

Supporting Information

Oxidative Heck Reaction of Glycals and Aryl Hydrazines: A Palladium-Catalyzed C-Glycosylation

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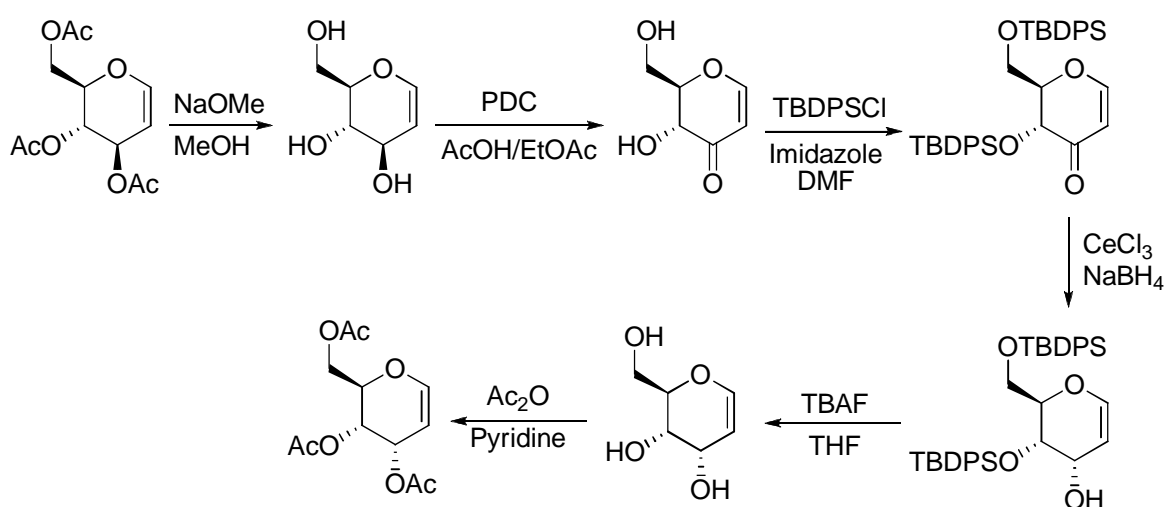
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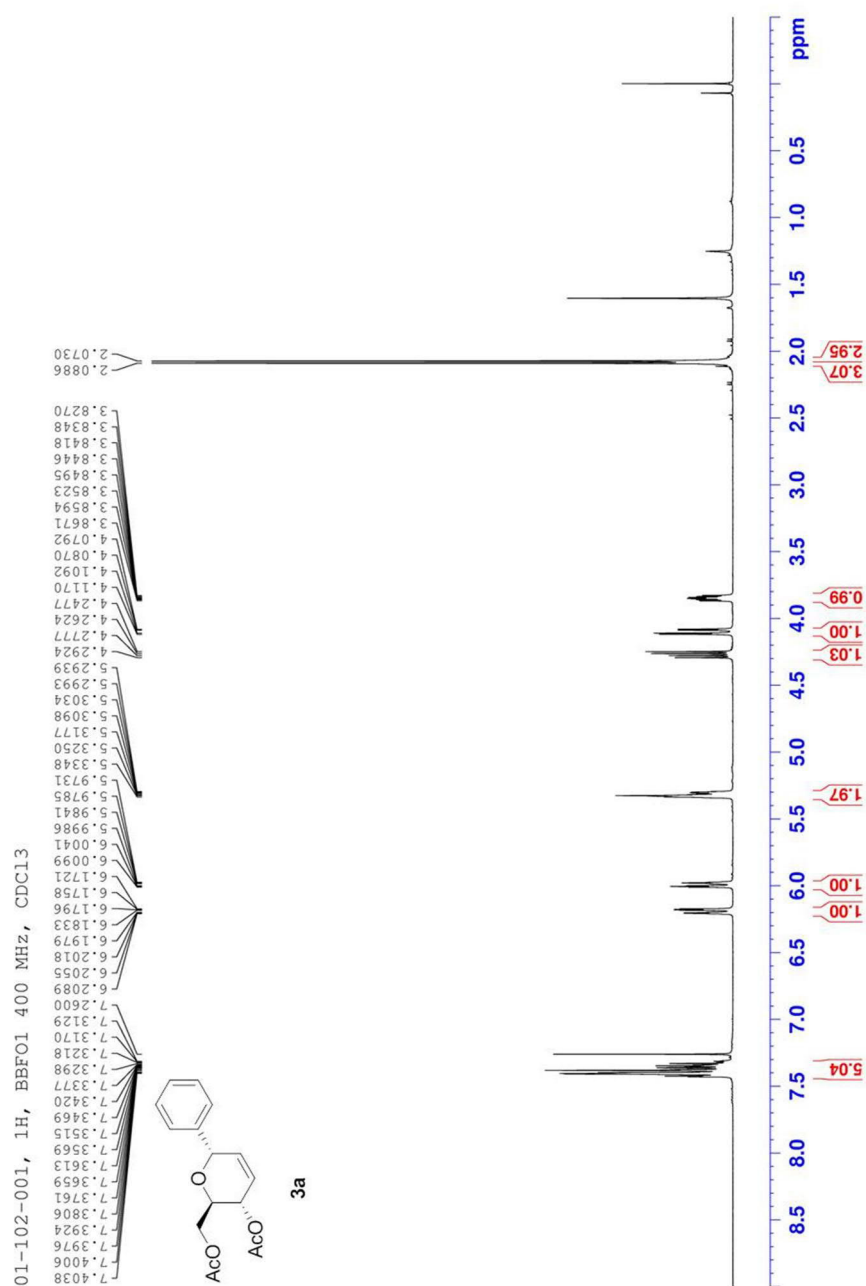
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General: All the reactions were carried out in a flame or oven dried glassware with freshly distilled dry solvents under anhydrous conditions unless otherwise indicated. Organic solutions were concentrated under reduced pressure by rotary evaporation with a water bath (temperature below 40 °C). Reactions were magnetically stirred and monitored by thin-layer chromatography (TLC) carried out on 0.25 mm silica gel plates using UV light at 254 nm as a visualizing agent and a KMnO_4 solution as stain. Product purification by flash column chromatography was accomplished using silica gel 60 (0.010–0.063 nm). Technical grade solvents were used for chromatography and were distilled prior to use. Optical rotations were measured in CHCl_3 with a 1 cm cell (c given in g/100 mL). NMR spectra were recorded at room temperature on 400 MHz. The residual solvent signals were taken as the reference (7.26 ppm for ^1H NMR spectra and 77.0 ppm for ^{13}C NMR spectra in CDCl_3). Sometimes the TMS signal at 0.0 ppm was used as an internal standard for ^1H NMR spectra. Chemical shift (δ) is reported in ppm, coupling constants (J) are given in Hz. The following abbreviations classify the multiplicity: s = singlet, d = doublet, t = triplet, m = multiplet or unresolved, br = broad signal. HR-MS (ESI) spectra were recorded on a Q-ToF mass spectrometer.

Synthesis of 3-inverted acetyl glucal ((2R,3S,4S)-2-(acetoxymethyl)-3,4-dihydro-2H-pyran-3,4-diyl diacetate) from commercially available acetyl glucal ((2R,3S,4R)-2-(acetoxymethyl)-3,4-dihydro-2H-pyran-3,4-diyl diacetate)

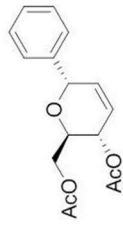


Copies of ^1H and ^{13}C NMR spectrum



01-102-001, ¹³C, BBO1 400 MHz, CDCl₃

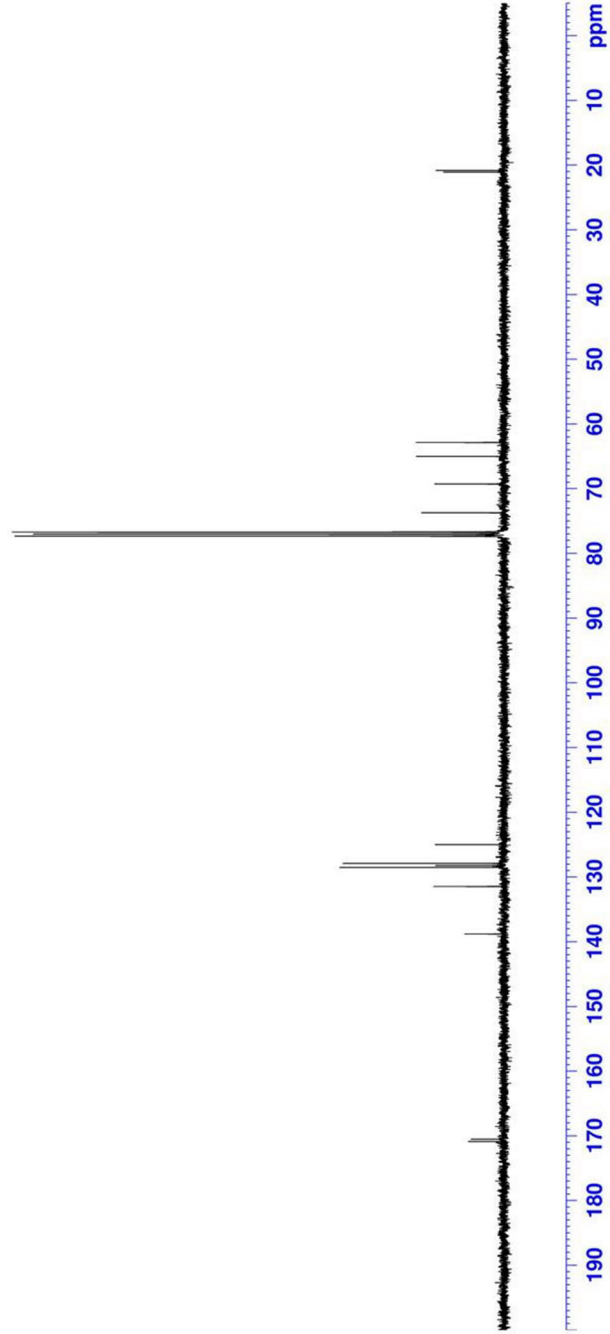
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170.49
138.82
131.48
128.53
128.25
127.91
125.02

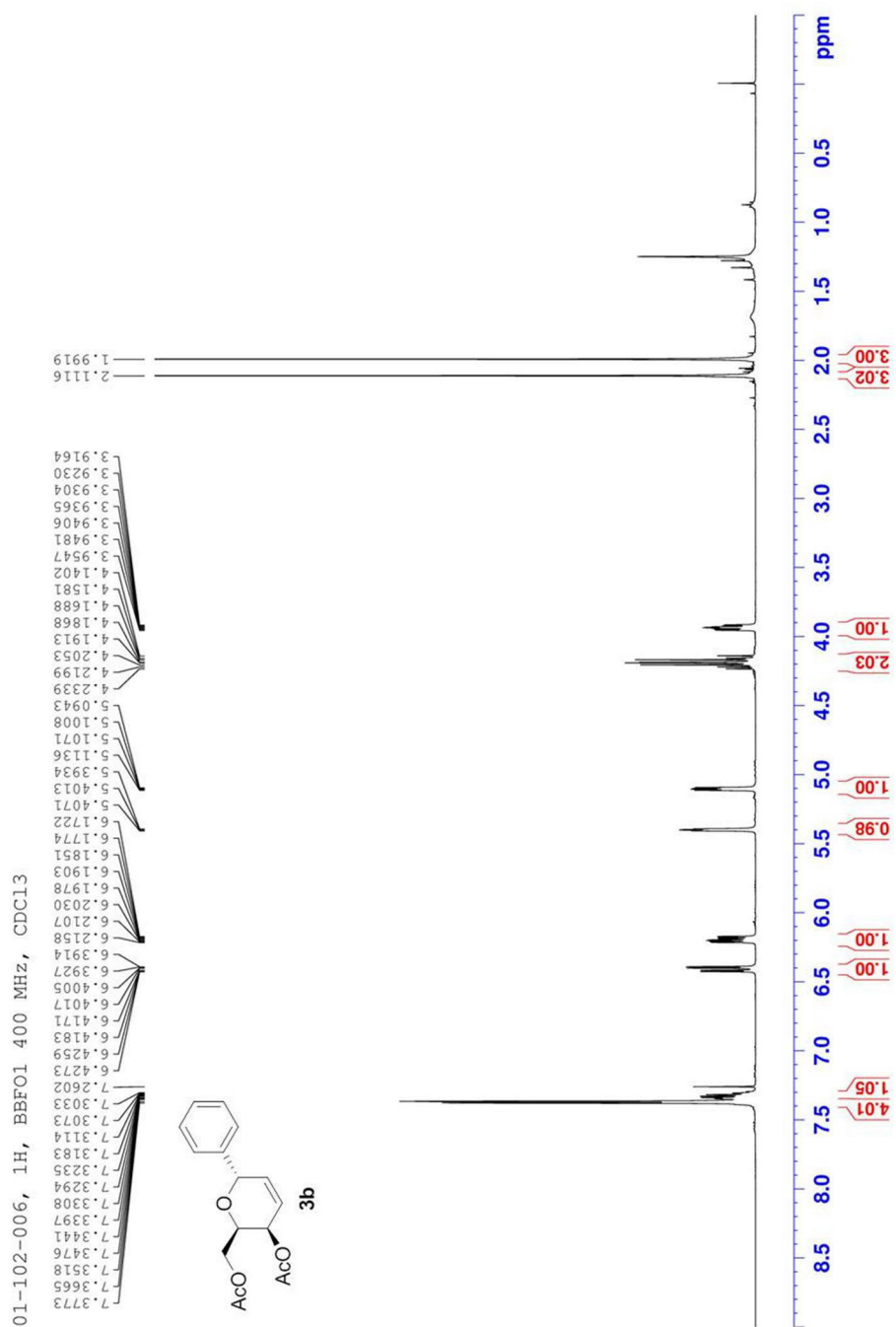


3a

73.73
69.29
65.02
62.87

21.09
20.82





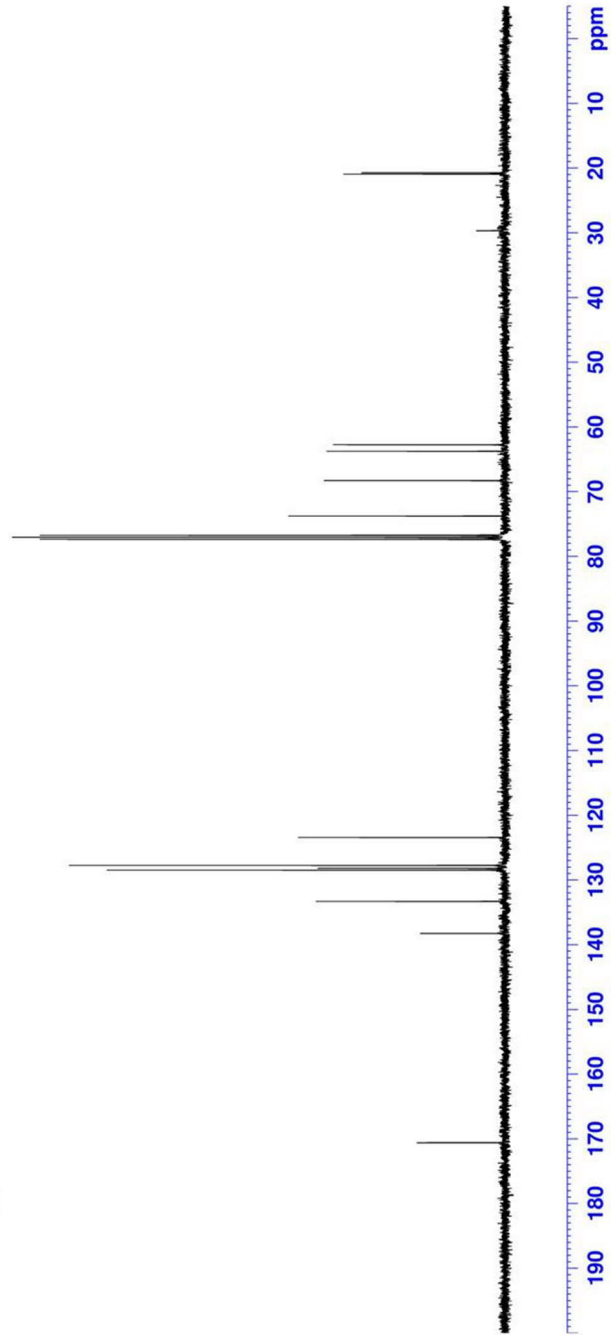
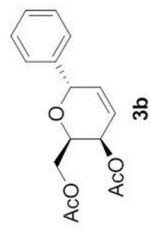
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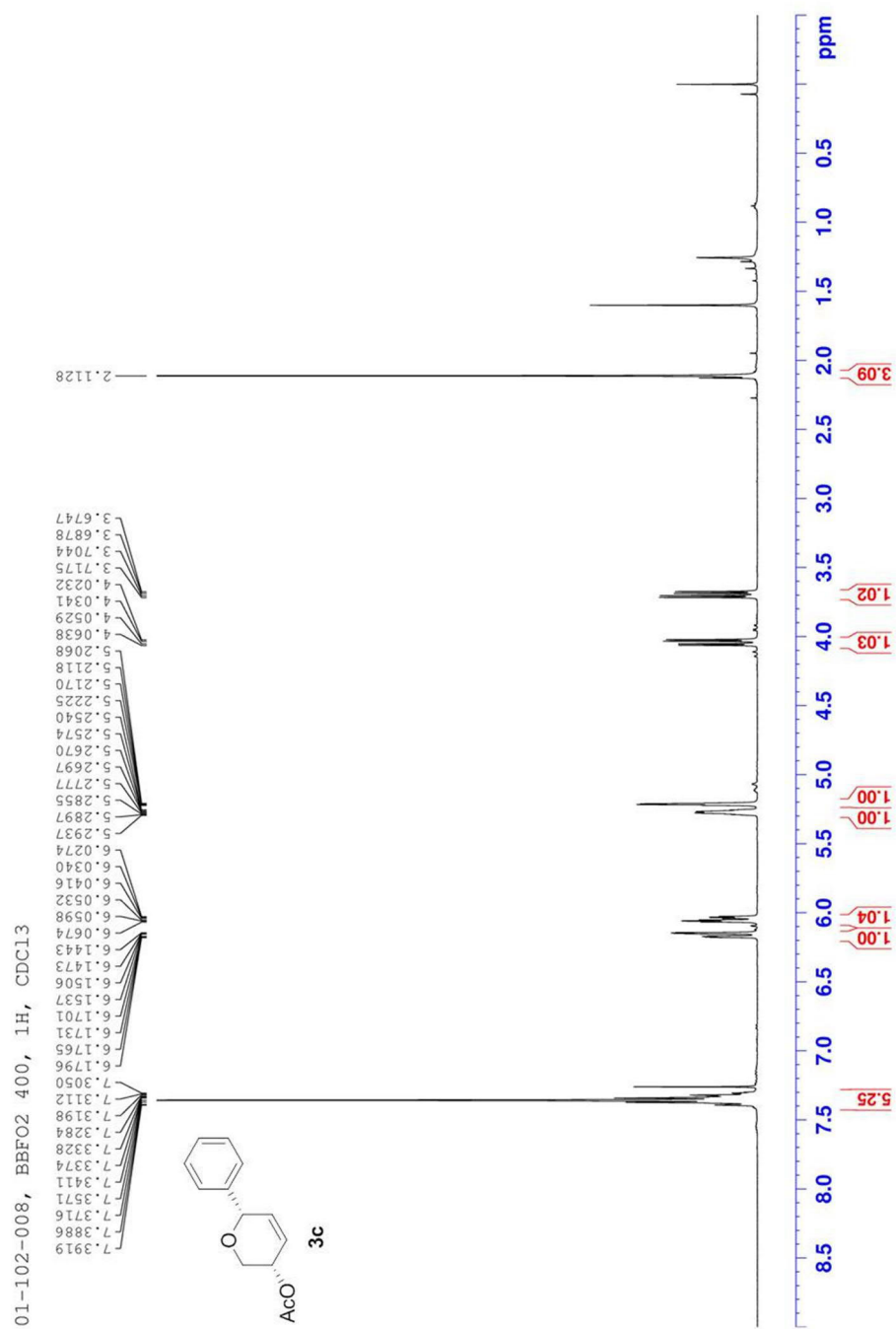
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170.56

138.29
133.33
128.49
128.21
127.76
123.45

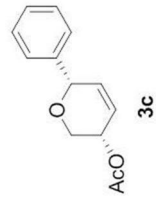
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68.30
63.76
62.75

20.94
20.73





01-102-008, BBFO2 400, 13C, CDCl3

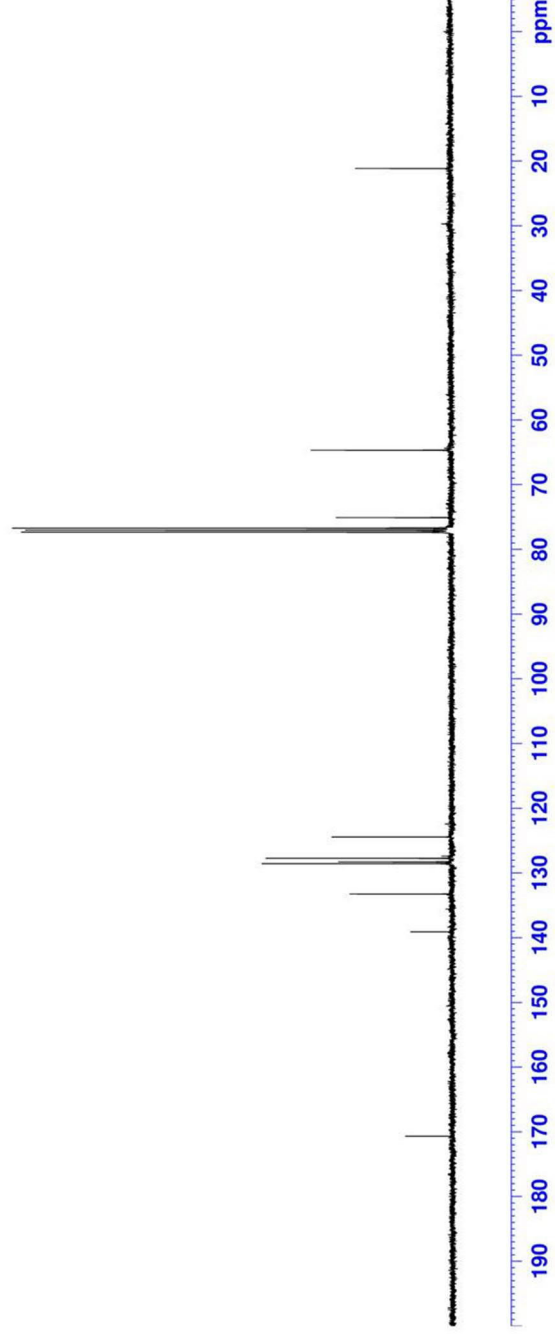


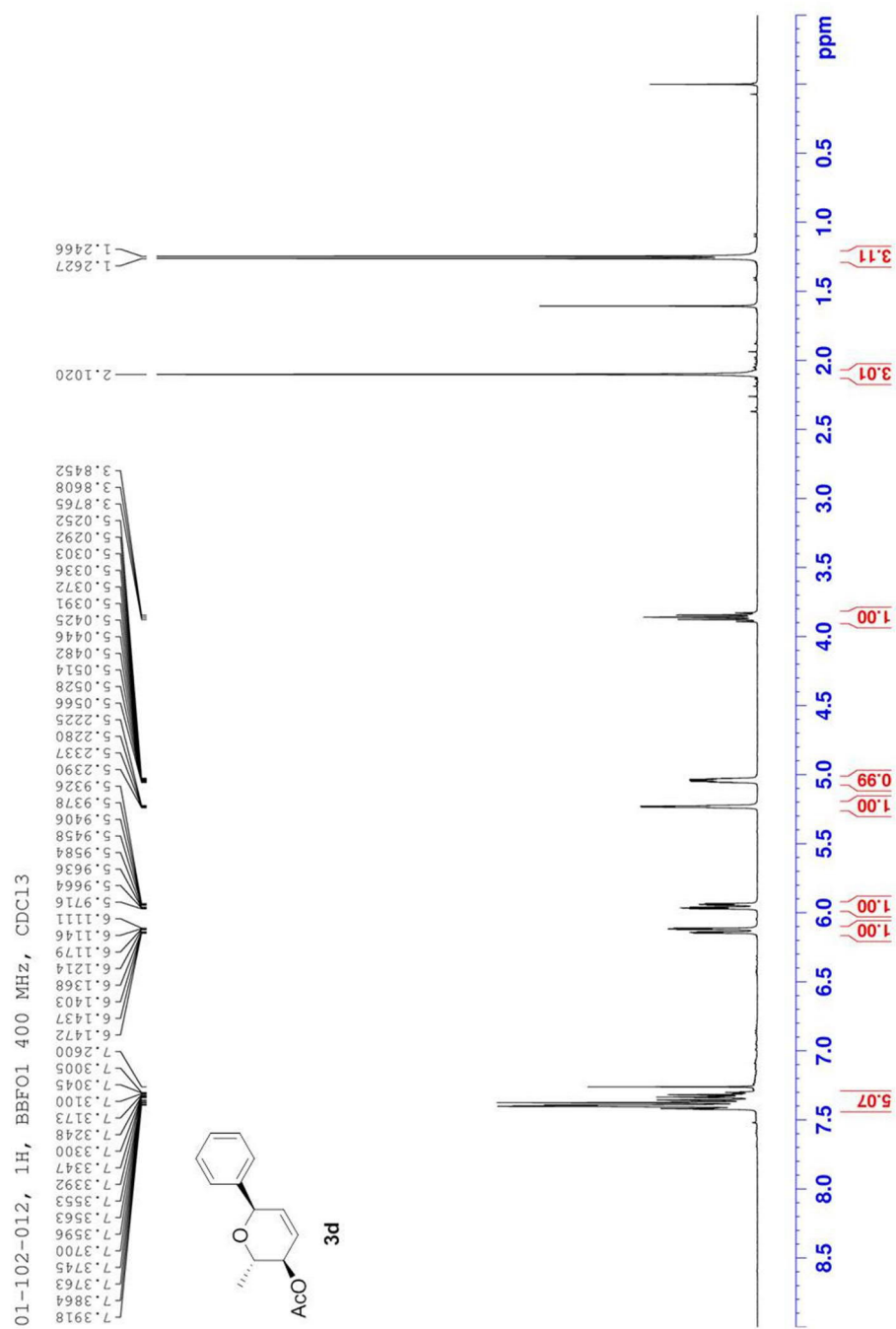
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139.11
133.27
128.57
128.29
127.76
124.45

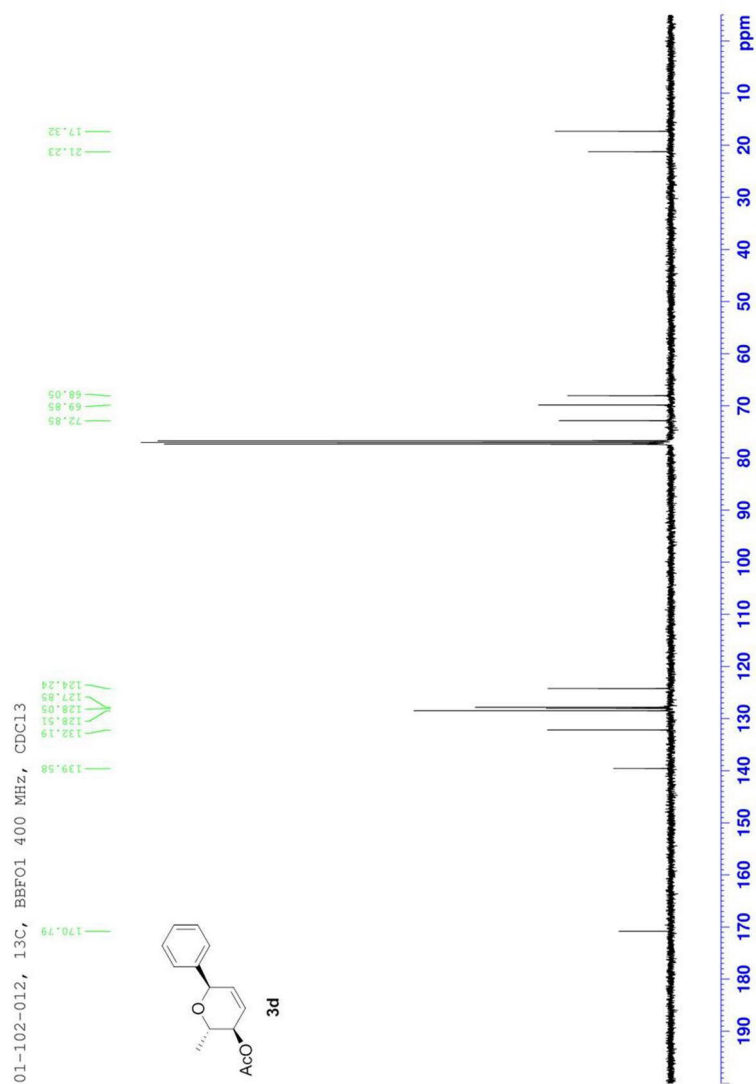
75.09

64.69
64.68

21.14

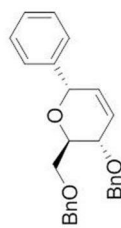




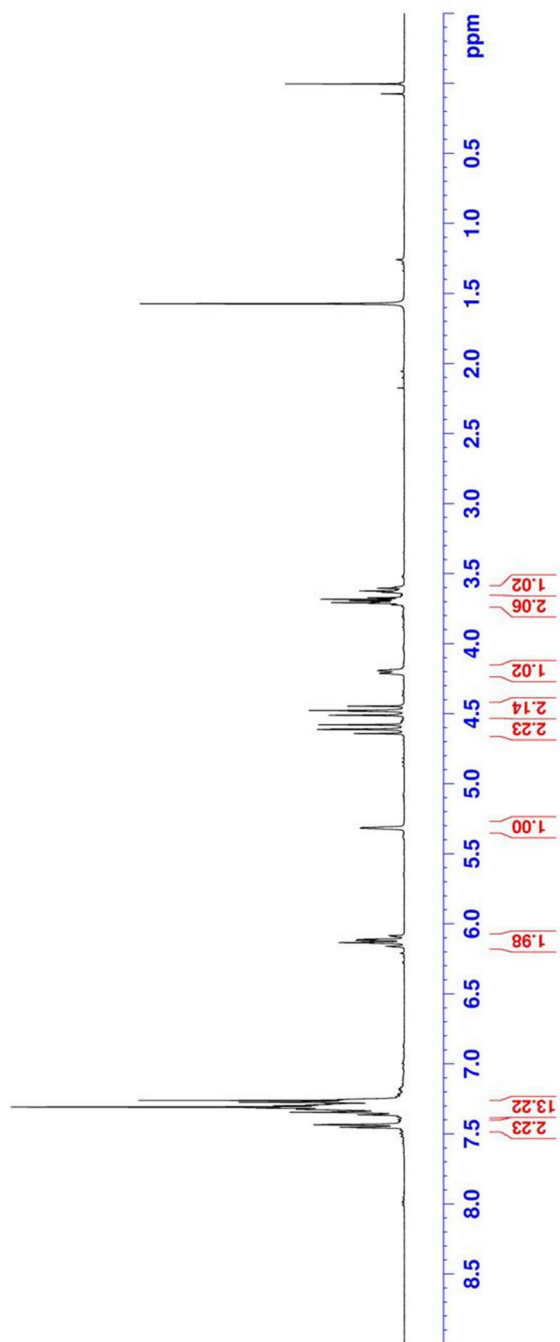


01-102-013, 1H, BBFO1 400 MHz, CDCl3

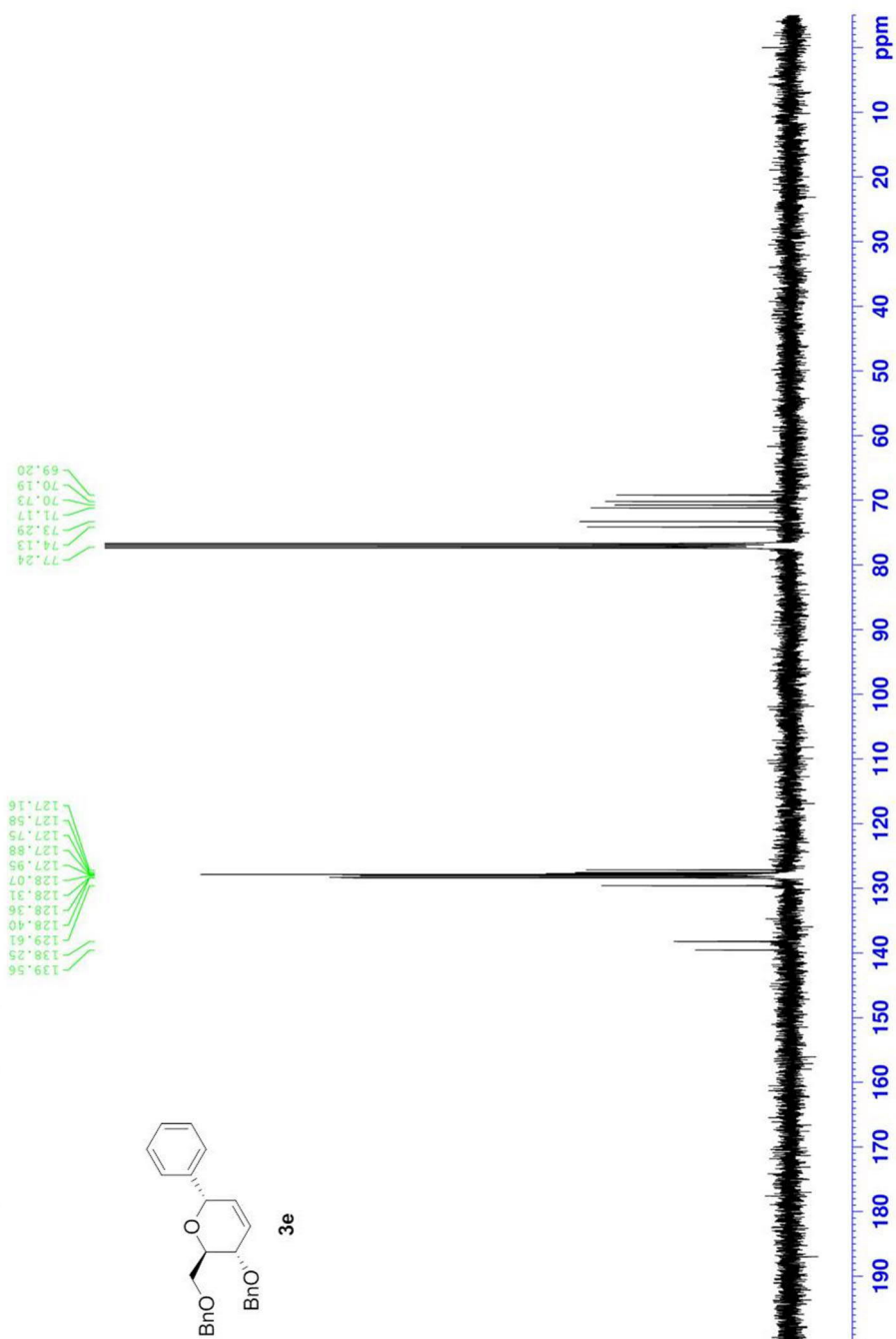
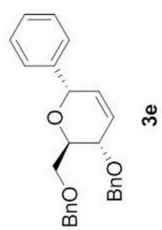
7.3410
7.3333
7.3267
7.3232
7.3201
7.3147
7.3077
7.2998
7.2984
7.2925
7.2882
7.2850
7.2733
7.2689
7.2537
6.1643
6.1600
6.1563
6.1382
6.1339
6.1301
6.1186
6.1156
6.1122
6.1086
6.0926
6.0896
6.0861
5.3169
5.3132
4.6410
4.6122
4.6092
4.5783
4.5105
4.4817
4.4762
4.4458
4.2123
4.2083
4.2033
4.1973
4.1932
4.1893
4.1843
3.7239
3.7193
3.7129
3.7084
3.7026
3.6946
3.6840
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3.6396
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3.6053
3.6003
3.5957

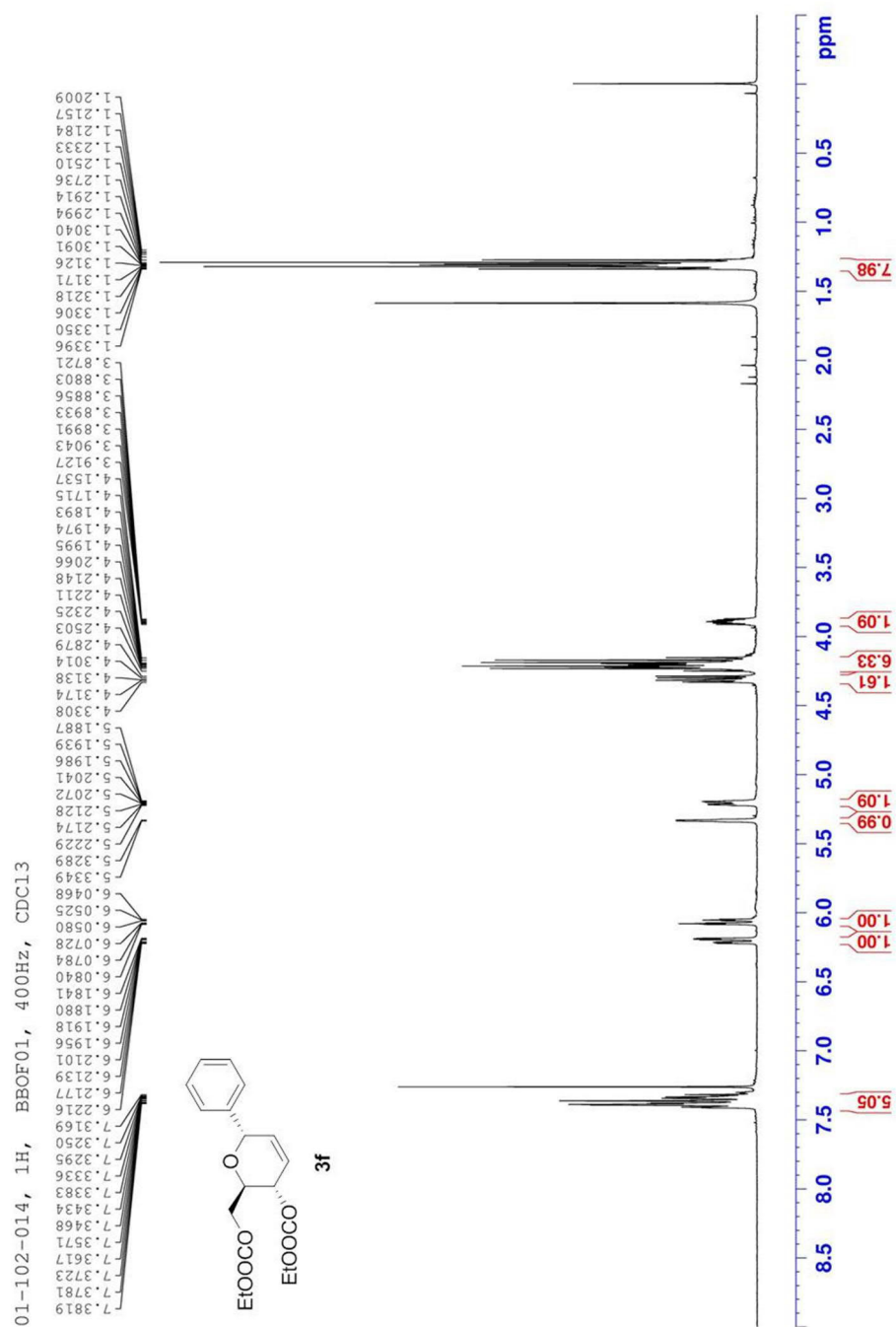


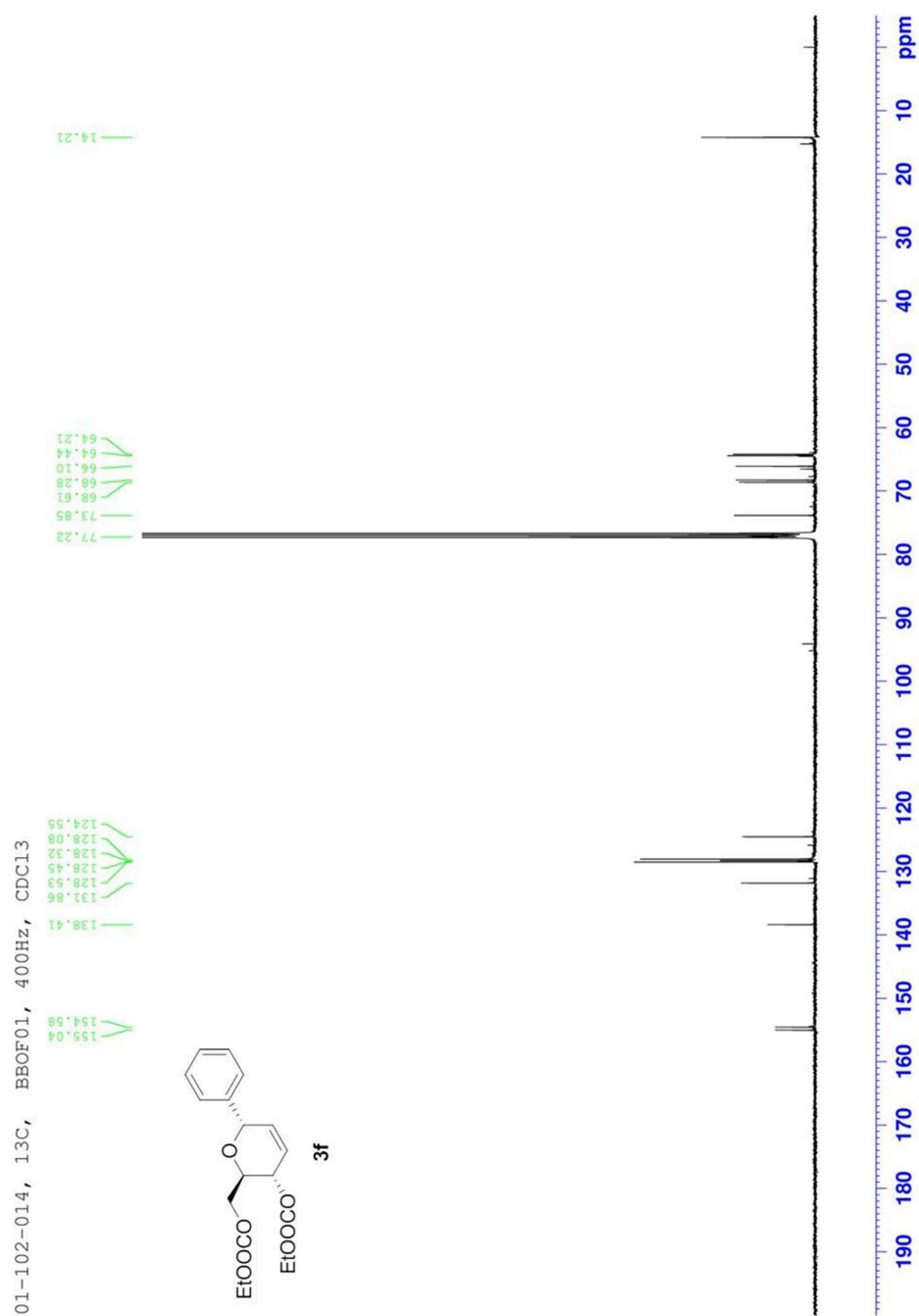
3e



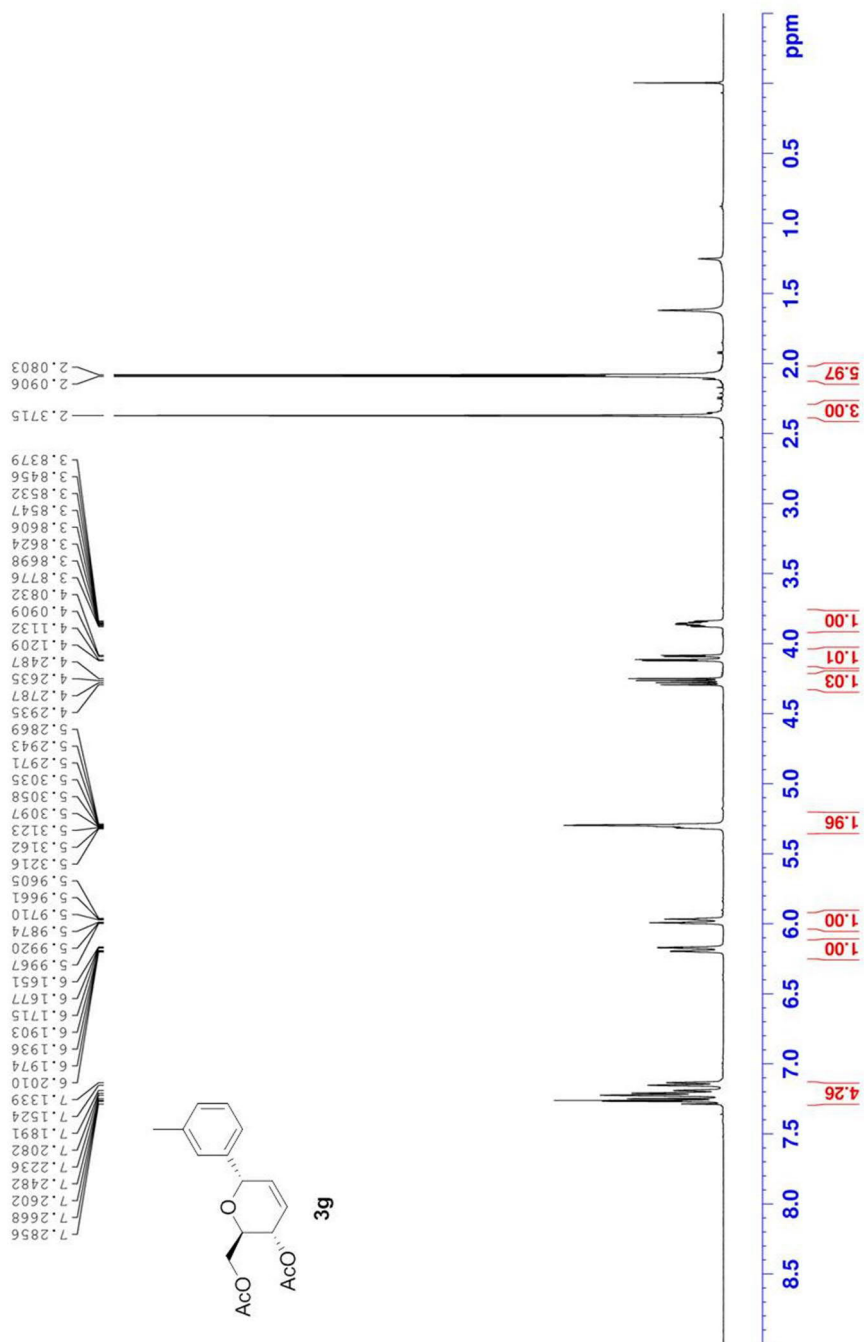
01-102-013, BBFO2 400, 13C, CDCl3



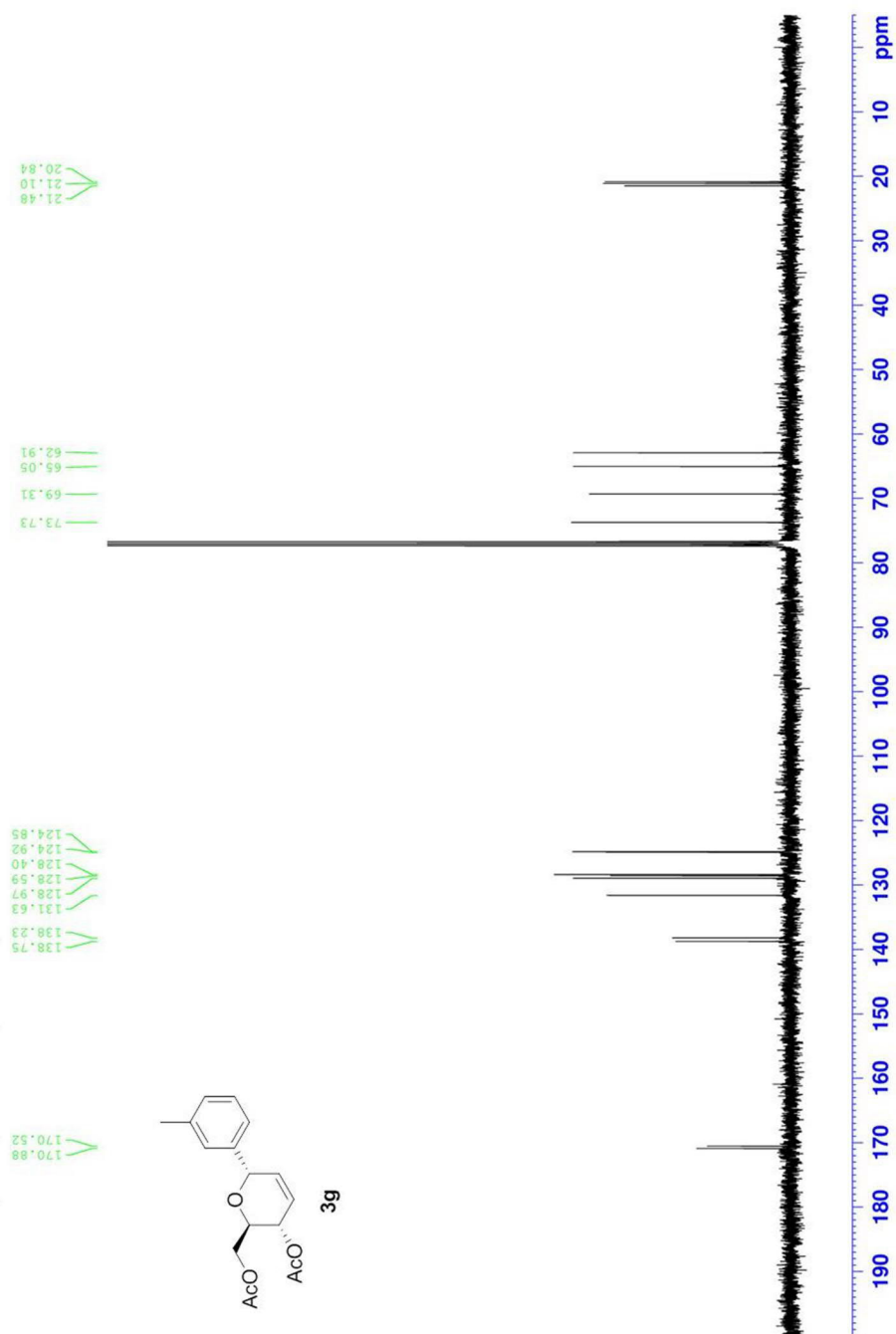




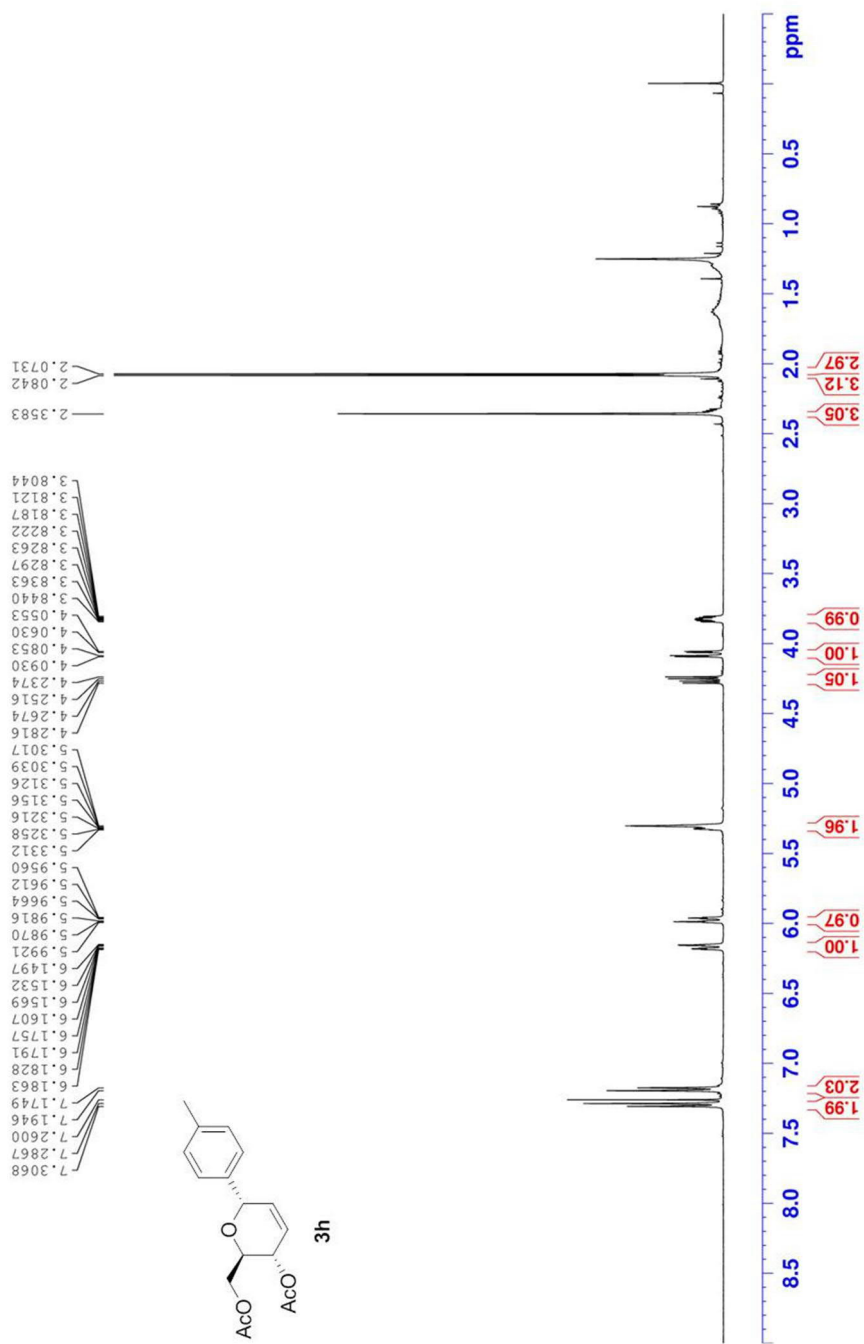
01-102-018, 1H, BBOF01, 400Hz, CDCl3



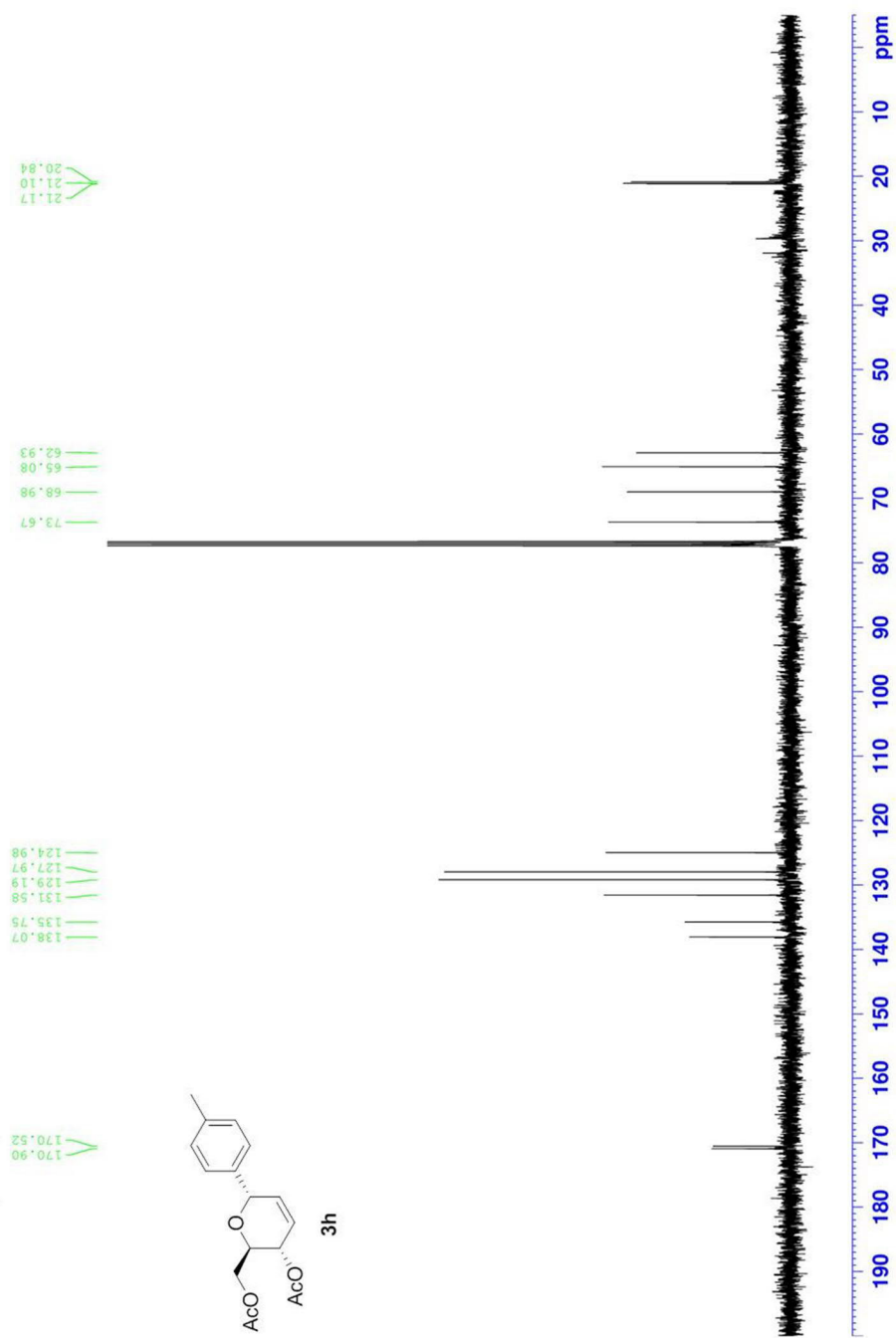
01-102-018, ¹³C, BBOF01, 400Hz, CDCl₃



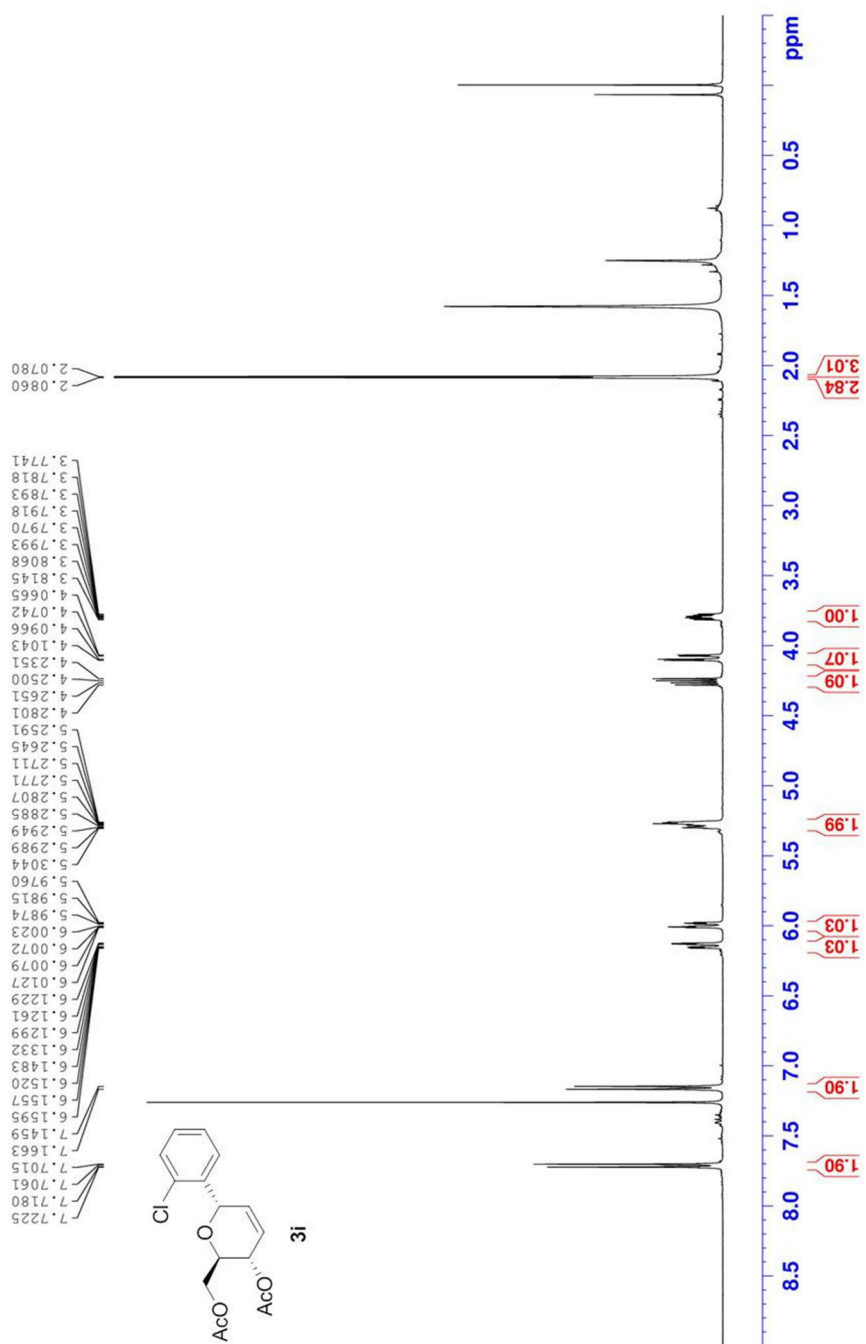
01-102-022, 1H, BEF01 CDCl3



01-102-022, ¹³C, BBO1 CDCl₃



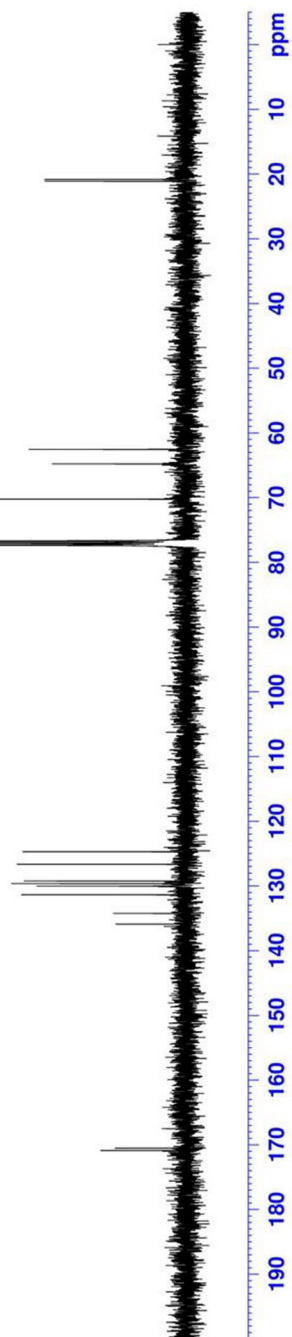
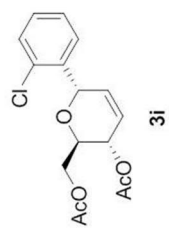
01-102-025, 1H, BBF01 CDCl3



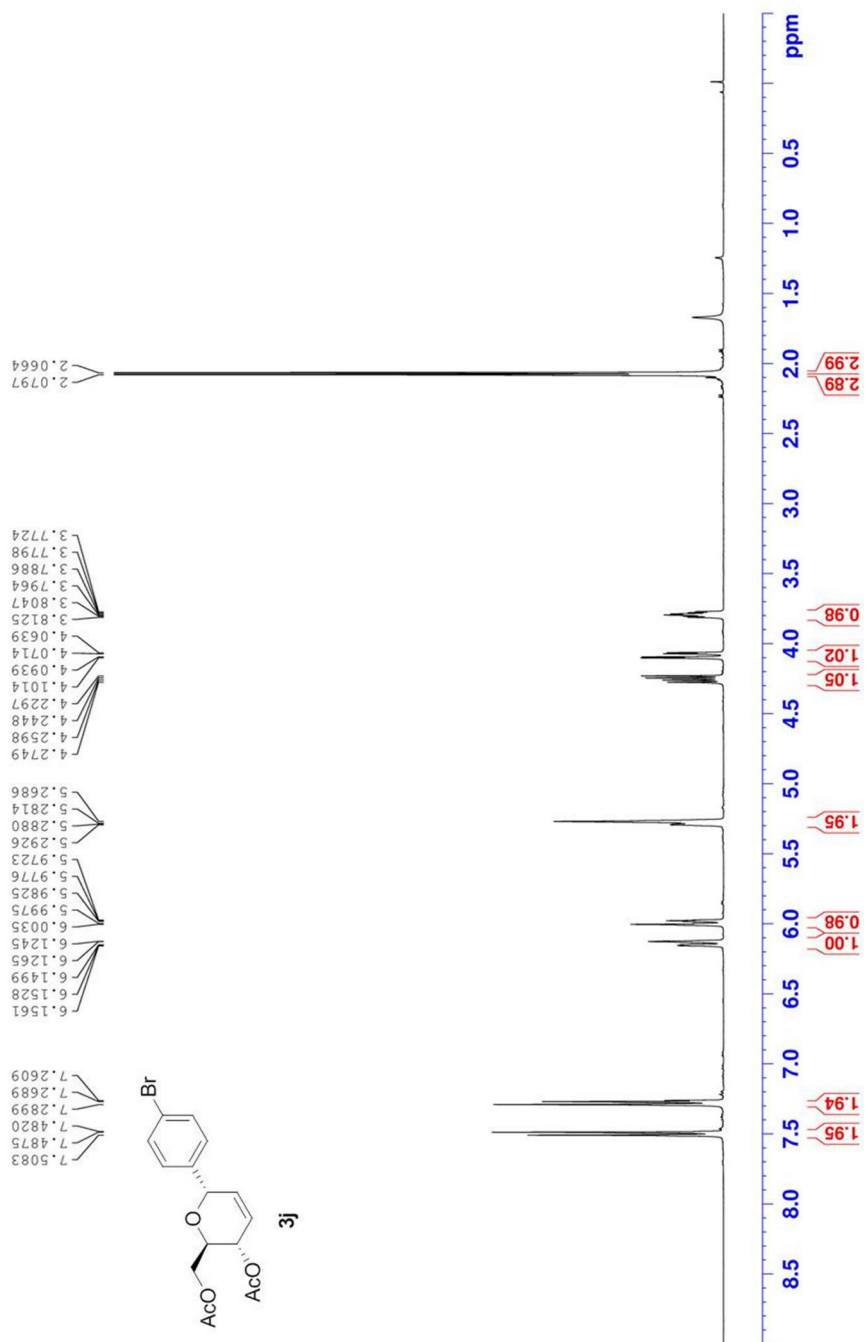
01-102-020, ¹³C, BBOF01, 400Hz, CDCl₃

170.86
170.50
135.91
134.24
131.37
130.03
129.65
129.25
126.66
124.72

77.23
70.23
64.78
62.54
21.11
20.83



01-102-028, AV 400 MHz, ¹H NMR, CDCl₃

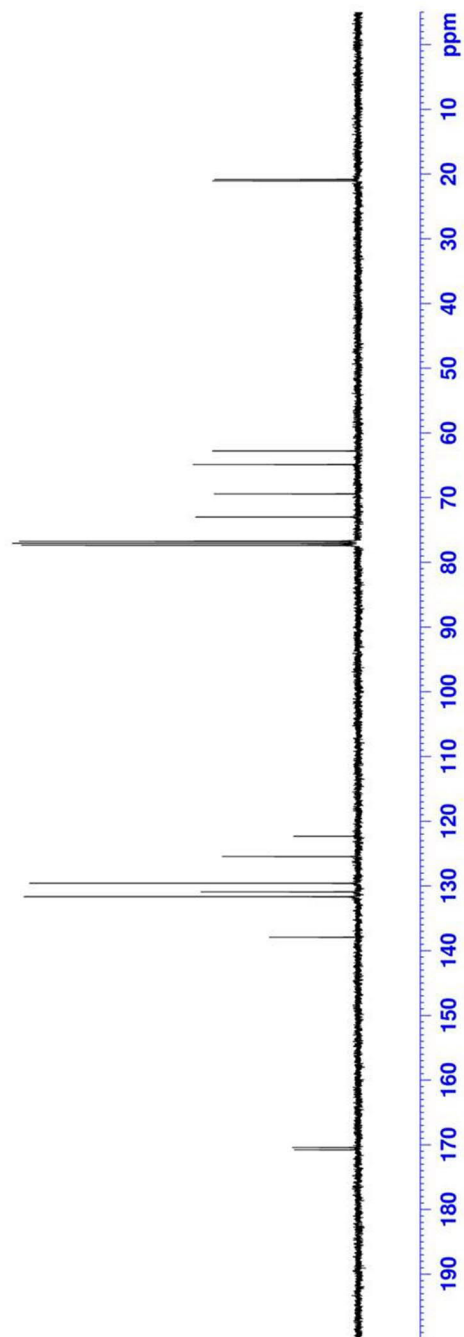
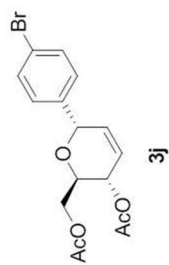


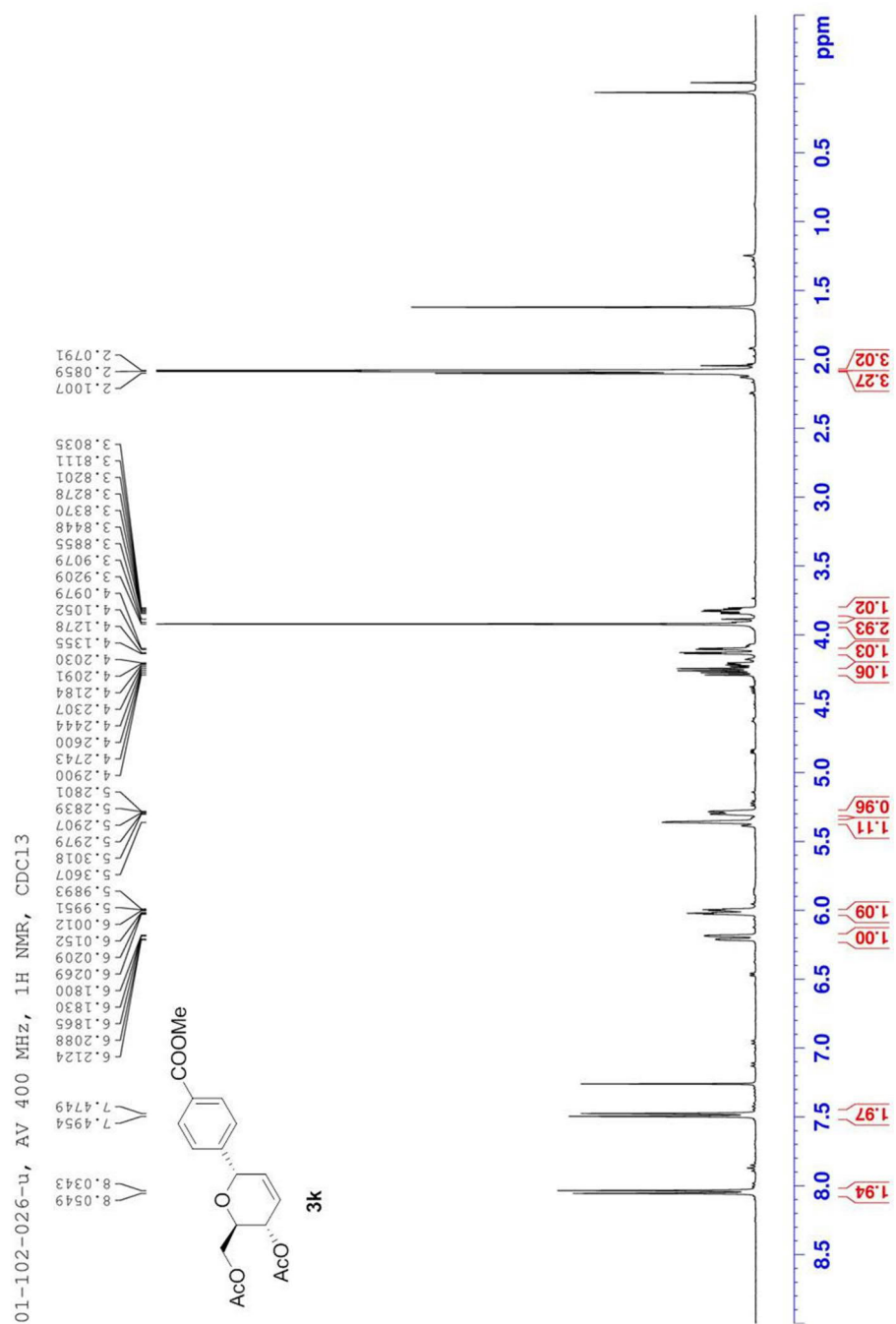
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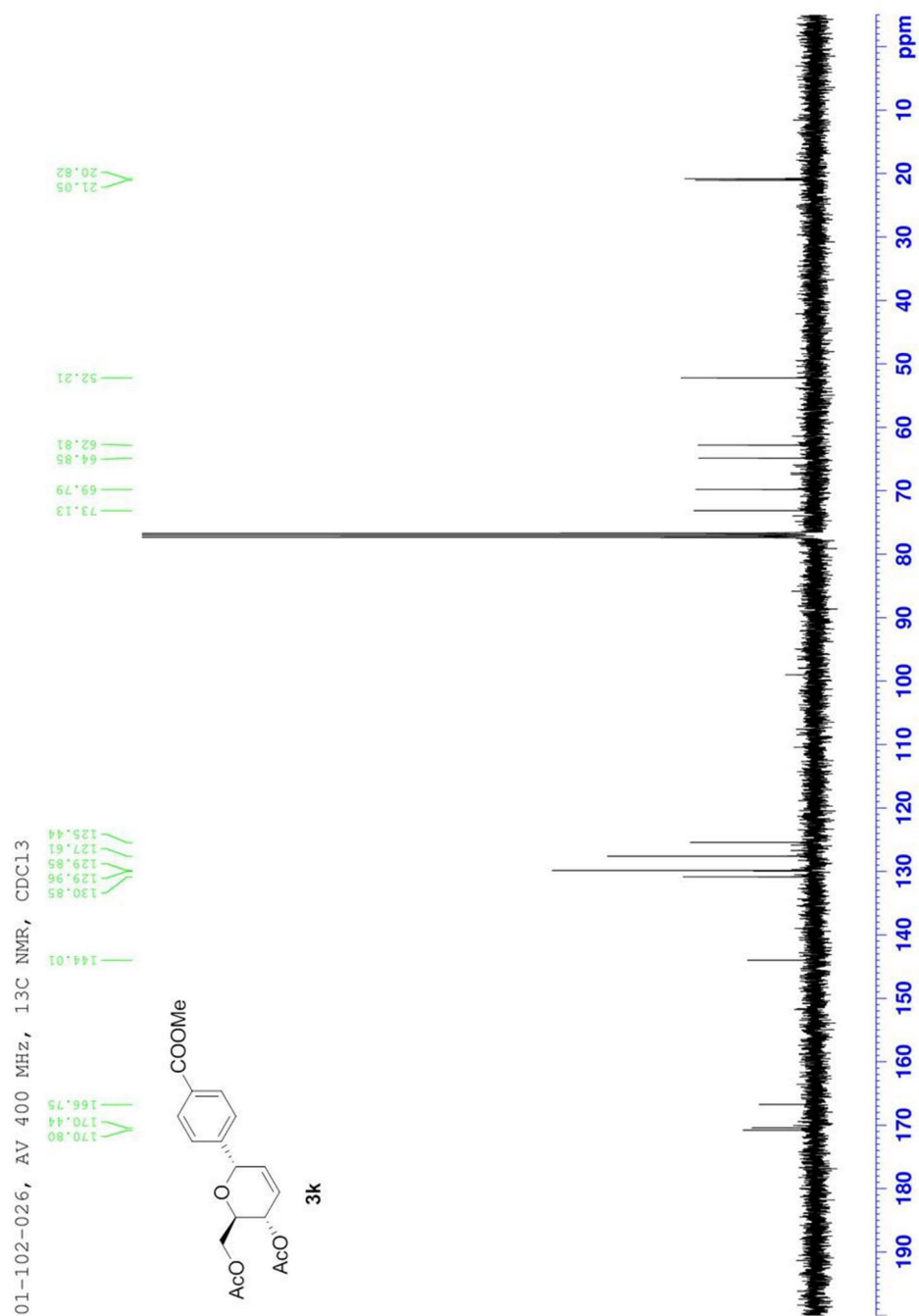
170.78
170.42
137.94
131.68
130.92
129.59
125.46
122.35

73.00
69.43
64.88
62.79

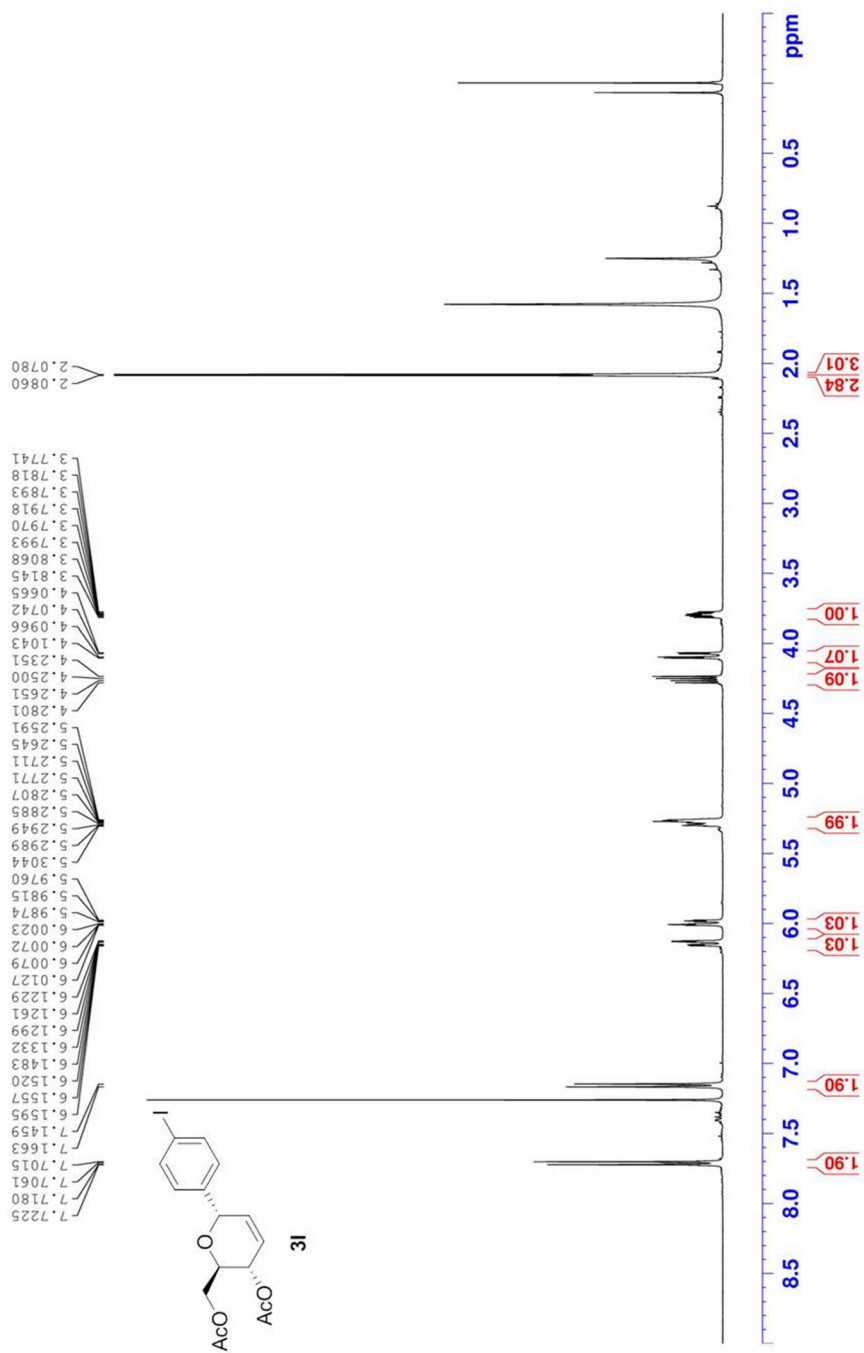
21.05
20.81



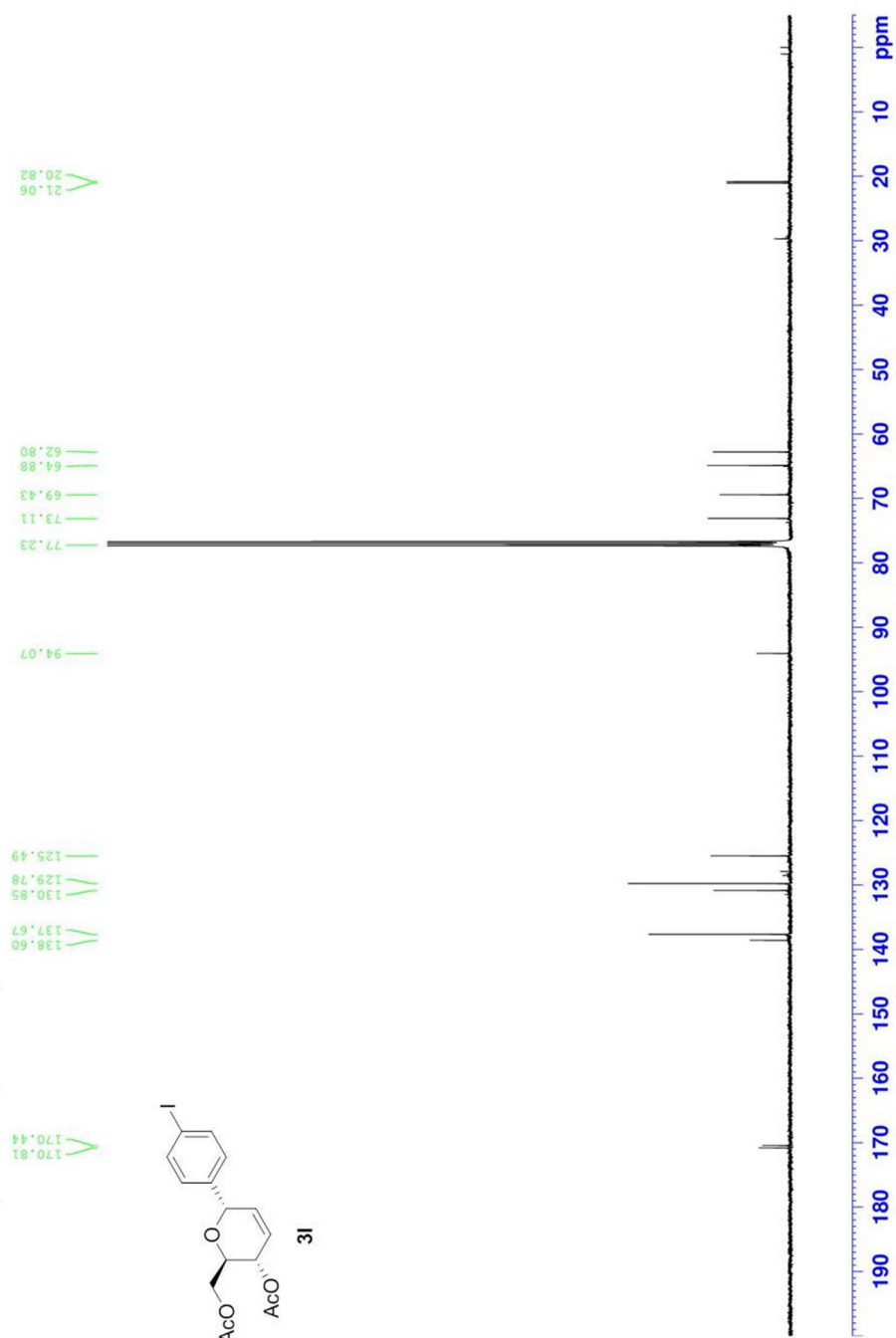


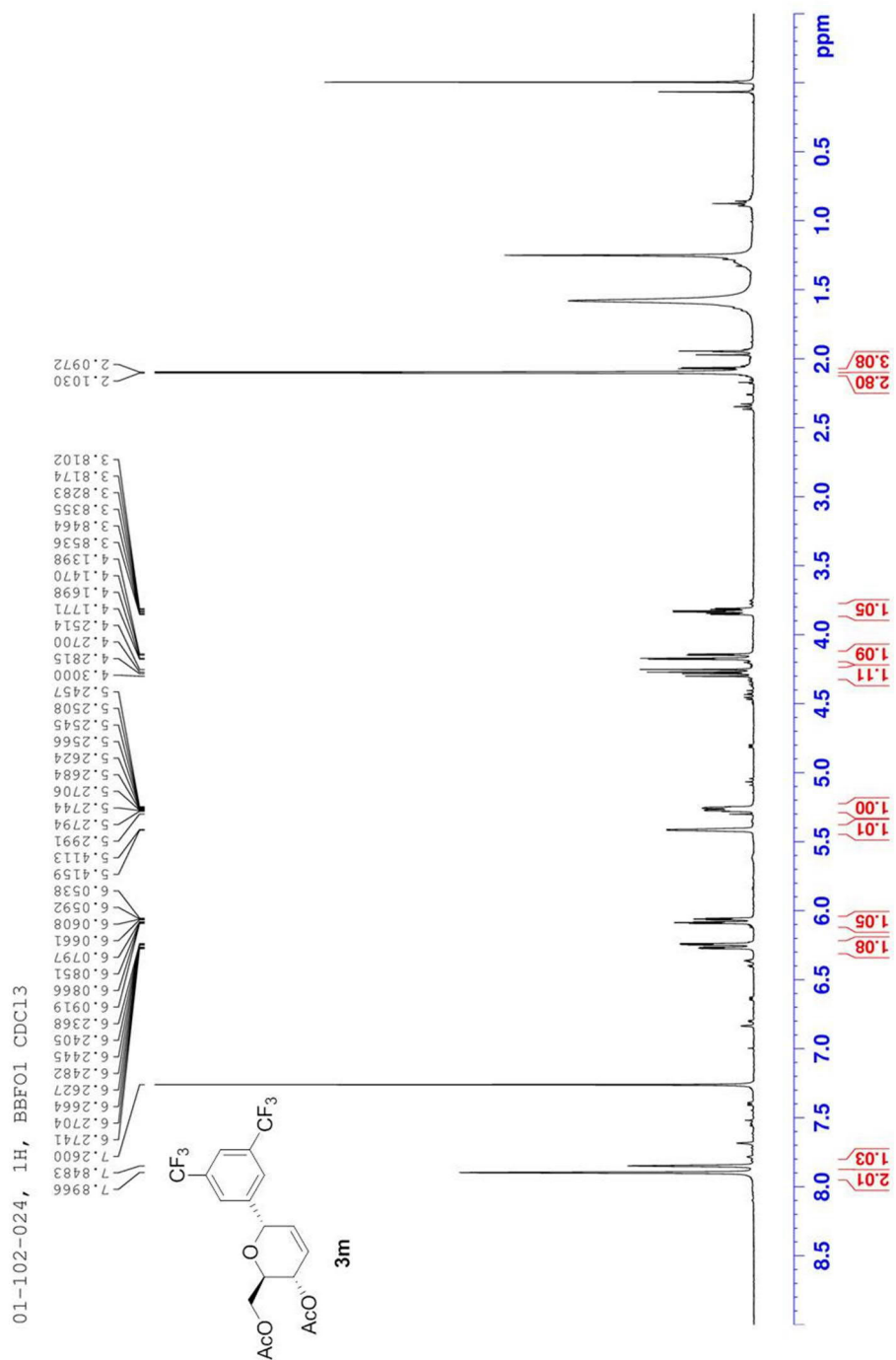


01-102-025, 1H, BBF01 CDCl3

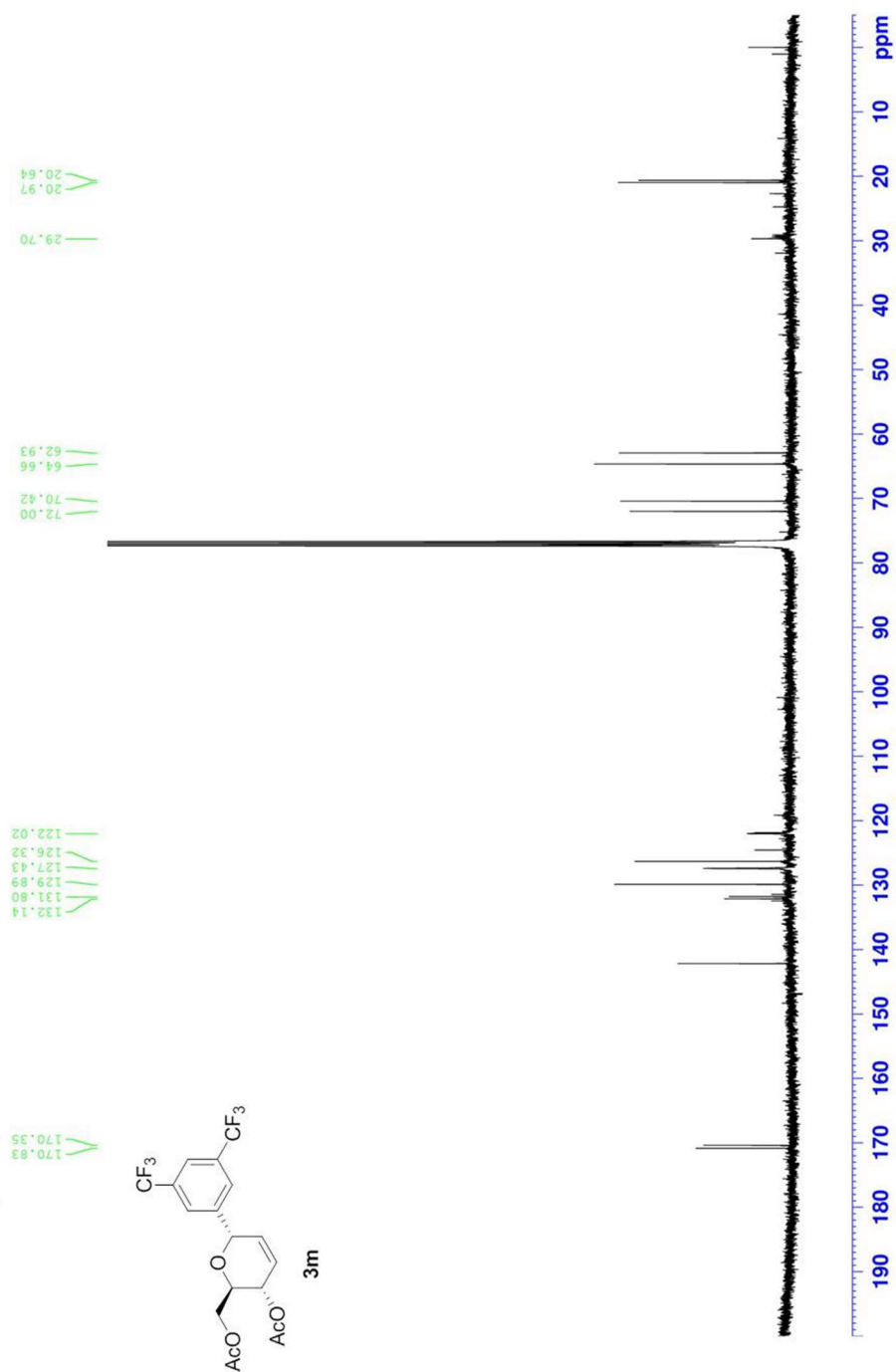


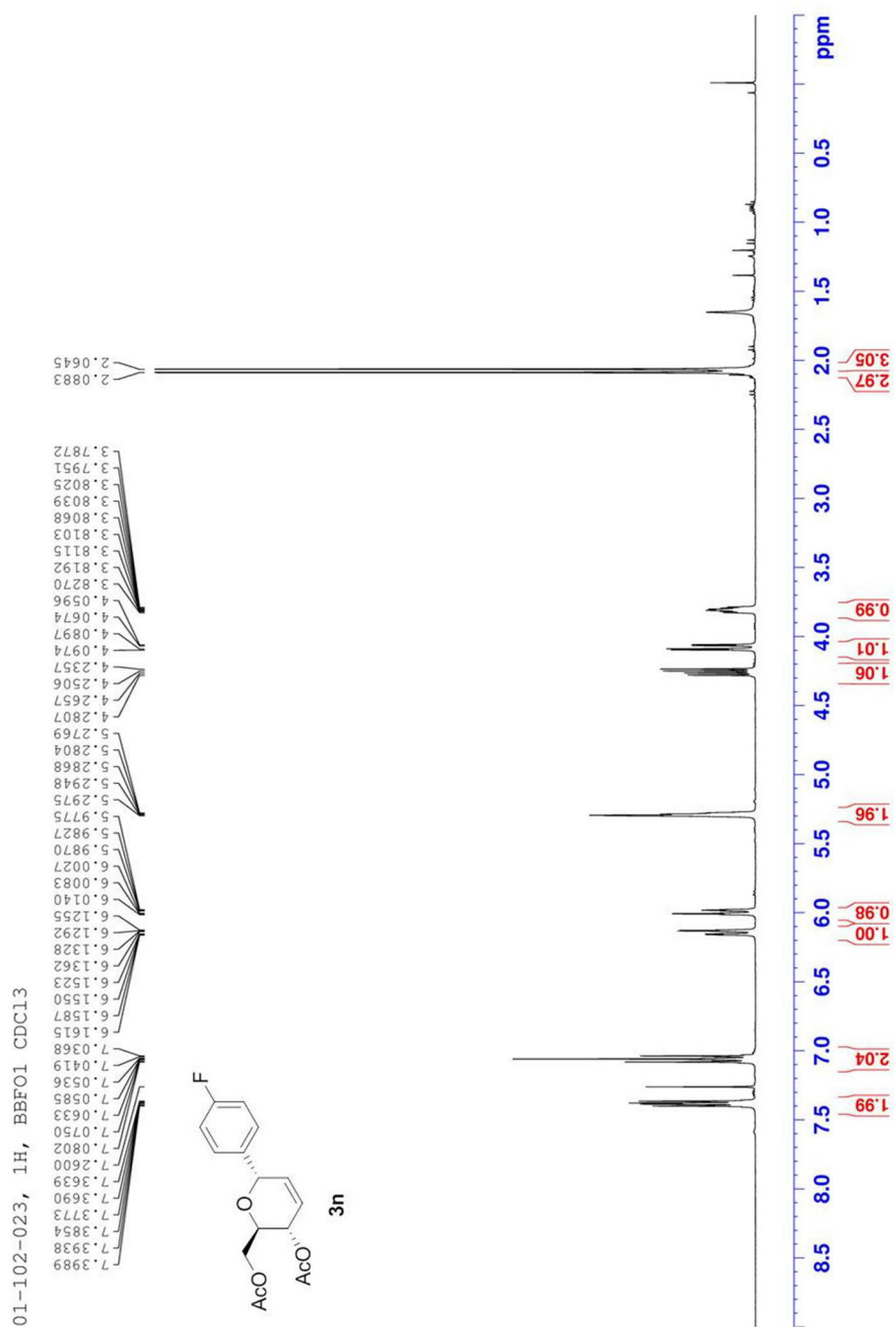
01-102-025; 400 MHz, ¹H NMR, CDCl₃



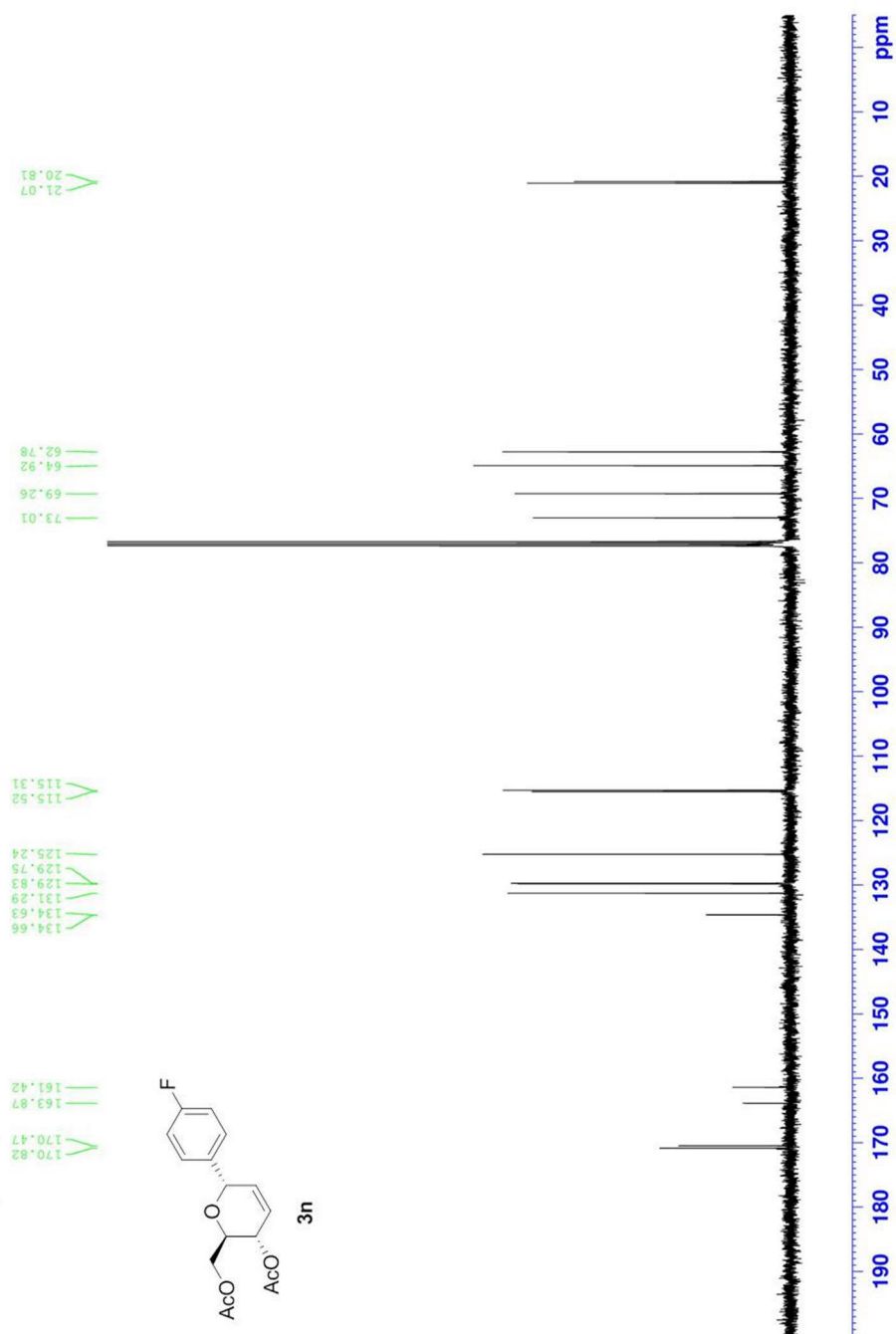


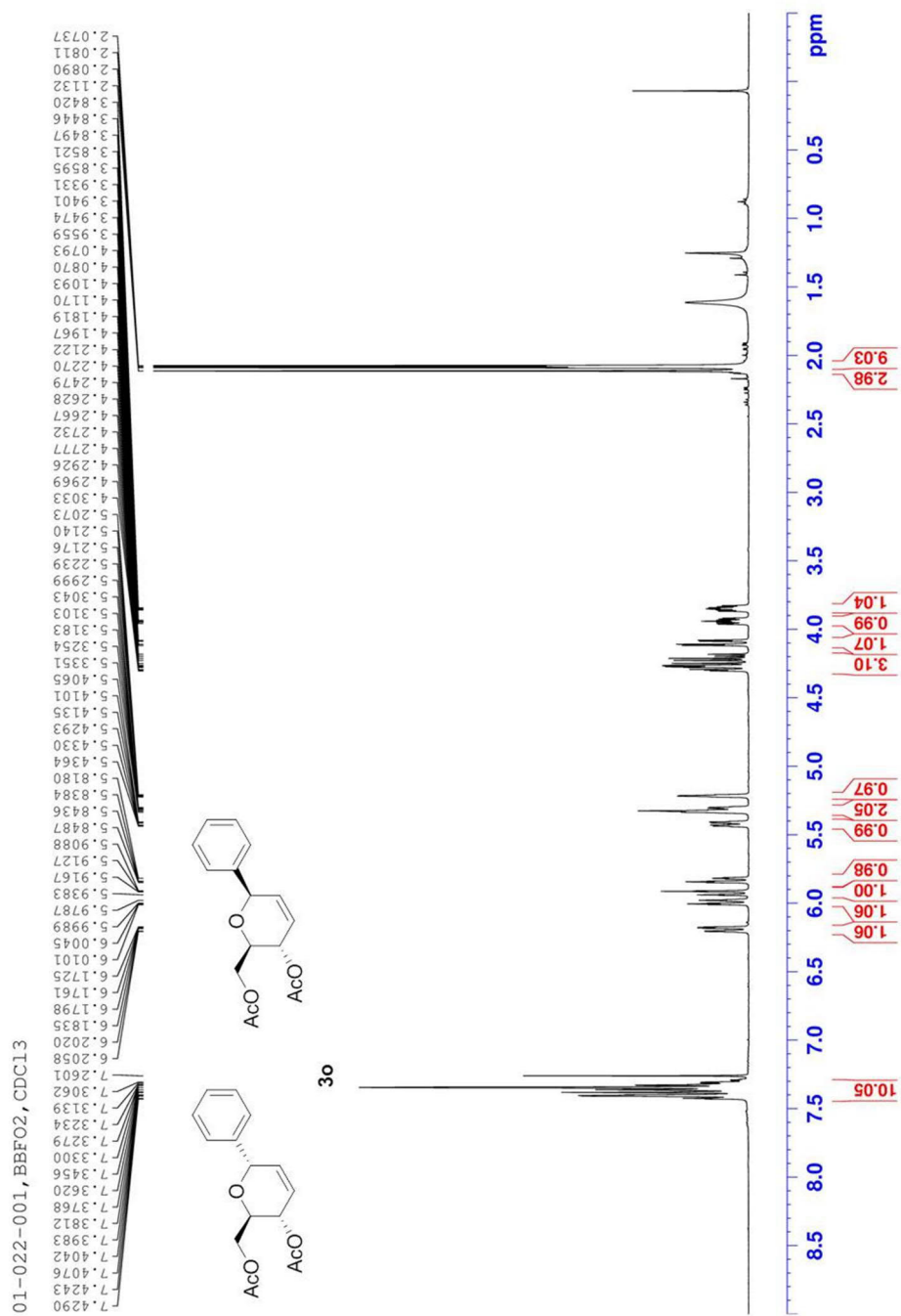
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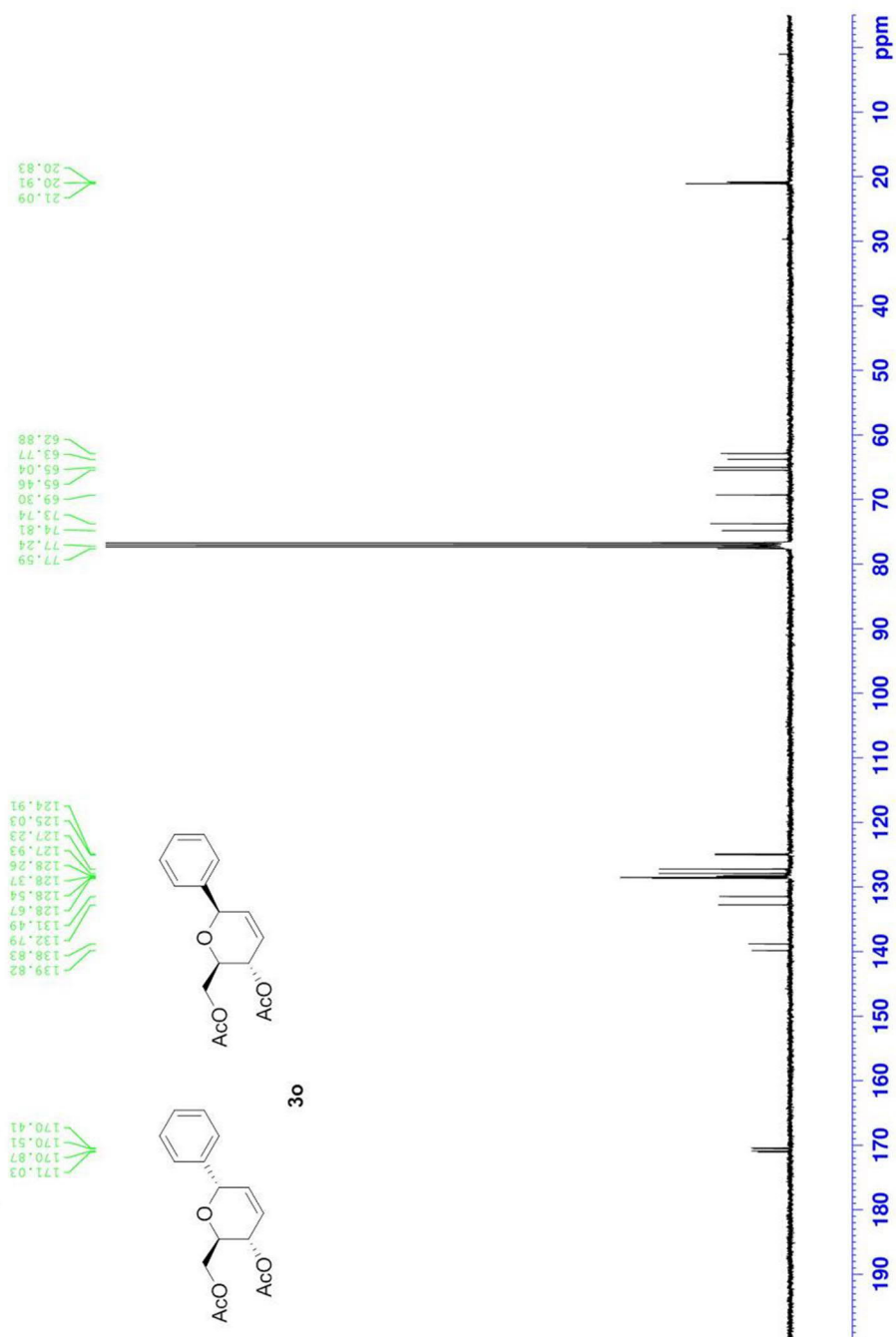


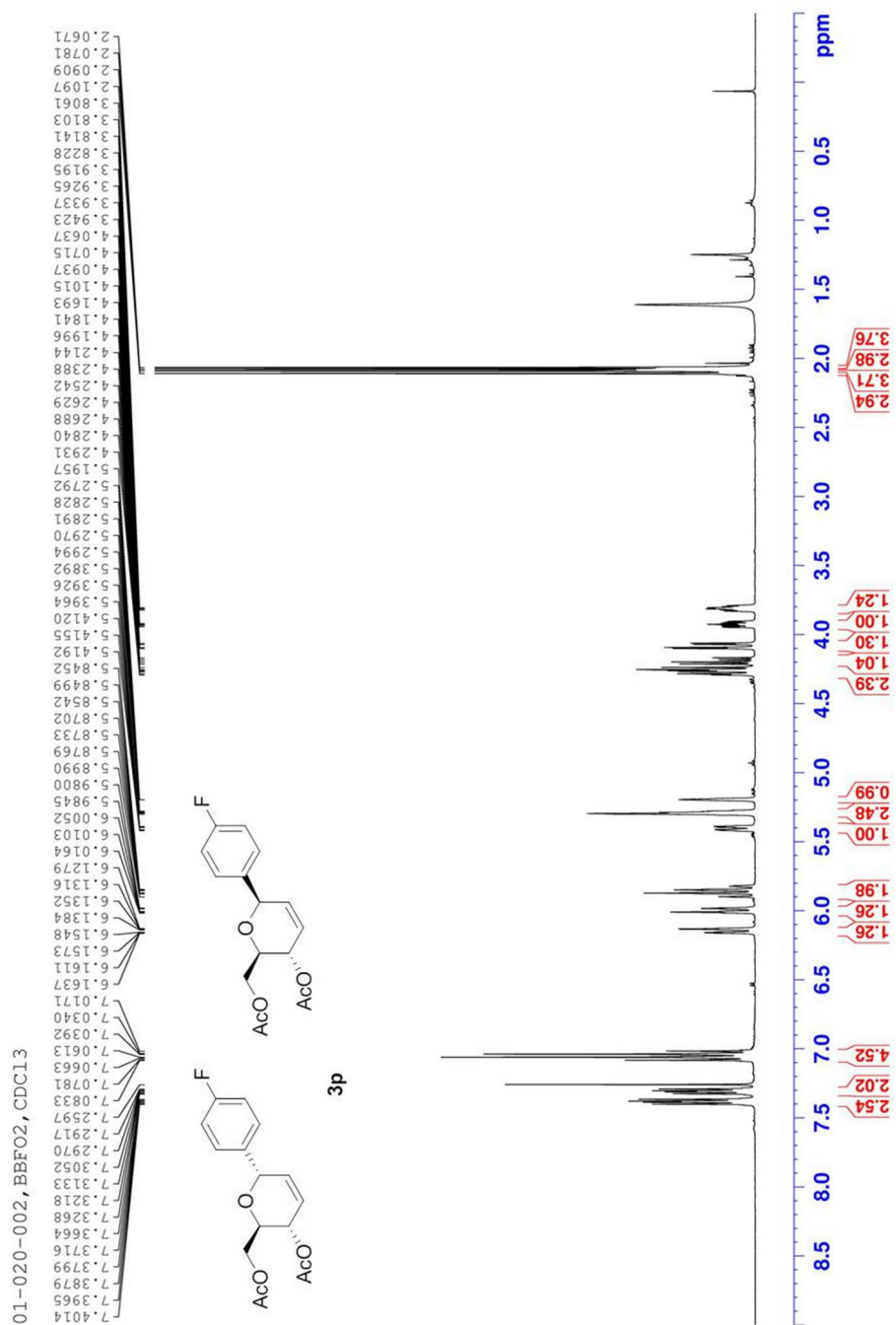
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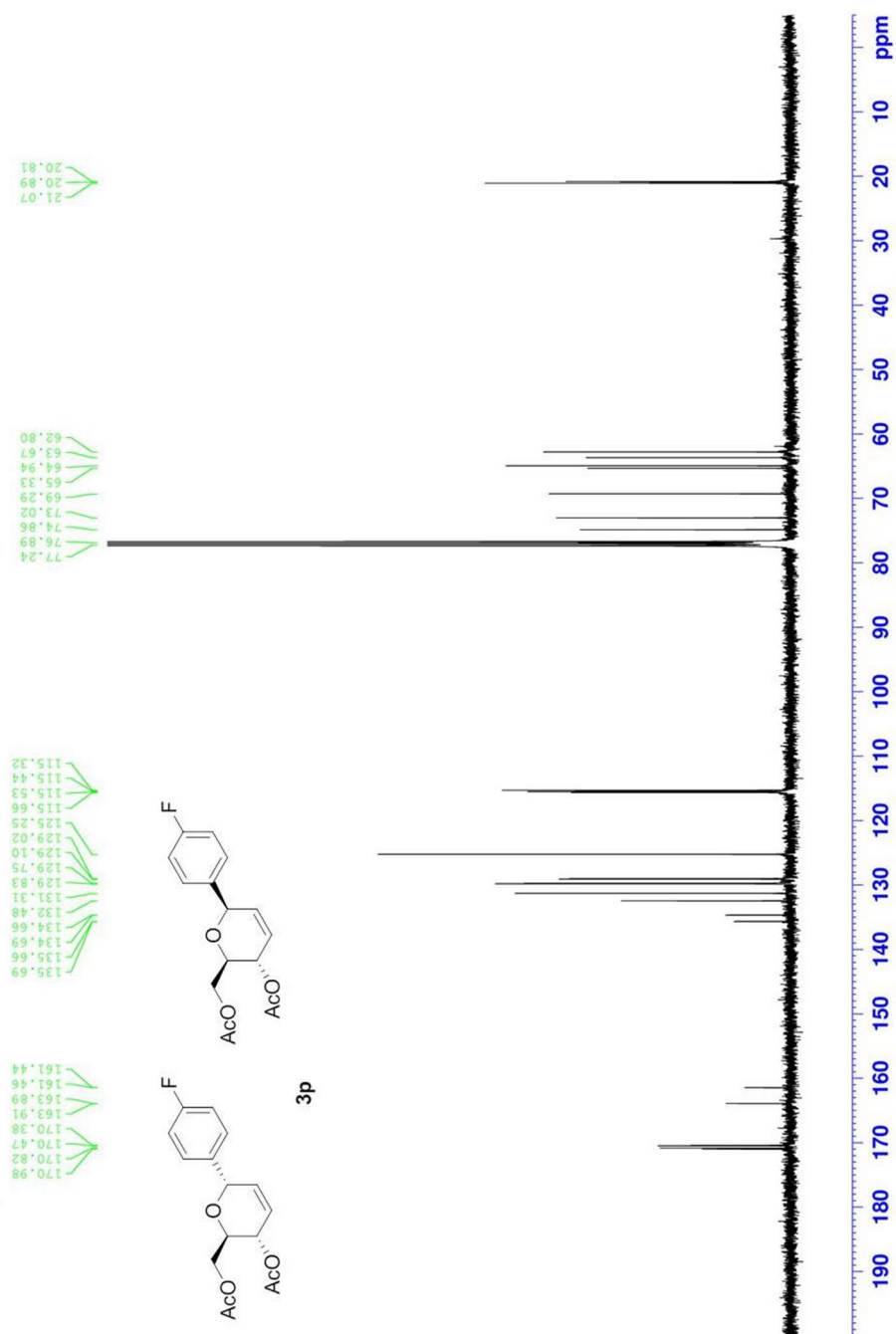


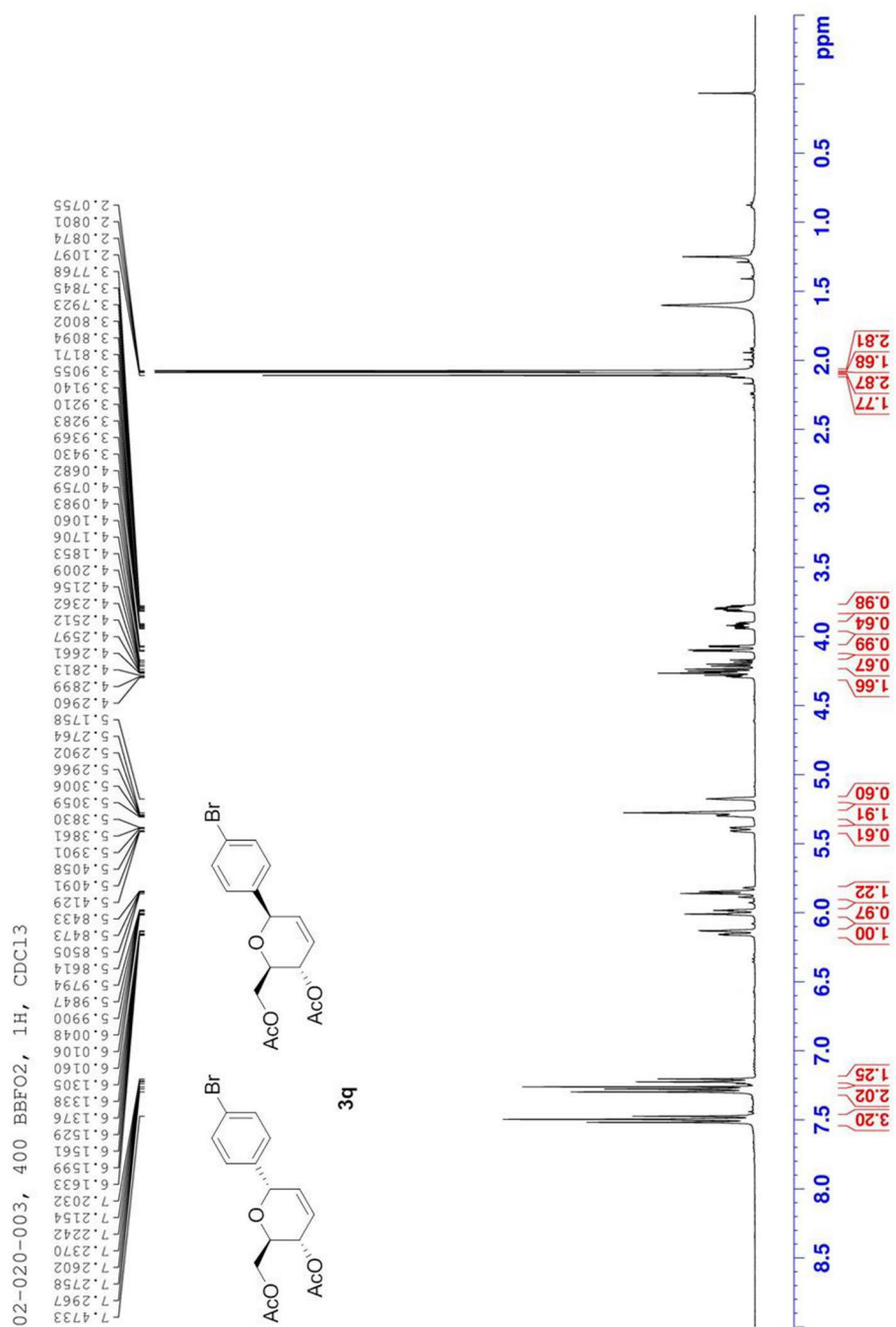
01-022-001, BBFO2, CDCl3



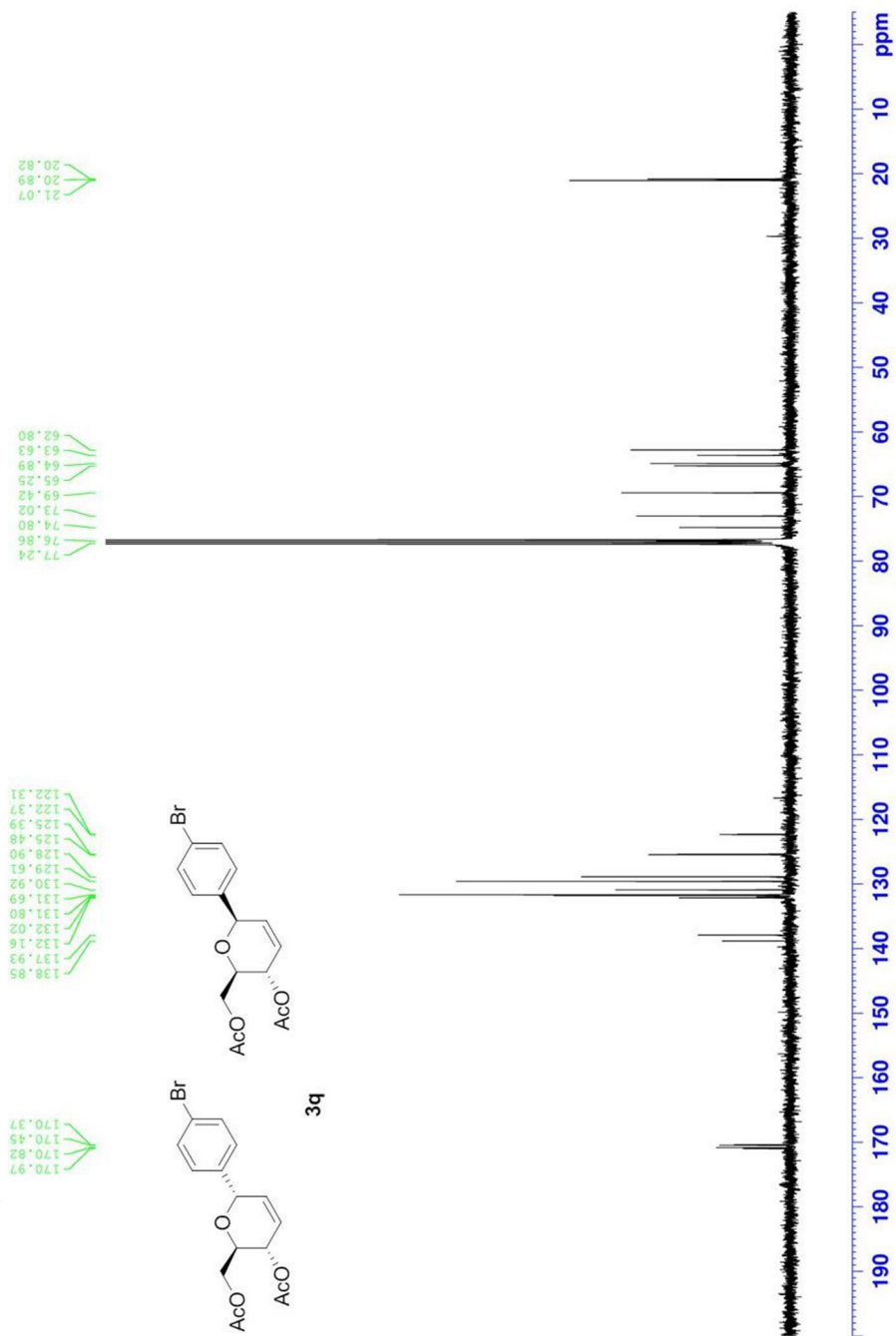


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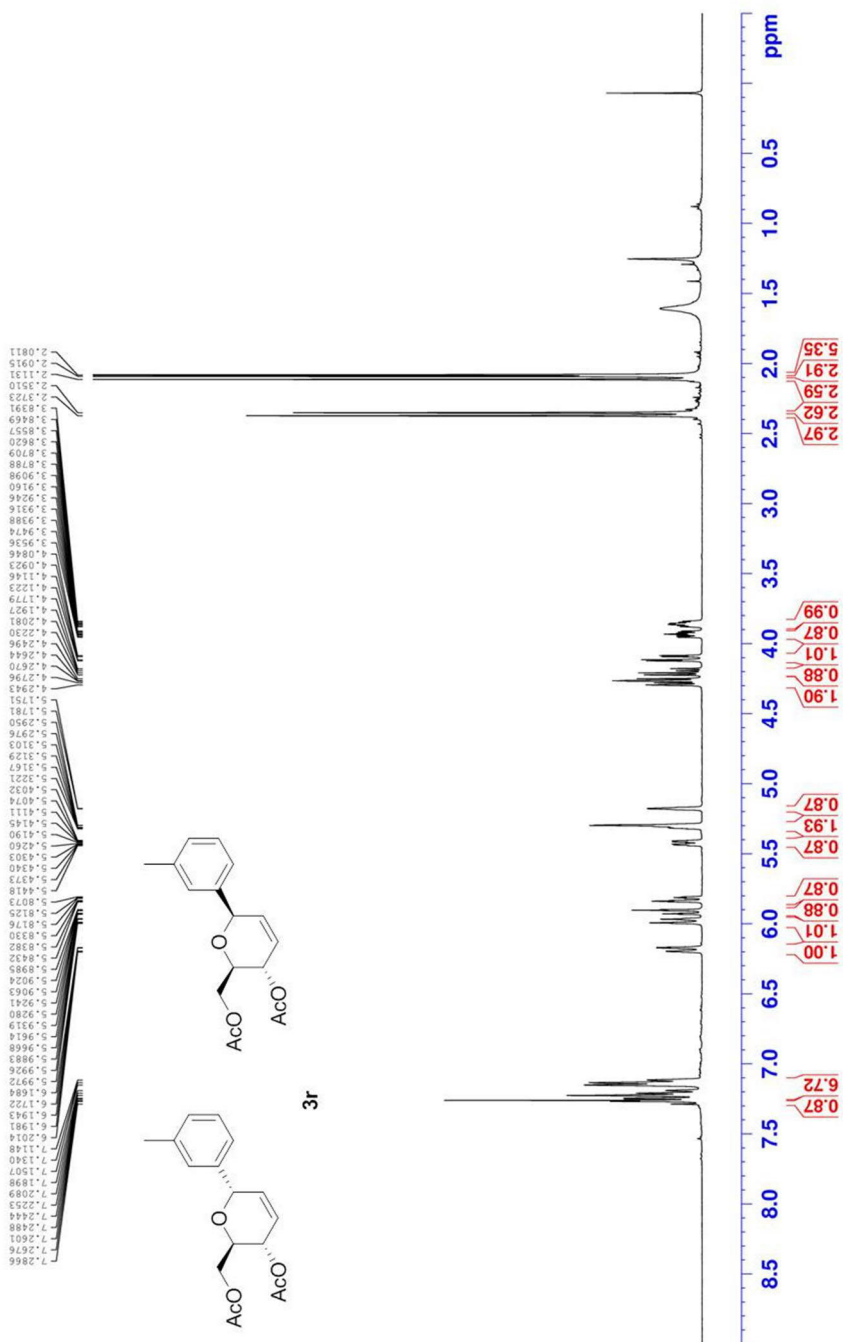




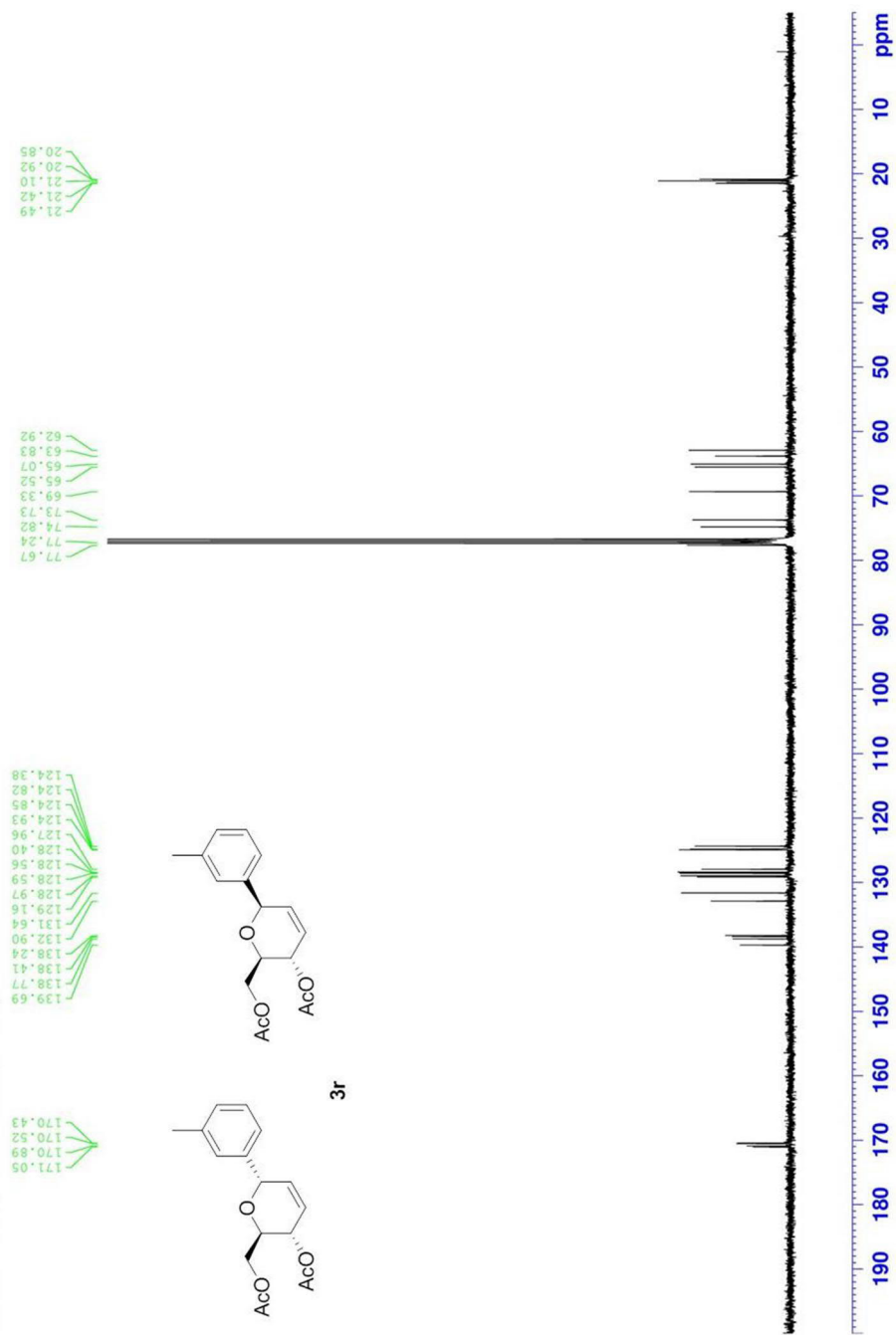
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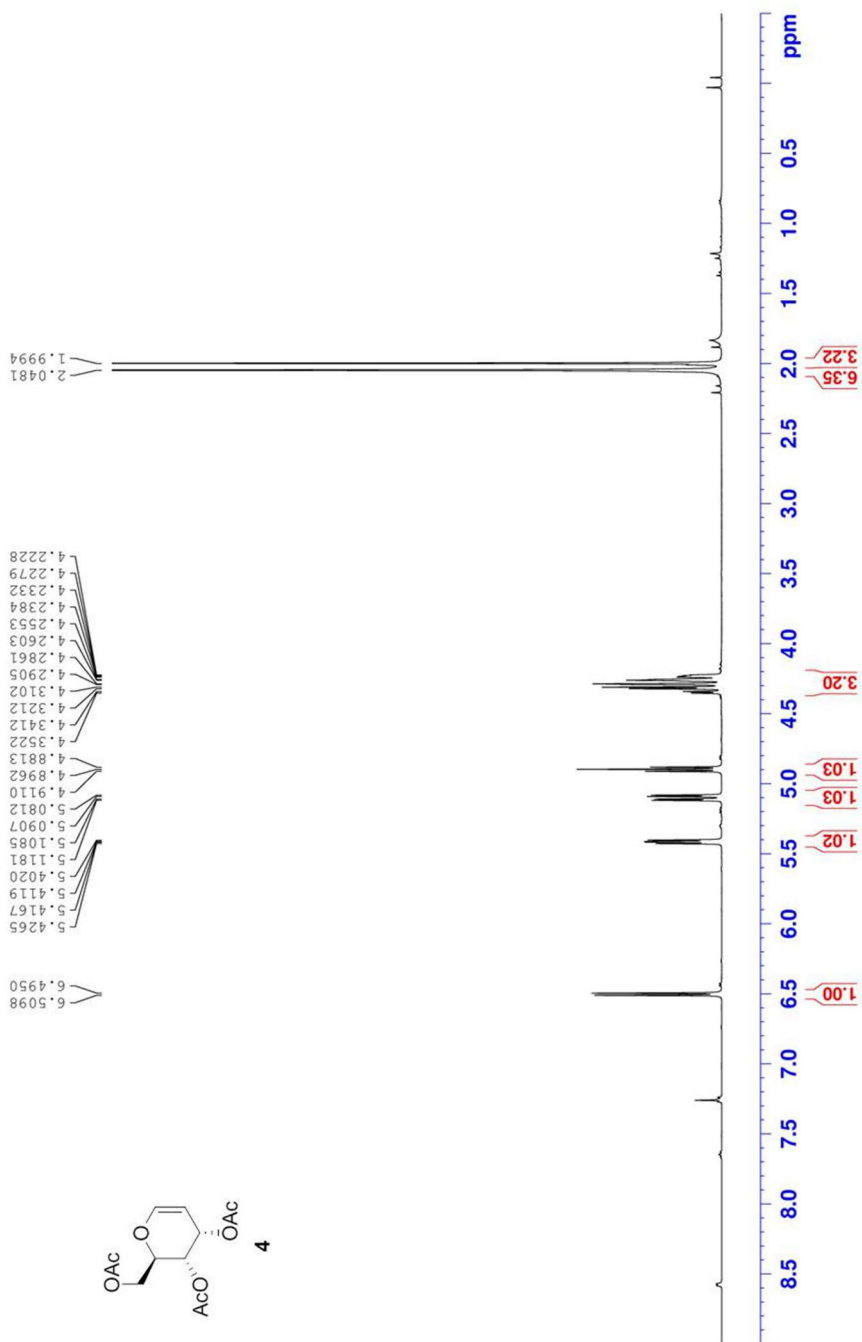
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02-020-004, 400 BBFO2, 13C, CDCl3



02-018-001, AV 400 MHz, ¹H NMR, CDCl₃



02-018-001, AV 400 MHz, ¹³C NMR, CDCl₃

