Supplementary Information: Mimicking Red Blood Cell Lipid Membrane to Enhance the Hemocompatibility of Large-Pore Mesoporous Silica

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The following SEM figures represent a quantification of the interactions between large pore mesoporous silica nanoparticles coated with either dipalmitoylphosphatidyl choline (DPPC-LP-MSNs, Fig. 1) or a mixture containing dipalmitoylphosphatidylcholine, dipalmitoylphosphatidylethanolamine and cholesterol in a weight ratio of 4:1:5 (mRBC-LP-MSNs Fig. 2). Due to the restrictions of the SEM images, only the red blood cells (RBCs) that are clearly visible (not obscured by other cells or particles) and have an obvious association with LP-MSNs were counted. Stomatocytic (cup-shaped) may be under-represented in our count do to the lack of 3-dimensional imaging that is available to other studies which utilize confocal microscopy to make morphological determinations. The "health factor" (HF) calculated for these studies is determined using the following formula where discocytic (D) morphologies are marked with green circles and stomatocytic and echinocytic (SE) morphologies are marked with red circles. The morphologies are identified using the work of Khairy, Foo, and Howard as guide.¹

$$\frac{D-ES}{Total \ Count} = HF$$
 eq. 1

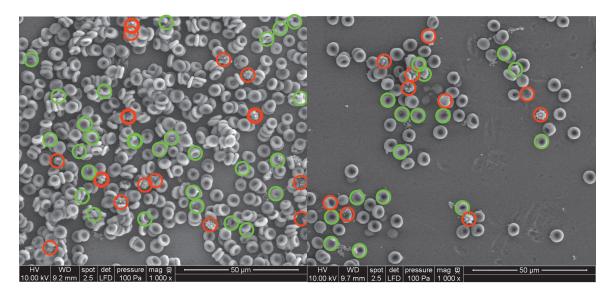


Figure 1. SEM images of RBCs exposed to DPPC-LP-MSNs at 50 μ g mL⁻¹ (left panel) and 100 μ g mL⁻¹ (right panel). The HF for the RBCs in these images is calculated to be 0.089 indicating that the counted interactions between DPPC-LP-MSNs and RBCs are divided equally between healthy cells (green circles) and cells with abnormal morphology (red circles).

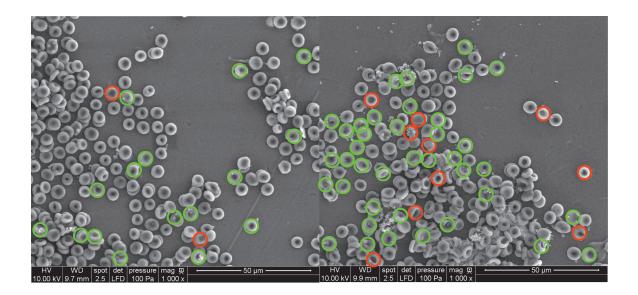


Figure 2. SEM images of RBCs exposed to mRBC-LP-MSN at 50 μ g mL⁻¹ (left panel) and 100 μ g mL⁻¹ (right panel). The HF for the RBCs in these images is calculated to be 0.64 indicating that the counted interactions between mRBC-LP-MSNs result predominately in cells with normal morphology (green circles) as opposed to abnormal morphologies (red circles).

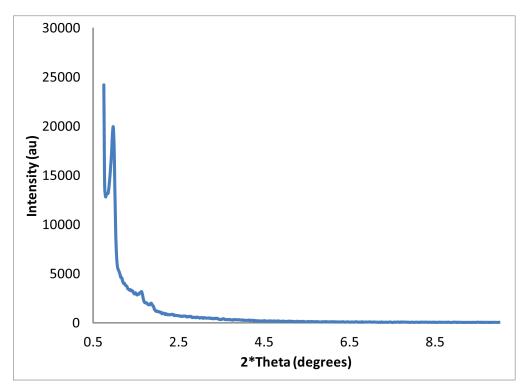


Figure 3. Powder X-ray diffraction pattern of *l*-MSN

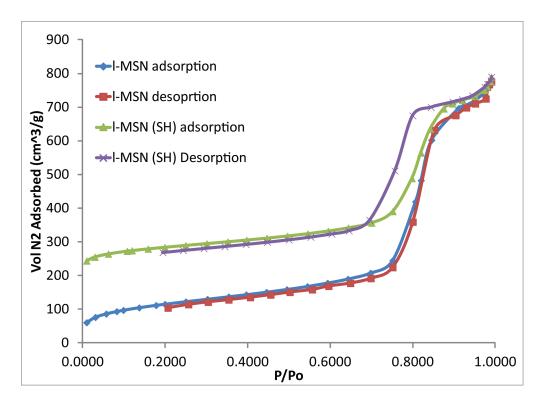
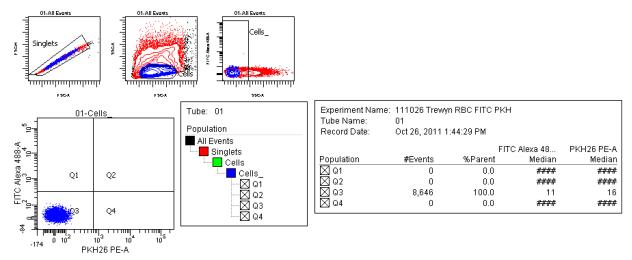
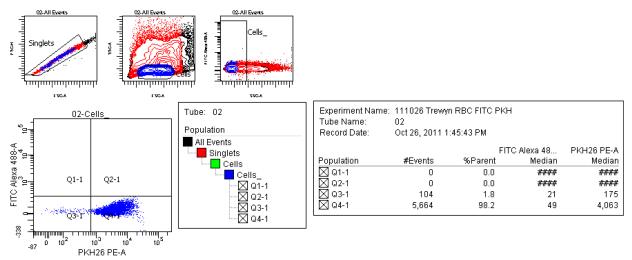


Figure 4. Nitrogen sorption isotherms of *l*-MSN (surfactant removed) and *l*-MSN that was post-synthetically grafted with 3-mercaptopropyltriethoxy silane (*l*-MSN (SH)).

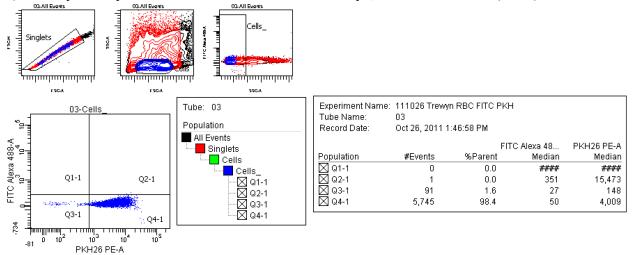


110614 Trewyn RBC FITC-1-Lipid Bilayer

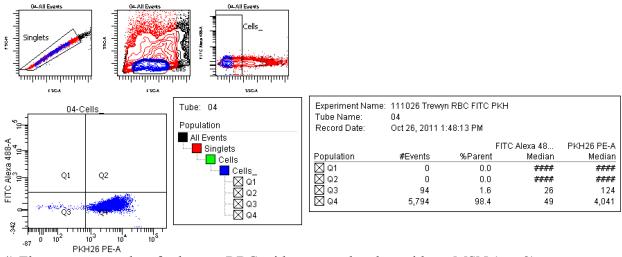
1) Flow cytometry data for human RBC without PKH26 dye or MSN.



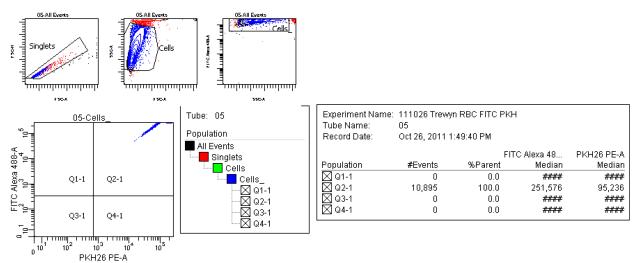
2) Flow cytometry data for human RBC with PKH26 dye, but without MSN (run 1).



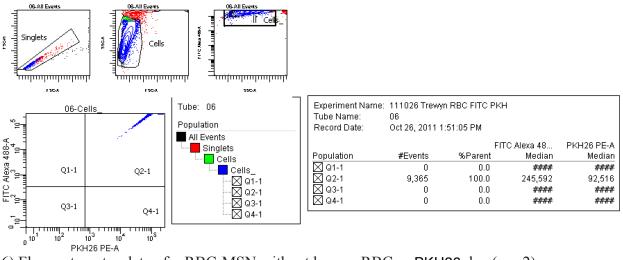
3) Flow cytometry data for human RBC with PKH26 dye, but without MSN (run 2).



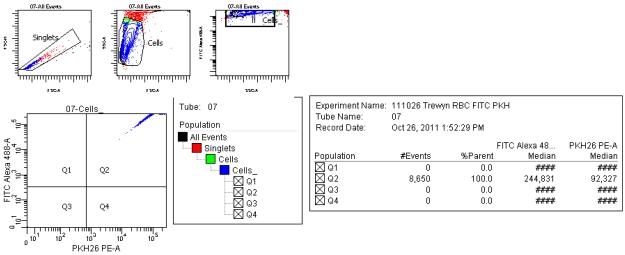
4) Flow cytometry data for human RBC with PKH26 dye, but without MSN (run 3).



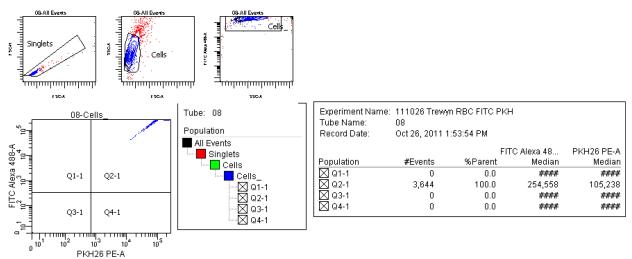
5) Flow cytometry data of mRBC-MSN without human RBC or PKH26 dye (run 1).



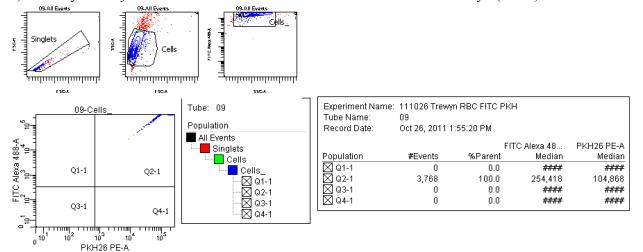
6) Flow cytometry data of mRBC-MSN without human RBC or PKH26 dye (run 2).



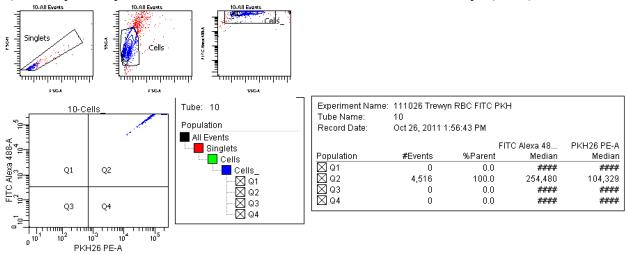
7) Flow cytometry data of mRBC-MSN without human RBC or PKH26 dye (run 3).



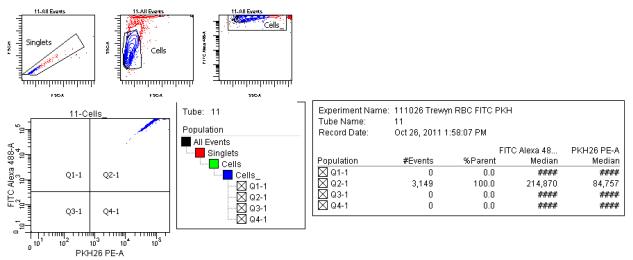
8) Flow cytometry data of DPPC-MSN without human RBC or PKH26 dye (run 1).



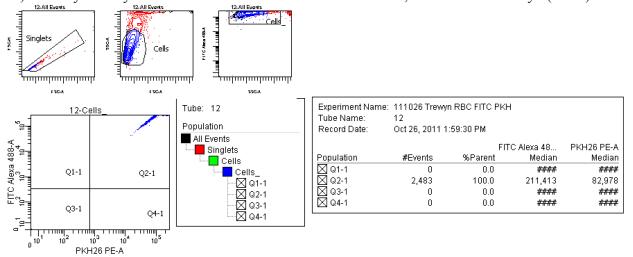
9) Flow cytometry data of DPPC-MSN without human RBC or PKH26 dye (run 2).



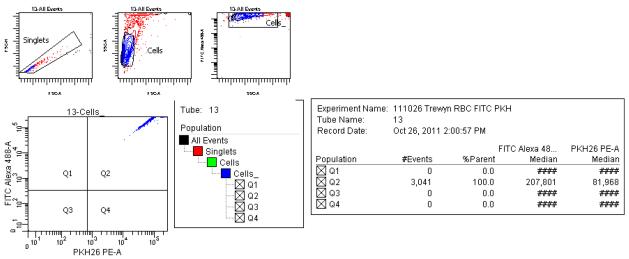
10) Flow cytometry data of DPPC-MSN without human RBC or PKH26 dye (run 3).



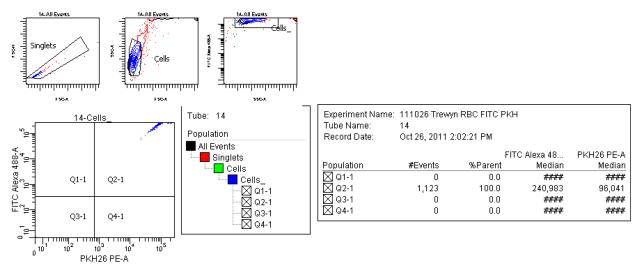
11) Flow cytometry data of mRBC-MSN without human RBC, but with PKH26 dye (run 1).



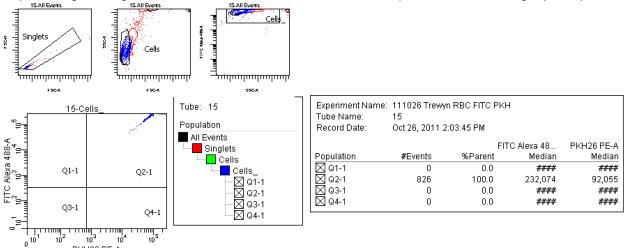
12) Flow cytometry data of mRBC-MSN without human RBC, but with PKH26 dye (run 2).



13) Flow cytometry data of mRBC-MSN without human RBC, but with PKH26 dye (run 3).

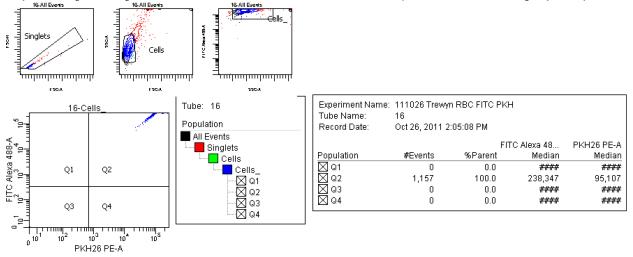


14) Flow cytometry data of DPPC-MSN without human RBC, but with PKH26 dye (run 1).

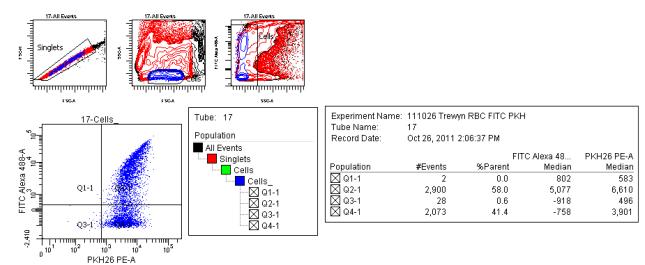


15) Flow cytometry data of DPPC-MSN without human RBC, but with PKH26 dye (run 2).

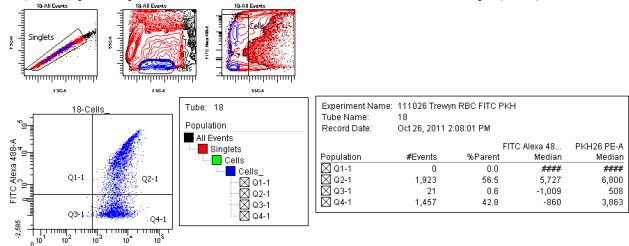
PKH26 PE-A



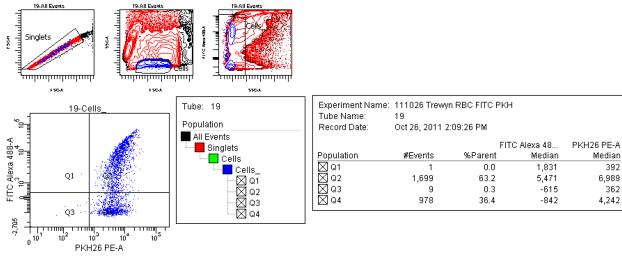
16) Flow cytometry data of DPPC-MSN without human RBC, but with PKH26 dye (run 3).



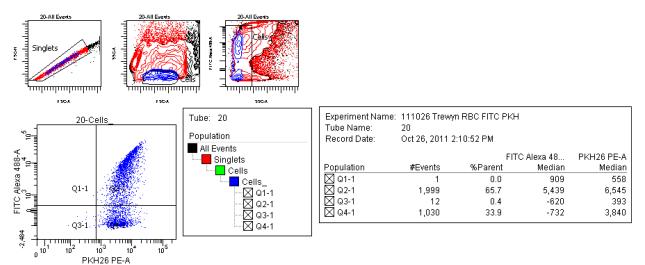
17) Flow cytometry data of mRBC-MSN with human RBC and PKH26 dye (run 1).



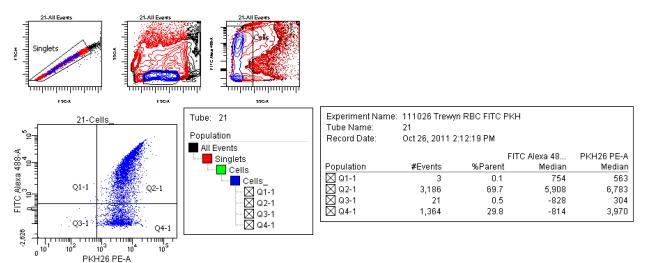
^{PKH26 PE-A} 18) Flow cytometry data of mRBC-MSN with human RBC and PKH26 dye (run 2).



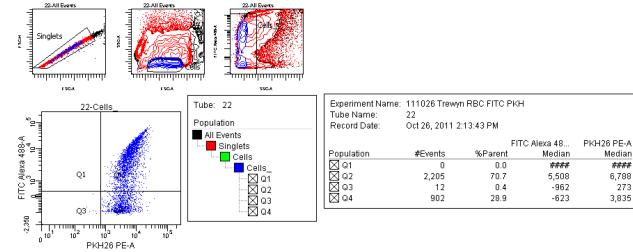
19) Flow cytometry data of mRBC-MSN with human RBC and PKH26 dye (run 3).



20) Flow cytometry data of DPPC-MSN with human RBC and PKH26 dye (run 1).

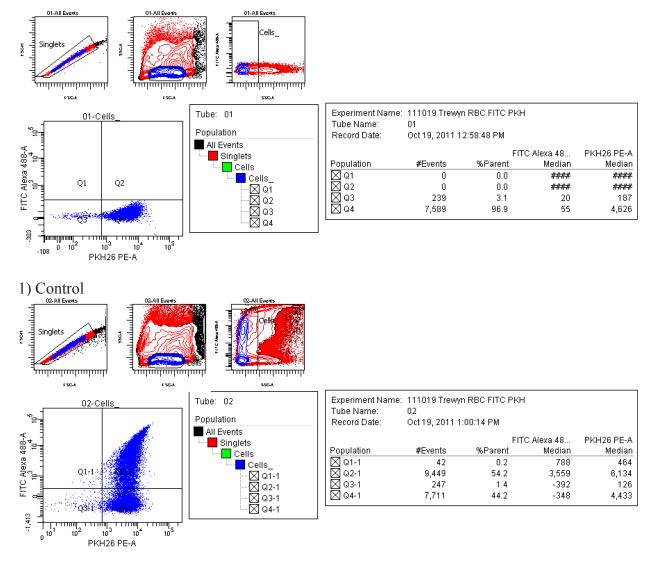


21) Flow cytometry data of DPPC-MSN with human RBC and PKH26 dye (run 2).

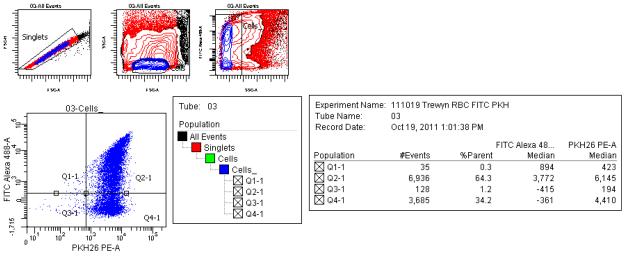


22) Flow cytometry data of DPPC-MSN with human RBC and PKH26 dye (run 3).

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2) Flow cytometry data of molecular DPPC with human RBC and PKH26 dye.



3) Flow cytometry data of molecular mRBC with human RBC and PKH26 dye.

(1) Khairy, K.; Foo, J.; Howard, J. *Cel. Mol. Bioeng.* **2008**, *1*, 173.