Supporting Information for the Article Mobility-Enhancing Coatings for Vitreoretinal Surgical Devices: Hydrophilic and Enzymatic Coatings Investigated by Microrheology

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Figure S1: Collagen gel polymerization over time. The collagen sample was kept under a microscope (5x magnification) connected to a camera (Optronis CamRecord CL600x2) during polymerization. Greyscale camera images were recorded at 2 fps for 45 min. The average intensity of all pixels in the images with bitmap format were calculated.



(a) Magnetic field gradient generated by applying $i_{\text{gradient}}=1A$. The field at the center was adjusted by k. Remark: the fields are scaled by i_{gradient} .

(b) The gradient variation around the center of magnetic workspace (Axial position 0 mm) where the gradient was $3.0 \frac{T}{m}$ for k=0 and k=0.3.

Figure S2: Magnetic fields and field gradients superpositioned.



Figure S3: The mean |G| measured in *ex vivo* porcine vitreous by probes coated by (a) HA (n=3), (b) PEG (n=5), (c) PVP (n=7), and (d) TiO₂ (n=4). The SeM values are in error bars.



Figure S4: |G| measured in *ex vivo* porcine vitreous by PEG-coated probes. The examples 1-2 demonstrate experimental variation that is affected by individual differences of the vitreous in different *ex vivo* porcine eyes.



Figure S5: |G| measured in collagen gel by collagenase-coated probes. The examples 1-2 demonstrate experimental variation that is affected by individual differences of the collagen gel samples.



Figure S6: Probe-coating-related mobility enhancement illustrated by the mean RD values and 95% confidence intervals in error bars.