Bacterial Sliding Clamp Inhibitors that Mimic the Sequential Binding Mechanism of Endogenous Linear Motifs

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Supplementary Results

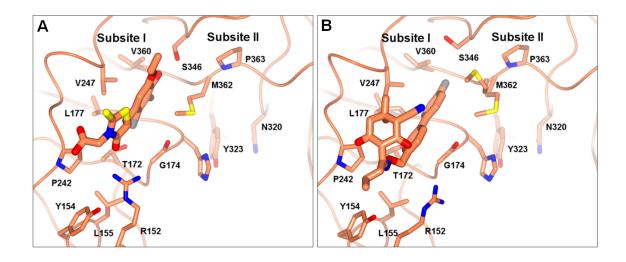


Figure S1. X-ray co-crystal structures of the *E. coli* SC in complex with (A) a thioxothiazolinine derivative (PDB entry 3D1G)¹ and (B) a biphenyloxime ether derivative (PDB entry 3QSB).² Carbon atoms are colored orange and non-carbon atoms follow CPK convention. Both inhibitors occupy subsite I.

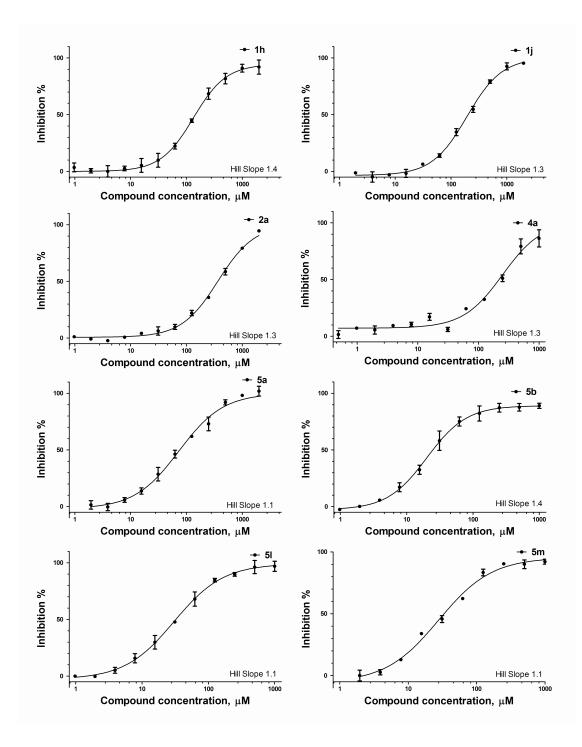


Figure S2. Dose-response curves of representative THC analogs inhibiting the *E. coli* SC, as measured by fluorescence polarization.

Table S1. Data collection and refinement statistics for X-ray co-crystal structures of the *E. coli* SC in complex with compounds 1e, 2a, 5b and 5l

Name	SC ^{1e}	SC ^{2a}	SC ^{5D}	SC⁵		
PDB Code	40VF	40VG	40VH	4PNU		
		Data collection				
Space group	P21					
	a, b, c (Å) / α, β, γ (°)	a, b, c (Å) / α, β, γ (°)	a, b, c (Å) / α, β, γ (°)	a, b, c (Å) / α, β, γ (°)		
Cell dimensions	79.87, 67.26, 81.19 /	80.08, 66.20, 80.62 /	80.03, 66.42, 80.79 /	79.89, 67.19, 81.02 /		
	90.00, 114.27, 90.00	90.00, 114.89, 90.00	90.00, 114.76, 90.00	90.00, 114.54, 90.00		
Resolution (Å)	50.00-2.05 (2.12-2.05)	40.00–1.90 (1.97–1.90)	30.00–2.25 (1.79–1.70)	30.00–1.90 (1.97–1.90)		
R _{merge} (%)	4.3 (26.1)	2.8 (17.1)	7.1 (45.6)	3.2 (38.0)		
No. of Reflections	169349	219352	116359	217379		
Unique Reflections	49386 (4716)	60330 (6013)	37099 (3029)	61732 (5986)		
Mean I/o(I)	27.4 (3.5)	41.5 (7.0)	16.6 (2.2)	36.1 (2.7)		
Completeness (%)	96.4 (95.9)	99.9 (100.0)	96.4 (82.4)	99.7 (97.4)		
Multiplicity	3.6 (3.5)	3.7 (3.5)	3.1 (2.6)	3.5 (3.1)		
		Refinement				
Resolution (Å)	34.61–2.05 (2.10–2.05)	21.62–1.90 (1.95–1.90)	29.82–2.24 (2.29–2.24)	28.27–1.90 (1.95–1.90)		
R _{work} / R _{free} (%)	21.0 (19.7) / 27.0 (29.9)	18.0 (21.9) / 22.6 (27.3)	19.2 (20.5) / 25.0 (32.1)	18.2 (25.0) / 23.0 (28.9)		
RMS deviations						
Bond lengths (Å)	0.0056	0.0108	0.0070	0.0071		
Bond angles (°)	1.0960	1.5211	1.2173	1.2654		
B-factors						
main chain	22.6	17.1	24.8	17.8		
sidechain & water	26.4	21.7	26.8	22.1		
ligand*	32.7	22.1	33.1	20.3		
Ramachandran	0.43%	0.29%	0.42%	0.29%		
Plot Outliers						

Values for data in the highest resolution shell are given in parentheses.

Diffraction data were collected using a Rigaku 007HF X-ray generator producing Cu K α X-rays (wavelength of 1.5418 Å) and Mar345dtb area detector. Diffraction data were processed using HKL2000.³

*Ligand refers to the compounds bound to SC chain A.

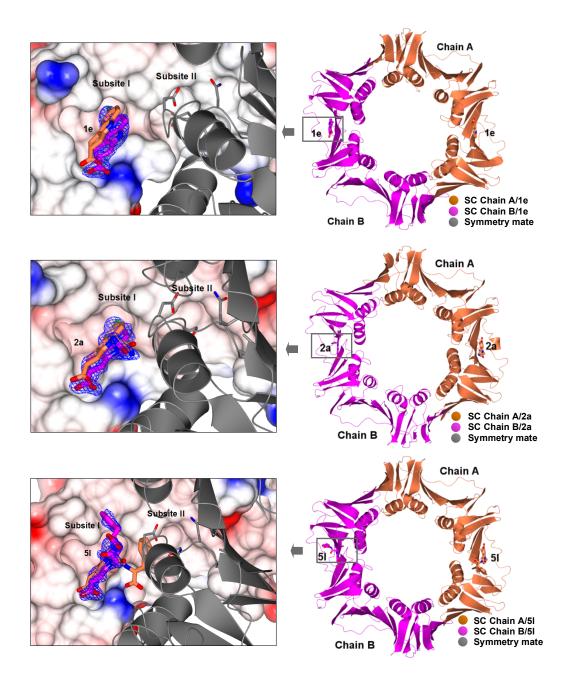


Figure S3. X-ray co-crystal structures of the *E. coli* SC showing chain B in complex with compounds **1e**, **2a** and **5l**. Carbon atoms of SC chain A/B and the bound compounds **1e**, **2a** and **5l** are colored orange/magenta respectively. All other atoms are colored according to the CPK convention. The SC symmetry partner is shown in gray. SC chain A is superimposed with chain B in the enlarged views and the electrostatic potential surfaces of chain B are shown with blue = positive and red = negative. Electron density maps $(2mF_o-DF_c)$ contoured at 1 σ are shown in blue wire-basket form.

Table S2. Data collection and refinement statistics for the X-ray crystal structures of apo-E. coli SC (SC^{apo}) and E. coli SC in complex with compound 5m (SC^{5m}), respectively

Structure Name	SC ^{apo}	SC ^{5m}	
PDB Code	4PNV	4PNW	
	Data collection		
Space group	P2	1	
	a, b, c (Å) / α, β, γ (°)	a, b, c (Å) / α, β, γ (°)	
Cell dimensions	80.14, 70.26, 84.47 /	79.81, 68.71, 83.06 /	
	90.00, 114.67, 90.00	90.00, 115.72, 90.00	
Resolution (Å)	30.00–1.86 (1.93–1.86)	30.00-2.00 (2.07-2.00)	
R _{merge} (%)	2.8 (31.9)	5.1 (45.6)	
No. of Reflections	251660	188410	
Unique Reflections	71719 (6779)	55049 (5267)	
Mean //σ(/)	41.1 (3.2)	24.0 (2.6)	
Completeness (%)	99.3 (95.3)	98.0 (96.2)	
Multiplicity	3.5 (3.1)	3.4 (3.2)	
	Refinement		
Resolution (Å)	27.55–1.86 (1.91–1.86)	28.01-2.00 (2.05-2.00)	
R _{work} / R _{free} (%)	18.3(26.0) / 23.3(33.8)	18.4(20.4) / 23.3(28.2)	
R.M.S. deviations			
Bond lengths (Å)	0.0067	0.0079	
Bond angles (°)	1.2017	1.3198	
B-factors			
mainchain	19.4	23.1	
sidechain & water	23.8	26.8	
ligand*	N/a	39.4	
Ramachandran Plot Outliers	0.29%	0.57 %	

Values for data in the highest resolution shell are given in parentheses.

Diffraction data were collected using a Rigaku 007HF X-ray generator producing Cu K α X-rays (wavelength of 1.5418 Å) and Mar345dtb area detector. Diffraction data were processed with HKL2000.³

*Ligand refers to compounds bound to SC Chain B.

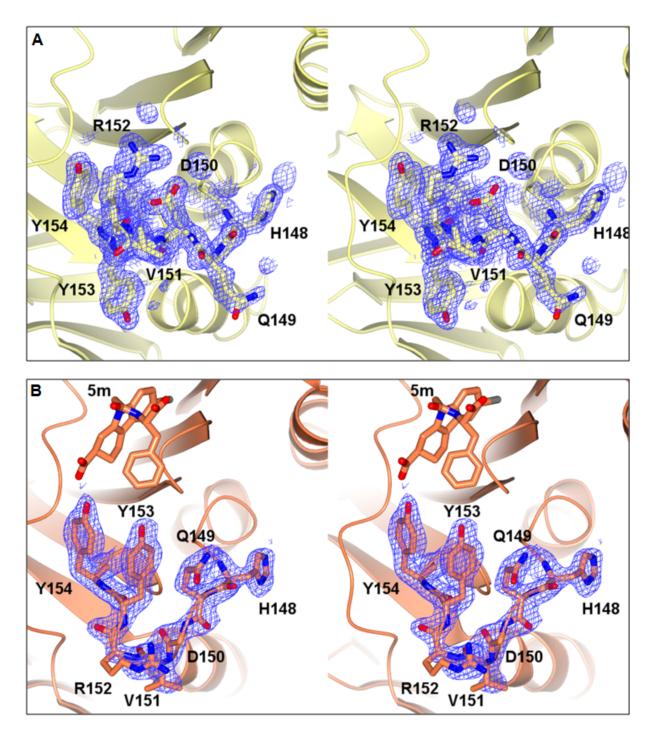


Figure S4. Stereo view of the loop rearrangement induced upon binding of compound **5m** to the *E. coli* SC. (A) The H148–Y154 loop region of SC^{apo}. Carbon atoms are shown in yellow. All other atoms are colored according to CPK convention. (B) The H148–Y154 loop region of SC^{5m}. Carbon atoms are colored orange. All other atoms are colored according to CPK convention. Electron density maps $(2mF_0-DF_c)$ contoured at 1 σ are shown in blue wire-basket form.

	10 20	30 40	50 60	70 80	90 100
E. coli	-MKFTVEREH LLKPLQQVSG PLGG	RPTLPI LGNLLLQVAD GTI	LSLTGTDL EMEMVARVAL	VQP HEPGATTVPA	RKFFDICRGL PEGAEIAVQL
A. baylyi	-MRLKIAKES LLNVLSHVVG AVER	RHTLNI LSNVKIQANA QAL	LTITGSDL EVELVASTTL	AEGAC IEAGETTVPA	RKLVDICKSL PSAALIDLQI
A. baumannii	-MRLKIAKES LLNVLSHVVG AVER	RHTLNI LSNVKIQTNA QAL	LTITGSDL EVELVASTAL	SEGAC LEAGETTVPA	RKLMEICKSL PTAALIDLQI
P. aeruginosa	-MHFTIQREA LLKPLQLVAG VVER	RQTLPV LSNVLLVVEG QQL	LSLTGTDL EVELVGRVVL	EDA AEPGEITVPA	RKLMDICKSL PNDVLIDI
S. aureus	MMEFTIKRDY FITQLNDTLK AISP	RTTLPI LTGIKIDAKE HEV	VILTGSDS EISIEITIPK	TVDGEDIVNI SETGSVVLPG	RFFVDIIKKL P-GKDVKLST
B. subtilis	-MKFTIQKDR LVESVQDVLK AVSS	RTTIPI LTGIKIVASD DGV	VSFTGSDS DISIESFIPK	EEGDKEIVTI EQPGSIVLQA	RFFSEIVKKL P-MATVEIEV
S. pneumoniae	MIHFSINKNL FLQALNITKR AISS	KNAIPI LSTVKIDVTN EGV	VTLIGSNG QISIENFISQ	KNEDAGL-LI TSLGSILLEA	SFFINVVSSL PDVTLDF
S. pyogenes	MIQFSINRTL FIHALNTTKR AIST	KNAIPI LSSIKIEVTS TGV	VTLTGSNG QISIENTIPV	SNENAGL-LI TSPGAILLEA	SFFINIISSL PDISINV
			8	2 22	2 25 1
		[][]			
	110 120	130 140		44 170 180	
E. coli	EGERMLV RSGRSRFSLS TLPA				
A. baylyi	TEDORCIL KSGNSRFVLG TLPA	_			
A. baumannii	TEDORCIL KSGNSRFVLG TLPA		-		
P. aeruginosa	R-VEEQKLLV KAGRSRFTLS TLPA				
S. aureus	NEQFQTLI TSGHSEFNLS GLDP				
B. subtilis	ONOYLTII RSGKAEFNLN GLDA		-		
S. pneumoniae	KEIEQNQIVL TSGKSEITLK GKDS		-		
S. pyogenes	KEIEOHOVVL TSGKSEITLK GKDV				
11 5					
				2 2	
				8 8	
				242	
	 210 220	230 240		·····	
E. coli		230 240	250 260	 270 4 280	290 300
E. coli A. baylyi	210 220	230 240 DNPLRV QIGSNNIRAH VGE	250 260 DFIFTSK	 ↓ 270 ↓ 280 LVDGRF <mark>P</mark> DYR R <mark>V</mark> LPKNPDKH) 290 300 LEAGCDLLKQ AFARAAILSN
	210 220 SLPSHSVIVP RKGVIELMRM LDGG AMQAVQAIVP RKAVGELQRL LSIE SSQLVQAIVP RKAVGELQRL LSIE	230 240 DNPLRV QIGSNNIRAH VGE DDQLSL LIGRELLNVT INI DEQLTL LIGRELLNVT INT	250 260 DFIFTSK IANRDKEQ HPITVRFTTK IPSRDKEQ GDITVRFTTK	LUDGKFPDYR RVIPRGGDKH) 290 300 LEAGCDLLKQ AFARAAILSN VQIAHDVFKQ SLQRVAILSN VLIGHDVFKQ SLQRVAILSN
A. baylyi	210 220 SLPSHSVIVP RKGVIELMRM LDGG AMQAVQAIVP RKAVGELQRL LSIE	230 240 DNPLRV QIGSNNIRAH VGE DDQLSL LIGRELLNVT INI DEQLTL LIGRELLNVT INT	250 260 DFIFTSK IANRDKEQ HPITVRFTTK IPSRDKEQ GDITVRFTTK	LUDGKFPDYR RVIPRGGDKH) 290 300 LEAGCDLLKQ AFARAAILSN VQIAHDVFKQ SLQRVAILSN VLIGHDVFKQ SLQRVAILSN
A. baylyi A. baumannii	210 220 SLPSHSVIVP RKGVIELMRM LDGG AMQAVQAIVP RKAVGELQRL LSIE SSQLVQAIVP RKAVGELQRL LSIE	230 240 DNPLRV QIGSNNIRAH VGE DDQLSL LIGRELLNVT INI DEQLTL LIGRELLNVT INT DGEVGI VLGQHHIRAT TGE	250 260 DFIFTSK IANRDKEQ HPITVRFTTK IPSRDKEQ GDITVRFTTK EFTFTSK) 290 300 LEAGCDLLKQ AFARAAILSN VQIAHDVFKQ SLQRVAILSN VLIGHDVFKQ SLQRVAILSN VVGDRQQLRE AFSRTAILSN
A. baylyi A. baumannii P. aeruginosa	210 220 SLPSHSVIVP RKGVIELMRM LDGG AMQAVQAIVP RKAVGELQRL LSIE SQLVQAIVP RKAVGELQRL LSIE SQDRHQVIVP RKGILELARL LTEQ VSENKNVIIP GKALAELNKI MSDNN EDRSYNVVP GKSLTELSKI LDDN	230 240 DNPLRV QIGSNNIRAH VGE DDQLSL LIGRELLNVT INI DGQLTL LIGRELLNVT INI DGEVGI VLGQHHIRAT TGE EEDLDI FFASNQVLFK VGR QELVDI VITETQVLFK AKN	250 260 DFIFTSK IANRDKEQ HPITVRFTTK PPSRDKEQ GDITVRFTTK EFIFTSK NVIFISR NVLFFSR	270 ■ 280 270 ■ 280 LVDGRFPDYR RVLPKNPDKH LIDGKFPDYR RVIPRGGDKH LVDGKFPDYE RVIPRGGDKL LLEGHYPDTT RLFPENYEIK LLDGNYPDTT SLIPQDSKTE) 290 300 LEAGCDLLKQ AFARAAILSN VQIAHDVFKQ SLQRVAILSN VVGDRQQLRE AFSRTAILSN LSIDNGEFYH AIDRASLLAR IVNTKEFLQ AIDRASLLAR
A. baylyi A. baumannii P. aeruginosa S. aureus B. subtilis S. pneumoniae	210 220 SLPSHSVIVP RKGVIELMRM LDGG AMQAVQAIVP RKAVGELQRL LSIE SSQLVQAIVP RKAVGELQRL LSIE SQDRHQVIVP RKGILELARL LTEQ VSENKNVIIP GKALAELNKI MSDNI EDRSYNVVIP GKSLTELSKI LDDNN NSDDFDVVIP SRSLREFSAV FTDD	230 240 DNPLRV QIGSNNIRAH VGE DDQLSL LIGRELLNVT INI DEQLTL LIGRELLNVT INT DGEVGI VLGQHHIRAT TGE EEDIDI FFASNQVLFK VGN QELVDI VITETQVLFK ARN IETVEI FFANNQILFR SEN	250 260 DFIFTSK IANRDKEQ HPITVRFTTK TPSRDKEQ GDITVRFTTK EFIFTSK NVNFISR NUFFSR NISFYTR) 290 300 LEAGCDLLKQ AFARAAILSN VQIAHDVFKQ SLQRVAILSN VLIGHDVFKQ SLQRVAILSN VVGDRQQLRE AFSRTAILSN LSIDNGEFYH AIDRASLLAR IIVNTKEFLQ AIDRASLLAR ITFNVVNLRQ SMERARLLSS
A. baylyi A. baumannii P. aeruginosa S. aureus B. subtilis	210 220 SLPSHSVIVP RKGVIELMRM LDGG AMQAVQAIVP RKAVGELQRL LSIE SQLVQAIVP RKAVGELQRL LSIE SQDRHQVIVP RKGILELARL LTEQ VSENKNVIIP GKALAELNKI MSDNN EDRSYNVVP GKSLTELSKI LDDN	230 240 DNPLRV QIGSNNIRAH VGE DDQLSL LIGRELLNVT INI DEQLTL LIGRELLNVT INT DGEVGI VLGQHHIRAT TGE EEDIDI FFASNQVLFK VGN QELVDI VITETQVLFK ARN IETVEI FFANNQILFR SEN	250 260 DFIFTSK IANRDKEQ HPITVRFTTK TPSRDKEQ GDITVRFTTK EFIFTSK NVNFISR NUFFSR NISFYTR) 290 300 LEAGCDLLKQ AFARAAILSN VQIAHDVFKQ SLQRVAILSN VLIGHDVFKQ SLQRVAILSN VVGDRQQLRE AFSRTAILSN LSIDNGEFYH AIDRASLLAR IIVNTKEFLQ AIDRASLLAR ITFNVVNLRQ SMERARLLSS
A. baylyi A. baumannii P. aeruginosa S. aureus B. subtilis S. pneumoniae	210 220 SLPSHSVIVP RKGVIELMRM LDGG AMQAVQAIVP RKAVGELQRL LSIE SSQLVQAIVP RKAVGELQRL LSIE SQDRHQVIVP RKGILELARL LTEQ VSENKNVIIP GKALAELNKI MSDNI EDRSYNVVIP GKSLTELSKI LDDNN NSDDFDVVIP SRSLREFSAV FTDD	230 240 DNPLRV QIGSNNIRAH VGE DDQLSL LIGRELLNVT INI DEQLTL LIGRELLNVT INT DGEVGI VLGQHHIRAT TGE EEDIDI FFASNQVLFK VGN QELVDI VITETQVLFK ARN IETVEI FFANNQILFR SEN	250 260 DFIFTSK IANRDKEQ HPITVRFTTK PPSRDKEQ GDITVRFTTK EVNFISR NVNFISR NVLFFSR N ISFYTR H ISFYTR	270 270 280 LVDGRFPDYR RVLPKNPDKH LIDGKFPDYR RVIPRGGDKH LVDGKFPDYR RVIPRGGDKL LUCGKPDYT RLFPENYEIK LLEGNYPDTT SLIPQDSKTE LLEGNYPDTD RLIPTDFNTT LLEGNYPDTD RLIPTDFNTT	290 300 LEAGCDLLKQ AFARAAILSN VQIAHDVFKQ SLQRVAILSN VVGDRQQLRE AFSRTAILSN LSIDNCEFYH AIDRASLLAR IIVNTKEFLQ AIDRASLLAR ITFNVVNLRQ SMERARLLSS VVFNTQSLRH AMERAFLISN
A. baylyi A. baumannii P. aeruginosa S. aureus B. subtilis S. pneumoniae	210 220 SLPSHSVIVP RKGVIELMRM LDGG AMQAVQAIVP RKAVGELQRL LSIE SSQLVQAIVP RKAVGELQRL LSIE SQDRHQVIVP RKGILELARL LTEQ VSENKNVIIP GKALAELNKI MSDN EDRSYNVVIP GKSLTELSKI LDDM NSDDFDVVIP SRSLREFSAV FTDD TSADLMVVLP SKSLREFSAV FTDD	230 240 DNPLRV QIGSNNIRAH VGE DDQLSL LIGRELLNVT INI DEQLTL LIGRELLNVT INT DGEVGI VLGQHHIRAT TGE EEDIDI FFASNQVLFK VGN QELVDI VITETQVLFK AKN IETVEI FFANNQILFR SEN IETVEV FFSPSQILFR SEH	250 260 DFIFTSK IANRDKEQ HPITVRFTTK TPSRDKEQ GDITVRFTTK EVNFISR NVNFISR N	270 280 270 280 270 280 270 280 280 280 280 280 280 280 280	290 300 LEAGCDLLKQ AFARAAILSN VQIAHDVFKQ SLQRVAILSN VVGDRQQLRE AFSRTAILSN LSIDNGEFYH AIDRASLLAR IIVNTKEFLQ AIDRASLLAR ITFNVVNLRQ SMERARLLSS VVFNTQSLRH AMERAFLISN
A. baylyi A. baumannii P. aeruginosa S. aureus B. subtilis S. pneumoniae	210 220 SLPSHSVIVP RKGVIELMRM LDGG AMQAVQAIVP RKAVGELQRL LSIE SSQLVQAIVP RKAVGELQRL LSIE SQDRHQVIVP RKGILELARL LTEQ VSENKNVIIP GKALAELNKI MSDN EDRSYNVVIP GKSLTELSKI LDDN NSDDFDVVIP SRSLREFSAV FTDD TSADLMVVLP SKSLREFSAV FTDD	230 240 DNPLRV QIGSNNIRAH VGE DDQLSL LIGRELLNVT INI DGQLTL LIGRELLNVT INT DGEVGI VLGQHHIRAT TGE EEDIDI FFASNQVLFK VGN QELVDI VITETQVLFK AKN IETVEI FFANNQILFR SEN IETVEV FFSPSQILFR SEH	250 260 DFIFTSK IANRDKEQ HPITVRFTTK PPSRDKEQ GDITVRFTTK EVNFISR N	270 280 270 280 270 280 280 280 280 280 280 280 280	0 290 300 LEAGCDLLKQ AFARAAILSN VQIAHDVFKQ SLQRVAILSN VVGDRQQLRE AFSRTAILSN LSIDNGEFYH AIDRASLLAR IIVNTKEFLQ AIDRASLLAR ITFNVVNLRQ SMERARLLSS VVFNTQSLRH AMERAFLISN 000000000000000000000000000000000000
 A. baylyi A. baumannii P. aeruginosa S. aureus B. subtilis S. pneumoniae S. pyogenes 	210 220 SLPSHSVIVP RKGVIELMRM LDGG AMQAVQAIVP RKAVGELQRL LSIE SQDRHQVIVP RKGILELARL LTEQ SQDRHQVIVP RKGILELARL LTEQ VSENKNVIP GKSLTELSKI LDDN NSDDFDVVIP SKSLREFSAV FTDD TSADLMVVLP SKSLREFSAV FTDD 310	230 240 DNPLRV QIGSNNIRAH VGE DDQLSL LIGRELLNVT INI DGLVGL LIGRELLNVT INT DGEVGI VLGQHHIRAT TGE EEDIDI FFASNQVLFK VGN QELVDI VITETQVLFK ARN IETVEI FFANNQILFR SEN IETVEV FFSPSQILFR SEH	250 260 DFIFTSK IANRDKEQ HPITVRFTTK PPSRDKEQ GDITVRFTTK EVNFISR NVNFISR N	270 270 280 LVDGRFPDYR RVLPRGGDKH LIDGKFPDYR RVLPRGGDKH LVDGRFPDYE RVLPRGGDKL LVGRFPDYE RVLPRGGDKL LLEGNYPDTT SLFPONYEIK LLEGNYPDTD RLIPTDFNTT LLEGNYPDTD RLIPTDFNTT LLEGNYPDTD RLIPTDFNTT LLEGNYPDTD RLIPTOFNTT LLEGNYPDTD RLIPTOFNTT LLEGNYPDT RVI A A A A A A A A A A A A A A A A A A A	290 300 LEAGCDLLKQ AFARAAILSN VQIAHDVFKQ SLQRVAILSN VVIGHDVFKQ SLQRVAILSN VVGDRQQLRE AFSRTAILSN LSIDNGEFYH AIDRASLLAR IIVNTKEFLQ AIDRASLLAR ITFNVVNLRQ SMERARLLSS VVFNTQSLRH AMERAFLISN 8
 A. baylyi A. baumannii P. aeruginosa S. aureus B. subtilis S. pneumoniae S. pyogenes E. coli 	210 220 SLPSHSVIVP RKGVIELMRM LDGG AMQAVQAIVP RKAVGELQRL LSIE SQLVQAIVP RKAVGELQRL LSIE SQDRHQVIVP RKGILELARL LTEQ VSENKNVIIP GKALAELNKI MSDNN EDRSYNVVP GKSLTELSKI LDDN NSDDFDVVIP SRSLREFSAV FTDD TSADLMVVLP SKSLREFSAV FTDD 310 320 E-KFRGVRLY VSE-NQLKIT ANNER	230 240 DNPLRV QIGSNNIRAH VGE DDQL5L LIGRELLNVT INI DGQL7L LIGRELLNVT INT DGEVGI VLGQHHIRAT TGE EEDLDI FFASNQVLFK VGN QELVDI VITETQVLFK AKN IETVEI FFANNQILFR SEN IETVEV FFSPSQILFR SEH	250 260 DFIFTSK IANRDKEQ HPITVRFTTK PSRDKEQ GDITVRFTTK EVNFISR NVNFISR N	270 ZOUR SUPERSTREETS STORESS	290 300 LEAGCDLLKQ AFARAAILSN VQIAHDVFKQ SLQRVAILSN VUGDRQQLRE AFSRTAILSN LSIDNGEFYH AIDRASLLAR IIVNTKEFLQ AIDRASLLAR ITFNVVNLRQ SMERARLLSS VVFNTQSLRH AMERAFLISN 8 8 0 0 0 0 0 0 0 0 0 0 0 0 0
 A. baylyi A. baumannii P. aeruginosa S. aureus B. subtilis S. pneumoniae S. pyogenes E. coli A. baylyi 	210 220 SLPSHSVIVP RKGVIELMRM LDGG AMQAVQAIVP RKAVGELQRL LSIE SSQLVQAIVP RKAVGELQRL LSIE SQDRHQVIVP RKGILELARL LTEQ VSENKNVIIP GKALAELNKI MSDN EDRSYNVVIP GKSLTELSKI LDDN NSDDFDVVIP SKSLREFSAV FTDD TSADLMVVLP SKSLREFSAV FTDD 	230 240 DNPLRV QIGSNNIRAH VGE DDQLSL LIGRELLNVT INI DGEVGI VLGOHHIRAT TGE EEDIDI FFASNQVLFK VGN QELVDI VITETQVLFK ARN IETVEI FFANNQILFR SEN IETVEV FFSPSQILFR SEH 	250 260 DFIFTSK IANRDKEQ HPITVRFTTK TPSRDKEQ GDITVRFTTK E	270 270 280 LVDGRFDYR RVLPKNPDKH LIDGKFPDYR RVLPRGGDKH LUDGKFPDYR RVLPRGGDKL LVDGKFDYR RLFPENYEIK LLEGNYPDTT RLFPENYEIK LLEGNYPDTD RLFPDFNTT LLEGNYPDTD RLFDFNTT LLEGNYPDTD RLFDFNTT S70 380 CENVRMMITD SVSSVQIEDA GDVSMSMTE ANQSVLVQDA) 290 300 LEAGCDLLKQ AFARAAILSN VQIADVFKQ SLQRVAILSN VVGDRQQLRE AFSRTAILSN LSIDNGEFYH AIDRASLLAR IIVNTKEFLQ AIDRASLLAR ITFNVVNLRQ SMERARLLSS VVFNTQSLRH AMERAFLISN SOC ASQ-SAAYVV MEMRL- AHP-DQTYVV MEMRV-
 A. baylyi A. baumannii P. aeruginosa S. aureus B. subtilis S. pneumoniae S. pyogenes E. coli A. baylyi A. baumannii 	210 220 SLPSHSVIVP RKGVIELMRM LDGG AMQAVQAIVP RKAVGELQRL LSIE SSQLVQAIVP RKAVGELQRL LSIE SQDRHQVIVP RKGILELARL LTEQ VSENKNVIIP GKALAELNKI MSDN EDRSYNVVIP GKSLTELSKI LDDN NSDDFDVVIP SKSLREFSAV FTDD TSADLMVVLP SKSLREFSAV FTDD 320 E-KFRGVRLY VSE-NQLKIT ANNEP E-KLRGVFLN FNP-DVLQLR ANNEP E-KLRGVFLN FNP-DSLQLR ANNEP	230 240 DNPLRV QIGSNNIRAH VGE DDQLSL LIGRELLNVT INI DEQLTL LIGRELLNVT INI DGEVGI VLGOHHIRAT TGE EEDIDI FFASNQVLFK VGN QELVDI VITETQVLFK AKN IETVEI FFANNQILFR SEN IETVEV FFSPSQILFR SEH 	250 260 DFIFTSK IANRDKEQ HPITVRFTTK IPSRDKEQ GDITVRFTTK E		290 300 LEAGCDLLKQ AFARAAILSN VQIAHDVFKQ SLQRVAILSN VVGDRQQLRE AFSRTAILSN LSIDNGEFYH AIDRASLLAR IIVNTKEFLQ AIDRASLLAR ITFNVVNLRQ SMERARLLSS VVFNTQSLRH AMERAFLISN SOC ASQ-SAAYVV MPMRL- AHP-DQTYVV MPMRV-
 A. baylyi A. baumannii P. aeruginosa S. aureus B. subtilis S. pneumoniae S. pyogenes E. coli A. baylyi A. baumannii P. aeruginosa 	210 220 SLPSHSVIVP RKGVIELMRM LDGG AMQAVQAIVP RKAVGELQRL LSIE SQULVQAIVP RKAVGELQRL LSIE SQURQVIVP RKGILELARL LTEQ VSENKNVIIP GKALAELNKI MSDN EDRSYNVVIP GKSLTELSKI LDDM NSDDFDVVIP SRSLREFSAV FTDD TSADLMVVLP SKSLREFSAV FTDD 	230 240 DNPLRV QIGSNNIRAH VGE DDQLSL LIGRELLNVT INI DGQUTL LIGRELLNVT INI DGEVGI VLGQHHIRAT TGE EEDIDI FFASNQVLFK VGN QELVDI VITETQVLFK AKN IETVEI FFASNQILFR SEN IETVEV FFSPSQILFR SEN I 330 340 EQEEAE EILD-VTYSG AEM EQDEAI EDIA-IQYQD ASI EQDEAI EDIA-IQYQS AFI EQDEAE EEVQ-VEYNG GNI	250 260 DFIFTSK IANRDKEQ HPITVRFTTK PPSRDKEQ GDITVRFTTK EVNFISR N		290 300 LEAGCDLLKQ AFARAAILSN VQIAHDVFKQ SLQRVAILSN VVGDRQQLRE AFSRTAILSN LSIDNGEFYH AIDRASLLAR IIVNTKEFLQ AIDRASLLAR ITFNVVNLRQ SMERARLSS VVFNTQSLRH AMERAFLISN
 A. baylyi A. baumannii P. aeruginosa S. aureus B. subtilis S. pneumoniae S. pyogenes E. coli A. baylyi A. baumannii P. aeruginosa S. aureus 	210 220 SLPSHSVIVP RKGVIELMRM LDGG AMQAVQAIVP RKAVGELQRL LSIE SQDRQVIVP RKAVGELQRL LSIE SQDRHQVIVP RKGILELARL LTEQ VSENKNVIP GKSLTELSKI LDDN NSDDFDVVIP SKSLREFSAV FTDD TSADLMVVLP SKSLREFSAV FTDD 310 320 E-KFRGVRLY VSE-NQLKIT ANNPI E-KLRGVFLN FNP-DVLQLR ANNPI E-KLRGVFLN FNP-DVLQLR ANNPI E-KLRGVFLN SLSN-GLLKIQ ANNPI E-KLRGVFLN SCS-QLVQLKIQ ANNPI	230 240 DNPLRV QIGSNNIRAH VGE DDQLSL LIGRELLNVT INI DGQLTL LIGRELLNVT INT DGEVGI VLGQHHIRAT TGE EEDIDI FFASNQVLFK VGN QELVDI VITETQVLFK ARN IETVEI FFANNQILFR SEN IETVEV FFSPSQILFR SEN 	250 260 DFIFTSK IANRDKEQ HPITVRFTTK PPSRDKEQ GDITVRFTTK EVNFISR NVNFISR N ISFYTR H ISFYTR H 350 350 350 MEIGFNVS YVLDVLNALK LEMSFNAQ YLLDVLSVLD LEMSFNAQ YLLDVLGVLD LEMSFNASK YMMDALKAID		290 300 LEAGCDLLKQ AFARAAILSN VQIAHDVFKQ SLQRVAILSN VVGBRQQLRE AFSRTAILSN LSIDNCEFYH AIDRASLLAR IIVNTKEFLQ AIDRASLLAR ITFNVVNLRQ SMERARLSS VVFNTQSLRH AMERAFLISN 890
 A. baylyi A. baumannii P. aeruginosa S. aureus B. subtilis S. pneumoniae S. pyogenes E. coli A. baylyi A. baumannii P. aeruginosa S. aureus B. subtilis 	210 220 SLPSHSVIVP RKGVIELMRM LDGG AMQAVQAIVP RKAVGELQRL LSIE SQDVQAVVP RKAVGELQRL LSIE SQDRQVIVP RKGILELARL LTEQ VSENKNVIIP GKALAELNKI MSDN EDRSYNVVIP GKSLTELSKI LDDN NSDDFDVVIP SRSLREFSAV FTDD TSADLMVVLP SKSLREFSAV FTDD 	230 240 DNPLRV QIGSNNIRAH VGE DDQLSL LIGRELLNVT INI DGLVGI VLGQHHIRAT TGE EEDIDI FFASNQVLFK VGN QELVDI VITETQVLFK ARN IETVEI FFANNQILFR SEN IETVEV FFSPSQILFR SEN 	250 260 DFIFTSK IANRDKEQ HPITVRFTTK PPSRDKEQ GDITVRFTTK EVNFISR NVNFISR N	270 270 280 LVDGRFPDYR RVLPKNPDKH LIDGKFPDYR RVLPRGGDKH LVDGKFPDYR RVLPRGGDKL LVDGKFPDYE RVLPRGGDKL LLEGNYPDTT RLFPENYEIK LLEGNYPDTT RLFPENYEIK LLEGNYPDTD RLIPTDFNTT LLEGNYPDTD RLIPTDFNTT LLEGNYPDTD RLIPTDFNTT LLEGNYPDTD RLIPTDFNTT LLEGNYPDTD RLIPTDFNTT AUGULTUNSMTE ANGSVLVQDA GDDVSMSMTE ANGSVLVQDA GDVSMSMTE ANGSVLVQDA GDVSMSMTE ANGSVLVQDA GDVSMSMTE ANGSVLVQDA GDVSMSMTE ANGSVLVQDA GDVSMSMTE ANGSVLVQDA GDVSMSMTE ANGSVLVQDA GDVSMSMTE ANGSVLVQDA GDVSMSMTE ANGSVLVQA GDVSMSMTE ANGSVLVQA GDVSMSMTE ANGSVLVQA GDVSMSMTE ANGSVLVQA GDVSMSMTE ANGSVLVQA GDVSMSMTE ANGSVLVQA GDVSMSMTE ANGSVLVQA GDVSMSMTE ANGSVLVQA GDVSMSMTE ANGSVLVQA GDVSMSMTE ANGSVLVA GDVSMSMTE ANGSVLVA GDVSMS) 290 300 LEAGCDLLKQ AFARAAILSN VQIAHDVFKQ SLQRVAILSN VUGDRQQLRE AFSRTAILSN LSIDNCEFYH AIDRASLLAR IIVNTKEFLQ AIDRASLLAR ITFNVVNLRQ SMERARLLSS VVFNTQSLRH AMERAFLISN 8 8 90 ASQ-SAAYVV MPMRL- AHP-DQTYVV MPMRV- AHP-DQTYVV MPMRV- AHP-DQTYVV MPMRV- AHP-DQTYVV MPMRL- IND-DSAYV MPML- BIRTY NDE-TIVQLI LFURTY
 A. baylyi A. baumannii P. aeruginosa S. aureus B. subtilis S. pneumoniae S. pyogenes E. coli A. baylyi A. baumannii P. aeruginosa S. aureus 	210 220 SLPSHSVIVP RKGVIELMRM LDGG AMQAVQAIVP RKAVGELQRL LSIE SQDRQVIVP RKAVGELQRL LSIE SQDRHQVIVP RKGILELARL LTEQ VSENKNVIP GKSLTELSKI LDDN NSDDFDVVIP SKSLREFSAV FTDD TSADLMVVLP SKSLREFSAV FTDD 310 320 E-KFRGVRLY VSE-NQLKIT ANNPI E-KLRGVFLN FNP-DVLQLR ANNPI E-KLRGVFLN FNP-DVLQLR ANNPI E-KLRGVFLN SLSN-GLLKIQ ANNPI E-KLRGVFLN SCS-QLVQLKIQ ANNPI	230 240 DNPLRV QIGSNNIRAH VGE DDQLSL LIGRELLNVT INI DGEVGI VLGOHHIRAT TGE EEDIDI FFASNQVLFK VGN QELVDI VITETQVLFK ARN IETVEI FFANNQILFR SEN IETVEV FFSPSQILFR SEH 330 340 EQEEAE EILD-VTYSG AEM EQDEAI EDIA-IQYQD ASI EQDEAI EDIA-IQYQC API EQEEAE EEVQ-VEYNG GNI EIGEVV EAUVADQIEG EEL EVGKVN EEIDTDQVTG EDI	250 260 DFIFTSK IANRDKEQ HPITVRFTTK TPSRDKEQ GDITVRFTTK E) 290 300 LEAGCDLLKQ AFARAAILSN VQIAHDVFKQ SLQRVAILSN VVGDRQQLRE AFSRTAILSN LSIDNGEFYH AIDRASLLAR IIVNTKEFLQ AIDRASLLAR ITFNVVNLRQ SMERARLLSS VVFNTQSLRH AMERAFLISN SOUND SALAN AMERAFLISN SOUND SALAN SOUND SALAN SOU

Figure S5. Sequence Alignment of SCs from four representative Gram-negative (*Escherichia coli, Acinetobacter baylyi, Acinetobacter baumannii* and *Pseudomonas aeruginosa*) and four Gram-positive bacterial species (*Staphylococcus aureus, Bacillus subtilis, Streptococcus pneumoniae* and *Streptococcus pyogenes*). Red box indicates the H148–Y154 loop region. The switching pairs of residues, i.e., Q149–D150 and R152–Y153, are highlighted in purple and V151 in green for the Gram-negative species. Residues comprising subsites I and II are highlighted in yellow. Numbering is based on the *E. coli* SC sequence.

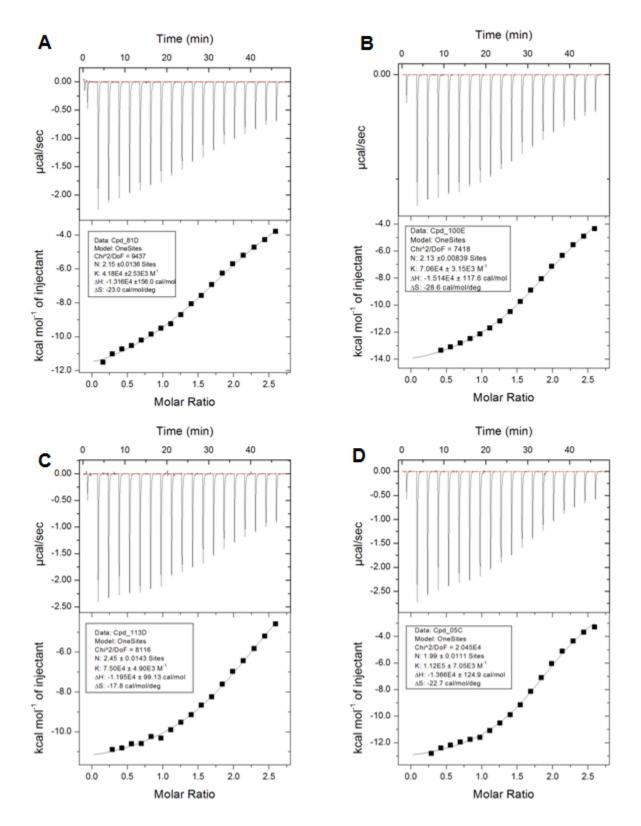


Figure S6. Isothermal titration calorimetry data for the binding of (A) 5a, (B) 5b, (C) 5I and (D) 5m to the *E. coli* SC.

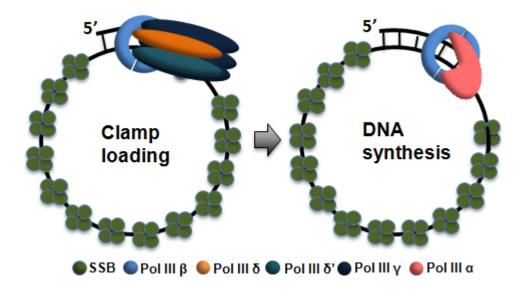


Figure S7. Schematic representation of the *in vitro* DNA replication assay.

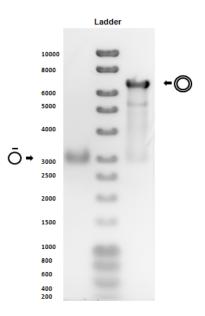


Figure S8. Control *in vitro* DNA replication assay. Molecular sizes (in bp) corresponding to bands in the DNA ladder are shown. The circle with dashed line above represents primed ssDNA template. Two concentric circles represent completed dsDNA replication products.

Supplementary Methods

Chemo-informatics. LogD values at *p*H 7.2 were calculated using Accord for Excel 6.2 (Accelrys). Calculations of Ligand Lipophilicity Efficiency (LLE_{AT}) followed the published methods⁴ and are summarized below:

 $\Delta G^* = \Delta G - \Delta G_{\text{lipo}}$

 $= RT \ln(K_i) + RT \log D$

 $LLE_{AT} = 0.11 - \Delta G^*/HAC$

 ΔG : difference in Gibbs free energy; K_i : inhibition constant; HAC: heavy atom count; LogD: distribution coefficient at *p*H 7.2.

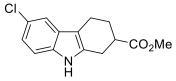
Bioinformatics. Sequence alignments of bacterial sliding clamps (NCBI IDs: YP_859300.1, WP_004930066.1, ZP_08441263.1, NP_064722.1, NP_373240.1, WP_003242509.1, YP_815419.1 and AAF98349.2) were carried out with COBALT.⁵

Chemistry – General. ¹H and ¹³C NMR spectra were acquired on a Varian Mercury 300 MHz, Varian Inova 500 MHz or VNMRS 500 MHz spectrometer. Chemical shifts (δ) are reported in ppm relative to the solvent and coupling constants (*J*) are in Hz. Electrospray ionisation (EI) low resolution mass spectra (LRMS) were recorded on a Waters Micromass Platform LCZ spectrometer. High resolution mass spectra (HRMS) were recorded on a Waters Xevo spectrometer using either an El source or atmospheric solids analysis probe (ASAP). Melting points were recorded using a Gallenkamp (Griffin) melting point apparatus and are uncorrected. Optical rotations were measured on a Jasco P-2000 polarimeter. TLC analysis was performed using pre-coated Merck silica gel 60 PF₂₅₄ aluminium sheets. Flash column chromatography was performed using Davisil silica gel (40-63 µm). Petrol refers to petroleum spirits of bp 40–60°C. All compounds examined showed ≥ 95% purity by ¹H NMR and HPLC-MS.

(±)-6-Chloro-2,3,4,9-tetrahydrocarbazole-2-carboxylic acid (1a).^{6,7} To a solution of 4-chlorophenylhydrazine hydrochloride (305 mg, 1.70 mmol) in glacial acetic acid (2 mL) was added 3-oxo-cyclohexanecarboxylic acid (237 mg, 1.67 mmol) in glacial acetic acid (2 mL) and the mixture heated at reflux overnight. The reaction mixture was cooled and the resulting precipitate collected by vacuum filtration and washed with cold water. The resultant solid was purified by silica gel column

chromatography (25:75:0.5 to 50:50:0.5 Et₂O/petrol/AcOH) followed by recrystallization from EtOH/H₂O to give **1a** (42 mg, 10% yield) as a yellow powder: mp 242–244°C (lit.⁶ 249–250°C); ¹H NMR (DMSO-*d*₆, 500 MHz): δ 1.80–183 (1H, m), 2.14–2.16 (1H, m), 2.56–2.92 (5H, m), 6.97 (1H, d, *J* = 8.0 Hz), 7.24 (1H, d, *J* = 8.5 Hz), 7.34 (1H, s), 10.90 (1H, s), 12.32 (1H, s.); ¹³C NMR (DMSO-*d*₆, 125 MHz): δ 19.4, 25.1, 25.7, 39.3, 107.6, 112.0, 116.5, 119.9, 122.8, 128.1, 134.3, 135.2, 175.97; LRMS (ES⁺) *m/z*: 272.2 [M+Na]⁺; HRMS (ASAP⁺) calcd. for C₁₃H₁₃NO₂CI [M+H]⁺ 250.0635, found 250.0628.

(±)-Methyl 6-chloro-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylate (1b).⁶ To a solution of 1a (50 mg,



0.20 mmol) in MeOH (1.5 mL) was added concentrated H_2SO_4 (200 μ L) and the mixture heated at reflux overnight. The resulting solution was cooled, concentrated to dryness, treated with saturated aqueous NaHCO₃ and extracted with EtOAc (3 x 10 mL). The combined organic extracts were

washed with water (10 mL) and brine (10 mL), dried over anhydrous MgSO₄ and concentrated. The crude residue was recrystallized from MeOH/H₂O to give **1b** (16 mg, 31% yield) as a beige powder: mp 164–166 °C (lit.⁶ 175–176 °C); ¹H NMR (DMSO-*d*₆, 500 MHz): δ 1.81–1.86 (1H, m), 2.15–2.17 (1H, m), 2.59–2.71 (2H, m), 2.89–2.96 (3H, m), 3.65 (3H, s), 6.98 (1H, dd, *J* = 18.5, 2.0 Hz), 7.26 (1H, d, *J* = 9.0 Hz), 7.35 (1H, d, *J* = 1.0 Hz), 10.94 (1H, brs); ¹³C NMR (DMSO-*d*₆, 125 MHz): δ 19.4, 25.1, 25.7, 39.1, 51.6, 107.6, 112.0, 116.6, 120.0, 122.8, 128.0, 134.3, 134.8, 174.7; LRMS (ES⁺) *m/z*: 264.1 [M+H]⁺; HRMS (ES⁺) calcd. for C₁₄H₁₅NO₂CI [M+H]⁺ 264.0791, found 264.0791.

(±)-6-Chloro-2,3,4,9-tetrahydro-1H-carbazole-2-carboxamide (1c).⁷ To a solution of 1a (75 mg, 0.30 mmol) in dry CH_2CI_2 (2 mL) containing a few drops of DMF was added *N*-hydroxysuccinimide (59 mg, 0.51 mmol) and *N*-(3-dimethylaminopropyl)-*N'*-ethylcarbodiimide hydrochloride (97 mg, 0.51 mmol) and the mixture stirred for 2.5 h at room temperature. The resulting suspension was diluted with water and extracted with CH_2CI_2 (3 x 10 mL). The combined organic extracts were

dried over anhydrous MgSO₄ and concentrated. The resultant residue was redissolved in THF (1 mL), NH₄OH (28%; 1 mL) added and the mixture stirred for 2 h. The resulting orange solution was diluted with water and extracted with EtOAc (3 x 20 mL). The combined organic extracts were washed with brine (10 mL), dried over anhydrous MgSO₄ and concentrated. Trituration of the residue with CH₂Cl₂ gave **1c** (23 mg, 31% yield) as an off-white powder: mp 194–196°C (lit.⁶ 203–204 °C); ¹H NMR (DMSO-*d*₆, 500 MHz): δ 1.69–1.75 (1H, m), 2.05–2.09 (1H, m), 2.53–2.64 (2H, m), 2.71–2.87 (3H, m), 6.87 (1H, br s), 6.97 (1H, dd, *J* = 8.0, 1.3 Hz), 7.24 (1H, d, *J* = 8.5 Hz), 7.35 (1H, s), 7.42 (1H, br s), 10.90 (1H, s); ¹³C NMR (DMSO-*d*₆, 125 MHz): δ 19.9, 25.5, 26.7, 40.4, 107.6, 112.0, 116.5, 119.9, 122.8, 128.1, 134.3, 135.7, 176.5; LRMS (ES⁺) *m/z*: 271.11 [M+Na]⁺; HRMS (ES⁺) calcd. for C₁₃H₁₃N₂OCINa [M+Na]⁺ 271.0614, found 271.0616.

(±)-(6-Chloro-2,3,4,9-tetrahydro-1H-carbazol-2-yl)methanol (1d).⁶ To a solution of 1a (75 mg, 0.30 mmol) in dry THF (2 mL) was added portion-wise LiAlH₄ (35 mg, 0.93 mmol) and the mixture stirred at room temperature for 3 h. A saturated solution of sodium potassium tartrate in water was then added and the mixture stirred for 30 min before being extracted with EtOAc (3 x 20 mL). The combined organic extracts were washed with brine (10 mL), dried over anhydrous MgSO₄ and concentrated. The resulting residue was triturated with CH₂Cl₂ to give 1d (30 mg,

20% yield) as a yellow powder: mp 152–154°C (lit.⁶ 168–169 °C); ¹H NMR (CD₃OD, 500 MHz): δ 1.50–1.52 (1H, m), 2.04–2.05 (2H, m), 2.43–2.48 (1H, m), 2.59–2.61 (1H, m), 2.71–2.74 (1H, m), 2.81–2.85 (1H, m), 3.55–3.62 (2H, m), 6.94 (1H, d, *J* = 8.0 Hz), 7.17 (1H, d, *J* = 8.5 Hz), 7.29 (1H, s); ¹³C NMR (CD₃OD, 125 MHz): δ 21.0, 27.2, 27.4, 38.4, 67.5, 109.8, 112.4, 117.6, 121.2, 125.0, 130.0, 136.2, 136.8; LRMS (ES⁺) *m/z*: 258.0 [M+Na]⁺; HRMS (ES⁺) calcd. for C₁₃H₁₃NO₂Cl [M+H]⁺ 236.0842, found 236.0831.

(*R*)-6-Chloro-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylic acid (1e). To a solution of (*R*)-3-oxo-cyclohexane-1-carboxylic acid (104 mg, 0.12 mmol) in glacial acetic acid (0.5 mL), 4-chlorophenylhydrazine hydrochloride (136 mg, 0.76 mmol) was added in glacial acetic acid (1.0 mL) and the resulting suspension heated at reflux overnight. The reaction mixture was cooled, diluted with water (15 mL) and extracted with EtOAc (3 x 15 mL). The combined organic extracts were washed with water (10 mL), brine (10 mL), dried over anhydrous MgSO₄ and

concentrated. The resulting residue was purified by silica gel column chromatography (20:80:0.5 to 50:50:0.5 Et₂O/petrol/acetic acid) followed by trituration with petrol to give **1e** (31 mg, 17% yield) as a yellow crystalline solid: mp 242–244°C (lit.⁶ 249–251°C); ¹H NMR (CD₃OD, 500 MHz): δ 1.86–1.94 (1H, m),

2.25-2.27 (1H, m), 2.60-2.66 (1H, m), 2.73-2.78 (1H, m), 2.80-2.86 (1H, m), 2.91 (2H, br d, J = 7.5 Hz), 6.95 (1H, dd, J = 8.5, 1.8 Hz), 7.17 (1H, d, J = 8.5 Hz), 7.29 (1H, s), 10.2 (1H, br s); ¹³C NMR (CD₃OD, 125 MHz): δ 20.9, 26.5, 27.5, 41.3, 109.2, 112.5, 117.7, 121.5, 125.1, 129.8, 135.7, 136.2, 178.9; LRMS (ES⁻) *m*/z: 248.0 [M-H]⁻; HRMS (ES⁻) calcd. for C₁₃H₁₁NO₂Cl [M-H]⁻ 248.0478, found 248.0486; $[\alpha]_{589}^{25}$ +47.7 (c 1.02, MeOH).

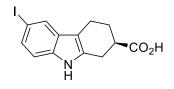
(S)-6-Chloro-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylic acid (1f). The compound was prepared according to the method described for 1e from (S)-3-oxo-cyclohexane-1carboxylic acid (101 mg, 0.71 mmol) and 4-chlorophenylhydrazine hydrochloride (133.0 mg, 0.74mmol). 1f (59 mg, 33% yield) was obtained as a yellow crystalline solid: mp 244–246°C (lit⁶ 249–250°C); ¹H NMR (CD₃OD, 500 MHz): δ 1.87–1.95 (1H, m), 2.25–2.28 (1H, m), 2.61–2.67 (1H, m), 2.74–2.79 (1H. m), 2.81–2.86 (1H, m), 2.95 (2H, d, J = 7.0Hz), 6.95 (1H, dd, J = 8.5, 1.5 Hz), 7.17 (1H, d, J = 9.0 Hz), 7.29 (1H, d, J = 1.5 Hz), 10.19 (1H, br s); ¹³C NMR (CD₃OD, 125 MHz): δ

20.9, 26.6, 27.5, 41.4, 109.3, 112.6, 117.7, 121.5, 125.2, 129.8, 135.8, 136.3, 178.9; LRMS (ES⁺) m/z: 250.0 $[M+H]^{+}$; HRMS (ASAP⁺) calcd. for C₁₃H₁₃NO₂Cl $[M+H]^{+}$ 250.0635, found 250.0643; $[\alpha]_{589}^{25}$ -45.2 (c 1.02. MeOH).

(R)-6-Fluoro-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylic acid (1g). The compound was prepared according to the method described for 1e from (R)-3-oxo-cyclohexane-1carboxylic acid (127 mg, 0.89 mmol) and 4-fluorophenylhydrazine hydrochloride (122 mg, 0.75 mmol). 1g (51 mg, 29% yield) was obtained as a yellow powder: mp 244–246°C; ¹H NMR (CD₃OD, 300 MHz): δ 1.87–1.96 (1H, m), 2.24–2.28 (1H, m), 2.58–2.86 (3H, m), 2.95 (2H, d, J = 7.2 Hz), 6.75 (1H, t, J = 9.0 Hz), 6.98 (1H, d, J = 9.9 Hz), 7.14–7.18 (1H, m), 10.09 (1H, br s); ¹³C NMR (CD₃OD,

75 MHz): δ 21.0, 26.6, 27.6, 41.4, 103.1 (d, J = 24.0 Hz), 109.1 (d, J = 26.3 Hz), 109.5 (d, J = 4.6 Hz), 112.0 (d, J = 10.3 Hz), 128.9 (d, J = 10.3 Hz), 134.3, 136.0, 158.8 (d, J = 230.0 Hz), 179.0; LRMS (ES⁻) *m*/z: 232.0 [M-H]⁻; HRMS (ES⁻) calcd. for C₁₃H₁₁NO₂F [M-H]⁻ 232.0774, found 232.0769; $[\alpha]_{589}^{25}$ +16.2 (c 0.52, MeOH).

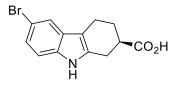
(R)-6-lodo-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylic acid (1h). The compound was prepared



according to the method described for 1e from (R)-3-oxo-cyclohexane-1carboxylic acid (84 mg, 0.59 mmol) and 4-iodophenylhydrazine (107 mg, 0.46 mmol). 1h (24 mg, 15% yield) was obtained as a brown powder: mp 202-204°C; ¹H NMR (CD₃OD, 500 MHz): δ 1.88–1.95 (1H, m), 2.25–2.28 (1H, m), 2.60-2.67 (1H, m), 2.73-277 (1H, m), 2.81-2.86 (1H, m), 2.95 (2H, d, J = 7.0 Hz), 7.05 (1H, d, J = 9.0 Hz), 7.25 (1H, d, J = 8.0 Hz), 7.65 (1H, s), 10.22 (1H,

br s); ¹³C NMR (CD₃OD, 125 MHz): δ 20.9, 26.5, 27.5, 41.4, 82.2, 108.8, 113.6, 127.3, 129.8, 131.3, 135.2, 136.9, 178.9; LRMS (ES⁻) *m/z*: 340.0 [M-H]⁻; HRMS (ES⁻) calcd. for C₁₃H₁₁NO₂I [M-H]⁻ 339.9835, found 339.9830; $[\alpha]_{589}^{25}$ –27.7 (c 0.28, MeOH).

(R)-6-Bromo-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylic acid (1i). The compound was prepared



according to the method described for 1e from (R)-3-oxo-cyclohexane-1carboxylic acid (568 mg, 3.99 mmol) and 4-bromophenylhydrazine hydrochloride (908 mg, 4.06 mmol). 1i (943 mg, 80% yield) was obtained as a yellow powder: mp 230-232°C;¹H NMR (CD₃OD, 500 MHz): δ 1.87-1.95 (1H, m), 2.25–2.28 (1H, m), 2.61–2.67 (1H, m), 2.74–2.87 (2H, m), 2.96 (2H, d, J = 7.0 Hz), 7.08 (1H, d, J = 8.5 Hz), 7.14 (1H, d, J = 8.5 Hz), 7.45 (1H, s); ¹³C NMR (CD₃OD, 125 MHz): δ 20.9, 26.5, 27.5, 41.4, 109.1, 112.6, 113.0, 120.9, 124.1, 130.5, 135.6, 136.4, 178.9; LRMS (ES⁻) m/z: 294.0 [M-H]⁻; HRMS (ES⁻) calcd. for $C_{13}H_{11}NO_2Br$ [M-H]⁻ 291.9973, found 291.9964; $[\alpha]_{589}^{25}$ +49.0 (c 0.51, MeOH).

Br

C

CI

(S)-6-Bromo-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylic acid (1). The compound was prepared according to the method described for 1e from (S)-3-oxo-cyclohexane-1carboxylic acid (241 mg, 1.70 mmol) and 4-bromophenylhydrazine hydrochloride (405 mg, 1.81 mmol). 1j (212 mg, 42% yield) was obtained as a CO₂H yellow powder: mp 236–238°C; ¹H NMR (CD₃OD, 500 MHz): δ 1.89–1.90 (1H, m), 2.24 - 2.26 (1H, m), 2.61 - 2.67 (1H, m), 2.72 - 2.95 (4H, m), 7.07 (1H, br d, J = 8.0 Hz), 7.13 (1H, d, J = 7.5 Hz), 7.44 (1H, s); ¹³C NMR (CD₃OD,

125MHz): δ 20.9, 26.5, 27.5, 41.4, 109.1, 112.5, 113.0, 120.9, 124.1, 130.4, 135.6, 136.4, 178.9; LRMS (ES⁻) m/z: 292.0 [M-H]⁻; HRMS (ES⁻) calcd. for C₁₃H₁₁NO₂Br [M-H]⁻ 291.9973, found 291.9980; $[\alpha]_{589}^{25}$ – 39.6 (c 0.52, MeOH).

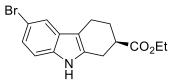
Ethyl (R)-6-chloro-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylate (1k). To a solution of 4-chlorophenylhydrazine hydrochloride (974 mg, 6.85 mmol) in glacial acetic acid (8 mL) was added (R)-3-oxo-1-cyclohexane carboxylic acid (702 mg, 4.94 mmol) and the suspension heated at reflux for 6 h. The reaction mixture was cooled, diluted with water and extracted with EtOAc (3 x 50 mL). The combined organic CO₂Et extracts were washed with water (50 mL), brine (50 mL), dried over anhydrous MgSO₄ and concentrated. The resulting crude residue was

redissolved in absolute ethanol (5 mL) containing concentrated H₂SO₄ (100 µL) and the reaction heated at reflux overnight. The solution was cooled, concentrated, made alkaline with saturated NaHCO_{3(aq)} and the mixture extracted with EtOAc (3 x 40 mL). The combined organic fractions were washed with brine (40 mL), dried over anhydrous MgSO₄ and concentrated. The resulting residue was purified by silica gel column chromatography (10:0-8:2 Et₂O/petrol) to give **1k** (877 mg, 64% yield) as a yellow powder: mp 130-132°C; ¹H NMR (CDCl₃, 500 MHz): δ 1.29 (3H, t, J = 7.3 Hz), 1.91–1.98 (1H, m), 2.28–2.31 (1H, m), 2.64–2.70 (1H, m), 2.77–3.05 (4H, m), 4.16–4.21 (2H, m), 7.06 (1H, d, J = 8.0 Hz), 7.16 (1H, d, J = 8.5 Hz), 7.39 (1H, s), 7.83 (1H, br s); ¹³C NMR (CDCl₃, 125 MHz): δ 14.4, 20.1, 25.7, 26.3, 40.3, 60.9, 109.5, 111.5, 117.6, 121.5, 125.1, 128.6, 134.0, 134.4, 175.1; LRMS (ES⁺) *m/z*: 278.0 [M+H]⁺; HRMS (ES⁺) calcd. for $C_{15}H_{17}NO_2CI [M+H]^+ 278.0948$, found 278.0938; $[\alpha]_{589}^{25} + 48.4$ (c 1.05, MeOH).

Ethyl (S)-6-chloro-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylate (11). The compound was prepared according to the method described for 1k from (S)-3-oxocyclohexane-1carboxylic acid (272 mg, 1.91 mmol) and 4-chlorophenylhydrazine hydrochloride (340 mg, 1.89 mmol). 11 was obtained (184 mg, 35% yield) as a pale 'CO₂Et yellow solid: mp 130–132°C; ¹H NMR (CDCl₃, 500 MHz): δ 1.29 (3H, t, *J* = 7.3 Hz), 1.92–1.98 (1H, m), 2.29–2.31 (1H, m), 2.65–2.71 (1H, m), 2.77–3.05 (4H, m), 4.17–4.23 (2H, m), 7.06 (1H, dd, J = 8.0, 1.5 Hz), 7.17 (1H, d, J = 9.0 Hz),

7.40 (1H, s), 7.81 (1H, br s); ¹³C NMR (CDCl₃, 125 MHz): δ 14.4, 20.1, 25.7, 26.3, 40.3, 60.9, 109.5, 111.5, 117.6, 121.6, 125.1, 128.6, 134.0, 134.4, 175.1; LRMS (ES⁻) m/z: 276.0 [M-H]⁻; HRMS (ES⁻) calcd. for $C_{15}H_{15}NO_2CI [M-H]^- 276.0791$, found 276.0780; $[\alpha]_{589}^{25} - 42.8$ (c 1.03, MeOH).

Ethyl (R)-6-bromo-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylate (1m). The compound was prepared according to the method described for 1k from (R)-3-oxocyclohexane-1-carboxylic acid (2.28 g, 10.18



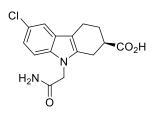
mmol) and 4-bromophenylhydrazine hydrochloride (1.45 g, 10.17 mmol). **1m** (1.99 g, 58% yield) was obtained as a yellow powder: mp 144–146°C; ¹H NMR (CDCl₃, 500 MHz): δ 1.29 (3H, t, *J* = 7.0 Hz), 1.92–1.98 (1H, m), 2.28–2.31 (1H, m), 2.65–2.70 (1H, m), 2.77–3.05 (4H, m), 4.17–4.23 (2H, m, CH₂), 7.11 (1H, d, *J* = 8.5 Hz), 7.18 (1H, d, *J* = 8.5 Hz), 7.55 (1H, s), 7.84 (1H, br s);

¹³C NMR (CDCl₃, 125 MHz): δ 14.5, 20.1, 25.7, 26.4, 40.3, 61.0, 109.5, 112.1, 112.7, 120.7, 124.2, 129.3, 133.9, 134.8, 175.2; LRMS (ES⁺) *m/z*: 362.0 [M+K]⁺; HRMS (ES⁺) calcd. for C₁₅H₁₇NO₂Br [M+H]⁺ 322.0443, found 322.0443; $[\alpha]_{589}^{25}$ +49.5 (c 1.04, MeOH).

Ethyl (S)-6-bromo-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylate (1n). The compound was prepared according to the method described for 1k from (S)-3-oxocyclohexane-1carboxylic acid (227 mg, 1.60 mmol) and 4-bromophenylhydrazine hydrochloride (387 mg, 1.73 mmol). 1n (183 mg, 35% yield) was obtained as a yellow powder: mp 136–138°C; ¹H NMR (CDCl₃, 500 MHz): δ 1.29 (3H, d, *J* = 7.3 Hz), 1.90–1.98 (1H, m), 2.28–2.31 (1H, m), 2.64–2.70 (1H, m), 2.76–3.04

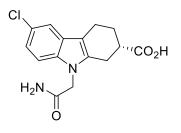
(4H, m), 4.16–4.23 (2H, m, CH₂), 7.11 (1H, d, J = 8.5 Hz), 7.18 (1H, dd, J = 8.0, 1.5 Hz), 7.55 (1H, s), 7.84 (1H, br s); ¹³C NMR (CDCl₃, 125 MHz): δ 14.4, 20.1, 25.6, 26.3, 40.3, 60.9, 109.4, 112.0, 112.6, 120.7, 124.1, 129.3, 133.8, 134.7, 175.1; LRMS (ES⁻) *m/z*: 320.0 [M-H]⁻; HRMS (ES⁻) calcd. for C₁₅H₁₅NO₂Br [M-H]⁻ 320.0286, found 320.0288; $[\alpha]_{589}^{25}$ –39.3 (c 0.98, MeOH).

(R)-9-(2-Amino-2-oxoethyl)-6-chloro-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylic acid (2a). To a



solution of **1k** (163 mg, 0.59 mmol) in dry DMF (1.0 mL) was added Cs_2CO_3 (304 mg, 0.93 mmol) and the suspension stirred at room temperature for 15 min. A solution of iodoacetamide (152 mg, 0.82 mmol) in dry DMF (0.5 mL) was added drop wise and the mixture stirred at 60°C under N₂ overnight. The reaction was quenched with 1 M HCI (20 mL) and extracted with EtOAc (3 x 20 mL). The combined extracts were washed with 0.5 M HCI (2 x 20 mL), brine (20 mL), dried over anhydrous MgSO₄ and concentrated. The crude residue was purified by

silica gel column chromatography (6:4:0–9:0:1 Et₂O/petrol/MeOH) to give ethyl (*R*)-9-(2-amino-2-oxoethyl)-6-chloro-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylate as a yellow powder. The compound was dissolved in absolute EtOH (1 mL) and an aqueous solution of NaOH (2 M, 400 µL) added. The resulting suspension was stirred at room temperature overnight. The reaction mixture was concentrated, diluted with water and washed with CH₂Cl₂ (10 mL). The aqueous layer was acidified with 1 M HCl and extracted with EtOAc (3 x 10 mL). The combined organic extracts were washed with water (10 mL), brine (10 mL), dried over anhydrous MgSO₄ and concentrated. The resulting residue was triturated with Et₂O to give **2a** (11 mg, 6% yield) as a white solid: mp > 260°C (dec.); ¹H NMR (DMSO-*d*₆, 500 MHz): δ 1.74–1.76 (1H, m), 2.17–2.20 (1H, m), 2.60–2.63 (1H, m), 2.73–2.92 (4H, m), 4.68 (2H, s), 7.05 (1H, d, *J* = 9.0 Hz), 7.23 (1H, br s), 7.33 (1H, d, *J* = 8.5 Hz), 7.41 (1H, s), 7.53 (1H, br s); ¹³C NMR (DMSO-*d*₆, 125 MHz): δ 19.7, 24.0, 25.6, 39.40, 45.2, 107.9, 110.5, 116.7, 120.1, 123.3, 127.7, 135.4, 136.7, 169.4, 176.0; LRMS (ES⁺) *m/z*: 329.0 [M+Na]⁺; HRMS (ES⁺) calcd. for C₁₅H₁₆N₂O₃Cl [M+H]⁺ 307.0849, found 307.0847; [*a*]²⁵₅₈₉ +10.4 (c 0.6, DMSO).



(S)-9-(2-Amino-2-oxoethyl)-6-chloro-2,3,4,9-tetrahydro-1H-carbazole-2-

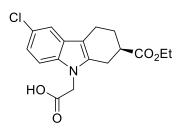
carboxylic acid (2b). The compound was prepared according to the method described for **2a** from **1I** (170 mg, 0.61 mmol), iodoacetamide (104 mg, 0.56 mmol) and Cs_2CO_3 (260 mg, 0.80 mmol) in dry DMF (1.5 mL). Following ethyl ester hydrolysis, **2b** (7.4 mg, 4% yield) was obtained as a white powder: mp >

260°C (dec.);¹H NMR (DMSO-*d*₆, 500 MHz): δ 1.74–1.76 (1H, m), 2.18–2.20 (1H, m), 2.61–2.62 (1H, m), 2.77–2.91 (4H, m), 4.68 (2H, s, CH₂), 7.05 (1H, d, *J* = 8.5 Hz), 7.23 (1H, br s), 7.34 (1H, d, *J* = 8.0 Hz), 7.40 (1H, s), 7.53 (1H, br s); ¹³C NMR (DMSO-*d*₆, 125 MHz): δ 19.7, 24.0, 25.7, 39.5, 45.2, 107.9, 110.6, 116.7, 120.1, 123.3, 127.7, 135.4, 136.7, 169.5, 176.0; LRMS (ES⁺) *m/z*: 329.0 [M+Na]⁺; HRMS (ES⁺) calcd. for C₁₅H₁₅N₂O₃ClNa [M+Na]⁺ 329.0669, found 329.0683; [α]²⁵⁸₅₈₉ –9.2 (c 0.29, DMSO).

(*R*)-2-(6-Bromo-2-(ethoxycarbonyl)-1,2,3,4-tetrahydro-9H-carbazol-9-yl)acetic acid (3a). To a solution Br CO_2Et N HO_2 HO_2 HO_2

(R)-9-(2-(tert-butoxy)-2-oxoethyl)-6-chloro-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylate, which was treated with TFA (1 mL). Following reaction for 1 h, the TFA was evaporated under a stream of N₂ to afford **3a** (402 mg, 83% yield) as a yellow powder: mp 180–182°C; ¹H NMR (DMSO-*d*₆, 500 MHz): δ 1.22 (3H, t, *J* = 6.8 Hz, CH₃), 1.77–1.78 (1H, m), 2.17–2.19 (1H, m), 2.63–2.94 (5H, m), 4.11–4.15 (2H, m), 4.92 (2H, s), 7.17 (1H, d, *J* = 8.0), 7.35 (1H, d, *J* = 8.5 Hz), 7.55 (1H, br s), 13.09 (1H, br s); ¹³C NMR (DMSO-*d*₆, 125 MHz): δ 14.1, 19.4, 23.7, 25.6, 39.3, 44.1, 60.2, 108.1, 110.7, 116.8, 120.3, 123.5, 127.7, 135.3, 136.1, 170.3, 174.2; LRMS (ES⁻) *m/z*: 380.0 [M-H]⁻; HRMS (ES⁻) calcd. for C₁₇H₁₇NO₄Br 378.0341 [M-H]⁻, found. 378.0324; [*α*]²⁵₅₈₉ +25.7 (c 1.00, MeOH).

2-(6-Chloro-2-(ethoxycarbonyl)-3,4-dihydro-1H-carbazol-9(2H)-yl)acetic acid (3c). The compound was



prepared according to the method described for **3a** from **1k** (642 mg, 2.31 mmol), *tert*-butyl bromoacetate (375 μ L, 495 mg, 2.54 mmol) and Cs₂CO₃ (1.03 g, 3.15 mmol) in DMF (4 mL). Following *tert*-butyl ester deprotection **3c** (331 mg, 43% yield) was obtained as an pale yellow powder: mp 180–182°C; ¹H NMR (DMSO-*d*₆, 500 MHz): δ 1.22 (3H, t, *J* = 7.0 Hz), 1.75–1.80 (1H, m), 2.17–2.19 (1H, m), 2.60–2.65 (1H, m), 2.73–2.94 (4H, m), 4.13 (2H, t, *J* = 6.8 Hz), 4.92 (2H, d, *J* = 2.0 Hz), 7.05 (1H, dd, *J* = 8.5, 1.8 Hz), 7.39 (1H, d, *J* = 8.5 Hz), 7.41 (1H, d, *J* = 2.0 Hz), 13.00 (1H, br s); ¹³C NMR (DMSO-*d*₆, 125

anhydrous MgSO₄ and concentrated. The resulting crude residue was

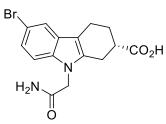
MHz): δ 14.1, 19.5, 23.7, 25.6, 39.3, 44.1, 60.2, 108.1, 110.7, 116.8, 120.3, 123.5, 127.7, 135.3, 136.1, 170.3, 174.2; LRMS (ES⁺) *m/z*: 358.0 [M+Na]⁺; HRMS (ES⁺) calcd. for C₁₇H₁₈NO₄ClNa [M+Na]⁺ 358.0822, found 358.0815; $[\alpha]_{589}^{25}$ +2.0 (c 0.97, MeOH).

(*R*)-9-(2-Amino-2-oxoethyl)-6-bromo-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylic acid (4a). To a Br R solution of **3a** (87.9 mg, 0.23 mmol) in dry CH₂Cl₂ (2 mL) was added NHS (41 mg, 0.36 mmol) and EDC (79 mg, 0.41mmol) and the reaction mixture stirred for 1 h at room temperature. The CH₂Cl₂ was evaporated under nitrogen and the resulting residue redissolved in THF (2 mL). Aqueous NH₄OH solution (28%; 1 mL) was added and the mixture stirred overnight at room temperature. The resulting precipitate was collected by vacuum filtration (washing with water) and then triturated with Et₂O. The resulting ethyl (*R*)-9-

(2-amino-2-oxoethyl)-6-bromo-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylate (29 mg, 0.08 mmol) was dissolved in absolute EtOH (2 mL), an aqueous NaOH solution (2 M; 500 μ L) added and the suspension stirred at room temperature overnight. The reaction mixture was concentrated, diluted with water, acidified

with 1 M HCl and extracted with EtOAc (3 x 10 mL). The combined extracts were washed with water (10 mL), brine (10 mL), dried over anhydrous MgSO₄ and concentrated. The resulting crude residue was triturated with Et₂O to give **4a** (12 mg, 14% yield) as a yellow powder: mp > 250°C (dec.); ¹H NMR (DMSO*d*₆, 300 MHz): δ 1.70–1.76 (1H, m), 2.16–2.20 (1H, m), 2.60–2.92 (5H, m), 4.69 (2H, s), 7.16 (1H, d, *J* = 8.7 Hz), 7.25 (1H, br s), 7.29 (1H, d, *J* = 8.7 Hz), 7.54 (2H, br s), 12.44 (1H, br s); ¹³C NMR (DMSO-*d*₆, 75 MHz): δ 19.7, 23.9, 25.6, 39.3, 45.2, 107.8, 111.1, 111.2, 119.7, 122.7, 128.4, 135.6, 136.5, 169.4, 176.0; LRMS (ES⁺) *m/z*: 373.0 [M+Na]⁺; HRMS (ES⁺) calcd. for C₁₅H₁₅N₂O₃BrNa [M+Na]⁺ 373.0164, found 373.0182; [α]²⁵⁸/₆₈₉ +12.9 (c 0.64, DMSO).

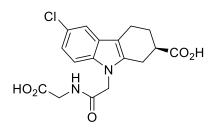
(S)-9-(2-amino-2-oxoethyl)-6-bromo-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylic acid (4b). (S)-2-(6-



bromo-2-(ethoxycarbonyl)-1,2,3,4-tetrahydro-9H-carbazol-9-yl)acetic acid (**3b**) was prepared according to the method described for **3a** from **1n** (145 g, 0.45 mmol), Cs_2CO_3 (220 mg, 0.67 mmol) and *tert*-butyl bromoacetate (115 mg, 0.59 mmol). Following TFA deprotection of the *tert*-butyl ester, (*S*)-2-(6-bromo-2-(ethoxycarbonyl)-1,2,3,4-tetrahydro-9H-carbazol-9-yl)acetic acid (92 mg, 0.24 mmol) was subsequently reacted with NHS (40 mg, 0.34 mmol) and EDC (72 mg, 0.38 mmol) according to the method described for **4a**. Ethyl ester

hydrolysis in an aqueous solution of NaOH (2 M; 300 μ L) in absolute ethanol (4 mL) yielded **4b** (28 mg, 33% yield) as a yellow powder: mp > 250°C (dec.); ¹H NMR (DMSO-*d*₆, 500 MHz): δ 1.75–1.76 (1H, m), 2.17–2.20 (1H, m), 2.59–2.62 (1H, m), 2.73–2.92 (4H, m), 4.69 (2H, s, CH₂), 7.16 (1H, d, *J* = 8.5 Hz), 7.23 (1H, br s), 7.29 (1H, d, *J* = 8.5 Hz), 7.52 (1H, br s), 7.54 (1H, s), 12.42 (1H, br s); ¹³C NMR (DMSO-*d*₆, 125 MHz): δ 19.7, 23.9, 25.6, 39.3, 45.2, 107.8, 111.1, 111.2, 119.7, 122.7, 128.3, 135.6, 136.5, 169.4, 175.9; LRMS (ES⁺) *m/z*: 373.0 [M+Na]⁺; HRMS (ES⁺) calcd. for C₁₅H₁₅N₂O₃BrNa [M+Na]⁺ 373.0164, found 373.0174; [α]²⁵₅₈₉ +4.4 (c 0.71, DMSO).

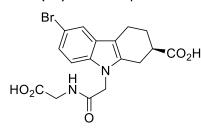
(R)-9-(2-((carboxymethyl)amino)-2-oxoethyl)-6-chloro-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylic



acid (5a). To a solution containing 3c (147 mg, 0.44 mmol), glycine methyl ester hydrochloride (69 mg, 0.55 mmol) and HATU (221 mg, 0.58 mmol) in dry DMF (1 mL) was added DIPEA (250 μ L, 1.44 mmol) dropwise and the resulting mixture stirred at room temperature overnight under N₂. The reaction mixture was diluted water and extracted with EtOAc (3 x 15 mL). The combined extracts were washed with 0.5 M HCl (3 x 15 mL), sat. NaHCO₃ (15 mL) and brine (15 mL) before being dried over anhydrous MgSO₄ and concentrated. The resulting residue

containing ethyl (*R*)-6-chloro-9-(2-((2-methoxy-2-oxoethyl)amino)-2-oxoethyl)-2,3,4,9-tetrahydro-1Hcarbazole-2-carboxylate was dissolved in absolute EtOH (1 mL), an aqueous solution of NaOH (2 M, 400 μ L) added and the suspension stirred at room temperature overnight. The reaction mixture was concentrated, diluted with water and then washed with CH₂Cl₂ (15 mL). The aqueous layer was acidified (1 M HCl) and extracted with EtOAc (3 x 15 mL). The combined organic layers were washed with water (15 mL) and brine (15 mL) before being dried over anhydrous MgSO₄ and concentrated. The resulting residue was purified by silica gel column chromatography (0:10:0–1:9:0.005 MeOH/CH₂Cl₂/acetic acid) to give **5a** (96 mg, 60% yield) as an off-white powder: mp 220–222°C; ¹H NMR (CH₃OD, 500 MHz): δ 1.92–1.99 (1H, m), 2.26–2.29 (1H, m), 2.64–2.70 (1H, m), 2.76–2.79 (1H, m), 2.89–2.98 (3H, m), 3.89 (2H, s), 4.77 (2H, s), 7.04 (1H, d, *J* = 8.5 Hz), 7.23 (1H, d, *J* = 9.0 Hz), 7.35 (1H, s), 7.96 (1H, br s); ¹³C NMR (CH₃OD, 125 MHz): δ 20.9, 25.2, 27.1, 41.2, 41.8, 46.7, 110.5, 111.0, 118.2, 122.1, 126.1, 129.8, 137.0, 137.2, 171.4, 172.6, 178.6; LRMS (ES⁻) *m/z*: 363.0 [M-H]⁻; HRMS (ES⁻) calcd. for C₁₇H₁₆N₂O₅Cl [M-H]⁻ 363.0748, found 363.0733; [α]²⁵₅₈₉ –5.7 (c 0.50, MeOH).

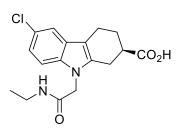
(*R*)-6-bromo-9-(2-((carboxymethyl)amino)-2-oxoethyl)-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylic acid (5b). The compound was prepared according to the method described for 5a from 3a (108 mg, 0.28



mmol), glycine methyl ester hydrochloride (49 mg, 0.39 mmol), HATU (137 mg, 0.36 mmol) and DIPEA (160 μ L, 0.92 mmol) in dry DMF (1 mL). Following ester deprotection in an aqueous solution of NaOH (2 M; 300 μ L) and absolute ethanol (1 mL), **5b** (46 mg, 40% yield) was obtained as an off white powder: mp 238–240°C; ¹H NMR (CH₃OD, 500 MHz): δ 1.92–1.95 (1H, m), 2.26–2.28 (1H, m), 2.64–2.68 (1H, m), 2.76–2.79 (1H, m), 2.89–2.98 (3H, m), 3.89 (2H, s), 4.76 (2H, s), 7.17–7.18 (2H, m), 7.50

(1H, s), 8.01 (1H, br s); ¹³C NMR (CH₃OD, 125 MHz): δ 20.9, 25.1, 27.2, 41.1, 41.8, 46.6, 110.4, 111.5, 113.5, 121.3, 124.7, 130.4, 137.1, 137.3, 171.3, 172.7, 178.6; LRMS (ES⁻) *m/z*: 407.0 [M-H]⁻; HRMS (ES⁻) calcd. for C₁₇H₁₆N₂O₅Br [M-H]⁻407.0243, found 407.0256; $[\alpha]_{589}^{25}$ +23.1 (c 0.59, MeOH).

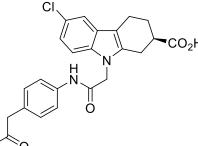
(R)-6-chloro-9-(2-(ethylamino)-2-oxoethyl)-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylic acid (5c).



The compound was prepared according to the method described for **5a** from **3c** (140 mg, 0.42 mmol), ethylamine hydrochloride (46 mg, 0.56 mmol), HATU (211 mg, 0.56 mmol) and DIPEA (200 μ L, 1.15 mmol) in dry DMF (1 mL). Ester deprotection in an aqueous solution of NaOH (2 M; 400 μ L) and absolute ethanol (1 mL) afforded **5c** (54 mg, 38% yield) as an off-white powder: mp 246–248°C; ¹H NMR (DMSO-*d*₆, 500 MHz): δ 1.02 (3H, t, *J* = 7.0 Hz), 1.75–1.76 (1H, m), 2.17–2.20 (1H, m), 2.61–2.62 (1H, m), 2.77–2.93 (4H, m), 3.08–3.12 (2H, m), 4.69 (2H, s), 7.05 (1H, d, *J* = 8.0 Hz), 7.34 (1H, d, *J* =

9.0 Hz), 7.40 (1H, s), 8.12 (1H, s), 12.43 (1H, br s); ¹³C NMR (DMSO- d_6 , 125 MHz): δ 14.6, 19.7, 24.1, 25.6, 33.5, 39.6, 45.6, 108.0, 110.6, 116.7, 120.1, 123.3, 127.7, 135.4, 136.7, 167.0, 176.0; LRMS (ES⁺) *m/z*: 357.0 [M+Na]⁺; HRMS (ES⁺) calcd. for C₁₇H₁₉N₂O₃ClNa [M+Na]⁺ 357.0982, found 357.0970; [α]²⁵₅₈₉ +2.7 (c 0.52, MeOH).

(R)-9-(2-((4-(2-amino-2-oxoethyl)phenyl)amino)-2-oxoethyl)-6-chloro-2,3,4,9-tetrahydro-1H-

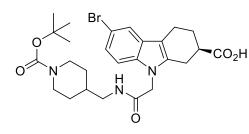


carbazole-2-carboxylic acid (5d). The compound was prepared according to the method described for **5a** from **3c** (125 mg, 0.37 mmol), 2-(4-aminophenyl)acetamide (73 mg, 0.48 mmol), HATU (181 mg, 0.48 mmol) and DIPEA (230 µL, 1.32 mmol) in dry DMF (1 mL). Ester deprotection in an aqueous solution of NaOH (2 M; 140 µL) and absolute ethanol (1 mL) gave **5d** (29 mg, 31% yield) as a white powder: mp > 260°C (dec.);¹H NMR (DMSO-*d*₆, 500 MHz): δ 1.76–1.78 (1H, m), 2.18–2.20 (1H, m), 2.63–2.96 (5H, m), 3.31 (2H, s), 4.95 (2H, s), 6.82 (2H, s), 7.06 (1H, d, *J* = 7.5 Hz), 7.19

(2H, d, J = 8.0 Hz), 7.39–7.42 (2H, m), 7.49 (2H, d, J = 8.0 Hz), 10.34 (1H, s), 12.50 (1H, br s); ¹³C NMR (DMSO- d_6 , 125 MHz): δ 19.7, 24.0, 25.7, 39.3, 41.7, 46.0, 108.1, 110.6, 116.8, 119.1, 120.2, 123.4, 127.7, 129.4, 131.7, 135.5, 136.8, 136.9, 166.1, 172.2, 176.0; LRMS (ES⁻) m/z: 438.0 [M-H]⁻; HRMS (ES⁻) calcd. for C₂₃H₂₁N₃O₄Cl [M-H]⁻ 438.1221, found 438.1215; $[\alpha]_{589}^{25}$ +3.77 (c 0.53, DMSO).

(R)-6-bromo-9-(2-(((1-(BOC)piperidin-4-yl)methyl)amino)-2-oxoethyl)-2,3,4,9-tetrahydro-1H-

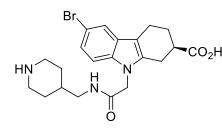
carbazole-2-carboxylic acid (5e). The compound was prepared according to the method described for **5a** from **3a** (216 mg, 0.57 mmol), *tert*-butyl 4-(aminomethyl)piperidine-1-carboxylate (159 mg, 0.74 mmol), HATU (301 mg, 0.79 mmol) and DIPEA (350 μ L, 2.01 mmol) in dry DMF (2 mL). Ester deprotection in an aqueous solution of NaOH (2 M; 300 μ L) and absolute ethanol (1 mL) provided **5e** (68 mg, 36% yield) as a



pale yellow powder: mp 158-160°C; ¹H NMR (DMSO-d₆, 500 MHz): δ 0.95–0.99 (2H, m), 1.39 (9H, s), 1.57–1.59 (3H, m), 1.74-1.77 (1H, m), 2.17-2.20 (1H, m), 2.61-2.64 (3H, m), 2.73-2.91 (3H, m), 2.94–2.97 (3H, m), 3.91 (2H, br d, J = 9.5 Hz), 4.72 (2H, s), 7.16 (1H, dd, J = 9.0, 1.8 Hz), 7.30 (1H, d, J = 9.0 Hz), 7.54 (1H, d, J = 1.5 Hz), 8.16 (1H, br t, J = 5.8 Hz), 12.44 (1H, br s); ¹³C NMR (DMSO-*d*₆, 125 MHz): δ 19.7, 24.0, 25.7, 28.1, 29.4, 35.7, 39.4, 44.0, 45.5, 78.5, 107.9, 111.2, 111.3, 119.8, 122.8,

128.4, 135.6, 136.6, 153.8, 167.5, 176.0; LRMS (ES⁺) m/z: 570.0 [M+Na]⁺; HRMS (ES⁺) calcd. for $C_{26}H_{34}N_3O_5BrNa[M+Na]^+ 570.1580$, found 570.1604; $[\alpha]_{589}^{25}$ +8.5 (c 0.52, MeOH).

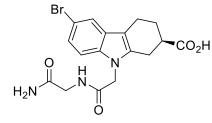
(R)-6-bromo-9-(2-oxo-2-((piperidin-4-ylmethyl)amino)ethyl)-2,3,4,9-tetrahydro-1H-carbazole-2-



carboxylic acid.TFA salt (5f). Neat TFA (1 mL) was added to 5e (42 mg, 0.076 mmol) and the mixture stirred at room temperature for 1 h. The TFA was evaporated under a stream of N₂ and the resulting residue triturated with petrol to give 5f (39 mg, 91% yield) as a yellow powder: mp 212-214°C; ¹H NMR (CH₃OD, 500 MHz): δ 1.29-1.37 (2H, m), 1.77-1.84 (3H, m), 1.98-1.99 (1H, m), 2.27-2.30 (1H, m), 2.70-2.81 (2H, m), 2.89-2.94 (5H, m), 3.13 (2H, d, J = 6.0 Hz), 3.31-3.36 (2H, m), 4.76 (2H, d, J = 3.0 Hz), 7.19 (2H, br s), 7.54 (1H, s); ¹³C

NMR (CH₃OD, 125 MHz): δ 20.8, 25.3, 27.1, 27.6, 35.2, 41.0, 44.8, 45.0, 46.8, 110.4, 111.4, 113.5, 121.4, 124.7, 130.5, 137.1, 137.3, 171.0, 178.5; LRMS (ES⁺) m/z: 448.0 [M+H]⁺; HRMS (ES⁺) calcd. for $C_{21}H_{27}N_3O_3Br[M+H]^+448.1236$, found 448.1222; $[\alpha]_{589}^{25}$ +4.6 (c 0.51, MeOH).

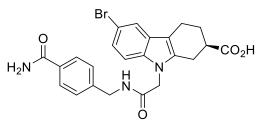
(R)-9-(2-((2-amino-2-oxoethyl)amino)-2-oxoethyl)-6-bromo-2,3,4,9-tetrahydro-1H-carbazole-2-



carboxylic acid (5g). The compound was prepared according to the method described for 5a from 3a (122 mg, 0.32 mmol), glycinamide hydrochloride (60 mg, 0.55 mmol), HATU (166 mg, 0.44 mmol) and DIPEA (230 µL, 1.32 mmol) in dry DMF (1 mL). Ester hydrolysis in an aqueous solution of NaOH (2 M; 300 µL) and absolute ethanol (1 mL) afforded 5g (8.5 mg, 6.5% yield) as an off-white powder: mp 232-234°C; ¹H NMR (DMSO-*d*₆, 500 MHz): δ 1.74–1.76 (1H, m), 2.17–2.19

(1H, m), 2.61–2.63 (1H, m), 2.73–2.95 (4H, m), 3.66 (2H, d, J = 5.5 Hz, CH₂), 4.80 (2H, s, CH₂), 7.05 (1H, s), 7.15 (1H, d, J = 8.5 Hz), 7.31–7.35 (2H, m), 7.54 (1H, s), 8.26 (1H, br s), 12.40 (1H, br s); ¹³C NMR (DMSO-*d*₆, 125 MHz): δ 19.6, 23.9, 25.7, 39.8, 41.8, 45.3, 107.9, 111.2, 119.7, 122.7, 128.4, 135.6, 136.6, 167.7, 170.4, 175.9; LRMS (ES⁻) m/z: 408.0 [M-H]⁻; HRMS (ES⁻) calcd. for C₁₇H₁₇N₃O₄Br [M-H]⁻406.0402, found 406.0404; $[\alpha]_{589}^{25}$ –9.6 (c 0.19, DMSO).

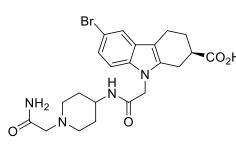
(R)-6-bromo-9-(2-((4-carbamoylbenzyl)amino)-2-oxoethyl)-2,3,4,9-tetrahydro-1H-carbazole-2-



carboxylic acid (5h). The compound was prepared according to the method described for 5a from 3a (155 mg, 0.41 mmol), 4-(aminomethyl)benzamide (74 mg, 0.49 mmol), HATU (227 mg, 0.60 mmol) and DIPEA (250 µL, 1.44 mmol) in dry DMF (1 mL). Following ester deprotection in an aqueous solution of NaOH (2 M; 300 µL) and absolute ethanol (1 mL), the mixture was acidified and the resulting precipitate collected by vacuum filtration. The precipitate was basified with 1 M NaOH, dissolved in water and filtered through a 1 cm plug

of reverse phase silica, washing with water. The aqueous solution was acidified and the precipitate collected by vacuum filtration, washing with ethanol and petrol to give **5h** (7.0 mg, 4% yield) as an off-white powder: mp > 230°C (dec); ¹H NMR (DMSO-*d*₆, 500 MHz): δ 1.75–1.77 (1H, m), 2.18–2.20 (1H, m), 2.62–2.63 (1H, m), 2.73–2.97 (4H, m), 4.34 (2H, s), 4.83 (2H, s), 7.17 (1H, br s), 7.28–7.38 (4H, m), 7.55 (1H, s), 7.82 (2H, d, *J* = 7.5 Hz), 7.93 (1H, br s), 8.78 (1H, br s), 12.44 (1H, br s); ¹³C NMR (DMSO-*d*₆, 125 MHz): δ 19.7, 24.1, 25.7, 40.2, 42.0, 45.6, 108.1, 111.2, 111.4, 119.8, 122.8, 126.9, 127.6, 128.5, 132.9, 135.7, 136.6, 142.5, 167.7, 176.1; LRMS (ES⁻) *m/z*: 482.0 [M-H]⁻; HRMS (ES⁻) calcd. for C₂₃H₂₁N₃O₄Br [M-H]⁻ 482.0715, found 482.0717; [*a*]²⁵₅₈₉ +7.4 (c 0.48, DMSO).

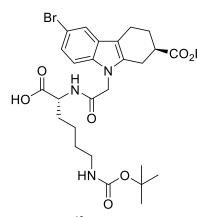
(R)-9-(2-((1-(2-amino-2-oxoethyl)piperidin-4-yl)amino)-2-oxoethyl)-6-bromo-2,3,4,9-tetrahydro-1H-



carbazole-2-carboxylic acid potassium salt (5i). The compound was prepared according to the method described for **5a** from **3a** (83 mg, 0.22 mmol), 2-(4-amino-1-piperidinyl) acetatmide dihydrochloride (70 mg, 0.25 mmol), HATU (111 mg, 0.29 mmol) and DIPEA (200 μ L, 1.15 mmol) in dry DMF (1 mL). Following deprotection in an aqueous solution of KOH (1 M, 38 μ L, 0.038 mmol) in THF:EtOH:H₂O (1:1:1; 1 mL) the mixture was concentrated, washed with CH₂Cl₂ (2 x 2 mL) and the aqueous

layer lyophilized. The resulting residue was triturated with ether, redissolved in methanol, filtered and concentrated to give the potassium salt of **5i** (13 mg, 64% yield) as an off-white gum: ¹H NMR (DMSO-*d*₆, 500 MHz): δ 1.47–1.52 (2H, m), 1.68–1.71 (3H, m), 2.06–2.11 (3H, m), 2.27–2.29 (1H, m), 2.48–2.50 (1H, m), 2.65–2.73 (5H, m), 2.81 (2H, s), 3.49–3.51 (1H, m), 4.67 (2H, d, *J* = 3.5 Hz), 7.09–7.11 (2H, m), 7.16 (1H, br s), 7.25 (1H, d, *J* = 8.5 Hz), 7.46 (1H, d, *J* = 2.0 Hz), 8.28 (1H, br d, *J* = 7.0 Hz); ¹³C NMR (DMSO-*d*₆, 125 MHz): δ 20.4, 25.6, 27.2, 31.2, 43.0, 45.5, 45.8, 52.1, 61.4, 108.1, 110.9, 119.4, 122.0, 128.8, 135.0, 139.2, 166.7, 171.9, 177.4; LRMS (ES⁺) *m/z*: 491.0 [M-K+H]⁺; HRMS (ES⁺) calcd. for C₂₂H₂₈N₄O₄Br [M-K+H]⁺ 491.1294, found 491.1310; [*a*]²⁵₅₈₉ –4.3 (c 1.28, MeOH).

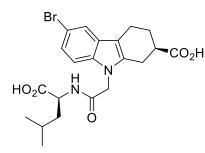
(R)-6-bromo-9-(2-(((R)-5-((tert-butoxycarbonyl)amino)-1-carboxypentyl)amino)-2-oxoethyl)-2,3,4,9-



tetrahydro-1H-carbazole-2-carboxylic acid (5j). The compound was prepared according to the method described for **5a** with the following modification. Compound **3a** (96 mg, 0.25 mmol) was pre-activated with HATU (136 mg, 0.36 mmol) and DIPEA (160 μL, 0.92 mmol) in dry CH₂Cl₂ (1 mL) containing a few drops of DMF, before a solution of NH₂-(D)-Lys-(BOC)-O^tBu.HCl (107 mg, 0.32 mmol) in dry CH₂Cl₂ (1 mL) was added. Ester deprotection with LiOH.H₂O (27 mg, 0.63 mmol) in THF:EtOH:H₂O (1:1:1; 1 mL) yielded **5j** (98 mg, 57% yield) as a yellow solid: mp 78–80°C; ¹H NMR (DMSO-*d*₆, 500 MHz): δ 1.28 (4H, br s), 1.38 (9H, s), 1.59–1.61 (1H, m), 1.71–1.72 (2H, m), 2.17–2.18 (1H, m), 2.61–2.96 (7H, m), 4.14 (1H, br s), 4.74–4.80 (2H, m), 6.75 (1H, br s), 7.16 (1H, d, *J* = 8.5 Hz), 7.31 (1H, t), 7.54 (1H, s), 8.48 (1H, t), 12.54

(1H, br s); ¹³C NMR (DMSO-*d*₆, 75 MHz): δ 19.7, 22.7, 24.0, 25.7, 28.3, 29.1, 30.8, 39.4, 39.8, 45.2, 52.1, 77.4, 107.9, 111.2, 111.3, 119.7, 122.7, 128.4, 135.6, 136.6, 155.6, 167.5, 173.4, 176.0; LRMS (ES⁻) *m/z*: 580.0 [M-H]⁻; HRMS (ES⁻) calcd. for C₂₆H₃₃N₃O₇Br [M-H]⁻ 578.1502, found 578.1504; [α]²⁵₅₈₉ +2.4 (c 2.84, MeOH).

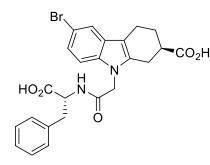
(*R*)-6-bromo-9-(2-(((S)-1-carboxy-3-methylbutyl)amino)-2-oxoethyl)-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylic acid (5k). The compound was prepared according to the method described for 5a from 3a



(249 mg, 0.654 mmol), NH₂-(L)-Leu-(O^tBu).HCl (194 mg, 0.87 mmol), HATU (339 mg, 0.891 mmol) and DIPEA (450 μ L, 2.58 mmol) in dry DMF (5 mL). Following ethyl ester hydrolysis in an aqueous solution of NaOH (2 M; 500 μ L) in EtOH:H₂O (1:1; 3 mL), the resulting residue was treated with TFA (1 mL) and the mixture stirred for 1 h. Removal of the TFA under a stream of N₂ and trituration of the residue with toluene gave **5k** (39 mg, 30% yield); ¹H NMR (CD₃OD, 500 MHz): δ 0.82–0.90 (6H, m), 1.25 (1H, br s), 1.52–1.59 (2H, m), 1.86–1.93 (1H, m), 2.25–2.28 (1H,

m), 2.63–2.69 (1H, m), 2.74–2.78 (1H, m) 2.86–2.93 (3H, m), 4.23–4.37 (1H, m), 4.76 (2H, s), 7.13 (1H, d, J = 9.0 Hz), 7.16 (1H, d, J = 8.5 Hz), 7.48 (1H, s); ¹³C NMR (CD₃OD, 125 MHz): δ 21.0, 23.5, 25.4, 26.1, 27.1, 41.5, 46.6, 49.2, 52.1, 110.3, 111.7, 113.5, 121.5, 124.5, 130.5, 137.2, 137.4, 170.8, 175.6, 178.6; LRMS (ES⁻) *m/z*: 465.0 [M-H]⁻; HRMS (ES⁻) calcd. for C₂₂H₂₀N₆OBr [M–H]⁻ 463.0882, found 463.0879; $[\alpha]_{589}^{25}$ +5.22 (c 0.46, MeOH).

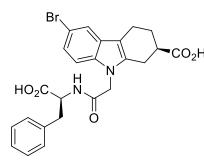
(R)-6-bromo-9-(2-(((R)-1-carboxy-2-phenylethyl)amino)-2-oxoethyl)-2,3,4,9-tetrahydro-1H-carbazole-



2-carboxylic acid (5I). The compound was prepared according to the method described for **5a** from **3a** (134 mg, 0.35 mmol), NH₂-(D)-Phe-(OMe).HCI (92 mg, 0.43 mmol), HATU (173 mg, 0.46 mmol) and DIPEA (230 μ L, 1.32 mmol) in dry DMF (1 mL). Global ester deprotection in an aqueous solution of NaOH (2 M; 300 μ L) and absolute ethanol (2 mL) afforded **5I** (69 mg, 39% yield) as an off-white powder: mp 194-196 °C; ¹H NMR (DMSO-*d*₆, 500 MHz): δ 1.72-1.73 (1H, m), 2.15 (1H, m), 2.58-2.86 (5H, m), 2.86-2.90 (1H, m), 3.06-3.09 (1H, m), 4.41 (1H, br s), 4.65-4.77 (2H, m), 7.11-7.25 (7H, m), 7.52 (1H, s), 8.36 (1H, br s), 12.70

(1H, br s); ¹³C NMR (DMSO-*d*₆, 125 MHz): δ 19.6, 23.8, 25.6, 36.7, 45.1, 53.4, 53.5, 107.9, 111.2, 111.3, 1119.7, 122.7.7, 126.5, 128.2, 128.4, 129.1, 135.5, 136.4, 137.3, 167.3, 172.6, 175.9; LRMS (ES⁺) *m/z*: 522.9 [M+Na]⁺; HRMS (ES⁺) calcd. for C₂₄H₂₃N₂O₅BrNa [M+Na]⁺ 521.0688, found 521.0686; [α]²⁵₅₈₉ -22.9 (c 0.52, MeOH).

(*R*)-6-bromo-9-(2-(((*S*)-1-carboxy-2-phenylethyl)amino)-2-oxoethyl)-2,3,4,9-tetrahydro-1H-carbazole-2-carboxylic acid (5m). The compound was prepared according to the method described for 5a from 3a



(129 mg, 0.34 mmol), NH₂-(L)-Phe-(OMe).HCl (84 mg, 0.39 mmol), HATU (163 mg, 0.43 mmol) and DIPEA (200 μ L, 1.15 mmol) in dry DMF (1 mL). Global ester deprotection in an aqueous solution of NaOH (2 M; 300 μ L) and H₂O:EtOH (1:1; 2 mL) gave **5m** (98 mg, 57% yield) as a yellow solid: mp 208–210°C; ¹H NMR (DMSO-*d*₆, 500 MHz): δ 1.73–1.75 (1H, m), 2.16 (1H, m), 2.59–2.60 (1H, m), 2.70–2.76 (3H, m), 2.87–2.93 (2H, m), 3.08–3.11 (1H, m), 4.47–4.84 (1H, m, CH), 4.66–4.78 (2H, m), 7.12–7.29 (7H, m), 7.53 (1H, s), 8.44–8.46 (1H, m), 12.63 (1H, br s); ¹³C NMR (DMSO-*d*₆, 125 MHz): δ 19.6, 23.8, 25.7, 36.7, 39.4, 45.1, 53.3,

107.9, 111.2, 111.3, 119.7, 122.7, 126.5, 128.2, 128.4, 129.1, 135.5, 136.5, 137.3, 167.3, 172.6, 175.9; LRMS (ES⁺) m/z: 521.0 [M+Na]⁺; HRMS (ES⁺) calcd. for C₂₄H₂₃N₂O₅BrNa [M+Na]⁺ 521.0688, found 521.0710; [α]²⁵₅₈₉ +17.8 (c 0.48, MeOH).

Supplementary References

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