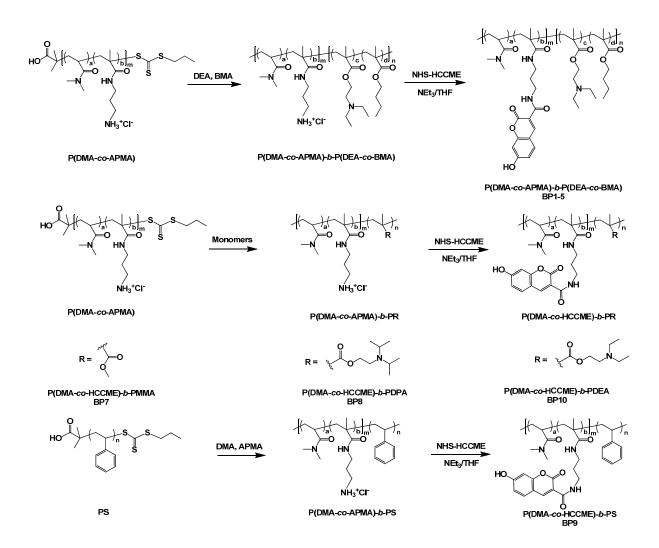
Supporting Information

Spatiotemporal Monitoring Endocytic and Cytosolic pH Gradients with Endosomal Escaping pH-Responsive Micellar Nanocarriers

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Scheme S1. Synthetic routes employed for the preparation of HCCME-labeled diblock copolymers, P(DMA-*co*-HCCME)-*b*-P(DEA-*co*-BMA) with varying BMA molar fractions (**BP1-BP5**), P(DMA-*co*-HCCME)-*b*-PMMA (**BP7**), P(DMA-*co*-HCCME)-*b*-PDPA (**BP8**), P(DMA-*co*-HCCME)-*b*-PS (**BP9**), and P(DMA-*co*-HCCME)-*b*-PDEA (**BP10**) by sequential RAFT polymerizations.

Entry	$< D_h > (nm)^a$	PDI ^a	pK _a ^b	$\xi (mV)^{c}$
BP1	34.8	0.054	6.73	-2.7
BP2	30.4	0.054	6.51	-3.7
BP3	41.6	0.099	6.12	-4.5
BP4	37.6	0.10	5.92	-6.2
BP5	35.6	0.20	5.57	-5.6
BP6	24.2	0.11	6.12	-5.8
BP7	108.0	0.21	/	-24.0
BP8	32.8	0.13	6.30	-6.7
BP9	31.2	0.23	/	-21.4
BP10	35.4	0.044	7.07	5.8

Table S1. Intensity-Average Hydrodynamic Diameter, $\langle D_h \rangle$, Polydispersity Index, p K_a , and Zeta Potentials (ξ) of Micelles Self-Assembled from Diblock Copolymers.

^{*a*} Determined by DLS at a copolymer concentration of 1.0 g/L and pH 7.4; ^{*b*} Determined by pH titration experiments; ^{*c*} Zeta potentials were determined by Malvern Zetasizer Nano ZS at copolymer concentrations of 0.5 g/L in PBS buffer (pH 7.4). Note: all data were averaged over three measurements.

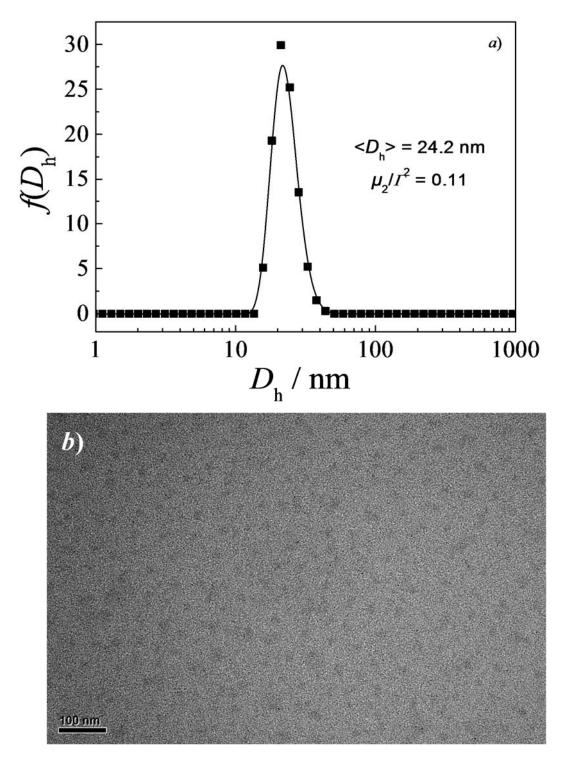


Figure S1. Typical (a) hydrodynamic diameter distribution, $f(D_h)$, and (b) TEM image recorded for **BP6** micelles (0.2 g/L, pH 7.4).

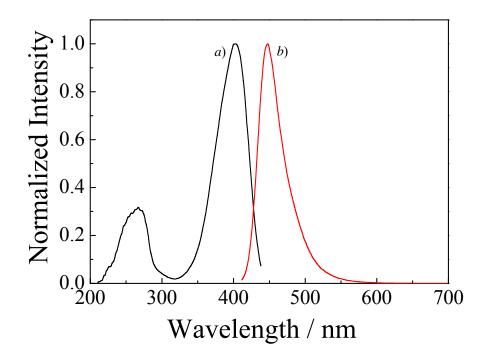


Figure S2. Normalized fluorescence (a) excitation and (b) emission spectra recorded for the aqueous solution of **P1** (pH 9.0; 0.018 g/L, [HCCME] = 1.0×10^{-6} M; slit widths: Ex. 2.5 nm, Em. 2.5 nm).

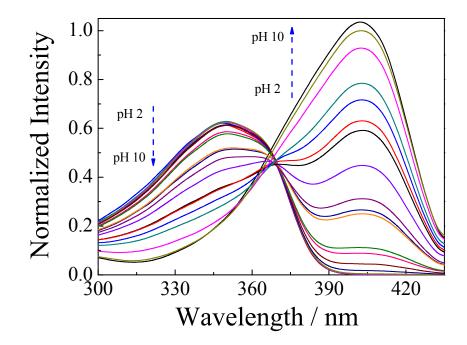


Figure S3. Normalized fluorescence excitation spectra ($\lambda_{em} = 447$ nm, slit widths: Ex. 2.5 nm, Em. 2.5 nm) recorded for the aqueous solution of **P1** (0.018 g/L, [HCCME] = 1.0×10^{-6} M) in the pH range of 2-10.

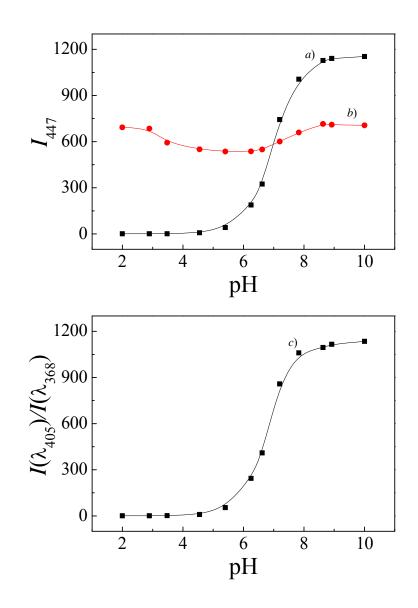


Figure S4. Relative fluorescence emission intensity changes excited at (a) 405 nm ($\lambda_{em} = 447$ nm) and (b) 368 nm ($\lambda_{em} = 447$ nm) recorded for the aqueous solution of **P1** (0.018 g/L, [HCCME] = 1.0×10^{-6} M) in the pH range of 2-10. (c) Normalized fluorescence intensity ratio changes, $I(\lambda_{405})/I(\lambda_{368})$, recorded for the aqueous solution of **P1** in the pH range of 2-10. $I(\lambda_{405})$ and $I(\lambda_{368})$ refer to the fluorescence intensities at 447 nm when excited at 405 nm and 368 nm, respectively.

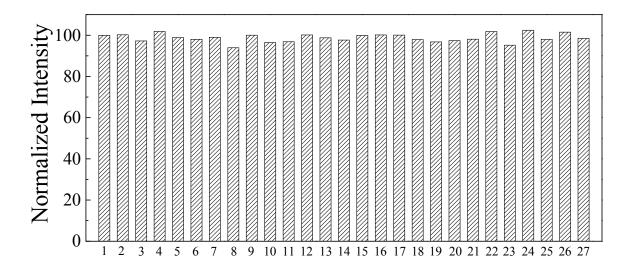


Figure S5. Fluorescent emission intensities recorded at 447 nm for aqueous solutions of **P1** (0.018 g/L, [HCCME] = 1.0×10^{-6} M) in the presence of 1 mM (2) Ag⁺, (3) Al³⁺, (4) Ca²⁺, (5) Cu²⁺, (6) Mg²⁺, (7) Pb²⁺, (8) Zn²⁺, (9) H₂O₂, (10) *L*-cysteine, (11) *L*-phenylalanine, (12) *L*-alanine, (13) glycine, (14) *L*-lysine, (15) *L*-tyrosine, (16) *L*-tryptophan, (17) *L*-aspartic acid, (18) *L*-histidine, (19) glucose, and (20) mannose, respectively, or in the presence of 0.01 g/L (21) lipase, (22) glucose oxidase, (23) acid phosphatase, (24) amylase, (25) alkaline phosphase, (26) lysozyme, and (27) trypsin. The mixture was incubated at 37 °C for 1 h prior to fluorescence measurement. Column 1 was taken as the control.

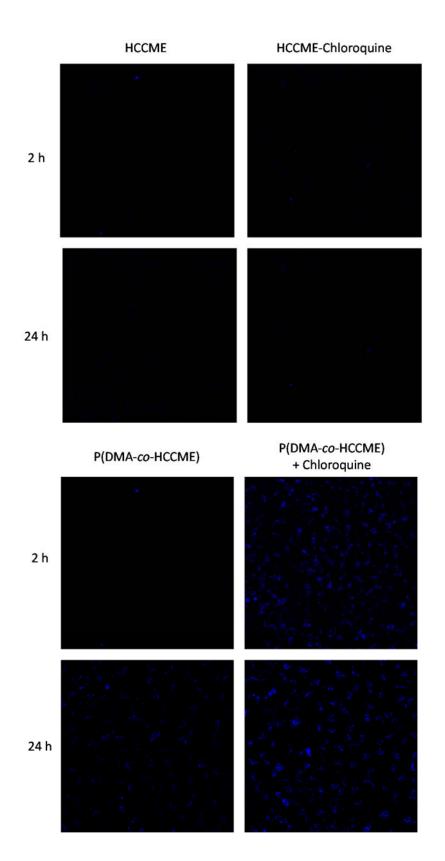


Figure S6. CLSM images recorded for live HepG2 cells after incubating with (top) HCCME and (bottom) **P1** (0.018 g/L, [HCCME] = 1.0×10^{-6} M) in the (left) absence and (right) presence of chloroquine (100 μ M) at 37 °C for 2 h and 24 h, respectively.

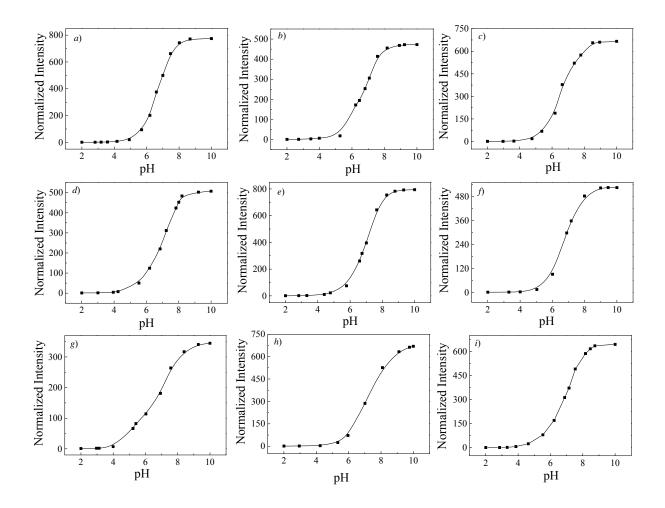


Figure S7. Normalized fluorescence emission intensity changes ($\lambda_{ex} = 405 \text{ nm}$, $\lambda_{em} = 447 \text{ nm}$) recorded for aqueous solutions of (a) **BP1**, (b) **BP2**, (c) **BP3**, (d) **BP4**, (e) **BP5**, (f) **BP7**, (g) **BP8**, (h) **BP9**, and (i) **BP10** in the pH range of 2-10, respectively. In all cases, the concentration of HCCME moieties was fixed at 1.0×10^{-6} M.

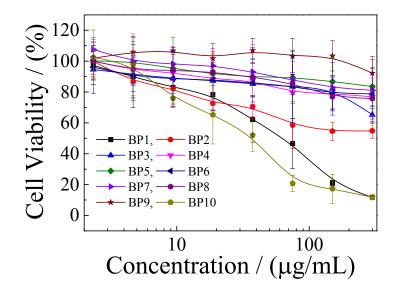


Figure S8. *In vitro* cytotoxicity of **BP1-10** micelles determined by MTT assay against HepG2 cells. The data represent a single experiment performed in quadruple with the error bars denoting the standard deviation.

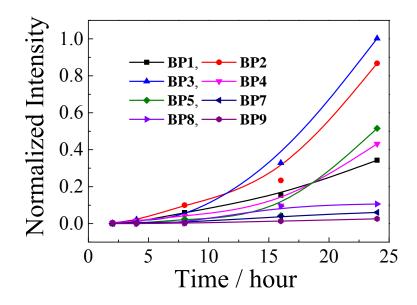


Figure S9. Time-dependent relative changes in intracellular fluorescence intensities recorded for BP1-5 and BP7-9 micellar dispersions, respectively. In all cases, the concentration of HCCME moieties was fixed at 1.0×10^{-6} M.

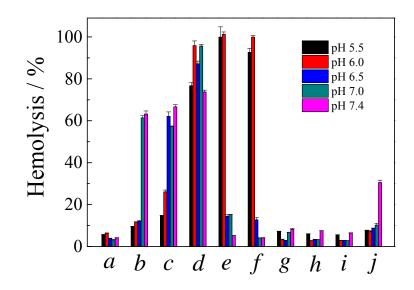


Figure S10. Hemolysis dose-response curves recorded for aqueous solutions ($20 \mu g/mL$) of (a) **P1,** (b) **BP1** (c) **BP2,** (d) **BP3,** (e) **BP4,** (f) **BP5,** (g) **BP7,** (h) **BP8,** (i) **BP9,** and (j) **BP10** at varying pH conditions. PBS (10 mM phosphate, 150 mM NaCl, pH 7.4) and Triton X-100 (1% v/v) were used as negative and positive controls, respectively. The data represent a single experiment performed in triplicate with the error bars denoting the standard deviation.

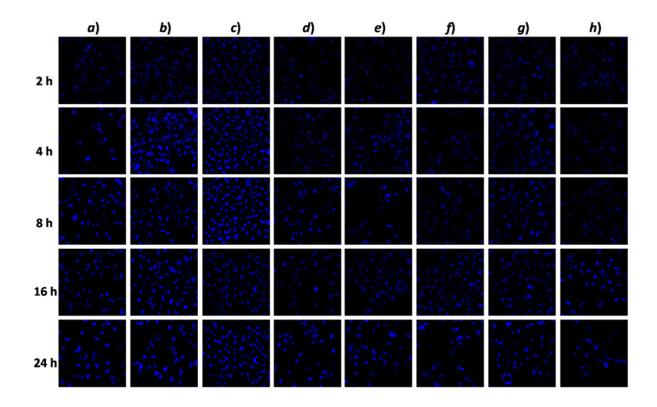


Figure S11. CLSM images recorded for live HepG2 cells incubated at 37 °C for varying times with (a) **BP1**, (b) **BP2**, (c) **BP3**, (d) **BP4**, (e) **BP5**, (f) **BP7**, (g) **BP8**, and (h) **BP9**, respectively. The cells were further treated with chloroquine (100 μ M) and incubated at 37 °C for 1 h before recording fluorescence images. In all cases, the concentration of HCCME moieties was fixed at 1.0 × 10⁻⁶ M.

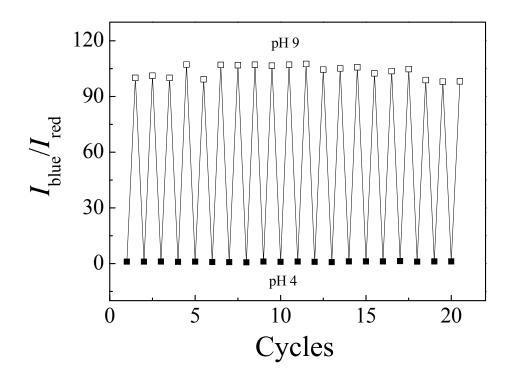


Figure S12. Emission intensity ratio changes (I_{blue}/I_{red}) recorded for the aqueous solution (0.049 g/L) of **BP6** ($\lambda_{ex} = 405$ nm for HCCME and $\lambda_{ex} = 543$ nm for TMR; slit widths: Ex. 2.5 nm, Em. 2.5 nm; [HCCME] = 1.0×10^{-6} M, [TMR] = 1.6×10^{-6} M) when the solution pH was cycled between 4 and 9.

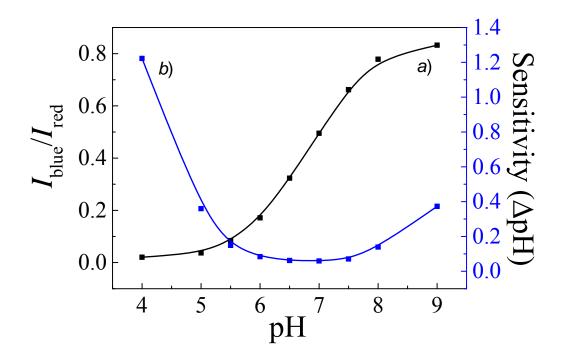


Figure S13. (a) Changes in fluorescence intensity ratios (I_{blue}/I_{red}) and (b) detection sensitivity for **BP6** aqueous solution (0.049 g/L, [HCCME] = 1.0×10^{-6} M, [TMR] = 1.6×10^{-6} M) with excitation at 405 nm and collected between 420-470 nm for the blue channel and excitation at 543 nm and collected between 555-595 nm for the red channel.