

Supporting Information for

Does the choice of NOEC or EC10 affect the hazardous concentration for 5% of the species?

Yuichi Iwasaki^{*,†} Kensuke Kotani[‡], Shosaku Kashiwada[†], and Shigeki Masunaga[†]

^{*}Research Center for Life and Environmental Sciences, Toyo University, 1-1-1

Izumino, Itakura, Oura, Gunma 374-0193, Japan

[‡]Graduate School of Environment and Information Sciences, Yokohama National

University, 79-7 Tokiwadai, Hodogaya, Yokohama 240-8501, Japan

[†]Faculty of Environment and Information Sciences, Yokohama National University,

79-7 Tokiwadai, Hodogaya, Yokohama 240-8501, Japan

Number of pages: 13

Number of tables: 5

Number of figures: 2

Table S1. Summary table for zinc (the unit for concentration = µg/L)

Test species	Group	Endpoint	Duration (d)	NOEC	LOEC	Reduction percentage		EC5	EC10 (SE)	EC20	Best model ^b	Reference
						NOEC	LOEC					
<i>Chroococcus paris</i>	Algae	Pop. growth	10	400	1000	2.7%	17.3%	539	757 (84)	1078	W1.3	Les & Walker 1984 ¹
<i>Daphnia magna</i>	Crustaceans	Reproduction	21	100	150	5.4%	53.1%	98.7	110 (14)	124	W1.3	Munzinger & Monicelli 1991 ²
<i>Daphnia magna</i>	Crustaceans	Reproduction	21	100	150	0.0%	62.6%	137	140 (153)	143	W1.3	Munzinger & Monicelli 1991 ²
<i>Daphnia magna</i>	Crustaceans	Reproduction	21	100	150	23.3%	54.2%	53.9	70.7 (34)	93.7	W1.3	Munzinger & Monicelli 1991 ²
<i>Daphnia magna</i>	Crustaceans	Reproduction	50	25	50	9.2%	23.2%	18.1	26.3 (15)	43.6	W2.3	Paulauskis & Winner 1988 ³
<i>Daphnia magna</i>	Crustaceans	Reproduction	50	75	100	6.7%	27.7%	72.1	79.8 (4.7)	91.6	W2.3	Paulauskis & Winner 1988 ³
<i>Daphnia magna</i>	Crustaceans	Reproduction	50	150	175	6.8%	13.9%	141	163 (27)	191	LL.3	Paulauskis & Winner 1988 ³
<i>Hyalella azteca</i>	Crustaceans	Survival	70	42	108	4.1%	39.6%	45.5	59.7 (21)	79.3	W1.3	Borgmann et al. 1993 ⁴
<i>Ephoron virgo^a</i>	Insects	Survival	10	720	NA	NA	NA	NA	720 (NA)	NA	NA	van der Geest et al. 2001 ⁵
<i>Dreissena polymorpha</i>	Molluscans	Survival	70	382	1266	2.9%	54.5%	377	517 (109)	719	W1.2	Kraak et al. 1994 ⁶
<i>Potamopyrgus jenkinsi</i>	Molluscans	Growth	77-112	72	115	22.0%	53.2%	35.7	48.9 (2.6)	67.8	W1.2	Dorgelo et al. 1995 ⁷
<i>Danio rerio</i>	Fish	Time to hatch	16	360	710	27.1%	28.8%	56.9	187 (NA)	617	W2.4	Dave et al. 1987 ⁸
<i>Jordanella floridae</i>	Fish	Growth	100	26	51	7.1%	11.5%	13.3	37.4 (1.5)	110	W1.3	Spehar 1976 ⁹
<i>Oncorhynchus mykiss</i>	Fish	Survival	21 mo	320	640	1.2%	5.1%	634	1005 (48)	1881	W2.2	Sinley et al. 1974 ¹⁰
<i>Oncorhynchus mykiss</i>	Fish	Survival	>21 mo	140	260	0.0%	5.2%	258	301 (5.8)	370	W2.3	Sinley et al. 1974 ¹⁰
<i>Phoxinus phoxinus</i>	Fish	Reproduction	>137	50	130	3.6%	75.8%	52.1	57.6 (12)	66.1	W2.3	Bengtsson 1974 ¹¹
<i>Pimephales promelas</i>	Fish	Reproduction	240	78	145	10.9%	77.1%	71.5	77.2 (15)	85.6	W2.3	Benoit & Holcombe 1978 ¹²

SE = standard error

^a EC10 estimated by the original study was used as NOEC. NA = not available.^b LL.3 = three-parameter log-logistic model; W1.2 and W1.3 = two- and three-parameter Weibull-1 model; W2.2, W2.3, and W2.4 = two-, three-, and four- parameter Weibull-2 model.

Table S2. Summary table for lead (the unit for concentration = $\mu\text{g/L}$)

Test species	Group	Endpoint	Duration (d)	Reduction percentage				EC10 (SE)	EC20	Best model ^b	Reference	
				NOEC	LOEC	NOEC	LOEC					
<i>Ankistrodesmus Falcatus</i> ^a	Algae	Pop. growth	10	1018	NA	NA	NA	681	1018 (78)	1549	W1.2	Devi Prasad & Devi Pradas 1982 ¹³
<i>Chlorocoecum spp</i> ^a	Algae	Pop. growth	10	1302	NA	NA	NA	903	1302 (79)	1934	LL.2	Devi Prasad & Devi Pradas 1982 ¹³
<i>Scenedesmus obliquus</i> ^a	Algae	Pop. growth	10	1287	NA	NA	NA	913	1287 (78)	1867	LL.2	Devi Prasad & Devi Pradas 1982 ¹³
<i>Ceriodaphnia dubia</i>	Crustaceans	Reproduction	7	28.8	52.1	2.2%	7.2%	43.3	61.5 (30)	88.9	W1.3	Spehar et al. 1986 ¹⁴
<i>Daphnia magna</i>	Crustaceans	Reproduction	NA	210.6	729.0	6.1%	43.4%	187	281 (99)	430	W1.3	Enserink et al. 1995 ¹⁵
<i>Hyalella azteca</i>	Crustaceans	Survival	70	2.6	11.6	7.2%	58.3%	1.99	3.11 (3.1)	4.96	W1.3	Borgmann et al. 1993 ⁴
<i>Dreissena polymorpha</i>	Molluscans	Survival	70	85	364	7.8%	35.1%	58.2	105 (88)	198	LL.3	Kraak et al. 1994 ⁶
<i>Catostomus commersonii</i>	Fish	Survival	60	119	253	0.0%	75.8%	178	190 (557)	204	LL.3	Sauter et al. 1976 ¹⁶
<i>Danio rerio</i>	Fish	Survival time	NA	30	60	11.5%	17.1%	7.68	23.7 (7.1)	80.3	W1.4	Dave & Xiu 1991 ¹⁷
<i>Esox Lucius</i>	Fish	Survival	50	253	483	57.2%	85.7%	109	124 (57)	149	W2.3	Sauter et al. 1976 ¹⁶
<i>Ictalurus punctatus</i>	Fish	Survival	60	75	136	2.3%	84.3%	82.0	89.0 (27)	97.2	LL.3	Sauter et al. 1976 ¹⁶
<i>Lepomis macrochirus</i>	Fish	Survival	60	70	120	39.0%	81.5%	46.5	51.0 (9.4)	57.9	W2.3	Sauter et al. 1976 ¹⁶
<i>Oncorhynchus mykiss</i>	Fish	Deformity	570	18.2	32.6	0.5%	69.8%	20.5	21.6 (0.4)	23.3	W2.2	Davies et al. 1976 ¹⁸
<i>Oncorhynchus mykiss</i>	Fish	Deformity	570	4.1	7.6	0.3%	4.8%	7.67	8.99 (0.04)	10.7	LL.2	Davies et al. 1976 ¹⁸
<i>Oncorhynchus mykiss</i>	Fish	Deformity	570	7.2	14.6	0.9%	40.9%	8.73	9.76 (0.4)	11.4	W2.2	Davies et al. 1976 ¹⁸
<i>Oncorhynchus mykiss</i>	Fish	Survival	60	71	146	1.7%	49.3%	87.8	101 (21)	117	W1.3	Sauter et al. 1976 ¹⁶
<i>Salvelinus fontinalis</i>	Fish	Deformity	455	39.44	84.49	0.0%	34.0%	67.3	72.5 (0.6)	78.6	LL.2	Holcombe et al. 1976 ¹⁹
<i>Salvelinus namaycush</i>	Fish	Survival	60	48	83	2.6%	8.5%	64.9	89.3 (25)	125	W1.3	Sauter et al. 1976 ¹⁶
<i>Sander vitreus</i>	Fish	Survival	30	237	397	51.7%	74.0%	25.8	47.1 (147)	88.2	W1.3	Sauter et al. 1976 ¹⁶

SE = standard error

^a NOEC and LOEC were not indicated in the reference and EC10 was used as NOEC. NA = not available.^b LL.3 = three-parameter log-logistic model; W1.2 and W1.3 = two- and three-parameter Weibull-1 model; W2.2, W2.3, and W2.4 = two-, three-, and four- parameter Weibull-2 model.

Table S3. Summary table for nonylphenol (the unit for concentration = µg/L)

Test species	Group	Endpoint	Duration (d)			Reduction percentage		EC5	EC10 (SE)	EC20	Best model ^a	Reference
				NOEC	LOEC	NOEC	LOEC					
<i>Pseudokirchneriella subcapitata</i>	Algae	Biomass	4	694	1480	11.4%	69.8%	549	668 (107)	826	LL.3	Brooke 1993 ²⁰
<i>Ceriodaphnia dubia</i>	Crustaceans	Reproduction	7	125	250	0.4%	13.4%	206	236 (10)	272	W1.3	Tatarazako et al. 2002 ²¹
<i>Daphnia magna</i>	Crustaceans	Reproduction	21	116	215	0.0%	48.7%	180	186 (305)	195	W2.3	Brooke 1993 ²⁰
<i>Daphnia magna</i>	Crustaceans	Reproduction	21	24	39	1.8%	4.2%	42.9	63.6 (52)	94.7	W1.4	Comber et al. 1993 ²²
<i>Chironomus tentans</i>	Insects	Survival	20	42	91	0.0%	49.3%	73.0	77.2 (79)	82.1	LL.3	Kahl et al. 1997 ²³
<i>Danio rerio</i>	Fish	Survival	NA	30	100	12.3%	39.4%	17.6	26.0 (11)	44.3	W2.3	Hill & Janz 2003 ²⁴
<i>Oncorhynchus mykiss</i>	Fish	Growth	90	6	10.3	2.0%	32.1%	6.75	7.57 (0.3)	8.85	W2.3	Brooke 1993 ²⁰
<i>Oryzias latipes</i>	Fish	Survival	40	8.2	17.7	5.8%	14.1%	7.39	12.7 (4.3)	26.6	W2.3	Yokota et al. 2001 ²⁵
<i>Pimephales promelas</i>	Fish	Survival	33	7.4	14	10.6%	23.6%	4.88	7.13 (1.2)	12.0	W2.3	Ward & Boeri 1991 ²⁶

NA = not available, SE = standard error.

^aLL.3 = three-parameter log-logistic model; W1.2 and W1.3 = two- and three-parameter Weibull-1 model; W2.2, W2.3, and W2.4 = two-, three-, and four- parameter Weibull-2 model.

Table S4. Summary table for 3,4-dichlorobenzenamine (the unit for concentration = $\mu\text{g/L}$)

Test species	Group	Endpoint	Duration (d)	Reduction				EC5	EC10 (SE)	EC20	Best model ^a	Reference
				NOEC	LOEC	Percentage NOEC	Percentage LOEC					
<i>Brachionus calyciflorus</i>	Crustaceans	Pop. growth	4	2500	5000	12.0%	75.0%	2052	2474 (185)	3031	W1.4	Janssen et al. 1994 ²⁷
<i>Brachionus calyciflorus</i>	Crustaceans	Pop. growth	3	5000	10000	0.0%	90.1%	6094	6335 (4436)	6678	W2.3	Janssen et al. 1994 ²⁷
<i>Ceriodaphnia quadrangula</i>	Crustaceans	Pop. growth	21	2.97	6.15	13.5%	70.8%	2.08	2.66 (0.7)	3.44	W1.3	Kluttgen et al. 1996 ²⁸
<i>Daphnia magna</i>	Crustaceans	Pop. growth	42	10	25	5.4%	18.9%	18.2	30.7 (7.2)	54.1	LL.4	Kluttgen et al. 1996 ²⁸
<i>Daphnia magna</i>	Crustaceans	Reproduction	21	10	20	2.8%	93.5%	10.3	10.8 (1.5)	11.5	W2.3	Crossland & Hillaby 1985 ²⁹
<i>Daphnia magna</i>	Crustaceans	Reproduction	21	5	10	5.3%	23.2%	4.89	6.64 (0.5)	9.25	LL.3	Guilhermino et al 1994 ³⁰
<i>Daphnia magna</i>	Crustaceans	Reproduction	21	6	12	5.7%	18.7%	5.85	8.72 (0.2)	13.2	W1.4	Elendt 1990 ³¹
<i>Daphnia magna</i>	Crustaceans	Survival	14	2.5	5	9.6%	23.0%	1.53	2.58 (0.8)	4.46	W1.3	Diamantino et al. 1997 ³²
<i>Daphnia magna</i>	Crustaceans	Reproduction	21	5	10	7.3%	26.3%	5.32	7.52 (0.6)	12.0	W2.4	Baird 1991 ³³
<i>Gammarus pulex</i>	Crustaceans	Growth	25	80	240	1.7%	28.9%	107	136 (32)	189	W2.3	Taylor et al. 1994 ³⁴
<i>Chironomus riparius</i>	Insects	Growth	12	760	2160	11.1%	37.2%	405	699 (759)	1236	W1.3	Taylor et al. 1994 ³⁴
<i>Chironomus riparius</i>	Insects	Growth	10	170	970	0.2%	9.6%	626	998 (38)	1885	W2.3	Taylor et al. 1991 ³⁵
<i>Brachydanio rerio</i>	Fish	Survival	180	20	100	0.1%	30.8%	58.4	71.1 (14)	87.2	W1.3	Nagel et al. 1991 ³⁶
<i>Oncorhynchus mykiss</i>	Fish	Growth	28	19	39	1.0%	2.9%	13.4	20.8 (12)	33.0	W1.4	Crossland 1985 ³⁷
<i>Perca fluviatilis</i>	Fish	Survival	18	20	200	0.0%	36.4%	107	130 (350)	161	LL.3	Schafers & Nagel 1993 ³⁸
<i>Pimephales promelas</i>	Fish	Growth	28	5.1	7.1	1.1%	2.7%	9.49	14.2 (5.2)	24.4	W2.3	Call 1987 ³⁹

SE = standard error.

^aLL.3 = three-parameter log-logistic model; W1.2 and W1.3 = two- and three-parameter Weibull-1 model; W2.2, W2.3, and W2.4 = two-, three-, and four- parameter Weibull-2 model.

Table S5. Summary table for lindane (the unit for concentration = $\mu\text{g/L}$)

Test species	Group	Endpoint	Duration (d)			Reduction percentage		EC5	EC10 (SE)	EC20	Best model ^b	Reference
				NOEC	LOEC	NOEC	LOEC					
<i>Brachionus calyciflorus</i>	Crustaceans	Pop. growth	3	2500	5000	12.9%	31.4%	1258	2069 (218)	3476	W2.3	Janssen et al. 1994 ²⁷
<i>Brachionus calyciflorus</i>	Crustaceans	Pop. growth	NA	10000	15000	3.4%	54.6%	10285	10943 (781)	11908	W2.3	Ferrando et al. 1993 ⁴⁰
<i>Daphnia magna</i> ^a	Crustaceans	Growth	16	150	NA	NA	NA	NA	330 (NA)	NA	NA	Deneer et al. 1988 ⁴¹
<i>Daphnia magna</i>	Crustaceans	Survival	21	160	250	24.9%	49.4%	66.0	95.6 (19)	140.7	W1.2	Ferrando et al. 1995 ⁴²
<i>Gammarus fasciatus</i>	Crustaceans	Survival	120	4.3	8.6	67.6%	85.4%	0.909	1.11 (0.5)	1.46	W2.3	Macek et al. 1976 ⁴³
<i>Gammarus pulex</i>	Crustaceans	Growth	14	2.67	6.11	7.0%	15.5%	2.03	3.73 (1.1)	8.57	W2.3	Blockwell et al. 1996 ⁴⁴
<i>Chironomus riparius</i>	Insects	Growth	10	0.09	0.20	3.5%	6.8%	0.138	0.316 (0.06)	0.746	W1.3	Taylor et al. 1991 ³⁵
<i>Chironomus riparius</i>	Insects	Emergence	NA	1.1	9.9	4.6%	89.1%	1.12	1.34 (0.03)	1.70	W2.3	Taylor et al. 1993 ⁴⁵
<i>Chironomus tentas</i>	Insects	Emergence	NA	5	7.3	36.6%	99.8%	4.69	4.76 (1.0)	4.86	W2.3	Macek et al. 1976 ⁴³
<i>Brachydanio rerio</i>	Fish	Growth	35	40	80	0.7%	3.2%	96.8	132 (17)	183	W1.3	Gorge & Nagel 1990 ⁴⁶
<i>Brachydanio rerio</i>	Fish	Survival	14	40	80	8.3%	18.4%	26.3	47.0 (23)	86.3	W1.3	Ensenbach & Nagel 1995 ⁴⁷
<i>Pimephales promelas</i>	Fish	Survival	301	9.1	23.5	1.5%	22.5%	13.8	17.5 (4.2)	22.5	W1.3	Macek et al. 1976 ⁴³
<i>Salvelinus fontinalis</i>	Fish	Growth	261	8.8	16.6	23.6%	28.5%	0.080	0.619 (0.9)	5.20	W1.3	Macek et al. 1976 ⁴³

SE = standard error.

^a EC10 was directly obtained from the reference. NA = not available.^b LL.3 = three-parameter log-logistic model; W1.2 and W1.3 = two- and three-parameter Weibull-1 model; W2.2, W2.3, and W2.4 = two-, three-, and four- parameter Weibull-2 model.

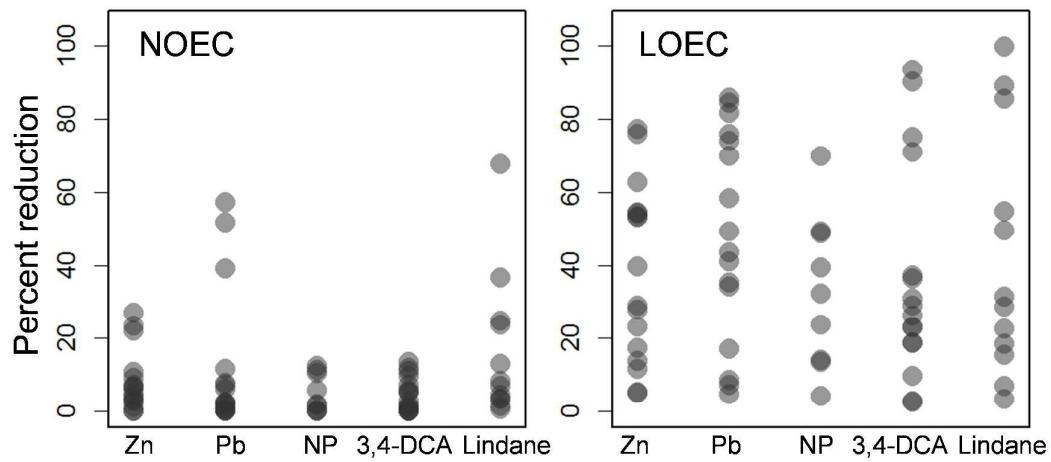


Figure S1. Percent reduction of endpoint response estimated at the no observed effect concentration (NOEC) and the lowest observed effect concentration (LOEC) for the five chemicals evaluated in this study. A Kruskal-Wallis test was performed to determine if median percent-reductions estimated at NOECs (or LOECs) were different among the five chemicals. Because results were insignificant ($p = 0.31$ and 0.72 , respectively), we pooled all the data (i.e., percent reductions) for NOECs as well as LOECs (i.e., treated as a single population of values) and used for the further analyses (see the text).

NP = Nonylphenol. 3,4-DCA = 3,4-dichlorobenzenamine.

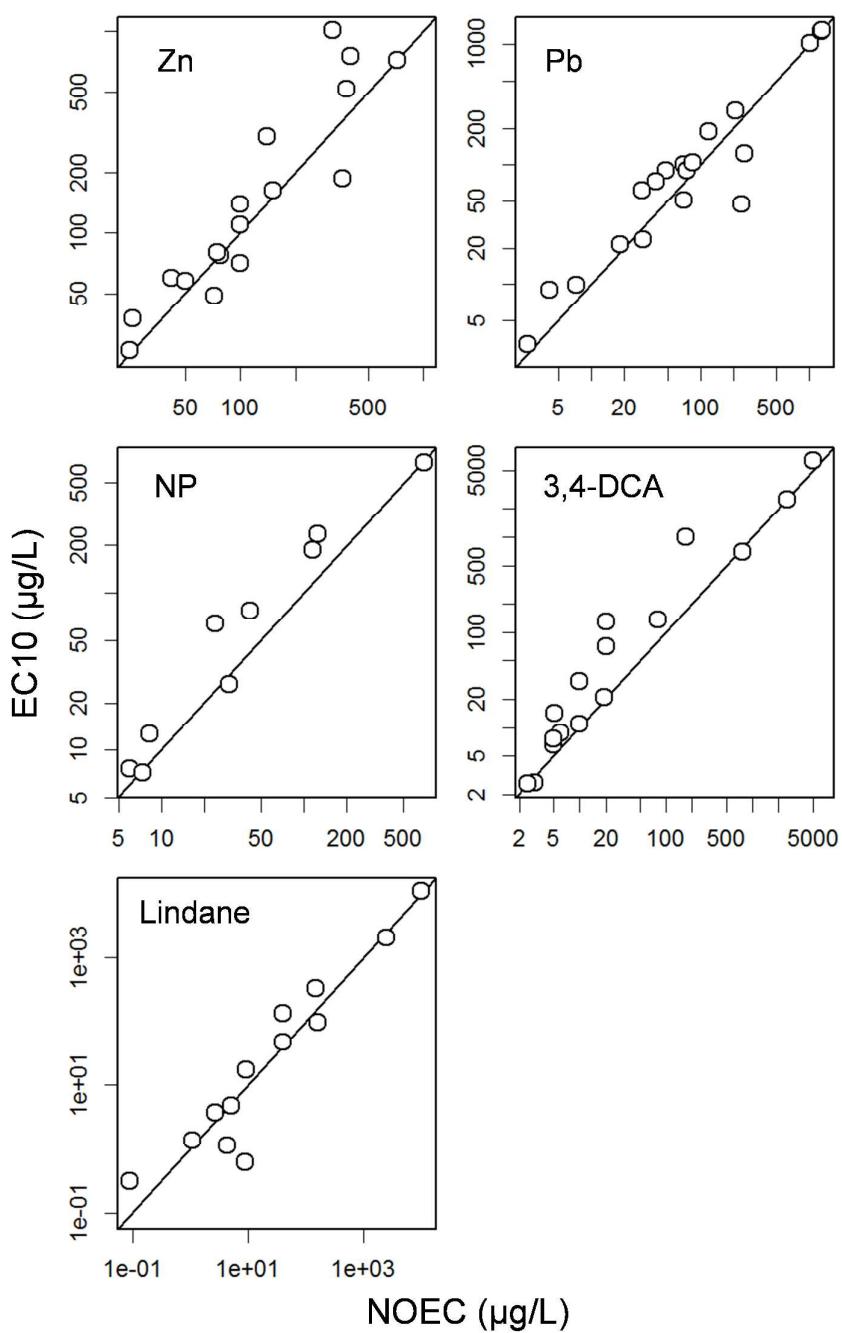


Figure S2. Relationships between the no observed effect concentration (NOEC) and the 10% effect concentration (EC10) for the five chemicals evaluated in this study.
 NP = Nonylphenol. 3,4-DCA = 3,4-dichlorobenzenamine.

References

- (1) Les, A.; Walker, R. W. Toxicity and binding of copper, zinc, and cadmium by the blue-green alga, *Chroococcus paris*. *Water Air Soil Pollut.* **1984**, *23* (2), 129–139.
- (2) Münzinger, A.; Monicelli, F. A comparison of the sensitivity of three *Daphnia magna* populations under chronic heavy metal stress. *Ecotoxicol. Environ. Saf.* **1991**, *22* (1), 24–31.
- (3) Paulauskis, J. D.; Winner, R. W. Effects of water hardness and humic acid on zinc toxicity to *Daphnia magna* Straus. *Aquat. Toxicol.* **1988**, *12* (3), 273–290.
- (4) Borgmann, U.; Norwood, W. P.; Clarke, C. Accumulation, regulation and toxicity of copper, zinc, lead and mercury in *Hyalella Azteca*. *Hydrobiologia* **1993**, *259* (2), 79–89.
- (5) van der Geest, H. G.; de Haas, E. M.; Boivin, M. E.; Admiraal, W. *Effects of zinc on larvae of the mayfly Ephoron virgo*, Report for European Zinc Industry; Universiteit van Amsterdam: 2001.
- (6) Kraak, M. H. S.; Wink, Y. A.; Stuijfzand, S. C.; Buckert-de Jong, M. C.; de Groot, C. J.; Admiraal, W. Chronic ecotoxicity of Zn and Pb to the zebra mussel *Dreissena polymorpha*. *Aquat. Toxicol.* **1994**, *30* (1), 77–89.
- (7) Dorgelo, J.; Meester, H.; Velzen, C. Effects of diet and heavy metals on growth rate and fertility in the deposit-feeding snail *Potamopyrgus jenkinsi* (Smith) (Gastropoda: Hydrobiidae). *Hydrobiologia* **1995**, *316* (3), 199–210.
- (8) Dave, G.; Damgaard, B.; Grande, M.; Martelin, J. E.; Rosander, B.; Viktor, T. Ring test of an embryo-larval toxicity test with zebrafish (*Brachydanio rerio*) using chromium and zinc as toxicants. *Environ. Toxicol. Chem.* **1987**, *6* (1), 61–71.
- (9) Spehar, R. L. *Cadmium and zinc toxicity to Jordanella floridae*; U. S. Environmental Protection Agency: Duluth, Minnesota, USA, 1976.
- (10) Sinley, J. R.; Goettl, J. P.; Davies, P. H. The effects of zinc on rainbow trout (*Salmo gairdneri*) in hard and soft water. *Bull. Environ. Contam. Toxicol.* **1974**, *12* (2), 193–201.
- (11) Bengtsson, B.-E. The effects of zinc on the mortality and reproduction of the minnow, *Phoxinus phoxinus* L. *Arch. Environ. Contam. Toxicol.* **1974**, *2* (4), 342–355.
- (12) Benoit, D. A.; Holcombe, G. W. Toxic effects of zinc on fathead minnows *Pimephales promelas* in soft water. *J. Fish Biol.* **1978**, *13* (6), 701–708.
- (13) Devi Prasad, P. V.; Devi Prasad, P. S. Effect of cadmium, lead and nickel on three

- freshwater green algae. *Water. Air. Soil Pollut.* **1982**, *17* (3), 263–268.
- (14) Spehar, R. L.; Fiandt, J. T. Acute and chronic effects of water quality criteria-based metal mixtures on three aquatic species. *Environ. Toxicol. Chem.* **1986**, *5* (10), 917–931.
- (15) Enserink, E. L.; Kerkhofs, M. J. J.; Baltus, C. A. M.; Koeman, J. H. Influence of food quantity and lead exposure on maturation in *Daphnia magna*; Evidence for a trade-off mechanism. *Funct. Ecol.* **1995**, *9* (2), 175–185.
- (16) Sauter, S.; Buxton, K. S.; Macek, K. J.; Petrocelli, S. R. *Effects of exposure to heavy metals on selected freshwater fish: toxicity of copper, cadmium, chromium and lead to eggs and fry of seven fish species*, EPA-600/3-76-105; U. S. Environmental Protection Agency: Duluth, MN, 1976; p 74.
- (17) Dave, G.; Xiu, R. Toxicity of mercury, copper, nickel, lead, and cobalt to embryos and larvae of zebrafish, *Brachydanio rerio*. *Arch. Environ. Contam. Toxicol.* **1991**, *21* (1), 126–134.
- (18) Davies, P. H.; Goettl, J. P.; Sinley, J. R.; Smith, N. F. Acute and chronic toxicity of lead to rainbow trout *Salmo gairdneri*, in hard and soft water. *Water Res.* **1976**, *10* (3), 199–206.
- (19) Holcombe, G. W.; Benoit, D. A.; Leonard, E. N.; McKim, J. M. Long-term effects of lead exposure on three generations of brook trout (*Salvelinus fontinalis*). *J. Fish. Res. Bd. Can.* **1976**, *33* (8), 1731–1741.
- (20) Brooke, L. T. *Acute and chronic toxicity of nonylphenol to ten species of aquatic organisms* (U.S. EPA Contract No. 68-CI-0034); Lake Superior Research Institute, University of Wisconsin-Superior: Superior, Wisconsin, 1993.
- (21) Tatarazako, N.; Takao, Y.; Kishi, K.; Onikura, N.; Arizono, K.; Iguchi, T. Styrene dimers and trimers affect reproduction of daphnid (*Ceriodaphnia dubia*). *Chemosphere* **2002**, *48* (6), 597–601.
- (22) Comber, M. H. I.; Williams, T. D.; Stewart, K. M. The effects of nonylphenol on *Daphnia magna*. *Water Res.* **1993**, *27* (2), 273–276.
- (23) Kahl, M. D.; Makynen, E. A.; Kosian, P. A.; Ankley, G. T. Toxicity of 4-Nonylphenol in a life-cycle test with the midge *Chironomus tentans*. *Ecotoxicol. Environ. Saf.* **1997**, *38* (2), 155–160.
- (24) Hill, R. L.; Janz, D. M. Developmental estrogenic exposure in zebrafish (*Danio rerio*): I. Effects on sex ratio and breeding success. *Aquat. Toxicol.* **2003**, *63* (4), 417–

- (25) Yokota, H.; Seki, M.; Maeda, M.; Oshima, Y.; Tadokoro, H.; Honjo, T.; Kobayashi, K. Life-cycle toxicity of 4-nonylphenol to medaka (*Oryzias latipes*). *Environ. Toxicol. Chem.* **2001**, *20* (11), 2552–2560.
- (26) Ward, T. J.; Boeri, R. L. *Early life stage toxicity of nonylphenol to the fathead minnow (Pimephales promelas)*, EnviroSystems Study Number 8979-CMA; EnviroSystems Division, Resource Analysts, Inc.: Hampton, New Hampshire, 1991.
- (27) Janssen, C. R.; Persoone, G.; Snell, T. W. Cyst-based toxicity tests. VIII. Short-chronic toxicity tests with the freshwater rotifer *Brachionus calciflorus*. *Aquat. Toxicol.* **1994**, *28* (3-4), 243–258.
- (28) Klutgen, B.; Kuntz, N.; Ratte, H. T. Combined effects of 3,4-dichloroaniline and food concentration on life-table data of two related cladocerans, *Daphnia magna* and *Ceriodaphnia quadrangula*. *Chemosphere* **1996**, *32* (10), 2015–2028.
- (29) Crossland, N. O.; Hillaby, J. M. Fate and effects of 3,4-dichloroaniline in the laboratory and in outdoor ponds: II. chronic toxicity to *Daphnia* spp. and other invertebrates. *Environ. Toxicol. Chem.* **1985**, *4* (4), 489–499.
- (30) Guilhermino, L.; Lopes, M. C.; Donato, A. M.; Silveira, L.; Carvalho, A. P.; Soares, A. M. V. M. Comparative study between the toxicity of 3,4-dichloroaniline and sodium bromide with 21-days chronic test and using lactate dehydrogenase activity of *Daphnia magna* Straus. *Chemosphere* **1994**, *28* (11), 2021–2027.
- (31) Elendt, B. P. Influence of water composition on the chronic toxicity of 3,4-dichloroaniline to *Daphnia magna*. *Water Res.* **1990**, *24* (9), 1169–1172.
- (32) Diamantino, T. C.; Ribeiro, R.; Goncalves, F.; Soares, A. M. V. M. METIER (modular ecotoxicity tests incorporating ecological relevance) for difficult substances. 4. Test chamber for cladocerans in flow-through conditions. *Environ. Toxicol. Chem.* **1997**, *16* (6), 1234–1238.
- (33) Baird, D. J.; Barber, I.; Soares, A. M. V. M.; Calow, P. An early life-stage test with *Daphnia magna* straus: An alternative to the 21-day chronic test? *Ecotoxicol. Environ. Saf.* **1991**, *22* (1), 1–7.
- (34) Taylor, E. J.; Maund, S. J.; Bennett, D.; Pascoe, D. Effects of 3,4-dichloroaniline on the growth of two freshwater macroinvertebrates in a stream mesocosm. *Ecotoxicol. Environ. Saf.* **1994**, *29* (1), 80–85.

- (35) Taylor, E. J.; Maund, S. J.; Pascoe, D. Evaluation of a chronic toxicity test using growth of the insect *Chironomus riparius* Meigen. In *Bioindicators and Environmental Management: Proceedings, 6th International Bioindicators Symposium*, Jeffrey, D. W.; Madden, B., Eds. Academic Press: London, UK, 1991; Vol. 343–352, pp 343–352.
- (36) Nagel, R.; Bresch, H.; Caspers, N.; Hansen, P. D.; Markert, M.; Munk, R.; Scholz, N.; ter Höfte, B. B. Effect of 3,4-dichloroaniline on the early life stages of the zebrafish (*Brachydanio rerio*): Results of a comparative laboratory study. *Ecotoxicol. Environ. Saf.* **1991**, *21* (2), 157–164.
- (37) Crossland, N. O. A method to evaluate effects of toxic chemicals on fish growth. *Chemosphere* **1985**, *14* (11–12), 1855–1870.
- (38) Schäfers, C.; Nagel, R. Toxicity of 3,4-dichloroaniline to perch (*Perca fluviatilis*) in acute and early life stage exposures. *Chemosphere* **1993**, *26* (9), 1641–1651.
- (39) Call, D. J.; Poirier, S. H.; Knuth, M. L.; Harting, S. L.; Lindberg, C. A. Toxicity of 3,4-dichloroaniline to fathead minnows, *Pimephales promelas*, in acute and early life-stage exposures. *Bull. Environ. Contam. Toxicol.* **1987**, *38* (2), 352–358.
- (40) Ferrando, M. D.; Janssen, C. R.; Andreu, E.; Persoone, G. Ecotoxicological studies with the freshwater rotifer *Brachionus calyciflorus* II. An assessment of the chronic toxicity of lindane and 3,4-dichloroaniline using life tables. *Hydrobiologia* **1993**, *255–256* (1), 33–40.
- (41) Deneer, J. W.; Seinen, W.; Hermens, J. L. M. Growth of *Daphnia magna* exposed to mixtures of chemicals with diverse modes of action. *Ecotoxicol. Environ. Saf.* **1988**, *15* (1), 72–77.
- (42) Ferrando, M. D.; Sancho, E.; Andreu-Moliner, E. Effects of lindane on *Daphnia magna* during chronic exposure. *Journal of Environmental Science and Health, Part B* **1995**, *30* (6), 815–825.
- (43) Macek, K. J.; Buxton, K. S.; Derr, S. K.; Dean, J. W.; Sauter, S. *Chronic toxicity of lindane to selected aquatic invertebrates and fishes*, US-EPA 600/3-76-046.; U. S. Environmental Protection Agency: 1976.
- (44) Blockwell, S. J.; Pascoe, D.; Taylor, E. J. Effects of lindane on the growth of the freshwater amphipod *Gammarus pulex* (L.). *Chemosphere* **1996**, *32* (9), 1795–1803.
- (45) Taylor, E. J.; Blockwell, S. J.; Maund, S. J.; Pascoe, D. Effects of lindane on the life-cycle of a freshwater macroinvertebrate *Chironomus riparius* Meigen (Insecta:

- Diptera). *Arch. Environ. Contam. Toxicol.* **1993**, *24* (2), 145–150.
- (46) Görge, G.; Nagel, R. Toxicity of lindane, atrazine, and deltamethrin to early life stages of zebrafish (*Brachydanio rerio*). *Ecotoxicol. Environ. Saf.* **1990**, *20* (3), 246–255.
- (47) Ensenbach, U.; Nagel, R. Toxicity of complex chemical mixtures: Acute and long-term effects on different life stages of zebrafish (*Brachydanio rerio*). *Ecotoxicol. Environ. Saf.* **1995**, *30* (2), 151–157.