

# Topological effects and binding modes operating with multivalent iminosugar-based glycoclusters and mannosidases

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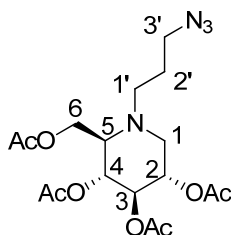
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**Materials** NMR spectra were recorded at room temperature with a Bruker Avance 300 Ultra Shield or eBruker Avance III 400 spectrometer and chemical shifts are reported in parts per million relative to tetramethylsilane or a residual solvent peak (CHCl<sub>3</sub>: <sup>1</sup>H: δ=7.26, <sup>13</sup>C: δ=77.2; DMSO-d<sub>6</sub>: <sup>1</sup>H: δ=2.54, <sup>13</sup>C: δ=40.4). Peak multiplicity is reported as: singlet (s), doublet (d), triplet (t), quartet (q), multiplet (m), and broad (br). High resolution mass spectra HRMS were obtained by Electrospray Ionisation (ESI) on a Micromass-Waters Q-TOF Ultima Global or with a Bruker Autoflex III SmartBeam spectrometer (MALDI). Low-resolution mass spectra (MS) were recorded with a Thermo electron DSQ spectrometer. All reagents were purchased from Acros Organics or Aldrich and were used without further purification. Column chromatography was conducted on silica gel Kieselgel SI60 (40-63 μm) from Merck. Reactions requiring anhydrous conditions were performed under argon. Dichloromethane was distilled from calcium hydride under nitrogen prior to use. Microwave experiments were conducted in sealed vials in commercial microwave reactors especially designed for synthetic chemistry. (MultiSYNTH, Milestone). The instrument features a special shaking system that ensures high homogeneity of the reaction mixtures. Optical rotations were measured on a 343 PERKIN ELMER at 20°C in a 1cm cell in the stated solvent; [α]<sub>D</sub> values are given in 10<sup>-1</sup> deg.cm<sup>2</sup> g<sup>-1</sup> (concentration c given as g/100 mL).

**N-(3-Azidopropyl)-2,3,4,6-tetra-O-acetyl-1,5-dideoxy-1,5-imino-D-glucitol (14)**



Conventional catalytic hydrogenolysis of **11** (760 mg, 1.25 mmol) was carried out with Pd(OH)<sub>2</sub> (875 mg, 1.25 mmol) in MeOH-1M aq. HCl solution (8 mL, 3:1) at 1 atm for 15 h. Then, the catalyst was filtered over celite and the solvent removed under reduced pressure to give **12**(quant) as a hygroscopic brown gum.

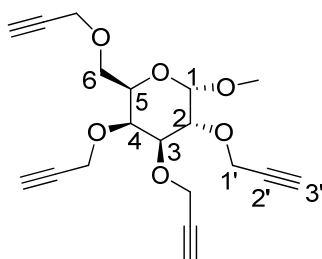
(For analytical data see: J. D. Diot, I. G. Moreno, G. Twigg, C. O. Mellet, K. Haupt, T. D. Butters, J. Kovensky, S. G. Gouin, *J. Org. Chem.*; **2011**, 76, 7757-7768.)

Imidazole-1-sulfonyl azide hydrochloride **13** (326 mg, 1.25 mmol) was added to the crude mixture (275 mg, 1.25 mmol) with K<sub>2</sub>CO<sub>3</sub> (207 mg, 1.5 mmol) and CuSO<sub>4</sub>·5H<sub>2</sub>O (3.2 mg, 12.5 μmol) in MeOH (15 mL) and the mixture was stirred overnight at room temperature.

The solvent was evaporated under reduced pressure and crude **14** was dissolved in pyridine-acetic anhydride (1:1, 14 mL) was stirred for 24 hours. Water (9mL) was added slowly at 0°C to quench the reaction. The aqueous phase was extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 x 15 mL). The combined organic layers were then washed with an aqueous solution of HCl (1M, 2 x 10 mL) and a saturated aqueous NaHCO<sub>3</sub> solution (1 x 15 mL), dried (MgSO<sub>4</sub>), filtered and concentrated. The resulting residue was purified by flash chromatography (AcOEt - petroleum spirit 4 - 6) to give **14** as yellow oil (180 mg, 35% for three steps).

[α]<sub>D</sub> = +3.8 (c= 1, Chloroform); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ = 5.04-4.92 (m, 2H, H-3 and H-4), 4.92-4.80 (m, 1H, H-2), 4.10 (d, *J* = 2.7 Hz, 2H, H-6), 3.28 (t, *J* = 6.6 Hz, 2H, H-3'), 3.12 (dd, *J* = 11.4 and 5 Hz, 1H, H-1<sub>a</sub>), 2.89-2.75 (m, 1H, H-1'), 2.63-2.43 (m, 2H, H-1'<sub>b</sub>,5), 2.22 (dd, *J* = 1.2, *J* = 11.4, 1H, H-1<sub>b</sub>), 2.00, 1.95, 1.93 (3s, 12H, C=OCH<sub>3</sub>), 1.71-1.53 (m, 2H, H-2'); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ = 170.7, 170.2, 170.0, 169.6 (C=O), 74.4 (C-4), 69.2 (C-2), 69.1 (C-3), 61.9 (C-5), 59.5 (C-6), 52.7 (C-1), 49.0 (C-3'), 48.4 (C-1'), 25.2 (C-2'), 20.7 (CH<sub>3</sub>).; HRMS (MALDI): Found 415.1816 C<sub>9</sub>H<sub>19</sub>O<sub>4</sub>N<sub>4</sub>Na requires 415.1823.

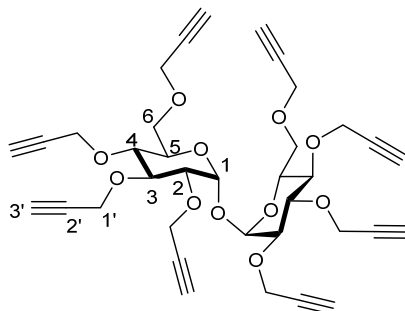
### Methyl 2,3,4,6-tetra-*O*-propargyl- $\alpha$ -D-galactopyranoside (**18**)



NaH (412 mg, 10.3 mmol, dispersed in oil, 60%) was added in a small portion at 0°C to a solution of methyl  $\alpha$ -D-galactopyranoside (200 mg, 1.03 mmol) in anhydrous DMF (10 mL). The suspension was stirred for one hour at room temp. Propargyl bromide (690  $\mu$ L, 6.18 mmol, dispersed in toluene 80%) was added dropwise, and the reaction mixture was maintained for 3 hours. The mixture was cooled, MeOH was added 10 mL, and the solvent removed under reduced pressure after 30 min. Water 10 mL was added and the compound was extracted with diethyl ether (2 x 20 mL). The organic phase was dried (MgSO<sub>4</sub>), filtered, and the solvent were evaporated under reduced pressure. The crude material was purified by column chromatography (8-2 EtOAc - cyclohexane) to give **18** as a yellow oil (306 mg, 86%).

$[\alpha]_D = +31$  (c= 0.5, CHCl<sub>3</sub>); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  = 4.94 (1 H, d,  $J$  = 3.6 Hz, H-1), 4.49 (2 H, d,  $J$  = 2.3 Hz, CH<sub>2</sub>CCH), 4.41(4 H, m, CH<sub>2</sub>CCH), 4.24-4.18 (2 H, dd,  $J$  = 2.4 Hz,  $J$  = 7.9 Hz, CH<sub>2</sub>CCH), 4.12-4.07 (1 H, m, H-4), 4.03-3.88 (3 H, m, H-2,3,5), 3.78 (1 H, dd,  $J$  = 5.7 Hz,  $J$  = 9.6 Hz, H-6<sub>A</sub>), 3.66 (1 H, dd,  $J$  = 6.6 Hz, H-6<sub>B</sub>), 3.42 (3 H, s, OMe), 2.44 (4 H, m, CCH).; <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  = 98.8 (C-1), 80.4, 80.2, 79.7 (C-2'), 78.6 (C-3), 76.2 (C-2), 74.7 (C-3'), 74.6 (C-4), 69.3 (C-6), 69.1 (C-5), 59.9, 59.1, 58.9, 58.8 (C-1'), 55.5 (OMe); HRMS (MALDI) [M+Na]<sup>+</sup>: Found 369.1305 C<sub>19</sub>H<sub>22</sub>O<sub>6</sub>Na requires 369.1309.

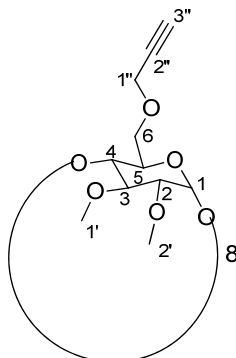
### Propargyl- $\alpha$ -D-glucopyranosyl- $\alpha$ -D-glucopyranoside (**20**)



NaH (280 mg, 7.01 mmol, dispersed in oil, 60%) was added in a small portion at 0°C to a solution of  $\alpha$ -D-glucopyranosyl- $\alpha$ -D-glucopyranoside (150 mg, 0.44 mmol) in anhydrous DMF (10 mL). The suspension was stirred for one hour at room temp. Propargyl bromide (490  $\mu$ L, 4.38 mmol, dispersed in toluene 80%) was added dropwise, and the reaction mixture was stirred for 6 hours. mixture was cooled, MeOH was added 10 mL, and the solvent removed under reduced pressure after 30 min. Water 10 mL was added and the compound was extracted with diethyl ether (2 x 20 mL). The organic phase was dried (MgSO<sub>4</sub>), filtered, and the solvent were evaporated under reduced pressure. The crude material was purified by column chromatography (EtOAc - cyclohexane, 7-3) to give **20** as a yellow oil (130 mg, 46%).

$[\alpha]_D = +125$  (c= 0.3, CHCl<sub>3</sub>); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  = 5.18 (2 H, d,  $J$  = 3.6 Hz, H-1), 4.56-4.35 (8 H, br, H-1'), 4.35-4.10 (8 H, br, H-1'), 4.13-4.01 (2 H, m, H-5), 3.83 (2 H, dd,  $J$  = 3.5 Hz, 10.5 Hz, H-6<sub>A</sub>), 3.75 (2 H, t,  $J$  = 9.3 Hz, H-3-3'), 3.70 (2 H, dd,  $J$  = 2.1 Hz,  $J$  = 10.5 Hz, H-6<sub>B</sub>), 3.56 (2 H, dd,  $J$  = 9.5, 3.6 Hz, H-2), 3.48 (2 H, m, H-4), 2.49-2.38 (8 H, m, H-1'); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  = 94.0 (C-1), 81.3 (C-3), 80.2-80.1-79.7-79.5 (C-3'), 78.9 (C-2), 76.6 (C-4), 75.0 (C-2'), 74.3 (C-3'), 69.9 (C-5), 67.8 (C-6), 60.5, 60.3, 58.6, 58.4 (C-1'); HRMS (MALDI)  $[M+Na]^+$ : Found 669.2317 C<sub>36</sub>H<sub>38</sub>O<sub>11</sub>Na requires 669.2306.

## Octakis(6-O-propargyl-2,3-di-O-methyl)cyclomaltooctaose (**21**)



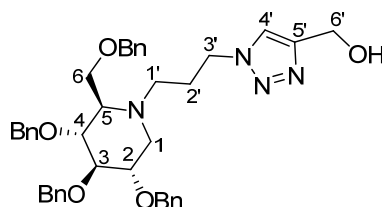
NaH (105 mg, 2.62 mmol, dispersed in oil, 60%) was added in small portions at 0°C to a solution of octakis(2,3-di-*O*-methyl)cyclomaltooctaose (200 mg, 0.131 mmol - K. Takeo, H. Mitoh, K. Uemura, *Carbohydrate Res.* **1989**, 187, 203-221) in anhydrous DMF (15 mL). The suspension was magnetically stirred for one hour at room temp. Propargyl bromide (180  $\mu$ L, 1.57 mmol,) dispersed in toluene 80%, was then added dropwise, and the reaction mixture was stirred for 6 hours. The mixture was cooled, MeOH was added (5 mL), and the solvent removed under reduced pressure after 30 min. Water (5 mL) was added and the compound was extracted with diethyl ether (2 x 15 mL). The organic phase was dried (MgSO<sub>4</sub>), filtered, and the solvent were evaporated under reduced pressure. The crude material was purified by column chromatography (EtOAc - cyclohexane 96 - 4) to give **21** as a brown gum (192 mg, 80%).

$[\alpha]_D = +101$  (c= 0.5, CHCl<sub>3</sub>); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  = 5.23 (8 H, d,  $J$  = 3.6 Hz, H-1), 4.22 (16 H, dq,  $J$  = 2.61 Hz,  $J$  = 15.6 Hz, H-1'), 3.96-3.78 (24 H, m, H-5,-6), 3.75-3.66 (8 H, m, H-5), 3.72-3.58 (32 H, m, H-1',-4), 3.58-3.46 (32 H, m, H-2', -3), 3.20 (8 H, dd,  $J$  = 3.4 Hz,  $J$  = 9.66 Hz, H-2,), 2.49 (8 H, t,  $J$  = 2.3 Hz, H-3''); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  = 98.3 (C-1), 82.2 (C-3), 82.0 (C-2), 79.8 (C-2''), 79.0 (C-4), 75.1 (C-3''), 70.9 (C-5), 68.7 (C-6), 61.6 (C-1'), 58.9 (C-2'), 58.6 (C-1''); HRMS (MALDI)  $[M+Na]^+$ : Found 1847.7832 C<sub>88</sub>H<sub>128</sub>O<sub>40</sub>Na requires 1847.7874.

## General procedure for CuAAC

To a solution of the **18** (30 mg, 86.6  $\mu$ mol) and the azido-derivative (230 mg, 0.38 mmol) in dioxane-H<sub>2</sub>O (5 mL, 4–1) copper sulfate (65 mg, 0.26 mmol) and sodium ascorbate (103 mg, 0.52 mmol) were added and the mixture was irradiated at 80 °C for 45 min in a sealed vessel. The mixture was poured into a NH<sub>4</sub>Cl satd. solution (20 mL) and extracted with ethyl acetate. The organic layer was dried (MgSO<sub>4</sub>), filtered and the solvent removed under reduced pressure. The crude product was purified by flash chromatography on silica gel (AcOEt – MeOH, 99 - 1), colorless oil, 66% yield.

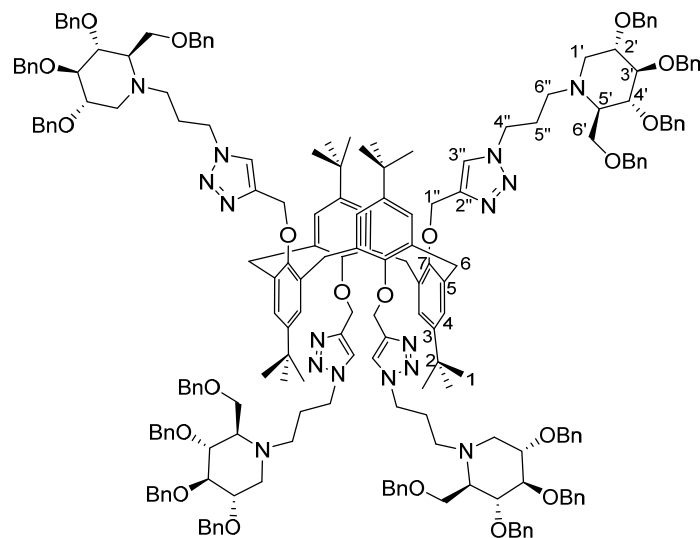
## Compound (22)



**22** was purified by flash chromatography on silica gel (AcOEt – cyclohexane, 99 - 1), colorless oil, 92% yield.

$[\alpha]_D = +1$  ( $c = 0.3$ , CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  = 7.33 (1 H, s, H-4'), 7.31-7.05 (20 H, m, 20  $\times$  Har), 4.95-4.71 (3 H, m, CHHPh), 4.71-4.55 (4 H, m, 2  $\times$  CH<sub>2</sub>Ph, 2  $\times$  H-6'), 4.45-4.28 (3 H, m, CHHPh), 4.25-4.05 (2 H, m, H-3'), 3.61 (1 H, dd,  $J = 3.7$  Hz,  $J = 10.8$  Hz, H-6A), 3.55-3.37 (4 H, m, H-2, -3, -4, -6B), 2.93 (1 H, dd,  $J = 4.14$  Hz,  $J = 11.5$  Hz, H-1A), 2.83-2.65 (1 H, m, H-1'A), 2.52-2.35 (1 H, m, H-1'B), 2.35-2.25 (1 H, m, H-5), 2.08 (t,  $J = 10.5$  Hz, 1 H, H-1B), 2.01-1.85 (m, 2H, H-2'); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  = 147.7 (C-5'), 139.0, 138.5, 137.9 (Car), 128.6-127.5 (CHar), 122.1 (C-4'), 87.1 (C-3), 78.6 (C-2), 78.4 (C-4), 75.4, 75.3, 73.3, 72.9 (CH<sub>2</sub>Ph), 66.6 (C-6), 64.7 (C-5), 56.5 (C-6'), 54.4 (C-1), 49.3 (C-1'), 48.5 (C-3'), 26.2 (C-2'); HRMS (MALDI)  $[M+Na]^+$ : Found 663.3533 C<sub>40</sub>H<sub>46</sub>N<sub>4</sub>O<sub>5</sub> Na requires 663.3541.

## Compound (23)

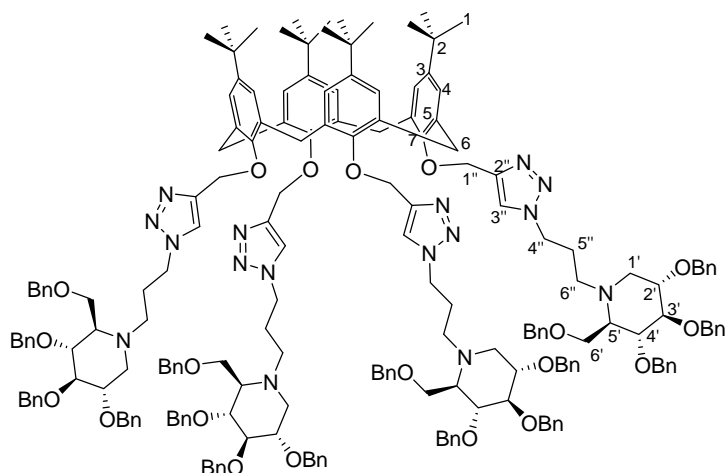


**23** was purified by flash chromatography on silica gel (AcOEt – cyclohexane, 95 - 5), colorless oil, 75% yield.

$[\alpha]_D = +4$  ( $c = 0.5$ ,  $\text{CHCl}_3$ );  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.33\text{--}7.05$  (80 H, m, Har), 6.92–6.84 (12 H, m, 8 x H-4, 4 x H-3''), 5.02–4.77 (12 H, m,  $\text{CH}_2\text{Ph}$ ), 4.72–4.60 (8 H, m,  $\text{CH}_2\text{Ph}$ ), 4.54–4.37 (20 H, m, 12 x  $\text{CH}_2\text{Ph}$ , 4 x H-1''), 4.33–4.11 (8 H, m, H-4''), 3.74–3.41 (28 H, m, 4 x H-2', 3', 4', 6', 8 x H-6), 3.08 (4 H, dd,  $J = 3.7$  Hz,  $J = 10.8$  Hz, H-1'A), 2.93–2.79 (4 H, m, H-6''A), 2.70–2.58 (4 H, m, H-6''B), 2.40–2.30 (4 H, m, H-5'), 2.22 (4 H, t,  $J = 10.5$  Hz, H-1'B), 2.09–1.90 (8 H, m, H-5''), 1.09 (36 H, s, H-1);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ ):  $\delta = 153.5$  (C<sub>IV</sub>), 144.8, 144.7 (C-2''), 139.0, 138.6, 137.9 (Car), 133.6 (C<sub>IV</sub>), 128.8–127.3 (CHar), 126.8 (C-4), 123.0 (C-3''), 87.2 (C-3'), 78.7 (C-2'), 78.5 (C-4'), 75.4, 75.3, 73.3, 72.8 ( $\text{CH}_2\text{Ph}$ ), 66.4 (C-6'), 64.5 (C-1''), 64.4 (C-5'), 54.7 (C-1'), 49.5 (C-6''), 48.3 (C-4''), 38.6 (C-6), 33.9 (C-2), 31.6 (C-1), 25.9 (C-5''); HRMS (ES)  $[\text{M}+2\text{H}]^{2+}$ : Found 1076.2614  $\text{C}_{204}\text{H}_{234}\text{N}_{16}\text{O}_{20}$  requires 1076.2616.



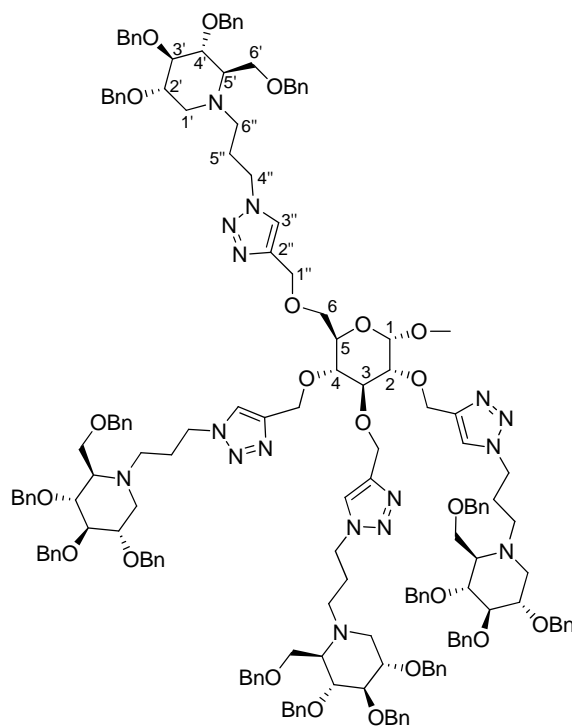
## Compound (24)



**24** was purified by flash chromatography on silica gel (AcOEt – cyclohexane, 98 - 2), colorless oil, 64% yield.

$[\alpha]_D = +5$  ( $c = 1$ ,  $\text{CHCl}_3$ );  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.67$  (4 H, m, 4 x H-3''), 7.31-7.05 (80 H, m, Har), 6.72 (8 H, s, H-4), 5.05 (8 H, m, H-1''), 4.97-4.67 (12 H, m,  $\text{CH}_2\text{Ph}$ ), 4.69-4.59 (8 H, m,  $\text{CH}_2\text{Ph}$ ), 4.45-4.31 (12 H, m,  $\text{CH}_2\text{Ph}$ ), 4.29-4.09 (12 H, m, 4 x H-6<sub>B</sub>, 8 x H-4''), 3.68-3.41 (20 H, m, H-2', 3', 4', 6'), 3.04 (4 H, dd,  $J = 4.8$  Hz,  $J = 11.1$  Hz, H-1'<sub>A</sub>), 2.87 (4 H, d,  $J = 12.7$  Hz, 4 x H-6<sub>B</sub>), 2.82-2.69 (4 H, m, H-6''<sub>A</sub>), 2.61-2.46 (4 H, m, H-6''<sub>B</sub>), 2.36-2.27 (4 H, m, H-5'), 2.16 (4 H, t,  $J = 10.8$  Hz, H-1'<sub>B</sub>), 2.04-1.87 (8 H, m, H-5''), 1.06 (36 H, s, H-1);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ ):  $\delta = 152.2$  ( $\text{C}_{\text{IV}}$ ), 145.1, 144.6 ( $\text{C}-2''$ ), 139.1, 138.7, 137.9 (Car), 134.2 ( $\text{C}_{\text{IV}}$ ), 128.4-127.3 ( $\text{CHar}$ ), 125.1 ( $\text{C}-4$ ), 124.5 ( $\text{C}-3''$ ), 87.3 ( $\text{C}-3'$ ), 78.7 ( $\text{C}-2'$ ), 78.5 ( $\text{C}-4'$ ), 75.4, 75.2, 73.3, 72.8 ( $\text{CH}_2\text{Ph}$ ), 66.5 ( $\text{C}-6'$ ,  $\text{C}-1''$ ), 64.4 ( $\text{C}-5'$ ), 54.6 ( $\text{C}-1'$ ), 49.4 ( $\text{C}-6''$ ), 48.4 ( $\text{C}-4''$ ), 34.0 ( $\text{C}-2$ ), 31.6 ( $\text{C}-6$ ), 31.4 ( $\text{C}-1$ ), 26.1 ( $\text{C}-5''$ ); HRMS (ES)  $[\text{M}+2\text{H}]^{2+}$ : Found 1613.8887  $\text{C}_{204}\text{H}_{234}\text{N}_{16}\text{O}_{20}$  requires 1613.8854.

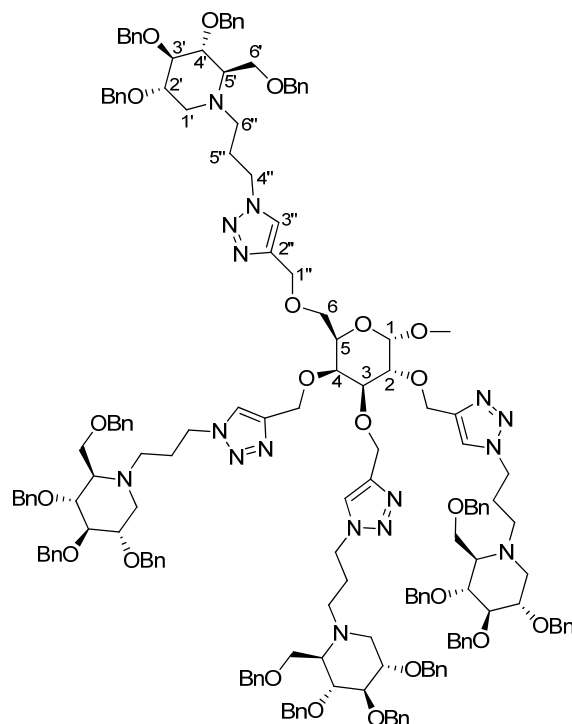
## Compound (25)



**25** was purified by flash chromatography on silica gel (AcOEt – MeOH, 99 - 1), colorless oil, 69% yield.

$[\alpha]_D = +28$  ( $c = 0.5$ ,  $\text{CHCl}_3$ );  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.96, 7.78, 7.76, 7.54$  (4 H, s, 4 x H-3''), 7.45-7.12 (80 H, m, Har), 5.1-4.75 (18 H, m, 12 x CHHPh, 5 x H-1'', H-1), 4.78-4.63 (11 H, m, 4 x  $\text{CH}_2\text{Ph}$ , 3 x H-1''), 4.50-4.32 (12 H, m, CHHPh), 4.34-4.13 (8 H, m, H-4''), 3.92 (1 H, t,  $J = 9.2$  Hz, H-3), 3.84 (1 H, dd,  $J = 3.7$  Hz,  $J = 10.5$  Hz, H-6<sub>A</sub>), 3.78-3.45 (24 H, m, H-2', 3', 4', 6', H-2,4,5,6<sub>B</sub>), 3.38 (3 H, s,  $\text{OCH}_3$ ), 2.93-2.78 (4 H, m, H-1'<sub>A</sub>), 2.69-2.59 (4 H, m, H-6''<sub>A</sub>), 2.46-2.29 (4 H, m, H-6''<sub>B</sub>), 2.20-2.09 (4 H, m, H-5'), 2.05-1.92 (4 H, m, H-1'<sub>B</sub>), 1.89-1.71 (8 H, m, H-5'');  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta = 145.0, 144.8, 144.7, 144.6$  (C-2''), 139.0, 138.5, 137.8 (Car), 128.7-127.5 (CHar), 123.8, 123.5, 123.0 (C-3''), 97.8 (C-1), 87.2 (C-3'), 81.6 (C-3), 79.8 (C-2), 78.6 (C-2'), 78.4 (C-4'), 77.5 (C-4), 75.3, 75.2, 73.3, 72.8 ( $\text{CH}_2\text{Ph}$ ), 70.0 (C-5), 68.8 (C-6), 66.4 (C-1''), 64.8 (C-6'), 64.4 (C-5'), 55.2 ( $\text{OCH}_3$ ), 54.4 (C-1'), 49.2 (C-6''), 48.4 (C-4''), 25.9 (C-5''); HRMS (MALDI)  $[\text{M}+\text{Na}]^+$ : Found 2794.4184  $\text{C}_{167}\text{H}_{190}\text{N}_{16}\text{O}_{22}\text{Na}$  requires 2794.4133.

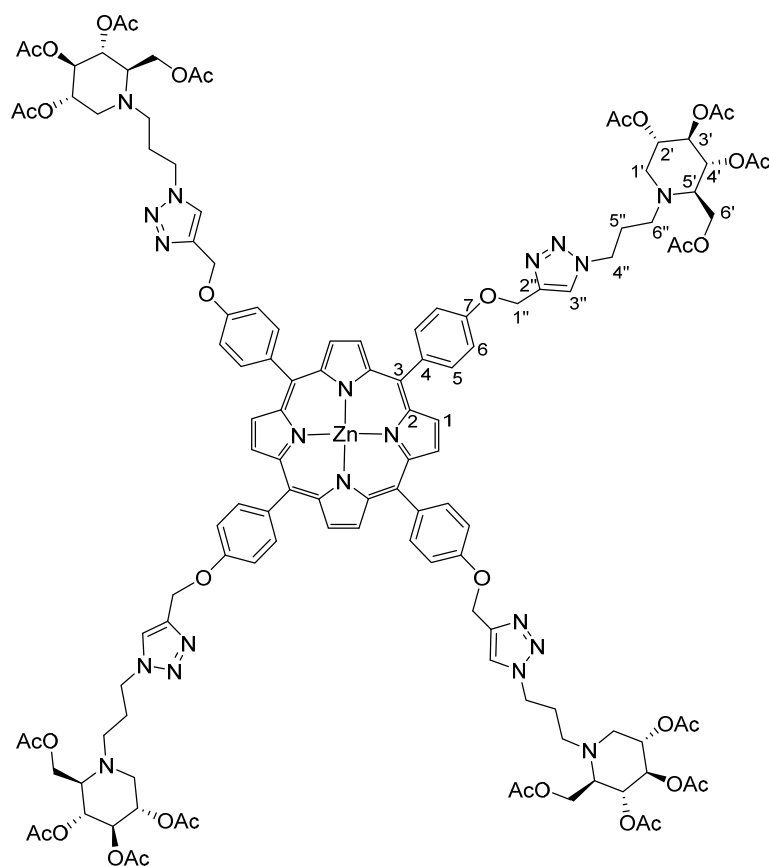
## Compound (26)



**26** was purified by flash chromatography on silica gel (AcOEt – MeOH, 99 - 1), colorless oil, 66% yield.

$[\alpha]_D = +14$  ( $c = 0.5$ ,  $\text{CHCl}_3$ );  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.75, 7.61, 7.58, 7.48$  (4 H, s, H-3''), 7.40-7.08 (80 H, m, Har), 5.02-4.72 (18 H, m, 12 x  $\text{CHHPH}$ , 5 x H-1'', H-1), 4.72-4.50 (11 H, m, 4 x  $\text{CH}_2\text{Ph}$ , 3 x H-1''), 4.49-4.32 (12 H, m,  $\text{CH}_2\text{Ph}$ ), 4.32-4.08 (8 H, m, H-4''), 4.04-4.96 (2 H, m, H-2, -3), 4.93-4.83 (2 H, m, H-4, -5), 3.70-3.40 (22 H, m, 4 x H-2', 3', 4', 6', H-6), 3.34 (3 H, s,  $\text{OCH}_3$ ), 3.12-2.98 (4 H, m, H-1'A), 2.83-2.71 (4 H, m, H-6''A), 2.64-2.46 (4 H, m, H-6''B), 2.39-2.28 (4 H, m, H-5'), 2.23-2.10 (4 H, m, H-1'B), 2.07-1.90 (8 H, m, H-5'');  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ ):  $\delta = 145.0, 144.9, 144.5$  (C-2''), 139.0, 138.5, 137.8 (Car), 128.7-127.5 ( $\text{CHar}$ ), 123.2-122.9 (C-3''), 98.5 (C-1), 87.1 (C-3'), 79.3 (C-3), 78.9 (C-2), 78.6 (C-2'), 78.4 (C-4'), 77.4 (C-4), 75.3, 75.0, 73.3, 72.8 ( $\text{CH}_2\text{Ph}$ ), 69.2 (C-5), 69.0 (C-6), 66.2 (C-1''), 65.0 (C-6'), 64.5 (C-5'), 55.3 ( $\text{OCH}_3$ ), 54.4 (C-1'), 49.2 (C-6''), 48.3 (C-4''), 25.9 (C-5''); HRMS (MALDI)  $[\text{M}+\text{Na}]^+$ : Found 2794.4111  $\text{C}_{167}\text{H}_{190}\text{N}_{16}\text{O}_{22}\text{Na}$  requires 2794.4133.

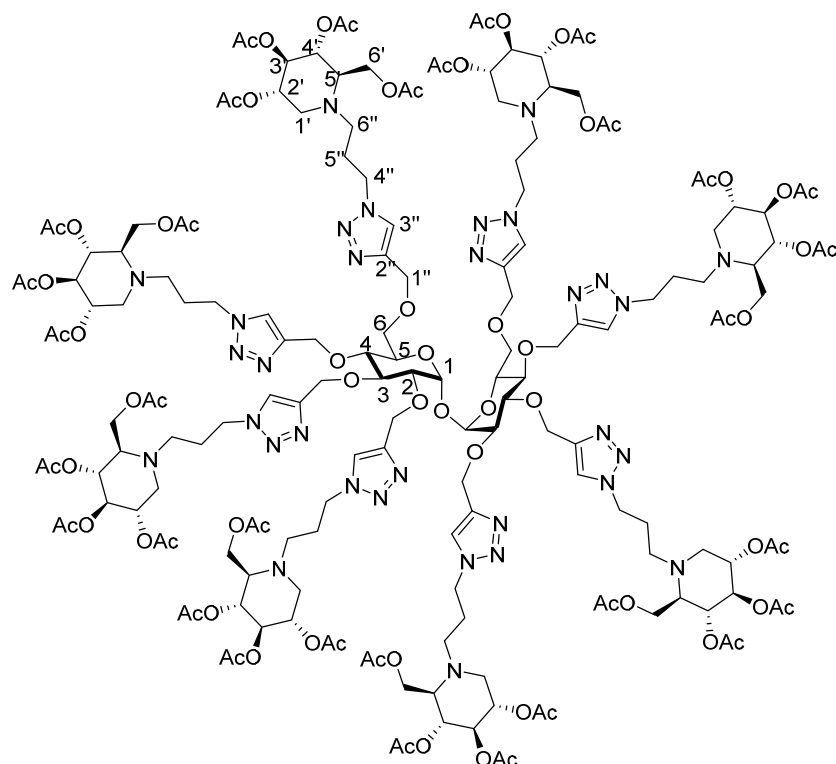
## Compound (27)



**27** was purified by flash chromatography on silica gel (DCM – MeOH, 8 - 2), purple solid, 66% yield.

$[\alpha]_D$  = too dark for analysis;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  = 8.93 (8 H, s, H-1), 8.11 (8 H, d,  $J$  = 8.3 Hz, H-5), 7.43 (4 H, s, H-3''), 7.22 (8 H, d,  $J$  = 8.3 Hz, H-6), 5.09-4.97 (8 H, m, H-3', -4'), 4.97-4.83 (4 H, m, H-2'), 4.67 (8 H, s, H-1''), 4.22-3.99 (16 H, m, H-6', -4''), 3.11 (4 H, dd,  $J$  = 4.9 Hz,  $J$  = 11.8 Hz, H-1'A), 2.77-2.65 (4 H, m, H-6''B), 2.61-2.51 (4 H, m, H-5'), 2.42-2.28 (4 H, m, H-6''A), 2.25-2.15 (4 H, m, H-1'B), 2.06-1.96 (48 H, m,  $\text{CH}_3$ ), 1.94-1.84 (8 H, m, H-5'');  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  = 170.7-169.8 (C=O), 157.9, 150.5, 143.9, 136.5 ( $\text{C}_{\text{IV}}$ ), 135.9 (C-5), 131.8 (C1), 122.5 (C-2''), 120.4 ( $\text{C}_{\text{IV}}$ ), 112.9 (C-6), 74.4 (C-4'), 69.4 (C-3'), 69.2 (C-2'), 62.4 (C-5'), 61.9 (C-1''), 59.6 (C-6'), 52.7 (C-1'), 48.4 (C-6''), 47.9 (C-4''), 27.1 (C-5''), 20.9 ( $\text{CH}_3$ ); HRMS (ES)  $[\text{M}+2\text{H}]^{2+}$ : Found 1275.4588  $\text{C}_{124}\text{H}_{142}\text{N}_{20}\text{O}_{36}\text{Zn}$  requires 1275.4577.

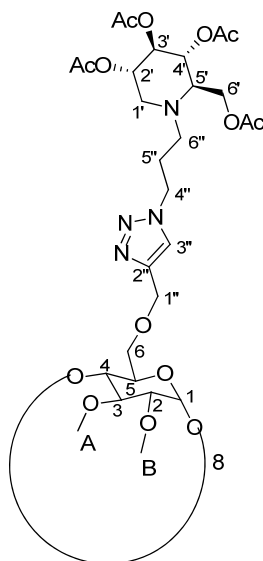
## Compound (28)



**28** was purified by flash chromatography on silica gel (DCM – MeOH, 98 - 2), colorless oil, 58% yield.

$[\alpha]_D = +24$  ( $c = 1$ ,  $\text{CHCl}_3$ );  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta = 8.07, 8.04, 7.98, 7.74$  (8 H, s, H-3''), 5.18 (2 H, d,  $J = 3.46$  Hz, H-1), 5.08-4.98 (16 H, m, H-3', -4'), 4.98-4.74 (16 H, m, 8 x H-2', 8 x H-1''), 4.73-4.57 (8 H, m, H-1''), 4.50-4.28 (16 H, m, H-4''), 4.19-4.01 (18 H, m, 16 x H-6', 2 x H-5), 3.80 (2 H, t,  $J = 9.3$  Hz, H-3), 3.75-3.65 (4 H, m, H-6), 3.56-3.41 (4 H, m, H-2, -4), 3.26-3.13 (8 H, m, H-1'A), 2.97-2.79 (8 H, m, H-6''), 2.72-2.47 (16 H, m, H-5', -6''), 2.41-2.22 (8 H, m, H-1'B), 2.18-1.89 (112 H, m, H-5'',  $\text{CH}_3$ );  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta = 170.3$ -169.7 (C=O), 145.2-144.3 (C-2''), 124.3-123 (C-3''), 93.6 (C-1), 81.1 (C-3), 79.4 (C-2), 77.3 (C-4), 74.5 (C-4'), 70.7 (C-5), 69.4 (C-6,2',3'), 66.4, 66.0, 64.8, 64.4 (C-1''), 62.1 (C-5'), 59.8 (C-6'), 52.8 (C-1'), 48.6 (C-6''), 48.0 (C-4''), 26.9 (C-5''), 20.9 ( $\text{CH}_3$ ); HRMS (MALDI)  $[\text{M}+4\text{H}]^{4+}$ : Found 990.9178  $\text{C}_{172}\text{H}_{250}\text{N}_{32}\text{O}_{75}$  requires 990.9216.

## Compound (29)



**29** was purified by flash chromatography on silica gel (DCM – MeOH, 99 - 1), colorless oil, 57% yield.

$[\alpha]_D = +57$  ( $c = 1$ ,  $\text{CHCl}_3$ );  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta = 7.70$  (8 H, s, H-3''), 5.19 (8 H, d,  $J = 3.43$  Hz, H-1), 5.08-4.99 (16 H, m, H-3', -4'), 4.98-4.87 (8 H, m, H-2'), 4.64-4.50 (16 H, m, H-1''), 4.45-4.30 (16 H, m, H-4''), 4.14 (16 H, d,  $J = 2.2$  Hz, H-6'), 4.01-3.90 (8 H, m, H-6<sub>A</sub>), 3.82-3.44 (80 H, m, H-3, -4, -5, -6<sub>B</sub>, A, B), 3.25-3.08 (16 H, m, H-2, -1'<sub>A</sub>), 2.94-2.83 (8 H, m, H-6''<sub>A</sub>), 2.71-2.53 (16 H, m, H-5', -6''<sub>B</sub>), 2.36-2.26 (8 H, m, H-1'<sub>B</sub>), 2.08-1.97 (112 H, m, H-5'',  $\text{CH}_3$ );  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta = 170.6$ -169.7 (C=O), 145.0 (C-2''), 123.0 (C-3''), 98.2 (C-1), 82.1 (C-2,-3), 78.6 (C-4), 74.4 (C-4'), 71.2 (C-5), 69.2 (C-6, -2', -3'), 64.9 (C-1''), 61.9 (C-5'), 61.4 (C-A), 59.6 (C-6'), 58.8 (C-B), 52.7 (C-1'), 48.6 (C-6''), 47.9 (C-4''), 26.8 (C-5''), 20.7 ( $\text{CH}_3$ ).; HRMS (ES)  $[\text{M}+5\text{H}]^{5+}$ : Found 1028.6470  $\text{C}_{224}\text{H}_{341}\text{N}_{32}\text{O}_{104}$  requires 1028.6488.

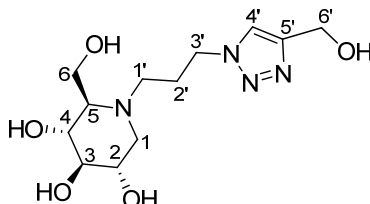
## General procedure for benzyl hydrogenolysis

Catalytic hydrogenation of compound **26** (80 mg, 28.8  $\mu$ mol) was carried out with Pd(OH)<sub>2</sub> (73 mg) in MeOH-1M aq. HCl solution (10 mL, 8:2) at 1 atm for 48 h. The catalyst was filtered over celite and the solvent removed under reduced pressure to give compound **5** (quant) as a sticky solid.

## General procedure for acetates deprotection

Compound **29** (30 mg, 0.0058 mmol) was dissolved in MeOH-H<sub>2</sub>O (1-1, 5 mL). Amberlite resin IRA 400 OH- 1.25 meq/mL (780 mg) was added to the solution, and the mixture was stirred overnight at room temperature. The resin was filtered off and washed with methanol and water. The solvent was evaporated under reduced pressure to give compound **8** (20 mg, 91%) as pale yellow solid.

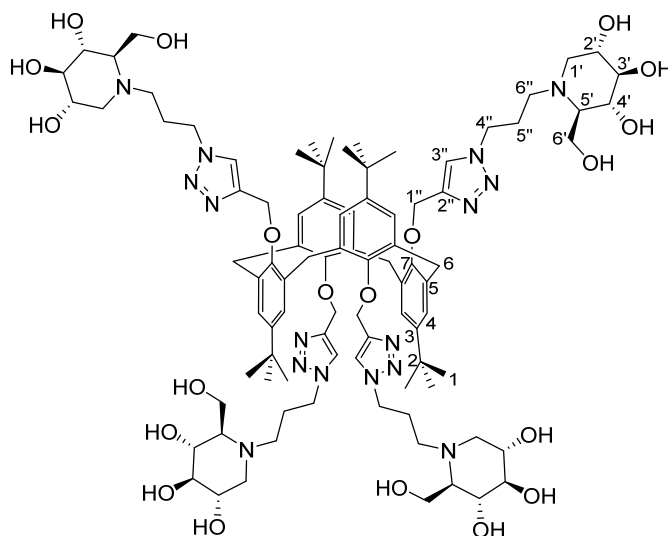
## Compound (1)



**1** was purified by flash chromatography on silica gel (AcOEt – cyclohexane, 99 - 1), white solid, 92% yield.

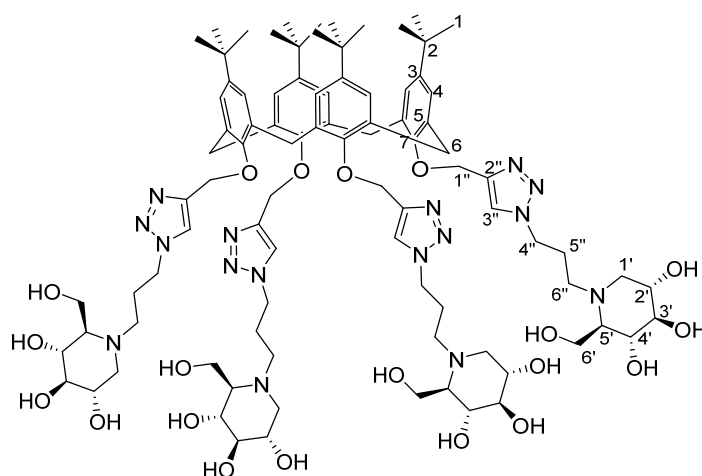
$[\alpha]_D = -8$  ( $c = 1$ , H<sub>2</sub>O); <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O)  $\delta$  = 8.05 (1 H, s, H-4'), 4.79 [under solvent peak] (2 H, H-6'), 4.57-4.47 (2 H, m, H-3'), 3.82 (2 H, d,  $J = 2.57$  Hz, H-6), 3.62-3.54 (1 H, m, H-2), 3.42 (1 H, t,  $J = 9.4$  Hz, H-4), 3.31 (1 H, t,  $J = 9.2$  Hz, H-3), 3.02 (1 H, dd,  $J = 4.96$  Hz,  $J = 11.3$  Hz, H-1<sub>A</sub>), 2.87-2.77 (1 H, m, H-1'<sub>A</sub>), 2.75-2.55 (1 H, m, H-1'<sub>B</sub>), 2.39-2.27 (2 H, m, H-1<sub>B</sub>, -5), 2.24-2.14 (2 H, m, H-2'); <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O):  $\delta$  = 147.0 (C-5'), 124.1 (C-4'), 78.3 (C-3), 70.0 (C-4), 68.9 (C-2), 64.8 (C-5), 57.6 (C-6), 55.3 (C-1), 54.7 (C-6'), 49.3 (C-3'), 48.6 (C-1'), 23.2 (C-2'); HRMS (MALDI)  $[M+H]^+$ : Found 303.1655 C<sub>12</sub>H<sub>22</sub>N<sub>4</sub>O<sub>5</sub> requires 303.1663.

## Compound (2)



White solid,  $[\alpha]_D = -4$  ( $c = 0.3$ ,  $H_2O$ );  $^1H$  NMR (400 MHz,  $D_2O$ )  $\delta = 7.99$  (4 H, s, H-3''), 7.03 (8 H, m, H-4), 4.75-4.72 (8 H, m, H-1''), 4.70-4.59 (8 H, m, H-4''), 4.08 (4 H, d,  $J = 13$  Hz, H-6'A), 4.02-3.94 (4 H, m, H-6'B), 3.93-3.85 (4 H, m, H-2'), 3.77-3.70 (4 H, m, H-4'), 3.70-3.52 (20 H, m, H-6, -1'A, -3', -6'A), 3.49-3.37 (4 H, m, H-6'B), 3.29 (4 H, d, H-5'), 3.19 (4 H, t,  $J = 11.8$  Hz, H-1'B), 2.62-2.45 (8 H, m, H-5''), 1.06 (36 H, s, H-1);  $^{13}C$  NMR (100 MHz,  $D_2O$ ):  $\delta = 153.6$  (C<sub>IV</sub>), 145.0, 144.4 (C-2''), 133.4 (C<sub>IV</sub>), 127.1 (C-4), 125.9 (C-3''), 75.8 (C-3'), 67.1 (C-4'), 65.9 (C-2'), 65.7 (C-5'), 63.8 (C-1''), 53.9 (C-6'), 53.3 (C-1'), 50.1 (C-6''), 47.3 (C-4''), 36.8 (C-6), 33.4 (C-2), 30.8 (C-1), 23.6 (C-5''); HRMS (ES)  $[M+2Na]^{2+}$ : Found 915.4951  $C_{92}H_{136}N_{16}Na_2O_{20}$  requires 915.4968.

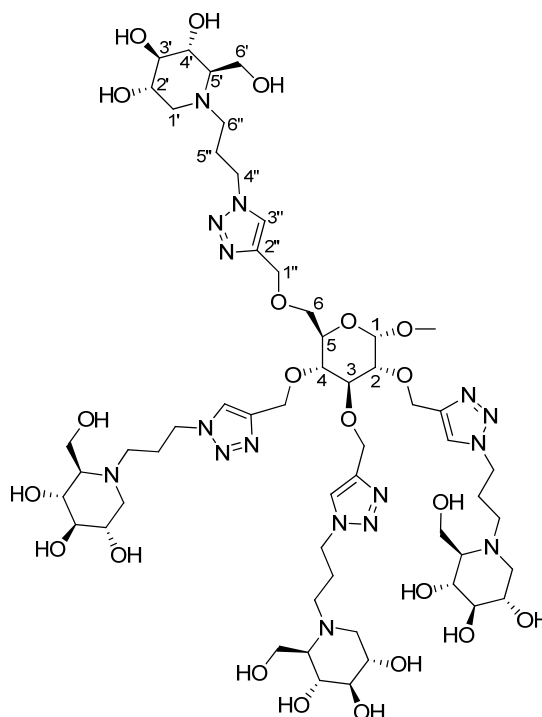
## Compound (3)





White solid,  $[\alpha]_D = -8$  ( $c = 0.1$ , MeOH);  $^1\text{H}$  NMR (400 MHz, MeOD)  $\delta = 8.08$  (4 H, s, H-3''), 6.79 (8 H, s, H-4), 5.05 (8 H, s, H-1''), 4.51-4.43 (8 H, m, H-4''), 4.22 (4 H, d, H-6<sub>A</sub>), 3.86-3.75 (8 H, m, H-6'), 3.50-3.42 (4 H, m, H-2'), 3.36 (4 H, t,  $J = 9.2$  Hz, H-4'), 3.15 (4 H, t,  $J = 9$  Hz, H-3'), 3.03-2.94 (8 H, m, H-6<sub>B</sub>, -1'<sub>A</sub>), 2.91-2.78 (4 H, m, H-6''<sub>A</sub>), 2.53-2.43 (4 H, m, H-6''<sub>B</sub>), 2.15-2.00 (16 H, m, H-1'<sub>B</sub>, -5', -5''), 0.97 (36 H, s, H-1);  $^{13}\text{C}$  NMR (100 MHz, MeOD):  $\delta = 153.4$ , 146.5, 145.7, 135.5 (C<sub>IV</sub>), 126.6 (C-3''), 126.3 (C-4), 80.5 (C-3'), 72.1 (C-4'), 70.8 (C-2'), 67.9 (C-5'), 67.4 (C-1'') 59.6 (C-6'), 57.8 (C-1'), 50.3 (C-6''), 49.5 (C-4''), 34.8 (C-2), 32.9 (C-6), 32.0 (C-1), 27.3 (C-5''); HRMS (ES)  $[\text{M}+2\text{Na}]^{2+}$ : Found 915.4951 cf comp prec C<sub>92</sub>H<sub>136</sub>N<sub>16</sub>Na<sub>2</sub>O<sub>20</sub> requires 915.4949.

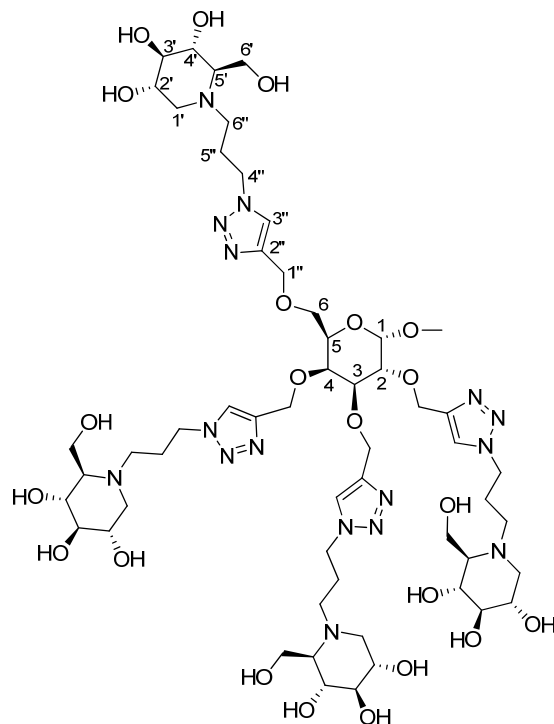
#### Compound (4)



White solid,  $[\alpha]_D = -15$  ( $c = 1$ , H<sub>2</sub>O);  $^1\text{H}$  NMR (400 MHz, D<sub>2</sub>O)  $\delta = 8.17$  (3 H, s, H-3''), 8.12 (1 H, s, H-3''), 4.91-4.55 (17 H, m, H-1'', -4'', -1), 4.05 (4 H, d,  $J = 12.9$  Hz, H-6'<sub>A</sub>), 3.93-3.69 (12 H, m, H-2, -5, -6, -H-4', -6'<sub>B</sub>), 3.69-3.42 (18 H, m, H-3, -4, -1', -2', -3', -6''<sub>A</sub>), 3.36 (3 H, s, OCH<sub>3</sub>), 3.35-3.18 (8 H, m, H-5', -6''<sub>B</sub>), 3.12 (4 H, t,  $J = 11.6$  Hz, H-1'), 2.56-2.35 (8 H, m, H-5'');  $^{13}\text{C}$  NMR (100 MHz, D<sub>2</sub>O):  $\delta = 143.6$ -143.4 (C-2''), 125.8-125.4 (C-3''), 97.0 (C-1), 80.4 (C-2), 78.9 (C-3), 76.7 (C-4), 75.7 (C-3'), 69.1 (C-5), 68.0 (C-6), 67.0 (C-2'), 66.0 (C-4'), 65.5 (C-5'), 64.9, 64.7, 62.9, 62.8 (C-1''), 55.1 (OCH<sub>3</sub>), 53.8 (C-6'), 53.3 (C-1'), 50.1

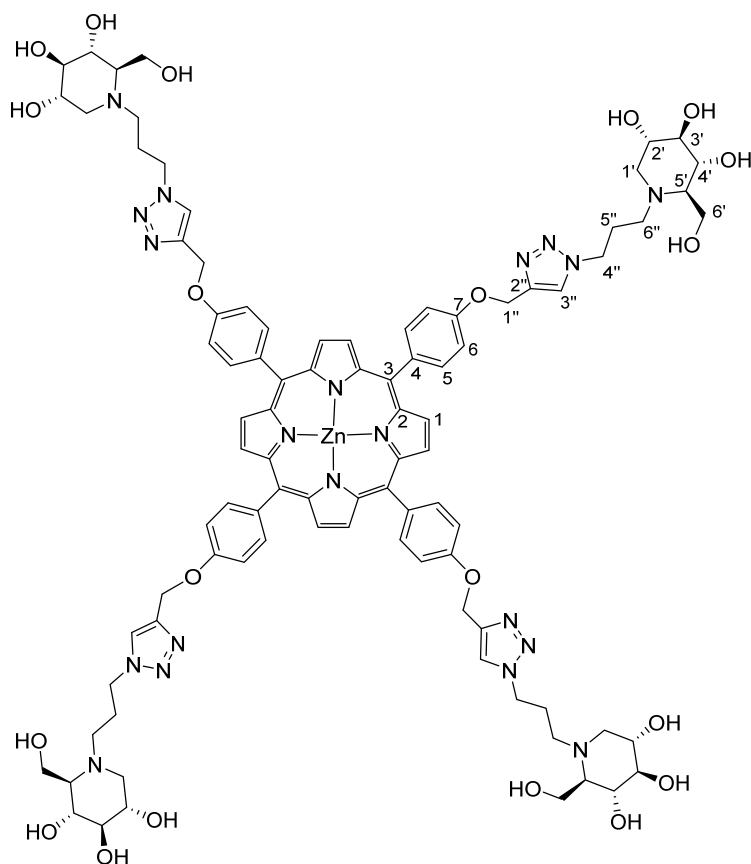
(C-6''), 47.6 (C-4''), 23.3 (C-5''); HRMS (MALDI)  $[M+Na]^+$ : Found 1353.6590  $C_{92}H_{136}N_{16}NaO_{20}$  requires 1353.6621.

### Compound (5)



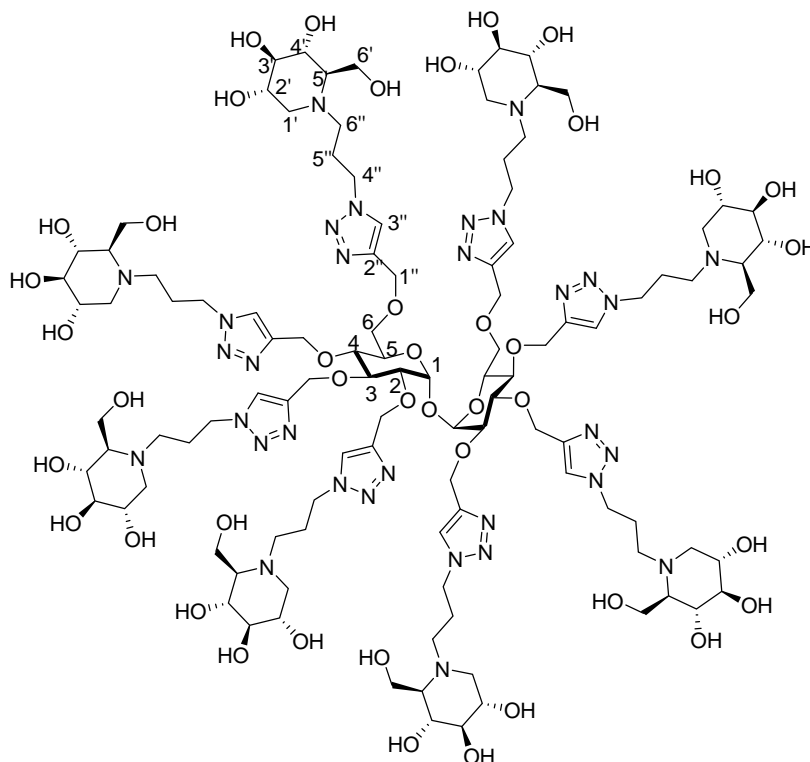
White solid,  $[\alpha]_D = +32$  ( $c = 1$ ,  $H_2O$ );  $^1H$  NMR (400 MHz,  $D_2O$ )  $\delta = 8.15, 8.12$  (4 H, 2 s, H-3''), 4.93-4.66 (9 H, m, H-1'', H-1), 4.64-4.61 (8 H, m, H-4''), 4.23-4.18 (1 H, m, H-3), 4.10-4.04 (1 H, m, H-5), 4.00-3.82 (10 H, m, H-2, -4, -6'), 3.76-3.61 (5 H, m, H-6<sub>A</sub>, -2'), 3.59-3.49 (5 H, m, H-6<sub>B</sub>, -4'), 3.47-3.35 (7-H, m, H-3', OMe), 3.32-3.22 (4 H, m, H-1'<sub>A</sub>), 3.17-3.05 (4 H, m, H-6''<sub>A</sub>), 3.03-2.87 (4 H, m, H-6''<sub>B</sub>), 2.75-2.06 (8 H, m, H-1'<sub>B</sub>, -5'), 2.41-2.24 (8 H, m, H-5'');  $^{13}C$  NMR (100 MHz,  $D_2O$ ):  $\delta = 144.1$  (C-2''), 125.3 (C-3''), 97.7 (C-1), 77.5 (C-3), 77.3 (C-3'), 75.7 (C-2), 74.8 (C-4), 68.9 (C-6, 2'), 68.5 (C-5), 67.7 (C-4'), 65.1 (C-5'), 65.0, 63.5, 63.4, 63.0 (C-1''), 56.1 (OCH<sub>3</sub>), 55.1 (C-6'), 54.5 (C-1'), 49.2 (C-6''), 48.2 (C-4''), 23.8 (C-5''); HRMS (MALDI)  $[M+Na]^+$ : Found 1353.6659  $C_{92}H_{136}N_{16}NaO_{20}$  requires 1353.6621.

## Compound (6)



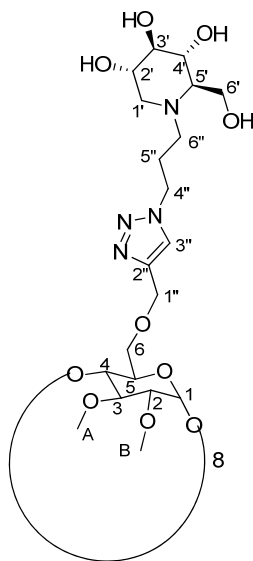
Purple solid,  $[\alpha]_D$  = Too dark for analysis;  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  = 8.73 (8 H, s, H-1), 8.26 (4 H, s, H-3''), 7.95 (8 H, s, H-5), 7.30 (8 H, s, H-6), 5.26 (8 H, s, H-1''), 4.55-4.30 (8 H, m, H-4''), 3.80-3.58 (8 H, m, H-6'), 3.38-3.31 (4 H, m, H-2'), 3.19 (4 H, t,  $J$  = 9.4 Hz, H-4'), 3.04 (4 H, t,  $J$  = 9 Hz, H-3'), 3.00-2.84 (8 H, m, H-1'A, H-6''A), 2.71-2.58 (4 H, m, H-6''B), 2.30-2.18 (8 H, m, H-1'B, -5'), 2.17-2.03 (8 H, m, H-5'');  $^{13}\text{C}$  NMR (100 MHz,  $\text{DMSO}-d_6$ ):  $\delta$  = 158.2, 150.4, 143.7 ( $\text{C}_{\text{IV}}$ ), 136.1 (C-5), 132.3 (C1), 125.7 (C-3''), 120.8 ( $\text{C}_{\text{IV}}$ ), 113.6 (C-6), 78.3 (C-3'), 69.8 (C-4'), 68.5 (C-2'), 66.7 (C-5'), 61.8 (C-1''), 57.4 (C-6'), 55.9 (C-1'), 49.8 (C-6''), 48.5 (C-4''), 25.4 (C-5''); HRMS (ES)  $[\text{M}+2\text{H}]^{2+}$ : Found 939.3743  $\text{C}_{92}\text{H}_{110}\text{N}_{20}\text{O}_{20}\text{Zn}$  requires 939.3767.

## Compound (7)



White solid,  $[\alpha]_D = +15$  ( $c = 0.5$ ,  $H_2O$ );  $^1H$  NMR (400 MHz,  $D_2O$ )  $\delta = 8.10-8.04$  (8 H, s, H-3''), 5.26 (2 H, d,  $J = 3.3$  Hz, H-1), 4.99-4.58 (16 H, m, H-1''), 4.57-4.35 (16 H, m, H-4''), 4.07-3.89 (4 H, m, H-3, -5), 3.88-3.74 (16 H, m, H-6'), 3.73-3.52 (16 H, m, H-2, -4, -6, -2'), 3.46-3.36 (8 H, m, H-4'), 3.34-3.25 (8 H, m, H-3'), 3.07-2.95 (8 H, m, H-1'A), 2.90-2.57 (16 H, m, H-6''), 2.38-2.21 (16 H, m, H-1'B, -5'), 2.21-2.04 (16 H, m, H-5'');  $^{13}C$  NMR (100 MHz,  $D_2O$ ):  $\delta = 144.3-143.7$  (C-2''), 125.2-124.8 (C-3''), 92.4 (C-1), 80.4 (C-3), 78.5 (C-2), 78.3 (C-3'), 76.7 (C-4), 70.2 (C-5), 70.1 (C-4'), 68.9 (C-2'), 67.8 (C-6), 65.5 (C-1'), 64.9 (C-5'), 63.6, 63.3 (C-1''), 57.6 (C-6'), 55.4 (C-1'), 48.6 (C-4'', 6''), 24.0 (C-5''); HRMS (ES)  $[M+2Na]^{2+}$ : Found 1330.6411  $C_{108}H_{182}N_{32}Na_2O_{43}$  requires 1330.6445.

## Compound (8)

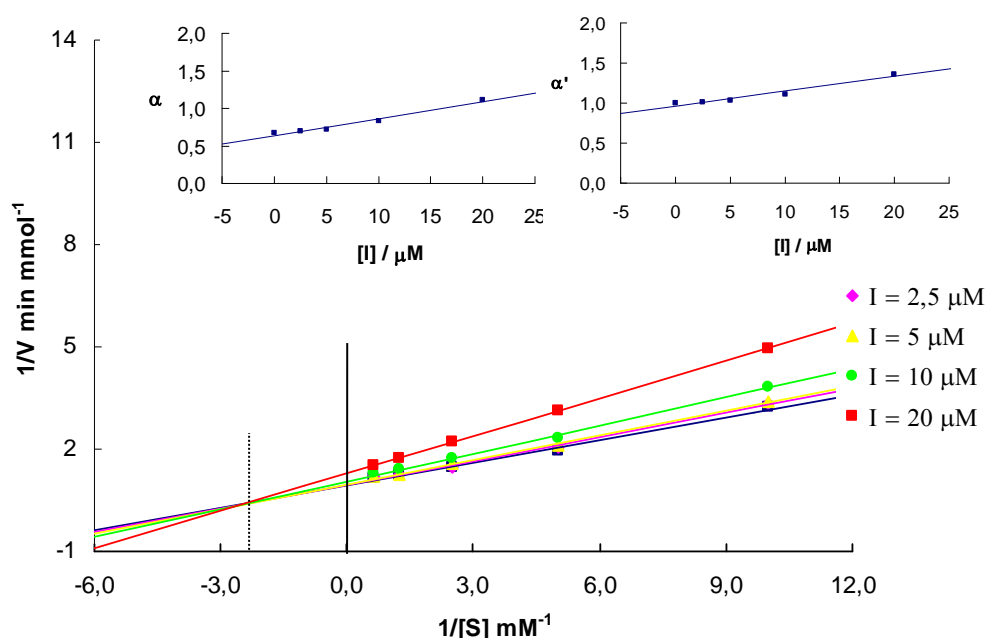


White solid,  $[\alpha]_D = +37$  ( $c = 0.5$ ,  $H_2O$ );  $^1H$  NMR (400 MHz,  $D_2O$ )  $\delta = 8.11$  (8 H, s, H-3''), 5.29 (8 H, s, H-1), 4.75-4.54 (16 H, m, H-1''), 4.54-4.38 (16 H, m, H-4''), 4.09-3.51 (112 H, m, H-3, -4, -5, -6, -2', -6', -7'A, -7'B), 3.48-3.37 (8 H, m, H-4'), 3.37-3.25 (16 H, m, H-2, -3'), 3.02 (8 H, dd,  $J = 4.75$  Hz,  $J = 11$  Hz, H-1'A), 2.94-2.78 (8 H, m, H-6''A), 2.77-2.64 (8 H, m, H-6''B), 2.42-2.26 (m, 16 H, H-1'B, 5'), 2.22-2.07 (16 H, m, H-5'');  $^{13}C$  NMR (100 MHz,  $D_2O$ ):  $\delta = 144.0$  (C-2''), 125.0 (C-3''), 96.6 (C-1), 81.4-79.8 (C-3,2), 78.4 (C-3'), 75.8 (C-4), 70.0 (C-5,4'), 68.8 (C-6,2'), 65.0 (C-5'), 63.6 (C-1'), 59.8-58.8 (C-A), 58.4 (C-B), 57.7 (C-6'), 55.5 (C-1'), 48.8 (C-6''), 48.6 (C-4''), 24.2 (C-5'').

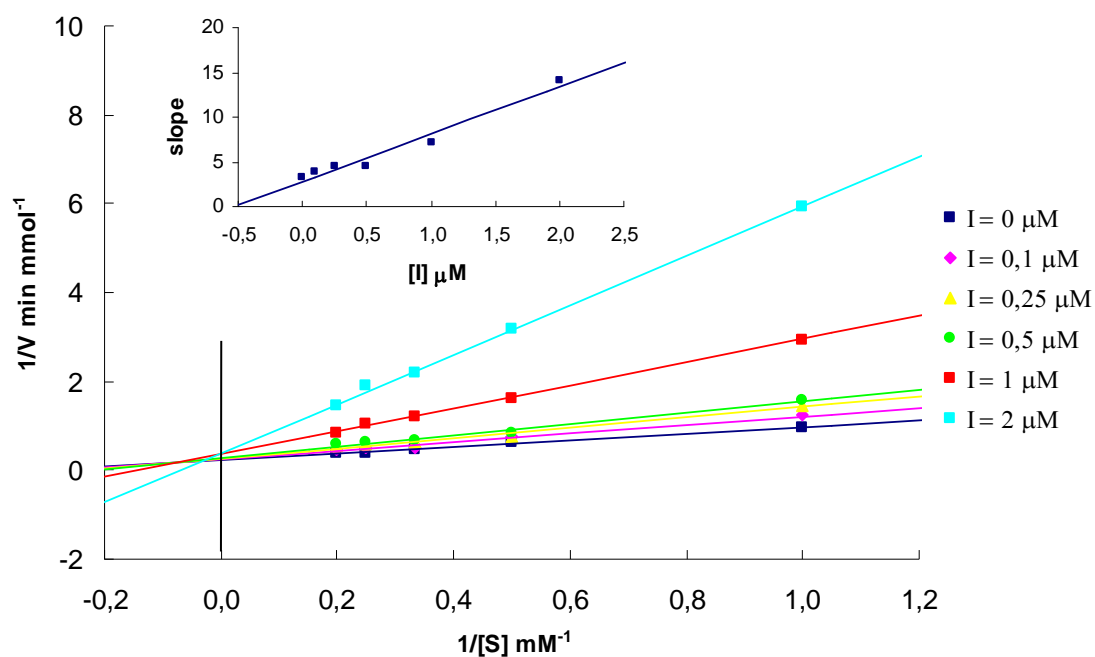
## General procedure for the inhibition assays

Inhibition constant ( $K_i$ ) values were determined by spectrophotometrically measuring the residual hydrolytic activities of the glycosidases against the respective *o*- (for  $\beta$ -gluco/ $\beta$ -galactosidase from bovine liver) or *p*-nitrophenyl  $\alpha$ - or  $\beta$ -D-glycopyranoside (for other glycosidases) or  $\alpha,\alpha'$ -trehalose (for trehalase). Each assay was performed in phosphate buffer or phosphate-citrate buffer (for  $\alpha$ - or  $\beta$ -mannosidase and amyloglucosidase) at the optimal pH for the enzymes. The reactions were initiated by addition of enzyme to a solution of the substrate in the absence or presence of various concentrations of inhibitor. The mixture was incubated for 10-30 min at 37 °C or 55 °C (for amyloglucosidase) and the reaction was quenched by addition of 1 M Na<sub>2</sub>CO<sub>3</sub>. Reaction times were appropriate to obtain 10-20% conversion of the substrate in order to achieve linear rates. The absorbance of the resulting mixture was determined at 405 nm. Approximate values of  $K_i$  were determined using a fixed concentration of substrate (around the  $K_M$  value for the different glycosidases) and various concentrations of inhibitor. Full  $K_i$  determinations and enzyme inhibition mode were determined from the slope of Lineweaver-Burk plots and double reciprocal analysis.

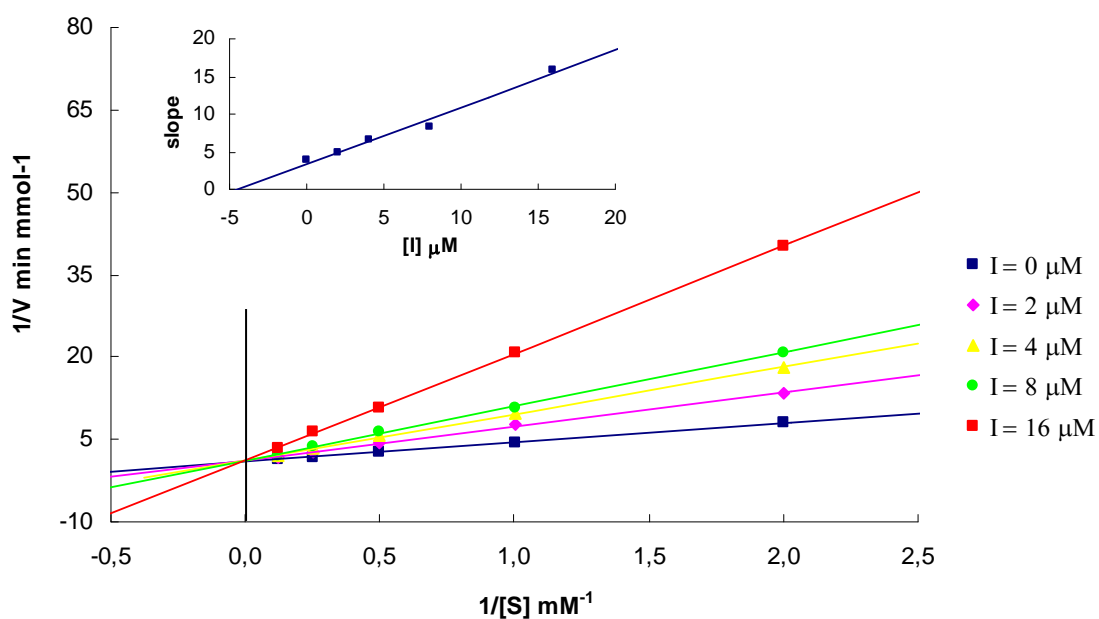
## Representative Lineweaver-Burk Plot



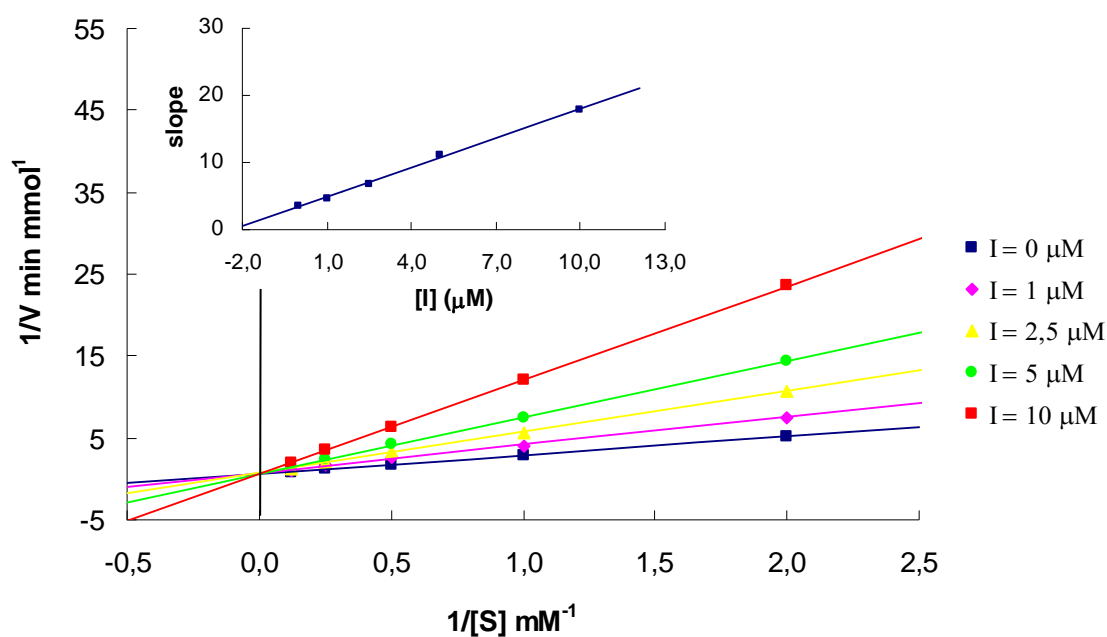
**Figure S1.** Lineweaver-Burk Plot for  $K_i$  determination (45  $\mu\text{M}$ ) and  $K_i'$  determination (54  $\mu\text{M}$ ) of **2** against baker yeast  $\alpha$ -glucosidase (pH 6.8).



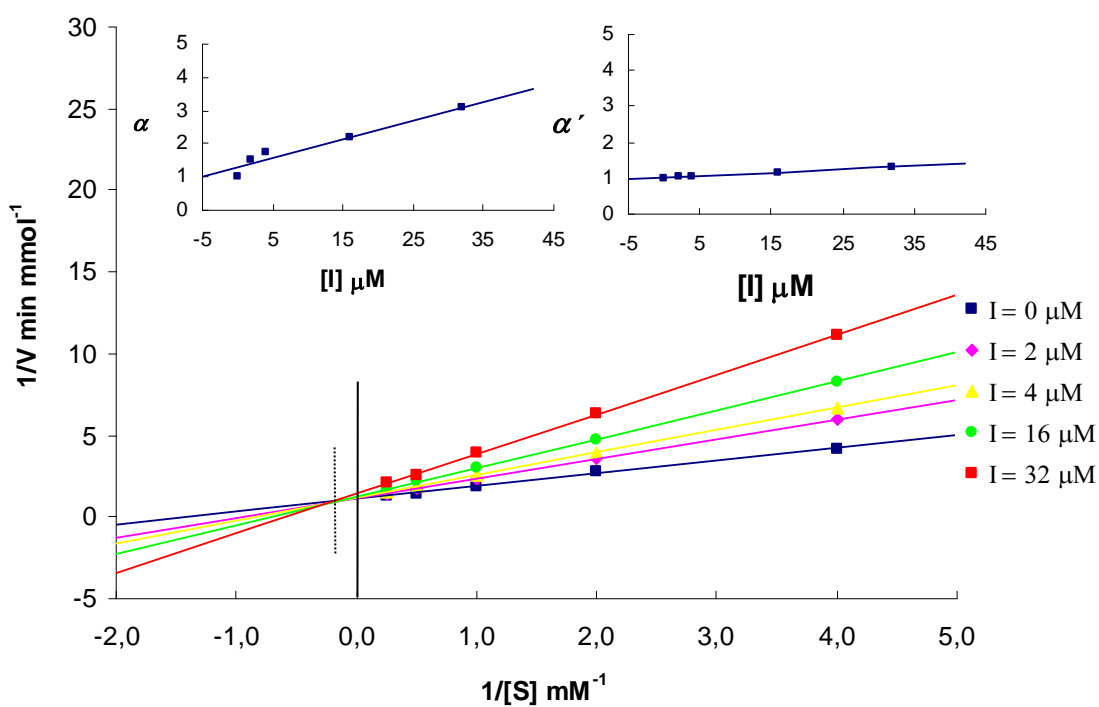
**Figure S2.** Lineweaver-Burk Plot for  $K_i$  determination (0.5  $\mu\text{M}$ ) of **6** against *Jack bean*  $\alpha$ -mannosidase (pH 5.5).



**Figure S3.** Lineweaver-Burk Plot for  $K_i$  determination (4.6  $\mu\text{M}$ ) of **7** against *Aspergillus niger* amyloglucosidase (pH 5.5).

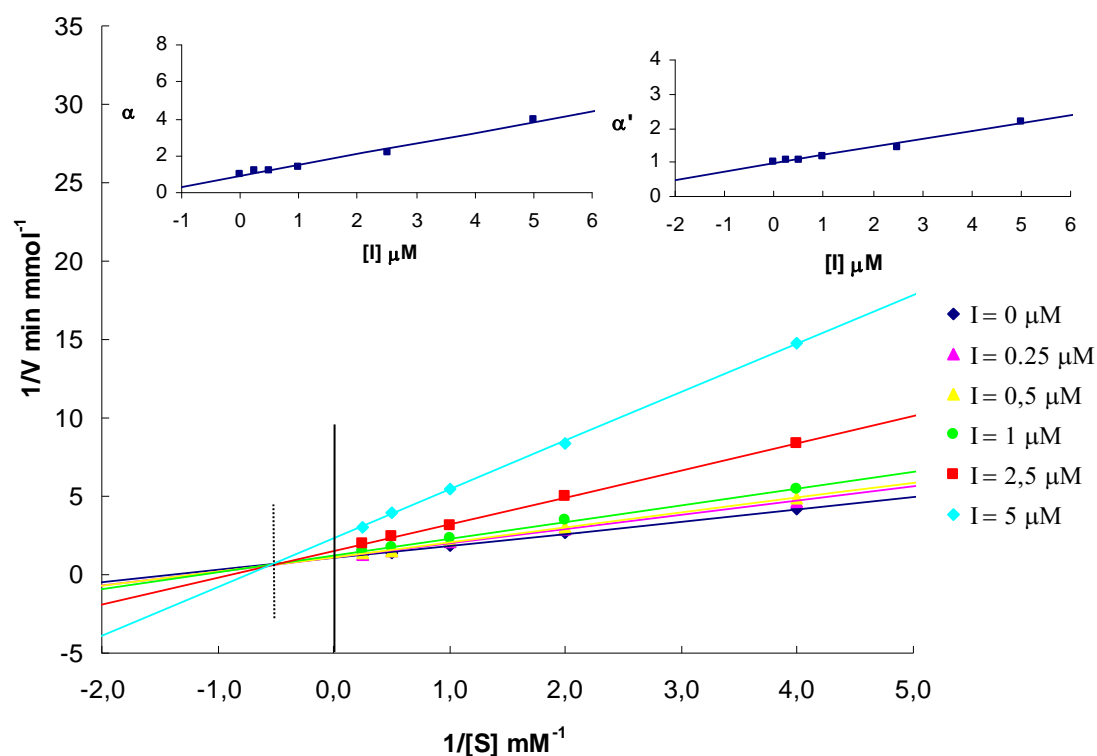


**Figure S4.** Lineweaver-Burk Plot for  $K_i$  determination (2.3  $\mu\text{M}$ ) of **5** against *Aspergillus niger* amyloglucosidase (pH 5.5).



**Figure S5.** Lineweaver-Burk Plot for  $K_i$  determination (18  $\mu\text{M}$ ) and  $K_i'$  determination (108  $\mu\text{M}$ ) of **6** against green coffee  $\alpha$ -galactosidase (pH 6.8).





**Figure S6.** Lineweaver-Burk Plot for  $K_i$  determination ( $1.7 \mu\text{M}$ ) and  $K_i'$  determination ( $4.3 \mu\text{M}$ ) of **7** against green coffee  $\alpha$ -galactosidase (pH 6.8).

### GM and LM inhibition

GM and LM inhibition were measured as previously described (I. Nemčovičová, S. Šesták, D. Rendić, M. Plšková, J. Mucha, I. B. H. Wilson, *Glycoconj. J.* **2013**, *in press*). Briefly, mannosidase activities were measured with p-nitrophenyl- $\alpha$ -D-mannopyranoside (pNP-Man; Sigma) as substrate at a concentration of 2 mM (diluted from a 100 mM stock solution in dimethylsulfoxide) in 100 mM of acetate buffer at pH 6.0 or at pH 5.2 for GM and LM, respectively. 0.5–2  $\mu\text{l}$  of enzyme and 10  $\mu\text{l}$  of the inhibitor in 50% DMSO (or the same volume of 50% DMSO in a control reaction) were incubated for 3 h at 37 °C (total volume of 50  $\mu\text{l}$ ). In case of GM, the reaction was supplemented with 0.2 mM  $\text{CoCl}_2$  (final concentration). The reactions were terminated with ten volumes (0.5 ml) of a 100 mM sodium carbonate solution. The formation of p-nitrophenol was measured at 410 nm with a spectrophotometer.

### Sample preparation for AFM imaging

JB $\alpha$ Man was suspended in 20 mM citrate buffer pH 5.5. The multivalent iminosugars were suspended either in citrate or citrate/DMSO (or methanol) mixtures. Mixtures of JB $\alpha$ Man/DNJ 1:1 (mol/mol) in 20 mM citrate buffer pH 5.5 (final concentration of JB $\alpha$ Man and DNJ 4.54 nM) and incubated for 1 h at room temperature. Freshly cleaved mica squares (16 mm<sup>2</sup>) were glued onto steel sample discs (Agar Scientific, England) using Epotek 377 (Polytec, France). A 150  $\mu$ L portion of the JB $\alpha$ Man/DNJ suspensions were then deposited onto the mica samples, and were allowed to adsorb on the solid surface for 20 h at room temperature. Subsequently, samples were rinsed 3 times with ultrapure water and they were dried in a dessicator under vacuum for 1 h. The samples were then left overnight under ambient conditions before AFM imaging.

### AFM imaging

The samples were investigated using a commercial AFM (NanoScope III MultiMode AFM, Veeco Metrology LLC, Santa Barbara, CA) equipped with a 125  $\mu$ m  $\times$  125  $\mu$ m  $\times$  5  $\mu$ m scanner (J-scanner). Topographic images were recorded in air in Tapping<sup>TM</sup> mode using RTESP cantilevers (Veeco Metrology Group, Santa Barbara, CA) with a nominal spring constant of 40 N/m (manufacturer specified), with a minimal applied force (<200 pN), at a scan rate of 2 Hz and a drive amplitude of  $\sim$ 300 kHz. The curvature radius of silicon nitride tips was  $\sim$ 10 nm. Images were obtained at room temperature (21–22°C) in air. All images (512  $\times$  512 pixels) shown in this paper are flattened raw data.

## RESULTS

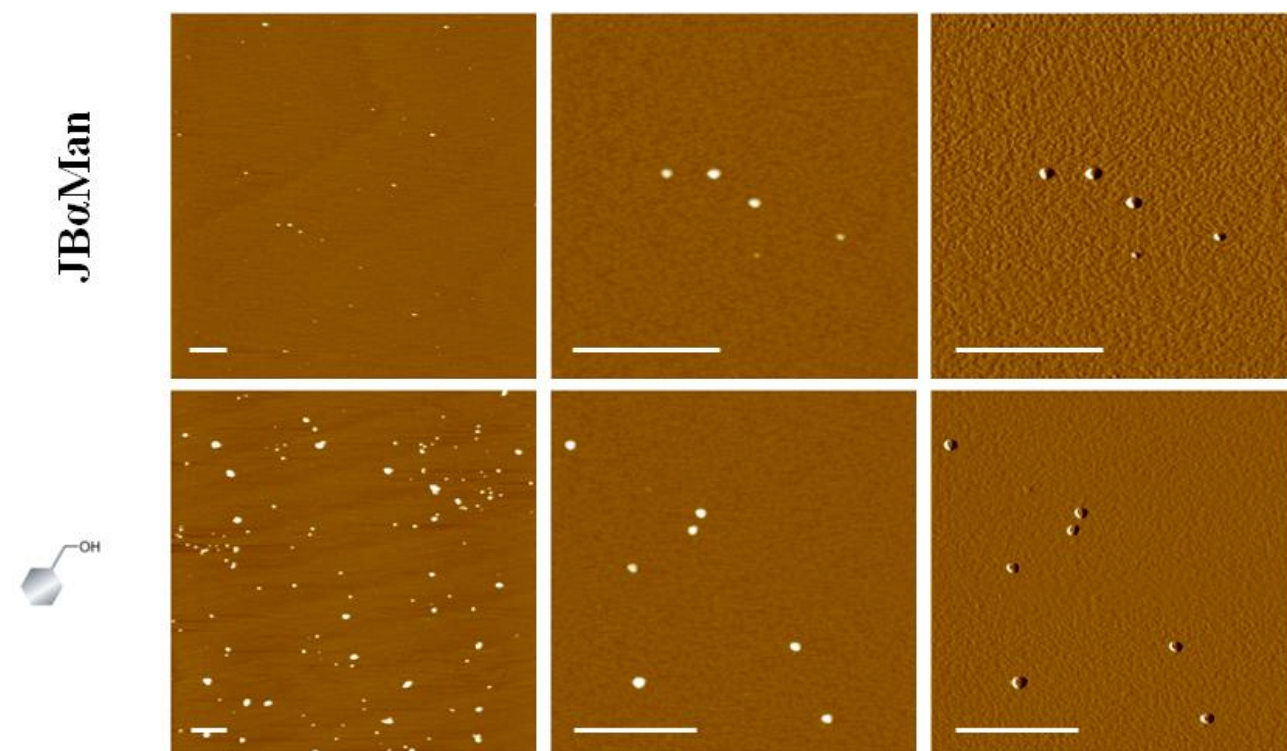


Figure S7: AFM height (left and middle) and deflection (right column) images of JB $\alpha$ Man alone or interacting with monovalent ligand ( $z=10$  nm in height images ; scale bar is 500 nm).



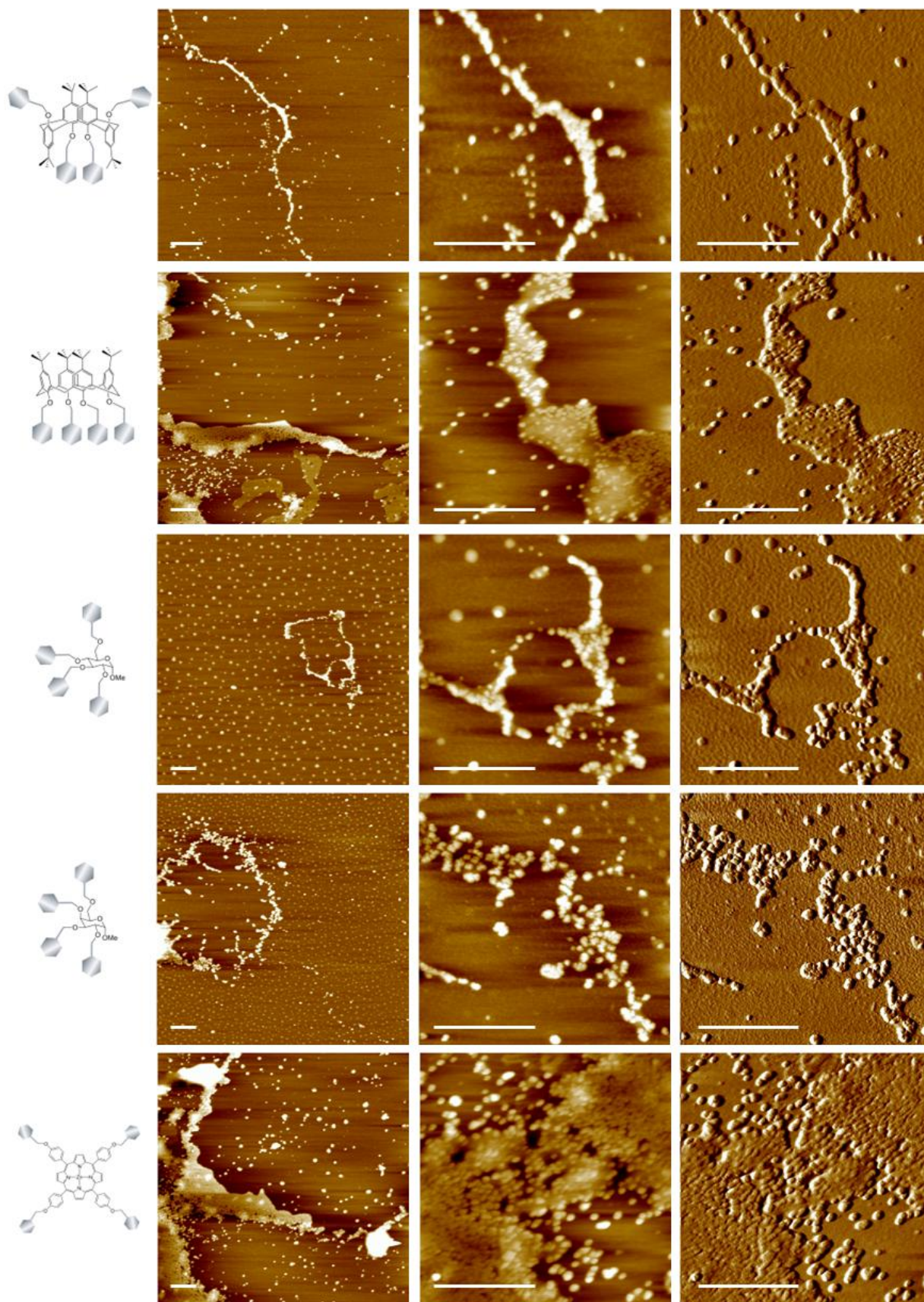


Figure S8: AFM height (left and middle) and deflection (right column) images of JBoMan interacting with tetraivalent ligands ( $z=10$  nm in height images ; scale bar is 500 nm).



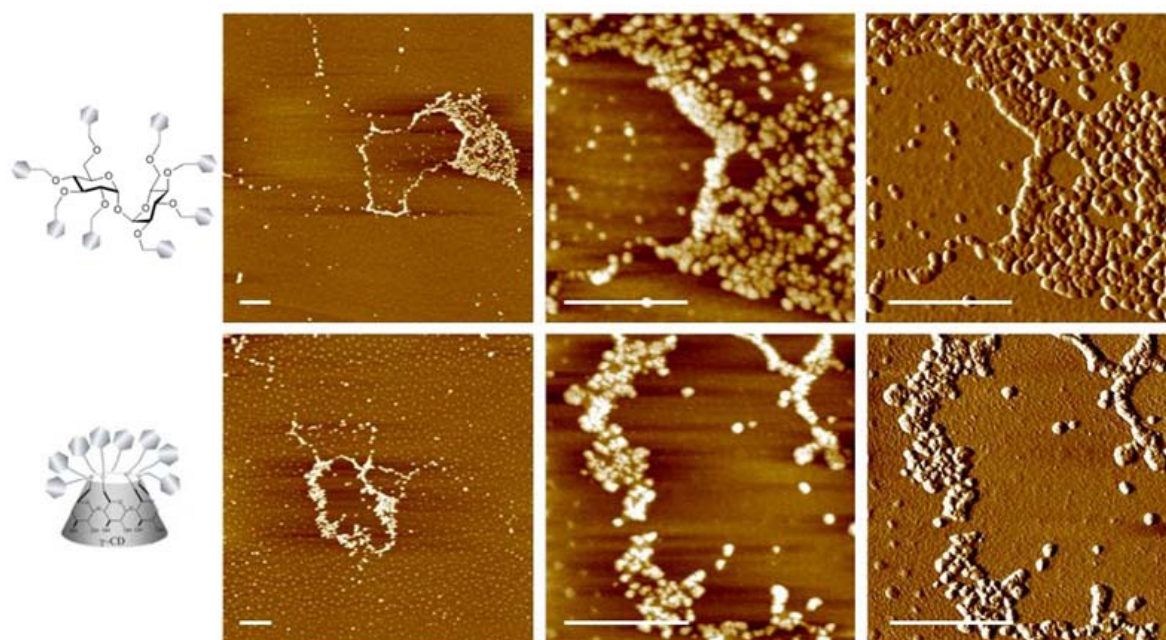


Figure S9: AFM height (left and middle) and deflection (right column) images of JB $\alpha$ Man interacting with mannose octavalent ligands ( $z=10$  nm in height images ; scale bar is 500 nm).

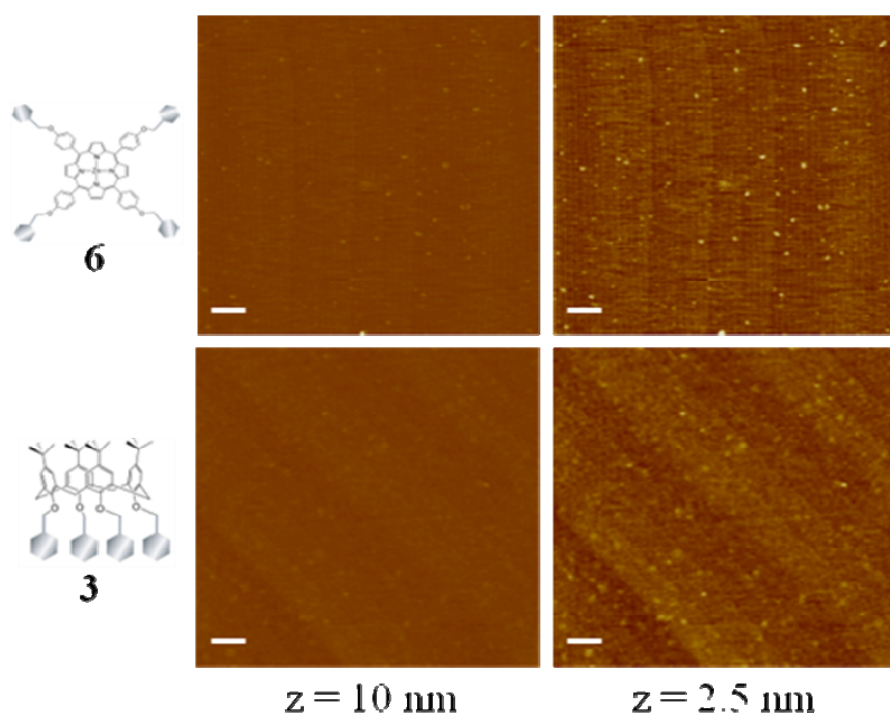


Figure S10: AFM height images tetraivalent ligands **3** and **6** adsorbed on mica without JB $\alpha$ Man (scale bar is 500 nm). Note that left and right column are the same images. Indeed, images of the left column are presented with the same  $z$ -scale as the ones with proteins (see previous figures) and the  $z$ -scale was adjusted at 2.5 nm to reveal the presence of very thin structures (right column).

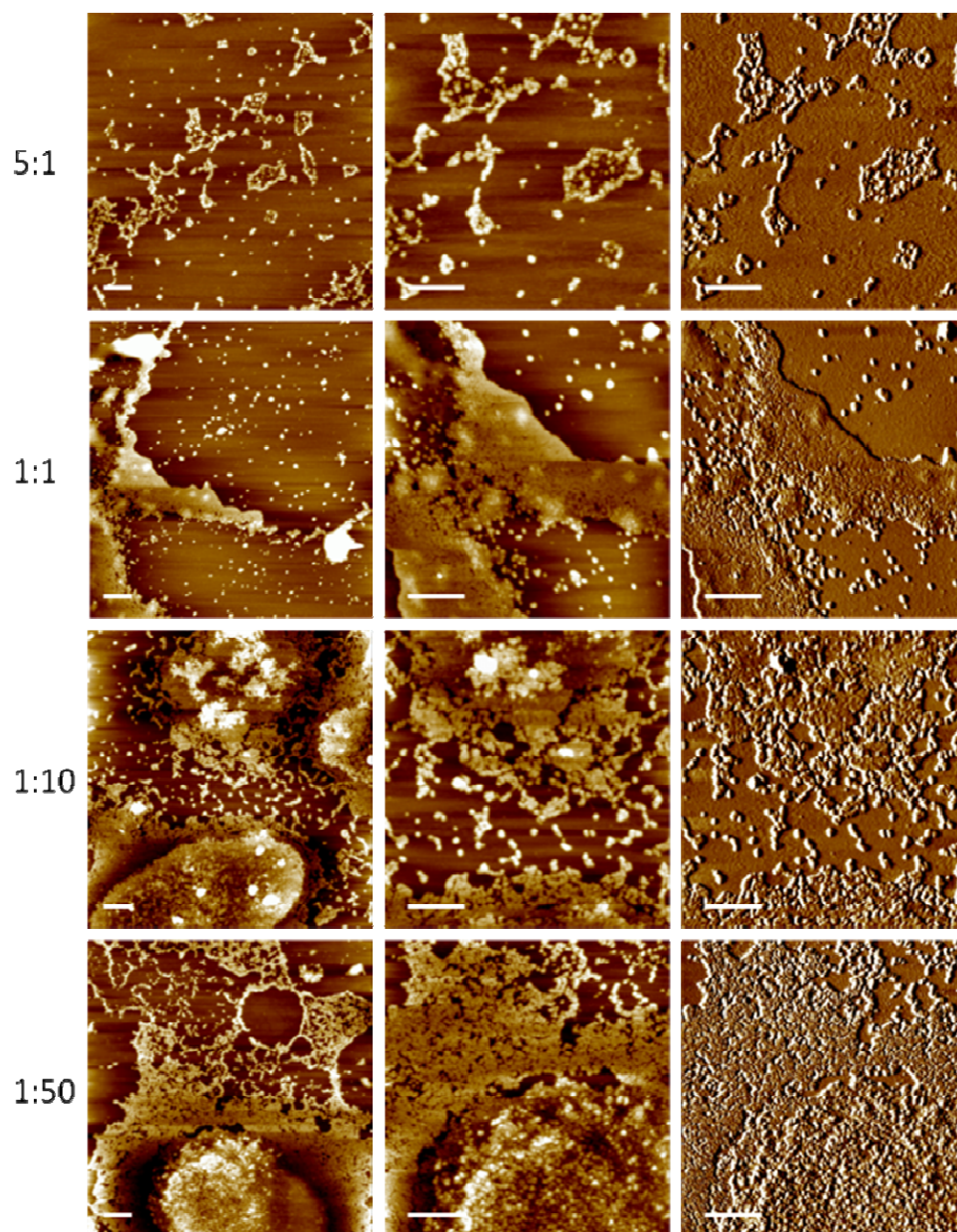


Figure S11: AFM height (left and middle) and deflection (right column) images of JB $\alpha$ Man interacting with tetravalent ligands at different JB $\alpha$ Man/ligand molar ratios ( $z=10$  nm in height images ; scale bar is 500 nm). By increasing the amount of ligand, the surface covered with aggregates increases in AFM images.

## DLS measurements

Dynamic light scattering (DLS) was used to determine the hydrodynamic diameter of protein/ligands complexes in solution (20 mM citrate buffer pH 5.5). Samples were prepared by incubating the protein with the different ligands (1:10, molar ratio) at 4.54  $\mu$ M during one hour at ambient temperature under gentle agitation. The measurements were performed with the Zetasizer (Nano ZS, Malvern Instruments Ltd., UK). The mean hydrodynamic diameter was determined from the autocorrelation function of the intensity of light scattered from the particles. The software used was DTS Nano version 5.03, supplied by the manufacturer (Malvern Instruments Ltd.).

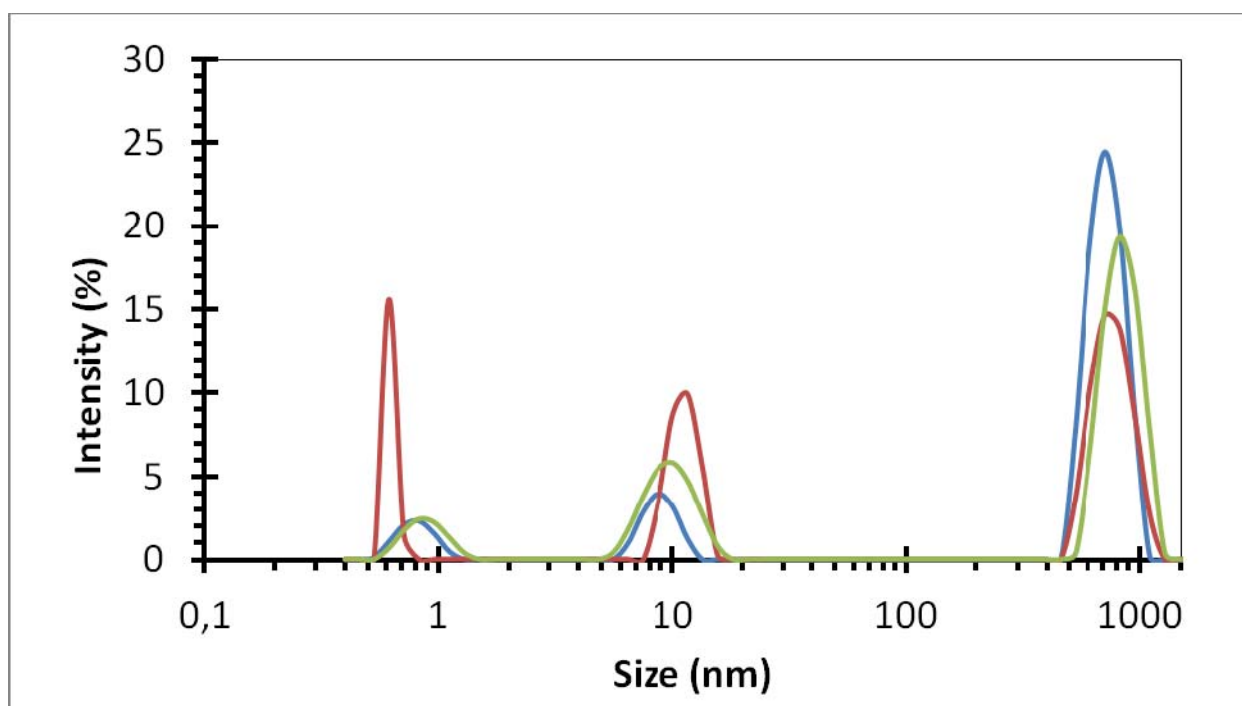


Figure S12: Typical size distribution graph obtained by DLS on mixtures of JB $\alpha$ Man/**6** in 1:10 molar ratio. These results show the coexistence of large complexes ( $731.9 \pm 161.9$ ) with the free protein ( $10.4 \pm 2.2$ ) and the free ligand **6** (less than 1 nm).

