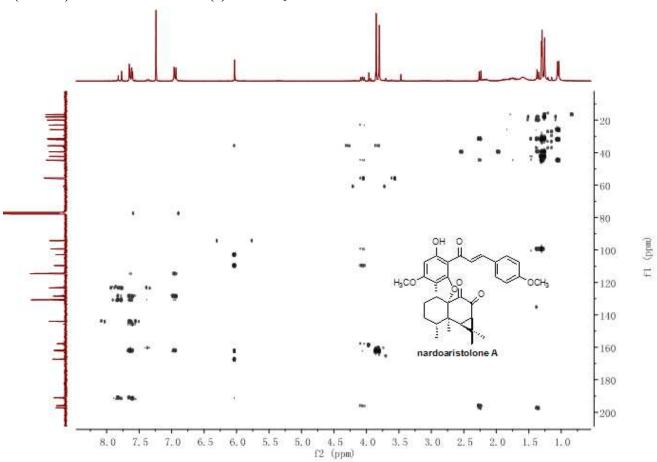
<sup>13</sup>C NMR spectrum (AV-400, 100 MHz) of nardoaristolone A (1) in CDCl<sub>3</sub>



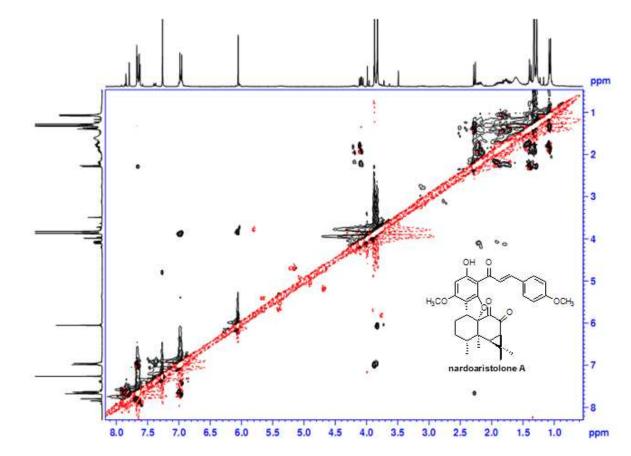
HSQC spectrum (AV-400) of nardoaristolone A (1)1 in CDCl<sub>3</sub>



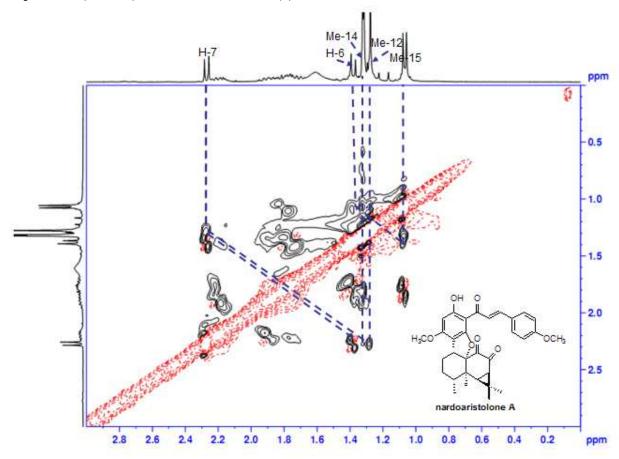
<sup>1</sup>H<sup>-1</sup>H COSY spectrum (AV-400) of nardoaristolone A (1) in CDCl<sub>3</sub>



HMBC spectrum (AV-400) of nardoaristolone A (1) in CDCl<sub>3</sub>

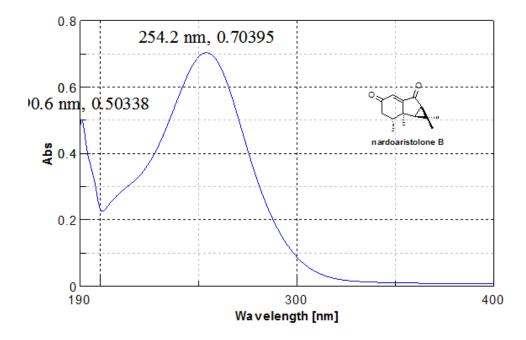


ROESY spectrum (AV-400) of nardoaristolone A (1) in CDCl<sub>3</sub>

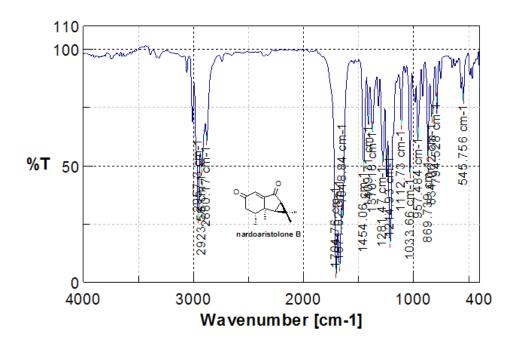


Enlarged ROESY spectrum (AV-400) of nardoaristolone A (1) in CDCl<sub>3</sub>

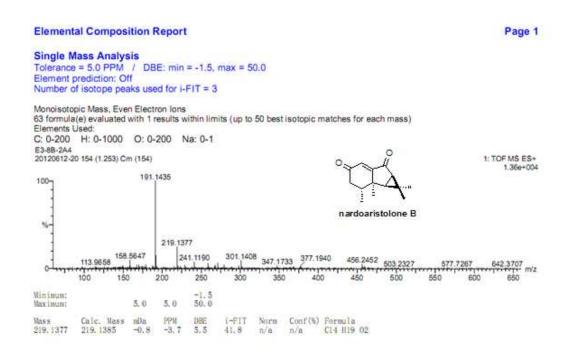
# UV spectrum of nardoaristolone B (2) in CH<sub>3</sub>OH.

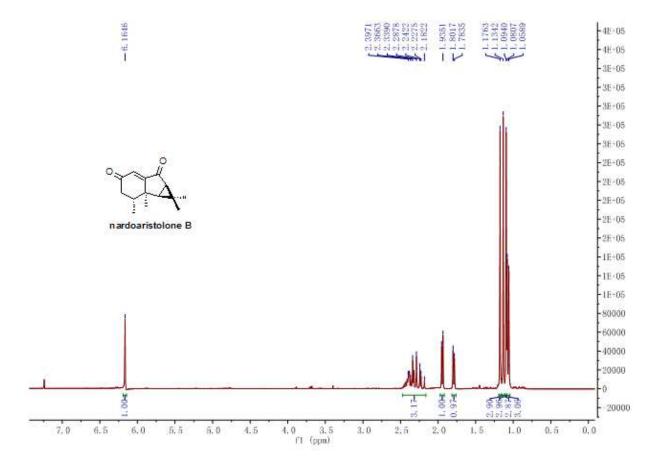


IR (KBr disc) spectrum of nardoaristolone B (2).

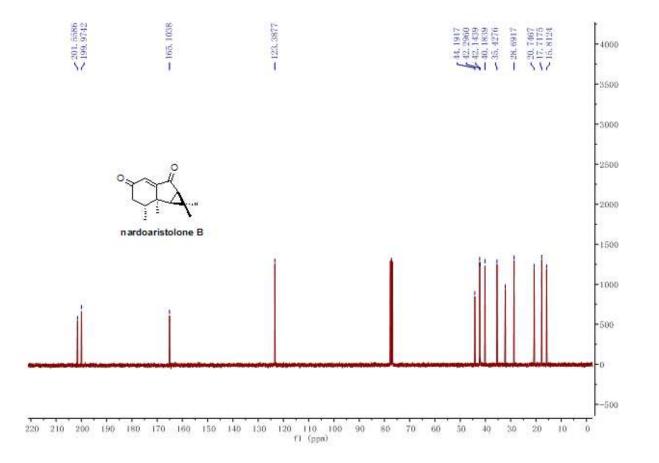


### HR-ESI-MS spectrum of nardoaristolone B (2).

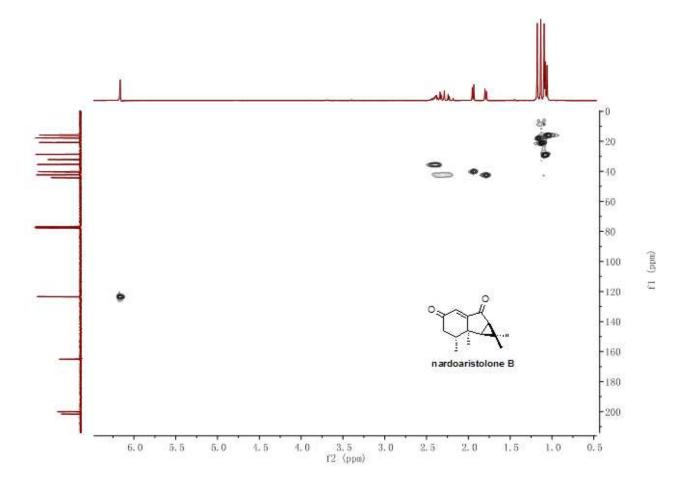




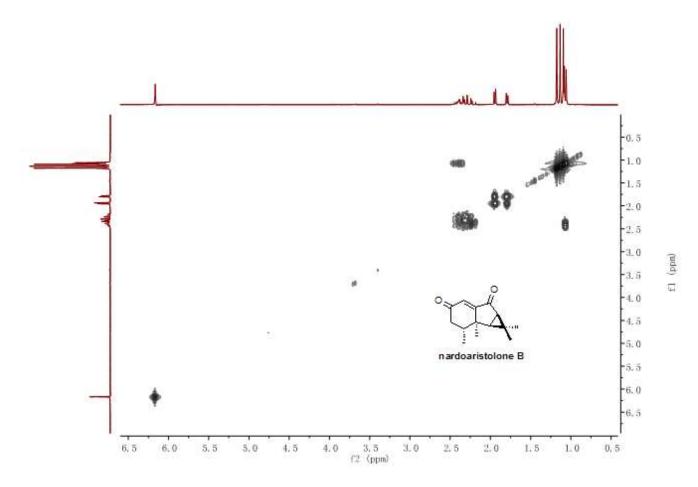
<sup>1</sup>H NMR (AV-300, 300 MHz) spectrum of nardoaristolone B (2) in CDCl<sub>3</sub>



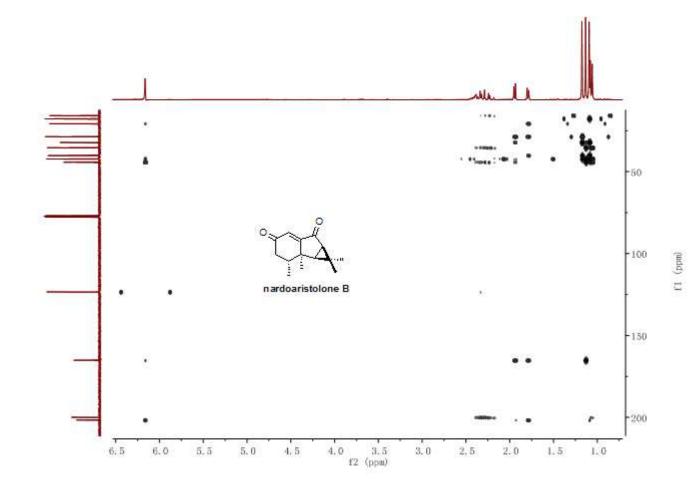
<sup>13</sup>C NMR spectrum (AV-300, 75 MHz) of nardoaristolone B (2) in CDCl<sub>3</sub>



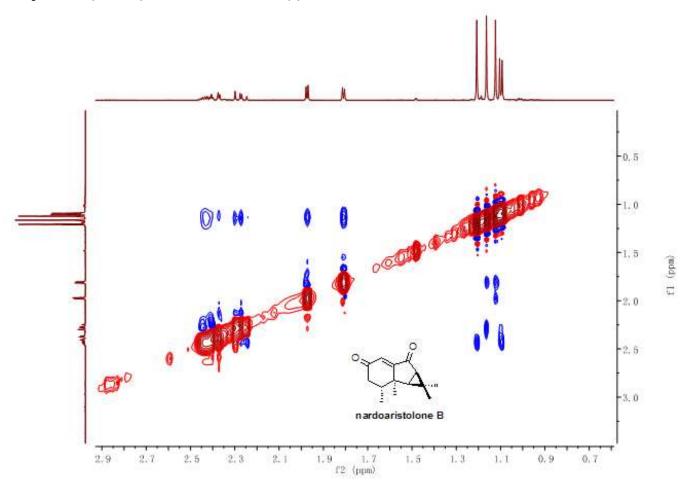
HSQC spectrum (AV-300) of nardoaristolone B (2)1 in CDCl<sub>3</sub>



<sup>1</sup>H<sup>-1</sup>H COSY spectrum (AV-300) of nardoaristolone B (2) in CDCl<sub>3</sub>



HMBC spectrum (AV-300) of nardoaristolone B (2) in CDCl<sub>3</sub>



Enlarged NOESY spectrum (AV-600) of nardoaristolone B (2) in CDCl<sub>3</sub>

### Isolation and culture of cardiomyocytes

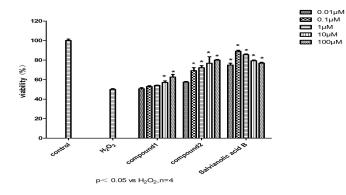
The neonatal rat cardiomyocytes were prepared from one to three-day-old Wistar rats by trypsin as described previously<sup>1,2</sup>. The ventricular myocardium was minced in DMEM (Dulbeco's Modified Eagle Medium, Hyclone, USA), which contains 25 mM D-glucose and 4 mM L-Glutamine. After the successive digestion with 0.25% trypsin, the cells were suspended in DMED containning 10% FBS (Fetal Bovine Serum, Hyclone, USA) and then centrifuged. Pooled cells were planted onto a 50 mL-cell culture bottle and incubated for 1.5-2 h at 37 °C in a humidified air with 5% CO<sub>2</sub>. The fibroblasts were depleted afterward with differential velocity adherent technique. Finally, the supernatant cells containing non-adherent cells were planted onto the 96-well plates at a density of  $10^5$  cells·cm<sup>-2</sup> with 0.1 mM bromodeoxyuridine (Sigma, USA), and incubated for 72 h at the same condition as described above.

### **Experimental Classification**

The cultured cells were divided into control, hydrogen peroxide ( $H_2O_2$ ), compound **1**, compound **2** and Salvianolic acid B groups. After starved in serum-free DMEM for 12 h, the cells were treated with 0.01, 0.1, 1, 10, 100  $\mu$ M compound **1** for 12 h, respectively (as well as compound **2** and salvianolic acid B), then all groups were treated with DMEM containing 10% FBS and 100  $\mu$ M H<sub>2</sub>O<sub>2</sub> for 3 h except control group.

### MTT assay for cell viability

[3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT)] (Roche Molecular Biochemicals, Laval, PQ, Canada) was used to quantify the cell survival of each group. After 3 h H<sub>2</sub>O<sub>2</sub> injury, the supernatants of each well were replaced with 120 uL serum DMEM containing 20 uL MTT solution (5mg/ml), and the cells were incubated for 4 h at 37 °C. The cell layers were dissolved with 150 uL DMSO after the supernatants were removed. The optical density of each well was measured at 490 nm using a Model 680 Microplate Reader (Bio-Rad, USA).



Protective effects on H<sub>2</sub>O<sub>2</sub>-induced myocardial injury of compounds 1 and 2.

### **References:**

(1) Lu, Y.; Zhang, Y.; Wang, N.; Pan, Z.; Gao, X.; Zhang, F.; Shan, H.; Luo, X.; Bai, Y.; Sun, L.; Song, W.; Xu, C.; Wang, Z.; Yang, B. *Circulation*. **2010**, *122*, 2378-2387.
(2) Lu, Y.; Zhang, Y.; Shan, H.; Pan, Z.; Li, X.; Li, B.; Xu, C.; Zhang, B.; Zhang, F.; Dong, D.; Song, W.; Qiao, G.; Yang, B. *Cardiovasc Res* **2009**, *84*, 434-441.