## **Supporting Information to**

# Air-Stable Nickel Precatalysts for Fast and Quantitative Cross-Coupling of Aryl Sulfamates with Aryl Neopentylglycolboronates at Room Temperature

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#### 1. Materials

Tricyclohexylphosphine, NiCl<sub>2</sub>6H<sub>2</sub>O, 1,5-cyclooctadiene, sodium, pyridine, 4-iodobenzoic acid, borane-dimethylsulfide complex, 2-methoxyphenol, anthracene, potassium phosphate tribasic trihydrate, potassium phosphate anhydrous, and Mg turnings were used as received from commercial sources. THF, and toluene were distilled from sodium and benzophenone. Hexanes were distilled from sodium. Triethylamine was distilled from calcium hydride. Methanol was distilled from Mg turnings. Triphenyl phosphine was recrystallized from hexanes. 9-Chloroanthracene, 9-chlorophenanthrene, 5-chloro-dihydroacenaphthene, methyl 4-(5,5dimethyl-1,3,2-dioxaborinan-2-yl)benzoate,<sup>3</sup> 1-naphthyl 4-methylbenzenesulfonate, 2-naphthyl 4methylbenzenesulfonate, 1-naphthyl methanesulfonate, 1-naphthyl methanesulfonate and 2methoxyphenyl dimethylsulfamate were synthesized according to literature procedures.<sup>4</sup> Ni<sup>II</sup>Cl(1-Naphthyl)(PPh<sub>3</sub>)<sub>2</sub>, and Ni<sup>II</sup>Br(1-Naphthyl)(PPh<sub>3</sub>)<sub>2</sub> were synthesized according to a literature procedure. 5 C<sub>6</sub>D<sub>6</sub> was distilled from sodium and benzophenone and kept inside a nitrogen filled glove box. Chlorobenzene was dried over P<sub>2</sub>O<sub>5</sub> for 4 h and then distilled from molecular sieves (0.3 nm) under N<sub>2</sub>. Phenanthrene and acenaphthene were recrystallized from ethanol. Solvents for column chromatography were used as received from commercial sources. 1-Naphthol and 2naphthol were sublimed prior use in the synthesis of their mesylates and tosylates.

#### 2. Instrumentation

<sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were recorded at 500 or 360 MHz on a Bruker DRX (500 MHz) or Bruker DMX (360 MHz). All NMR spectra were measured at 25°C in the indicated deuterated solvents. Proton and carbon chemical shifts (δ) are reported in ppm and coupling constants (J) are reported in Hertz (Hz). The resonance multiplicities in the <sup>1</sup>H NMR spectra are described as "s" (singlet), "d" (doublet), "t" (triplet), "quint" (quintuplet) and "m" (multiplet) and broad resonances are indicated by "br". Residual protic solvent of CDCl<sub>3</sub> (<sup>1</sup>H, 7.27 ppm; <sup>13</sup>C, 77.16 ppm (central resonance of the triplet)), and tetramethylsilane (TMS) were used as the internal reference. <sup>31</sup>P NMR (203 MHz) spectra were recorded using H<sub>3</sub>PO<sub>4</sub> in D<sub>2</sub>O as external standard. A GC coupled with an FID detector and column HP 19091J-413 (5%-phenyl)-methylpolysiloxane 30m length 0.32mm internal diameter was used to follow the reaction conversions and to assess purity of final compounds complementary to the NMR technique. The crude reaction mixtures were diluted with distilled THF. Evolution of the reaction, when indicated, was monitored by analytical thin-layer chromatography using silica gel 60 F254 precoated plates (E. Merck). Compounds were visualized by 254 nm light. Purifications by flash column chromatography were performed using flash silica gel from Silicycle (60 Å, 40-63 μm)

with the indicated eluent. The purity of the products was determined by a combination of thin-layer chromatography (TLC) on silica gel coated aluminum plates (with F253 indicator; layer thickness, 200 μm; particle size, 2-25 μm; pore size, 60 Å). Detection was done by UV absorbance at 254 nm. Accurate mass measurements (HRMS) were performed on a high-resolution double focusing chemical ionization mass spectrometer (Mass Spectrometry Facility, University of Pennsylvania). Dechlorinated [M-Cl]<sup>+</sup>, protonated molecular ions [M+nH]<sup>n+</sup> or sodium adducts [M+Na]<sup>+</sup> were used for empirical formula confirmation. LC/MS were performed with a Waters LCT Premier XE LC/MS system and a Waters GC-TOF Premier. The LCT Premier was equipped with a high-resolution orthogonal time-of-flight (oa-TOF) analyzer using ESI<sup>+</sup> ionization condition. Dechlorinated [M-Cl]<sup>+</sup>, [M-OTs]<sup>+</sup>, protonated molecular ions [M+nH]<sup>n+</sup> or sodium adducts [M+Na]<sup>+</sup> were used for empirical formula confirmation. Elemental analyses were performed at M-H-W Laboratories in Phoenix, AZ.

#### 3. List of Abbreviations

COD 1, 5-cyclooctadiene

dppe 1, 2-bis(diphenylphosphino)ethane

dppf 1, 1'-bis(diphenylphosphino)ferrocene

dppp 1, 3-dis(diphenylphosphino)propane

PCy<sub>3</sub> tricyclohexylphosphine

PPh<sub>3</sub> triphenylphosphine

THF tetrahydrofuran

Tol toluene

#### 4. Experiments

## 4.1 Syntheses of Coupling Reagents

**Synthesis of 2-Methoxyphenyl dimethylsulfamate (1).** 2-Methoxyphenyl dimethylsulfamate was synthesized according to a literature procedure.<sup>6</sup> A flame-dried round bottom flask was charged with a stirring bar and sodium hydride (0.864 g, 36 mmol, 1.2 equiv). The flask was degassed by evacuation for 5 min, backfilled with nitrogen for 1 min for three cycles. 2-Methoxyphenol (3.724 g, 30 mmol, 1 equiv) in dry THF (20 mL) was added dropwise *via* 

syringe. The flask was cooled in an ice bath for 10 min. *N*,*N*-dimethyl sulfamoyl chloride (3.54 mL, 33 mol, 1.1 equiv) in dry THF (20 mL) was added dropwise to the solution. The solution was warmed to room temperature and reacted for 1 h. The crude was quenched by addition of deionized water (20 mL). Ethyl acetate (30 mL) was added to extract the organic phase. The aqueous phase was back extracted with DCM (20 mL) twice. The organic phases were combined, dried over MgSO<sub>4</sub> and then filtered. The filtrate was concentrated on a rotary evaporator and purified by silica gel column chromatography with 40% ethyl ether in hexanes as eluent. The off-white solid was recrystallized from methanol to give a white solid. Yield (5.49 g, 23.73 mmol, 79%), m. p. 47-48.5 °C. ¹H NMR (500 MHz, Chloroform-*d*)  $\delta$  7.37 (d, J = 8.0 Hz, 1H, Ar*H*), 7.23 (t, J = 7.9 Hz, 1H, Ar*H*), 6.96 (dd, J = 13.5, 6.0 Hz, 2H, 2Ar*H*), 3.90 (s, 3H, OC*H*<sub>3</sub>), 2.97 (d, J = 1.3 Hz, 6H, N(C*H*<sub>3</sub>)<sub>2</sub>). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  151.7(Ar*C*), 139.5(Ar*C*), 127.7(Ar*C*), 123.9(Ar*C*), 121.0(Ar*C*), 113.0(Ar*C*), 56.1(OCH<sub>3</sub>), 38.8 (N(CH<sub>3</sub>)<sub>2</sub>). <sup>1</sup>H and <sup>13</sup>C NMR spectra match with literature data.<sup>7</sup>

Synthesis of Methyl 4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)benzoate (2). Methyl 4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)benzoate was synthesized according to a literature procedure.<sup>8</sup>

#### **Preparation of Neopentylglycolborane**

Neopentylglycol (recrystallized from DCM, 1. 25 g, 12 mmol) and a stirring bar were added to a 50 mL round bottom flask. The flask was evacuated for 5 min and then refilled with  $N_2$  three times. Toluene (distilled, 6 mL) was added *via* syringe. The flask was cooled to 0 °C in an ice/water bath for 10 min. Borane dimethyl sulfide complex (1.14 mL, 12 mmol) was added dropwise *via* a syringe. The reaction was kept at 0 °C for 0. 5 h then 23 °C for 1.5 h until the bubbling ceased.

#### Neopentylglycolborylation of Methyl 4-Iodobenzoate

A 50 mL round bottom flask was charged with a stirring bar, methyl 4-iodobenzoate (1. 572 g, 6 mmol), NiCl<sub>2</sub>(dppp) (163 mg, 0. 3 mmol, 0.05 equiv), PPh<sub>3</sub> (157 mg, 0.6 mmol, 0.1 equiv). The flask was degassed for 10 min and refilled with nitrogen. The process was repeated two more times. Then, toluene (distilled, 6 mL) was added *via* syringe. The reaction was left stirring for 5 min. Et<sub>3</sub>N (dry, 2.5 mL, 18 mmol, 3 equiv) was added *via* a syringe. The neopentylglycolborane prepared in step 1 was transferred to the flask *via* a syringe right after the addition of Et<sub>3</sub>N. The flask was heated to 100 °C for 1 h. Then the reaction mixture was cooled to 23 °C, quenched by addition of saturated ammonium chloride solution (20 mL) and ethyl acetate (20 mL). The

organic phase was washed with saturated ammonium chloride solution (20 mL) twice. The aqueous phase was combined and extracted with ethyl acetate (25 mL) three times. The organic phase was collected, combined and dried over brine and then anhydrous MgSO<sub>4</sub>. The solution was filtered and the filtrate was concentrated under vacuum. The crude product was purified by silica gel column chromatography with DCM/hexanes mixture, gradient from DCM/hexanes = 7:3 to DCM to DCM/ethyl acetate = 17:3 to obtain a white solid. The white solid was recrystallized from methanol to give a colorless flat crystal, m.p. 113-115.5 °C, 1.35 g, 91%.  $^{1}$ H NMR (500 MHz, Chloroform-d)  $\delta$  8.01 (d, J = 8.4 Hz, 2ArH), 7.87 (d, J = 8.3 Hz, 2ArH), 3.93 (s, 3COOCH<sub>3</sub>), 3.79 (s, 4H, 2(OCH<sub>2</sub>)), 1.04 (s, 6H, 2CH<sub>3</sub>).  $^{13}$ C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  167.2(COOCH<sub>3</sub>), 133.7(ArC), 131.7(ArC), 128.4(ArC), 72.3(OCH<sub>2</sub>), 52.0(COOCH<sub>3</sub>), 31.8(C(CH<sub>2</sub>O<sub>2</sub>(CH<sub>3</sub>)<sub>2</sub>), 21.8(CH<sub>3</sub>).  $^{1}$ H and  $^{13}$ C NMR spectra match with literature data.  $^{9}$ 

#### 4.2 Syntheses of Precatalysts

## **4.2.1** Syntheses of Precursors for Precatalysts

Synthesis of 9-Chloroanthracene. 9-Chloroanthracene was synthesized according to a literature procedure. A 250 mL round bottom flask was charged with a stirring bar, anthracene (3.56 g, 20 mmol, 1 equiv), CuCl<sub>2</sub> (anhydrous, dried in a 180 °C oven for 24 h prior to use, 5.44 g, 40 mmol, 2 equiv) and chlorobenzene (100 mL). The reaction mixture was heated in a silicone oil bath from 23 °C to 150 °C in 30 min and was allowed to cool in the oil bath to 23 °C in approximately 1 h. The solid cuprous chloride was removed by filtration. The filtrate was passed through 40 g basic alumina column to remove the copper residue. The column was eluted with chlorobenzene (50 mL), hexanes (100 mL) and dichloromethane (100 mL). The solution was collected and concentrated first in a rotary evaporator. Subsequently the chlorobenzene was removed by vacuum distillation. The solid was purified by silica gel column chromatography with hexanes. The first portion ( $R_f = 0.7$ ) was 9,10-dichloroanthracene, the second portion ( $R_f = 0.6$ ) was 9-chloroanthracene and the last portion was anthracene ( $R_f = 0.5$ ). 9-Chloroanthracene was recrystallized from ethanol prior to being used.

**9-Chloroanthracene.** Lemon yellow needle crystals (1.465 g, 70%) were obtained by recrystallization from ethanol. m.p. 108-109 °C, lit. 104-106 °C. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.52 (d, J = 8.8, 2H, 2Ar*H*-1), 8.41 (s, 1H, Ar*H*-10), 8.02 (d, J = 8.4, 2H, 2Ar*H*-4), 7.74 – 7.59

(m, 2H, 2Ar*H*-2), 7.59 - 7.50 (m, 2H, 2Ar*H*-3). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  128.7 (2Ar*C*CCl), 127.0 (Ar*C*Cl), 126.2 (2Ar*C*), 125.8 (Ar*C*H), 124.9 (Ar*C*H). M. p. matches with literature data. <sup>1</sup>

## Chlorination of Phenanthrene and 1, 2-Dihydroacenaphthene.

9-Chlorophenanthrene and 5-chloro-dihydroacenaphthene were synthesized according to a literature procedure.<sup>2</sup> CuCl<sub>2</sub> on basic alumina was prepared by dissolving CuCl<sub>2</sub> (10g) in water and adding it to basic alumina (20 g). The solid was dried in a rotary evaporator and then at 180 °C for 24 h prior to use. A round bottom flask was charged with a stirring bar, 1,2-dihydroacenaphthene (1.386 g, 9 mmol), CuCl<sub>2</sub>/Al<sub>2</sub>O<sub>3</sub> (22 g) and chlorobenzene (90 mL). The mixture was heated to 130 °C for 2 h. The mixture was filtered, washed with 10 mL chlorobenzene, and concentrated by vacuum distillation. An orange solid (972.6 mg, 57%) was obtained after purification by silica gel column chromatography with hexanes.

**5-Chloro-dihydroacenaphthene.** An orange solid (972.6 mg, 57%) was obtained after purification by silica gel column chromatography with hexanes. m. p. 71 °C, lit.<sup>10</sup> 70. 5 °C. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.86 (d, J = 8.3, 1H, Ar*H*-6), 7.66 – 7.54 (m, 1H, Ar*H*-7), 7.49 (d, J = 7.3, 1H, Ar*H*-8), 7.35 (d, J = 6.8, 1H, Ar*H*-4), 7.18 (d, J = 7.3, 1H, Ar*H*-3), 3.56 – 3.40 (m, 2H, ArC*H*<sub>2</sub>-2), 3.40 – 3.26 (m, 2H, ArC*H*<sub>2</sub>-1). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  150.6 (Ar*C*-CH<sub>2</sub>), 146.2 (Ar*C*-CH<sub>2</sub>), 145.2 (Ar*C*), 140.3 (Ar*C*), 129.5 (Ar*C*Cl), 128.9 (Ar*C*H), 127.4 (Ar*C*H), 126.9 (Ar*C*H), 120.2 (Ar*C*H), 119.6 (Ar*C*H), 119.4 (Ar*C*H), 30.8 (Ar*C*H<sub>2</sub>), 29.9 (Ar*C*H<sub>2</sub>). m. p. matches with literature data.<sup>10</sup>

**9-Chlorophenanthrene.**<sup>2</sup> A round bottom flask was charged with a stirring bar, phenanthrene (1.604 g, 9 mmol), CuCl<sub>2</sub>/Al<sub>2</sub>O<sub>3</sub> (22 g) and chlorobenzene (90 mL). The reaction mixture was heated to 130 °C for 2 h. The mixture was lifted out of the oil bath, cooled to 23 °C, filtered, washed with 10 mL chlorobenzene, and concentrated by vacuum distillation. An off-white solid (1.3844 g, 72.3%) was obtained after purification by silica gel column chromatography with hexanes. m. p. 49 °C. The solid was recrystallized from aqueous ethanol (95% ethanol in water) two times to give white needle-like crystals (0.374 g, 27.6%) with m. p. 51-52 °C. lit. 51-52 °C. <sup>1</sup>H NMR matches with literature data. H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.72 (d, J = 8.0, 1H, Ar*H*-5), 8.67 (d, J = 8.4, 1H Ar*H*-4), 8.41 (d, J = 8.6, 1H, Ar*H*-8), 7.89 (s, 1H, Ar*H*-10), 7.82 (d, J =

7.7, 1H, ArH-1), 7.79 – 7.70 (m, 2H, 2ArH-3, 6), 7.67 (t, J = 7.5, 1H, ArH-7), 7.62 (t, J = 7.4, 1H, ArH-2). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  131.7 (ArC), 131.3 (ArC), 130.5 (ArC-Cl), 129.4 (ArC), 129.3 (ArC), 127.8 (ArCH), 127.3 (ArCH), 127.2 (ArCH), 127.2 (ArCH), 126.8 (ArCH), 126.4 (ArCH), 125.2 (ArCH), 122.8 (ArCH), 122.6 (ArCH).

1-Naphthyl 4-methylbenzenesulfonate. Under nitrogen, to an oven dried round bottom flask charged with a stirring bar was added the sublimed 1-naphthol (3.0 g, 2.08 x 10-2 mol), toluenesulfonyl chloride (5.95 g, 3.12 x10<sup>-2</sup> mol) and freshly distilled dichloromethane (20mL). The flask was cooled for 10 min in an ice water bath. Anhydrous pyridine (25 mL, 0.31 mol) was added. The reaction was allowed to proceed with stirring in the ice water bath for 4 h after which the ice water bath was removed and the reaction was allowed to warm to room temperature and stirring was continued until complete consumption of the starting material was observed by TLC. The reaction was quenched by addition of water (40 mL) and the organic phase was separated. The aqueous phase was further extracted with dichloromethane (3 x 20mL) and all the organic layers were combined and washed successively with 15% HCl (2 x 20 mL) and Brine (3 x 20mL) then dried over anhydrous MgSO<sub>4</sub>. Following filtration the solvent was removed under reduced pressure and purified by silica gel column chromatography with dichloromethane to yield a pale yellow crystal (4.37 g, 70%). Recrystalization from methanol gives white crystals, m.p. 87 °C. lit. 11 89-91 °C. If 1-naphthol used in this synthesis is not sublimed, the resulting tosylate crystals are pale yellow. <sup>1</sup>H NMR (500 MHz, Chloroform-d)  $\delta$  7.93 (dd, J = 8.6, 1.2 Hz, 1H), 7.81 (dd, J= 12.5, 8.0 Hz, 3H), 7.75 (d, J = 8.3 Hz, 1H), 7.46 (dt, J = 21.4, 6.9 Hz, 2H), 7.38 (t, J = 7.9 Hz, 1H), 7.29 (d, J = 8.1 Hz, 2H), 7.23 (dd, J = 7.6, 1.1 Hz, 1H), 2.43 (s, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  145.7, 145.3, 134.6, 132.7, 129.7, 128.4, 127.6, 127.2, 127.0, 126.6, 126.6, 125.0, 121.7, 118.3, 21.6.

**2-Naphthyl 4-methylbenzenesulfonate.** Under nitrogen, to an oven dried round bottom flask charged with a stirring bar was added the sublimed 2-naphthol (5.48 g, 3.8 x 10-<sup>2</sup> mol), toluenesulfonyl chloride (8.68 g, 4.56 x10-<sup>2</sup> mol) and freshly distilled dichloromethane (31mL). The flask was cooled for 10 min in an ice water bath. Anhydrous pyridine (15g, 0.19 mol) was added. The reaction was allowed to proceed with stirring in the ice water bath for 4 h after which the ice water bath was removed and the reaction was allowed to warm to room temperature and stirring was continued until complete consumption of the starting material was observed by TLC.

The reaction was quenched by addition of water (40 mL) and the organic phase was separated. The aqueous phase was further extracted with dichloromethane (3 x 20mL) and all organic layers were combined and washed successively with 15% HCl (2 x 20 mL) and Brine (3 x 20mL) then dried over anhydrous MgSO<sub>4</sub>. Following filtration the solvent was removed under reduced pressure and purified by silica gel column chromatography with dichloromethane to yield pale yellow crystals (5.6 g, 80%). Recrystalization from methanol gives white crystals, m.p. 125 °C. lit. 11 119-120 °C. 1H NMR (500 MHz, Chloroform-d)  $\delta$  7.82 (d, J = 9.0 Hz, 1H), 7.75 (t, J = 8.8 Hz, 4H), 7.54 - 7.44 (m, 3H), 7.31 (d, J = 8.1 Hz, 2H), 7.11 (dd, J = 8.9, 2.1 Hz, 1H), 2.45 (s, 3H).  $^{13}$ C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  147.1, 145.3, 133.4, 132.4, 131. 8, 129.7, 129.6, 128.5, 127.8, 127.6, 126.7, 126.3, 121.1, 119.9, 21.6.

1-Naphthyl methanesulfonate. A flame-dried 50mL round bottom flask was charged with a stirring bar, sublimed 1-naphthol (2.88 g, 20 mmol, 1 equiv), and DMAP (244 mg, 2 mmol, 0.1 equiv). The flask was sealed with a rubber septum and then purged with nitrogen for 10 min. Dichloromethane (20 ml) was added via syringe followed by pyridine (8 mL, 100 mmol, 5 equiv). The flask was placed in a 0°C ice bath and stirred for 10 min. Methanesulfonylchloride (2.4 mL, 30 mmol, 1.5 equiv) was added dropwise via syringe. The reaction was allowed to run for 1h and then the ice bath was removed. After 10 hours, the reaction was quenched by addition of 10 mL of 1M HCl. The contents of the flask were transferred to a 125 mL separatory funnel and then washed three times with 10 mL of 1M HCl. The organic layer was dried by washing two times with 15 mL brine solution and then stirred over anhydrous MgSO<sub>4</sub>. The solution was filtered and then the solvent was removed on a rotary evaporator. Column chromatography on silica gel with dichloromethane yielded a solid that was recrystallized from methanol to obtain colorless crystals. Yield (4.08g, 92%), m.p. 36.5°C, lit. 11 35-36 °C. 1H NMR (500 MHz, Chloroform-d)  $\delta$  8.16 - 8.11 (m, 1H), 7.89 (d, J = 8.1 Hz, 1H), 7.81 (dd, J = 8.1, 1.1 Hz, 1H), 7.62 - 7.51 (m, 3H), 7.47 (td, J = 7.9, 1.0 Hz, 1H), 3.20 (s, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$ 145.4, 135.0, 128.2, 127.5, 127.4, 127.1, 125.5, 121.5, 118.5, 38.1.

**2-Naphthyl methanesulfonate.** A flame-dried 50mL round bottom flask was charged with a stirring bar, sublimed 2-naphthol (2.88 g, 20 mmol, 1 equiv), and DMAP (244 mg, 2

mmol, 0.1 equiv). The flask was sealed with a rubber septum and then purged with nitrogen for 10 minutes. Dichloromethane (20 ml) was added *via* syringe followed by pyridine (8 mL, 100 mmol, 5 equiv). The flask was placed in a 0°C ice bath and stirred for 10 min. Methanesulfonylchloride (2.4 mL, 30 mmol, 1.5 equiv) was added dropwise *via* syringe. The reaction was allowed to run for 1 h and then the ice bath was removed. After 10 h, the reaction was quenched by addition of 10 mL of 1M HCl. The contents of the flask were transferred to a 125 mL separatory funnel and then washed three times with 10 mL of 1 M HCl. The organic layer was dried by washing two times with 15 mL brine solution and then stirred over anhydrous MgSO<sub>4</sub>. The solution was filtered and then the solvent was removed on a rotary evaporator. Column chromatography on silica gel with dichloromethane yielded a solid that was recrystallized from MeOH to obtain colorless crystals. Yield (3.20g, 72%) m. p. 105-106.5 °C, lit.<sup>11</sup> 101-102 °C. ¹H NMR (500 MHz, Chloroform-*d*)  $\delta$  7.93 - 7.83 (m, 3H), 7.78 (s, 1H), 7.55 (p, J = 6.8 Hz, 2H), 7.42 (d, J = 9.0 Hz, 1H), 3.19 (s, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  146.8, 133.5, 132.0, 130.2, 127.8, 127.8, 127.1, 126.5, 120.7, 119.3, 37.3.

Ni(COD)<sub>2</sub>. Ni(COD)<sub>2</sub> was synthesized via a modified literature procedure. <sup>12</sup> NiCl<sub>2</sub>(pyridine)<sub>4</sub> (20.58 g) was prepared by refluxing of  $NiCl_2(H_2O)_6$  (10.8 g) in pyridine (125 mL) for 3 h. The bright blue solid was collected by vacuum filtration, and then allowed to air dry. A 250 mL round bottom flask was charged with a stirring bar and NiCl<sub>2</sub>(pyridine)<sub>4</sub> (8.92 g, 20 mmol). The flask was evacuated (10 min) and refilled with nitrogen three times. 1,5-Cyclooctadiene (7.4 mL, 60 mmol) and THF (12 mL, distilled immediately before addition) were introduced via syringes. The mixture was left stirring for 5 min. Na (in small pieces, 0.92 g) was added quickly. The solution was cooled down to - 78 °C for 10 min. The flask was evacuated (15 s), refilled with nitrogen (1 min) and the procedure was repeated two more times. The mixture was warmed to 23 °C and kept stirring vigorously at 23 °C for 3 h. MeOH (24 mL, distilled from Mg turnings under nitrogen immediately before addition) was added to induce the precipitation of Ni(COD)2. Stirring was halted, and the yellow precipitate was allowed to settle. After all sodium was consumed, the black upper layer was removed by syringe. MeOH (12 mL) was added again to rinse the crystals, and the upper layer was removed by syringe. The procedure was repeated 4 more times until the upper layer was clear. The Ni(COD)<sub>2</sub> (3.3 g, 70%, yellow solid) was dried overnight under vacuum for 12 h. Ni(COD)<sub>2</sub> (1 g) was dissolved in toluene (distilled within 1 day of use) at 23 °C inside the glove box, and then the solution was filtered through celite inside the glove box. The

filtrate was kept in a screw cap vial (20 mL) at -78 °C for 12 h until a bright yellow solid formed in the bottom of the vial. The bright yellow crystals (350 mg) were washed with ethyl ether (distilled right before use) two times, hexanes (distilled right before use) two times and dried under vacuum for 2 h. The NMR tube was prepared inside a nitrogen filled glove box with  $C_6D_6$ .  $^1H$  NMR (500 MHz,  $C_6D_6$ )  $\delta$  4.40 (s, 4H, 4CH), 2.17 (s, 8H, 4CH<sub>2</sub>).  $^{13}C$  NMR (126 MHz,  $C_6D_6$ )  $\delta$  89.7 (4CH), 30.9 (4CH<sub>2</sub>).  $^{14}H$  NMR matches with literature data.  $^{12}$ 

#### **4.2.2** Syntheses of Precatalysts

General Procedure for the Synthesis of Ni<sup>II</sup>X(Aryl)(PCy<sub>3</sub>)<sub>2</sub>. Nickel sigma complexes used as precatalysts were synthesized according to a literature procedure.<sup>13</sup> All nickel sigma complexes were synthesized and purified in a nitrogen filled glove box.

General Procedure for the Synthesis of Ni<sup>II</sup>Cl(Aryl)(PCy<sub>3</sub>)<sub>2.</sub>

Oxidative Addition of Aryl Cchlorides to Ni(COD)<sub>2</sub>. In a nitrogen filled glove box, a stirring bar, Ni(COD)<sub>2</sub> (100 mg, 0.36 mmol), PCy<sub>3</sub> (300 mg, 1.07 mmol), and THF (1 mL) were added to a 20 mL vial. The solution was left stirring for 1 min, and then 9-chloroanthracene (76 mg, 0.36 mmol) was added. The reaction was left stirring inside a glove box with inert atmosphere for 6 h. At this time the oxidative addition reaction was complete as demonstrated by NMR. The reaction mixture was filtered through a membrane (0.22  $\mu$ m) inside the glovebox, the solid was collected, washed 5 times with distilled hexanes (2 mL), then dried for 12 h under vacuum. Yield: 200 mg, 70%.

**Ligand exchange.** Ni<sup>II</sup>Cl(1-naphthyl)(PPh<sub>3</sub>)<sub>2</sub> (0.2 mmol, 149 mg), PCy<sub>3</sub> (0.6 mmol, 148 mg) and 10 mL EtOH (anhydrous) were added to a Schlenk tube inside the glovebox with a stirring bar. The tube was sealed, brought outside the glovebox, and then heated to 80 °C for 2 h. The reaction mixture was then cooled to 23 °C. The Schlenk tube was brought inside the glove box. After filtration, the solid was washed with ethyl ether three times inside the glovebox. The solid was recrystallized from a DCM/hexanes = 2:8 mixture to yield a yellow solid (106 mg, 68%).

(6) The reaction was carried out following the general procedure from 1-chloronaphthalene (50 μL, 0.36 mmol). Yield: by oxidative addition, from 1-chloronaphthalene (50 μL, 0.36 mmol), Ni(COD)<sub>2</sub>(100 mg, 0.36 mmol), 200mg, 70%; by ligand exchange method, from Ni<sup>II</sup>Cl(1-naphthyl)(PPh<sub>3</sub>)<sub>2</sub> (0.2 mmol, 149 mg), PCy<sub>3</sub> (0.6 mmol, 148 mg), yield 106 mg, 68%. Yellow solid, m.p. 190.4 °C (decompose). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 10.18 (d, J = 8.6, 1H), 7.50 (d, J = 8.4, 1H), 7.47 – 7.42 (m, 1H), 7.40 (s, 1H), 7.23 (d, J = 7.6, 2H), 7.05 – 6.97 (m,

1H), 2.10-0.72 (br, 76H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  153.8, 141.6, 136.7, 135.1, 132.5, 127.8, 124.5, 124.3, 122.4, 121.4, 33.4, 30.2, 29.4, 28.1, 28.0, 27.7, 26.6. <sup>31</sup>P NMR (203 MHz, CDCl<sub>3</sub>)  $\delta$  10.66. [M-Cl]<sup>+</sup> calcd for C<sub>46</sub>H<sub>73</sub>NiP<sub>2</sub>, 745.4541, HRMS found 745. 4360.

CI Cy<sub>3</sub>P-Ni-PCy<sub>3</sub>

chloride (59 mg, 0.36 mmol, 1 equiv), with Ni(COD)<sub>2</sub> (100 mg, 0.36 mmol, 1 equiv). Yield: 147 mg, 52%. Yellow solid, m.p. 147-148 °C (decompose).  $^{1}$ H NMR (500 MHz,  $C_{6}D_{6}$ )  $\delta$  8.22 (d, J = 7.8, 1H), 8.18 (s, 1H), 7.77 (d, J = 7.4, 2H), 7.71 (d, J = 7.9, 1H), 7.52 (d, J = 7.8, 1H), 7.31 (s, 1H), 1.07-2.26 (br, 66 H).  $^{31}$ P NMR (203 MHz,  $C_{6}D_{6}$ )  $\delta$  12.36.  $^{13}$ C NMR (126 MHz,  $C_{6}D_{6}$ )  $\delta$  137.8, 137.6, 132.6, 130.3, 128.5, 128.2, 125.2, 125.0, 123.2, 122.9, 33.9, 30.1, 27.8, 26.6.  $^{1}$ H and  $^{31}$ P NMR spectra match literature data.  $^{14}$  The catalyst was also recrystallized from toluene/hexanes = 1:10 mixture. In an oven dried screw cap vial were added Ni $^{11}$ Cl(2-Naphthyl)(PCy<sub>3</sub>)<sub>2</sub> (100.1 mg), the catalyst was dissolved in 4 mL boiling toluene/hexanes = 1:10 mixture. The dark red solution was filtered through a 0.22  $\mu$ m membrane filter and cooled in -10 °C freezer for 4 h. The solid was collected, washed with cold hexanes and dried in vacuum. Yield: 77.8 mg. The  $^{1}$ H,  $^{31}$ P NMR spectra as well as kinetic results are identical with that of the non-recrystalized precatalyst.

CI Cy<sub>3</sub>P-Ni-PCy<sub>3</sub>

(12) The reaction was carried out following the general procedure from 5-chloro-1,2-dihydroacenaphthene (68.6 mg, 0.36 mmol). Yield: 110mg, 39%. Yellow-brown solid, m.p. 151-153 °C (decompose). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.92 (d, J = 7.0, 1H), 7.75 (d, J = 6.5, 2H), 7.32 (d, J = 6.4, 2H), 3.33 (s, 2H), 3.31 (s, 2H), 2.48-0.91 (br, 66H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  146.1, 141.0, 139.4, 138.4, 137.6, 129.9, 129.2, 125.4, 119.1, 118.5, 34.0, 30.8, 30.0, 28.4, 28.1, 27.1. <sup>31</sup>P NMR (203 MHz, CDCl<sub>3</sub>)  $\delta$  11.69. [M-Cl]<sup>+</sup> calcd for C<sub>48</sub>H<sub>75</sub>NiP<sub>2</sub>, 771.470, LC/MS found 771.500. Anal. calcd for C<sub>48</sub>H<sub>75</sub>ClNiP<sub>2</sub>: C, 71.33; H, 9.35. Found: C 71.63 H, 8.97.

CI Cy<sub>3</sub>P-Ni-PCy<sub>3</sub>

(13)In a nitrogen filled glove box, 1-chloro-2methoxynaphthalene (154 mg, 0.8 mmol, 1.1 equiv), Ni(COD)<sub>2</sub> (200 mg, 0.72 mmol, 1 equiv) and PCy<sub>3</sub> (610 mg, 2.16 mmol, 3 equiv) were added to a 20 mL vial charged with a stirring bar. THF (distilled, 1 mL) was added via syringe. The solution was left stirring for 18 h, transferred to a Schlenk flask and brought outside the glove box. The dark red solution was concentrated under reduced pressure and brought inside

the glove box. Distilled hexanes (1 mL) were added to induce precipitation. The precipitation was collected and washed with distilled hexanes (0.5 mL) five times. The pink solid was collected and dried under vacuum for 6 h. Then, the pink solid (150 mg) was transferred to a 4 mL vial. Dichloromethane/hexanes 1:7 solution was heated until boiling and added to the solid until the solid was fully dissolved. (In the case of slow solvation, a maximum of 4 mL of mixed solvent was added and the vial was sealed and heated until homogenous.) The solution was filtered while still hot to generate a red solution. The red solution was left in a -10 °C freezer for 12 h to produce a pink solid. The pink solid was collected, washed with distilled hexanes (0.5 mL) three times and dried under vacuum to give the COD free catalyst. (57.8 mg, yield: 10%) m.p. 193 °C.  $^{1}$ H NMR (500 MHz, Benzene- $d_6$ )  $\delta$  10.70 (d, J = 8.4 Hz, 1H), 7.81-7.68 (m, 2H), 7.51 (d, J = 8.7 Hz, 1H), 7.35 (d, J = 7.6 Hz, 1H), 6.71 (d, J = 8.5 Hz, 1H), 3.72 (s, 3H), 2.38 (s, 6H), 2.06 (s, 6H), 1.86 (m, 18H), 1.75-1.59 (m, 18H), 1.26 (s, 12H), 1.07 - 0.90 (m, 12H).  $^{31}$ P NMR (203 MHz,  $C_6D_6$ )  $\delta$  11.76.  $^{13}$ C NMR (126 MHz,  $C_6D_6$ )  $\delta$  159. 7, 143.7, 134.2, 128.8, 128.2, 124.3, 123.2, 122.5, 109.1, 53.2, 36.0, 34.6, 34.0, 30.4, 28.2, 28.0, 26.7, 25.3, 20.5, 11.3. [M-CI]<sup>+</sup> calcd for  $C_{47}H_{75}$ ONiP<sub>2</sub>, 775.4647, HRMS found 775.4639.

CI Cy<sub>3</sub>P-Ni-PCy<sub>3</sub>

(14) In a nitrogen filled glove box, 9-chlorophenanthrene (61 mg, 0.29 mmol, 1.1 equiv), Ni(COD)<sub>2</sub> (70 mg, 0.26 mmol, 1 equiv) and PCy<sub>3</sub> (280 mg, 0.78 mmol, 3 equiv) were added to a 20 mL vial charged with a stirring bar. THF (distilled, 0.5 mL) was added via a syringe. The solution was left stirring for 6 h. The solid was collected, washed with hexanes (1 mL) five times. The yellow solid was dried under vacuum for 6 h to remove the solvent. Then, the yellow solid (97 mg) was transferred to a 4 mL vial. Dichloromethane/hexanes 1:5 solution was heated until boiling and added to the solid until the solid was fully dissolved. (In the case of slow solvation, maximum of 4 mL of mixed solvent was added and the vial was sealed and heated till homogenous) The solution was filtered while still hot to generate a red solution. The red solution was concentrated under reduced pressure to produce a yellow solid. The yellow solid was washed 3 times with distilled hexanes to remove the COD. The yellow solid was collected and dried under vacuum for 12 h. (59.2 mg, yield: 27.4%) Yellow solid, m.p. 158 °C (decompose). H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  11.13 (d, J = 8.0 Hz, 1H), 8.61 (dd, J = 31.5, 8.3 Hz, 2H), 7.99 (s, 1H), 7.91 (t, J = 7.4 Hz, 1H), 7.75 (d, J = 7.9 Hz, 1H), 7.61 (t, J = 7.5 Hz, 1H), 7.51  $(t, J = 7.4 \text{ Hz}, 1\text{H}), 7.46-7.38 \text{ (m, 1H)}, 4.38 \text{ (s, 1.2 H, C}_2\text{Cl}_2), 2.55-0.72 \text{ (m, 80H)}.$  <sup>13</sup>C NMR (91) MHz,  $CDCl_3$ )  $\delta$  141.2, 137.4, 136.1, 135.1, 132.2, 128.5, 127.1, 126.3, 125.6, 125.4, 123.9, 122.6, 122.5, 34.7, 33.8, 30.5, 29.7, 28.1, 27.8, 26.7.  $^{31}P$  NMR (203 MHz, CDCl<sub>3</sub>)  $\delta$  9.75. [M-Cl]<sup>+</sup> calcd for  $C_{50}H_{75}NiP_2$ , 795.470, LC-MS found 795.509. Anal. calcd for  $C_{50}H_{75}NiClP_2(CH_2Cl_2)_{0.55}$ : C, 69.08; H, 8.73. Found: C 68.69 H, 8.48. The ratio between  $CH_2Cl_2$  and the nickel complex was determined by  $^{1}H$  NMR.

CI Cy<sub>3</sub>P-Ni-PCy<sub>3</sub>

(15) The reaction was carried out following the general procedure, from 9-chloroanthracene (77 mg, 0.36 mmol). Yield: 248 mg, 82%. Orange solid, m.p. 170 °C (decompose).  $^{1}$ H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  10.21 (d, J = 7.6, 2H), 7.79 (s, 1H), 7.73 (d, J = 7.2, 2H), 7.48 – 7.39 (m, 2H), 7.39 – 7.31 (m, 2H), 1.92 – 0.50 (br, 68H).  $^{13}$ C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  140.0, 136.3, 130.6, 128.6, 124.6, 121.4, 121.4, 121.0, 77.4, 77.2, 76.9, 34.1, 34.0, 34.0, 30.1, 28.1, 26.6.  $^{31}$ P NMR (146 MHz, CDCl<sub>3</sub>)  $\delta$  8.82. [M-Cl]<sup>+</sup> calcd for C<sub>50</sub>H<sub>75</sub>NiP<sub>2</sub>, 795.4697, HRMS found 795.4698.

General Procedure for the Synthesis of Ni<sup>II</sup>OTs(Aryl)(PCy<sub>3</sub>)<sub>2</sub> or Ni<sup>II</sup>OMs(Aryl)(PCy<sub>3</sub>)<sub>2</sub>. A stirring bar, Ni(COD)<sub>2</sub> (100 mg, 0.36 mmol), PCy<sub>3</sub> (300 mg, 1.07 mmol), and 1 mL THF were added to a 20 mL vial. The solution was left stirring for 1 min, and then 1-naphthyl 4-methylbenzenesulfonate (107 mg, 0.36 mmol) was added. The reaction was left stirring inside a glove box with inert atmosphere for 6 h - until the oxidative addition reaction was complete as demonstrated by NMR. The reaction mixture was dissolved in THF, filtered through a 22 μm membrane and collected. The filtrate was collected in a 50 mL Schlenk flask and dried under vacuum outside the glove box. The side arm of the flask was degassed prior to application of the vacuum. Once the filtrate had dried, the flask was brought inside the glove box and the residue was washed with hexanes 5 times. The solid was collected and dried under vacuum overnight.

OTs Cy<sub>3</sub>P-Ni-PCy<sub>3</sub>

(8) The reaction was carried out following the general procedure from 1-naphthyl 4-methylbenzenesulfonate (108.5 mg, 0. 36 mmol), Ni(COD)<sub>2</sub> (100 mg, 0.36 mmol). Yield: 212 mg, 64%. The catalyst contained a small amount of Ni<sup>II</sup>OTs(2-Naphthyl)(PCy<sub>3</sub>)<sub>2</sub>, as indicated by a small peak in <sup>31</sup>P NMR. Yellow solid, m.p. 149 °C. <sup>1</sup>H NMR (500 MHz,  $C_6D_6$ )  $\delta$  10.86 (d, J = 15.3, 1H), 8.18 (d, J = 7.9, 2H), 7.77 (d, J = 6.9, 2H), 7.56 (d, J = 8.0, 1H), 7.48 – 7.35 (m, 2H), 7.31 (d, J = 7.6, 1H), 7.11 (t, J = 7.5, 1H), 7.02 (d, J = 7.8, 2H), 2.51-2.13 (br, 6H), 2.13 (s, 3H), 2.13 – 0.80 (br, 60H). <sup>31</sup>P NMR (203 MHz,  $C_6D_6$ )  $\delta$  11.04. <sup>13</sup>C NMR (126 MHz,  $C_6D_6$ )  $\delta$  143.4, 141.3, 139.4, 136.5, 135.4, 132.5, 128.3, 128.2, 128.0, 127.1, 125.3, 123.7, 123.5, 122.2, 34.0, 31.6, 30.6, 30.0, 27.7, 26.5, 22.7, 20.7. [M+H]<sup>+</sup> calcd for  $C_{53}H_{81}NiO_3P_2S$ , 917.473, LC/MS found

917.057. [M-OTs]<sup>+</sup> calcd for  $C_{46}H_{73}NiP_2$ , 745.4541, HRMS found 745.4539, [M-OTs + MeCN]<sup>+</sup> calcd for  $C_{48}H_{76}NNiP_2$ , 786.4806, HRMS found 786.4839.

OTs Cy<sub>3</sub>P-Ni-PCy<sub>3</sub>

(9) The reaction was carried out following the general procedure from 2-naphthyl 4-methylbenzenesulfonate (108. 5 mg, 0. 36 mmol), with Ni(COD)<sub>2</sub> (100 mg, 0.36 mmol). Yield: 182 mg, 55%. Yellow solid, m.p. 112-113 °C. ¹H NMR (500 MHz,  $C_6D_6$ )  $\delta$  8.40 (d, J = 7.0, 1H), 8.07 (d, J = 6.5, 2H), 7.79 (s, 1H), 7.59 (dd, J = 21.9, 7.6, 2H), 7.32 (d, J = 4.7, 1H), 7.26 (t, J = 7.3, 1H), 6.92 (d, J = 6.9, 2H), 2.35-1.99 (br, 11H), 1.99 (s, 3H), 1.99-0.83 (br, 63H). ³¹P NMR (203 MHz,  $C_6D_6$ )  $\delta$  10.99. ¹³C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  143.4, 139.5, 137.1, 136.9, 132.4, 130.6, 128.4, 128.0, 127.0, 125.3, 125.0, 123.6, 122.7, 34.1, 31.6, 30.2, 29.9, 27.5, 26.5, 20.7. [M-OTs]<sup>+</sup> calcd for  $C_{46}H_{73}NiP_2$ , 745.454, LC/MS found 745. 611. [M+MeCN+Na]<sup>+</sup> calcd for  $C_{55}H_{83}NNaNiO_3P_2S$ , 980.482, LC/MS found 980.470. [M-OTs]<sup>+</sup> calcd for  $C_{46}H_{73}NiP_2$ , 745.4541, HRMS found 745.4555.

OMs Cy<sub>3</sub>P-Ni-PCy<sub>3</sub>

(10) The reaction was carried out following the general procedure from 1-naphthyl methylsulfonate (163 mg, 0. 72 mmol), with Ni(COD)<sub>2</sub> (200 mg, 0.72 mmol). Yield: 482 mg, 79%. Yellow solid, m.p. 139-143 °C. The catalyst had small amount of impurity after 24 h of drying under vacuum by <sup>31</sup>P NMR. The crude was used for reactivity tests. <sup>1</sup>H NMR (500 MHz, Benzene- $d_6$ )  $\delta$  10.96 (d, J = 8.7 Hz, 1H), 7.74 (s, 1H), 7.64 (s, 1H), 7.54 (s, 1H), 7.39 (d, J = 9.3 Hz, 1H), 7.10 (s, 1H), 3.67 (s, 2.8H), 2.87 (s, 3H), 1.10 (s, 77H). <sup>31</sup>P NMR (203 MHz, Benzene- $d_6$ )  $\delta$  10.01, 9.67. <sup>13</sup>C NMR (126 MHz, C<sub>6</sub>D6)  $\delta$  141.27, 136.29, 135.12, 132.30, 128.19, 127.17, 125.24, 123.85, 123.60, 122.16, 67.46, 40.34, 33.84, 31.40, 31.30, 30.53, 29.84, 27.80, 27.68, 26.55, 25.46. Anal. calcd for C<sub>47</sub>H<sub>76</sub>NiO<sub>3</sub>P<sub>2</sub>S(C<sub>4</sub>H<sub>8</sub>O)<sub>0.7</sub>(C<sub>6</sub>H<sub>14</sub>)<sub>0.5</sub>: C, 67.85; H, 9.48. Found: C 67.67; H, 9.16. The ratio of THF and hexanes to the nickel complex was determined by <sup>1</sup>H NMR spectroscopy.

OMs Cy<sub>3</sub>P-Ni-PCy<sub>3</sub>

(11) The reaction was carried out following the general procedure from 2-naphthyl methylsulfonate (163 mg, 0. 72 mmol), with Ni(COD)<sub>2</sub> (200 mg, 0.72 mmol). Yield: 458 mg, 75%. Yellow solid, m. p. 123 °C. <sup>1</sup>H NMR (500 MHz, Benzene- $d_6$ )  $\delta$  8.41 (s, 1H), 7.74 (s, 1H), 7.66 – 7.44 (m, 2H), 7.26 (m, 2H), 3.57 (s, 3H, THF), 2.74 (s, 3H), 1.73-1.13 (m, 74H). <sup>31</sup>P NMR (203 MHz, Benzene- $d_6$ )  $\delta$  9.84. <sup>13</sup>C NMR (126 MHz, Benzene- $d_6$ )  $\delta$  136.6, 136.5, 132.5, 128.2,

128.0, 126.3, 125.9, 125.3, 125.0, 123.6, 122.7, 34.6, 33.8, 30.2, 30.0, 29.8, 27.6, 26.5, 25.5. Anal. calcd for  $C_{47}H_{76}NiO_3P_2S(C_4H_8O)_{0.7}$ : C, 67.09; H, 9.15. Found: C 66.99; H, 8.90. The ratio of THF to the nickel complex was determined by  $^1H$  NMR spectroscopy.

#### **4.3** General Procedure for Kinetic Experiments

Sampling Kinetic Experiment Without Additional PCy<sub>3</sub>. In an oven dried test tube (15 mm x 85 mm) charged with a stirring bar (5/8" x 5/16") were added 2-methoxyphenyl dimethylsulfamate (69.40  $\pm$  0.10 mg, 0. 3 mmol, 1 equiv), methyl 4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)benzoate (78.15  $\pm$  0.10 mg, 0.315 mmol, 1.05 equiv), Ni<sup>II</sup>Cl(1-Naphthyl)(PCy<sub>3</sub>)<sub>2</sub> (11.73  $\pm$  0.0510 mg, 0.015 mmol, 5% catalyst loading) and K<sub>3</sub>PO<sub>4</sub>(H<sub>2</sub>O)<sub>n</sub> (191.00  $\pm$  1.00 mg,  $\sim$  9 mmol,  $\sim$  3 equiv). The test tube was brought into a nitrogen filled glove box (moisture level < 10 ppm) through three degassing cycles. Distilled THF (1 mL) was added inside the glove box and the test tube was sealed by a rubber septum and left stirring for 60 min. A sample was taken by syringe and transferred outside the glove box. The sample was diluted by distilled THF (0.2 mL) and filtered through a short column of silica gel. The filtrate was concentrated and the GC analysis was carried out.

Sampling Kinetic Experiment with Additional PCy<sub>3</sub> (10%). Inside a nitrogen filled glove box, a 20 mL vial was charged with a stirring bar, PCy<sub>3</sub> (37.1 mg) and distilled THF (4.5 mL). The vial was screw closed and kept stirring. The solution was prepared prior to the kinetic experiments and kept inside the glove box for less than 4 h to prevent the oxidation of PCy<sub>3</sub> and evaporation of THF. Outside the glove box, in an oven dried test tube (15 mm x 85 mm) charged with a stirring bar (5/8" x 5/16") were added 2-methoxyphenyl dimethylsulfamate (69.40  $\pm$  0.10 mg, 0. 3 mmol, 1 equiv), methyl 4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)benzoate (78.15 + 0.10 mg, 0.315 mmol, 1.05 equiv),  $Ni^{II}Cl(1-Naphthyl)(PCy_3)_2$  (11.73 ± 0.050 mg, 0.015 mmol, 5% catalyst loading) and  $K_3PO_4(H_2O)_n$  (191.00 ± 1.00 mg, ~ 9 mmol, ~ 3 equiv). The test tube was brought into a nitrogen filled glove box (moisture level < 10 ppm) through three degassing cycles. PCy<sub>3</sub> in THF (1 mL) was added inside the glove box. The test tube was sealed by a rubber septum and left stirring for 60 min. A sample was taken by syringe and transferred outside the glove box. The sample was diluted by distilled THF (0.2 mL) and filtered through a short column of silica gel. The filtrate was concentrated and the GC analysis was carried out. The results are summarized in Table SI 1-8. The product was isolated by silica gel column chromatography with 10% ethyl acetate in hexanes.

Sampling Kinetic Experiments with as Received THF. As received, wet THF was used from a commercial source (Fisher certified grade), containing ~0.025% butylated hydroxytoluene, about five months after opening and purged by  $N_2$  for 30 min prior to use. In an oven dried test tube (15 mm x 85 mm) charged with a stirring bar (5/8'' x 5/16'') were added 2-methoxyphenyl dimethylsulfamate (69.40  $\pm$  0.10 mg, 0. 3 mmol, 1 equiv), methyl 4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)benzoate (78.15  $\pm$  0.10 mg, 0.315 mmol, 1.05 equiv),  $Ni^{II}Cl(1-Naphthyl)(PCy_3)_2$  (11.73 $\pm$  0.10 mg, 0.015 mmol, 5% catalyst loading) and  $K_3PO_4(H_2O)_n$  (191.00  $\pm$  1.00 mg,  $\sim$  9 mmol,  $\sim$  3 equiv). The test tube was brought into a nitrogen filled glove box (moisture level < 10 ppm) through three degassing cycles. Wet THF (1 mL) was added inside the glove box and the test tube was sealed by a rubber septum and left stirring for 60 min. A sample was taken by syringe and transferred outside the glove box. The sample was diluted by distilled THF (0.2 mL) and filtered through a short column of silica gel. The filtrate was concentrated and the GC analysis was carried out.

COOMe

3: Isolated after kinetics experiments. Purified by silica gel column chromatography with hexanes to 10% ethyl acetate in hexanes gradient. White solid, m.p. 47.5-49 °C.  $^{1}$ H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.79 (d, J = 7.6, 1H), 7.50 (t, J = 7.5, 1H), 7.37 (t, J = 8.3, 2H), 7.25 (d, J = 8.6, 2H), 6.94 (d, J = 8.5, 2H), 3.85 (s, 3H), 3.67 (s, 3H).  $^{13}$ C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  169.5, 159.1, 131.3, 131.0, 130.9, 129.9, 129.6, 127.0, 113.7. Spectroscopy data matches with literature data.

Isolated yields are listed in Table SI 9. <sup>1</sup>H and <sup>13</sup>C NMR spectra are listed in section 5.

#### Sample Procedures for Drying K<sub>3</sub>PO<sub>4</sub>(H<sub>2</sub>O)<sub>n</sub> and Measuring Water Content:

## Measuring Water Content in K<sub>3</sub>PO<sub>4</sub>(H<sub>2</sub>O)<sub>n</sub> by Weight Loss:

In a tared round bottom flask (47.6791 g, average of three measurements within instrumental error) fitted with an adapter were added  $K_3PO_4(H_2O)_n$  (48.9636 g for flask and base). The round bottom flask was heated by a Fischer burner until the visible water was evaporated. The flask was then connected to vacuum and flame dried until constant mass (flask + base: 48.6924 g, average of three measurements within instrumental error). The value of n is determined by the following equation:

$$n = \frac{mole (H_2O)}{mole (K_3PO_4)} = \frac{m(H_2O)/M_w(H_2O)}{m(K_3PO_4)/M_w(K_3PO_4)} = \frac{\frac{(48.9636 - 48.6924)g}{18.02g/mol}}{\frac{(48.6924 - 47.6791)g}{212.27g/mol}} = 3.2$$

The water content determination is more accurate with large amount of base. To minimize error induced by dehydration of glassware, water content determination with smaller round bottom flask (25 mL) are more accurate compared to large ones (250 mL) when the water content of base is determined.

#### Drying $K_3PO_4(H_2O)_n$ to Predetermined n:

The amount of weight loss desired is calculated from following equation:

$$m(mass\ loss) = m_1 \frac{18.02(n_1 - n_2)}{(212.27 + 18.02 \times n_1)}$$

where: m<sub>1</sub> is the mass of the K<sub>3</sub>PO<sub>4</sub>(H<sub>2</sub>O)n<sub>1</sub> from commercial source;

 $n_1$  is the water content determined by weight loss in commercial source

 $n_2$  is the desired water content.

Drying procedure for base with n = 3.2:

From the equation above, for 23.8093 g of  $K_3PO_4(H_2O)_7$ , the desired weight loss was calculated to be 4.9 g. In a tared round bottom flask (250 mL) fitted with an adapter were added a stirring bar,  $K_3PO_4(H_2O)_7$  (23.8093 g, hydrous, tribasic from commercial source) were added and dried at 40 °C under vacuum. After 6 h, the mass of the flask with the base was weighed and 4.8841 g water was lost. The base was ground and the exact amount of water was then verified to be 3.2 by the method above.

Kinetics experiments were carried out with bases dried by these procedures and the results are provided in Table SI 2.

## 4.4 Supporting Data for Kinetic Experiments

Table SI 1. Supporting Data for Table  $\mathbf{1}^a$ 

entry	catalyst	mixed-ligand (%)	time (h)	convn/yield (%)	compound number
1	$Ni(COD)_2$	PPh <sub>3</sub> (10)	90	6.9	
2	$Ni(COD)_2$	$PCy_{3}(10)$	1	71/71	3a
3	$Ni(COD)_2$	PCy <sub>3</sub> (20)	1	92/89	<b>3b</b>
			1	61	
4	ÇI Ph₃P−Ni−PPh₃	DC (10)	24	95	
4		$PCy_3$ (10)	48	96	
			70	100/96	3c
	ÇI Ph₃P−Ņi−PPh₃		48	99	
5		PCy <sub>3</sub> (15)	56	100	
6	CI Ph <sub>3</sub> P-Ni-PPh <sub>3</sub>	PCy <sub>3</sub> (20)	24	100/95	<b>3</b> d
	Br Ph₃P−Ni−PPh₃		48	96	
7		PCy <sub>3</sub> (10)	70	100/94	<b>3e</b>
_	Br Ph <sub>3</sub> P-Ni-PPh <sub>3</sub>		48	99	
8		PCy <sub>3</sub> (15)	57	100	
	CI Cy₃P−Ni−PCy₃		0.7	98	
9			0.9	100	
	CI Cy₃P−Ņi−PCy₃		1	44	
10		COD (10)	2	57/50	3f
	CI Cy₃P−Ni−PCy₃		1	53	
11		PPh <sub>3</sub> (10)	2	60/48	<b>3</b> g

<sup>a</sup>Conversion by GC, isolated yield, both as percentages.

**Table SI 2. Supporting Data for Table 2** 

entry	catalyst	THF	n	time (min)	convn (%)
	CI Cy <sub>3</sub> P-Ni-PCy <sub>3</sub>			70	16
1	Cy <sub>3</sub> P-NI-PCy <sub>3</sub>	deri	1.0	270	45
1		dry	1.0	990	92
				1620	100
	ÇI Cy₃P−Ni−PCy₃				
2	Sy3 1 Sy3	dry	3.0	60	100
	CI Cy₃P−Ni−PCy₃			50	97
3	<b>5</b> ,3 5,3	dry	3.1	60	98
				70	99
	CI Cy <sub>3</sub> P-Ni-PCy <sub>3</sub>			42	97
4	Sy3. III 1 Sy3	dry	3.2	52	100
				32	100
	CI Cy <sub>3</sub> P-Ni-PCy <sub>3</sub>			40	92
5	Cy3: 11 1 Cy3	dry	3.7	50	95
				60	98
	CI Cy₃P−Ni−PCy₃			52	100
6	Cy <sub>3</sub> F-NI-FCy <sub>3</sub>	desi	4.7	70	97
O		dry		80	99
				90	99
	CI Cy <sub>3</sub> P-Ni-PCy <sub>3</sub>			60	94
7		dry	7.0	90	97
				168	99
	Cy₃P CI	dry	2.8	38	75
8	PCy <sub>3</sub>			58	85
o				78	95
				116	98
9	Cy₃P CI	desi	3.2	50	99
9	PCy <sub>3</sub>	dry	3.2	60	100/95 ( <b>3h</b> )
	CI Cy₃P−Ni−PCy₃	۸ -		58	85
10	Cy3: 11 1 Cy3	As received	1.0	78	95
		received		116	98
	CI Cy₃P−Ni−PCy₃			50	94
11	Cy3F NI FCy3	As	2.2	70	96
11		received	3.2	120	99
				160	100
	CI Cy <sub>3</sub> P-Ni-PCy <sub>3</sub>	٨		60	93
12	Cy <sub>3</sub> P-NI-PCy <sub>3</sub>	As	7.0	120	98
		received		180	99
<sup>a</sup> Conversion by GC, isolated yield, both as percentages.					

Table SI 3. Supporting Data for Table 3 with 0% PCy<sub>3</sub>

entry	catalyst	time (min)	convn/ yield (%)	time (min)	convn/ yield (%)	time (min)	convn/ yield (%)
	CI Cy <sub>3</sub> P-Ni-PCy <sub>3</sub>	42	98	50	94	50	97
1	Cy <sub>3</sub> P-Ni-PCy <sub>3</sub>	52	100	60	100	60	98
		-	-	70	100	70	99
		50	99	40	98	50	98
2	Cy₃P\ CI	60	100	50	99	60	99
2	PCy <sub>3</sub>	70	100/95	60	100	70	00
		70	(3h)	60	100	70	99
	QTs	47	97	50	100	50	100
3	Cy₃P−Ni−PCy₃	57	100	60	100	60	100
l		-	-	70	100	70	100
	Cv. D	47	98.0	50	99	50	98
4	Cy <sub>3</sub> P\ OTs	57	100	60	100	60	99
		-	-	70	100	70	100
		50	99	50	98	50	97
~	OMs Cy <sub>3</sub> P-Ni-PCy <sub>3</sub>	60	99	60	99	60	98
5		70	100/91		00	70	00
	<b>~ ~</b>	70	(3i)	70	99	70	99
		42	99	40	95	40	96
	Cy₃P∖ OMs	52	100/95	50	06	50	00
6	PCy <sub>3</sub>	52	(3j)	50	) 96	50	99
		62	100	60	98	60	100

<sup>&</sup>lt;sup>a</sup>Conversion by GC as percentages. Reactions were duplicated by three different chemists.

Table SI 4. Supporting Data for Table 3 with 5% PCy<sub>3</sub>

$$\begin{array}{c} \text{OSO}_2\text{NMe}_2 \\ \text{OMe} \\ \text{1} \end{array} \begin{array}{c} \text{MeOOC} \\ \text{2} \\ \text{0} \\ \text{2} \\ \text{OMe} \\ \text{S% Ni catalyst, 5% PCy}_3 \\ \text{K}_3\text{PO}_4(\text{H}_2\text{O})_{3.2}, \text{ THF, 23 °C} \end{array} \begin{array}{c} \text{OMe} \\ \text{3} \\ \text{OMe} \\ \text{3} \end{array}$$

entry	catalyst	time(min)	convn(%)	time(min)	convn(%)
	CI Cy <sub>3</sub> P-Ni-PCy <sub>3</sub>	30	97	40	96
1	Cy <sub>3</sub> P-NI-PCy <sub>3</sub>	40	99	50	97
		-	-	60	99
	Cv-P	37	99	40	99
2	Cy <sub>3</sub> P, Cl	47	100	50	99
		-	-	60	100
	OTs	43	98	40	100
3	Cy <sub>3</sub> P-Ni-PCy <sub>3</sub>	53	100	50	100
		-	-	60	100
	Cv B	-	-	40	99
4	Cy <sub>3</sub> P\ OTs	-	-	50	100
		-	-	60	100
	ОМs	-	-	40	96
5	Cy <sub>3</sub> P-Ni-PCy <sub>3</sub>	-	-	50	97
		-	-	60	99
	Cv D	-	-	40	99
6	Cy <sub>3</sub> P\ OMs	-	-	50	99
		-	-	60	100

<sup>a</sup>Conversion by GC as percentages. Reactions were duplicated by two different chemists.

Table SI 5. Supporting Data for Table 3 with 10% PCy<sub>3</sub>

ent	ry catalyst	time (min)	convn/ yield (%)	time (min)	convn( %)	time (min)	convn( %)
	CI Cy₃P−Ni−PCy₃	38	99	40	98	40	99
1	Cy <sub>3</sub> P-N <sub>1</sub> -PCy <sub>3</sub>	48	100	50	100	50	99
		-	-	60	100	60	100
	Cy₃P、 CI	37	99	40	97	40	99
2	Ni PCy <sub>3</sub>	47	100	50	99	60	100
		-	-	60	100	-	-
	ОТs	40	94	40	94	40	100
3	3 Cy <sub>3</sub> P-Ni-PCy <sub>3</sub>	50	100	50	99	50	100
	-	-	60	100	60	100	
	0.5	42	97	40	96	40	100
4	Cy <sub>3</sub> P, OTs	53	100	50	99	50	100
		-	-	60	100	60	100
		40	100/98	40	07	40	100
_	OMs Cy <sub>3</sub> P-Ni-PCy <sub>3</sub>	40	(3k)	40	97	40	100
5		50	100	50	98	50	100
	~ ~	-	-	60	99	60	100
	Cv-P	40	98	40	95	40	100
6	Cy <sub>3</sub> P, OMs	50	100	50	96	50	100
		-	-	60	98	60	100

<sup>a</sup>Conversion by GC, isolated yields, both as percentages. Reactions were duplicated by three different chemists.

Table SI 6. Supporting Data for Table 3 with 15%  $PCy_3$ 

entry	catalyst	time(min)	convn(%)
1	CI Cy₃P−Ni−PCy₃ I	30	99
1		40	100
2	Cy <sub>3</sub> P <sub>\</sub> Cl	34	98
2	PCy <sub>3</sub>	44	100
2	OTs Cy₃P−Ni−PCy₃	57	100
3		67	100
	Cy <sub>3</sub> P, OT-	45	97
4	Cy <sub>3</sub> P <sub>\</sub> OTs Ni PCy <sub>3</sub>	55	100
	<b>* *</b>	65	100

<sup>a</sup>Conversion by GC as percentages.

Table SI 7. Supporting Data for Table 3 with 20%  $PCy_3$ 

entry	catalyst	time(min)	convn(%)	time(min)	convn(%)
	ÇI Cy₃P−Ni−PCy₃	40	97/97 ( <b>3l</b> )	40	97
1		50	99	50	97
	* *	-	-	60	97
	CvaP	50	99	40	98
2	Cy <sub>3</sub> P, Cl Ni, Cl PCy <sub>3</sub>	60	100	50	98
		-	-	60	99
	OTs Cy <sub>3</sub> P-Ni-PCy <sub>3</sub>	40	100/99		
3	Cy <sub>3</sub> P-Ni-PCy <sub>3</sub>	P-Ni-PCy <sub>3</sub> 40	(3m)		
		50	100		
	CvoP.	40	100		
4	Cy <sub>3</sub> P, OTs	50	100/96		
		30	(3n)		
		40	100	30	100
5	OMs Cy <sub>3</sub> P-Ni-PCy <sub>3</sub>	50	100/98	40	100
		50	(3o)	40	100
	• •				
6	Cy <sub>3</sub> P, OMs	30	100	30	100
6	PCy <sub>3</sub>	40	100	40	100

<sup>a</sup>Conversion by GC as percentages. Reactions were duplicated by two different chemists.

**Table SI 8. Supporting Data for Table 4** 

entry	catalyst	$PCy_3$	time (min)	convn (%)
			40	97
		0	50	99
	ÇI		70	99/95 ( <b>3p</b> )
	CI Cy <sub>3</sub> P-Ni-PCy <sub>3</sub>		50	97
1		10	60	98
			70	99
			30	95
		20	40	99
			50	100
	CI Cy₃P−Ni−PCy₃		40	85
2	0	0	60	95
			70	97
			60	97
		0	70	99
	CI Cy₃P−Ni−PCy₃		80	99
	Cy₃P-Ni-PCy₃		50	98
3		10	60	99
			70	100
	~		40	96
		20	50	98
			60	99
			40	97
		0	50	99
			60	100
	CI Cy₃P−Ni−PCy₃		50	99
4	Cy3, 111 1 Cy3	10	60	100
			70	100
	•		40	96
		20	50	97
			60	99
<sup>a</sup> Conve	ersion was dete	ermined	by GC as percen	tages.

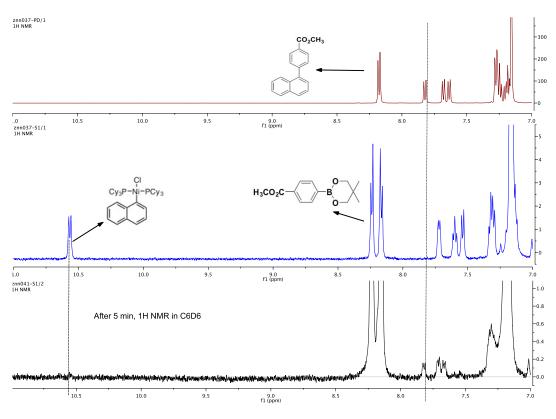
Table SI 9. Isolated Yields and Spectroscopy Data for Kinetics Experiments

entry	compound number	isolated yield (mg)	isolated yield (%)
1	3a	51.41	71
2	<b>3</b> b	64.45	89
3	3c	69.80	96
4	<b>3d</b>	68.97	95
5	<b>3</b> e	68.32	94
6	3f	36.56	50
7	<b>3</b> g	34.59	48
8	3h	69.19	95
9	3i	70.52	91
10	3ј	68.79	95
11	3k	71.09	98
12	31	70.52	97
13	3m	72.11	99
14	3n	69.42	96
15	30	71.06	98
16	<b>3</b> p	69.28	95

#### 4.5 Experiments to Study the Activation of Precatalyst 6

### Procedure for Monitoring the Activation of Precatalyst 6 by <sup>1</sup>H NMR

In an oven dried screw cap vial (4 mL) charged with a stirring bar were added methyl 4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)benzoate (78.15 mg, 0.315 mmol, 1.05 equiv), and  $K_3PO_4(H_2O)_n$  (191.02 mg, ~ 9 mmol, ~ 3 equiv), in a separate oven dried screw cap vial (4 mL) was added Ni<sup>II</sup>Cl(1-Naphthyl)(PCy<sub>3</sub>)<sub>2</sub> (11.72 mg, 0.015 mmol, 5% catalyst loading). The vials were brought into a nitrogen filled glove box (moisture level < 10 ppm) through three degassing cycles (5 min each). Distilled THF (1 mL) was added inside the glove box to the second vial to dissolve Ni<sup>II</sup>Cl(1-naphthyl)(PCy<sub>3</sub>)<sub>2</sub> and the solution was transferred to the first vial. The vial was capped and left stirring for 5 min. A sample (100  $\mu$ L) was taken and injected into an NMR tube. The tube was brought outside the glove box.  $C_6D_6$  (0.5 mL) was added outside the glove box and <sup>1</sup>H NMR was carried out right away. The spectra are shown in Figure SI 1.



**Figure SI 1.** Activation of precatalyst **6** monitored by <sup>1</sup>H NMR in C<sub>6</sub>D<sub>6</sub>.

## **Procedure for Model Reaction of Activation of Precatalyst 6**

In an oven dried screw cap vial (4 mL) charged with a stirring bar were added methyl 4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)benzoate (49.60 mg, 0.2 mmol, 2 equiv), Ni<sup>II</sup>Cl(1-naphthyl)(PCy<sub>3</sub>)<sub>2</sub> (78.31 mg, 0.1 mmol) and K<sub>3</sub>PO<sub>4</sub>(H<sub>2</sub>O)<sub>n</sub> (63.62 mg, ~ 0.3 mmol, ~ 3 equiv). The vial was brought into a nitrogen filled glove box (moisture level < 10 ppm) through three degassing cycles. Distilled THF (1 mL) was added inside the glove box, the vial was capped, and then left stirring for 12 h. The reaction mixture was dissolved in DCM, filtered and concentrated. The crude product was purified by silica gel column chromatography with DCM/hexanes = 1:1 eluent twice. Methyl 4-(naphthalen-1-yl)benzoate (16) was isolated to yield: 21.4 mg, 88%. White solid, m.p. 70 °C. ¹H NMR (500 MHz, Chloroform-*d*)  $\delta$  8.17 (dd, J = 8.3, 2.4 Hz, 2H), 7.90 (dd, J = 13.8, 8.2 Hz, 2H), 7.84 (d, J = 8.4 Hz, 1H), 7.63 - 7.48 (m, 4H), 7.44 (dd, J = 12.8, 7.2 Hz, 2H), 3.97 (s, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  167.2, 145.7, 139.3, 133.9, 131.38, 130.3, 129.7, 129.2, 128.5, 128.4, 127.1, 126.5, 126.1, 125.8, 125.5, 52.3. NMR spectra match with literature data. <sup>15</sup>

## 5. Characterization of the Reaction Products

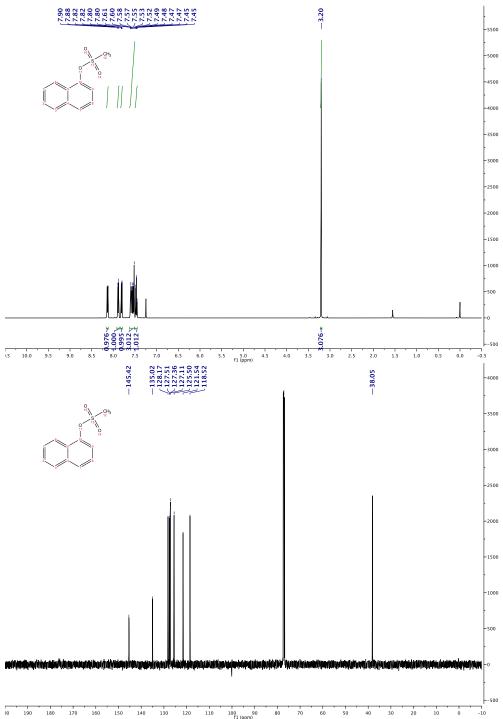
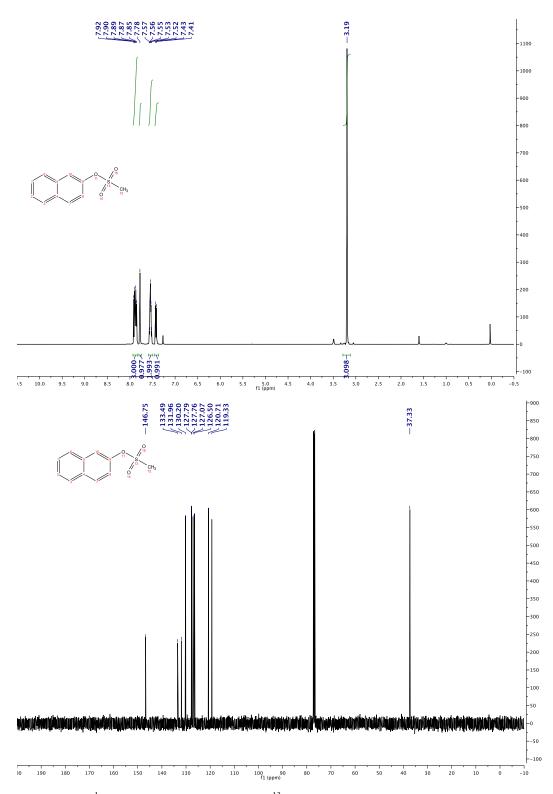
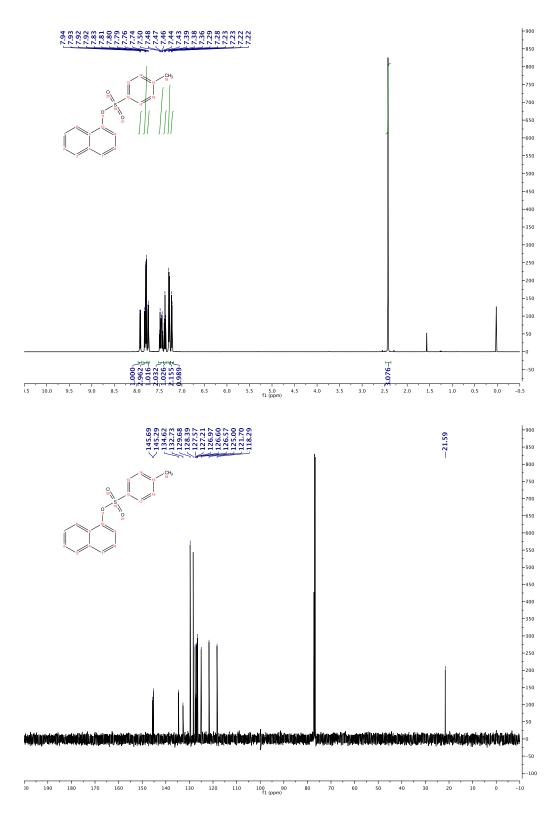


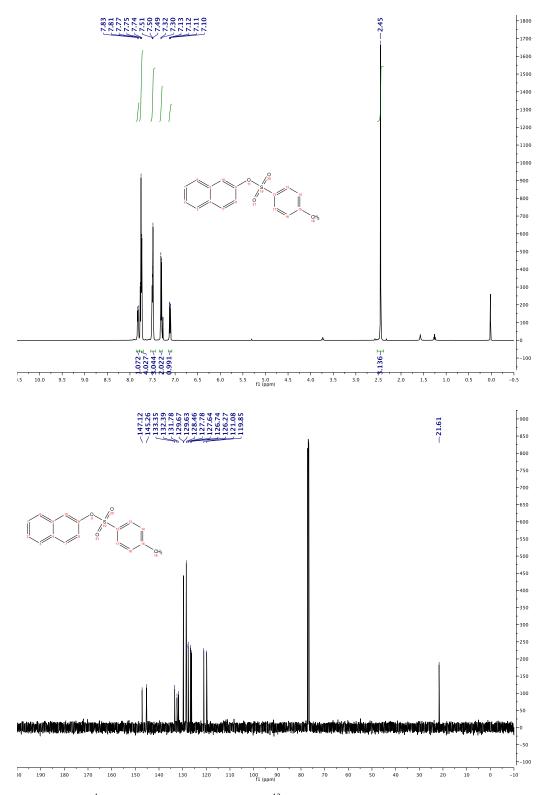
Figure SI 2. <sup>1</sup>H NMR (top, 500 MHz), and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of 1-naphthyl methanesulfonate in CDCl<sub>3</sub>.



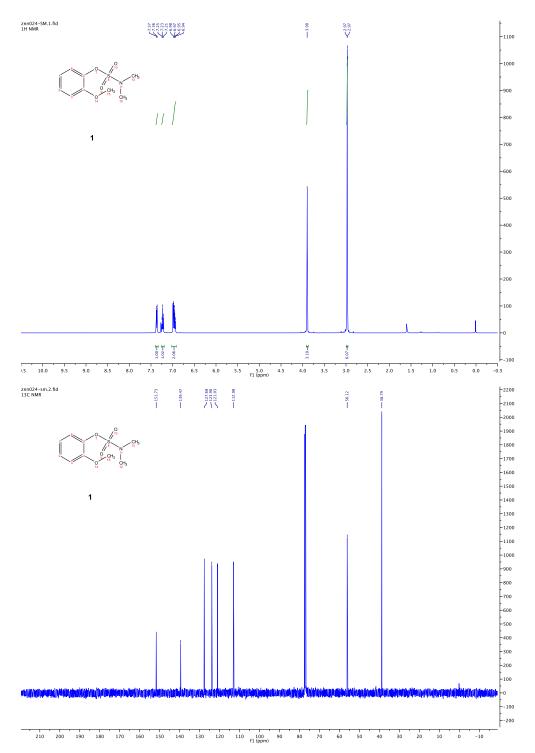
**Figure SI 3.** <sup>1</sup>H NMR (top, 500 MHz), and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of 2-naphthyl methanesulfonate in CDCl<sub>3</sub>.



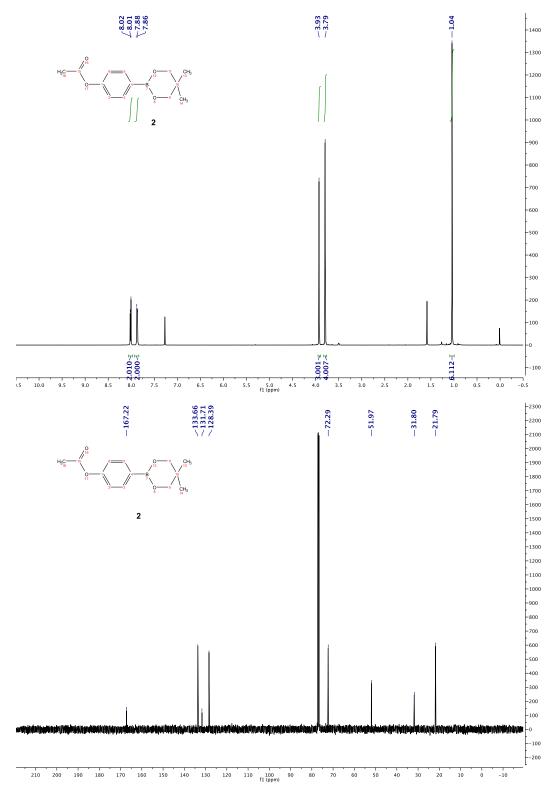
**Figure SI 4.** <sup>1</sup>H NMR (top, 500 MHz), and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of 1-naphthyl 4-methylbenzenesulfonate in CDCl<sub>3</sub>.



**Figure SI 5.** <sup>1</sup>H NMR (top, 500 MHz), and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of 2-naphthyl 4-methylbenzenesulfonate in CDCl<sub>3</sub>.



**Figure SI 6.** <sup>1</sup>H NMR (top, 500 MHz), and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of 2-methoxyphenyl dimethylsulfamate in CDCl<sub>3</sub>.



**Figure SI 7.** <sup>1</sup>H NMR (top, 500 MHz), and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of 4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)phenyl acetate in CDCl<sub>3</sub>.

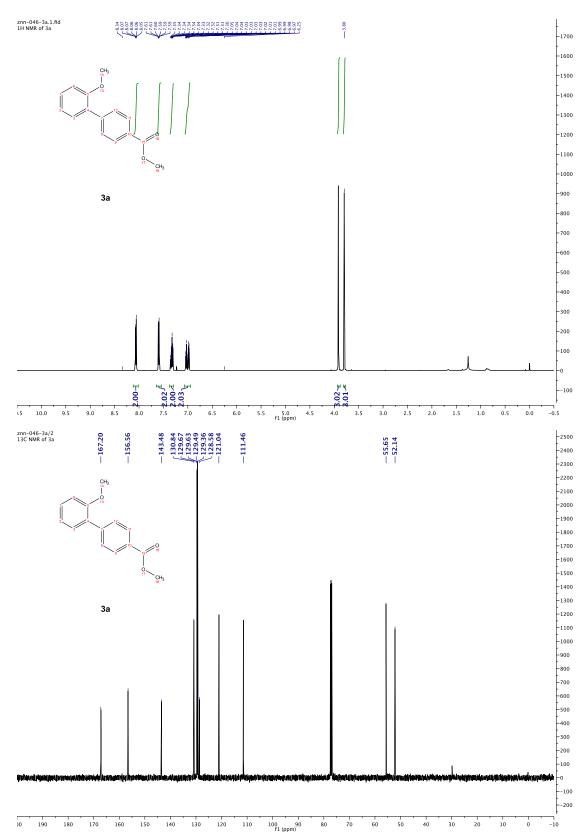


Figure SI 8. <sup>1</sup>H NMR (top, 500 MHz), and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of 3a in CDCl<sub>3</sub>.

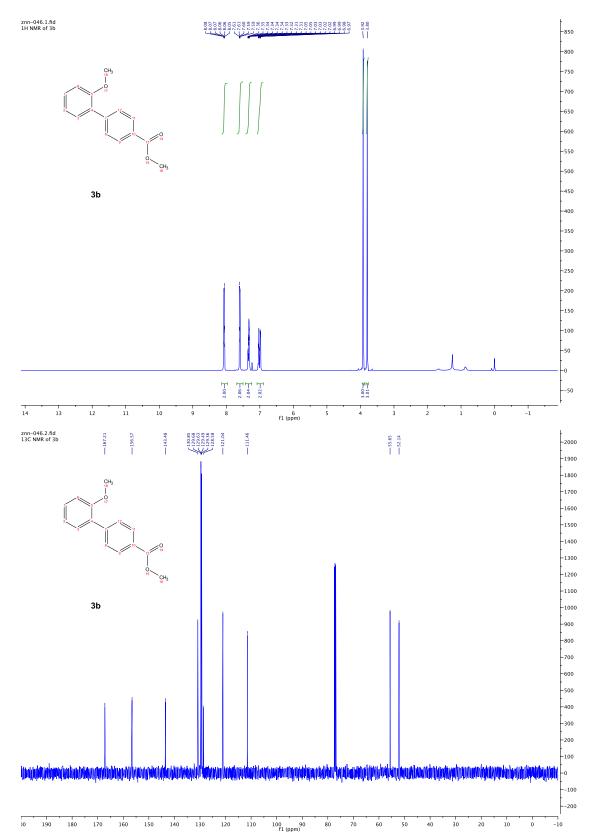


Figure SI 9. <sup>1</sup>H NMR (top, 500 MHz), and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of **3b** in CDCl<sub>3</sub>.

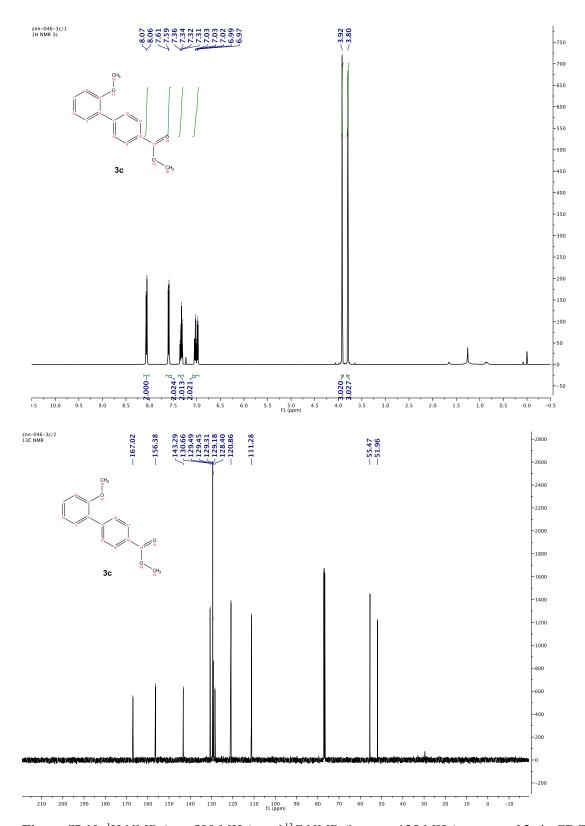
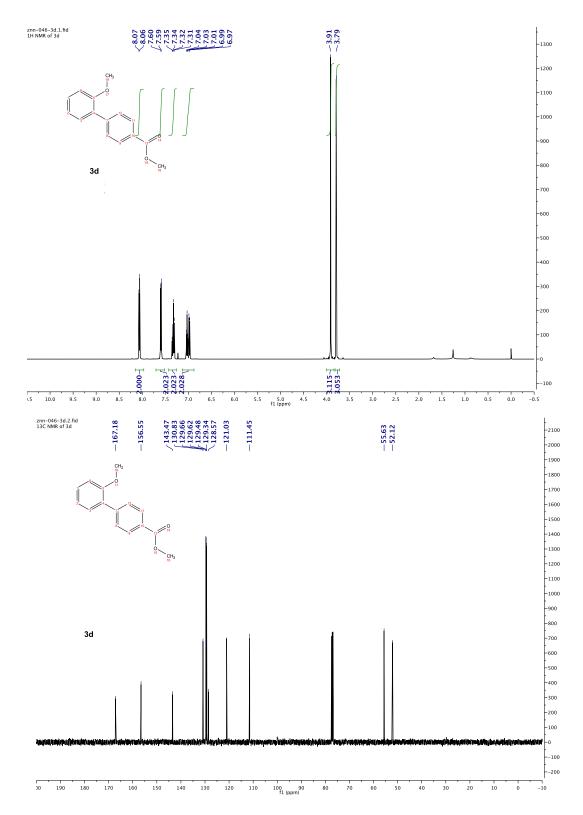


Figure SI 10. <sup>1</sup>H NMR (top, 500 MHz), and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of 3c in CDCl<sub>3</sub>.



**Figure SI 11.**  $^{1}$ H NMR (top, 500 MHz), and  $^{13}$ C NMR (bottom, 125 MHz) spectra of **3d** in CDCl<sub>3</sub>.

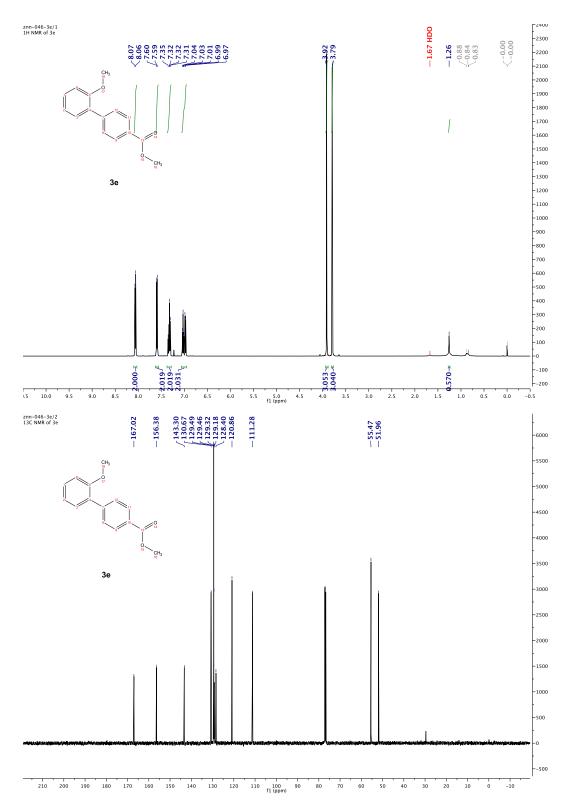


Figure SI 12. <sup>1</sup>H NMR (top, 500 MHz), and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of 3e in CDCl<sub>3</sub>.

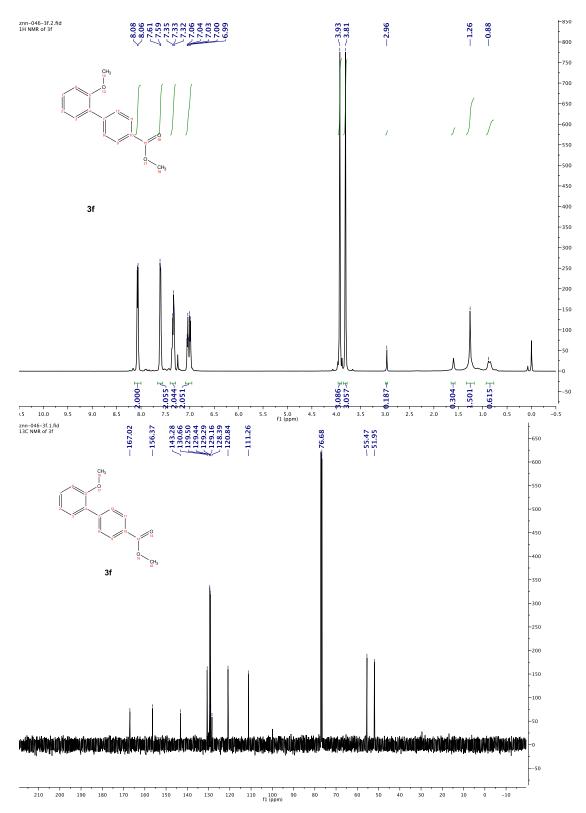
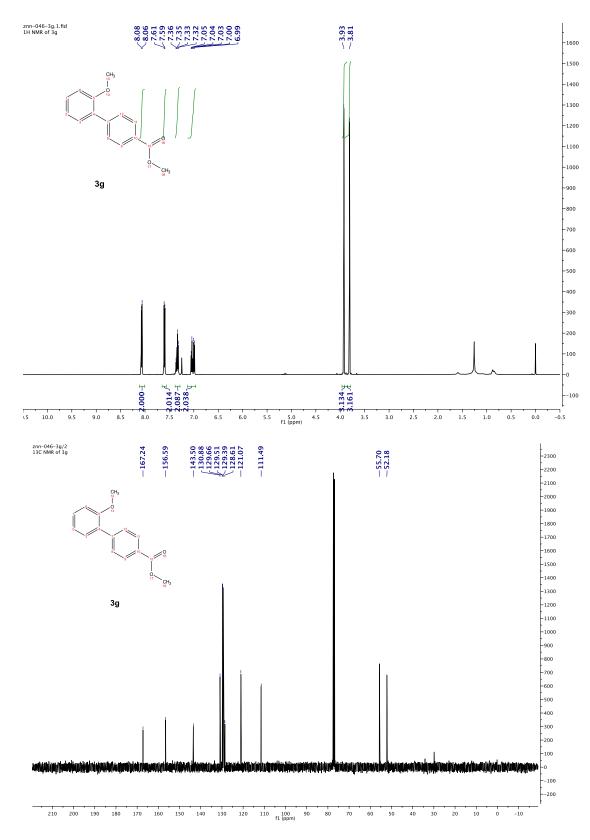
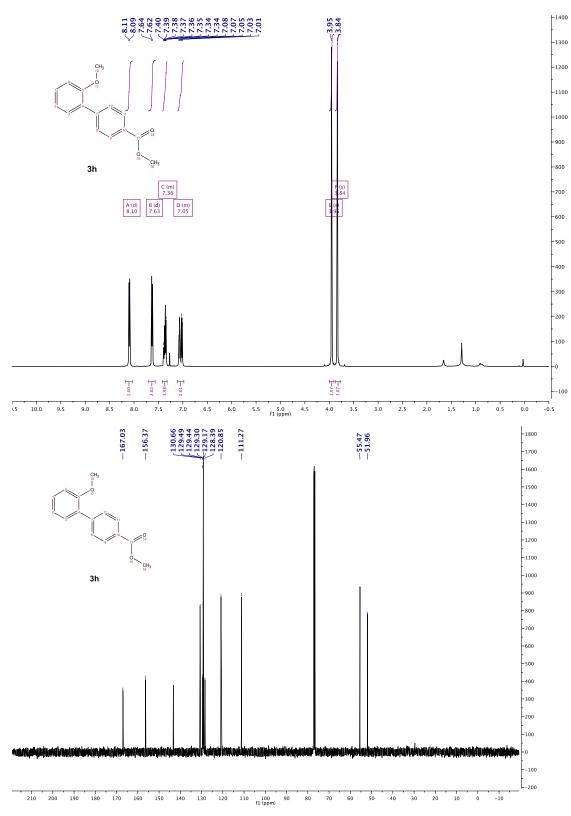


Figure SI 13. <sup>1</sup>H NMR (top, 500 MHz), and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of 3f in CDCl<sub>3</sub>.



**Figure SI 14.**  $^{1}$ H NMR (top, 500 MHz), and  $^{13}$ C NMR (bottom, 125 MHz) spectra of **3g** in CDCl<sub>3</sub>.



**Figure SI 15.** <sup>1</sup>H NMR (top, 500 MHz), and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of **3h** in CDCl<sub>3</sub>.

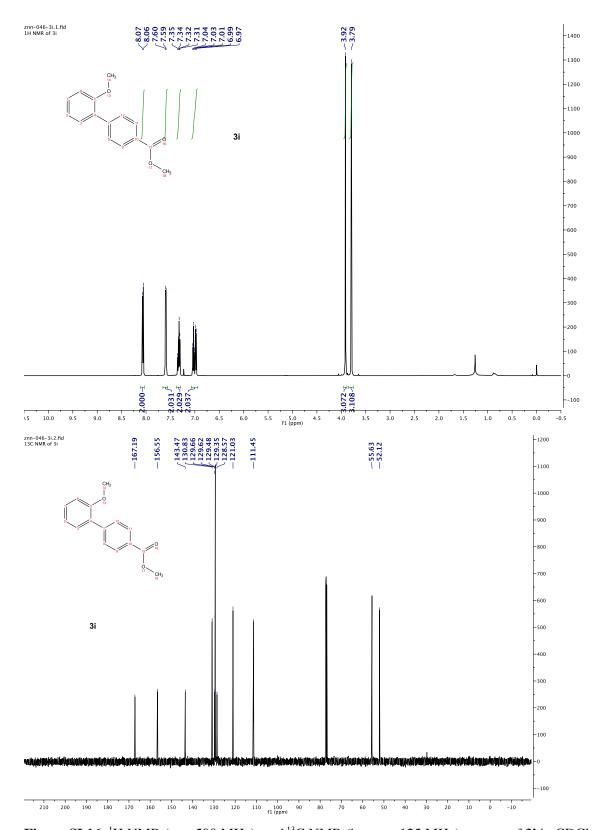


Figure SI 16. <sup>1</sup>H NMR (top, 500 MHz), and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of 3i in CDCl<sub>3</sub>.

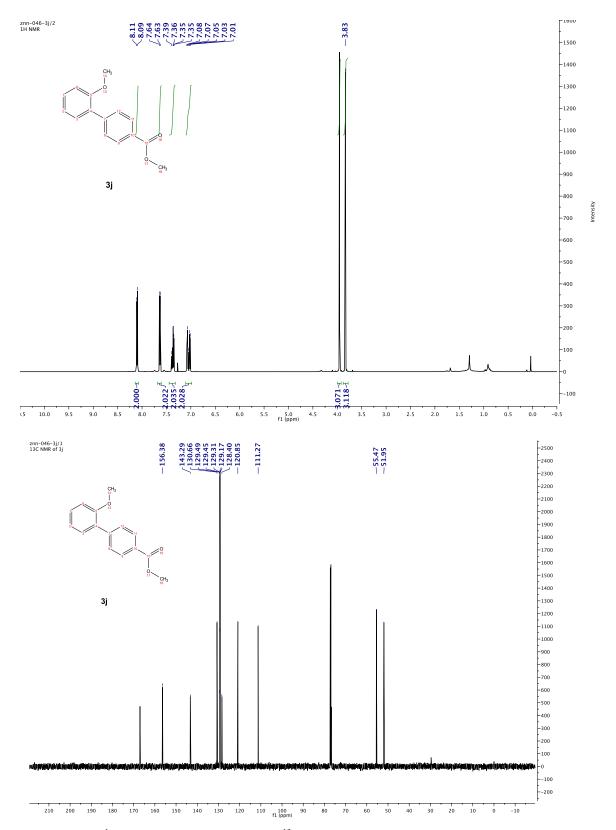
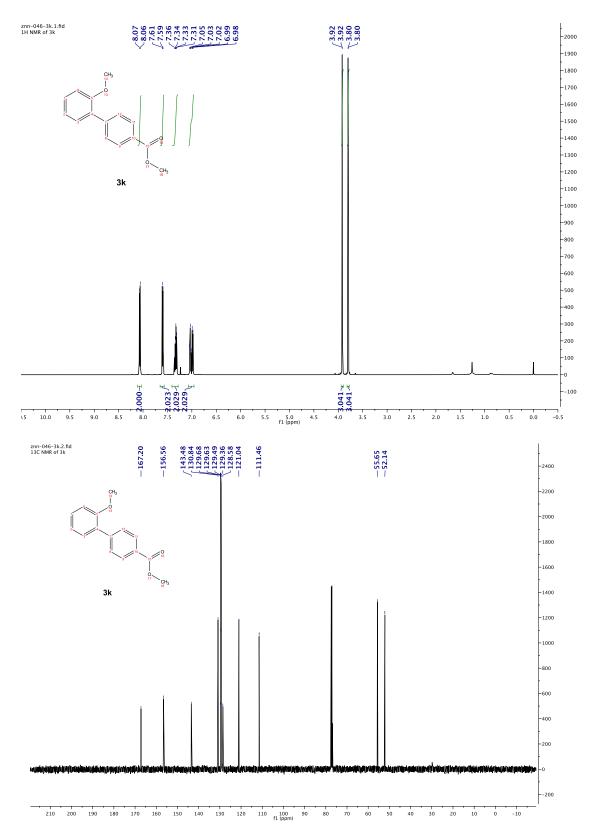


Figure SI 17. <sup>1</sup>H NMR (top, 500 MHz), and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of 3j in CDCl<sub>3</sub>.



**Figure SI 18.**  $^{1}$ H NMR (top, 500 MHz), and  $^{13}$ C NMR (bottom, 125 MHz) spectra of **3k** in CDCl<sub>3</sub>.

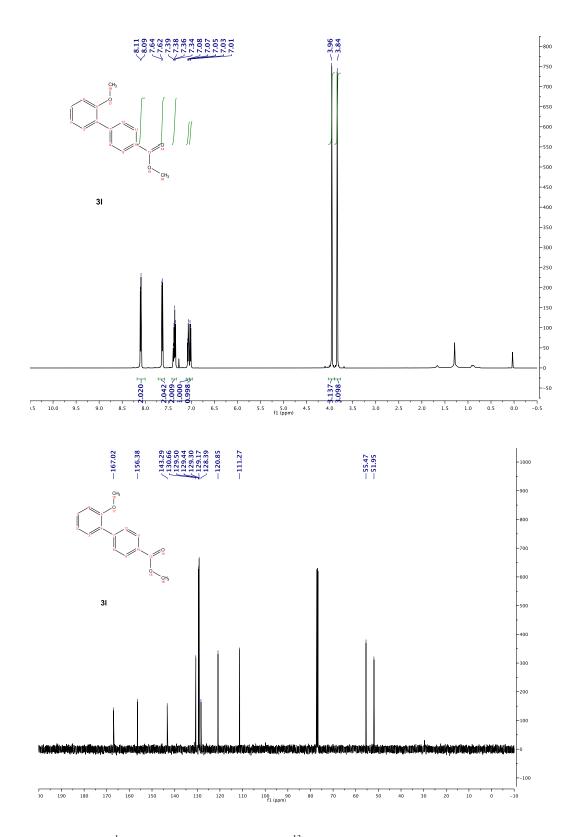
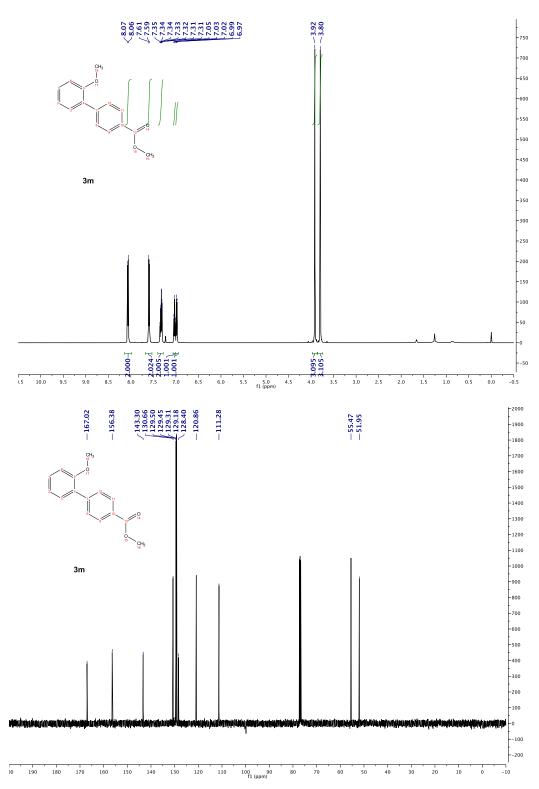
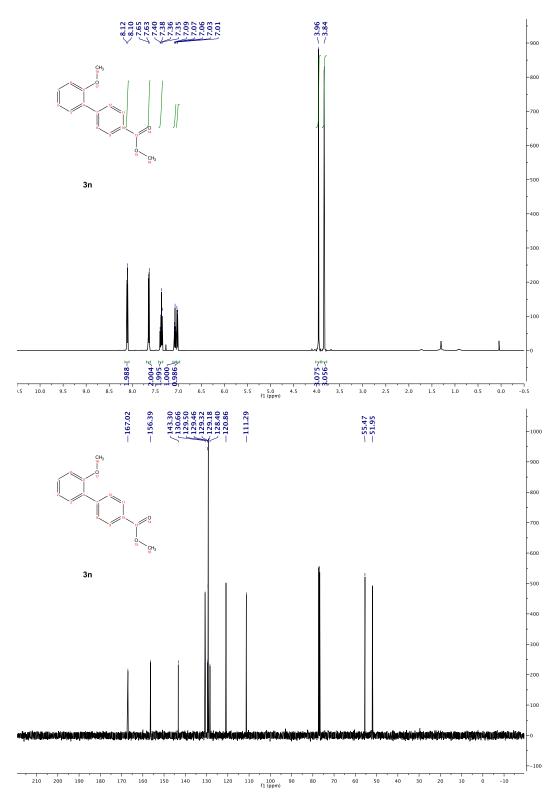


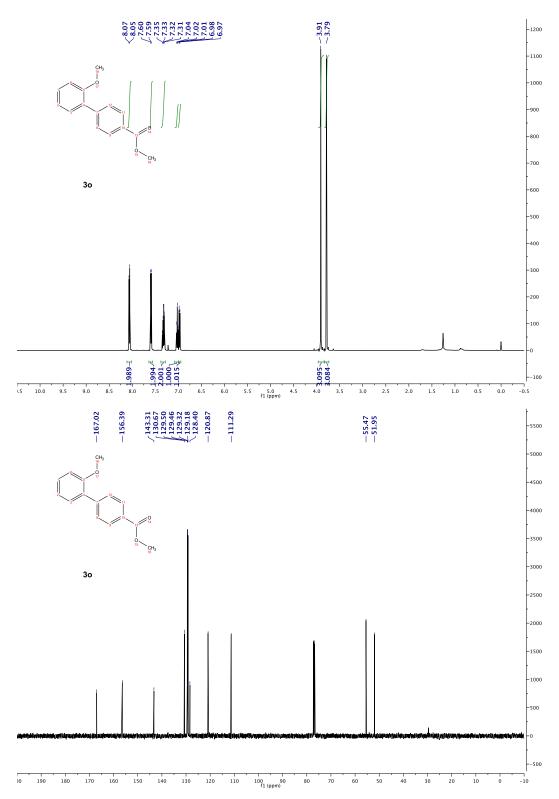
Figure SI 19. <sup>1</sup>H NMR (top, 500 MHz), and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of 3l in CDCl<sub>3</sub>.



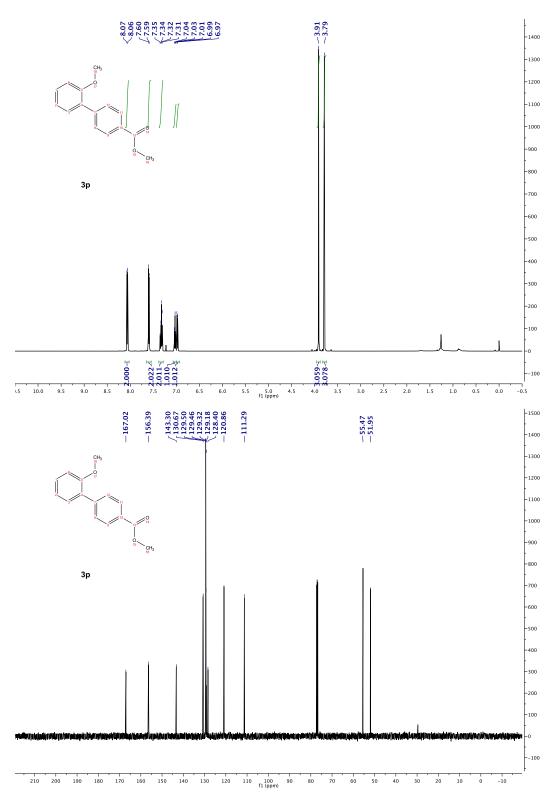
**Figure SI 20.** <sup>1</sup>H NMR (top, 500 MHz), and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of **3m** in CDCl<sub>3</sub>.



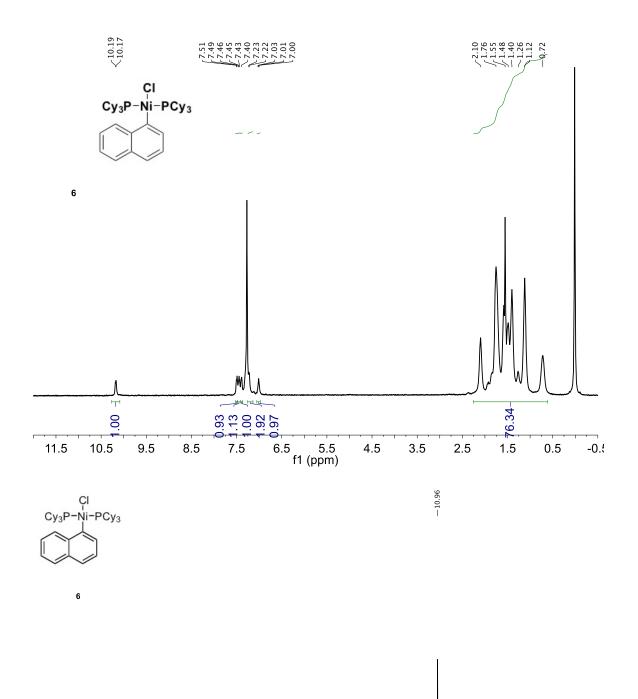
**Figure SI 21.**  $^{1}$ H NMR (top, 500 MHz), and  $^{13}$ C NMR (bottom, 125 MHz) spectra of **3n** in CDCl<sub>3</sub>.

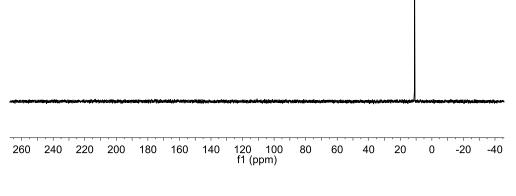


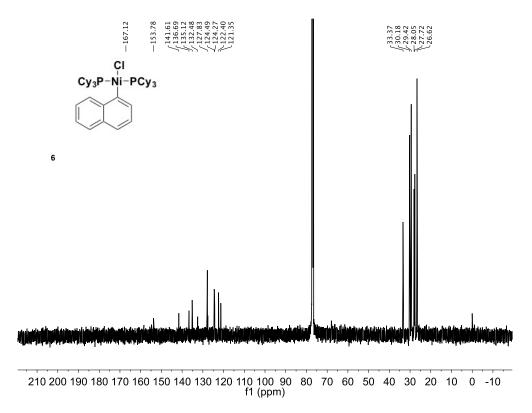
**Figure SI 22.**  $^{1}$ H NMR (top, 500 MHz), and  $^{13}$ C NMR (bottom, 125 MHz) spectra of **30** in CDCl<sub>3</sub>.



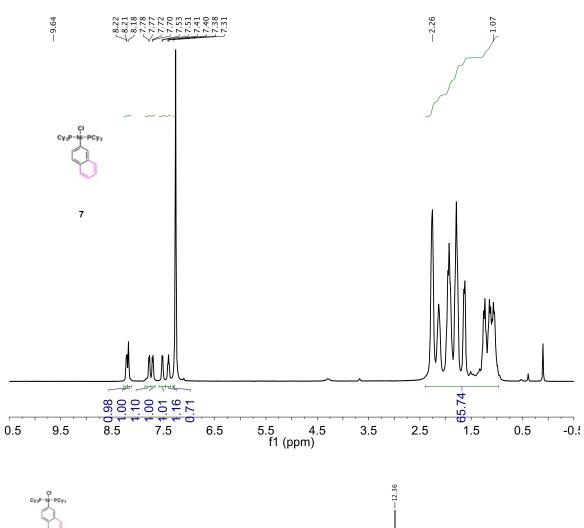
**Figure SI 23.**  $^{1}$ H NMR (top, 500 MHz), and  $^{13}$ C NMR (bottom, 125 MHz) spectra of **3p** in CDCl<sub>3</sub>.

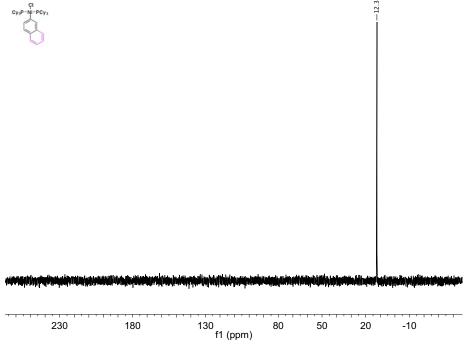


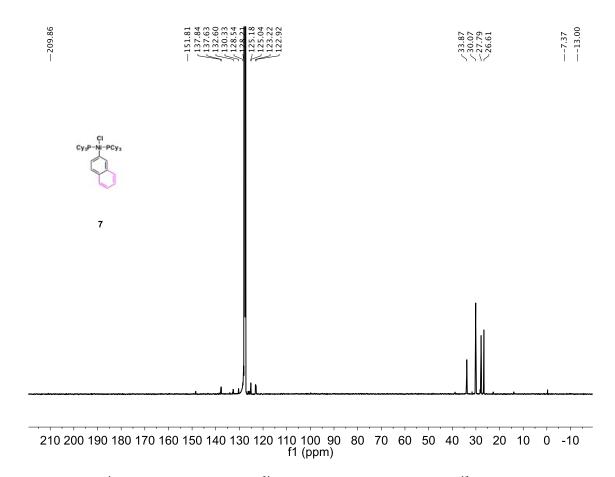




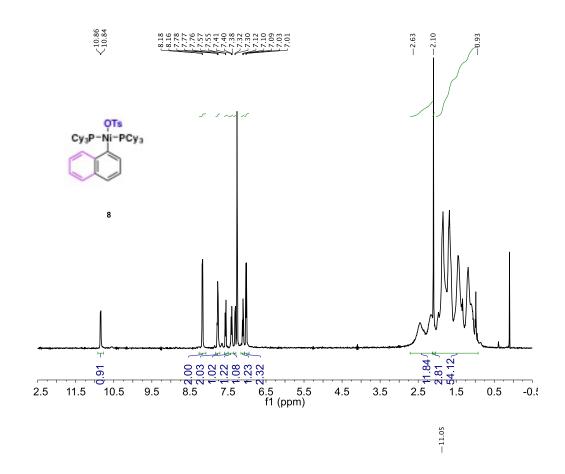
 $\label{eq:Figure SI 24.} \ ^{1}\text{H NMR (top, 500 MHz), } \ ^{3}\text{P NMR (middle, 203 MHz) and } \ ^{13}\text{C NMR (bottom, 125 MHz) spectra of Ni} \ ^{1}\text{Cl}(1\text{-naphthyl})(PCy_{3})_{2} \ \text{in } C_{6}D_{6}.$ 

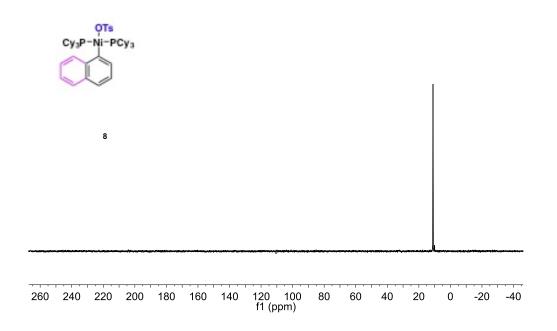


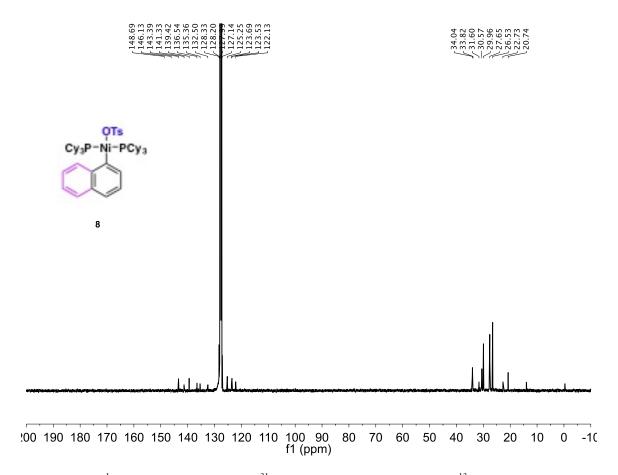




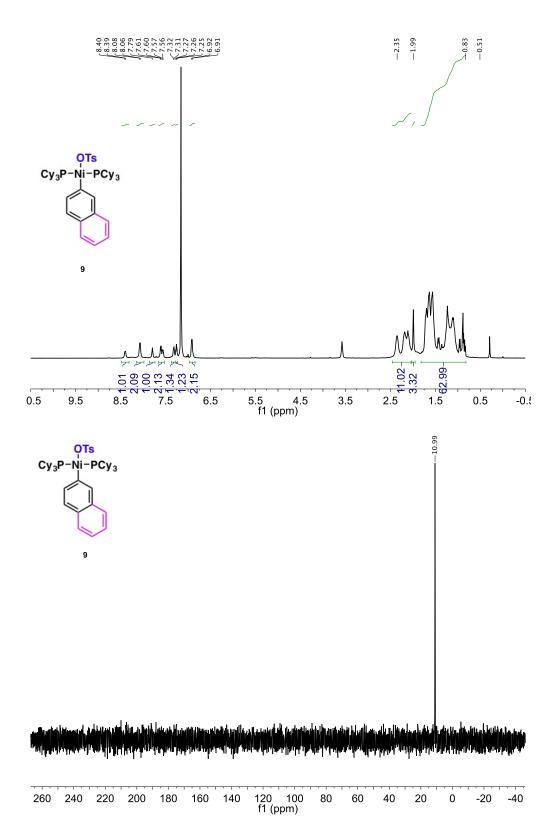
 $\label{eq:Figure SI 25.} \ ^{1}H\ NMR\ (top, 500\ MHz), \ ^{31}P\ NMR\ (middle, 203\ MHz)\ and \ ^{13}C\ NMR\ (bottom, 125\ MHz)\ spectra\ of\ Ni \ ^{11}Cl(2-naphthyl)(PCy_3)_2\ in\ C_6D_6.$ 

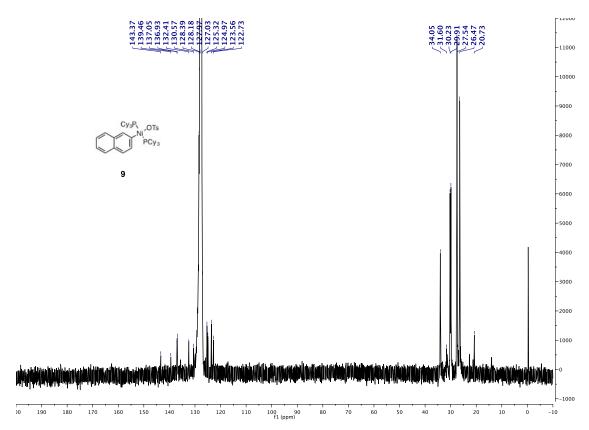




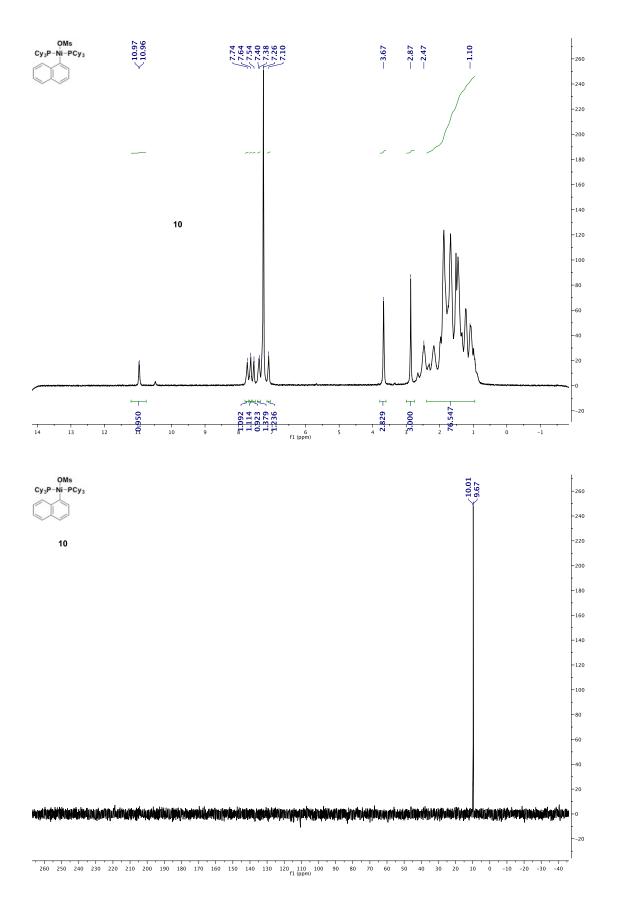


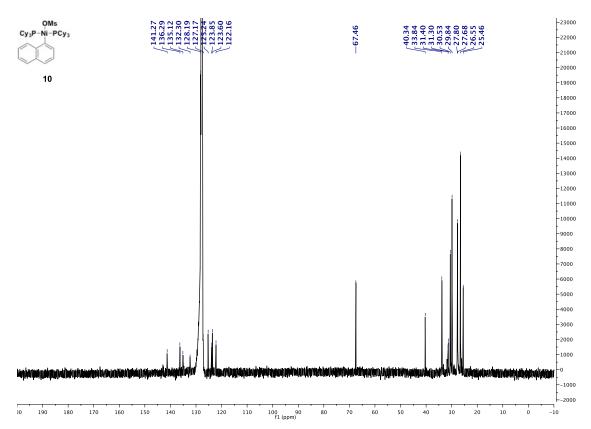
 $\label{eq:Figure SI 26.} ^{1}H \ NMR \ (top, 500 \ MHz), ^{31}P \ NMR \ (middle, 203 \ MHz) \ and ^{13}C \ NMR \ (bottom, 125 \ MHz) \ spectra of \ Ni ^{II}OTs (1-naphthyl) (PCy_3)_2 \ in \ C_6D_6.$ 



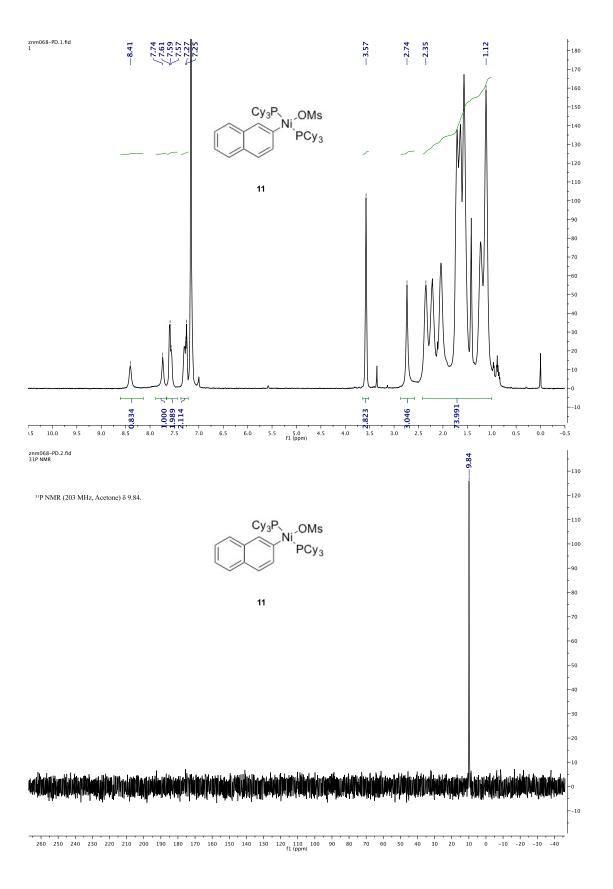


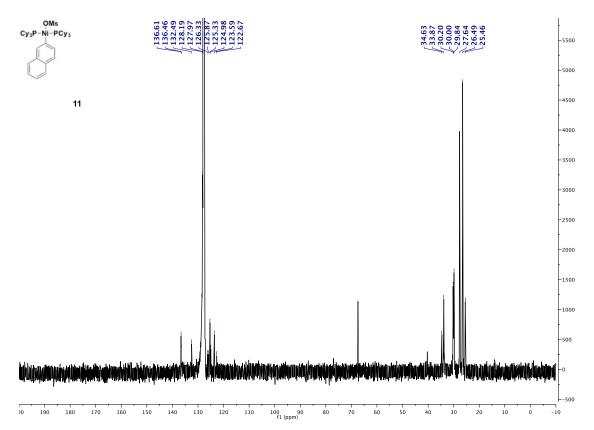
**Figure SI 27.** <sup>1</sup>H NMR (top, 500 MHz), <sup>31</sup>P NMR (middle, 203 MHz) and <sup>13</sup>C NMR (bottom, 126 MHz) spectra of Ni<sup>II</sup>OTs(2-naphthyl)(PCy<sub>3</sub>)<sub>2</sub> in  $C_6D_6$ .



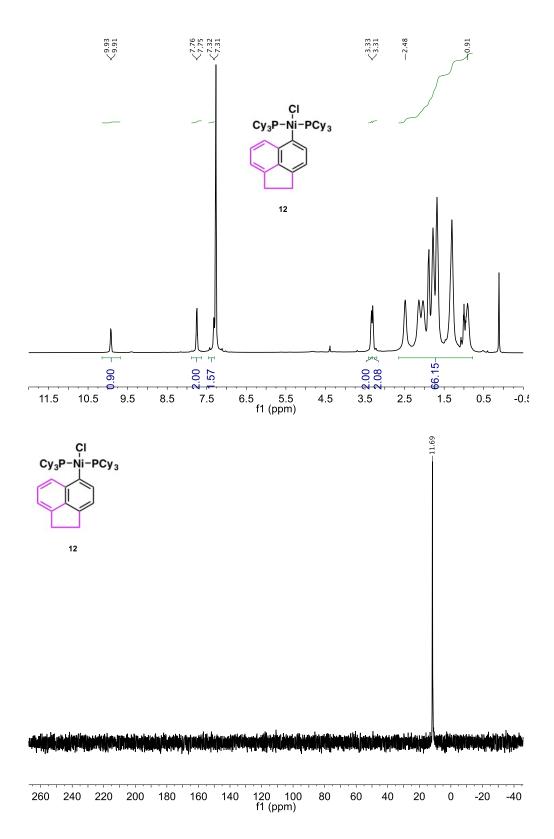


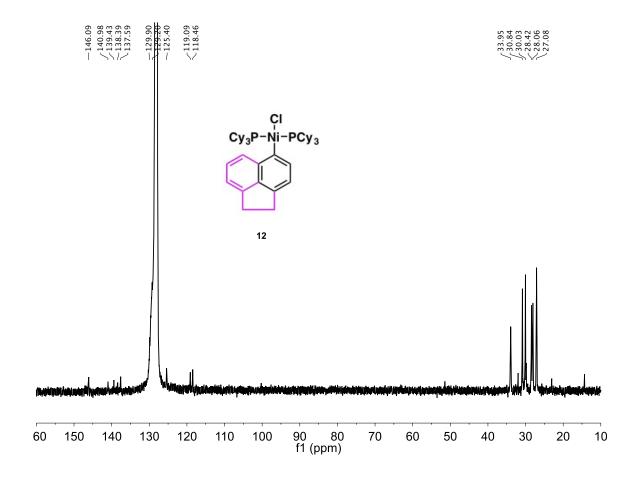
**Figure SI 28.** <sup>1</sup>H NMR (top, 500 MHz), and <sup>31</sup>P NMR (bottom, 203 MHz) spectra of Ni<sup>II</sup>OMs(1-naphthyl)(PCy<sub>3</sub>)<sub>2</sub> in  $C_6D_6$ .



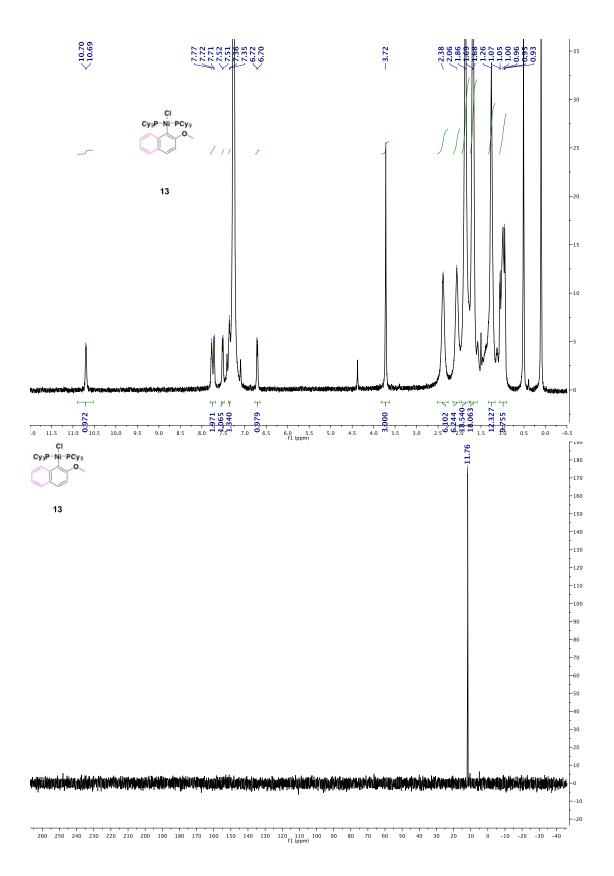


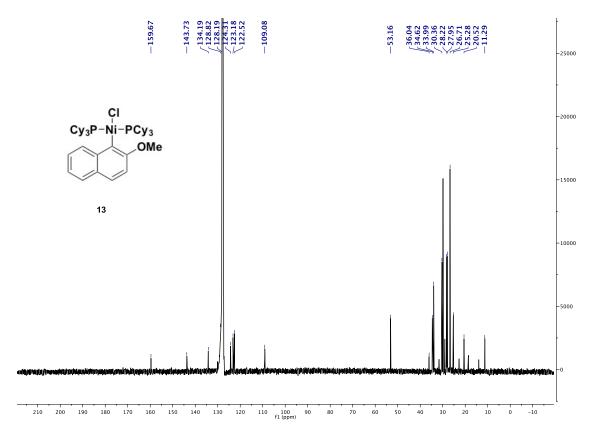
**Figure SI 29.**  $^{1}$ H NMR (top, 500 MHz), and  $^{31}$ P NMR (bottom, 203 MHz) spectra of Ni<sup>II</sup>OMs(2-naphthyl)(PCy<sub>3</sub>)<sub>2</sub> in  $C_6D_6$ .



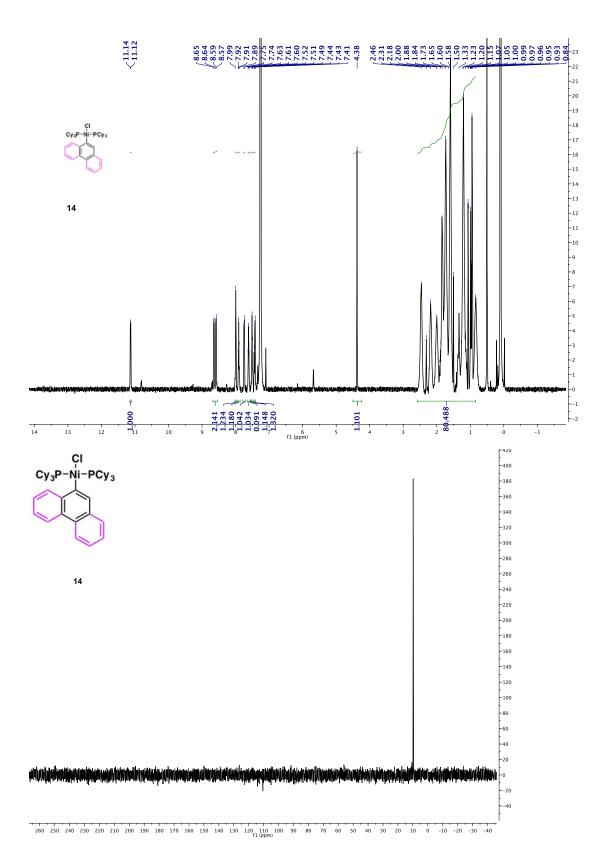


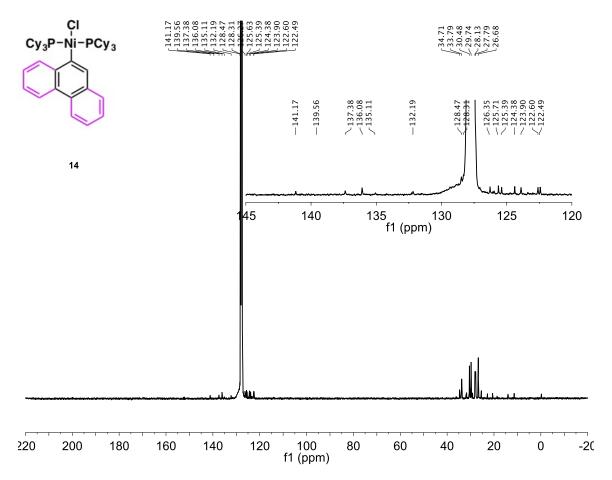
**Figure SI 30.** <sup>1</sup>H NMR (top, 500 MHz), <sup>31</sup>P NMR (middle, 203 MHz) and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of Ni<sup>II</sup>Cl(5-acenaphthyl)(PCy<sub>3</sub>)<sub>2</sub> in CDCl<sub>3</sub>.



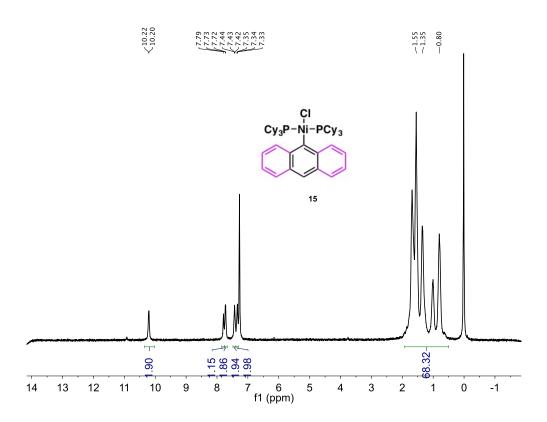


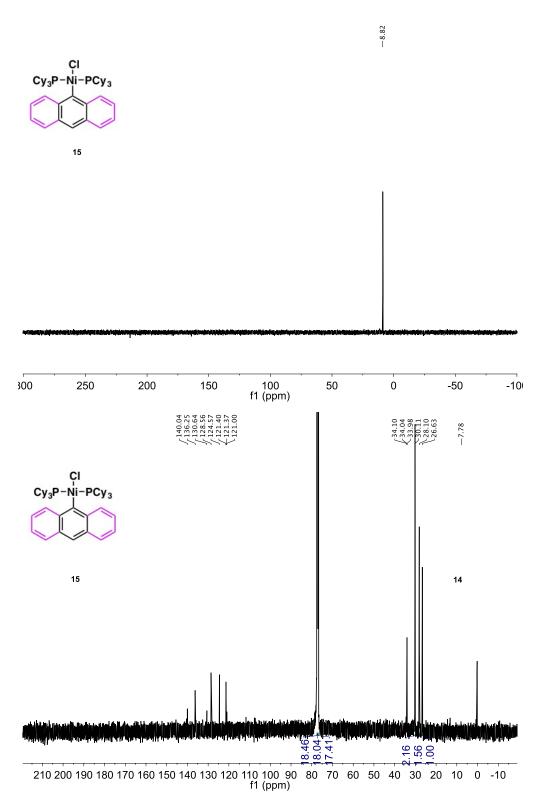
 $\label{eq:Figure SI 31.} \ ^{1}H\ NMR\ (top, 360\ MHz), \ ^{31}P\ NMR\ (middle, 203\ MHz)\ and \ ^{13}C\ NMR\ (bottom, 91\ MHz)\ spectra\ of\ Ni \ ^{11}Cl(2-OMeNaphthyl)(PCy_3)_2\ in\ C_6D_6.$ 



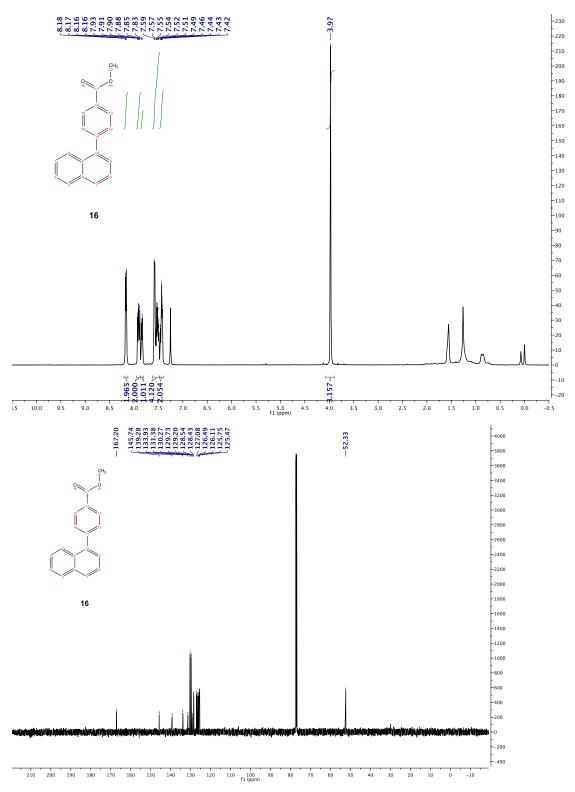


**Figure SI 32.** <sup>1</sup>H NMR (top, 360 MHz), <sup>31</sup>P NMR (middle, 203 MHz) and <sup>13</sup>C NMR (bottom, 91 MHz) spectra of Ni<sup>II</sup>Cl(9-phenathrenyl)(PCy<sub>3</sub>)<sub>2</sub> in  $C_6D_6$  and CDCl<sub>3</sub>.





**Figure SI 33.** <sup>1</sup>H NMR (top, 500 MHz), <sup>31</sup>P NMR (middle, 203 MHz) and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of Ni<sup>II</sup>Cl(9-anthracyl)(PCy<sub>3</sub>)<sub>2</sub> in CDCl<sub>3</sub>.



**Figure SI 34.** <sup>1</sup>H NMR (top, 500 MHz), <sup>31</sup>P NMR (middle, 203 MHz) and <sup>13</sup>C NMR (bottom, 125 MHz) spectra of methyl 4-(naphthalen-1-yl)benzoate in CDCl<sub>3</sub>.

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