Highly Enantioselective Ruthenium/PNNP-Catalyzed Imine Aziridination: Evidence of Carbene Transfer from a Diazoester Complex

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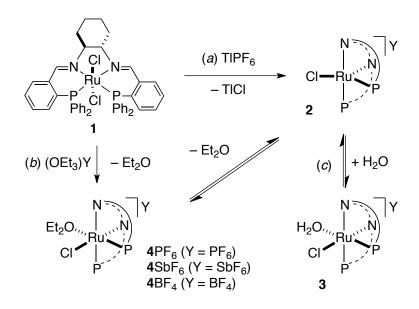
SUPPORTING INFORMATION

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Low-Temperature NMR Spectra of $[RuCl(OEt_2)(PNNP)]Y$ (Y = PF₆, BF₄, or SbF₆).

As previously reported, [RuCl(OEt₂)(PNNP)]PF₆ (**4**PF₆) gives a broad ³¹P NMR signal at δ 41 at room temperature, which decoalesces upon cooling and eventually gives a sharp AX system (δ 55.5 and 36.9) at low temperature.¹ This behavior has been explained with the dissociation of [RuCl(OEt₂)(PNNP)]⁺ to give OEt₂ and the 16-electron species [RuCl(PNNP)]⁺ (**2**), which is a fast equilibrium on the NMR time scale at room temperature and is frozen out at -40 °C (Scheme 3 of main paper):



The low-temperature ³¹P NMR spectra of the BF_4^- and SbF_6^- salts of 4 contain an additional, broad signal at ca. δ 49, which is nearly indistinguishable in the spectrum of $4PF_6$. Interestingly, the nature of the counterion affects the temperature at which this signal appears (-80, -60, and -20 °C for $4PF_6$, $4BF_4$, and $4SbF_6$, respectively), as well as its intensity at -80 °C (5, 26, and 45% of the total integrated intensity, see Figures S1-S4). The chemical shift of this signal (δ 49) is indicative of an octahedral complex of the type *trans*-[RuCl(Y)(PNNP)],¹ in which the counterion Y (Y = BF_4^- or SbF_6^-) is associated with the 16-electron complex $[RuCl(PNNP)]^+$ in the low-polar CD₂Cl₂ solvent. We have previously suggested¹ that the inequivalent phosphines of complexes of the type *trans*-[RuCl(Y)(PNNP)] resonate in a δ region centered at about δ 48, which is the chemical shift of *trans*-[RuCl₂(PNNP)],² because they are *trans* to the same donor type, that is, imine. Therefore, these complexes typically give relatively tight AB spin systems. An example thereof is the alkyl complex trans-[RuCl(CH₂COOEt)(PNNP)] (11) (see Figure S6 below). In contrast, in *cis*- β complexes such as $[RuCl(OEt_2)(PNNP)]^+$ (4), one of the phosphines resonates at a much higher frequency (*ca*. δ 63) than the other one (ca. δ 45). This indicates that these P donors are *trans* to ligands with a largely different *trans* influence, such as agua and imine.¹ The same pattern has been observed for the ³¹P NMR chemical shifts in $[Ru(OH_2)_2(PNNP)]^{2+.3}$.

As the signal at δ 49 observed in the low-temperature spectra of [RuCl(OEt₂)(PNNP)]Y is broad even at -80 °C, which prevents further studies by 2D NMR spectroscopy, the formulation of the corresponding species as *trans*-[RuCl(Y)(PNNP)] remains tentative. Furthermore, the lowtemperature ¹⁹F NMR spectra show the essentially unperturbed signals of the free anions at δ -73 (PF₆⁻) and -152 (BF₄⁻) (unsurprisingly, no signal was observed for SbF₆⁻).⁴ Overall, the lowtemperature NMR spectra suggest that the dissociation of the Et₂O adduct **4** into five-coordinate **2** and Et₂O is the main dynamic process in solution and fail to give conclusive evidence of the cation/anion interactions evoked by the counterion effect observed in catalysis. **Figure S1.** Effect of the counterion Y on the NMR spectra of $[RuCl(OEt_2)(PNNP)]Y$ (Y = PF₆, 4PF₆; BF₄, 4BF₄; or SbF₆, 4SbF₆) at -80 °C (202 MHz, CD₂Cl₂). In the spectra of 4BF₄ and 4SbF₆, "A" denotes the unknown signal at δ 49. The signals of the aqua complex *trans*-[RuCl(OH₂)(PNNP)]⁺ (present in traces in 4PF₆ and 4SbF₆) are marked "*". The other signals belong to unknown impurities.

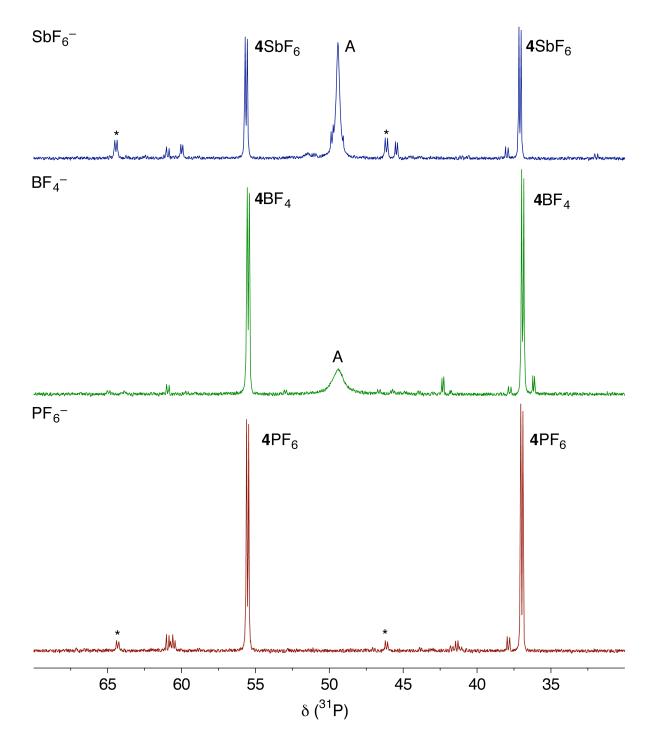
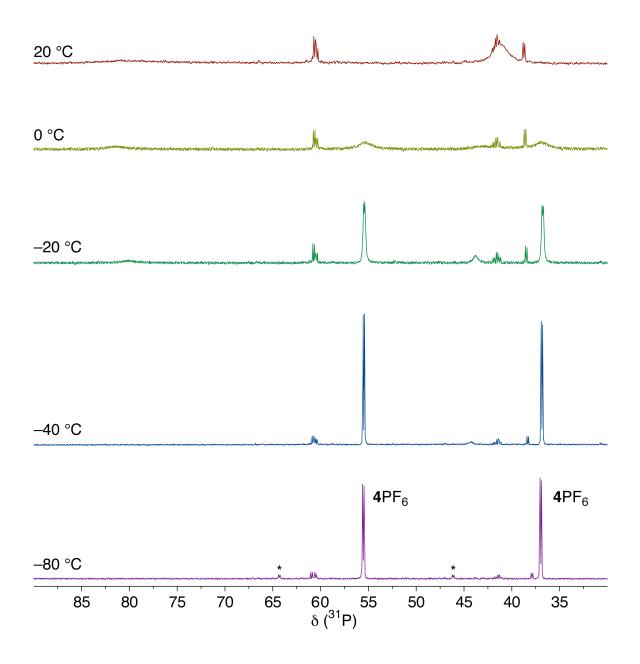


Figure S2. ³¹P NMR spectra of [RuCl(OEt₂)(PNNP)]PF₆ (**4**PF₆) at different temperatures (202 MHz, CD₂Cl₂). The signals of the aqua complex *trans*-[RuCl(OH₂)(PNNP)]⁺ (traces) are marked "*". The other signals belong to unknown impurities.



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Figure S3. ³¹P NMR spectra of [RuCl(OEt₂)(PNNP)]BF₄ (4BF₄) at different temperatures (202 MHz, CD₂Cl₂). "A" denotes the unknown signal at δ 49. The other signals belong to unknown impurities.

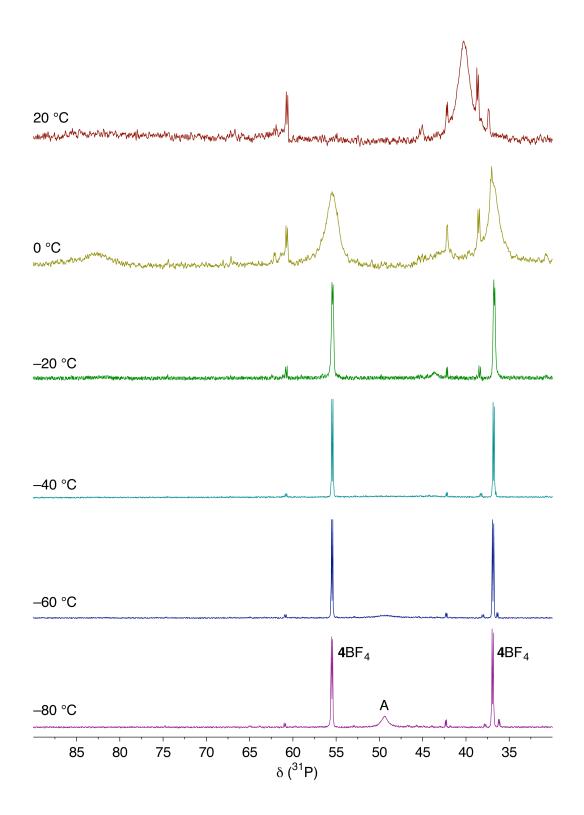
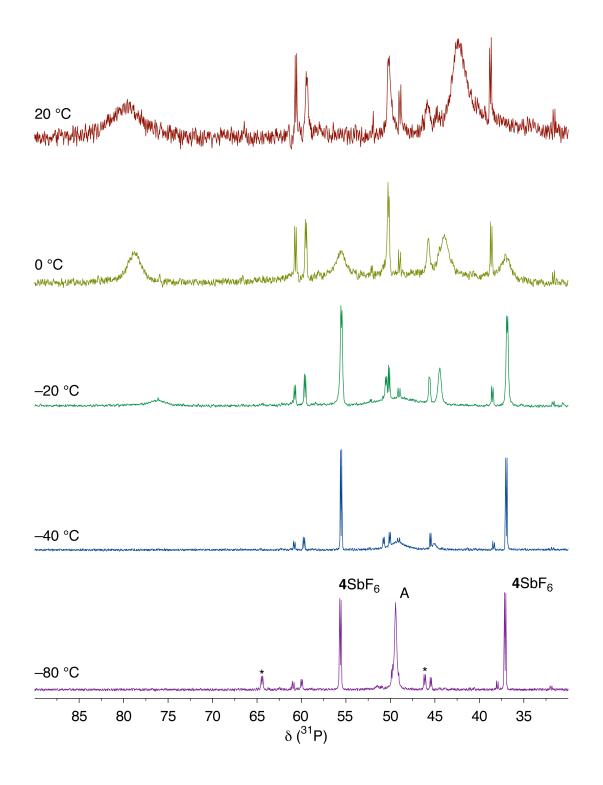


Figure S4. ³¹P NMR spectra of [RuCl(OEt₂)(PNNP)]SbF₆ (4SbF₆) at different temperatures (202 MHz, CD₂Cl₂). "A" denotes the unknown signal at δ 49. The signals of the aqua complex *trans*-[RuCl(OH₂)(PNNP)]⁺ (traces) are marked "*". The other signals belong to unknown impurities.



Synthesis of ¹³C- and ¹⁵N-Labeled EDA

Ethyl 2-¹³C-Glycine Hydrochloride.⁵ 2-¹³C-glycine (98% 2-¹³C, 0.50 g, 4.85 mmol) was suspended in ethanol, and the mixture cooled down to -20 °C (ice-salt bath). SOCl₂ (0.58 mL, 8.00 mmol) was added, the temperature raised to room temperature, and another equivalent of solid 2-¹³C-glycine (0.50 g, 4.85 mmol) was slowly added. The mixture was refluxed for 2 h. After cooling the colorless solution to room temperature, the solvent was evaporated under reduced pressure. The resulting white solid was dried in high vacuum for 2 h and recrystallized from ethanol. Yield: 1.10 g, 95 %. m.p. = 145–147 °C.

Synthesis of N₂¹³CHCO₂Et (¹³C-EDA).⁶ Ethyl 2-¹³C-glycine hydrochloride (1.00 g, 7.1 mmol) was mixed with H₂O (2 mL) and CH₂Cl₂ (4 mL) in a two-necked flask equipped with septum, argon inlet, and internal thermometer. The colorless mixture was cooled down to -5 °C, and an ice-cold solution of NaNO₂ (0.59 g, 8.5 mmol) in H₂O (2 mL) was added. The resulting mixture was cooled to -9 °C, and a 5% (w/w) H₂SO₄ solution (0.679 g) was slowly added. As higher temperature might decrease the yield, the temperature was never let to above +1 °C during the addition. Thereafter, the mixture was stirred for 20 min between -9 °C and +1 °C, and then poured into an ice-cold separating funnel. The yellow organic layer was recovered, and the water phase was extracted with CH₂Cl₂ (2 × 3 mL). The combined organic phase was washed with a 5% ice-cold NaHCO₃ solution (6 mL), the organic phase was separated, and the water phase was removed under reduced pressure. The resulting yellow oil was dried in vacuum for 15 min, and the product distilled with cold distillation under high vacuum. Yield: g (0.74 g, 81%). ¹H NMR (500 MHz, CD₂Cl₂, 25 °C): δ 4.80 (*d*, 1H, ¹J_{CH} = 205 Hz, N₂¹³CH), 4.23

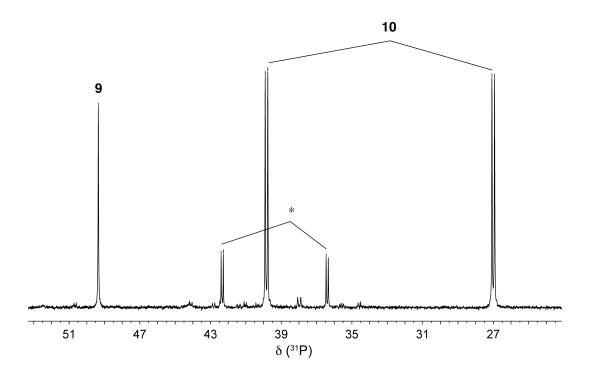
 $(q, {}^{2}J_{H,H'} = 7.1, 2H, OCH_{2}CH_{3}), 1.30 (t, {}^{2}J_{H,H'} = 7.1, 3H, OCH_{2}CH_{3}). C NMR (126 MHz, CD_{2}Cl_{2}, 25 °C): \delta 46.3 (s, N_{2}CH). Low-temperature data: {}^{1}H NMR (500 MHz, CD_{2}Cl_{2}, -78 °C): \delta 5.01 (d, 1H, {}^{1}J_{C,H} = 205 Hz, N_{2}{}^{13}CH), 4.20 (q, {}^{2}J_{H,H'} = 7.1, 2H, OCH_{2}CH_{3}), 4.17 (q, {}^{2}J_{H,H'} = 7.1, 2H, OCH_{2}CH_{3}), 1.29 (t, {}^{2}J_{H,H'} = 7.1, 3H, OCH_{2}CH_{3}), 1.25 (t, {}^{2}J_{H,H'} = 7.1, 3H, OCH_{2}CH_{3}), 1.25 (t, {}^{2}J_{H,H'} = 7.1, 3H, OCH_{2}CH_{3}), 1.30 (t, {}^{2}CH_{2}, -78 °C): \delta 47.5 (s, N_{2}CH), 46.5 (s, N_{2}CH).$

Synthesis of ¹⁵*N*NCHCO₂Et. (¹⁵N-EDA).⁶ Terminally ¹⁵N-labeled ¹⁵N-EDA was prepared analogously to ¹³C-EDA from ethyl glycine hydrochloride (1.00 g, 7.1 mmol) and Na¹⁵NO₂ (98% ¹⁵N, 0.60 g, 8.5 mmol). ¹H NMR (500 MHz, CD₂Cl₂, 25 °C): δ 4.80 (*br s*, 1H, N₂C*H*), 4.23 (*q*, ²*J*_{H,H'} = 7.1, 2H, OC*H*₂CH₃), 1.30 (*t*, ²*J*_{H,H'} = 7.1, 3H, OCH₂C*H*₃). ¹⁵N NMR (50.7 MHz, CD₂Cl₂, 25 °C): δ 4.05 (*br s*, 1N, ¹⁵*N*NC). Low-temperature data: ¹H NMR (500 MHz, CD₂Cl₂, -78 °C): δ 5.04 (*s*, 1H, N₂C*H*), 4.74 (*s*, 1H, N₂C*H*), 4.20 (*q*, ²*J*_{H,H'} = 7.1, 2H, OC*H*₂CH₃), 4.17 (*q*, ²*J*_{H,H'} = 7.1, 2H, OC*H*₂CH₃), 1.29 (*t*, ²*J*_{H,H'} = 7.1, 3H, OCH₂C*H*₃), 1.25 (*t*, ²*J*_{H,H'} = 7.1, 3H, OCH₂C*H*₃). ¹⁵N NMR (50.7 MHz, CD₂Cl₂, -78 °C): δ 7.6 (*s*, 1N, ¹⁵*N*NC), -1.29 (*s*, 1N, ¹⁵*N*NC).

NMR Spectroscopic Studies: General. The reactions described below were run under argon in NMR tubes fitted with serum septa and were monitored by NMR spectroscopy as detailed below. Additions of reagents were performed by microsyringe. A 2-PrOH bath at the appropriate temperature was used to keep the sample temperature at the values indicated below during all manipulations and transfers from and to the spectrometer.

Experiment 1: [RuCl(OEt₂)(PNNP)]PF₆ (4PF₆) + EDA, then imine 5a (1:1:1). Complex 4PF₆ was prepared by treating [RuCl₂(PNNP)] (1) (28.0 mg, 0.034 mmol) with (Et₃O)PF₆ (8.4 mg, 0.034 mmol) in CD₂Cl₂ (0.5 mL). After stirring the solution at room temperature overnight, the formation of 4PF₆ was confirmed by the ³¹P and ¹H NMR spectra at 298 and -78 °C. Then, EDA (3.7 µL, 0.034 mmol) was added at -78 °C, and quantitative conversion of $4PF_6$ to give the carbene complex *trans*-[RuCl(CHCOOEt)(PNNP)]⁺ (10) (71%, δ : 39.8 and 27.0, ${}^2J_{P,P'} = 29.8$ Hz)),⁷ dinitrogen complex 9 (15%, δ 49.2, see below), and the AB pattern (marked "*") of an unkown impurity (14%, δ 42.3 and 36.3 (d, ${}^2J_{P,P'} = 24.8$ Hz)) was observed (Figure S5).

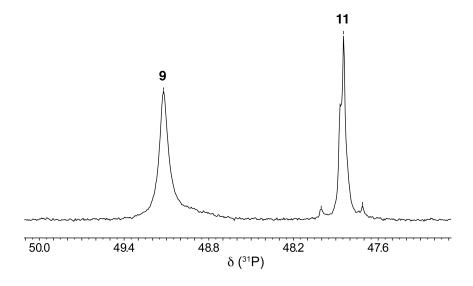
Figure S5. ³¹P NMR spectrum (202 MHz, CD_2Cl_2 , -78 °C) immediately after the addition of EDA (1 equiv) to $4PF_6$ at -78 °C, showing carbene complex **10** (71%) along with dinitrogen complex **9** (15%) and an unknown species (14%) whose signals are marked "*".



After adding imine **5a** (0.0091 g, 0.034 mmol) to this solution at -78 °C, the ³¹P NMR spectrum remained unchanged in the temperature range between -78 °C and room temperature. After 4 h at room temperature, a (¹³C,¹H)-HMQC experiment showed that no aziridine had formed. The ³¹P NMR spectrum of the reaction solution shows no signals attributable to the

carbene complex **10** (Figure S6). The newly appeared tight AB pattern at δ 47.9 (*d*, *J* = 28.2 Hz) and 47.8 (*d*, *J* = 28.2 Hz) (202 MHz, CD₂Cl₂, 25 °C) was assigned to the alkyl complex *trans*-[RuCl(CH₂COOEt)(PNNP)] (**11**) on the basis of the ¹H NMR signals of the RuCH₂COOEt moiety, which were identified by means of (³¹P,¹H)-HMQC and (¹³C,¹H)-HMQC experiments: ¹H NMR (500 MHz, CD₂Cl₂, 25 °C): δ 3.79 (*d*, 1H, *J* = 11.2 Hz, RuCHH'COOEt), 3.36 (*d*, 1H, *J* = 11.2 Hz, RuCHH'COOEt.

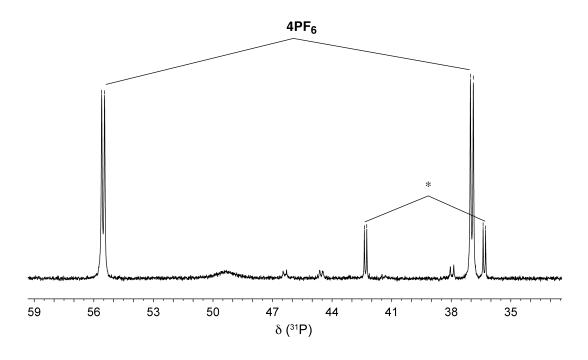
Figure S6. ³¹P NMR spectrum (202 MHz, CD_2Cl_2 , 25 °C) of the same reaction solution after 4 h at 25 °C showing the signals of *trans*-[RuCl(CH₂COOEt)(PNNP)] (**11**), the decay product of 10, and of the N₂ complex **9**.



Experiment 2: [RuCl(OEt₂)(PNNP)]PF₆ (4PF₆) + imine 5a, then EDA (1:1:1). Complex 4PF₆ was prepared by treating [RuCl₂(PNNP)] (1) (30.0 mg, 0.036 mmol) with (Et₃O)PF₆ (9.0 mg, 0.036 mmol) in CD₂Cl₂ (0.5 mL). After stirring the solution at room temperature overnight, the formation of $4PF_6$ was confirmed by the ³¹P and ¹H NMR spectra at 298 and -78 °C. Imine 5a (9.8 mg, 0.036 mmol) was added to the mixture at room temperature,

and the ³¹P and ¹H NMR spectra were recorded at 25 °C and at -78 °C. Along with unreacted **4PF**₆, the signals (marked "*") of the unknown product described above were observed (Figure S7). This species is not an imine complex, as confirmed by (¹H,¹H)-NOESY analysis and by the observation that it is formed in small amounts also in the reaction of **4PF**₆ with EDA, that is, in the absence of imine (see Experiment 1, Figure S5). After extracting the sample from the NMR spectrometer, EDA (9.0 mL, 0.036 mmol) was added by microsyringe to the solution at -78 °C. The sample was transferred immediately to the precooled NMR spectrometer (-78 °C) and the ³¹P and ¹H NMR spectra were recorded.

Figure S7. ³¹P NMR spectrum (202 MHz, CD_2Cl_2 , -78 °C) of the reaction solution of $4PF_6$ with imine 5a (1 equiv). Unreacted $4PF_6$ is the main species in solution, along with the unknown species marked "*".

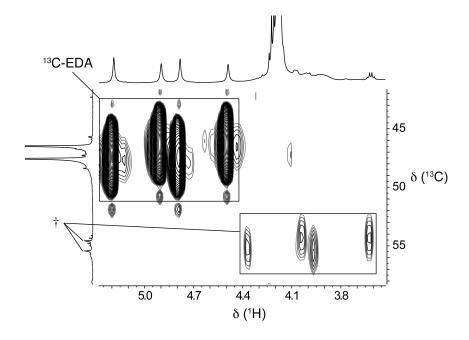


The ³¹P NMR spectrum of the reaction solution (not shown) indicates that $4PF_6$ was quantitatively converted to the carbene complex *trans*-[RuCl(CHCOOEt)(PNNP)]⁺ (10) (74%)⁷ and to the dinitrogen complex 9 (15%). Upon warming to room temperature in 20 °C steps, the composition of the solution did not change, and no aziridine was formed, as indicated by (¹³C,¹H)-HMQC experiments. The signals of impurity (signal "*", 11%) remained unchanged up to room temperature and disappeared within 4 h time. After 4 h at room temperature, all the species had converted to dinitrogen complex 9 (60%) and to the alkyl complex [RuCl(CH₂COOEt)(PNNP)] (11, 40%, δ 47.8, AB system) already observed in Experiment 1.

Experiment 3: [RuCl(OEt₂)(PNNP)]PF₆ (4PF₆) + imine 5a, then ¹³C-EDA (1:1:10). Complex 4PF₆ was prepared by treating [RuCl₂(PNNP)] (1) (24.3 mg, 0.029 mmol) with (Et₃O)PF₆ (7.3 mg, 0.029 mmol) in CD₂Cl₂ (0.5 mL). After stirring the solution at room temperature overnight, the formation of 4PF₆ was confirmed by the ³¹P and ¹H NMR spectra at 25 °C and -78 °C. Then, imine 5a (7.9 mg, 0.029 mmol) was added to the solution, which was cooled again. EDA (32.2 µL, 0.293 mmol) was added at -78 °C, and the ¹³C, ¹H, and ³¹P NMR spectra were run at the same temperature, as well as a (¹³C,¹H)-HMQC experiment. The (¹³C,¹H)-HMQC correlation showed the signals of unreacted ¹³C-EDA as major product (Figure S8).

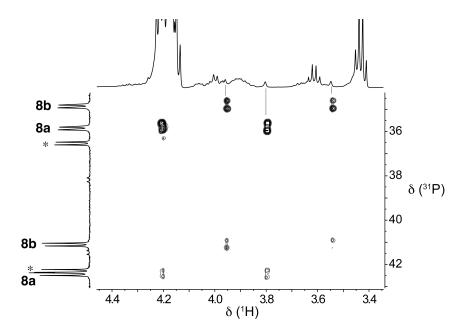
The signal (marked "†") of an additional ¹³C-containing species with a $J_{C,H}$ comparable to that of ¹³C-EDA was present, but disappeared after heating to -20 °C. As this signal has never been observed at temperatures higher than -20 °C, we deem it immaterial for the further discussion. The ³¹P NMR spectrum showed the quantitative conversion of the Et₂O adduct **4PF**₆ to several unknown species. Outside the spectral range shown in Figure S8, traces of *trans*-[RuCl(¹³CHCOOEt)(PNNP)]⁺ (**10**) and of ¹³C-labeled dietyl maleate (**7**) were detected.

Figure S8. Section of a (${}^{13}C, {}^{1}H$)-HMQC experiment after EDA addition (10 equiv) to a solution containing complex **4PF**₆ and imine **5a** (1 equiv) at -78 °C (500 MHz (${}^{1}H$), CD₂Cl₂). The signal marked "†" belongs to an unknown species (see footnote 25 of paper).



As no aziridine **6a** was observed at -80 °C, the sample was carefully warmed up to -20 °C. At this temperature, a (¹³C,¹H)-HMQC correlation experiment indicated that a small amount of ¹³C-aziridine had formed. To slow down the reaction, the sample was cooled to -60 °C, at which temperature a (¹³C,¹H)-HMQC experiment revealed new signals that we assign to coordinated ¹³C-EDA in [RuCl(¹³C-EDA)(PNNP)]PF₆ (**8**) (see Figure 2 of main paper). At the same temperature, the ³¹P NMR spectrum shows the signals of the dinitrogen complex **9** (31%) and the same AB system observed in Experiment 2 upon addition of imine to the Et₂O adduct **4PF**₆ (signal marked "*", 9%) (see Figure 3 of main paper). The main feature of the spectrum consists of two AB patterns in equal ratio, **8a** (31%) and **8b** (31%) (δ (**8a**) 42.4 (*d*, ²*J*_{P,P'} = 25.3 Hz)) and 35.8 (*d*, ²*J*_{P,P'} = 25.2 Hz); δ (**8b**) 41.1 (*d*, ²*J*_{P,P'} = 25.3 Hz), 34.8 (*d*, ²*J*_{P,P'} = 25.4 Hz)), which we assign to the diazoester complex *trans*-[RuCl(N₂¹³CHCOOEt)(**1a**)]⁺ (¹³C-**8**).

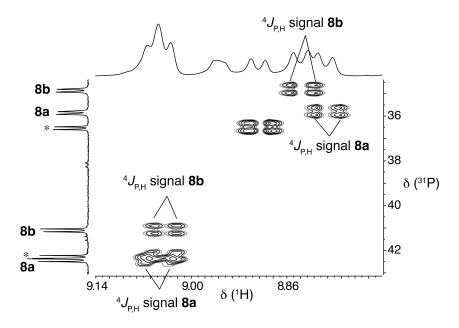
Figure S9. Section of $({}^{31}P, {}^{1}H)$ -HMQC NMR spectrum of EDA complex ${}^{13}C$ -8 (500 MHz (${}^{1}H$), CD₂Cl₂, -60 °C). The signals of ${}^{13}C$ -8 are labeled "8a" and "8b", those marked with "*" belong to the impurity seen in Figures S5 and S7.



Despite the fact that no NOESY contacts were detected between the sp^2 diazoester proton (N₂¹³C-*H*) and any other signal of the PNNP ligand, the diazoester complex **8** was identified unambiguously by (³¹P,¹H)-HMQC and by the use of ¹⁵N labeled EDA (see Experiment 4 below). The (³¹P,¹H)-HMQC spectrum showed cross peaks between the ³¹P signals and the N₂¹³C-*H* proton of the coordinated diazoester in **8a** and **8b**, which had been previously identified by the (¹³C,¹H)-HMQC spectrum of **8** at -60 °C (Figure S9). Additionally, this spectrum shows a ⁴J_{P,H} coupling constant of about 18 Hz for both imine H atoms (despite their signals are overlapped with those of the other Ru/PNNP complexes in solution (Figure S10). This is diagnostic of two *trans*-P–Ru–N moieties and hence of the *trans* configuration.¹ Finally, (¹H-¹H)-NOESY and (³¹P,¹H)-HMQC experiments indicate that the species giving signals **8a** and **8b** are exchanging

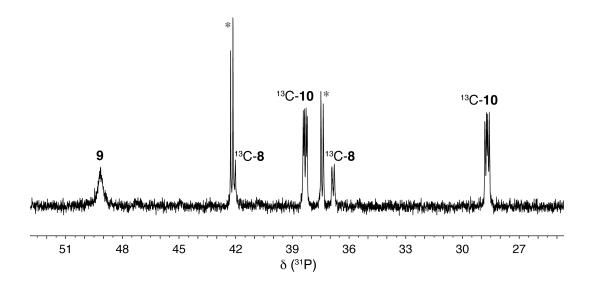
with each other even at -40 °C. Again, we attribute this observation to the interconversion between the *s*-*cis* and *s*-*trans* isomers of the CHCOOEt moiety of complex **8** (see above).

Figure S10. Section of the $({}^{31}P, {}^{1}H)$ -HMQC spectrum of *trans*-[RuCl(EDA)(PNNP)]⁺ (8) (500 MHz, CD₂Cl₂, -60 °C) showing coupling of both imine H atoms to phosphorus.



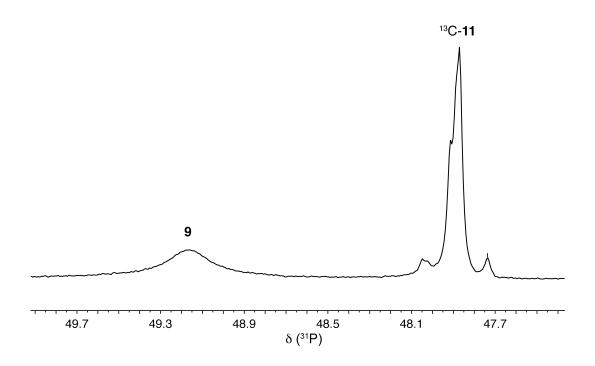
Upon raising the temperature in 20 °C-steps, the ³¹P NMR signals of the rotamers of the EDA complex **8** coalesced at –20 °C to give the a single well-resolved AB system at 20 °C (**8**, δ 42.1 (d, ² $J_{P,P'}$ = 24.2 Hz), 36.9 (d, ² $J_{P,P'}$ = 24.2 Hz)) (see Figure 4 of paper). In the temperature interval between 253 and 20 °C, imine **5a** was fully converted to aziridine **6a**, and the signals of free ¹³C-EDA disappeared from the ¹H and ¹³C NMR spectra. After few minutes at 20 °C, the ¹³C-EDA complex **8** was converted to the carbene complex *trans*-[RuCl(¹³CHCOOEt)(PNNP)]⁺ (¹³C-**10**) (Figure S11).

Figure S11. ³¹P NMR spectrum of the reaction solution of $4PF_6$ with imine **5a** (1 equiv) and ¹³C-EDA (10 equiv) after few minutes at room temperature (202 MHz, CD₂Cl₂, 25 °C). The main species in solution is the carbene complex ¹³C-**10**. The other signals belong to the EDA complex ¹³C-**8** (in the fast exchange regime), dinitrogen complex **9**, and to the unknown impurity "*" (see Figures S5 and S7).



The ³¹P and ¹³C NMR spectra indicated that the conversion of **8** to **10** begins after the disappearance of free ¹³C-EDA from the reaction solution and is quantitative after 15 min. Then, the *trans*-carbene complex **10** decomposes within 4 h to the alkyl derivative [RuCl(¹³CH₂COOEt)(PNNP)] (**11**). The main signals in the ³¹P NMR spectrum after 10 h at 20 °C are those of the dinitrogen complex **9** (55%) at δ 49.2 and of alkyl complex **11** at ca. δ 47.9 (45%, AB part of an ABX system, where X is ¹³C) (Figure S12). As previously observed in Experiment 1, the alkyl complex **10** was detected as the main product after 3 days at 25 °C. At present, we have no explanation for its formation from *trans*-carbene **9**.

Figure S12. ³¹P NMR spectrum of the reaction solution in Figure S11 after 4 h at room temperature (202 MHz, CD_2Cl_2 , 25 °C) showing the signals of alkyl complex **11** and of the N₂ complex **12**.



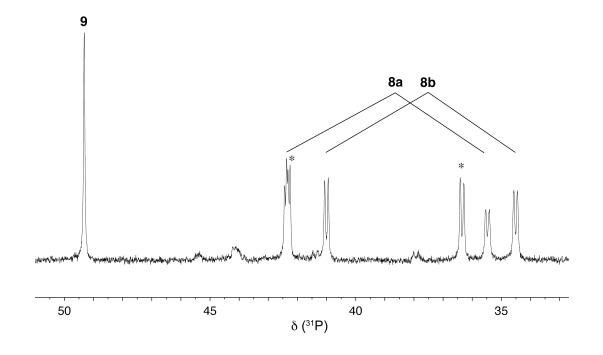
Experiment 3 was repeated three times with essentially the same results. In the last run, the ¹³C NMR signals of the coordinated diazoester of the EDA adduct **8** at δ 58.0 were irradiated at 0 °C, which left the intensity of the signal of free N₂¹³CHCOOEt unchanged, indicating that the exchange between free and coordinated EDA is slow on the NMR time scale at this temperature.

Experiment 4: [RuCl(OEt₂)(PNNP)]PF₆ (4PF₆) + imine 5a, then ¹⁵N-EDA (1:1:10). To prove the coordination of EDA to ruthenium, the former experiment was repeated with ¹⁵Nlabeled EDA (10 equiv) instead of ¹³C-EDA. The Ru:imine:¹⁵N-EDA ratio was 1:1:10. [RuCl₂(PNNP)] (1) (21.5 mg, 0.026 mmol) and (Et₃O)PF₆ (6.4 mg, 0.026 mmol) were dissolved in CD₂Cl₂ (0.5 mL) and stirred overnight at room temperature, and ³¹P and ¹H NMR spectra were recorded at 25 °C and –78 °C. Imine **5a** (7.0 mg, 0.026 mmol) was added to the solution at room temperature. Then, after cooling the sample to -78 °C, ¹⁵N-EDA (28.4 µL, 0.259 mmol) was added at -78 °C, and the sample was transferred to the precooled NMR spectrometer.

After warming to -20 °C for 15 min to ensure aziridine formation, the sample was cooled at -60 °C. A (¹³C,¹H)-HMQC experiment confirmed the formation of the aziridine. The ³¹P NMR spectrum at the same temperature (-60 °C) showed that the Et₂O adduct **4PF**₆ was quantitatively converted to the diazoester complex **8** (signals **8a+8b**), the unknown impurity at δ 42.3 and 36.3, and to the dinitrogen complex **9** with the same pattern observed in Experiment 3. As no P,N coupling was detected, the sample was further cooled down to -80 °C. At this temperature, the high-frequency ³¹P NMR signals of *trans*-[RuCl(¹⁵N₂CHCOOEt)(PNNP)]⁺ (¹⁵N-**8**) showed coupling to ¹⁵N (δ 42.4 and 41.1, ²*J*_{P,P} = 25.3, ²*J*_{P,N} = 2.4 Hz for both) (see Figure 5 of paper). Additionally, the ¹⁵N NMR spectrum at -60 °C showed two broad signals corresponding to the two isomers of ¹⁵N-**8** along with free ¹⁵N-EDA, ¹⁵NN, and coordinated ¹⁵NN (see Figure 6 of paper).

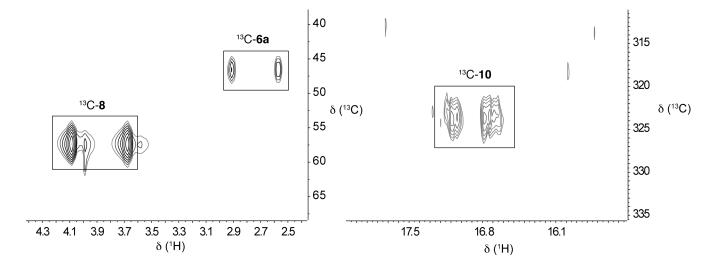
Experiment 5: [**RuCl(OEt**₂)(**PNNP**)]**PF**₆ (**4PF**₆) + ¹³**C-EDA**, then **5a** (1:10:1). The goal of this experiment was to check whether aziridine **5a** is formed in the presence of the diazoester complex **8** *after quantitative consumption of EDA*. [**RuCl**₂(**PNNP**)] (**1**) (21.8 mg, 0.026 mmol) and (Et₃O)**PF**₆ (6.5 mg, 0.026 mmol) were dissolved in CD₂Cl₂ (0.5 mL) and stirred at room temperature overnight. The formation of **4PF**₆ was confirmed by ³¹P and ¹H NMR spectroscopy at 25 °C and -78 °C. Then, ¹³C-EDA (28.8 µL, 0.262 mmol) was added at -78 °C, and the mixture was warmed to 0 °C. After 30 min, the ¹H and ¹³C NMR signals of free EDA had disappeared. Then, the mixture was cooled to -78 °C, and ³¹P NMR spectrum showed the signals of the EDA adduct **8** (**8a+8b**), N₂ complex **9**, and of the unknown impurity (marked "*") (Figure S13), with the same pattern observed in the presence of imine **5a** (see Experiment 2).

Figure S13. ³¹P NMR spectrum (202 MHz, CD_2Cl_2 , -78 °C) recorded just after the addition of ¹³C-EDA (10 equiv) to **4PF**₆ showing the signals of the EDA complex ¹³C-**8** (**8a** and **8b**). The other signals are those of the dinitrogen complex **9** and of the unknown impurity (marked "*", see Figure S5).



Then, imine **5a** (7.1 mg, 0.026 mmol) was added to the solution at -78 °C. The sample temperature was increased in 20 °C-steps. At -20 °C, a ($^{13}C, ^{1}H$)-HMQC experiment (Figure S14) indicated the formation of aziridine **6a** and the decomposition of the diazoester complex **8** to the carbene complex **10** as usually observed after the consumption of free EDA.

Figure S14. Section of the ($^{13}C, ^{1}H$)-HMQC experiment after addition of imine **5a** (1 equiv) to a solution containing the EDA adduct **8** at -78 °C (500 MHz (^{1}H), CD₂Cl₂).

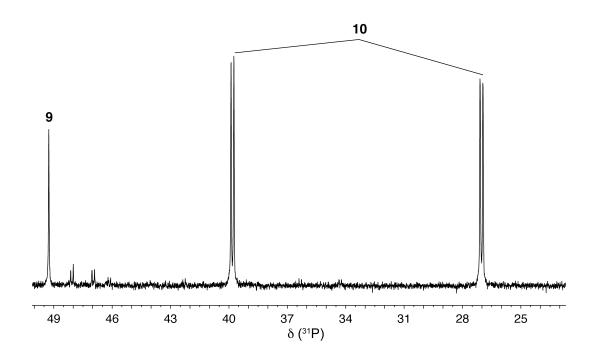


Experiment 6: Non-labeled [RuCl(CHOOEt)(PNNP)]⁺ (10) + ¹³C-EDA (1:2). [RuCl₂(PNNP)] (1) (19.8 mg, 0.024 mmol) and TlPF₆ (8.3 mg, 0.024 mmol) were dissolved in CD₂Cl₂ (0.5 mL) and stirred overnight at room temperature, The ³¹P and ¹H NMR spectra at 25 °C showed the formation of the five-coordinate complex [RuCl(PNNP)]PF₆ (2). Then, EDA (5.2 μ L, 0.024 mmol) was added at room temperature, and the mixture was cooled down to -78 °C. The ³¹P and ¹H NMR spectra showed full conversion of [RuCl(PNNP)]PF₆ (2) to [RuCl(CHCOOEt)(1a)]⁺ (10) (85%) and to the dinitrogen complex 9 (15%) (Figure S15).

Upon addition of ¹³C-EDA (10.4 μ L, 0.048 mmol) at -78 °C, the (¹³C,¹H)-HMQC and ¹H NMR spectra showed the signals of the diazoester complex [RuCl(N₂¹³CHCOOEt)(PNNP)]⁺ (**8**), traces of [RuCl(¹³CHCOOEt)(PNNP)]⁺ (¹³C-**10**), and the signals of different isotopomers of diethyl maleate. The isotopic distribution, as determined by integration of the ¹H NMR spectrum, was 41% diethyl 2-(¹³C)-maleate, 52% diethyl 2,3-bis(¹³C)-maleate, and 7% diethyl maleate.

Upon increasing the temperature, the ratio between labeled [RuCl(13 CHCOOEt)(PNNP)]PF₆ (13 C-**10**) and the nonlabeled analogue [RuCl(CHCOOEt)(PNNP)]PF₆ (**10**) gradually increased.

Figure S15. ³¹P NMR spectrum of the reaction solution of [RuCl(PNNP)]PF₆ (2) with EDA (1 equiv) at room temperature (202 MHz, CD_2Cl_2 , -78 °C). The products are the carbene complex 10 (84%) and the dinitrogen complex 9 (16%).



Summary of NMR Spectroscopic Data

$cis-\beta$ -[RuCl(OEt₂)(PNNP)]⁺ (4PF₆):

³¹**P NMR** (202 MHz, CD_2Cl_2 , -78 °C): δ 55.5 (*d*, ²*J*_{P,P} = 29.5 Hz), 36.9 (*d*, ²*J*_{P,P} = 29.5 Hz).

Diethylmaleate (7):

¹**H NMR** (500 MHz, CD₂Cl₂, -78 °C): δ 6.29 (*s*, 2H).

2-¹³C-diethylmaleate:

¹H NMR data (500 MHz, CD₂Cl₂, -78 °C): δ 6.29 (*dd*, 1H, ²*J*_{C,H} = 2.0 Hz, ²*J*_{H,H} = 11.9 Hz, *H*C), 6.29 (*dd*, 1H, ¹*J*_{C,H} = 167 Hz, ²*J*_{H,H} = 11.9 Hz, *H*¹³C).

2,3-bis(¹³C)-diethylmaleate

¹H NMR data (500 MHz, CD_2Cl_2 , -78 °C): δ 6.29 (AA' of an AA'XX' system, 2H, ¹ $J_{C,H}$ = 166 Hz, ² $J_{C,H}$ = 16.7 Hz, ³ $J_{C,H}$ = 6.81).

trans-[RuCl(N₂¹³CHCOOEt)(PNNP)]⁺ (¹³C-8):

³¹**P** NMR (202 MHz, CD₂Cl₂, -60 °C): δ 42.4 (*d*, ²*J*_{P,P'} = 25.3 Hz), 41.1 (*d*, ²*J*_{P,P'} = 25.3 Hz), 35.8 (*d*, ²*J*_{P,P'} = 25.2 Hz), 34.8 (*d*, ²*J*_{P,P'} = 25.4 Hz). 25 °C: δ 42.1 (*d*, ²*J*_{P,P'} = 24.2 Hz), 36.9 (*d*, ²*J*_{P,P'} = 24.2 Hz). ¹**H** NMR (500 MHz, CD₂Cl₂, -60 °C): δ 3.97 (*d*, 1H, ¹*J*_{C,H} = 203 Hz, RuN₂CHCOOEt) 3.72 (*d*, 1H, ¹*J*_{C,H} = 205 Hz, RuN₂CHCOOEt).

¹³C NMR (126 MHz, CD₂Cl₂, -60 °C): δ 58.1 (*s*, RuN₂CHCOOEt), 58.0 (*s*, RuN₂CHCOOEt).

trans-[RuCl(¹⁵NNCHCOOEt)(PNNP)]⁺ (¹⁵N-8):

¹⁵N NMR (50.7 MHz, CD₂Cl₂, -60 °C): δ -24.9 (*s*, 1N, ¹⁵*N*NC), -25.3 (*s*, 1N, ¹⁵*N*NC). ³¹P NMR (202 MHz, CD₂Cl₂, -80 °C): δ 42.4 (*dd*, ²*J*_{P,P} = 25.3 Hz, ²*J*_{P,N} = 2.4 Hz), 41.1 (²*J*_{P,P} = 25.3 Hz, ²*J*_{P,N} = 2.4 Hz), 41.1 (²*J*_{P,P} = 25.3 Hz, ²*J*_{P,N} = 2.4 Hz), 35.8 (*d*, ²*J*_{P,P} = 25.2 Hz), 34.8 (*d*, ²*J*_{P,P} = 25.4 Hz).

Dinitrogen Complex [RuCl(N₂)(PNNP)] (9):

³¹**P NMR** (202 MHz, CD₂Cl₂, -78 °C): δ 49.2 (*s*). 20 °C: δ 49.2 (*br s*).

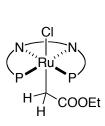
¹⁵N NMR (50.7 MHz, CD_2Cl_2 , -60 °C): δ -89.9 (*bt*, 1N, Ru-¹⁵*N*N), -40.2 (*s*, 1N, Ru-N¹⁵*N*).

trans-[RuCl(¹³CHCOOEt)(PNNP)]⁺ (¹³C-10):

³¹**P** NMR (202 MHz, CD₂Cl₂, 25 °C): 38.3 (*dd*, ${}^{2}J_{P,C} = 13.9$ Hz, ${}^{2}J_{P,P'} = 30.4$ Hz), 28.7 (*dd*, ${}^{2}J_{P,C} = 13.9$ Hz, ${}^{2}J_{P,P'} = 30.4$ Hz).

trans-[RuCl(CH₂COOEt)(PNNP)] (11):

³¹**P** NMR (202 MHz, CD₂Cl₂, 25 °C): 47.9 (d, ²J_{P,P} = 28.2 Hz), 47.8 (d, ²J_{P,P} = 28.2 Hz). ¹**H** NMR (500 MHz, CD₂Cl₂, 25 °C): δ 3.79 (d, 1H, ²J_{H,H} = 11.2 Hz, RuCHH'COOEt), 3.36 (d, 1H, ²J_{H,H} = 11.2 Hz, RuCHH'COOEt).



COOEt

(¹³C-Labeled) Unknown Species containing a X=C(H)Y moiety observed below -20

°C (see Figure S8 and footnote 25 of main paper):

¹**H** NMR (500 MHz, CD_2Cl_2 , -78 °C): δ 4.21 (*d*, 1H, ¹ $J_{C,H}$ = 206 Hz), 4.21 (*d*, 1H, ¹ $J_{C,H}$ = 213 Hz).

¹³C NMR (126 MHz, CD₂Cl₂, -78 °C): δ 55.4 (*s*), 54.6 (*s*).

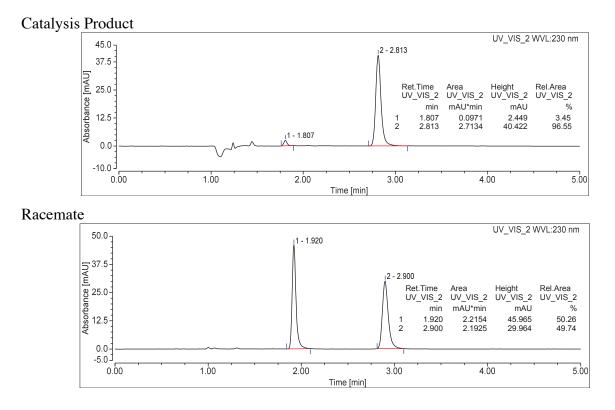
	x	Ph N ← Ph		Br 5e	Ph N Ph	Ph N Ph 5f	
entry	Y	imine	EDA	crude 6^{b}		isolated <i>cis</i> - 6	
			(equiv)	yield (%)	cis:trans	yield (%) °	ee (%) ^d
1	PF_6	5a	1	22	77:23	13	80
2	PF_6	5 a	4	35	79:21	30	53 °
3	BF_4	5 a	1	32	78:22	24	78
4	BF_4	5a	4	53	81:19	20	78
5	SbF_6	5a	1	32	85:15	24	93
6	SbF_6	5a	4	58	78:22	40	67
7	BF_4	5b	1	36	86:14	23	70
8	BF_4	5b	4	50	74:26	33	25
9	SbF_6	5b	1	18	85:15	14	91
10	BF_4	5c	1	24	79:21	16	68
11	BF_4	5c	4	20	65:35	20	17
12	SbF_{6}	5c	1	16	84:16	9	75
13	SbF_6	5d	1	30	cis only	24	79
14	SbF_{6}	5e	1	46	93:7	34	83
15	BF_4	5f	1	32	81:19	24	70
16	BF_4	5f	4	53	77:23	38	60
17	SbF_6	5f	1	40	90:10	33	93
18	SbF_6	5g	1	11	cis only	4	57
19	SbF_6	5h	1	25	90:10	17	63

Table S1. Optimization of the Asymmetric Aziridination with Selected Imines and Catalysts.^a

^a Reaction conditions: EDA (0.48 mmol, 1 equiv, or 1.92 mmol, 4 equiv, neat) was added in one portion to a CH₂Cl₂ solution (3 mL) containing the imine (0.48 mmol) and the catalyst (10 mol%) prepared by activation of [RuCl₂(PNNP)] (1) (0.048 mmol) with (Et₃O)Y (0.048 mmol). The total reaction time was 24 h at 0 °C. ^b Based on the imine, determined by ¹H NMR spectroscopy. ^c Isolated yield. ^d The absolute configuration of **6b-6h** was not assigned. ^e The isolated aziridine was contaminated with variable amounts of diethyl maleate.

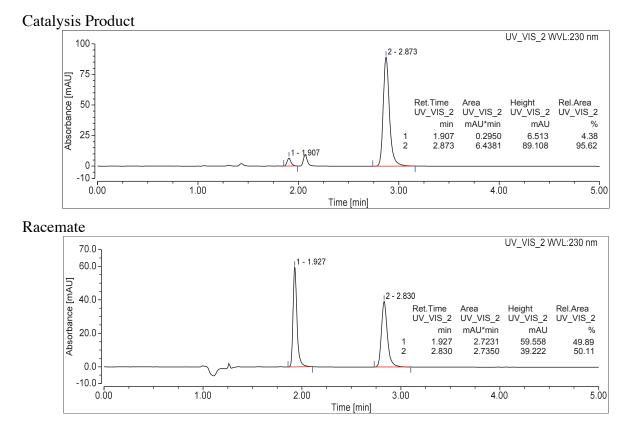
(2*R*,3*R*)-Ethyl 1-Benzhydryl-3-phenylaziridine-2-carboxylate (6a). The reaction of 5a and EDA gave aziridine 6a as a white solid after workup (see above). ¹H NMR and ¹³C NMR spectroscopic data are in agreement with published values.⁸ ¹H NMR (CDCl₃, 300.2 MHz): δ 1.00 (*t*, 3H, *J*= 7.11 Hz, *CH*₃), 2.71 (*d*, 1H, *J*= 6.84 Hz, NCHPh), 3.25 (*d*, 1H, *J*= 6.84 Hz, NCHCOOEt), 3.96 (*q*, 2H, *J*= 7.14 Hz, COOCH₂), 3.99 (*s*, 1H, *CH*Ph₂), 7.18 – 7.64 (*m*, 14H, H_{arom}). ¹³C NMR (CDCl₃, 125.8 MHz): δ 14.0, 46.5, 48.1, 60.6, 77.8, 127.3 – 128.6, 135.1, 134.0, 142.5, 142.6, 167.8. Chiral HPLC: IB column, 3 µm, eluent: hexane/2-propanol (95:5), flow rate 2.0 mL/min, *R*_t (min) = 1.8 (minor, (2*S*,3*S*)-6a), 2.8 (major, (2*R*,3*R*)-6a). [α]₀²⁰ = 22.9 ± 1 (c = 1.5, CHCl₃) @ 93% ee (Table S1, entry 5). Absolute configuration assigned on the basis of the sign of the reported optical rotation.⁸ HRMS (MALDI): Calcd. for C₂₄H₂₄NO₂ *m/z* 358.1802 found *m/z* 358.1801.





Ethyl 1-Benzhydryl-3-(4-chlorophenyl)aziridine-2-carboxylate (6b). The reaction of **5b** and EDA gave 6b as a white solid after workup (see above). ¹H NMR (CDCl₃, 500.2 MHz): δ 1.06 (*t*, 3H, *J*= 6.95 Hz, H13), 2.72 (*d*, 1H, *J*= 6.35 Hz, H1), 3.20 (*d*, 1H, *J*= 6.45 Hz, H2), 3.97 (*s*, 1H, H15), 3.99 (*q*, 2H, *J*= 6.55 Hz, H12), 3.97 (*s*, 1H, H15), 7.22 – 7.62 (*m*, 14H, H_{arom}). ¹³C NMR (CDCl₃, 125.8 MHz): δ 14.4, 46.9, 47.7, 61.1, 78.1, 127.6 – 129.6, 133.6, 134.0, 142.6, 142.8, 167.9. Chiral HPLC: IB column, 3 µm; eluent: hexane/2-propanol (95:5); flow rate: 2.0 mL/min; *R*₁ (min) = 1.9 (minor), 2.8 (major), 91% ee (Table S1, entry 9). [α]_D²⁰ = 21.4 ± 0.1 @ 91% ee (c = 0.368, CHCl₃). The absolute configuration was not assigned. HRMS (MALDI): Calcd. for C₂₄H₂₃ClNO₂ *m*/*z* 392.1412 found *m*/*z* 392.1412.





S27

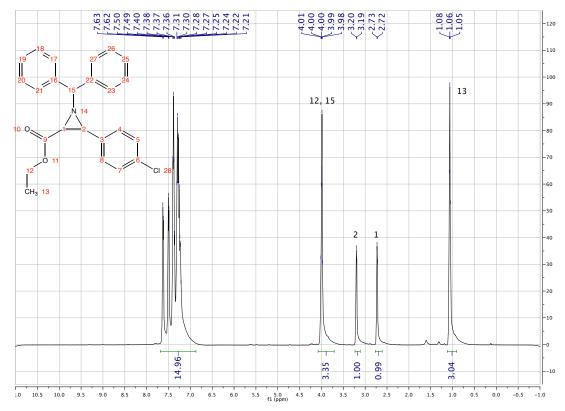
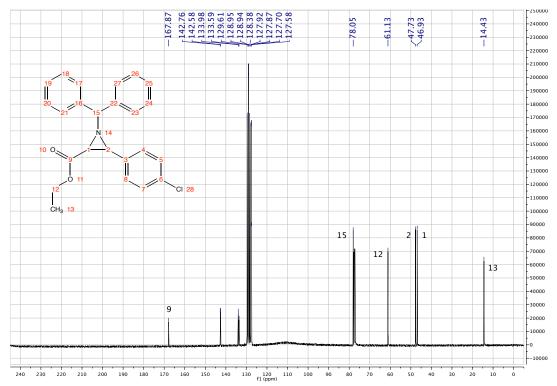


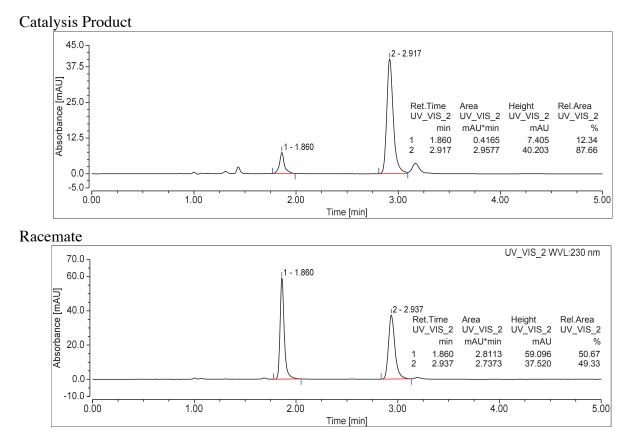
Figure S18. ¹H NMR Spectrum of 6b (500.2 MHz, CDCl₃).

Figure S19. ¹³C NMR Spectrum of 6b (125.8 MHz, CDCl₃).



Ethyl 1-Benzhydryl-3-(4-fluorophenyl)aziridine-2-carboxylate (6c). The reaction of 5c and EDA gave 6c as a white solid after workup (see above). ¹H NMR (CDCl₃, 500.2 MHz): δ 1.05 (*t*, 3H, *J*= 7.10 Hz, H13), 2.71 (*d*, 1H, *J*= 6.80 Hz, H1), 3.22 (*d*, 1H, *J*= 6.85 Hz, H2), 3.97 (*s*, 1H, H15), 3.99 (*q*, 2H, *J*= 6.60 Hz, H12), 6.96 – 7.64 (*m*, 14H, H_{arom}). ¹³C NMR (CDCl₃, 125.8 MHz): δ 14.4, 46.8, 47.7, 61.1, 78.1, 115.0, 115.2, 127.9 – 129.0, 142.7, 142.9, 161.7, 163.6, 168.0. Chiral HPLC: IB column, 3 µm; eluent: hexane/2-propanol (95:5); flow rate: 2.0 mL/min; *R*₁ (min) = 1.8 (minor); 2.9 (major), 75% ee (Table S1, entry 12). [α]_D²⁰ = 39 ± 0.1 @ 75% ee (c = 0.232, CHCl₃). The absolute configuration was not assigned. HRMS (MALDI): Calcd. for C₂₄H₂₃FNO₂ *m/z* 376.1707 found *m/z* 376.1707.





S29

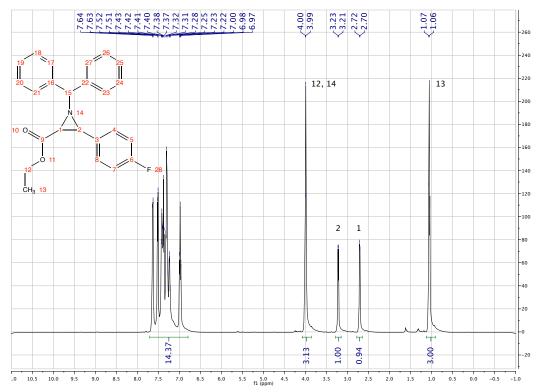
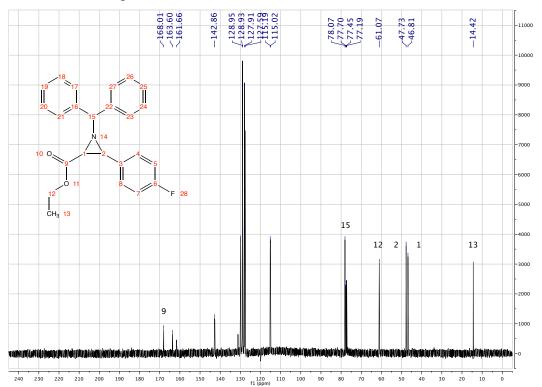


Figure S21. ¹H NMR Spectrum of **6c** (500.2 MHz, CDCl₃).

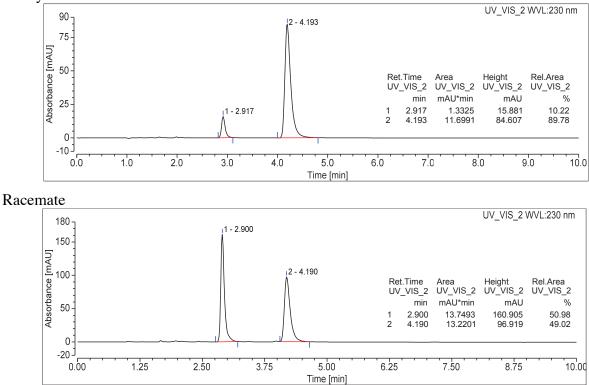
Figure S22. ¹³C NMR Spectrum of 6c (125.8 MHz, CDCl₃).



Ethyl 1-Benzhydryl-3-(4-(methoxycarbonyl)phenyl)aziridine-2-carboxylate (6d). The reaction of 5d and EDA gave 6d as a white solid after workup (see above). ¹H NMR (CDCl₃, 400 MHz): δ 1.02 (t, J= 7.2 Hz, 3H, H17), 2.77 (d, J= 6.8 Hz, 1H, H1), 3.26 (d, J= 6.8 Hz, 1H, H2), 3.91 (s, 3H, H10), 3.96 (qd-like, AB part of ABX₃ system, 2H, ³J= 7.2, 2H, H16), 4.00 (s, 1H, H19), 7.18 – 7.64 (m, 12H, H_{arom}), 7.97 (m, 2H, H_{arom}). ¹³C NMR (CDCl₃, 101 MHz): δ 14.02, 46.72 47.62, 52.02, 60.76, 77.62, 127.17, 127.34, 127.44, 127.53, 127.89, 128.56, 129.12, 129.23, 140.31, 142.15, 142.30, 166.96, 167.32. Chiral HPLC: IB column, 3 μm; eluent: hexane/2-propanol (95:5); flow rate: 2.0 mL/min; R_t (min) = 2.9 (minor); 4.2 (major), 79% ee (Table S1, entry 13). [α]_D²⁰ = 7.7 ± 0.1 @ 79% ee (c = 1.16, CHCl₃). The absolute configuration was not assigned. HRMS (MALDI): Calcd. for C₂₆H₂₆NO₄ m/z 416.1856 found m/z 416.1857.







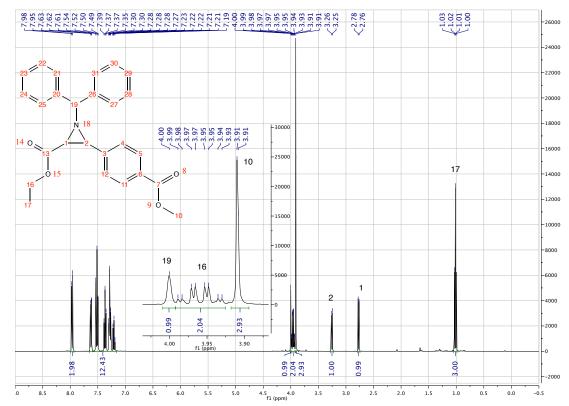
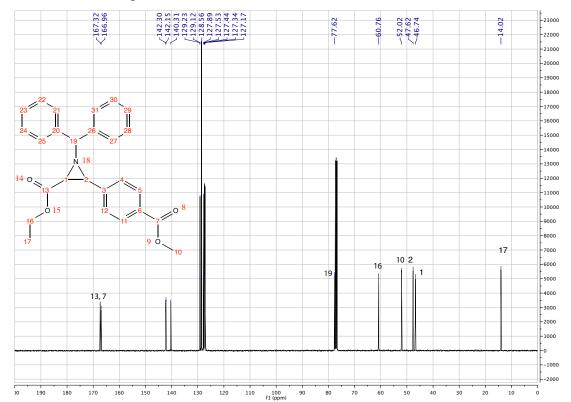


Figure S24. ¹H NMR Spectrum of 6d (400 MHz, CDCl₃).

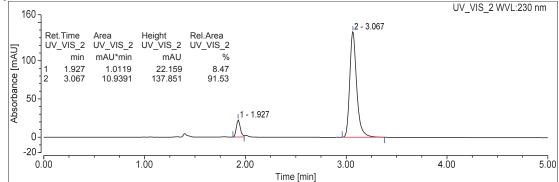
Figure S25. ¹³C NMR Spectrum of 6d (101 MHz, CDCl₃).



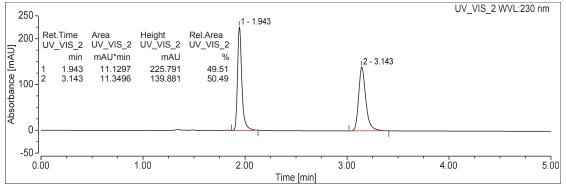
Ethyl 1-Benzhydryl-3-(3-bromophenyl)aziridine-2-carboxylate (6e). The reaction of 5e and EDA gave 6e as a white solid after workup (see above). ¹H NMR (CDCl₃, 500.2 MHz): δ 1.08 (*t*, 3H, *J*= 7.1 Hz, H14), 2.75 (*d*, 1H, *J*= 6.8 Hz, H1), 3.20 (*d*, 1H, *J*= 6.8 Hz, H2), 4.0 (s, 1H, H1), 4.02 (*qd* like, AB part of ABX₃ system, 2H, ³*J*= 7 Hz, H13), 7.14 – 7.66 (*m*, 14H, H_{arom}). ¹³C NMR (CDCl₃, 125.8 MHz): δ 14.05, 46.63, 47.20, 60.80, 77.66, 121.91, 126.59, 127.21, 127.35, 127.51, 127.58, 128.57, 128.61, 129.42, 130.54, 130.90, 137.51, 142.16, 142.33, 167.45. Chiral HPLC: IB column, 3µm; eluent: hexane/2-propanol (95:5); flow rate: 2.0 mL/min; *R*_t (min) = 1.9 (minor); 3.1 (major), 83% ee (Table S1, entry 14). $[\alpha]_D^{20} = 30.4 \pm 0.2$ @ 83% ee (c = 1.44, CHCl₃). The absolute configuration was not assigned. HRMS (MALDI): Calcd. for C₂₄H₂₃BrNO₂ *m/z* 436.0907 found *m/z* 436.0906.

Figure S26. HPLC traces of 6e.





Racemate



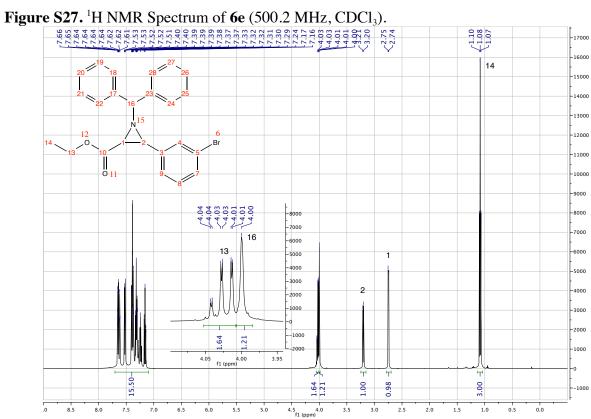
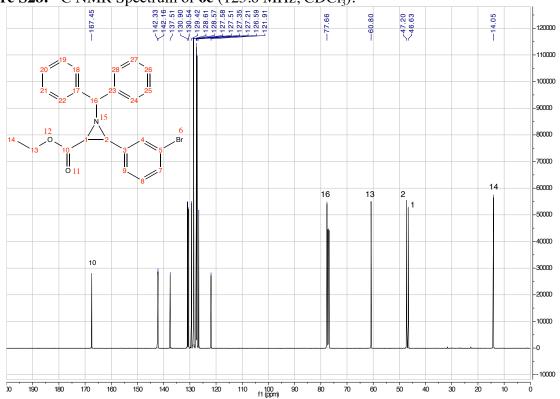


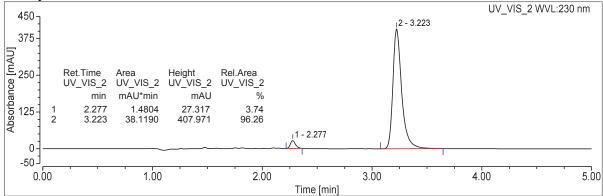
Figure S28. ¹³C NMR Spectrum of 6e (125.8 MHz, CDCl₃).



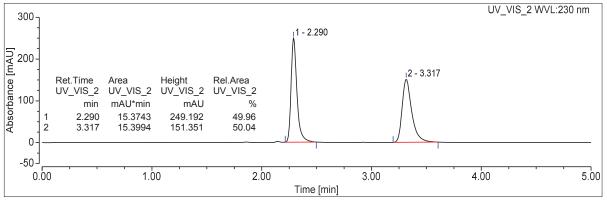
Ethyl 1-Benzhydryl-3-(2-naphthalene-2-yl)aziridine-2-carboxylate (6f). The reaction of 5f and EDA gave 6f as a white solid after workup (see above). ¹H NMR (CDCl₃, 500.2 MHz): $\delta 0.98 (t, 3H, J= 7.4 \text{ Hz}, H13), 2.80 (d, 1H, J= 6.85 \text{ Hz}, H1), 3.40 (d, 1H, J= 6.80 \text{ Hz}, H2), 3.94$ $(q, 2H, J= 7.50 \text{ Hz}, H12), 4.06 (s, H15) 7.19 - 7.92 (m, 14H, H_{arom}).$ ¹³C NMR (CDCl₃, 125.8 MHz): $\delta 14.1, 46.9, 48.4, 60.8, 77.9, 125.8 - 128.7, 132.8, 133.0, 133.1, 142.5, 142.6, 167.9.$ Chiral HPLC: IB column, 3µm; eluent: hexane/2-propanol (95:5); flow rate: 2.0 mL/min; R_t (min) = 2.7 (minor), 3.2; (major), 93% ee (Table S1, entry 17). $[\alpha]_D^{20} = 10.0 \pm 0.3$ @ 93% ee (c = 0.81, CHCl₃). The absolute configuration was not assigned. HRMS (MALDI): Calcd. for $C_{28}H_{26}NO_2 m/z$ 408.1958 found m/z 408.1958.











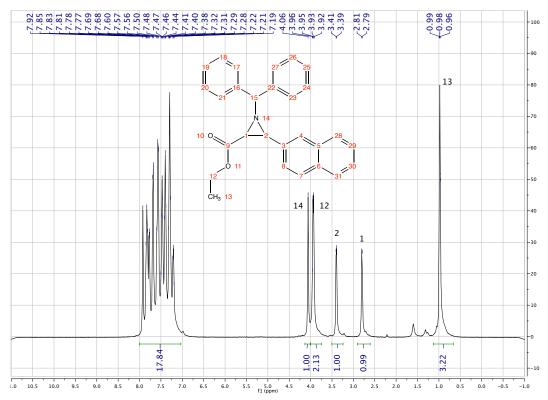
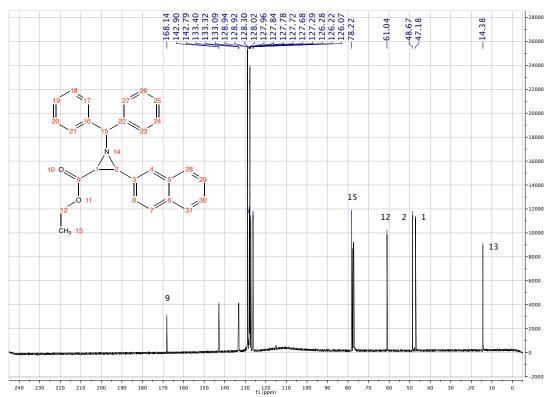


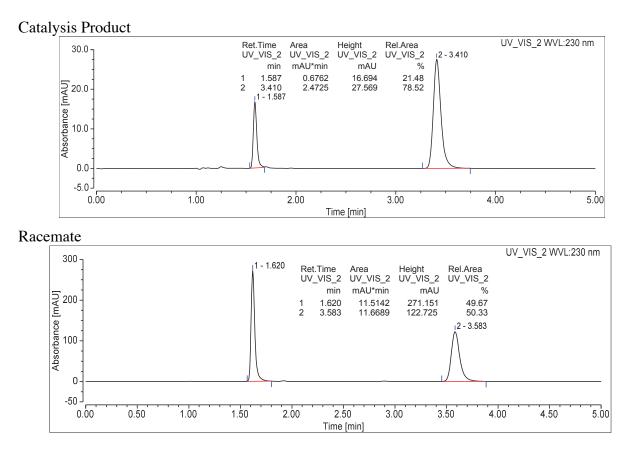
Figure S30. ¹H NMR Spectrum of 6f (500.2 MHz, CDCl₃).

Figure S31. ¹³C NMR Spectrum of 6f (125.8 MHz, CDCl₃).



Ethyl 1-Benzhydryl-3-(4-isopropylphenyl)aziridine-2-carboxylate (6g). The reaction of 5g and EDA gave 6g as a white solid after workup (see above). ¹H NMR (CDCl₃, 500.2 MHz): δ 0.99 (*t*, 3H, *J*= 6.95 Hz, H13), 1.24 (*d*, 6H, *J*= 6.95 Hz, H29 and H30), 2.67 (*d*, 1H, *J*= 6.75 Hz, H1), 2.88 (*sep*, 1H, *J*= 6.90 Hz, H28), 3.23 (*d*, 1H, *J*= 6.85 Hz, H2), 3.99 (*q*, 2H, *J*= 7.3 Hz, H12), 3.97 (*s*, 1H, H15), 7.13 – 7.64 (*m*, 14H, H_{arom}). ¹³C NMR (CDCl₃, 125.8 MHz): δ 14.30, 24.3, 24.4, 34.2, 46.7, 48.4, 60.9, 77.9, 126.3 - 128.9, 132.8, 143.0, 148.3, 168.3. Chiral HPLC: IB column, 3µm; eluent: hexane/2-propanol (95:5); flow rate: 2.0 mL/min; *R*₁ (min) = 1.6 (minor); 3.4 (major), 57% ee (Table S1, entry 18). The absolute configuration was not assigned. HRMS (MALDI): Calcd. for C₂₇H₃₀NO₂ *m/z* 400.2271 found *m/z* 400.2271.





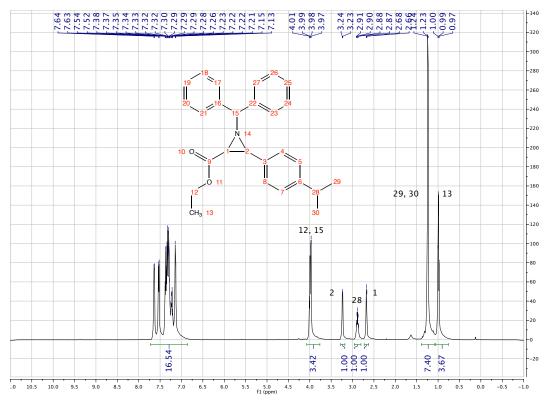
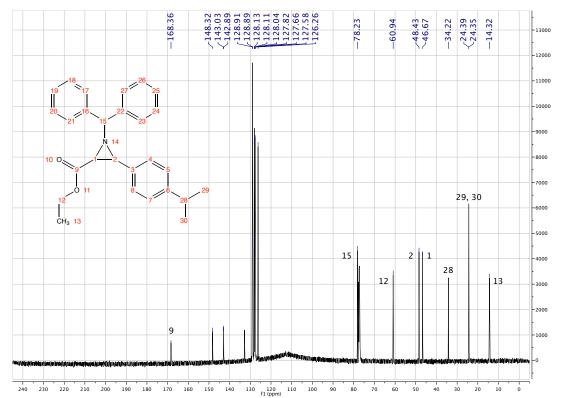


Figure S33. ¹H NMR Spectrum of 6g (500.2 MHz, CDCl₃).

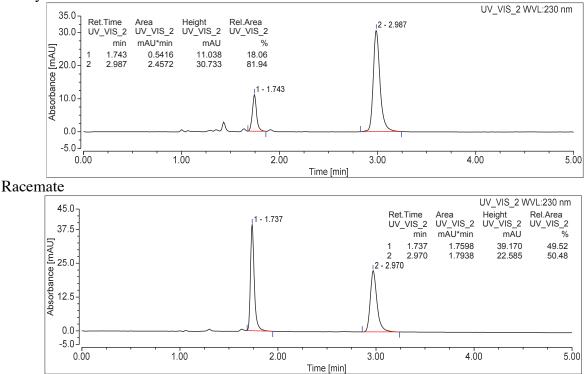
Figure S34. ¹³C NMR Spectrum of 6g (125.8 MHz, CDCl₃).



Ethyl 1-Benzhydryl-3-(*p*-tolyl)aziridine-2-carboxylate (6h). The reaction of 5g and EDA gave 6g as a white solid after workup (see above). ¹H NMR and ¹³C NMR spectroscopic data are in agreement with published values.⁹ ¹H NMR (CDCl₃, 300.2 MHz): δ 1.00 (*t*, 3H, *J*= 7.1 Hz, COOCH₂CH₃), 2.27 (*s*, 3H, PhCH₃), 2.63 (*d*, 1H, *J*= 6.8 Hz, NCHPh), 3.17 (*d*, 1H, *J*= 6.84 Hz, NCHCOOEt), 3.93 (*s*, 1H, CHPh₂), 3.94 (*q*, *J*= 7.1 Hz, COOCH₂CH₃), 7.03 – 7.60 (*m*, 14H, H_{arom}). ¹³C NMR (CDCl₃, 75 MHz): δ 14.06, 21.20, 46.42, 48.11, 60.59, 77.83, 127.23, 127.30, 127.42, 127.60, 127.73, 128.52, 132.06, 136.96, 142.51, 142.62, 167.88. Chiral HPLC: IB column, 3 µm, eluent: hexane/2-propanol (95:5), flow rate 2.0 mL/min, R_1 (min) = 1.7 (minor), 3.0 (major), 63 % ee (Table S1, entry 19). $[\alpha]_D^{20} = 23.6 \pm 0.3$ @ 63% ee (c = 0.364, CHCl₃). The absolute configuration was not assigned. HRMS (MALDI): Calcd. for C₂₅H₂₆NO₂ *m/z* 372.1958.

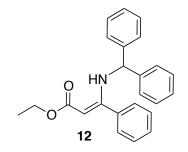
Figure S35. HPLC traces of 6h.





Synthesis of Racemic Aziridines 6a-6h. The racemic aziridines 6a-6h were prepared according to a published procedure¹⁰ and used as references for the chiral HPLC determination of the enantiomeric purity of the catalysis products. The corresponding HPLC traces are shown above. Boron trifluoride ethyl etherate (0.11 mmol, 0.1 equiv) was added to a CH_2Cl_2 solution (10 ml) of the imine 5a-5h (1.1 mmol, 1 equiv) and EDA (1 equiv), and the mixture was stirred for 2 h. The solvent was evaporated under reduced pressure, and the oily residue was subject to flash chromatography on silica (hexane/ethyl acetate 95:5) and cystallized from hexane. Yields were in the range 70–90%.

(Z)-Ethyl 3-(Benzhydrylamino)-3-phenylacrylate (12). The title compound was prepared as authentic sample to rule out its formation in imine aziridination with catalyst $4PF_4$ following a published procedure.¹¹ Diphenylmethaneamine (7.5 mL, 43.2 mmol, 5 equiv), ethyl 3-oxo3-phenylpropanoate (1.5 mL, 8.6 mmol, 1 equiv) and glacial acetic acid (2.5 mmol, 43.2 mmol, 5 equiv) were mixed at room temperature. Immediately, a light yellow precipitate formed. The identity of **12** was confirmed by the ¹H NMR spectrum, which showed the diagnostic broad doublet of the NH group. ¹H NMR (CDCl₃, 300.1 MHz): δ 5.61 (*d*, 1H, *J*= 10.06 Hz, CHPh₂), 9.42 (*bd*, 1H, *J*= 10.08 Hz, NH).



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