

Organocatalytic Asymmetric Michael Addition of 5H-Oxazol-4-Ones to Nitroolefins

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1. General Information

General Procedures and Methods

Experiments involving moisture and/or air sensitive components were performed under a positive pressure of nitrogen in oven-dried glassware equipped with a rubber septum inlet. Dried solvents and liquid reagents were transferred by oven-dried syringes or hypodermic syringe cooled to ambient temperature in a desiccator. Reactions mixtures were stirred in 4 mL sample vial with Teflon-coated magnetic stirring bars unless otherwise stated. Moisture in non-volatile reagents/compounds was removed in high *vacuo* by means of an oil pump and subsequent purging with nitrogen. Solvents were removed in *vacuo* under ~30 mmHg and heated with a water bath at 30–35 °C using rotary evaporator with aspirator. The condenser was cooled with running ethanol at 0 °C.

All experiments were monitored by analytical thin layer chromatography (TLC). TLC was performed on pre-coated plates, 60 F₂₅₄. After elution, plate was visualized under UV illumination at 254 nm for UV active material. Further visualization was achieved by staining KMnO₄, ceric molybdate, or anisaldehyde solution. For those using the aqueous stains, the TLC plates were heated on a hot plate.

Columns for flash chromatography (FC) contained silica gel 200–300 mesh. Columns were packed as slurry of silica gel in petroleum ether and equilibrated solution using the appropriate solvent system. The elution was assisted by applying pressure of about 2 atm with an air pump.

Instrumentations

Proton nuclear magnetic resonance (¹H NMR) and carbon NMR (¹³C NMR) spectra were recorded in CDCl₃ otherwise stated. ¹H (400 MHz) and ¹³C (100 MHz) were performed on a Bruker AVANCE-III (400MHz) spectrometer. Chemical shifts are reported in parts per million (ppm), using the residual solvent signal as an internal standard: CDCl₃ (¹H NMR: δ 7.26, singlet; ¹³C NMR: δ 77.0, triplet). Multiplicities were given as: *s* (singlet), *d* (doublet), *t* (triplet), *q* (quartet), *quintet*, *m* (multiplets), *dd* (doublet of doublets), *dt* (doublet of triplets), and *br* (broad). Coupling constants (*J*) were recorded in Hertz (Hz). The number of proton

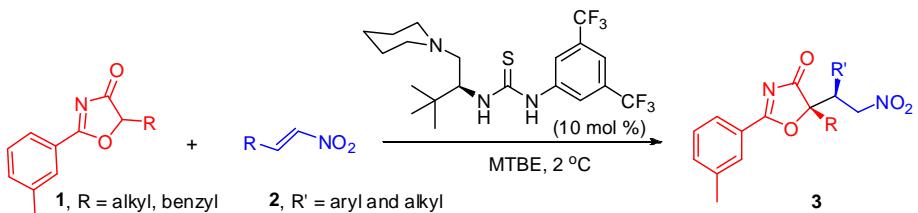
atoms (*n*) for a given resonance was indicated by *nH*. The number of carbon atoms (*n*) for a given resonance was indicated by *nC*. HRMS were reported in units of mass of charge ratio (m/z). Mass samples were dissolved in CH₃CN (HPLC Grade) unless otherwise stated. Optical rotations were recorded on a polarimeter with a sodium lamp of wavelength 589 nm and reported as follows; $[\alpha]_{\lambda}^{T^{\circ}C}$ (c = g/100 mL, solvent). Melting points were determined on a X-6 microscopic melting point apparatus.

Enantiomeric excesses were determined by chiral High Performance Liquid Chromatography (HPLC) analysis on Agilent HPLC 1260 series, including 1260 quaternary pump with degasser, 1260 thermostatted column compartment, 1260 series variable wavelength detector with manual injection valve. HPLC samples were dissolved in HPLC grade isopropanol (IPA) unless otherwise stated.

Materials

All commercial reagents were purchased from Sigma-Aldrich, J&K, Alfa-Aesar, TCI and Aladdin of the highest purity grade. They were used without further purification unless specified. All solvents used, mainly petroleum ether (PE) and ethyl acetate (EtOAc), were distilled. Anhydrous CH₂Cl₂ and MeCN were freshly distilled from CaH₂ and stored under N₂ atmosphere. THF, Et₂O, CPME, MTBE and toluene were freshly distilled from sodium/benzophenone before use. Anhydrous methanol was distilled from Mg. All compounds synthesized were stored in a -20 °C freezer and light-sensitive compounds were protected with aluminium foil.

2. General Procedure for The Asymmetric Addition of 5H-Oxazol-4-ones **1 to Nitroolefins **2** Catalyzed by **IV****

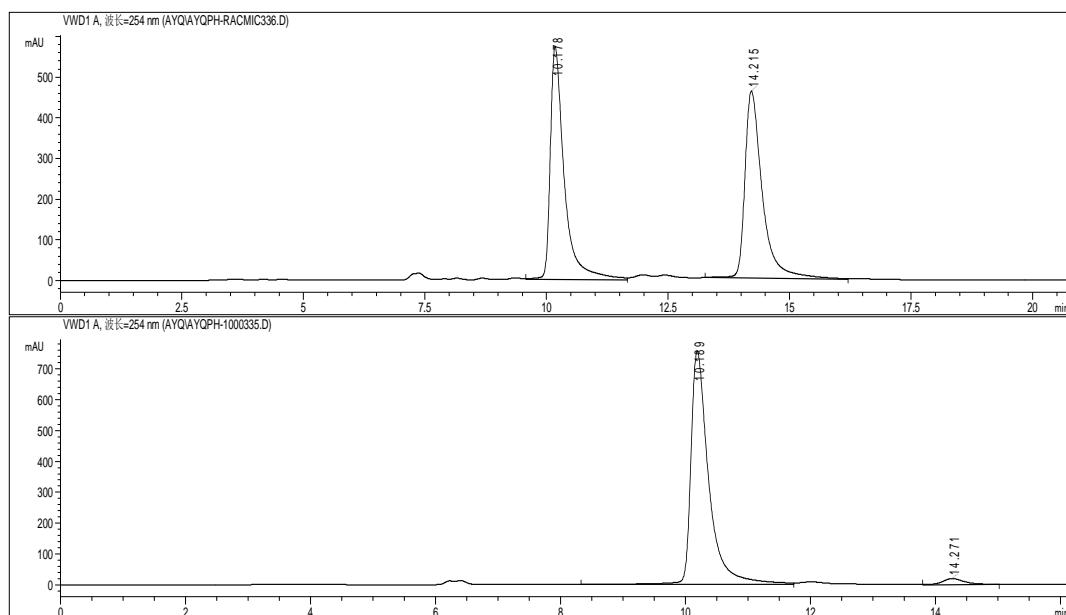


5*H*-Oxazol-4-ones **1** (0.1 mmol, 1.0 equiv.) and **IV** (0.01 mmol, 0.1 equiv.) were dissolved in MTBE (1.0 mL). The reaction mixture was stirred at 2 °C for 30 minutes, nitroolefin **2** (0.11 mmol, 1.1 equiv.) was added in one portion to the above mixture. The reaction mixture was stirred at 2 °C for 40 h and monitored by TLC. Upon complete consumption of 5*H*-oxazol-4-ones, the reaction mixture was concentrated under reduced pressure. The crude material was analyzed by ¹H NMR in order to determine diastereoselectivity. When the dr value is determined more than 19:1 by ¹H NMR, it is described in paper as >19:1 dr due to the accuracy of the measurement. The recovered crude material was subsequently purified by flash column chromatography on silica gel with PE/EtOAc mixture (20:1–5:1 ratio, the crude material was completely dissolved in CH₂Cl₂/PE before loaded on silica gel). After removing the solvent in *vacuo*, the major diasteromer of **3** could be obtained.

3. Characterization of Adducts

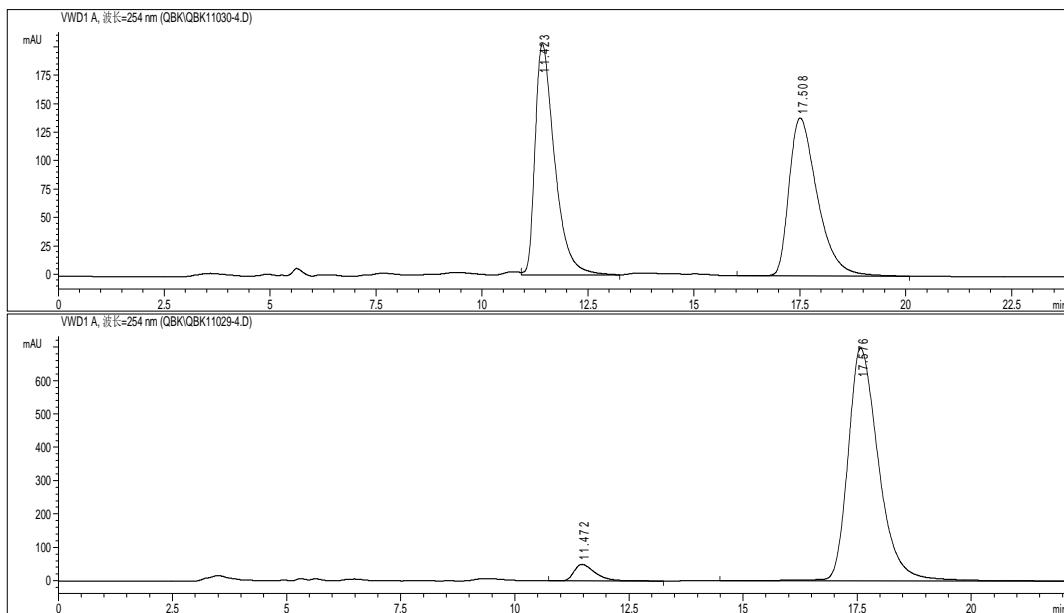
3ea, White solid; Mp 139.1–139.9 °C; 94% ee; dr >19:1; 93% yield (major diastereomer); $[\alpha]_D^{26} -96.5$ (*c* 1.70, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.97–7.94 (m, 2H), 7.54–7.52 (m, 1H), 7.45–7.44 (m, 1H), 7.35–7.29 (m, 5H), 4.85 (dd, *J* = 13.2, 10.9 Hz, 1H), 4.68 (dd, *J* = 13.2, 4.5 Hz, 1H), 4.09 (dd, *J* = 10.9, 4.5 Hz, 1H), 2.44 (s, 3H), 1.51 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 192.3, 185.7, 139.1, 136.7, 133.1, 130.7, 129.0, 128.9, 128.8, 127.2, 124.7, 87.5, 74.8, 49.4, 21.6, 21.2; HRMS (ESI) m/z 339.1339 (M+H⁺), calc. for C₁₉H₁₉N₂O₄ 339.1345.

The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 10.2 min (major) and 14.3 min (minor).

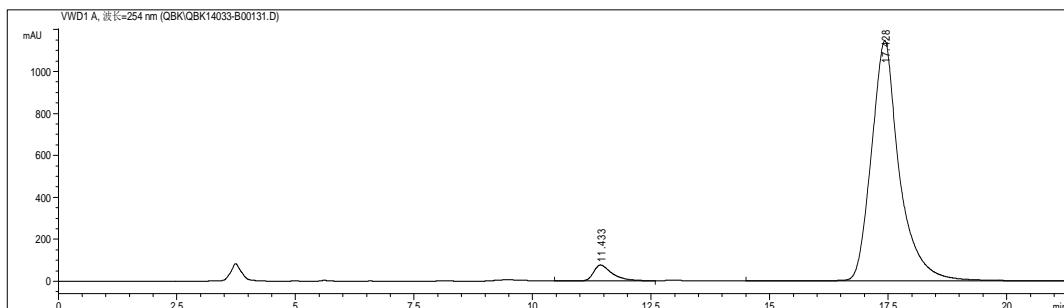


3eb, White solid; Mp 74–75 °C; 91% ee; dr >19:1; 91% yield (major diastereomer); $[\alpha]_D^{26} -74.2$ (*c* 1.70, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.93–7.90 (m, 2H), 7.64 (d, *J* = 8.4 Hz, 2H), 7.55 (br d, *J* = 7.6 Hz, 1H), 7.45 (dd, *J* = 14.6, 7.9 Hz, 3H), 4.91 (dd, *J* = 13.6, 11.1 Hz, 1H), 4.75 (dd, *J* = 13.6, 4.4 Hz, 1H), 4.18 (dd, *J* = 11.1, 4.4 Hz, 1H), 2.45 (s, 3H), 1.55 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 191.6, 185.8, 139.3, 138.6, 137.1, 132.7, 130.7, 129.7, 129.1, 127.2, 124.4, 118.0, 113.0, 86.7, 74.1, 49.2, 21.6, 21.2; HRMS (ESI) m/z 364.1297 (M+H⁺), calc. for C₂₀H₁₈N₃O₄ 364.1297.

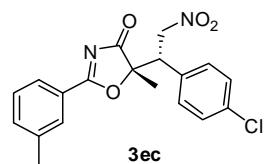
The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 75/25; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 11.5 min (minor) and 17.6 min (major).



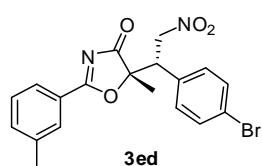
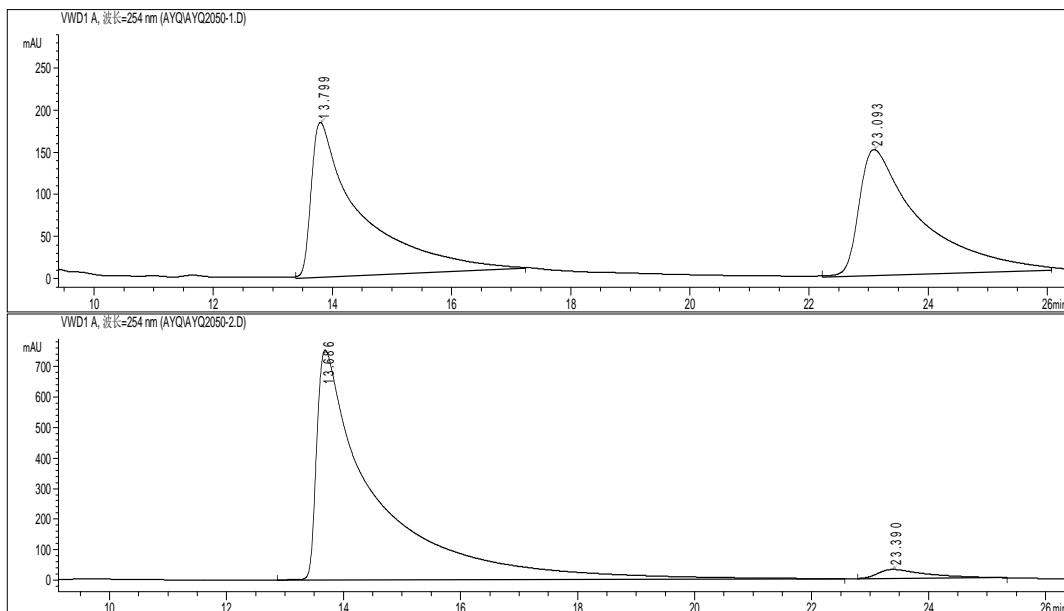
The **3eb** was obtained with 78% yield in 92% ee and >99:1 dr when the reaction was stopped after 18 hours.



3ec, White solid; Mp 55–56 °C; 94% ee; dr >19:1; 89% yield (major diastereomer); $[\alpha]_D^{26} -84.2$ (*c* 2.83, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.96–7.93 (m, 2H), 7.55–7.53 (m, 1H), 7.44 (t, *J* = 7.6 Hz, 1H), 7.33–7.27 (m, 4H), 4.82 (dd, *J* = 13.3, 11.1 Hz, 1H), 4.67 (dd, *J* = 13.3, 4.5 Hz, 1H), 4.08 (dd, *J* = 11.1, 4.5 Hz, 1H), 2.45 (s, 3H), 1.51 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 192.0, 185.7, 139.2, 136.8, 134.9, 131.7, 130.7, 130.2, 129.2, 129.0, 127.2, 125.0, 87.1, 74.6, 48.8, 21.6, 21.2; HRMS (ESI) m/z 373.0947 ($\text{M}+\text{H}^+$), calc. for $\text{C}_{19}\text{H}_{18}\text{ClN}_2\text{O}_4$ 373.0955.

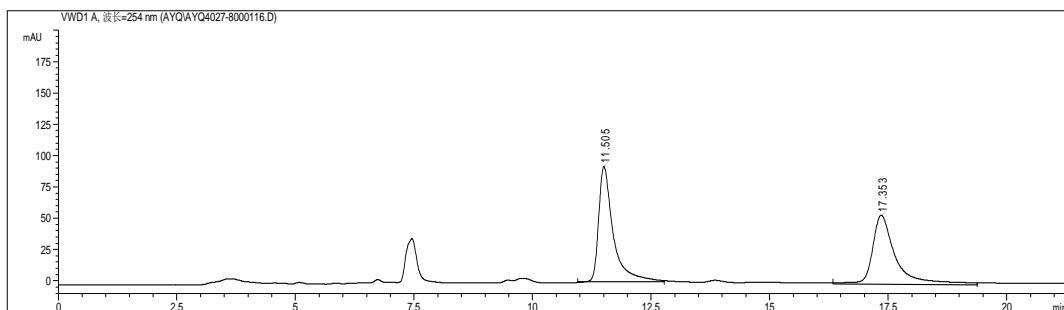


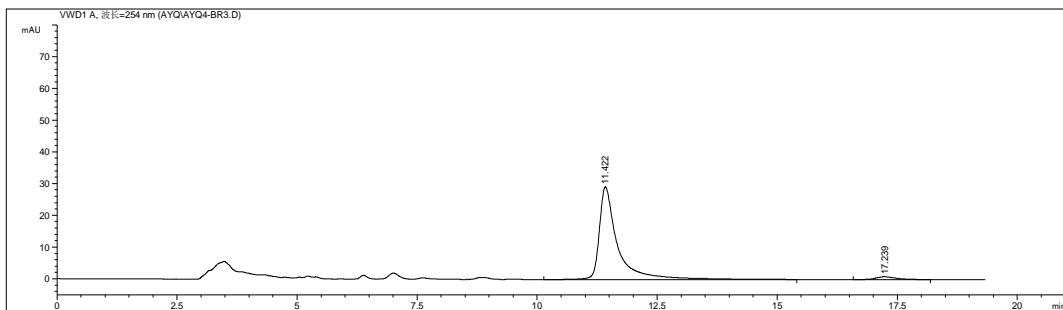
The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 85/15; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 13.7 min (major) and 23.4 min (minor).



3ed, white solid; Mp 54–55 °C; 94% ee; dr >19:1; 85% yield (major diastereomer); $[\alpha]_D^{26} -81.7$ (*c* 2.80, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.96–7.93 (m, 2H), 7.55–7.51 (m, 1H), 7.45 (dd, *J* = 16.5, 8.1 Hz, 3H), 7.21 (d, *J* = 8.4 Hz, 2H), 4.81 (dd, *J* = 13.3, 11.0 Hz, 1H), 4.67 (dd, *J* = 13.3, 4.4 Hz, 1H), 4.06 (dd, *J* = 11.0, 4.4 Hz, 1H), 2.45 (s, 3H), 1.51 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 191.9, 185.8, 139.2, 136.8, 132.3, 132.2, 130.7, 130.5, 129.1, 127.2, 124.7, 123.1, 87.1, 74.6, 48.9, 21.6, 21.2; HRMS (ESI) m/z 417.0443 (M+H⁺), calc. for C₁₉H₁₇BrN₂O₄ 417.0450.

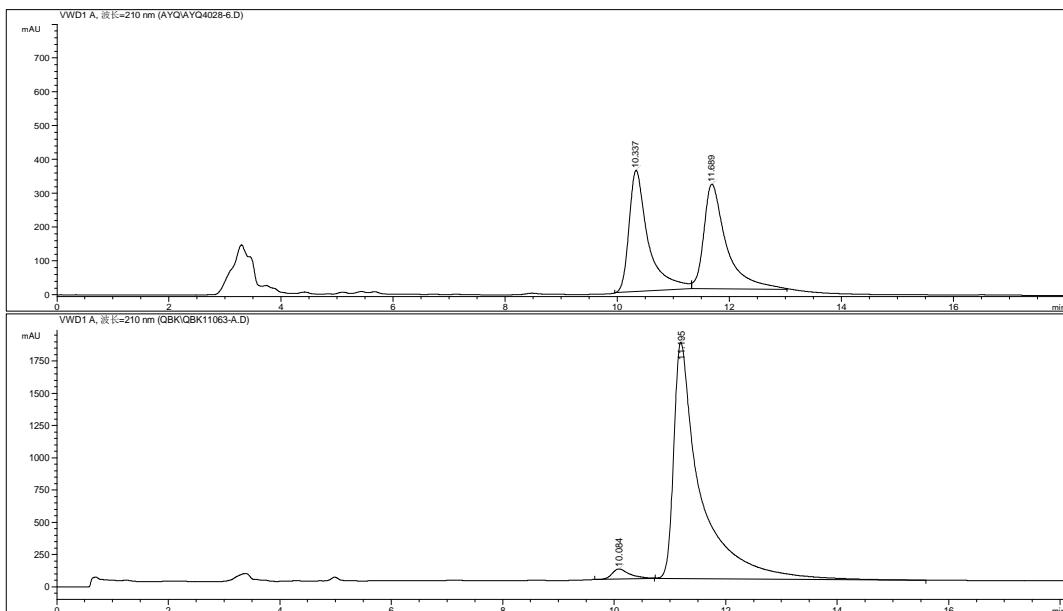
The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 75/25; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 11.4 min (major) and 17.2 min (minor).





3ee, white solid; Mp 61–62 °C; 95% ee; dr >19:1; 79% yield (major diastereomer); $[\alpha]_D^{26} -78.9$ (*c* 2.40, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.95–7.92 (m, 2H), 7.66 (m, 1H), 7.63–7.52 (m, 3H), 7.46 (dt, *J* = 11.2, 7.7 Hz, 2H), 4.89 (dd, *J* = 13.6, 11.2 Hz, 1H), 4.73 (dd, *J* = 13.6, 4.4 Hz, 1H), 4.15 (dd, *J* = 11.1, 4.4 Hz, 1H), 2.45 (s, 3H), 1.54 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 191.5, 185.8, 139.4, 137.1, 135.0, 132.7, 132.6, 130.0, 129.2, 127.3, 124.4, 118.0, 113.3, 86.7, 74.1, 49.0, 21.6, 21.2; HRMS (ESI) m/z 364.1293 (M+H⁺), calc. for C₂₀H₁₇N₃O₄ 364.1297.

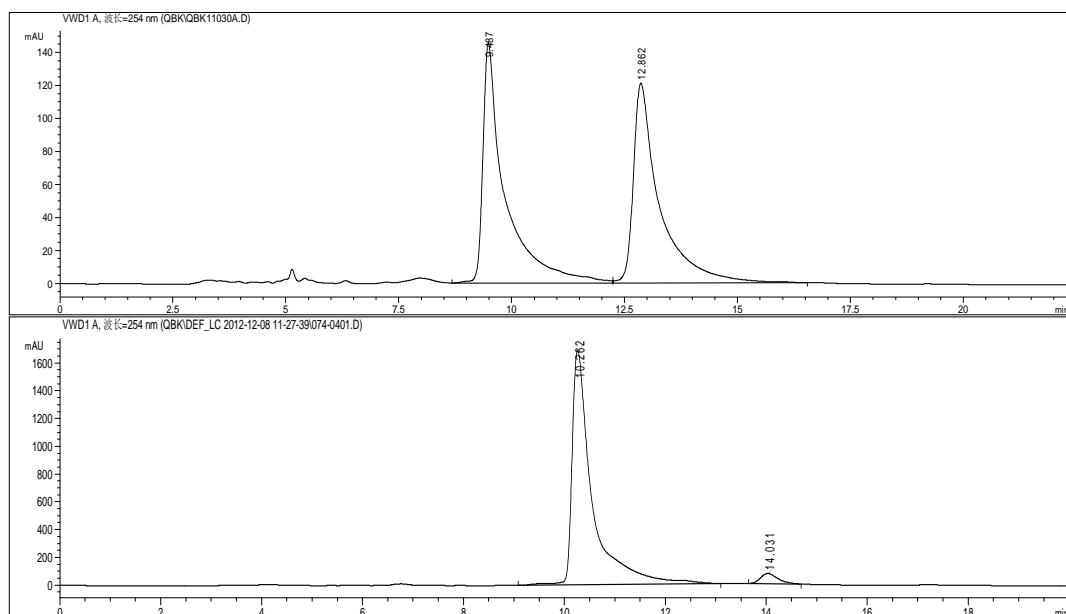
The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 75/25; flow rate 1.0 ml/min; 25 °C; 210 nm; retention time: 10.1 min (minor) and 11.2 min (major).



3ef, white solid; Mp 55–56 °C; 93% ee; dr >19:1; 87% yield (major diastereomer); $[\alpha]_D^{26} -84.2$ (*c* 2.70, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.97–7.95 (m, 2H), 7.55 (d, *J* = 7.6 Hz, 1H), 7.45 (t, *J* = 7.6 Hz, 1H), 7.34–7.28 (m, 3H), 7.25–7.21 (m, 1H), 4.82 (dd, *J* = 13.4, 11.0 Hz, 1H), 4.67 (dd, *J* = 13.4,

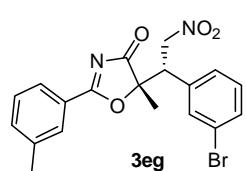
4.4 Hz, 1H), 4.07 (dd, J = 11.0, 4.4 Hz, 1H), 2.45 (s, 3H), 1.52 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 191.9, 185.8, 139.2, 136.9, 135.2, 134.8, 130.7, 130.3, 129.3, 129.2, 129.1, 127.3, 126.8, 124.6, 87.0, 74.4, 49.0, 21.6, 21.2; HRMS (ESI) m/z 373.0944 ($\text{M}+\text{H}^+$), calc. for $\text{C}_{19}\text{H}_{18}\text{N}_2\text{O}_4\text{Cl}$ 373.0955.

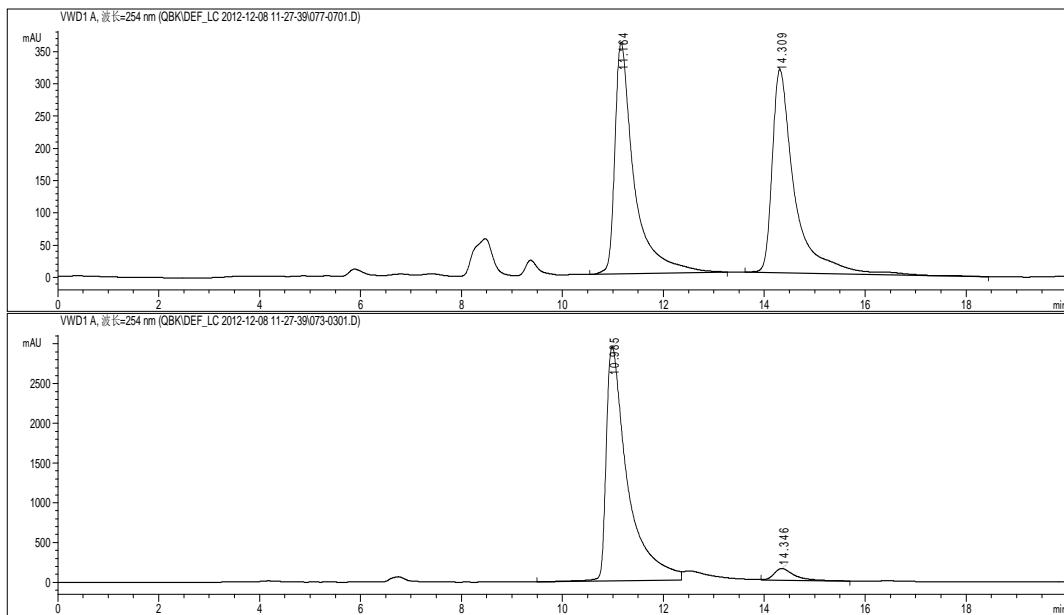
The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 75/25; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 10.3 min (major) and 14.0 min (minor).

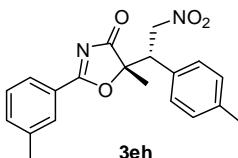


3eg, white solid; Mp 108–109 °C; 91% ee; dr >19:1; 91% yield (major diastereomer); $[\alpha]_D^{26} -152.3$ (c 0.22, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.97–7.94 (m, J = 9.3 Hz, 2H), 7.55 (d, J = 7.6 Hz, 1H), 7.50 (t, J = 1.6 Hz, 1H), 7.44 (ddd, J = 6.6, 4.5, 4.0 Hz, 2H), 7.28 (m, 1H), 7.22 (t, J = 7.8 Hz, 1H), 4.82 (dd, J = 13.4, 11.0 Hz, 1H), 4.67 (dd, J = 13.5, 4.4 Hz, 1H), 4.06 (dd, J = 11.0, 4.4 Hz, 1H), 2.45 (s, 3H), 1.52 (s, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 191.8, 185.7, 139.2, 136.8, 135.5, 132.3, 132.0, 130.6, 130.5, 129.1, 127.3, 127.1, 124.6, 122.9, 87.0, 74.4, 49.0, 21.6, 21.2; HRMS (ESI) m/z 417.0443 ($\text{M}+\text{H}^+$), calc. for $\text{C}_{19}\text{H}_{18}\text{N}_2\text{O}_4\text{Br}$ 417.0450.

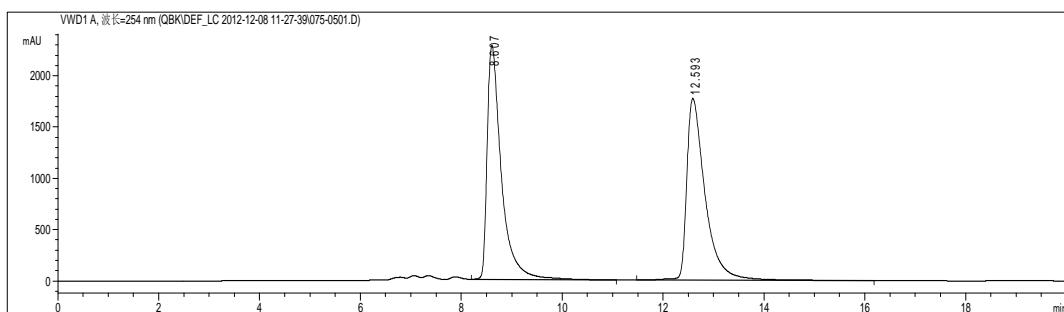
The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 75/25; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 11.0 min (major) and 14.3 min (minor).

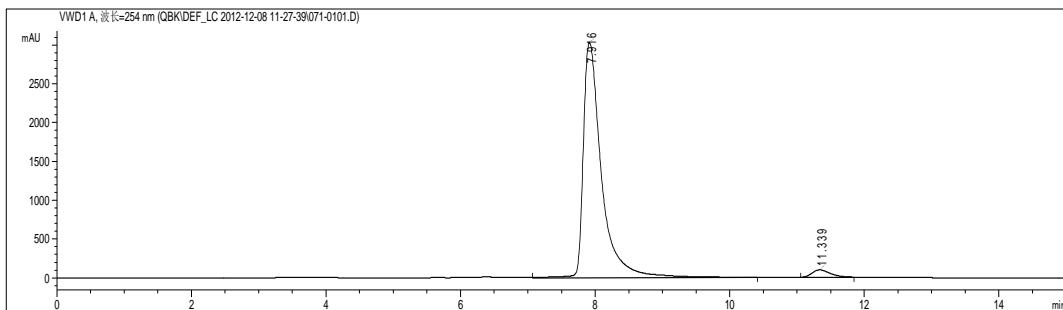




3eh, white form; 94% ee; dr >19:1; 87% yield (major diastereomer);

 $[\alpha]_D^{26} -103.5$ (*c* 2.20, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.98–7.96 (m, Hz, 2H), 7.54 (d, *J* = 7.6 Hz, 1H), 7.44 (t, *J* = 7.6 Hz, 1H), 7.22 (d, *J* = 8.1 Hz, 2H), 7.14 (d, *J* = 7.9 Hz, 2H), 4.81 (dd, *J* = 13.0, 11.0 Hz, 1H), 4.64 (dd, *J* = 13.0, 4.5 Hz, 1H), 4.04 (dd, *J* = 11.0, 4.5 Hz, 1H), 2.45 (s, 3H), 2.30 (s, 3H), 1.49 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 192.4, 185.7, 139.2, 138.7, 136.7, 130.7, 130.0, 129.7, 129.0, 128.7, 127.2, 124.9, 87.6, 74.9, 49.0, 21.6, 21.2, 21.1; HRMS (ESI) *m/z* 353.1497 (M+H⁺), calc. for C₂₀H₂₁N₂O₄ 353.1501.

The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 75/25; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 7.9 min (major) and 11.3 min (minor).

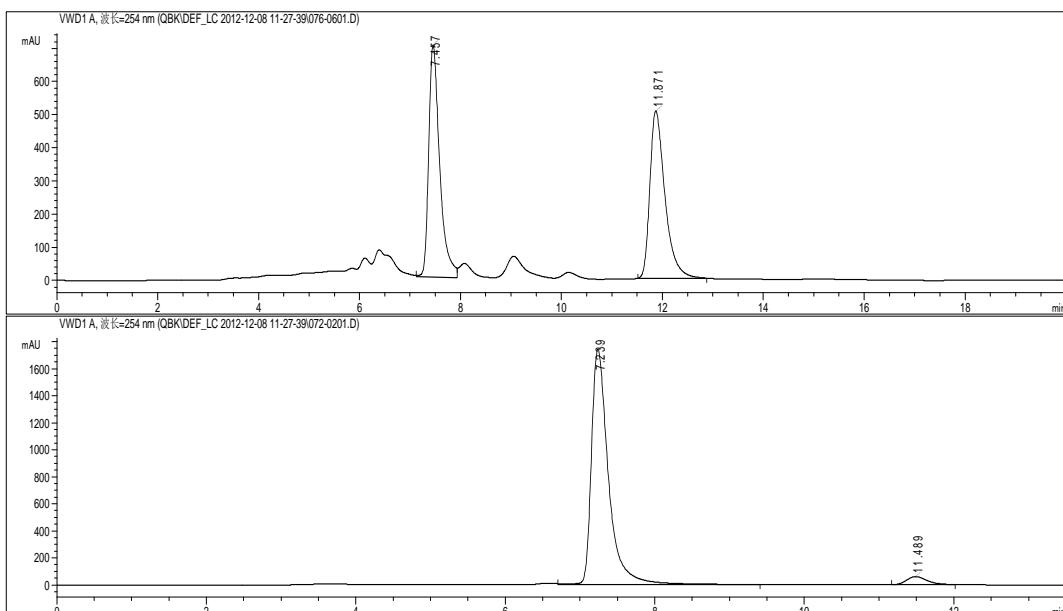




3ei

3ei, white solid; Mp 35–36 °C; 92% ee; dr >19:1; 99% yield; $[\alpha]_D^{26}$ -122.9 (*c* 0.80, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.97–7.94 (m, 2H), 7.53 (d, *J* = 7.6 Hz, 1H), 7.44 (t, *J* = 7.6 Hz, 1H), 7.24 (d, *J* = 8.3 Hz, 2H), 7.18 (d, *J* = 8.2 Hz, 2H), 4.81 (dd, *J* = 13.1, 11.0 Hz, 1H), 4.64 (dd, *J* = 13.1, 4.5 Hz, 1H), 4.06 (dd, *J* = 10.9, 4.5 Hz, 1H), 2.84 (dq, *J* = 13.9, 6.9 Hz, 1H), 2.45 (s, 3H), 1.50 (s, 3H), 1.17 (dd, *J* = 6.9, 2.9 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 192.4, 185.8, 149.5, 139.1, 136.6, 130.7, 130.3, 129.0, 128.8, 127.2, 127.0, 124.9, 87.7, 74.8, 49.1, 33.7, 23.7, 21.6, 21.2. HRMS (ESI) m/z 381.1804 (M+H⁺), calc. for C₂₂H₂₅N₂O₄ 381.1814.

The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 75/25; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 7.2 min (major) and 11.5 min (minor).

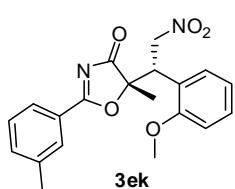
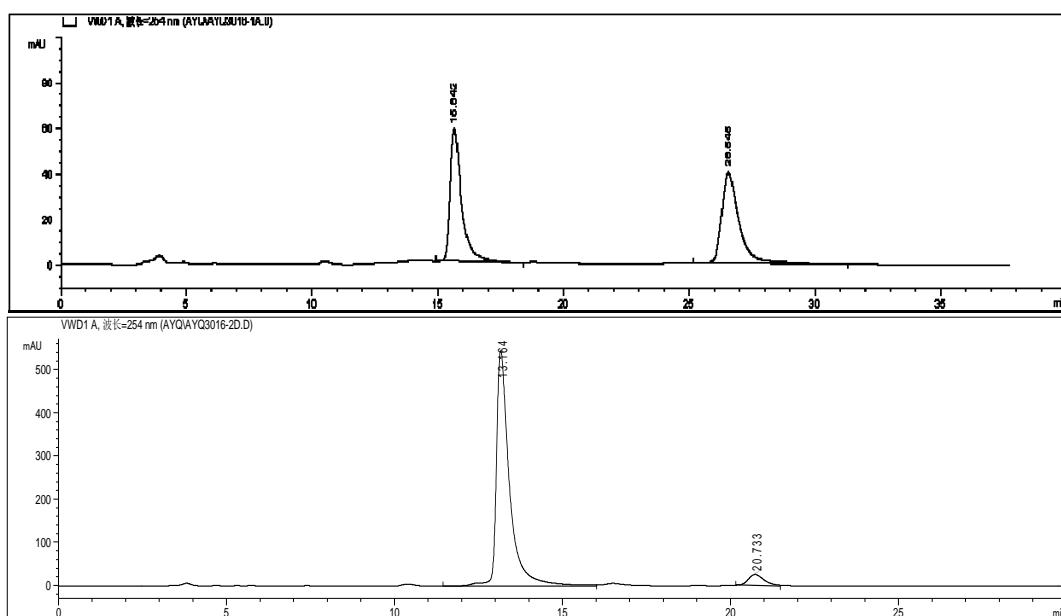


3ej

3ej, white solid; Mp 128–129 °C; 90% ee; dr >19:1; 99% yield; $[\alpha]_D^{26}$ -114.9 (*c* 1.68, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.98–7.96 (m, 2H), 7.54 (d, *J* = 7.6 Hz, 1H), 7.44 (t, *J* = 7.6 Hz, 1H), 7.22 (d, *J* =

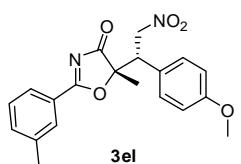
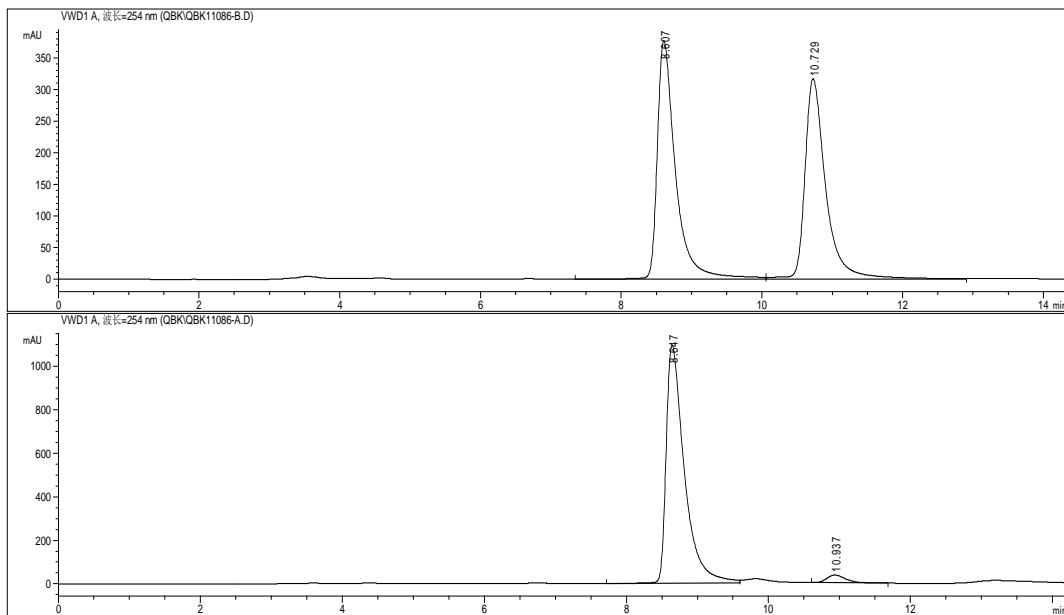
8.1 Hz, 2H), 7.14 (d, J = 7.9 Hz, 2H), 4.81 (dd, J = 13.0, 11.0 Hz, 1H), 4.64 (dd, J = 13.0, 4.5 Hz, 1H), 4.04 (dd, J = 11.0, 4.5 Hz, 1H), 2.45 (s, 3H), 2.30 (s, 3H), 1.49 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 192.4, 185.7, 139.2, 138.7, 136.7, 130.7, 130.0, 129.7, 129.0, 128.7, 127.2, 124.9, 87.6, 74.9, 49.0, 21.6, 21.2, 21.1; HRMS (ESI) m/z 353.1504 ($\text{M}+\text{H}^+$), calc. for $\text{C}_{20}\text{H}_{21}\text{N}_2\text{O}_4$ 353.1501.

The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 13.2 min (major) and 20.7 min (minor).



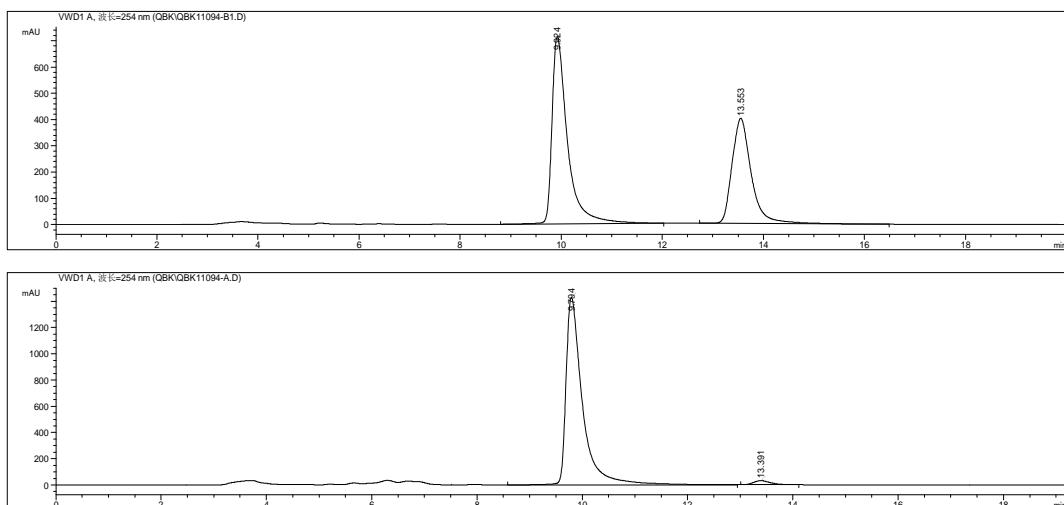
3ek, white solid; Mp 133–135 °C; 94% ee; dr >19:1; 99% yield; $[\alpha]_D^{26}$ −100.0 (c 2.50, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.95 (m, 2H), 7.52–7.40 (m, 2H), 7.27–7.23 (m, 2H), 6.91 (dd, J = 37.4, 7.3 Hz, 2H), 4.76 (d, J = 49.4 Hz, 3H), 3.85 (s, 3H), 2.43 (s, 3H), 1.52 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 192.4, 185.4, 157.9, 139.0, 136.4, 130.6, 129.9, 128.9, 127.1, 125.1, 121.7, 120.9, 111.2, 74.4, 55.6, 21.2; HRMS (ESI) m/z 369.1443 ($\text{M}+\text{H}^+$), calc. for $\text{C}_{20}\text{H}_{21}\text{N}_2\text{O}_5$ 369.1450.

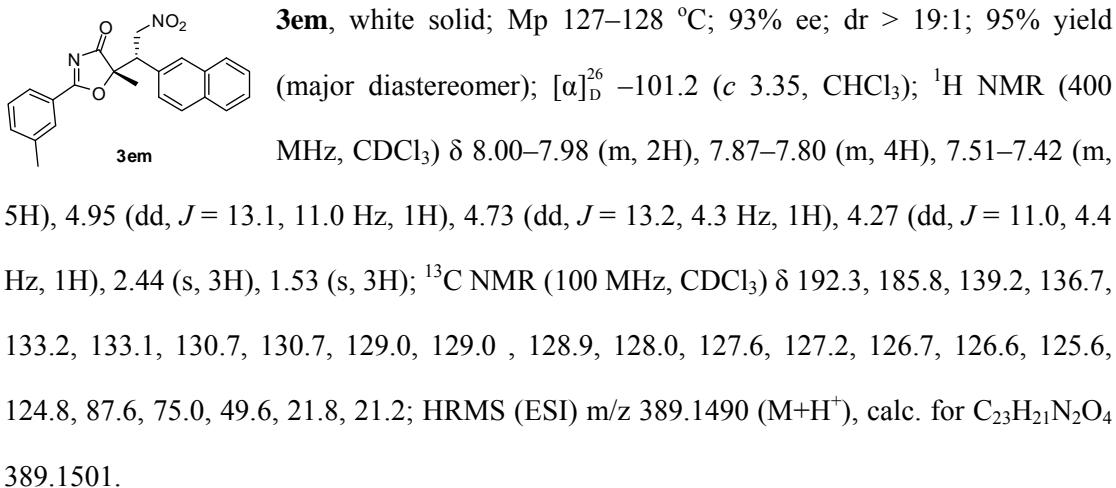
The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 75/25; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 8.6 min (major) and 10.9 min (minor).



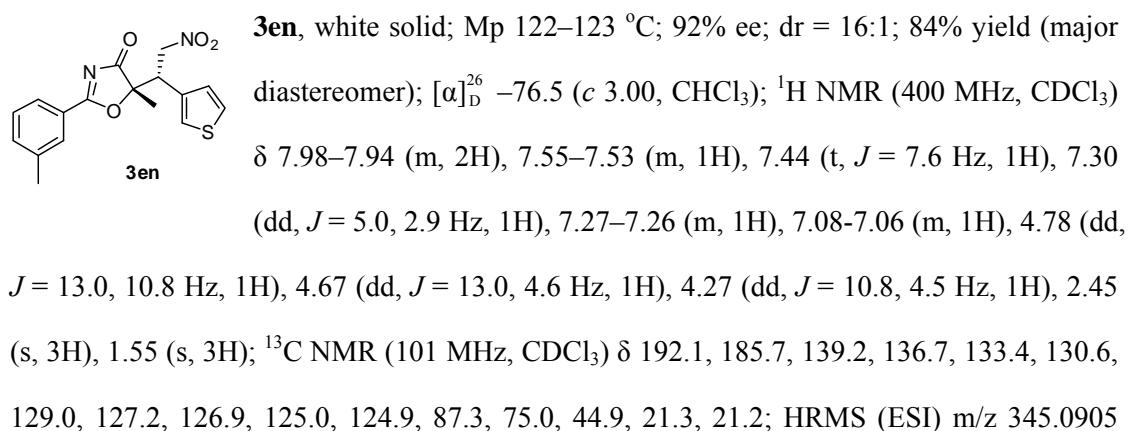
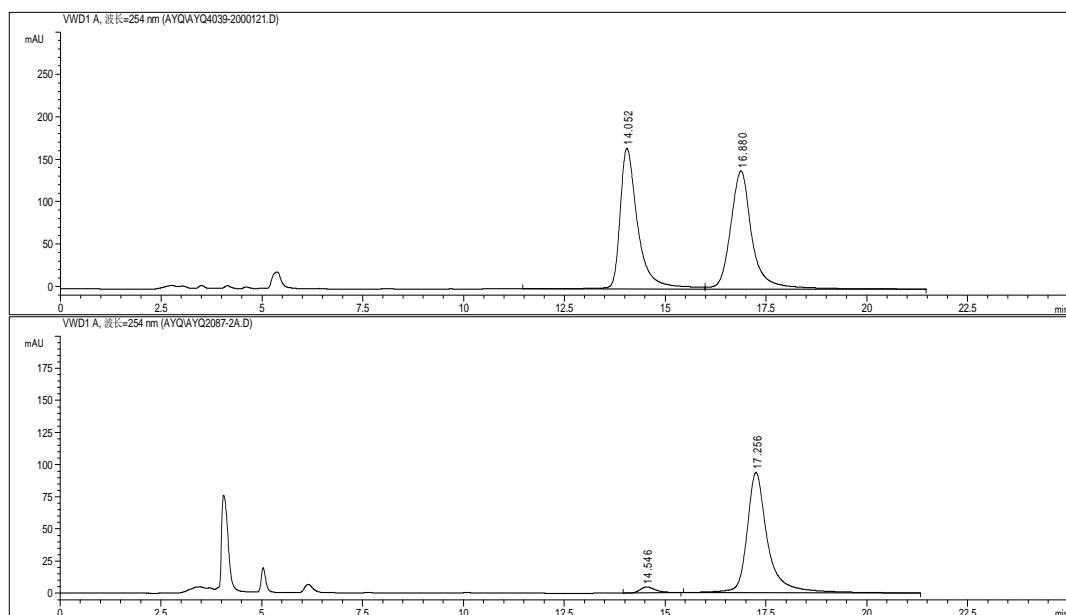
3el, white solid; Mp 32–33 °C; 96% ee; dr >19:1; 96% yield (major diastereomer); $[\alpha]_D^{26} -87.7$ (*c* 2.97, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.97–7.94 (m, 2H), 7.53–7.52 (m, 1H), 7.43 (t, *J* = 7.6 Hz, 1H), 7.25–7.22 (m, 2H), 6.85 (d, *J* = 8.7 Hz, 2H), 4.81 (dd, *J* = 12.9, 11.1 Hz, 1H), 4.65 (dd, *J* = 13.0, 4.5 Hz, 1H), 4.03 (dd, *J* = 11.0, 4.5 Hz, 1H), 3.75 (s, 3H), 2.44 (s, 3H), 1.50 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 192.4, 185.7, 159.7, 139.1, 136.6, 130.7, 130.0, 129.0, 127.2, 124.9, 124.9, 114.3, 87.7, 75.0, 55.2, 48.8, 21.6, 21.2; HRMS (ESI) m/z 369.1444 (M+H⁺), calc. for C₂₀H₂₁N₂O₅ 369.1450.

The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 75/25; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 9.7 min (major) and 13.4 min (minor).



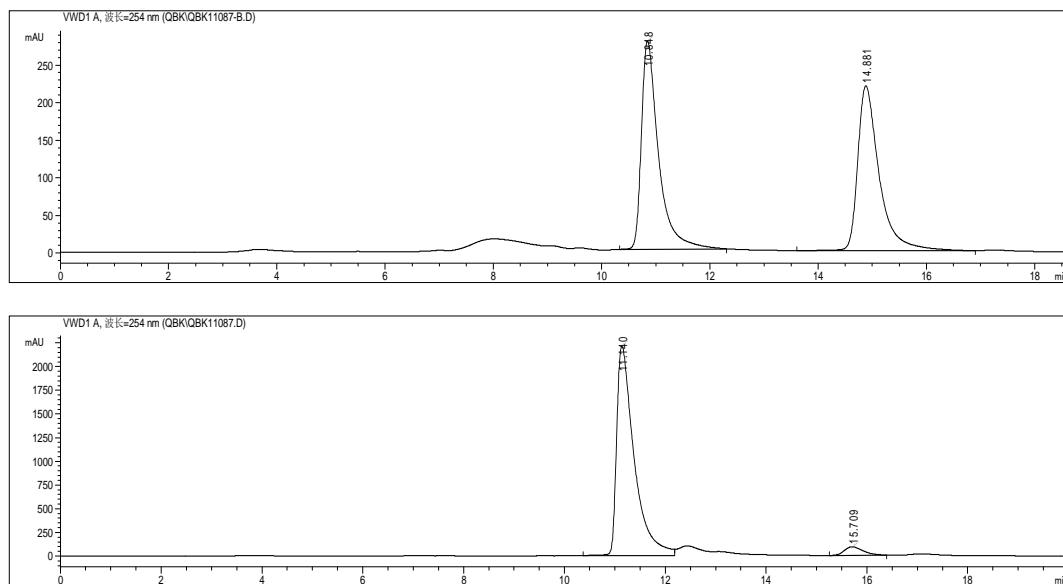


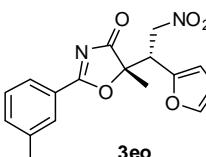
The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 75/25; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 14.5 min (major) and 17.3 min (minor).



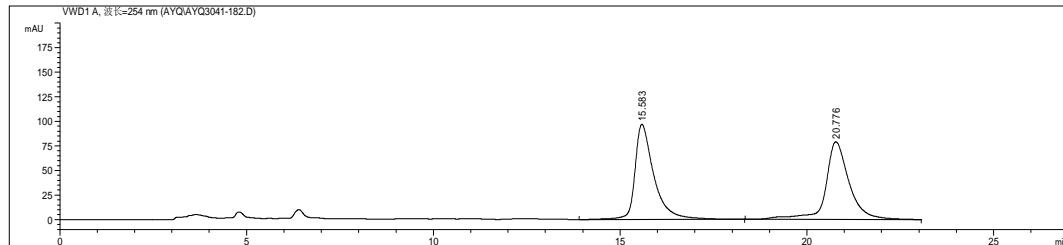
(M+H⁺), calc. for C₁₇H₁₇N₂O₄S 345.0909.

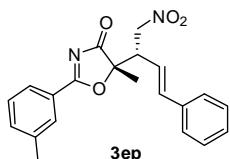
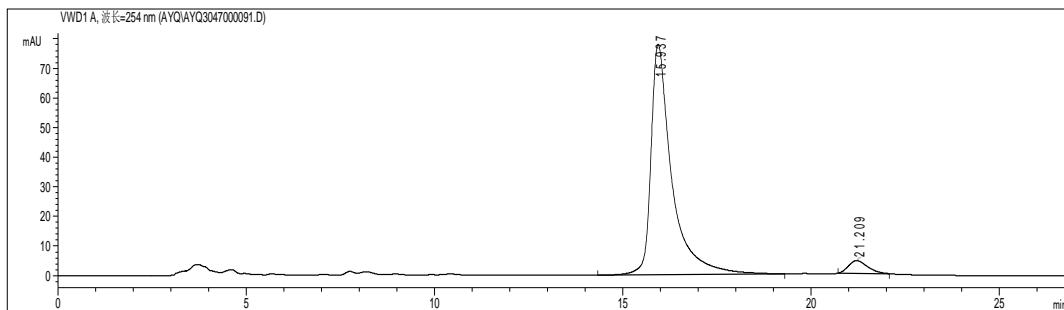
The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 75/25; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 11.1 min (major) and 15.7 min (minor).



3eo  white solid; Mp 111–112 °C; 90% ee; dr = 10:1; 84% yield (major diastereomer); $[\alpha]_D^{26} -75.9$ (*c* 1.43, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.99–7.97 (m, 2H), 7.53 (br d, *J* = 7.6 Hz, 1H), 7.44 (t, *J* = 7.6 Hz, 1H), 7.37 (d, *J* = 1.1 Hz, 1H), 6.39–6.25 (m, 2H), 4.88 (dd, *J* = 13.4, 10.5 Hz, 1H), 4.65 (dd, *J* = 13.4, 4.4 Hz, 1H), 4.25 (dd, *J* = 10.5, 4.4 Hz, 1H), 2.45 (s, 3H), 1.58 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 191.6, 185.8, 146.8, 143.4, 139.1, 136.7, 130.7, 129.0, 127.4, 124.9, 110.6, 110.5, 86.2, 73.0, 43.3, 21.2, 21.1; HRMS (ESI) m/z 329.1135 (M+H⁺), calc. for C₁₇H₁₇N₂O₅ 329.1137.

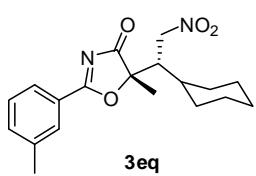
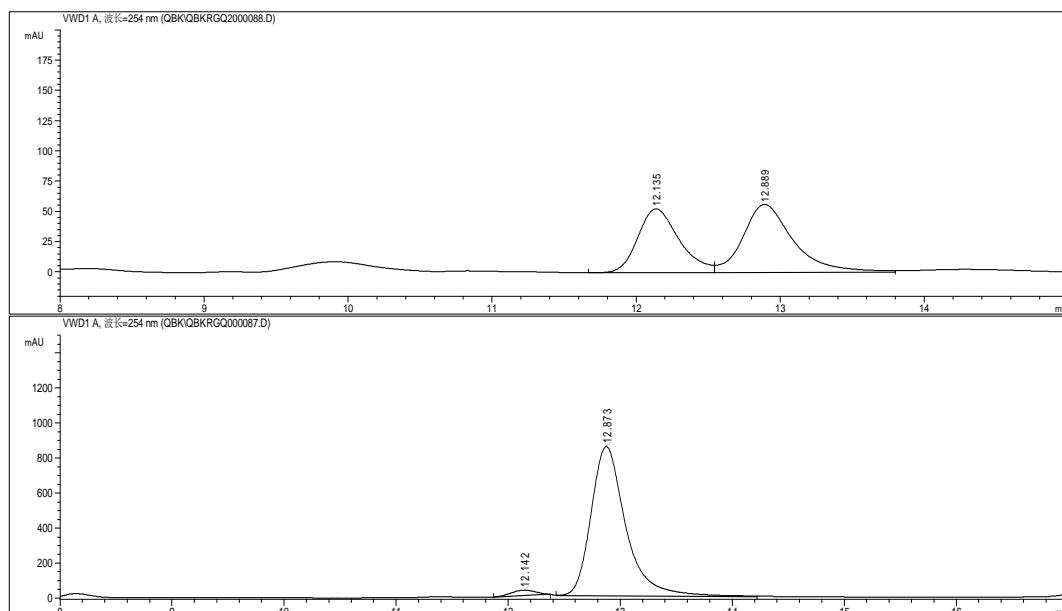
The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 15.9 min (major) and 21.2 min (minor).





3ep, yellow solid; Mp 117–118 °C; 95% ee; dr >19:1; 89% yield (major diastereomer); $[\alpha]_D^{26} -77.9$ (*c* 2.45, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.02–8.00 (m, 2H), 7.53 (br d, *J* = 7.7 Hz, 1H), 7.45–7.42 (m, 1H), 7.34–7.25 (m, 5H), 6.66 (d, *J* = 15.7 Hz, 1H), 6.02 (dd, *J* = 15.7, 9.7 Hz, 1H), 4.52 (qd, *J* = 12.2, 7.5 Hz, 2H), 3.63 (td, *J* = 9.9, 4.7 Hz, 1H), 2.44 (s, 3H), 1.65 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 191.9, 185.9, 139.1, 138.3, 136.7, 135.5, 130.7, 129.0, 128.6, 128.5, 127.4, 126.7, 124.9, 120.0, 86.8, 74.9, 47.8, 21.2; HRMS (ESI) m/z 365.1496 (M+H⁺), calc. for C₂₁H₂₀N₂O₄ 365.1501.

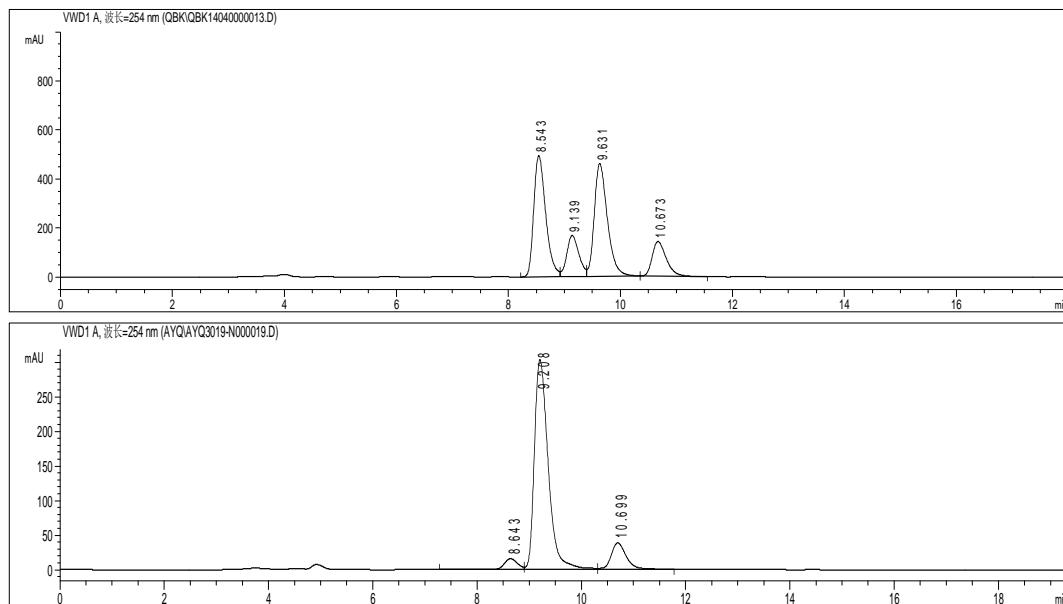
The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 12.1 min (major) and 12.9 min (minor).



3eq, as a slightly sticky white solid; 78% ee; dr >19:1; 42% yield (major diastereomer); $[\alpha]_D^{26} -67.4$ (*c* 0.35, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.01–7.96 (m, 2H), 7.54 (d, *J* = 7.5 Hz, 1H), 7.45 (t,

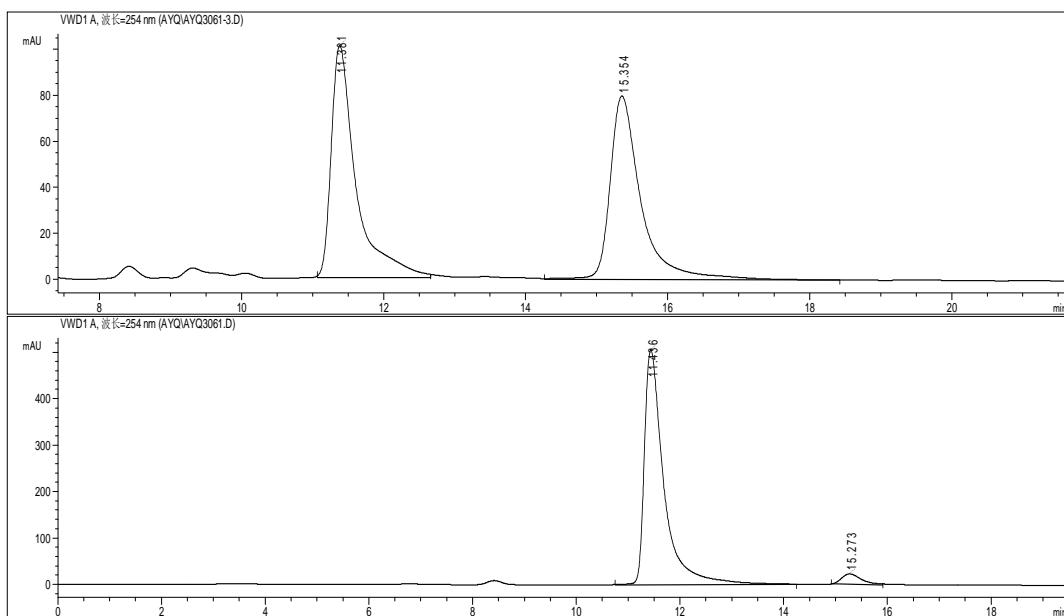
J = 7.7 Hz, 1H), 4.64 (dd, *J* = 14.2, 6.5 Hz, 1H), 4.48 (dd, *J* = 14.2, 5.1 Hz, 1H), 2.45 (s, 3H), 1.88 (t, *J* = 11.8 Hz, 1H), 1.62 (m, 4H), 1.35–0.81 (m, 10H); ^{13}C NMR (101 MHz, CDCl_3) δ 192.9, 185.4, 139.2, 136.5, 130.8, 129.0, 127.3, 125.1, 87.1, 72.1, 48.1, 36.20, 32.2, 28.2, 26.6, 26.3, 25.8, 21.2; HRMS (ESI) m/z 345.1808 ($\text{M}+\text{H}^+$), calc. for $\text{C}_{19}\text{H}_{25}\text{N}_2\text{O}_4$ 345.1814.

The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 8.6 min (minor), 9.2 min (major) and 10.7 min (minor).



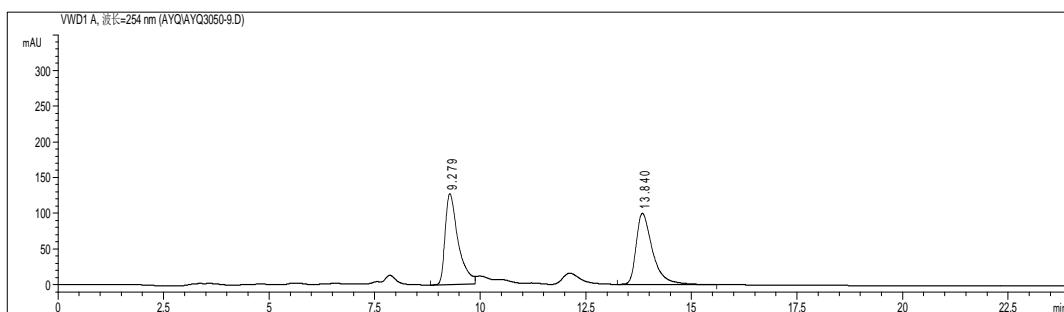
3ha, white solid; Mp 113–114 °C; 91% ee; dr >19:1; 94% yield; $[\alpha]_D^{26}$ −76.8 (*c* 1.93, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.98–7.95 (m, 2H), 7.55–7.53 (br d, *J* = 7.7 Hz, 1H), 7.44 (t, *J* = 7.6 Hz, 1H), 7.30 (dt, *J* = 8.2, 3.9 Hz, 5H), 4.85 (dd, *J* = 13.1, 11.2 Hz, 1H), 4.68 (dd, *J* = 13.1, 4.3 Hz, 1H), 4.11 (dd, *J* = 11.1, 4.3 Hz, 1H), 2.45 (s, 3H), 1.96 (dq, *J* = 14.7, 7.4 Hz, 1H), 1.82 (dq, *J* = 14.6, 7.3 Hz, 1H), 0.80 (t, *J* = 7.4 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 192.0, 186.3, 139.2, 136.7, 133.3, 130.7, 129.0, 128.9, 128.9, 128.8, 127.2, 124.6, 91.1, 74.9, 48.9, 28.3, 21.2, 6.9; HRMS (ESI) m/z 375.1317 ($\text{M}+\text{Na}^+$), calc. for $\text{C}_{20}\text{H}_{20}\text{N}_2\text{O}_4\text{Na}$ 375.1321.

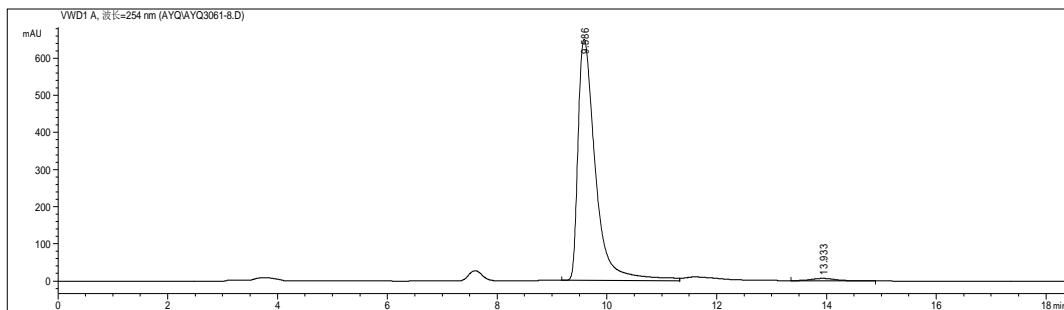
The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 11.4 min (major) and 15.3 min (minor).

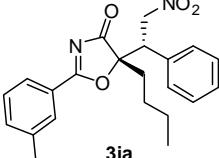


3ia, white solid; Mp 134–135 °C; 98% ee; dr >19:1; 70% yield; $[\alpha]_D^{26}$ –54.1 (*c* 0.50, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.90–7.89 (m, 2H), 7.52–7.50 (m, 1H), 7.41 (t, *J* = 7.8 Hz, 1H), 7.34–7.32 (m, 2H), 7.29–7.27 (m, 1H), 7.25–7.20 (m, 2H), 5.01 (dd, *J* = 13.1, 11.6 Hz, 1H), 4.70 (dd, *J* = 13.1, 3.9 Hz, 1H), 4.29 (dd, *J* = 11.5, 3.9 Hz, 1H), 2.43 (s, 3H), 2.28–2.15 (m, 1H), 1.09 (d, *J* = 6.8 Hz, 3H), 1.00 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 191.9, 186.2, 139.1, 136.5, 133.0, 130.6, 128.9, 128.8, 128.8, 127.0, 124.7, 92.8, 74.8, 46.7, 32.1, 21.2, 16.1, 15.8; HRMS (ESI) m/z 367.1656 (M+H⁺), calc. for C₂₁H₂₃N₂O₄ 367.1658.

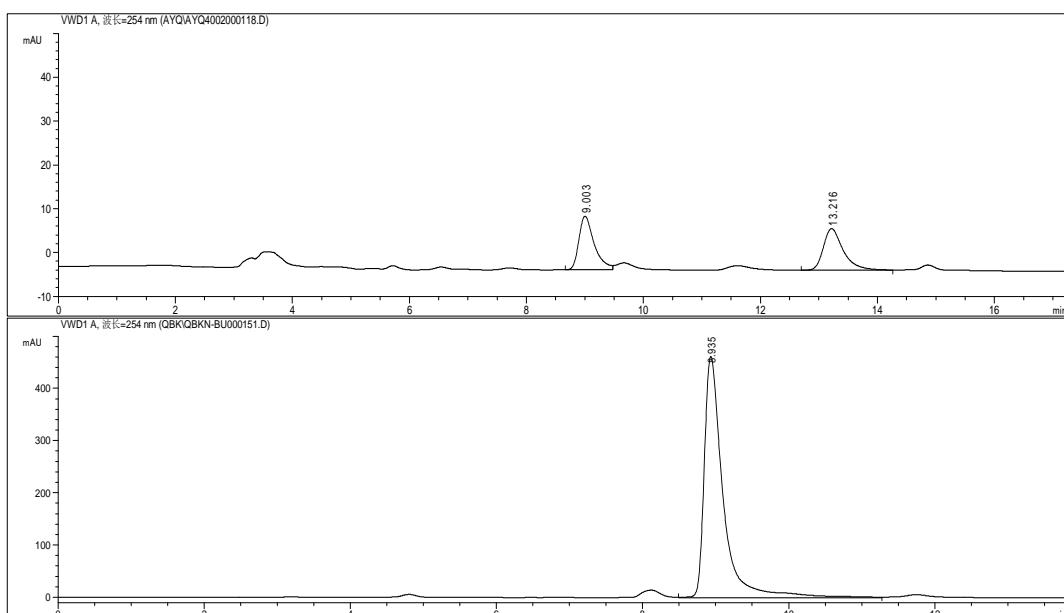
The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 9.6 min (major) and 13.9 min (minor).



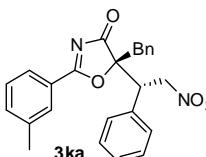


3ja, white solid; Mp 148.9–149.2 °C; 99% ee; dr >19:1; 99% yield;

 $[\alpha]_D^{26} -77.3$ (*c* 0.90, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.97–7.94 (m, 2H), 7.54–7.53 (m, 1H), 7.44 (t, *J* = 7.7 Hz, 1H), 7.32–7.28 (m, 5H), 4.87 (dd, *J* = 13.1, 11.1 Hz, 1H), 4.69 (dd, *J* = 13.1, 4.3 Hz, 1H), 4.10 (dd, *J* = 7.6, 3.4 Hz, 1H), 2.45 (s, 3H), 1.95–1.86 (m, 1H), 1.83–1.72 (m, 1H), 1.24–1.11 (m, 4H), 0.78 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 192.1, 186.2, 139.2, 136.7, 133.2, 130.7, 129.0, 128.9, 128.8, 127.2, 124.7, 90.7, 74.9, 49.1, 34.7, 24.6, 22.3, 21.2, 13.7; HRMS (EI) m/z 381.1806 (M+H⁺), calc. for C₂₂H₂₅N₂O₄ 381.1814.

The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 ml/min; 25 °C; 254 nm; retention time: 8.9 min (major) and 13.0 min (minor).

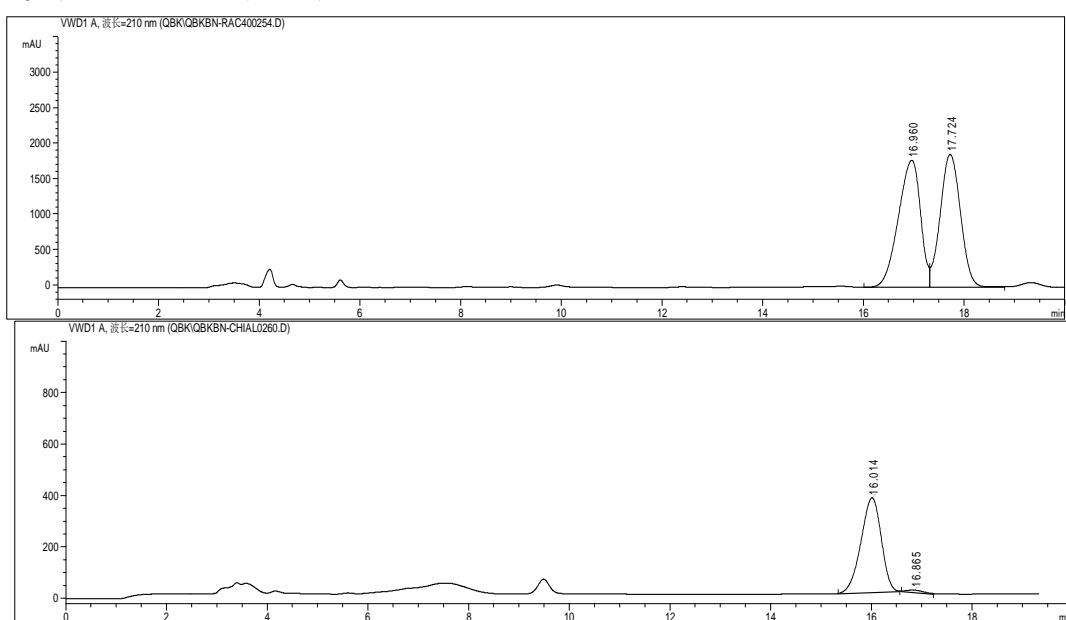


3ka, white solid; Mp 135–136 °C; 95% ee; dr >19:1; 85% yield; $[\alpha]_D^{26}$ −70.0 (*c* 0.10, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.81–7.79 (m, 2H), 7.49–7.47 (m, 1H), 7.41–7.32 (m, 6H), 7.11–7.06 (m, 5H), 4.88 (dd, *J* =



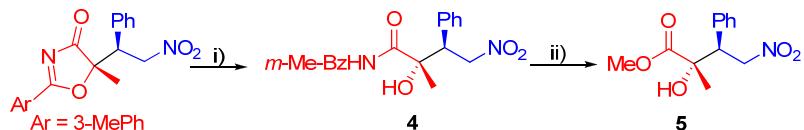
13.0, 11.3 Hz, 1H), 4.72 (dd, J = 13.1, 4.2 Hz, 1H), 4.22 (dd, J = 11.3, 4.2 Hz, 1H), 3.18 (d, J = 14.0 Hz, 1H), 3.02 (d, J = 14.0 Hz, 1H), 2.40 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 191.4, 185.9, 139.0, 136.5, 133.3, 131.9, 130.5, 129.9, 129.1, 128.9, 128.9, 128.4, 127.7, 126.8, 124.6, 90.5, 75.0, 49.0, 41.1, 21.2; HRMS (ESI) m/z 415.1662 ($\text{M}+\text{H}^+$), calc. for $\text{C}_{25}\text{H}_{23}\text{N}_2\text{O}_4$ 415.1658.

The ee was determined by HPLC analysis. CHIRALCEL ADH (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 85/15; flow rate 1.0 ml/min; 25 °C; 210 nm; retention time: 16.0 min (major) and 16.9 min (minor).



4. Synthesis and Characterization of 5 and 6

a) Synthesis and Characterization of 5

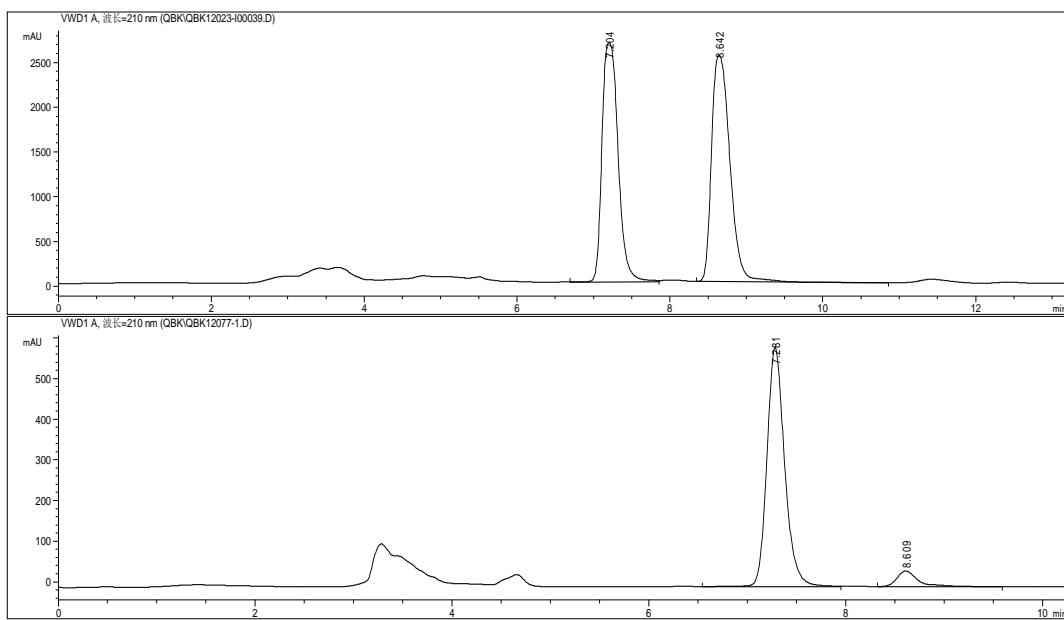


i) 0.5 mL NaOH aqueous solution (1.0 N, 5.0 equiv.) was added dropwise to a stirred solution of **3ea** (33.8 mg, 0.10 mmol, 1.0 equiv.) in THF (3mL) at -15 °C. After stirring at the same temperature for 6 hours, 1.5 mL HCl aqueous solution (2.0 N) was added to the reaction mixture. Then the mixture was extracted three times with EtOAc, and the combined organic phase was washed with brine, dried (Na_2SO_4) and concentrated under reduced pressure. The residue was purified by silica gel chromatography (PE/EtOAc = 2/1), affording product **4** (26.0 mg, 73% yield) as a white solid.

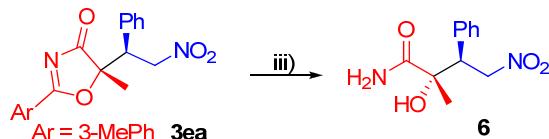
ii) To a solution of **4** (17.8 mg, 0.05 mmol) in MeOH (0.5 mL) was added $\text{Er}(\text{OTf})_3$ (30.7 mg, 0.05 mmol). The resulting mixture was stirred for 24 hours at 60 °C. The solvent was removed under reduced pressure, and the residue was purified by flash chromatography on silica gel (PE/EtOAc = 5/1) to afford compound **5** (9.0 mg) as a white solid in 71% yield and 94% ee.

5 White solid, Mp 148.2–148.7 °C; 94% ee; 52% yield (two steps); $[\alpha]_{D}^{26}$ +150.0 (*c* 0.20, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.34 (m, 5H), 4.85 (dd, *J* = 13.1, 9.9 Hz, 1H), 4.60 (dd, *J* = 13.2, 4.7 Hz, 1H), 3.90 (d, *J* = 4.8 Hz, 1H), 3.87 (s, 3H), 3.34 (s, 1H), 1.20 (s, 3H); ^{13}C NMR (100 MHz, DMSO) δ 175.0, 135.8, 129.5, 128.1, 127.7, 76.8, 75.3, 52.3, 50.7, 24.0; HRMS (EI) *m/z* 254.1031 ($\text{M}+\text{H}^+$), calc. for $\text{C}_{12}\text{H}_{16}\text{NO}_5$ 254.1028.

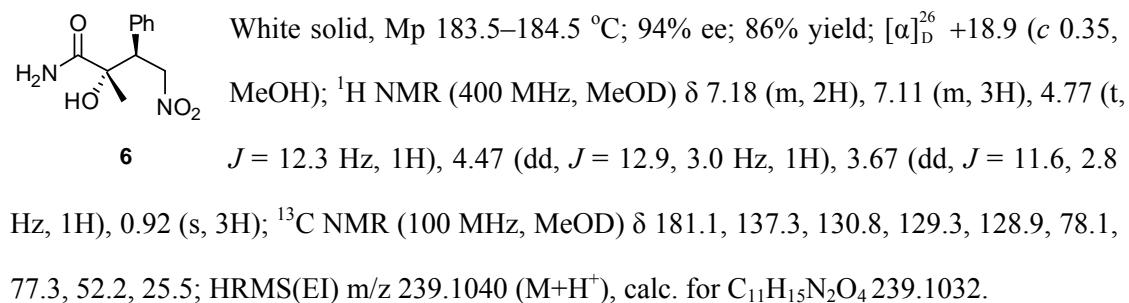
The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 ml/min; 25 °C; 210 nm; retention time: 7.3 min (major), 8.6 min (minor).



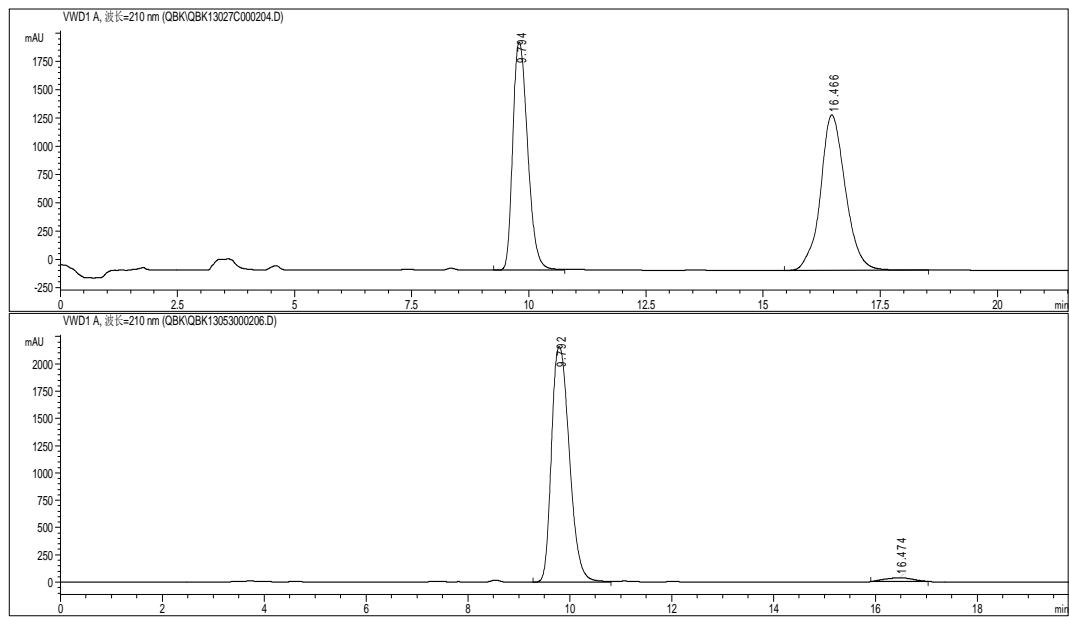
b) Synthesis and Characterization of **6**



To a solution of **3ea** (33.8 mg, 0.1 mmol) in EtOH (0.5 mL) was added 0.5 mL NaOH aqueous solution (2.0 N, 10 equiv.). The resulting mixture was stirred for 1 hours at 60 °C , then the reaction mixtures were directly loaded onto silica gel column, followed by gradient elution with PE/EtOAc (20/1–2/1 ratio) and EtOAc/MeOH (100/1–10/1 ratio). Removing the solvent in *vacuo*, afforded products **6** (20.5 mg) as a white solid in 86% yield and 94% ee.

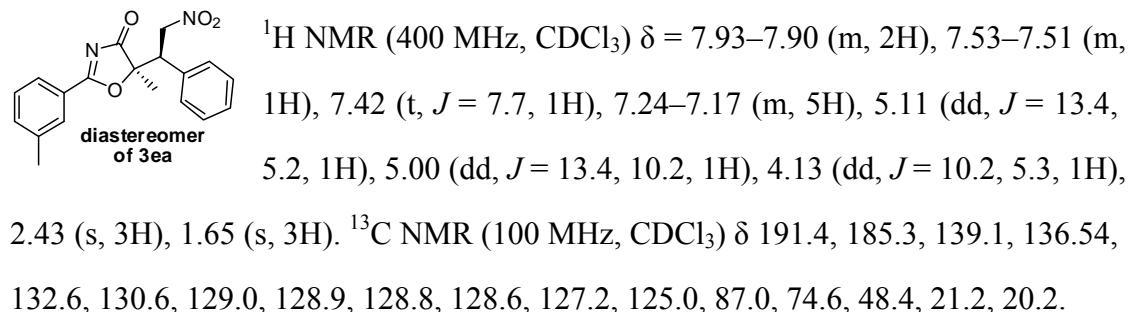


The *ee* was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/20; flow rate 1.0 ml/min; 25 °C; 210 nm; retention time: 9.8 min (major), 16.5 min (minor).



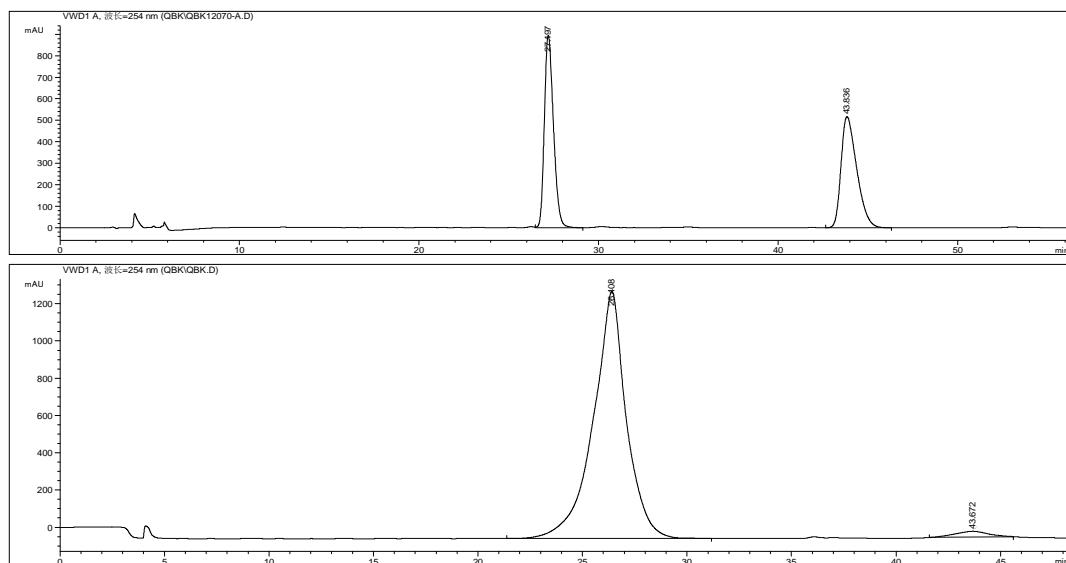
5. Determination of the Absolute Configuration

a. From ^1H NMR of **3ea**, chemical shifts and coupling constants of all peaks are same as literature¹ reported, indicating that the obtained major diastereomer of **3ea** has the same relative configuration. We also isolated another diastereomer and checked with ^1H NMR as well as ^{13}C NMR, and the results were different which confirmed our proposed relative configuration.



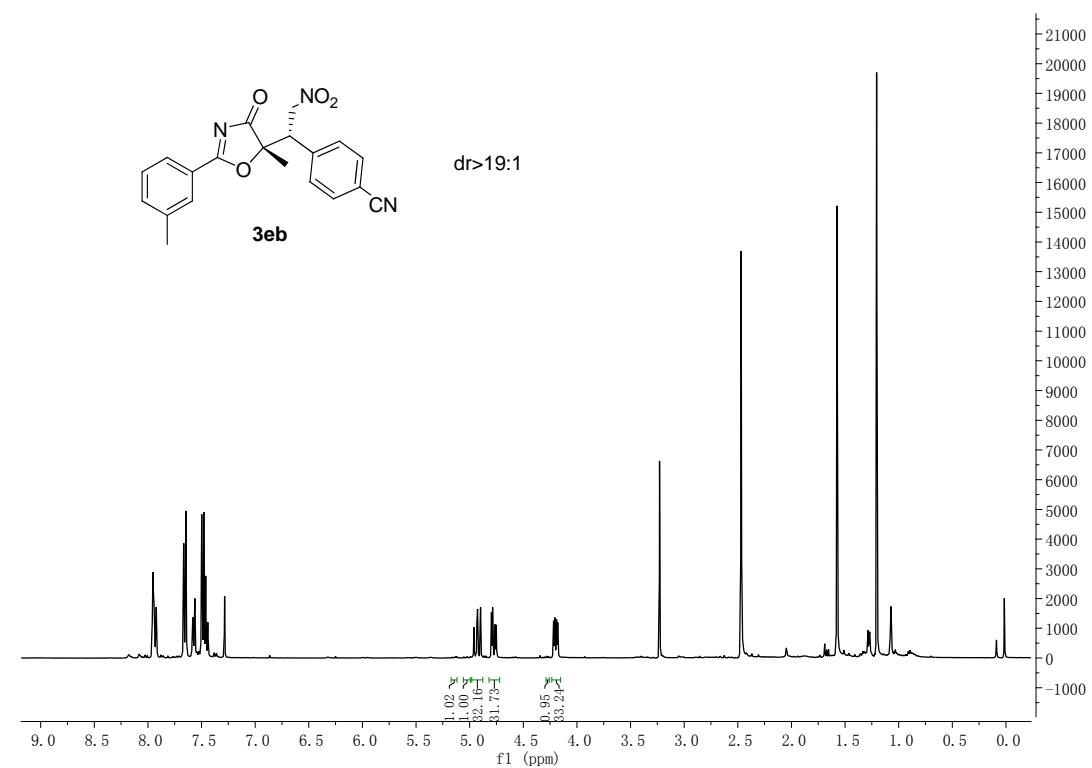
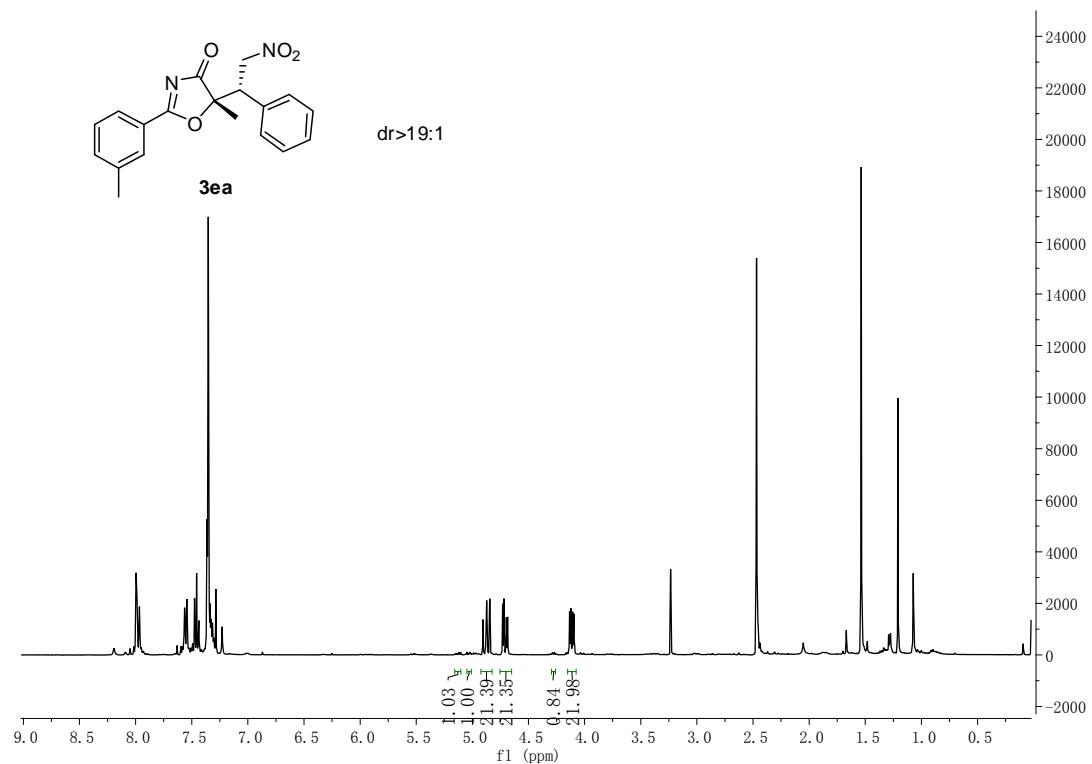
b. Under the same HPLC conditions, we found that **3ea** has opposite absolute configuration comparing with literature from the retention times.

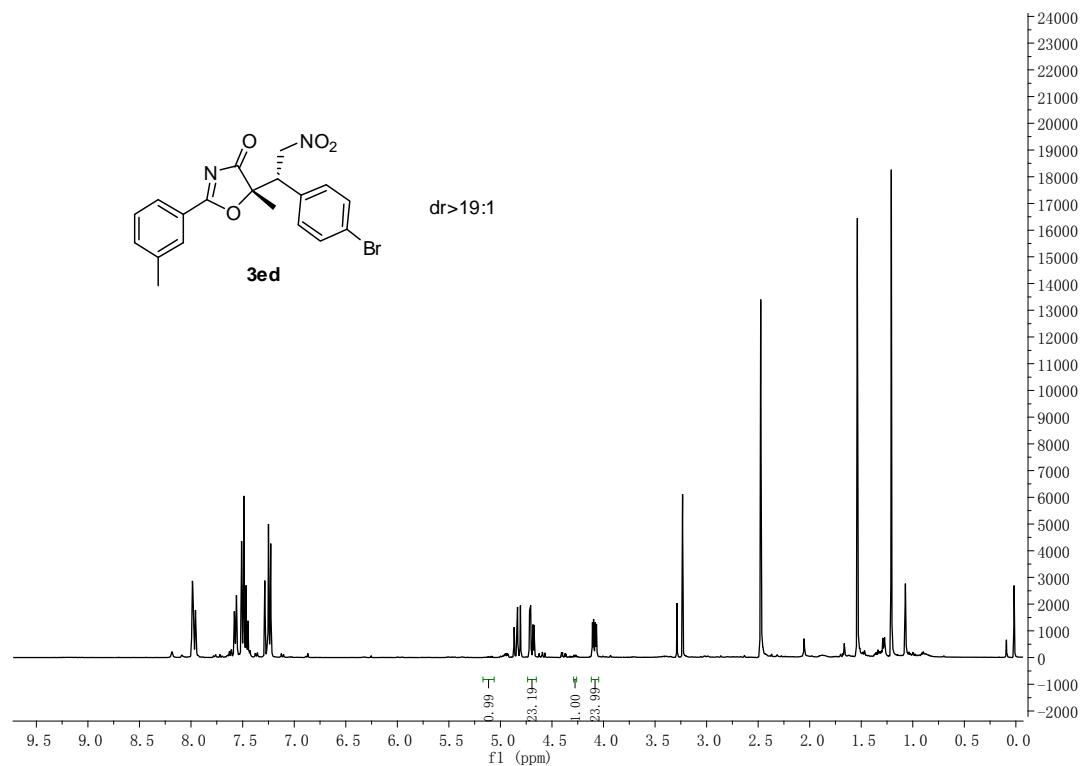
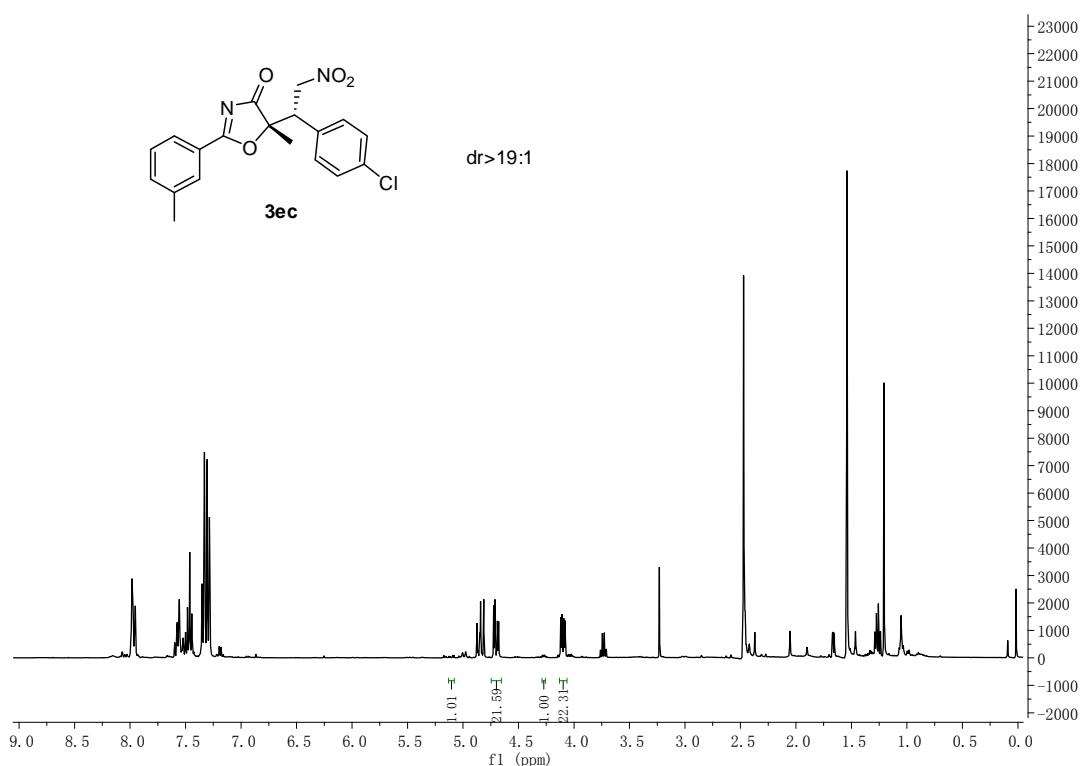
The ee was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. x 250 mm); Hexane/Ethyl acetate = 88/12; flow rate 0.8 ml/min; 25 °C; 254 nm; retention time: 26.4 min (major) and 43.7 min (minor).

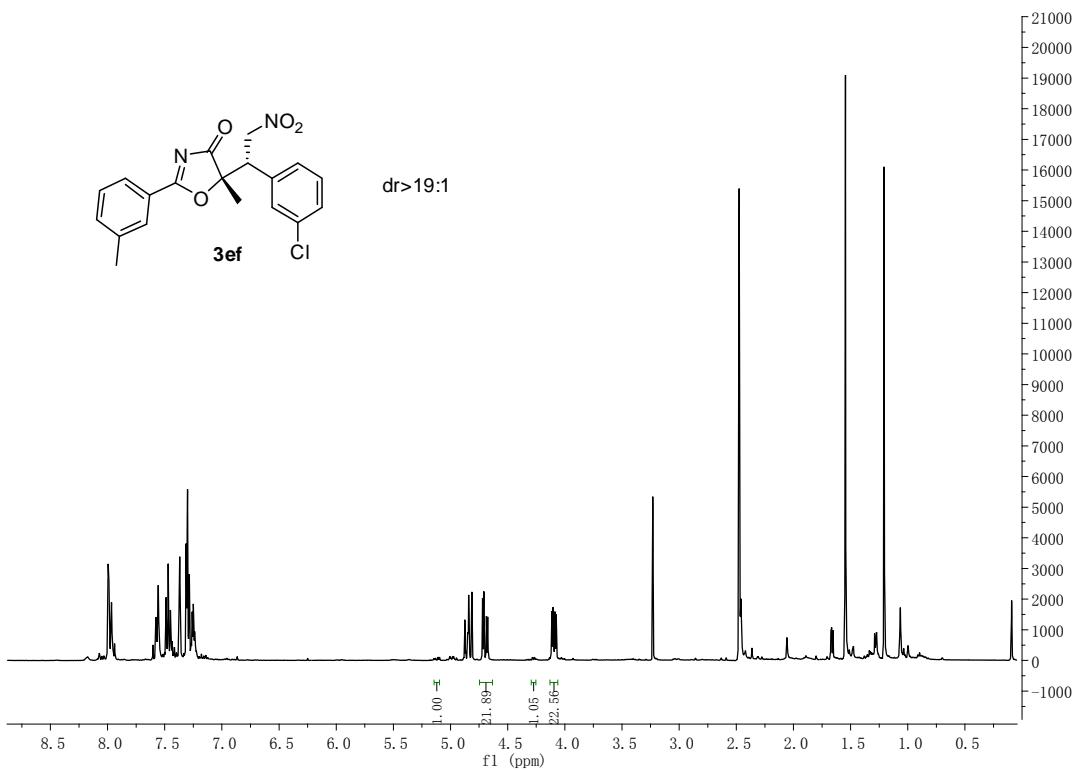
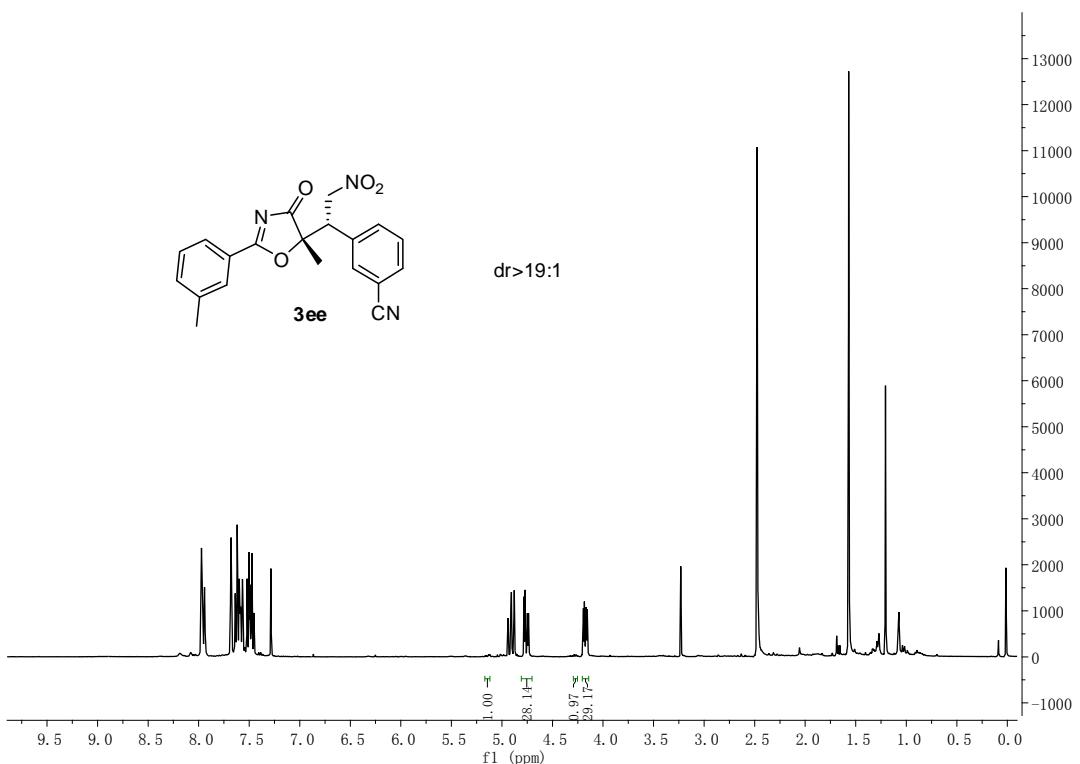


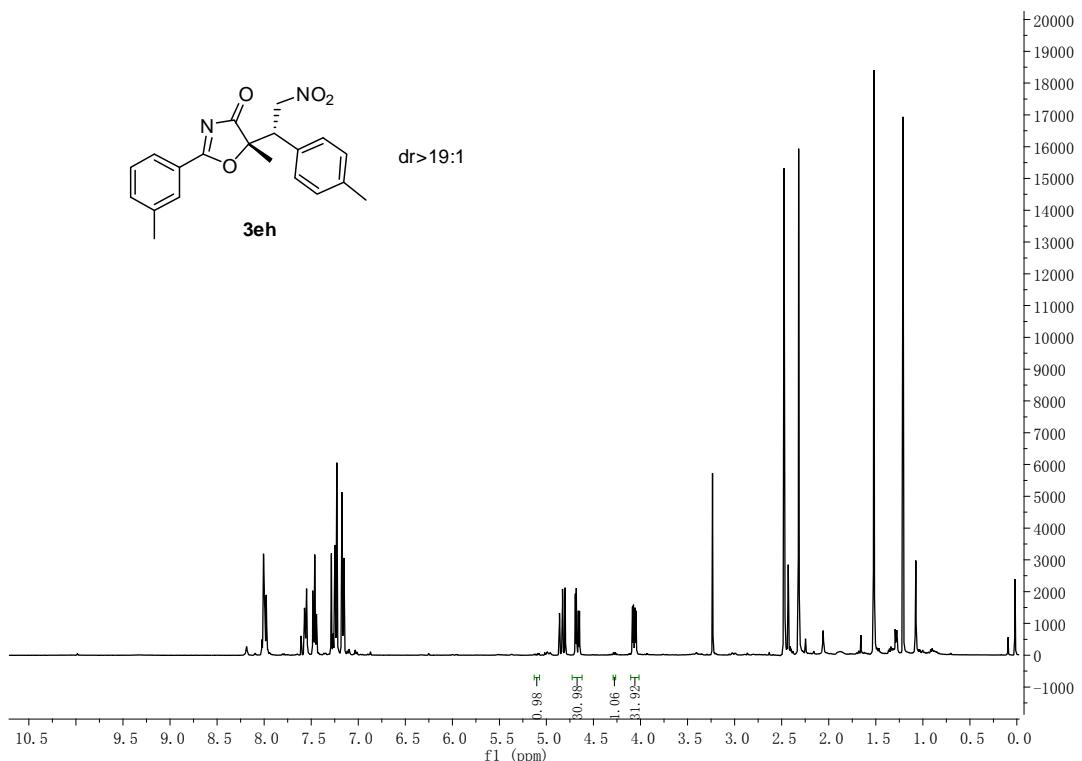
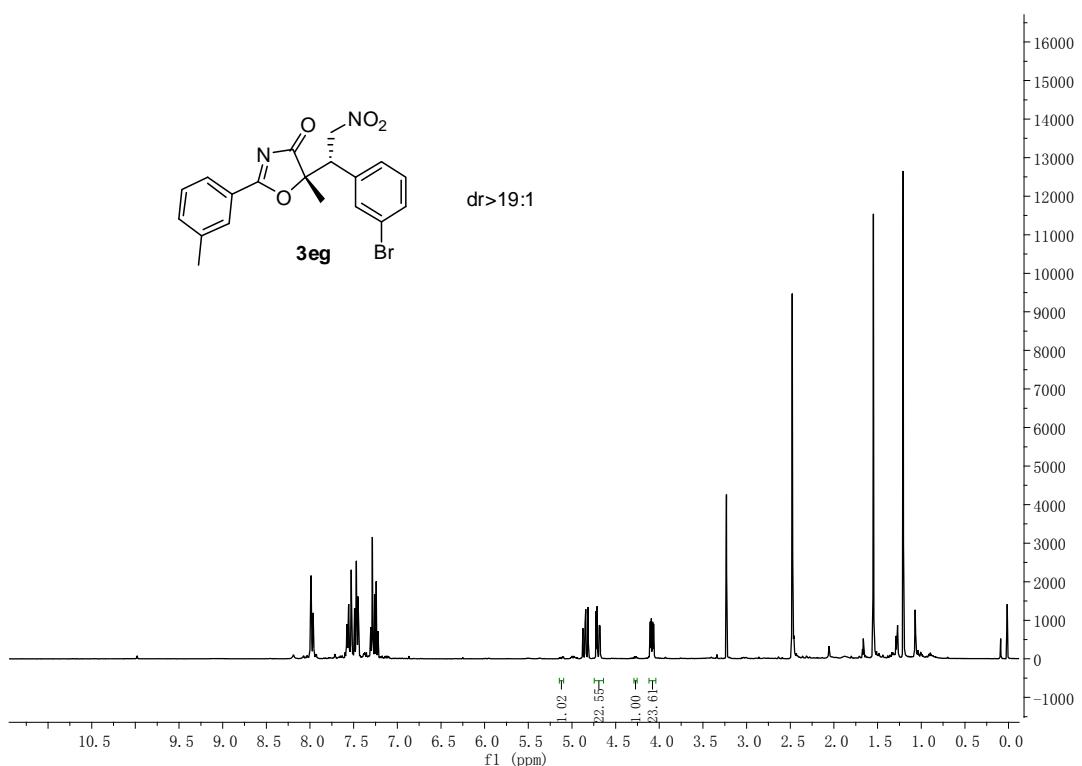
¹ B. M. Trost, K. Hirano, *Angew. Chem. Int. Ed.* 2012, **51**, 6480.

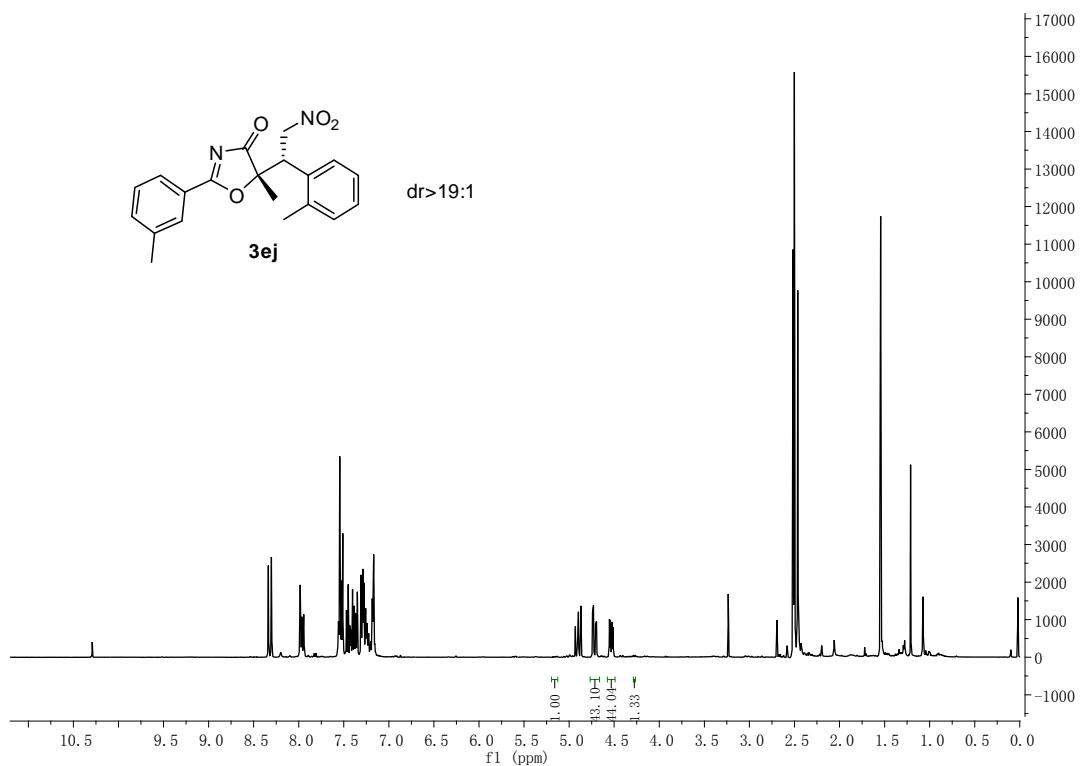
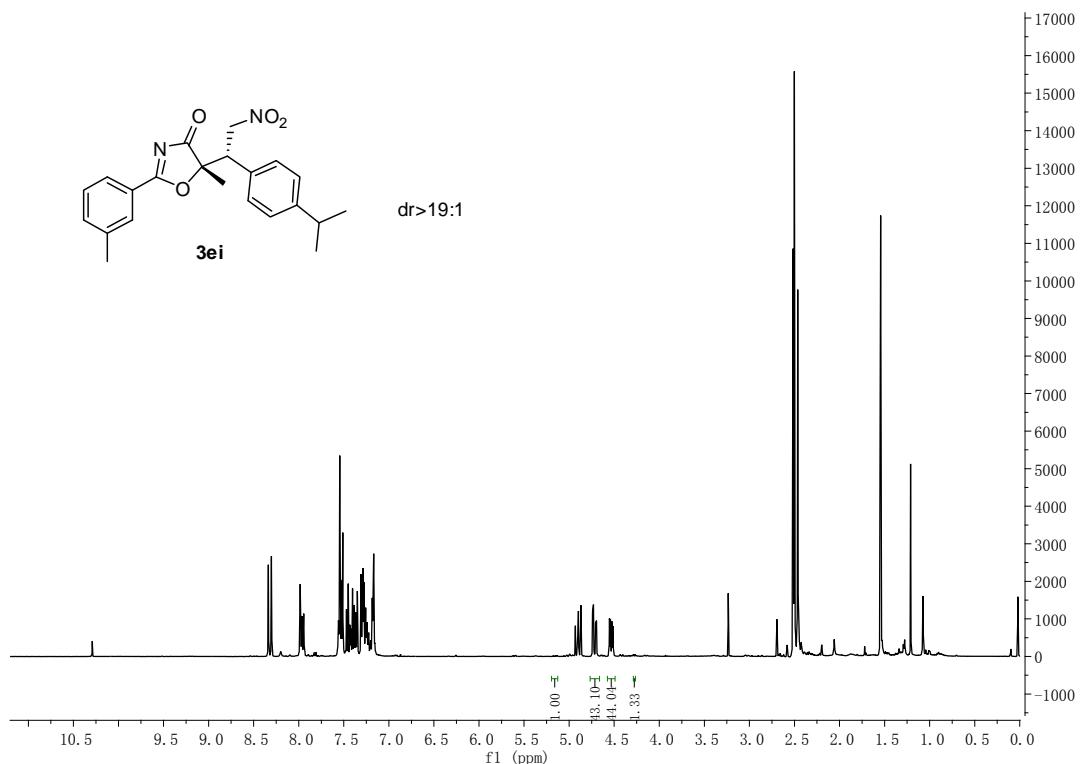
6. Crude ^1H NMR spectra to determine dr

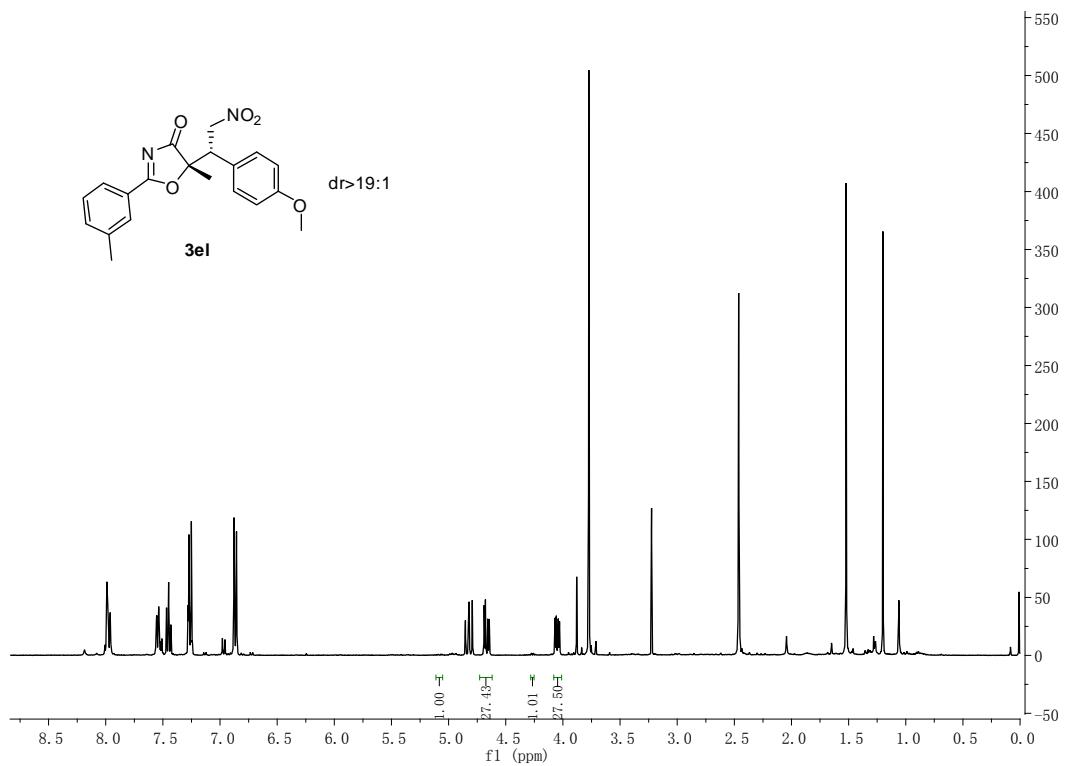
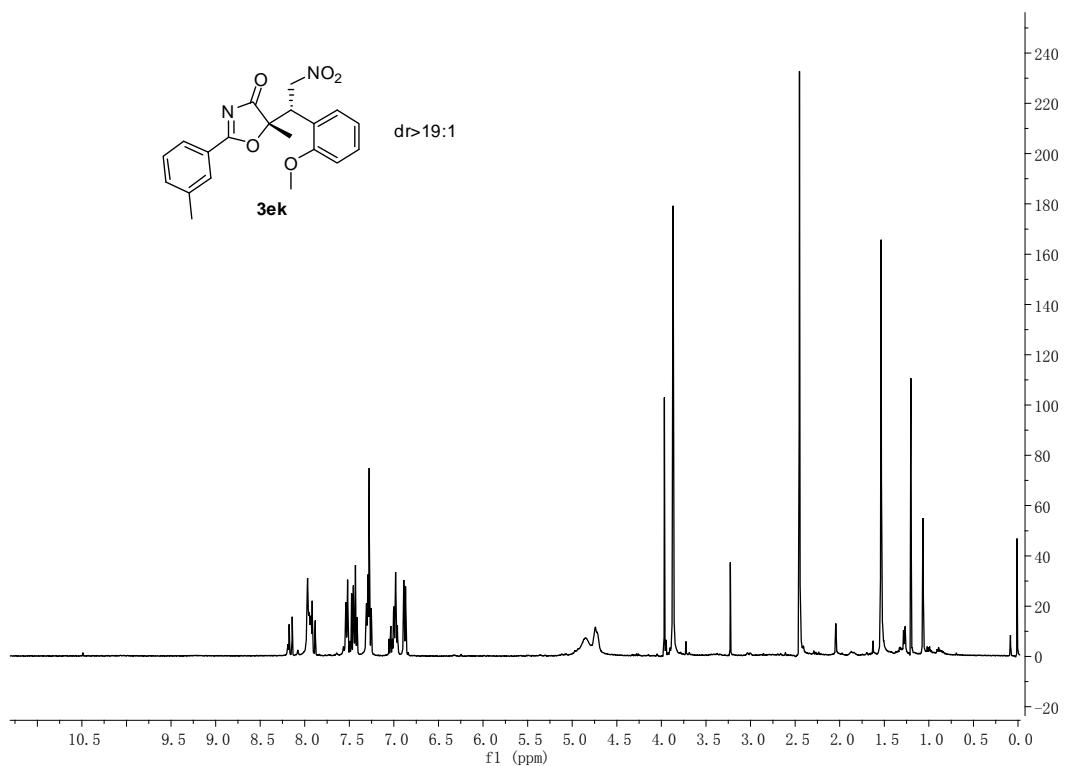


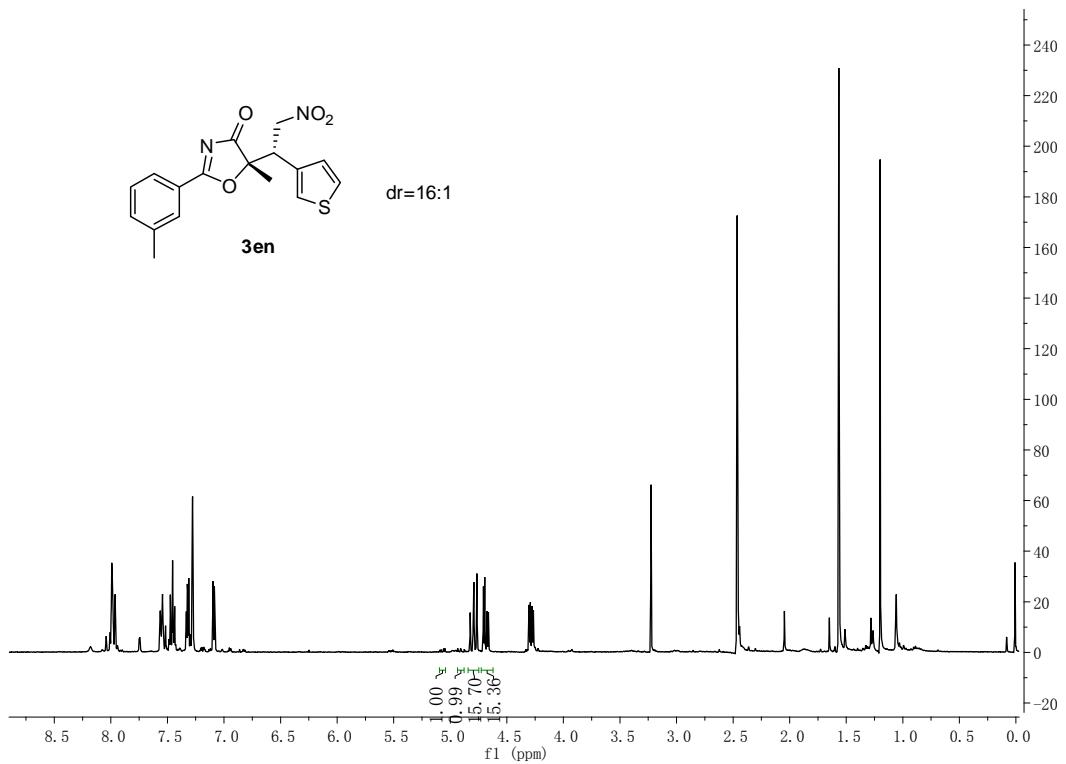
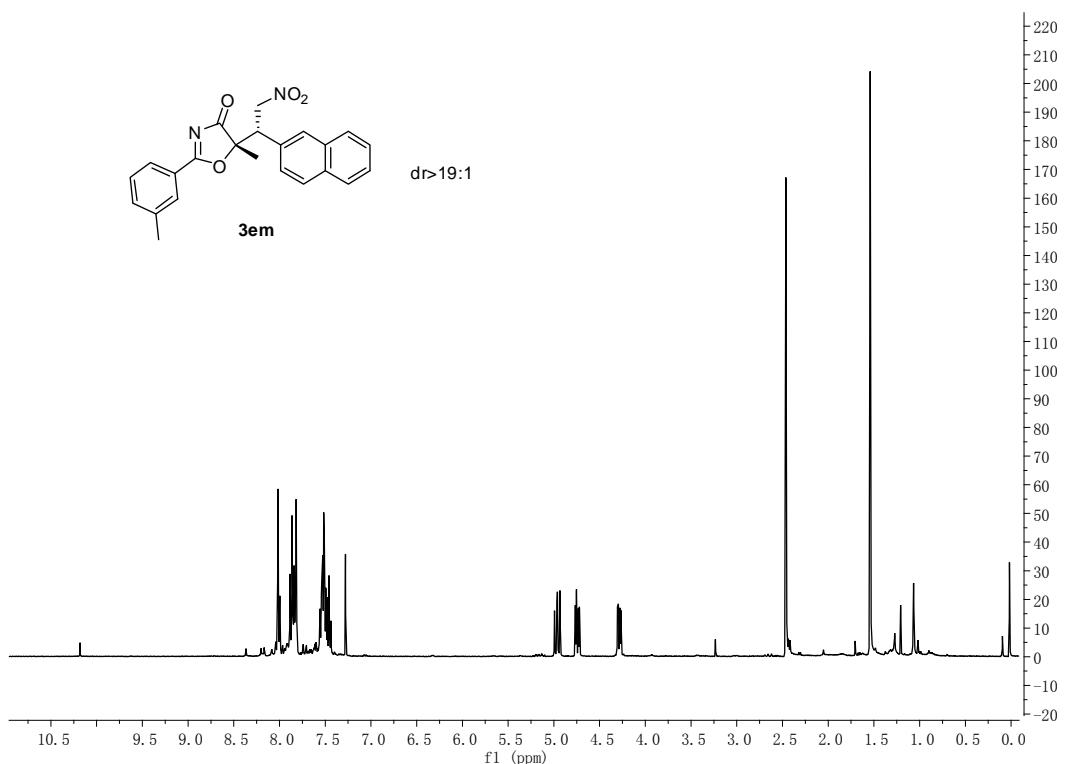


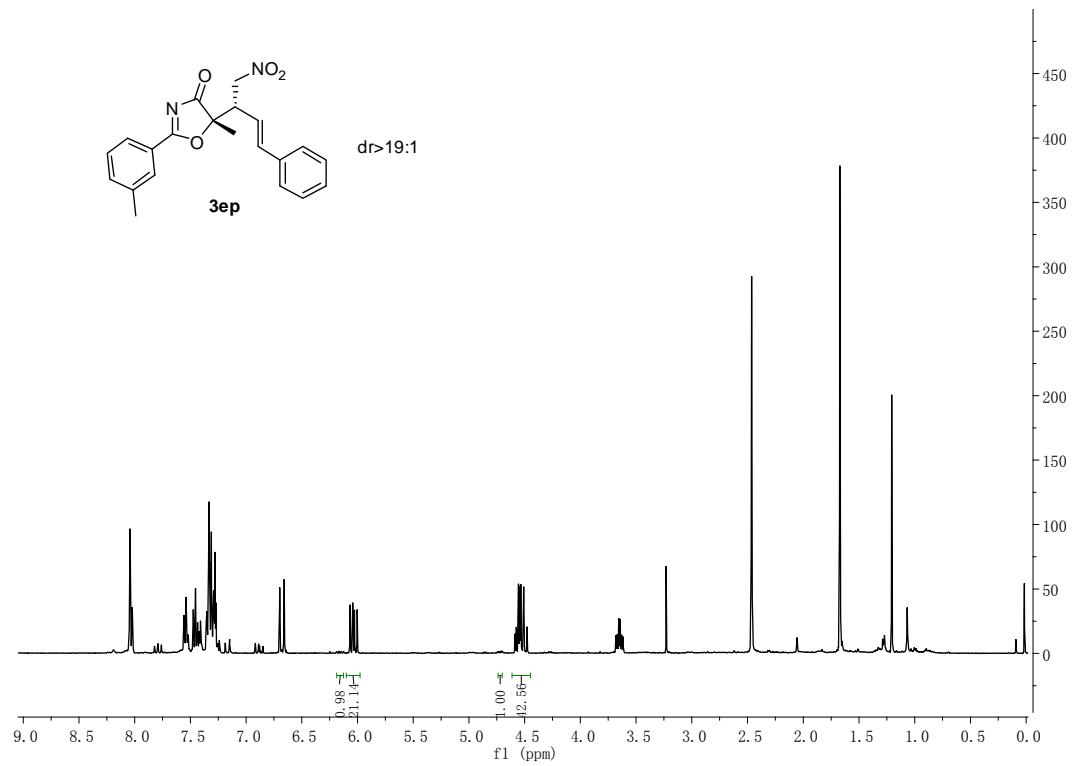
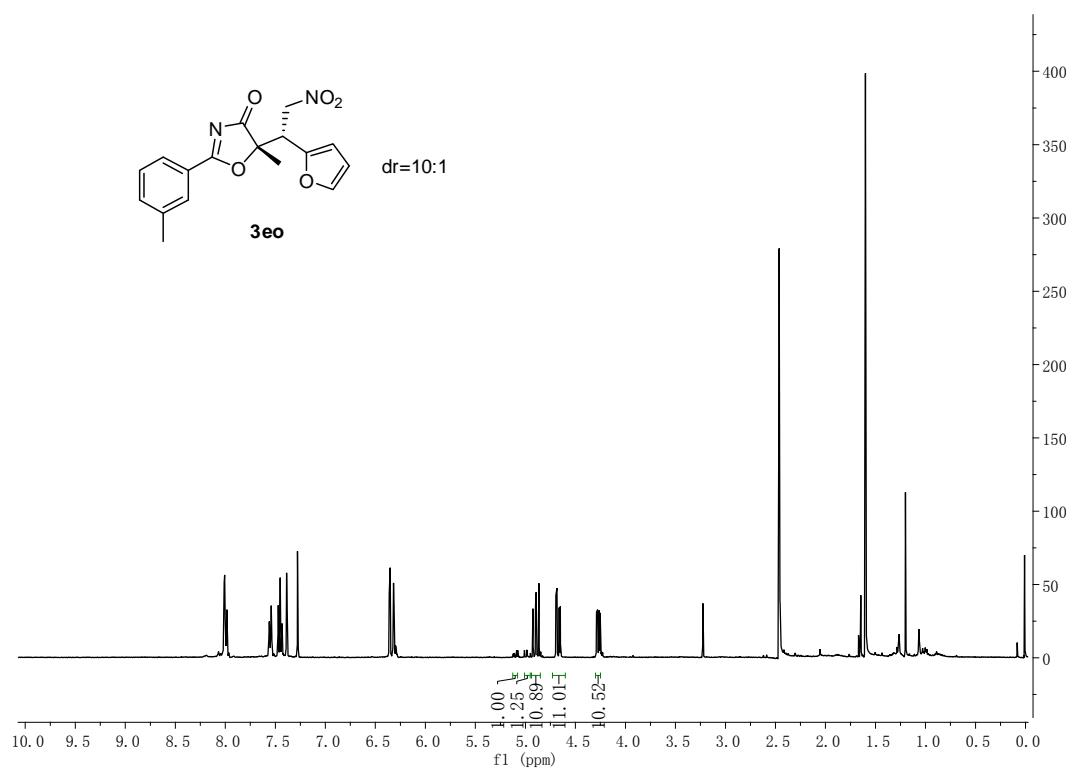


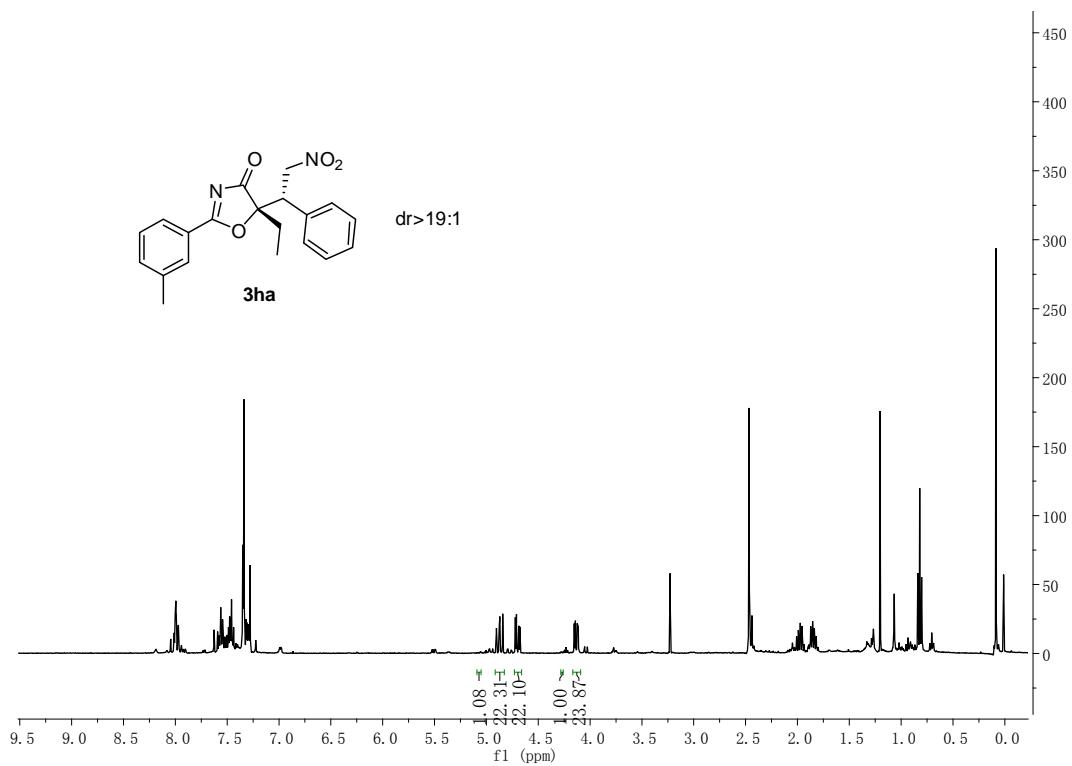
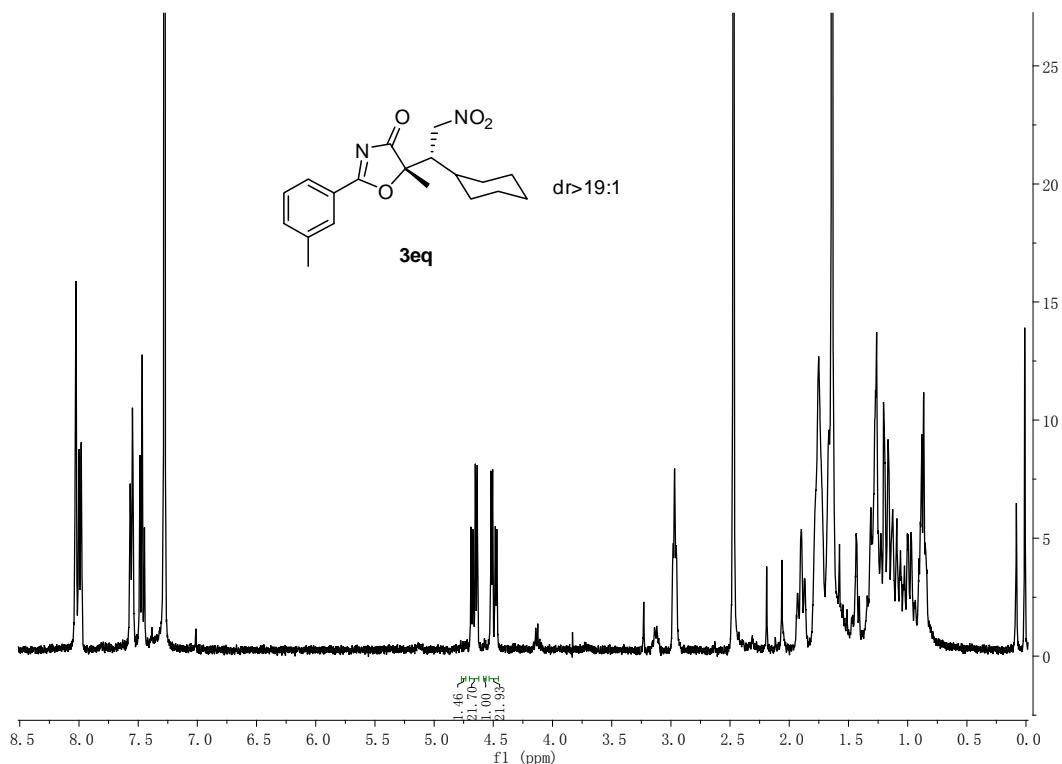


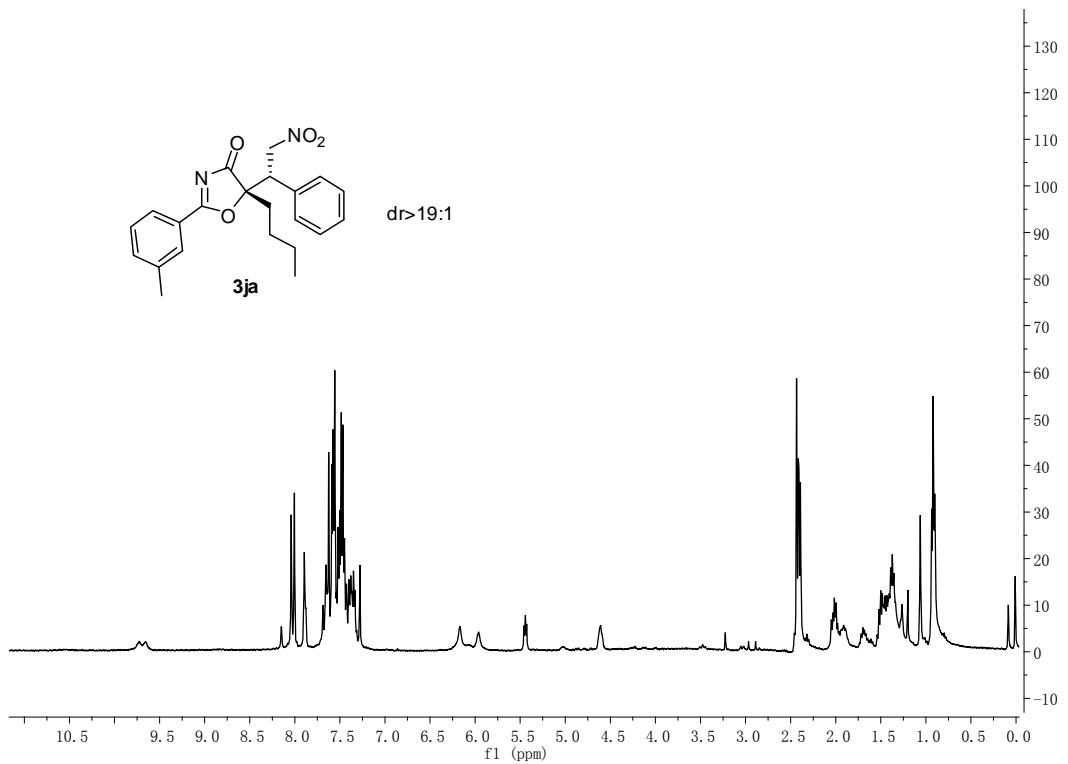
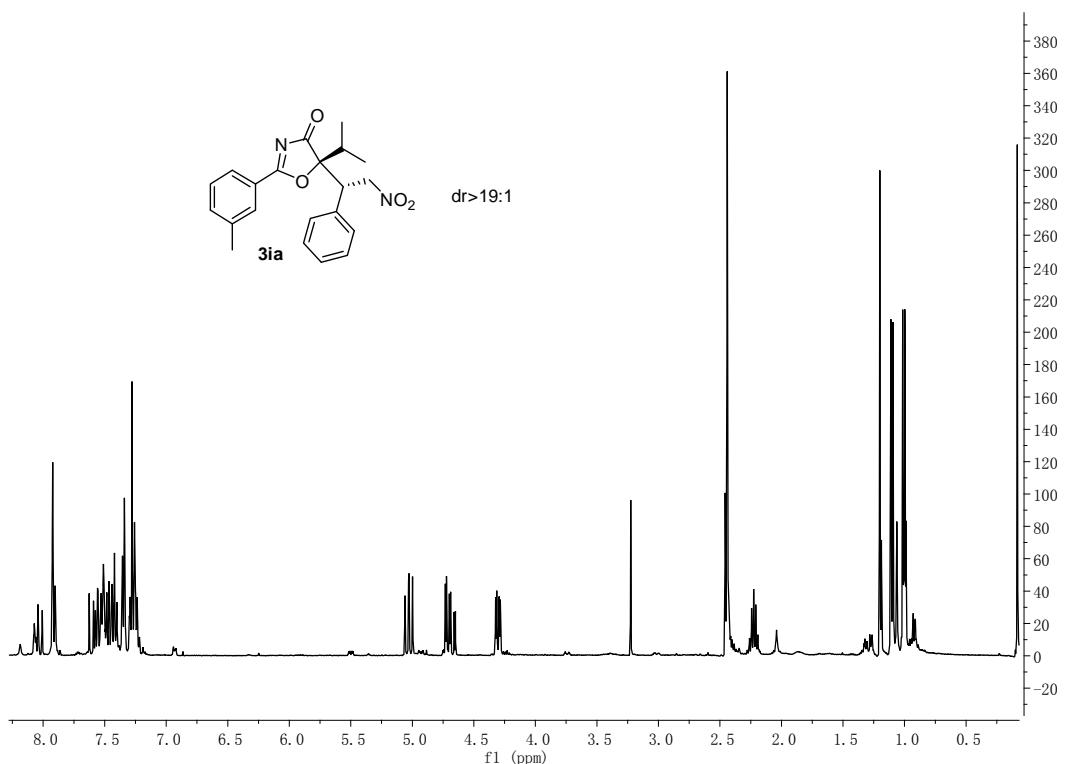


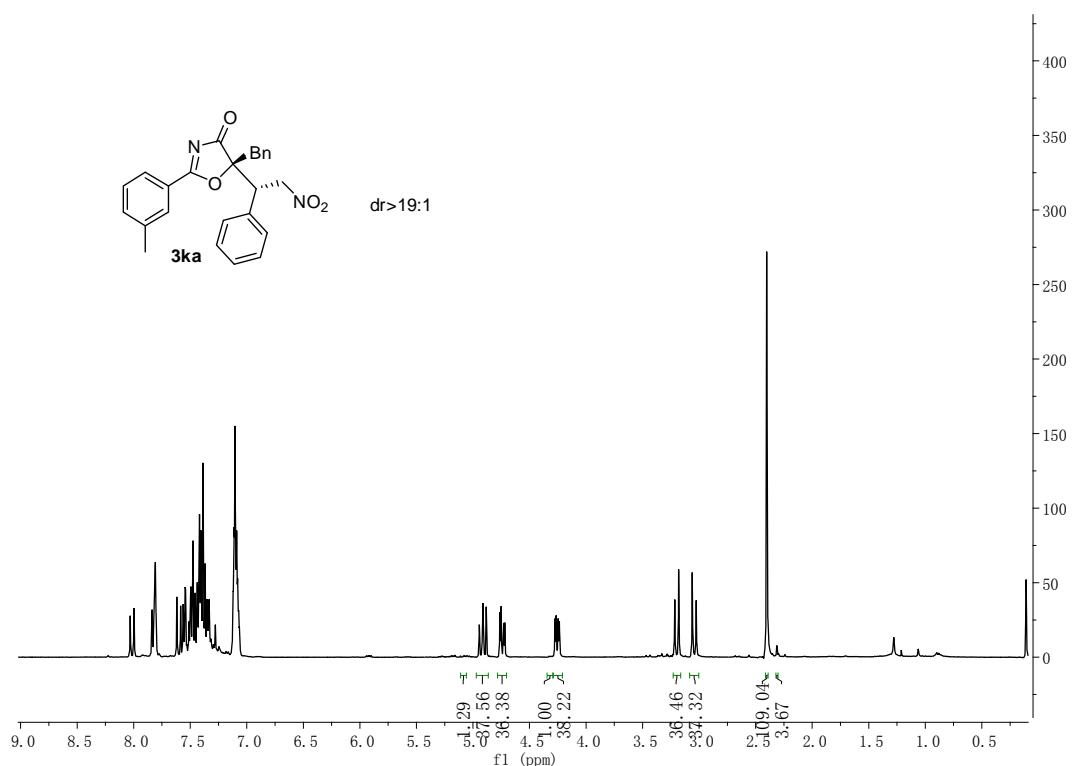












7. Copies of NMR Spectra

