

## Supporting Information

### Selective Small Molecule Probes for the Hypoxia Inducible Factor (HIF) Prolyl Hydroxylases

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## Supplementary Schemes

Supplementary Scheme 1. Catalysis of histone demethylation by 2OG dependent histone demethylases (KDMs).

Supplementary Scheme 2. Syntheses of 4HQ derivatives.

## Spectroscopic data for Tested Compounds

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Supplementary Figure 1. Principles of the Amplified Luminescent Proximity Homogeneous Assay (AlphaScreen) for PHD2.

Supplementary Figure 2. Stereo-views from crystal structures of PHD2 in complex with **4** (A and B), **3** (C) and **15** (D). showing the *Fo-Fc* OMIT map (contoured to 3 $\sigma$ ) for the ligands.

Supplementary Figure 3. Comparison of binding modes for PHD inhibitors.

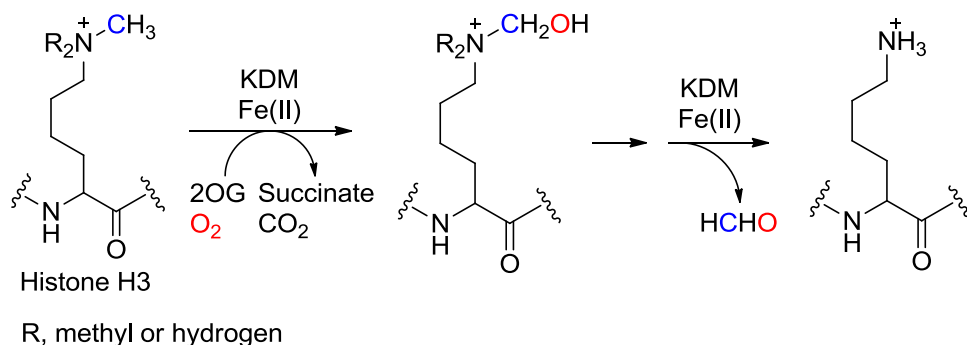
## Supplementary Tables

Supplementary Table 1. Crystallographic data processing, refinement statistics of tPHD2 complexes with **3**, **4** and **10**.

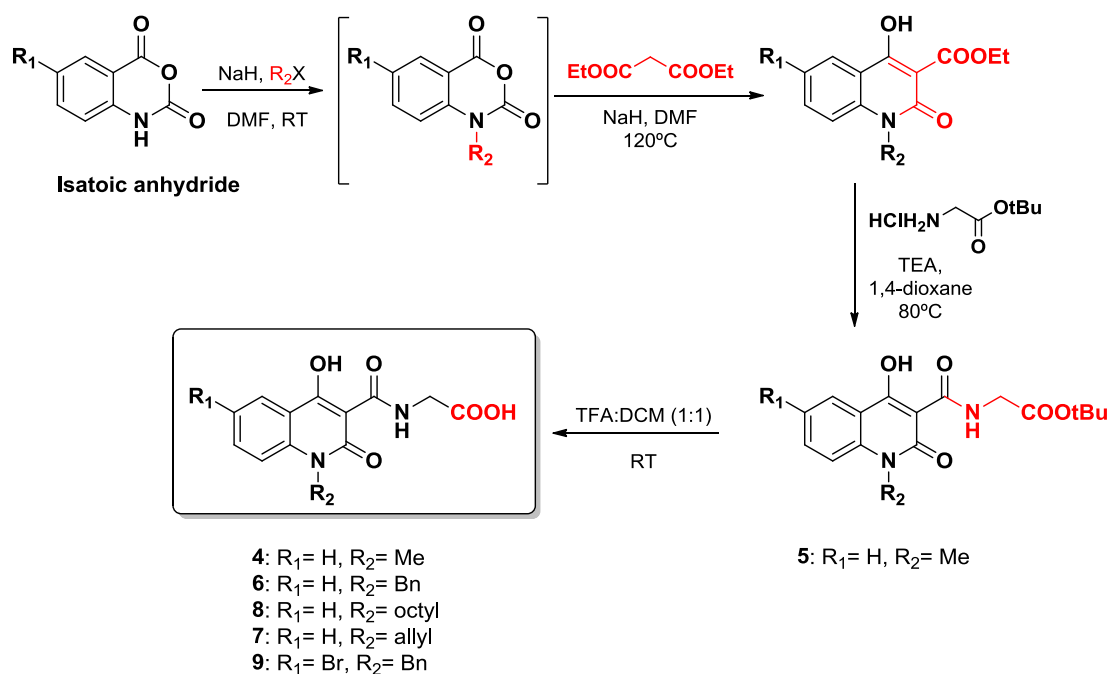
## Additional supplementary references

## Supplementary Schemes

**Supplementary Scheme 1.** Catalysis of histone demethylation by 2OG dependent histone demethylases (KDMs).



**Supplementary Scheme 2.** Syntheses of 4HQ derivatives. Compounds **4**, **6**, **8**, **7** and **9** were synthesized as described in three steps starting from the commercially available isatoic anhydride or its 5-bromo derivative.<sup>1</sup> One pot reaction with benzyl/alkyl bromides, followed by treatment with diethyl malonate, under basic conditions, led to the dihydroquinoline derivatives. Coupling with *tert*-butyl glycinate hydrochloride at reflux and subsequent *tert*-butyl ester deprotection with  $CF_3COOH$  gave the desired compounds.



## Spectroscopic Data for Tested Compounds

### *2-(4-Hydroxy-1-methyl-2-oxo-1,2-dihydroquinoline-3-carboxamido)acetic acid (4)*

<sup>1</sup>H NMR (500 MHz, DMSO-d<sub>6</sub>): δ = 16.98 (s, 1H, OH), 12.94 (bs, 1H, COOH), 10.56 (t, 1H, *J* = 5.5 Hz, NH), 8.08 (dd, 1H, *J* = 1.2, 8.0 Hz, ArCH), 7.81 (ddd, 1H, *J* = 1.4, 7.3, 8.6 Hz, ArCH), 7.62 (d, 1H, *J* = 8.5 Hz, ArCH), 7.38 (t, 1H, *J* = 7.4 Hz, ArCH), 4.14 (d, 2H, *J* = 5.6 Hz, CH<sub>2</sub>), 3.63 (s, 3H, CH<sub>3</sub>) ppm; <sup>13</sup>C NMR (125 MHz, DMSO-d<sub>6</sub>): δ = 171.0, 170.7, 170.4, 161.4, 139.8, 134.4, 124.5, 122.5, 115.4, 114.9, 96.0, 40.9, 29.0, ppm; HRMS (ESI<sup>-</sup>) calcd for C<sub>13</sub>H<sub>12</sub>N<sub>2</sub>NaO<sub>5</sub> (M-H)<sup>-</sup> 299.0644; found, 299.0638.

### *tert-Butyl 2-(4-hydroxy-1-methyl-2-oxo-1,2-dihydroquinoline-3-carboxamido)acetate (5)*

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ = 16.61 (s, 1H, OH), 10.77 (bs, 1H, NH), 8.21 (dd, 1H, *J* = 1.6, 8.0 Hz, ArCH), 7.69 (ddd, 1H, *J* = 1.6, 7.2, 8.6 Hz, ArCH), 7.36 (d, 1H, *J* = 8.6 Hz, ArCH), 7.30 (t, 1H, *J* = 7.6 Hz, ArCH), 4.14 (d, 2H, *J* = 5.4 Hz, CH<sub>2</sub>), 3.70 (s, 3H, CH<sub>3</sub>), 1.51 (s, 9H, C(CH<sub>3</sub>)<sub>3</sub>) ppm; <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>): δ = 171.6, 171.1, 168.3, 162.6, 140.0, 133.8, 125.5, 122.3, 116.0, 114.2, 96.9, 82.3, 41.8, 29.2, 28.0 (3C) ppm; HRMS (ESI<sup>-</sup>) calcd for C<sub>17</sub>H<sub>20</sub>N<sub>2</sub>O<sub>5</sub> (M-H)<sup>-</sup> 331.1294; found, 331.1299.

### *2-(4-Hydroxy-1-benzyl-2-oxo-1,2-dihydroquinoline-3-carboxamido)acetic acid (6)*

<sup>1</sup>H NMR (500 MHz, DMSO-d<sub>6</sub>): δ = 12.97 (bs, 1H, COOH), 10.54 (t, 1H, *J* = 5.3 Hz, NH), 8.13 (d, 1H, *J* = 8.1 Hz, ArCH), 7.71 (t, 1H, *J* = 8.0 Hz, ArCH), 7.48 (d, 1H, *J* = 8.7 Hz, ArCH), 7.39-7.20 (m, 6H, ArCH), 5.56 (s, 2H, PhCH<sub>2</sub>), 4.16 (d, 2H, *J* = 5.5 Hz, CH<sub>2</sub>) ppm; <sup>13</sup>C NMR (125 MHz, DMSO-d<sub>6</sub>): δ = 171.5, 170.7, 170.4, 161.7, 139.2, 136.6, 134.4, 128.7 (2C), 127.1, 126.4 (2C), 124.8, 122.7, 115.7, 115.4, 95.9, 44.6, 41.0 ppm; HRMS (ESI<sup>-</sup>) calcd for C<sub>19</sub>H<sub>16</sub>N<sub>2</sub>O<sub>5</sub> (M-H)<sup>-</sup> 351.0981; found, 351.0986.

### *2-(4-Hydroxy-1-allyl-2-oxo-1,2-dihydroquinoline-3-carboxamido)acetic acid (7)*

<sup>1</sup>H NMR (500 MHz, DMSO-d<sub>6</sub>): δ = 17.13 (bs, 1H, OH), 10.53 (s, 1H, NH), 8.12 (dd, 1H, *J* = 1.2, 8.2 Hz, ArCH), 7.79 (t, 1H, *J* = 7.8 Hz, ArCH), 7.54 (d, 1H, *J* = 8.5 Hz, ArCH), 7.38 (t, 1H, *J* = 7.4 Hz, ArCH), 5.97 (dddd, 1H, *J* = 4.5, 5.0, 10.5, 17.2 Hz, =CH), 5.16 (dd, 1H, *J* = 1.3, 10.5 Hz, =CH<sub>2</sub>), 5.01 (dd, 1H, *J* = 1.3, 17.2 Hz, =CH<sub>2</sub>), 4.94 (bs, 2H, CH<sub>2</sub>), 4.12 (d, 2H, *J* = 5.5 Hz, CH<sub>2</sub>) ppm; <sup>13</sup>C NMR (125 MHz, DMSO-d<sub>6</sub>): δ = 171.4, 170.6, 170.3, 161.2, 139.1, 134.3, 132.4, 124.6, 122.6, 116.4, 115.8, 115.2, 95.8, 43.6, 41.2 ppm; HRMS (ESI<sup>-</sup>) calcd for C<sub>15</sub>H<sub>14</sub>N<sub>2</sub>O<sub>5</sub> (M-H)<sup>-</sup> 301.0825; found, 301.0830.

### *2-(4-Hydroxy-1-octyl-2-oxo-1,2-dihydroquinoline-3-carboxamido)acetic acid (8)*

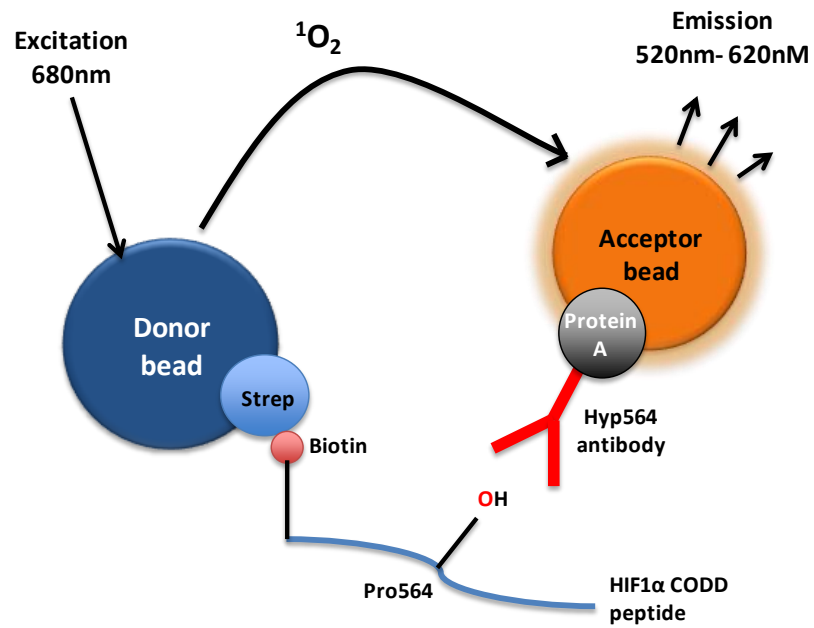
<sup>1</sup>H NMR (500 MHz, DMSO-d<sub>6</sub>): δ = 16.98 (s, 1H, OH), 12.94 (bs, 1H, COOH), 10.58 (t, 1H, *J* = 5.6 Hz, NH), 8.09 (dd, 1H, *J* = 1.4, 8.1 Hz, ArCH), 7.81 (ddd, 1H, *J* = 1.5, 7.5, 8.7 Hz, ArCH), 7.63 (d, 1H, *J* = 8.7 Hz, ArCH), 7.36 (t, 1H, *J* = 7.5 Hz, ArCH), 4.24 (t, 2H, *J* = 7.9 Hz, CH<sub>2</sub>), 4.13 (d, 2H, *J* = 5.6 Hz, CH<sub>2</sub>), 1.61 (quintet, 2H, *J* = 7.6 Hz, CH<sub>2</sub>), 1.45-1.20 (m, 10H, CH<sub>2</sub>), 0.85 (t, 3H, *J* = 7.1 Hz, CH<sub>3</sub>) ppm; <sup>13</sup>C NMR (125 MHz, DMSO-d<sub>6</sub>): δ = 171.0, 170.7, 170.4, 161.2, 139.0, 134.4, 124.7, 122.4, 115.2, 115.1, 95.9, 41.5, 40.9, 31.2, 28.7, 28.6, 27.2, 26.3, 22.0, 13.9 ppm; HRMS (ESI<sup>-</sup>) calcd for C<sub>20</sub>H<sub>26</sub>N<sub>2</sub>O<sub>5</sub> (M-H)<sup>-</sup> 397.1739; found, 397.1734.

### *2-(6-bromo-4-Hydroxy-1-benzyl-2-oxo-1,2-dihydroquinoline-3-carboxamido)acetic acid (9)*

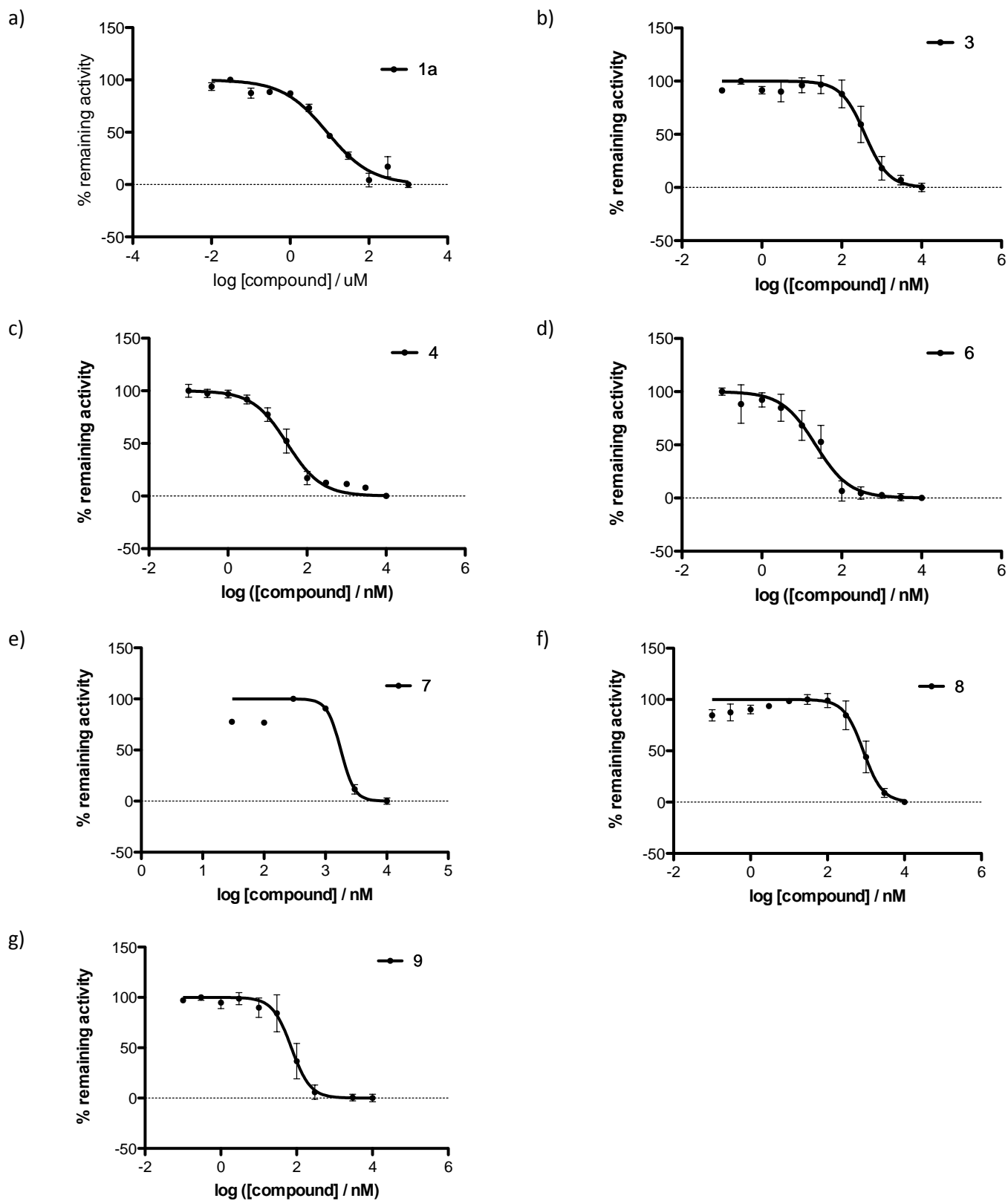
<sup>1</sup>H NMR (500 MHz, DMSO-d<sub>6</sub>): δ = 17.30 (bs, 1H, OH), 10.47 (t, 1H, *J* = 5.3 Hz, NH), 8.18 (d, 1H, *J* = 2.3 Hz, ArCH), 7.86 (dd, 1H, *J* = 2.3, 9.2 Hz, ArCH), 7.43 (d, 1H, *J* = 9.1 Hz, ArCH), 7.32 (t, 2H, *J* = 7.6 Hz, ArCH), 7.25 (t, 1H, *J* = 7.2 Hz, ArCH), 7.20 (d, 2H, *J* = 7.6 Hz, ArCH), 5.54 (bs, 2H, PhCH<sub>2</sub>), 4.15 (d, 2H, *J* = 5.5 Hz, CH<sub>2</sub>) ppm; <sup>13</sup>C NMR (125 MHz, DMSO-d<sub>6</sub>): δ = 170.5, 170.4, 170.2, 161.5, 138.3, 136.7, 136.2, 128.7 (2C), 127.2, 126.6, 126.3 (2C), 118.2, 117.1, 114.8, 96.5, 44.8, 41.1 ppm; HRMS (ESI<sup>-</sup>) calcd for C<sub>19</sub>H<sub>15</sub>BrN<sub>2</sub>O<sub>5</sub> (M-H)<sup>-</sup> 429.0086; found, 429.0092.

## Supplementary Figures

**Supplementary Figure 1.** Principles of the Amplified Luminescent Proximity Homogeneous Assay (AlphaScreen) for PHD2.

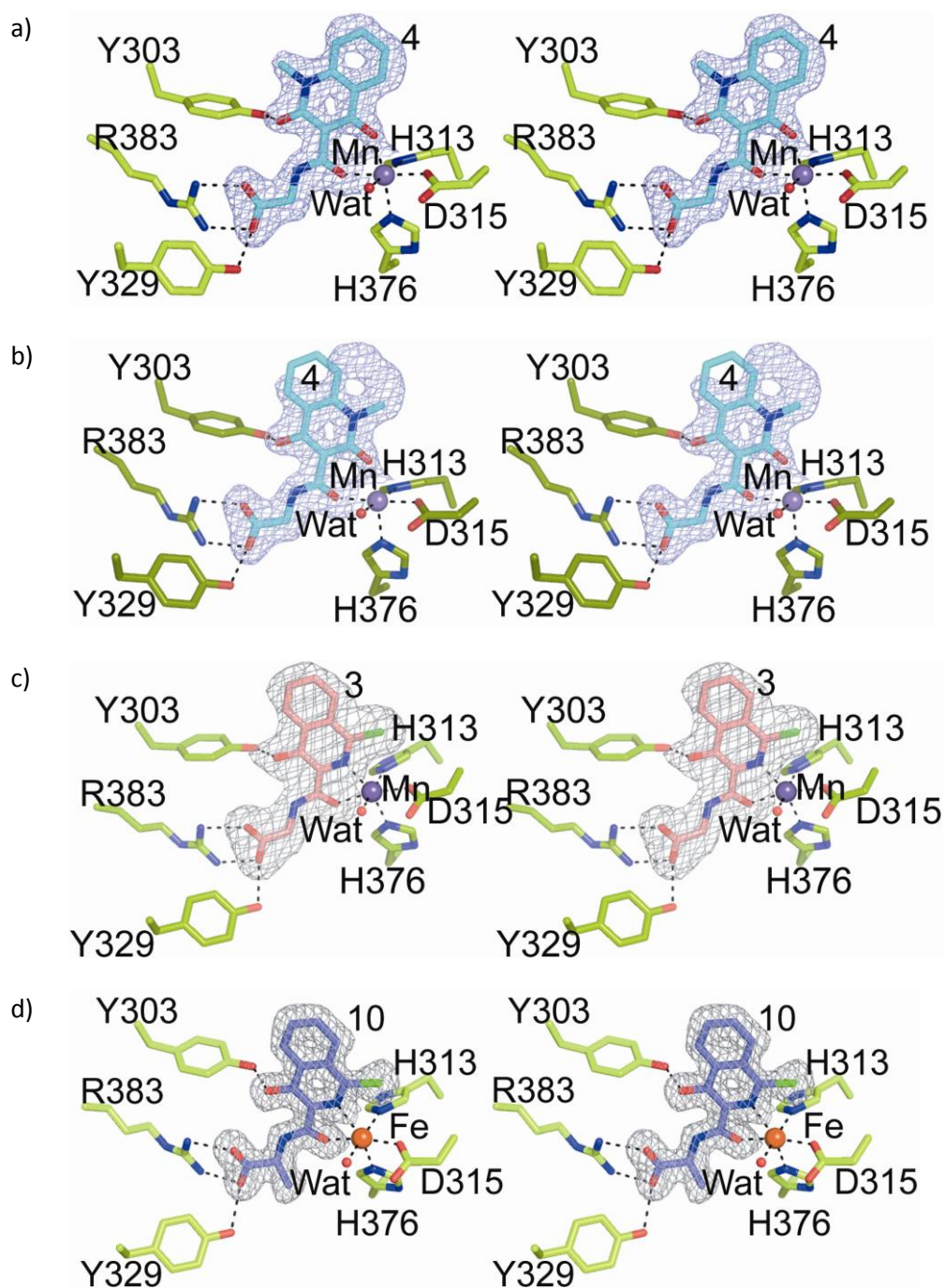


**Supplementary Figure 2.** Representative inhibition plots for compounds **1a**, **3**, **4**, **6**, **7**, **8** and **9** obtained using the PHD2 AlphaScreen assay. Values for each data point are averages  $\pm$  standard deviation ( $n \geq 3$ ).



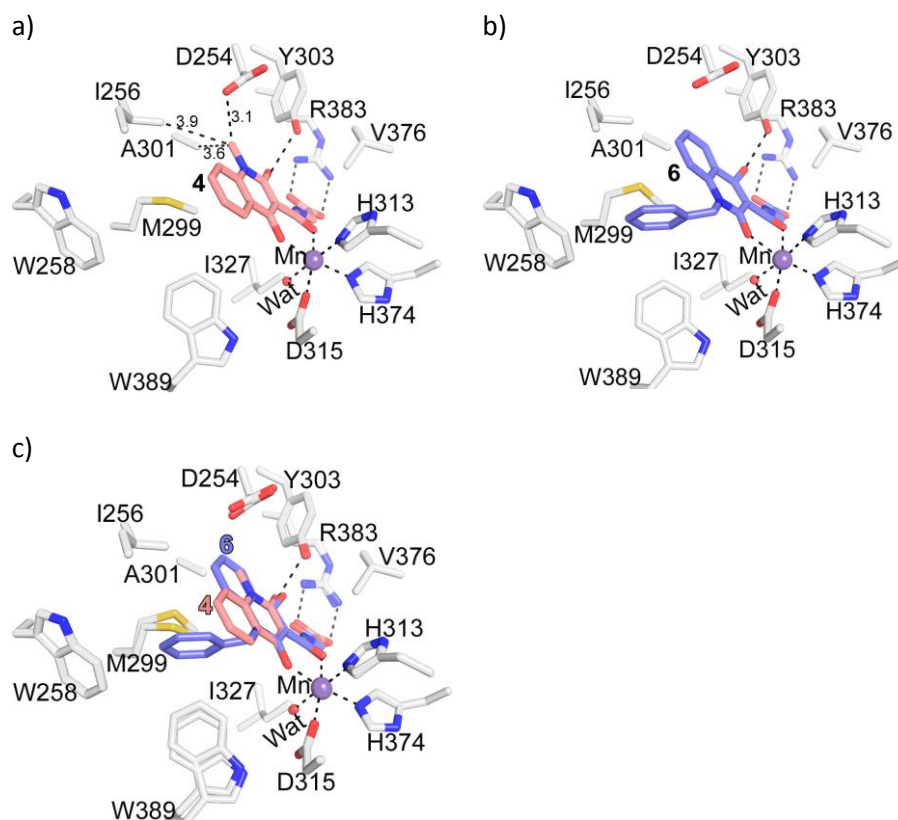
**Supplementary Figure 3.** Stereo-views from crystal structures of PHD2 in complex with **4** (a and b), **3** (c) and **15** (d)

showing the *Fo*-*Fc* OMIT map (contoured to  $3\sigma$ ) for the ligands. Because NMR evidence reveals that compound **4** can bind to the metal in the PHD2 active site in two modes (named 'X' and 'F' orientations) in solution,<sup>2</sup> attempts were made to model **4** in both these possible orientations ('X' as in S3A and 'F' as in S3B). Simulated annealed OMIT map analyses show that **4** only fits the map when in the 'X' orientation (as in S1a, map contoured to  $3\sigma$ ). Dotted lines indicate apparent hydrogen-bonds/ polar interactions.



**Supplementary Figure 4.** Comparison of binding modes for PHD inhibitors. View from a crystal structure of PHD2 in complex with **4** (a). A model predicting the binding mode for PHD2 in complex with Mn(II) and **6** (shown in b) was

generated using PHD2.Mn(II).**4** structure as the template. c) View from the superimposed PHD2.Mn(II).**4** crystal structure and a PHD2.Mn(II).**6** model. Parameter and topology files for **6** were generated using PRODRG.<sup>3</sup> The PHD2.Mn(II).**6** model was conjugate energy minimized using CNS (version 1.3)<sup>4</sup> without applying external energy terms. Note that the benzyl group of **6** is predicted to locate in a hydrophobic region.



**Supplementary Table 1. Crystallographic data processing, refinement statistics of tPHD2 complexes with 3, 4 and 10.**



Measurement	tPHD2.Mn(II).3	tPHD2.Mn(II).4	tPHD2.Fe(II).10
PDB acquisition codes	4BQX	4BQW	4BQY
<b>Data collection</b>			
Space Group	$P6_3$	$P6_3$	$P6_3$
Cell dimensions a,b,c (Å)	109.600	109.914	110.885
	109.600	109.914	110.885
	39.330	39.409	40.432
Resolution (Å)	54.8 – 1.79 (1.89 – 1.79)*	23.8 – 1.79 (1.85 – 1.79)*	36.3 – 1.55 (1.62 – 1.55)*
No. of unique reflections	25470 (3690)*	25705 (2568)*	43534 (4284)*
Completeness (%)	99.0 (99.9)*	98.9 (99.9)*	95.5 (94.7)*
Redundancy	5.9 (5.9)*	5.3 (3.5)*	2.5 (2.3)*
$R_{\text{sym}}$ **	0.067 (0.859)*	0.055 (0.681)*	0.051 (0.439)*
Mean $I/\sigma(I)$	11.3 (2.1)*	22.4 (2.1)*	16.1 (1.8)*
Wilson B value (Å <sup>2</sup> )	27.1	26.2	22.5
<b>Refinement</b>			
$R_{\text{factor}}$	0.211	0.195	0.212
$R_{\text{free}}$	0.229	0.201	0.227
R.m.s. deviation			
Bond length, Å	0.006	0.006	0.01
Bond angle, °	1.2	1.5	1.5

\*Highest resolution shell shown in parenthesis.

\*\* $R_{\text{sym}} = \sum |I - \langle I \rangle| / \sum I$ , where  $I$  is the intensity of an individual measurement and  $\langle I \rangle$  is the average intensity from multiple observations.

$R_{\text{factor}} = \sum_{\text{hkl}} |F_{\text{obs}}(\text{hkl})| - k |F_{\text{calc}}(\text{hkl})| / \sum_{\text{hkl}} |F_{\text{obs}}(\text{hkl})|$  for the working set of reflections;  $R_{\text{free}}$  is the  $R_{\text{factor}}$  for ~5% of the reflections excluded from refinement.

## Additional supplementary references

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- (3) Schuttelkopf, A. W., and van Aalten, D. M. (2004) PRODRG: a tool for high-throughput crystallography of protein-ligand complexes, *Acta Crystallogr. D Biol. Crystallogr.* **60**, 1355-1363.
- (4) Brunger, A. T., Adams, P. D., Clore, G. M., DeLano, W. L., Gros, P., Grosse-Kunstleve, R. W., Jiang, J. S., Kuszewski, J., Nilges, M., Pannu, N. S., Read, R. J., Rice, L. M., Simonson, T., and Warren, G. L. (1998) Crystallography & NMR system: A new software suite for macromolecular structure determination, *Acta Crystallogr. D Biol. Crystallogr.* **54**, 905-921.