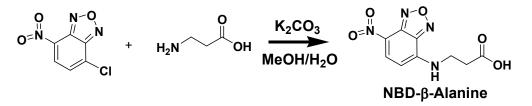
Environment-sensitive fluorescent supramolecular nanofibers for imaging applications

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Experimental supporting information

Preparation of NBD-β-Alanine:

The preparation of NBD- β -Alanine was according to Scheme S-1. To a 15 mL water solution of 980 mg of β -Alanine (1.1 equiv.) and 4.14 g of K₂CO₃ (3 equiv.), 2 g of NBD-Cl (1 equiv., 10 mmol) in 20 mL of MeOH was added dropwisely in the N₂ atmosphere (low yields if without nitrogen protection). The reaction mixture was stirred at room temperature for about 3h (LC-MS detection). After the 3h reaction, MeOH was removed by a rotary evaporator. The obtained aqueous solution was acidified to around pH 3 by conc. HCl. The acidic aqueous solution was then extracted by ether for 3 times. The combined organic solution was dried over MgSO₄ and then concentrated by a rotary evaporator. The resulting yellowish solid (NBD- β -Alanine) was directly used for solid phase peptide synthesis.

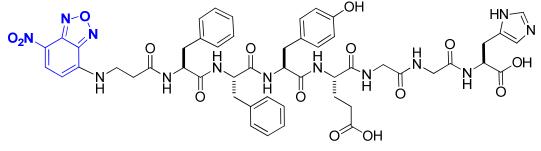


Scheme S-1. The synthetic route for NBD-β-Alanine

Peptide synthesis: The peptide derivative was prepared by solid phase peptide synthesis (SPPS) using 2-chlorotrityl chloride resin and the corresponding N-Fmoc protected amino acids with side chains properly protected by a tert-butyl group or Pbf group. 20% piperidine in anhydrous N,N'-dimethylformamide (DMF) was used during deprotection of Fmoc group. Then the next Fmoc-protected amino acid was coupled to the free amino group using O-(Benzotriazol-1-yl)-N,N,N',N'-tetramethyluroniumhexafluorophosphate (HBTU) as the coupling reagent. The growth of the peptide chain was according to the established Fmoc SPPS protocol. In the last coupling step, NBD-β-Alanine was used to produce NBD-peptides. After the last coupling step, excessive reagents were removed by a single DMF wash for 5 minutes (5 mL per gram of resin), followed by five steps of washing using DCM for 2 min (5 mL per gram of resin). The peptide derivative was cleaved using 95% of trifluoroacetic acid with 2.5% of trimethylsilane

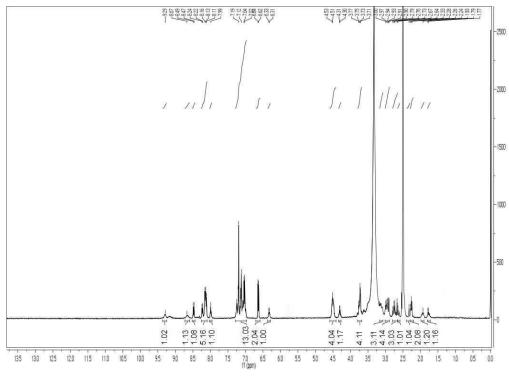
(TMS) and 2.5% of H_2O for 30 minutes. 20 mL per gram of resin of ice-cold diethylether was then added to cleavage reagent. The resulting precipitate was centrifuged for 10 min at 4 ^{0}C at 10,000 rpm. Afterward the supernatant was decanted and the resulting solid was dissolved in DMSO for HPLC separation using MeOH and H_2O containing 0.05% of TFA as eluents.

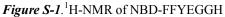
Characterization of the peptides



Compound1:NBD-FFYEGGH

Scheme S-2. Chemical structure of NBD-FFYEGGH





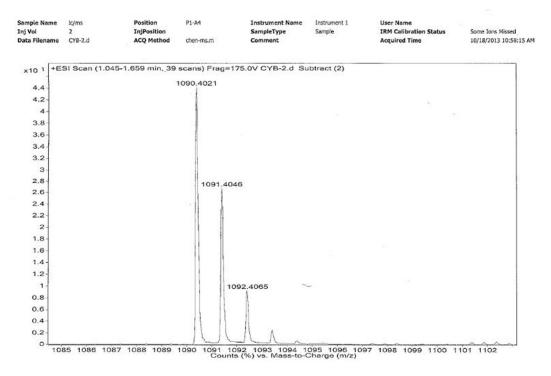
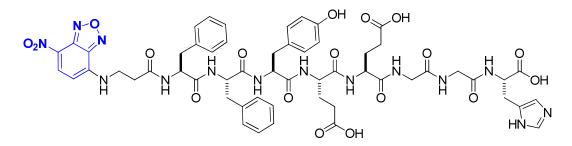


Figure S-2.HR-MS of NBD-FFYEGGH



Compound 2:NBD-FFYEEGGH

Scheme S-3. Chemical structure of NBD-FFYEEGGH

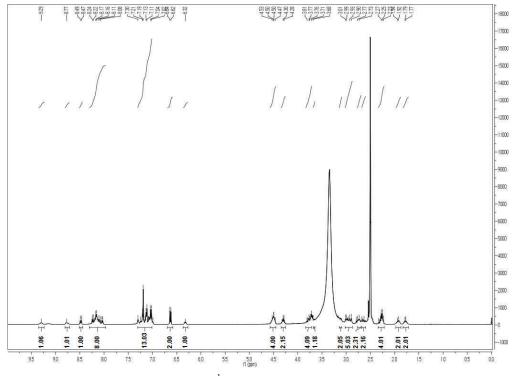
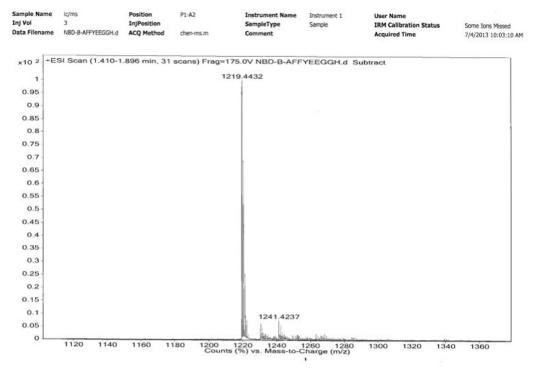
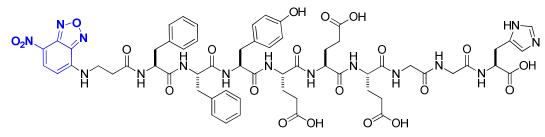


Figure S-3. ¹H-NMR of NBD-FFYEEGGH

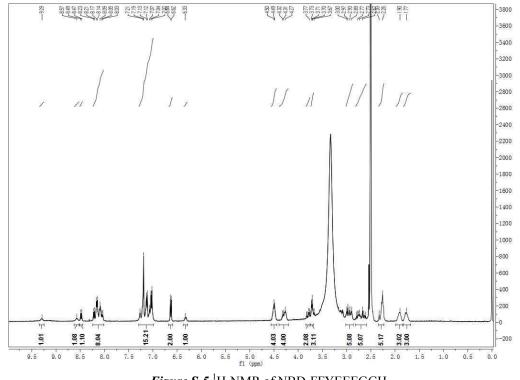


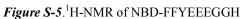




Compound3:NBD-FFYEEEGGH

Scheme S-4. Chemical structure of NBD-FFYEEEGGH





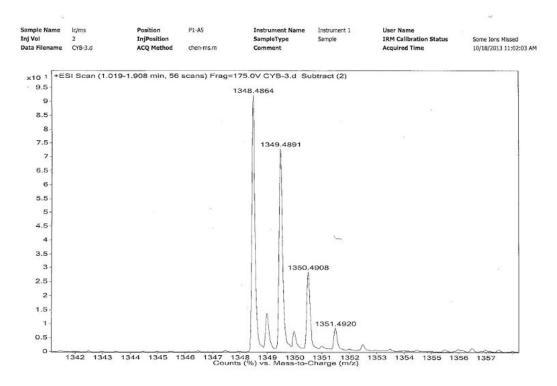
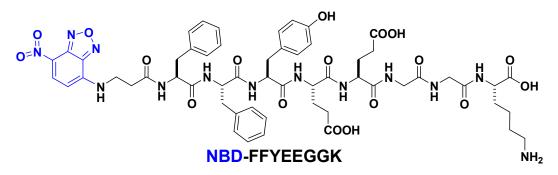
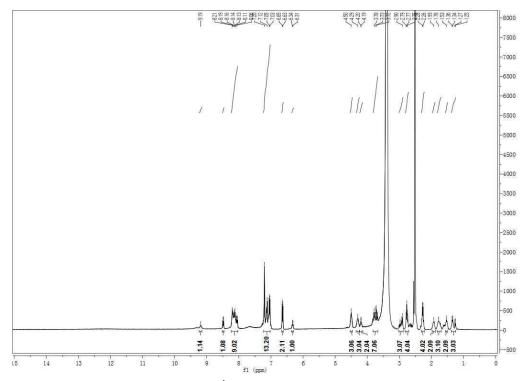
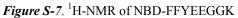


Figure S-6.HR-MS of NBD-FFYEEEGGH



Scheme S-5. Chemical structure of NBD-FFYEEGGK





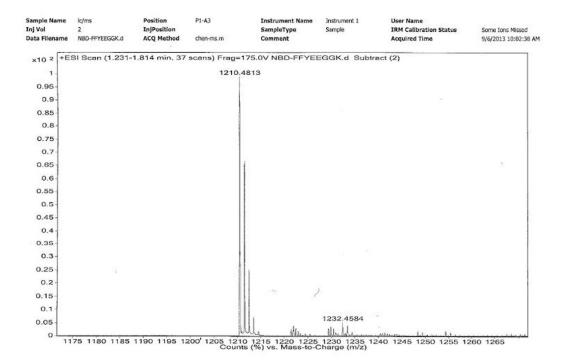
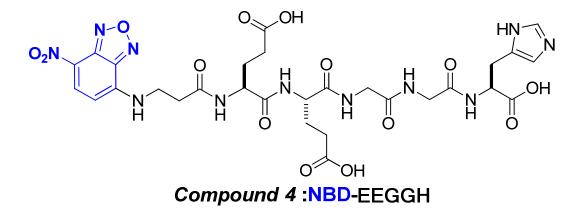


Figure S-8. HR-MS of NBD-FFYEEGGK



Scheme S-6. Chemical structure of NBD-EEGGH

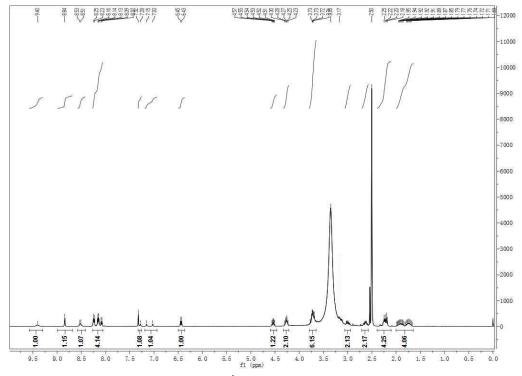
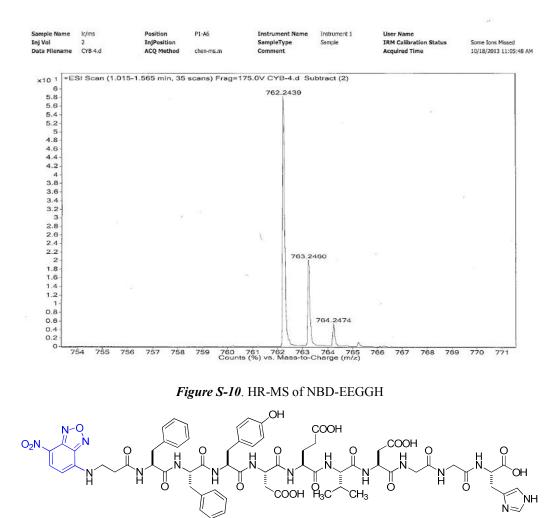
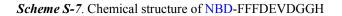
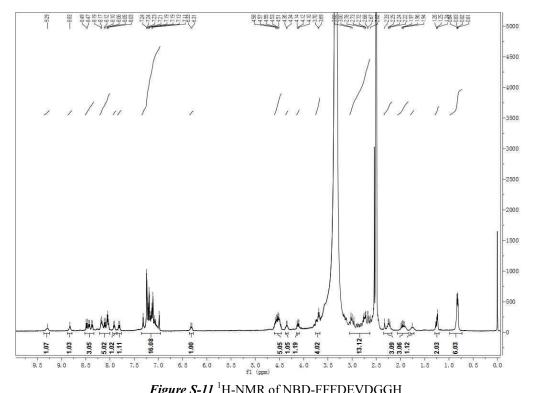


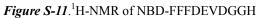
Figure S-9. ¹H-NMR of NBD-EEGGH

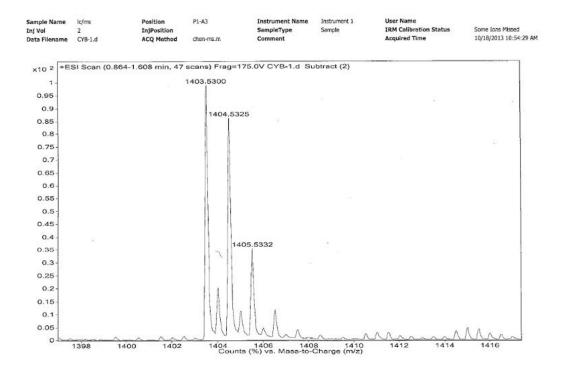


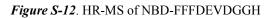


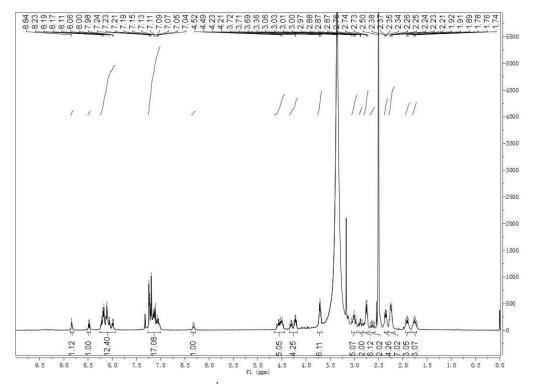


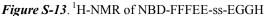


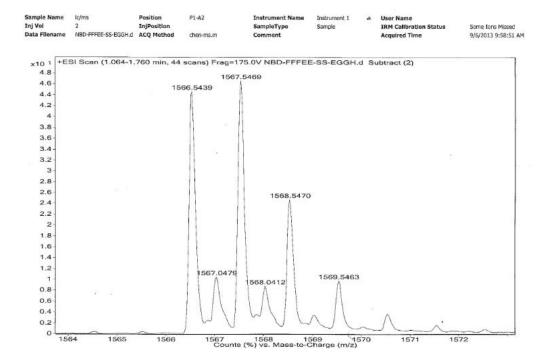












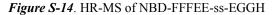




Figure S-15. An optical image of water solution of NBD-FFYEEGGK with 1 equiv. of Cu²⁺

Hydrogelation test of 2 by adding different metal ions: To a water solution of **2** (1.0 wt%, pH = 7.4, adjusted by Na₂CO₃), an equal volume of water solution containing 1 equiv. of different kinds of metal ions (Cu²⁺, Mn²⁺, Ni²⁺, Co²⁺, Fe²⁺, Zn²⁺, Ca²⁺, Sr²⁺, Ba²⁺, Na⁺, and K⁺) was added (pH = 7.4). The hydrogelation could only be observed for Cu²⁺.

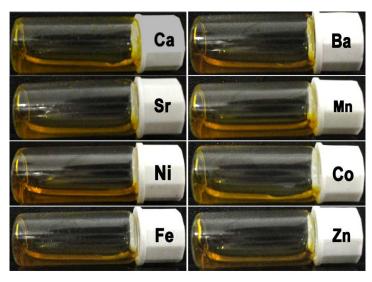


Figure S-16. Optical images of water solution of 2 with 1 equiv. of different metal ions (pH = 7.4)

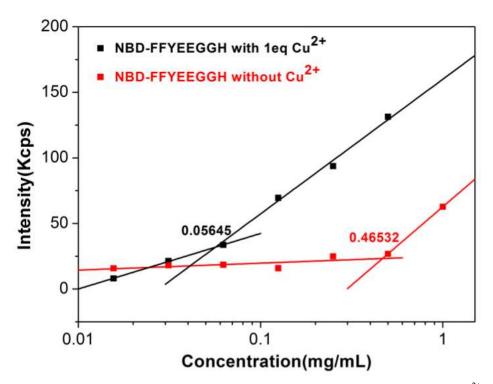


Figure S-17. CMC of compound 2 in the absence and presence of 1 equiv. of Cu^{2+}

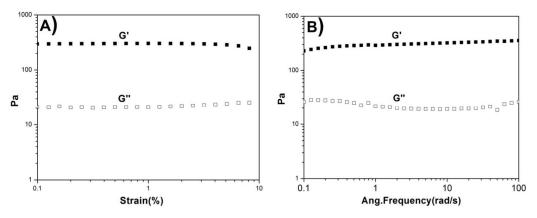


Figure S-18. Rheological measurements A) in dynamic strain sweep mode at the frequency of 1 rad/s and B) in dynamic frequency sweep at the strain of 1% for hydrogel containing 0.5% compound 2 and 1 equiv. of Cu²⁺

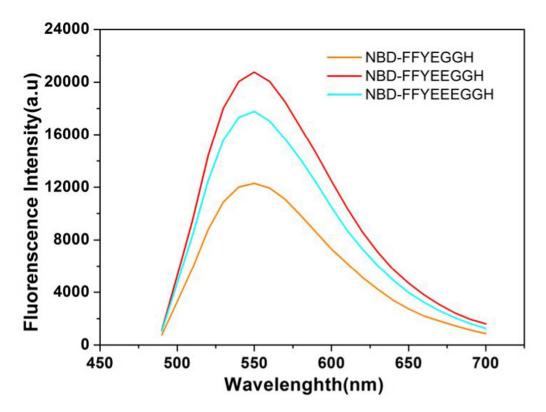


Figure S-19. Fluorescence spectra of aqueous solution containing 0.05 wt% of 1-3.

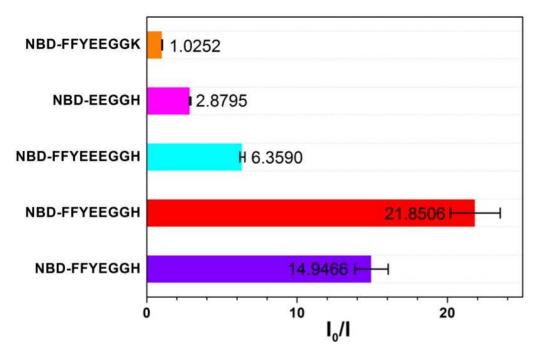


Figure S-20. I_0/I value of different compounds (the data were expressed as the mean \pm standard error of the mean (SEM, N = 3))

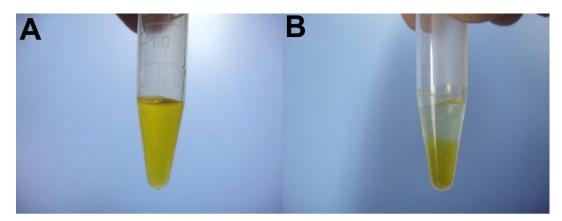


Figure S-21. Optical images of solutions of compound *I* with or without Cu²⁺: A) 0.5 wt% of compound *I* without Cu²⁺; B) 0.5 wt% of compound *I* with one equiv. of Cu²⁺

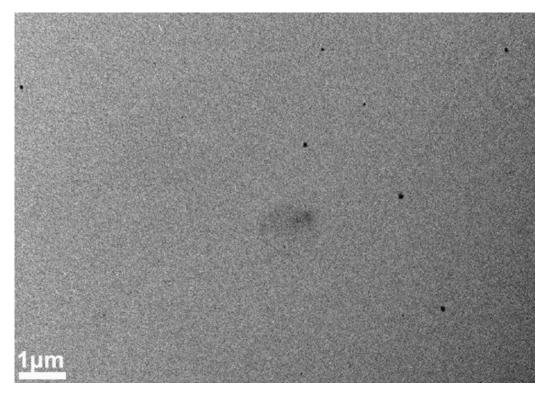


Figure S-22. Transmission electron microscopy (TEM) images of solution of 4 with one equiv. of Cu^{2+}

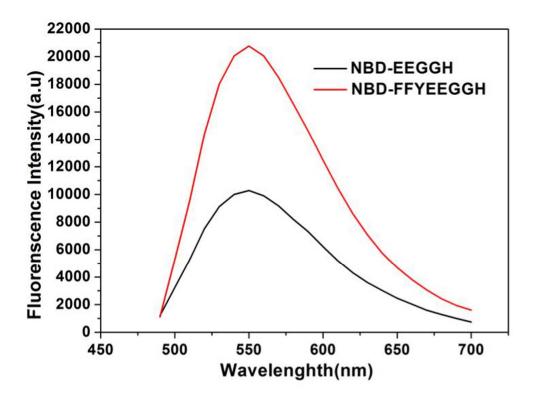


Figure S-23. Fluorescence spectra of aqueous solution containing compound 2 or compound 4 $(410 \ \mu\text{M}, \text{pH} = 7.4)$

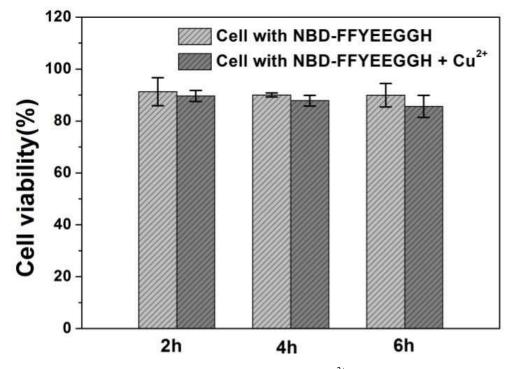


Figure S-24. Cell viability of Compounds 2 with 100 μ M Cu²⁺ by MTT assay for 2 to 6 h (the data were expressed as the mean \pm standard error of the mean (SEM, N = 3))

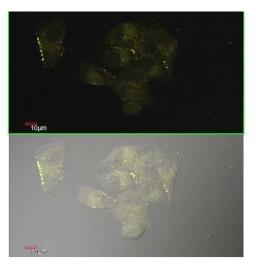




Figure S-25. Confocal images of HeLa cells treated with 2 (0.05 wt%) at 2 h time point (top left: fluorescence image, top right: bright field image, and down: overlay image)

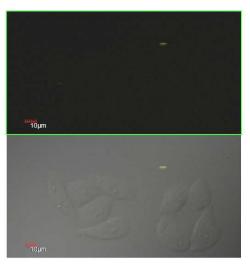




Figure S-26. Confocal images of HeLa cells pre-treated with 100 μM of Cu²⁺ and then treated with 2 (0.05 wt%) at 6 h time point (top left: fluorescence image, top right: bright field image, and down: overlay image)

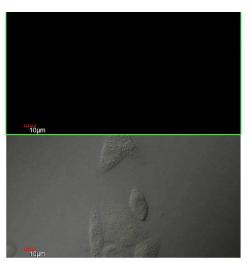




Figure S-27. Confocal images of HeLa cells treated with 2 (0.05 wt%) for 2h and then treated with 100 μ M of Cu²⁺ for another 2h (top left: fluorescence image, top right: bright field image, and down: overlay image)

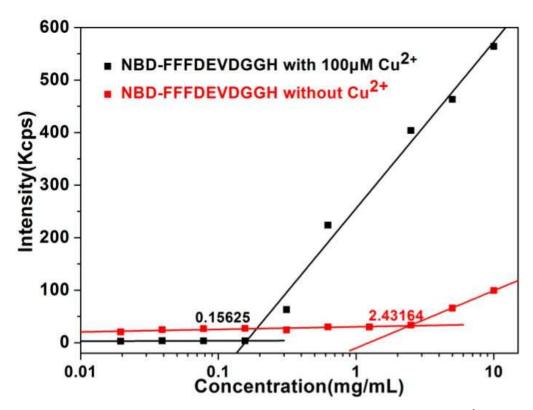


Figure S-28. CMC of compound 4 in the absence and presence of 100 μ M of Cu²⁺

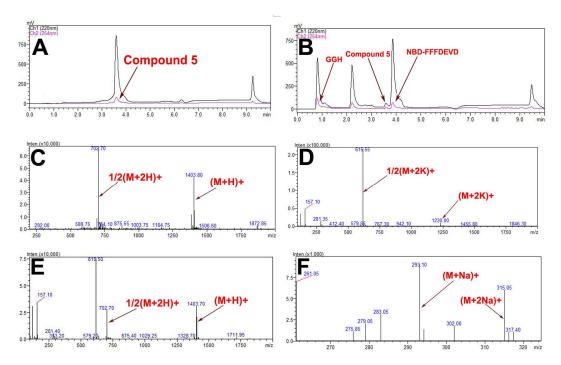


Figure S-29. A) HPLC of compound 5; B) HPLC of compound 5 with 1 U/mL caspase-3 for 1 h;
C) MS of compound 5 in A); D) MS of NBD-FFFDEVD cleaved from compound 5 by caspase-3 in B); E) MS of compound 5 uncleaved by caspase-3 in B); F) MS of GGH cleaved from compound 5 by caspase-3 in B).

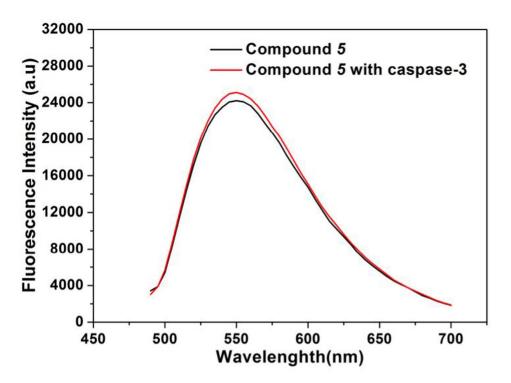


Figure S-30 Fluorescence spectra of aqueous solution containing compound 5 (0.05 wt%) before and after adding Caspase-3 in the absence of Cu^{2+}

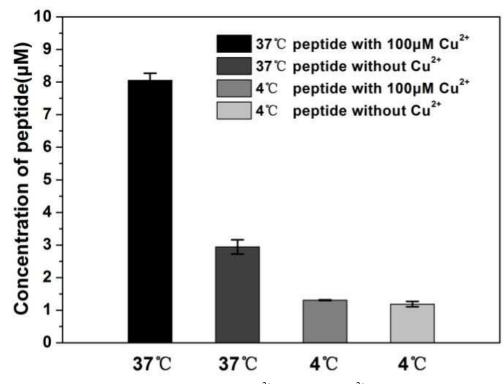


Figure S-31. The amount of compound 4 with Cu^{2+} and without Cu^{2+} within HeLa cells at 4°C and 37 °C determined by LC-MS (the data were expressed as the mean \pm standard error of the mean (SEM, N = 3))

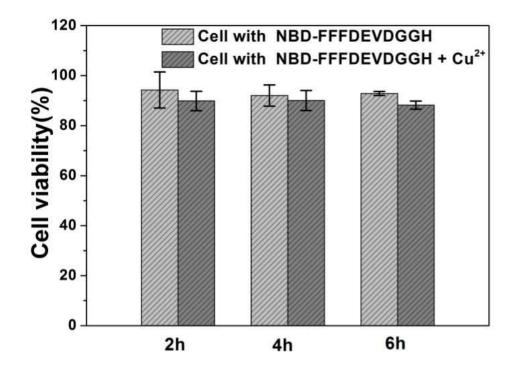


Figure S-32. Cell viability of Compound *4* with 100 μ M Cu²⁺ by MTT assay for 2 to 6h (the data were expressed as the mean \pm standard error of the mean (SEM, N = 3))

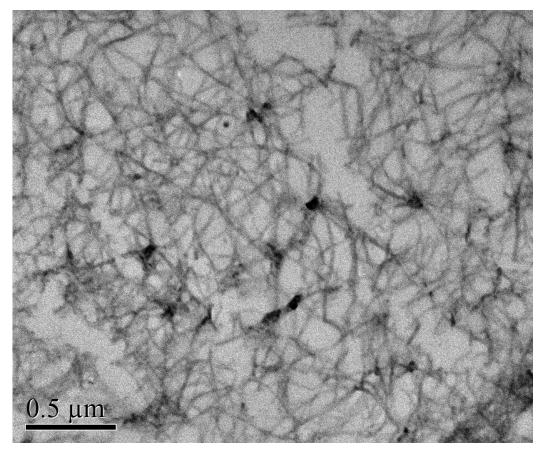


Figure S-33. Transmission electron microscopy (TEM) images of solution of 6 with one equiv. of Cu²⁺ and 4 equiv. of GSH

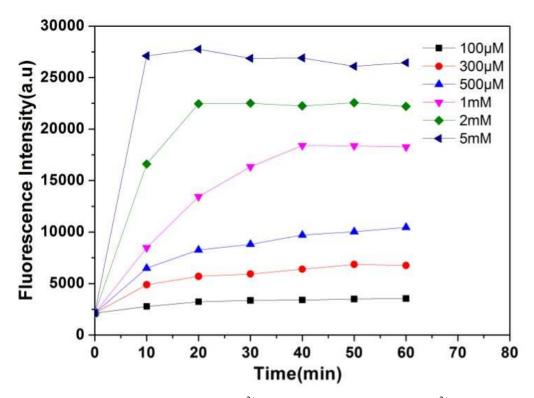


Figure S-34. Fluorescence values of $6:Cu^{2+}(0.05 \text{ wt\% } 6 \text{ with one equiv. of } Cu^{2+})$ treated with different concentrations of GSH at 0-60mins.

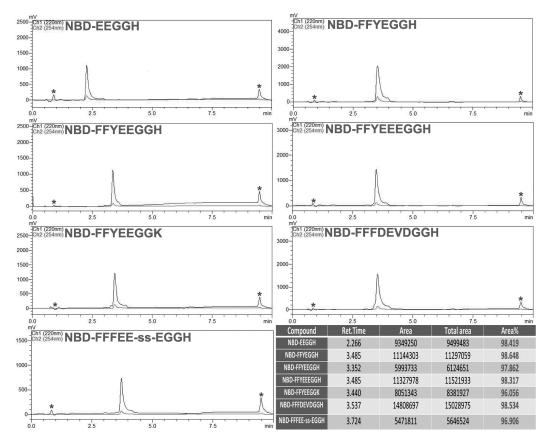


Figure S-35. The purity of peptides obtained from LC-MS data (the peaks assigned by * in every figure are the system peaks).

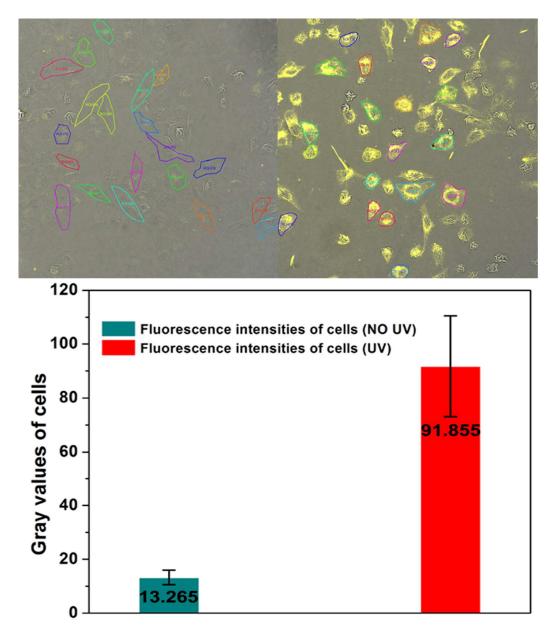


Figure S-36. The mean value of fluorescence intensities of cells from CLSM images (the data were expressed as the mean \pm standard error of the mean (SEM, N = 20)).

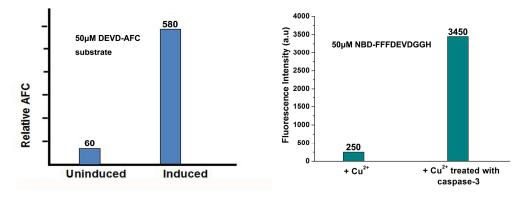


Figure S-36. The comparative data of the commercial probe with our probe (left: the change of fluorescence intensity of the commercial probe, the ratio of two histograms= 9.67; right: the change of fluorescence intensity of our probe, the ratio of two histograms=13.8).