

Supporting Information

Solid-Phase Synthesis of Peptide Thioureas and Thiazole-Containing Macrocycles through Ru-Catalyzed Ring-Closing Metathesis

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Abbreviations

Boc: *N*-*tert*-butoxycarbonyl; HATU: *N*-[(dimethylamino)-1*H*-1,2,3-triazolo-[4,5-*b*]pyridin-1-ylmethylene]-*N*-methylmethanaminium hexafluorophosphate *N*-oxide; HOAt: 1-hydroxy-7-azabenzotriazole; HMBA: 4-hydroxymethylbenzoic acid; MeIm: 1-Methylimidazole; MSNT: 1-(mesitylene-2-sulfonyl)-3-nitro-1,2,4-triazole; NEM: *N*-ethyl morpholine; Pbf: 2,2,4,6,7-penta-methyldihydrobenzofuran-5-sulfonyl; PEGA: Polyethylene glycol dimethyl acrylamide; TBTU: *O*-(benzotriazol-1-yl)-*N,N,N',N'*-tetramethyluronium tetrafluoroborate; Trt: Trityl

General Methods

All reagents used were commercially available and all solvents were of HPLC grade. Solid-phase synthesis was carried out using syringe techniques in flat-bottomed PE-syringes fitted with PPE filters. The syringes were placed in a special setup equipped with Teflon tubing and Teflon valves, which allow suction to be applied to the syringes. Products were analyzed using a Waters Alliance 2695 HPLC system equipped with a Symmetry® C-18 column 3.5 μ m, 4.6 x 75 mm, column temp; 25 °C, (1 ml/min.) with detection at 215 nm and 254 nm using a diode array detector. Eluent A (0.1% TFA in H₂O) and eluent B (0.1% TFA in acetonitrile) were used with a linear gradient (100% A to 100% B) with a run time of 13 min. Analytical LC-MS analysis was performed on a Waters AQUITY UPLC system equipped with PDA and SQD MS detector; column: AQUITY UPLC BEH C18 1.7 μ m, 2.1 x 50mm; column temp: 65 °C; solvent A: 0.1% formic acid (aq); solvent B: 0.1% formic acid (acetonitrile); gradient: 5% B to 100% B in 2.4 min, hold for 0.1 min, total run-time ca. 2.6 min. LC-DAD-HRMS was performed on a Agilent 1100 LC system with a diode array detector (Agilent Technologies, Waldbronn, Germany) equipped with a 50 mm x 2 mm i. d. 3 μ m Luna C₁₈ (2) column (Phenomenex, Torrance, CA, USA). Separation was achieved using a linear reverse phase gradient system at 40 °C and a flow rate of 400 μ L/min. The gradient was started at 20% acetonitrile, which was increased to 100% in 8 min (holding this level for 2 min), then reverted to 20% in 1 min (holding this for 4 min.). Both water and acetonitrile contained 20mM formic acid. The LC was coupled to a Micromass LCT orthogonal time-of-flight mass spectrometer equipped with Lock Mass probe and operated in positive electrospray mode. All compounds were analyzed by UPLC-MS/HRMS and ¹H NMR. Compound sufficient for NMR analysis was obtained by cleaving a resin sample of ca. 100 mg. ¹H NMR 300 Mhz were recorded on a Varian Mercury 300 BB spectrometer at room temperature. ¹H NMR 500 MHz spectra were recorded using a Varian Unity Inova-500 spectrometer. All NMR spectra were recorded using CDCl₃ or DMSO-*d*₆.

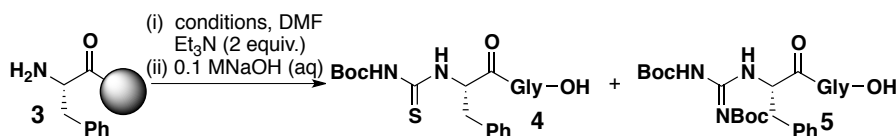
Solid-Phase Synthesis

Attachment of the HMBA linker to the amino-functionalized ChemMatrix (0.6 mmol/g) resin was carried out by premixing TBTU (2.88 equiv.), NEM (4 equiv.) and HMBA (3 equiv.) for 5 min. in DMF. The solution was then added to the preswollen resin in DMF and allowed to react for 2 hours followed by washing with DMF (x 6). Coupling of the first amino acid to the HMBA handle was carried out by premixing the Fmoc-AA-OH (3 equiv.), MSNT (3 equiv.) and MeIm (2.25 equiv.) in dry CH₂Cl₂. The solution was added to the preswollen resin in dry CH₂Cl₂ and allowed to react for 1 hour. The resin was washed with dry CH₂Cl₂ (x 3) and the coupling was repeated once. The resin was washed with DMF (x 6) after the coupling. Peptide synthesis was carried out as described for the attachment of HMBA to ChemMatrix. Fmoc-deprotection was accomplished with treatment of the resin with 20% piperidine (DMF) for 2 min. and then 18 min, followed by washing with DMF (x 6). Prior to thiourea formation, the deprotected resin was washed with DMF (x 6), CH₂Cl₂ (x 6) and dry CH₂Cl₂ (x 3). Formation of the resin-bound Boc-protected thioureas was carried out by mixing *N,N'*-di-Boc-thiourea (2 equiv.), Et₃N (3 equiv.) and Mukaiyama's reagent (2 equiv.) in dry CH₂Cl₂ for 15 min before filtering off any solids. The filtrate was added to the preswollen resin in dry CH₂Cl₂ and allowed to react for 2 hours at room temperature followed by washing of the resin with DMF (x 6) and CH₂Cl₂ (x 6). Prior to guanylation, the deprotected resin was washed with DMF (x 6), CH₂Cl₂ (x 6), dry CH₂Cl₂ (x 3) and dry DMF (x 6). Formation of the resin-bound bis(Boc)-protected guanidines was achieved by adding *N,N'*-di-Boc-thiourea (1.5 equiv.) and Et₃N (9 equiv.) in dry DMF to the preswollen resin in dry DMF for 5 min. HgCl₂ (3 equiv.) in dry DMF was added to the resin and allowed to react for 2 hours at room temperature followed by washing of the resin with DMF (x 6), 20% piperidine (DMF) (x 6), DMF (x 6) and CH₂Cl₂ (x 6). Peptides with Trp(Boc), Arg(Pfp), Asn(Trt) or Gln(Trt) as the last residue needed to be subjected to the guanylation conditions twice to achieve full conversion. Deprotection of resin-bound thioureas (except tryptophan) was performed by adding 0.1 M SnCl₄ (dry CH₂Cl₂) to resin for 2 x 30 min, followed by washing with CH₂Cl₂ (x 6), 20% piperidine

(DMF) ($\times 6$), DMF ($\times 6$) and CH_2Cl_2 ($\times 6$). Deprotection of resin-bound guanidines (and thiourea-modified tryptophan derivatives) was performed by adding 0.25 M SnCl_4 (dry EtOAc) to the resin for 3 $\times$ 2 hours, followed by washing with CH_2Cl_2 ($\times 6$), 20% piperidine (DMF) ($\times 6$), DMF ($\times 6$) and CH_2Cl_2 ($\times 6$). Synthesis of solid supported thiazoles was achieved by adding the α -halo ketones or masked aldehydes (5 equiv.) in EtOH/ H_2O 3:1 to the solid supported deprotected thiourea in a glass vial. The vials were sealed and heated to 60 $^\circ\text{C}$ for 16 hours on a shaker, after which the solution was cooled to room temperature. The resin was transferred to plastic syringes fitted with PPE filters and washed with DMF ($\times 6$) and CH_2Cl_2 ($\times 6$). Coupling of alkene functionalized amines to **25** was achieved by adding HATU (2 equiv.), HOAt (1 equiv.) and NEM (2 equiv.) in DMF to **25** for 5 min followed by addition of the alkene functionalized amine HCl salt (4 equiv.) and NEM (4 equiv.) in DMF. The reaction was allowed to proceed for 1 hour. The resin was washed with DMF ($\times 3$) and the coupling repeated. The on-bead ring closing metathesis was performed by adding 30 mol% of the metathesis catalysts in dry degassed CH_2Cl_2 to give a final concentration of 0.02 M. The syringes were sealed and placed on a shaker for 24 hours at rt. The resin was washed with DMF ($\times 3$), CH_2Cl_2 ($\times 3$) and dry degassed CH_2Cl_2 ($\times 3$) before the addition of another portion of catalysts. The syringes were sealed again and placed on the shaker for additional 24 hours. The resin was washed with DMF ($\times 3$), CH_2Cl_2 ($\times 3$), 10% TFA (aq) (3 $\times$ 10 min), CH_2Cl_2 ($\times 3$), 20% piperidine (DMF) (3 $\times$ 10 min), DMF ($\times 6$), MeOH ($\times 6$) and CH_2Cl_2 ($\times 3$). Release of products from the HMBA functionalized resin for HPLC analysis was achieved by treatment with aqueous NaOH (0.1 M) for 2 hours, followed by neutralization with aqueous HCl (0.1 M) and washing with MeCN ($\times 4$) and CH_2Cl_2 ($\times 2$). Release of preparative amounts from 100-200 mg resin was achieved by treatment with aqueous NaOH (0.1 M) for 24 hours followed by neutralization with aqueous HCl (0.1 M) and washings with MeCN ($\times 4$) and CH_2Cl_2 ($\times 2$).

Screening of Reaction Conditions

Table S1. Screening of reaction conditions for the solid-phase synthesis of phenylalanine-derived thiourea and guanidine derivatives.



entry	conditions	conversion (%) ^a	product distribution (%) ^a	
			4	5
1	9 (2 equiv.), 24 h.	0	-	-
2	10 (2 equiv.), 2 h.	25	-	100
3	10 (2 equiv.), 4 h.	80	-	100
4	10 (2 equiv.), 8 h.	80	-	100
5	NIS (1 equiv.), 1 (1 equiv.), 4 h.	87	83	17
6 ^b	CuCl ₂ (2 equiv.), 1 (2 equiv.), 2 h. ^c	69	34	66
7 ^b	CuCl ₂ (2 equiv.), 1 (2 equiv.), 2 h. ^d	84	79	21

^a The conversion and product distribution was determined by RP-HPLC (215 nm). ^b 3 equiv. of Et₃N. ^c Drop-wise addition of CuCl₂ to a mixture of **3**, **1** and Et₃N in dry DMF. ^d A mixture of CuCl₂, **1** and Et₃N in DMF was allowed to react for 15 min before being added to the resin.

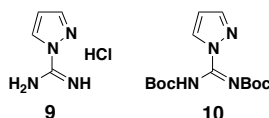
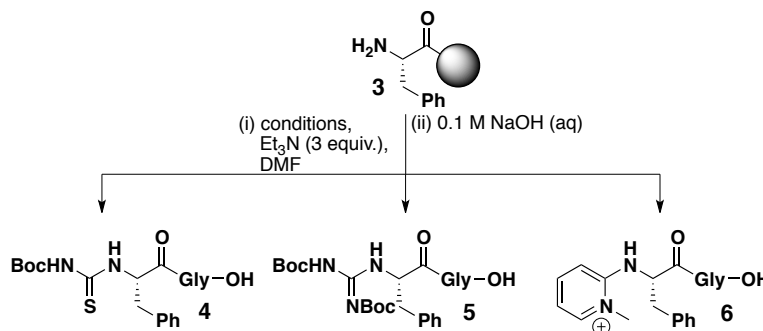


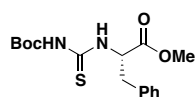
Table S2. Screening of reaction conditions for the HgO-mediated guanylation of solid-supported phenylalanine derivative.



entry	conditions	conversion (%) ^a	product distribution (%) ^a		
			4	5	6
1	HgO (2 equiv.), 1 (2 equiv.), 2 h.	33	0	100	0
2	HgO (5 equiv.), 1 (2 equiv.), 4 h. ^b	95	0	86	14
3	HgO (1.5 equiv.), 1 (1.5 equiv.), 4 h.	87	4	85	11
4	HgO (1.5 equiv.), 1 (1.5 equiv.), 7 h. ^b	89	7	77	16
5	HgO (1.5 equiv.), 1 (1.5 equiv.), 2x2 h.	>95	0	100	0

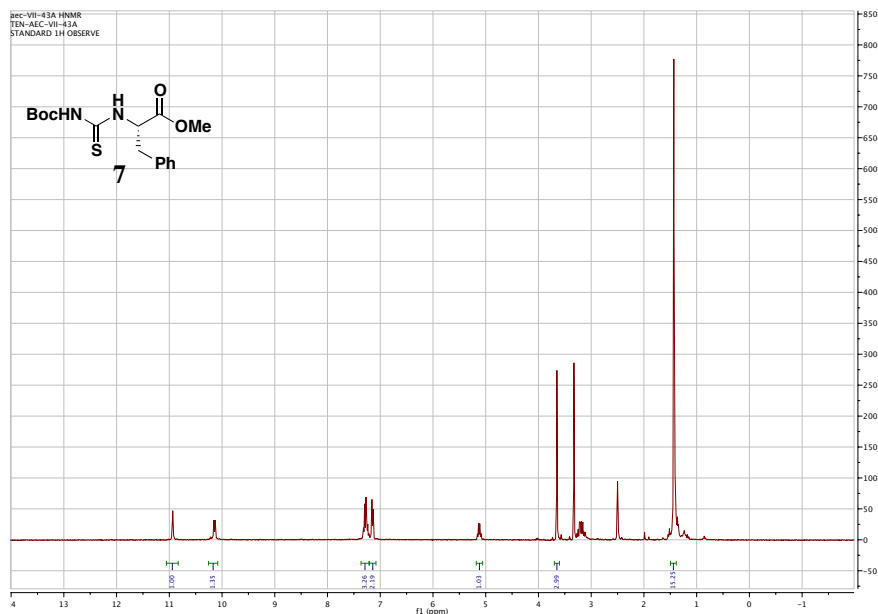
^a The conversion and product distribution was determined by RP-HPLC (215 nm). ^b Overall product purity was measured to be <30%. The product distribution shows the distribution of identified products.

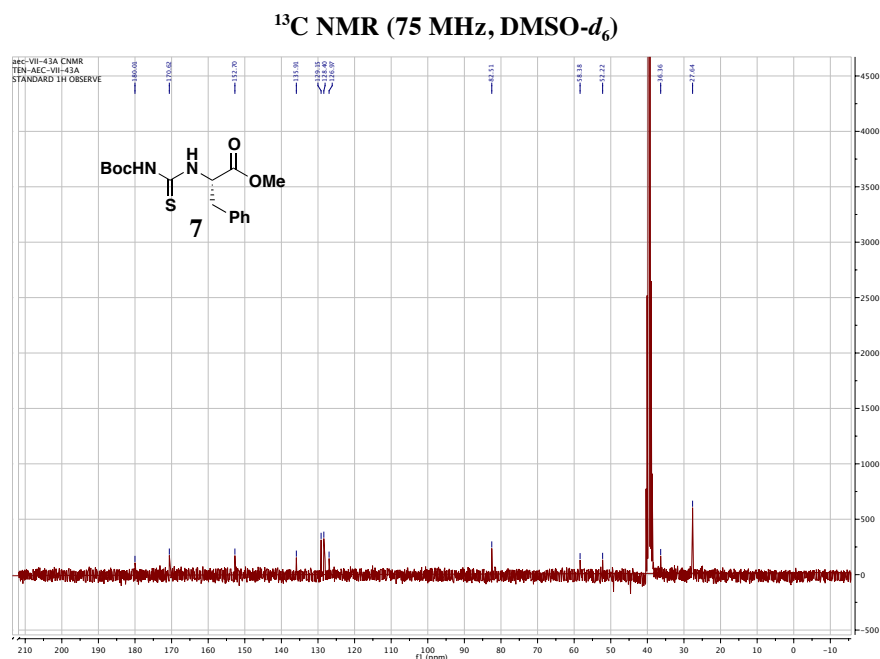
(S)-Methyl 2-(3-(Boc)thioureido)-3-phenylpropanoate (7)



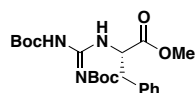
N,N'-di-Boc-thiourea (0.13 g, 0.46 mmol), Et₃N (96 μ l, 0.70 mmol) and Mukaiyama's reagent (0.12 g, 0.46 mmol) were stirred for 15 min in dry CH₂Cl₂ (1.3 ml), before filtering off any solids. The filtrate was added to a solution of (S)-H-Phe-OMe HCl (0.05 g, 0.23 mmol) in dry CH₂Cl₂ (1 ml) in a flame dried flask under argon. The reaction mixture was stirred at room temperature for 5 h., then concentrated *in vacuo* and the product purified by flash chromatography (EtOAc/heptane 1:3) to give **7** as a clear oil (8.4 mg, 11%). ¹H NMR (300 MHz, DMSO-*d*₆): δ 10.94 (1H, s), 10.14 (1H, d, *J* = 7.3 Hz), 7.42 – 7.07 (5H, m), 5.12 (1H, q, *J* = 6.6 Hz), 3.65 (3H, s), 3.24 (1H, dd, *J* = 6.3 Hz, 13.6 Hz), 3.15 (1H, dd, *J* = 6.8 Hz, 13.9 Hz), 1.43 (9H, s). ¹³C NMR (75 MHz, DMSO-*d*₆) δ 180.0, 170.6, 152.7, 135.9, 129.2 (2C), 128.4 (2C), 127.0, 82.5, 58.4, 52.2, 36.4, 27.6 (3C). HRMS (ESI) calcd for C₁₆H₂₃N₂O₄S [M + H]⁺ 339.1373, found 339.1397.

¹H NMR (300 MHz, DMSO-*d*₆)





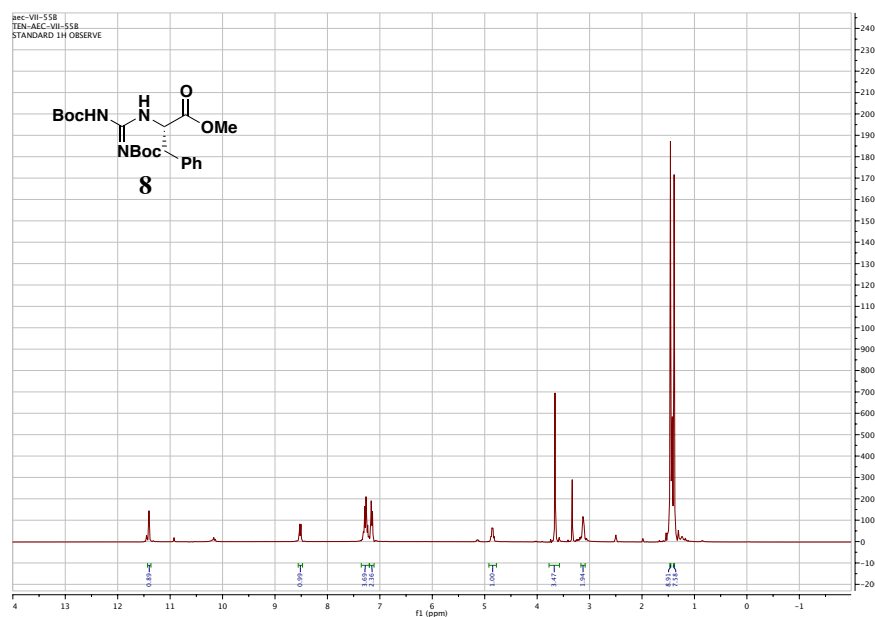
(S)-Methyl 2-(2,3-bis(Boc)guanidino)-3-phenylpropanoate (8**)**



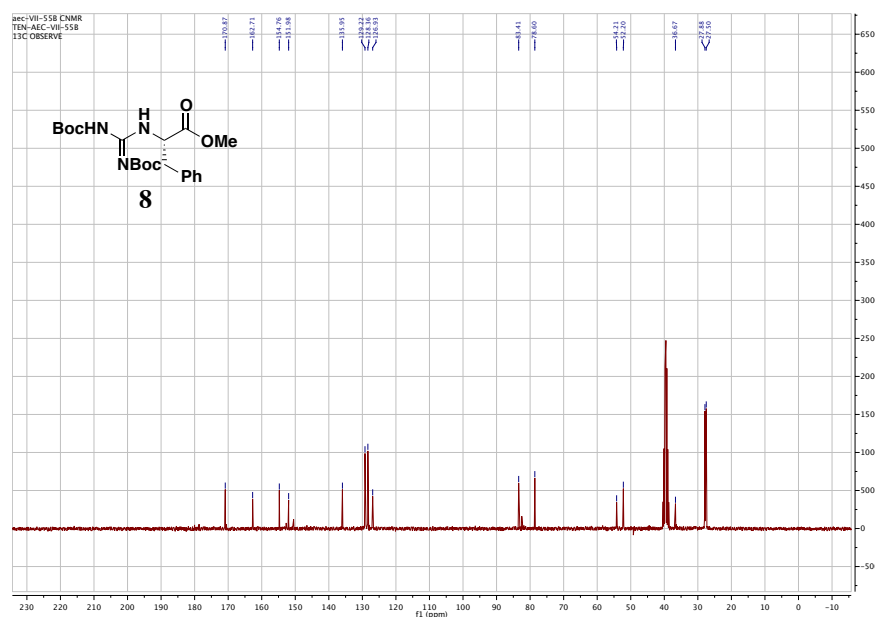
(S)-H-Phe-OMe HCl (0.05 g, 0.23 mmol), *N,N'*-di-Boc-thiourea (0.13 g, 0.46 mmol) and Et_3N (96 μl , 0.70 mmol) were added to dry DMF (1 ml) in a flame dried flask under argon. Mukaiyama's reagent (0.12 g, 0.46 mmol) in dry DMF (1.3 ml) was added drop-wise to the resulting solution. The reaction mixture was stirred at room temperature for 5 h. The reaction mixture was concentrated *in vacuo* and the product purified by flash chromatography (EtOAc/heptane 1:3) to give **8** as a clear oil (58.3 mg, 60%). ^1H NMR (300 MHz, DMSO- d_6): δ 11.40 (1H, s), 8.52 (1H, d, $J = 7.5$ Hz), 7.44 – 7.06 (5H, m), 4.85 (1H, q, $J = 6.8$ Hz), 3.66 (3H, s), 3.16 – 3.09 (2H, m), 1.46 (9H, s), 1.39 (9H, s). ^{13}C NMR (75 MHz, DMSO- d_6) δ 170.8, 162.7, 154.7, 152.0, 136.0, 129.2 (2C), 128.4 (2C), 126.9, 83.4, 78.6, 54.2, 52.2, 36.7, 27.9 (3C), 27.5 (3C). HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{32}\text{N}_3\text{O}_6$ $[\text{M} + \text{H}]^+$ 422.2286, found 422.2294.¹

[1] Balakrishnan, S.; Zhao, C.; Zondlo, N. J. Convergent and Stereospecific Synthesis of Molecules Containing α -Functionalized Guanidiniums via α -Guanidino Acids. *J. Org. Chem.* **2007**, 72, 9834-9837

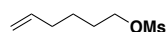
¹H NMR (300 MHz, DMSO-*d*₆)



^{13}C NMR (75 MHz, DMSO- d_6)

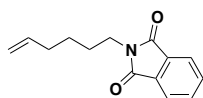


Hex-5-en-1-yl-methanesulfonate (37)



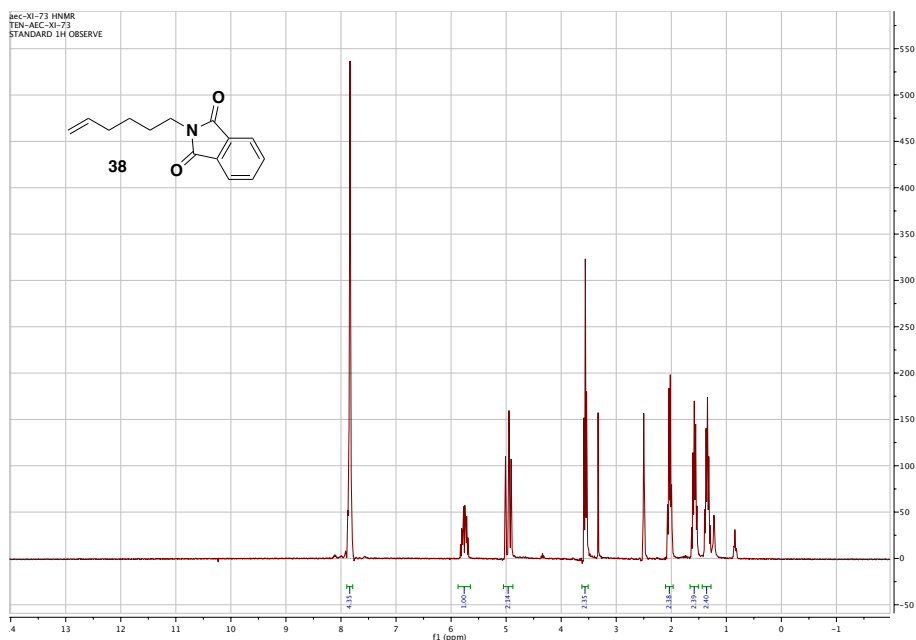
5-Hexen-1-ol (3 ml, 25 mmol) and methanesulfonyl chloride (2.7 ml, 35 mmol) was dissolved in dry CH_2Cl_2 (25 ml) and cooled to 0 °C. Et_3N (4.2 ml, 30 mmol) was slowly added and the solution was stirred for 1 h. The reaction mixture was washed with water (2 × 25 ml). The water phase was extracted with EtOAc (25 ml). The combined organic phases were dried over anhydrous Na_2SO_4 and concentrated *in vacuo* (the product was used straight away).

2-Hex-5-enyl-isoindole-1,3-dione (38)



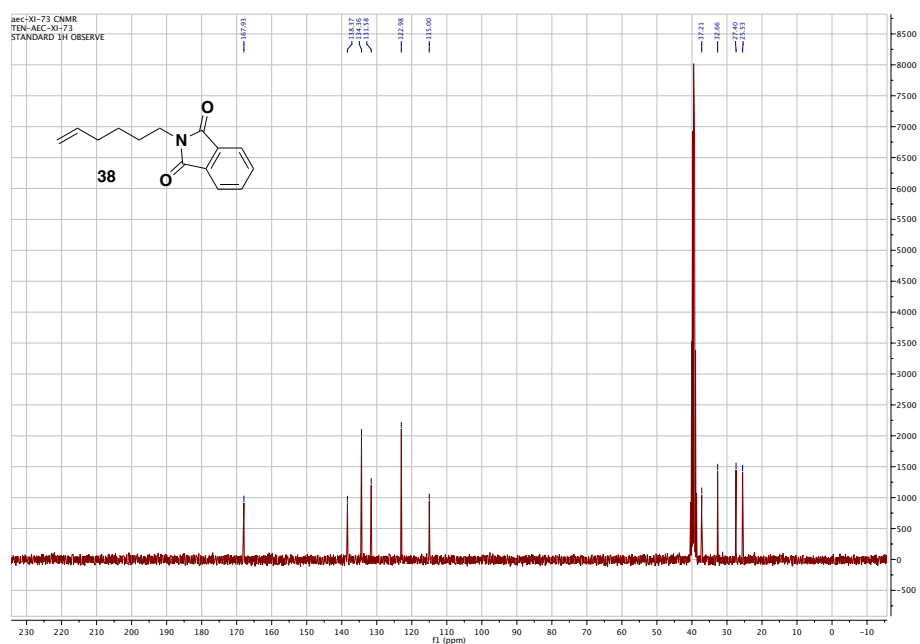
Mesylate **37** (4.5 g, 25 mmol) was dissolved in dry DMF. Potassium phthalimide (6.9 g, 37 mmol) was added and the reaction mixture was heated to 80 °C for 2 h. The solution was cooled to rt. and transferred to a separation funnel with EtOAc (200 ml). The organic solution was washed with NH₄Cl (sat) (2 × 200 ml) and NaHCO₃ (sat) (2 × 200 ml). The organic phase was dried over anhydrous Na₂SO₄ and concentrated *in vacuo*. The product was purified by flash chromatography (EtOAc/heptane 1:4) to yield the title product **38** as a clear oil (4.1 g, 82% over two steps). ¹H NMR (300 MHz, DMSO-*d*₆): δ 7.91 – 7.76 (4H, m), 5.86 – 5.68 (1H, m), 5.03 – 4.89 (2H, m), 3.56 (2H, t, *J* = 7.0 Hz), 2.03 (2H, q, *J* = 7.1 Hz), 1.69 – 1.52 (2H, m), 1.43 – 1.28 (2H, m). ¹³C NMR (75 MHz, DMSO-*d*₆) δ 167.9, 138.4, 134.4 (2C), 131.6 (2C), 123.0 (2C), 115.0, 37.2, 32.7, 27.4, 25.5. UPLC-MS (ESI) calcd for C₁₄H₁₆NO₂ [M + H]⁺ 230.3, found 230.2.²

¹H NMR (300 MHz, DMSO-*d*₆)

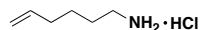


[2] Collins, B. S. L.; Suero, M. G. Copper-Catalyzed Arylative Meyer–Schuster Rearrangement of Propargylic Alcohols to Complex Enones Using Diaryliodonium Salts. *Angew. Chem. Int. Ed.* **2013**, 52, 5799-5802.

¹³C NMR (75 MHz, DMSO-*d*₆)

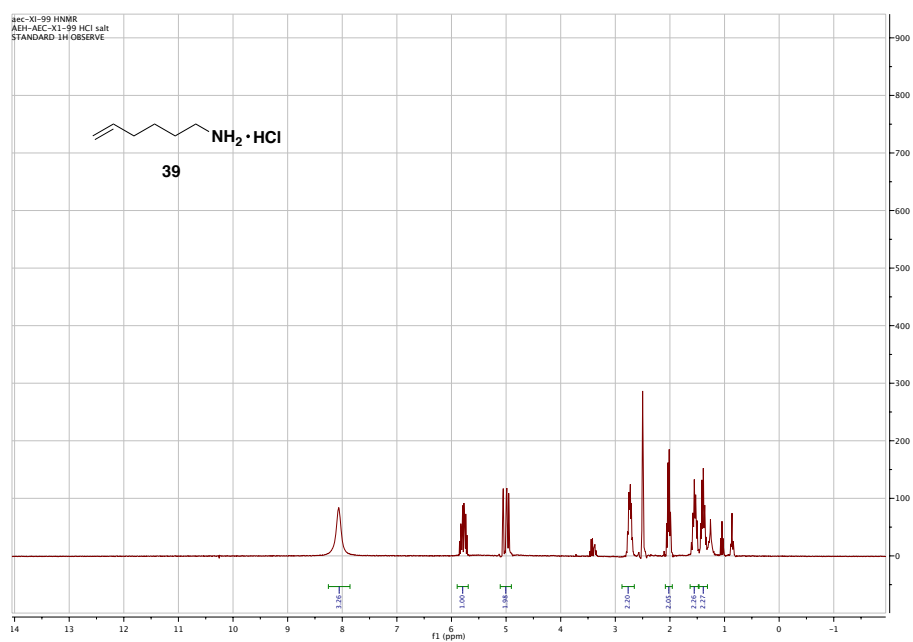


Hex-5-en-1-amine•HCl (39**)**

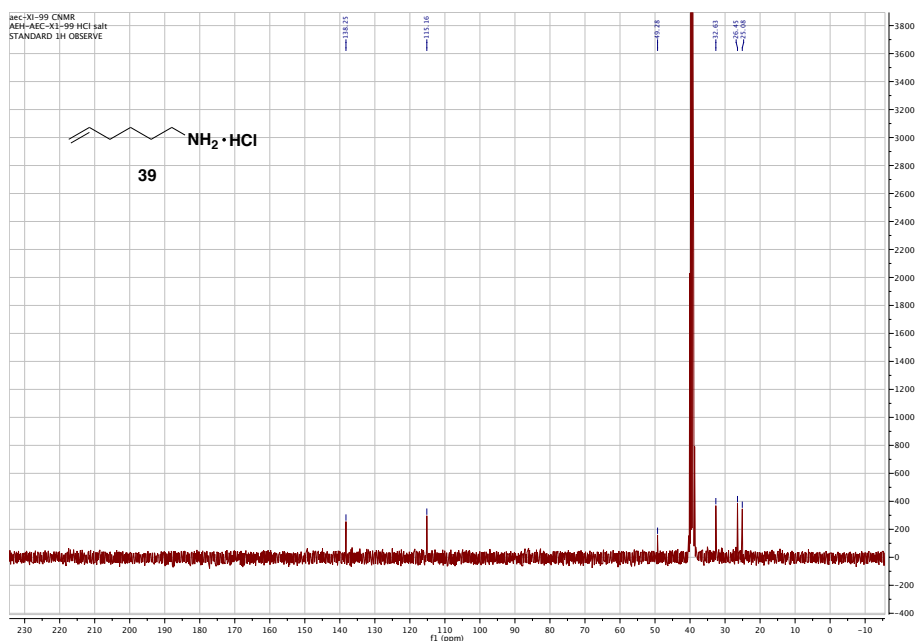


2-Hex-5-enyl-isoindole-1,3-dione **38** (2.5 g, 11 mmol) was dissolved in EtOH (90 ml). Hydrazine mono hydrate (3.2 ml, 33 mmol) was added and the solution was heated to 50 °C for 2 h. The reaction mixture was cooled to rt. The white precipitation was filtered off and washed with EtOH. The acidity of the solution was adjusted to pH 1-2 by addition of conc. HCl. The solution was concentrated *in vacuo* to dryness. The product was dissolved in water (25 ml) and the pH was adjusted to 10-11 by addition of 1 M NaOH. The product was extracted with Et₂O (4 × 50 ml). The organic phase was dried over anhydrous Na₂SO₄ and concentrated *in vacuo* at 0 °C. The product was distilled by use of Kugelrohr. The distillate was acidified by addition of 1M HCl (Et₂O) and concentrated *in vacuo* to give the title product **39** (0.74 g, 49 %). ¹H NMR (300 MHz, DMSO-*d*₆): δ 8.07 (3H, bs), 5.78 (1H, ddt, *J* = 16.9, 10.2, 6.6 Hz), 5.10- 4.87 (2H, m), 2.85 – 2.63 (2H, m), 2.06 – 1.96 (2H, m), 1.64 – 1.48 (2H, m), 1.48 – 1.31 (2H, m). ¹³C NMR (75 MHz, DMSO-*d*₆) δ 138.3, 115.2, 49.3, 32.6, 26.5, 25.1.

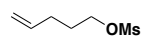
¹H NMR (300 MHz, DMSO-*d*₆)



^{13}C NMR (75 MHz, DMSO- d_6)

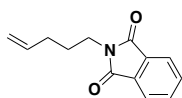


Pent-4-en-1-yl-methanesulfonate (40)



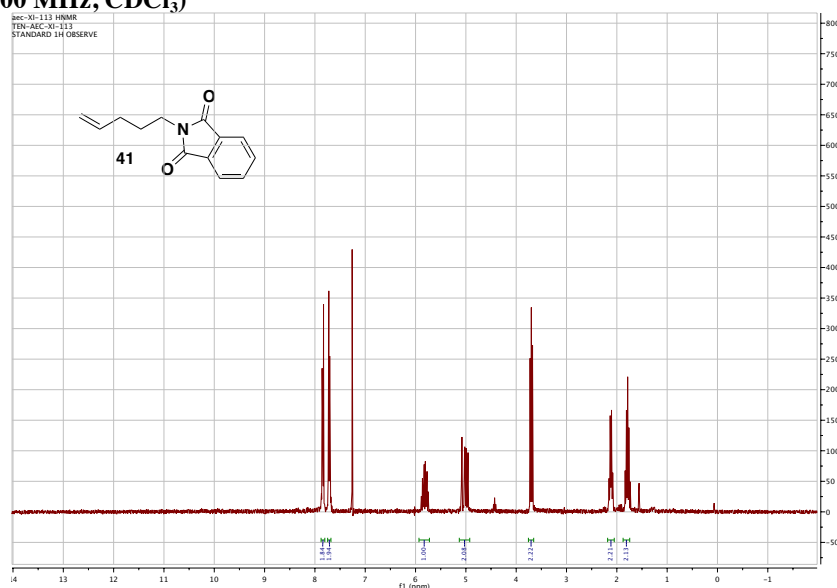
4-Penten-1-ol (3.6 ml, 35 mmol) and methanesulfonyl chloride (3.8 ml, 49 mmol) was dissolved in dry CH_2Cl_2 (35 ml) and cooled to 0 °C. Et_3N (5.8 ml, 42 mmol) was slowly added and the solution was stirred for 1 h. The reaction mixture was washed with water (2 × 50 ml). The water phase was extracted with EtOAc (50 ml). The combined organic phases were dried over anhydrous Na_2SO_4 and concentrated in vacuo (the product was used straight away).

2-Pent-4-enyl-isoindole-1,3-dione (41)

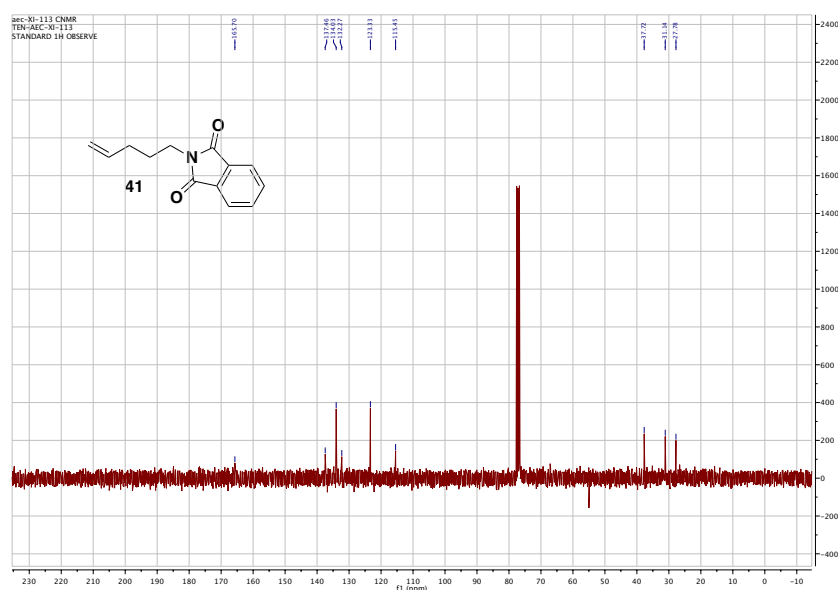


The mesylate **40** (5.7 g, 35 mmol) was dissolved in dry DMF (70 ml). Potassium phthalimide (9.7 g, 52 mmol) was added and the solution was heated to 80 °C for 2 h. The reaction was cooled to rt. and washed with NH₄Cl (sat) (2 × 200 ml) and NaHCO₃ (sat) (2 × 200 ml). The organic phase was dried over anhydrous Na₂SO₄ and concentrated *in vacuo*. The product was purified by flash chromatography (EtOAc/heptane 1:4) to give **41** as a clear colourless oil (5.1 g, 68%). ¹H NMR (300 MHz, CDCl₃): δ 7.84 (2H, dd, *J* = 5.5, 3.0 Hz), 7.71 (2H, dd, *J* = 5.4, 3.1 Hz), 5.82 (1H, ddt, *J* = 16.9, 10.3, 6.6 Hz), 5.05 (1H, ddd, *J* = 17.2, 3.4, 1.7 Hz), 4.98 (1H, dd, *J* = 10.2, 1.8 Hz), 3.77 – 3.63 (2H, m), 2.12 (2H, dd, *J* = 14.6, 6.8 Hz), 1.80 (2H, dd, *J* = 14.7, 7.5 Hz). ¹³C NMR (75 MHz, CDCl₃) δ 165.7, 137.5, 134.0 (2C), 132.3 (2C), 123.3 (2C), 115.5, 37.7, 31.1, 27.8.³

¹H NMR (300 MHz, CDCl₃)

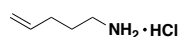


¹³C NMR (75 MHz, CDCl₃)



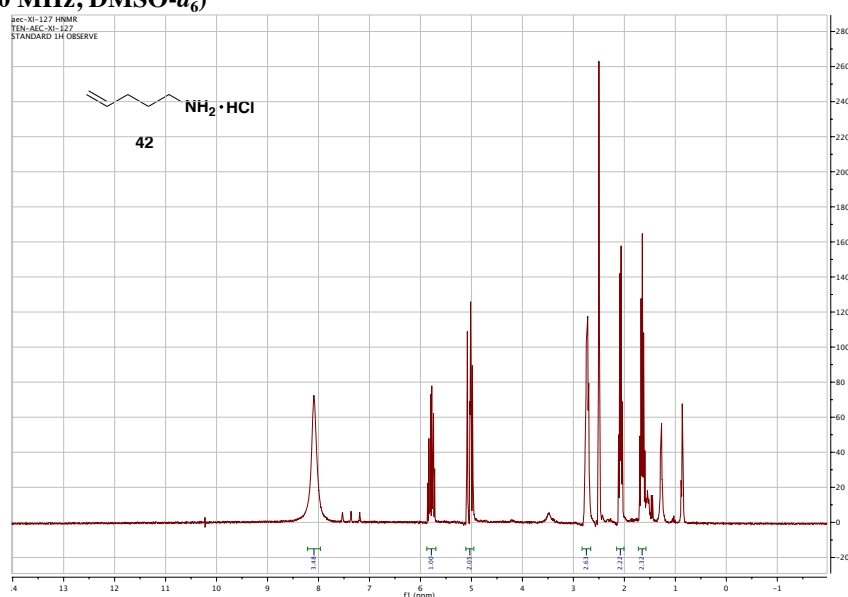
[3] Whittaker, A. M.; Lalic, G. Monophasic Catalytic System for the Selective Semireduction of Alkynes. *Org. Lett.* **2013**, *15*, 1112-1115.

Pent-4-en-1-amine•HCl (42)

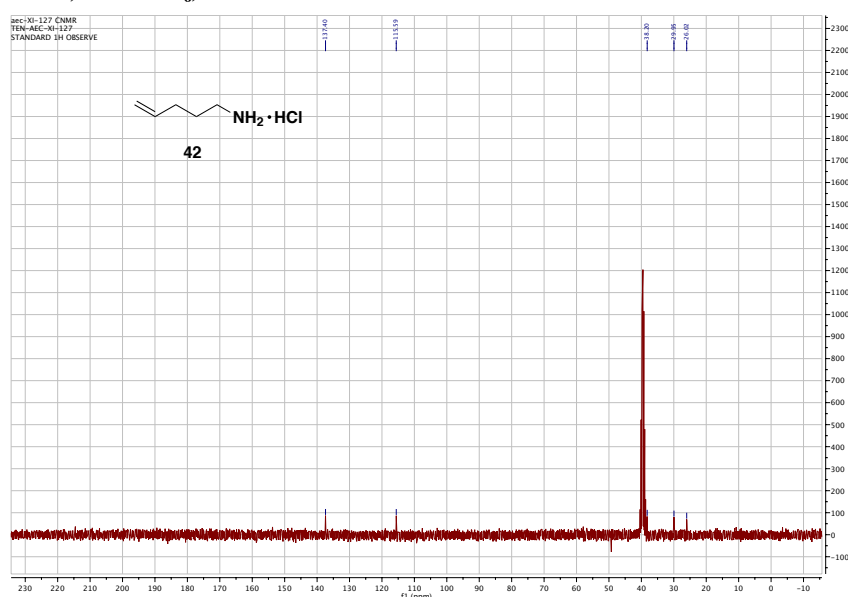


2-Pent-4-enyl-isoindole-1,3-dione **41** (3 g, 14 mmol) was dissolved in EtOH (120 ml). Hydrazine mono hydrate (4.1 ml, 42 mmol) was added and the reaction mixture was heated 50 °C for 19 h. The solution was cooled to rt. The white precipitation was filtered off and washed with EtOH. The acidity of the solution was adjusted to pH 1-2 by addition of conc. HCl. The solution was concentrated *in vacuo* to dryness. The product was dissolved in water (25 ml) and the pH was adjusted to 10-11 by addition of 1 M NaOH. The product was extracted with Et₂O (4 × 50 ml). The organic phase was dried over anhydrous Na₂SO₄ and concentrated *in vacuo* at 0 °C. The product was distilled by use of Kugelrohr. The distillate was acidified by addition of 1M HCl (Et₂O) and concentrated *in vacuo* to give the title product **42** (0.45 g, 27 %). ¹H NMR (300 MHz, DMSO-*d*₆): δ 8.09 (3H, bs), 5.79 (1H, ddt, *J* = 16.8, 10.2, 6.5 Hz), 5.10 – 4.96 (2H, m), 2.83 – 2.64 (2H, m), 2.08 (2H, q, *J* = 6.9 Hz), 1.73 – 1.58 (2H, m). ¹³C NMR (75 MHz, DMSO-*d*₆) δ 137.4, 115.6, 38.2, 30.0, 26.0.⁴

¹H NMR (300 MHz, DMSO-*d*₆)

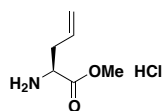


¹³C NMR (75 MHz, DMSO-*d*₆)



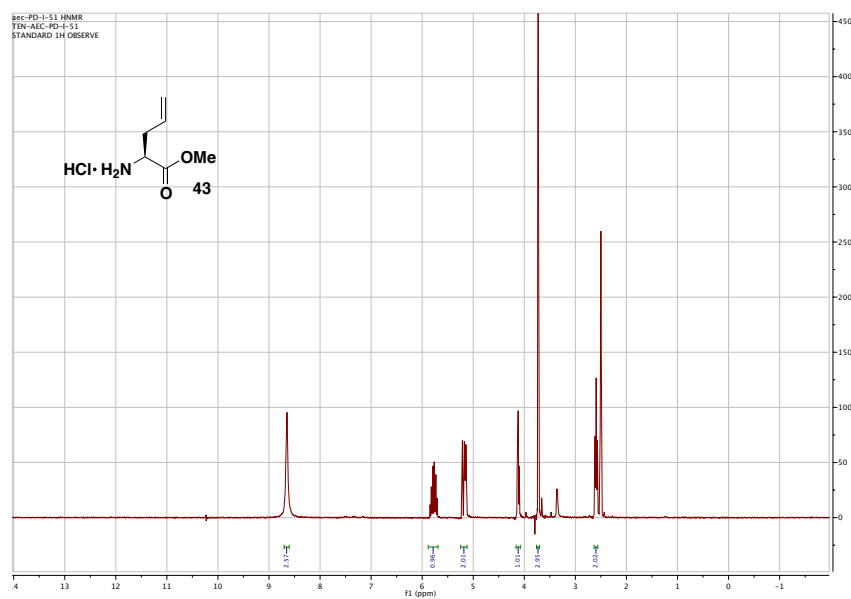
[4] Sohn, J-H.; Kim, K. H.; Lee, H-Y.; No, Z. S.; Ihee, H. Initial Catalyst–Substrate Association Step in Enyne Metathesis Catalyzed by Grubbs Ruthenium Complex Probed by Time-Dependent Fluorescence Quenching. *J. Am. Chem. Soc.* **2008**, *130*, 16506-16507.

L-Allylglycine methyl ester•HCl (43)

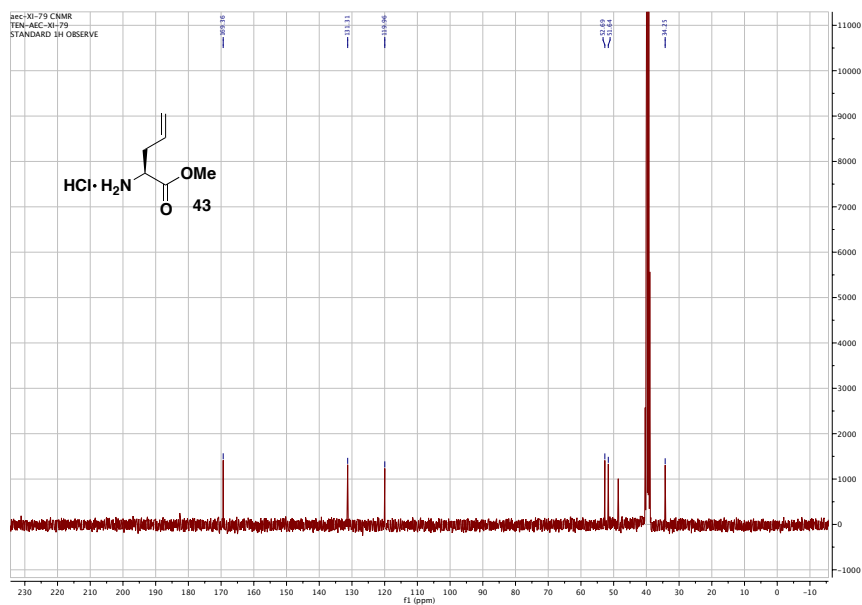


L-H-allylglycine (0.5 g, 4.3 mmol) was dissolved in MeOH (9 ml) and cooled to 0 °C. Thionyl chloride (0.47 ml, 6.5 mmol) was slowly added and the solution was stirred for 22 h at rt. The solution was concentrated *in vacuo*, dissolved in CH₂Cl₂ and concentrated again *in vacuo* to give the title product **43** (0.72 g, quant). ¹H NMR (300 MHz, DMSO-*d*₆): δ 8.65 (3H, bs), 5.78 (1H, ddt, *J* = 17.2, 10.1, 7.2 Hz), 5.23 – 5.12 (2H, m), 4.12 (1H, t, *J* = 6.1 Hz), 3.73 (3H, s), 2.59 (2H, t, *J* = 6.6 Hz). ¹³C NMR (75 MHz, DMSO-*d*₆) δ 169.4, 131.3, 120.0, 52.7, 51.6, 34.3.⁵

¹H NMR (300 MHz, DMSO-*d*₆)

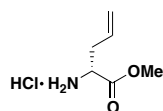


¹³C NMR (75 MHz, DMSO-*d*₆)



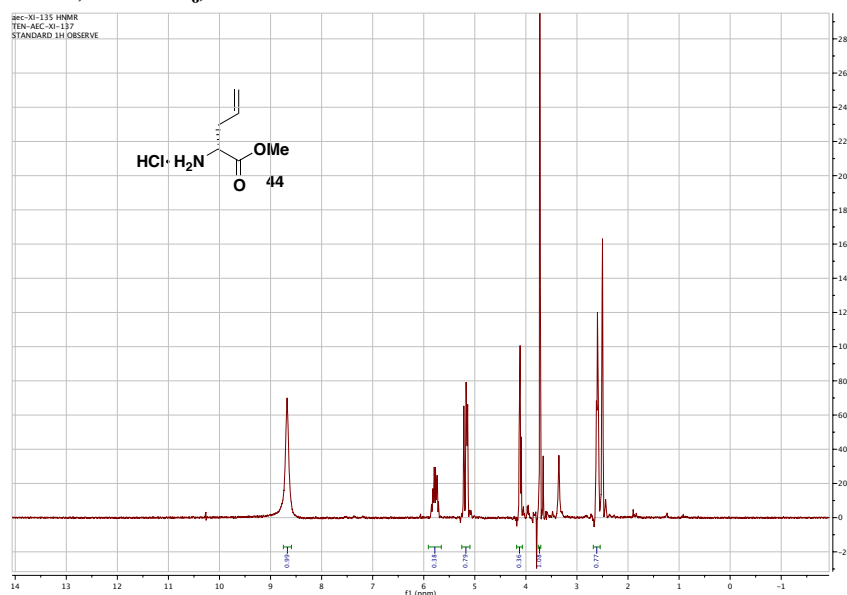
[5] Kaul, R.; Surprenant, S.; Lubell, W. D. Systematic Study of the Synthesis of Macrocyclic Dipeptide β -Turn Mimics Possessing 8-, 9-, and 10- Membered Rings by Ring-Closing Metathesis. *J. Org. Chem.* **2004**, *70*, 3838-3844.

D-Allylglycine methyl ester•HCl (44)

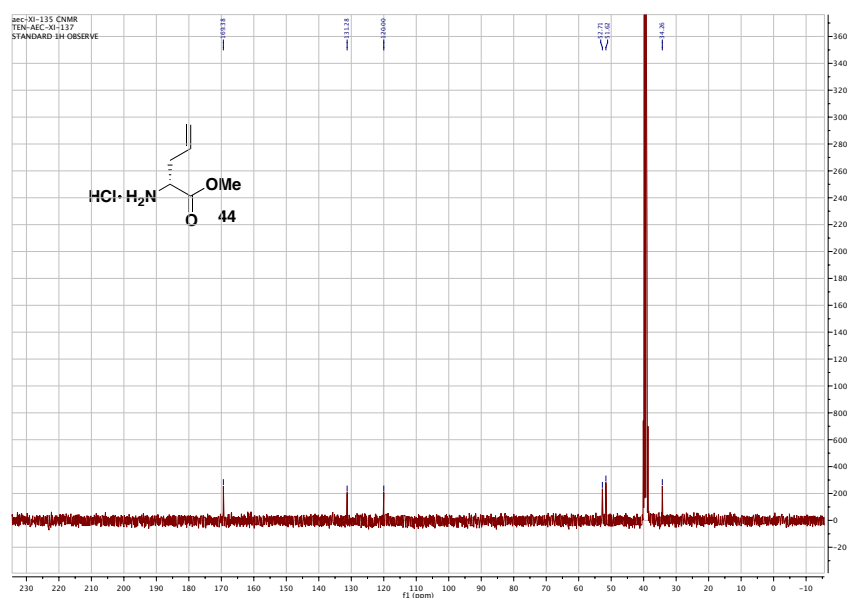


D-H-allylglycine (0.5 g, 4.3 mmol) was dissolved in MeOH (9 ml) and cooled to 0 °C. Thionyl chloride (0.47 ml, 6.5 mmol) was slowly added and the solution was stirred for 22 h at rt. The solution was concentrated *in vacuo*, dissolved in CH₂Cl₂ and concentrated again *in vacuo* to give the title product **44** (0.72 g, quant). ¹H NMR (300 MHz, DMSO-*d*₆): δ 8.67 (3H, bs), 5.92 – 5.61 (1H, m), 5.35 – 5.12 (2H, m), 4.11 (1H, t, *J* = 6.0 Hz), 3.73 (3H, s), 2.60 (2H, t, *J* = 6.0 Hz). ¹³C NMR (75 MHz, DMSO-*d*₆) δ 169.4, 131.3, 120.0, 52.7, 51.6, 34.3.⁶

¹H NMR (300 MHz, DMSO-*d*₆)



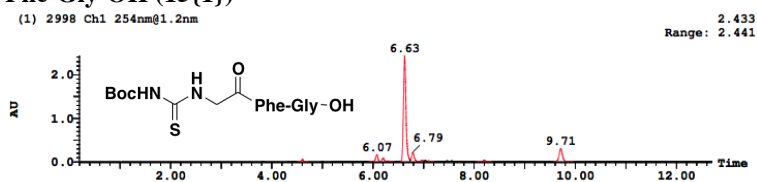
¹³C NMR (75 MHz, DMSO-*d*₆)



[6] Goudrea, N.; Brochu, C.; Cameron, D. R.; Duceppe, J-S.; Faucher, A-M.; Ferland, J-M.; Grand-Maître, C.; Poirier, M.; Simoneau, B.; Tsantrizos, Y. S. Potent Inhibitors of the Hepatitis C Virus NS3 Protease: Design and Synthesis of Macrocyclic Substrate-Based β -Strand Mimics. *J. Org. Chem.* **2004**, *69*, 6185-6201.

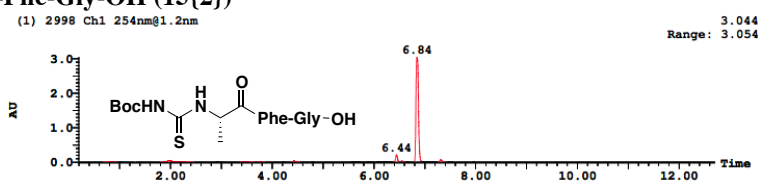
Synthesis of Boc-Protected Thiourea and Guanidino Peptides

BocNHCS-Gly-Phe-Gly-OH (15{1})



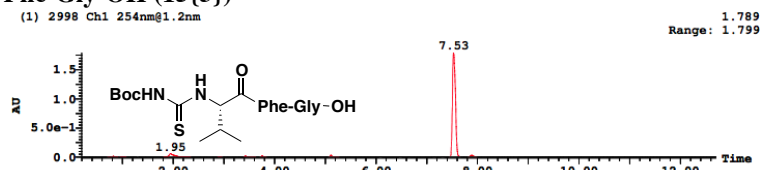
Purity: 86%, R_f = 6.63 min. UPLC-MS (ESI) calcd for $C_{19}H_{27}N_4O_6S$ $[M + H]^+$ 439.2, found 439.3.

BocNHCS-Ala-Phe-Gly-OH (15{2})



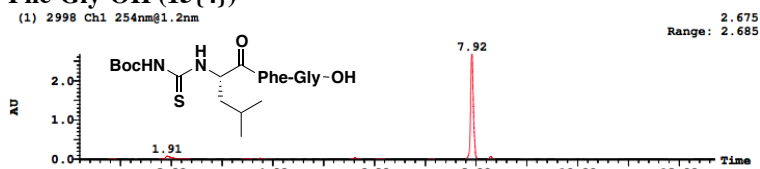
Purity: >95%, R_f = 6.84 min. HRMS (ESI) calcd for $C_{20}H_{29}N_4O_6S$ $[M + H]^+$ 453.1802, found 453.1804.

BocNHCS-Val-Phe-Gly-OH (15{3})



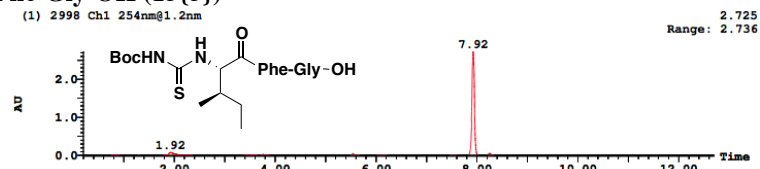
Purity: >95%, R_f = 7.53 min. UPLC-MS (ESI) calcd for $C_{22}H_{33}N_4O_6S$ $[M + H]^+$ 481.2, found 481.4.

BocNHCS-Leu-Phe-Gly-OH (15{4})

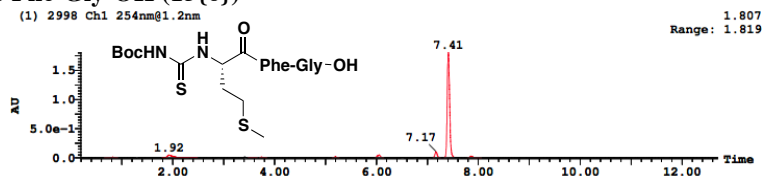


Purity: >95%, R_f = 7.92 min. UPLC-MS (ESI) calcd for $C_{23}H_{35}N_4O_6S$ $[M + H]^+$ 495.2, found 495.4.

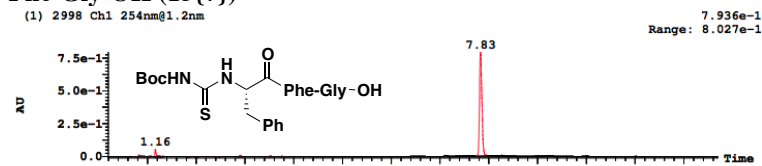
BocNHCS-Ile-Phe-Gly-OH (15{5})



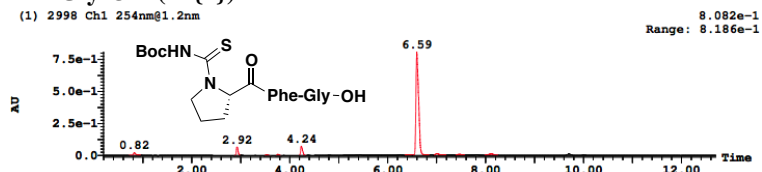
Purity: >95%, R_f = 7.92 min. UPLC-MS (ESI) calcd for $C_{23}H_{35}N_4O_6S$ $[M + H]^+$ 495.2, found 495.4.

BocNHCS-Met-Phe-Gly-OH (15{6})

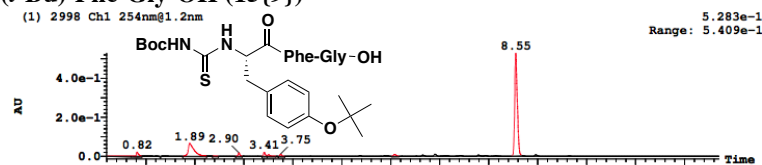
Purity: >95%, $R_f = 7.41$ min. UPLC-MS (ESI) calcd for $C_{22}H_{33}N_4O_6S_2$ $[M + H]^+$ 513.2, found 513.4.

BocNHCS-Phe-Phe-Gly-OH (15{7})

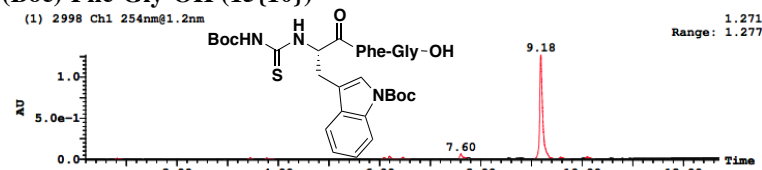
Purity: >95%, $R_f = 7.83$ min. UPLC-MS (ESI) calcd for $C_{26}H_{33}N_4O_6S$ $[M + H]^+$ 529.2, found 529.4.

BocNHCS-Pro-Phe-Gly-OH (15{8})

Purity: 87%, $R_f = 6.59$ min. UPLC-MS (ESI) calcd for $C_{22}H_{31}N_4O_6S$ $[M + H]^+$ 479.2, found 479.4

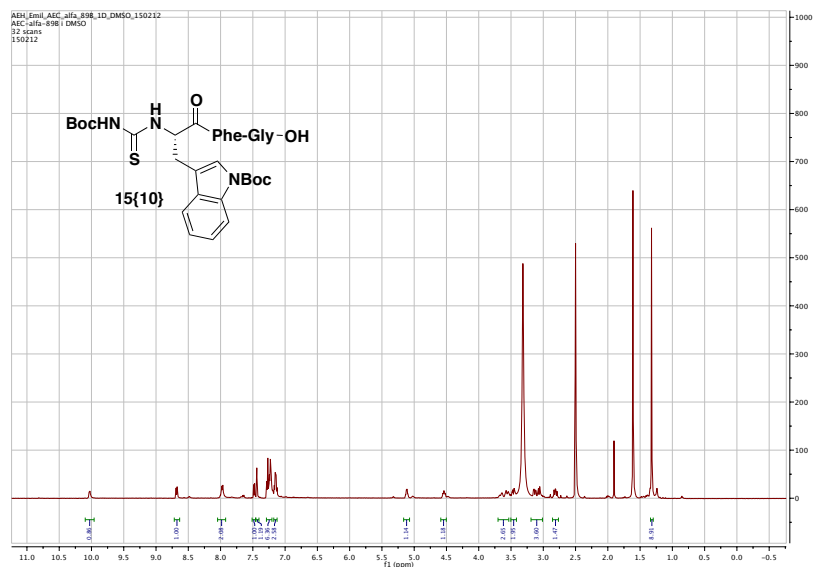
BocNHCS-Tyr(*t*-Bu)-Phe-Gly-OH (15{9})

Purity: >95%, $R_f = 8.55$ min. UPLC-MS (ESI) calcd for $C_{30}H_{41}N_4O_7S$ $[M + H]^+$ 601.3, found 601.5.

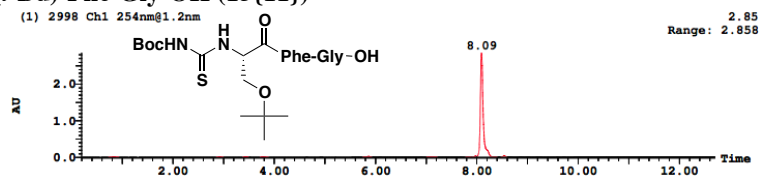
BocNHCS-Trp(Boc)-Phe-Gly-OH (15{10})

Purity: >95%, $R_f = 9.18$ min. 1H NMR (500 MHz, $DMSO-d_6$): δ 10.03 (1H, d, $J = 7.3$ Hz), 8.69 (1H, d, $J = 8.4$ Hz), 8.02 – 7.91 (2H, m), 7.48 (1H, d, $J = 7.7$ Hz), 7.44 (1H, s), 7.30 – 7.09 (8H, m), 5.12 (1H, d, $J = 5.9$ Hz), 4.54 (1H, td, $J = 4.1$ Hz, 9.4 Hz), 3.67 – 3.50 (2H, m), 3.46 (1H, dd, $J = 4.8$ Hz, 14.7 Hz), 3.13 (1H, dd, $J = 5.5$ Hz, 14.8 Hz), 3.06 (1H, dd, $J = 3.9$ Hz, 13.8 Hz), 2.81 (1H, dd, $J = 10.1$ Hz, 13.8 Hz), 1.33 (9H, s). HRMS (ESI) calcd for $C_{33}H_{42}N_5O_8S$ $[M + H]^+$ 668.2749, found 668.2750.

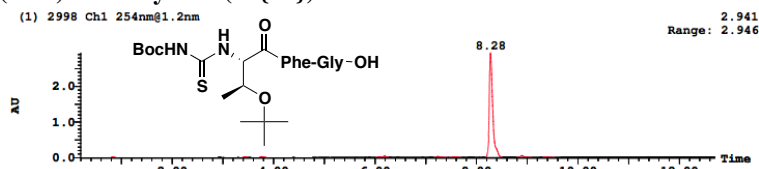
¹H NMR (500 MHz, DMSO-*d*₆)



BocNHCS-Ser(*t*-Bu)-Phe-Gly-OH (15{11})

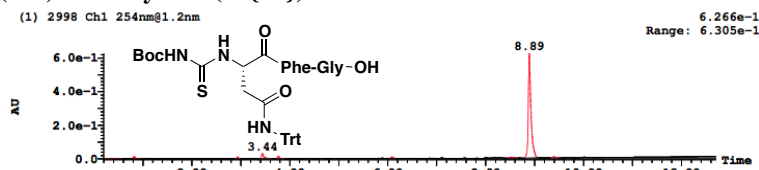
Purity: >95%, R_f = 8.09 min. HRMS (ESI) calcd for $C_{24}H_{37}N_4O_7S$ $[M + H]^+$ 525.2377, found 525.2369.

BocNHCS-Thr(*t*-Bu)-Phe-Gly-OH (15{12})



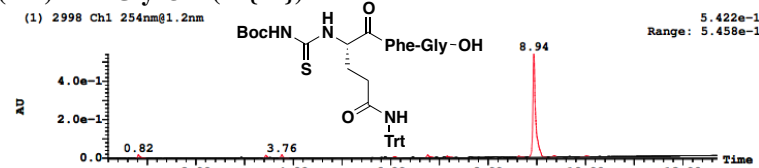
Purity: >95%, $R_f = 8.28$ min. UPLC-MS (ESI) calcd for $C_{25}H_{39}N_4O_7S$ $[M + H]^+$ 539.3, found 539.5.

BocNHCS-Asn(Trt)-Phe-Gly-OH (15{13})

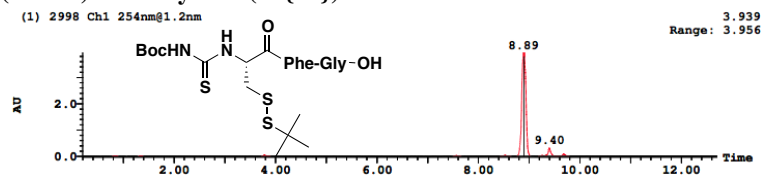


Purity: >95%, R_f = 8.89 min. UPLC-MS (ESI) calcd for $C_{40}H_{44}N_5O_7S$ $[M + H]^+$ 738.3, found 738.4.

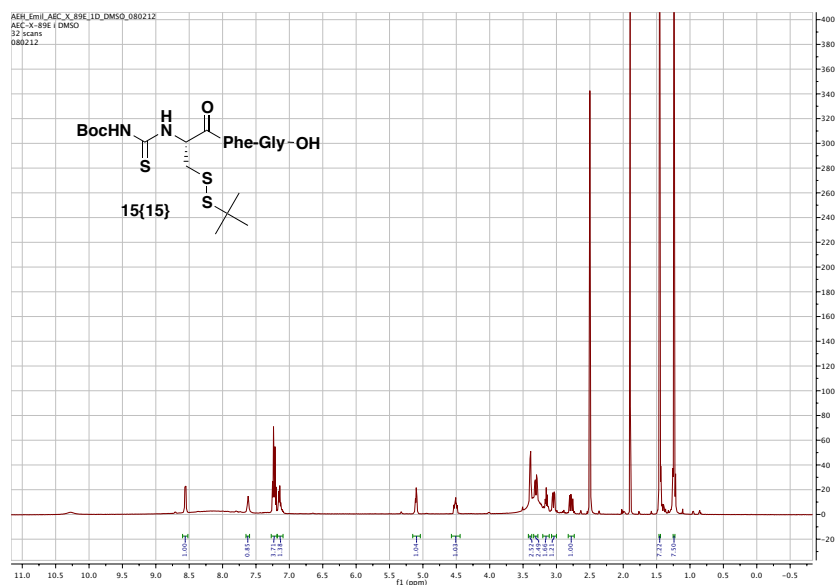
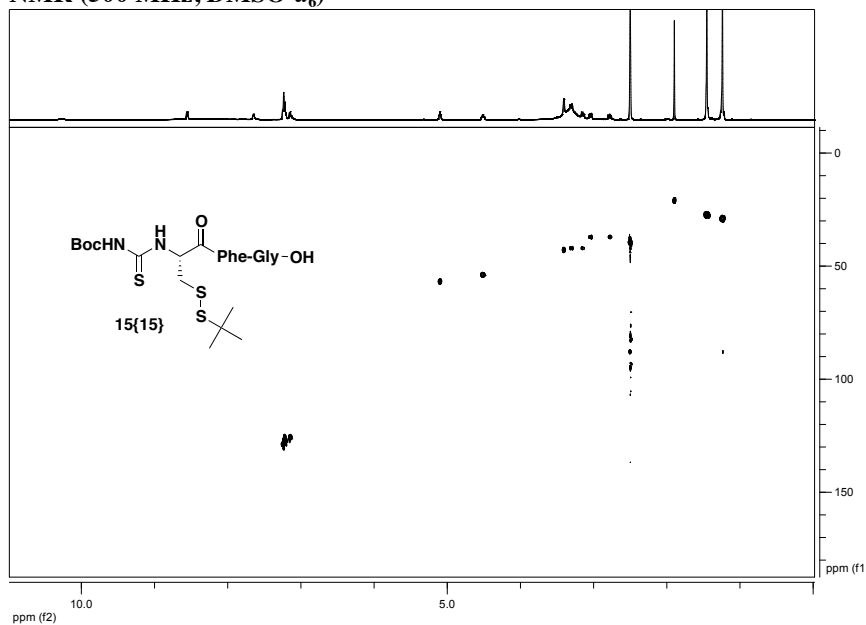
BocNHCS-Gln(Trt)-Phe-Gly-OH (15{14})



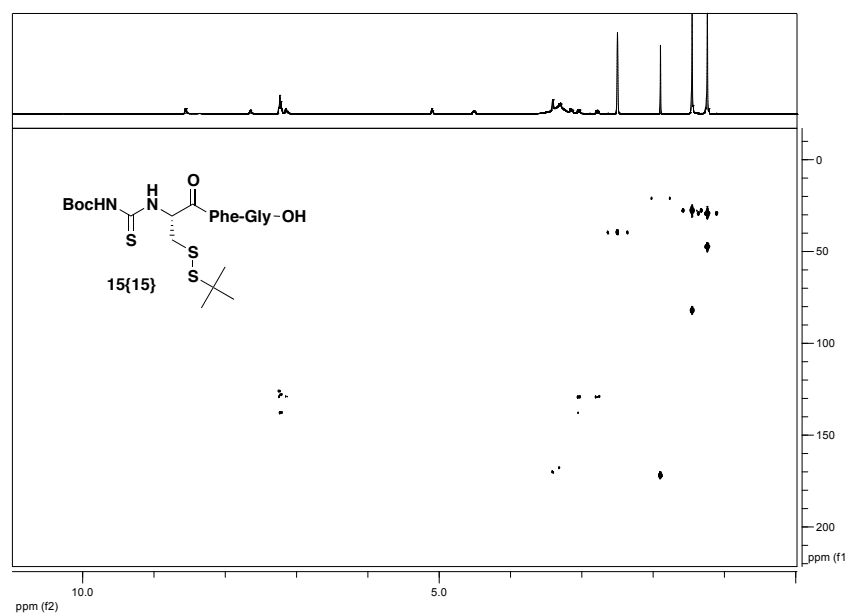
Purity: >95%, R_f = 8.94 min. UPLC-MS (ESI) calcd for C₄₁H₄₆N₅O₇S [M + H]⁺ 752.3, found 752.4.

BocNHCS-Cys(S-*t*-Bu)-Phe-Gly-OH (15{15})

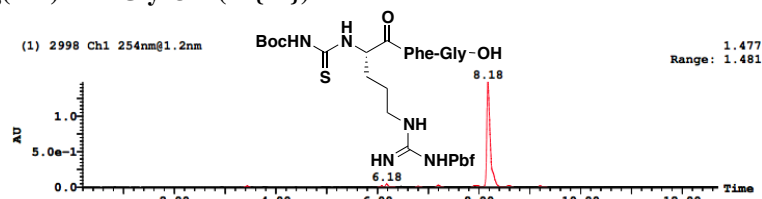
Purity: >95%, R_f = 8.89 min. ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δ 8.56 (1H, d, J = 8.5 Hz), 7.62 (1H, bs), 7.27 – 7.19 (m, 4H), 7.14 (1H, dd, J = 5.7 Hz, 12.6 Hz), 5.10 (1H, t, J = 5.3 Hz), 4.51 (1H, td, J = 4.6 Hz, 9.3 Hz), 3.39 (2H, d, J = 4.5 Hz), 3.31 (1H, dd, J = 4.6 Hz, 13.4 Hz), 3.15 (1H, dd, J = 6.2 Hz, 13.5 Hz), 3.04 (1H, dd, J = 4.4 Hz, 13.8 Hz), 2.78 (1H, dd, J = 9.8 Hz, 13.8 Hz), 1.45 (9H, s), 1.24 (9H, s). ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$): δ 179.5, 170.2, 169.7, 167.9, 137.8, 128.7 (2C), 127.8 (2C), 126.0, 82.1, 57.0, 54.0, 47.5, 43.0, 42.1, 37.2, 27.7 (3H), 29.1 (3H). UPLC-MS (ESI) calcd for $\text{C}_{24}\text{H}_{37}\text{N}_4\text{O}_6\text{S}_2$ $[\text{M} + \text{H}]^+$ 573.2, found 573.4.

 ^1H NMR (500 MHz, $\text{DMSO}-d_6$)**2D HSQC NMR (500 MHz, $\text{DMSO}-d_6$)**

2D HMBC NMR (500 MHz, DMSO-*d*₆)

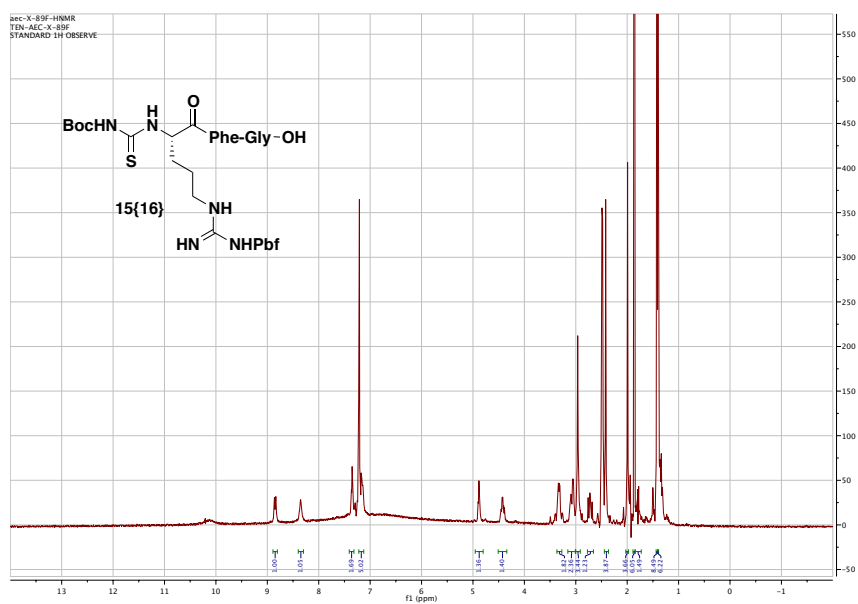


BocNHCS-Arg(Pbf)-Phe-Gly-OH (15{16})

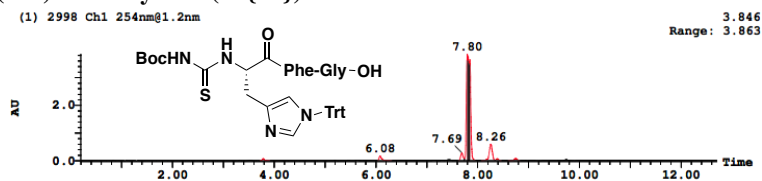


Purity: >95%, $R_f = 8.18$ min. ^1H NMR (300 MHz, DMSO-*d*₆): δ 8.86 (1H, d, $J = 8.3$ Hz), 8.36 (1H, bs), 7.36 (1H, t, $J = 4.0$ Hz), 7.28 – 7.11 (5H, m), 4.89 (1H, t, $J = 5.2$ Hz), 4.49 – 4.38 (1H, m), 3.44 – 3.25 (2H, m), 3.19 – 3.04 (2H, m), 2.97 (2H, bs), 2.83 – 2.65 (1H, m), 2.43 (3H, s), 2.00 (3H, s), 1.87 (6H, s), 1.85 – 1.77 (2H, m), 1.43 (9H, s), 1.41 (6H, s). HRMS (ESI) calcd for $\text{C}_{36}\text{H}_{52}\text{N}_7\text{O}_9\text{S}_2$ $[\text{M} + \text{H}]^+$ 790.3262, found 790.3287.

^1H NMR (300 MHz, DMSO-*d*₆)

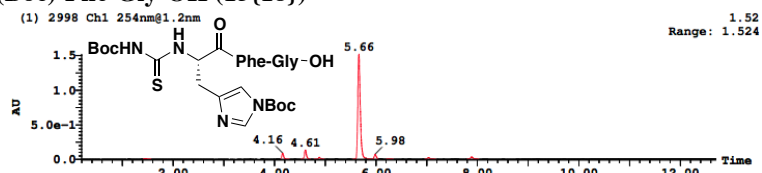


BocNHCS-His(Trt)-Phe-Gly-OH (15{17})



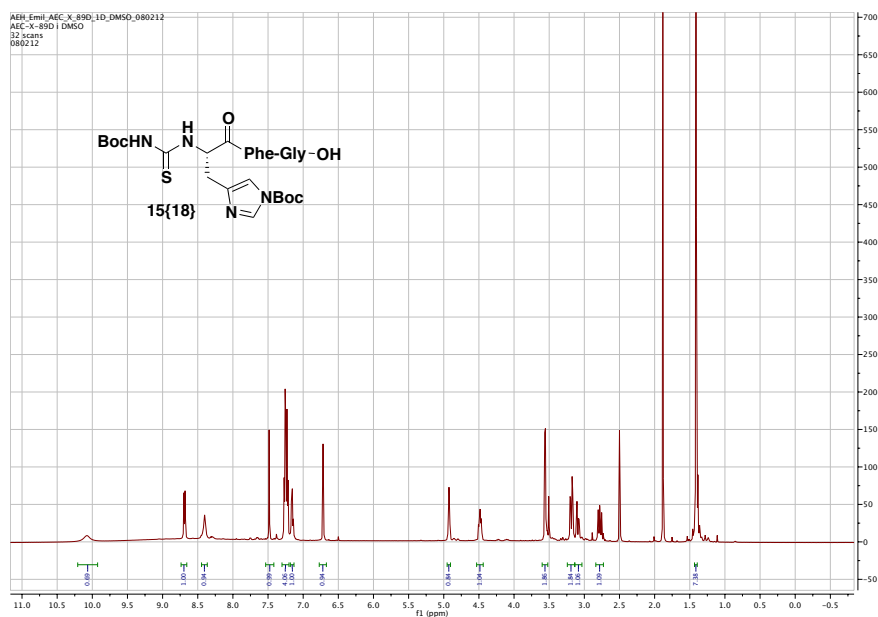
Purity: 83%, R_f = 7.80 min. UPLC-MS (ESI) calcd for $C_{42}H_{45}N_6O_6S$ $[M + H]^+$ 761.3, found 761.6.

BocNHCS-His(Boc)-Phe-Gly-OH (15{18})

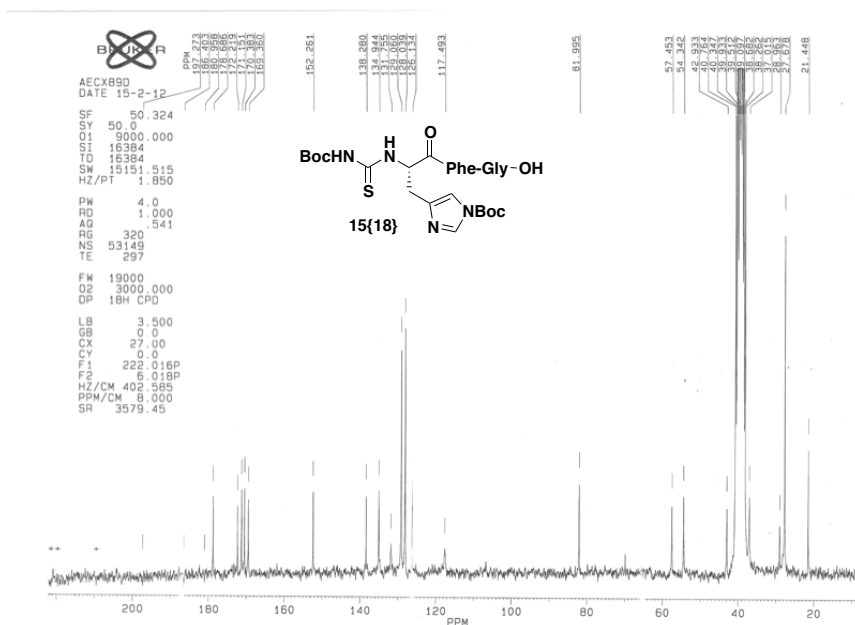


Purity: 92%, R_f = 5.66 min. (The side-chain Boc group was lost during cleavage from the resin). ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δ 10.08 (1H, s), 8.69 (1H, d, J = 8.2 Hz), 8.40 (2H, s), 7.48 (1H, s), 7.29 – 7.21 (5H, m), 7.15 (1H, t, J = 7.0 Hz), 6.72 (1H, s), 4.93 (1H, t, J = 4.8 Hz), 4.54 – 4.44 (1H, m), 3.56 (2H, d, J = 4.9 Hz), 3.25 – 3.14 (1H, m), 3.09 (2H, dd, J = 4.7 Hz, 14.8 Hz), 2.78 (1H, dd, J = 10.6 Hz, 13.9 Hz), 1.41 (9H, s). ^{13}C NMR (50 MHz, $\text{DMSO}-d_6$): δ 181.0, 172.2, 171.2, 170.4, 169.4, 152.3, 138.3, 134.9, 131.8, 129.1 (2C), 128.0 (2C), 126.1, 117.5, 82.0, 57.4, 54.3, 42.9, 37.0, 29.0, 27.7. HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{31}\text{N}_6\text{O}_6\text{S} [\text{M} - \text{Boc}]^+$ 519.2020, found 519.2043.

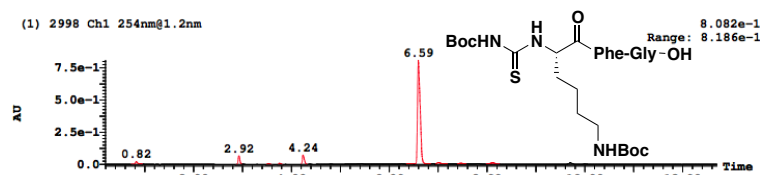
¹H NMR (500 MHz, DMSO-*d*₆)



^{13}C NMR (50 MHz, DMSO- d_6)

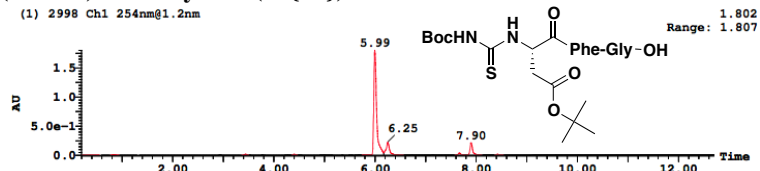


BocNHCS-Lys(Boc)-Phe-Gly-OH (15{19})



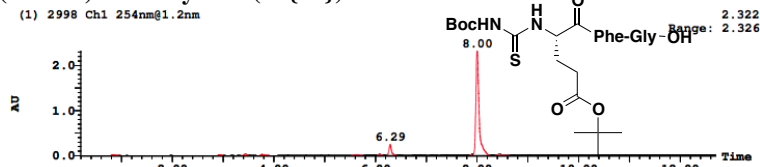
Purity: >95%, R_f = 7.82 min. UPLC-MS (ESI) calcd for $\text{C}_{28}\text{H}_{44}\text{N}_5\text{O}_8\text{S}$ $[\text{M} + \text{H}]^+$ 610.3, found 610.5.

BocNHCS-Asp(O-*t*-Bu)-Phe-Gly-OH (15{20})

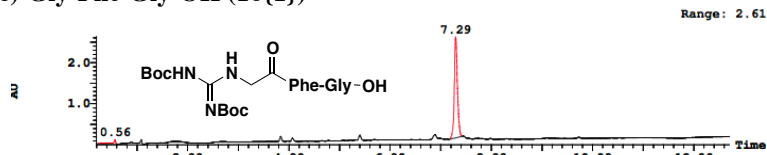


Purity: 78%, R_f = 5.99 min. (The O-*t*-Bu group was lost during cleavage from the resin). UPLC-MS (ESI) calcd for $\text{C}_{21}\text{H}_{29}\text{N}_4\text{O}_8\text{S}$ $[\text{M} - \text{t-Bu}]^+$ 497.2, found 497.4.

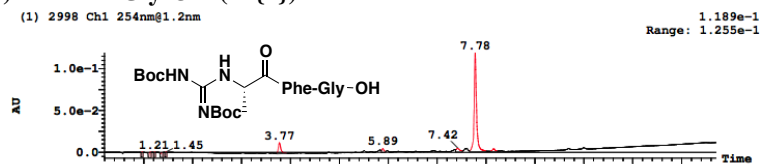
BocNHCS-Glu(O-*t*-Bu)-Phe-Gly-OH (15{21})



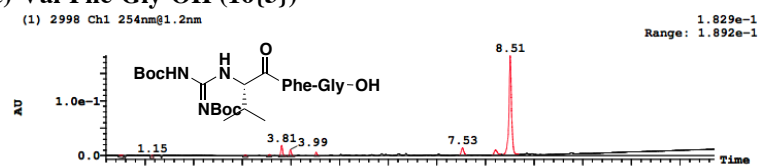
Purity: 92%, R_f = 8.00 min. UPLC-MS (ESI) calcd for $\text{C}_{26}\text{H}_{39}\text{N}_4\text{O}_8\text{S}$ $[\text{M} + \text{H}]^+$ 567.2, found 567.4.

BocNHCN(Boc)-Gly-Phe-Gly-OH (16{1})

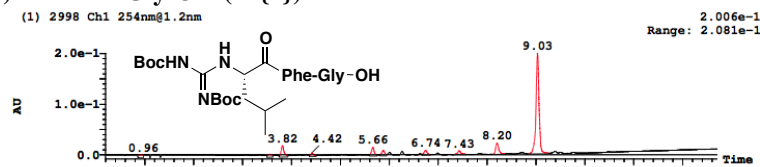
Purity: >95%, R_f = 7.29 min. UPLC-MS (ESI) calcd for $C_{24}H_{36}N_5O_8$ $[M + H]^+$ 522.3, found 522.5.

BocNHCN(Boc)-Ala-Phe-Gly-OH (16{2})

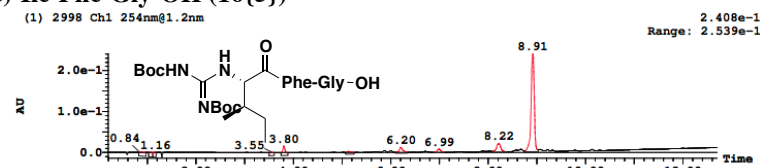
Purity: 90%, R_f = 7.78 min. UPLC-MS (ESI) calcd for $C_{25}H_{38}N_5O_8$ $[M + H]^+$ 536.3, found 536.5.

BocNHCN(Boc)-Val-Phe-Gly-OH (16{3})

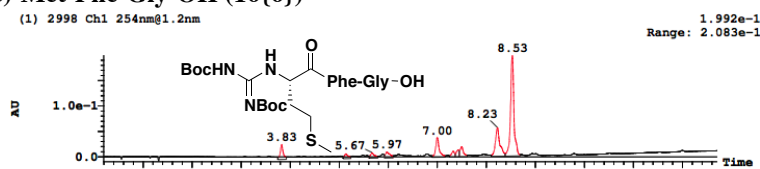
Purity: 87%, R_f = 8.51 min. HRMS (ESI) calcd for $C_{27}H_{42}N_5O_8$ $[M + H]^+$ 564.3028, found 564.3065.

BocNHCN(Boc)-Leu-Phe-Gly-OH (16{4})

Purity: >95%, R_f = 9.03 min. UPLC-MS (ESI) calcd for $C_{28}H_{44}N_5O_8$ $[M + H]^+$ 578.3, found 578.5.

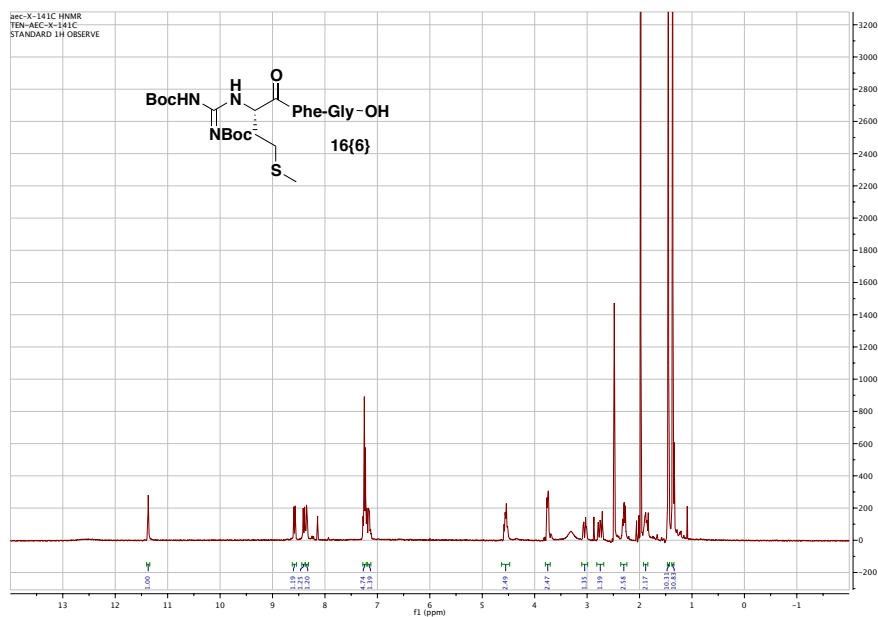
BocNHCN(Boc)-Ile-Phe-Gly-OH (16{5})

Purity: >95%, R_f = 8.91 min. UPLC-MS (ESI) calcd for $C_{28}H_{44}N_5O_8$ $[M + H]^+$ 578.3, found 578.6.

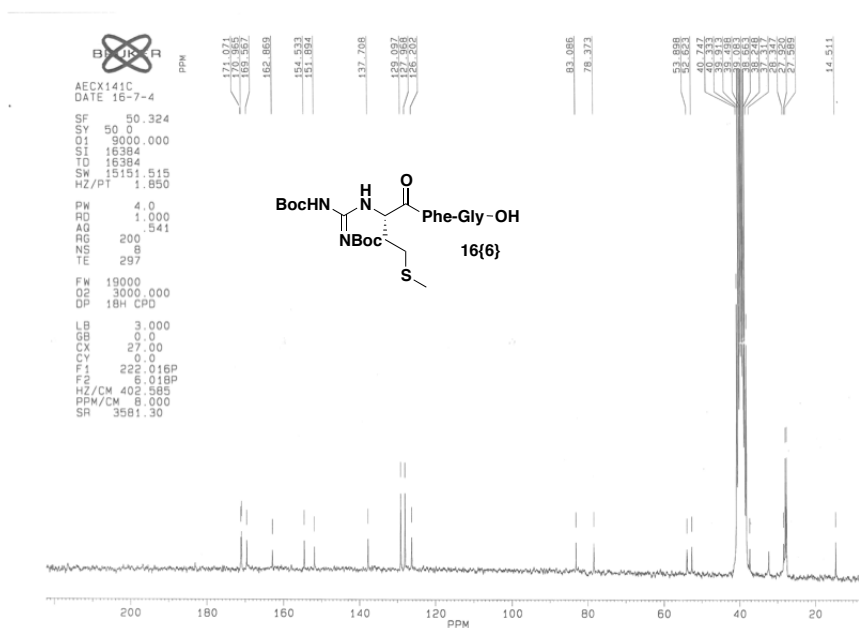
BocNHCN(Boc)-Met-Phe-Gly-OH (16{6})

Purity: 75%, R_f = 8.53 min. 1H NMR (300 MHz, $DMSO-d_6$): δ 11.39 (1H, s), 8.60 (1H, d, J = 7.6 Hz), 8.42 (1H, d, J = 8.3 Hz), 8.37 (1H, t, J = 5.8 Hz), 7.34 – 7.10 (5H, m), 4.61 – 4.52 (2H, m), 3.77 (2H, dd, J = 2.7 Hz, 5.7 Hz), 3.06 (1H, dd, J = 4.1 Hz, 13.8 Hz), 2.77 (1H, dd, J = 9.9 Hz, 13.5 Hz), 2.31 (2H, dd, J = 6.6 Hz, 9.0 Hz), 1.95 – 1.85 (2H, m), 1.47 (9H, s), 1.39 (9H, s). ^{13}C NMR (50 MHz, $DMSO-d_6$): δ 171.1, 171.0, 169.6, 162.9, 154.5, 151.9, 137.7, 129.1 (2C) 128.0 (2C), 126.2, 83.1, 78.4, 53.9, 52.6, 37.3, 32.3, 28.4, 27.9 (3C), 27.6 (3C), 14.5. HRMS (ESI) calcd for $C_{27}H_{42}N_5O_8S$ $[M + H]^+$ 596.2749, found 596.2779.

¹H NMR (300 MHz, DMSO-*d*₆)

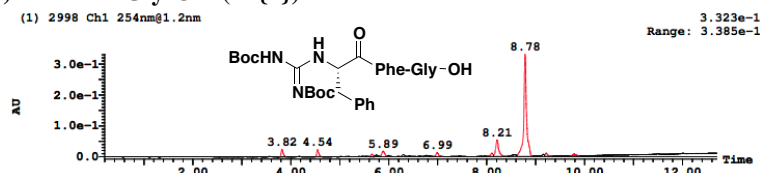


¹³C NMR (50 MHz, DMSO-*d*₆)

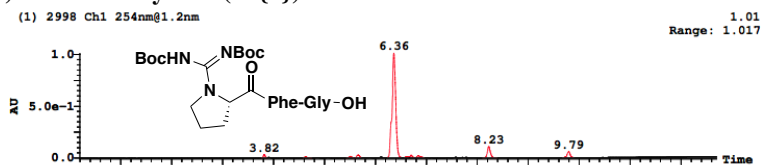


BocNHCN(Boc)-Phe-Phe-Gly-OH (16{7})

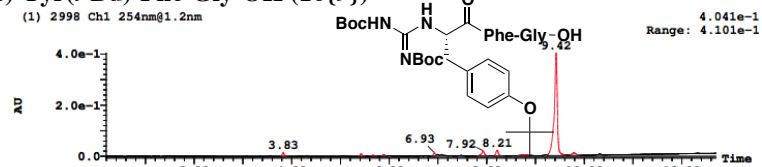
(1) 2998 Ch1 254nm@1.2nm



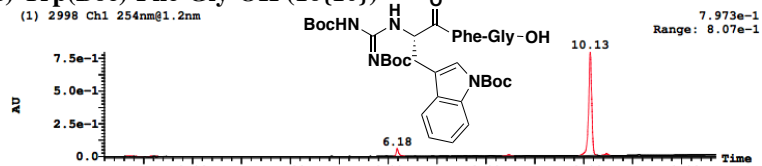
Purity: 74%, $R_f = 8.78$ min. UPLC-MS (ESI) calcd for $C_{31}H_{42}N_5O_8$ $[M + H]^+$ 612.3, found 612.5.

BocNHCN(Boc)-Pro-Phe-Gly-OH (16{8})

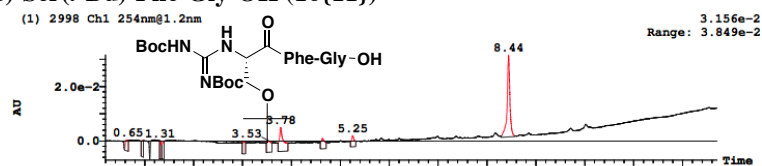
Purity: 90%, $R_f = 6.36$ min. UPLC-MS (ESI) calcd for $C_{27}H_{40}N_5O_8$ $[M + H]^+$ 562.3, found 562.5.

BocNHCN(Boc)-Tyr(*t*-Bu)-Phe-Gly-OH (16{9})

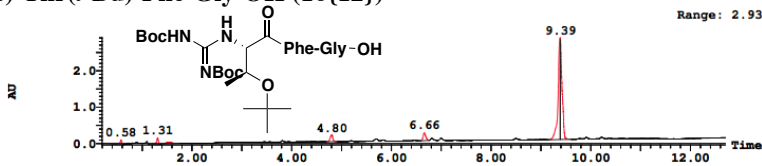
Purity: 85%, $R_f = 9.42$ min. UPLC-MS (ESI) calcd for $C_{35}H_{50}N_5O_9$ $[M + H]^+$ 684.4, found 684.6.

BocNHCN(Boc)-Trp(Boc)-Phe-Gly-OH (16{10})

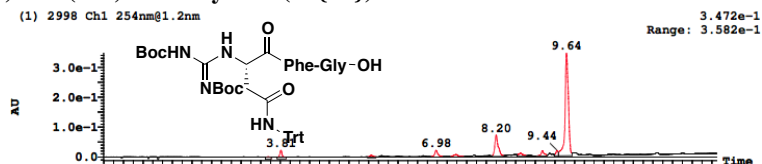
Purity: 94%, $R_f = 10.13$ min. UPLC-MS (ESI) calcd for $C_{38}H_{51}N_6O_{10}$ $[M + H]^+$ 751.4, found 751.6.

BocNHCN(Boc)-Ser(*t*-Bu)-Phe-Gly-OH (16{11})

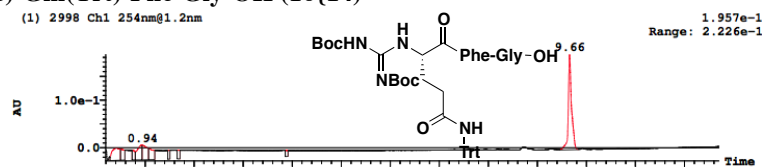
Purity: 90%, $R_f = 8.44$ min. UPLC-MS (ESI) calcd for $C_{29}H_{46}N_5O_9$ $[M + H]^+$ 608.3, found 608.6.

BocNHCN(Boc)-Thr(*t*-Bu)-Phe-Gly-OH (16{12})

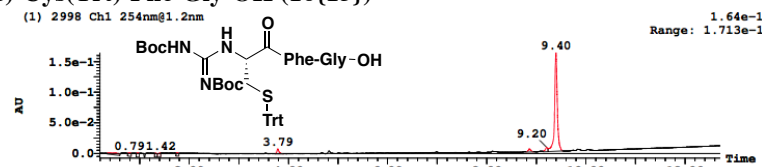
Purity: >95%, $R_f = 9.39$ min. UPLC-MS (ESI) calcd for $C_{30}H_{48}N_5O_9$ $[M + H]^+$ 622.3, found 622.5.

BocNHCN(Boc)-Asn(Trt)-Phe-Gly-OH (16{13})

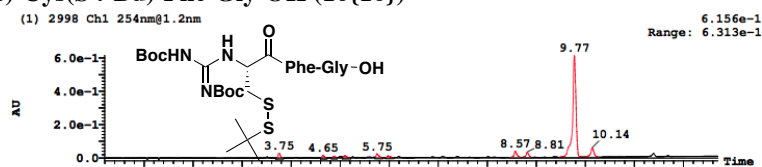
Purity: 78%, $R_f = 9.64$ min. UPLC-MS (ESI) calcd for $C_{45}H_{53}N_6O_9$ $[M + H]^+$ 821.4, found 821.6.

BocNHCN(Boc)-Gln(Trt)-Phe-Gly-OH (16{14})

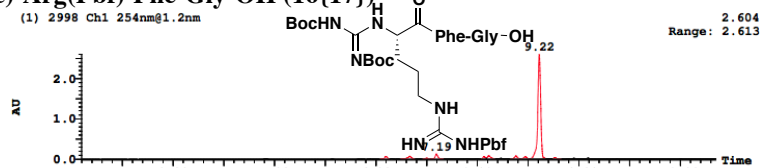
Purity: >95%, R_f = 9.66 min. UPLC-MS (ESI) calcd for $C_{46}H_{55}N_6O_9$ $[M + H]^+$ 835.4, found 835.7.

BocNHCN(Boc)-Cys(Trt)-Phe-Gly-OH (16{15})

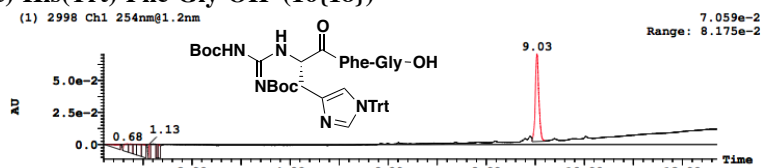
Purity: >95%, R_f = 9.40 min. UPLC-MS (ESI) calcd for $C_{44}H_{52}N_5O_8S$ $[M + H]^+$ 810.4, found 810.6.

BocNHCN(Boc)-Cys(S-*t*-Bu)-Phe-Gly-OH (16{16})

Purity: 79%, R_f = 9.77 min. HRMS (ESI) calcd for $C_{29}H_{46}N_5O_8S_2$ $[M + H]^+$ 656.2782, found 656.2806.

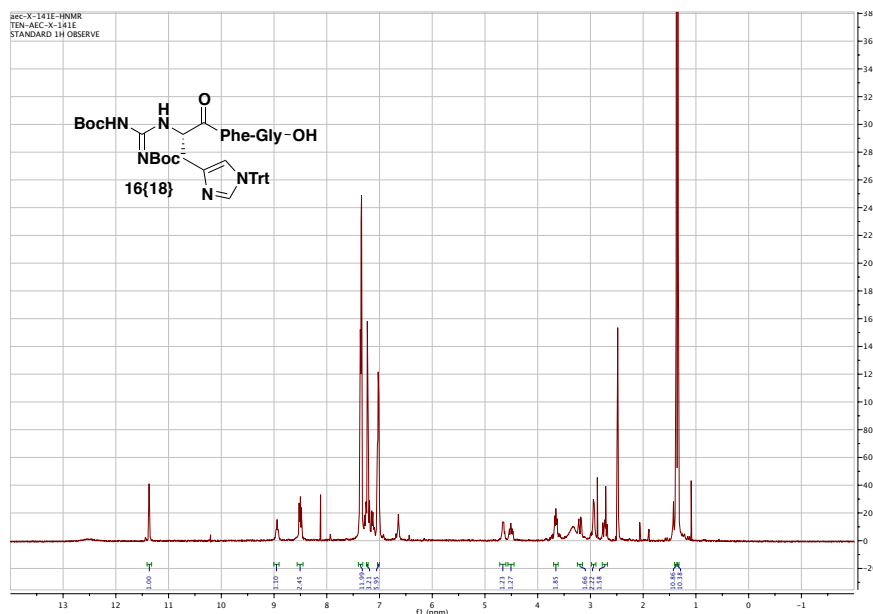
BocNHCN(Boc)-Arg(Pbf)-Phe-Gly-OH (16{17})

Purity: >95%, R_f = 9.22 min. UPLC-MS (ESI) calcd for $C_{41}H_{61}N_8O_{11}S$ $[M + H]^+$ 873.4, found 873.7.

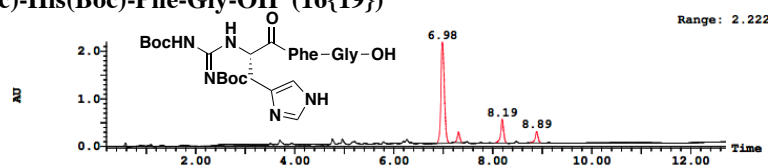
BocNHCN(Boc)-His(Trt)-Phe-Gly-OH (16{18})

Purity: >95%, R_f = 9.03 min. 1H NMR (500 MHz, DMSO- d_6): δ 11.39 (1H, s), 8.96 (1H, t, J = 5.2 Hz), 8.60 – 8.44 (2H, m), 7.42 – 7.33 (1H, m), 7.27 – 7.21 (3H, m), 7.04 (6H, dd, J = 2.9 Hz, 6.7 Hz), 4.67 (1H, d, J = 7.5 Hz), 4.52 (1H, dd, J = 5.6 Hz, 13.7 Hz), 3.67 (2H, t, J = 6.5 Hz), 3.29 (1H, dd, J = 3.8 Hz, 13.8 Hz), 2.98 – 2.93 (2H, m), 2.74 (1H, dd, J = 10.4 Hz, 13.8 Hz), 1.39 (9H, s), 1.36 (9H, s). HRMS (ESI) calcd for $C_{47}H_{54}N_7O_8$ $[M + H]^+$ 844.4028, found 844.4077.

¹H NMR (300 MHz, DMSO-*d*₆)

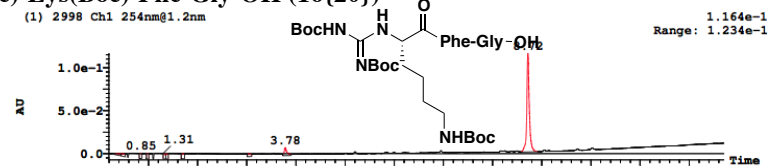


BocNHCN(Boc)-His(Boc)-Phe-Gly-OH (16{19})



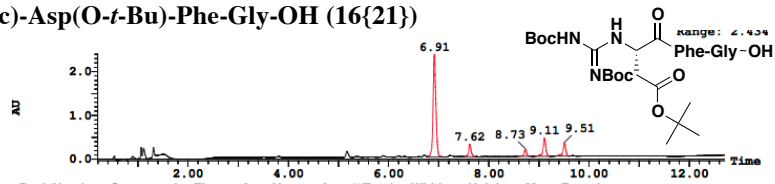
Purity: 89%, R_f = 6.98 min. (The side-chain Boc group was lost during cleavage from the resin).
UPLC-MS (ESI) calcd for $C_{33}H_{48}N_7O_{10}$ $[M - Boc]^+$ 702.3, found 702.6.

BocNHCN(Boc)-Lys(Boc)-Phe-Gly-OH (16{20})



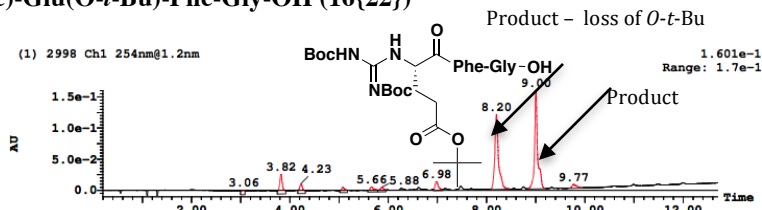
Purity: 91%, R_f = 8.72 min. UPLC-MS (ESI) calcd for $C_{33}H_{53}N_6O_{10}$ $[M + H]^+$ 693.4, found 693.6.

BocNHCN(Boc)-Asp(O-*t*-Bu)-Phe-Gly-OH (16{21})



Purity: 75%, R_f = 6.91 min. (The O-*t*-Bu group was lost during cleavage from the resin). HRMS (ESI) calcd for $C_{26}H_{38}N_5O_{10}$ $[M - tBu]^+$ 580.2613, found 580.2621.

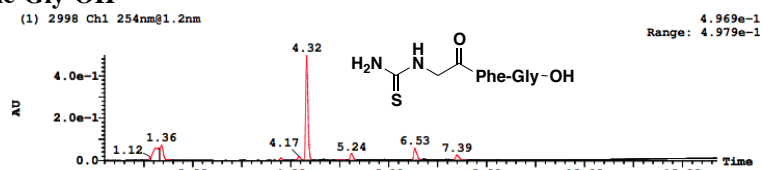
BocNHCN(Boc)-Glu(*O*-*t*-Bu)-Phe-Gly-OH (16{22})



Purity: 90%, $R_f = 9.01$ min. UPLC-MS (ESI) calcd for $C_{31}H_{48}N_5O_{10}$ $[M + H]^+$ 650.3, found 650.6.

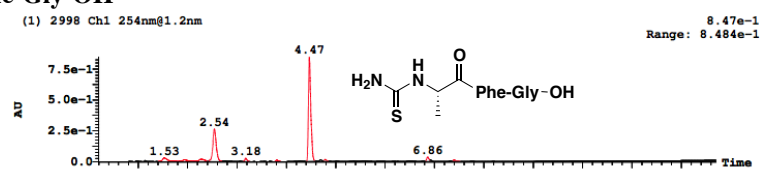
Synthesis of Thiourea and Guanidino Peptides

NH_2CS -Gly-Phe-Gly-OH



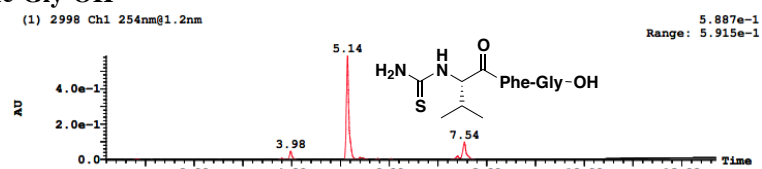
Purity: 76%, $R_f = 4.32$ min. UPLC-MS (ESI) calcd for $C_{14}H_{19}N_4O_4S$ $[M + H]^+$ 339.1, found 339.3.

NH_2CS -Ala-Phe-Gly-OH



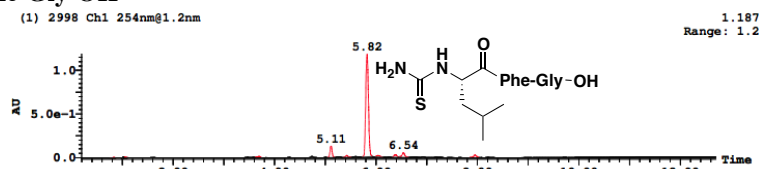
Purity: 93%, $R_f = 4.47$ min. UPLC-MS (ESI) calcd for $C_{15}H_{21}N_4O_4S$ $[M + H]^+$ 353.1, found 353.3.

NH_2CS -Val-Phe-Gly-OH



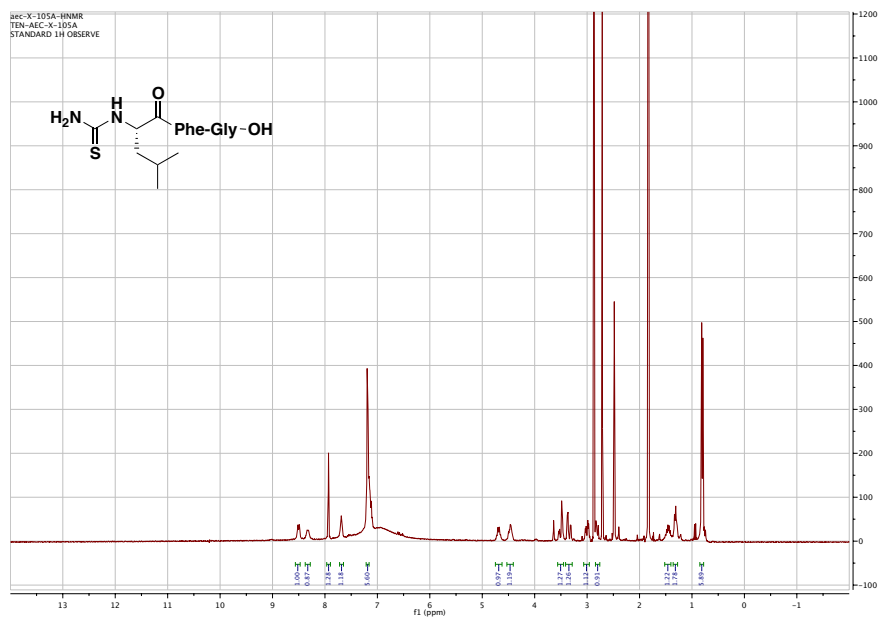
Purity: 88%, $R_f = 5.14$ min. UPLC-MS (ESI) calcd for $C_{17}H_{25}N_4O_4S$ $[M + H]^+$ 381.2, found 381.3.

NH_2CS -Leu-Phe-Gly-OH

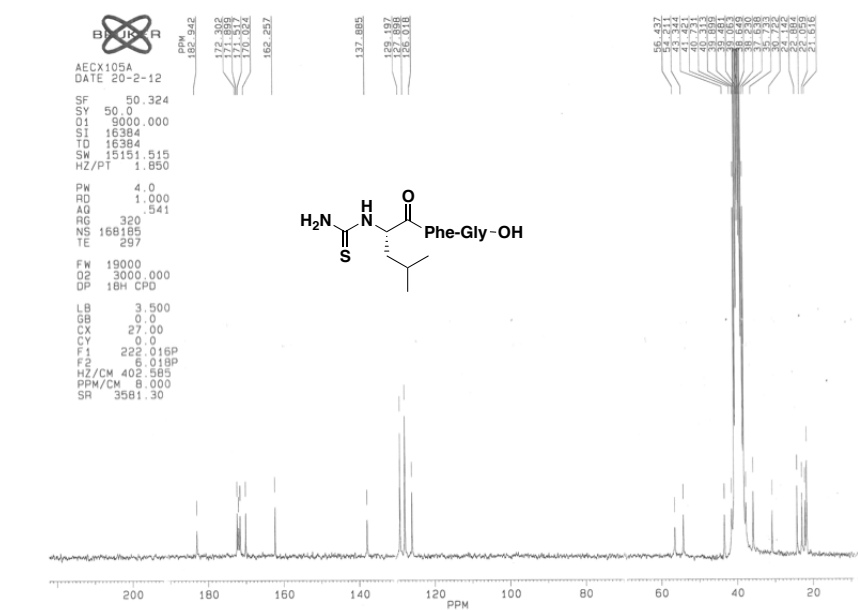


Purity: 92%, $R_f = 5.82$ min. 1H NMR (500 MHz, DMSO- d_6): δ 8.52 (1H, d, $J = 7.6$ Hz), 8.36 (1H, bs), 7.95 (2H, s), 7.71 (1H, bs), 7.28 – 7.10 (5H, m), 4.71 (1H, d, $J = 7.7$ Hz), 4.48 (1H, d, $J = 4.1$ Hz), 3.52 (1H, dd, $J = 5.2$ Hz, 16.8 Hz), 3.36 (1H, dd, $J = 4.1$ Hz, 17.1 Hz), 3.02 (1H, dd, $J = 4.3$ Hz, 13.7 Hz), 2.88 – 2.82 (1H, m), 1.47 (2H, p, $J = 6.2$ Hz), 1.38 – 1.28 (1H, m), 0.80 (6H, d, $J = 6.3$ Hz). ^{13}C NMR (50 MHz, DMSO- d_6): δ 182.9, 172.3, 171.5, 170.0, 137.9, 129.2 (2C), 127.9 (2C), 126.0, 56.4, 54.2, 43.3, 41.4, 37.7, 24.1, 22.1, 21.6. HRMS (ESI) calcd for $C_{18}H_{27}N_4O_4S$ $[M + H]^+$ 395.1748, found 395.1748.

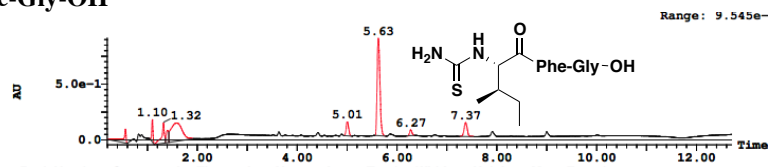
¹H NMR (300 MHz, DMSO-*d*₆)



^{13}C NMR (50 MHz, DMSO- d_6)



NH₂CS-Ile-Phe-Gly-OH

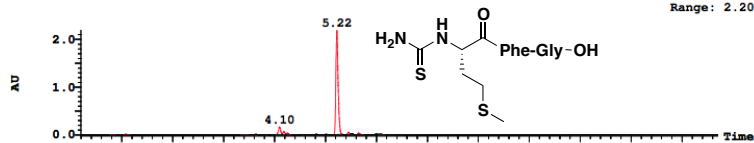


Purity: 91%, R_f = 5.63 min. UPLC-MS (ESI) calcd for $C_{18}H_{27}N_4O_4S$ $[M + H]^+$ 395.2, found 395.4.

NH₂CS-Met-Phe-Gly-OH

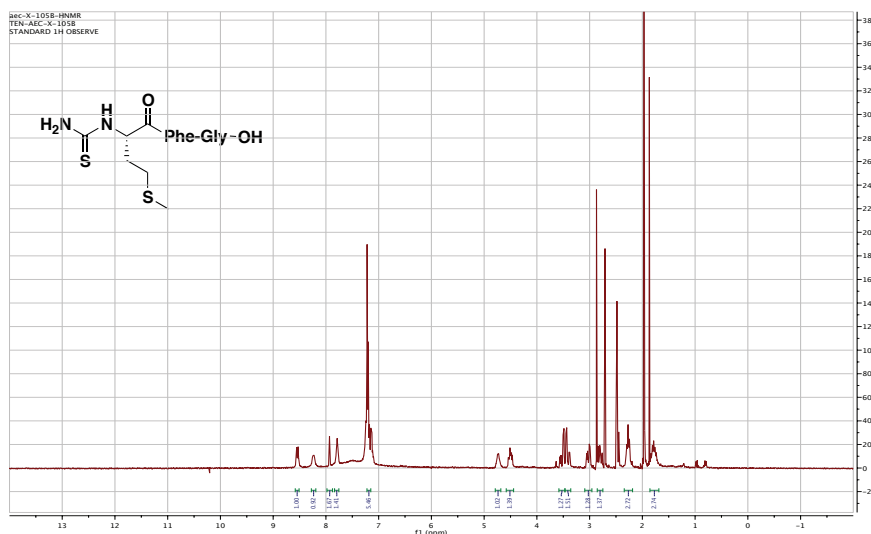
(1) 2998 Ch1 254nm@1.2nm

2.19
Range: 2.204

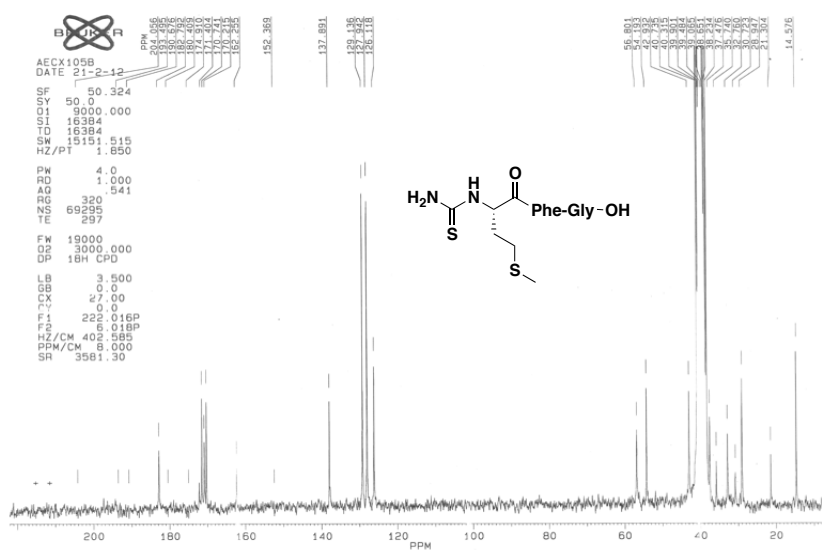


Purity: 92%, $R_f = 5.22$ min. ¹H NMR (500 MHz, DMSO-*d*₆): δ 8.56 (1H, d, $J = 8.4$ Hz), 8.24 (1H, bs), 7.95 (2H, s), 7.80 (1H bs), 7.33 – 7.11 (5H, m), 4.74 (1H, d, $J = 6.8$ Hz), 4.52 (1H, td, $J = 4.2$ Hz, 9.5 Hz), 3.54 (1H, dd, $J = 5.1$ Hz, 17.1 Hz), 3.43 (1H, dd, $J = 4.5$ Hz, 17.0 Hz), 3.04 (1H, dd, $J = 4.4$ Hz, 13.9 Hz), 2.82 (1H, dd, $J = 10.2$ Hz, 13.8 Hz), 2.34 – 2.24 (2H, m), 1.93 (3H, s), 1.88 – 1.71 (2H, m). ¹³C NMR (50 MHz, DMSO-*d*₆): δ 182.8, 171.4, 170.7, 170.2, 137.9, 129.1 (2C), 127.9 (2C), 126.1, 56.8, 54.2, 42.9, 37.5, 32.8, 29.0, 14.6. HRMS (ESI) calcd for C₁₇H₂₅N₄O₄S₂ [M + H]⁺ 413.1312, found 413.1310.

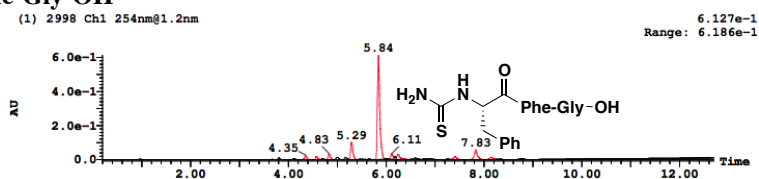
¹H NMR (300 MHz, DMSO-*d*₆)



¹³C NMR (50 MHz, DMSO-*d*₆)

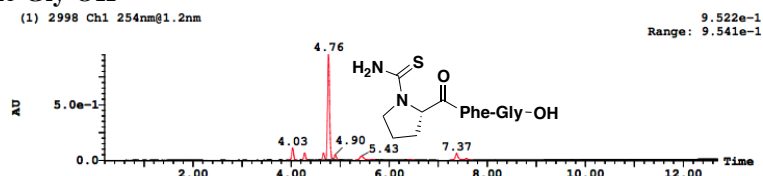


NH₂CS-Phe-Phe-Gly-OH



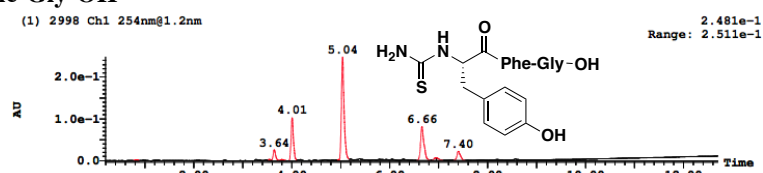
Purity: 79%, $R_f = 5.84$ min. UPLC-MS (ESI) calcd for C₂₁H₂₅N₄O₄S [M + H]⁺ 429.2, found 429.4.

NH₂CS-Pro-Phe-Gly-OH



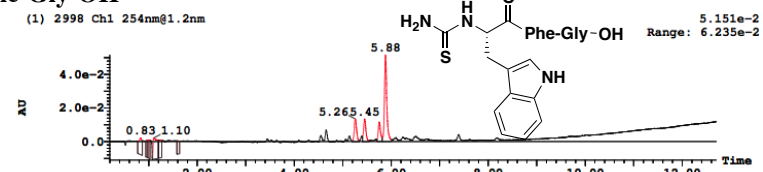
Purity: 91%, $R_f = 4.76$ min. UPLC-MS (ESI) calcd for C₁₇H₂₃N₄O₄S [M + H]⁺ 379.1, found 379.3.

NH₂CS-Tyr-Phe-Gly-OH



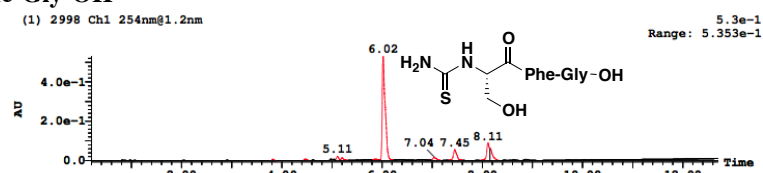
Purity: 76%, $R_f = 5.04$ min. UPLC-MS (ESI) calcd for C₂₁H₂₅N₄O₅S [M + H]⁺ 445.2, found 445.3.

NH₂CS-Trp-Phe-Gly-OH



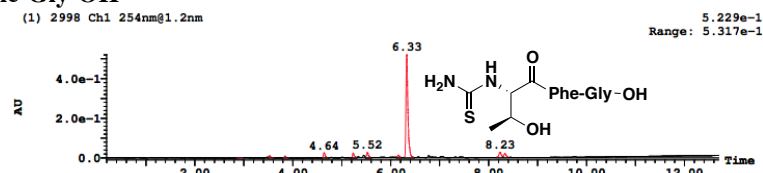
Purity: 75%, $R_f = 5.88$ min. UPLC-MS (ESI) calcd for C₂₃H₂₆N₅O₄S [M + H]⁺ 468.2, found 468.3.

NH₂CS-Ser-Phe-Gly-OH



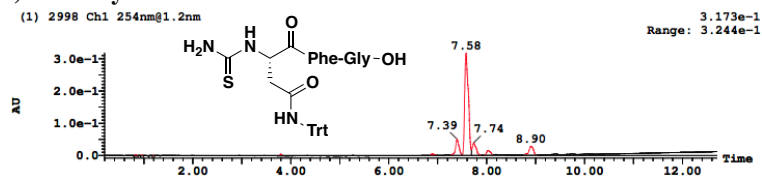
Purity: 76%, $R_f = 6.02$ min. UPLC-MS (ESI) calcd for C₁₅H₂₁N₄O₅S [M + H]⁺ 369.1, found 369.3.

NH₂CS-Thr-Phe-Gly-OH



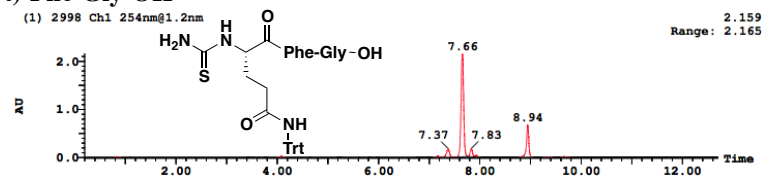
Purity: 83%, $R_f = 6.33$ min. UPLC-MS (ESI) calcd for C₁₆H₂₃N₄O₅S [M + H]⁺ 383.1, found 383.3.

NH₂CS-Asn(Trt)-Phe-Gly-OH



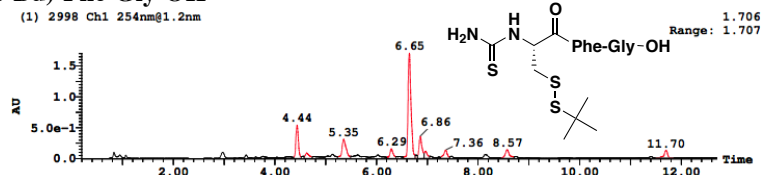
Purity: 94%, $R_f = 7.58$ min. HRMS (ESI) calcd for C₃₅H₃₆N₅O₅S [M + H]⁺ 638.2432, found 638.2432.

NH₂CS-Gln(Trt)-Phe-Gly-OH



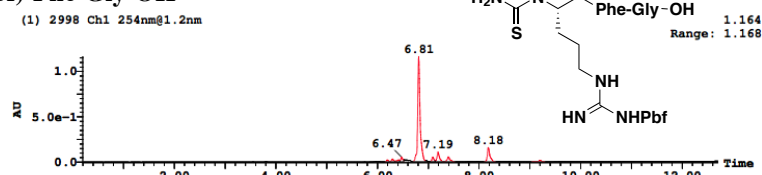
Purity: 72%, $R_f = 7.66$ min. UPLC-MS (ESI) calcd for C₃₆H₃₈N₅O₅S [M + H]⁺ 652.3, found 652.3.

NH₂CS-Cys(S-*t*-Bu)-Phe-Gly-OH



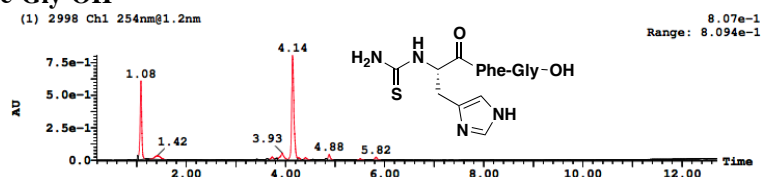
Purity: 71%, $R_f = 6.65$ min. HRMS (ESI) calcd for C₁₉H₂₉N₄O₄S₃ [M + H]⁺ 473.1345, found 473.1353.

NH₂CS-Arg(Pbf)-Phe-Gly-OH



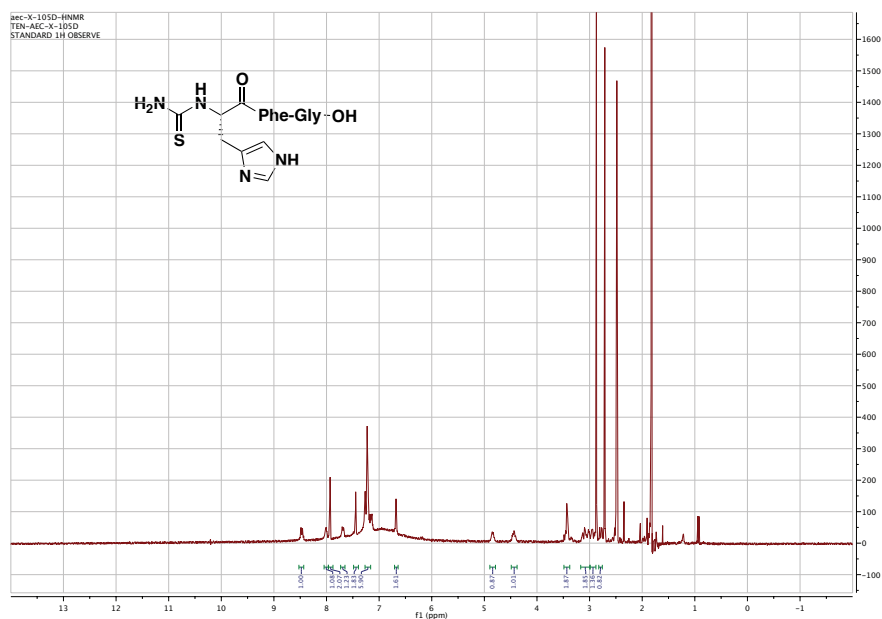
Purity: 78%, $R_f = 6.81$ min. UPLC-MS (ESI) calcd for C₃₁H₄₄N₇O₇S₂ [M + H]⁺ 690.3, found 690.5.

NH₂CS-His-Phe-Gly-OH

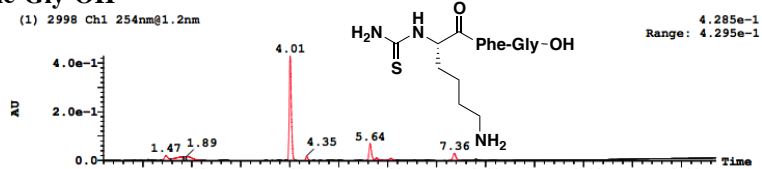


Purity: 83%, $R_f = 4.14$ min. ¹H NMR (500 MHz, DMSO-*d*₆): δ 8.45 (1H, d, $J = 7.7$ Hz), 8.03 (1H, bs), 7.94 (2H, s), 7.65 (1H, d, $J = 6.5$ Hz), 7.45 (1H, s), 7.30 – 7.10 (5H, m), 6.69 (1H, s), 4.90 – 4.81 (1H, m), 4.48 – 4.42 (1H, m), 3.49 (2H, dd, $J = 4.3$ Hz), 3.16 – 3.00 (1H, m), 3.00 – 2.91 (2H, m), 2.83 – 2.76 (1H, m). HRMS (ESI) calcd for C₁₈H₂₃N₆O₄S [M + H]⁺ 419.1496, found 419.1507.

¹H NMR (300 MHz, DMSO-*d*₆)

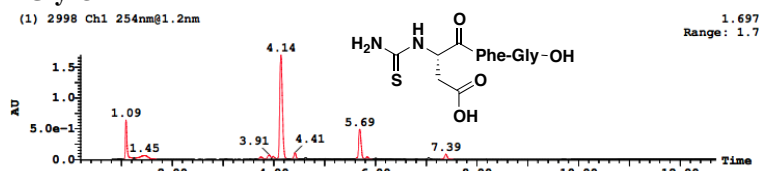


NH₂CS-Lys-Phe-Gly-OH



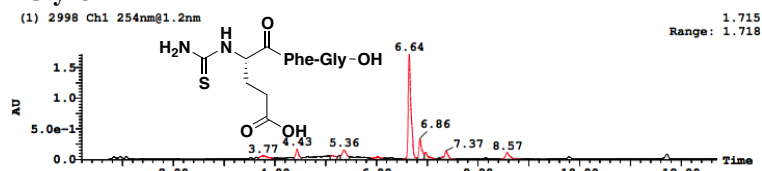
Purity: 90%, *R*_f = 4.01 min. UPLC-MS (ESI) calcd for C₁₈H₂₈N₅O₄S [M + H]⁺ 410.2, found 410.4.

NH₂CS-Asp-Phe-Gly-OH



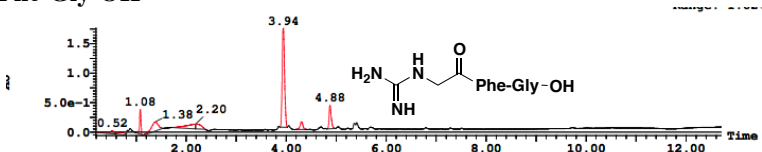
Purity: 76%, *R*_f = 4.14 min. UPLC-MS (ESI) calcd for C₁₆H₂₁N₄O₆S [M + H]⁺ 397.1, found 397.3.

NH₂CS-Glu-Phe-Gly-OH



Purity: 76%, *R*_f = 6.64 min. UPLC-MS (ESI) calcd for C₁₇H₂₃N₅O₆ [M + H]⁺ 411.1, found 411.3.

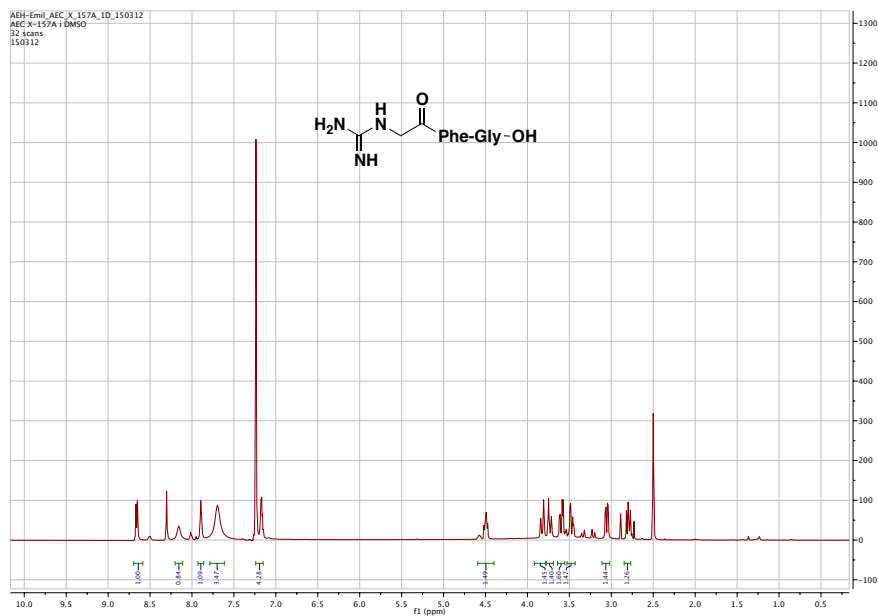
NH₂CNH-Gly-Phe-Gly-OH



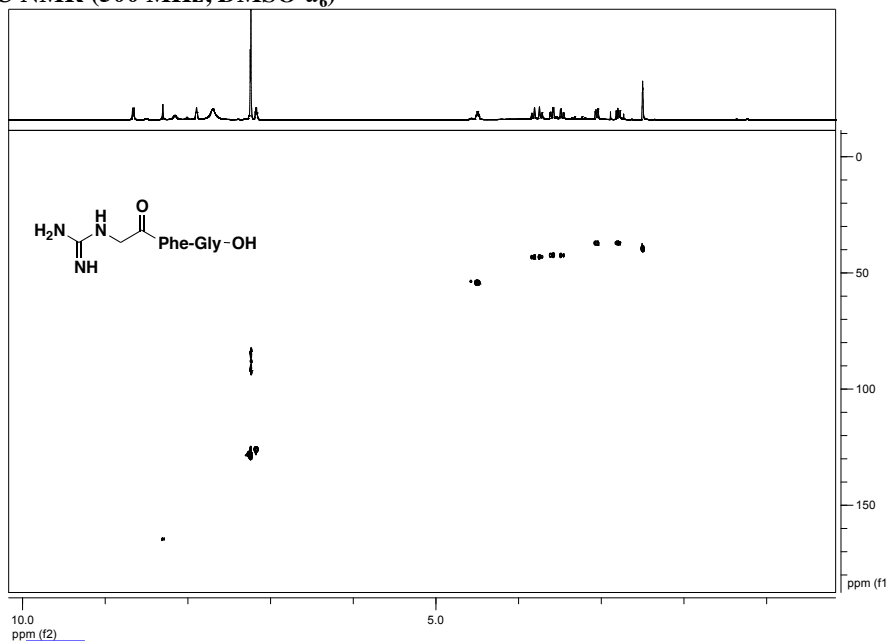
Purity: 83%, *R*_f = 3.94 min. ¹H NMR (500 MHz, DMSO-*d*₆): δ 8.66 (1H, d, *J* = 8.4 Hz), 8.16 (1H, bs), 7.89 (1H, t, *J* = 4.6 Hz), 7.70 (3H, bs), 7.31 – 7.13 (5H, m), 4.50 (1H, td, *J* = 4.4 Hz, 9.3 Hz), 3.82 (1H, ...)

d, $J = 16.8$ Hz), 3.73 (1H, d, $J = 16.8$ Hz), 3.60 (1H, dd, $J = 5.4$ Hz, 17.1 Hz), 3.47 (1H, dd, $J = 4.7$ Hz, 17.0 Hz), 3.06 (1H, dd, $J = 4.3$ Hz, 13.8 Hz), 2.80 (1H, dd, $J = 9.9$ Hz, 13.7 Hz). ^{13}C NMR (125 MHz, $\text{DMSO-}d_6$): δ 172.1, 170.4, 167.4, 157.7, 138.0, 129.4 (2C), 127.9 (2C), 126.0, 54.3, 43.2, 42.5, 37.3. HRMS (ESI) calcd for $\text{C}_{14}\text{H}_{20}\text{N}_5\text{O}_4$ $[\text{M} + \text{H}]^+$ 322.1510, found 322.1513.

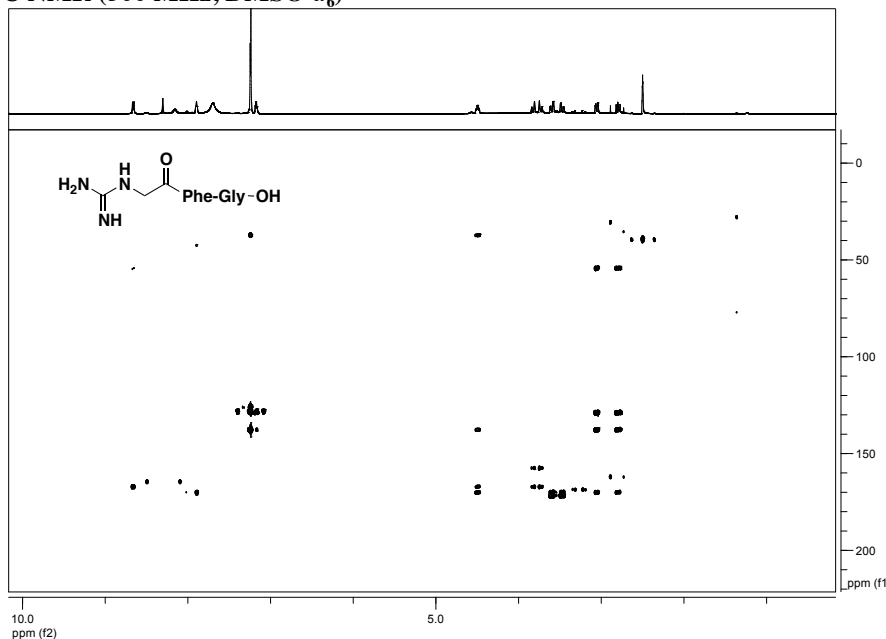
^1H NMR (500 MHz, $\text{DMSO-}d_6$)



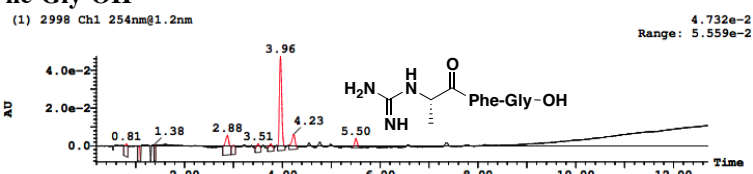
2D HSQC NMR (500 MHz, $\text{DMSO-}d_6$)



2D HMBC NMR (500 MHz, DMSO-*d*₆)

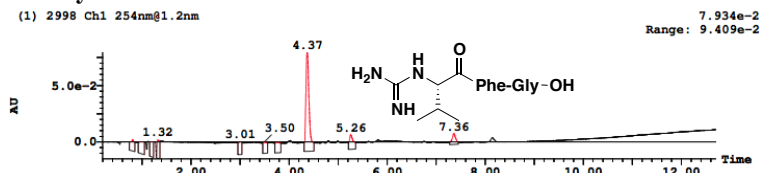


NH₂CNH-Ala-Phe-Gly-OH



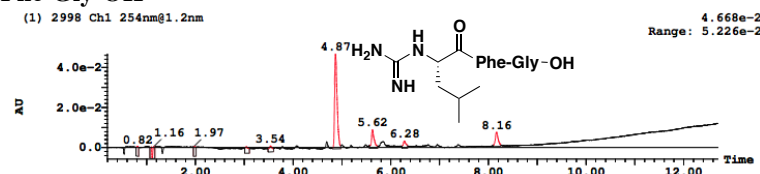
Purity: 74%, $R_f = 3.96$ min. UPLCMS (ESI) calcd for C₁₅H₂₂N₅O₄ [M + H]⁺ 336.2, found 336.4.

NH₂CNH-Val-Phe-Gly-OH



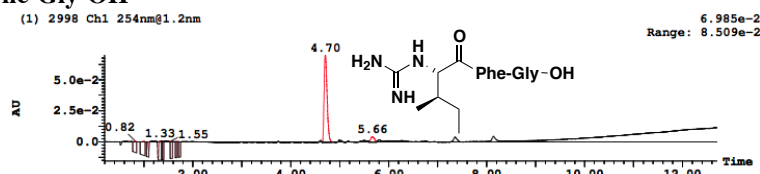
Purity: 76%, $R_f = 4.37$ min. UPLC-MS (ESI) calcd for C₁₇H₂₆N₅O₄ [M + H]⁺ 364.2, found 364.4.

NH₂CNH-Leu-Phe-Gly-OH



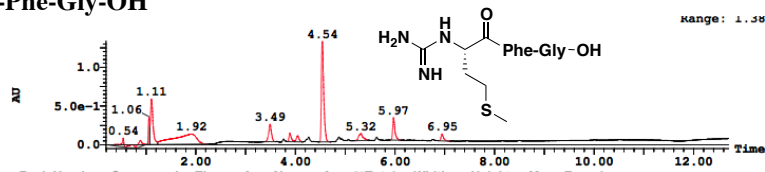
Purity: 84%, $R_f = 4.87$ min. UPLC-MS (ESI) calcd for C₁₈H₂₈N₅O₄ [M + H]⁺ 378.2, found 378.4.

NH₂CNH-Ile-Phe-Gly-OH



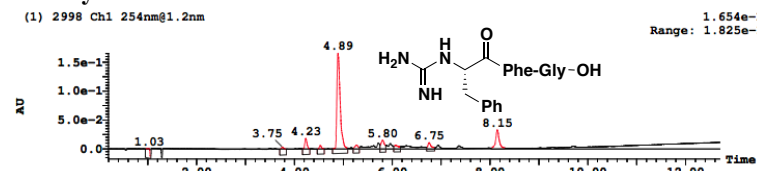
Purity: >95%, $R_f = 4.70$ min. UPLC-MS (ESI) calcd for C₁₈H₂₈N₅O₄ [M + H]⁺ 378.2, found 378.3.

NH₂CNH-Met-Phe-Gly-OH



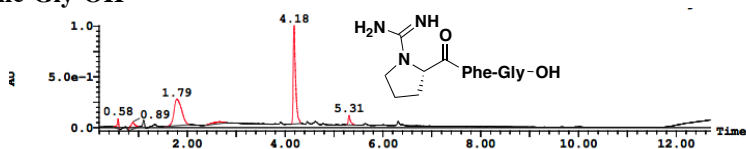
Purity: 71%, $R_f = 4.54$ min. UPLC-MS (ESI) calcd for C₁₇H₂₆N₅O₄S [M + H]⁺ 396.2, found 396.4.

NH₂CNH-Phe-Phe-Gly-OH



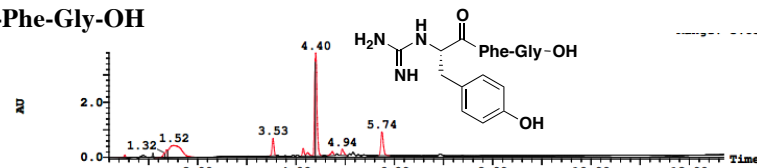
Purity: 75%, $R_f = 4.89$ min. UPLC-MS (ESI) calcd for C₂₁H₂₆N₅O₄ [M + H]⁺ 412.2, found 412.3.

NH₂NH-Pro-Phe-Gly-OH



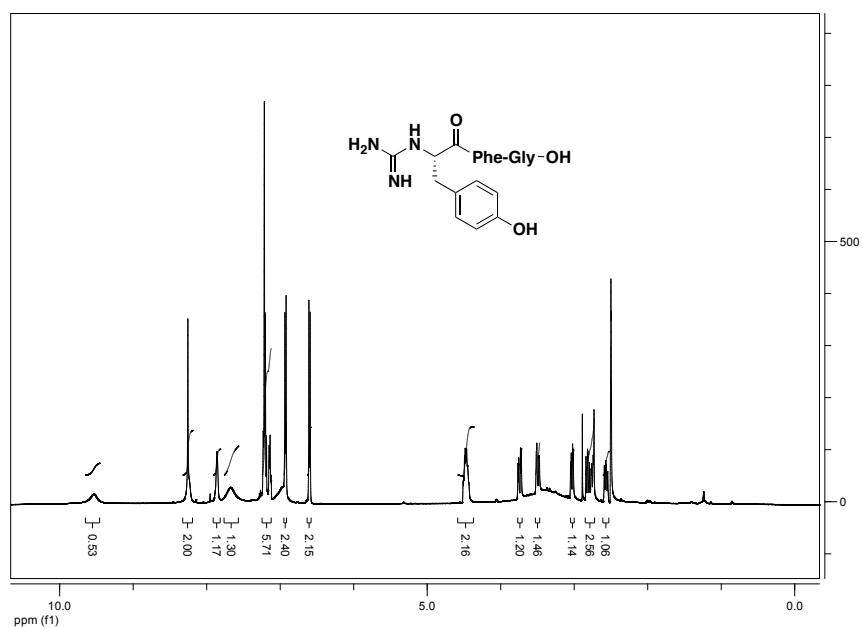
Purity: 69%, $R_f = 4.18$ min. UPLC-MS (ESI) calcd for C₁₇H₂₄N₅O₄ [M + H]⁺ 362.2, found 362.3.

NH₂CNH-Tyr-Phe-Gly-OH

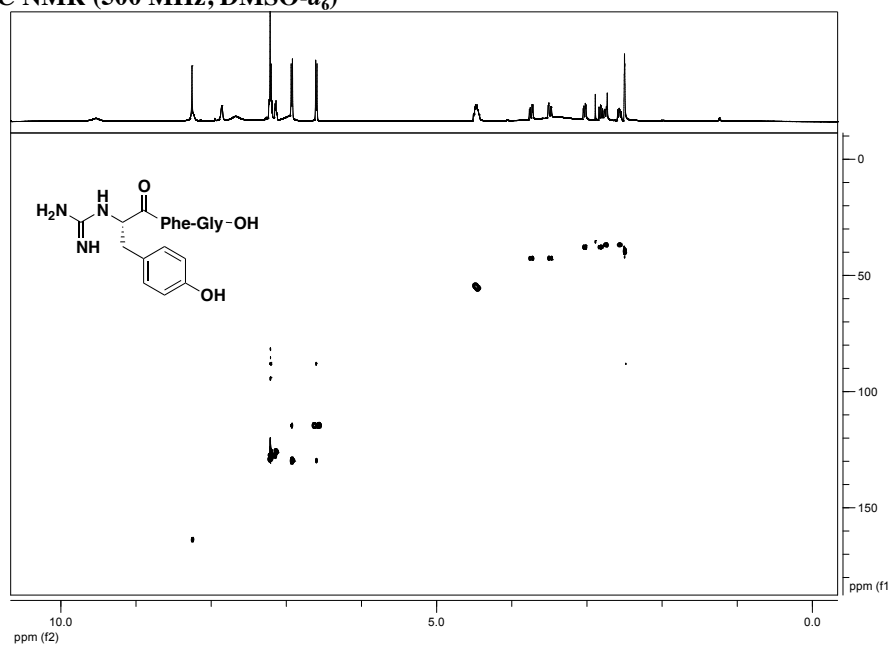


Purity: 65%, $R_f = 4.40$ min. ¹H NMR (500 MHz, DMSO-*d*₆): δ 9.54 (1h, bs), 8.30 – 8.07 (2H, m), 7.86 (1H, bs), 7.68 (1H, bs), 7.32 – 7.10 (m, 6H), 7.08 – 6.89 (3H, m), 6.60 (2H, d, *J* = 8.3 Hz), 4.59 – 4.39 (2H, m), 3.74 (1H, dd, *J* = 5.8 Hz, 17.2 Hz), 3.49 (1H, dd, *J* = 2.8 Hz, 16.8 Hz), 3.03 (2H, dd, *J* = 4.3 Hz, 13.5 Hz), 2.86 – 2.71 (2H, m), 2.57 (1H, dd, *J* = 9.1 Hz, 13.8 Hz). ¹³C NMR (125 MHz, DMSO-*d*₆): δ 172.1, 170.0, 169.5, 169.0, 155.7, 137.6, 129.7 (2C), 128.2 (2C), 126.4, 127.4 (2C), 125.9, 114.4 (2C), 55.4, 54.4, 42.5, 37.5, 36.7. HRMS (ESI) calcd for C₂₁H₂₆N₅O₅ [M + H]⁺ 428.1928, found 428.1951.

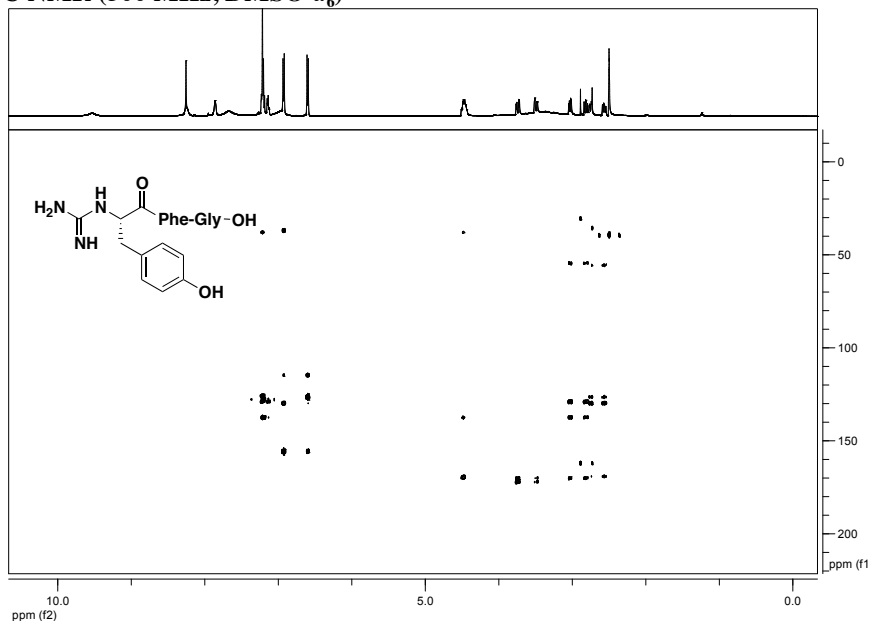
¹H NMR (500 MHz, DMSO-*d*₆)



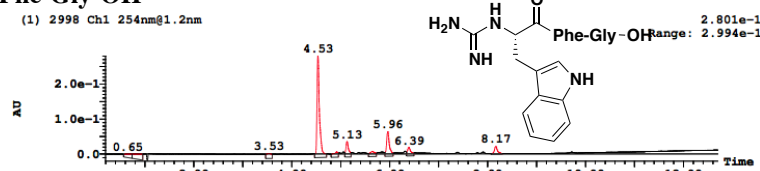
2D HSQC NMR (500 MHz, DMSO-*d*₆)



2D HMBC NMR (500 MHz, DMSO-*d*₆)

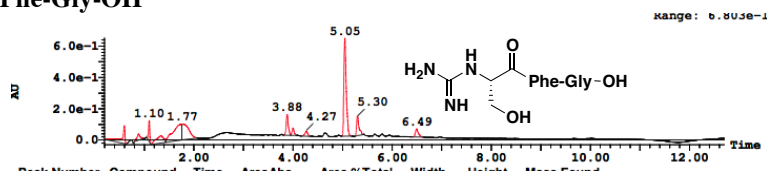


NH₂CNH-Trp-Phe-Gly-OH



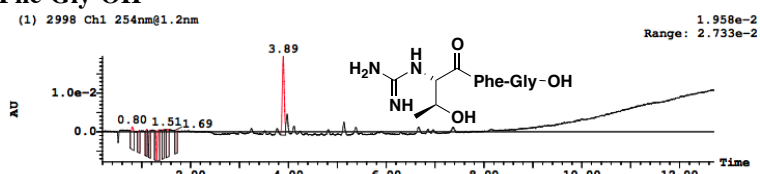
Purity: 72%, R_f = 4.53 min. UPLC-MS (ESI) calcd for C₂₃H₂₇N₆O₄ [M + H]⁺ 451.2, found 451.4.

NH₂CNH-Ser-Phe-Gly-OH



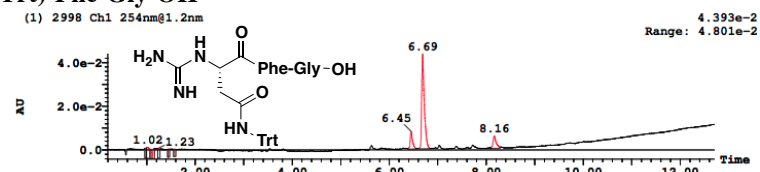
Purity: 72%, R_f = 5.05 min. UPLC-MS (ESI) calcd for C₁₅H₂₂N₃O₅ [M + H]⁺ 352.2, found 352.4.

NH₂CNH-Thr-Phe-Gly-OH



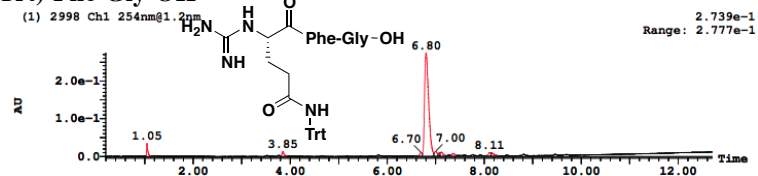
Purity: 90%, R_f = 3.89 min. UPLC-MS (ESI) calcd for C₁₆H₂₄N₃O₅ [M + H]⁺ 366.2, found 366.4.

NH₂CNH-Asn(Trt)-Phe-Gly-OH



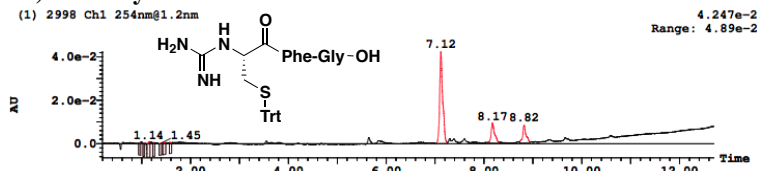
Purity: 85%, R_f = 6.69 min. UPLC-MS (ESI) calcd for C₃₅H₃₇N₆O₅ [M + H]⁺ 621.3, found 621.5.

NH₂CNH-Gln(Trt)-Phe-Gly-OH



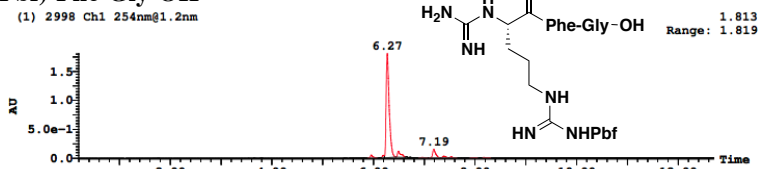
Purity: 85%, $R_f = 6.80$ min. HRMS (ESI) calcd for C₃₆H₃₉N₆O₅ [M + H]⁺ 635.2976, found 635.2993.

NH₂CNH-Cys(Trt)-Phe-Gly-OH



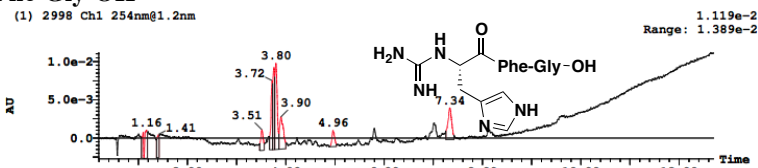
Purity: 89%, $R_f = 7.12$ min. HRMS (ESI) calcd for C₃₄H₃₆N₅O₄S [M + H]⁺ 610.2483, found 610.2478.

NH₂CNH-Arg(Pbf)-Phe-Gly-OH



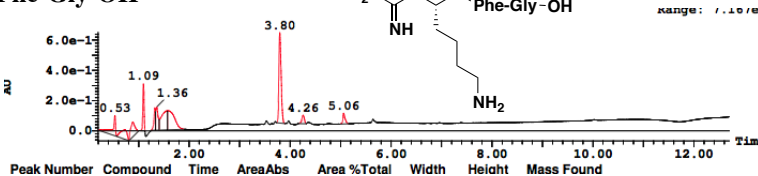
Purity: 92%, $R_f = 6.27$ min. HRMS (ESI) calcd for C₃₁H₄₅N₈O₇S [M + H]⁺ 673.3126, found 673.3131.

NH₂CNH-His-Phe-Gly-OH



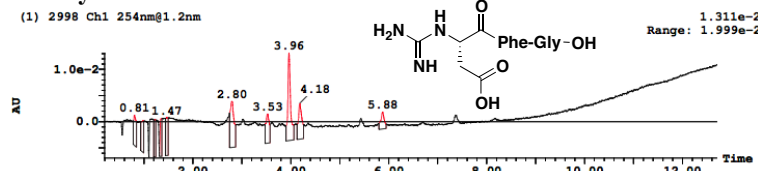
Purity: 73%, $R_f = 3.81$ min. UPLC-MS (ESI) calcd for C₁₈H₂₄N₇O₄ [M + H]⁺ 402.2, found 402.4.

NH₂CNH-Lys-Phe-Gly-OH



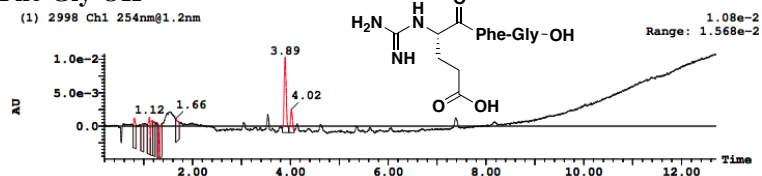
Purity: 82%, $R_f = 3.80$ min. UPLC-MS (ESI) calcd for C₁₈H₂₉N₆O₄ [M + H]⁺ 393.2, found 393.3.

NH₂CNH-Asp-Phe-Gly-OH



Purity: 72%, $R_f = 3.96$ min. UPLC-MS (ESI) calcd for C₁₆H₂₂N₅O₆ [M + H]⁺ 380.2, found 380.4.

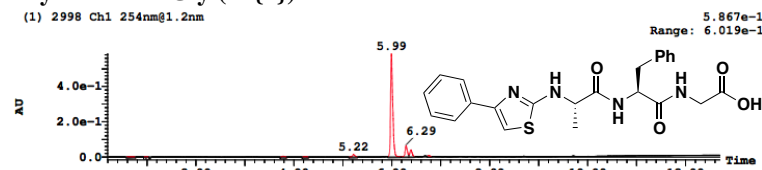
NH₂CNH-Glu-Phe-Gly-OH



Purity: 88%, $R_f = 3.89$ min. UPLC-MS (ESI) calcd for C₁₇H₂₄N₅O₆ [M + H]⁺ 394.2, found 394.4

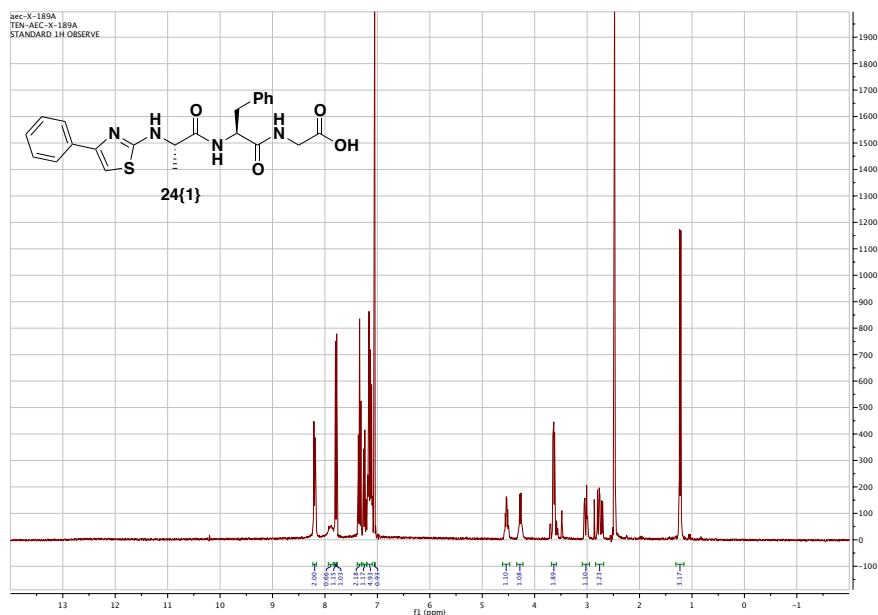
Synthesis of Thiazoles

4-Phenylthiazol-2-yl-Ala-Phe-Gly (24{1})

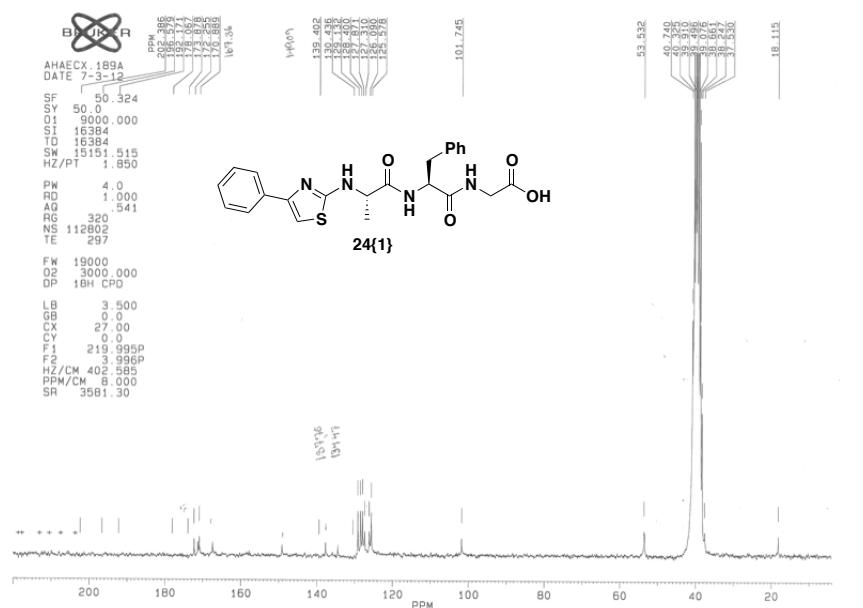


Purity: 92%, $R_f = 5.99$ min. ¹H NMR (300 MHz, DMSO-*d*₆): δ 8.26 – 8.18 (2H, m), 7.90 (1H, bs), 7.81 (2H, dd, $J = 1.3$ Hz, 8.3 Hz), 7.42 – 7.32 (2H, m), 7.27 (1H, tt, $J = 1.8$ Hz, 4.6 Hz), 7.22 – 7.11 (5H, m), 7.07 (1H, s), 4.56 (1H, td, $J = 4.4$ Hz, 9.4 Hz), 4.30 (1H, q, $J = 6.8$ Hz), 3.66 (2H, dd, $J = 4.3$ Hz, 5.7 Hz), 3.05 (1H, dd, $J = 4.4$ Hz, 13.8 Hz), 2.79 (1H, dd, $J = 9.7$ Hz, 13.8 Hz), 1.25 (3H, d, $J = 7.0$ Hz). ¹³C NMR (50 MHz, DMSO-*d*₆): δ 172.3, 171.3, 170.9, 167.4, 149.1, 137.8, 134.5, 130.4 (2C), 129.1 (2C), 128.4 (2C), 127.3, 126.1, 125.6 (2C), 101.7, 53.5 (2C), 37.6, 18.1. HRMS (ESI) calcd for C₂₃H₂₅N₄O₄S [M + H]⁺ 453.1591, found 453.1607.

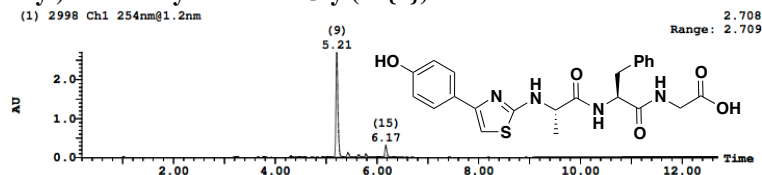
¹H NMR (300 MHz, DMSO-*d*₆)



¹³C NMR (50 MHz, DMSO-*d*₆)

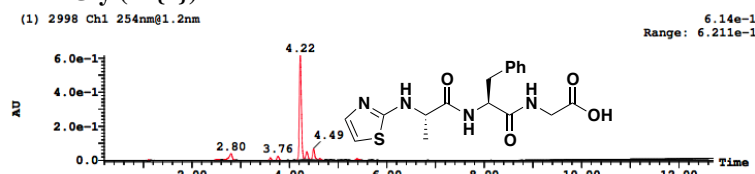


4-(4-hydroxyphenyl)thiazol-2-yl-Ala-Phe-Gly (24{2})



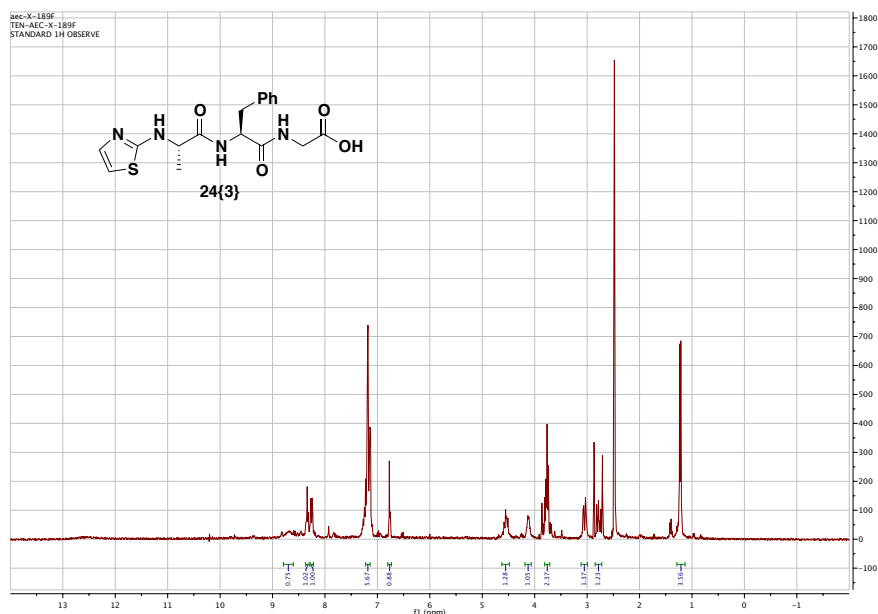
Purity: 90%, $R_f = 5.21$ min. UPLC-MS (ESI) calcd for $C_{23}H_{25}N_4O_5S$ $[M + H]^+$ 469.2, found 469.2.

Thiazol-2-yl-Ala-Phe-Gly (24{3})

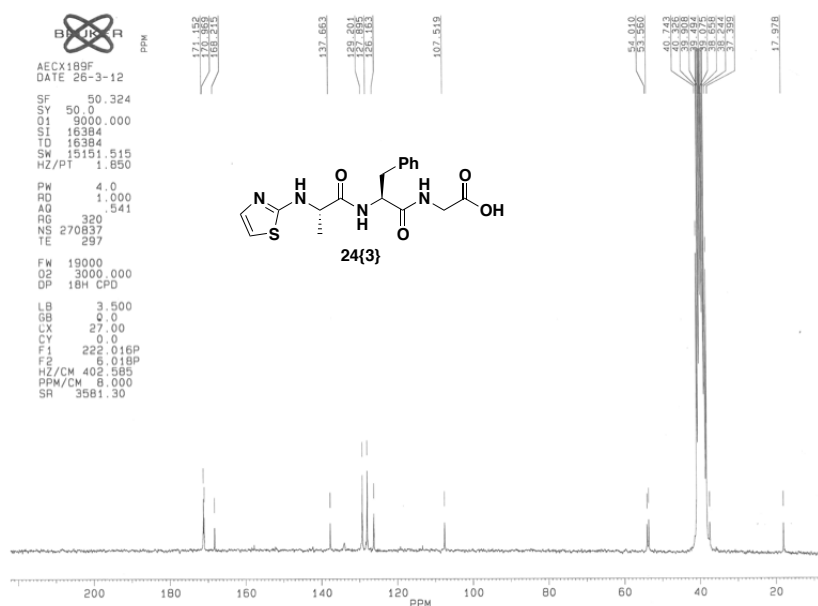


Purity: 87%, $R_f = 4.22$ min. ¹H NMR (300 MHz, DMSO-*d*₆): δ 8.70 (1H, bs), 8.36 (1H, t, $J = 5.7$ Hz), 8.28 (1H, d, $J = 8.3$ Hz), 7.37 – 7.08 (5H, m), 6.79 (1H, d, $J = 3.9$ Hz), 4.57 (1H, td, $J = 4.2$ Hz, 9.3 Hz), 4.15 (1H, bs), 3.78 (2H, t, $J = 6.0$ Hz), 3.07 (1H, dd, $J = 4.1$ Hz, 13.8 Hz), 2.80 (1H, dd, $J = 9.9$ Hz, 13.8 Hz), 1.24 (3H, d, $J = 6.9$ Hz). ¹³C NMR (50 MHz, DMSO-*d*₆): δ 171.2 (2C), 171.0, 168.2, 137.7, 129.2 (2C), 127.9 (2C), 126.2, 107.5, 54.1, 53.6, 37.4, 18.0. HRMS (ESI) calcd for $C_{17}H_{21}N_4O_4S$ $[M + H]^+$ 377.1278, found 377.1285.

¹H NMR (300 MHz, DMSO-*d*₆)

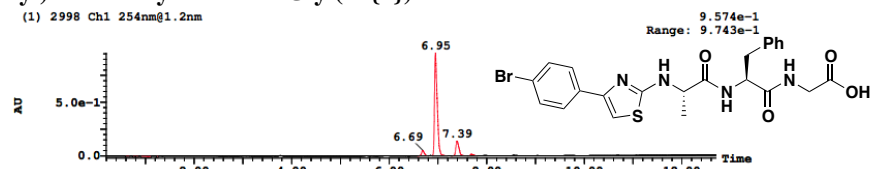


^{13}C NMR (50 MHz, DMSO- d_6)



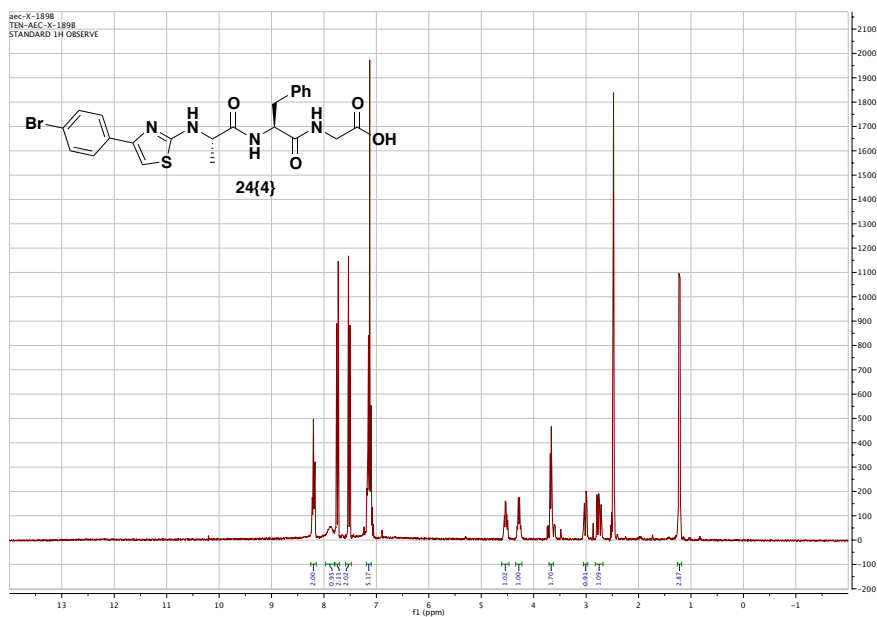
4-(4-Bromophenyl)thiazol-2-yl-Ala-Phe-Gly (24{4})

(1) 2998 Ch1 254nm@1.2nm

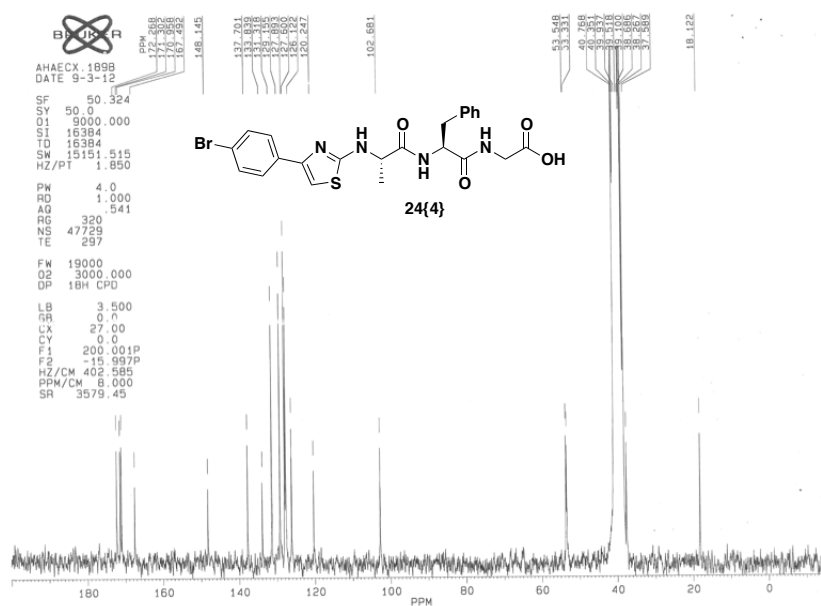


Purity: 87%, R_f = 6.95 min. ^1H NMR (300 MHz, $\text{DMSO}-d_6$): δ 8.26 – 8.18 (2H, m), 7.89 (1H, bs), 7.82 – 7.73 (2H, m), 7.60 – 7.49 (2H, m), 7.25 – 7.10 (5H, m), 4.56 (1H, td, J = 4.3 Hz, 9.2 Hz), 4.30 (1H, dd, J = 7.0 Hz), 3.69 (2H, dd, J = 4.7 Hz, 5.5 Hz), 3.04 (1H, dd, J = 4.3 Hz, 13.8 Hz), 2.78 (1H, dd, J = 9.7 Hz, 13.8 Hz), 1.24 (3H, d, J = 7.0 Hz, 3H). ^{13}C NMR (50 MHz, $\text{DMSO}-d_6$): δ 172.3, 171.3, 171.0, 167.5, 148.1, 137.7, 133.8, 131.3 (2C), 129.2 (2C), 127.9 (2C), 127.6 (2C), 126.1, 120.2, 102.7, 53.5, 53.3, 37.6, 18.1. HRMS (ESI) calcd for $\text{C}_{27}\text{H}_{24}\text{BrN}_4\text{O}_4\text{S}$ [$\text{M} + \text{H}$] $^+$ 531.0696, found 531.0698.

¹H NMR (300 MHz, DMSO-*d*₆)

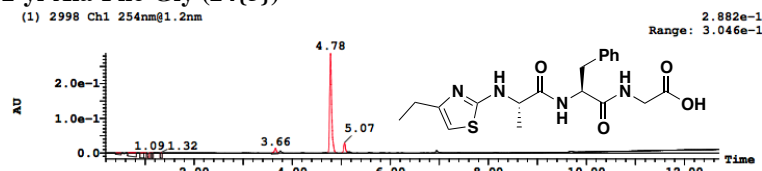


¹³C NMR (50 MHz, DMSO-*d*₆)



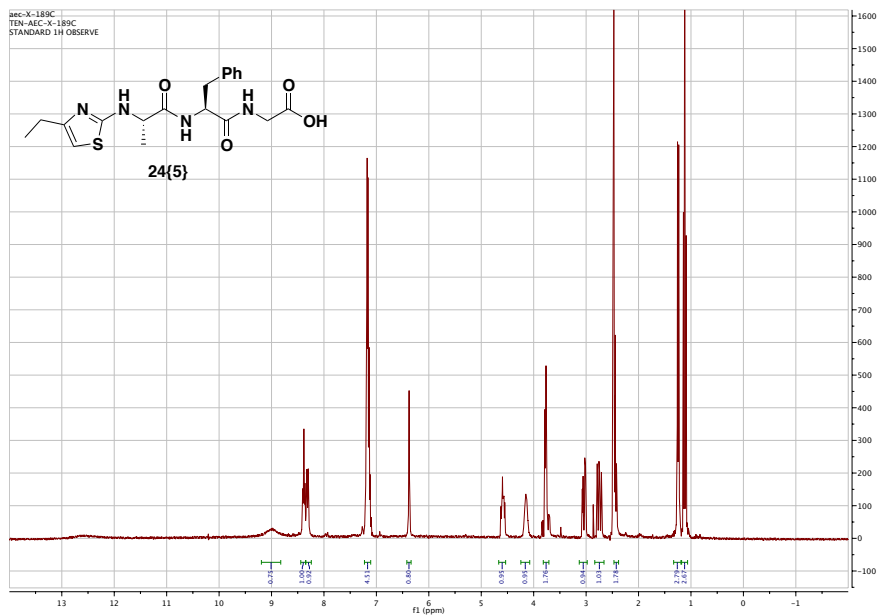
4-Ethylthiazol-2-yl-Ala-Phe-Gly (24{5})

(1) 2998 Ch1 254nm@1.2nm

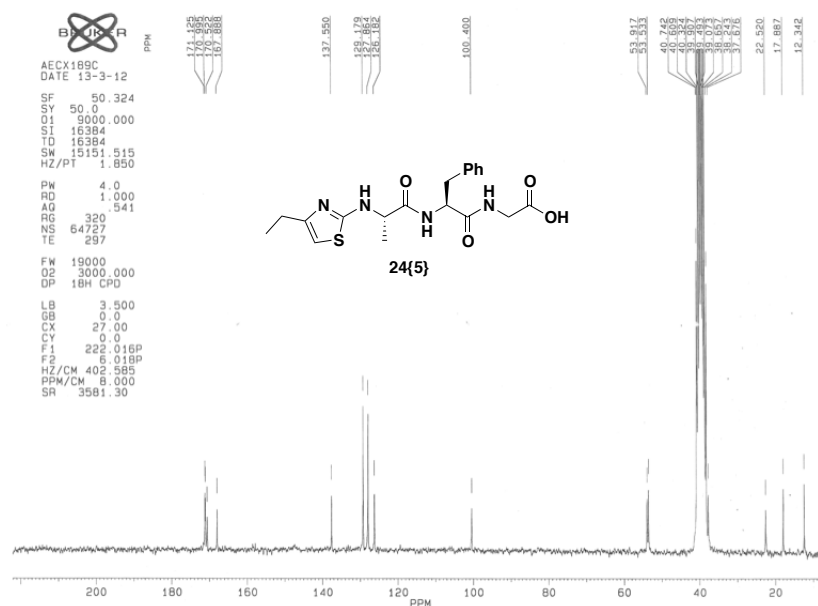


Purity: 93%, R_f = 4.78 min. ¹H NMR (300 MHz, DMSO-*d*₆): δ 9.02 (1H, bs), 8.41 (1H, t, J = 5.9 Hz), 8.34 (1H, d, J = 8.4 Hz), 7.27 – 7.11 (5H, m), 6.40 (1H, s), 4.61 (1H, td, J = 4.3 Hz, 9.5 Hz), 4.17 (1H, bs, 1H), 3.79 (2H, dd, J = 3.7 Hz, 5.7 Hz), 3.06 (1H, dd, J = 4.2 Hz, 13.8 Hz), 2.77 (1H, dd, J = 10.3 Hz, 14.0 Hz), 2.53 – 2.43 (2H, m), 1.27 (3H, d, J = 6.9 Hz), 1.14 (3H, t, J = 7.5 Hz). ¹³C NMR (50 MHz, DMSO-*d*₆): δ 171.1, 171.0, 170.5, 167.9, 137.6, 129.2 (2C), 127.9 (2C), 126.2, 100.4, 53.9, 53.5, 37.7, 22.5, 17.9, 12.3. HRMS (ESI) calcd for C₁₉H₂₅N₄O₄S [M + H]⁺ 405.1591, found 405.1613.

¹H NMR (300 MHz, DMSO-*d*₆)

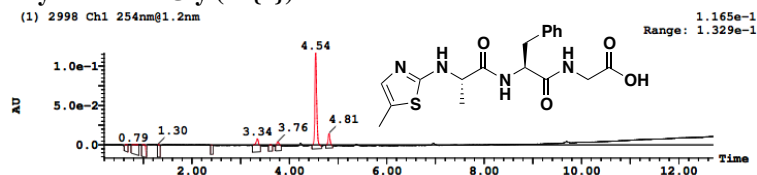


¹³C NMR (50 MHz, DMSO-*d*₆)



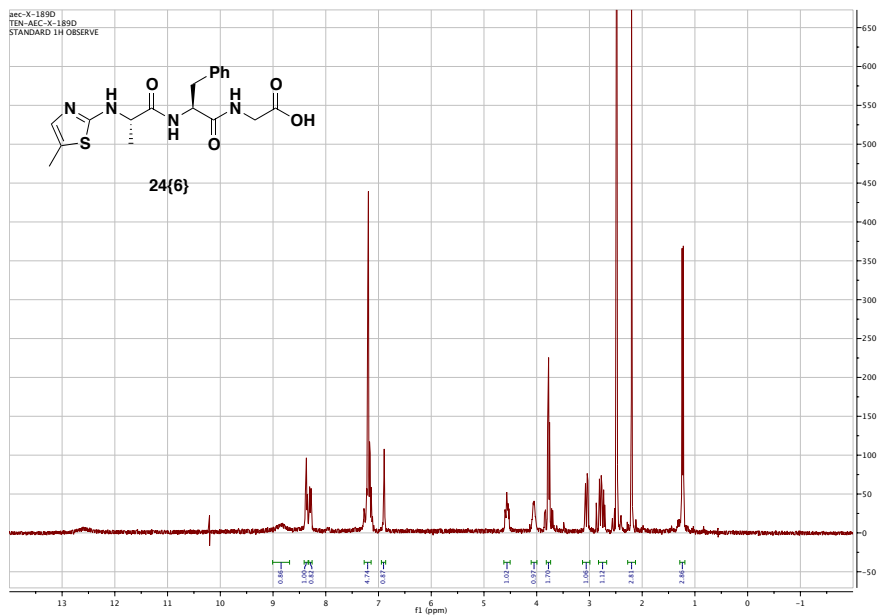
5-Mehtylthiazol-2-yl-Ala-Phe-Gly (24(6))

(1) 2998 Ch1 254nm@1.2nm

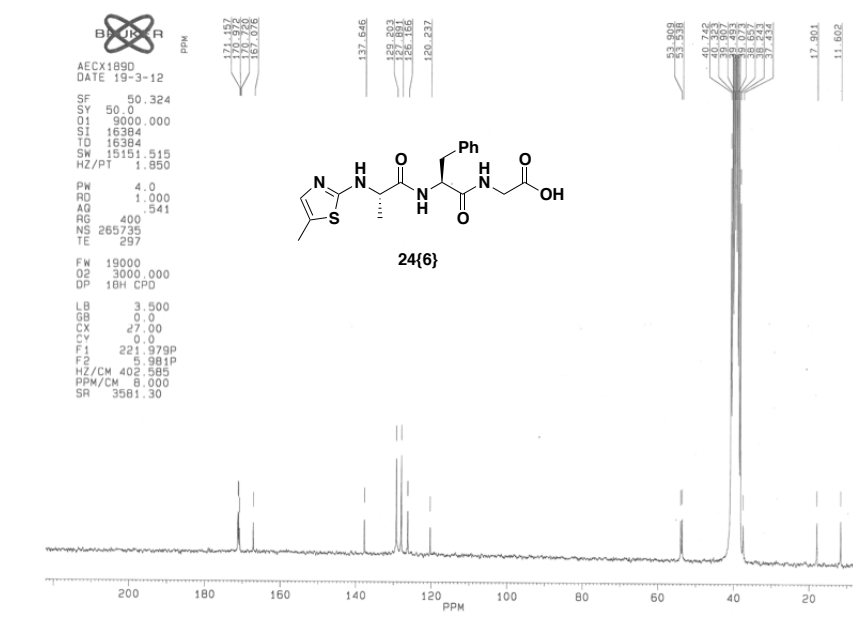


Purity: 84%, R_f = 4.54 min. ¹H NMR (300 MHz, DMSO-*d*₆): δ 8.85 (1H, bs), 8.38 (1H, t, J = 5.8 Hz), 8.31 (1H, d, J = 8.3 Hz), 7.28 – 7.11 (5H, m), 6.91 (1H, s), 4.58 (1H, td, J = 4.2 Hz, 9.8 Hz), 4.07 (1H, bs), 3.79 (2H, t, J = 5.6 Hz), 3.08 (1H, dd, J = 4.1 Hz, 13.8 Hz), 2.79 (1H, dd, J = 10.1 Hz, 13.9 Hz), 2.22 (3H, d, J = 1.4 Hz), 1.25 (3H, d, J = 6.9 Hz). ¹³C NMR (50 MHz, DMSO-*d*₆): δ 171.2, 171.0, 170.7, 167.1, 137.6 (2C), 129.2 (2C), 127.9 (2C), 126.2, 120.2, 53.9, 53.5, 37.4, 17.9, 11.6. HRMS (ESI) calcd for C₁₈H₂₃N₄O₄S [M + H]⁺ 391.1435, found 391.1439.

¹H NMR (300 MHz, DMSO-*d*₆)

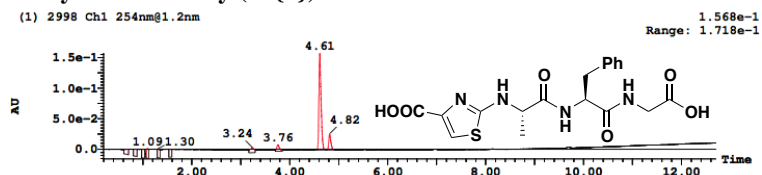


¹³C NMR (50 MHz, DMSO-*d*₆)



4-Carboxythiazol-2-yl-Ala-Phe-Gly (24{7})

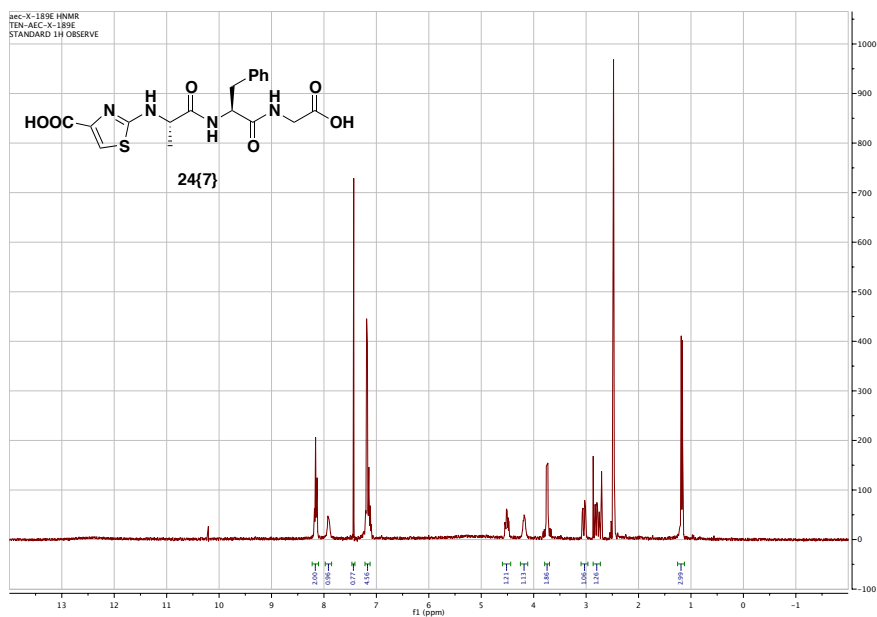
(1) 2998 Ch1 254nm@1.2nm



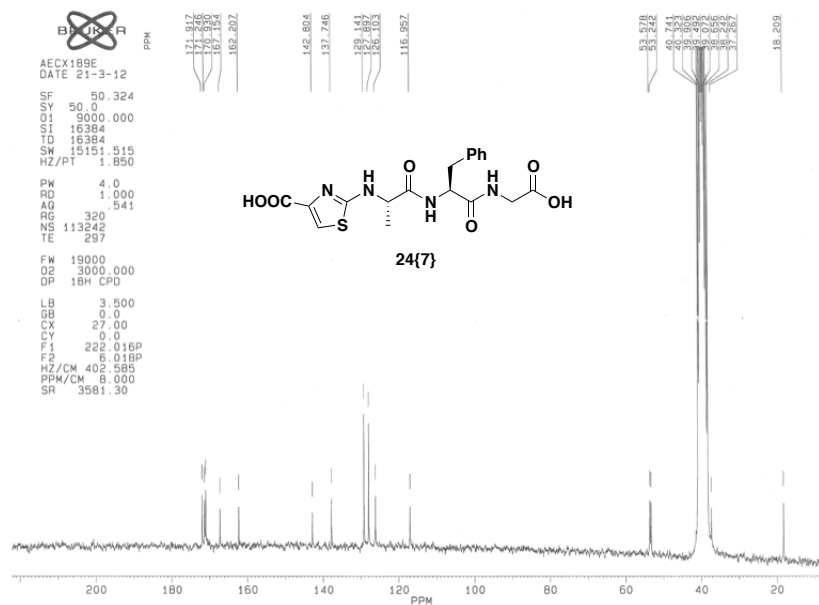
Purity: 88%, R_f = 4.61 min. ¹H NMR (300 MHz, DMSO-*d*₆): δ 8.22 – 8.14 (2H, m), 7.95 (1H, bs), 7.45 (1H, s), 7.28 – 7.10 (5H, m), 4.54 (1H, td, J = 4.1 Hz, 9.0 Hz), 4.20 (1H, bs), 3.77 (2H, dd, J = 3.3 Hz, 5.7 Hz), 3.07 (1H, dd, J = 4.2 Hz, 13.9 Hz), 2.81 (1H, dd, J = 9.8 Hz, 13.9 Hz), 1.20 (3H, d, J = 7.0 Hz). ¹³C NMR (50 MHz, DMSO-*d*₆): δ 171.9, 171.2, 170.9, 167.2, 162.2, 142.8, 137.7, 129.1 (2C),

127.9 (2C), 126.1, 117.0, 53.6, 53.2, 37.3, 18.2. HRMS (ESI) calcd for $C_{18}H_{21}N_4O_6S$ $[M + H]^+$ 421.1176, found 421.1171.

1H NMR (300 MHz, DMSO- d_6)



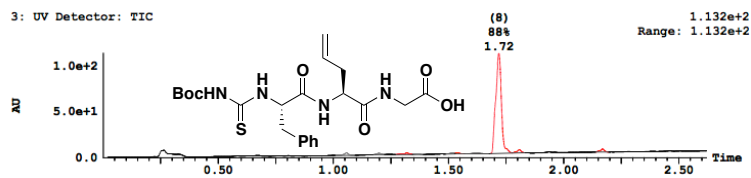
^{13}C NMR (50 MHz, DMSO- d_6)



Synthesis of Thiazole-Containing Macrocycles

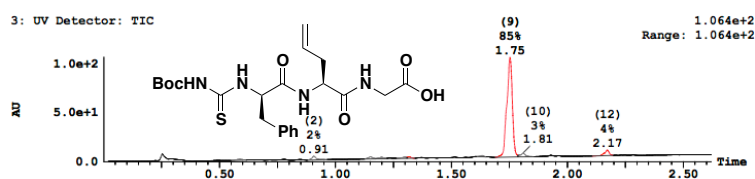
Precursors

BocNHCS-L-Phe-L-Gly(allyl)-Gly-OH



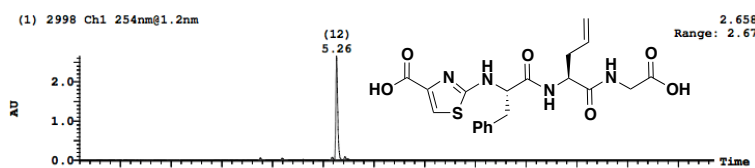
Purity: >95%, $R_f = 1.72$ min. (LCMS) HRMS (ESI) calcd for $C_{22}H_{31}N_4O_6S$ $[M + H]^+$ 479.1959, found 479.1963.

BocNHCS-D-Phe-L-Gly(allyl)-Gly-OH



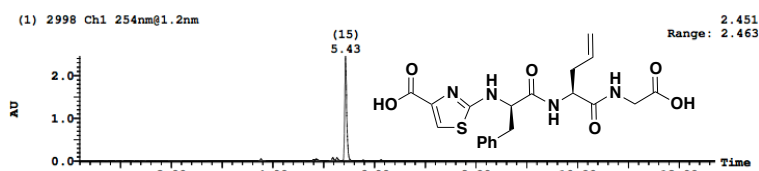
Purity: 91%, $R_f = 1.75$ min. (LCMS) UPLC-MS (ESI) calcd for $C_{22}H_{31}N_4O_6S$ $[M + H]^+$ 479.2, found 479.3.

4-Carboxythiazol-2-yl-L-Phe-L-Gly(allyl)-Gly-OH (26{1} (S,S))



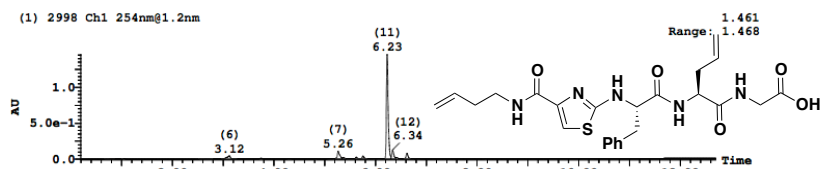
Purity: >95%, $R_f = 5.26$ min. UPLC-MS (ESI) calcd for $C_{20}H_{23}N_4O_6S$ $[M + H]^+$ 447.1, found 447.2.

4-Carboxythiazol-2-yl-D-Phe-L-Gly(allyl)-Gly-OH (26{2} (R,S))



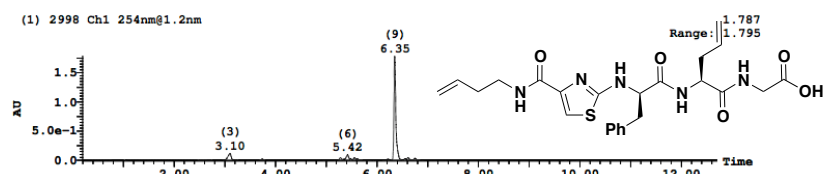
Purity: >95%, $R_f = 5.43$ min. HRMS (ESI) calcd for $C_{20}H_{23}N_4O_6S$ $[M + H]^+$ 447.1333, found 447.1340.

(4-(But-3-en-yl)-4-carbamoyl)-thiazol-2-yl-L-Phe-L-Gly(allyl)-Gly-OH



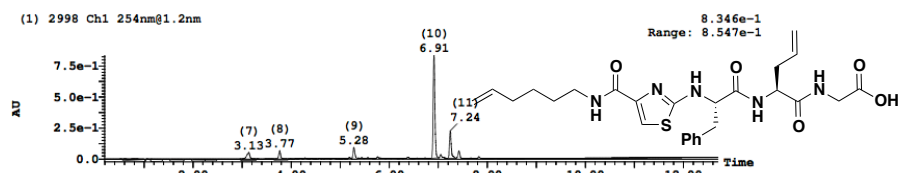
Purity: 78%, $R_f = 6.23$ min. UPLC-MS (ESI) calcd for $C_{24}H_{30}N_5O_5S$ $[M + H]^+$ 500.2, found 500.3.

4-(But-3-en-yl)-4-carbamoyl-thiazol-2-yl-D-Phe-L-Gly(allyl)-Gly-OH



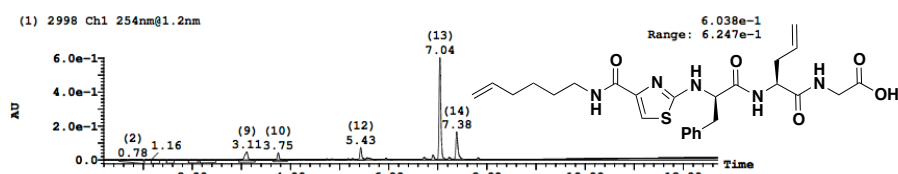
Purity: 83%, R_f = 6.35 min. UPLC-MS (ESI) calcd for $C_{24}H_{30}N_5O_5S$ $[M + H]^+$ 500.2, found 500.3.

4-(Hex-5-en-yl)-4-carbamoyl-thiazol-2-yl-L-Phe-L-Gly(allyl)-Gly-OH



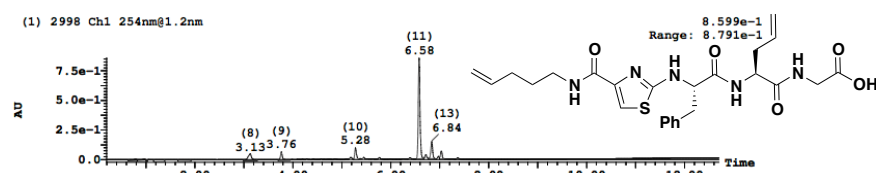
Purity: 63%, R_f = 6.91 min. UPLC-MS (ESI) calcd for $C_{26}H_{34}N_5O_5S$ $[M + H]^+$ 528.2, found 528.3.

4-(Hex-5-en-yl)-4-carbamoyl-thiazol-2-yl-D-Phe-L-Gly(allyl)-Gly-OH



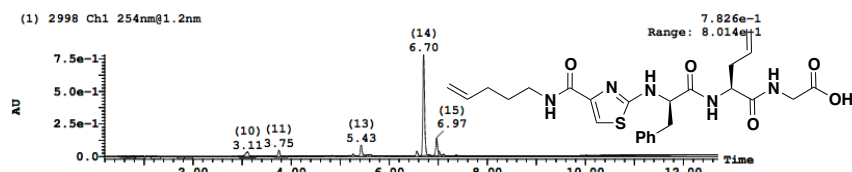
Purity: 60%, R_f = 7.04 min. HRMS (ESI) calcd for $C_{26}H_{34}N_5O_5S$ $[M + H]^+$ 528.2275, found 528.2280.

4-(Pent-4-en-yl)-4-carbamoyl-thiazol-2-yl-L-Phe-L-Gly(allyl)-Gly-OH



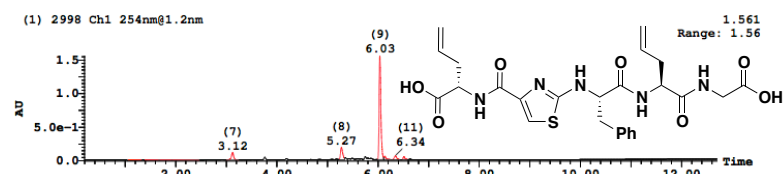
Purity: 70%, R_f = 6.58 min. UPLC-MS (ESI) calcd for $C_{25}H_{32}N_5O_5S$ $[M + H]^+$ 514.2, found 514.3.

4-(Pent-4-en-yl)-4-carbamoyl-thiazol-2-yl-D-Phe-L-Gly(allyl)-Gly-OH



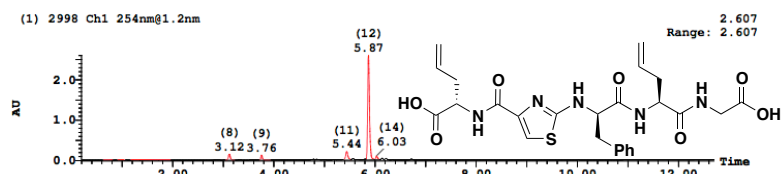
Purity: 68%, R_f = 6.70 min. HRMS (ESI) calcd for $C_{25}H_{32}N_5O_5S$ $[M + H]^+$ 514.2119, found 514.2112.

L-Gly(allyl)-4-carbamoyl-thiazol-2-yl-L-Phe-L-Gly(allyl)-Gly-OH



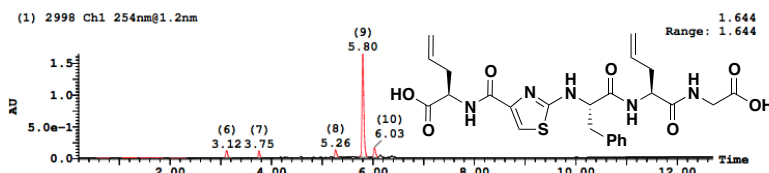
Purity: 83%, R_f = 6.03 min. UPLC-MS (ESI) calcd for $C_{25}H_{30}N_5O_7S$ $[M + H]^+$ 544.2, found 544.3.

L-Gly(allyl)-4-carbamoyl-thiazol-2-yl-D-Phe-L-Gly(allyl)-Gly-OH



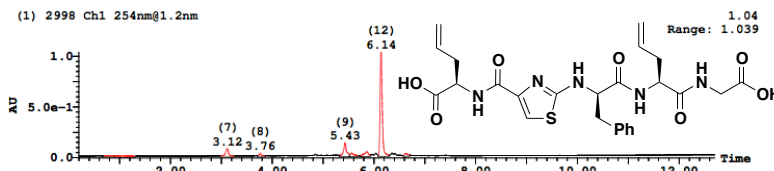
Purity: 88%, R_f = 5.87 min. UPLC-MS (ESI) calcd for $C_{25}H_{30}N_5O_7S$ $[M + H]^+$ 544.2, found 544.3.

D-Gly(allyl)-4-carbamoyl-thiazol-2-yl-L-Phe-L-Gly(allyl)-Gly-OH



Purity: 84%, R_f = 5.80 min. UPLC-MS (ESI) calcd for $C_{25}H_{30}N_5O_7S$ $[M + H]^+$ 544.2, found 544.3.

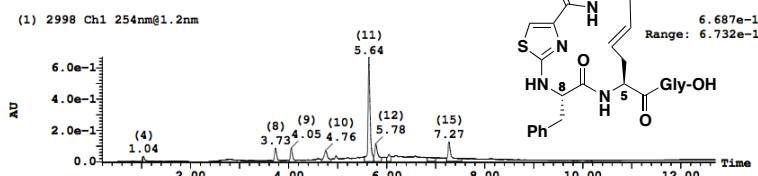
D-Gly(allyl)-4-carbamoyl-thiazol-2-yl-D-Phe-L-Gly(allyl)-Gly-OH



Purity: 87%, R_f = 6.14 min. UPLC-MS (ESI) calcd for $C_{25}H_{30}N_5O_7S$ $[M + H]^+$ 544.2, found 544.3.

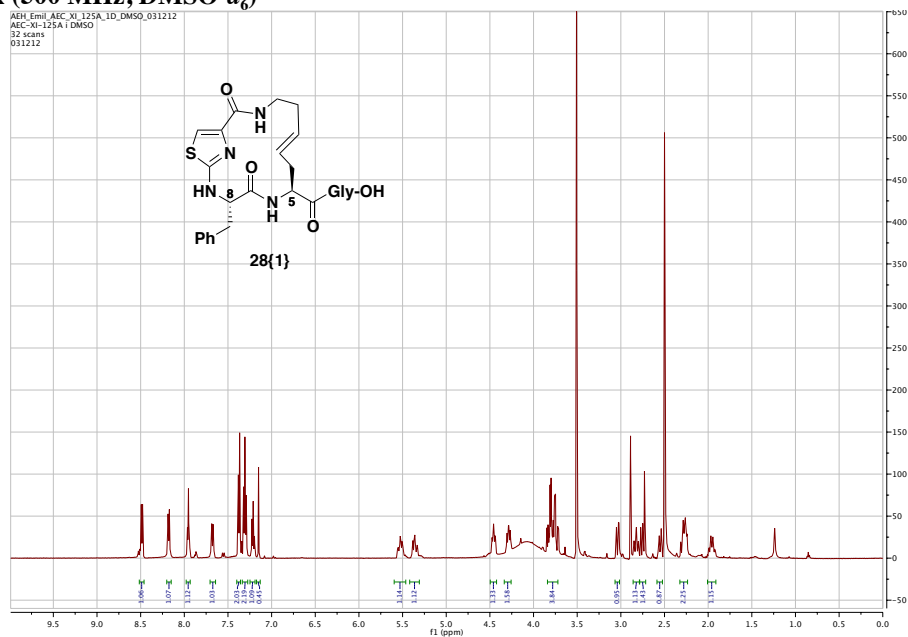
Macrocycles

Macrocycle 28{1} (5S, 8S)

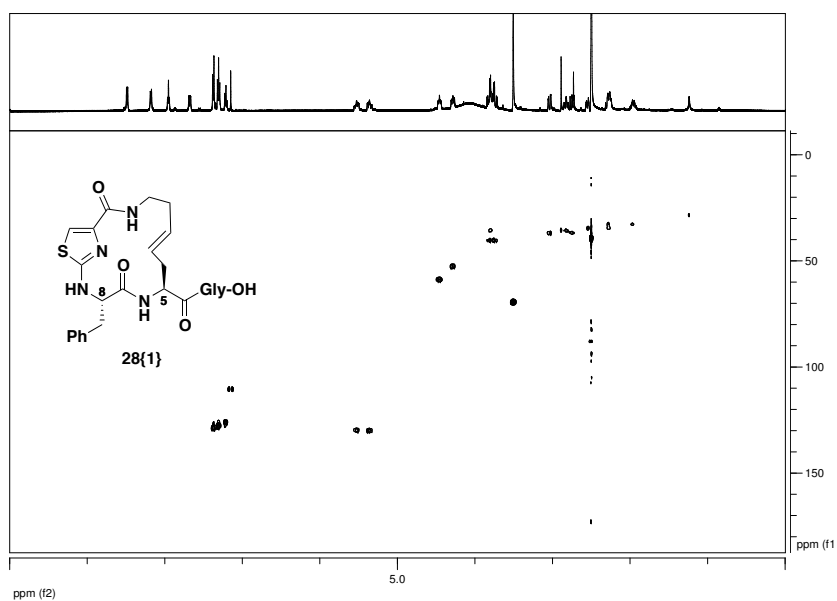


Purity: 71%, R_f = 5.64 min. 1H NMR (500 MHz, $DMSO-d_6$): δ 8.49 (1H, d, J = 7.4 Hz), 7.95 (1H, t, J = 5.8 Hz), 7.68 (1H, dd, J = 9.8, 2.1 Hz), 7.37 (2H, d, J = 7.3 Hz), 7.31 (2H, t, J = 7.6 Hz), 7.21 (1H, t, J = 7.3 Hz), 7.15 (1H, s), 5.53 (1H, ddd, J = 15.1, 10.0, 2.8 Hz), 5.36 (1H, ddd, J = 14.9, 10.0, 2.3 Hz), 4.46 (1H, ddd, J = 11.1, 7.6, 2.4 Hz), 4.29 (1H, ddd, J = 12.0, 7.4, 2.6 Hz), 3.85–3.70 (3H, m), 3.04 (1H, dd, J = 14.2, 2.7 Hz), 2.83 (1H, dd, J = 17.7, 7.0 Hz), 2.79 – 2.70 (1H, m), 2.58 – 2.52 (1H, m), 2.33 – 2.22 (2H, m), 2.01 – 1.91 (1H, m). ^{13}C NMR (125 MHz, $DMSO-d_6$): δ 172.8, 171.7, 171.2, 166.2, 159.8, 144.8, 137.9, 129.8, 129.7, 128.6 (2C), 127.6 (2C), 125.8, 110.4, 58.7, 52.7, 40.3, 36.9, 35.6, 34.6, 32.7, HRMS (ESI) calcd for $C_{22}H_{26}N_5O_5S$ $[M + H]^+$ 472.1649, found 472.1650.

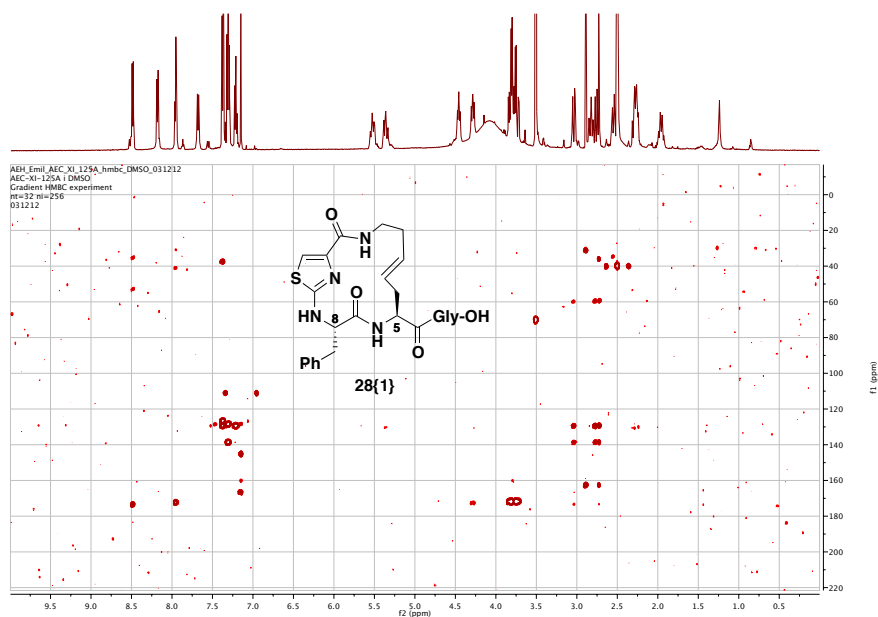
¹H NMR (500 MHz, DMSO-*d*₆)



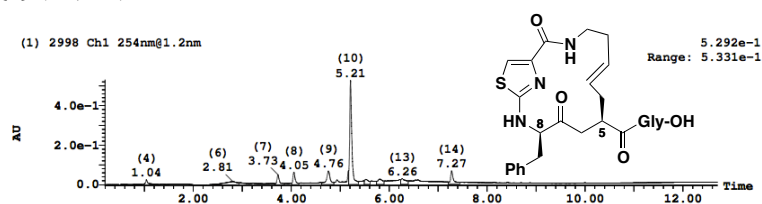
2D HSQC (500 MHz, DMSO-*d*₆)



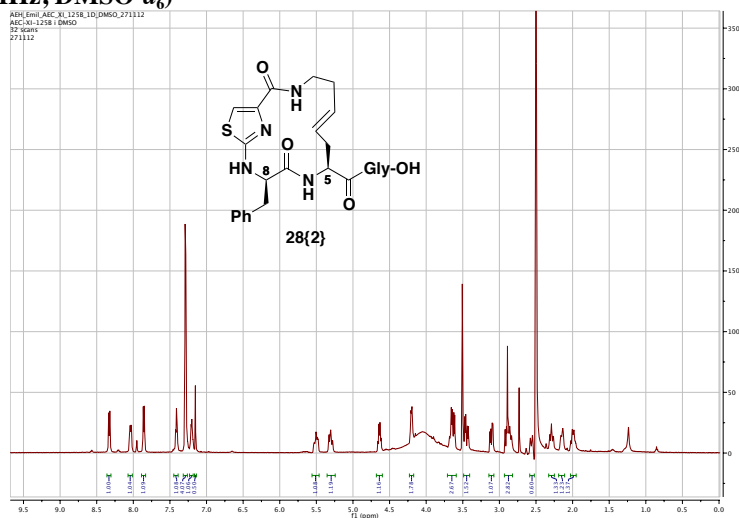
2D HMBC (500 MHz, DMSO-*d*₆)



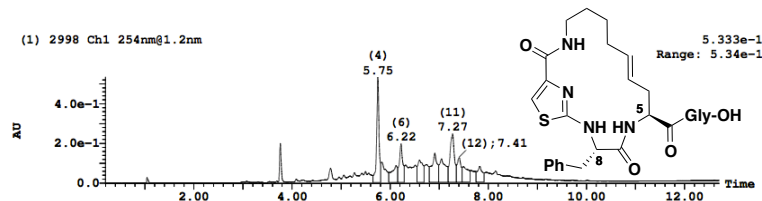
Macrocycle 28{2} (5*S*, 8*R*)



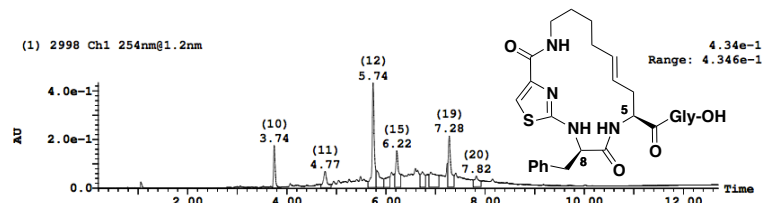
Purity: 74%, $R_f = 5.21$ min. ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δ 8.33 (1H, d, $J = 7.6$ Hz), 8.04 (1H, dd, $J = 8.7, 3.7$ Hz), 7.86 (1H, d, $J = 6.9$ Hz), 7.41 (1H, t, $J = 5.8$ Hz), 7.29 (4H, d, $J = 4.2$ Hz), 7.24 – 7.18 (1H, m), 7.15 (1H, s), 5.51 (1H, ddd, $J = 15.0, 10.0, 3.6$ Hz), 5.30 (1H, ddd, $J = 15.0, 9.9, 3.0$ Hz), 4.46 (1H, dd, $J = 14.6, 7.6$ Hz), 4.25 – 4.13 (1H, m), 3.72 – 3.60 (2H, m), 3.45 (1H, dd, $J = 17.5, 5.8$ Hz), 3.11 (1H, dd, $J = 14.0, 6.2$ Hz), 2.99 – 2.81 (2H, m), 2.56 (1H, d, $J = 14.1$ Hz), 2.28 (1H, dd, $J = 17.4, 7.1$ Hz), 2.15 (1H, d, $J = 12.0$ Hz), 2.04 – 1.94 (1H, m). UPLC-MS (ESI) calcd for $\text{C}_{22}\text{H}_{24}\text{N}_5\text{O}_5\text{S}$ [$\text{M} + \text{H}$] $^+$ 471.2, found 471.3.

¹H NMR (500 MHz, DMSO-*d*₆)

Macrocycle 34{1} (5S, 8S)

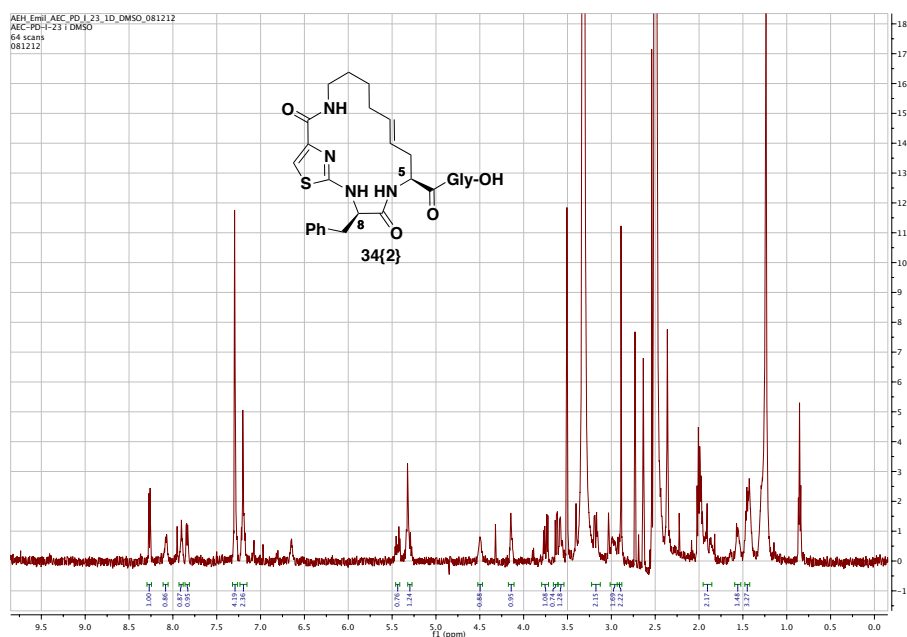
Purity: 51%, R_f = 5.75 min. HRMS (ESI) calcd for $C_{24}H_{30}N_5O_5S$ $[M - H]^-$ 500.1962, found 500.1965.

Macrocycle 34{2} (5*S*, 8*R*)

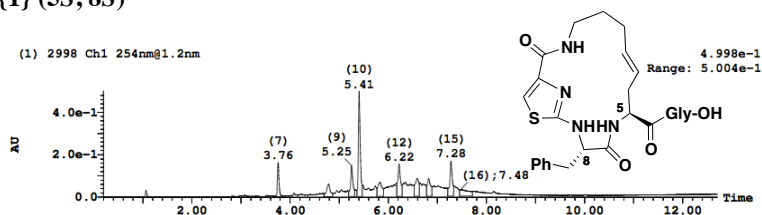


Purity: 65%, $R_f = 5.74$ min. ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δ 8.27 (1H, d, $J = 7.1$ Hz), 8.10 – 8.06 (1H, m), 7.92 – 7.89 (1H, m), 7.86 – 7.82 (1H, m), 7.33 – 7.25 (4H, m), 7.22 – 7.17 (2H, m), 5.47 – 5.41 (1H, m), 5.34 – 5.27 (1H, m), 4.53 – 4.47 (1H, m), 4.17 – 4.12 (1H, m), 3.75 (1H, dd, $J = 17.5$, 6.0 Hz), 3.60 (1H, dd, $J = 17.5$, 5.5 Hz), 3.60 – 3.53 (1H, m), 3.18 (1H, dd, $J = 14.1$, 5.1 Hz), 3.02 – 2.93 (1H, m), 2.93 – 2.86 (1H, m), 1.96 – 1.81 (2H, m), 1.59 – 1.52 (1H, m), 1.48 – 1.38 (3H, m). UPLC-MS (ESI) calcd for $\text{C}_{24}\text{H}_{28}\text{N}_5\text{O}_5\text{S} [\text{M} - \text{H}]^-$ 498.2, found 498.3

^1H NMR (500 MHz, DMSO- d_6)

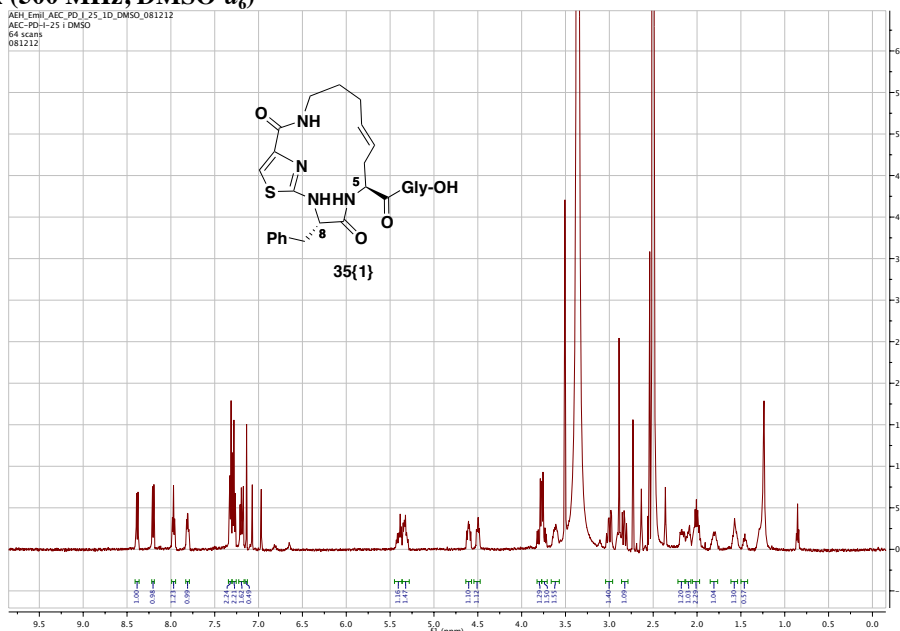


Macrocycle 35{1} (5*S*, 8*S*)

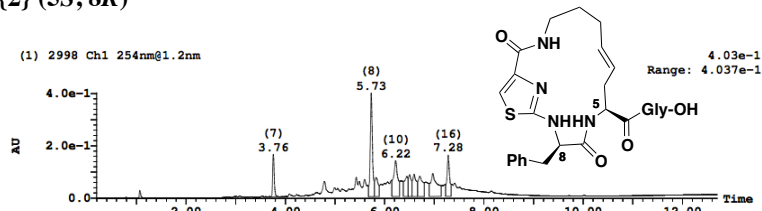


Purity: 69%, $R_f = 5.41$ min. ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δ 8.38 (1H, d, $J = 8.5$ Hz), 8.20 (1H, d, $J = 8.0$ Hz), 7.97 (1H, t, $J = 5.9$ Hz), 7.81 (1H, dd, $J = 7.3, 4.8$ Hz), 7.34 – 7.25 (4H, m), 7.23 – 7.16 (1H, m), 7.14 (1H, s), 5.44 (1H, dt, $J = 15.4, 5.8$ Hz), 5.31 (1H, dt, $J = 15.4, 6.3$ Hz), 4.60 (1H, ddd, $J = 10.0, 8.0, 4.8$ Hz), 4.54 – 4.46 (1H, m), 3.80 (1H, dd, $J = 17.5, 5.9$ Hz), 3.74 (1H, dd, $J = 17.5, 5.9$ Hz), 3.66 – 3.58 (2H, m), 2.99 (1H, dd, $J = 13.9, 4.5$ Hz), 2.83 (1H, dd, $J = 14.0, 10.1$ Hz), 2.22 – 2.13 (1H, m), 2.13 – 2.06 (1H, m), 2.03 – 1.95 (2H, m), 1.85 – 1.74 (1H, m), 1.61 – 1.52 (1H, m), 1.52 – 1.42 (1H, m). UPLC-MS (ESI) calcd for $\text{C}_{23}\text{H}_{26}\text{N}_5\text{O}_5\text{S}$ $[\text{M} - \text{H}]^-$ 484.2, found 484.3.

^1H NMR (500 MHz, $\text{DMSO}-d_6$)

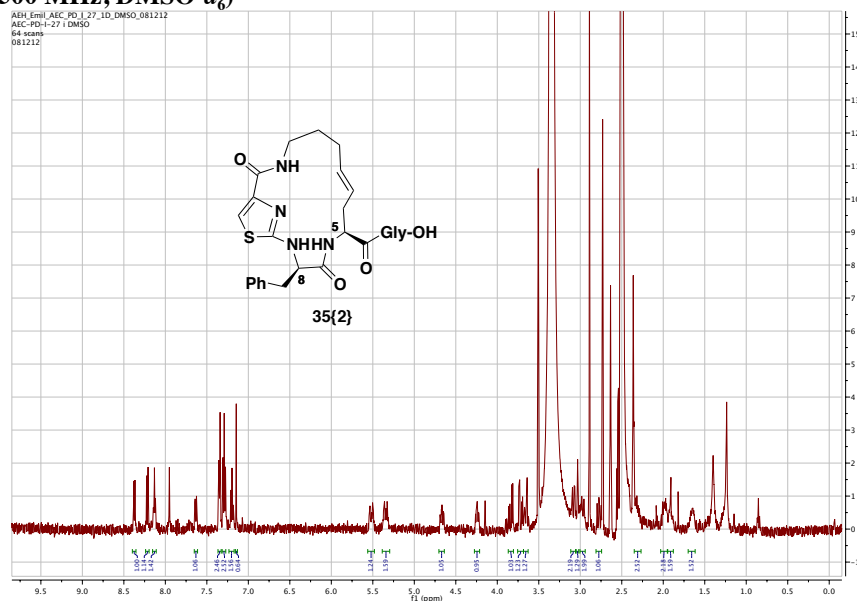


Macrocycle 35{2} (5*S*, 8*R*)

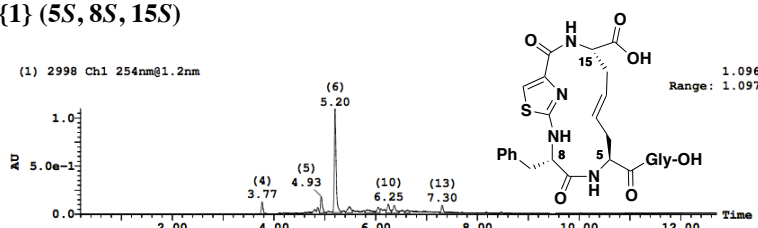


Purity: 71%, $R_f = 5.73$ min. ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δ 8.37 (1H, d, $J = 7.2$ Hz), 8.22 (1H, d, $J = 8.6$ Hz), 8.13 (1H, t, $J = 5.8$ Hz), 7.63 (1H, d, $J = 7.8$ Hz), 7.35 (2H, d, $J = 7.4$ Hz), 7.29 (2H, t, $J = 7.6$ Hz), 7.20 (1H, t, $J = 7.3$ Hz), 7.15 (1H, s), 5.52 (1H, dt, $J = 15.4, 5.5$ Hz), 5.34 (1H, dt, $J = 15.3, 6.5$ Hz), 4.71 – 4.63 (1H, m), 4.27 – 4.21 (1H, m), 3.84 (1H, dd, $J = 17.5, 6.1$ Hz), 3.72 (1H, dd, $J = 17.5, 5.5$ Hz), 3.69 – 3.63 (1H, m), 3.08 (1H, dd, $J = 14.0, 4.0$ Hz), 3.03 (1H, t, $J = 3.2$ Hz), 2.99 – 2.95 (1H, m), 2.77 (1H, dd, $J = 13.9, 10.9$ Hz), 2.37 – 2.27 (2H, m), 2.03 – 1.87 (2H, m), 1.70 – 1.60 (1H, m), 1.44 – 1.37 (1H, m). UPLC-MS (ESI) calcd for $\text{C}_{23}\text{H}_{26}\text{N}_5\text{O}_5\text{S}$ $[\text{M} - \text{H}]^-$ 484.2, found 484.3.

¹H NMR (500 MHz, DMSO-*d*₆)

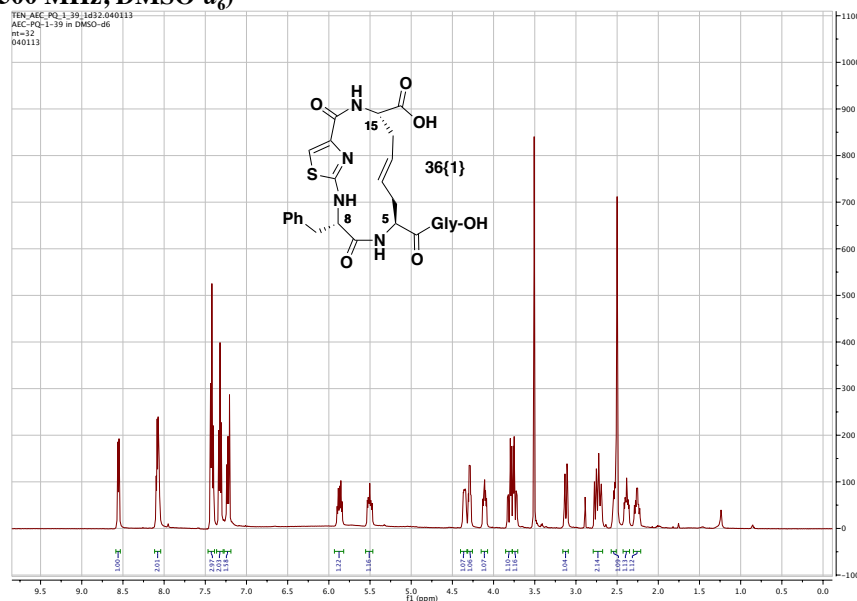


Macrocycle 36{1} (5*S*, 8*S*, 15*S*)

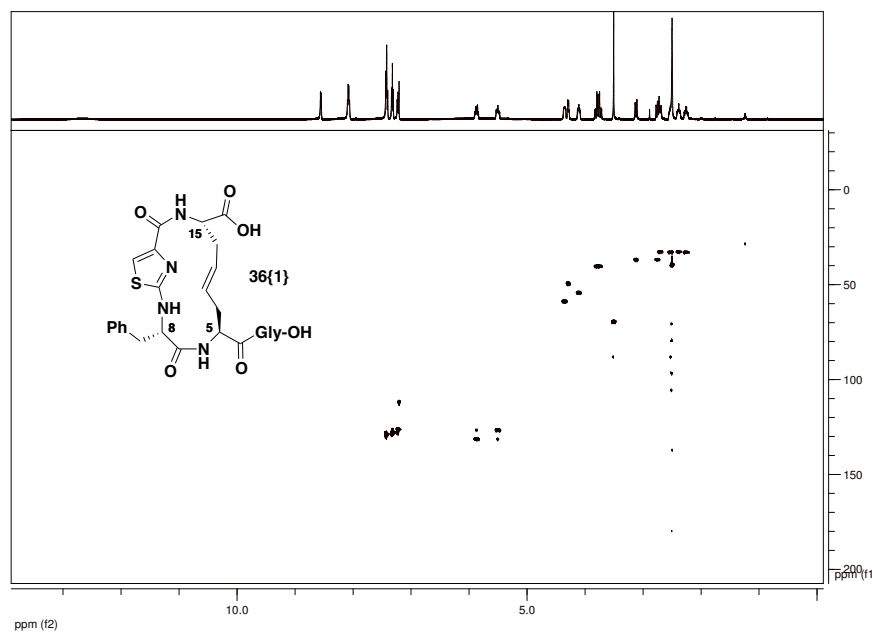


Purity: 76%, R_f = 5.20 min. ¹H NMR (500 MHz, DMSO-*d*₆): δ 8.58 (1H, d, J = 6.9 Hz), 8.12 (1H, t, J = 5.6 Hz), 8.08 (1H, d, J = 6.9 Hz), 7.42 (3H, t, J = 7.1 Hz), 7.32 (2H, t, J = 7.5 Hz), 7.23 (1H, d, J = 7.4 Hz), 7.21 (1H, s), 5.84 (1H, dt, J = 15.4, 7.6 Hz), 5.47 (1H, ddd, J = 15.5, 9.6, 4.8 Hz), 4.42 – 4.25 (2H, m), 4.10 (1H, ddd, J = 11.3, 6.6, 4.3 Hz), 3.81 (1H, dd, J = 17.8, 5.3 Hz), 3.73 (1H, dd, J = 17.7, 5.8 Hz), 3.12 (1H, d, J = 12.6 Hz), 2.83 – 2.64 (2H, m), 2.61 – 2.51 (1H, m), 2.43 – 2.32 (1H, m), 2.32 – 2.16 (1H, m). ¹³C NMR (125 MHz, DMSO-*d*₆): δ 172.1, 171.8, 171.5, 171.0, 166.2, 160.0, 144.1, 137.7, 131.2, 128.9 (2C), 127.8 (2C), 126.5, 126.1, 111.7, 58.8, 54.2, 49.4, 40.3, 36.8, 32.8, 32.7. UPLC-MS (ESI) calcd for C₂₃H₂₄N₅O₇S [M - H]⁻ 514.1, found 514.2.

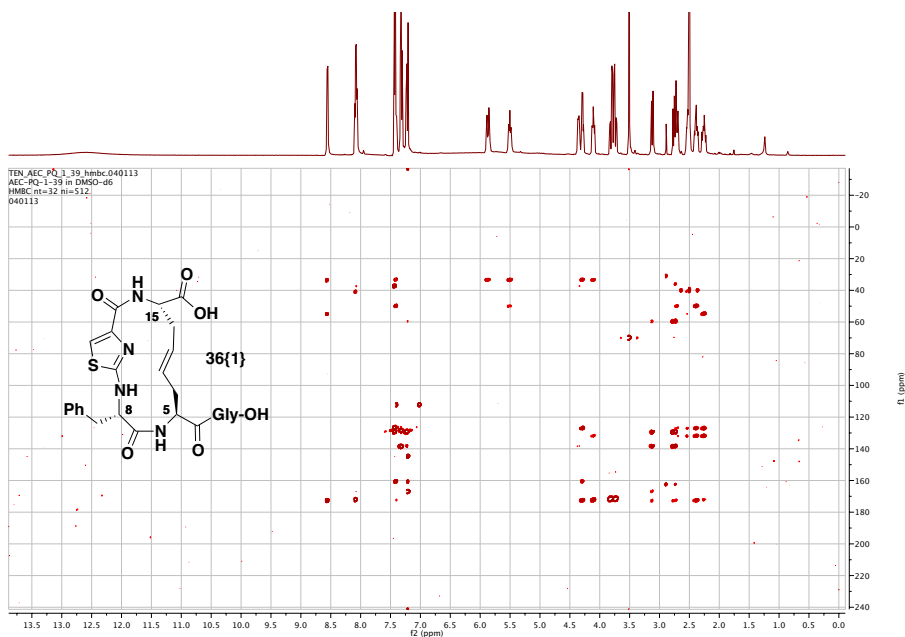
¹H NMR (500 MHz, DMSO-*d*₆)



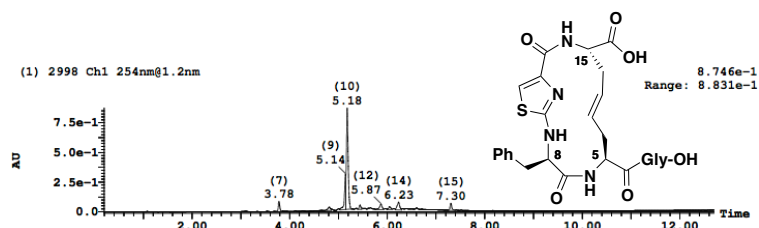
2D HSQC (500 MHz, DMSO-*d*₆)



2D HMBC (500 MHz, DMSO-*d*₆)

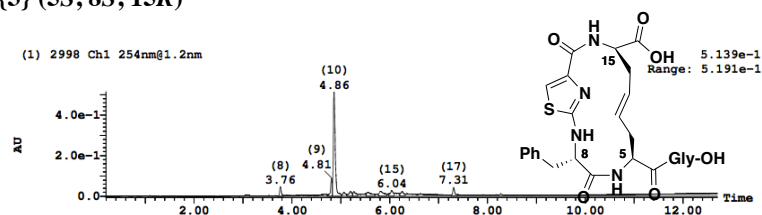


Macrocycle 36{2} (5*S*, 8*R*, 15*S*)

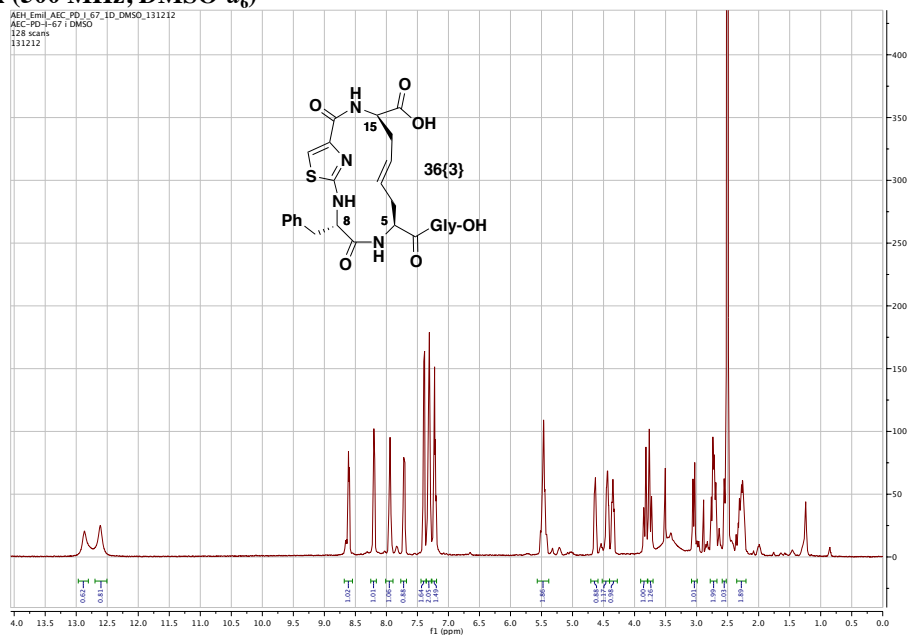


Purity: 86%, R_f = 5.18 min. Olefine isomeric ratio 2/1. UPLC-MS (ESI) calcd for $C_{23}H_{24}N_5O_7S$ [M - H]⁻ 514.1, found 514.2.

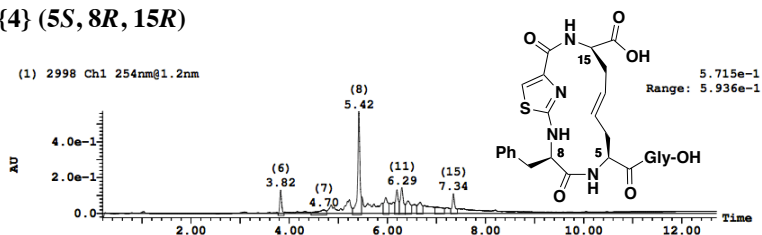
Macrocycle 36{3} (5*S*, 8*S*, 15*R*)



¹H NMR (500 MHz, DMSO-*d*₆)



Macrocycle 36{4} (5*S*, 8*R*, 15*R*)



Purity: 77%, *R*_f = 5.42 min. ¹H NMR (300 MHz, DMSO-*d*₆): δ 8.26 (1H, d, *J* = 6.8 Hz), 8.01 (1H, d, *J* = 7.4 Hz), 7.95 (1H, d, *J* = 7.6 Hz), 7.75 – 7.67 (1H, m), 7.33 – 7.16 (6H, m), 5.58 – 5.47 (1H, m), 5.47 – 5.33 (1H, m), 4.49 – 4.39 (1H, m), 4.24 (1H, dd, *J* = 11.8, 5.5 Hz), 4.16 – 4.06 (1H, m), 3.72 (1H, dd, *J* = 17.7, 6.0 Hz), 3.63 (1H, dd, *J* = 17.7, 5.4 Hz), 3.23 – 3.01 (2H, m), 2.56 – 2.45 (2H, m), 2.46 – 2.32 (2H, m). UPLC-MS (ESI) calcd for $C_{23}H_{24}N_5O_7S$ [M - H]⁻ 514.1, found 514.2.

¹H NMR (500 MHz, DMSO-*d*₆)

