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## **Increased Frequency of Nontuberculous Mycobacteria Detection at Potable Water Taps** within the United States

Number of Pages: 2 (includes coversheet)

Table S1: Summary of sampling process used by each NTM occurrence study

	Study	von Reyn et al., 1993	Glover et al., 1994	Covert <i>et al.,</i> 1999	Falkinham et al., 2001	Hilborn et al., 2006	This study
Sample collection	Flush time	First pull	First pull	1 to 2 min	Not reported	First pull	15 sec
	Sample preservative	Sodium thiosulfate	Sodium thiosulfate	Sodium thiosulfate	Sodium thiosulfate	Sodium thiosulfate	Not added
Sample treatment	Decontamination	None	CPC* %0.04	CPC %0.04	None	CPC %0.04	CPC %0.04
	Decontamination time		24 hr.	30 min		30 min	30 min
Sample concentrating	Concentration method	Centrifugation@ 5,000 g, Spread plate 0.1 mL	Vacuum filtration 500 mL	Vacuum filtration 500 mL	Centrifugation@ 5,000 g, Spread plate 0.1 mL	Vacuum filtration 500 mL	Vacuum filtration 500 mL
	Volume of water represented in NTM recovery	2.5 mL/plate 3 plates 7.5 mL analyzed	500 mL	500 mL	30 mL/plate 5 plates 150 mL analyzed	500 mL	500 mL
NTM isolation	Media	Tsukamura minimal- Tween 80 cycloheximide agar media	Middlebrook 7 H10	Middlebrook 7 H10	Middlebrook 7H10	Middlebrook 7H10	Middlebrook 7H10
	Incubation Time	Eight weeks	Eight weeks,	Eight weeks	Three weeks	Eight weeks	Eight weeks
	Incubation Temperature	37 °C	37 °C	37 °C	37 °C	37 °C	37 °C
Identification	Presumptive NTM	Acid fast	Acid fast	Acid fast	Acid fast	Acid fast	
	MAC identification	DNA probes (SNAP or Gen-Probe)	DNA probes (Syngene)	16S partial sequencing	Multiplex PCR	Multi locus electrophoresis	16S partial and hsp65 genes
	NTM identification excluding MAC isolates	Not done	Biochemical screening	16S partial sequencing	Endonuclease digest hsp65 (BstEll and Haelll)	HPLC mycolic acid	16S partial and hsp65 genes

<sup>\*</sup>CPC, cetylpyridinium chloride

Table S1: Summary of sampling approach used by each NTM occurrence study.