

Supplemental material for

**Mapping peptide thiol accessibility in membranes using
a quaternary ammonium isotope-coded mass tag
(ICMT)**

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RUNNING TITLE: Peptide thiol ICMT maps membrane topology

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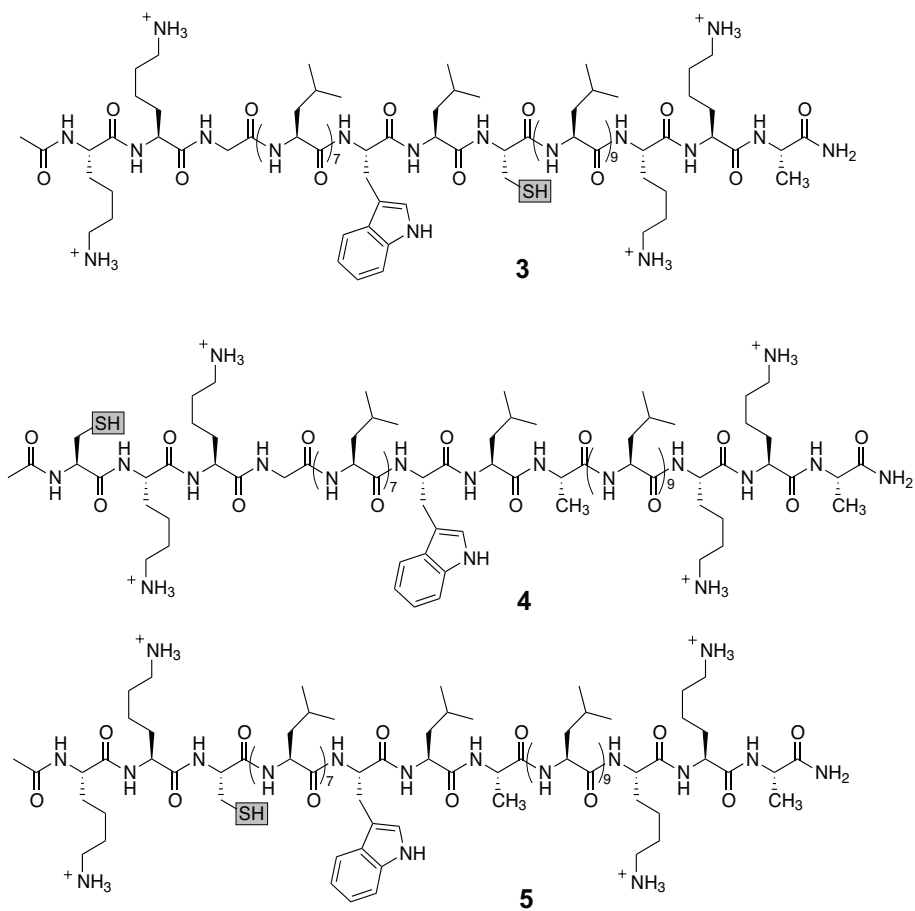


Figure S1. Structures of model α -helical peptides. Peptide **3** (cysteine inaccessible), peptide **4** (cysteine accessible), and peptide **5** (cysteine partially accessible).

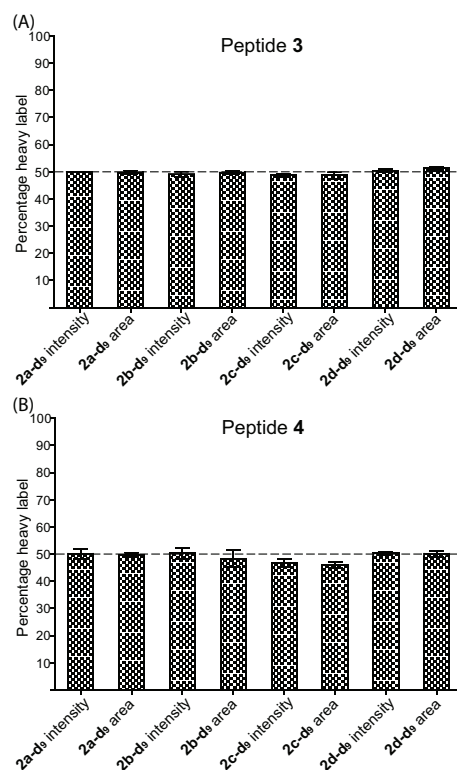


Figure S2. Accuracy and precision of labeling peptide 3 and peptide 4 with ICMT probes. Peptide 3 (panel A) or peptide 4 (50 μ M) (panel B) dissolved in 0.5% (w/v) SDS, 50 mM phosphate, pH 7.0 was labeled with a mixture of light (9 mM) and heavy (9 mM) ICMT probe. The percentage of peptide labeled with heavy probe was determined from either the average intensity of the isotopomer in each envelope or the area under the entire isotopic envelope. Average values from three replicate experiments and the standard deviations are shown. The dashed line represents the expected value of 50%.

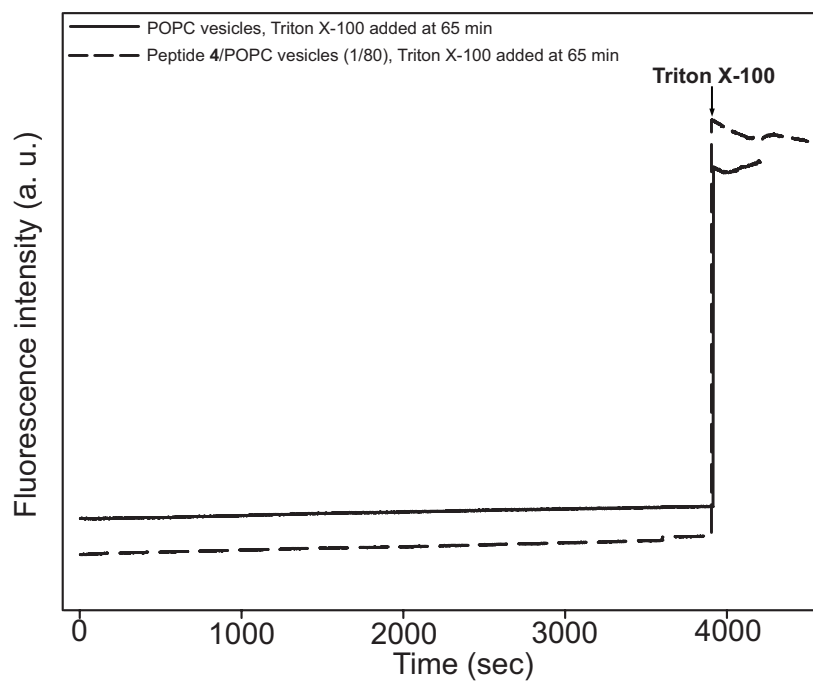


Figure S3. Vesicle permeability assay. Carboxyfluorescein (100 mM) was encapsulated in peptide **4**/POPC (1/80) vesicles or POPC vesicles (285 μ M total lipid). Fluorescence emission intensities ($\lambda_{\text{ex}} = 492$ nm, $\lambda_{\text{em}} = 517$ nm) were monitored for 70 min, and Triton X-100 was added to completely lyse vesicles at 65 min.

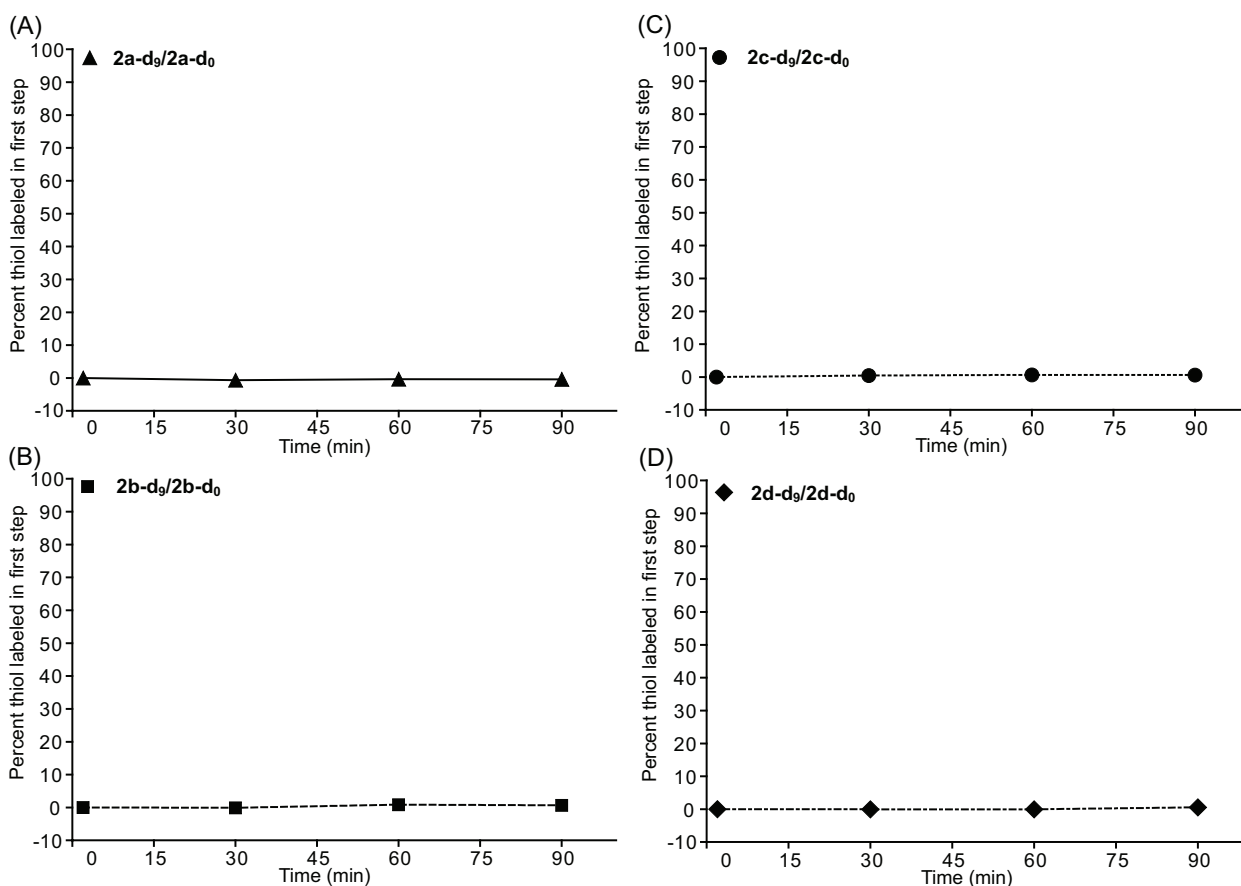


Figure S4. ICMT permeability tests with peptide 4/POPC (1/80) vesicles. Tri-peptide NB-ECD-OMe (3 mM) was encapsulated in peptide 4/POPC (1/80) vesicles. Intact vesicles were labeled with 6 mM $2a-d_9$, $2b-d_9$, $2c-d_9$, or $2d-d_9$ heavy probe for 2, 30, 60 or 90 minutes, quenched with 7 mM DTT, 0.03% Triton X-100 to lyse vesicles, and were then labeled with 15 mM $2a-d_0$, $2b-d_0$, $2c-d_0$, or $2d-d_0$ light probe for 1 hour. The reactions were analyzed by LC/MS and extracted ion chromatograms were integrated. Percentage heavy label indicates the percent of total labeling that occurred with intact vesicles. Average values from three replicate experiments and the standard deviations are shown. Tests with (A) $2a-d_9/2a-d_0$ probe. (B) $2b-d_9/2b-d_0$ probe. (C) $2c-d_9/2c-d_0$ probe. (D) $2d-d_9/2c-d_0$ probe.

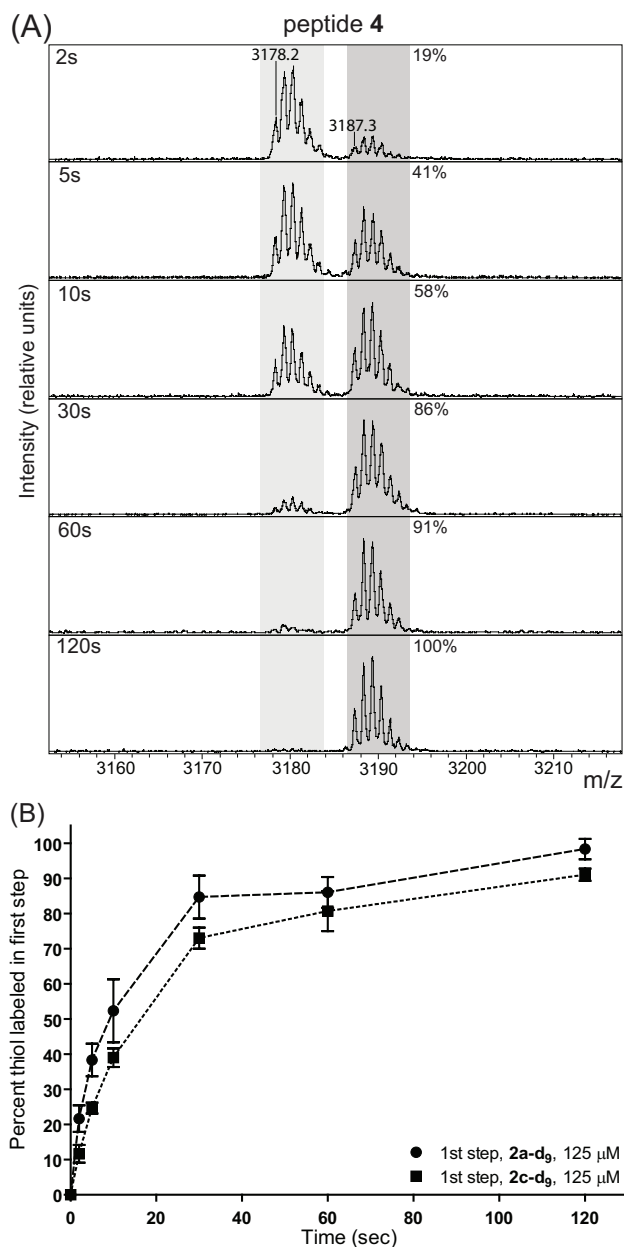
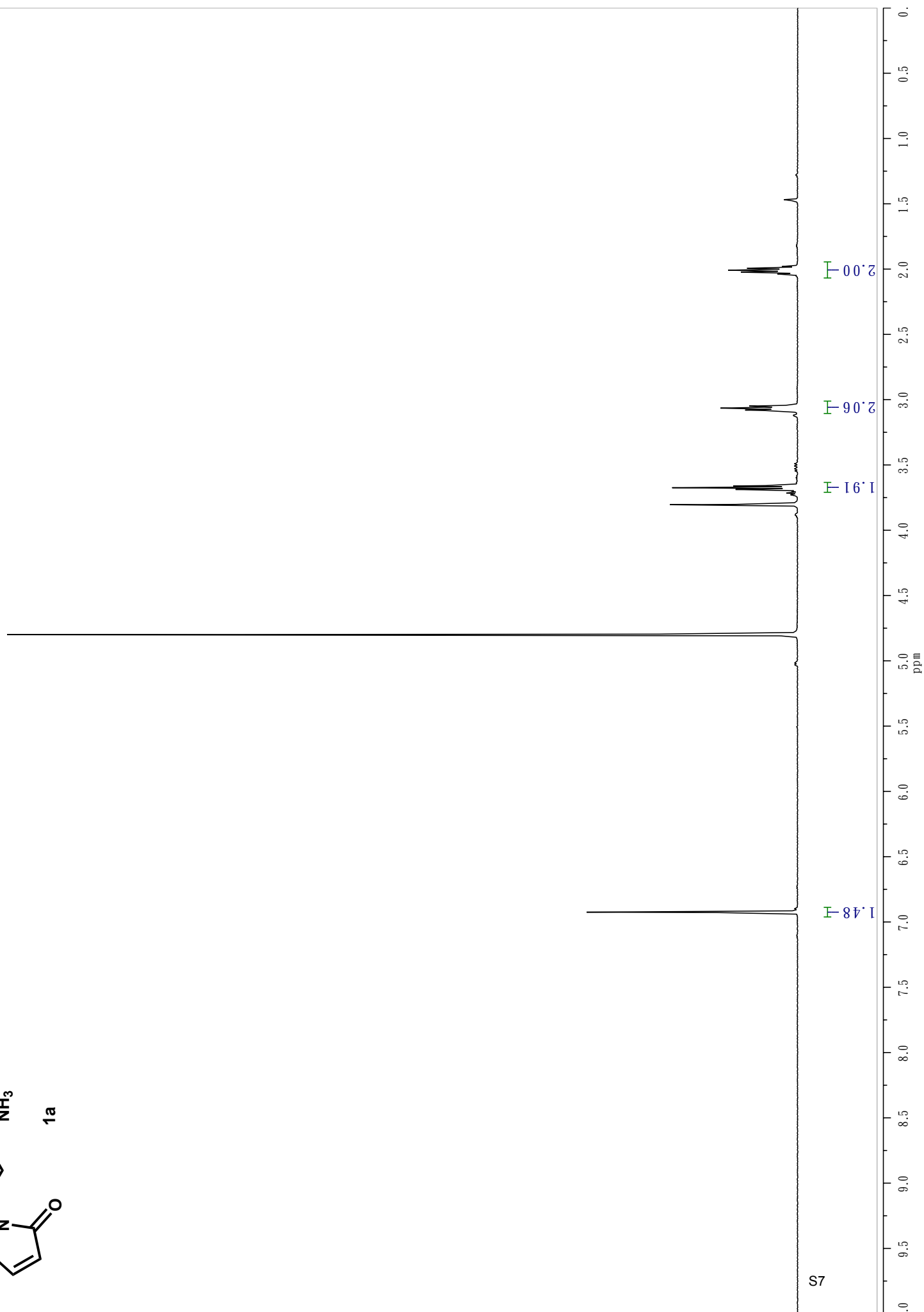
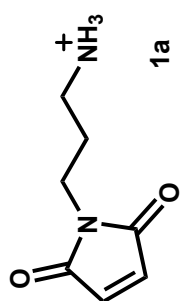
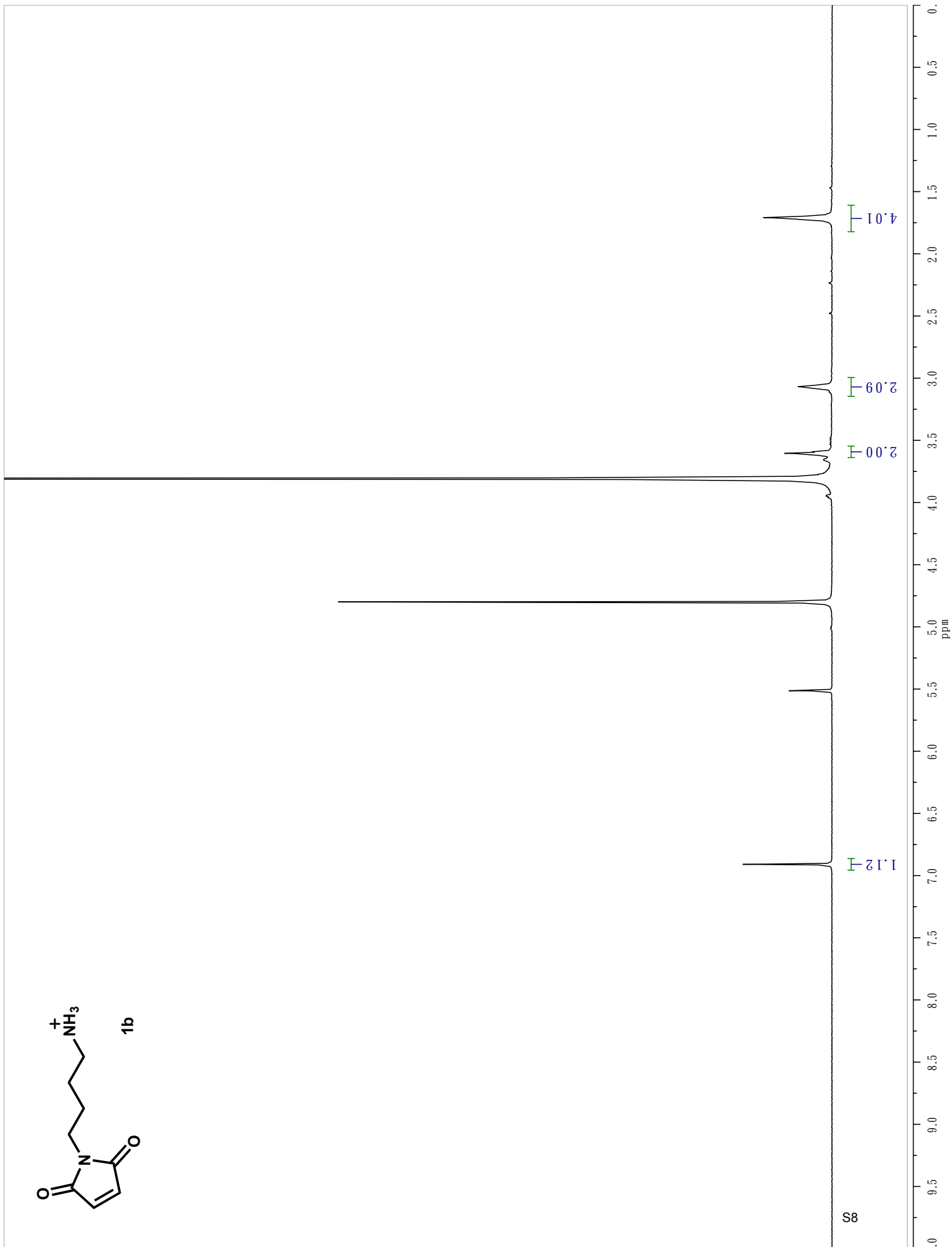
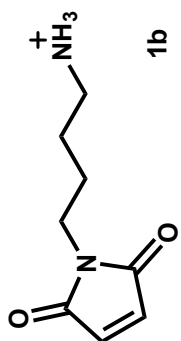
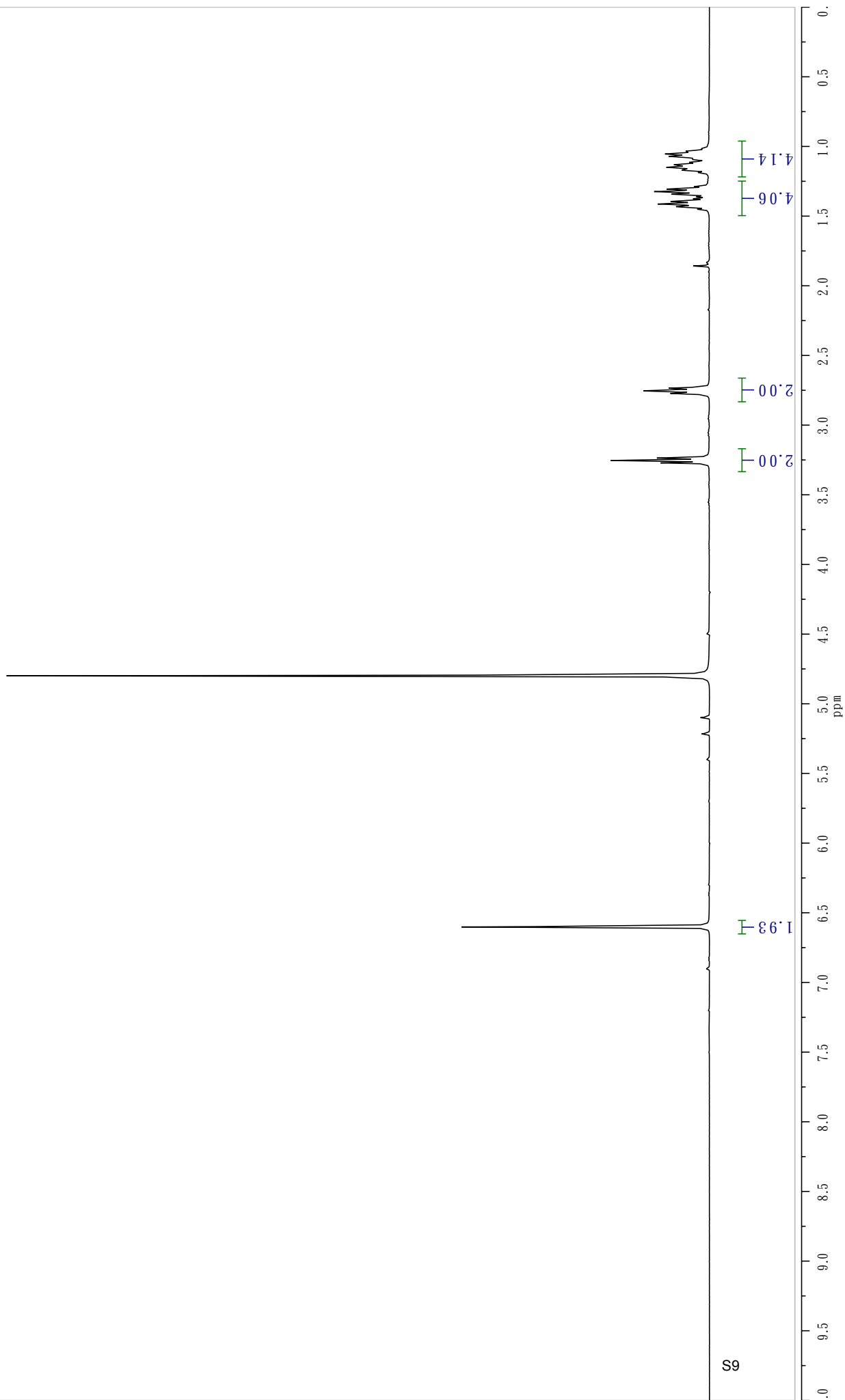
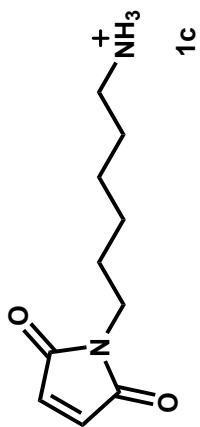
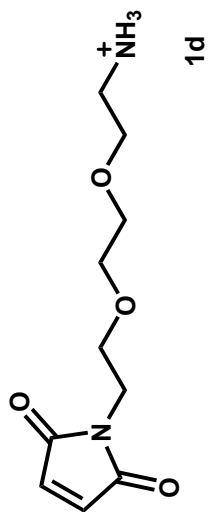


Figure S5. Kinetics of labeling peptide 4/POPC vesicles (1/80). The vesicles were labeled with 125 μ M **2a-d₉** or **2c-d₉**, after 2, 5, 10, 30, 60, or 120 s followed by 1.25 mM **2a-d₀** or **2c-d₀**, respectively. (A) Mass spectra of peptide 4 labeled with **2a-d₉**/**2a-d₀** at different time points. (B) Time courses for the two labeling experiments. Integrated areas under the isotopic envelope were used to calculate percent labeling.









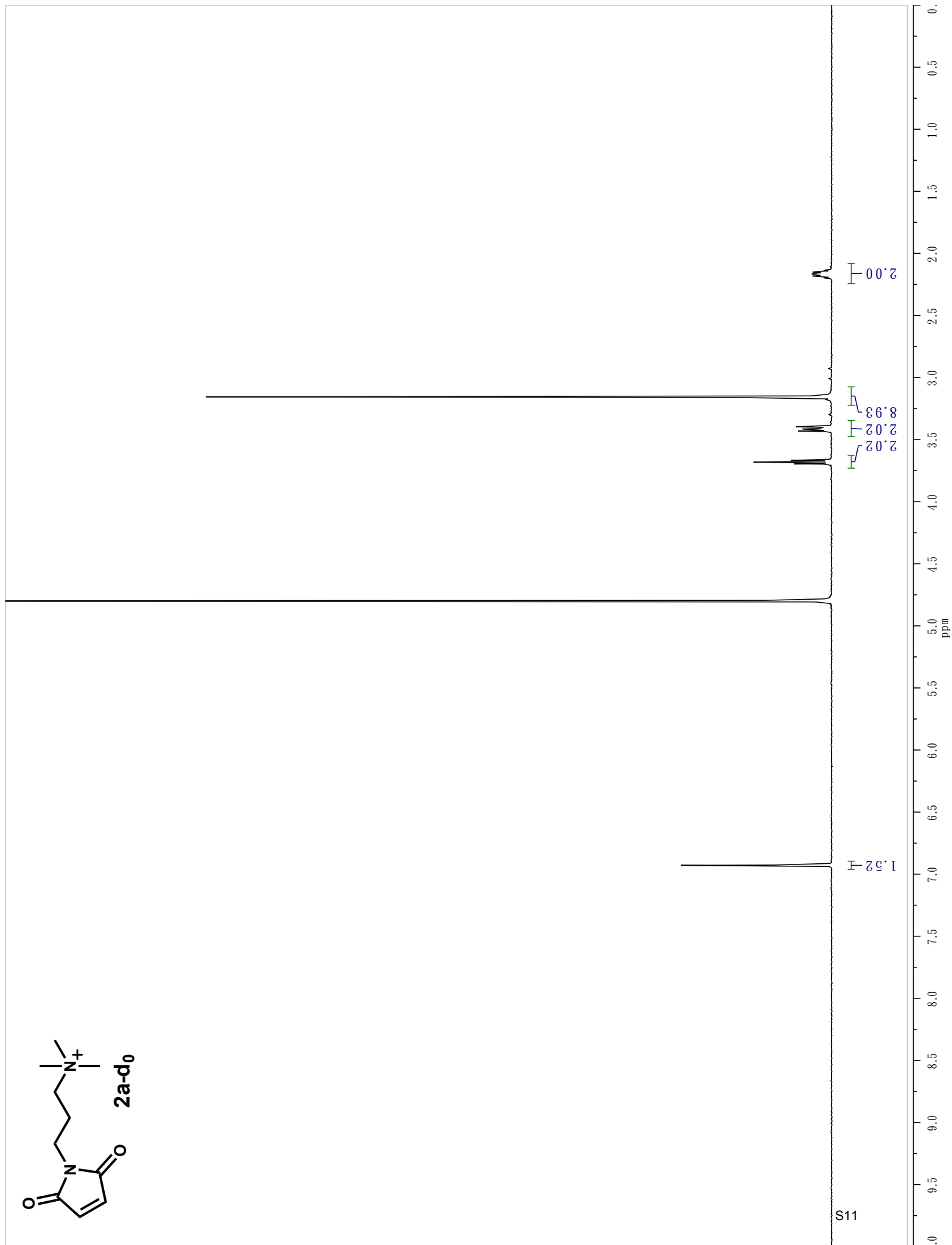
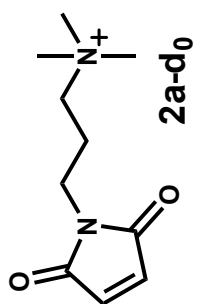
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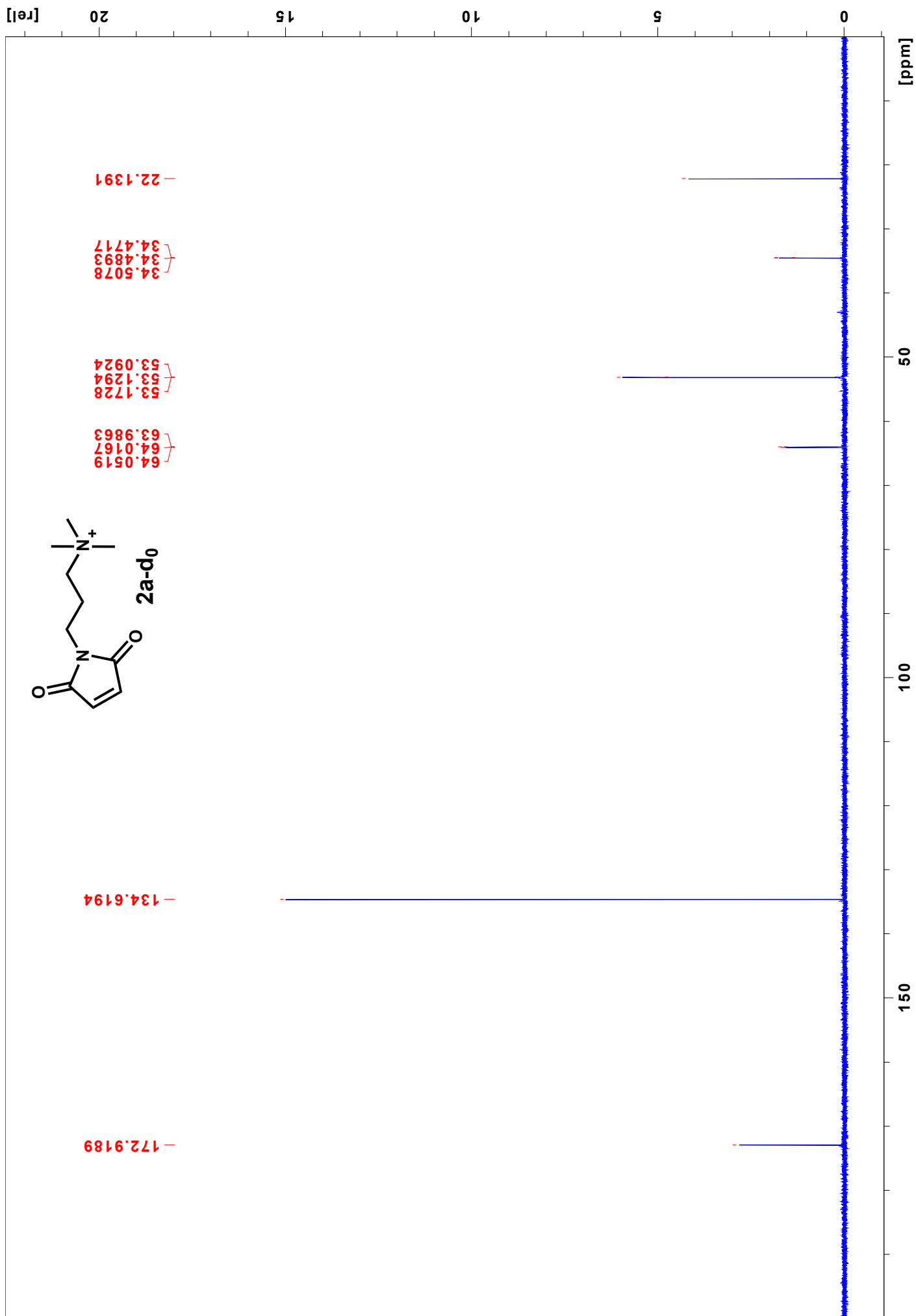
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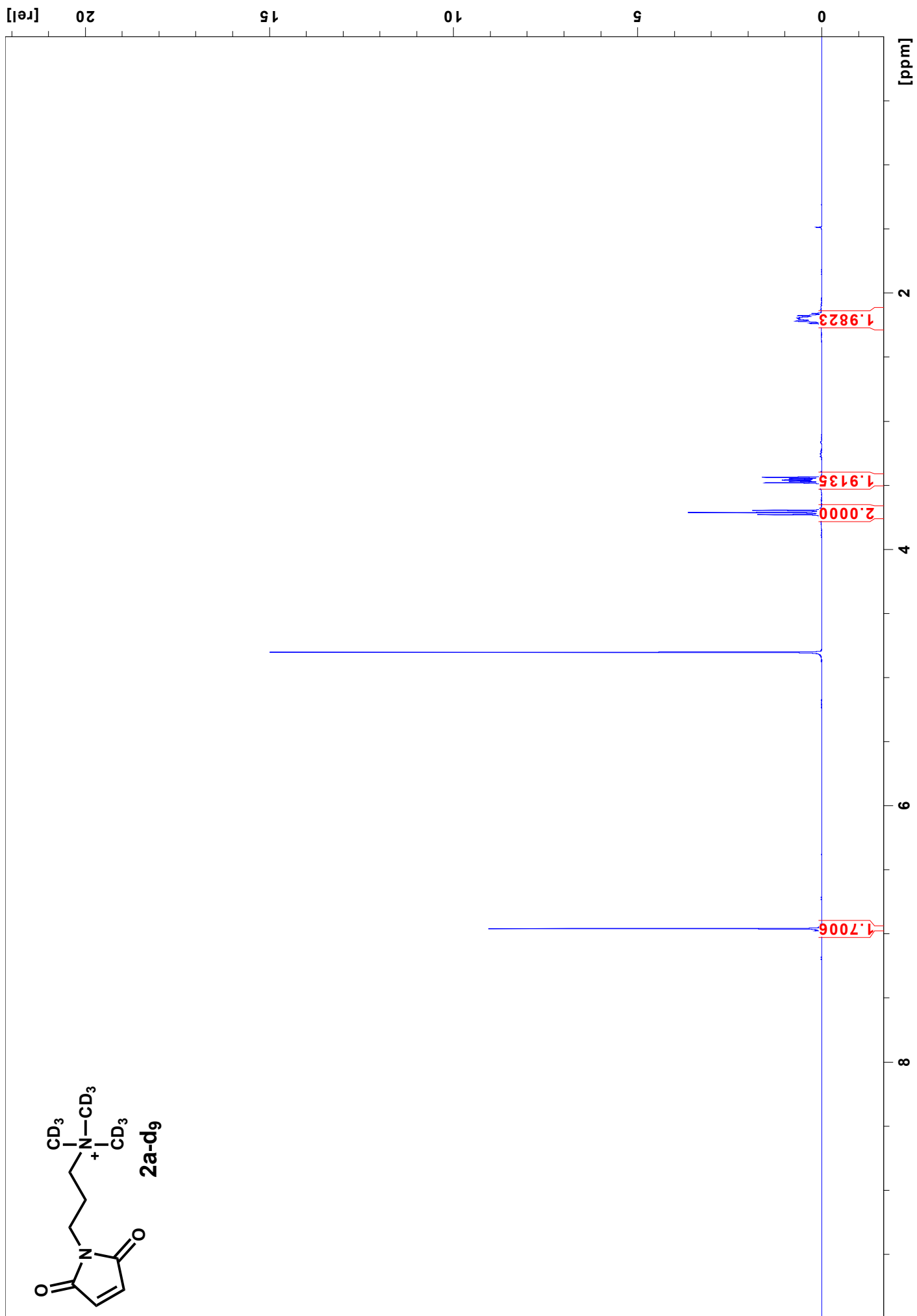
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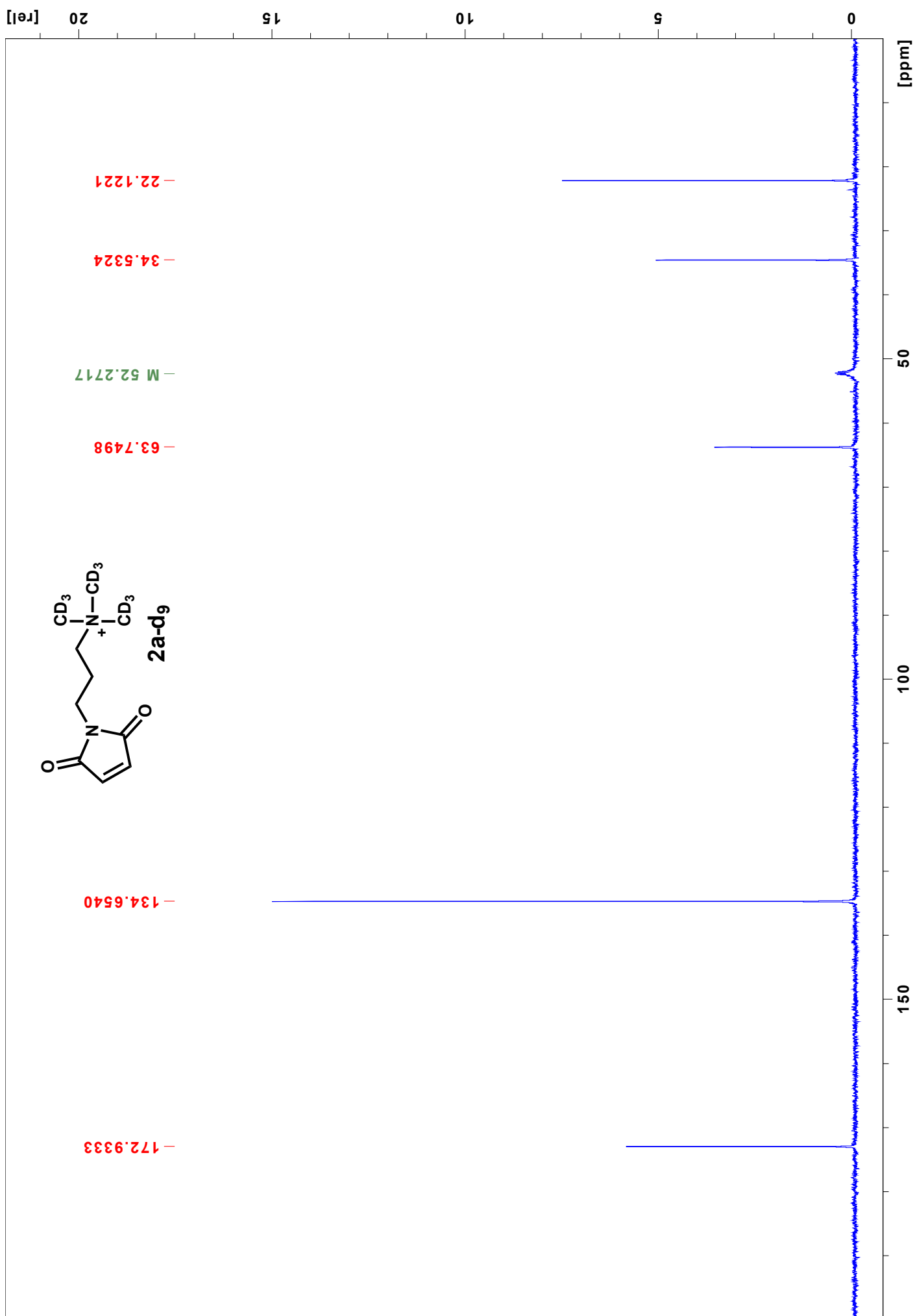
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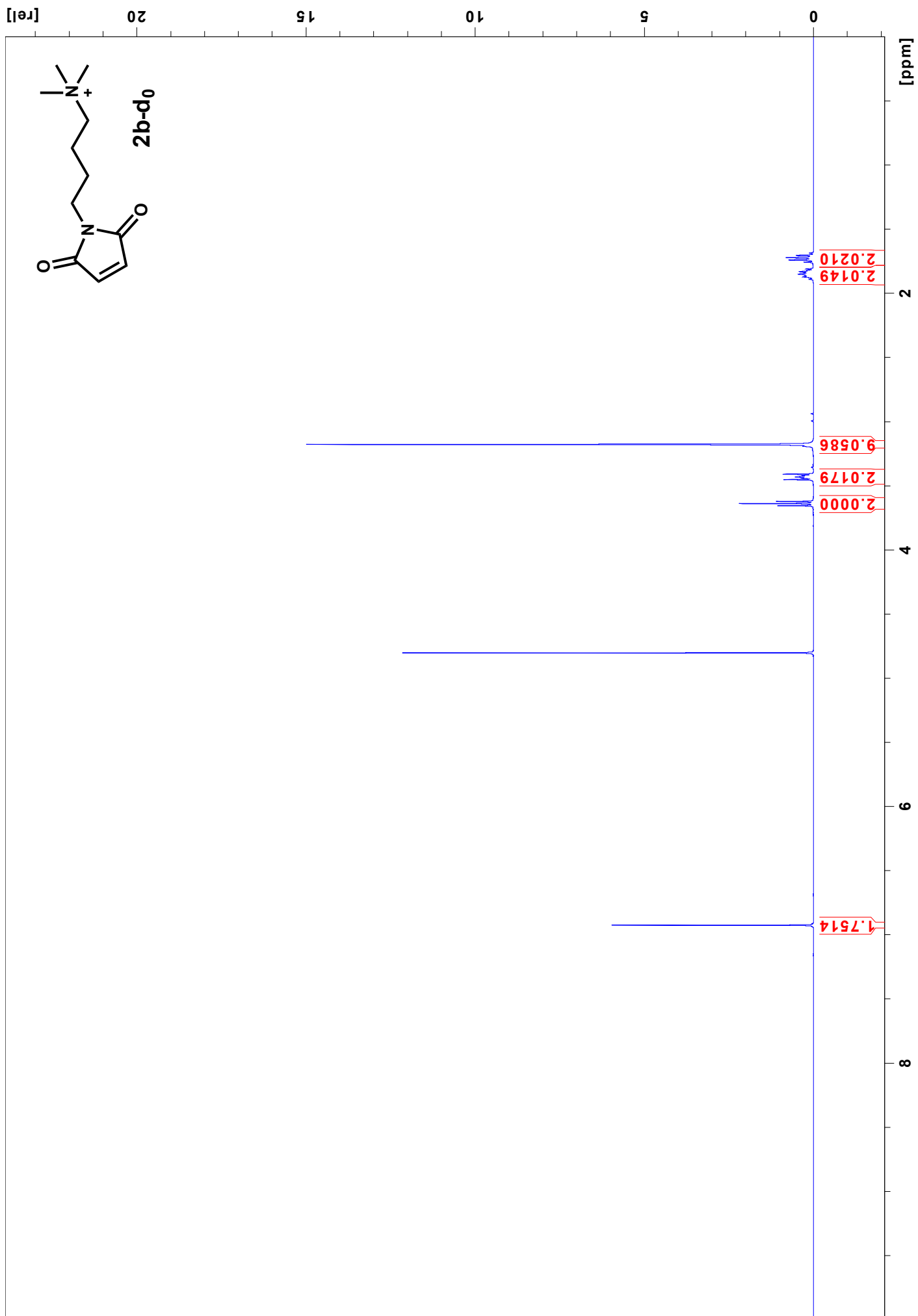
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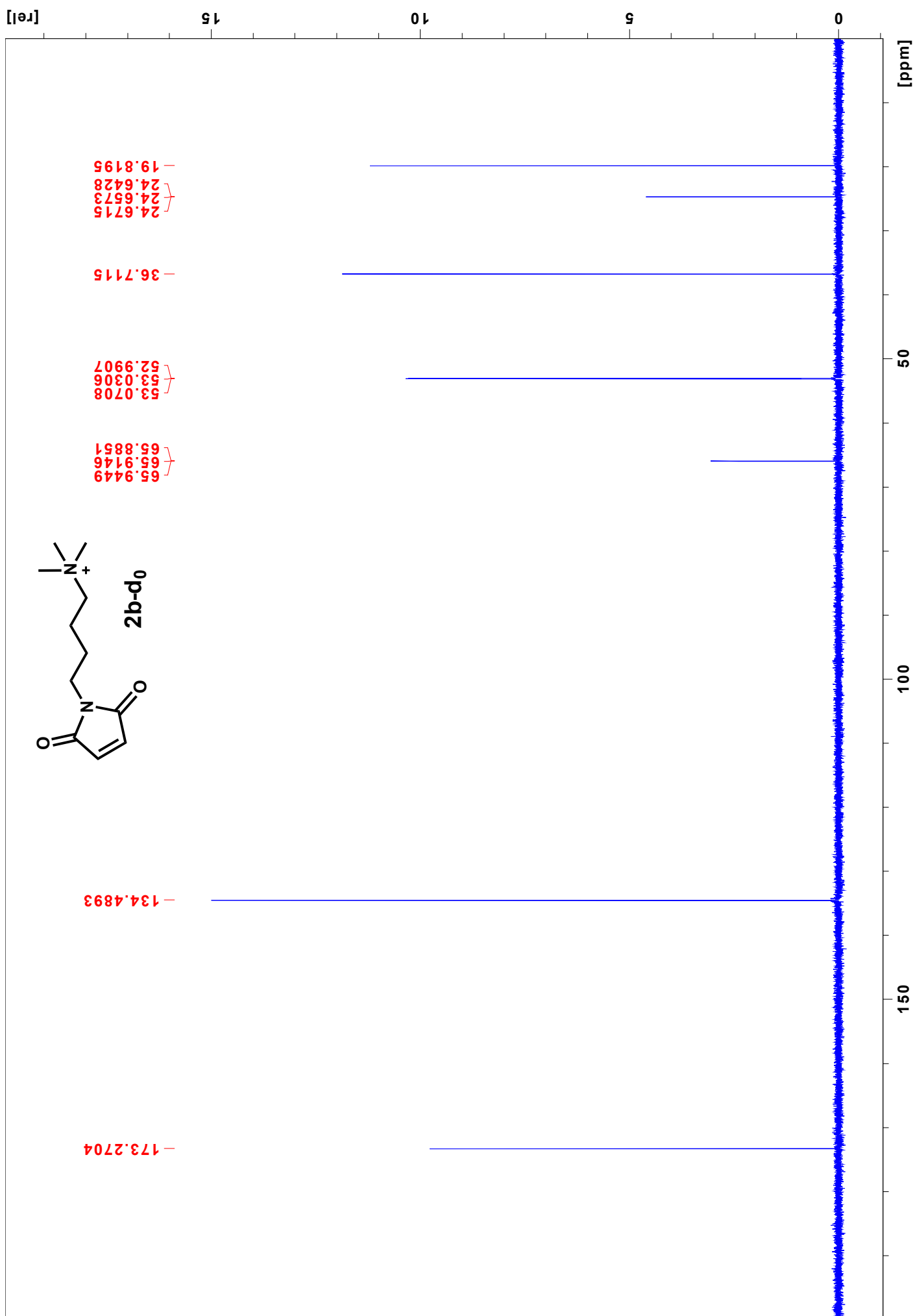


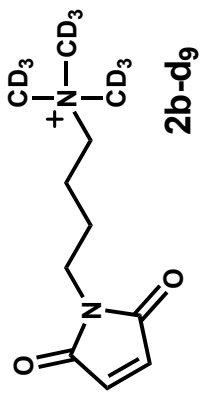












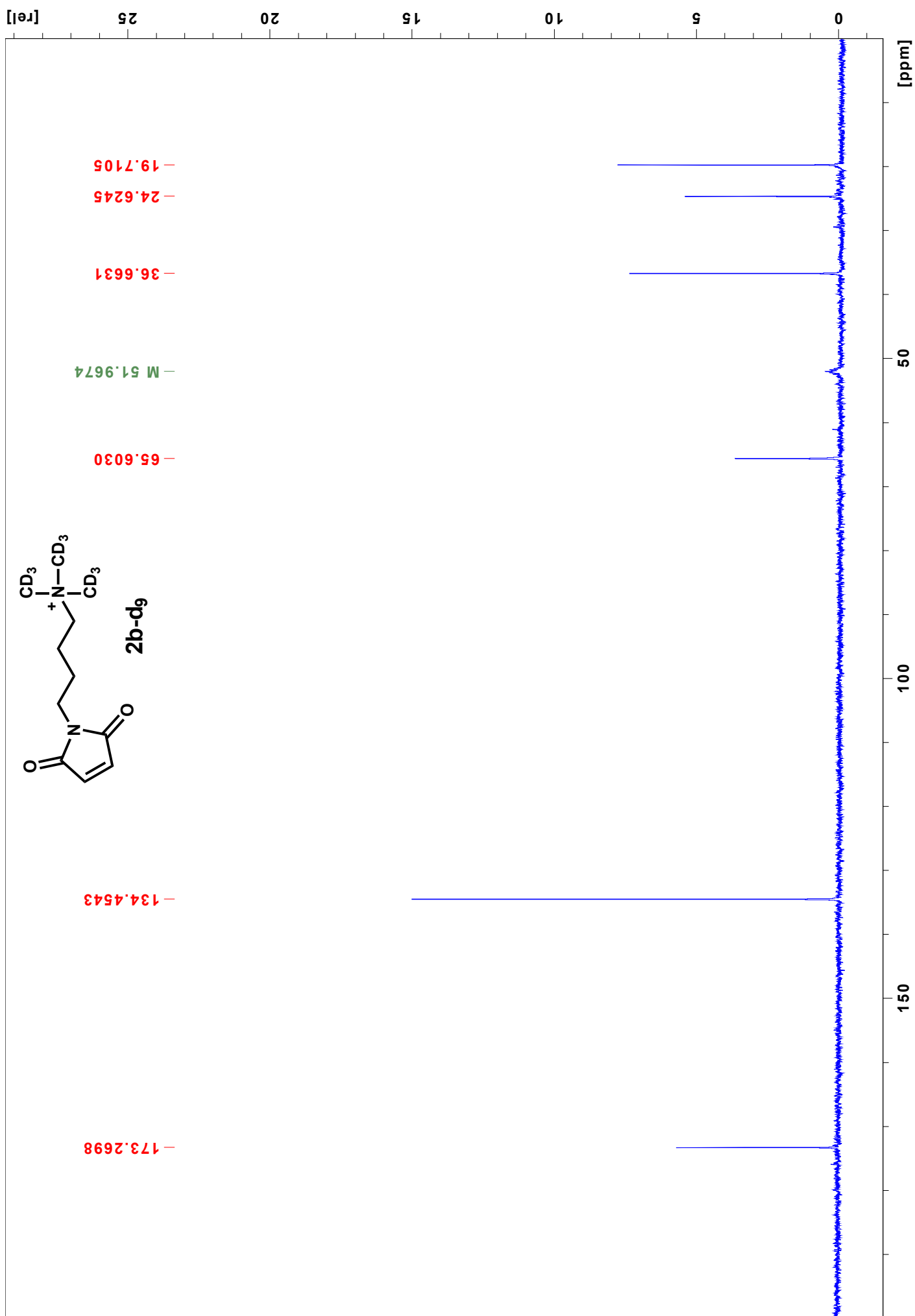
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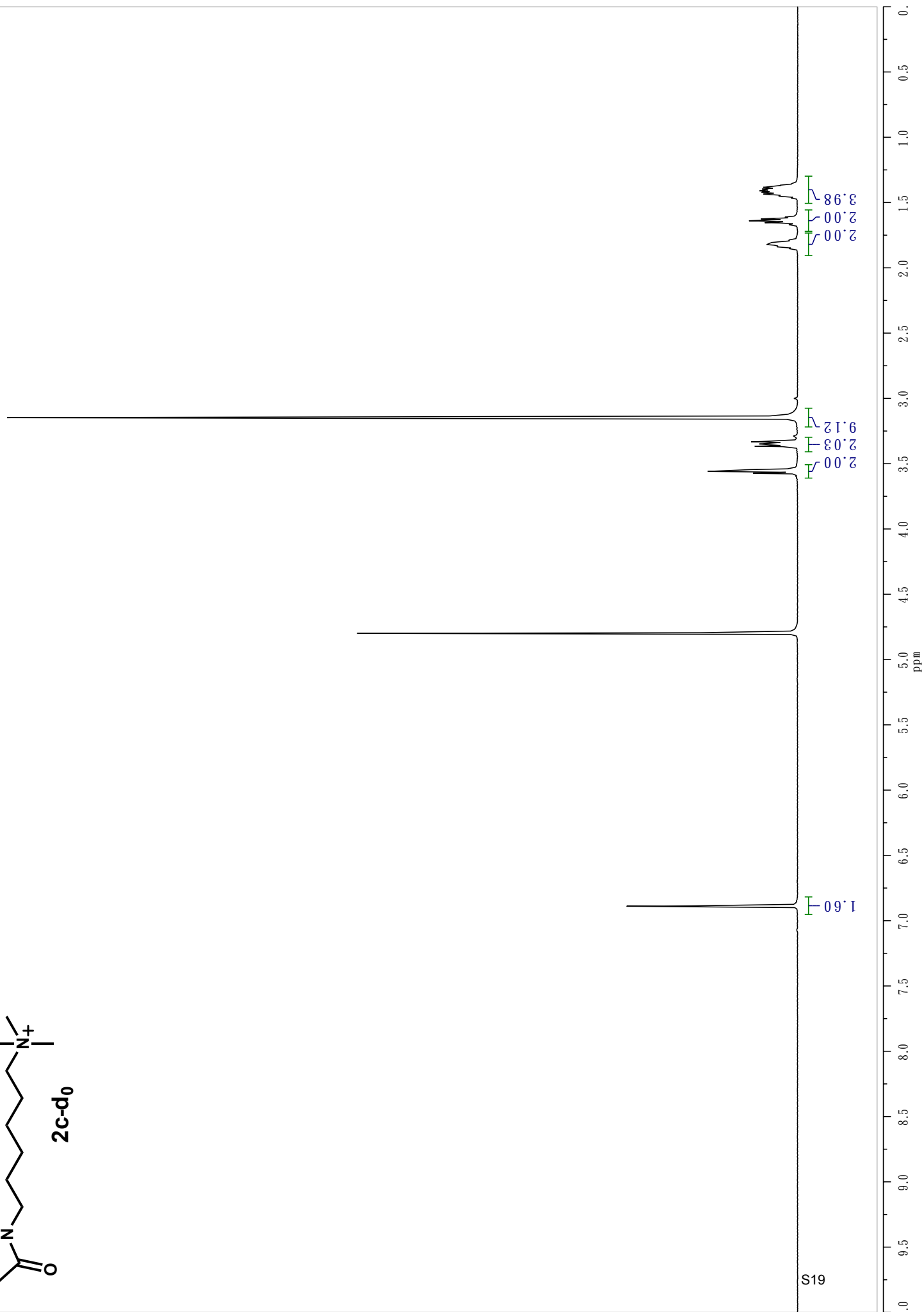
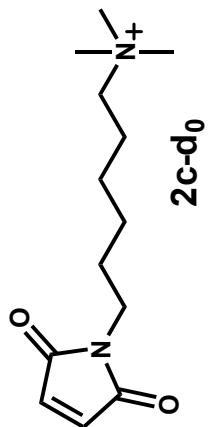
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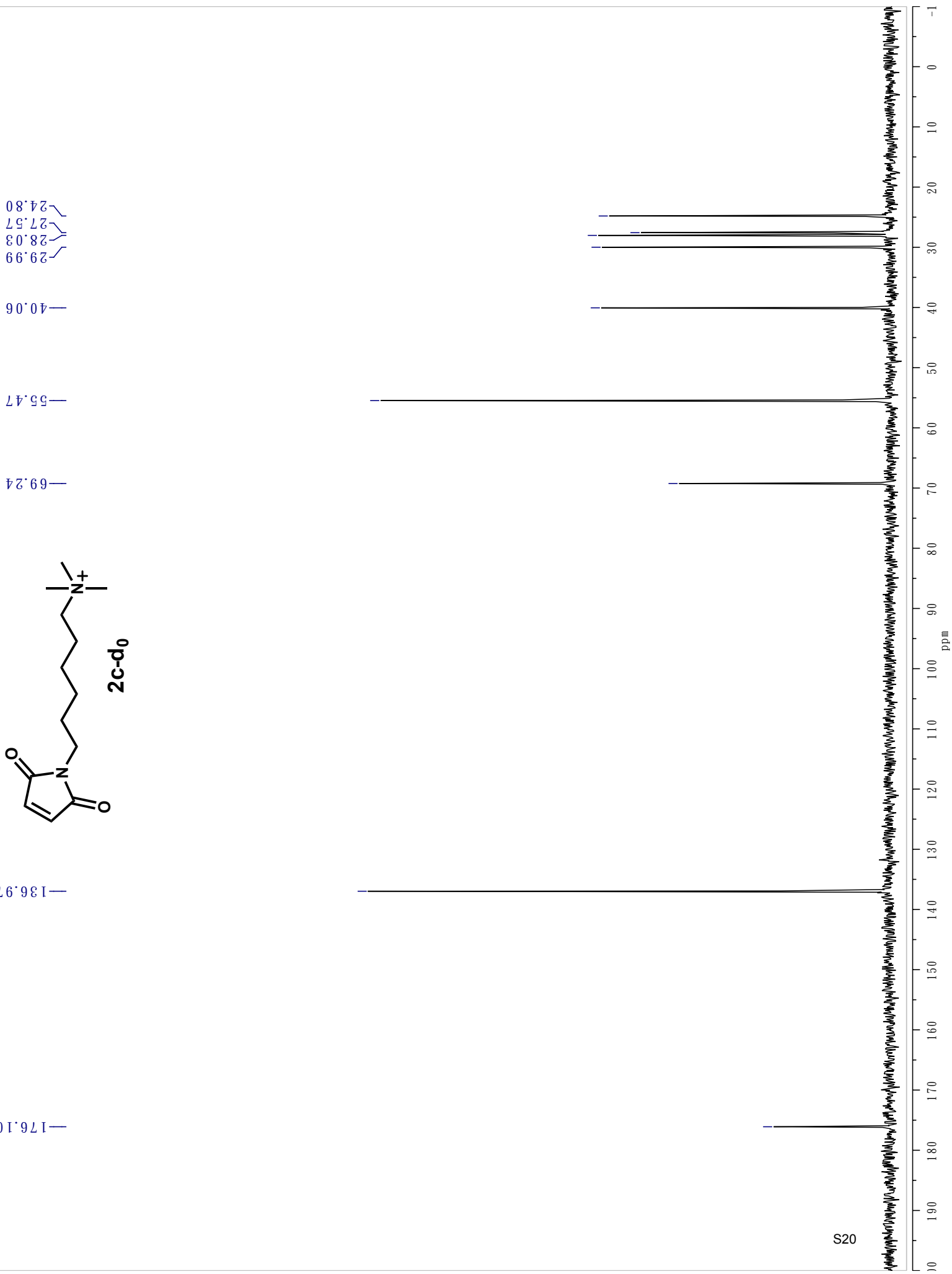
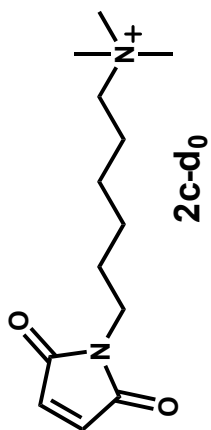
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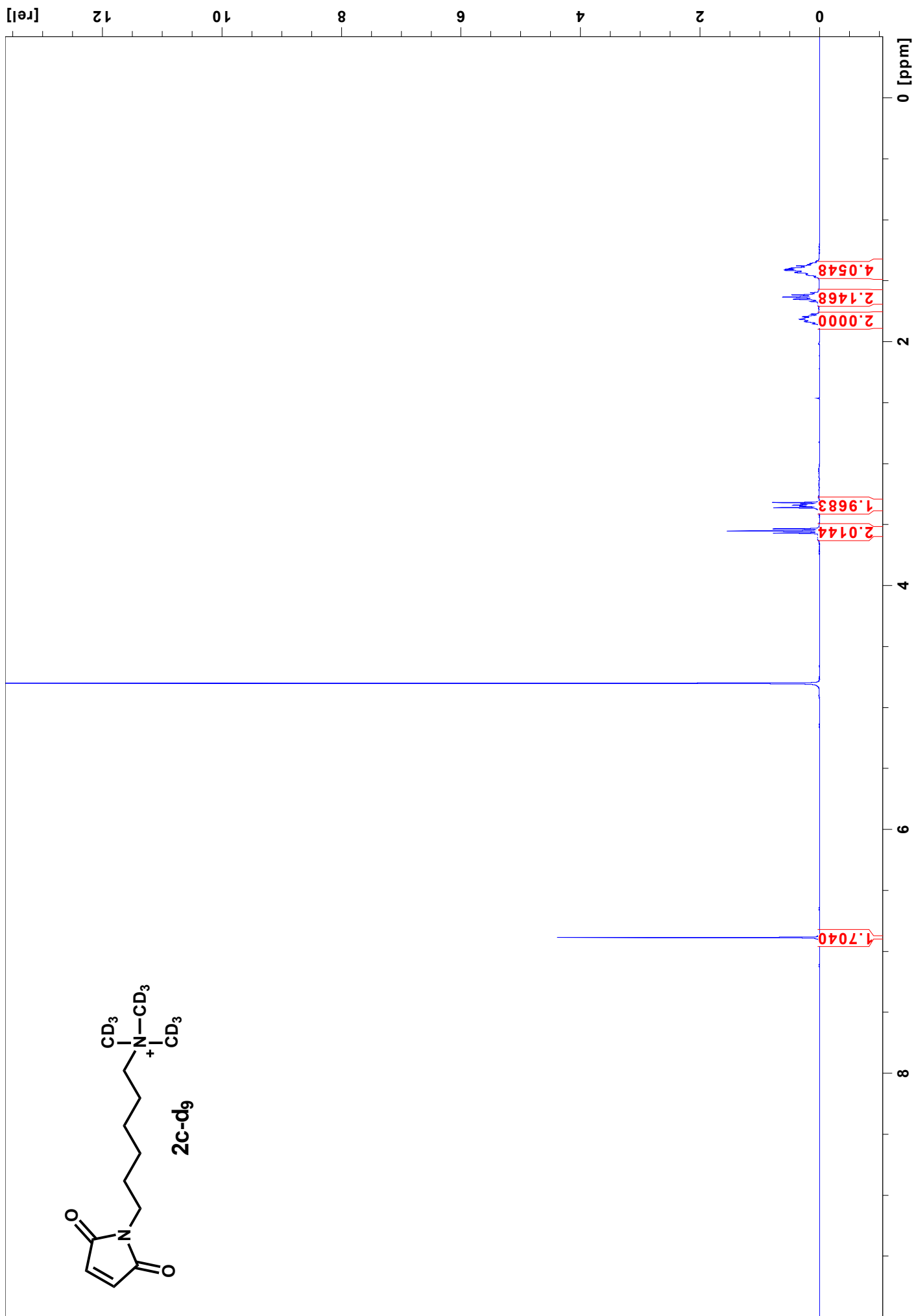
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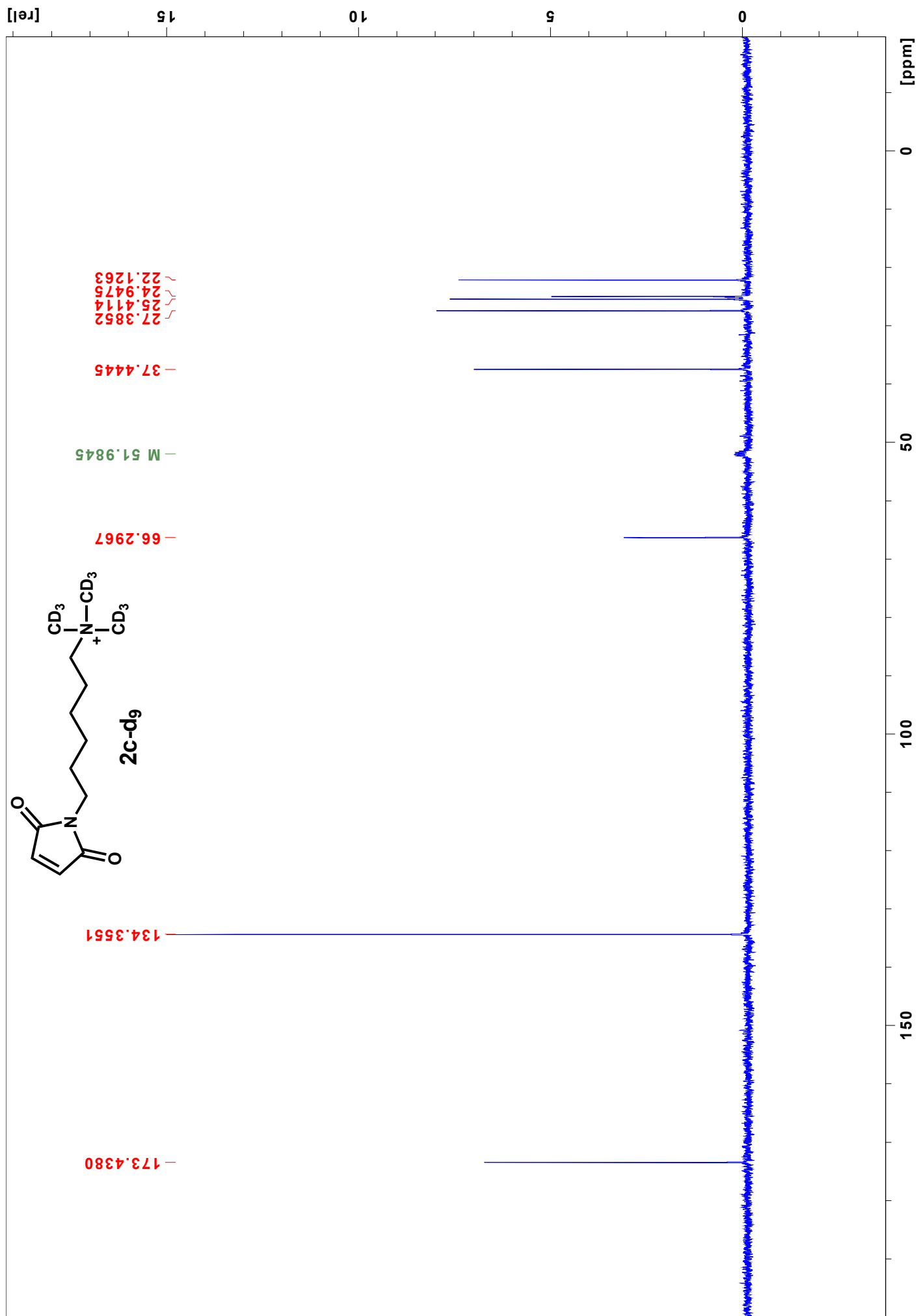
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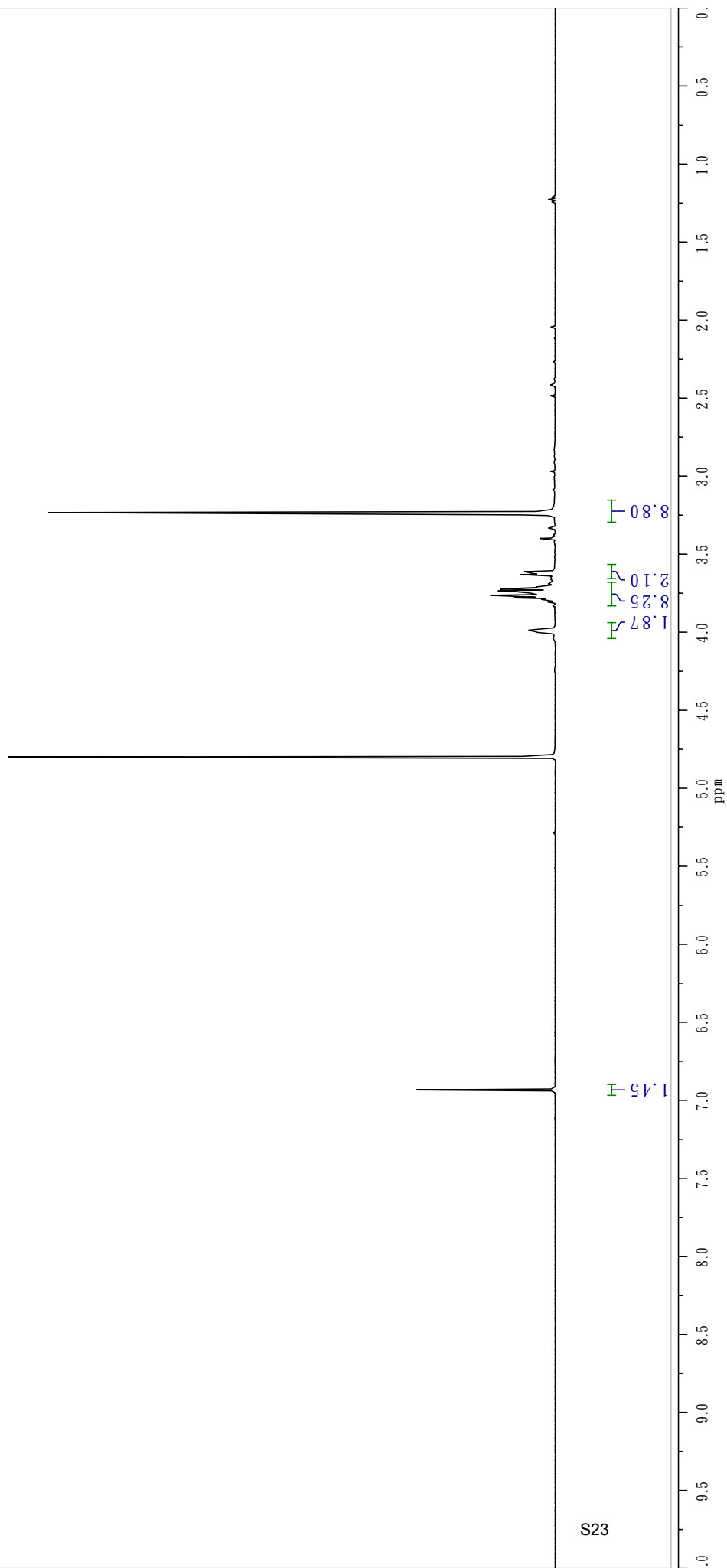
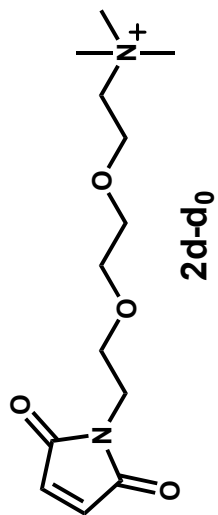
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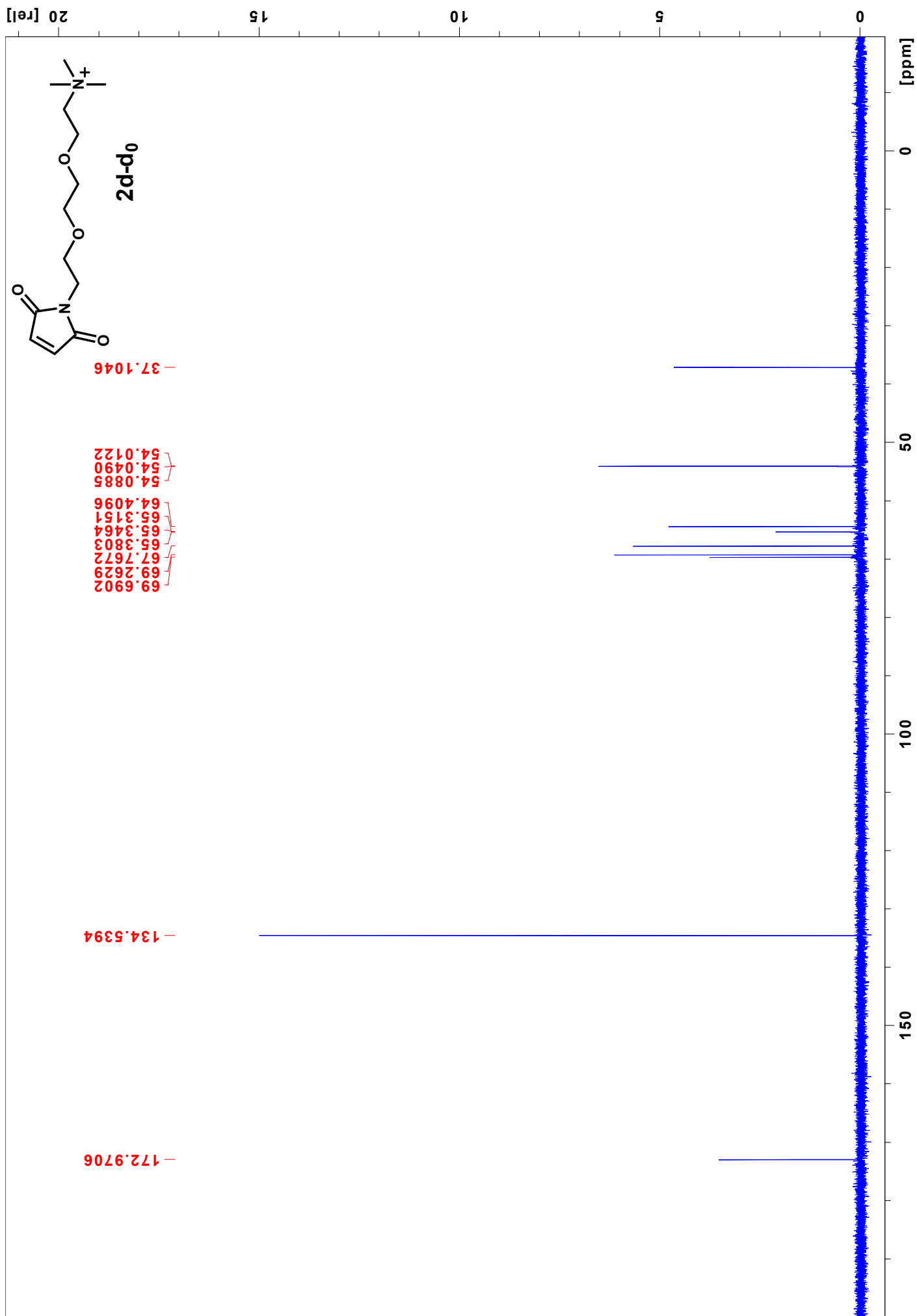
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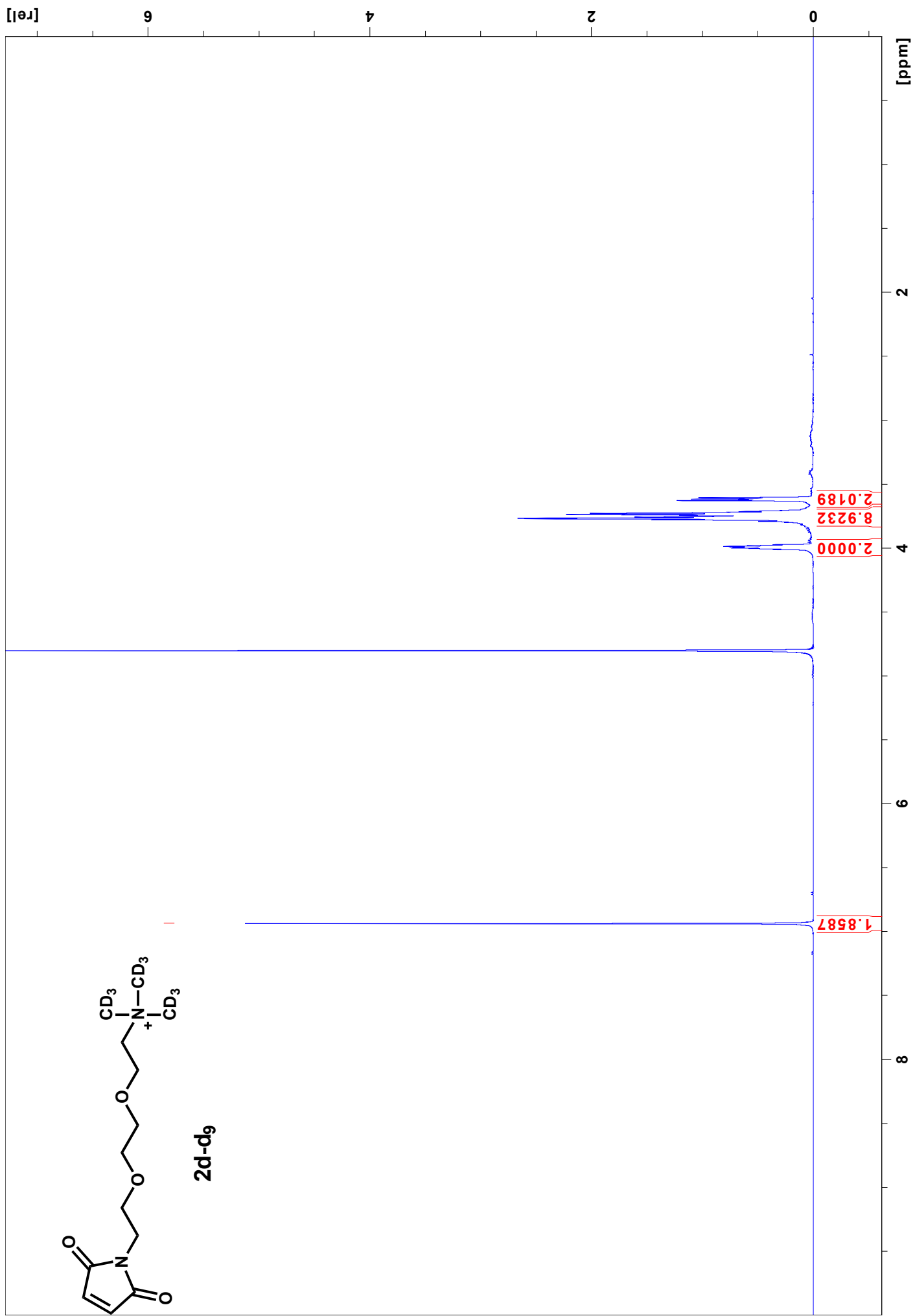


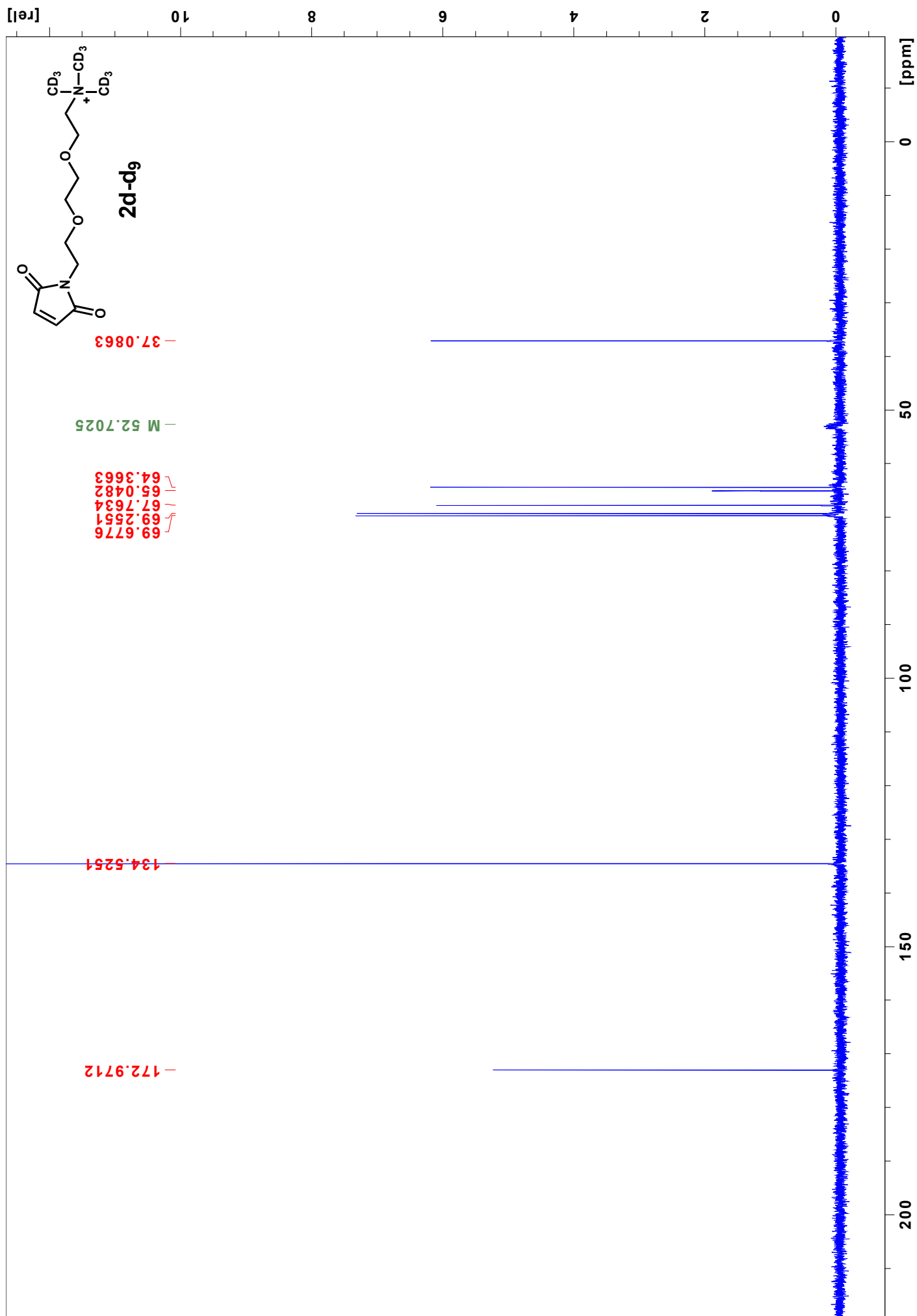


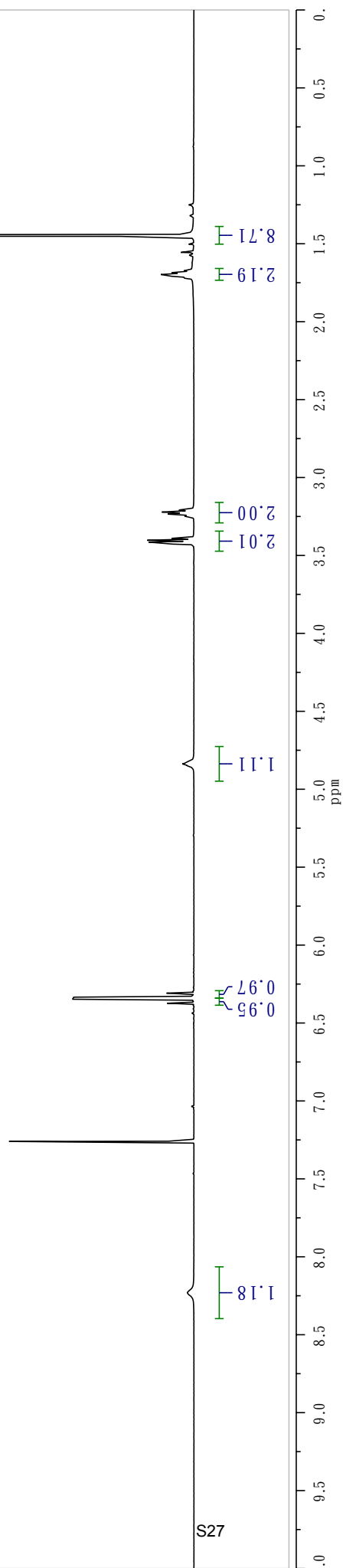
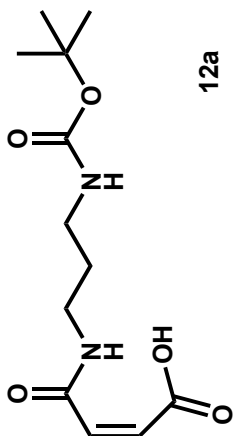


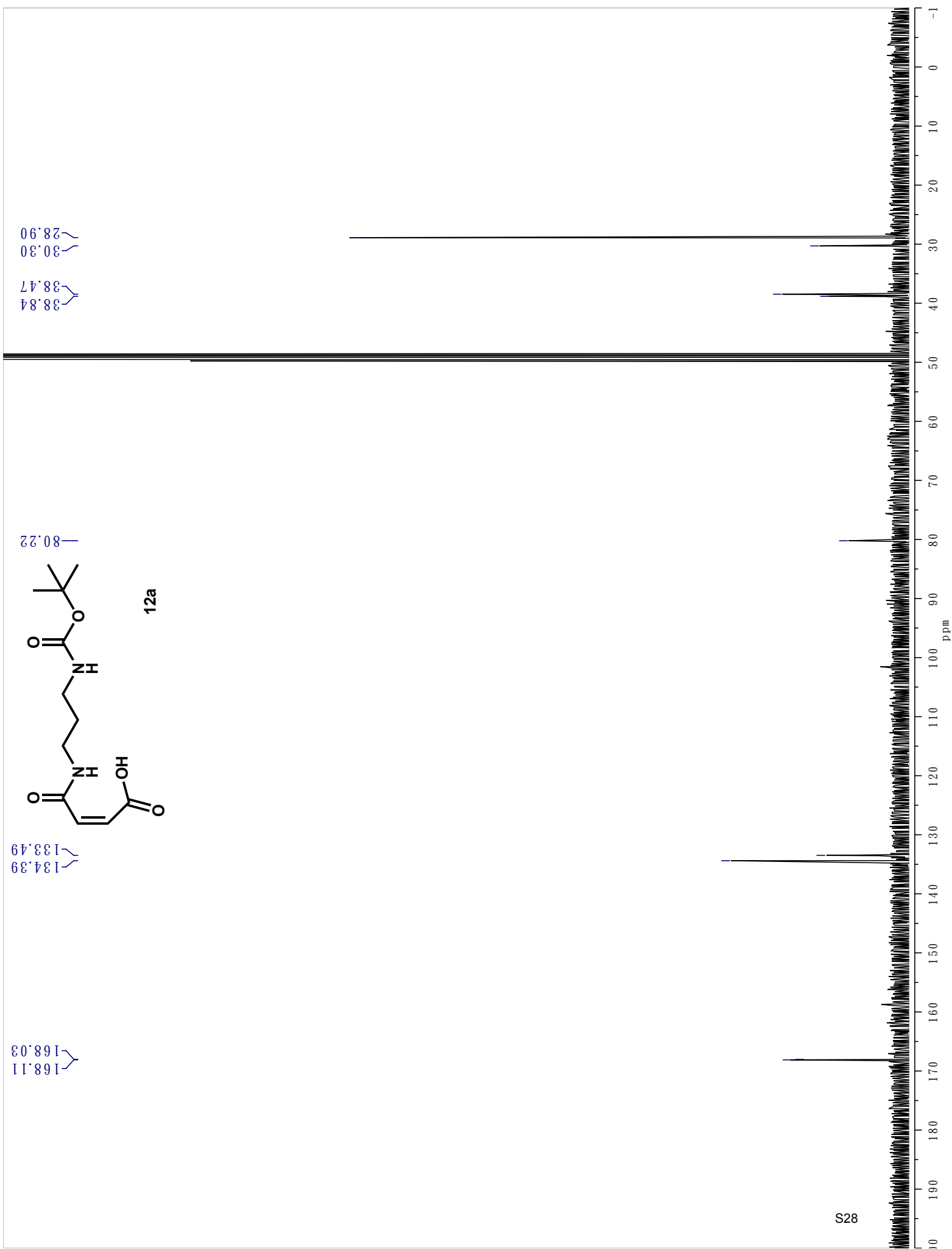


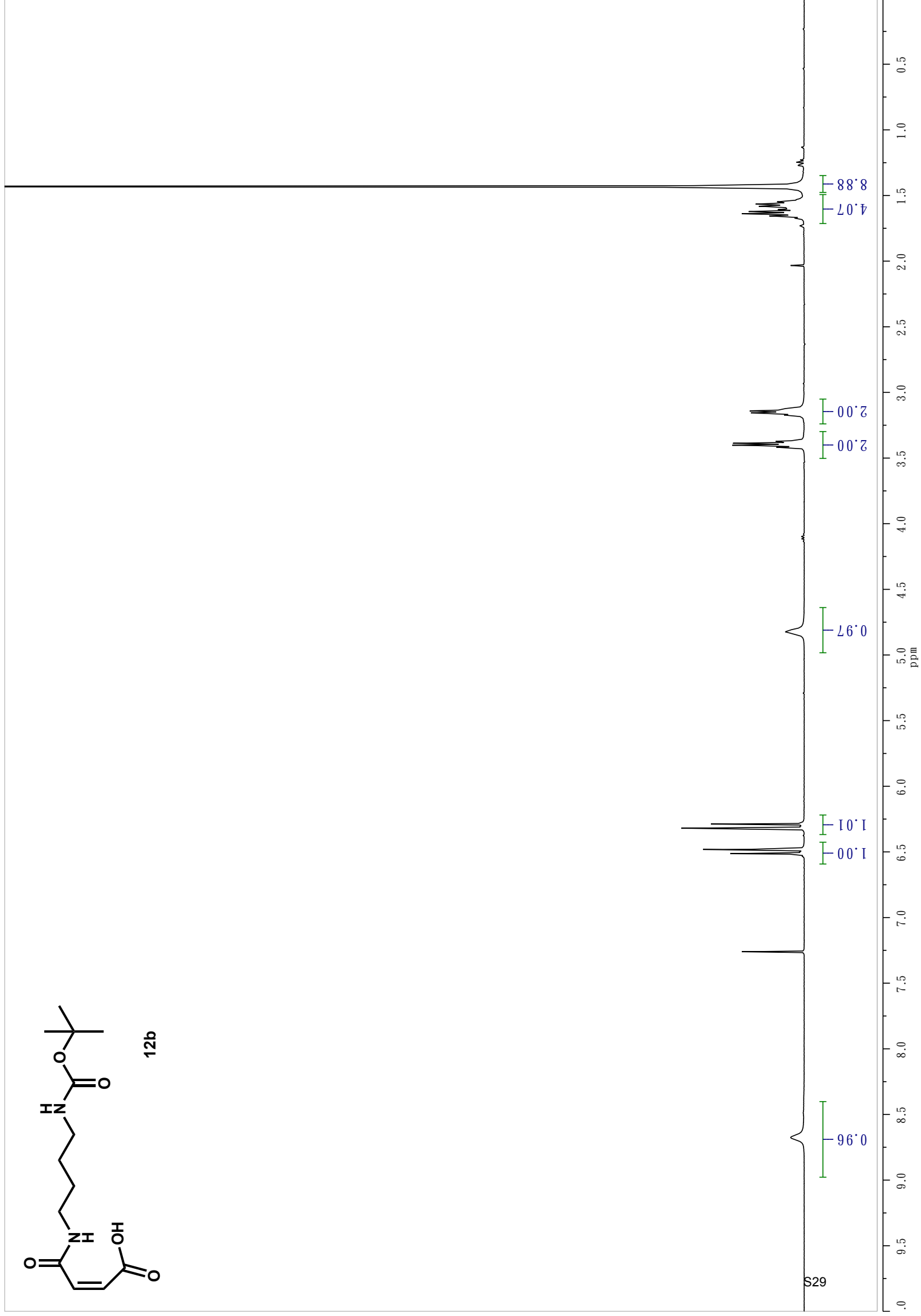
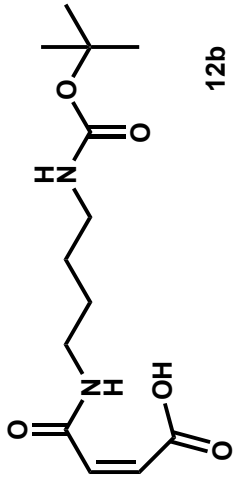


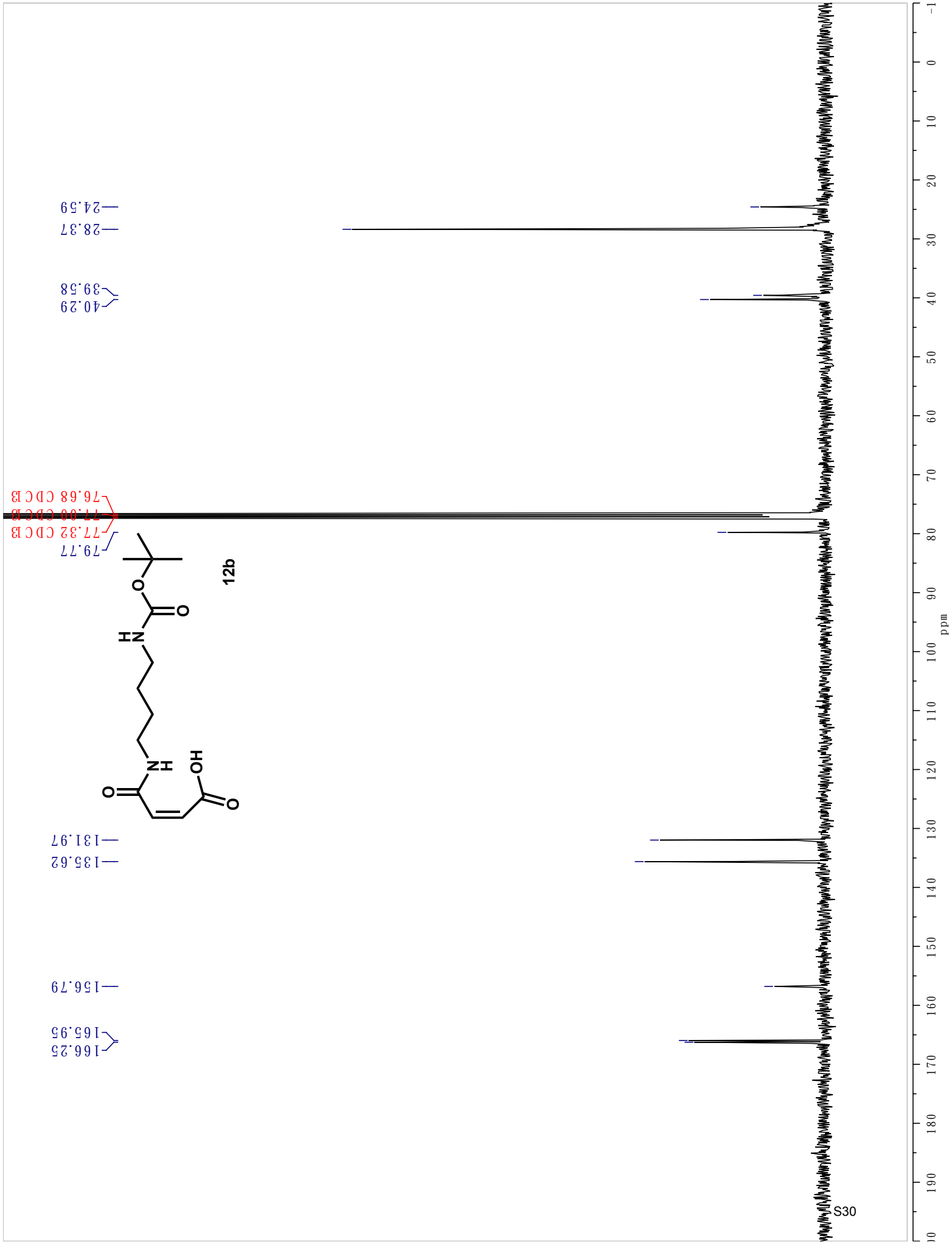


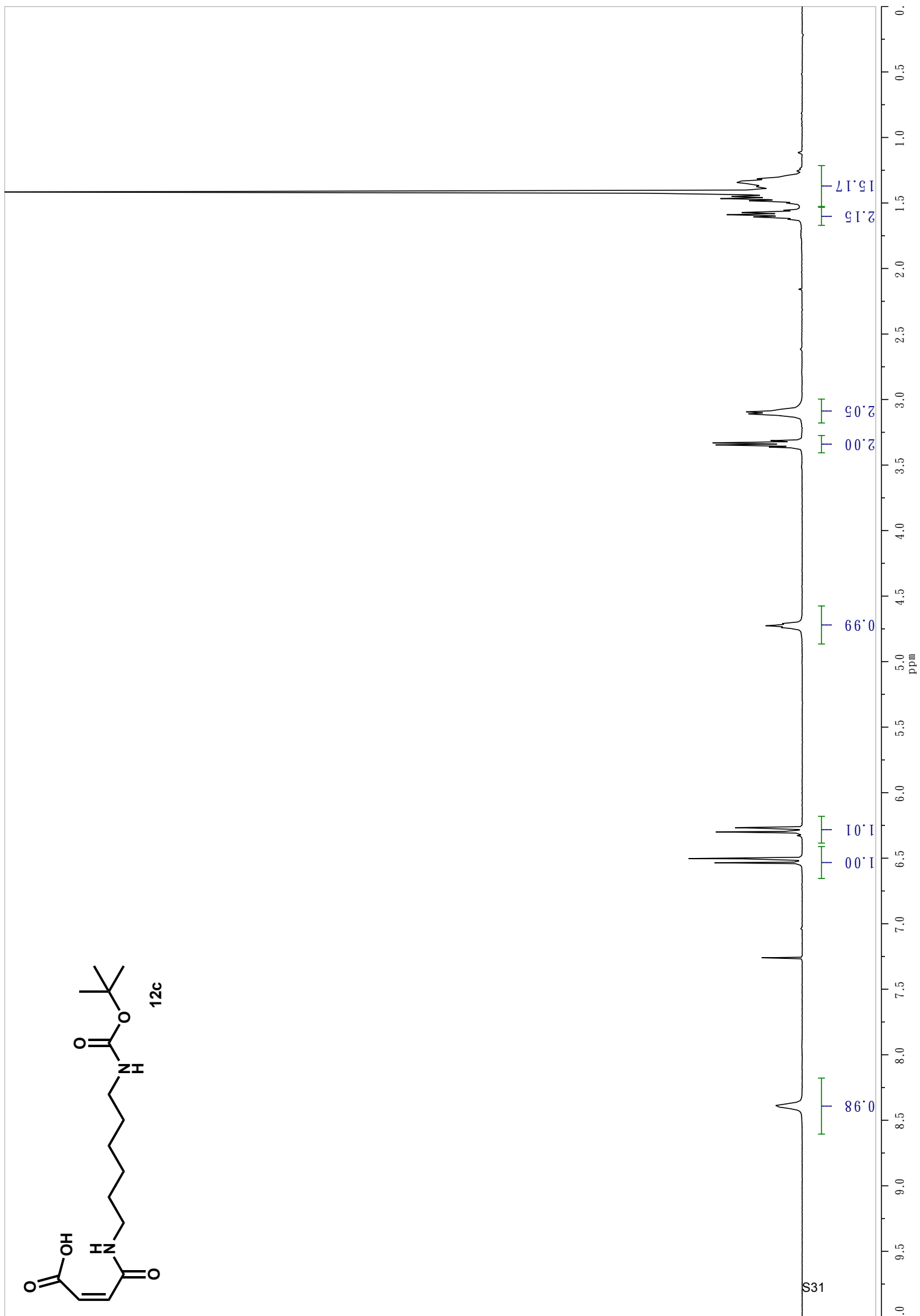
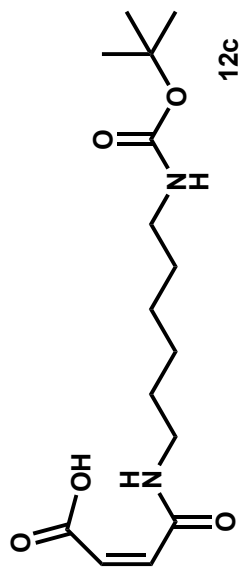


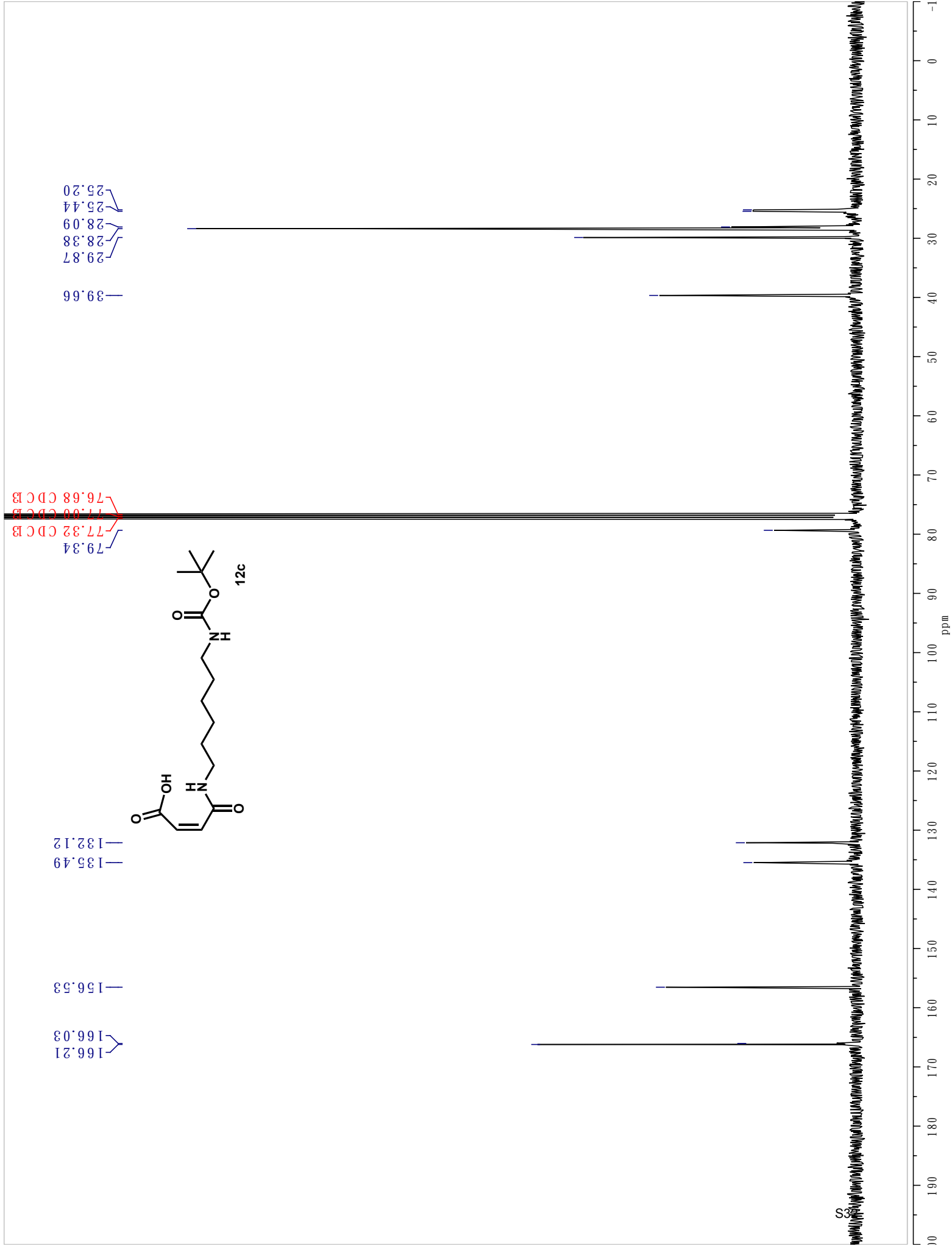


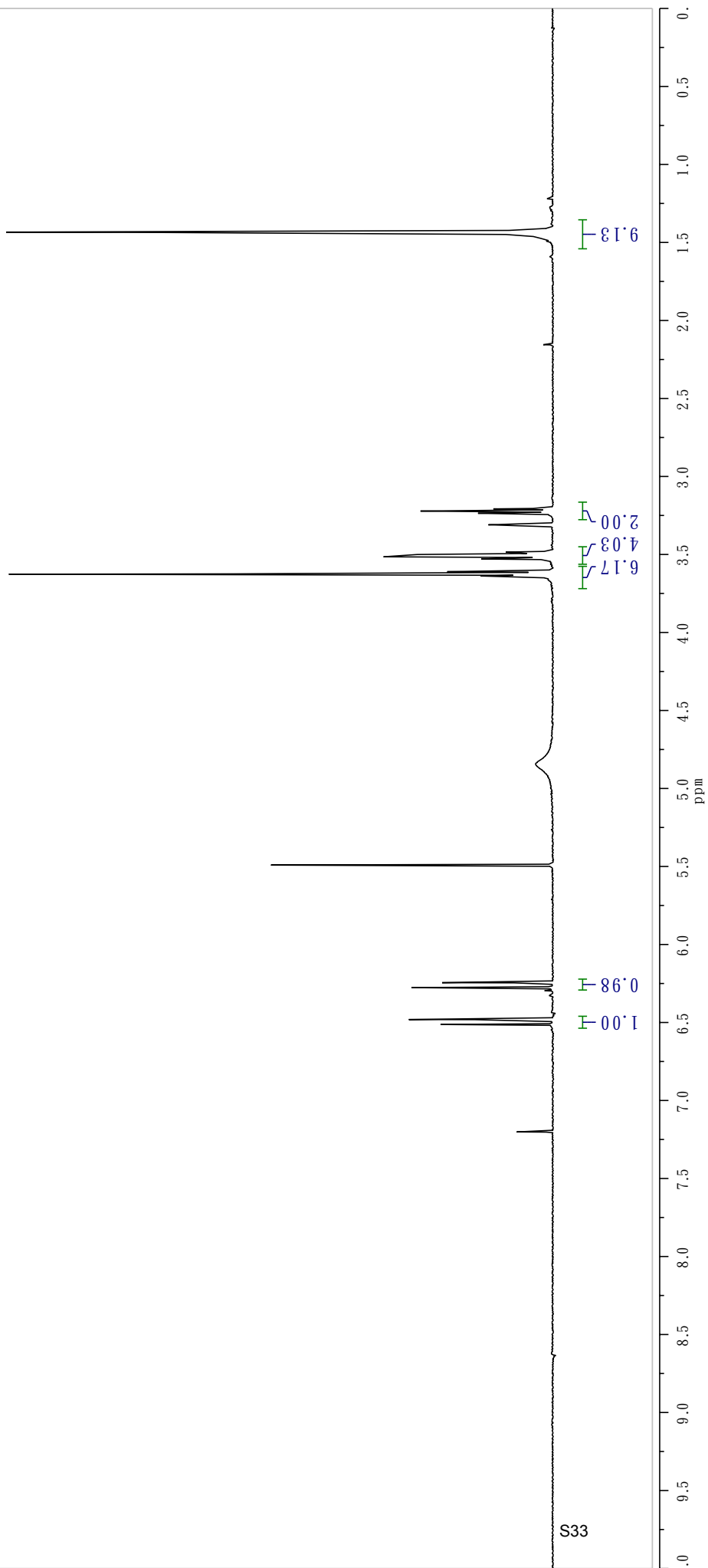
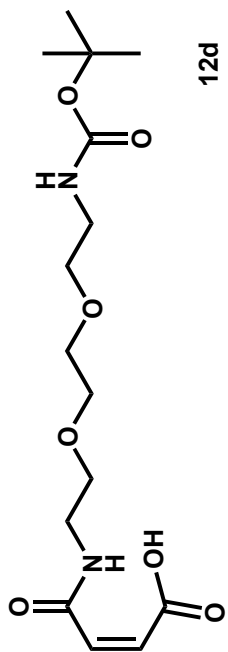


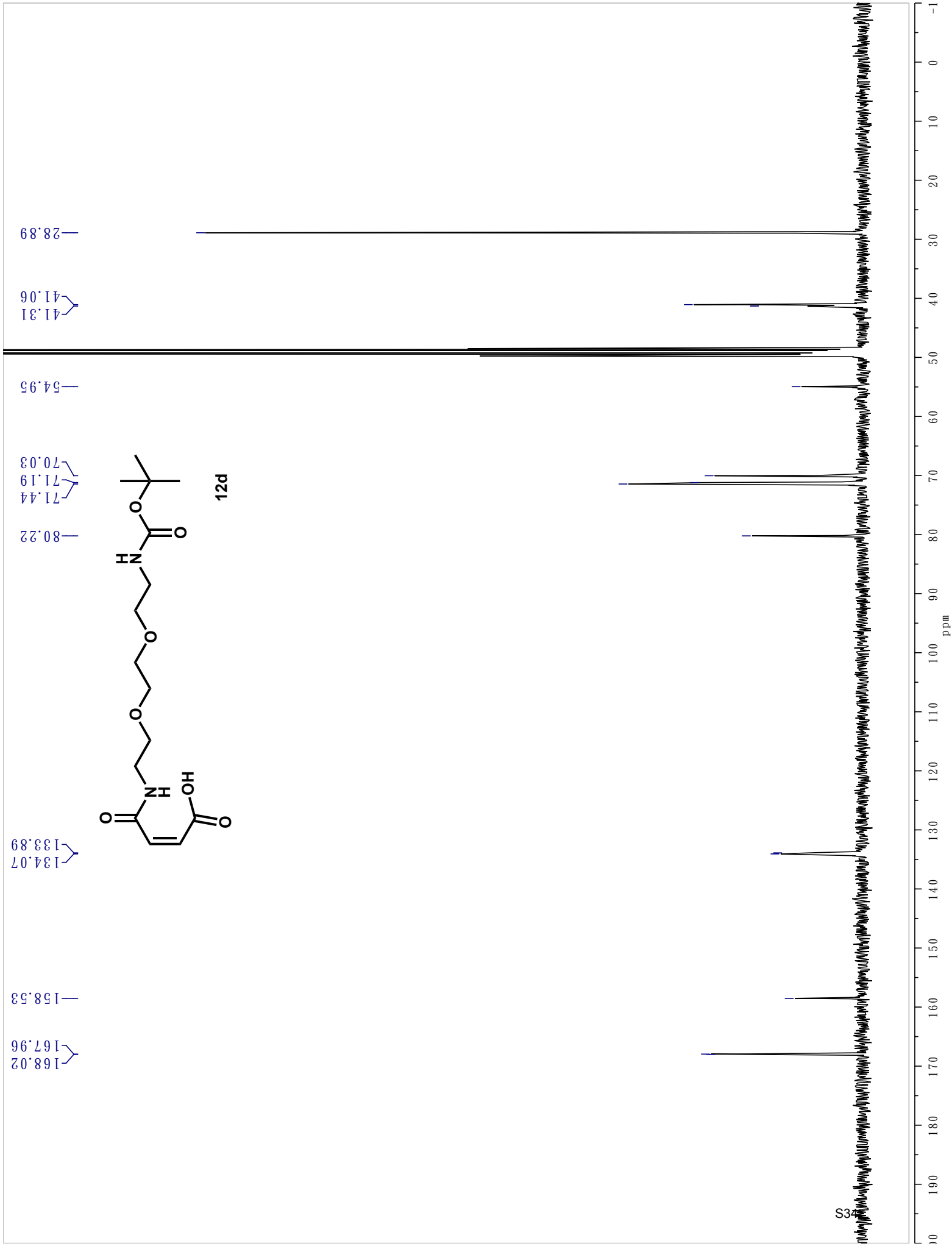


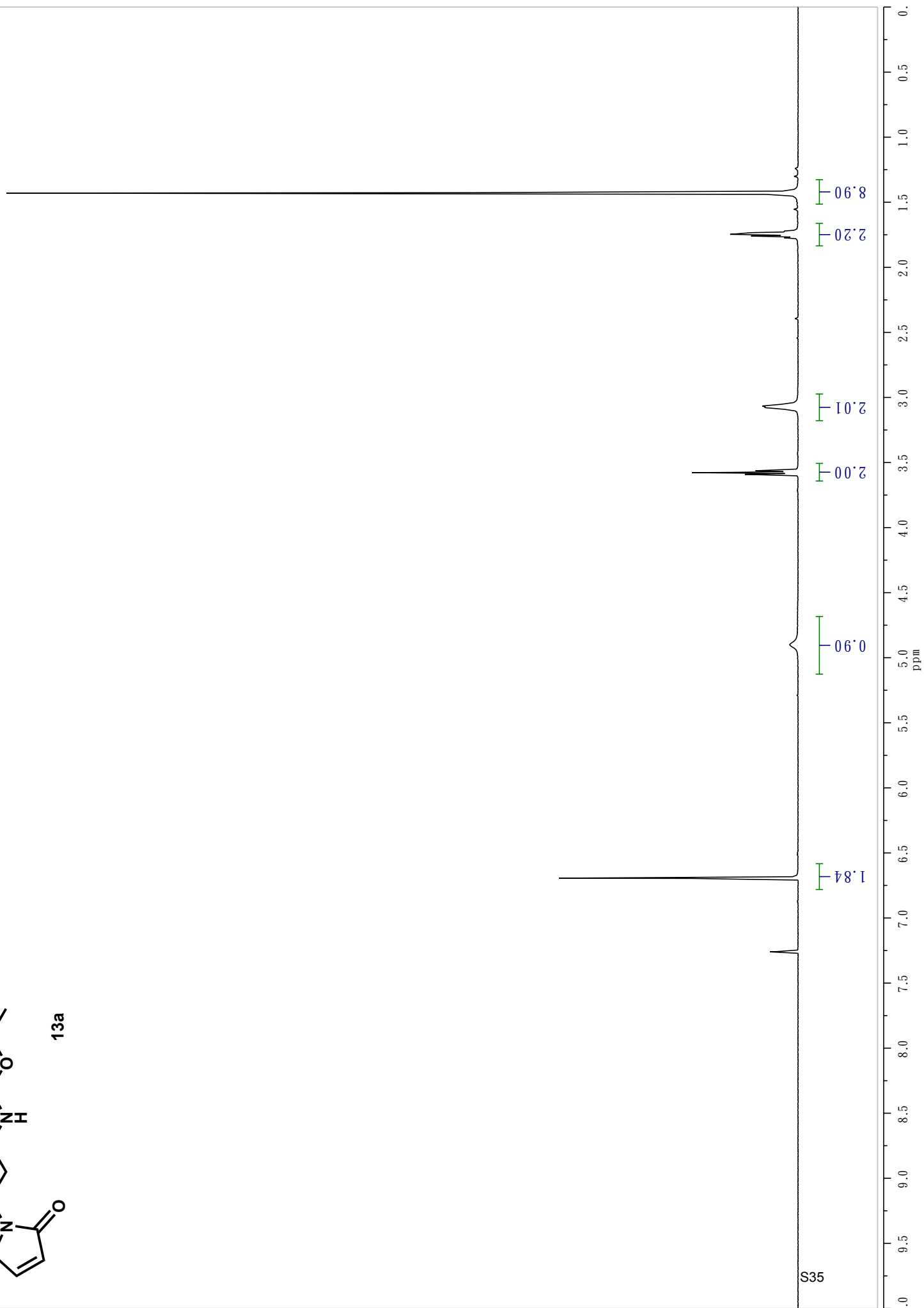
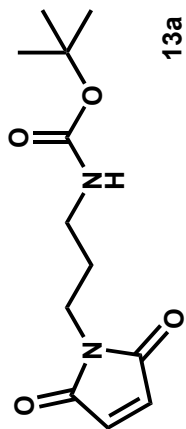


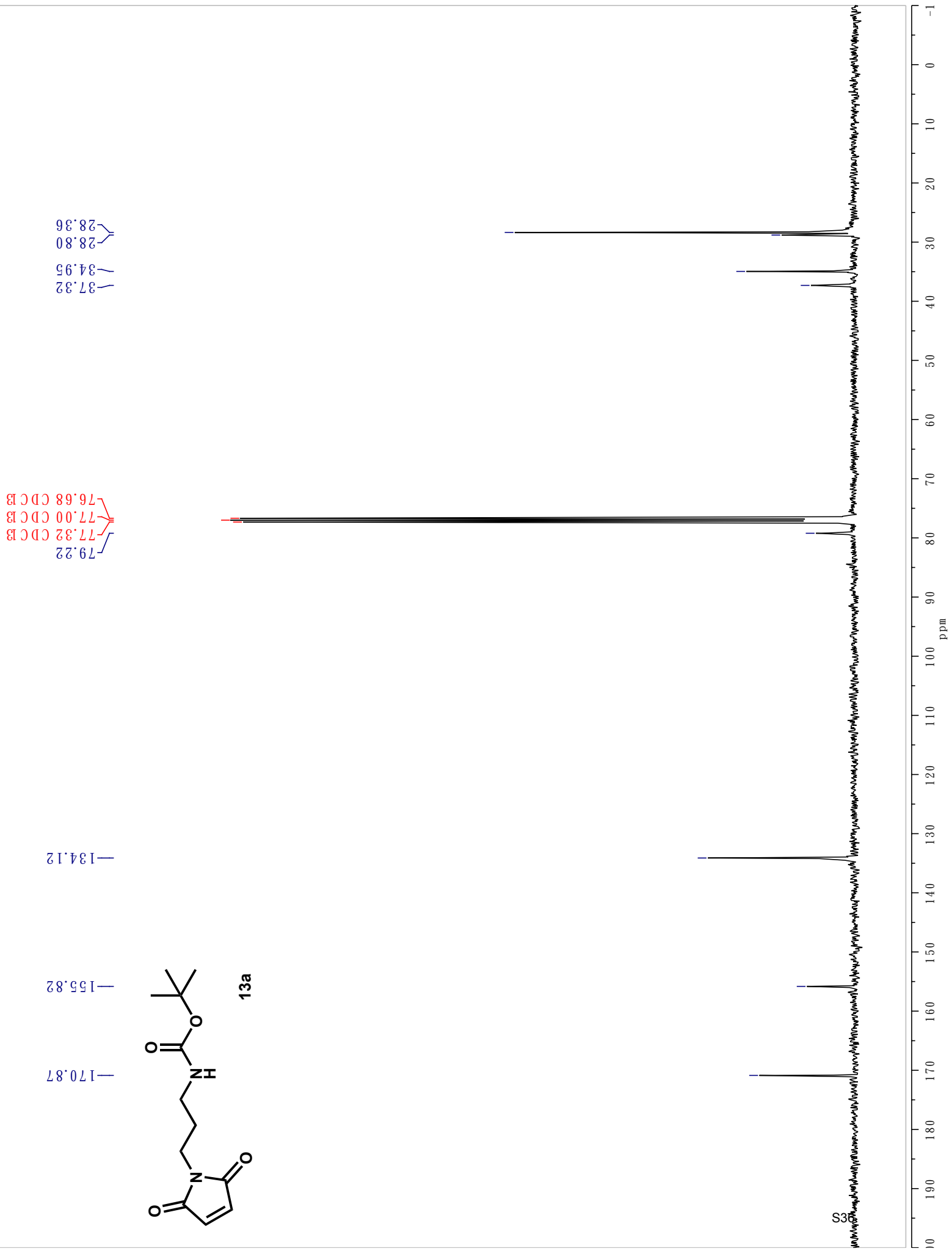


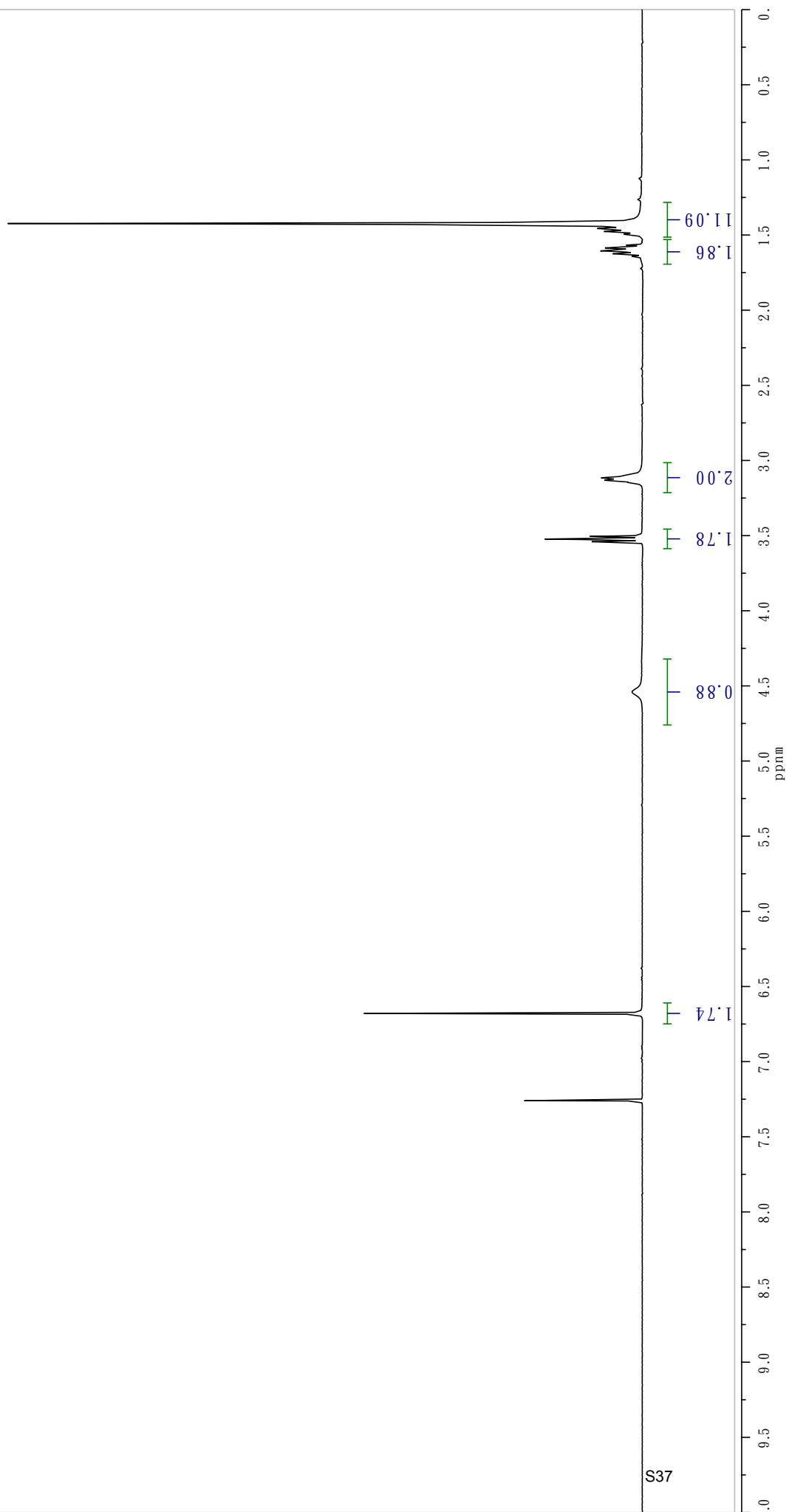
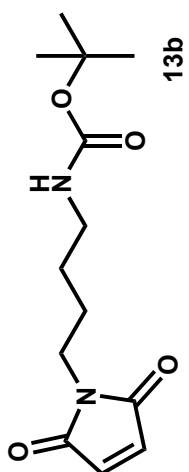


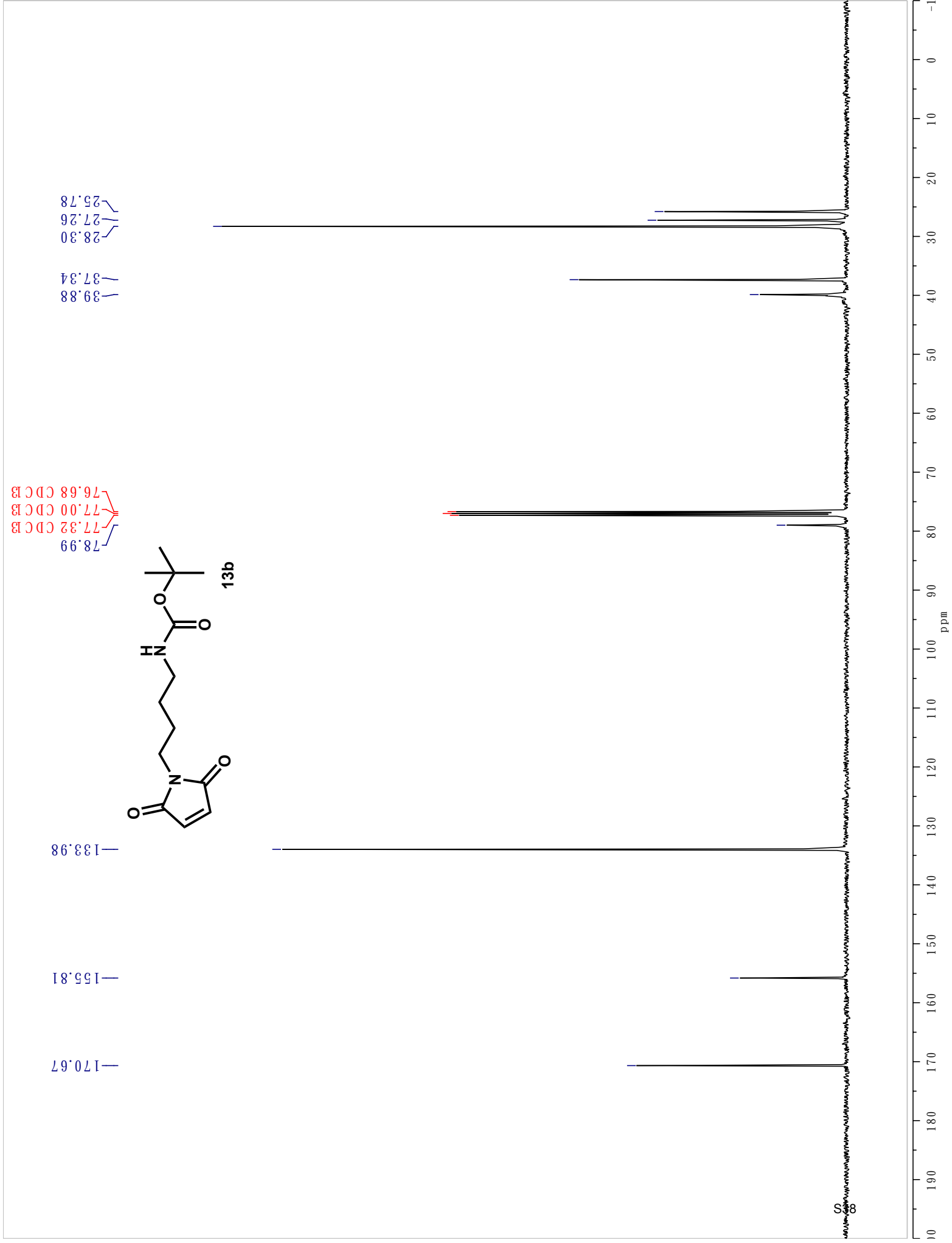


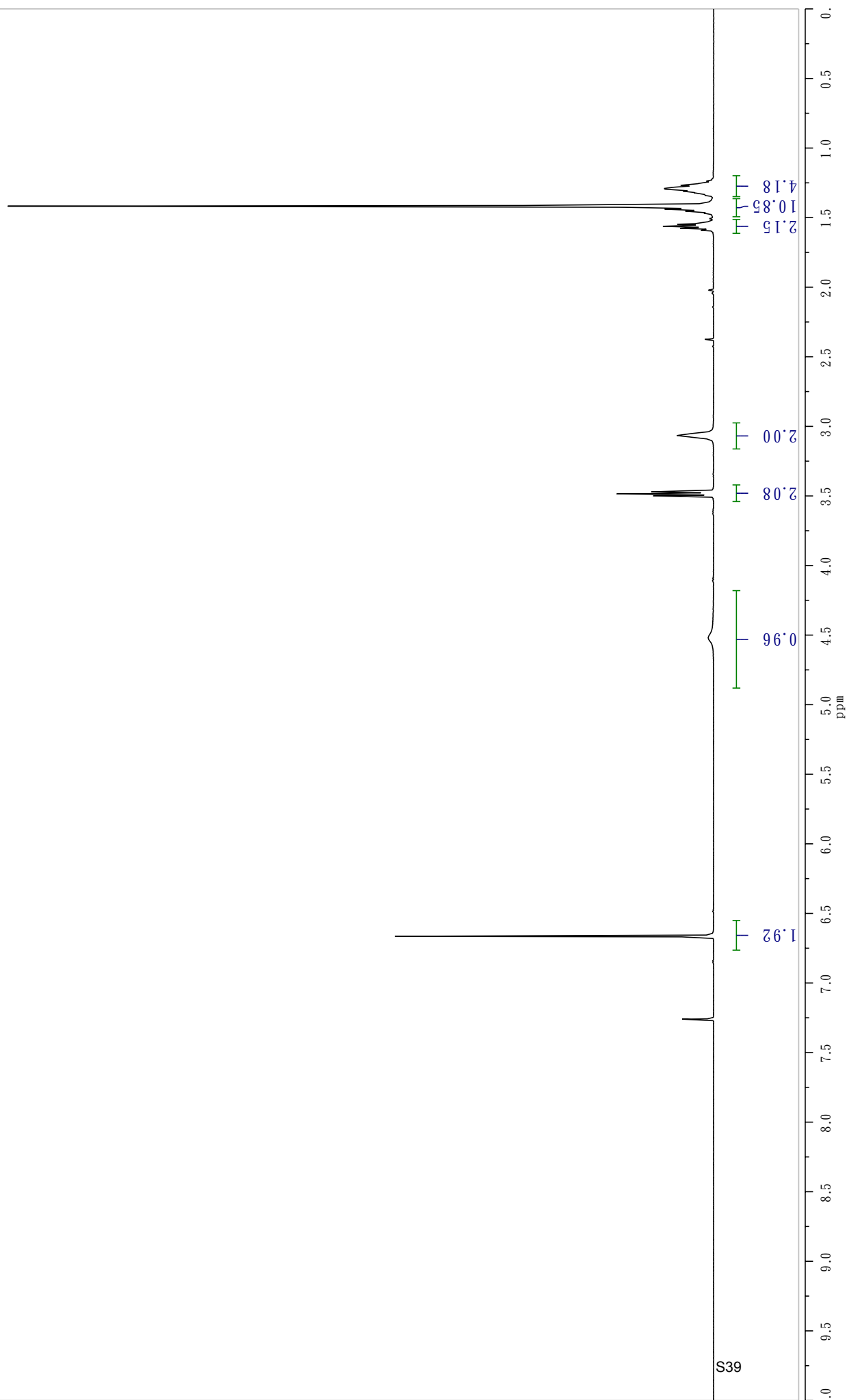
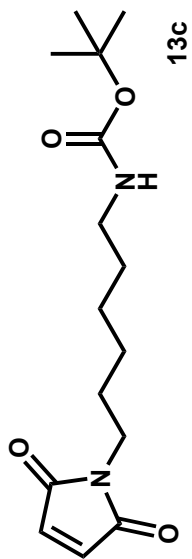


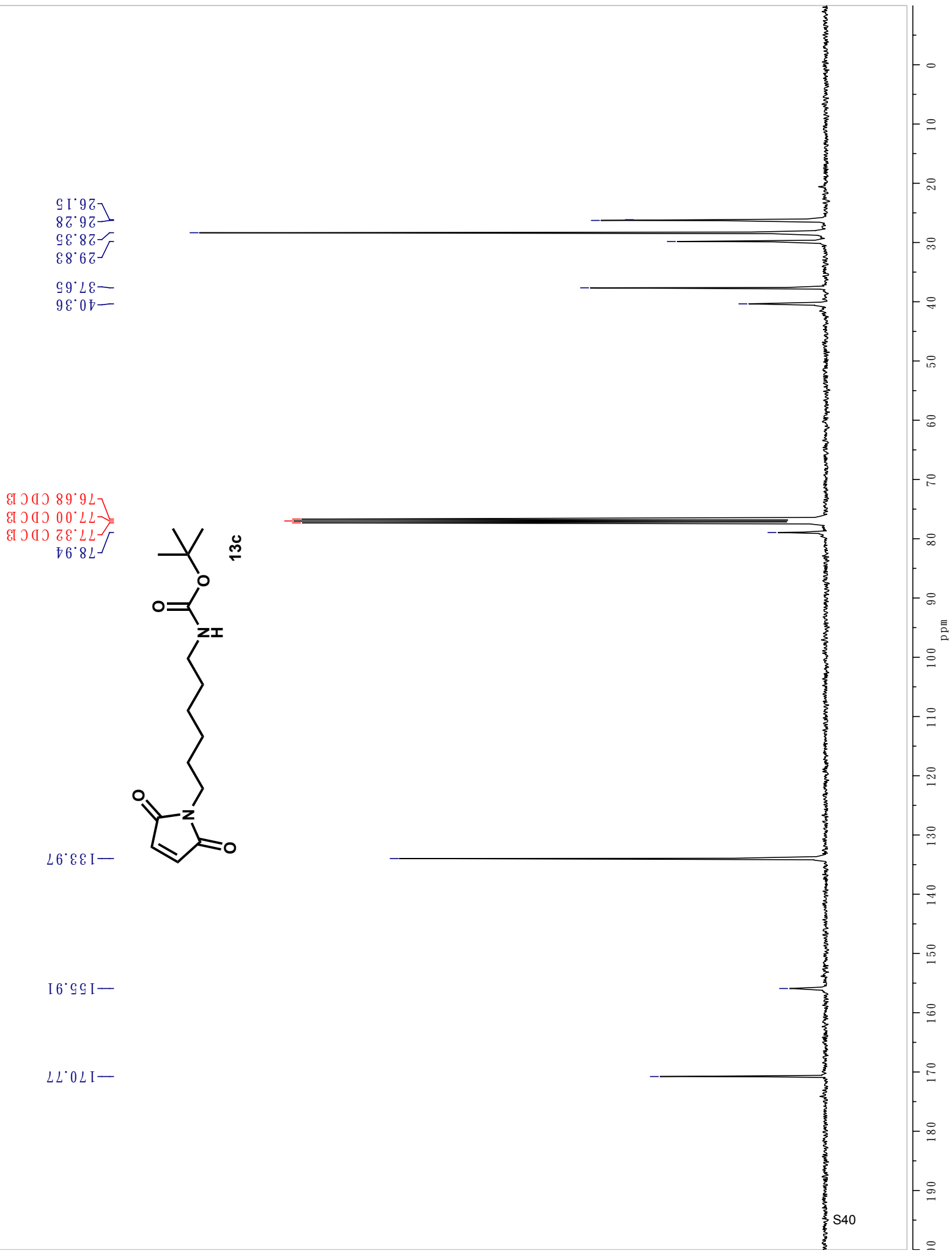


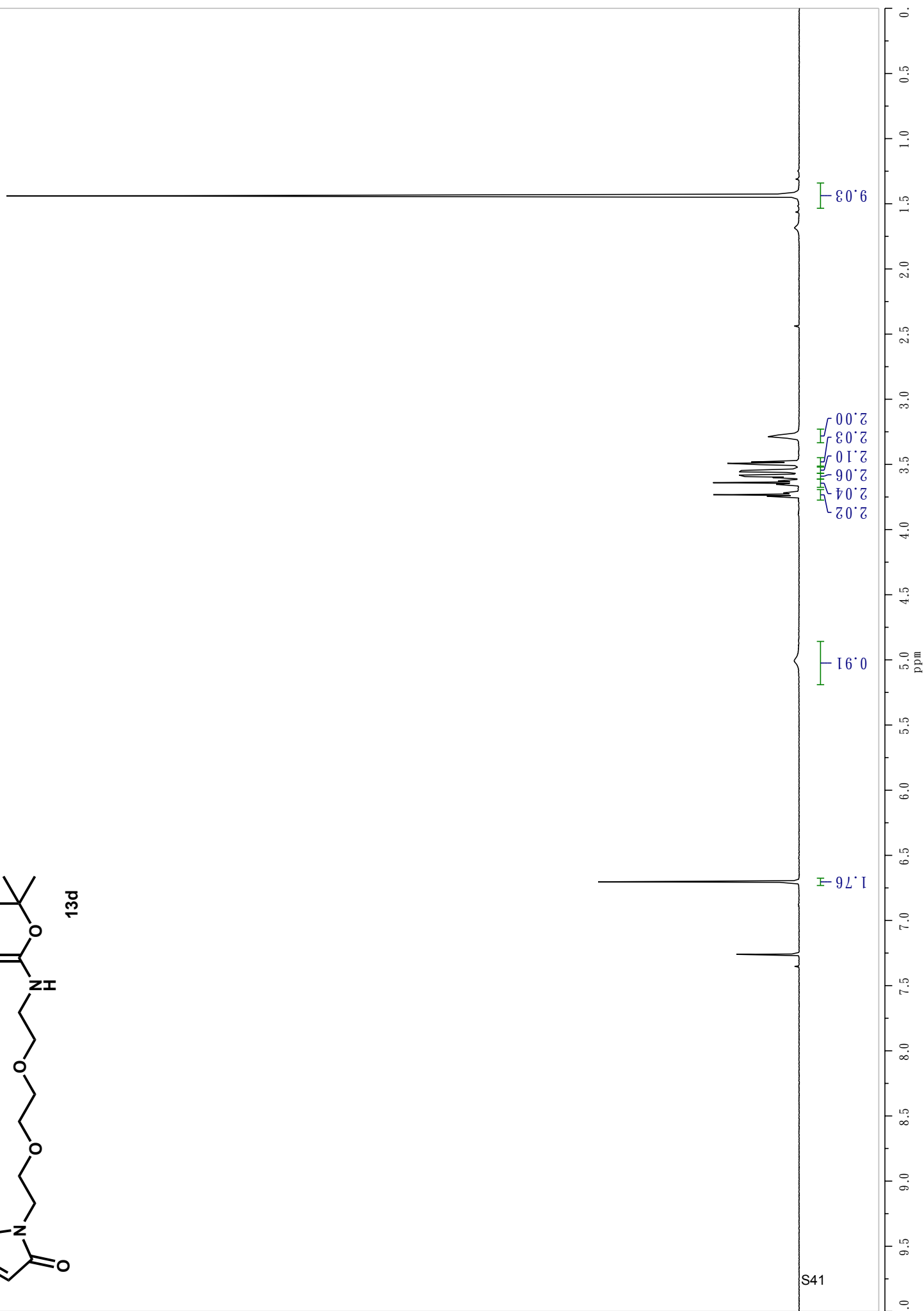
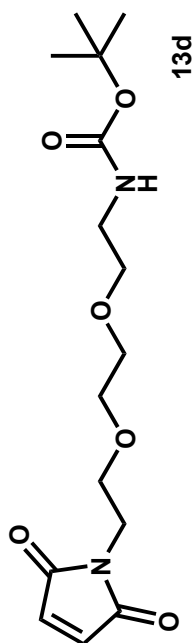


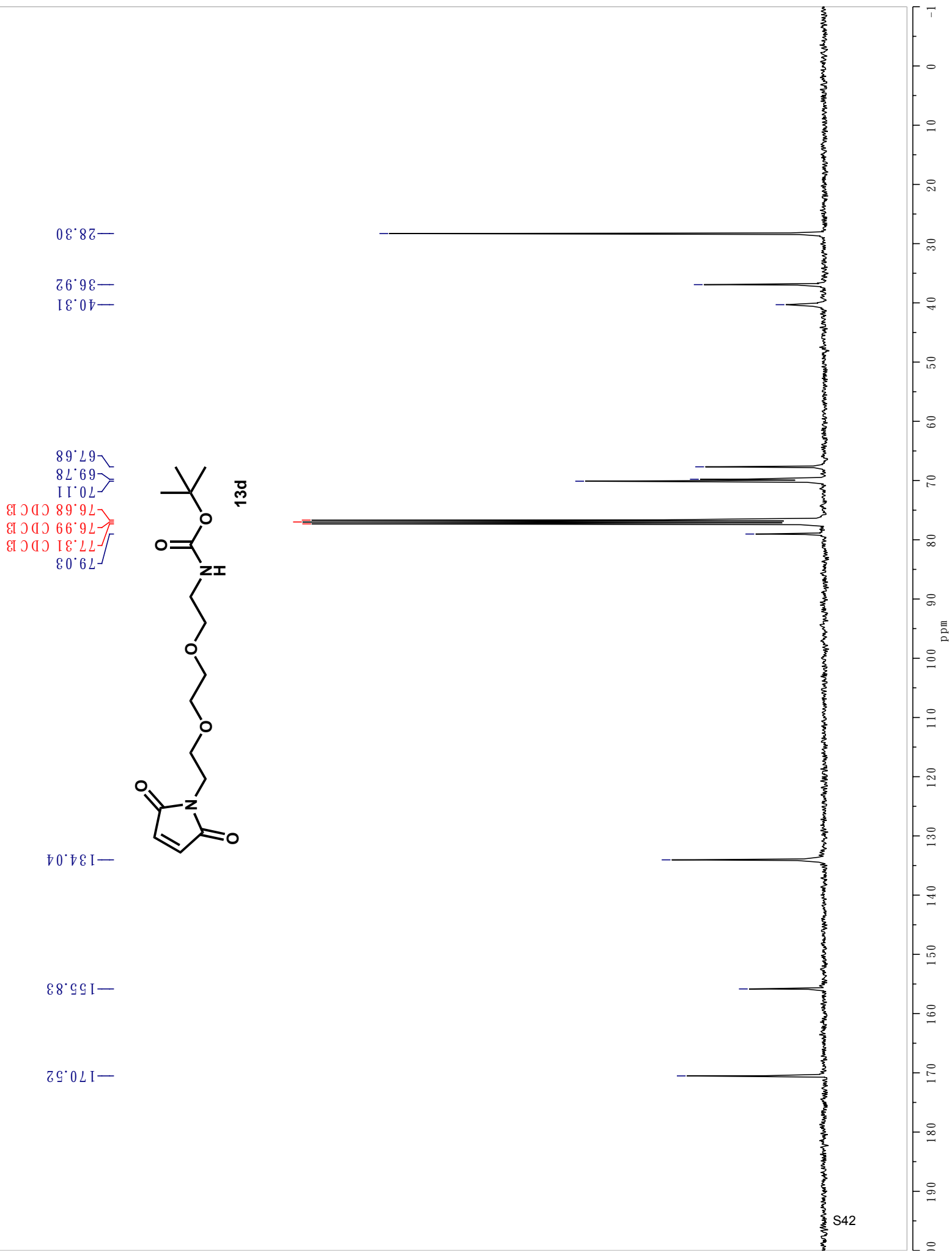




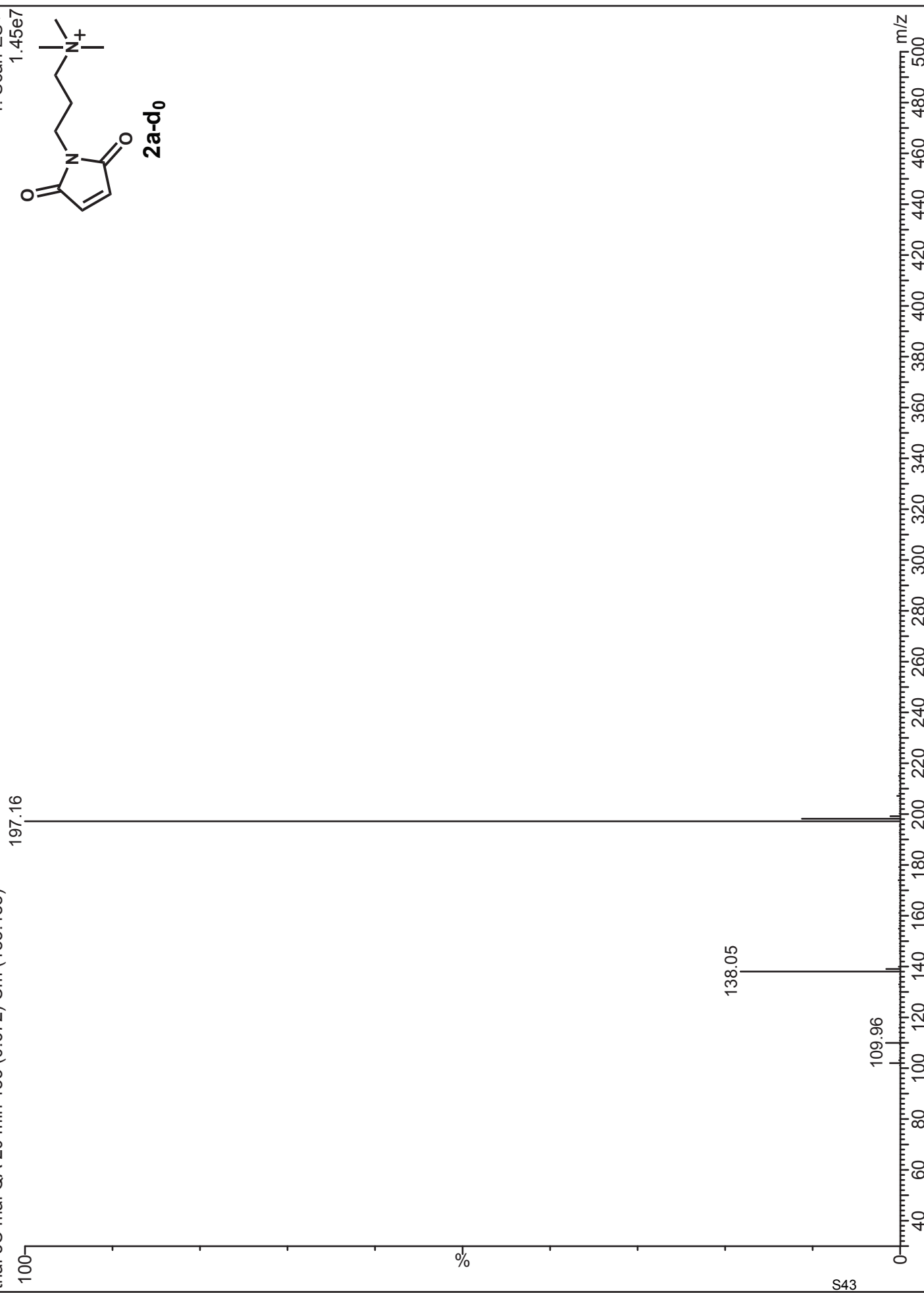
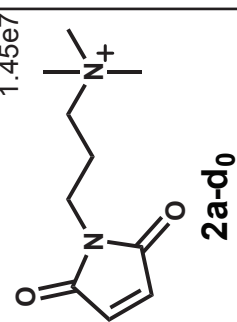








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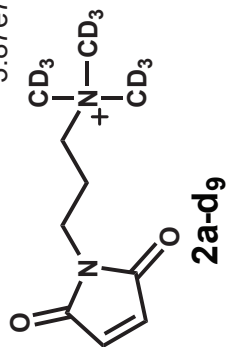
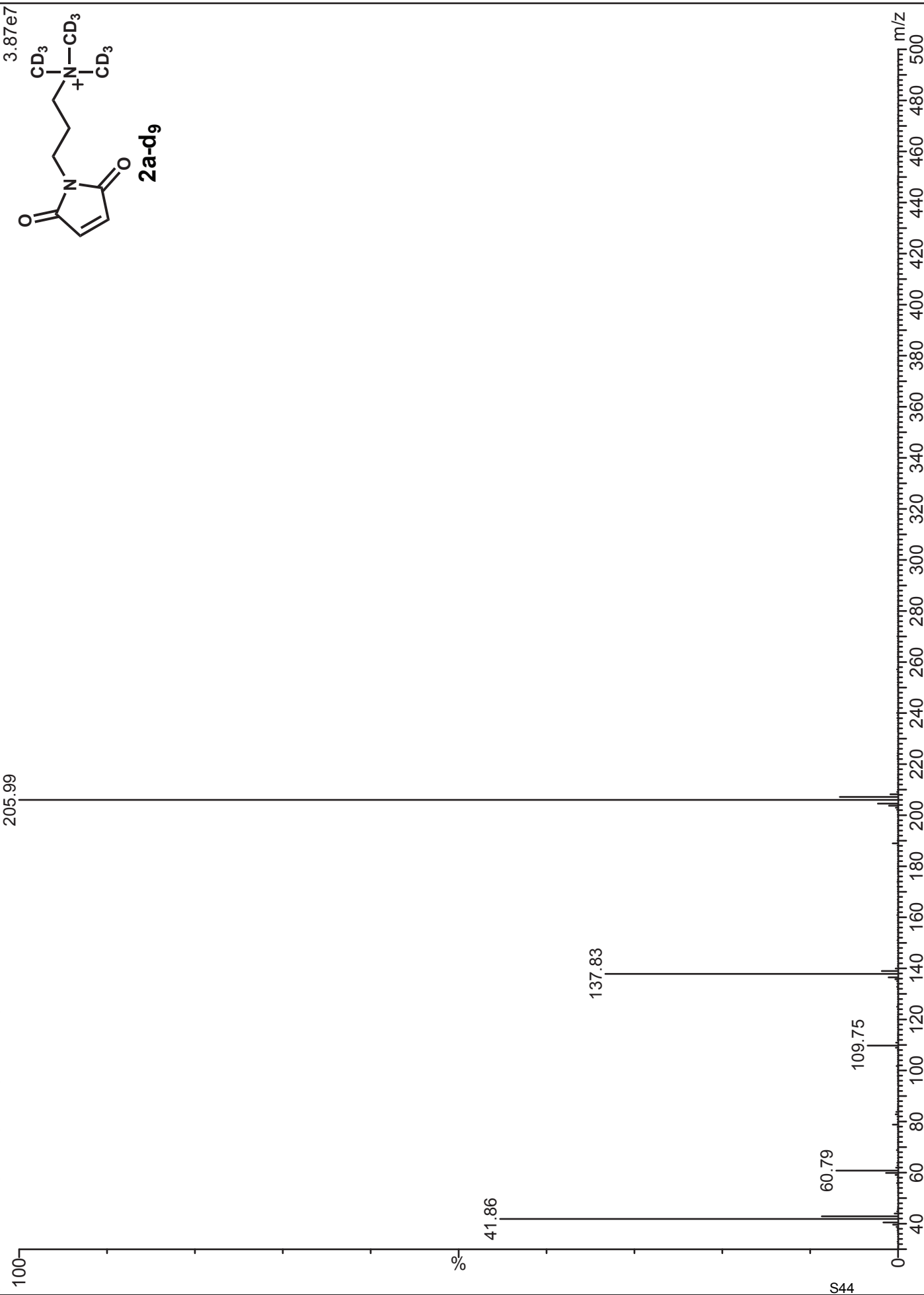
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17-Jan-2012

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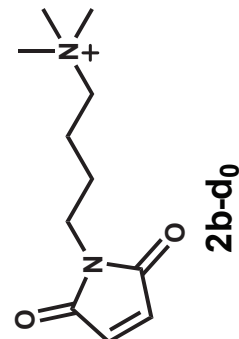
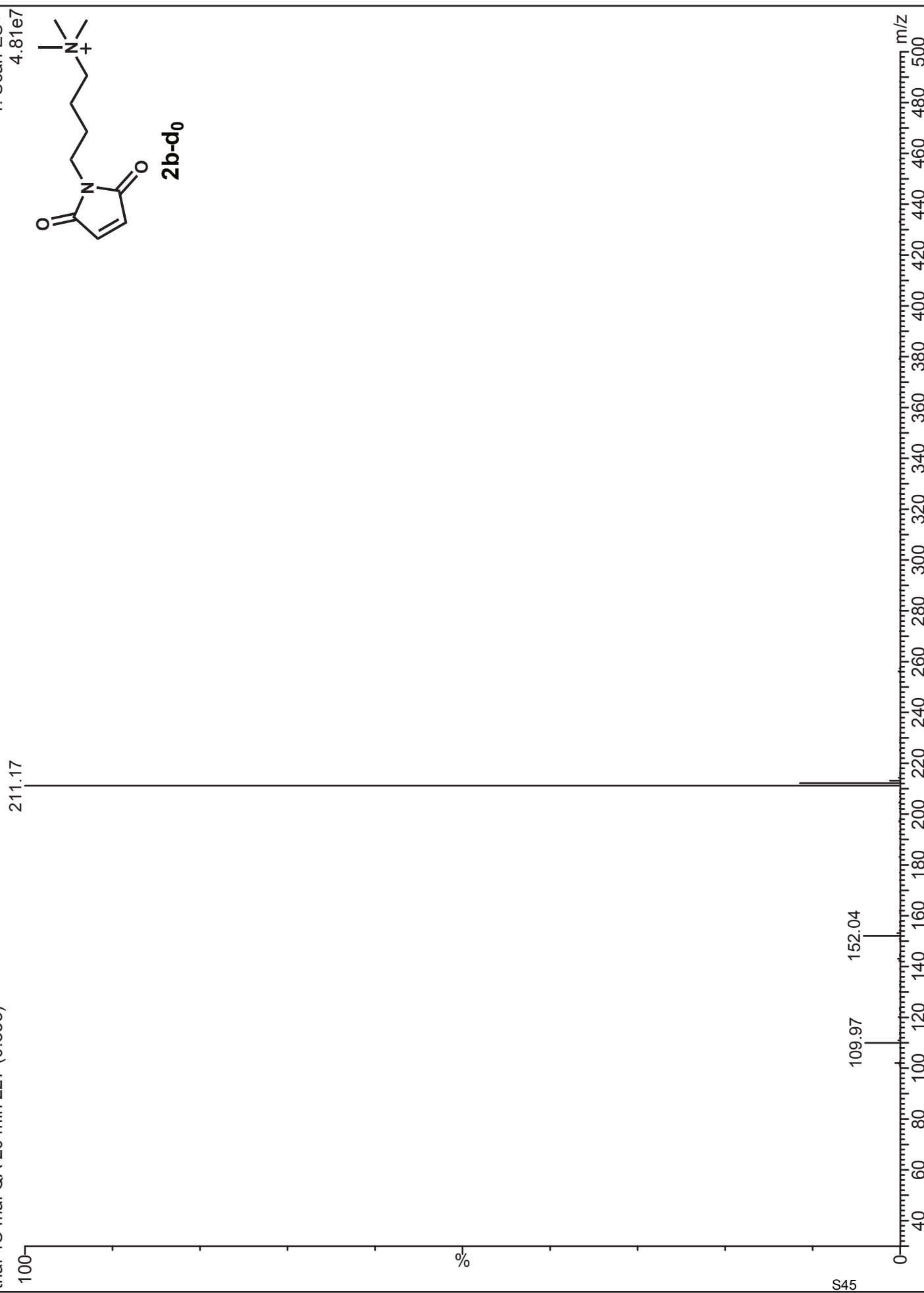
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S45



Benson

18-Jan-2012

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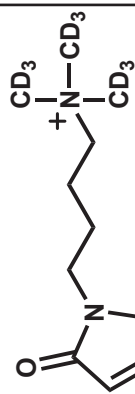
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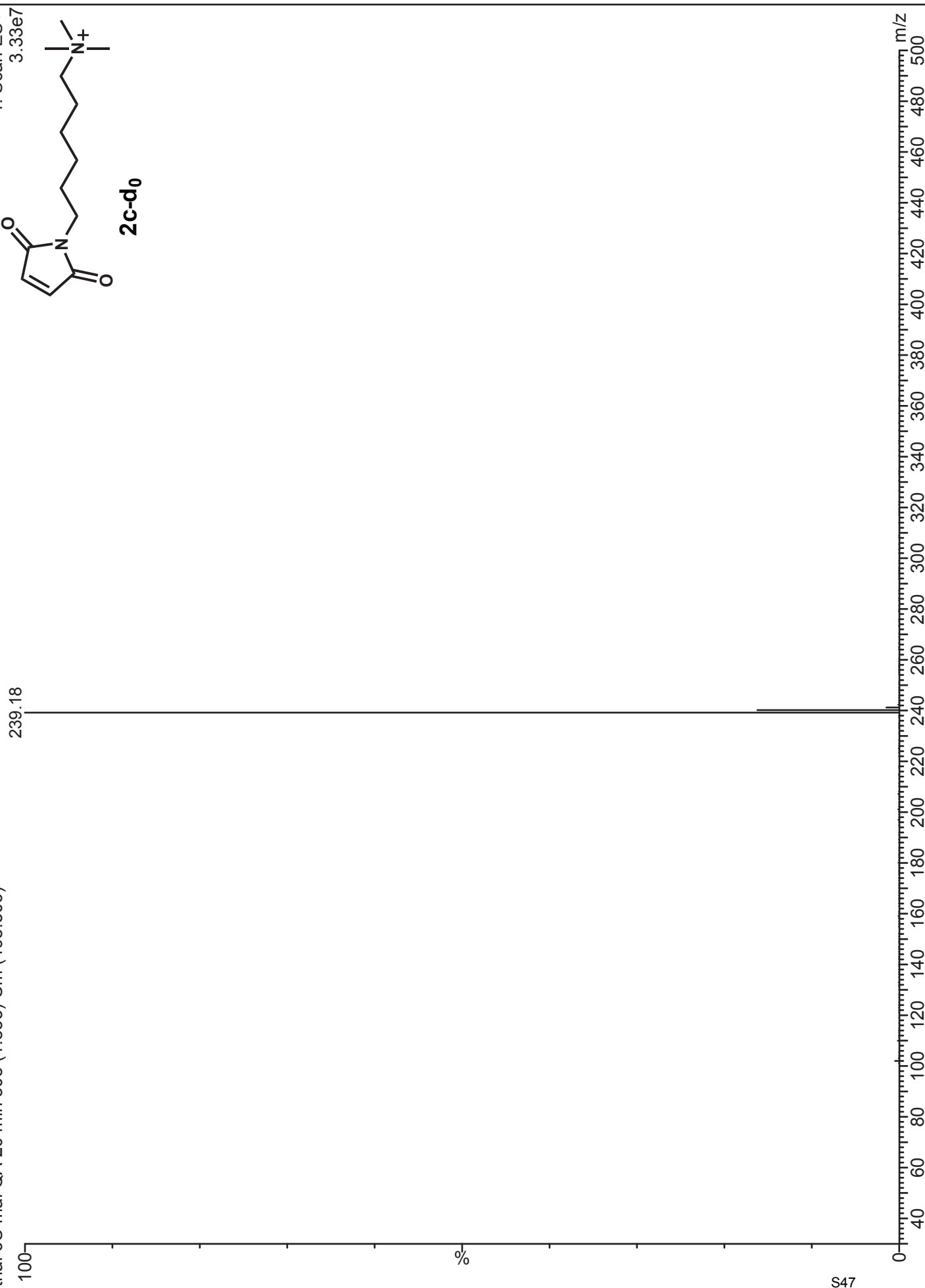
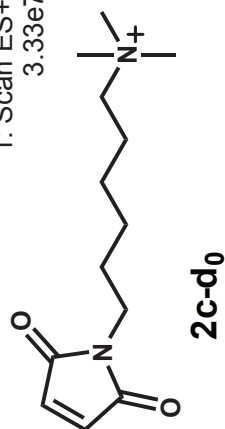


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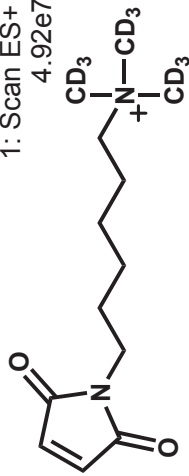
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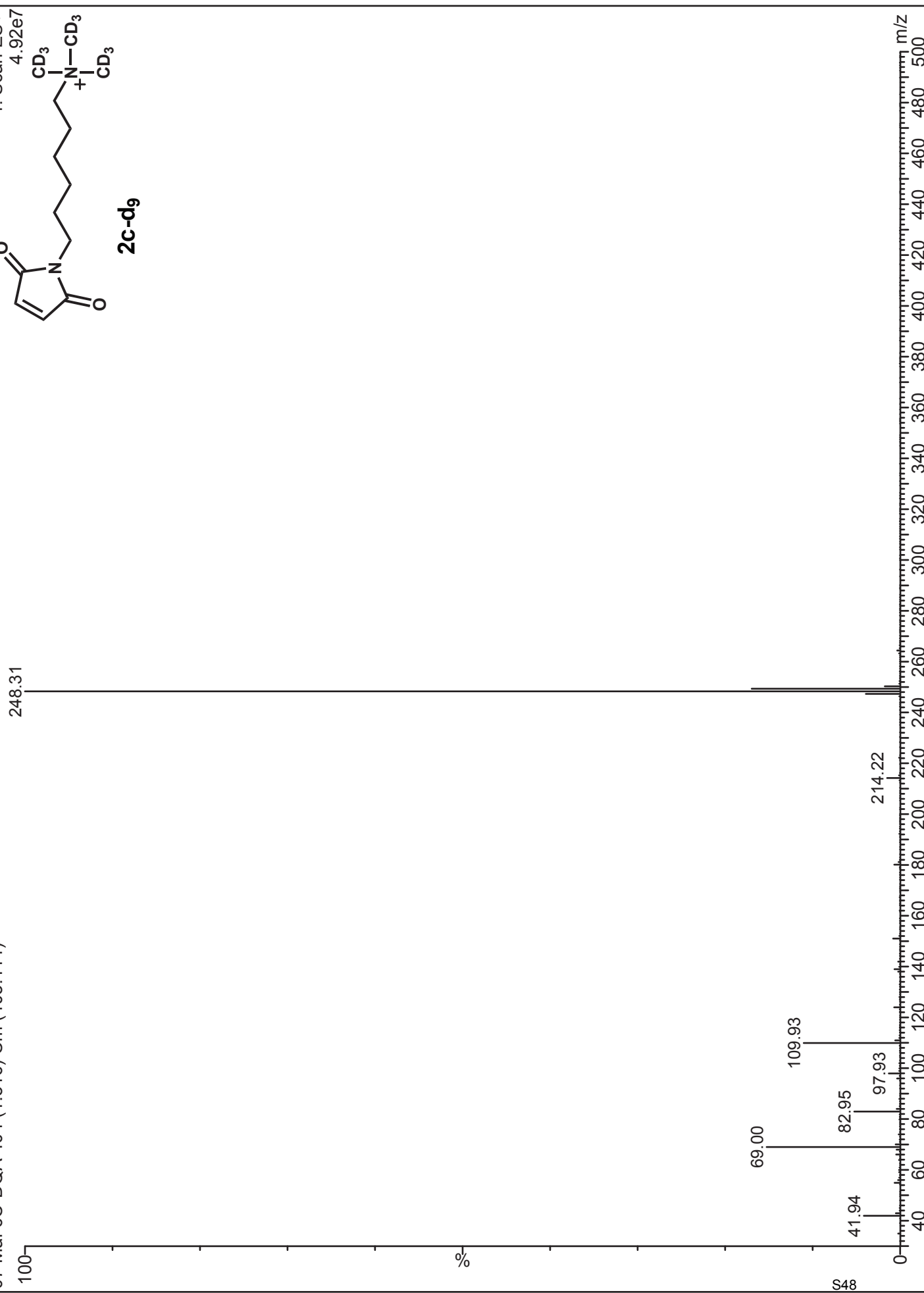
25-Aug-2012

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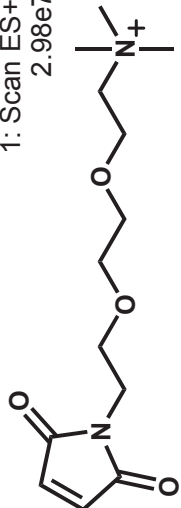
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m/z

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25-Aug-2012

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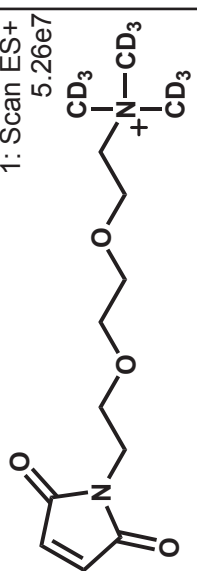
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S50

m/z