Supporting Information

Plasmid DNA Mono-Ion Complex Stabilized by Hydrogen Bond for *In Vivo* Diffusive Gene Delivery

Shoichiro Asayama, * Atsushi Nohara, Yoichi Negishi, and Hiroyoshi Kawakami



Figure S1. The pDNA used in agarose gel retardation assay. The pDNA was electrophoresed with size standard maker. The solid arrowhead indicates the well where the marker and pDNA were loaded.



Figure S2. ¹H NMR spectrum of APe-Im-PEG in D₂O.



Figure S3. Analysis of the MIC formation between pDNA and APe-Im-PEG by agarose gel electrophoresis in the presence of urea. The mixing ratios relative to the ω -amide-pentylimidazolium group of APe-Im-PEG per phosphate group of pDNA ([ω -amide-pentylimidazolium]_{APe-Im-PEG}/[phosphate]_{pDNA}) are indicated. The solid arrowhead indicates the well where each sample was loaded.



Figure S4. Serum stability of the APe-Im-PEG in FBS. The turbidity was measured by monitoring the absorbance at 500 nm of the mixture of the APe-Im-PEG (\bullet) and FBS (50%) during the incubation. As a control, *in vivo* transfection reagent poly(ethyleneimine) (*in vivo*-jetPEITM) (\circ) was used.



Figure S5. Effect of the APe-Im-PEG on the viability of HepG2 cells. Symbols and error bars represent the mean and standard deviation of the measurements made in paired wells (n = 5).



Figure S6. Transfection of luciferase gene into the skeletal muscles by APe-Im-PEG/pDNA MIC as well as Bu-Im-PEG/pDNA MIC at a positive/negative charge ratio (+/-) of 1. Gene expression was determined relative light unit (RLU) normalized by the protein concentration. Symbols and error bars represent the mean and standard deviation (n = 2).

+/-	Particle diameter / nm	Zeta potential / mV
0.5	(40.7±7.0)	N.D. ^{a)}
0.8	(42.6 ± 8.3)	(0.58)
1.0	(31.1 ± 4.1)	(-3.36)
1.5	(32.9 ± 5.6)	(-0.46)

Table S1. Particle size and zeta potentialof APe-Im-PEG/pDNA MICs.

^{a)} N.D.: not determined.