

Supporting information

Table S1 Plasmids used in the study

| Plasmid | Description | Reference |
|------------------|--|-----------|
| pZElac | <i>E. coli</i> vector, amp ^R | 1 |
| pZElac-his | The pZElac inserted with the T5 promoter/lacO::6Xhis, amp ^R | 2 |
| pZElac_GDH | pZElac inserted with <i>E.coli gdhA</i> wild type, amp ^R | This work |
| pZElac_GDH F18 | pZElac inserted with <i>E.coli gdhA</i> F18 mutant, amp ^R | This work |
| pZElac_GDH F18TA | pZElac inserted with <i>E.coli gdhA</i> F18 with T195A mutation, amp ^R | This work |
| pZElac_GDH F18TV | pZElac inserted with <i>E.coli gdhA</i> F18 with T195V mutation, amp ^R | This work |
| pZElac_GDH F18TL | pZElac inserted with <i>E.coli gdhA</i> F18 with T195L mutation, amp ^R | This work |
| pZElac_GDH F18TF | pZElac inserted with <i>E.coli gdhA</i> F18 with T195F mutation, amp ^R | This work |
| pZElac_aspC | pZElac inserted with <i>E.coli aspC</i> , amp ^R | This work |
| pZElac_tyrB | pZElac inserted with <i>E.coli tyrB</i> , amp ^R | This work |
| pZElac_pheDH | pZElac inserted with <i>Thermoactinomyces intermedius pdh</i> , amp ^R | This work |
| pZElac_hisGDH | for his tag- <i>E.coli</i> GDH expression, amp ^R | This work |
| pZElac_hisF18 | for his tag- <i>E.coli</i> GDH F18 mutant expression, amp ^R | This work |
| pZElac_hisF18TA | for his tag- <i>E.coli</i> GDH F18 T195A expression, amp ^R | This work |
| KAVS library | pZElac vector harboring <i>E. coli gdhA</i> with K92, A166, V377 and S380 residues randomized by NNK codon | This work |

Reference:

1. Zhang, K., Li, H., Cho, K. M., Liao, J. C. 2010. Expanding metabolism for total biosynthesis of the nonnatural amino acid L-homoalanine. *Proc Natl Acad Sci U S A* 107: 6234-6239
2. Li, H., Liao, J. C. 2013. Engineering a cyanobacterium as the catalyst for the photosynthetic conversion of CO₂ to 1,2-propanediol. *Microb Cell Fact* 12: 4

Table S2 Primers used in the study

| Primer | sequence (5'-3') |
|------------------|---|
| GDH Acc65I fwd | agaaaggattaccatggatcagacatattctctggagtc |
| GDH XbaI rev | gatgcctctagattaaatcacaccctgcgccagcatcg |
| aspC Acc65I fwd | gagaaaggattaccatgttggagaacattaccggccgtc |
| aspC XbaI rev | gatgcctctagattacagcaactgcccacaatcgctcg |
| tyrB Acc65I fwd | gagaaaggattaccatgttcaaaaagttgacgcctacg |
| tyrB XbaI rev | gatgcctctagattacatcaccgcagcaaacgcctttgc |
| PheDH Acc65I fwd | gagaaaggattaccatgataagaatgggaaggcatgaaaatc |
| PheDH XbaI rev | atgcctctagattacccttcgtcgctgttgcgggg |
| T195A fwd | caacaataccgcctgcgttgcgggttaagggccttcattggcg |
| T195A rev | cgc当地aaatgaaaggcccttacccgcgaagacgcaggcggatttttg |
| T195L fwd | caacaataccgcctgcgttctcgtggtaagggccttcattggcg |
| T195L rev | cgc当地aaatgaaaggcccttacccaggaagacgcaggcggatttttg |
| T195V fwd | caacaataccgcctgcgttctcgtggtaagggccttcattggcg |
| T195V rev | cgc当地aaatgaaaggcccttacccacgaagacgcaggcggatttttg |
| T195F fwd | caacaataccgcctgcgttctcgtggtaagggccttcattggcg |
| T195F rev | cgc当地aaatgaaaggcccttacccgaagaagacgcaggcggatttttg |
| K92 fwd | gcagttcagctctgcatcgccccgtacnnkggcgatgcgttccatccgtcagttaa |
| K92 rev | ttaactgacggatggaagcgcataccgcnnngtacggccgtatggcagagctgaactgc |
| A166 fwd | gccacctggcgccgataccgcacgtccgnnkggatatcgggttggcgtgaag |
| A166 rev | cttcacgaccaccaaccccgatataccmnncgaaacgtcggtatccgcgcccaggcggc |
| VS fwd | gtaaaggcggtaaatgctggcnnkgtacannkggctggaaatggcacaacacgtg |
| VS rev | cagcgtttgtgcattccaggccmnntgtagcmnngccaccagattccgccttac |

Cofactor specificity of GDH variants

E. coli GDH is naturally NADPH-dependent. To determine the cofactor specificity of the engineered GDH variants toward the final target 2-OPBA, enzyme assays were performed using purified proteins (Figure S1). The results suggested that both GDH F18 and F18 T195A are highly NADPH-specific for this reaction. The NADH-dependent activity was very minimal.

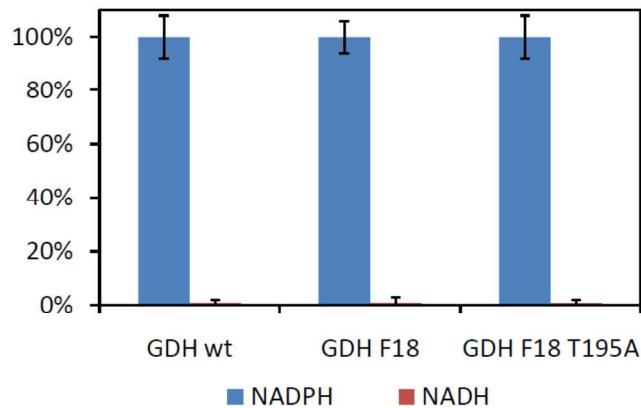


Figure S1 Cofactor specificity of GDH variants. 2-OPBA was the keto acid substrate used in the enzyme assays. The error bars represent the standard deviation of three repeats. (n=3)

Performance of the second generation GDH variant, F18 T195A, in keto acid feeding experiments

To test the enzyme's performance in converting the keto acid 2-OPBA to L-homophenylalanine inside the cells, GDH F18 T195A was overexpressed in *E. coli* and the keto acid feeding experiments were performed as described in the “methods” session. The results showed that F18 T195A produced higher level of L-homophenylalanine than the first generation variant F18. The increase was relatively small (~10%) but significant.

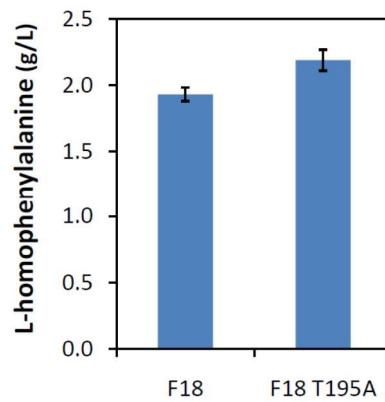


Figure S2 Performance of F18 T195A GDH variant in the keto acid feeding experiments. The error bars represent the standard deviation of three repeats. (n=3)