

SUPPORTING INFORMATION

Beyond a fluorescent probe: Inhibition of cell division protein FtsZ by *mant*-GTP elucidated by NMR and biochemical approaches

Sonia Huecas,^{†#} Filipa Marcelo,^{†‡#} Almudena Perona,^{||} Laura B. Ruiz-Ávila,[†] Antonio Morreale,^{||} F. Javier Cañada,[†] Jesús Jiménez Barbero,^{*†} and José M. Andreu^{*†}

[†]Centro de Investigaciones Biológicas, CIB-CSIC, Ramiro de Maeztu 9, 28040 Madrid, Spain

[‡]UCIBIO, REQUIMTE, Dept. de Química, Faculdade de Ciências e Tecnologia, UNL, 2829-516 Caparica, Portugal

^{||}Unidad de Bioinformática, Centro de Biología Molecular, CBMSO-CSIC, Cantoblanco, 28049 Madrid, Spain

CONTENTS: Figures S1 to S6. Molecular dynamics Movies M1_2O'-mant-GTP, M2_3O'-mant-GTP and M3_GTP, which are available free of charge via the Internet at <http://pubs.acs.org>

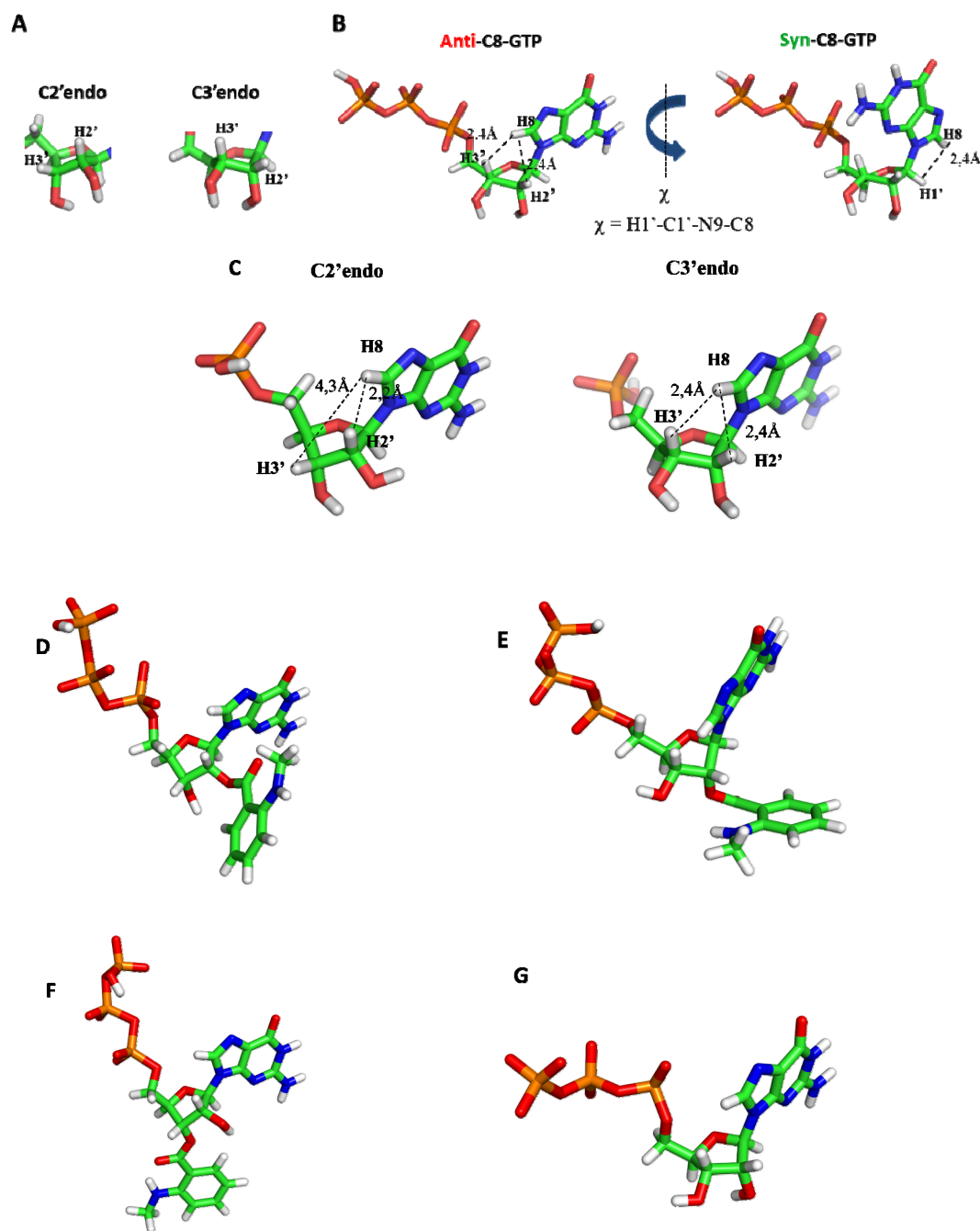


Figure S1, related to Figure 1 and Figure 2A. Guanosine nucleotide conformations. **A.** ribose puckering: C2'endo vs C3'endo. **B.** glycosidic torsion angle χ (H1'-C1'-N9-C8) and H/H proton distances for anti or syn-type conformations. **C.** H8-H2' and H8-H3' proton distances for the anti-type conformation and their relation with the C2'endo and C3'endo ribose puckering geometries. **D-F.** Selected mant-GTP structures in agreement with the NMR data that populate the major local minima found by molecular mechanics calculations. **D.** 2'-mant-GTP, anti-type, 2'endo; **E.** 2'-mant-GTP, anti-type, 3'endo; **F.** 3'-mant-GTP, anti-type, 2'endo; **G.** Non substituted GTP, anti-type, 2'endo.

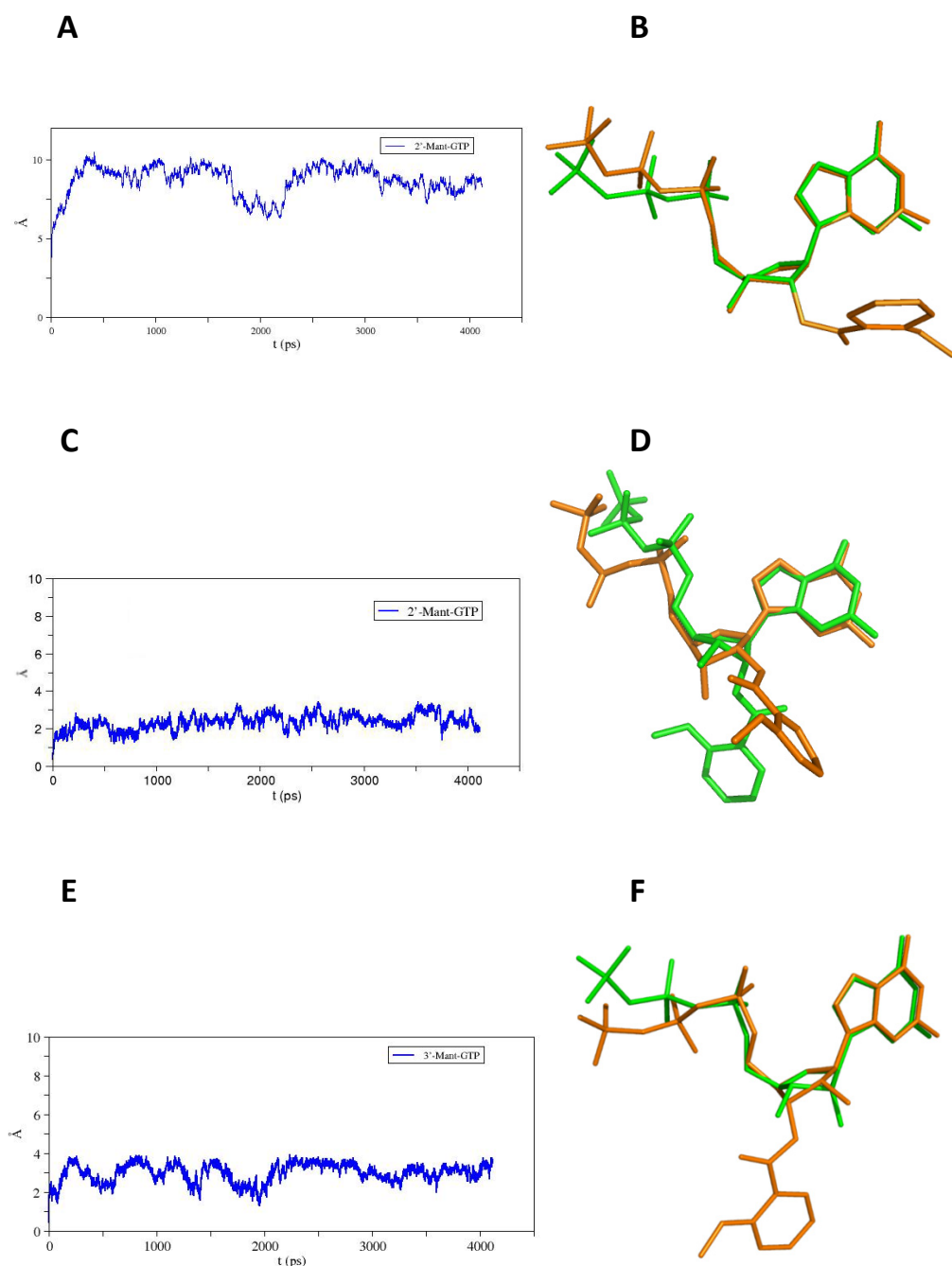


Figure S2. Molecular dynamics simulations of free mant-GTP, related to main text. **A.** RMSD value (Å) evolution along the MD simulation of 2'-mant-GTP C3'-endo in water. 2'-mant-GTP remained in the C3'-endo conformation. **B.** Superposition of the final MD model structure of free 2'-mant-GTP (orange) with the crystal structure of GTP (green) bound to Mj-FtsZ (PDB entry: 1w5a) **C.** RMSD evolution along the MD simulation of 2'-mant-GTP C2' endo, which remained in the C2' endo conformation. **D.** Superposition of the 3'endo (green) and 2'endo (orange) conformations of 2'-mant-GTP. **E.** .RMSD evolution along the MD simulation of 3'-mant-GTP in water. During the first 2 ns, 3'-mant-GTP kept the initial C3'-endo puckering, then it changed to C2'-endo for the remaining time. **F.** Superposition of the final MD model structure of free 3'-mant-GTP (orange, C2'-endo ribose puckering) with the crystal structure of GTP (green, C3'-endo) bound to Mj-FtsZ (PDB: 1w5a).

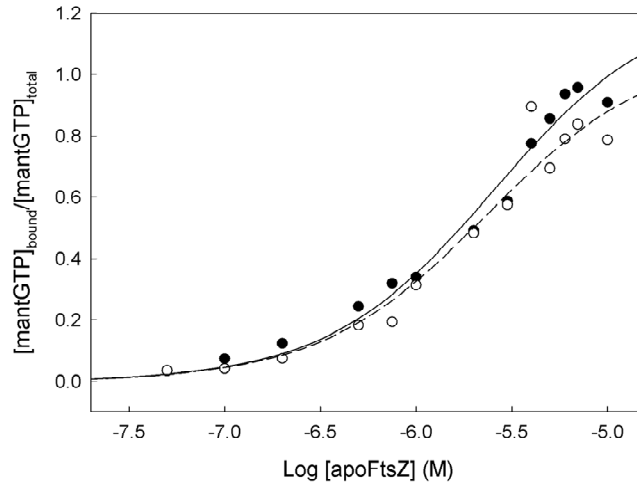
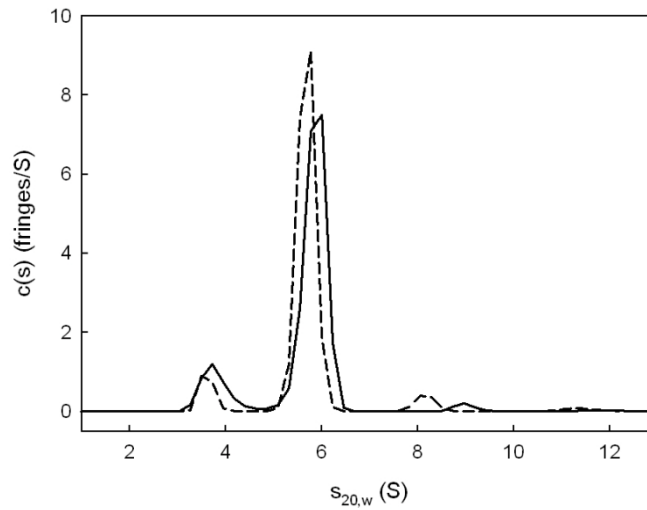
A**B**

Figure S3, related to Figure 2B-C. Biochemical characterization of FtsZ-*mant*-GTP interactions under NMR experimental conditions. A. Binding of *mant*-GTP by FtsZ. Titrations of *mant*-GTP (50 nM) binding by nucleotide-devoid Mj-FtsZ in Tris-H₂O buffer (○) and in D₂O buffer (●), both with 10 mM MgCl₂ added, measured by fluorescence anisotropy at 25 °C. The lines in each case correspond to the best single site model fits to the data: K_b $(4.3 \pm 1.7) \times 10^5 \text{ M}^{-1}$ in aqueous buffer and $(3.9 \pm 2.3) \times 10^5 \text{ M}^{-1}$ in deuterated buffer, with 1.1 ± 0.1 and 1.2 ± 0.1 binding sites respectively. **B. FtsZ association state with *mant*-GTP and GTP.** AUC sedimentation velocity profiles of Mj-FtsZ (30 μM) in Tris-D₂O buffer (no MgCl₂) at 25°C, in the presence of *mant*-GTP (0.6 mM, solid line) or GTP (1.2 mM, dashed line). *Mant*-GTP: 82.2% $s_{20,w}=5.8\text{S}$, 14.5% $s_{20,w}=3.8$ and 1.9% $s_{20,w}=8.9\text{S}$. GTP: 86.7% $s_{20,w}=5.7\text{S}$, 7.8% $s_{20,w}=3.5\text{S}$ and 4% $s_{20,w}=8$.

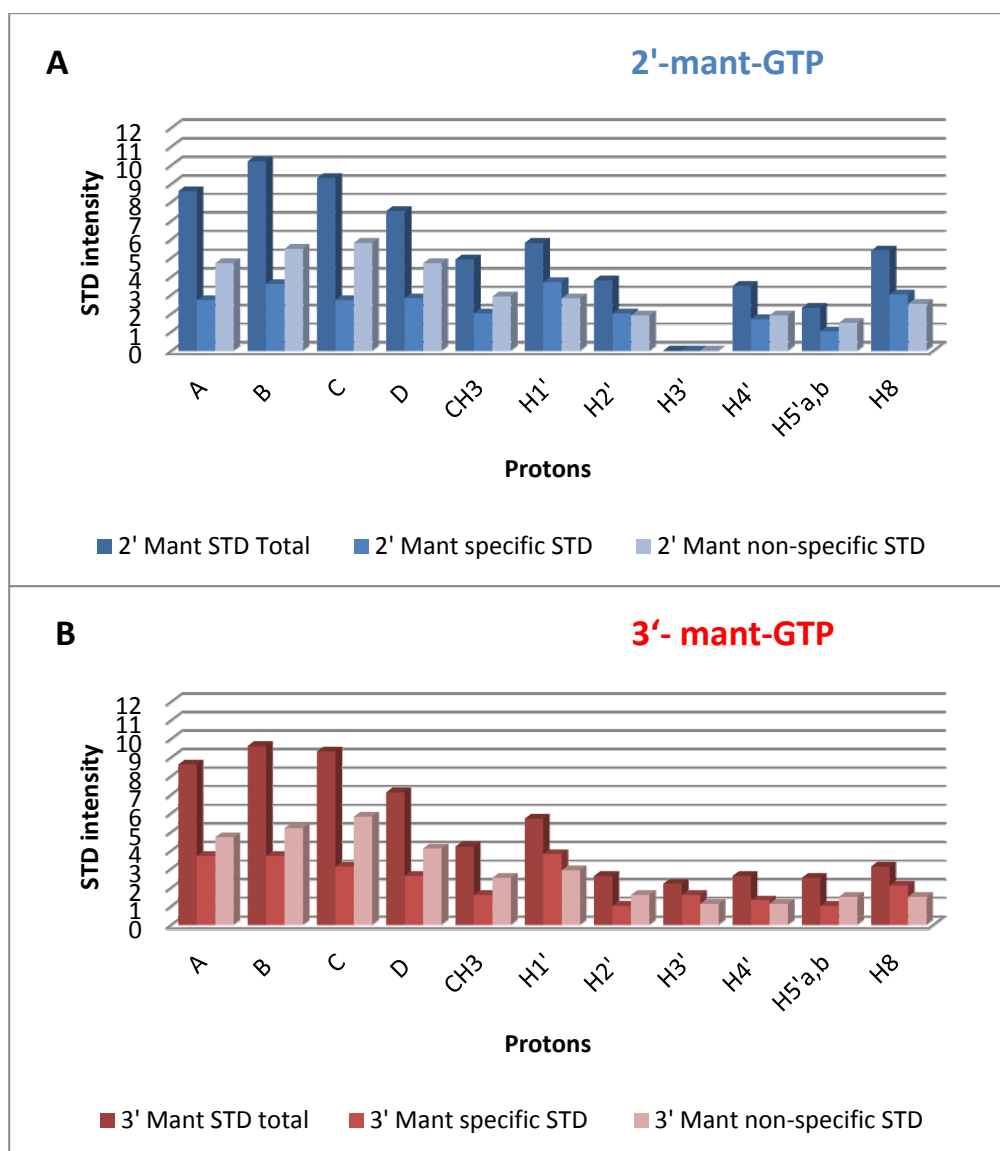


Figure S4, related to Figure 3. STD-NMR total, specific and non-specific intensities in binding experiments of mant-GTP to Mj-FtsZ. A. 2'- mant-GTP B. 3'-mant-GTP. Non specific intensities were determined in a 4-fold GTP excess over mant-GTP and subtracted from the total intensities to provide the specific intensities. For both isomers the non-specific contribution mostly arises from the aromatic fluorescent tag moiety. STD intensity values are percentages relative to each off-resonance signal. In the absence of Mj-FtsZ STD intensities were below 0.5% for any of the *mant*-GTP protons measured.

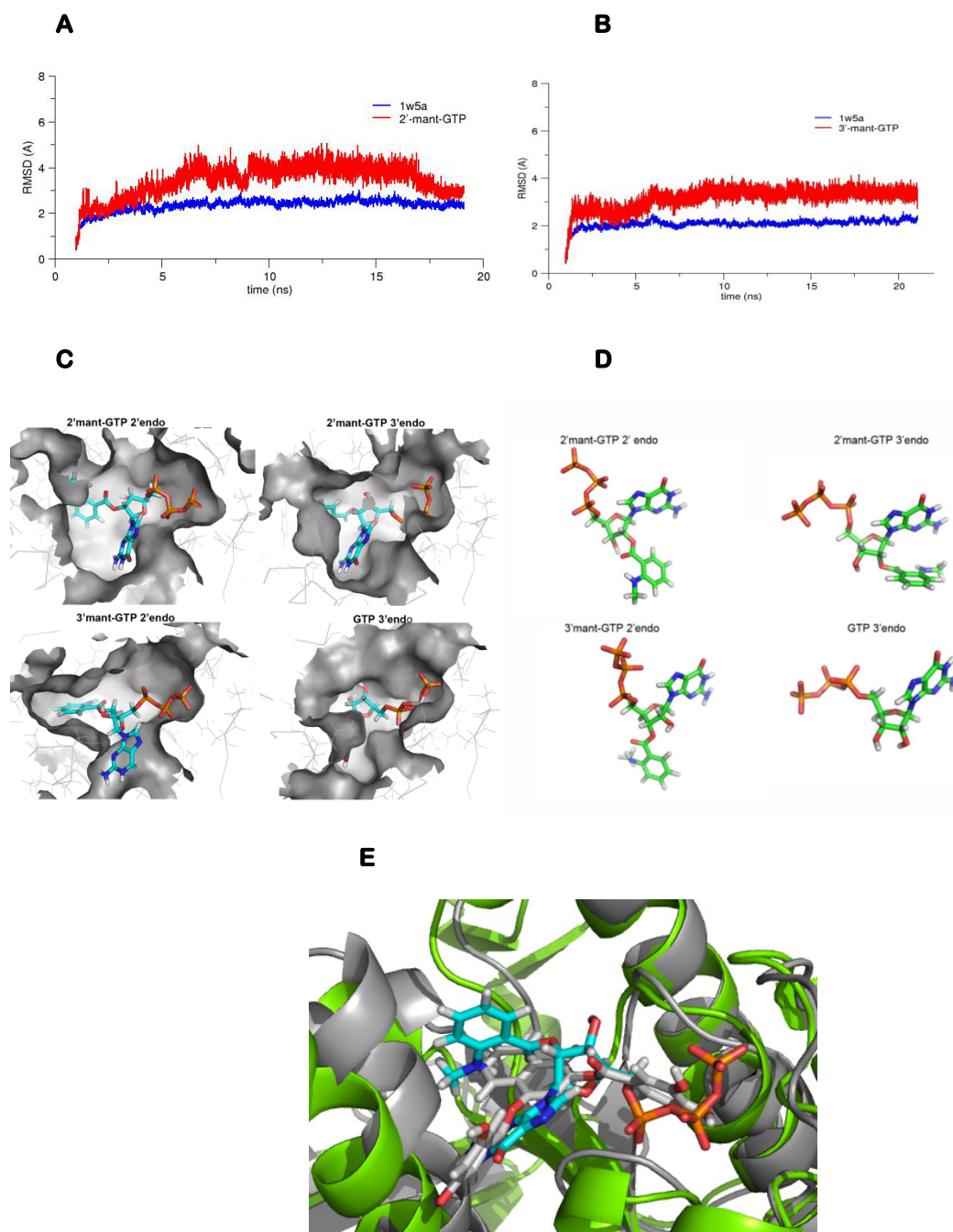
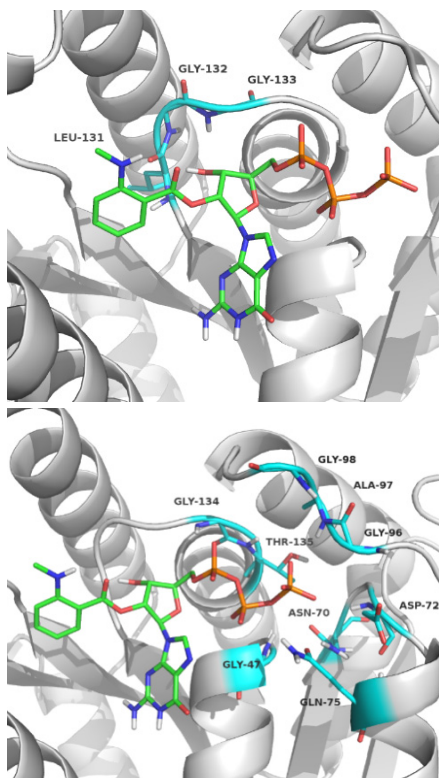
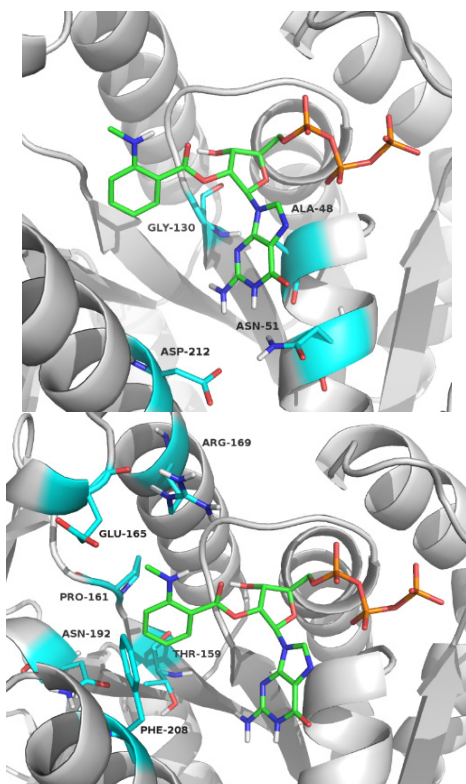
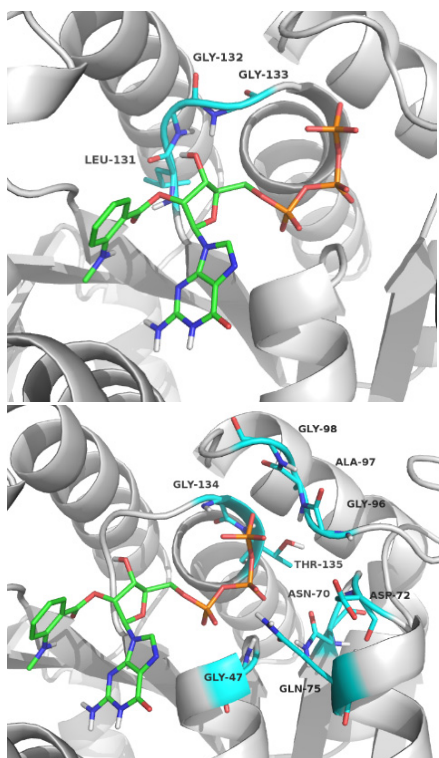
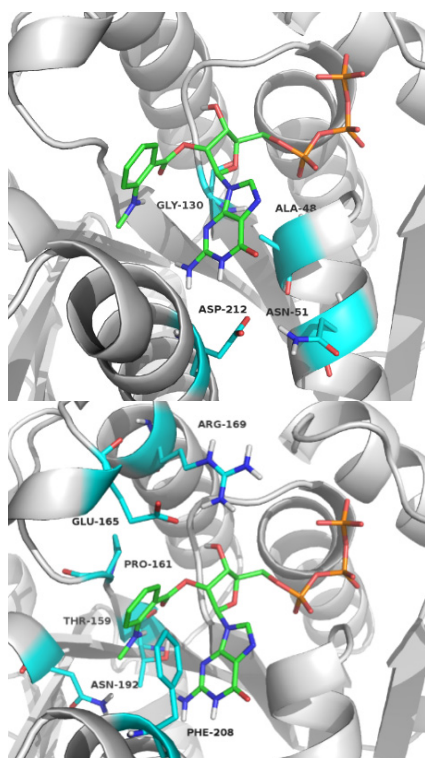


Figure S5, related to Figure 4. MD simulations of FtsZ-mant-GTP complexes. **A.** RMSD (Å) evolution along the simulation of the FtsZ-2'-mant-GTP complex. The protein RMSD is depicted in blue and the ligand RMSD is in red. **B.** Corresponding RMSD (Å) values along the simulation of the FtsZ-3'-mant-GTP complex. **C.** 2'-mant-GTP, 3'-mant-GTP, and GTP (atom-colored sticks) bound into the nucleotide binding site of Mj-FtsZ (grey surface representation; from the models in Figure 4). **D.** The different conformations of 2'-mant-GTP, 3'-mant-GTP, and GTP bound to Mj-FtsZ, extracted from the MD models in Figure 4; note that the ligands are colored and presented here in a view similar to the free ligands in Figure S1D-G. **E.** 2'-mant-GTP in the 2'endo conformation bound to a Mj-FtsZ monomer (green protein, cyan ligand; from Figure 4B) aligned to the model complex of Bs-FtsZ with the synthetic inhibitor UCM44 (grey protein and ligand, see main text).

A**B**

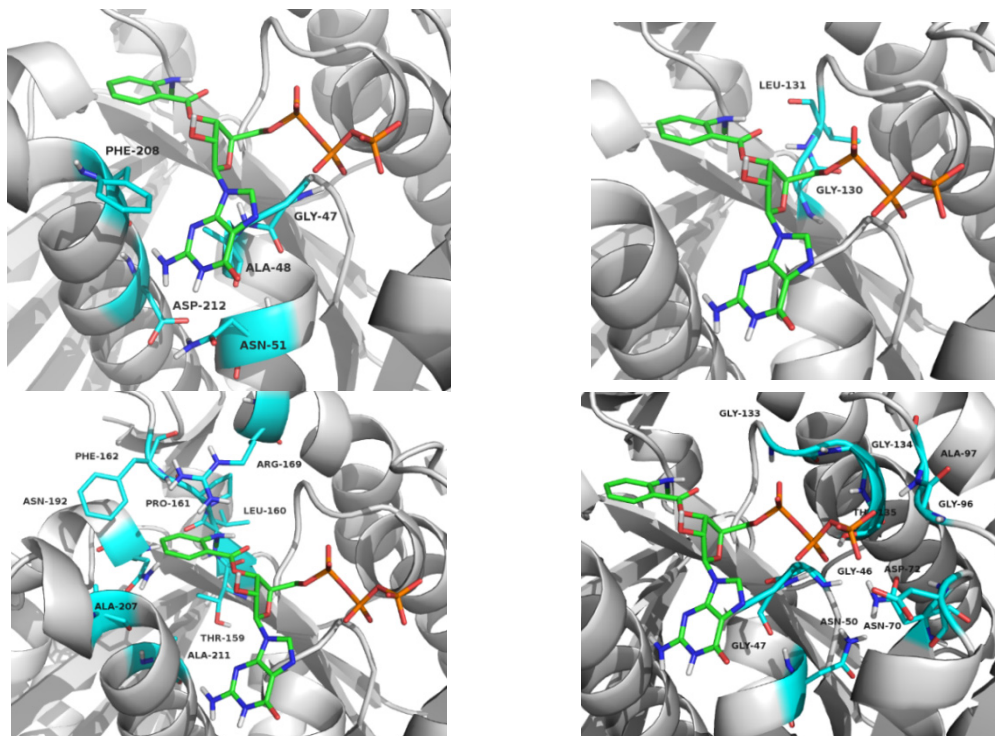
C

Figure S6, related to Figure 4. A. Aminoacid residues of Mj-FtsZ interacting with 2'-mant-GTP in 2'endo conformation during MD simulations. The main residues contacting the base, phosphates and the *mant* tag are highlighted in blue. **B.** Residues interacting with 2'-mant-GTP in 3'endo conformation. **C.** Residues of Mj-FtsZ interacting with 3'-mant-GTP in 2'endo conformation.

Movies 1, 2 and 3, related to Figure 4, show the molecular dynamics of FtsZ in complex with 2O'-mant-GTP, 3O'-mant-GTP and GTP. They can be found with this supplemental information online