Supporting Information

Simple Amine-Directed meta-Selective C-H Arylation via Pd/Norbornene Catalysis

Zhe Dong¹, Jianchun Wang¹, Guangbin Dong^{1*}

Table of Contents

I.	Materials and optimization tables		
II.	Synthesis of benzylamine derivatives and standard products		
III.	meta-Selective C-H activation protocol and conversion of the	S8	
	directing group		
IV.	Characterization of products	S10	
V.	¹ H- and ¹³ C-NMR spectra	S21	
VI.	References	S56	

¹Department of Chemistry, University of Texas at Austin, Austin, TX 78712, United States

^{*}To whom correspondence should be addressed at e-mail: gbdong@cm.utexas.edu

I. Materials and methods

Unless stated otherwise, all reactions were run in vials sealed with PTFE lined caps, purchased from Qorpak. Chlorobenzene (99% HPLC level) was directly used as received from Arcos. Triphenylarsine (AsPh₃, 99%), Copper acetate monohydrate (99%) and palladium acetate (98%) were purchased from Strem and used as received. Cesium acetate (CsOAc) and lithium acetate dihydrate were purchased from Alfa and used as received. All commercially available substrates were used without further purification. Thin layer chromatography (TLC) analysis was run on silica gel plates purchased from EMD Chemical (silica gel 60, F254). Gas chromatography (GC) data was obtained from Agilent 7820A GC system, equipped with Agilent 19091J-413 column and a FID detector. GC yield of 3a and 3a' was determined using standard curves with dodecane as internal standard. Mass spectra were recorded on an Autospec or Agilent 6150. Accurate masses from high-resolution mass spectra were reported for the molecular ion [M+Na]⁺, [M]⁺ or [M+H]⁺. ¹H NMR and ¹³C NMR spectra were recorded on a Varian Gemini (400 MHz for ¹H, 100 MHz for ¹³C). Chemical shifts are reported as parts per million (ppm) using residual solvent signals as internal standard (CDCl₃, $\delta = 7.26$ ppm for ¹H NMR, $\delta = 77.00$ ppm for ¹³C NMR; DMSO-d₆, $\delta = 2.50$ ppm for ¹H NMR, $\delta = 39.50$ ppm for ¹³C NMR; MeCN-d₃, $\delta = 1.94$ ppm for ¹H NMR, $\delta = 1.32$, 118.26 ppm for ¹³C NMR). Data for ¹H NMR were presented as following: chemical shifts (δ, ppm), multiplicity (br = broad, s = singlet, d = doublet, t = triplet, q = quartet, dd = doublet of doublets, tt = triplet of triplets, td = triplet of doublets, m = multiplet), coupling constant (Hz), and integration. The chemical shifts of peaks found were reported for ¹³NMR spectra. Infrared spectra were obtained from a Nicolet iS5 FTIR spectrometer.

General procedure for reaction-condition screening

The reaction was run at a 0.2 mmol scale based on the limiting reagent. A 4 mL vial was charged with palladium salt, ligand, additives, benzylamine derivative, aryl iodide, chlorobenzene (1.8 mL) and HOAc. The vial was sealed with a PTFE lined cap and heated in a pie-block at 100 °C for 36 hours under stirring. Then the mixture was allowed to cool to room temperature. The saturated potassium carbonate aqueous solution (2 mL) was carefully added to the vial to make the whole solution basic. The vial were kept stirring at room temperature for 15 min before the solid was filtered. The filter cake was further washed with ethyl acetate (containing 8% triethylamine), filtrate were collected together with the aqueous phase. The appropriate amount of dodecane (~10 mg) was added as the internal standard to the filtrate. The mixture was stirred for an additional 5 min. The filtrate was directly used for GC analysis.

GC instrument conditions: inlet temperature: 250 °C, detector temperature: 300 °C, hydrogen flow: 40 mL/min, air flow: 400 mL/min, column + makeup flow: 30 mL/min. Method: 50 °C hold for 0 min, followed by a temperature increase of 10°C/min to 320 °C, hold 0 min (total run time: 27 min). Yields of the products and byproducts are calculated using standard curves with dodecane as the internal standard. Full details of the control reactions are listed below (Tables S1-S4).

Table S1. Full details of salts effect:^a

Additives	yield of 3a	yield of 3a'	Total yield
None	7%	53%	60%
50 mol% Cu(OAc) ₂ •H ₂ O	8%	48%	54%
50 mol% LiOAc•2H ₂ O	10%	54%	64%
50 mol% Cu(OAc) ₂ •H ₂ O and 50 mol% LiOAc•2H ₂ O	9%	60%	69%
KOAc instead of CsOAc	8%	46%	54%
NaOAc instead of CsOAc	9%	42%	51%
50 mol% of Zn(OAc) ₂	10%	47%	57%
None 130°C 18h	13%	47%	60%

^aAll yields were determined by GC using dodecane as the internal standard.

Table S2. Selected ligand and solvent effect: a

Change from above conditions	yield of 3a	yield of 3a'	Total yield
None	2%	60%	62%
tAmyOH instead of PhCI	15.5%	26.5%	42%
TFE instead of PhCI	0%	0%	0%
PPh ₃ instead of AsPh ₃	15%	12%	27%
SPhos instead of AsPh ₃	13%	12%	25%

^aAll yields were determined by GC using dodecane as the internal standard.

Table S3. Acetic acid equivalent effect: a

^aAll yields were determined by GC using dodecane as the internal standard.

Table S4. The salts equivalent effect: a

100 C, 3611	yield of 3a	yield of 3a'	Total yield
No Cu(OAc) ₂ •H ₂ O and LiOAc•2H ₂	O 4%	54%	58%
No Cu(OAc) ₂ •H ₂ O No LiOAc•2H ₂ O with 1.0 equiv Cs	OAc 4%	48%	33%
50 mol% Cu(OAc) ₂ •H ₂ O 50 mol% LiOAc•2H ₂ O	2%	67%	69%
50 mol% Cu(OAc) ₂ •H ₂ O No LiOAc•2H ₂ O	2%	63%	65%
No Cu(OAc) ₂ •H ₂ O 50 mol% LiOAc•2H ₂ O	6%	53%	59%
100 mol% Cu(OAc) ₂ •H ₂ O 50 mol% LiOAc•2H ₂ O	2%	66%	68%
50 mol% Cu(OAc) ₂ •H ₂ O 100 mol% LiOAc•2H ₂ O	2%	72%	74%
100 mol% Cu(OAc) ₂ •H ₂ O 100 mol% LiOAc•2H ₂ O	1%	70%	71%
50 mol% Cu(OAc) ₂ •H ₂ O 50 mol% LiOAc•2H ₂ O Under N ₂	15%	43%	58%

^aAll yields were determined by GC using dodecane as the internal standard.

II. Synthesis of benzylamine derivatives

Most benzylamine substrates used in this work are known compounds, and substrates 11 and 1m. 1a, 1d and 1j are commercially available. The known benzylamine were all synthesized from corresponding benzyl halide and dimethyl amine. We used a slight modified protocol.¹

To a solution of benzylic bromide (1.0 equiv, 10 mmol) in ether (1 M) was added 50 wt.% aqueous dimethylamine solution (5 equiv) at room temperature. After stirring for 12 hours at the same temperature, the resulting mixture was transferred to a separatory funnel. The aqueous phase was removed and the organic phase was extracted with 10 wt.% aqueous citric acid solution twice. The combined aqueous extracts were treated with sodium hydroxide to keep pH slightly above 12. The mixture was extracted with ether and the combined organic extracts were washed with brine. The solution was dried over mageniusm sulfate and concentrated. The crude product were found pure based on NMR, was used without further purification. Further vacuum distillation can be adapted to remove all the colorful impurity.

11 and 1m were synthesized from corresponding benzaldehyde through reductive amination:

To solution of 1-tosyl-1*H*-pyrrole-2-carbaldehyde² or 2-chloroisonicotinaldehyde (10 mmol) in 20 mL tetrahydrofuran was added 2.0M dimethylamine solution in THF (10 ml, 20 mmol), acetic acid (1.2 g, 20 mmol). After being cooled in an ice bath for 5 min, sodium triacetoxyborohydride (4.2 g, 20 mmol) was added portion wise with stirring. The solution was further stirred at room temperature overnight. After monitored by TLC till full conversion, the solution was concentrated under vacuum. The residue was then dissolved by diethyl ether (20 mL) and extracted by 10% citric acid solution (30 mL x 3). The aqueous layer was neutralized by adding potassium hydroxide at zero degree and extracted by diethyl ether (30 mL x 3). The Combined organic layer was dried over magnesium sulfate and concertrated under vacuum to give the desired product 11 as a pale brown solid or 1m as an oil which were analytically pure and directly used as in the C-H activation reaction.

General Procedure for synthesis ortho and para products.

$$\begin{array}{c} \text{HNMe}_2 \text{ HCI} \\ \text{NaOAc} \\ \text{HOAc} \\ \text{S1} \end{array} \\ \begin{array}{c} \text{NaBH(OAc)}_3 \\ \text{THF r.t. overnight} \end{array} \\ \begin{array}{c} \text{Me} \\ \text{N} \\ \text{Me} \\ \text{N} \\ \text{CO}_2 \text{Me} \end{array} \\ \end{array} \\ \begin{array}{c} \text{Me} \\ \text{N} \\ \text{Ne} \\ \text{N$$

To solution of aldehyde S1³ or S3⁴ (0.55 g, 2.29 mmol) in 20 mL tetrahydrofuran was added dimethylamine hydrochloride (0.37g, 4.58 mmol), sodium acetate (0.18 g, 3.66 mmol), acetic acid (0.07 g, 1.15 mmol). After being cooled in an ice bath for 5 minutes, sodium triacetoxyborohydride (1.07 g, 5.04 mmol) was added portionwise with stirring. The solution was further stirred at room temperature overnight. After monitored by TLC till a full conversion, the solution was concentrated under vacuum. The residue was then dissolved by diethyl ether (20 mL) and extracted by 10% citric acid solution (30 mL x 3). The aqueous layer was neutralized by adding potassium hydroxide at zero degree and extracted by diethyl ether (30 mL x 3). The combined organic layer was dried over magnesium sulfate and concertrated under vacuum to give the desired product S2 or S4 as an oil which were analytically pure and directly used as the standard for GC-analysis.

III.General Procedure for the meta-Arylation of benzylamine derivative with simple aryl iodide.

Unless stated otherwise, an 8 mL vial was charged with benzyl amine (0.4 mmol, 1.0 equiv), aryl iodide (1.6 mmol, 4.0 equiv), Pd(OAc)₂ (9.0 mg, 0.1 equiv), norbornene (75.7 mg, 2.0 equiv), triphenylarsine (30.6 mg, 0.25 equiv), AgOAc (300 mg, 4.5 equiv), Cu(OAc)₂ H₂O (40.2 mg, 0.5 equiv.), LiOAc 2H₂O (40.1 mg, 1.0 equiv), CsOAc (235 mg, 1.2 equiv), HOAc (360 μL, 15.0 equiv) and PhCl (3.5 mL). The vial was sealed with a PTFE lined cap and heated in a pie-block at 100 °C for 36 hours under stirring. The mixture was allowed to cool to room temperature. The saturated potassium carbonate aqueous solution (4 mL) was carefully added to the vial to make the whole solution alkaline. The vial was kept stirring at room temperature for 15 min before the solid was filtered. The filter cake was further washed with ethyl acetate (containing 8% triethylamine, roughly about 20 mL), the filtrate were collected together with aqueous phase. The aqueous phase was isolated and further extracted with dichloromethane three times (8 mL x 3). The organic phase were combined and dried with magnesium sulfate. The magnesium sulfate were removed by filtration and solution were concentrated under vacuum, the residue was purified by chromatography on silica gel.

For other substrates in the Table 2 and Table 3, the equivalent of the aryl iodide, the equivalent of AgOAc and the reaction temperature were adopted as indicated in the manuscript. The rest of procedure was exactly the same as what we described here.

Conversion of the amine DG to other FGs:

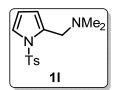
Chlorination:

In a 4 mL vial, a solution of **3a'** (72.1 mg, 0.179 mmol) and cyanuric chloride (12.9 mg, 0.07 mmol) in 1 mL dicholoromethane was sealed and heated at 55 °C for 18 hours. When the reaction showed a full conversion monitored by TLC, the solvent was removed under vacuum. The residue was directly purified by silica gel flash column chromatography (from hexane: ethyl acetate = 20:1 to hexane: ethyl acetate = 10:1) to give product **4** (61.1 mg, 87% yield) as a colorless oil.

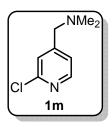
Oxidation:

In a 4 mL vial, 3a' (150.1 mg, 0.40 mmol) and tetrabutylammonium iodide (15.0 mg, 0.04 mmol) were dissolved in 1.5 mL dimethylacetamide. A solution of 30% hydrogen peroxide (0.33 mL, 3.2 mmol) was quickly added to the vial. The vial was then sealed and heated with stirring at 100 °C for 24 hours. (Positive pressure was generated in the vial, thus be careful when opening it.) The reaction was worked up with aqueous NaHCO₃, and then extracted with ethyl acetate (10 mL x 4). The combined organic layer was dried over magnesium sulfate, concentrated under vacuum, and purified by silica gel column chromatography (from hexane : ethyl acetate = 10:1 to hexane : ethyl acetate = 5:1) to give product 5 in 74.5 mg (54% yield) as a colorless oil.

IV. Characterization of new compound

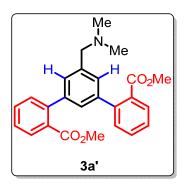


(11): brown solid, mp = 71.5 °C to 73.0 °C. $R_f = 0.37$ (Hex/EA = 3:1 with 3% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.76 (d, J = 8.4 Hz, 2H), 7.30 (dd, J = 3.4, 1.8 Hz, 1H), 7.24 (d, J = 8.1 Hz, 2H), 6.19 (t, J = 3.3 Hz, 1H), 6.13 – 6.10 (m, 1H), 3.50 (s, 2H), 2.38 (s, 3H), 2.04 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 144.16, 136.89, 132.73, 129.21, 127.11, 123.16, 114.67, 110.78, 55.00, 44.53, 21.53. IR (KBr, cm⁻¹) 2942, 2816, 2774, 1366, 1175, 1148, 1131, 1091, 1054. HRMS calcd $C_{14}H_{19}N_2O_2S$ [M+H]⁺: 279.1162. Found: 279.1164.



(1m): yellow oil. $R_f = 0.38$ (Hex/EA = 3:1 with 3% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 8.28 (d, J = 5.1Hz, 1H), 7.32–7.27 (m, 1H), 7.18–7.14 (m, 1H), 3.38 (s, 2H), 2.22 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 151.82, 151.66, 149.47, 124.00, 122.41, 62.51, 45.45.IR (KBr, cm⁻¹) 2970,2870, 2823, 2776, 1594, 1551, 1384, 1143, 913. HRMS calcd $C_8H_{12}ClN_2$ [M+H]⁺: 171.0684. Found: 171.0681.

(3a): 23% yield (reaction time: 12 hours). Colorless oil. $R_f = 0.33$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.82 – 7.75 (m, 1H), 7.50 – 7.44 (m, 1H), 7.38 – 7.23 (m, 5H), 7.21 – 7.16 (m, 1H), 3.59 (s, 3H), 3.44 (s, 2H), 2.23 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 168.81, 142.10, 141.00, 138.20, 130.91, 130.62, 130.45, 129.45, 128.87, 127.76, 127.73, 126.84, 63.94, 51.59, 45.02. IR (KBr, cm⁻¹) 2947, 2815, 2769, 1724, 1288, 1253, 1125, 1090, 1050. HRMS calcd $C_{17}H_{20}NO_2$ [M+H]⁺: 270.1489. Found: 270.1490.



(3a'): 70% (reaction time: 36 hours). Yellow oil. $R_f = 0.19$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.81 (ddd, J = 7.7, 1.5, 0.6 Hz, 2H), 7.56 – 7.48 (m, 2H), 7.44 – 7.36 (m, 4H), 7.25 (d, J = 1.8 Hz, 2H), 7.17 (t, J = 1.7 Hz, 1H), 3.65 (s, 6H), 3.49 (s, 2H), 2.27 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 169.12, 142.08, 141.12, 138.43, 131.12, 130.98, 130.73, 129.71, 128.02, 127.14, 127.04, 64.18, 51.90, 45.34.. IR (KBr, cm⁻¹) 2948, 2815, 2771, 1727, 1293, 1254, 1126, 1097, 1074. HRMS calcd $C_{25}H_{26}NO_4$ [M+H]⁺: 404.1862. Found: 404.1859.

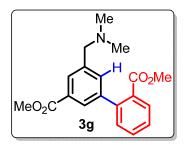
(3b): 73% yield. Yellow oil. $R_f = 0.29$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.80 – 7.76 (m, 1H), 7.53 – 7.46 (m, 1H), 7.41 – 7.34 (m, 2H), 7.14 (s, 1H), 7.03 (s, 2H), 3.63 (s, 3H), 3.41 (s, 2H), 2.38 (s, 3H), 2.25 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 169.27, 142.35, 141.03, 138.59, 137.62, 131.02, 130.93, 130.61, 129.54, 128.70, 127.69, 126.92, 126.28, 64.29, 51.84, 45.39, 21.29. IR (KBr, cm⁻¹) 2947, 2815, 2771, 1724, 1598, 1433, 1291, 1253, 1125, 1098, 1071, 1042. HRMS calcd $C_{18}H_{22}NO_2$ [M+H]⁺: 284.1645. Found: 284.1647.

(3c): 72% yield. Colorless oil. $R_f = 0.36$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.83 (ddd, J = 7.7, 1.5, 0.5 Hz, 1H), 7.51 (td, J = 7.5, 1.5 Hz, 1H), 7.41 (td, J = 7.6, 1.4 Hz, 1H), 7.37 – 7.28 (m, 1H), 7.34 – 7.28 (m, 1H), 7.20 (t, J = 1.8 Hz, 1H), 7.12 (t, J = 1.5 Hz, 1H), 3.65 (s, 3H), 3.41 (s, 2H), 2.24 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 168.52, 142.85, 141.04, 140.62, 133.69, 131.30, 130.55, 130.54, 129.90, 127.66, 127.54, 127.24, 126.95, 63.65, 51.92, 45.31. IR (KBr, cm⁻¹) 2947, 2818, 2772, 1730, 1577, 1456, 1435, 1289, 1254, 1126, 1095, 1065. HRMS calcd $C_{17}H_{19}CINO_2$ [M+H]⁺: 304.1104. Found: 304.1101.

(3d): 65% yield. Colorless oil. $R_f = 0.39$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.84 (dd, J = 7.7, 1.5 Hz, 1H), 7.52 (td, J = 7.5, 1.5 Hz, 1H), 7.48 – 7.46 (m, 1H), 7.42 (td, J = 7.6, 1.3 Hz, 1H), 7.36 (t, J = 1.8 Hz, 1H), 7.34 (dd, J = 7.7, 1.4 Hz, 1H), 7.17 (t, J = 1.6 Hz, 1H), 3.66 (s, 3H), 3.42 (s, 2H), 2.25 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 168.57, 143.15, 140.96, 140.87, 131.35, 130.62, 130.60, 130.57, 129.96, 129.88, 127.77, 127.61, 121.93, 63.65, 51.99, 45.36. IR (KBr, cm⁻¹) 2947, 2818, 2772, 1730, 1456, 1434, 1289, 1254, 1126, 1095, 1064, 1042. HRMS calcd $C_{17}H_{19}BrNO_2$ [M+H]⁺: 348.0594. Found: 348.0597

(3e): 80% yield. Colorless oil. $R_f = 0.21$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.81 – 7.76 (m, 1H), 7.53 – 7.47 (m, 1H), 7.42 – 7.36 (m, 2H), 6.88 (s, 1H), 6.83 (s, 1H), 6.77 (s, 1H), 3.82 (s, 3H), 3.63 (s, 3H), 3.42 (s, 2H), 2.25 (s, 6H). ¹³C NMR (101 MHz, CDCl₃₃) δ 169.16, 159.36, 142.34, 142.06, 140.15, 131.04, 130.96, 130.50, 129.51, 127.10, 121.65, 113.15, 112.75, 64.29, 55.24, 51.90, 45.36. IR (KBr, cm⁻¹) 2947, 2816, 2773, 1727, 1594, 1456, 1292, 1255, 1212, 913. HRMS calcd C₁₈H₂₁NNaO₃ [M+Na]⁺: 322.1414. Found: 322.1404.

(3f): 55% yield. Colorless oil. $R_f = 0.41$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, acetonitrile- d_3) δ 7.83 (ddd, J = 7.7, 1.5, 0.5 Hz, 1H), 7.64 –7.58 (m, 2H), 7.53 – 7.46 (m, 3H), 7.42 (ddd, J = 7.6, 1.3, 0.5 Hz, 1H), 3.62 (s, 3H), 3.49 (s, 2H), 2.20 (s, 6H). ¹³C NMR (101 MHz, Acetonitrile- d_3) δ 169.23 , 143.05 , 141.98 , 141.52 , 133.60 (d, J = 1.5 Hz), 132.57 , 131.93 , 131.63 , 130.75 , 130.68 (q, J = 31.9 Hz), 128.98 , 125.44 (q, J = 271.6 Hz), 125.12 (q, J = 3.9 Hz), 124.51 (q, J = 3.8 Hz), 63.88 , 52.60 , 45.54 . ¹⁹F NMR (376 MHz, Acetonitrile- d_3) δ 62.83. IR (KBr, cm⁻¹) 2950, 2820, 2776, 1726, 1458, 1437, 1346, 1293, 1253, 1164, 1125. HRMS calcd $C_{18}H_{19}F_3NO_2$ [M+H]⁺: 338.1362. Found: 338.1364.



(3g): 45% yield. Colorless oil. $R_f = 0.41$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.96 (t, J = 1.7 Hz, 1H), 7.90 (t, J = 1.7 Hz, 1H), 7.86 (dd, J = 7.8, 1.5 Hz, 1H), 7.52 (td, J = 7.5, 1.4 Hz, 1H), 7.45 (t, J = 1.7 Hz, 1H), 7.42 (td, J = 7.6, 1.4 Hz, 1H), 7.36 (dd, J = 7.6, 1.3 Hz, 1H), 3.90 (s, 3H), 3.62 (s, 3H), 3.48 (s, 2H), 2.25 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 168.47, 166.94, 141.65, 141.57, 139.13, 133.54, 131.38, 130.77, 130.42, 130.01, 129.93, 128.89, 128.23, 127.49, 63.81, 52.05, 51.90, 45.32. IR (KBr, cm⁻¹) 2950, 2818, 2774, 1724, 1457, 1436, 1328, 1292, 1238, 1126, 1097, 913. HRMS calcd C₁₉H₂₂NO₄ [M+H]⁺: 328.1543. Found: 328.1541.

(3h): 47% yield. Colorless oil. $R_f = 0.46$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.85 – 7.80 (m, 1H), 7.54 – 7.46 (m, 1H), 7.43 – 7.37 (m, 1H), 7.34 (d, J = 7.6, 0.7 Hz, 1H), 7.29 (dd, J = 7.0, 2.4 Hz, 1H), 7.21 – 7.12 (m, 1H), 7.06 (t, J = 9.4, 8.4, 0.9 Hz, 1H), 3.65 (s, 3H), 3.56 – 3.42 (m, 2H), 2.28 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 168.68 , 160.71 (d, J = 246.8 Hz), 141.46 , 137.00 (d, J = 3.7 Hz), 131.41 (d, J = 4.7 Hz), 131.16 , 130.69 , 130.58 , 129.75 , 128.55 (d, J = 8.3 Hz), 127.12 , 124.87 (d, J = 15.2 Hz), 114.81 (d, J = 22.8 Hz), 56.53 (d, J = 1.6 Hz), 51.82 , 45.12. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -120.43. IR (KBr, cm⁻¹) 2948, 2820, 2778, 1729, 1503, 1479, 1289, 1249, 1126, 1087, 913. HRMS calcd $C_{17}H_{19}FNO_2$ [M+H]⁺: 288.1394. Found: 288.1395.

(3i): 32% yield. Colorless oil. $R_f = 0.21$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.88 (dd, J = 7.8, 1.5 Hz, 1H), 7.54 (td, J = 7.5, 1.4 Hz, 1H), 7.47 – 7.38 (m, 2H), 6.84 (d, J = 1.6 Hz, 1H), 6.75 (d, J = 1.5 Hz, 1H), 5.91 (s, 2H), 3.72 (s, 3H), 3.37 (s, 2H), 2.25 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 168.45, 147.13, 143.68, 136.35, 132.80, 131.58, 130.97, 130.80, 130.04, 127.62, 122.72, 122.15, 108.69, 100.94, 64.12, 52.00,

45.22. **IR** (KBr, cm⁻¹) 2948, 2617, 2772, 1726, 1420, 1291, 1256, 1192, 1126, 1096, 1046, 913. **HRMS** calcd $C_{19}H_{22}NO_4$ [M+H]⁺: 314.1389. Found: 314.1387.

(3j): 33% yield. Colorless oil. $R_f = 0.43$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.83 – 7.78 (m, 1H), 7.55 – 7.49 (m, 1H), 7.43 – 7.38 (m, 2H), 7.38 – 7.33 (m, 1H), 7.31 – 7.26 (m, 1H), 7.25 – 7.18 (m, 2H), 3.61 (s, 3H), 3.27 (q, J = 6.7 Hz, 1H), 2.21 (s, 6H), 1.39 (d, J = 6.7 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 169.21, 143.87, 142.42, 141.15, 131.08, 130.98, 130.65, 129.63, 127.95, 127.60, 127.02, 126.78, 126.37, 65.89, 51.86, 43.24, 20.34. **IR** (KBr, cm⁻¹) 2976, 2950, 2816, 2769, 1725, 1290, 1126, 1088, 1050, 931. **HRMS** calcd $C_{18}H_{21}NNaO_2$ [M+Na]⁺: 306.1465. Found: 306.1467.

(3j'): 31% yield. Colorless oil. $R_f = 0.23$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.80 (ddd, J = 7.7, 1.5, 0.6 Hz, 2H), 7.51 (ddd, J = 7.7, 7.2, 1.4 Hz, 2H), 7.44 – 7.36 (m, 4H), 7.22 (dd, J = 1.7, 0.5 Hz, 2H), 7.18 (t, J = 1.7 Hz, 1H), 3.65 (s, 6H), 3.30 (q, J = 6.7 Hz, 1H), 2.23 (s, 6H), 1.40 (d, J = 6.7 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 169.20, 143.70, 142.13, 141.11, 131.13, 131.07, 130.68, 129.66, 127.11, 126.76, 126.56, 65.81, 51.89, 43.26, 20.47.IR (KBr, cm⁻¹) 2950, 2817, 2769, 1727, 1595, 1433, 1293, 1254, 1126, 913. HRMS calcd $C_{26}H_{27}NNaO_4$ [M+Na]⁺: 440.1832. Found: 440.1837.

For chiral (S)-(-)-3j', it was directly synthesized from commercially available (S)-(-)-N,N-dimethyl-1-phenylethylamine by following the general procedure at 130°C.

Chiral HPLC (Chiralpak IA, hexane: isopropanol (with 0.5% V/V diethylamine) = 98.5:1.5, 1 mL/min, 254 nm): $t_s = 28.65 \text{ min}$, $t_r = 31.34 \text{ min}$. $[\alpha]_D^{20} = -13^\circ$ (c 1.0, CDCl₃) at >99 % ee.

(3k): 17% yield. Yellow oil. $R_f = 0.54$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.81 – 7.74 (m, 1H), 7.54 – 7.46 (m, 1H), 7.44 – 7.36 (m, 2H), 6.91 (dd, J = 2.5, 1.4 Hz, 1H), 6.86 (t, J = 1.5 Hz, 1H), 6.76 (dd, J = 2.5, 1.6 Hz, 1H), 3.82 (s, 3H), 3.64 (s, 3H), 3.62 (s, 2H), 2.58 – 2.47 (m, 4H), 1.83 – 1.74 (m, 4H). ¹³C NMR (101 MHz, CDCl₃) δ 169.25, 159.35, 142.32, 142.16, 140.62, 131.06, 131.03, 130.56, 129.53, 127.12, 121.51, 113.27, 112.48, 60.62, 55.30, 54.12, 51.95, 23.44. IR (KBr, cm⁻¹) 2951, 2784, 1727, 1594, 1292, 1252, 1211, 1126, 913. HRMS calcd $C_{20}H_{23}NNaO_3$ [M+Na]⁺: 348.1570. Found: 348.1559.

(3k): 48% yield. Brown oil. $R_f = 0.36$ (Hex/EA = 3:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.84 – 7.78 (m, 2H), 7.67 (ddd, J = 7.7, 1.5, 0.5 Hz, 1H), 7.47 - 7.43 (m, 1H), 7.41 - 7.37 (m, 2H), 7.35 – 7.29 (m, 1H), 7.28 – 7.24 (m, 2H), 6.21 (dd, J = 1.9, 0.9 Hz, 1H), 3.71 (s, 3H), 3.52 (s, 2H), 2.39 (s, 3H), 2.07 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 169.41, 144.27, 136.80, 133.63, 132.70, 131.04, 130.79, 129.93, 129.33, 129.24, 127.16, 126.88, 125.43, 120.57, 115.68, 54.95, 51.99, 44.50, 21.52. IR (KBr, cm⁻¹) 2949, 2869, 2818, 1727, 1366, 1291, 1175, 1133, 1103, 913. HRMS calcd $C_{22}H_{25}N_2O_4S$ [M+H]⁺: 413.1530. Found: 413.1535.

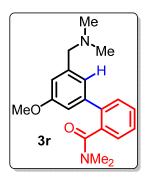
(3m): 66% yield. Brown oil. $R_f = 0.35$ (Hex/EA = 3:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.78 (ddd, J = 7.6, 1.4, 0.6 Hz, 1H), 7.61 – 7.51 (m, 2H), 7.49 – 7.44 (m, 1H), 7.39 (dd, J = 1.2, 0.6 Hz, 1H), 7.28 (dd, J = 1.3, 0.7 Hz, 1H), 3.75 (s, 3H), 3.46 (s, 2H), 2.28 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 169.05, 158.67, 152.06, 150.68, 139.04, 131.70, 131.00, 129.59, 128.72, 122.38, 121.19, 62.71, 52.14, 45.56. IR (KBr, cm⁻¹) 2979, 2947, 2869, 1728, 1595, 1548, 1291, 1258, 1127, 913. HRMS calcd $C_{16}H_{18}CIN_2O_2$ [M+H]⁺: 305.1051. Found: 305.1056.

(31): 70% yield. Colorless oil. $R_f = 0.21$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.59 (dt, J = 1.6, 0.7 Hz, 1H), 7.33 – 7.27 (m, 2H), 6.86 (dd, J = 2.5, 1.4 Hz, 1H), 6.81 (t, J = 1.5 Hz, 1H), 6.75 (dd, J = 2.5, 1.6 Hz, 1H), 3.81 (s, 3H), 3.63 (s, 3H), 3.41 (s, 2H), 2.40 (s, 3H), 2.24 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 169.34, 159.32, 142.27, 140.06, 139.18, 136.95, 131.75, 130.74, 130.40, 129.98, 121.67, 112.92, 112.78, 64.30, 55.19, 51.84, 45.34, 20.84.IR (KBr, cm⁻¹) 2946, 2816, 2772, 1732, 1594, 1456, 1434, 1296, 1253, 1204, 826. HRMS calcd $C_{19}H_{24}NO_3$ [M+H]⁺: 314.1751. Found: 314.1752.

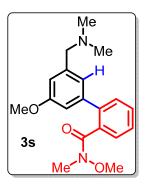
(3m): 71% yield. Colorless oil. $R_f = 0.19$ (Hex/EA = 3:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 8.01 (d, J = 1.7 Hz, 1H), 7.97 (dd, J = 8.0, 1.7 Hz, 1H), 7.74 (dd, J = 7.9, 1.4 Hz, 1H), 6.84 (s, 1H), 6.79 (s, 1H), 6.72 (s, 1H), 3.86 (s, 3H), 3.76 (s, 3H), 3.60 (s, 3H), 3.36 (s, 2H), 2.18 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 168.38, 165.91, 159.37, 141.93, 141.10, 140.40, 134.89, 132.04, 131.33, 129.32, 127.91, 121.31, 113.44, 112.54, 64.10, 55.11, 52.20, 52.02, 45.23.IR (KBr, cm⁻¹) 2950, 2817, 2773, 1727, 1594, 1456, 1434, 1284, 1250, 1209, 1116, 913. HRMS calcd $C_{20}H_{23}NO_5$ [M+H]⁺: 358.1649. Found: 358.1649.

(3n): 31% yield. Colorless oil. $R_f = 0.24$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.56 – 7.45 (m, 2H), 7.43 – 7.35 (m, 2H), 6.92 (s, 1H), 6.86 (s, 1H), 6.78 (s, 1H), 3.82 (s, 3H), 3.42 (s, 2H), 2.24 (s, 6H), 2.02 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 204.81, 159.80, 141.84, 141.03, 140.82, 140.32, 130.58, 130.03, 127.71, 127.44, 122.11, 113.78, 113.29, 64.14, 55.33, 45.35, 30.37. IR (KBr, cm⁻¹) 2941, 2816, 2772, 1688, 1592, 1456, 1361, 1332, 1276, 1238, 1211, 1146, 1030. HRMS calcd $C_{18}H_{21}NO_2$ [M+H]⁺: 284.1645. Found: 284.1646.

(30): 58% yield. Colorless oil. $R_f = 0.22$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.64 – 7.59 (m, 2H), 7.58 – 7.42 (m, 4H), 7.39 – 7.33 (m, 1H), 7.22 (s, 2H), 6.77 (t, J = 1.5 Hz, 1H), 6.71 – 6.68 (m, 1H), 6.67 – 6.61 (m, 1H), 3.67 (s, 3H), 3.25 (s, 2H), 2.09 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 198.83, 159.31, 141.18, 140.70, 140.45, 138.95, 137.21, 132.63, 130.21, 129.71, 129.63, 128.57, 127.92, 127.20, 122.25, 113.57, 113.40, 64.04, 55.16, 45.24. IR (KBr, cm⁻¹) 2942, 2817, 2771, 1728, 1667, 1594, 1456, 1285, 1254, 1212, 1148, 928. HRMS calcd $C_{23}H_{24}NO_2$ [M+H]⁺: 346.1802. Found: 346.1804.

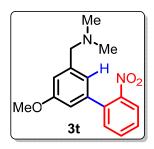


(3p): 60% yield. Colorless oil. $R_f = 0.29$ (Hex/EA = 1:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.44 – 7.33 (m, 4H), 6.97 (s, 1H), 6.93 – 6.90 (m, 1H), 6.89 – 6.85 (m, 1H), 3.80 (s, 3H), 3.46 – 3.35 (m, 2H), 2.85 (s, 3H), 2.44 (s, 3H), 2.23 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 171.25, 159.57, 141.06, 140.56, 138.45, 135.64, 129.29, 129.15, 127.66, 127.31, 121.56, 113.99, 112.47, 64.20, 55.32, 45.35, 37.99, 34.54.IR (KBr, cm⁻¹) 2939, 2856, 2816, 2772, 1633, 1593, 1456, 1394, 1210, 913. HRMS calcd $C_{19}H_{25}N_2O_2$ [M+H]⁺: 313.1911. Found: 313.1913.



(3q): 53% yield. Colorless oil. $R_f = 0.37$ (Hex/EA = 1:1 with 4% v/v Et₃N). This compound has several rotamers at r.t. in CDCl₃. Both proton and 13 carbon were taken in DMSO-d6 at 100°C.

¹**H NMR** (600 MHz, DMSO- d_6) δ 7.49 (dd, J = 7.4, 1.6 Hz, 1H), 7.43 (ddd, J = 14.6, 7.4, 1.4 Hz, 2H), 7.38 (dd, J = 7.6, 1.6 Hz, 1H), 6.94 (d, J = 1.7 Hz, 1H), 6.87 (dt, J = 7.8, 2.1 Hz, 2H), 3.79 (s, 3H), 3.41 (s, 2H), 3.06 – 2.90 (m, 6H), 2.20 (s, 6H). ¹³**C NMR** (151 MHz, DMSO- d_6) δ 158.84, 140.44, 140.18, 138.20, 134.67, 128.63, 128.57, 126.38, 126.19, 120.72, 113.04, 112.25, 62.94, 59.73, 54.71, 54.69, 44.32.**IR** (KBr, cm⁻¹): 2938, 2816, 2773, 1652, 1593, 1457, 1364, 1332, 1214. **HRMS** calcd $C_{19}H_{25}N_2O_3$ [M+H]⁺: 329.1860. Found: 329.1862.



(3r): 66% yield. Yellow oil. $R_f = 0.43$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.82 (dd, J = 8.5, 1.4 Hz, 1H), 7.62 – 7.55 (m, 1H), 7.49 – 7.41 (m, 2H), 6.91 (s, 1H), 6.83 (s, 1H), 6.79 – 6.73 (m, 1H), 3.81 (s, 3H), 3.41 (s, 2H), 2.24 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 159.70, 149.23, 140.96, 138.38, 136.11, 132.08, 131.78, 128.07, 123.88, 120.86, 113.95, 112.42, 64.11, 55.27, 45.33.IR (KBr, cm⁻¹) 2942, 2818, 2774, 1572, 1529, 1457, 1361, 1213, 913. HRMS calcd C₁₆H₁₉N₂O₃ [M+H]⁺: 287.1390. Found: 287.1390.

(3u): 4% yield. Pale yellow oil. $R_f = 0.41$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 8.12 – 8.05 (m, 2H), 7.69 – 7.62 (m, 2H), 7.16 (t, J = 1.5 Hz, 1H), 7.05 (dd, J = 2.5, 1.6 Hz, 1H), 6.92 (dd, J = 2.5, 1.3 Hz, 1H), 3.94 (s, 3H), 3.87 (s, 3H), 3.46 (s, 2H), 2.28 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 166.99, 160.10, 145.45,

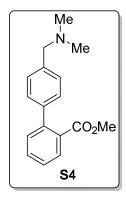
141.24, 129.98, 129.06, 128.94, 127.11, 120.40, 113.91, 111.91, 64.43, 55.43, 52.12, 45.48. **IR** (KBr, cm⁻¹) 2979, 2949, 2869, 1726, 1363, 1286, 1175, 1134, 913. **HRMS** calcd $C_{18}H_{22}NO_3$ [M+H]⁺: 300.1594. Found: 300.1599.

(4): 87% yield. Colorless oil. $R_f = 0.34$ (Hex/EA = 5:1). ¹H NMR (400 MHz, CDCl₃) δ 7.85 (ddd, J = 7.7, 1.4, 0.6 Hz, 2H), 7.54 (td, J = 7.5, 1.5 Hz, 2H), 7.46 – 7.39 (m, 4H), 7.33 (d, J = 1.7 Hz, 2H), 7.25 (t, J = 1.7 Hz, 1H), 4.65 (s, 2H), 3.68 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 168.90, 141.68, 141.46, 136.99, 131.30, 130.87, 130.64, 129.90, 128.37, 127.48, 52.04, 46.04. IR (KBr, cm⁻¹) 2950, 1725, 1596, 1434, 1294, 1255, 1127, 1098, 1074, 913. HRMS calcd $C_{23}H_{19}ClNaO_4$ [M+Na]⁺: 417.0864. Found: 417.0859.

(5): 55% yield. Colorless oil. $R_f = 0.23$ (Hex/EA = 5:1). ¹H NMR (400 MHz, CDCl₃) δ 10.08 (s, 1H), 7.92 (ddd, J = 7.8, 1.5, 0.5 Hz, 2H), 7.82 (d, J = 1.8 Hz, 2H), 7.56 (td, J = 7.5, 1.4 Hz, 2H), 7.50 (t, J = 1.7 Hz, 1H), 7.49 – 7.44 (m, 2H), 7.40 (ddd, J = 7.6, 1.4, 0.5 Hz, 2H), 3.68 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 191.95, 168.18, 142.14, 141.10, 135.98, 134.40, 131.59, 130.80, 130.34, 130.24, 128.35, 127.85, 52.03.IR (KBr, cm⁻¹) 2951, 1726, 1699, 1597, 1434, 1294, 1258, 1166, 1127, 913. HRMS calcd $C_{23}H_{18}NaO_{5}$ [M+Na]⁺: 397.1046. Found: 397.1047.

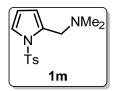
(S2): 43% yield. Colorless oil. $R_f = 0.44$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.96 (ddd, J = 7.8, 1.5, 0.5 Hz, 1H), 7.55 – 7.46 (m, 2H), 7.42 (td, J = 7.6, 1.4 Hz, 1H), 7.33 (td, J = 7.5, 1.5 Hz, 1H), 7.29 – 7.20 (m, 2H), 7.12 – 7.06 (m, 1H), 3.57 (s, 3H), 3.21 – 2.95 (m, 2H), 2.06 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 167.67, 142.15, 141.70, 136.36, 131.08, 131.04, 130.79, 129.70, 128.76, 128.71, 127.15, 127.07, 126.11,

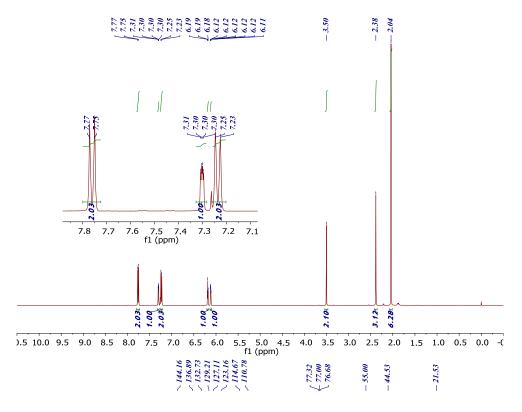
61.21, 51.70, 45.26.**IR** (KBr, cm⁻¹) 2947, 2815, 2769, 1732, 1456, 1289, 1253, 1127, 1085, 913. **HRMS** calcd $C_{17}H_{20}NO_2$ [M+H]⁺: 270.1489. Found: 270.1488.

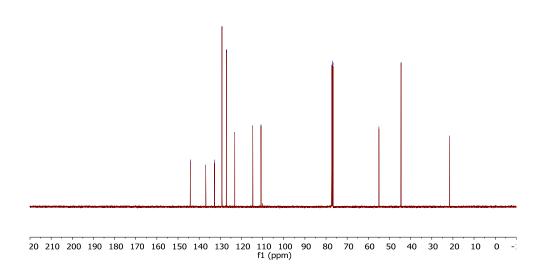


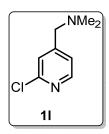
(S2): 45% yield. Colorless oil. $R_f = 0.35$ (Hex/EA = 5:1 with 4% v/v Et₃N). ¹H NMR (400 MHz, CDCl₃) δ 7.81 (ddd, J = 7.6, 1.4, 0.7 Hz, 1H), 7.52 (td, J = 7.5, 1.4 Hz, 1H), 7.44 – 7.37 (m, 2H), 7.36 – 7.31 (m, 2H), 7.27 (d, J = 7.0 Hz, 2H), 3.62 (s, 3H), 3.47 (s, 2H), 2.27 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 169.27 , 142.20 , 140.03 , 137.83 , 131.19 , 130.89 , 130.65 , 129.72 , 128.81 , 128.18 , 127.05 , 64.12 , 51.88 , 45.41 IR (KBr, cm⁻¹) 2947, 2816, 2769, 1727, 1447, 1433, 1287, 1253, 1126, 1089. HRMS calcd $C_{17}H_{20}NO_2$ [M+H]⁺: 270.1489. Found: 270.1481.

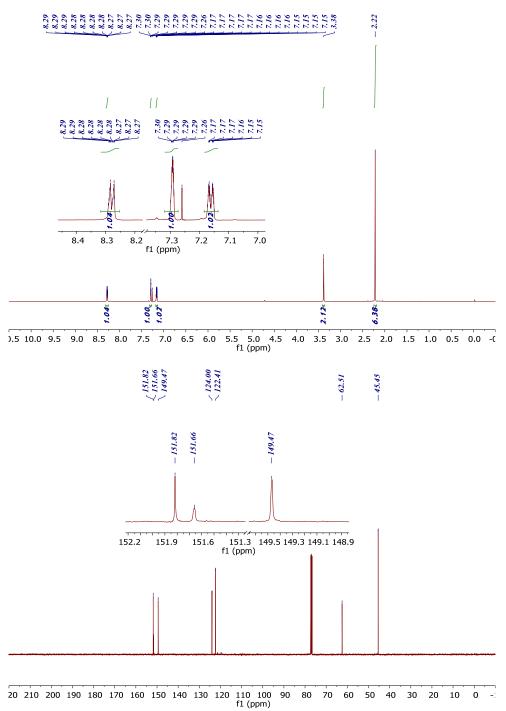
V. ¹H- and ¹³C-NMR spectra

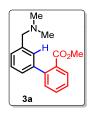


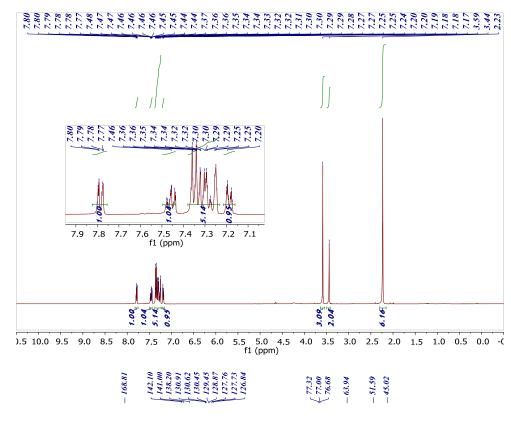


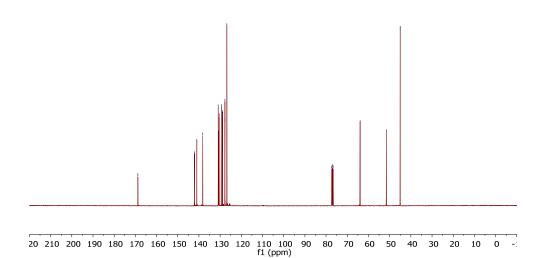


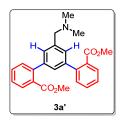


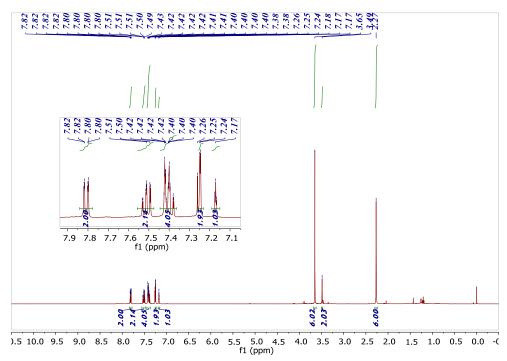


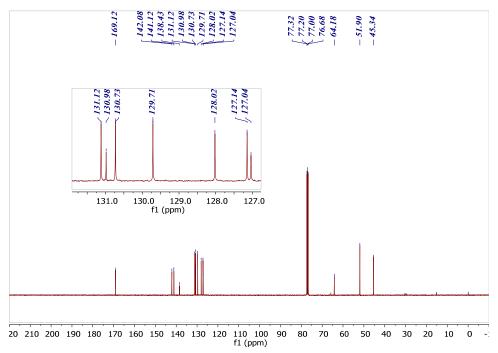


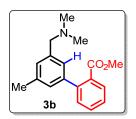


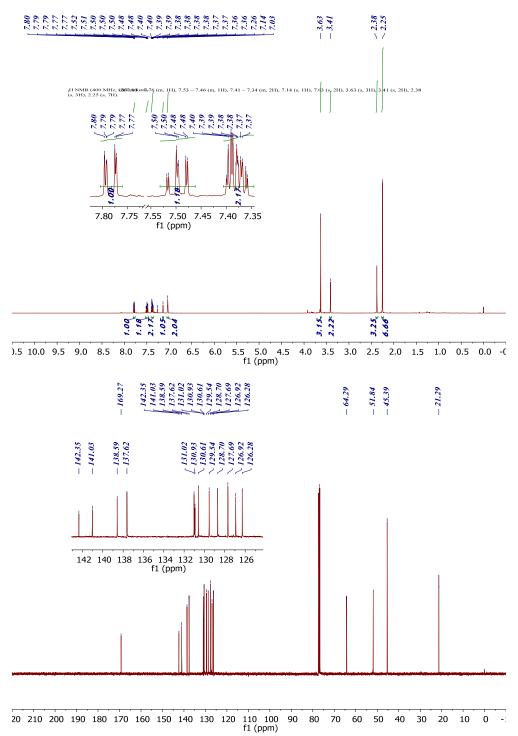




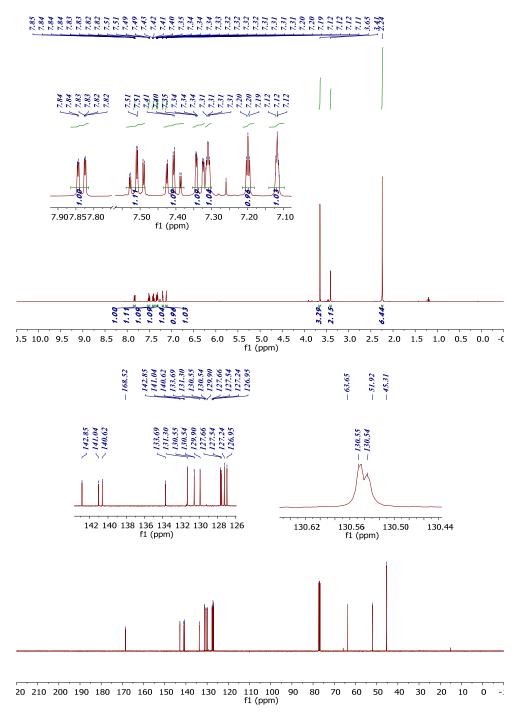


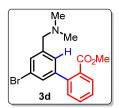


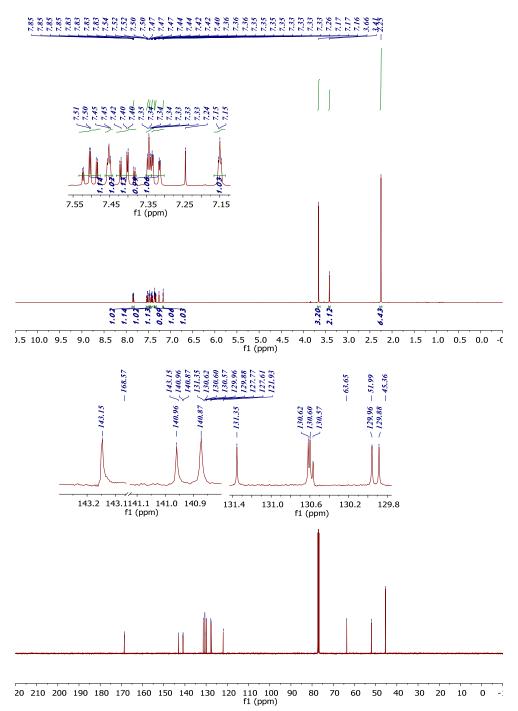


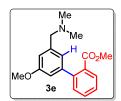


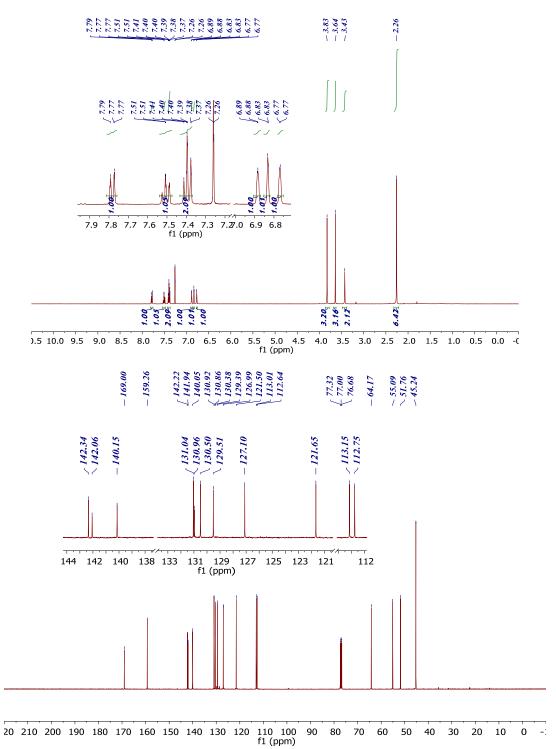


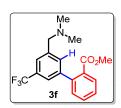


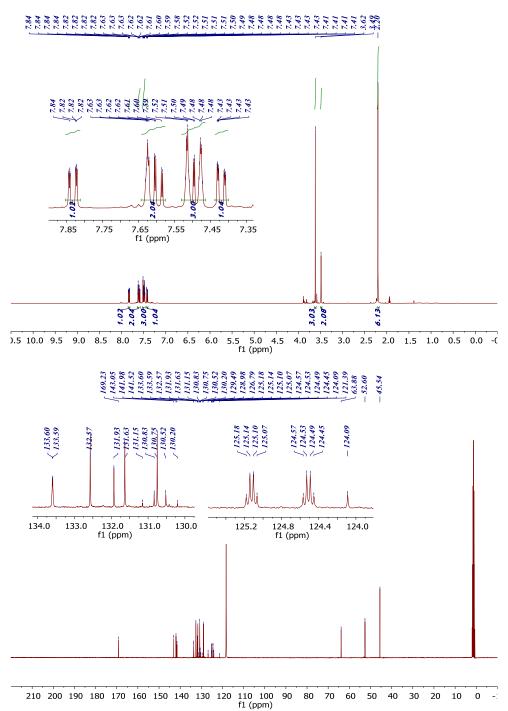


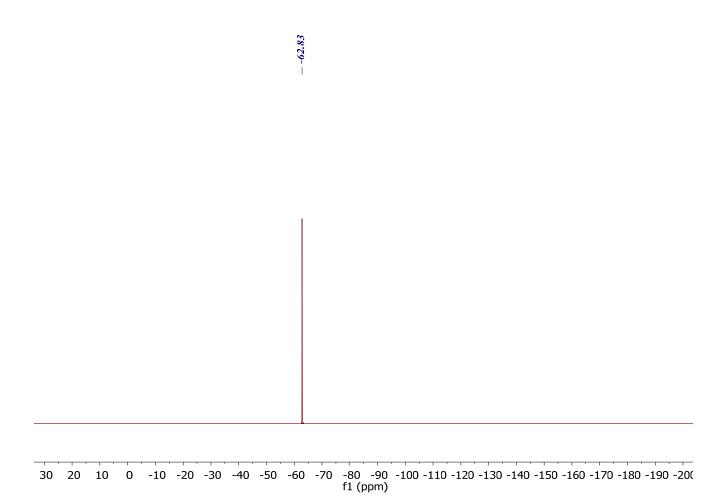


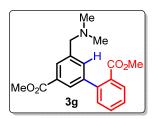


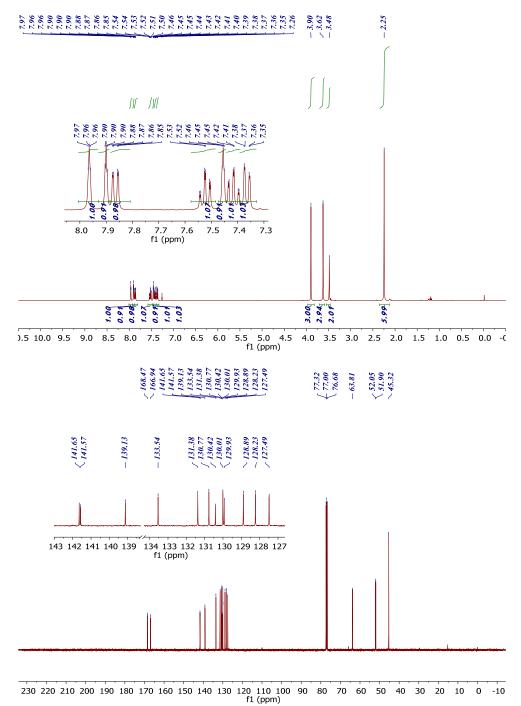


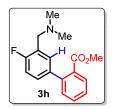


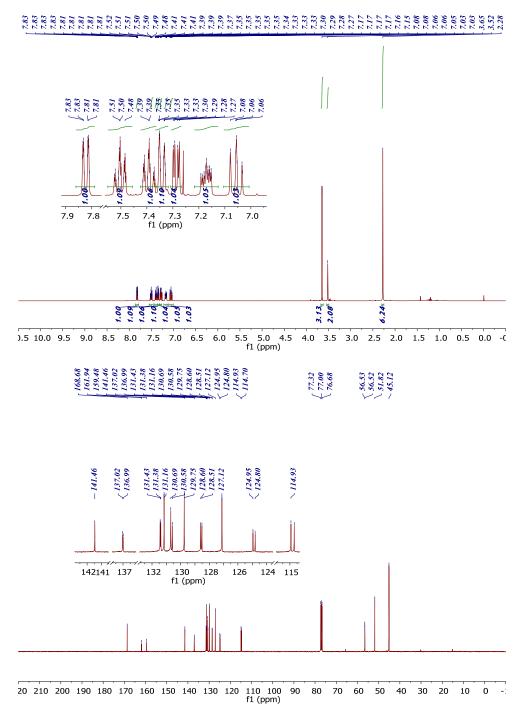




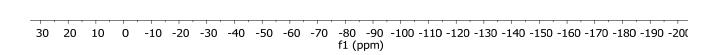


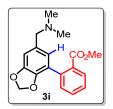


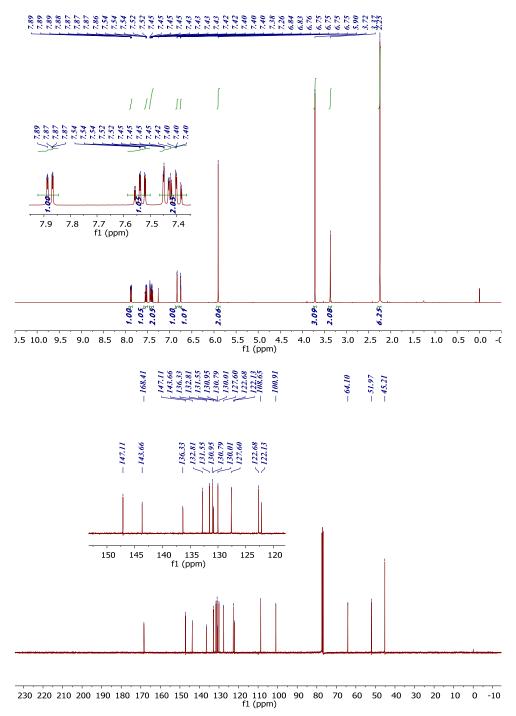


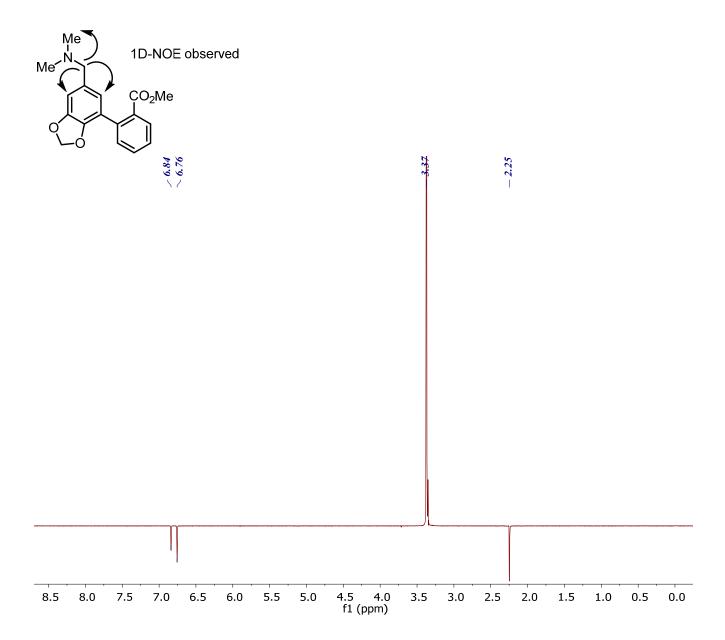


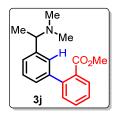


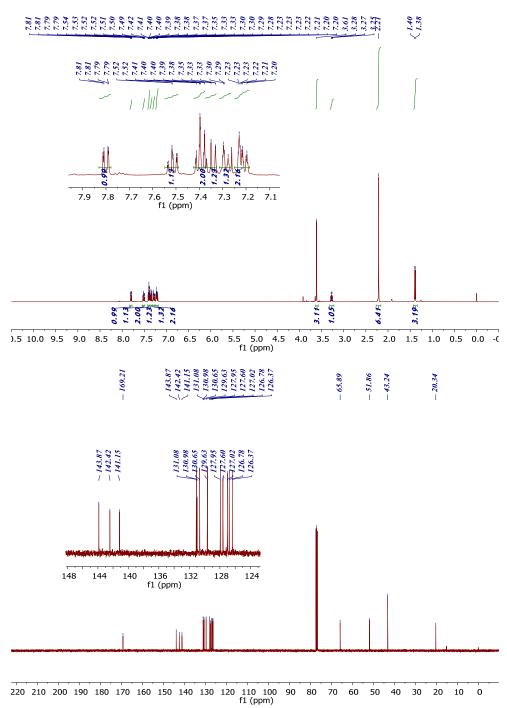


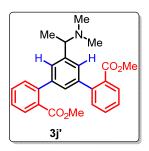


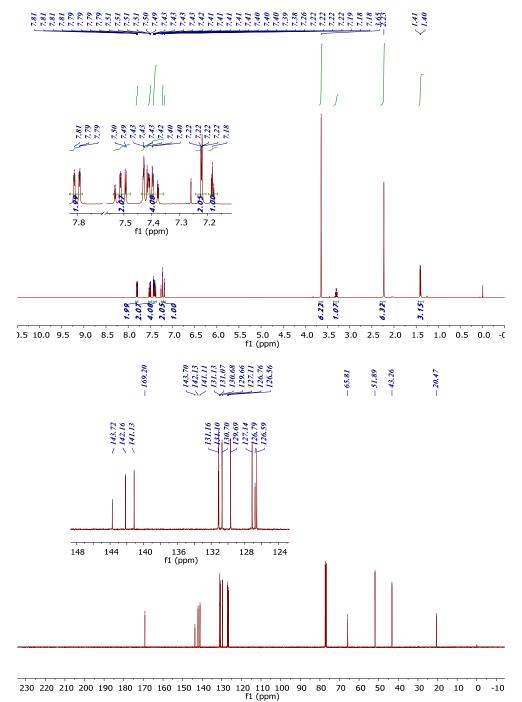




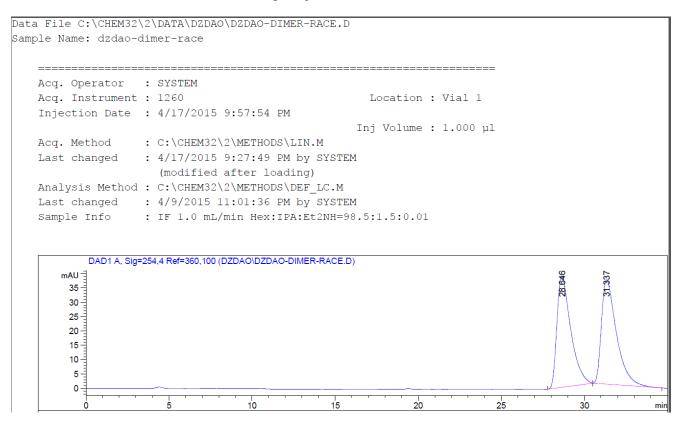








Chiral HPLC conditions for racemic sample 3j'



Data File C:\CHEM32\2\DATA\DZDAO\DZDAO-DIMER-RACE.D

Sample Name: dzdao-dimer-race

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak	RetTime Type	Width	Area	Height	Area
#	[min]	[min]	[mAU*s]	[mAU]	용
1	28.646 BB	0.8614	2246.63452	38.80727	49.5323
2	31.337 BBA	0.9459	2289.05762	36.49050	50.4677
Totals :			4535.69214	75.29777	

Chiral HPLC conditions for enantiopure sample 3j'

Data File C:\CHEM32\2\DATA\DZDAO\DZDAO-693-DIMER-CHIRAL.D

Sample Name: dzdao-693-dimer-chiral

Acq. Operator : SYSTEM

Acq. Instrument: 1260 Location: Vial 1

Injection Date : 4/17/2015 10:35:56 PM

Inj Volume : 1.000 µl

Acq. Method : C:\CHEM32\2\METHODS\LIN.M

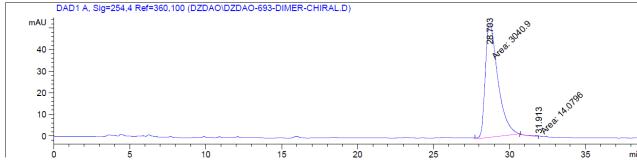
Last changed : 4/17/2015 9:27:49 PM by SYSTEM

(modified after loading)

Analysis Method : C:\CHEM32\2\METHODS\DEF_LC.M
Last changed : 4/9/2015 11:01:36 PM by SYSTEM

Sample Info : IF 1.0 mL/min Hex:IPA:Et2NH=98.5:1.5:0.01

Additional Info : Peak(s) manually integrated



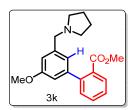
Data File C:\CHEM32\2\DATA\DZDAO\DZDAO-693-DIMER-CHIRAL.D

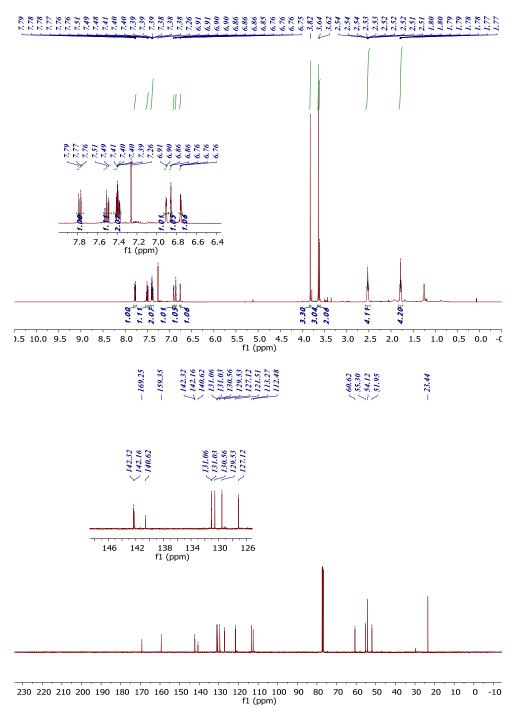
Sample Name: dzdao-693-dimer-chiral

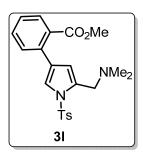
Signal 1: DAD1 A, Sig=254,4 Ref=360,100

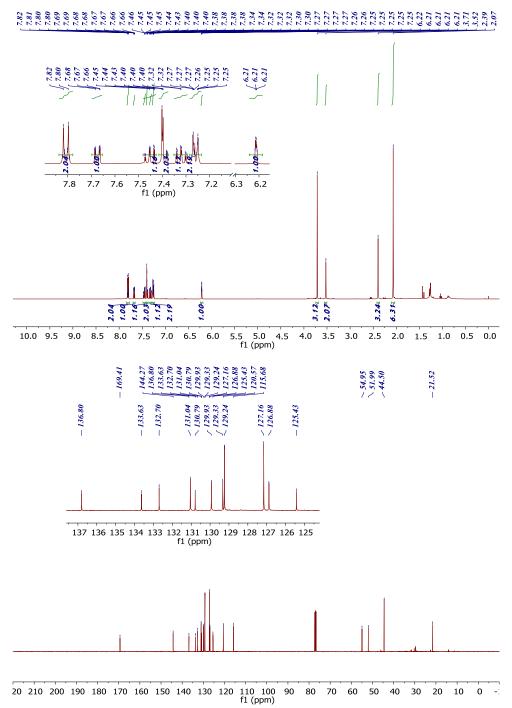
Peak RetTime	Type	Width	Area	Height	Area
# [min]		[min]	[mAU*s]	[mAU]	%
1 28.703	MM	0.9604	3040.89551	52.76996	99.5391
2 31.913	MM	0.6126	14.07957	3.83043e-1	0.4609

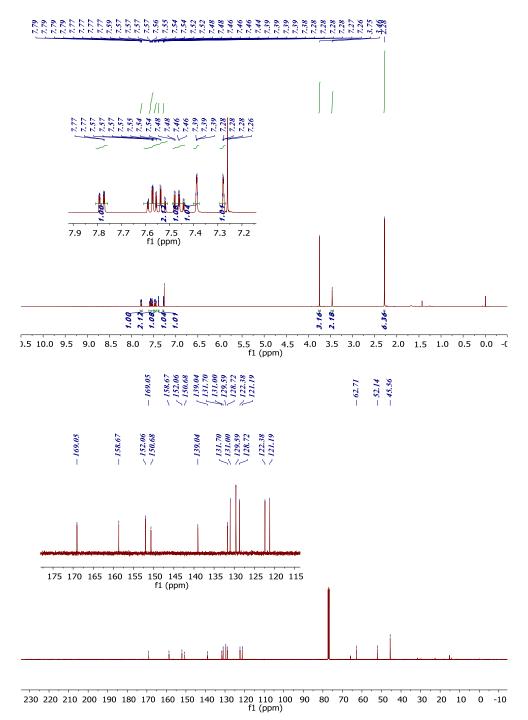
Totals: 3054.97507 53.15300

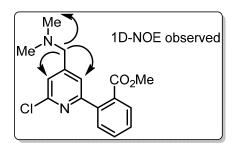


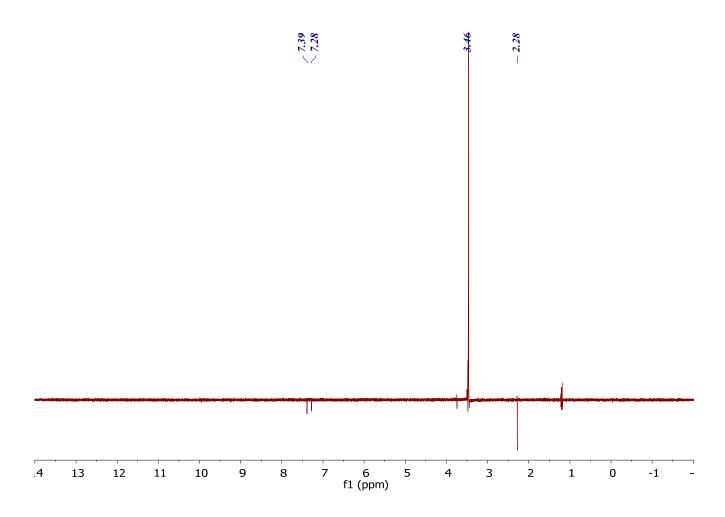


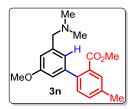


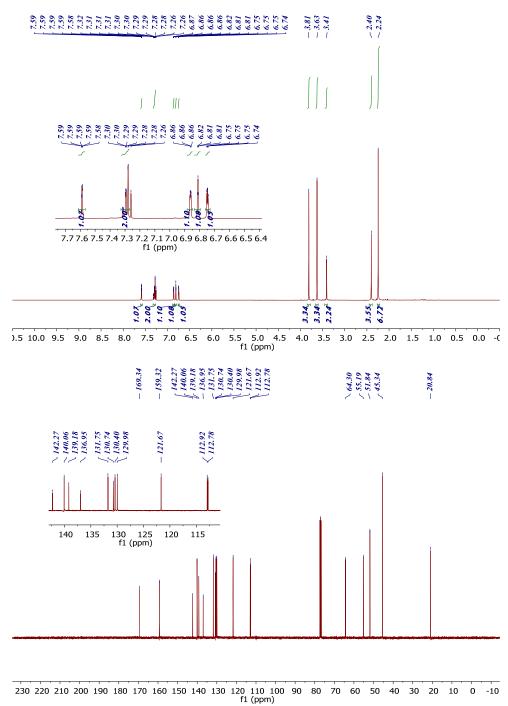


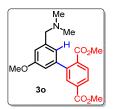


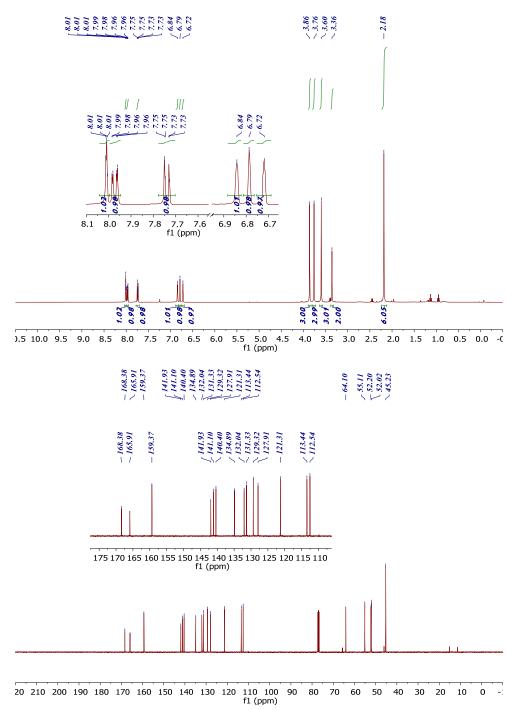




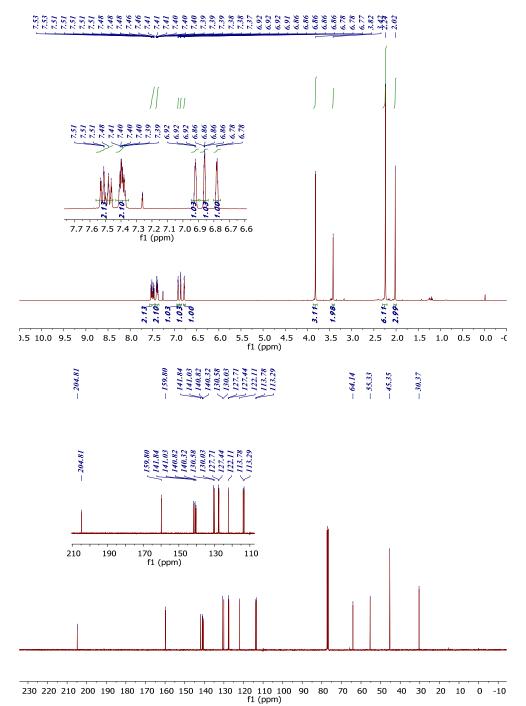




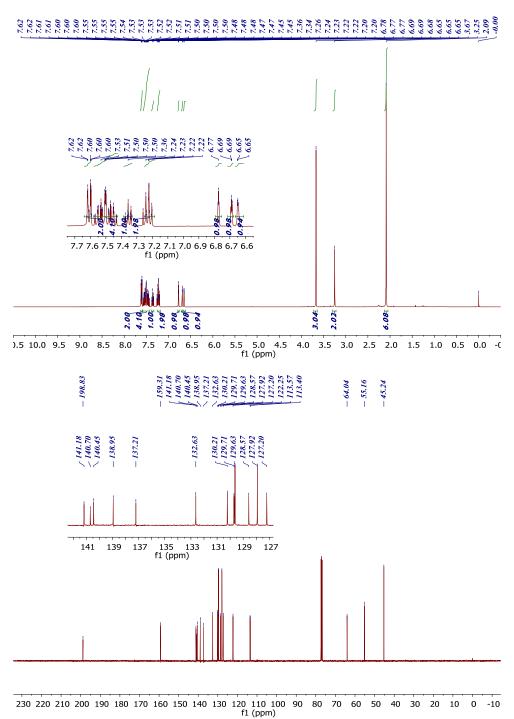


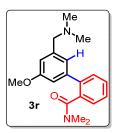


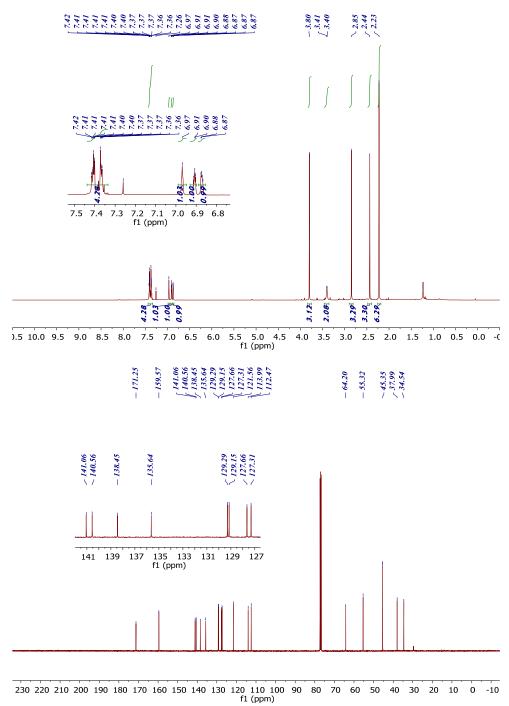


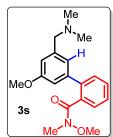




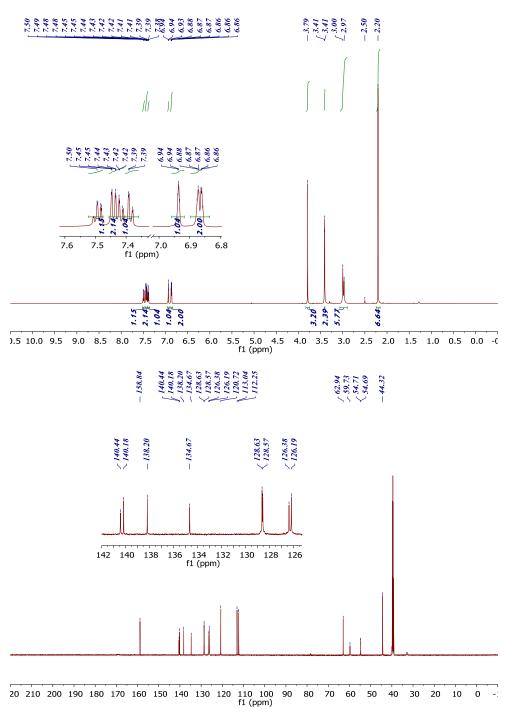




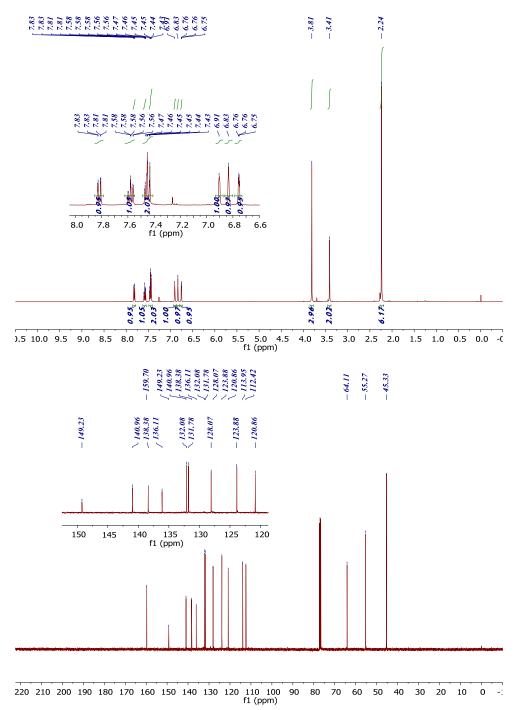


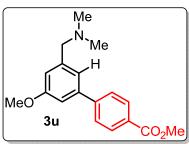


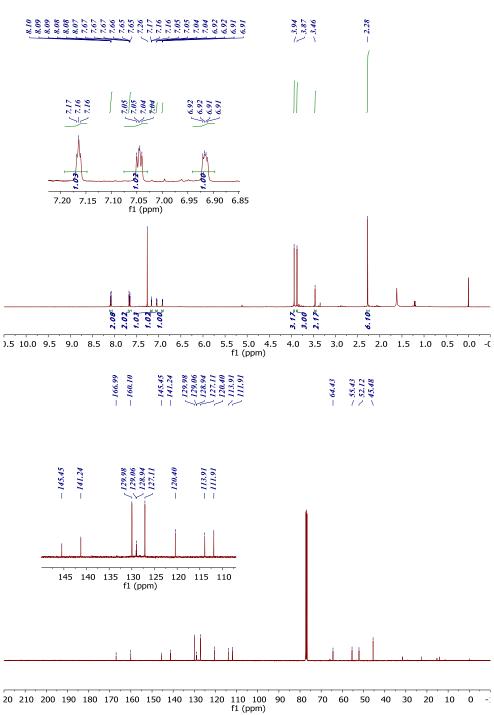
taken in DMSO-d6 at 100°C.

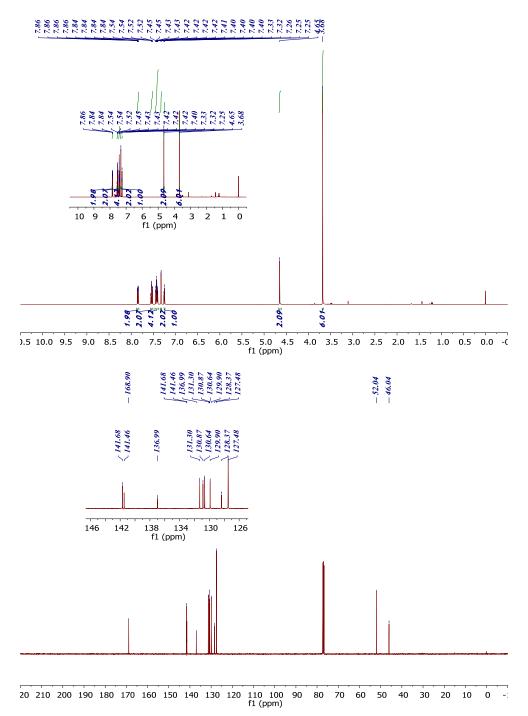


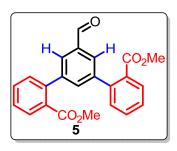


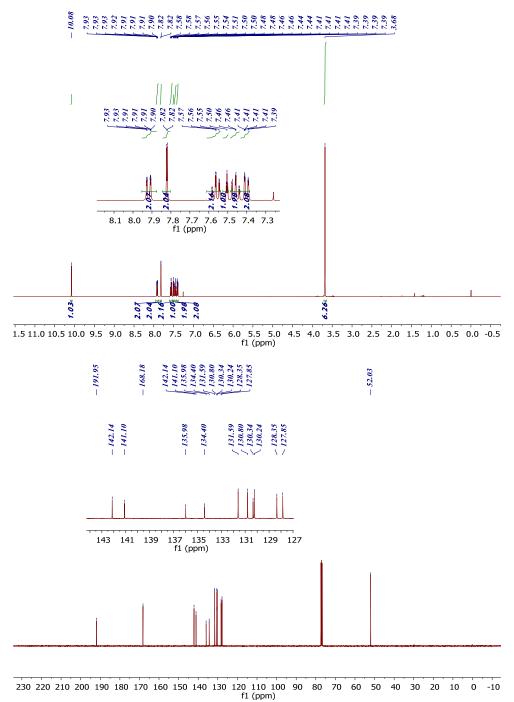


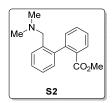


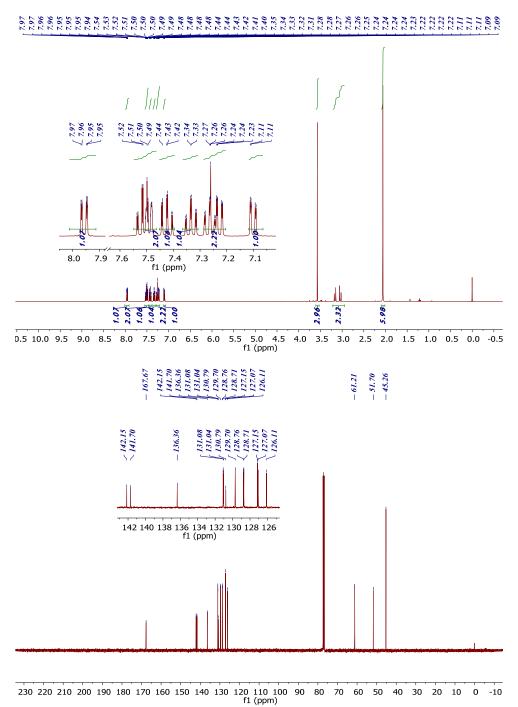


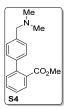


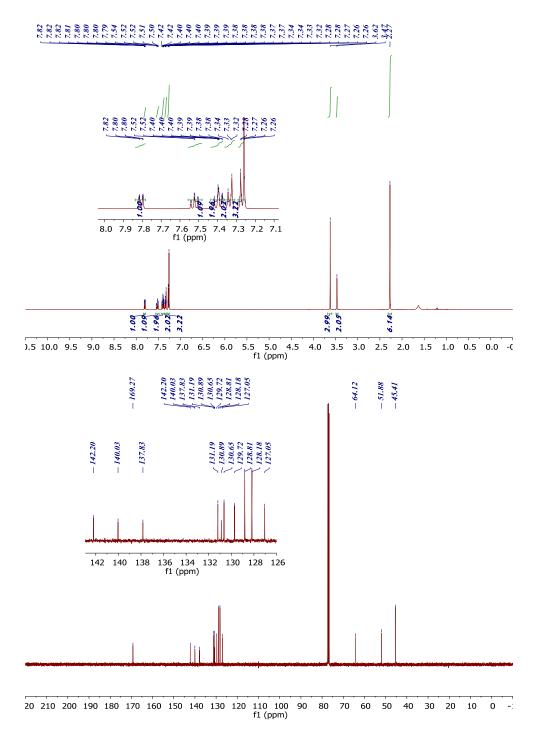












VI. References

- (1) Tayama, E.; Takedachi, K.; Iwamoto, H.; Hasegawa, E. Tetrahedron 2010, 66, 9389.
- (2) Waser, J.; Gaspar, B.; Nambu, H.; Carreira, E. M. J. Am. Chem. Soc. 2006, 128, 11693.
- (3) Knowles, J. P.; Oconnor, V. E.; Whiting, A. Org. Biomol. Chem. 2011, 9, 1876.
- (4) Wang, Y.; Gulevich, A. V.; Gevorgyan, V. Chem. Eur. J. 2013, 19, 15836.