

Supporting Information

Sensomics Analysis of Key Bitter Compounds in the Hard Resin of Hops (*Humulus lupulus* L.) and their Contribution to the Bitter Profile of Pilsner-type Beer

Michael Dresel, Andreas Dunkel, and Thomas Hofmann*

Chair of Food Chemistry and Molecular and Sensory Science, Technische Universität

München, Lise-Meitner-Str. 34, D-84354 Freising, Germany

Chromatographic Separation of MPLC-Fractions of ϵ -Resin. MPLC fractions were separated using an HPLC system (Varian, Middelburg, Niederlande) consisting of two ProStar 210 pumps, a 7725i type Rheodyne injection valve (Rheodyne, Bensheim, Germany), and a ProStar 330 diode array detector monitoring the effluent between 220 and 400 nm. Chromatography was performed on a 250 x 21.2 mm, 5 μ m, Luna Phenyl-Hexyl column (Phenomenex, Aschaffenburg, Germany) operated with a flow rate of 15 mL/min and using water (0.1 % formic acid) as solvent A and acetonitrile (0.1 % formic acid) as solvent B. Data acquisition was performed by means of Star Chromatography Workstation Software, Version 6.2.

MPLC fraction e2 (yield: 3.35%). Aliquots of fraction e2 (100 mg) dissolved in acetonitrile (1.6 mL; 12.5%, v/v) were separated using the following gradient: 15% solvent

24 B for 4.0 min, increasing solvent B to 22% within 20 min, and, then, increasing solvent B to
25 28% within 1.0 min. Solvent B was kept for 16 min at 28% and, finally, increased to 100%
26 within an additional minute, followed by isocratic elution for 1.5 min. After 45 min, solvent B
27 decreased again to 15% within 1.5 min and was kept for 5.0 min prior to the next injection.
28 A total of 14 fractions, namely fractions e2-1 to e2-14, were collected, separated from
29 solvent in vacuum, and was, then, subjected to a taste dilution analysis (**Figure 3**).

30 *MPLC fraction e3 (yield: 2.99%).* Aliquots of subfraction e3 (90 mg) dissolved in
31 acetonitrile (1.6 mL; 20%, v/v) were separated using following gradient: 30% solvent B for
32 19.5 min, increasing solvent B to 40% within 1.5 min and, then, to 50% within 6.0 min.
33 Solvent B was kept for 4.0 min at 50% and, finally, increased to 100% within an additional
34 minute, followed by isocratic elution for 1.5 min. After 33.5 min, solvent B decreased again
35 to 30% within 1.5 min and was kept for 5.0 min prior to the next injection. A total of 13
36 fractions, namely fractions e3-1 to e3-13, were collected, separated from solvent in
37 vacuum, and was, then, subjected to a taste dilution Analysis (**Figure 3**).

38 *MPLC fraction e4 (yield: 3.20%).* Aliquots of subfraction e4 (50 mg) dissolved in
39 acetonitrile (1.8 mL; 33%, v/v) were separated using the following gradient: 32.5% solvent
40 B for 2.0 min and then increasing solvent B to 47% within 14 min. Solvent B was kept at
41 47% for 3.0 min and, finally, increased to 100% within an additional minute, followed by
42 isocratic elution for 3.0 min. After 23 min, solvent B decreased again to 32.5% within 2.0
43 min and was kept for 5.0 min prior to the next injection. A total of 5 fractions, namely
44 fractions e4-1 to e4-5, were collected, separated from solvent in vacuum, and subjected to
45 a taste dilution analysis (**Figure 3**).

46 *MPLC fraction e5 (yield: 3.64%).* Aliquots of subfraction e5 (50 mg) dissolved in
47 acetonitrile (1.5 mL, 33%, v/v) were separated using the following gradient: 36% solvent B
48 for 25 min, increasing solvent B to 70% within 5.0 min and, then, to 100% within an
49 additional minute, followed by isocratic elution for 7.5 min. After 38.5 min, solvent B

50 decreased again to 30% within 2.5 min and was kept for 4.0 min prior to the next injection.
51 A total of 5 fractions, namely fractions e5-1 to e5-5, were collected, separated from solvent
52 in vacuum, and subjected to a taste dilution analysis (**Figure 3**).

53 *MPLC fraction e6 (yield: 3.16%).* Aliquots of subfraction e6 (50 mg) dissolved in
54 acetonitrile (1.7 mL, 30%, v/v) were separated using the following gradient: 38% solvent B
55 for 25 min and, then, increasing solvent B to 55% within 0.5 min. Solvent B was kept at
56 55% for 5 min and, finally, increased to 100% within an additional 1.5 min, followed by
57 isocratic elution for 7.0 min. After 39 min, solvent B decreased again to 38% within 2.0 min
58 and was kept for 4.0 min prior to the next injection. A total of 11 fractions, namely fractions
59 e6-1 to e6-11, were collected, separated from solvent in vacuum, and subjected to a taste
60 dilution analysis (**Figure 3**).

61 *MPLC fraction e7 (yield: 2.59%).* Aliquots of subfraction e7 (25 mg) dissolved in
62 acetonitrile (1.8 mL, 40%, v/v) were separated using a gradient as follows: 40% solvent B
63 for 3.0 min and, then, increasing solvent B to 43% within 1.0 min. Solvent B was kept for
64 11 min at 43% and, then, increasing solvent B to 65% within 13 min. and, finally, increased
65 to 100% within an additional 4.0 min, followed by isocratic elution for 3.0 min. After 35 min,
66 solvent B decreased again to 40% within 3.0 min and was kept for 2.0 min prior to the next
67 injection. A total of 16 fractions, namely fractions e7-1 to e7-16, were collected, separated
68 from solvent in vacuum, and subjected to a taste dilution analysis (**Figure 3**).

69 *MPLC fraction e8 (yield: 2.69%).* Aliquots of subfraction e8 (25 mg) dissolved in
70 acetonitrile (1.7 mL, 45%, v/v) were separated using the following gradient: 47% solvent B
71 for 14 min and, then, increasing solvent B to 55% within 0.5 min. Solvent B was kept for
72 5.0 min at 55% and, finally, increased to 100% within an additional 10 min, followed by
73 isocratic elution for 3.5 min. After 33 min, solvent B decreased again to 47% within 1.0 min
74 and was kept for 3.0 min prior to the next injection. A total of 8 fractions, namely fractions

75 e8-1 to e8-8, were collected, separated from solvent in vacuum, and subjected to a taste
76 dilution analysis (**Figure 3**).

77 *MPLC fraction e9 (yield: 18.38%).* Aliquots of subfraction e9 (20 mg) dissolved in
78 acetonitrile (1.7 mL, 35%, v/v) were separated using the following gradient: Starting with
79 48%, solvent B is increasing to 50% within 7.0 min, and was kept for 28.5 min at 50% and,
80 finally, increased to 100% within an additional 1.5 min, followed by isocratic elution for 7.0
81 min. After 44 min, solvent B decreased again to 48% within 2.0 min and was kept for 4.0
82 min prior to the next injection. A total of 9 fractions, namely fractions e9-1 to e9-9, were
83 collected, separated from solvent in vacuum, and subjected to a taste dilution analysis
84 (**Figure 3**).

85 *MPLC fraction e10 (yield: 31.33%).* Aliquots of subfraction e10 (50 mg) dissolved in
86 acetonitrile (1.7 mL, 40%, v/v) were separated using the following gradient: 57% solvent B
87 for 32 min and, then, increasing solvent B to 73% within 1.0 min. Solvent B was kept for
88 8.5 min at 73% and, finally, increased to 100% within an additional 0.5 min, followed by
89 isocratic elution for 3.0 min. After 45 min, solvent B decreased again to 57% within 1.5 min
90 and was kept for 3.5 min prior to the next injection. A total of 13 fractions, namely fractions
91 e10-1 to e10-13, were collected, separated from solvent in vacuum, and subjected to a
92 taste dilution analysis (**Figure 3**).

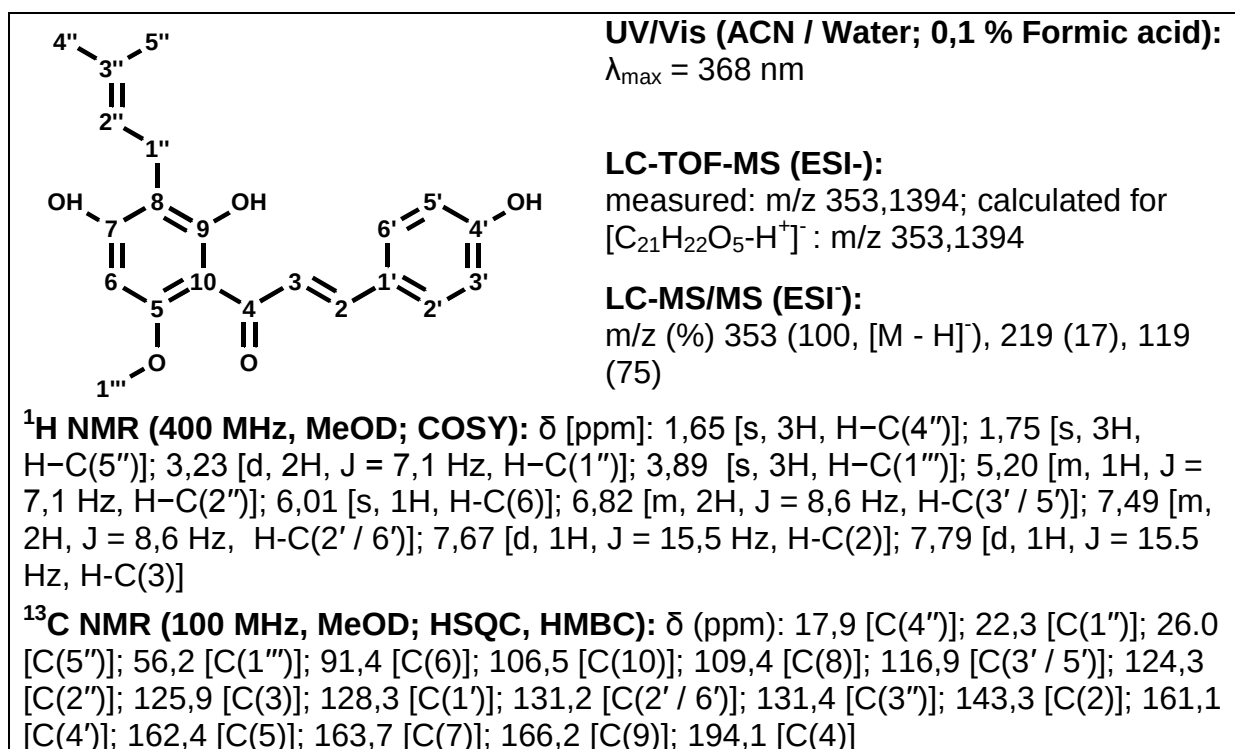
93 *MPLC fraction e11 (yield: 22,86%).* Aliquots of subfraction e11 (200 mg) dissolved
94 in acetonitrile (1.5 mL, 60%, v/v) were separated using the following gradient: 65% solvent
95 B for 1.0 min and, then, increasing solvent B to 68% within 7.0 min. Solvent B increased to
96 70% within 1 min and was kept for 1 min at 70% before solvent B increased to 90% within
97 28 min and, finally, increased to 100% within an additional 3.0 min, followed by isocratic
98 elution for 4.0 min. After 45 min, solvent B decreased again to 65% within 2.0 min and was
99 kept for 3.0 min prior to the next injection. A total of 15 fractions, namely fractions e11-1 to

100 e11-15, were collected, separated from solvent in vacuum, and subjected to a taste
101 dilution analysis (**Figure 3**).

102

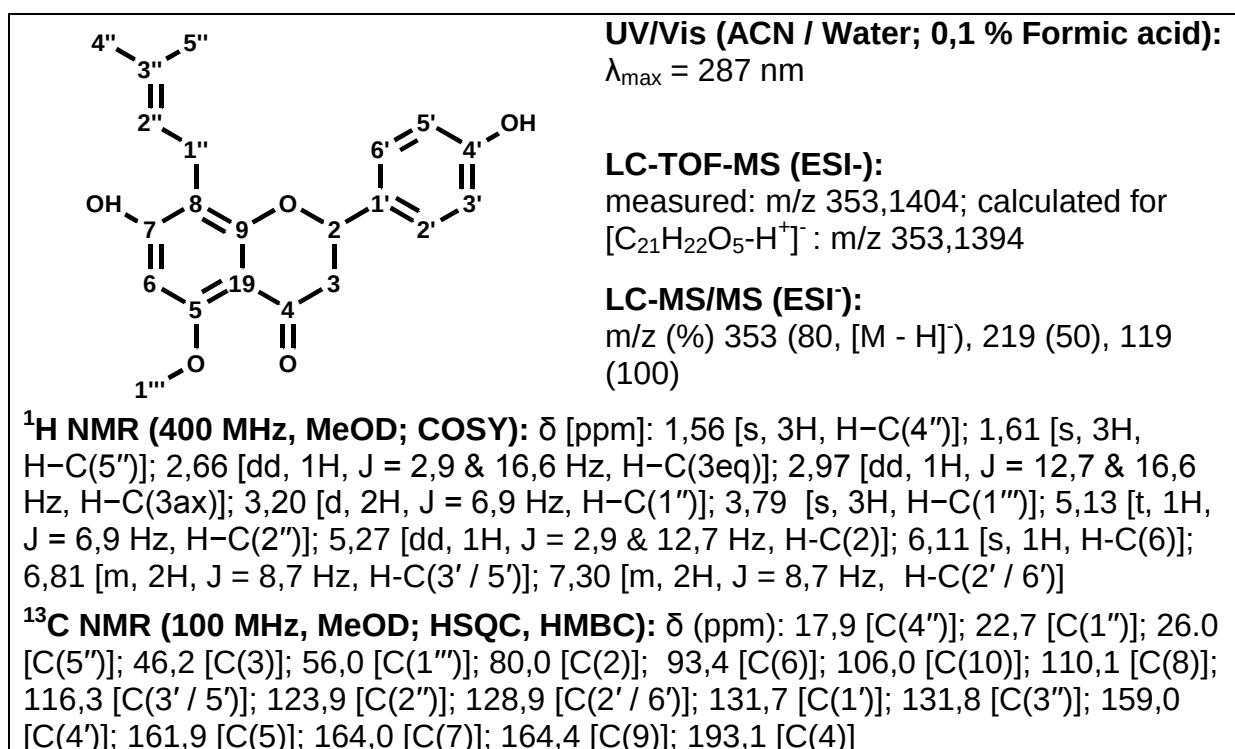
Spectroscopic data of all isolated and synthesized compounds.

103 Xanthohumol (5)

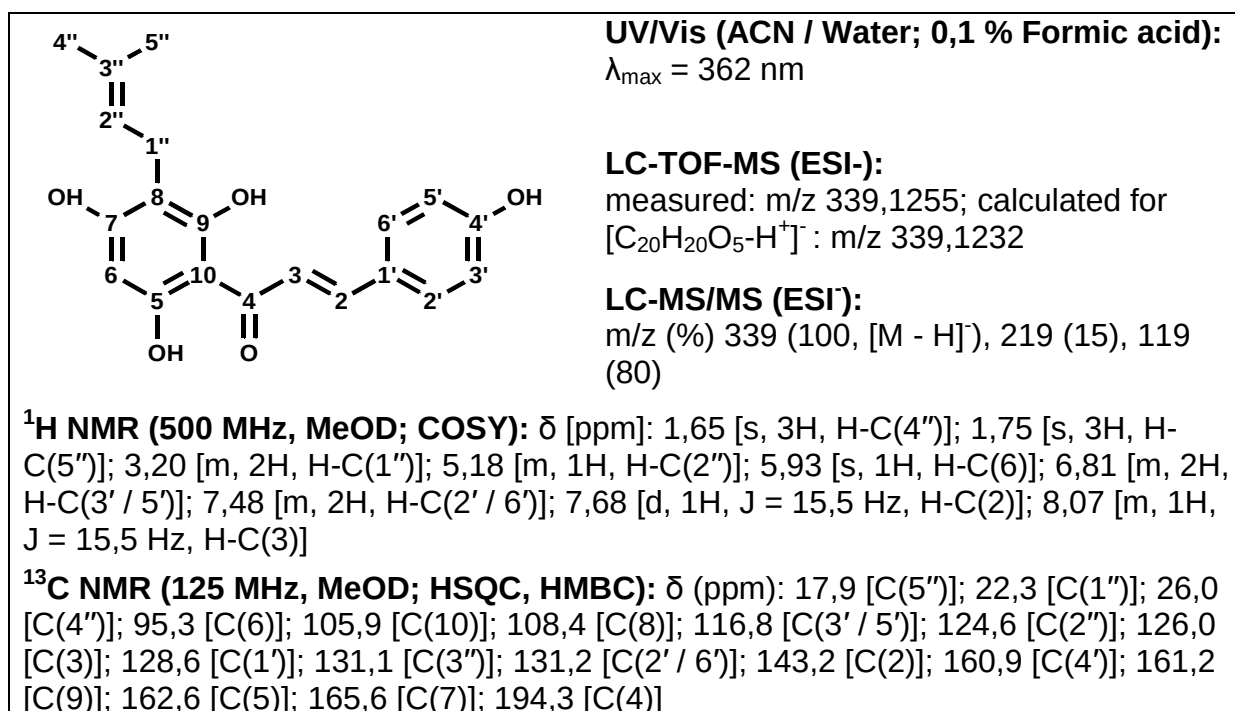


104

105 Isoxanthohumol (6)

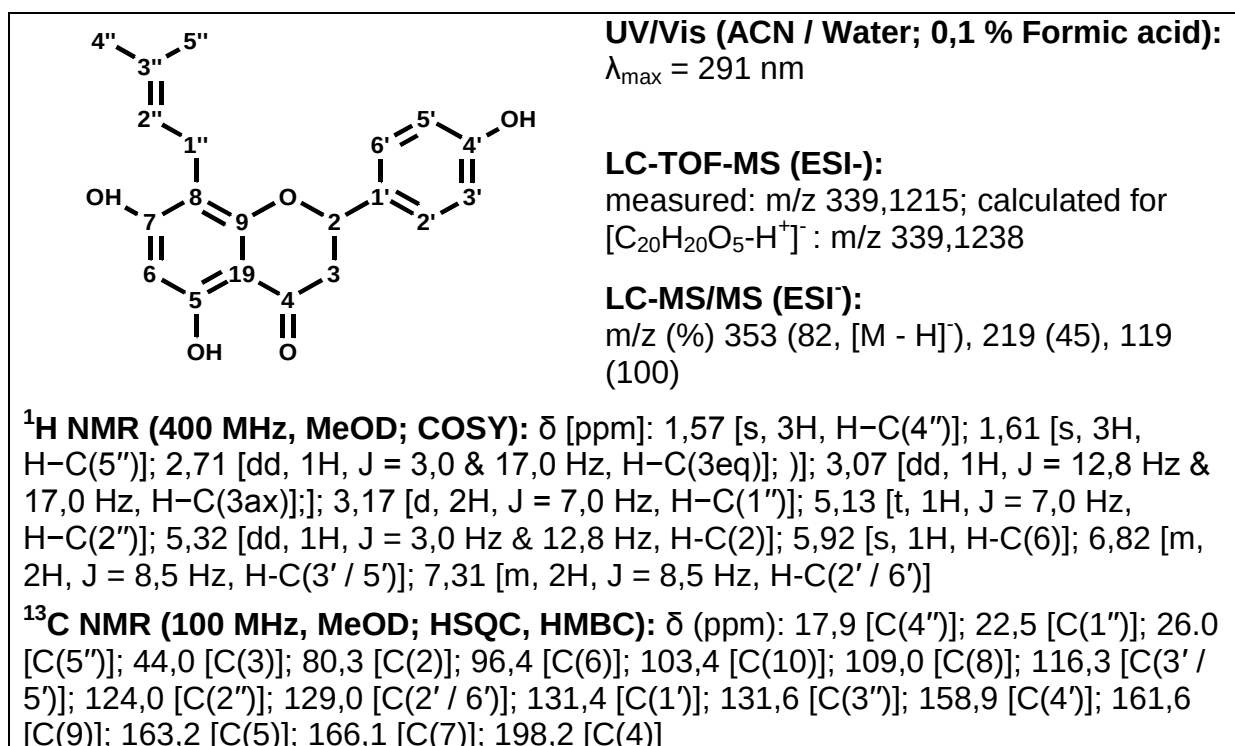


106 Desmethylxanthohumol (7)

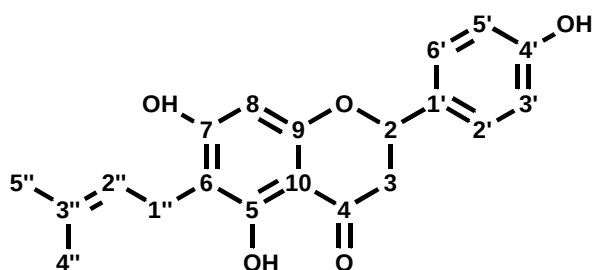


107

108 8-Prenylnaringenin (8)



109



UV/Vis (ACN / Water; 0,1 % Formic acid):

$\lambda_{\max} = 291 \text{ nm}$

LC-TOF-MS (ESI-):

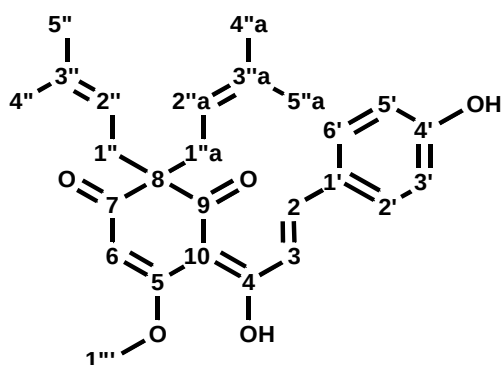
measured: m/z 339,1215; calculated for $[\text{C}_{20}\text{H}_{20}\text{O}_5\text{-H}^+]^-$: m/z 339,1238

LC-MS/MS (ESI-):

m/z (%) 353 (78, $[\text{M} - \text{H}]^-$), 219 (47), 119 (100)

^1H NMR (500 MHz, MeOD; COSY): δ [ppm]: 1,65 [s, 3H, H-C(4'')]; 1,75 [s, 3H, H-C(5'')]; 2,68 [dd, 1H, $J = 3,1 \text{ Hz} \& 16,9 \text{ Hz}$, H-C(3eq)];); 3,09 [dd, 1H, $J = 13,1 \& 16,9 \text{ Hz}$, H-C(3ax)]; 3,20 [d, 2H, $J = 6,9 \text{ Hz}$, H-C(1'')]; 5,18 [t, 1H, $J = 6,9 \text{ Hz}$, H-C(2'')]; 5,30 [dd, 1H, $J = 3,1 \text{ Hz} \& 13,1 \text{ Hz}$, H-C(2)]; 5,93 [s, 1H, H-C(6)]; 6,81 [m, 2H, $J = 8,5 \text{ Hz}$, H-C(3' / 5')]; 7,31 [m, 2H, $J = 8,5 \text{ Hz}$, H-C(2' / 6')]

^{13}C NMR (125 MHz, MeOD; HSQC, HMBC): δ (ppm): 17,8 [C(4'')]; 21,9 [C(1'')]; 26,0 [C(5'')]; 44,2 [C(3)]; 80,4 [C(2)]; 95,4 [C(6)]; 103,2 [C(10)]; 109,7 [C(8)]; 116,3 [C(3' / 5')]; 123,9 [C(2'')]; 129,0 [C(2' / 6')]; 131,3 [C(1')]; 131,6 [C(3'')]; 159,0 [C(4')]; 162,4 [C(9)]; 162,6 [C(5)]; 166,1 [C(7)]; 197,9 [C(4)]



UV/Vis (ACN / Water; 0,1 % Formic acid):

$\lambda_{\max} = 406 \text{ nm}$

LC-TOF-MS (ESI-):

measured: m/z 421,2014; calculated for $[\text{C}_{26}\text{H}_{30}\text{O}_5\text{-H}^+]^-$: m/z 421,2015

LC-MS/MS (ESI-):

m/z (%) 421 (12, $[\text{M} - \text{H}]^-$), 245 (1), 217 (2), 119 (100), 93 (3)

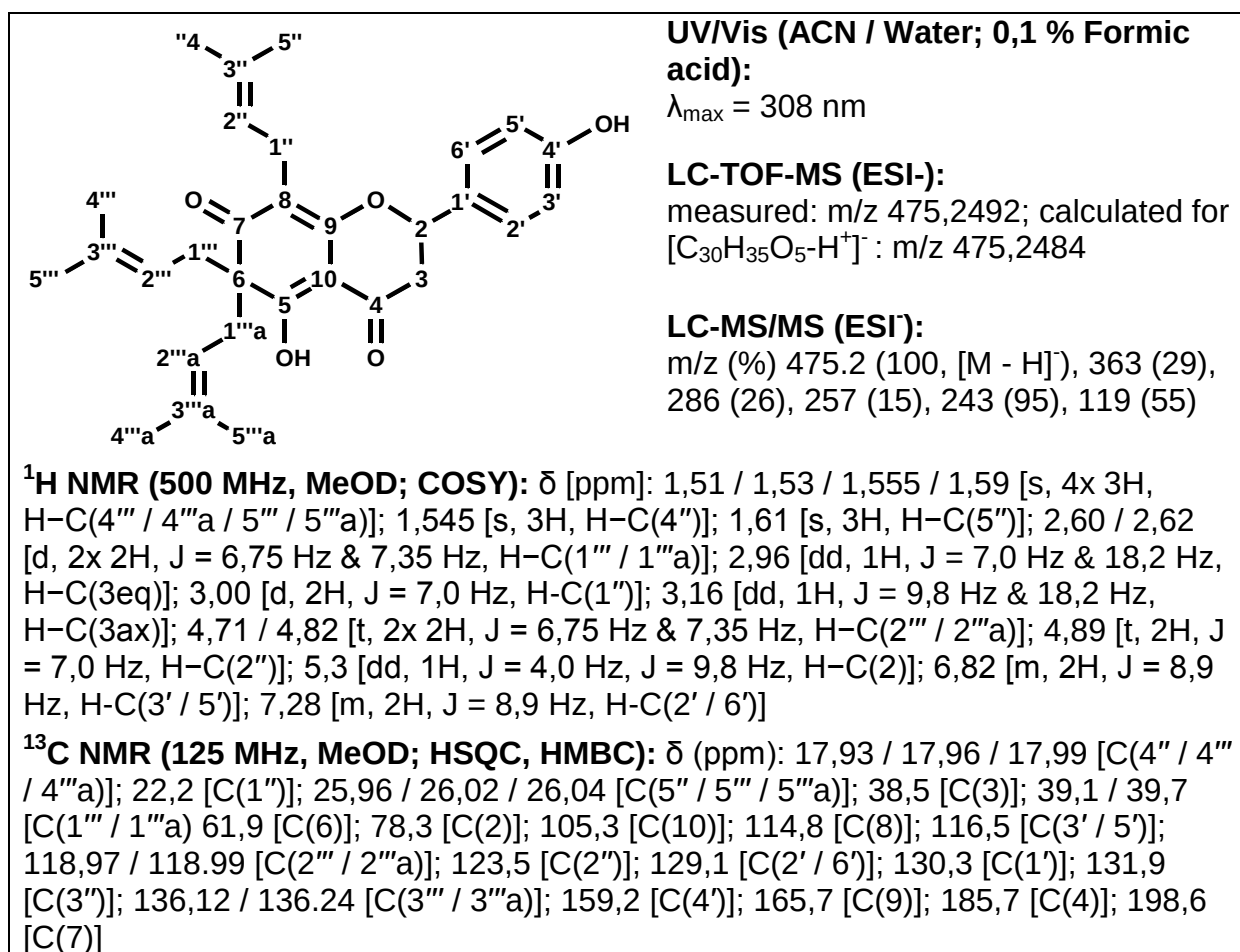
^1H NMR (500 MHz, MeOD; COSY): δ [ppm]: 1,56 [s, 12H, H-C(4'' / 4''a / 5'' / 5''a)]; 2,58 [m, 3H, H-C(1'' / 1''a)]; 3,99 [s, 3H, H-C(1''')]; 4,84 [m, 2H, H-C(2'' / 2''a)]; 5,56 [s, 1H, H-C(6)]; 6,85 [m, 2H, $J = 8,9 \text{ Hz}$, H-C(3' / 5')]; 7,54 [m, 2H, $J = 8,9 \text{ Hz}$, H-C(2' / 6')]; 7,61 [d, 1H, $J = 15,7 \text{ Hz}$, H-C(3)]; 7,88 [d, 1H, $J = 15,7 \text{ Hz}$, H-C(2)]

^{13}C NMR (125 MHz, MeOD; HSQC, HMBC): δ (ppm): 17,9 [C(5'' / 5''a)]; 25,8 [C(4'' / 4''a)]; 39,5 [C(1'' / 1''a)]; 57,1 [C(1''')]; 62,6 [C(8)]; 98,9 [C(6)]; 106,1 [C(10)]; 116,6 [C(3' / 5')]; 118,8 [C(2'' / 2''a)]; 119,1 [C(3)]; 128,1 [C(1')]; 131,8 [C(2' / 6')]; 136,2 [C(3'' / 3''a)]; 145,9 [C(2)]; 161,9 [C(4')]; 174,3 [C(5)]; 180,3 [C(4)]; 199,8 [C(7)]; 205,9 [C(9)]

^1H NMR (500 MHz, DMSO; COSY): δ [ppm]: 1,50 [s, 6H, H-C(4'' / 4''a)]; 1,52 [s, 6H, H-C(5'' / 5''a)]; 2,52 [m, 3H, $J = 7,5 \text{ Hz}$, H-C(1'' / 1''a)]; 3,94 [s, 3H, H-C(1''')]; 4,78 [t, 2H, $J = 7,5 \text{ Hz}$, H-C(2'' / 2''a)]; 5,52 [s, 1H, H-C(6)]; 6,86 [m, 2H, $J = 8,6 \text{ Hz}$, H-C(3' / 5')]; 7,59 [d, 1H, $J = 15,5 \text{ Hz}$, H-C(3)]; 7,63 [m, 2H, $J = 8,6 \text{ Hz}$, H-C(2' / 6')]; 7,84 [d, 1H, $J = 15,5 \text{ Hz}$, H-C(2)]

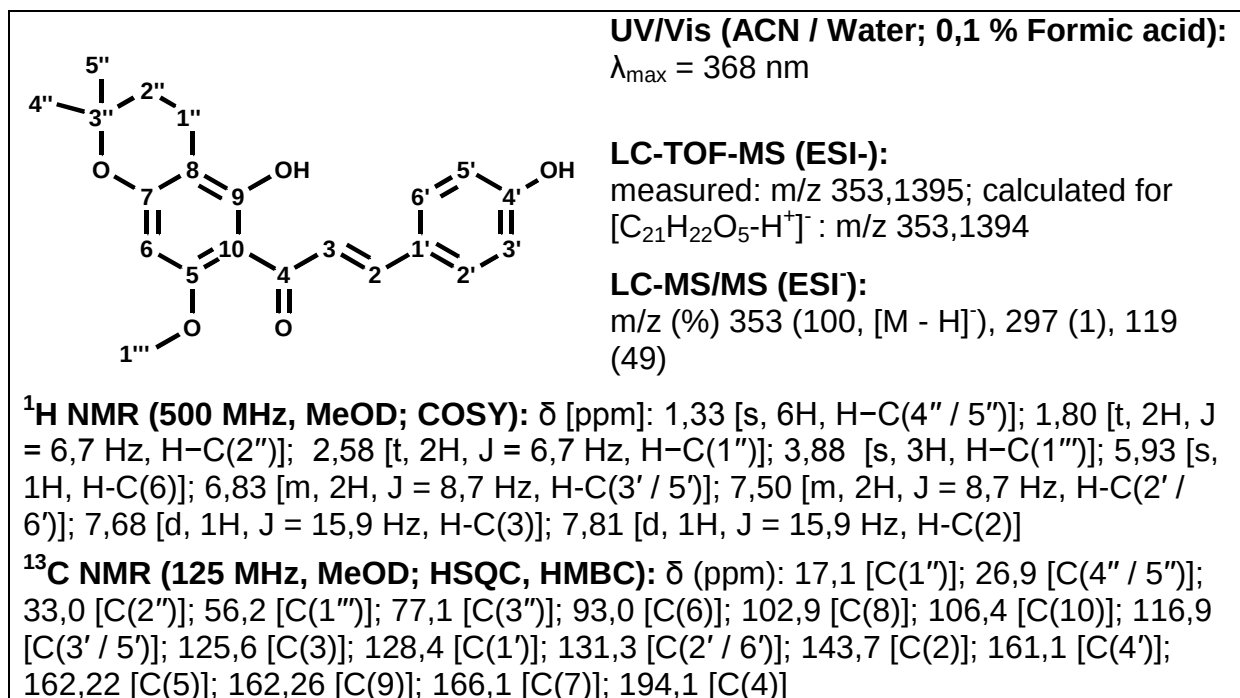
^{13}C NMR (125 MHz, DMSO; HSQC, HMBC): δ (ppm): 18,6 [C(5'' / 5''a)]; 26,6 [C(4'' / 4''a)]; 38,6 [C(1'' / 1''a)]; 57,6 [C(1''')]; 61,8 [C(8)]; 99,2 [C(6)]; 105,9 [C(10)]; 117,1 [C(3' / 5')]; 119,1 [C(2'' / 2''a)]; 119,3 [C(3)]; 126,8 [C(1')]; 132,1 [C(2' / 6')]; 135,2 [C(3'' / 3''a)]; 145,4 [C(2)]; 161,6 [C(4')]; 171,5 [C(5)]; 179,7 [C(4)]; 196,4 [C(7)]; 205,7 [C(9)]

114 Isoxantholupon (**11**)



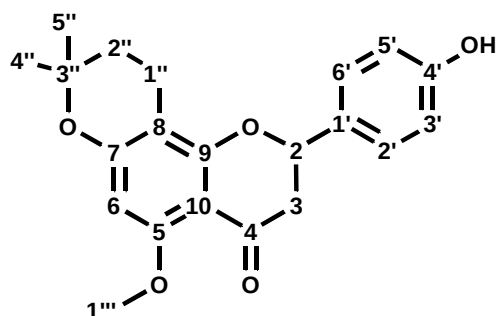
115

116 1'',2''-Dihydroxanthohumol C (**12**)



117

118 1'',2''-Dihydroisoxanthohumol C (13)



UV/Vis (ACN / Water; 0,1 % Formic acid):

$\lambda_{\max} = 286 \text{ nm}$

LC-TOF-MS (ESI-):

measured: m/z 369,1335; calculated for $[\text{C}_{21}\text{H}_{22}\text{O}_5\text{-H}^+]^-$: m/z 369,1338

LC-MS/MS (ESI-):

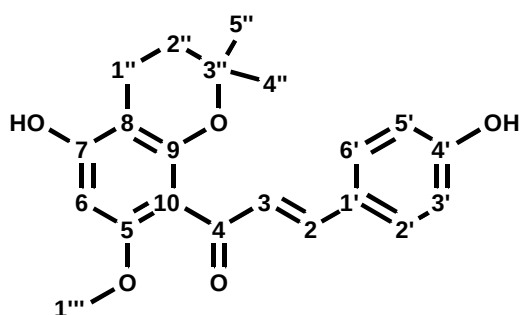
m/z (%) 369 (30, $[\text{M} - \text{H}]^-$), 119 (100)

^1H NMR (500 MHz, MeOD; COSY): δ [ppm]: 1,32 / 1,33 [s, 2x 3H, H-C(4'' / 5'')]; 1,78 [m, 2H, H-C(2'')]; 2,58 [dt, 1H, J = 1,1 Hz & 6,8 Hz, H-C(1'')]; 2,68 [dd, 1H, J = 3,2 Hz & 16,6 Hz, H-C(3eq)]; 3,00 [dd, 1H, J = 13,0 Hz & 16,6 Hz, H-C(3ax)]; 3,78[s, 3H, H-C(1''')]; 5,35 [dd, 1H, J = 3,2 Hz & 13,0 Hz, H-C(2)]; 6,04 [s, 1H, H-C(6)]; 6,82 [m, 2H, J = 8,5 Hz, H-C(3' / 5')]; 7,32 [m, 2H, J = 8,5 Hz, H-C(2' / 6')]

^{13}C NMR (125 MHz, MeOD; HSQC, HMBC): δ (ppm): 17,5 [C(1'')]; 26,4 / 27,2 [C(4'' / 5'')]; 329 [C(2'')]; 46,0 [C(3'')]; 56,1 [C(1''')]; 77,2 [C(3'')]; 80,1 [C(2)]; 94,9 [C(6)]; 103,2 [C(10)]; 105,9 [C(8)]; 116,3 [C(3' / 5')]; 128,8 [C(2' / 6')]; 158,9 [C(4')]; 161,7 [C(5)]; 162,7 [C(9)]; 163,6 [C(7)]; 192,7 [C(4)]

119

120 1'',2''-Dihydroxanthohumol K (14)



UV/Vis (ACN / Water; 0,1 % Formic acid):

$\lambda_{\max} = 325 \text{ nm}$

LC-TOF-MS (ESI-):

measured: m/z 353,1392; calculated for $[\text{C}_{21}\text{H}_{22}\text{O}_5\text{-H}^+]^-$: m/z 353,1394

LC-MS/MS (ESI-):

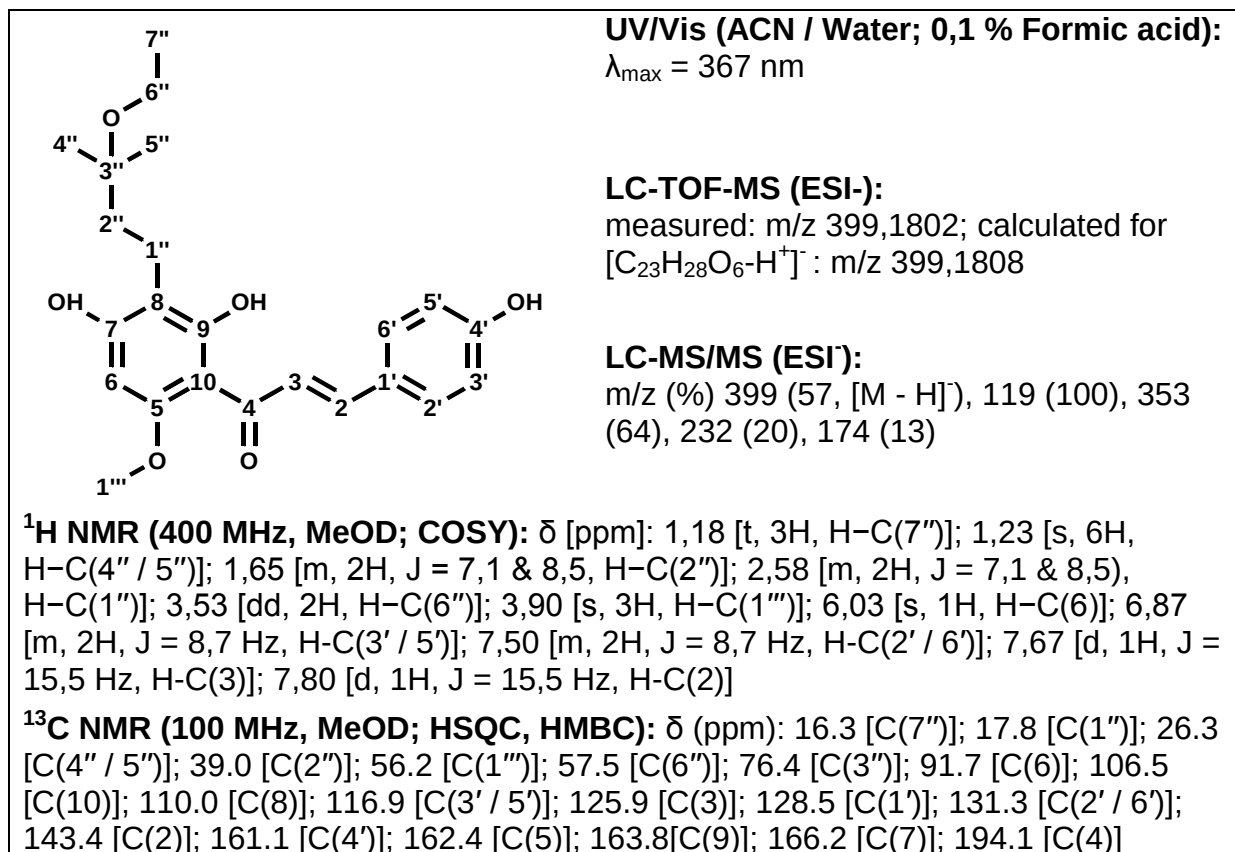
m/z (%) 353 (100, $[\text{M} - \text{H}]^-$), 233 (9), 119 (47)

^1H NMR (400 MHz, MeOD; COSY): δ [ppm]: 1,22 [s, 6H, H-C(4'' / 5'')]; 1,76 [t, 2H, J = 6,9 Hz, H-C(2'')]; 2,62 [t, 2H, J = 6,9 Hz, H-C(1'')]; 3,69 [s, 3H, H-C(1''')]; 6,11 [s, 1H, H-C(6)]; 6,77 [d, 1H, J = 16,0 Hz, H-C(3)]; 6,80 [m, 2H, J = 8,7 Hz, H-C(3' / 5')]; 7,23 [d, 1H, J = 16,0 Hz, H-C(2)]; 7,41 [m, 2H, J = 8,7 Hz, H-C(2' / 6')];

^{13}C NMR (100 MHz, MeOD; HSQC, HMBC): δ (ppm): 16,9 [C(1'')]; 26,1 [C(4'' / 5'')]; 32,3 [C(2'')]; 55,3 [C(1''')]; 75,0 [C(3'')]; 91,1 [C(6)]; 101,9 [C(8)]; 110,1 [C(10)]; 116,2 [C(3' / 5')]; 126,4 [C(3)]; 126,8 [C(1)]; 130,6 [C(2' / 6')]; 146,3 [C(2)]; 153,6 [C(7)]; 157,3 [C(5)]; 158,1 [C(9)]; 160,6 [C(4')]; 197,6 [C(4)]

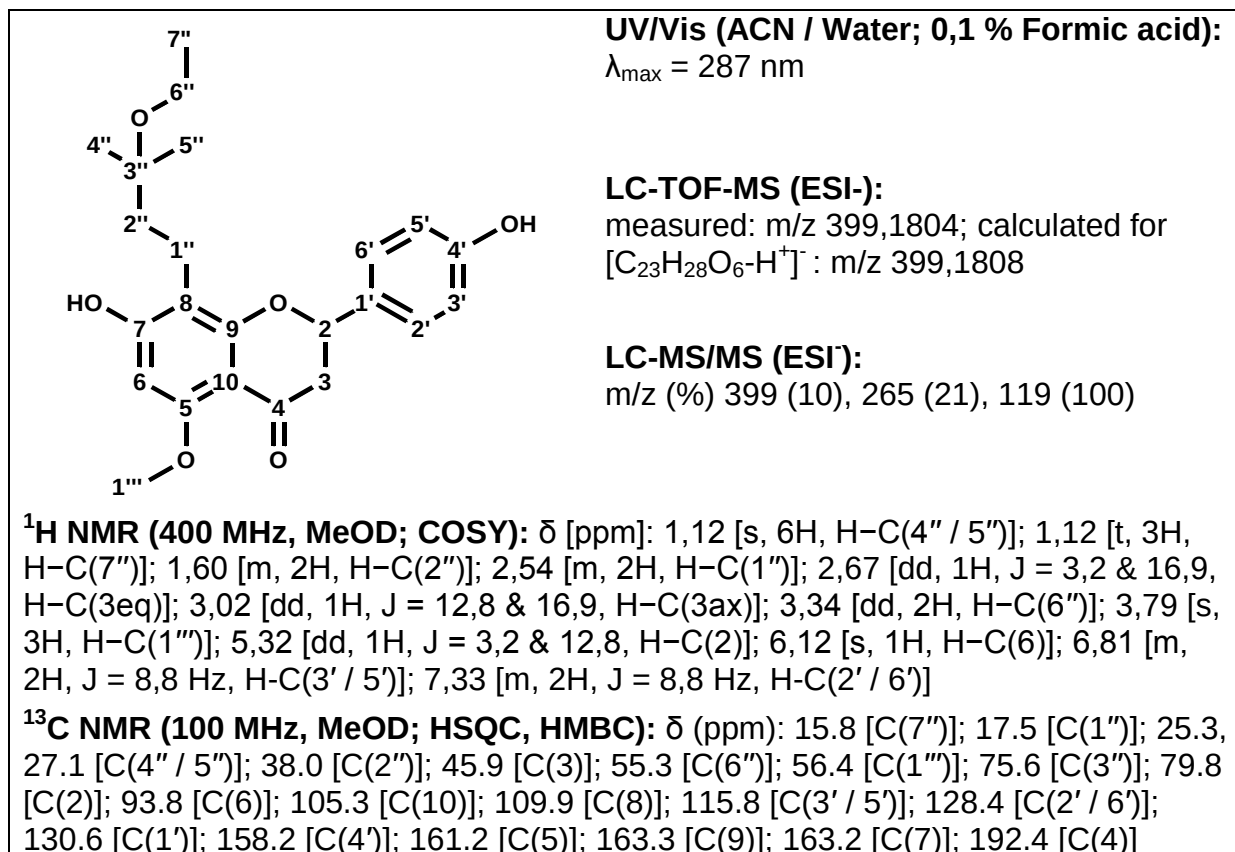
121

122 Xanthohumol P (15)

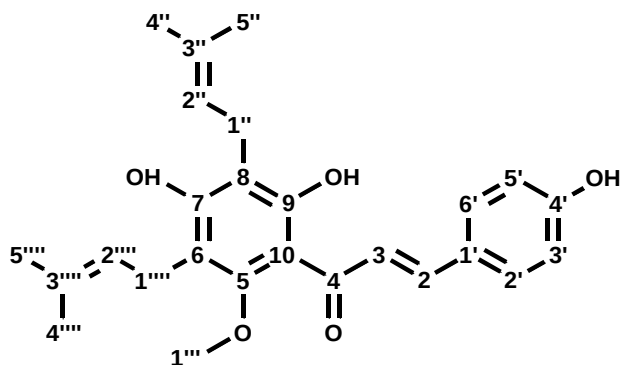


123

124 Isoxanthohumol P (16)



125



UV/Vis (ACN / Water; 0,1 % Formic acid):

$\lambda_{\max} = 368 \text{ nm}$

LC-TOF-MS (ESI-):

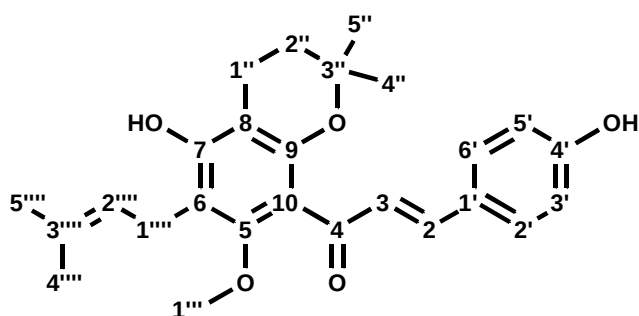
measured: m/z 421,2005; calculated for $[\text{C}_{26}\text{H}_{30}\text{O}_5\text{-H}^+]^-$: m/z 421,2015

LC-MS/MS (ESI-):

m/z (%) 421 (100, $[\text{M} - \text{H}]^-$), 301 (4), 286 (2), 243 (7), 119 (48)

^1H NMR (500 MHz, MeOD; COSY): δ [ppm]: 1,67 [s, 3H, H-C(5''')]; 1,70 [s, 3H, H-C(5'')]; 1,78 [s, 3H, H-C(4''')]; 1,80 [s, 3H, H-C(4'')]; 3,33 [d, 2H, $J = 7,0 \text{ Hz}$, H-C(1'')]; 3,36 [d, 2H, $J = 7,1 \text{ Hz}$, H-C(1''')]; 3,62 [s, 3H, H-C(1''')]; 5,16 [m, 1H, $J = 7,0 \text{ Hz}$, H-C(2'')]; 5,18 [m, 1H, $J = 7,1 \text{ Hz}$, H-C(2''')]; 6,84 [m, 2H, $J = 8,5 \text{ Hz}$, H-C(3' / 5')]; 7,53 [m, 2H, $J = 8,5 \text{ Hz}$, H-C(2' / 6')]; 7,78 [d, 1H, $J = 15,6 \text{ Hz}$, H-C(2)]; 7,83 [d, 1H, $J = 15,6 \text{ Hz}$, H-C(3)]

^{13}C NMR (125 MHz, MeOD; HSQC, HMBC): δ (ppm): 17,98 / 18,03 [C(4'' / 4''')]; 22,7 / 23,5 [C(1'' / 1''')]; 25,91 / 25,96 [C(5'' / 5''')]; 63,6 [C(1'')]; 109,8 [C(10)]; 113,1 [C(8)]; 115,8 [C(6)]; 117,0 [C(3' / 5')]; 123,43 [C(2''')]; 123,48 [C(2'')]; 124,5 [C(3)]; 128,2 [C(1')]; 131,5 [C(2' / 6')]; 132,4 [C(3'')]; 132,6 [C(3''')]; 144,7 [C(2)]; 160,3 [C(5)]; 161,3 [C(4')]; 161,6 [C(9)]; 162,9 [C(7)]; 194,6 [C(4)]



UV/Vis (ACN / Water; 0,1 % Formic acid):

$\lambda_{\text{max}} = 326 \text{ nm}$

LC-TOF-MS (ESI-):

measured: m/z 421,2008; calculated for $[\text{C}_{26}\text{H}_{30}\text{O}_5\text{-H}^+]^-$: m/z 421,2015

LC-MS/MS (ESI-):

m/z (%) 421 (100, $[\text{M} - \text{H}]^-$), 119 (98), 365 (10)

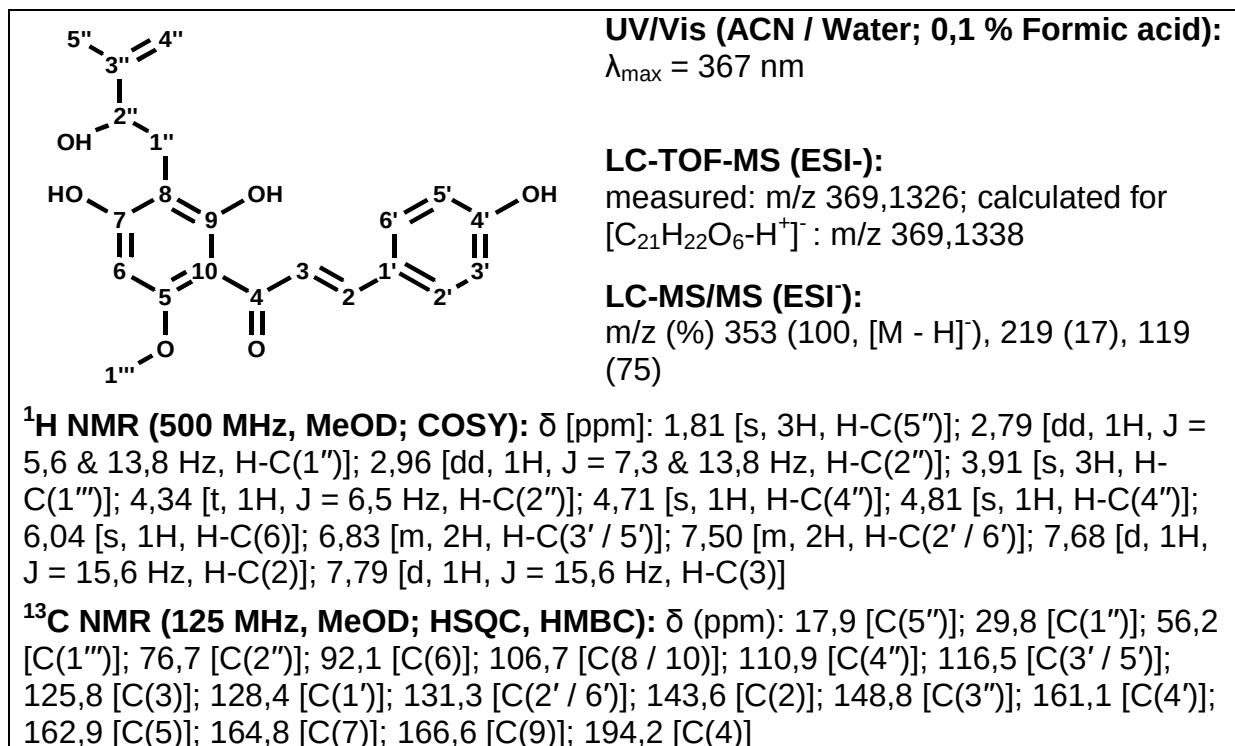
^1H NMR (500 MHz, MeOD; COSY): δ [ppm]: 1,22 [s, 6H, H-C(4'' / 5'')]; 1,68 [s, 3H, H-C(4''')]; 1,76 [s, 3H, H-C(5''')]; 1,79 [t, 2H, $J = 6,8 \text{ Hz}$, H-C(2'')]; 2,65 [t, 2H, $J = 6,8 \text{ Hz}$, H-C(1'')]; ~3,30 (unter Methanolsignal) [2H, H-C(1''')]; 3,61 [s, 3H, H-C(1''')]; 5,20 [m, 1H, H-C(2''')]; 6,80 [m, 2H, H-C(3' / 5')]; 6,82 [d, 1H, $J = 15,9 \text{ Hz}$, H-C(2)]; 7,25 [d, 1H, $J = 15,9 \text{ Hz}$, H-C(3)]; 7,43 [m, 2H, H-C(2' / 6')]

^{13}C NMR (125 MHz, MeOD; HSQC, HMBC): δ (ppm): 18,0 [C(5''')]; 18,2 [C(1'')]; 23,5 [C(1''')]; 25,9 [C(4''')]; 26,8 [C(4'' / 5'')]; 33,1 [C(2'')]; 63,1 [C(1''')]; 75,4 [C(3'')]; 106,6 [C(8)]; 115,1 [C(6)]; 116,4 [C(10)]; 117,0 [C(3' / 5')]; 124,8 [C(2''')]; 127,0 [C(1')]; 127,5 [C(3)]; 131,5 [C(2' / 6')]; 131,9 [C(3''')]; 147,6 [C(2)]; 151,7 [C(9)]; 156,3 [C(5)]; 156,4 [C(7)]; 161,5 [C(4')]; 198,3 [C(4)]

^1H NMR (500 MHz, DMSO; COSY): δ [ppm]: 1,15 [s, 6H, H-C(4'' / 5'')]; 1,63 [s, 3H, H-C(4''')]; 1,70 [s, 3H, H-C(5''')]; 1,72 [t, 2H, $J = 6,8 \text{ Hz}$, H-C(2'')]; 2,57 [t, 2H, $J = 6,8 \text{ Hz}$, H-C(1'')]; 3,20 [d, 2H, H-C(1''')]; 3,53 [s, 3H, H-C(1''')]; 5,14 [m, 1H, H-C(2''')]; 6,76 [d, 1H, $J = 15,9 \text{ Hz}$, H-C(2)]; 6,78 [m, 2H, H-C(3' / 5')]; 7,10 [d, 1H, $J = 15,9 \text{ Hz}$, H-C(3)]; 7,47 [m, 2H, H-C(2' / 6')]

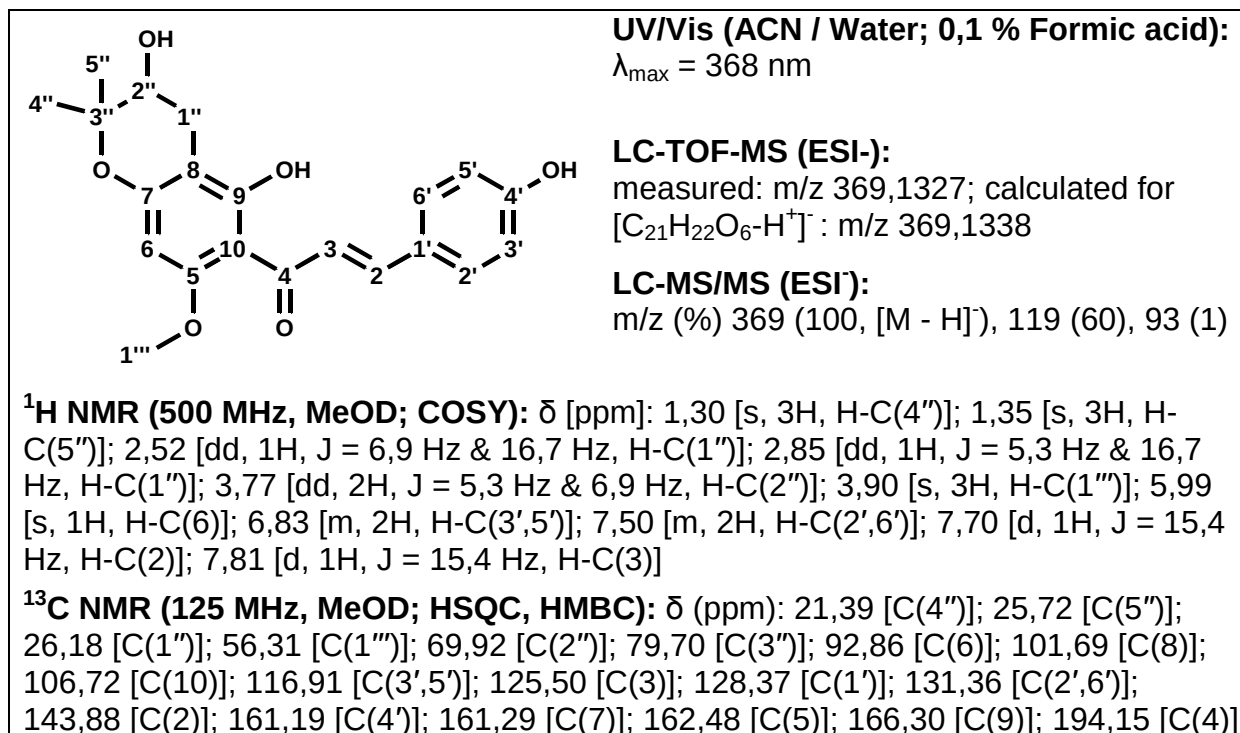
^{13}C NMR (125 MHz, DMSO; HSQC, HMBC): δ (ppm): 16,8 [C(1'')]; 17,9 [C(5''')]; 22,5 [C(1''')]; 25,6 [C(4''')]; 26,4 [C(4'' / 5'')]; 31,6 [C(2'')]; 62,3 [C(1''')]; 74,0 [C(3'')]; 105,2 [C(8)]; 113,5 [C(6)]; 115,5 [C(10)]; 116,1 [C(3' / 5')]; 124,0 [C(2''')]; 125,6 [C(1')]; 126,2 [C(3)]; 129,9 [C(2' / 6')]; 130,4 [C(3''')]; 143,9 [C(2)]; 149,9 [C(9)]; 154,5 [C(5)]; 154,7 [C(7)]; 159,9 [C(4')]; 193,9 [C(4)]

130 Xanthohumol D (19)



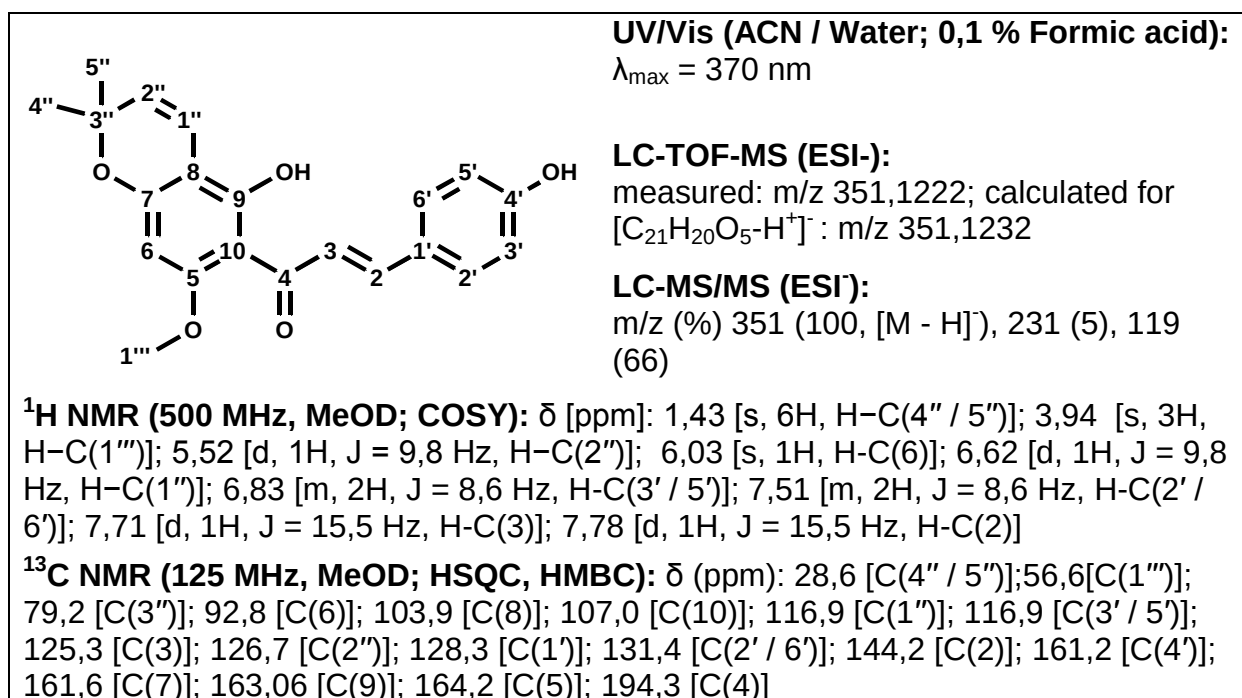
131

132 Xanthohumol B (20)



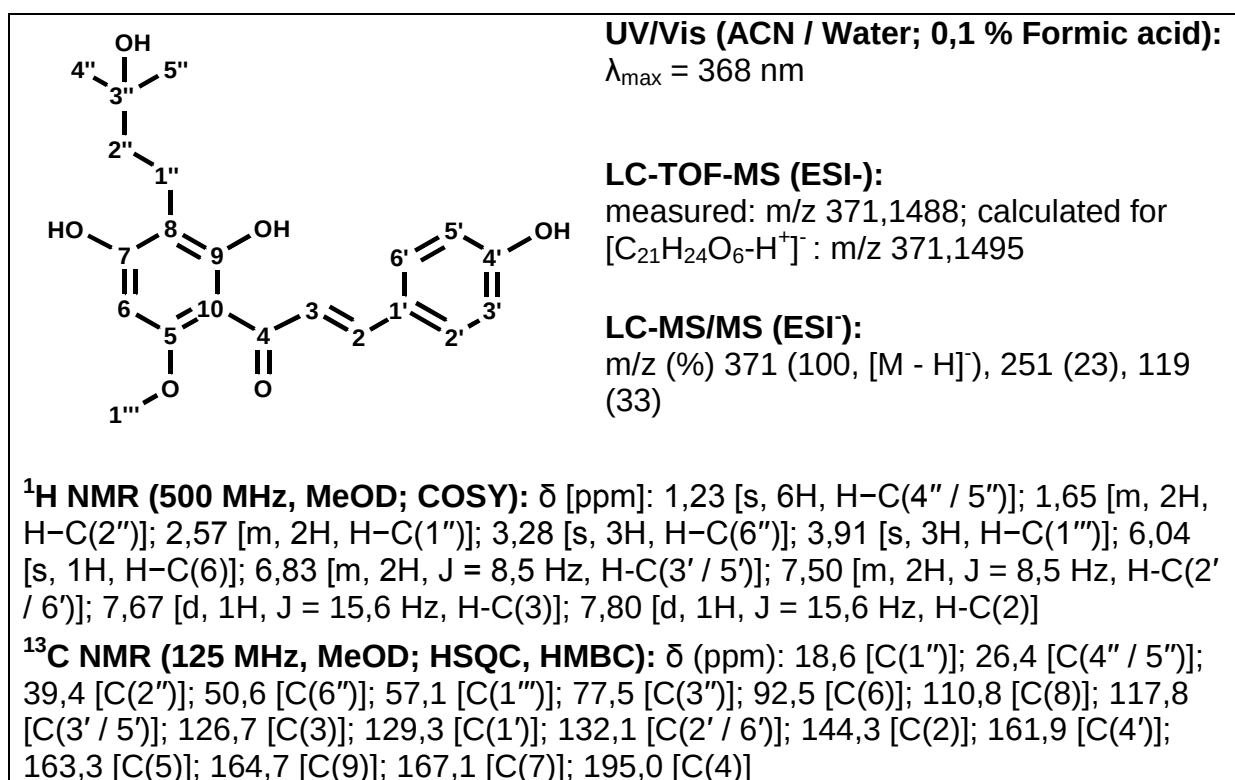
133

134 Xanthohumol C (21)

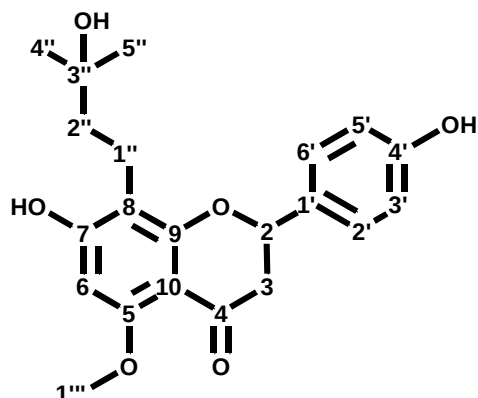


135

136 Xanthohumol H (22)



137



UV/Vis (ACN / Water; 0,1 % Formic acid):

$\lambda_{\max} = 286 \text{ nm}$

LC-TOF-MS (ESI-):

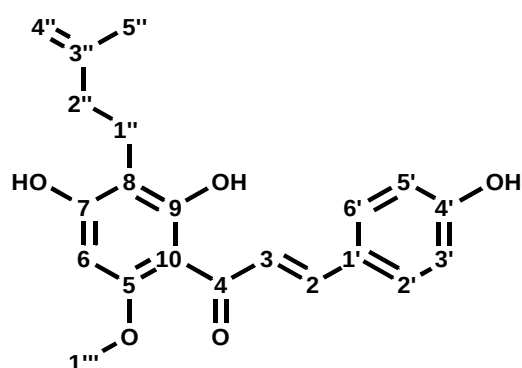
measured: m/z 371,1495; calculated for $[\text{C}_{21}\text{H}_{24}\text{O}_6\text{-H}^+]^-$: m/z 371,1495

LC-MS/MS (ESI-):

m/z (%) 371 (10), 251 (22), 163 (5), 119 (90)

^1H NMR (500 MHz, MeOD; COSY): δ [ppm]: 1,16 [s, 6H, H-C(4' / 5')]; 1,61 [m, 2H, H-C(2'')]; 2,61 [m, 2H, H-C(1'')]; 2,69 [dd, 1H, $J = 3,2$ & 16,8, H-C(3eq)]; 2,97 [dd, 1H, $J = 13,0$ & 16,8, H-C(3ax)]; 3,80 [s, 3H, H-C(1''')]; 5,31 [dd, 1H, $J = 3,2$ & 13,0, H-C(2)]; 6,12 [s, 1H, H-C(6)]; 6,81 [m, 2H, $J = 8,0$ Hz, H-C(3' / 5')]; 7,34 [m, 2H, $J = 8,0$ Hz, H-C(2' / 6')]

^{13}C NMR (125 MHz, MeOD; HSQC, HMBC): δ (ppm): 19,0 [C(1'')]; 28,8 [C(4' / 5')]; 43,8 [C(2'')]; 46,3 [C(3)]; 56,0 [C(1''')]; 71,7 [C(3')]; 79,9 [C(2)]; 93,5 [C(6)]; 105,9 [C(10)]; 110,7 [C(8)]; 116,3 [C(3' / 5')]; 128,7 [C(2' / 6')]; 131,6 [C(1')]; 158,8 [C(4')]; 161,8 [C(5)]; 163,9 [C(9)]; 164,4 [C(7)]; 193,0 [C(4)]



UV/Vis (ACN / Water; 0,1 % Formic acid):

$\lambda_{\max} = 368 \text{ nm}$

LC-TOF-MS (ESI-):

measured: m/z 353,1394; calculated for $[\text{C}_{21}\text{H}_{22}\text{O}_6\text{-H}^+]^-$: m/z 353,1394

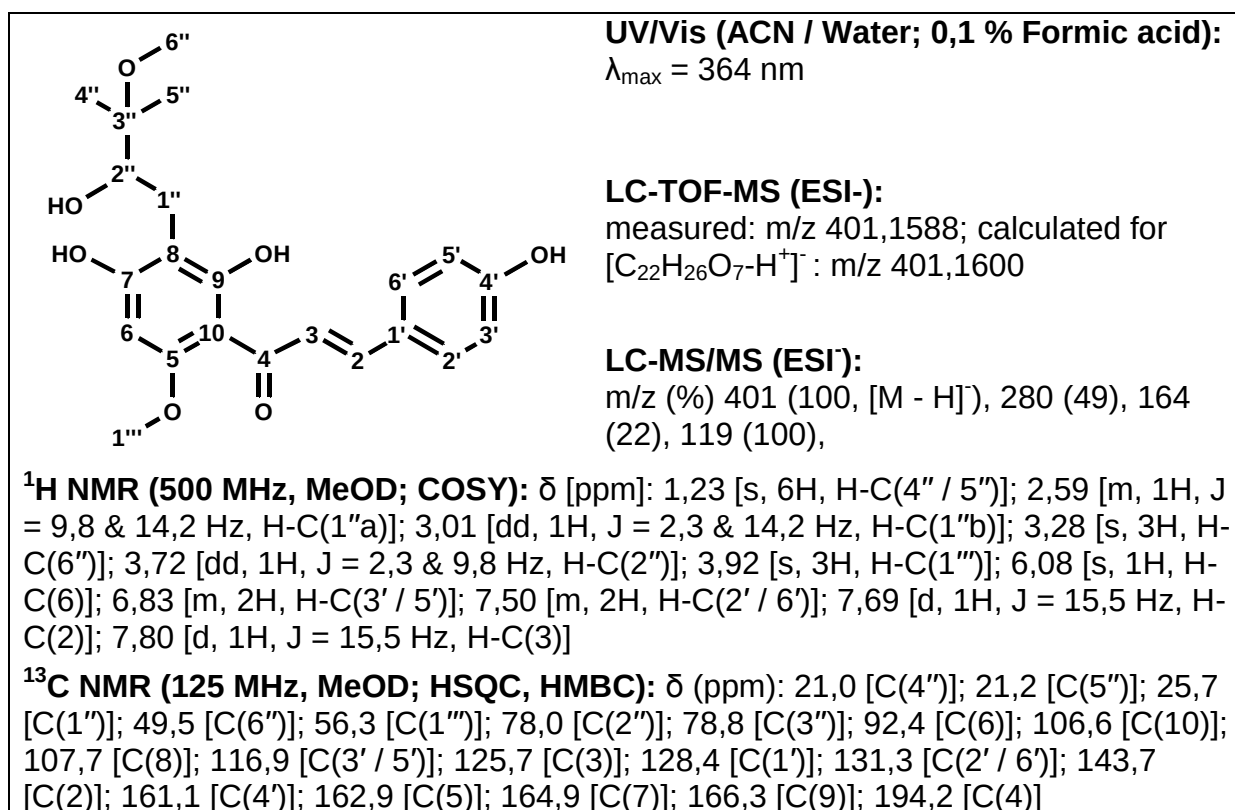
LC-MS/MS (ESI-):

m/z (%) 353 (100, $[\text{M} - \text{H}]^-$), 219 (23), 163 (40), 119 (85)

^1H NMR (500 MHz, MeOD; COSY): δ [ppm]: 1,79 [s, 3H, H-C(5'')]; 2,18 [m, 2H, $J = 8,1$, H-C(2'')]; 2,69 [m, 2H, $J = 8,1$, H-C(1'')]; 3,28 [s, 3H, H-C(6'')]; 3,90 [s, 3H, H-C(1''')]; 4,66 [s, 2H, H-C(4'')]; 6,03 [s, 1H, H-C(6)]; 6,83 [m, 2H, $J = 8,7$ Hz, H-C(3' / 5')]; 7,50 [m, 2H, $J = 8,7$ Hz, H-C(2' / 6')]; 7,67 [d, 1H, $J = 15,5$ Hz, H-C(3)]; 7,80 [d, 1H, $J = 15,5$ Hz, H-C(2)]

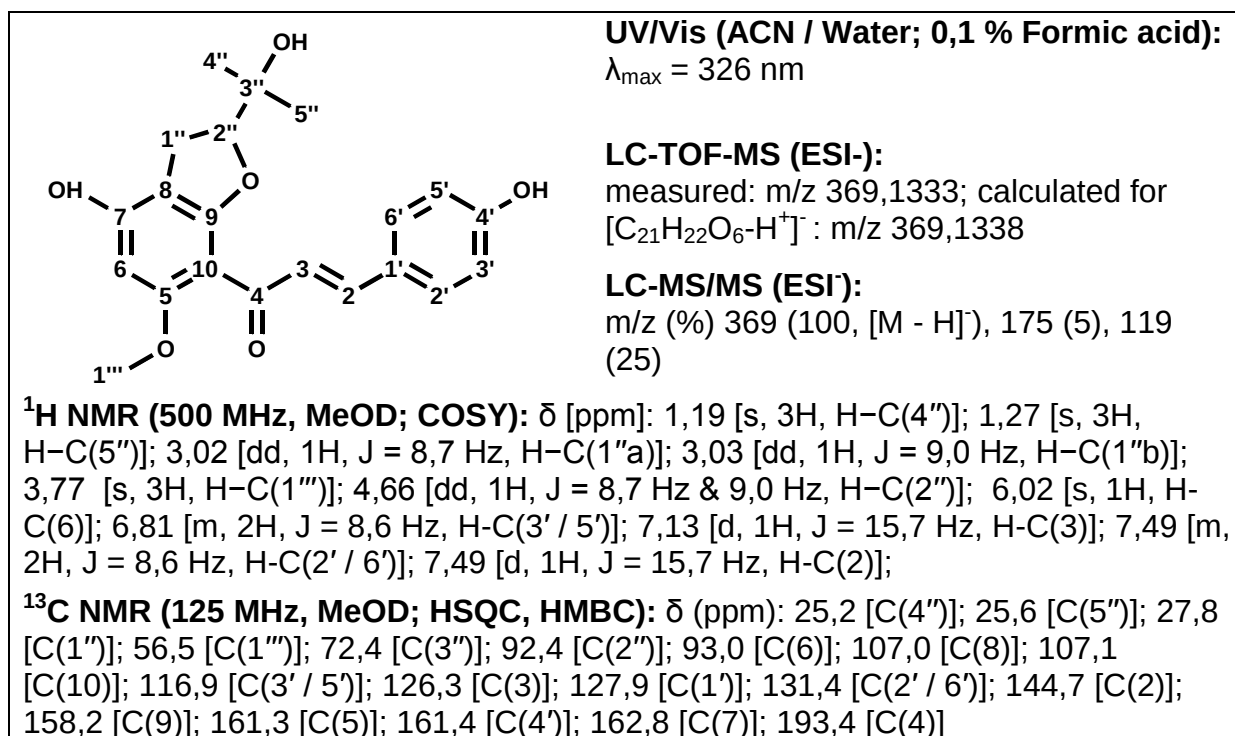
^{13}C NMR (125 MHz, MeOD; HSQC, HMBC): δ (ppm): 22,1 [C(1'')]; 22,6 [C(5'')]; 37,9 [C(2'')]; 56,2 [C(1''')]; 91,6 [C(6)]; 106,5 [C(10)]; 109,7 [C(8)]; 110,1 [C(4'')]; 116,9 [C(3' / 5')]; 125,9 [C(3)]; 128,5 [C(1')]; 131,2 [C(2' / 6')]; 143,3 [C(2)]; 161,0 [C(4')]; 162,5 [C(5)]; 163,9 [C(7)]; 166,2 [C(9)]; 194,1 [C(4)]

142 2''-Hydroxyxanthohumol M (25)



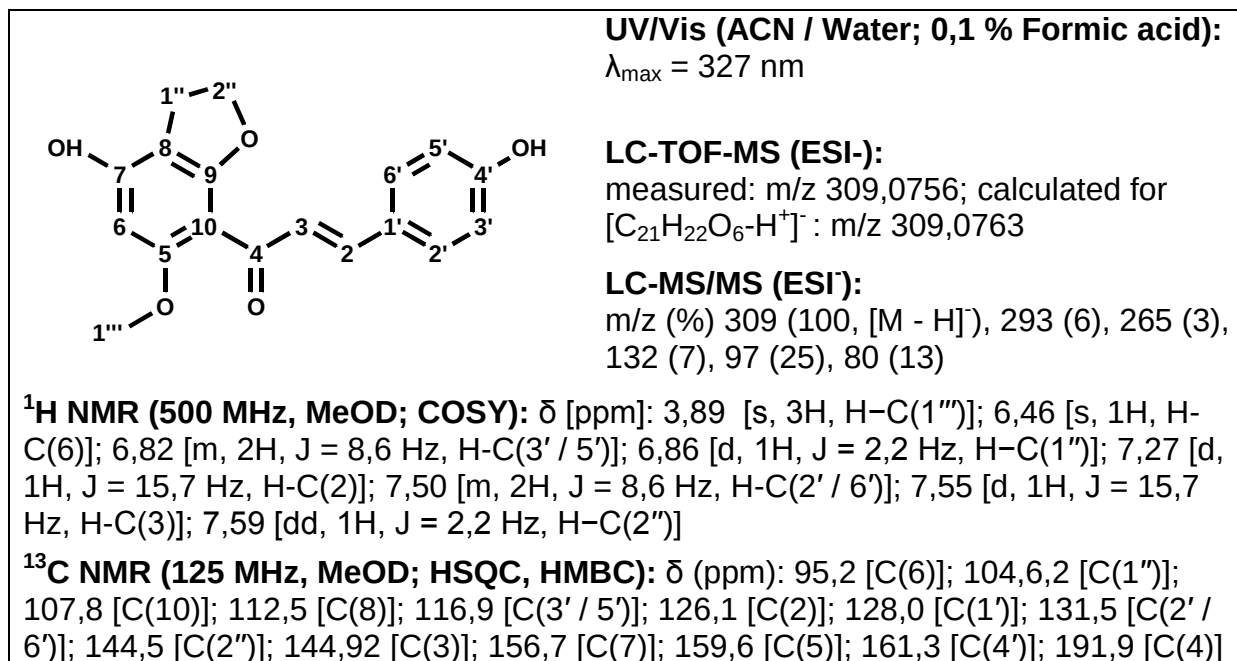
143

144 Xanthohumol I (26)



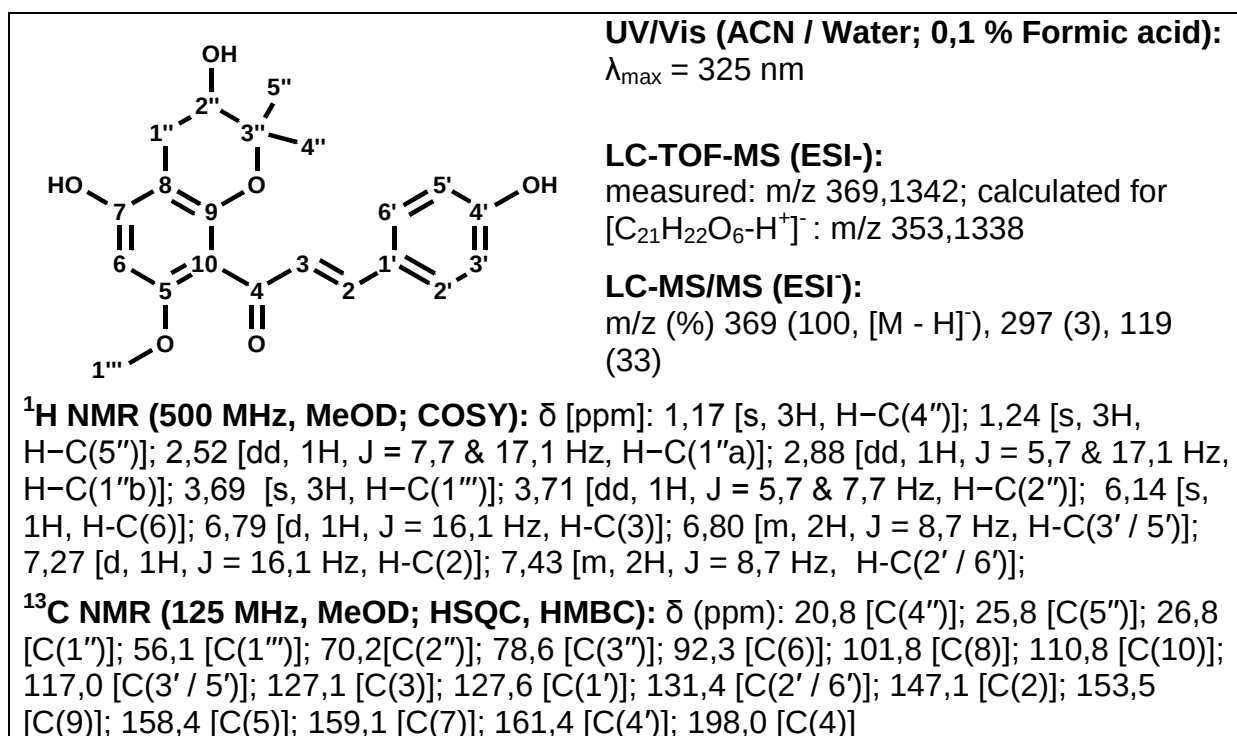
145

146 Xanthohumol O (27)



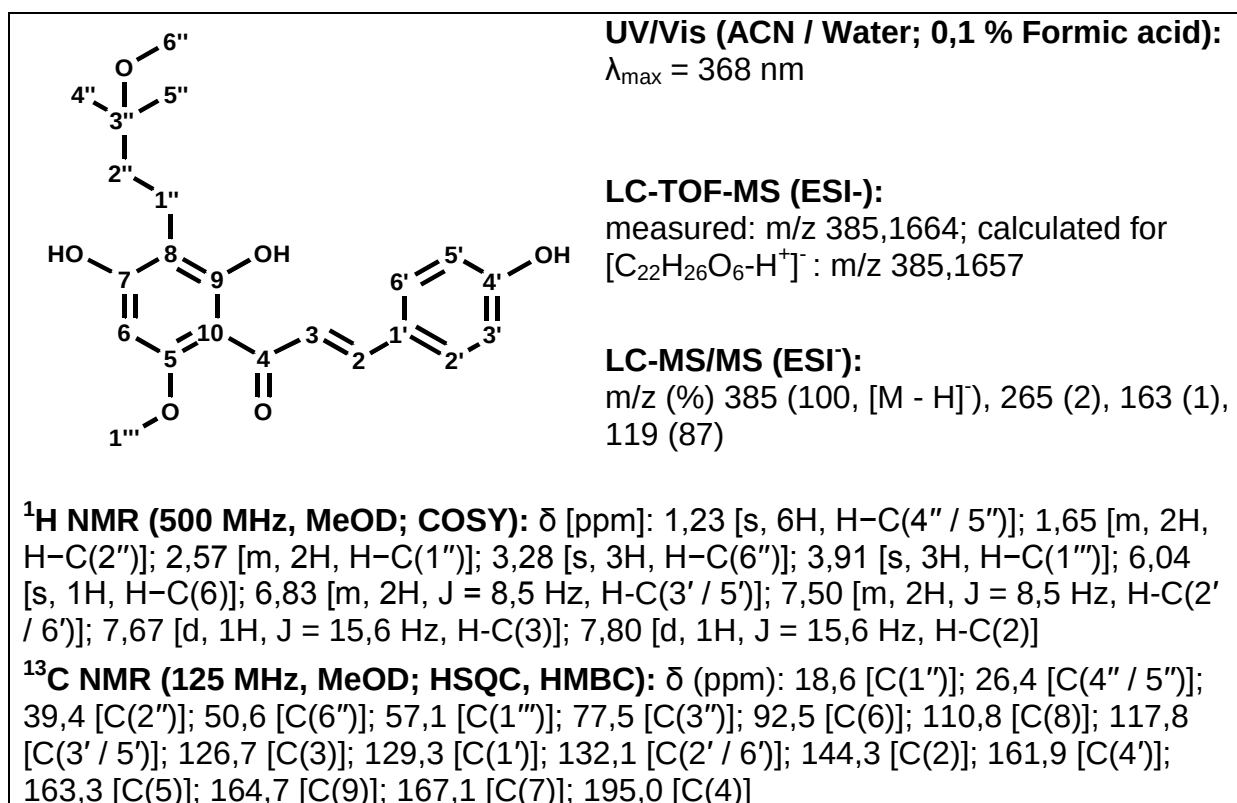
147

148 Xanthohumol L (28)

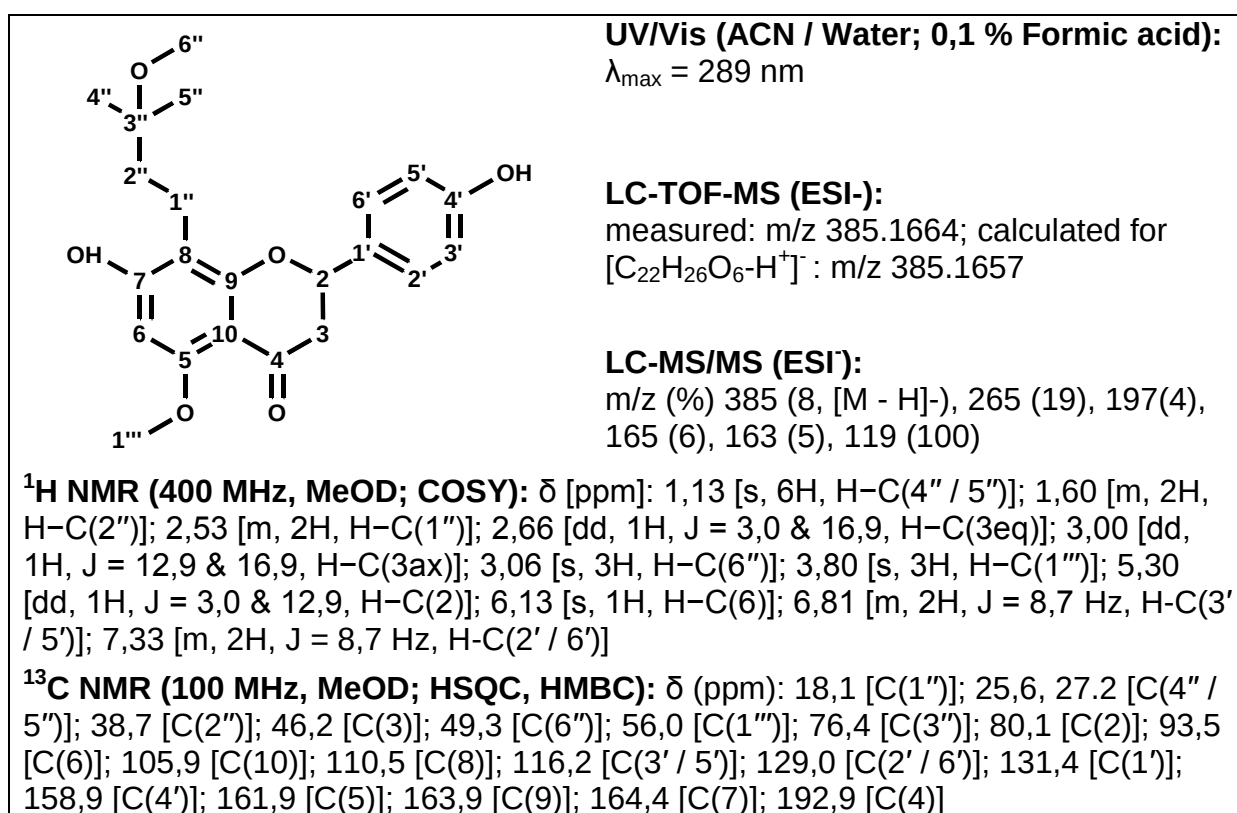


149

150 Xanthohumol M (29)

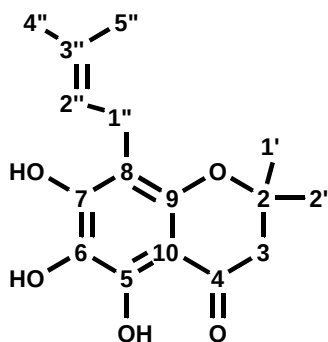


151 Isoxanthohumol M (30)
 152



153

154 2'',3''-Dehydrocyclohumulohydrochinon (**31**)



UV/Vis (ACN / Water; 0,1 % Formic acid):

$\lambda_{\text{max}} = 292 \text{ nm}$

LC-TOF-MS (ESI-):

measured: m/z 291,1222; calculated for $[\text{C}_{16}\text{H}_{20}\text{O}_5\text{-H}^+]^-$: m/z 291,1238

LC-MS/MS (ESI-):

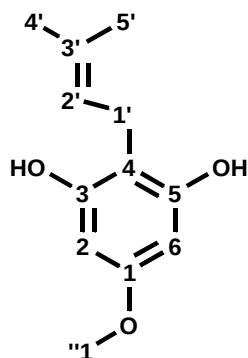
m/z (%) 291 (80, $[\text{M} - \text{H}]^-$), 207 (25), 235 (23), 136 (21), 135 (19)

^1H NMR (500 MHz, MeOD; COSY): δ [ppm]: 1,41 [s, 6H, H-C(1' / 2')]; 1,64 [s, 3H, H-C(4'')]; 1,76 [s, 3H, H-C(5'')]; 2,67 [s, 2H, H-C(3)]; 3,20 [d, 2H, $J = 7,3 \text{ Hz}$, H-C(1'')]; 5,15 [t, 1H, $J = 7,3 \text{ Hz}$, H-C(2'')]

^{13}C NMR (125 MHz, MeOD; HSQC, HMBC): δ (ppm): 16,6 [C(5'')]; 21,2 [C(1'')]; 24,5 [C(4'')]; 25,5 [C(1' / 2')]; 49,5 [C(3)]; 78,0 [C(2)]; 101,0 [C(10)]; 107,8 [C(8)]; 122,8 [C(2'')]; 129,9 [C(3'')]; 147,6 [C(5)]; 151,7 [C(9)]; 154,1 [C(7)]; 197,3 [C(4)]

155

156 1-Methoxy-4-prenylphloroglucinol (**32**)



UV/Vis (ACN / Water; 0,1 % Formic acid):

$\lambda_{\text{max}} = 292 \text{ nm}$

LC-TOF-MS (ESI-):

measured: m/z 207.1035; calculated for $[\text{C}_{12}\text{H}_{15}\text{O}_3\text{-H}^+]^-$: m/z 207.1027

LC-MS/MS (ESI-):

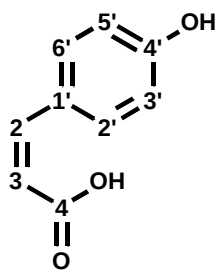
m/z (%) 207 (100, $[\text{M} - \text{H}]^-$), 137 (25), 121 (14), 109 (10), 149 (9)

^1H NMR (500 MHz, MeOD; COSY): δ [ppm]: 1.65 [s, 3H, H-C(4')]; 1.75 [s, 3H, H-C(5')]; 3.2 [d, 2H, $J = 7.3 \text{ Hz}$, H-C(1')]; 3.83 [s, 3H, H-C(1'')]; 5.21 [m, 1H, $J = 7.3 \text{ Hz}$, H-C(2')]; 5.42 [s, 1H, H-C(2)]; 6.04 [s, 1H, H-C(6)]

^{13}C NMR (125 MHz, MeOD; HSQC, HMBC): δ (ppm): 17.8 [C(5')]; 22.0 [C(1')]; 25.9 [C(4')]; 56.2 [C(1'')]; 93.4 [C(6)]; 99.8 [C(2)]; 105.9 [C(4)]; 123.2 [C(2'')]; 132.4 [C(3')]; 159.6 [C(1)]; 167.9 [C(5)]; 173.3 [C(3)]

157

158 *cis-p*-coumaric acid (**33a**)



UV/Vis (ACN / Water; 0,1 % Formic acid):

$\lambda_{\text{max}} = 301 \text{ nm}$

LC-TOF-MS (ESI-):

measured: m/z 163,0400; calculated for $[\text{C}_9\text{H}_8\text{O}_3\text{-H}^+]^-$: m/z 163,0395

LC-MS/MS (ESI-):

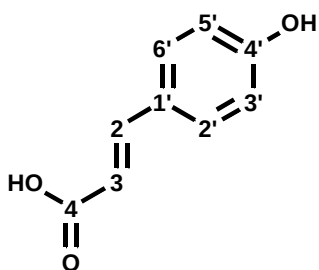
m/z (%) 163 (100, $[\text{M} - \text{H}]^-$), 119 (50), 93 (6)

^1H NMR (400 MHz, MeOD; COSY): δ [ppm]: 5,77 [d, 1H, $J = 12,8 \text{ Hz}$, H-C(3)]; 6,73 [m, 2H, $J = 8,5 \text{ Hz}$, H-C(3' / 5')]; 6,77 [d, 1H, $J = 12,8 \text{ Hz}$, H-C(2)]; 7,59 [m, 2H, $J = 8,5 \text{ Hz}$, H-C(2' / 6')];

^{13}C NMR (100 MHz, MeOD; HSQC, HMBC): δ (ppm): 115,8 [C(3' / 5')]; 118,2 [C(3)]; 128,0 [C(1')]; 133,2 [C(2' / 6')]; 142,7 [C(2)]; 159,6 [C(4')]; 171,1 [C(4)]

159

160 *trans-p*-coumaric acid (**33b**)



UV/Vis (ACN / Water; 0,1 % Formic acid):

$\lambda_{\text{max}} = 301 \text{ nm}$

LC-TOF-MS (ESI-):

measured: m/z 163,0400; calculated for $[\text{C}_9\text{H}_8\text{O}_3\text{-H}^+]^-$: m/z 163,0395

LC-MS/MS (ESI-):

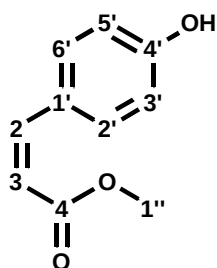
m/z (%) 163 (100, $[\text{M} - \text{H}]^-$), 119 (50), 93 (6)

^1H NMR (400 MHz, MeOD; COSY): δ [ppm]: 6,29 [d, 1H, $J = 15,9 \text{ Hz}$, H-C(3)]; 6,82 [m, 2H, $J = 8,5 \text{ Hz}$, H-C(3' / 5')]; 7,45 [m, 2H, $J = 8,5 \text{ Hz}$, H-C(2' / 6')]; 7,60 [d, 1H, $J = 15,9 \text{ Hz}$, H-C(2)]

^{13}C NMR (100 MHz, MeOD; HSQC, HMBC): δ (ppm): 115,7 [C(3)]; 116,9 [C(3' / 5')]; 127,2 [C(1')]; 131,0 [C(2' / 6')]; 146,7 [C(2)]; 161,1 [C(4')]; 171,1 [C(4)]

161

162 *cis-p*-coumaric acid methyl ester (**34a**)



UV/Vis (ACN / Water; 0,1 % Formic acid):

$\lambda_{\text{max}} = 294 \text{ nm}$

LC-TOF-MS (ESI-):

measured: m/z 177,0557; calculated for $[\text{C}_{10}\text{H}_{10}\text{O}_3\text{-H}^+]^-$: m/z 177,0552

LC-MS/MS (ESI-):

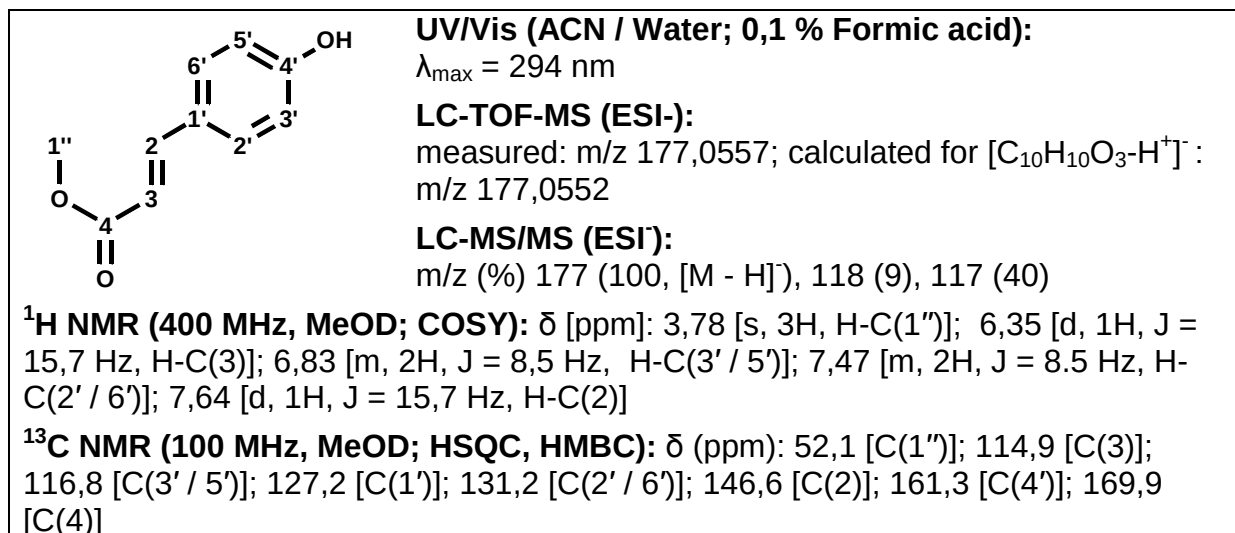
m/z (%) 177 (100, $[\text{M} - \text{H}]^-$), 118 (9), 117 (40)

^1H NMR (400 MHz, MeOD; COSY): δ [ppm]: 3,72 [s, 3H, H-C(1'')]; 5,79 [d, 1H, $J = 12,9 \text{ Hz}$, H-C(3)]; 6,77 [m, 2H, $J = 8,8 \text{ Hz}$, H-C(3' / 5')]; 6,88 [d, 1H, $J = 12,9 \text{ Hz}$, H-C(2)]; 7,64 [m, 2H, $J = 8,8 \text{ Hz}$, H-C(2' / 6')];

^{13}C NMR (100 MHz, MeOD; HSQC, HMBC): δ (ppm): 51,6 [C(1'')]; 115,9 [C(3' / 5')]; 116,4 [C(3)]; 127,5 [C(1')]; 133,6 [C(2' / 6')]; 145,0 [C(2)]; 160,0 [C(4')]; 168,9 [C(4)]

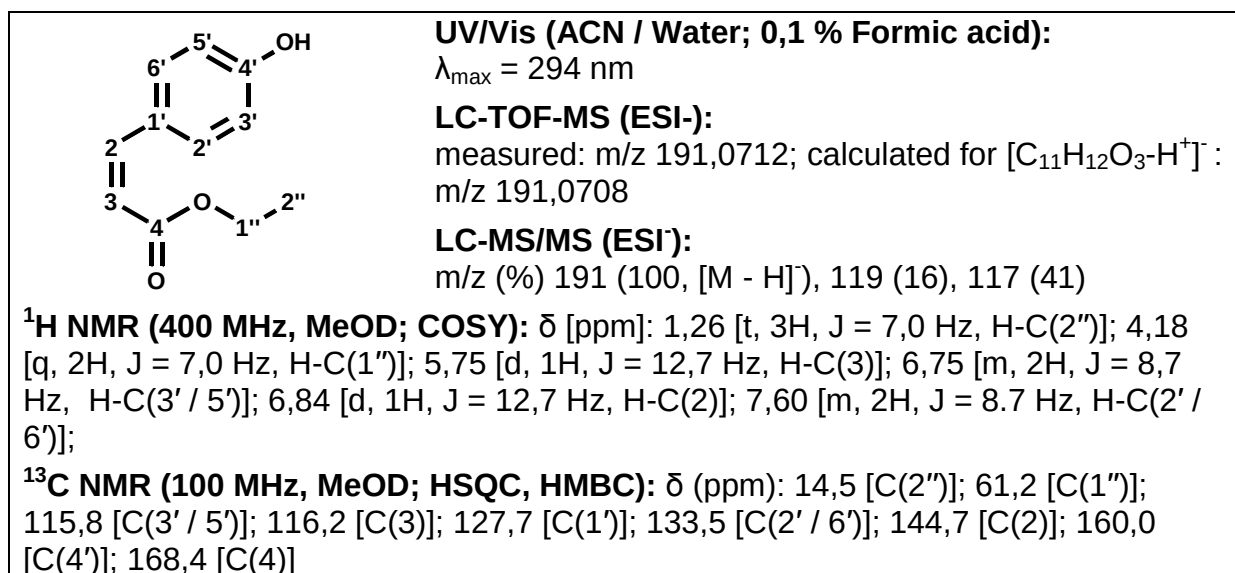
163

164 *trans-p*-coumaric acid methyl ester (**34b**)



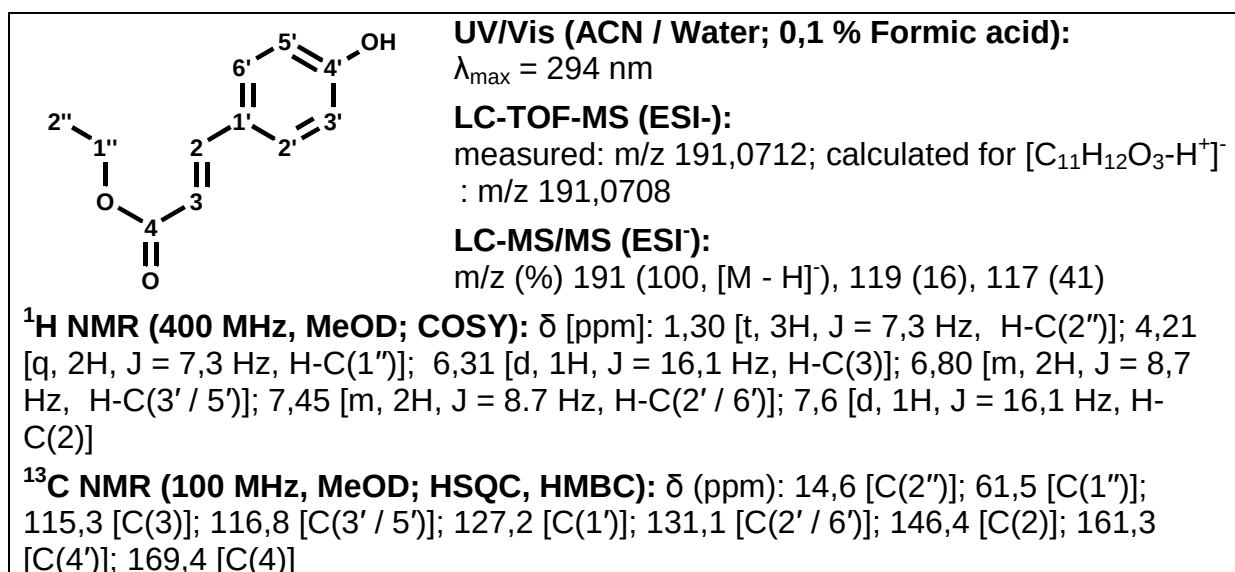
165

166 *cis-p*-coumaric acid ethyl ester (**35a**)



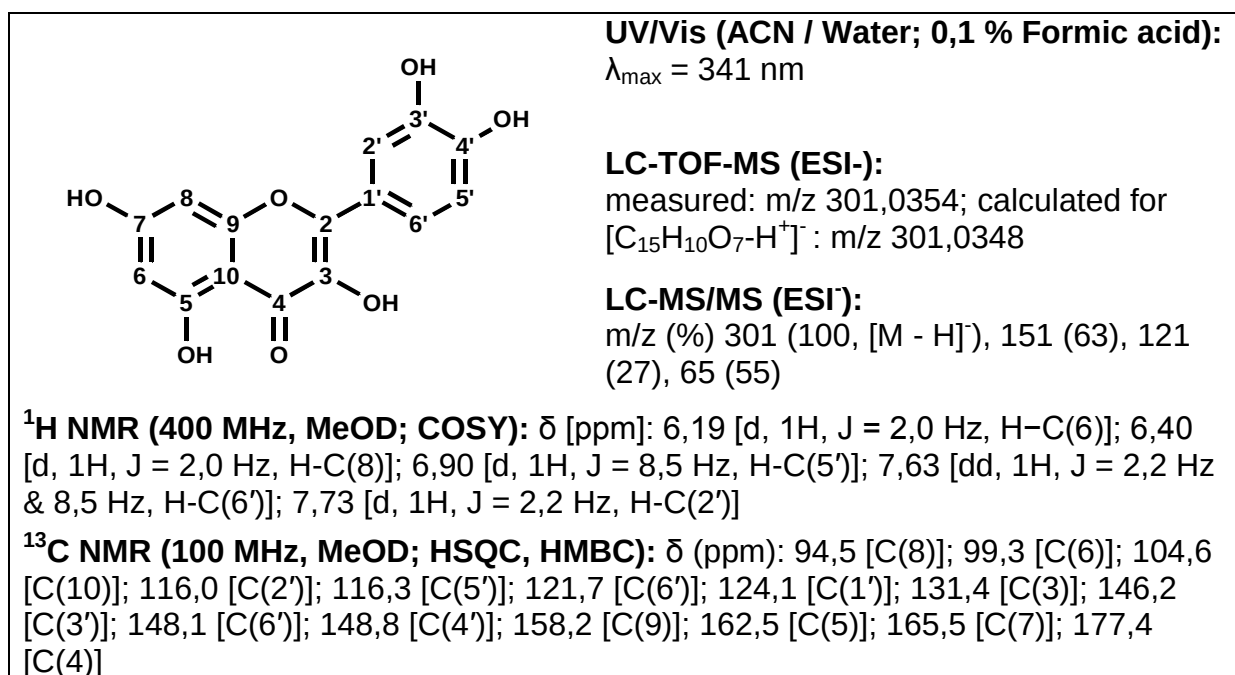
167

168 *trans-p*-coumaric acid ethyl ester (**35b**)



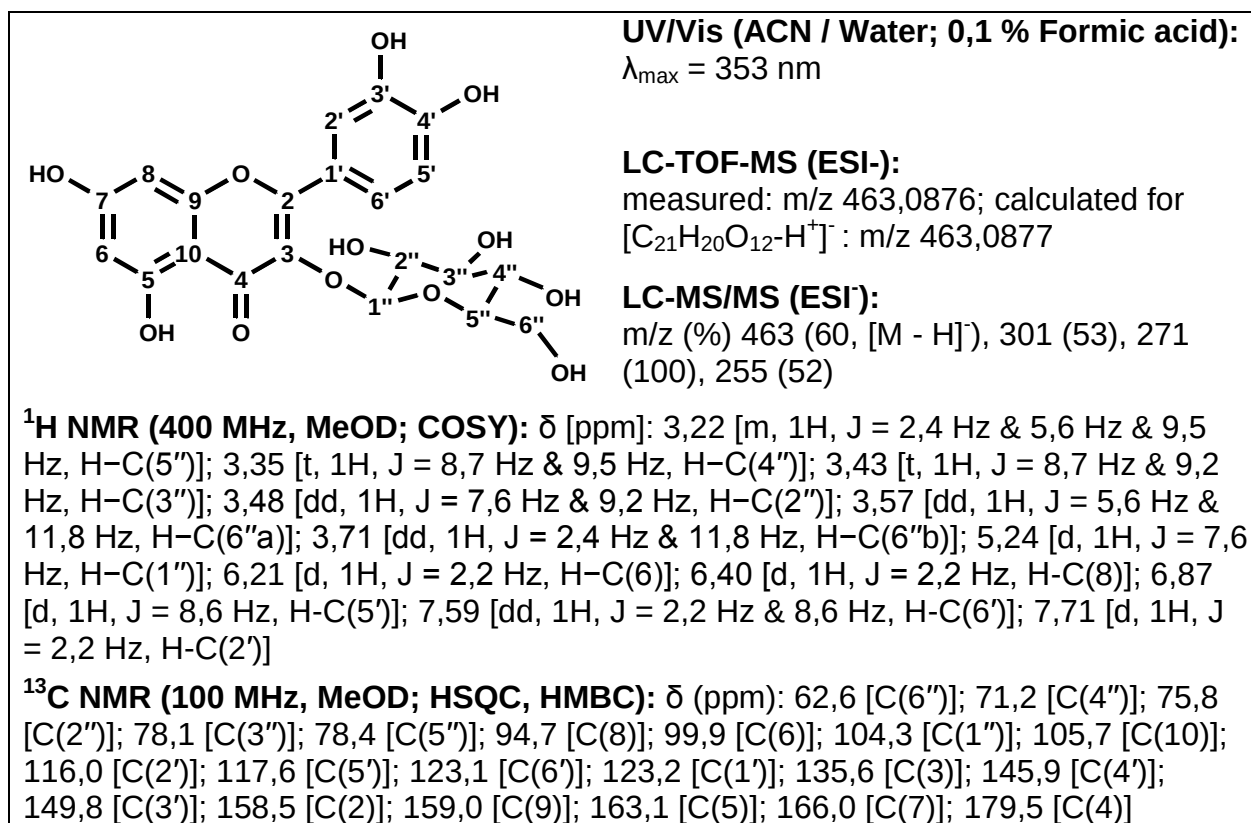
169

170 Quercetin (36)



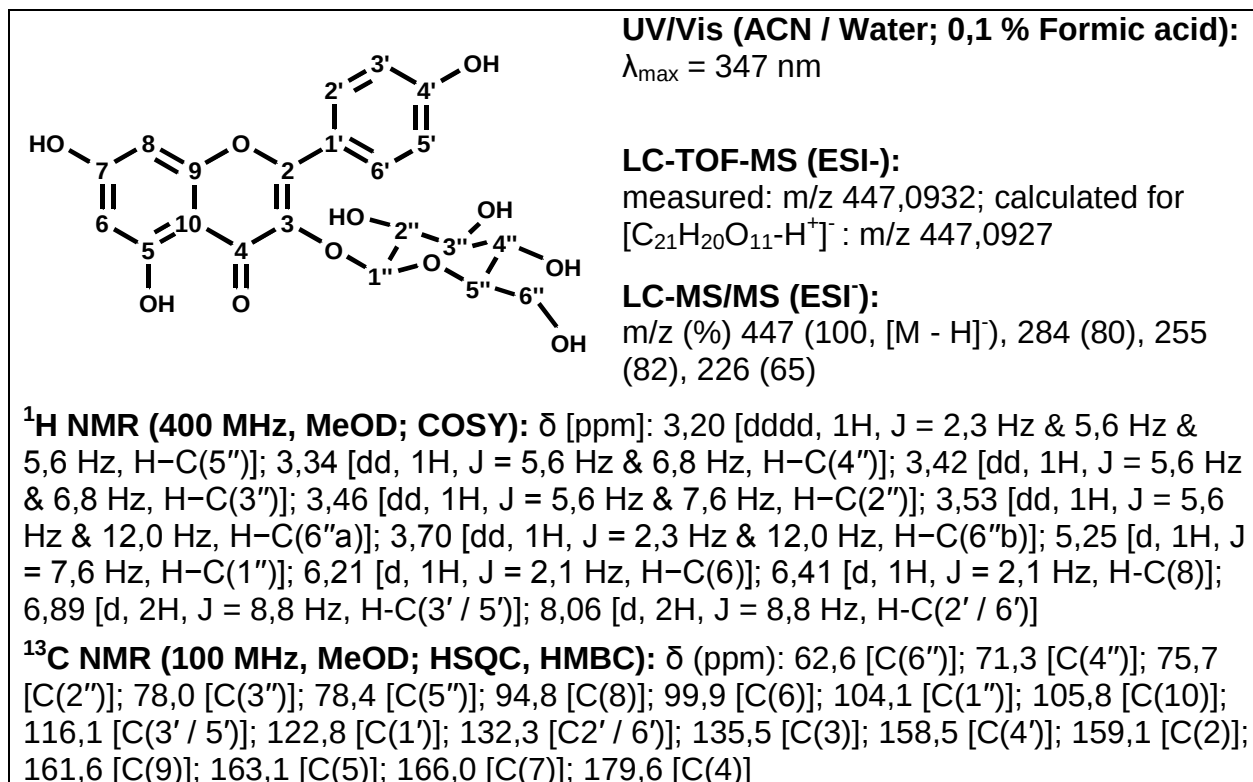
171

172 Quercetin-3-O- β -D-glucopyranoside (37)



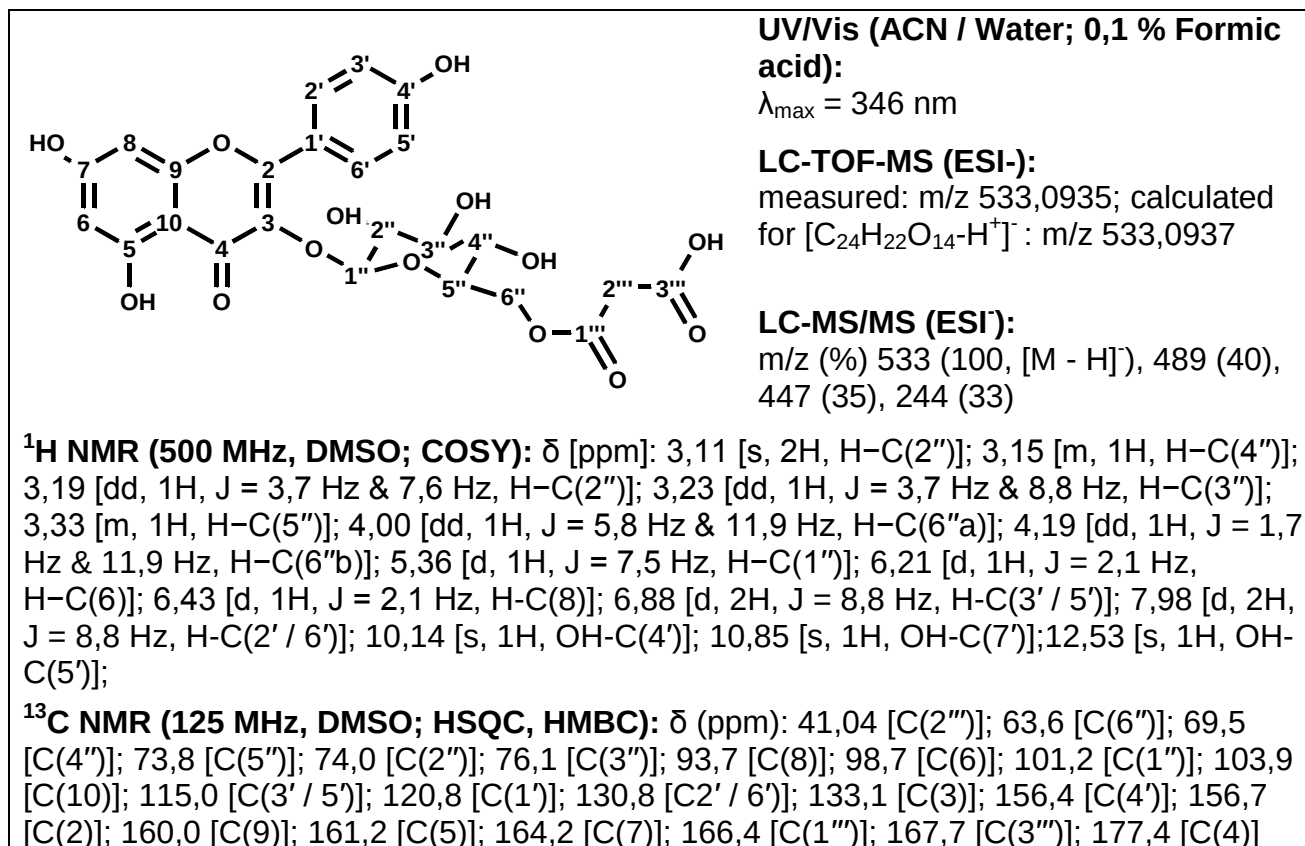
173

174 Kaempferol-3-O-β-D-glucopyranoside (**38**)



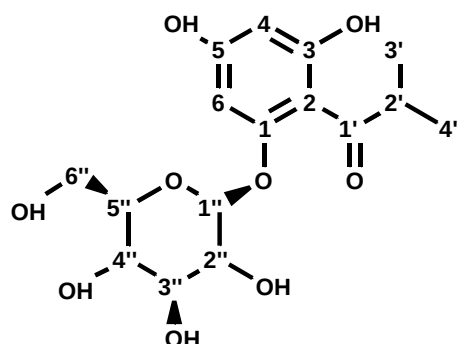
175

176 Kaempferol-3-O-β-D-(6'-malonyl)-glucopyranoside (**39**)



177

178 co-Multifidol-glucopyranoside (1-O-β-D-(2-Methyl-propanoyl)-phloroglucinol-
 179 glucopyranoside) (**40a**)



UV/Vis (ACN / Water; 0,1 % Formic acid):
 $\lambda_{\max} = 283 \text{ nm}$

LC-TOF-MS (ESI-):
 measured: m/z 357,1192; calculated for $[\text{C}_{16}\text{H}_{22}\text{O}_9\text{-H}^+]^-$: m/z 357,1186

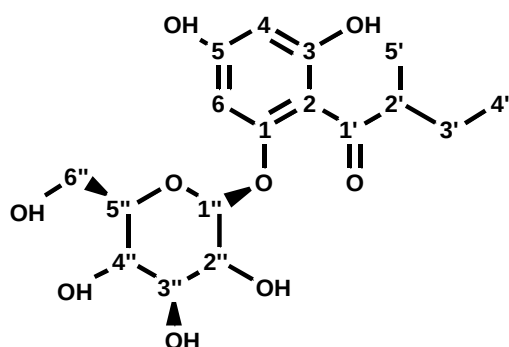
LC-MS/MS (ESI-):
 m/z (%) 357 (100, $[\text{M} - \text{H}]^-$), 195 (50), 151 (16), 111 (8)

^1H NMR (400 MHz, MeOD; COSY): δ [ppm]: 1,13 [d, 3H, $J = 6,7 \text{ Hz}$, H-C(3')]; 1,15 [d, 3H, $J = 6,7 \text{ Hz}$, H-C(4')]; 3,40 [t, 1H, $J = 9,1 \text{ Hz}$, H-C(4'')]; 3,46 [m, 1H, $J = 9,1 \text{ Hz}$, H-C(3'')]; 3,46 [m, 1H, H-C(5'')]; 3,50 [dd, 1H, $J = 7,5 \text{ Hz}$, H-C(2'')]; 3,72 [dd, 1H, $J = 5,5 \text{ Hz}$ & $12,1 \text{ Hz}$, H-C(6''a)]; 3,92 [dd, 1H, $J = 2,1 \text{ Hz}$ & $12,1 \text{ Hz}$, H-C(6''b)]; 3,99 [sept, 1H, $J = 6,7 \text{ Hz}$, H-C(2')]; 5,05 [d, 1H, $J = 7,5 \text{ Hz}$, H-C(1'')]; 5,97 [d, 1H, $J = 2,3 \text{ Hz}$, H-C(4)]; 6,18 [d, 1H, $J = 2,3 \text{ Hz}$, H-C(6)]

^{13}C NMR (100 MHz, MeOD; HSQC, HMBC): δ (ppm): 19,4 [C(3')]; 20,2 [C(4')]; 40,4 [C(2')]; 62,5 [C(6'')]; 71,2 [C(4'')]; 74,8 [C(2'')]; 78,3 [C(5'')]; 78,7 [C(3'')]; 95,5 [C(6)]; 98,3 [C(4)]; 101,5 [C(1'')]; 106,3 [C(2)]; 161,6 [C(1)]; 165,5 [C(5)]; 167,4 [C(3)]; 212,0 [C(1')]

180

181 ad-Multifidol-glucopyranoside (1-O-β-D-(2-methylbutyryl)-phloroglucinol-glucopyranoside)
 182 (**40c**)



UV/Vis (ACN / Water; 0,1 % Formic acid):
 $\lambda_{\max} = 281 \text{ nm}$

LC-TOF-MS (ESI-):
 measured: m/z 371,1346; calculated for $[\text{C}_{17}\text{H}_{24}\text{O}_9\text{-H}^+]^-$: m/z 377,1342

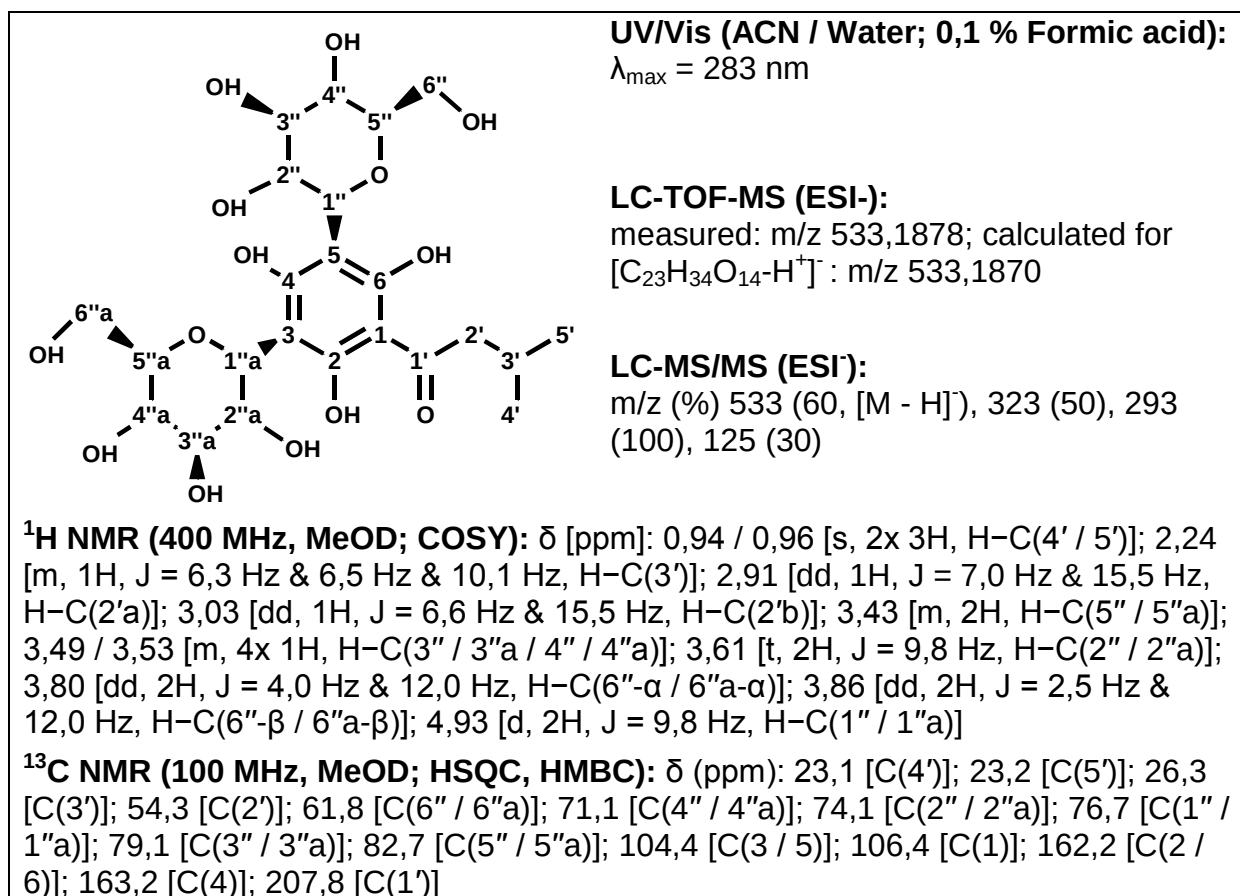
LC-MS/MS (ESI-):
 m/z (%) 371 (100, $[\text{M} - \text{H}]^-$), 209 (34), 165 (7), 125 (6)

^1H NMR (400 MHz, MeOD; COSY): δ [ppm]: 0,87 [dd, 3H, $J = 7,5 \text{ Hz}$ & $9,3 \text{ Hz}$, H-C(4')]; 1,13 [d, 3H, $J = 7,0 \text{ Hz}$, H-C(5')]; 1,38 [m, 1H, H-C(3'a)]; 1,13 [d, 1H, H-C(3'b)]; 3,38 [t, 1H, $J = 9,0 \text{ Hz}$, H-C(5'')]; 3,45 [m, 1H, H-C(3'')]; 3,45 [m, 1H, H-C(5'')]; 3,50 [dd, 1H, $J = 7,5 \text{ Hz}$ & $9,0 \text{ Hz}$, H-C(2'')]; 3,71 [dd, 1H, $J = 5,5 \text{ Hz}$ & $12,3 \text{ Hz}$, H-C(6''a)]; 3,92 [dd, 1H, $J = 2,1 \text{ Hz}$ & $12,3 \text{ Hz}$, H-C(6''b)]; 3,92 [m, 1H, H-C(2')]; 5,04 [d, 1H, $J = 7,5 \text{ Hz}$, H-C(1'')]; 5,95 [d, 1H, $J = 2,3 \text{ Hz}$, H-C(4)]; 6,18 [d, 1H, $J = 2,3 \text{ Hz}$, H-C(6)]

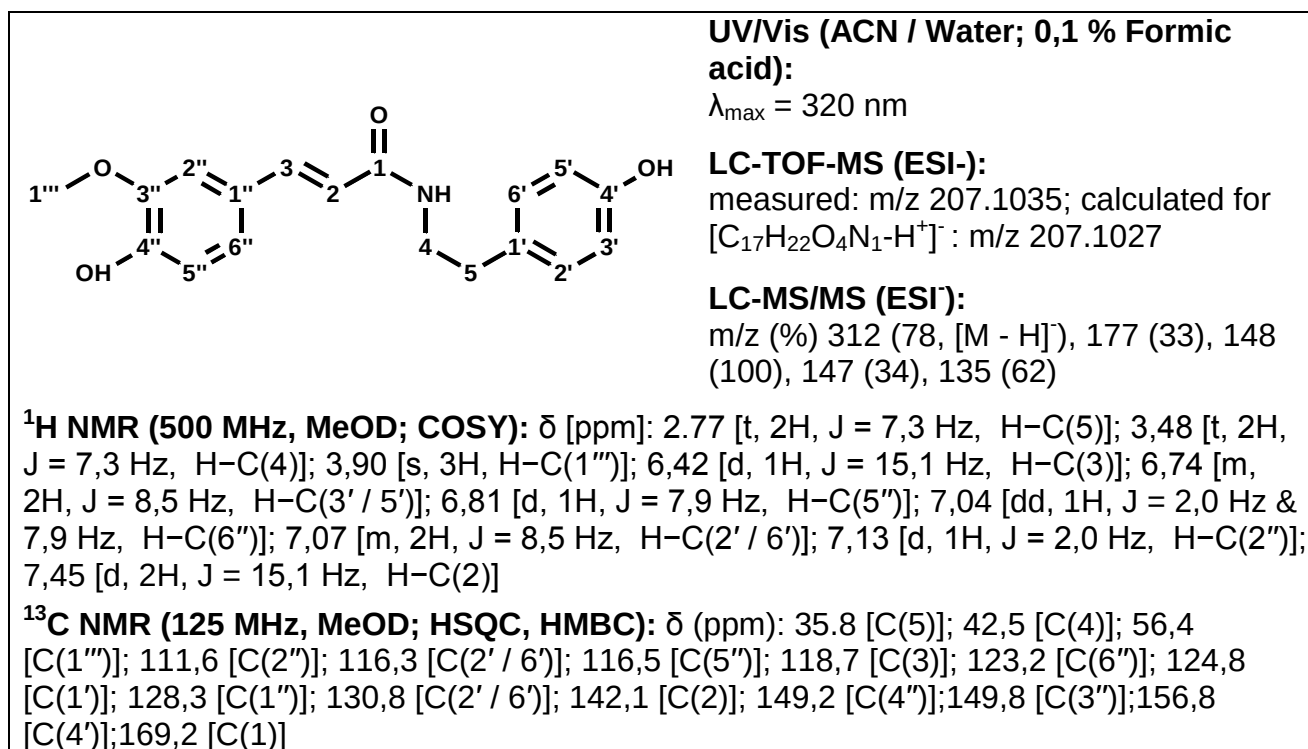
^{13}C NMR (100 MHz, MeOD; HSQC, HMBC): δ (ppm): 12,1 [C(4')]; 16,9 [C(5')]; 28,3 [C(3')]; 47,0 [C(2')]; 62,5 [C(6'')]; 71,2 [C(4'')]; 74,8 [C(2'')]; 78,3 [C(5'')]; 78,7 [C(3'')]; 95,4 [C(6)]; 98,3 [C(4)]; 101,6 [C(1'')]; 106,7 [C(2)]; 161,7 [C(1)]; 165,5 [C(5)]; 167,4 [C(3)]; 211,9 [C(1')]

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184 n-Multifidol-Di-C-glucopyranoside (Phloroisovalerophenon-3,5-Di-C-β-D-glucopyranoside)
 185 (41b)



186
 187 N-trans-Feruloyltyramine (42)



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Table S1: Optimized Mass Spectrometric Parameters for the Quantitative Analysis of Taste-active Hop-Derived Compounds by Means of LC-MS/MS Using Negative Electrospray Ionization (ESI-).

compd no. ^a	mass transition Q1 → Q3	DP ^b [V]	CE ^c [V]	CXP ^d [V]
1a	<i>m/z</i> 347.1 → 277.9	-65	-28	-17
1b, 1c	<i>m/z</i> 361.2 → 292.0	-65	-28	-17
2a	<i>m/z</i> 399.2 → 286.9	-80	-38	-15
2b, 2c	<i>m/z</i> 413.2 → 301.2	-80	-38	-15
3a, 4a	<i>m/z</i> 347.0 → 251.0	-90	-22	-11
3b, 3c, 4b, 4c	<i>m/z</i> 361.0 → 265.0	-90	-22	-11
5, 6	<i>m/z</i> 352.9 → 118.8	-95	-42	-7
4, 8, 9	<i>m/z</i> 339.1 → 218.9	-105	-30	-13
10	<i>m/z</i> 421.2 → 118.9	-90	-42	-7
11	<i>m/z</i> 475.2 → 243.1	-120	-50	-15
12, 13, 14	<i>m/z</i> 353.0 → 119.0	-105	-38	-7
15, 16	<i>m/z</i> 399.1 → 119.0	-30	-42	-9
17	<i>m/z</i> 421.2 → 118.9	-105	-44	-9
18	<i>m/z</i> 421.2 → 118.8	-120	-44	-1
19	<i>m/z</i> 369.1 → 163.9	-105	-40	-9
20	<i>m/z</i> 369.1 → 118.8	-105	-36	-7
21	<i>m/z</i> 351.0 → 118.9	-90	-32	-7
22, 23	<i>m/z</i> 371.1 → 118.9	-100	-52	-5
24	<i>m/z</i> 353.0 → 163.0	-100	-44	-9
25	<i>m/z</i> 401.2 → 118.9	-120	-46	-7
26	<i>m/z</i> 369.3 → 119.0	-115	-38	-7
27	<i>m/z</i> 309.2 → 293.1	-105	-40	-19
28	<i>m/z</i> 369.1 → 119.0	-110	-40	-5
29, 30	<i>m/z</i> 385.1 → 118.9	-105	-52	-5
31	<i>m/z</i> 291.1 → 207.1	-105	-36	-3
32	<i>m/z</i> 207.1 → 136.8	-125	-28	-7
33a, 33b	<i>m/z</i> 163.0 → 119.1	-50	-20	-1
34a, 34b	<i>m/z</i> 177.0 → 118.0	-60	-24	-7
35a, 35b	<i>m/z</i> 191.0 → 117.0	-75	-44	-7
36	<i>m/z</i> 301.1 → 153.0	-105	-30	-9
37	<i>m/z</i> 463.1 → 299.9	-115	-38	-19
38	<i>m/z</i> 447.1 → 283.9	-100	-38	-17
39	<i>m/z</i> 533.2 → 489.2	-50	-18	-23
40a	<i>m/z</i> 357.2 → 194.8	-90	-22	-15
40c	<i>m/z</i> 371.1 → 208.9	-105	-24	-13
41b	<i>m/z</i> 533.2 → 323.2	-85	-34	-9
42	<i>m/z</i> 312.1 → 147.8	-95	-36	-7

^aCompound numbering refers to the chemical structures given in Figure 1. ^bDeclustering potential. ^cCollision energy. ^dCell exit potential.