Synthesis, electrochemical and photophysical properties of calixarene based Ru(II) complexes as potential multivalent photoreagents

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Supporting Information

Syntheses of the intermediate compounds 5, 11, 12, 17, 18 and 23.

Figure S1. ¹H NMR spectrum (300 MHz, 298K) of compound 5 in CDCl₃.

Figure S2. ¹³C NMR spectrum (75 MHz, 298K) of compound 5 in CDCl₃.

Figure S3. COSY NMR spectrum (300MHz, 298K) of compound 5 in CDCl₃.

Figure S4. HSQC NMR spectrum (300MHz, 298K) of compound 5 in CDCl₃.

Figure S5. ¹H NMR spectrum (300 MHz, 298K) of compound 6 in CDCl₃.

Figure S6. ¹³C NMR spectrum (75 MHz, 298K) of compound 6 in CDCl₃.

Figure S7. COSY NMR spectrum (300MHz, 298K) of compound 6 in CDCl₃.

Figure S8. HSQC NMR spectrum (300MHz, 298K) of compound 6 in CDCl₃.

Figure S9. ¹H NMR spectrum (300 MHz, 298K) of compound 7 in CD₃OD.

Figure S10. ¹³C NMR spectrum (75 MHz, 298K) of compound 7 in CD₃OD.

Figure S11. COSY NMR spectrum (300MHz, 298K) of compound 7 in CD₃OD.

Figure S12. HSQC NMR spectrum (300MHz, 298K) of compound 7 in CD₃OD.

Figure S13. ¹H NMR spectrum (300 MHz, 298K) of compound 11 in CDCl₃.

Figure S14. ¹H NMR spectrum (600 MHz, 298K) of compound 12 in CDCl₃.

Figure S15. ¹³C NMR spectrum (75 MHz, 298K) of compound 12 in CDCl₃.

Figure S16. COSY NMR spectrum (300MHz, 298K) of compound 12 in CDCl₃.

Figure S17. HSQC NMR spectrum (300MHz, 298K) of compound 12 in CDCl₃.

Figure S18. HMBC NMR spectrum (600MHz, 298K) of compound 12 in CDCl₃.

Figure S19. ¹H NMR spectrum (300 MHz, 298K) of compound 13 in CDCl₃.

Figure S20. ¹³C NMR spectrum (75 MHz, 298K) of compound 13 in CDCl₃.

Figure S21. COSY NMR spectrum (300MHz, 298K) of compound 13 in CDCl₃.

Figure S22. HSQC NMR spectrum (300MHz, 298K) of compound 13 in CDCl₃.

Figure S23. HMBC NMR spectrum (400MHz, 298K) of compound 13 in CDCl₃.

Figure S24. ¹H NMR spectrum (300 MHz, 298K) of compound 14 in CDCl₃.

Figure S25. ¹³C NMR spectrum (100 MHz, 298K) of compound 14 in CDCl₃.

Figure S26. COSY NMR spectrum (400MHz, 298K) of compound 14 in CDCl₃.

Figure S27. HSQC NMR spectrum (400MHz, 298K) of compound 14 in CDCl₃.

Figure S28. HMBC NMR spectrum (400MHz, 298K) of compound 14 in CDCl₃.

Figure S29. ¹H NMR spectrum (300 MHz, 298K) of compound 17 in CDCl₃.

Figure S30. ¹³C NMR spectrum (75 MHz, 298K) of compound 17 in CDCl₃.

Figure S31. COSY NMR spectrum (300MHz, 298K) of compound 17 in CDCl₃.

Figure S32. HSQC NMR spectrum (300MHz, 298K) of compound 17 in CDCl₃.

Figure S33. ¹H NMR spectrum (400 MHz, 298K) of compound 18 in CDCl₃.

Figure S34. ¹³C NMR spectrum (75 MHz, 298K) of compound 18 in CDCl₃.

Figure S35. COSY NMR spectrum (400MHz, 298K) of compound 18 in CDCl₃.

Figure S36. HSQC NMR spectrum (400MHz, 298K) of compound 18 in CDCl₃.

Figure S37. HMBC NMR spectrum (400MHz, 298K) of compound 18 in CDCl₃.

Figure S38. ¹H NMR spectrum (400 MHz, 328K) of compound 19 in CDCl₃.

Figure S39. ¹³C NMR spectrum (100 MHz, 298K) of compound 19 in CDCl₃.

Figure S40. COSY NMR spectrum (400MHz, 298K) of compound 19 in CDCl₃.

Figure S41. HSQC NMR spectrum (400MHz, 298K) of compound 19 in CDCl₃.

Figure S42. HMBC NMR spectrum (400MHz, 298K) of compound 19 in CDCl₃.

Figure S43. ¹H NMR spectrum (300 MHz, 298K) of compound 23 in CDCl₃.

Figure S44. ¹³C NMR spectrum (75 MHz, 298K) of compound 23 in CDCl₃.

Figure S45. COSY NMR spectrum (300MHz, 298K) of compound 23 in CDCl₃.

Figure S46. HSQC NMR spectrum (300MHz, 298K) of compound 23 in CDCl₃.

Figure S47. HMBC NMR spectrum (400MHz, 298K) of compound 23 in CDCl₃.

Figure S48. ¹H NMR spectrum (400 MHz, 298K) of compound 24 in CD₃OD.

Figure S49. ¹³C NMR spectrum (75 MHz, 298K) of compound 24 in CD₃OD.

Figure S50. COSY NMR spectrum (300MHz, 298K) of compound 24 in CD₃OD.

Figure S51. HSQC NMR spectrum (300MHz, 298K) of compound 24 in CD₃OD.

Figure S52. HMBC NMR spectrum (400MHz, 298K) of compound 24 in CD₃OD.

Figure S53. Cyclic voltammetry: Reduction waves of [Ru(TAP)₂pytz'(diN₃C6)]²⁺(NO₃⁻)₂ **24**.

Figure S54. Absorption spectra (298K, air) in MeCN for complexes 6 (black), 7 (red), 13 (blue), 14 (green), 19 (orange) and 24 (pink).

Figure S55. Stern-Volmer experiment for $[Ru(TAP)_2phen]^{2+}$ 25 in acetonitrile, in presence of phenol.

Figure S56. Stern-Volmer experiment for $[Ru(TAP)_2phen]^{2+}$ **25** in acetonitrile, in presence of an acid (paratoluene sulfonic acid = PTSA).

Figure S57. Deuteration of the phenol moiety was checked by ¹H NMR (300 MHz, 298K) in CDCl₃.

Figure S58. Stern-Volmer experiment for $[Ru(TAP)_2phen]^{2+}$ **25** in acetonitrile, in presence of deuterated phenol.

Figure S59. Equation for ΔG_{PCET}^0 in the case of a PCET: (1.2) ET followed by HT or (1.3) ET and HT concerted.

Instrumentation. The ¹H NMR, ¹³C NMR and 2D NMR spectra were recorded with a Bruker Avance-300 instrument, a Varian-400 VNMRJ System and a Bruker Varian Unity-600. The chemical shifts are expressed in ppm and determined in comparison of the deutered solvent used as internal reference (CDCl₃ and CD₃OD). CDCl₃ was filtered over a short basic alumina column to remove traces of DCl. Most of the ¹H NMR spectra signals were attributed through 2D NMR analyses (COSY, HSQC, HMBC). The electrospray mass spectra were recorded with a Q-TOF 6520 Agilent Technology spectrometer (at the Organic Pharmaceutic Chemistry Lab). ATR-FTIR spectra were recorded, at room temperature, on a Bruker IFS55 FTIR spectrophotometer equipped with a liquid nitrogen-cooled mercurycadmium-telluride (MCT) detector at a nominal resolution of 2 cm⁻¹ and encoded every 1 cm⁻¹. The emission spectra were recorded with a Shimadzu RF-5301PC and the absorption spectra with a Perkin-Elmer Lambda UV-Vis spectrophotometer. The determination of the molar absorption coefficients were performed by weight and absorption measurements. Cyclic voltammetry was carried out on a platinum disk working electrode (approximate area = 3mm^2), in dried acetonitrile with tetrabutylammonium hexafluorophosphate (0.1 mol.L⁻¹) as supporting electrolyte. The potential of the working electrode was controlled by an Autolab PGSTAT 100 (Eco Chemie B.V., Utrecht, The Netherlands) potentiostat through a PC interface with a scan rate of 100 mV s⁻¹ between -2 and +2 V versus SCE. The counter electrode was a platinum disk and the reference electrode a Saturated Calomel Electrode (SCE). All measurements were performed in a single compartment cell. The emission lifetimes were measured by using the single-photon counting technique (SPC) with an Edinburgh Instruments FL900 spectrometer (Edinburgh, U.K.) equipped with a nitrogenfilled discharge lamp and a peltier-cooled Hamamatsu R955s photomultiplier tube. The emission

decays were analyzed with the Edinburgh Instruments software (version 3.0), based on nonlinear least-squares regressions using Marquardt algorithms.

The transients induced under pulsed illumination were measured by using as excitation a source composed of a frequency-tripled (355 nm) Nd:YAG Q-switched laser (Continuum Inc.) coupled with an optical parametric oscillator (Continuum Inc.) covering the wavelengths region 410-2300 nm with a maximum pulse energy from 10 to 120 mJ depending on the wavelength. The emission was detected perpendicularly by a photomultiplier (R928, Hamamatsu). The signal was recorded with a digital oscilloscope (HP 54200A), connected through the IEEE488 interface to a personal computer, and was averaged over at least 16 shots. The emission wavelength was selected via a grating Czerny-Turner monochromator (Spectra Pro 2300i, Acton Research Corp.). For the transient absorption, a cross-beam configuration was adapted by using as a probe source a 150 W xenon arc lamp producing a continuous spectral distribution ranging from 190 to 2600 nm. Complete transient absorption spectra were measured in the 200-650 nm spectral range with a 2 ns gated intensified CCD camera (PIMAX, 1024 x 256 pixels, Princeton Instruments). The time delays for probing, following the laser excitation pulse, were controlled by a programmable time generator (Princeton Instruments). For kinetic analyses, the transient absorption traces at selected individual wavelengths were recorded by the photomultiplier as emission lifetime measurement. Kinetic analyses of the absorption traces were performed by nonlinear leastsquares regression modified by Levenberg-Marquardt algorithms.¹

Synthesis of calix[4]monoamidephenanthroline 5. Calix[4]monoacide 4 (0.296 g, 0.419 mmol, 1 equiv.), DCC (0.131 g, 0.636 mmol, 1.5 equiv.) and HOBt (0.087 g, 0.644 mmol, 1.5 equiv.) were dissolved in DMF (7 mL). At 0°C, DIPEA (450 μ L, 2.59 mmol, 6.2 equiv.) was added to the mixture which was stirred for 30 min at 0°C under inert atmosphere. Then, 5-

glycinamido-1,10-phenanthroline 22 (0.296 g, 0.809 mmol, 2 equiv.) was added to the mixture which was stirred for 16h at room temperature under inert atmosphere. The reaction mixture was concentrated under reduced pressure and dissolved in CH₂Cl₂ (60 mL). The organic layer was washed with H₂O (3×35 mL) and the combined aqueous layers were extracted with CH₂Cl₂ (2×45 mL). The combined organic layers were concentrated under reduced pressure. Ether (3 mL) was added and DCU was filtered. The filtrate was concentrated under reduced pressure and the crude residue was purified by flash chromatography (CH₂Cl₂/MeOH 9:1) to yield the compound 5 as a clear yellow solid (0.352 g, 0.329 mmol, 78%). Rf = 0.42 (CH₂Cl₂/MeOH 9:1). m.p. 182-184°C (dec.); IR: v: 3319, 2966, 1683, 1485, 1298 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 298K): $\delta_{(ppm)} = 1.17$ (s, 9H, tBu), 1.22 (s, 18H, tBu), 1.23 (s, 9H, tBu), 3.46 (d, ²J = 14.1 Hz, 2H, ArCH_{2ea}), 3.51 (d, ²J = 13.5 Hz, 2H, ArCH_{2ea}), 4.21 (d, ²J = 13.8 Hz, 2H, ArCH_{2ax}), 4.25 (d, ²J = 13.5 Hz, 2H, ArCH_{2ax}), 4.52 (d, ${}^{3}J = 5.4$ Hz, 2H, NCH₂), 4.77 (s, 2H, OCH₂), 7.02 (d, ${}^{4}J = 2.4$ Hz, 2H, ArH), 7.07-7.09 (m, 6H, ArH), 7.56 (dd, 1H, ³J = 4.5 Hz, ³J = 8.4 Hz, H₃), 7.66 (dd, ³J = 4.5 Hz, ${}^{3}J = 8.1$ Hz, 1H, H₈), 8.25 (dd, ${}^{4}J = 1.2$ Hz, ${}^{3}J = 8.3$ Hz, 1H, H₇), 8.43 (s, 1H, H₆), 8.62 (d, ${}^{3}J = 8.7$ Hz, 1H, H₄), 9.05 (d, ${}^{3}J = 4.2$ Hz, 1H, H₂), 9.14 (d, ${}^{4}J = 1.2$ Hz, ${}^{3}J = 4.5$ Hz, 1H, H₉), 9.49 (s_b, 2H, OH), 10.06 (s_b, 1H, OH), 10.06-10.22 (s_b, 2H, NH_{phen} + NH_{amide}); ¹³C NMR (75 MHz, CDCl₃, 298K): $\delta_{(ppm)} = 31.2, 31.5(7), 31.6(2), 32.4, 33.1, 34.1, 34.2, 34.5, 45.5, 75.3, 34.5, 45.5, 75.3, 34.5, 45.5, 75.3, 35.5, 75.3, 7$ 118.1, 123.6, 123.9, 126.1, 126.4, 127.2, 127.4, 127.8, 128.2, 128.8, 131.4, 131.6, 132.9 (2C), 137.5, 144.3, 144.4, 145.6, 146.9, 147.0, 148.0, 148.9, 149.0, 149.7, 149.9, 170.0, 168.7, 170.6; HRMS (ESI-TOF) calcd for $C_{60}H_{68}N_4O_6 (M+H)^+$ 941.5217, found 941.5196.

Synthesis of calix[6]monoacid 11. Calix[6]monoamide **10** (0.628 g, 0.586 mmol, 1 equiv.) was dissolved in dioxane (30 mL). An aqueous HCl solution (6 M, 3 mL) was added dropwise and the mixture was stirred for 4 days at 56°C under inert atmosphere. The reaction mixture was

concentrated under reduced pressure and the residue was dissolved in CH_2Cl_2 (100 mL). The organic layer was washed with H_2O (4×50 mL) until pH = 5-6 and concentrated under reduced pressure to yield the compound **11** as a white solid (0.629 g, 0.586 mmol, quant.).

m.p. 171-172°C (dec.); IR: v: 2956, 1684, 1484 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 298K): $\delta_{(ppm)} = 0.98$ (s, 18H, *t*Bu), 1.06 (s, 18H, *t*Bu), 1.17 (s, 9H, *t*Bu), 1.38 (s, 9H, *t*Bu), 2.87 (s, 2H, OCH₂), 3.77-3.84 (m, 7H, OMe + ArCH₂), 3.84-3.94 (m, 10H, OMe + ArCH₂), 3.95-4.07 (m, 4H, ArCH₂), 6.69 (d, 2H, ⁴J = 2.2 Hz, ArH), 6.75 (d, 2H, ⁴J = 2.3 Hz, ArH), 6.96-6.98 (m, 4H, ArH), 7.01 (s, 2H, ArH), 7.24-7.28 (m, 2H, ArH); ¹³C NMR (75 MHz, CDCl3, 293K): $\delta_{(ppm)} =$ 169.8, 153.6, 152.9, 151.9, 149.3, 147.2, 146.6, 142.4, 133.5, 133.3, 132.8, 127.8, 127.5, 126.6, 126.4, 125.6, 125.3, 125.0, 124.8, 68.9, 61.8, 61.7, 34.5, 34.4, 34.3, 33.9, 31.9, 31.8, 31.6, 31.3, 30.1; HRMS (ESI-TOF) calcd for C₇₁H₉₂O₈ (M+H)⁺ 1073.6870, found 1073.6931.

Synthesis of calix[6]monoamidephenanthroline 12. Calix[6]monoacid 11 (0.326 g, 0.304 mmol, 1 equiv.), DCC (0.094 g, 0.456 mmol, 1.5 equiv.) and HOBt (0.062 g, 0.459 mmol, 1.5 equiv.) were dissolved in DMF (7 mL). At 0°C, DIPEA (320 μ L, 1.84 mmol, 6 equiv.) was added to the mixture which was stirred for 20min at 0°C under inert atmosphere. Then, 5-glycinamido-1,10-phenanthroline 20 was added to the mixture which was stirred for 16h at room temperature under inert atmosphere. The reaction mixture was concentrated under reduced pressure and dissolved in CH₂Cl₂ (60 mL). The organic layer was washed with H₂O (3×35 mL) and the combined aqueous layers were extracted with CH₂Cl₂ (2×45 mL). The combined organic layers were concentrated under reduced pressure and the crude residue was purified by flash chromatography (CH₂Cl₂/MeOH 95:5) to yield the compound 12 as a clear yellow solid (0.343 g, 0.263 mmol, 86%). Rf = 0.38 (CH₂Cl₂/MeOH 95:5).

m.p. 185 (dec.); IR: v: 3310, 2965, 1687, 1483, 1210 cm⁻¹; ¹H NMR (600 MHz, CDCl₃, 298K): $\delta_{(ppm)} = 0.90$ (s, 18H, *t*Bu), 1.16 (s, 9H, *t*Bu), 1.23 (s, 18H, *t*Bu), 1.32 (s, 9H, *t*Bu), 3.09 (s, 3H, OMe), 3.33 (s, 6H, OMe), 3.41 (d, ²J = 17.0 Hz, 2H, ArCH_{2eq}), 3.49 (d, ²J = 17.0 Hz, 2H, ArCH_{2eq}), 3.50 (d, ²J = 17.0 Hz, 2H, ArCH_{2eq}), 4.05-4.09 (m, 4H, ArCH_{2ax}), 4.17 (s, 2H, OCH₂), 4.33 (d, ³J = 4.8 Hz, 2H, NCH₂), 4.47 (d, ²J = 17.0 Hz, 2H, ArCH_{2ax}), 6.72 (s, 2H, ArH), 6.85 (s, 2H, ArH), 6.96 (d, ⁴J = 2.4 Hz, 2H, ArH), 7.02 (d, ⁴J = 1.8 Hz, 2H, ArH), 7.03 (s, 2H, ArH), 7.25 (s, 2H, ArH), 7.34 (s_b, 2H, OH), 7.36 (dd, ³J = 4.2 Hz, ³J = 8.4 Hz, 1H, H₃), 7.47 (dd, ³J = 4.2 Hz, ³J = 8.4 Hz, 1H, H₈), 7.77 (s_b, 1H, NH_{amide}), 7.81 (d, ³J = 7.8 Hz, 1H, H₇), 7.93 (s, 1H, H₆), 8.43 (d, ³J = 7.8 Hz, 1H, H₄), 9.05-9.07 (m, 2H, H_{2,9}), 9.22 (s_b, 1H, NH_{phen}); ¹³C NMR (75 MHz, CDCl₃, 298K): $\delta_{(ppm)} = 30.5$, 31.1 (2C), 31.2, 31.4, 31.6, 31.7, 34.0, 34.2, 34.3(6), 34.4(3), 45.0, 61.5, 61.7, 71.5, 120.8, 123.2 (2C), 124.7, 125.4, 126.0, 126.1, 126.3, 126.6, 127.3, 128.3, 131.0, 131.1, 132.4, 132.7, 132.9, 133.2, 136.0, 142.5 (2C), 144.8, 146.5, 146.7 (2C), 147.5, 147.6, 149.8, 170.1, 170.2, 171.7, 171.9, 172.7, 169.5, 171.5; HRMS (ESI-TOF) calcd for C₈₅H₁₀₂N₄O₈ (M+H)⁺ 1307.7776, found 1307.7762.

Synthesis of calix[6]monoazido 17. NaN₃ (0.285 g, 4.38 mmol, 4.9 equiv.) was suspended in CH₃CN (16 mL). Trifluoromethanesulfonic anhydride (Tf₂O) (0.510 mL, 3.07 mmol, 3.4 equiv.) was added drop by drop at 0°C. The mixture was stirred for 2h at 0°C under inert atmosphere. Calix[6]monoamine 16 (1.049 g, 0.896 mmol, 1 equiv.) was dissolved in toluene (16 mL). Triethylamine (0.790 mL, 5.63 mmol, 6.3 equiv.) and CuSO₄ (0.013 g, 0.0815 mmol, 0.09 equiv.) were added. At 0°C, the solution containing TfN₃ was added drop by drop to the reaction mixture which was stirred for 16h at room temperature under inert atmosphere. An aqueous NH₄OH solution (25%, 15 mL) was added to the mixture which was stirred for 40min and the aqueous layer was washed with CH₂Cl₂ (2×10mL). The combined organic layers were

concentrated under reduced pressure and dissolved in CH_2Cl_2 (20 mL). The organic layer was washed with an aqueous HCl solution (10%, 15 mL) and the mixture was stirred for 45min at room temperature. Then, the organic layer was washed with H₂O (3×15 mL) until pH = 7 and concentrated under reduced pressure to yield the compound **17** as a clear yellow solid (0.669 g, 0.616 mmol, 69%).

m.p. 166°C; IR: v: 2961, 2110, 1484, 1210 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 298K): $\delta_{(ppm)} = 0.60$ (s, 9H, *t*Bu), 1.02 (s, 18H, *t*Bu), 1.26 (s, 18H, *t*Bu), 1.39 (s, 9H, *t*Bu), 2,12 (t_b, ³J = 4.5 Hz, 2H, CH₂N₃), 2.67 (t_b, ³J = 4.5 Hz, 2H, OCH₂), 3.52 (s, 3H, OMe), 3.85-3.95 (m, 14H, ArCH₂ + OMe), 4.01-4.11 (m, 4H, ArCH₂), 6.22 (s_b, 2H, ArH), 6.85 (d, ⁴J = 2.4 Hz, 2H, ArH), 6.97 (d, 2H, ⁴J = 2.2 Hz, ArH), 7.02 (d, ⁴J = 2.4 Hz, 2H, ArH), 7.18 (d, ⁴J = 2.4 Hz, 2H, ArH), 7.23 (s, 2H, ArH), 7.28 (s, 2H, OH); ¹³C NMR (75 MHz, CDCl₃, 298K): $\delta_{(ppm)} = 29.7$, 30.9, 31.2(6), 31.3(4), 31.6(8), 31.7(4), 32.1, 33.8, 34.0, 34.3, 50.0, 60.4, 61.7, 72.3, 124.4, 125.0, 125.6, 126.3, 126.5, 126.7, 127.1, 128.1, 132.1(7), 132.2(2), 133.1, 133.3, 142.3, 145.5, 146.1, 147.3, 150.6, 151.4, 153.3, 153.8; HRMS (ESI-TOF) calcd for C₇₁H₉₃N₃O₆ (M+H)⁺ 1084.7143, found 1084.7109.

Caution! Although we have not encountered any problem, it is noted that small azide derivatives are potentially explosive and should be handled with appropriate precautions.³²

Synthesis of calix[6]monopytz 18. Calix[6]monoazido 17 (0.403 g, 0.371 mmol, 1 equiv.) and 2-(trimethylsilyl)ethynylpyridine (0.124 g, 0.709 mmol, 2 equiv.) were dissolved in a mixture of *t*-BuOH/H₂O (1:1) (7 mL) and in CH₂Cl₂ (7 mL). CuSO₄ (0.077 g, 0.484 mmol, 1.3 equiv.), sodium ascorbate (0.116 g, 0.586 mmol, 1.6 equiv.) and K₂CO₃ (0.057 g, 0,413 mmol, 1.1 equiv.) were successively added and the mixture was stirred vigorously for 2h30 at room temperature under inert atmosphere. CH₂Cl₂ (190 mL) and an aqueous NH₄OH solution (25%,

58 mL) were added and the mixture was stirred for 2h at room temperature. The organic layer was washed with an aqueous NH₄OH solution (25%, 50 mL) and the combined aqueous layers were extracted with CH₂Cl₂ (2×50 mL). The combined organic layers were washed with H₂O (4×150 mL) until pH = 7 and concentrated under reduced pressure. The crude residue was purified by flash chromatography (CH₂Cl₂/AcOEt 9:1) to yield the compound **18** as a white solid (0.316 g, 0.266 mmol, 72%). Rf = 0.28 (CH₂Cl₂/AcOEt 9:1).

m.p. 145-147°C (dec.); IR: v: 3384, 2961, 1483, 1209, 1011 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 298K): $\delta_{(ppm)} = 0.92$ (s, 9H, *t*Bu), 1.02 (s, 18H, *t*Bu), 1.15 (s, 18H, *t*Bu), 1.39 (s, 9H, *t*Bu), 2.98 (s, 3H, OMe), 3.06 (s_b, 2H, OCH₂), 3.49 (d, ²J = 15.6 Hz, 2H, ArCH_{2eq}), 3.53-3.60 (m, 4H, ArCH_{2eq}), 3.75 (s, 6H, OMe), 3.97 (s_b, 2H, NCH₂), 4.20-4.28 (m, 4H, ArCH_{2ax}), 4.41 (d, ²J = 15.6 Hz, 2H, ArCH_{2ax}), 6.67 (s, 2H, ArH), 6.82-6.89 (m, 4H, ArH), 7.02 (s, 2H, ArH), 7.11 (d, ⁴J = 1.6 Hz, 2H, ArCH_{2ax}), 6.67 (s, 2H, ArH), 6.82-6.89 (m, 4H, ArH), 7.02 (s, 2H, ArH), 7.75 (td, ⁴J = 1.2 Hz, ³J = 6.0 Hz, 1H, H_{py}), 7.82 (s_b, 2H, OH), 8.19 (d, ³J = 8.0 Hz, 1H, H_{py}), 8.38 (s_b, 1H, H_{tz}), 8.64 (d, ³J = 4.8 Hz, 1H, H_{py}); ¹³C NMR (75 MHz, CDCl₃, 298K): $\delta_{(ppm)} = 29.6$, 29.8, 31.0, 31.3, 31.4, 31.6, 31.7, 31.8, 33.9, 34.1, 34.2(9), 34.3(3), 50.3, 60.4, 61.5, 70.4, 120.5, 122.5 123.8, 124.5, 125.2, 125.4, 125.6, 126.0, 126.7, 126.9, 127.9, 132.5, 132.7, 133.1, 133.2, 137.0, 142.3, 146.2, 146.3, 146.9, 147.5, 149.1, 150.2, 150.9, 151.8, 152.8, 153.8; HRMS (ESI-TOF) calcd for C₇₈H₉₉N₄O₆ (M+H)⁺ 1188.7643, found 1188.7573.

Synthesis of calix[6]diazidomonopytz 23. Calix[6]monopytz **18** (0.206 g, 0.173 mmol, 1 equiv.) and NaH (60% in oil, 0.017 g, 0.425 mmol, 4.5 equiv.) were dissolved in a mixture of THF/DMF (1:1) (10 mL). The mixture was stirred for 20min at room temperature under inert atmosphere. 2-azidoethyl-4-methylbenzenesulfonate (0.201 g, 0.834 mmol, 4.8 equiv.) in THF (5 mL) was added and the reaction mixture was stirred for 16h at reflux under inert atmosphere.

The reaction was quenched through the addition of EtOH (2 mL) and the reaction mixture was concentrated under reduced pressure. The residue was dissolved in CH_2Cl_2 (50 mL) and the organic layer was washed with H_2O (3×20 mL). The combined aqueous layers were extracted with CH_2Cl_2 (30 mL) and the combined organic layers were concentrated under reduced pressure. The residue was purified by flash chromatography ($CH_2Cl_2/MeOH$ 9:1) to yield the compound **23** as a white solid (0.162 g, 0.122 mmol, 71%).

m.p. 210-212°C (dec.); IR: v: 3455, 2962, 2108, 1484, 1205, 1017 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 298K): $\delta_{(ppm)} = 0.92$ (s, 27H, *t*Bu), 1.29 (s, 18H, *t*Bu), 1.32 (s, 9H, *t*Bu), 2.49 (s, 6H, OMe), 2.57 (s, 3H, OMe), 3.36 (d, ²J = 15.0 Hz, 2H, ArCH_{2eq}), 3.44-3.53 (m, 4H, CH₂N₃), 3.58-3.70 (m, 4H, ArCH_{2eq}), 3.78-3.95 (m, 4H, OCH₂CH₂N₃), 4.23-4.31 (m, 8H, ArCH_{2ax} + OCH₂CH₂N), 4.80 (t_b, ³J = 5.1 Hz, 2H, CH₂N), 6.74-6.82 (m, 4H, ArH), 6.80 (s, 2H, ArH), 7.14-7.17 (m, 4H, ArH), 7.18-7.22 (m, 3H, H_{py} + ArH), 7.73 (td, ⁴J = 1.8 Hz, ³J = 7.8 Hz, 1H, H_{py}), 8.07 (d, ³J = 7.8 Hz, 1H, H_{py}), 8.44 (s, 1H, H_{tz}), 8.58 (d, ³J = 3.9 Hz, 1H, H_{py}); ¹³C NMR (75 MHz, CDCl₃, 298K): $\delta_{(ppm)} = 30.2(5)$, 30.3(4), 30.4, 30.5, 31.3(0), 31.3(3), 31.7 (2C), 34.2 (2C), 34.3, 34.4, 50.7, 51.2, 60.1 (2C), 70.5, 71.2, 120.5, 122.8, 123.7, 124.6, 124.7, 124.8, 127.4(8), 127.5 (2C), 133.0 (2C), 133.2, 133.4(8), 133.5(4), 133.6, 136.8, 145.9, 146.0, 146.3, 146.6, 148.5, 149.5, 150.4, 151.3, 151.7, 154.3(9), 154.4(4); HRMS (ESI-TOF) calcd for C₈₂H₁₀₄N₁₀O₆ (M+H)⁺ 1325.8219, found 1325.8192.

Figure S1. ¹H NMR spectrum (300 MHz, 298K) of compound 5 in CDCl₃. S = solvent, W = water, G = grease.



Figure S2. ¹³C NMR spectrum (75 MHz, 298K) of compound 5 in CDCl₃. S = solvent.







Figure S4. HSQC NMR spectrum (300MHz, 298K) of compound 5 in CDCl₃.



Figure S5. ¹H NMR spectrum (300 MHz, 298K) of compound 6 in CDCl₃. S = solvent, W = water, G = grease.



Figure S6. ¹³C NMR spectrum (75 MHz, 298K) of compound 6 in CDCl₃. S = solvent.







Figure S8. HSQC NMR spectrum (300MHz, 298K) of compound 6 in CDCl₃.



Figure S9. ¹H NMR spectrum (300 MHz, 298K) of compound 7 in CD₃OD. S = solvent, W = water, G = grease.



Figure S10. ¹³C NMR spectrum (75 MHz, 298K) of compound 7 in CD₃OD. S = solvent.



Figure S11. COSY NMR spectrum (300MHz, 298K) of compound 7 in CD₃OD.



Figure S12. HSQC NMR spectrum (300MHz, 298K) of compound 7 in CD₃OD.







Figure S14. ¹H NMR spectrum (600 MHz, 298K) of compound 12 in CDCl₃. S = solvent, W = water, G = grease.



Figure S15. ¹³C NMR spectrum (75 MHz, 298K) of compound 12 in CDCl₃. S = solvent.





Figure S16. COSY NMR spectrum (300MHz, 298K) of compound 12 in CDCl₃.

Figure S17. HSQC NMR spectrum (300MHz, 298K) of compound 12 in CDCl₃.





Figure S18. HMBC NMR spectrum (600MHz, 298K) of compound 12 in CDCl₃.

Figure S19. ¹H NMR spectrum (300 MHz, 298K) of compound 13 in CDCl₃. S = solvent, W = water, G = grease.



Figure S20. ¹³C NMR spectrum (75 MHz, 298K) of compound 13 in CDCl₃. S = solvent.





Figure S21. COSY NMR spectrum (300MHz, 298K) of compound 13 in CDCl₃.

Figure S22. HSQC NMR spectrum (300MHz, 298K) of compound 13 in CDCl₃.





Figure S23. HMBC NMR spectrum (400MHz, 298K) of compound 13 in CDCl₃.

Figure S24. ¹H NMR spectrum (300 MHz, 298K) of compound 14 in CDCl₃. S = solvent, W = water, G = grease.



Figure S25. ¹³C NMR spectrum (100 MHz, 298K) of compound 14 in CDCl₃. S = solvent.







Figure S27. HSQC NMR spectrum (400MHz, 298K) of compound 14 in CDCl₃.





Figure S28. HMBC NMR spectrum (400MHz, 298K) of compound 14 in CDCl₃.

Figure S29. ¹H NMR spectrum (300 MHz, 298K) of compound 17 in CDCl₃. S = solvent, W = water.



Figure S30. ¹³C NMR spectrum (75 MHz, 298K) of compound 17 in CDCl₃. S = solvent.





Figure S31. COSY NMR spectrum (300MHz, 298K) of compound 17 in CDCl₃.

Figure S32. HSQC NMR spectrum (300MHz, 298K) of compound 17 in CDCl₃.



Figure S33. ¹H NMR spectrum (400 MHz, 298K) of compound 18 in CDCl₃. S = solvent, G = grease.



Figure S34. ¹³C NMR spectrum (75 MHz, 298K) of compound 18 in CDCl₃. S = solvent.







Figure S36. HSQC NMR spectrum (400MHz, 298K) of compound 18 in CDCl₃.





Figure S37. HMBC NMR spectrum (400MHz, 298K) of compound 18 in CDCl₃.

Figure S38. ¹H NMR spectrum (400 MHz, 328K) of compound 19 in CDCl₃. S = solvent, W = water, G = grease.



Figure S39. ¹³C NMR spectrum (100 MHz, 298K) of compound 19 in CDCl₃. S = solvent.







Figure S41. HSQC NMR spectrum (400MHz, 298K) of compound 19 in CDCl₃.





Figure S42. HMBC NMR spectrum (400MHz, 298K) of compound 19 in CDCl₃.

Figure S43. ¹H NMR spectrum (300 MHz, 298K) of compound 23 in CDCl₃. S = solvent.



Figure S44. ¹³C NMR spectrum (75 MHz, 298K) of compound 23 in CDCl₃. S = solvent.







Figure S46. HSQC NMR spectrum (300MHz, 298K) of compound 23 in CDCl₃.





Figure S47. HMBC NMR spectrum (400MHz, 298K) of compound 23 in CDCl₃.

Figure S48. ¹H NMR spectrum (400 MHz, 298K) of compound 24 in CD₃OD. S = solvent and W = water.



Figure S49. ¹³C NMR spectrum (75 MHz, 298K) of compound 24 in CD_3OD . S = solvent.







Figure S51. HSQC NMR spectrum (300MHz, 298K) of compound 24 in CD₃OD.





Figure S52. HMBC NMR spectrum (400MHz, 298K) of compound 24 in CD₃OD.

Figure S53. Cathodic cyclic voltammetry of $[Ru(TAP)_2pytz'(diN_3C6)]^{2+}(NO_3^-)_2$ **24** (1.10⁻³ M) in dry deoxygenated MeCN, V versus SCE at room temperature, 0.1V/s, with 0.1M Bu₄N⁺PF₆⁻ as supporting electrolyte and a Pt working electrode.



Figure S54. Cathodic cyclic voltammetry of $[Ru(TAP)_2pytz'C6]^{2+}(NO_3^{-})_2$ **19** (1.10⁻³ M) in dry deoxygenated MeCN, V versus SCE at room temperature, 0.1V/s, with 0.1M Bu₄N⁺PF₆⁻ as supporting electrolyte and a Pt working electrode.



Figure S55. Cathodic cyclic voltammetry of $[Ru(TAP)_2phen'(C6)]^{2+}(NO_3^-)_2$ **14** (1.10⁻³ M) in dry deoxygenated MeCN, V versus SCE at room temperature, 0.1V/s, with 0.1M Bu₄N⁺PF₆⁻ as supporting electrolyte and a Pt working electrode.



Figure S56. Anodic cyclic voltammetry of $[Ru(TAP)_2pytz'(diN_3C6)]^{2+}(NO_3^{-})_2$ **24** (1.10⁻³ M) in dry deoxygenated MeCN, V versus SCE at room temperature, 0.1V/s, with 0.1M Bu₄N⁺PF₆⁻ as supporting electrolyte and a Pt working electrode.



Figure S57. Absorption spectra (298 K, air) in MeCN for complexes 6 (black), 7 (red), 13 (blue), 14 (green), 19 (orange) and 24 (pink).



Figure S58. Stern-Volmer experiments (luminescence intensities (green) and lifetimes (red)) for $[Ru(TAP)_2phen]^{2+}$ **25** (1.10⁻⁵ M) in acetonitrile, in presence of phenol.



Figure S59. Stern-Volmer experiments (luminescence intensities (green) and lifetimes (red)) for $[Ru(TAP)_2phen]^{2+}$ **25** (1.10⁻⁵ M) in acetonitrile, in presence of an acid (paratoluene sulfonic acid = PTSA).



Figure S60. Deuterated phenol was obtained after 30 minutes mixing in hot CD_3OD . Deuteration of the phenol moiety was checked by ¹H NMR (300 MHz, 298 K) in $CDCl_3$, A) ¹H NMR spectrum of phenol and B) ¹H NMR of deuterated phenol.



Figure S61. Stern-Volmer experiments (luminescence intensities (green) and lifetimes (red)) for $[Ru(TAP)_2phen]^{2+}$ **25** (1.10⁻⁵ M) in acetonitrile, in presence of deuterated phenol.



Figure S62. $[Ru(phen)_2phen'(C6)]^{2+}(NO_3^-)_2$ **13** in acetonitrile under inert atmosphere. Decay of the TA at 510 nm in a microseconds timescale.



Figure S63. Equation for ΔG_{PCET}^0 in the case of a PCET: (1.1) ET followed by a HT or (1.2) ET and HT concerted. The following equations cannot be applied in our case due to problems for determinations of important experimental values such as: precise E*_{red}, pKa in MeCN, pKa of the reduced and excited complex.

$$\Delta G_{PCET}^{0} = -nF \left[E_{red(Ru^{II}/Ru^{I})}^{*} - E_{ox(phOH/phOH^{\bullet+})} \right] + 0.0059 \times \{ pK_{a}(phOH^{\bullet+}) - pK_{a}(Ru^{I}) \}$$
(1.1)

$$\Delta G_{PCET}^{0} = -nF \left[E_{red(Ru^{II}/Ru^{I})}^{*} - E_{ox(phOH/phOH^{*+})} \right] + 0.0059 \times \{ pK_{a}(phOH^{*+}) - pK_{a}(Ru^{*II}) \} (1.2)$$

(1) Gans, P. Data Fitting in the Chemical Sciences: By the Method of Least Squares; John Wiley & Sons, 1992.