Supporting Information

Modulation of the fibrillogenesis inhibition properties of two transthyretin ligands by halogenation

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Figure S0. Selected TTR ligands from the Protein Data Bank:

- 1 thyroxine or T4, **ligand T44** in pdb 2ROX;
- 2 3,3',5,5'-tetraiodothyroacetic acid or T4Ac, **ligand T4A** in pdb1Z7J;
- 3 flurbiprofen, **ligand FLP** in pdb 1DVT;
- 4 flufenamic acid, **ligand FLF** in pdb 1BM7;
- 5 diclofenac, **ligand DIF**inpdb 1DVX;
- 6 retinoic acid, **ligand REA** in pdb 1TYR;
- 7 resveratrol, **ligand STL** in pdb1DVS;
- 8 diethylstilbestrol, ligand DES in pdb 1TT6;
- **9** 3,5,3',5'-tetrachlorobiphenyl-4,4'-diol, **ligand PCQ** in pdb 2G5U;
- **10** 2,4-dinitrophenol, **ligand DNF** in pdb 2B15;
- **11** 3',5'-difluorobiphenyl-4-carboxylic acid, ligand FBC in pdb 2B9A;
- 12 2',4'-dichloro-4-hydroxy-1,1'-biphenyl-3-carboxylic acid, ligand 3CA in pdb 2B77;
- 13 iododiflunisal, ligand FHI in pdb 1Y1D.



Figure S1. Chemical nature of the ligand binding site of TTR.

COMMENT 1:

Crystallographic properties of the TTR-ligand complexes. To properly compute the PDB crystal information it was important to take into account that, as the majority of human TTR-ligand crystal complexes, the ones considered in the present work belong to the orthorhombic space group $P2_12_12_2$. In all of them, the two binding sites located at the dimer-dimer interface are bisected by the crystallographic twofold axis (C_2) . Consequently, the electron density observed for non-symmetric ligands in each binding site is an average of the two symmetry-related positions. Therefore, ligands not possessing twofold symmetry that adopt one single conformation or binding mode bound to TTR, must have a 50% statistical disorder or 0.50 of maximal occupation in each binding site. Taking into account the symmetry-related conformer or binding mode the maximal occupation for the majority of these ligands results to be of 100%. (Table S1). Flufenamic acid (4), for which it has been possible to distinguish two conformations (cis and trans) in the binding site of TTR is an exception. In this case, the occupation factor of each conformer is 0.25 and by considering the two symmetry-related conformers associated to each one of them, the resulting maximal occupation for (4) in the binding site is 1. In addition, this maximal value of occupancy is not observed in the TTR complexes with iododiflunisal (13), retinoic acid (6), tetraiodothyroacetic (2), and diclofenac (5) (Table 1). For these ligands, deviation from the maximum occupancy value could be due to the fact that their structures were not refined to obtain a final occupancy of 0.50 for the ligand atoms.

LIGAND	PDB	LIGAND PDB code	Binding mode	LIGAND OCCUPANCY	Reference In PDB		
1	2ROX	T44	F	0.50	а		
2	1Z7J	T4A	F R	0.18 0.22	b		
3	1DVT	FLP	F	0.50	С		
4	1BM7	FLF	F	0.25	d		
5	1DVX	DIF	R	0.33	с		
6	1TYR	REA	F	0.25	e		
7	1DVS	STL	F	0.50	c		
8	1TT6	DES	F	0.50	f		
9	2G5U	PCQ	R	0.50	g		
10	2B15	DNF	R	0.50	h		
11	2B9A	FBC	R	0.50	i		
12	2B77	3CA	R	0.50	i		
13	1Y1D	FHI	F	0.29	j		

Table S1 Crystallographic properties of the ligands and ligand-TTR complexes. Forward and reverse binding modes are indicated by F and R, respectively.

References in PDB

- a) Wojtczak, A.; Cody, V.; Luft, J.R.; Pangborn, W. Structures of human transthyretin complexed with thyroxine at 2.0 A resolution and 3',5'-dinitro-N-acetyl-L-thyronine at 2.2 A resolution. *Acta Crystallogr. D Biol. Crystallogr.* **1996**, *52*(Pt 4):758-765.
- b) Neumann, P.; Cody, V.; Wojtczak, A. Ligand binding at the transthyretin dimer-dimer interface: structure of the transthyretin-T4Ac complex at 2.2 Angstrom resolution. *Acta Crystallogr. D Biol. Crystallogr.* 2005, 61(Pt 10):1313-1319.
- c) Klabunde, T.; Petrassi, H.M.; Oza, V.B.; Raman, P.; Nelly, J.W.; Sacchettini, J.C. Rational design of potent human transthyretin amyloid disease inhibitors. *Nat. Struct. Biol.* 2000, 7, 312-321.
- d) Peterson, S.A.; Klabunde, T.; Lashuel, H.A.; Purkey, H.; Sacchettini, J.C.; Kelly, J.W. Inhibiting transthyretin conformational changes that lead to amyloid fibril formation. *Proc. Natl. Acad. Sci. U S A.* **1998**, *95*, 12956-12960.
- e) Zanotti, G.; D'Acunto, M.R.; Malpeli, G.; Folli, C.; Berni, R. Crystal structure of the transthyretin--retinoic-acid complex. *Eur. J. Biochem.* **1995**, *234*, 563-569.
- f) Morais-de-Sá, E.; Pereira, P.J.; Saraiva, M.J.; Damas, A.M. The crystal structure of transthyretin in complex with diethylstilbestrol: a promising template for the design of amyloid inhibitors. *J. Biol. Chem.* **2004**, *279*, 53483-53490.
- g) Purkey, H.E.; Palaninathan, S.K.; Kent, K.C.; Smith, C.; Safe, S.H.; Sacchettini, J.C.; Kelly J.W. Hydroxylated polychlorinated biphenyls selectively bind transthyretin in blood and inhibit amyloidogenesis: rationalizing rodent PCB toxicity. *Chem. Biol.* 2004, 11, 1719-1728.

- h) Morais-de-Sá, E; Neto-Silva, R.M.; Pereira, P.J.; Saraiva, M.J.; Damas, A.M. The binding of 2,4-dinitrophenol to wild-type and amyloidogenictransthyretin. *Acta Crystallogr. D Biol. Crystallogr.* 2006, 62(Pt 5), 512-519.
- i) Adamski-Werner, S.L.; Palaninathan, S.K.; Sacchettini, J.C.; Kelly, J.W. Diflunisal analogues stabilize the native state of transthyretin. Potent inhibition of amyloidogenesis. *J. Med. Chem.* **2004**, *47*, 355-374.
- j) Gales, L.; Macedo-Ribeiro, S; Arsequell, G.; Valencia, G.; Saraiva, M.J.; Damas, A.M. Human transthyretin in complex with iododiflunisal: structural features associated with a potent amyloid inhibitor. *Biochem. J.* 2005, 388, 615-621.

COMMENT 2:

Hydrogen bonds involving Lys 15 can be either direct, such as in the case of (3), (4), (6) or (8) or mediated by a water molecule as in the case of (7), (10) or (13) (Figure S2)



Figure S2. Types of hydrogen bond involving Lys15

COMMENT 3:

Besides the conformational changes observed for the Lys15 residues, other important structural rearrangements produced by ligand binding occur at the inner part of the binding site. In the apo-TTR form, the HBP3 and HBP3' are occupied by water molecules, each one acting as a bridge between the hydroxyl groups of the Ser117 and Thr119 of each pocket. The placement of a ligand substituent in any of these pockets displaces the initial water molecule, leading to a rotation of the side chains of these residues. The conformation adopted by residues Ser117 and Thr119 upon ligand binding depends on the nature of the substituting group that, in turn, triggers two major effects. On the one hand, it allows the formation of additional intermonomer

interactions which may contribute to stabilize the TTR tetramer. On the other hand, it modifies the surface polarity of the HBP3/3' pockets.

Figures S3 and S4 schematically describe the conformational changes of residues Ser117 and Thr119 upon ligand binding. In the apo-TTR form, only Thr119 takes part in intermonomer interactions, forming a hydrogen bond to the hydroxyl group of Ser115 from the adjacent monomer. This interaction is conserved in the TTR-(**3**) complex which has no substituents in the HBP3/3' pockets. Upon ligand binding, this hydrogen bond is replaced by two new additional interactions: Thr119 A (OH) – (O) Tyr114 B and Thr119 A (OH) – water – (O) Asp18 A' (or Thr119 A' (OH) – (O) Tyr114 B' and Thr119 A' (OH) – water – (O) Asp18 A). In contrast to Thr119, Ser117 only takes part in the intermonomer interaction network upon ligand binding.







Figure S4

COMMENT 4



Figure S5: A) Some examples of the promiscuous ligand-binding properties of TTR. B) Selected ligands from TTR-ligand complexes available at the PDB (See Figure S0 for details on the related PDB structures)

A clear example of the lack of correlation between binding mode and biological activity is provided, in our case, by compounds (3) and (6).Both compounds bind to TTR in the forward mode, but their RA (%) are of 35% and 100% respectively. More generally, in spite of the large amount of information accumulated on the forward and reverse modes of many different families of ligands, a successful prediction of the ligand binding mode in the binding site of TTR is a very difficult task for several reasons: (a) very similar and structurally related compounds can bind to the protein in both modes. For example, (4) and (5) two bisarylamine compounds, bind to TTR in forward and reverse mode, respectively (Figure S5a); (b) ligands containing the same scaffold, *i.e.* biphenyls (3), (9) and (13), can be placed differently in the TTR binding site (Figure S5b); (c) some ligands adopt more than one orientation bound to TTR. This is the case of the thyroxine derivative 3,3',5,5'-tetraiodothyroacetic acid (2), which is able to bind transthyretin in both forward and reverse modes (Figure S5c); and (d) there are ligands that are able to adopt more than one conformation bound to TTR. This is the case of the crystallographic complex TTR-(4) which shows the ligand bound to the protein in both *cis* and *trans* conformations (Figure S5d).

LIGAND	PDB	LIGAND PDB code	Binding mode
			F
2	1Z7J	T4A	R
3	1DVT	FLP	F
4	1BM7	FLF	F
5	1DVX	DIF	R
6	1TYR	REA	F
9	2G5U	PCQ	R
13	1Y1D	FHI	F

COMMENT 5

Indeed, one of the clearest evidences pointing out that TTR constitutes an unusual system comes from the superposition of the ligand conformations deduced from the crystal structures that clearly demonstrate the impossibility to define a classical pharmacophore due to the promiscuous ligand-binding of TTR (Figure S5a, Comment 4).

Some examples of the promiscuous ligand-binding properties of TTR. (a) Very similar compounds (4) in fuchsia and (5)in blue bind to TTR in forward and reverse modes respectively; (b) The same compound (2) is able to bind to TTR in both forward (maroon) and reverse (grey) modes; (c) The same compound (4) is able to adopt both *cis* and *trans* conformations within the binding site of TTR.

The computational analysis performed on the series of 13 crystallographic TTR-ligand complexes considered in the present work provides good evidences about the promiscuous ligand-binding properties of TTR. This promiscuity is supported by: (1) the ability of TTR to accommodate a wide diversity of structures; (2) structurally related compounds can bind in forward or reverse mode; (3) structurally similar ligands can be placed rather differently within the binding site; (4) one single molecule can bind in both forward and reverse modes and, (5) a ligand can adopt more than one conformation into the binding site of TTR.

In addition and regardless of the structure, the chemical properties or the binding mode of the ligand to TTR, it is possible to deduce the following SAR correlations and binding features: (1) no single interaction can be qualitatively correlated with the fibrillogenesis inhibitory activity of the ligands; (2) A common feature in all these structures is that regardless of the ligand, there is always a contribution of the residues Lys15, Leu17, Ala108, Leu110, Ser117 and Thr119 of monomers A and A' to the van der Waals interaction energy between TTR and the ligand; (3) ligand binding allows residues Ser117 and Thr119 to make additional intermonomer interactions, reinforcing their contribution to the stability of the tetramer; (4) no relationship can be found between the binding mode or number of occupied HBPs and the inhibitory potency of a compound and finally, (5) the most active compounds have in common an halogen atom positioned in the HBP1/1' or HBP3/3' pockets.

The estimated interaction values for Ala108, Ser117 and Thr119 are generally lower and remain constant across the series. However, the involvement of Ser117 and Thr119 in the hydrogen bond network of the TTR-ligand complexes enhances their contribution to the stability of the complex. In contrast to these two residues, the contribution of residues Lys15 to ligand binding is highly variable but the most active compounds have a large contribution of the Lys15 residues to their binding (Figure S6).

			Van der Waals interaction														
Ligand	RA (%)	Monomer	13	15	16	17	54	106	107	108	109	110	117	118	119	120	121
FLP	25	A		1	i ii		j i									i – 1	
	35	A'			[
FBC	89	A															185
		A'		x - 5			(6	3 8				÷ 3				2 - 8	
304	90	A															
JLA	50	A'					(C)	÷ 8				à là				9 - X	
CHI	94	A	5				63 - 3 1	S 14								e #	
F III	34	A'			I Ü			j – j				Ì.				8 8	
PCO	100	A		ļ							()				<u>(</u>)	((
104	100	A'															
REA	110	A	ę - 5	:				s									- 33
ALA	110	A'		5 5			-	5 5			;;	5 - 3				2 - X	
DNE	80	A															
Dia	00	A'										i i				6 X	
TAA	76	A															
1.44		A'							<u> </u>								
	6	A	[]														
T4A	<mark>9</mark> 3	A'	ļ.—.,														
1.10		A		: 3			-		- 8							_	
a 10	1	A'		5 - 3			-					<u>i i</u>			s - 3	e 3	
DES	67	A															
DLU		A'						i i					i				
STI	77	A															
UIL		A'										_					
FLF	98	A															
100000	0.505	A'	. — .				8		- 22			4					
DIF	39	A	2		- 8		e	2 8				a - 13				<u> </u>	-
	0.53.56	A.	÷				a		- 63		(;)					;;;	

Figure S6. Van der Waals contribution to ligand binding made by individual residues of monomers A and A'. Colour intensity is directly related with the strength of the interaction.

Figure S7. TTR-iododiflunisal (13) complex. Residues Thr106, Ala108, Thr119, and Val121 shape the halogen binding pocket establishing van der Waals contacts with the iodine atom. The –COOH and –OH groups of IDIF are only available for a badly oriented salt bridge and a water mediated hydrogen bond with Lys15, respectively.



CHEMISTRY

Synthesis of analogues. Flufenamic acid analogues are *N*-aryl anthranilic acids. Conventionally, substituted anthranilic acids can be prepared from anilines via isatins through the Sandmeyer methodology or via Ullmann reactions, this is, either the condensation between a 2-halobenzoic acid and an aryl amine or an anthranilic acid and an aryl halide. Palladium-catalyzed C-N bond-forming methods are widely recognized as convenient and efficient means for obtaining *N*-arylation products.¹ Here we used Buchwald–Hartwig cross coupling reactions of o-halobenzoic acids with arylamines.² Synthesis of these halogenated flufenamic acid analogues was achieved via two step reaction. The key feature is first a palladium-catalyzed Buchwald–Hartwig cross coupling reaction followed by saponification of the formed methyl ester (Scheme 1).³

Briefly, the methyl anthranilate derivatives **18a-20a** were synthesised by palladium catalyzed crosscoupling reaction between commercially available methyl 2-bromobenzoate and the corresponding 5halogenated 3-trifluoromethyl substituted anilines (fluoro-, chloro- and bromo-). Briefly, the corresponding aniline (1.2 equiv), Cs_2CO_3 (120 mol%), BINAP (5 mol%) and $Pd_2(dba)_3$ (3 mol%) were added to a solution of methyl 2-bromobenzoate (1.2 equiv) in toluene (0.1 M) at room temperature. The reaction mixture was stirred at 120 °C for 4-48 h.

The iodinated anthranilic ester compound **21b** could not be prepared following the same pathway. A new pathway was explored using the bromo methyl ester **20a**. Heating **20a** in the presence of sodium iodide and copper iodide (I) in *N*-methyl pyrrolidone at 160 °C for 24h yielded a mixture (1:1) of the bromo and iodo analogues with a low conversion (58%) as analysed by HPLC. Thus, compound **21b** was synthesised in two steps from the brominated compound **20a** involving an organostanne as intermediate. The arylstannane intermediate was obtained by a Stille-type cross coupling reaction. Alternatively, treatment of the bromocompound **20a** with hexabutylditin in the presence of a catalytic amount of Pd(PPh₃)₄ afforded the desired aryl stannane**21a** containing a tributylstannyl group. The iodinated methyl anthranilate**21** was obtained by electrophilic iodination of the stannyl derivative with

a mixture of sodium iodide and *N*-chlorosuccinimide in equimolar amounts yielded. The final saponification step with lithium hydroxide in a mixture of dioxane/H₂O (5:1) at 80 °C was performed to obtain the final *N*-aryl anthranilic acid derivatives **18-21**.



Scheme 1.Synthesis of halogenated flufenamic acid analogues^a

^a Reagents and conditions: (a) 2,5 mol% Tris(dibenzylideneacetone)dipalladium(0) (Pd₂(dba)₃), 5 mol% *rac*-2,2'-bis(diphenylphosphino)-1,1'-binaphthyl (BINAP), Cs₂CO₃, toluene, reflux, 15h; (b) LiOH (5 equiv), dioxane/H₂O (5:1) 80 °C, 4h; (c) LiCl (5 equiv), Pd(PPh₃)₄ (0,3 equiv), hexabutylditin (SnBu₃)₂, dioxane, reflux, 4h; (d) NaI (1.1 equiv), NCS (1.1 equiv), acetonitrile, rt, 30 min.

Synthesis of halogenated diffunisal compounds 13, 16 and 17 was achieved from the commercially available NSAID drug diflunisal using different procedure involving halogenating reagents. First, the synthesis of the iododerivative 13 (iododiflunisal) was performed as already reported⁴ using bis(pyridine)iodonium(I) tetrafluoroborate or Barluenga's reagent.⁵ Solution-phase bromination of diffunisal using commercially available polymer bround brominating agent⁶ (poly(4-vinylpyridine hydrobromide perbromide) suspended in acetonitrile yielded the corresponding analogue 17. After unsuccessful attempts to directly fluorinate diffunisal using Selectfluor reagent⁷ (F-TEDA: 1-Chloromethyl-4fluoro-1,4-diazoniabicyclo[2.2.2]octane bis(tetrafluoroborate) in acetonitrile at rt, an alternative pathway was chosen starting with commercially available 3-fluorosalicylic acid (15a). A first step in this pathway implied the direct iodination of the salicylic compound 15a which afforded the iodo compound 15b in high yield. A second step was the palladiumcatalysed cross-coupling reaction (Suzuki reaction) in neat water between the iodosalicylic derivative and the 2,4-difluorophenyl boronic acid using palladium acetate as catalyst (1 mol%) and sodium carbonate as a base. The coupling reaction vielded the fluorinated analogue 15 in a 95% yield.

Scheme 2.Synthesis of halogenated diflunisal analogues^a



^aReagents and conditions: (a) for **13**: IPy_2BF_4 (Barluenga's reagent) (1.5 equiv), dichloromethane, rt, 1h; for 16: NCS (2.1 equiv), a drop of concentrated HCl, acetonitrile, rt, 1h; for 17: pyridine hydrobromide perbromide polymer-bound (PVPHP) (~1 mmol Br₃ per g of resin, 5 equiv), dichloromethane, rt, 18h; (b) from 3-fluorosalicylic acid (15a), then IPy_2BF_4 (Barluenga's reagent) (1.5 equiv), dichloromethane, rt, 1h; (c) 2,4-difluorophenyl boronic acid (1 equiv); 1 mol% Pd(OAc)₂, Na₂CO₃ (3 equiv), H₂0, rt, 3h.

ABBREVIATIONS

AcOH, acetic acid; IPy_2BF_4 , bis(pyridine)iodonium(I) tetrafluoroborate; NCS, *N*-chlorosuccimide; F-TEDA; 1-Chloromethyl-4-fluoro-1,4-diazoniabicyclo[2.2.2]octane bis(tetrafluoroborate); DMSO, dimethylsulfoxide; EtOAc, ethyl acetate; HRMS; high resolution mass spectrometry; HPLC, high performance liquid chromatography; 1H NMR, proton nuclear magnetic resonance; PVPHP, pyridine hydrobromideperbromide polymer-bound; $Pd(PPh_3)_4$; RT, retention time; Tetrakis(triphenylphosphine)palladium(0); (Pd₂(dba)₃), Tris(dibenzylideneacetone)dipalladium(0).

References for the synthesis of compounds

- BOOKS: a) Negishi E-i, de Meijere A., editors. Handbook of Organopalladium Chemistry for Organic Synthesis.Wiley; 2002. (b) Tsuji J, editor. Perspectives In Organopalladium Chemistry For The 21st Century. Elsevier; 1999. (c) De Meijere A, Diederich F, editors. Metal-Catalyzed Cross-Coupling Reactions.Wiley-VCH; 2nd; Weinheim 2004.
- (2) BUCHWALD-HARTWIG reaction: (a) Guram, A. S.; Buchwald, S. L. Palladium-Catalyzed Aromatic Aminations with in situ Generated Aminostannanes, J. Am. Chem. Soc. 1994, 116, 7901–7902. (b) Paul, F.; Patt, J.; Hartwig, J. F. Palladium-catalyzed formation of carbon-nitrogen bonds. Reaction intermediates and catalyst improvements in the hetero cross-coupling of aryl halides and tin amides. J. Am. Chem. Soc. 1994, 116, 5969–5970. (c) Surry, D.S.; Buchwald, S. L. Dialkylbiarylphosphines in Pd-catalyzed amination: a user's guide, Chem. Sci. 2011, 2, 27-50; (d) Hartwig, J. Evolution of a Fourth Generation Catalyst for the Amination and Thioetherification of Aryl Halides, Acc. Chem. Res. 2008, 41, 1534-1544; (e) Maiti, D.; Fors, B. P.; Henderson, J. L.; Nakamura, Y.; Buchwald, S. L. Palladium-Catalyzed Coupling of Functionalized Primary and Secondary Amines with Aryl and Heteroaryl Halides: Two Ligands Suffice in Most Cases, Chem. Sci.2011, 2, 57-68. (f) Muci, A.R.; Buchwald, S. L., Practical Palladium Catalysts for C-N and C-O Bond Formation. Topics in Curr. Chem. 2002, 219, 131–209; (g) Hartwig, J. F. Transition Metal Catalyzed Synthesis of Arylamines and Aryl Ethers from Aryl Halides and Triflates: Scope and Mechanism. Angew Chem. Int. Ed. 1998, 37, 2046–2067.
- (3) Synthesis of N-phenyl anthranilates as transthyretin amyloid fibril formation inhibitors: a) Oza, V. B.; Smith, C.; Raman, P.; Koepf, E. K.; Lashuel, H. A.; Petrassi, H. M.; Chiang, K. P.; Powers, E. T.; Sachettinni, J.; Kelly, J. W. Synthesis, structure, and activity of diclofenac

analogues as transthyretin amyloid fibril formation inhibitors. *J. Med. Chem.* **2002**, *45*, 321-32; b) Oza, V. B.; Petrassi, M.; Purkey, H. E.; Kelly, J. W.; Synthesis and Evolution of Anthranilic Acid-Based Transthyretin Amyloid Fibril Inhibitors. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 1-6.

- (4) Mairal, T.; Nieto, J.; Pinto, M.; Almeida, M. R.; Gales, L.; Ballesteros, A.; Barluenga, J.; Pérez, J. J.; Vázquez, J. T.; Centeno, N. B.; Saraiva, M. J.; Damas, A. M.; Planas, A.; Arsequell, G.; Valencia, G. Iodine atoms: a new molecular feature for the design of potent transthyretin fibrillogenesis inhibitors. *PLoS ONE* 2009;4(1):e4124. DOI: 10.1371/journal.pone.0004124. Publication date: January 6, 2009.
- (5) (a) Barluenga, J. Transferring iodine: more than a simple functional group exchange in organic synthesis, *Pure Appl. Chem.* 1999, 71, 431-436; (b) Muñiz, K. Bis(pyridine)iodonium(I)tetrafluoroborate, *Synlett* 1999, 1679; (c) Arsequell, G.; Espuña, G.; Valencia, G.; Barluenga, J.; Pérez-Carlón, R.; González, J. M. *Tetrahedron Lett.* 1998, 39, 7393-7396.
- (6) (a) Frechet, J. M. J.; Farrall, M. J.; Nuyens, L. J.; Polymeric reagents. II. Synthesis and applications of cross-linked poly(vinylpyridiniumhydrobromideperbromide) resins, *J. Macromol. Sci. Chem. A* 1977, *11*, 507-51; (b) Barbara, Z.; Marko, Z. Bromination of aromatic molecules with polymer-supported reagents, *Tetrahedron* 1989, *45*, 7869-7872.
- (7) Singh, R. P.; Shreeve, J. M. Recent highlights in electrophilic fluorination with 1-chloromethyl-4-fluoro-1,4-diazoniabicyclo[2.2.2]octane bis(tetrafluoroborate). *Acc. Chem. Res.* **2004**, *37*, 31-44.

CHEMICAL SYNTHESIS OF COMPOUNDS

TLC analyses were carried out on Merck Kieselgel 60 F₂₅₄ silica gel plates. Visualization was by UV light (254 nm). Chromatographic purification was carried out on Silica Gel columns 60 (70-200 µm, 70-230 mesh) from SDS. Solvents for synthesis purposes were used at GPR grade. Reverse-phase HPLC analyses were performed on a MERCK-HITACHI (D-6000) HPLC System with an UV L-4000 detector (λ =214 nm), a L-6200 pump, and an AS-2000 automatic injector, using a Merck LiChroCART 250-4 LiChrospher 100-5 RP-C18 column (250 mm × 4.6 mm) with injection volume at 10 µL and sample concentrations at 1-2 mg/0.5 mL in 100% water; the sample was detected at single wavelength of 214 nm with a mobile phase system composed with a mixture of acetonitrile:water each containing an 0.1 % of TFA at 1 mL/min flow. The column was maintained at room temperature. The gradient program used (GEN1) was as follows: from a initial (50:50) mixture of A:B to a (10:90) mixture of (A:B) in 25 min, then back to (50:50) in 15 min. All compounds reported here exhibited spectral data consistent with their proposed structures and had HPLC purities in excess of 97%. Chemical names were generated using ChemBioDraw version by CambridgeSoft. 1H and 13C NMR spectra were recorded at 400 MHz and 100,62 MHz, respectively, in CDCl₃ or (CD₃)₂SO on a Varian Mercury-400 spectrometer. Chemical shifts are reported relative in parts per million (ppm) relative to the residual signal of the deuterated solvent (δ 7.27 for ¹H; δ 77.23 for ¹³C) or deuterated or ¹³C labeled dimethyl sulfoxide (δ 2.50 for ¹H, δ 39.53 for ¹³C) as internal standards. 1H-NMR data are tabulated in the following order: multiplicity as follows (s, singlet; d, doublet; t, triplet; m, multiplet; br, broad), coupling constants (J) are reported in Hertz (Hz), and number of protons. HRMS spectra (UPLC-TOF/MS) were recorded on a Waters ACQUITY UPLC System with Waters LCT Premier XE Mass Spectrometer operating either in the positive ion electrospray mode or in negative electrospray mode. Water and acetonitrile were used as carrier solvents.

Compound characterization

All compounds were obtained in milligram quantities. Details of the synthesis and characterization of each compounds is described below. Selected spectra are included inn the Supporting Information. Structures of all reported compounds were confirmed by ¹H and ¹³C NMR and their mass confirmed by HRMS. Purity was determined by HPLC analysis. All reported compounds had >95% purity.

Synthesis of compounds

ANTHRANILIC SERIES



General procedure A: Pd⁰-catalyzed reaction between 3-halo-5-(trifluoromethyl)anilines and methyl 2-bromobenzoate.

A round bottom flask was charged with Cs_2CO_3 (1.20 mmol), *rac*-BINAP (0.05 mmol) and $Pd_2(dba)_3$ (0.025 mmol). The mixture was gently purged with argon, suspended in anhydrous toluene (8 mL) and refluxed for 5 min. Then, a solution of 3-halo-5-(trifluoromethyl)aniline (1.00 mmol) and methyl 2-bromobenzoate (1.10 mmol) in anhydrous toluene (2 mL) was added dropwise. The reaction mixture was refluxed overnight and then diluted with 50 mL of ethyl acetate, filtered over Celite and evaporated to dryness. The residue was then purified by silica gel column chromatography (hexane/ethyl acetate 85:15).

General procedure B: Saponification of methyl esters

To a stirred solution of the corresponding anthranilic methyl ester (1.00 mmol) in 1,4-dioxane (5 mL) was added a solution of LiOH (5.00 mmol) in water (1 mL). The mixture was then heated at 80 °C for 4 h, acidified with 0.25 M HCl (20 mL) and diluted with ethyl acetate (50 mL). The phases were



separated and the aqueous phase was back extracted with more ethyl acetate (25 mL). The organic layer was separated and washed with brine, dried over anhydrous Na_2SO_4 , and concentrated. The residue was purified by silica gel column chromatography (hexane/ethyl acetate (1:1)) to afford the pure acid.

MethylN-(3-fluoro-5-(trifluoromethyl)phenyl)anthranilate (18a):

General procedure A: From 3-fluoro-5-(trifluoromethyl)aniline (361 mg, 2.01 mmol) and methyl 2-bromobenzoate (337 μ L, 2.4 mmol). Silica gel column chromatography afforded **18a** (534 mg, 85%) as a white solid.

White solid.**RP-HPLC** (GEN1): RT = 23,85 min. ¹**H-NMR (500 MHz, CD₃COCD₃):** 3.91 (s, 3H), 6.98 (ddd, J = 16.5, 11.0, 3.5,1H), 7.04-7.12 (m, 1H), 7.34 (dt, J = 18.0, 3.5, 1H), 7.41 (m, 1H), 7.45-7.47 (m, 2H), 8.00 (ddd, J = 13.5, 2.5, 1.1, 1H), 9.63 (br s, 1H). ¹³**C-NMR (75 MHz, CD₃COCD₃):** 53.3, 107.1 (dq, J = 25.1, 3.5), 111.1 (d, J = 24.8), 114.0 (dq, J = 3.3, 3.3), 116.5, 117.7, 121.6, 125.1 (qd, J = 267.4, 5.2), 133.4, 133.8 (qd, J = 34.2, 12.3), 136.0, 146.5 (d, J = 10.7), 146.5, 165.1 (d, J = 244.5), 169.8. **HRMS (ESI⁺):** Calc. for [C₁₅H₁₁F₄NO₂+H]⁺ = 314.0799; found 314.0806.

N-(3-Fluoro-5-(trifluoromethyl)phenyl)anthranilic acid (18)

General procedure B from **18a** (200 mg, 0.64 mmol). Silica gel column chromatography afforded **18** (190 mg, 89%) as a white solid.



White solid.**RP-HPLC** (GEN1): $RT = 14,93 \text{ min.}^{1}H$ -NMR (500 MHz, DMSO-d₆): 6.99 (ddd, J = 9.0, 7.0, 1.0, 1H), 7.13 (dt, J = 8.5, 1.5, 1H),

7.30-7.38 (m, 2H), 7.40 (dd, J = 8.0, 1.0, 1H), 7.50 (ddd, J = 10.0, 6.5, 1.5, 1H), 7.94 (dd, J = 8.0, 2.0, 1H), 9.65 (s, 1H). ¹³**C-NMR (100 MHz, CDCI₃):** 104.8 (dq, J = 25.5, 3.8), 108.8 (d, J = 24.7), 111.4, 116.3, 116.7, 120.2, 123.3 (qd, J = 267.1, 3.4), 131.7 (qd, J = 34.2, 12.9), 131.8, 134.0, 143.9, 144.8, (d, J = 11.1), 162.8 (d, J = 243.4), 169.1. **HRMS (ESI'):** Calc. for $[C_{14}H_9F_4NO_2-H]^{-} = 298.0497$; found 298.0482.

Methyl N-(3-chloro-5-(trifluoromethyl)phenyl)anthranilate (19a)

General procedure A: From 3-chloro-5-(trifluoromethyl)aniline (215 mg, 1.10 mmol) and methyl 2-bromobenzoate (143 μ L, 1.00 mmol). Silica gel column chromatography afforded **19a** (106 mg, 32%)as an oil.



Colorless oil. R_f (hexane/EtOAc 70:30) = 0.65.¹H-NMR (400 MHz, CDCI₃): 3.92 (s, 3H), 6.89 (ddd, J = 8.1, 7.2, 1.2, 1H), 7.24 (s, 1H), 7.31-7.36 (m, 2H), 7.38-7.45 (m, 2H), 8.01 (dd, J = 8.0, 1.6, 1H), 9.65 (br s, 1H). ¹³C-NMR (100 MHz, CDCI₃): 52.1, 113.9, 115.7, 115.7 (quad, J = 3.8), 119.2 (quad, J = 3.9), 119.3, 123.1 (quad, J = 1.0), 123.2 COOMe (quad, J = 272.9), 131.9, 133.0 (quad, J = 32.9), 134.3, 135.7, 143.2,

145.5, 168.7. **HRMS (ESI⁺):** Calc. for $[C_{15}H_{11}CIF_3NO_2+H]^+ = 330.0504$, 332.0527; found 330.0528, 332.0507.

N-(3-Chloro-5-(trifluoromethyl)phenyl)anthranilic acid (19)

General procedure B: From **19a** (65 mg, 0.197 mmol). Silica gel column chromatography afforded **19** (41 mg, 66%) as a white solid.



White solid. **RP-HPLC** (GEN1): RT = 17,49 min. R_f (hexane/EtOAc 50:50) = 0.16. ¹H-NMR (400 MHz, DMSO-d_6): 6.97 (ddd, J = 8.1, 7.2, 1.1, 1H), 7.31 (m, 1H), 7.36 (dd, J = 8.4, 1.1, 1H), 7.45-7.48 (m, 1H), 7.50 (ddd, J = 7.2, 7.0, 1.6, 1H), 7.51-7.53 (m, 1H), 7.92 (dd, J = 7.9, 1.6, 1H), 9.58 (s, 1H), 13.21 (br s, 1H). ¹³C-NMR (100 MHz, CDCl₃): 113.9 (quad, J = 3.7),

116.6, 116.9, 117.4 (quad, J = 4.0), 120.5, 121.6, 123.3 (quad, J = 273.0), 131.5 (quad, J = 32.5), 132.0, 134.2, 134.9, 143.9, 144.4, 169.2. **HRMS (ESI'):** Calc. for $[C_{14}H_9CIF_3NO_2-H]^- = 314.0201$, 316.0171; found 314.0220, 316.0161.

Methyl N-(3-bromo-5-(trifluoromethyl)phenyl)anthranilate (20a)

General procedure A: From 3-bromo-5-(trifluoromethyl)aniline (240 mg, 1.000 mmol) and methyl 2-bromobenzoate (351 μL, 2.50 mmol). Silica gel column chromatography afforded **20a** (106 mg, 32%)as a yellowish wax.



Yellowish wax. **RP-HPLC** (GEN1): RT = 25,17 min. R_f (hexane/EtOAc 85:15) = 0.40. ¹H-NMR (400 MHz, CDCI₃): 3.92 (s, 3H), 6.89 (ddd, J = 8.1, 7.1, 1.2, 1H), 7.32 (dd, J = 8.5, 1.1, 1H), 7.39 (dd, J = 1.8, 0.7, 1H), 7.43 (dddd, J = 8.4, 7.2, 1.7, 0.5, 1H), 7.55 (tt, J = 1.9, 0.5, 1H), 8.01 (ddd, J = 8.0, 1.7, 0.4, 1H), 9.64 (s, 1H). ¹³C-NMR (100 MHz, CDCI₃): 52.1, 113.8, 114.8, 115.8 (quad, J = 3.8), 119.3 (quad, J = 1.0), 121.9

(quad, J = 3.8), 123.2 (quad, J = 272.9), 123.3, 126.0, 133.1 (quad, J = 32.9), 131.9, 134.3, 143.2, 145.5, 168.7. **HRMS (ESI⁺):** Calc. for $[C_{15}H_{11}BrF_3NO_2+H]^+ = 373.9999$, 375.9978; found 374.0025, 375.9985.

N-(3-Bromo-5-(trifluoromethyl)phenyl)anthranilic acid (20)

General procedure B: **From 20a** (100 mg, 0.27 mmol) silica gel column chromatography afforded **20** (83 mg, 86%) as a white solid.



RP-HPLC (GEN1): RT = 18,23 min. R_f (hexane/EtOAc 70:30) = 0.15. ¹H-NMR (400 MHz, DMSO-d_6): 6.97 (ddd, J = 7.4, 7.3, 1.1, 1H), 7.30 (dd, J = 13.0, 1.1, 1H), 7.39-7.42 (m, 1H), 7.44 (dd, J = 3.6, 1.7, 1H), 7.45-7.48 (m, 1H), 7.60-7.63 (m, 1H), 7.92 (t, J = 2.0, 1H), 9.70 (br s, 1H). ¹³C-NMR (100 MHz, DMSO-d_6): 114.1 (quad, J = 4.3), 116.7, 119.9 (quad, J = 3.8), 120.2, 120.3, 122.9, 123.0 (quad, J = 267.7), 124.4, 131.8 (quad, J = 3.8)

32.0), 131.9, 133.8, 143.9, 144.4, 169.2. **HRMS (ESI'):** Calc. for [C₁₄H₉BRf ₃NO₂-H]⁻ = 357.9696, 359.9675; found 357.9671, 359.9658.

Methyl N-(3-(tributylstannyl)-5-(trifluoromethyl)phenyl)anthranilate (21a)

A round bottom flask was charged with LiCl (31 mg, 0.720 mmol), Pd(PPh₃)₄ (50 mg, 0.043 mmol) and **20a** (54 mg, 0.144 mmol). The flask was purged with argon and anhydrous 1,4-dioxane (1.5 mL) was added. Then, $(SnBu_3)_2$ (144 μ L, 0.288 mmol) was added and the solution was refluxed for 4 h. Crude reaction mixture was adsorbed on silica gel and purified by silica gel column chromatography (hexane/Ethyl acetate 7:3) to afford **21a** (40 mg, 50%)as an oil.



Colorless oil. R_f (hexane/EtOAc 70:30) = 0.51. ¹H-NMR (400 MHz, CDCI₃): 0.92 (t, J = 7.3, 9H), 1.22-1.42 (m, 12H), 1.59-1.70 (m, 6H), 3.93 (s, 3H), 6.86 (ddd, J = 8.0, 7.0, 1.3, 1H), 7.32-7.44 (m, 2H), 7.46 (s, 1H), 7.53 (s, 1H), 7.57 (s, 1H), 8.01 (dd, J = 8.1, 1.6, 1H), 9.71 (br s, 1H).

Methyl N-(3-iodo-5-(trifluoromethyl)phenyl)anthranilate (21b)

Compound **21a** (73 mg, 0.126 mmol) was dissolved in Acetonitrile and Nal (21 mg, 0.140 mmol) and *N*-chlorosuccinimide (17 mg, 0.140 mmol) were added. The mixture was stirred for 30 min at rt and diluted with Ethyl acetate (20 mL), water (10 mL) and 5% sodium thiosulphate solution (3



mL). Phases were separated and the aqueous phase extracted again with Ethyl acetate (10 mL). Organic layer was dried over Na_2SO_4 anhydrous, filtered and evaporated to dryness. Purification by silica gel column chromatography (hexane/ethyl acetate (85:15)) afforded **21b** (42 mg, 79%)as an oil.

e Yellowish wax. R_f (hexane/EtOAc 85:15) = 0.40. ¹H-NMR (400 MHz, CDCI₃): 3.92 (s, 3H), 6.88 (ddd, J = 8.2, 7.1, 1.1, 1H), 7.30 (dd, J = 8.5,

1.1, 1H), 7.39-7.45 (m, 2H), 7.58 (s, 1H), 7.75 (s, 1H), 8.02 (dd, J = 8.0, 1.7, 1H), 9.61 (br s, 1H). **HRMS (ESI⁺):** Calc. for [C₁₅H₁₁F₃INO₂+H]⁺ = 421.9859; found 421.9873.

N-(3-lodo-5-(trifluoromethyl)phenyl)anthranilic acid (21)

General procedure B: From **21b** (42 mg, 0.100 mmol). Silica gel column chromatography afforded **21** (17 mg, 41%) as a white solid.



White solid. R_f (hexane/EtOAc 70:30) = 0.15. **RP-HPLC** (GEN1): RT = 19,27 min.¹H-NMR (400 MHz, CDCl₃): 6.89 (ddd, J = 8.1, 7.1, 1.2, 1H), 7.24-7.27 (m, 1H), 7.35-7.41 (m, 2H), 7.51 (s, 1H), 7.70 (s, 1H), 8.01 (dd, J = 8.0, 1.6, 1H), 9.68 (br s, 1H). : ¹H-NMR (400 MHz, DMSO-d₆): 6.98 (dt, J = 7.4, 1.1, 1H), 7.32 (dd, J = 12.9, 1.0, 1H), 7.45-7.48 (m, 1H), 7.48

(dd, J = 3.6, 1.7, 1H), 7.52-7.55 (m, 1H), 7.77-7.80 (m, 1H), 7.94 (t, J = 2.0, 1H), 9.67 (br s, 1H). ¹³**C**-**NMR (100 MHz, DMSO-d_6):** 95.9, 114.7 (quad, J = 4.1), 116.4, 120.4, 123.0 (quad, J = 272.9), 125.8 (quad, J = 3.7), 130.6, 131.8 (quad, J = 32.1), 131.9, 134.0, 143.9, 144.0, 169.2. **HRMS (ESI):** Calc. for [C₁₄H₉F₃INO₂-H] = 405.9557; found 405.9558.

DIFLUNISAL SERIES



3-fluoro-5-iodosalicylic acid (15b)

To a stirred solution of 3-fluorosalicylic acid (**15a**) (158 mg, 1.000 mmol) in CH_2Cl_2 (15 mL) was added, in small portions, IPy_2BF_4 (409 mg, 1.100 mmol). The mixture was stirred at rt for 2 h and then diluted with ethyl acetate (50 mL), washed with 0.5 M HCl (2 x 10 mL), with brine (15 mL) and dried over anhydrous Na₂SO₄, filtered and evaporated to dryness. The residue was further purified by silica gel column chromatography ($CH_2Cl_2/MeOH/AcOH$ (90:10:0.5)) to afford pure **15b** (264 mg, 94%) as a white solid.



2',4',5-trifluoro-4-hydroxy-[1,1'-biphenyl]-3-carboxylic acid (15)

Under Ar atmosphere degassed water (2.5 mL) was added over a mixture of 2,4difluorophenyl boronic acid (45 mg, 0.285 mmol), sodium carbonate (82 mg, 0.774 mmol), Pd(OAc)₂ (0.6 mg, 1 mol%) and compound **15b** (73 mg, 0.258 mmol). The suspension was stirred at rt for 3 h and then diluted with ethyl acetate (50 mL) and water (10 mL). The pH was adjusted to 2-3 with 0.5 M HCl and the phases separated. The aqueous layer was back-extracted with more ethyl acetate (2x10 mL) and the combined organic phases dried over Na₂SO₄ anhydrous, filtered and evaporated to dryness. The residue was purified by silica gel column chromatography (CH₂Cl₂/ethyl acetate/AcOH (50:50:0.5)) to afford pure **15** (71 mg, 100%).



White solid. R_f (CH₂Cl₂/MeOH 85:15) = 0.13. **RP-HPLC** (GEN1): RT = 10,27 min; ¹H-NMR (400 MHz, DMSO-d₆) δ (ppm): 7.19 (tdd, J = 8.6, 2.6, 1.0, 1H), 7.37 (ddd, J = 11.6, 9.3, 2.6, 1H), 7.62 (td, J = 9.0, 6.5, 1H), 7.70 (ddd, J = 11.9, 2.3, 1.3, 1H), 7.74 (dt, J = 2.5, 1.4, 1H). ¹³C- NMR (100 MHz, DMSO-d₆) δ (ppm): 104.6 (t, J = 26.0), 112.2 (dd, J = 21.2, 3.8), 115.8 (d, J = 3.9), 121.3 (dd, J = 19.3, 2.4), 122.5-123.1 (m), 124.4

(d, J = 6.7), 125.5 (d, J = 3.0), 131.7 (dd, J = 9.7, 4.4), 149.0 (d, J = 13.1), 150.7 (d, J = 243.6), 159.1 (dd, J = 248.7, 15.5), 161.7 (dd, 260.0, 12.3), 170.9 (d, J = 3.6). **HRMS (ESI'):** Calc. for [C₁₃H₇F₃O₃-H]⁻ = 268.0347; found 267.0295.

5-chloro-2',4'-difluoro-4-hydroxy-[1,1'-biphenyl]-3-carboxylic acid (16)

To a stirred solution of Diflunisal (50 mg, 0.200 mmol) in acetonitrile (2 ml) was added a solution of *N*-chlorosuccinimide (56 mg, 0.420 mmol) in acetonitrile (1.5 mL). Next, 1 drop of HCI (c) was added and the mixture was stirred for 1 h at rt The mixture was then diluted with CH_2Cl_2 (20 mL), washed with 5% solution of Na_2SO_3 (5 mL) and brine (5 mL) and dried over anhydrous Na_2SO_4 ,

filtered and evaporated to dryness. Further purification by silica gel column chromatography (CH₂Cl₂/MeOH/AcOH (95:5:0.5)) afforded pure **16** (27 mg, 47%) as a white solid.



White solid. R_f (CH₂Cl₂/MeOH/AcOH 95:5:0.5) = 0.11. **RP-HPLC** (GEN1): RT = 12,46 min.¹**H-NMR** (400 MHz, CDCl₃/CD₃OD 95:5) δ (ppm): 6.86-7.00 (m, 2H), 7.31-7.42 (m, 1H), 7.71 (s, 1H), 7.96 (s, 1H), 9.14 (br s, 1H). ¹³C-NMR (100 MHz, CDCl₃/CD₃OD 95:5) δ (ppm): 104.5 (dd, J =25.4, 26.5), 111.7 (dd, J = 21.2, 3.8), 113.9, 122.1, 123.1 (dd, J = 13.6, 4.0), 126.2, 129.3 (d, J = 2.6), 131.0 (dd, J = 9.5, 4.6), 135.9 (d, J = 3.2),

157.0, 159.6 (dd, J = 250.4, 11.9), 162.4 (dd, J = 249.7, 11.8), 178.0. **HRMS (ESI'):** Calc. for $[C_{13}H_7CIF_2O_3-H]^- = 282.9979$; found 282.998.

5-bromo-2',4'-difluoro-4-hydroxy-[1,1'-biphenyl]-3-carboxylic acid (17)

To 600 mg of the polymer bound brominating agent (pyridine hydrobromideperbromide polymer-bound, PVPHP, ~1 mmol Br₃ per g of resin, 3 equiv) in CH_2Cl_2 (5 ml) a solution of Diflunisal (50 mg, 0.200 mmol) in CH_2Cl_2 (1 ml) was added and the mixture was shaken o/n. After removing the resin, the filtrate was diluted with more CH_2Cl_2 (20 mL), and washed with 5% aq. solution of Na₂SO₃ (5 mL) and brine (5 mL) and dried over anhydrous Na₂SO₄, filtered and evaporated to dryness. Further purification by silica gel column chromatography ($CH_2Cl_2/MeOH/AcOH$ (95:5:0.5)) afforded pure **17** (34 mg, 52%) as a white solid.



RP-HPLC (GEN1): RT = 13,32 min. ¹**H-NMR** (400 MHz, DMSO-d₆) δ (ppm): 7.17 (ddd, J = 8.5, 8.4, 2.7, 1H), 7.35 (ddd, J = 11.6, 9.3, 2.7, 1H), 7.61 (ddd, J = 9.0, 9.0, 6.6, 1H), 7.92 (t, J = 1.9, 1H), 7.97 (t, J = 1.9, 1H). ¹³C-NMR (100 MHz, DMSO-d₆) δ (ppm): 104.6 (dd, J = 26.9, 25.9), 110.6, 112.2 (dd, J = 21.1, 3.7), 114.5, 122.6 (dd, J = 13.3, 3.7), 126.1,

129.8 (d, J = 3.5), 131.8 (dd, J = 9.7, 4.4), 138.2 (d, J = 2.3), 157.4, 159.0 (dd, J = 248.4, 11.6), 161.8 (dd, J = 259.7, 11.6), 171.3. **HRMS (ESI'):** Calc. for [C₁₃H₇BrF₂O₃-H] = 326.9473, 328.9453; found 326.9463, 328.9445.

2',4'-difluoro-4-hydroxy-5-iodo-[1,1'-biphenyl]-3-carboxylic acid or lododiflunisal(13)

To a solution of 200 mg (0.80 mmol) of diflunisal in 5 ml of dichloromethane, 357 mg (1.2 mmol) of IPy_2BF_4 were added to obtain a substrate/iodinating reagent ratio of (1:1.5) equivalents. The reaction was left under stirring at rt and monitored by HPLC until full conversion to the iodo derivative.



After 1,5 h the solution was diluted with dicloromethane and the organic phase was successively washed with HCl 1N and a 0.1N sodium thiosulfate solution. The organic phase was dried over MgSO₄ and after evaporation yielded a residue (> 98% purity) which was further purified by column chromatography on silica gel using Cl₃CH/MeOH gradient mixture or HPLC

White solid.**HPLC (GEN1) RT:** 14.32 min; ¹**H-NMR (500 MHz; DMSO-d₆)** δ (ppm): 7.12-7.16 (m, 1H). 7.27-7.31 (m, 1H), 7.57 (ddd, J = 9.0, 9.0, 6.5, 1H), 7.92-7.93 (m, 1H), 8.09-8.10 (m, 1H). ¹³**C-NMR (125.7 MHz; DMSO-d₆)** δ (ppm): 171.2, 161.6 (dd, J = 12.6, 247.6), 159.7, 158.9 (dd, J = 12.3, 248.5), 144.1, 131.5 (dd, J = 4.5, 9.5), 130.4, 126.8, 122.4 (dd, J = 3.6, 13.3), 113.2, 111.9 (d, J = 20.9), 104.3 (t, J = 26.4), 85.9. **HRMS (ESI⁻¹):** Calc. for [C13H7F2IO3-1] = 374,9408; found 374,9306.

Selected HPLC, NMR and HRMS data of target compounds:









Retention Time (min)



S26









Single Mass Analysis

Tolerance = 10.0 mDa / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3

Compound 19a

1: TOF MS ES+

Monoisotopic Mass, Even Electron Ions 45 formula(e) evaluated with 1 results within limits (up to 50 best isotopic matches for each mass) Elements Used: C: 0-15 H: 0-12 N: 0-1 O: 0-2 F: 0-3 CI: 0-1 LB025A 28071117 34 (0.413)









Elemental Composition Report



1: TOF MS ES-

Tolerance = 10.0 mDa / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3



Chemical Formula: C₁₄H₉ClF₃NO₂ Exact Mass: 315,0274

Monoisotopic Mass, Even Electron Ions 55 formula(e) evaluated with 1 results within limits (up to 50 best isotopic matches for each mass) Elements Used: C: 10-14 H: 5-10 N: 0-1 O: 0-2 F: 0-3 CI: 0-2 LB032A

28071115 35 (0.424)











Solvent B: ACN






Tolerance = 20.0 mDa / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3

Compound 20a

1: TOF MS ES+

Chemical Formula: C₁₅H₁₁BrF₃NO₂ Exact Mass: 372,9925

Monoisotopic Mass, Even Electron Ions

185 formula(e) evaluated with 2 results within limits (up to 50 best isotopic matches for each mass) Elements Used: C: 0-15 H: 11-12 N: 0-1 O: 0-2 F: 0-3 79Br: 0-8 81Br: 0-8







Tolerance = 10.0 mDa / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3 F₃C N COOH

Monoisotopic Mass, Even Electron Ions 137 formula(e) evaluated with 2 results within limits (up to 50 best isotopic matches for each mass) Elements Used: C: 0-15 H: 8-8 N: 0-1 O: 0-2 F: 0-3 79Br: 0-8 81Br: 0-8

Compound 20

Chemical Formula: C₁₄H₉BrF₃NO₂ Exact Mass: 358,9769

2: TOF MS ES-

Fia Gemma Br_CF3_ANTH_Acid 4 (0.208)





Tolerance = 10.0 mDa / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3 Compound 20

Chemical Formula: C₁₄H₉BrF₃NO₂ Exact Mass: 358,9769

1: TOF MS ES+

Monoisotopic Mass, Even Electron Ions 137 formula(e) evaluated with 2 results within limits (up to 50 best isotopic matches for each mass) Elements Used: C: 0-15 H: 10-10 N: 0-1 O: 0-2 F: 0-3 79Br: 0-8 81Br: 0-8

Fia Gemma

Br_CF3_ANTH_Acid 9 (0.408)







F₃C N COOH

Compound 20















Pump A Type: L-6200 Solvent A: Agua

Solvent B: ACN



Elemental Composition Report

Single Mass Analysis

Tolerance = 10.0 mDa / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions 43 formula(e) evaluated with 1 results within limits (up to 50 best isotopic matches for each mass) Elements Used: C: 10-14 H: 5-10 N: 0-1 O: 0-2 F: 0-3 127I: 0-1

LB038A

28071128 43 (0.524)





Compound 21

Chemical Formula: C₁₄H₉F₃INO₂ Exact Mass: 406,9630

1: TOF MS ES-



Single Mass Analysis

Tolerance = 20.0 mDa / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

45 formula(e) evaluated with 1 results within limits (up to 50 best isotopic matches for each mass) Elements Used: C: 0-14 H: 0-8 N: 0-1 O: 0-2 F: 0-3 I: 0-1



н

ĊOOH

 F_3





S53



S54





Pump A Type: L-6200 Solvent A: Agua





Elemental Composition Report

Tolerance = 10.0 mDa / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions 5 formula(e) evaluated with 1 results within limits (up to 50 best isotopic matches for each mass) Elements Used: C: 10-13 H: 5-8 O: 3-5 F: 0-3 LB031A 28071114 36 (0.434)



Compound 15

Chemical Formula: C₁₃H₇F₃O₃ Exact Mass: 268,0347

> 1: TOF MS ES-1.36e+005











COOH

C: 0-7 H: 0-4 O: 0-3 F: 0-1 I: 0-1

LB028A









S65



COOH

OH

Single Mass Analysis

Calc. Mass

282.9974

Mass

282,9976

S66

i-FIT

51.4

Formula

C13 H6 O3 F2 C1

DBE

9.5

PPM

0.7

mDa

0.2







Tolerance = 10.0 mDa / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions 229 formula(e) evaluated with 2 results within limits (up to 50 best isotopic matches for each mass) Elements Used: C: 0-14 H: 0-10 O: 0-3 F: 0-2 79Br: 0-8 81Br: 0-8



Compound 17

Chemical Formula: C₁₃H₇BrF₂O₃ Exact Mass: 327,9547

1: TOF MS ES-



30111204 58 (0.723)





Element prediction: Off Number of isotope peaks used for i-FIT = 3 Monoisotopic Mass, Even Electron Ions 18 formula(e) evaluated with 1 results within limits (up to 50 best isotopic matches for each mass) Elements Used: C: 0-15 H: 0-10 O: 0.3 E: 0.2 I: 0-1							F I Compound 13
							Chemical Formula: C ₁₃ H ₇ F ₂ IO ₃ Exact Mass: 375,9408
0. 0-13 11. 0-10	0.0-5 1.0-	2 1.0-1					
Fia Gemma IDIF 8 (0.386)							2: TOF MS ES- 3.08e+005
100	374.9306						
% 	375.9	9380	770 0	EOE 040 0470			
0-1111111111111111111111111111111111111	376	6.9385 <u>632</u>	2.9634	595 840.8478	1164	.7693 <u>1367.2454</u> 1	1762.6979 1762.1752 m/z
100 200	300 400	500 600) 700 8	800 900	1000 1100	1200 1300 1400 150	00 1600 1700 1800
Minimum: Maximum:		20.0	10.0	-1.5 50.0			
Mass Cal	c. Mass	mDa	PPM	DBE	i-FIT	Formula	
374.9306 374	.9330	-2.4	-6.4	9.5	112.0	C13 H6 O3 F2 I	

S72

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Single Mass Analysis Tolerance = 20.0 mDa / DBF: min = -1.5, max = 50.0