## **Supporting Information**

Biomimetic Self-Templated Hierarchical Structures of Collagenlike Peptide Amphiphiles

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**Figure S1.** AFM images of drop cast CLPA and Control-PA on Si substrates. **a-c**, 10  $\mu$ L of solution was dropped on a Si wafer and dried at room temperature.



Figure S2. AFM image and cross-sectional profile of CLPA nanofibers. The nanofibers were formed from a drop-cast CLPA solution (10  $\mu$ g/mL).



**Figure S3.** Polarized optical microscope image of the meniscus area of CLPA (5 mg/mL) filled in the capillary tube. As the meniscus receded the organized structures were deposited on the wall of the capillary tube.



Figure S4. Optical microscope images of twisted plywood-like structures created by pulling a substrate at: 30  $\mu$ m/min (a); and 70  $\mu$ m/min (b) from 1 mg/mL of CLPA solution.



**Figure S5**. Structural periodicity in twisted plywood-like phase structures (1 mg/mL) depends on pulling speed.



**Figure S6.** AFM images of the self-templated Control-PA structures. **a**, pulled at 30  $\mu$ m/min from 10 mg/mL of Control-PA solution. **b**, pulled at 50  $\mu$ m/min from 10 mg/mL of Control-PA solution. **c**, pulled at 30  $\mu$ m/min from 5 mg/mL of Control-PA solution.



**Figure S7.** Cell viability on the drop-casted CLPA film. MC-3T3 E1 cells were used at passage number 12 and seeded at a density of  $\sim 2 \times 10^2$  cells/cm<sup>2</sup> on top of the CLPA films (drop-cast). The viability of cells grown on the film for five days was measured using Live/Dead viability kit (Invitrogen, Carlsbad, CA) according to the manufacturer's instruction (live cells: green, dead cells: red). The CLPA supported over 95% cell viability throughout the experiment.



**Figure S8.** Microscope images of MC-3T3 cells cultured (five days) on glass (pre-treated with 3-aminopropyltriethoxysilane). (A) bright field, (B) merged fluorescence (actin:green, nuclei:blue) and (C) nuclei. The cells grew without preferred orientation on the glass substrates.



**Figure S9.** Self-templated CLPA mineralized by alternate dipping into  $Ca^{2+}$  (100 mM) and  $PO_4^{3-}$  (100 mM) solution 50 times. Between each dipping the film was rinsed with DI water. **a**, AFM image of the mineralized film showing the preserved micro-structural order. **b** & c, XPS analysis of the mineralized CLPA film showed a calcium to phosphorous atomic ratio of 1.64:1.

Features		Peptide amphiphile (PA)	M13 phage <sup>1</sup>
	Site-specific multi- functional peptide display	Not available	Display on the shaft of the phage body (pVIII) and at either tips (pIII, pVII, pIX)
Molecular	ar Displayable peptide with high density	- Net charge: (+), (-) (CLPA has (+) net charge)	- Net charge: (-) is favorable on pVIII
design		<ul> <li>Length: commonly 6-20 amino acids</li> <li>Amino acids (AAs): natural, unnatural and synthetic</li> <li>Crosslinkable AA: multiple cysteines</li> </ul>	<ul> <li>Length: up to 8-10 amino acids on pVIII</li> <li>Amino acids (AAs): only natural</li> <li>Crosslinkable AA: cysteine is disfavored on pVIII</li> </ul>
Building block production	Method	<ul> <li>Chemical method</li> <li>Use of organic solvents in synthesis</li> <li>Non-amplifiable</li> </ul>	<ul> <li>Biological method</li> <li>Use of water-based media in production</li> <li>Amplifiable (via E. coli infection)</li> </ul>
	Yield	~300 mg in a lab scale	~30 mg in a lab scale
Assembly property	Liquid crystalline behavior	<ul> <li>PA structure-LC behavior relationship has not been fully explored yet</li> <li>Concentration-dependent</li> </ul>	<ul> <li>Well studied and established</li> <li>Concentration-dependent (nematic, cholesteric, smectic, chiral smectic)</li> </ul>
	Self- templated structures	In CLPA case, - Twisted plywood-like phase (1 mg/mL) - Crimped filament-phase (3-5 mg/mL)	<ul> <li>Nematic orthogonal twists and cholesteric helical ribbons (1-2.0 mg/mL)</li> <li>Smectic helicoidal filaments (3-6 mg/mL)</li> </ul>

**Table S1.** Comparison of peptide amphiphile and M13 phage in self-templating assembly

## Reference

1. Chung W-J, *et al.* Biomimetic self-templating supramolecular structures. *Nature* **478**, 364-368 (2011).