SERBP1 is a component of the Liver Receptor Homolog-1 transcriptional complex

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Supplemental Table 1: QPCR Primer Sequences. Sequences of the forward and reverse QPCR primers were used in this study.

Supplemental Figure 1: Overexpression of both LRH1 isoforms using HaloTag. HEK293T cells were transfected with HaloLRH1. After transfection ( 24 hrs ), cells were harvested and a subcellular enrichment strategy was used to separate the nuclear and insoluble fractions. Cells were then labeled with TMRDirect Halo Ligand following Promega's Technical Manual protocol Part \# TM348. Proteins were separated by SDS-PAGE and the gel was scanned on a fluorescent scanner (Typhoon, GE Healthcare Biosciences).

Supplemental Figure 2: HaloLRH1 localizes to the nucleus. HEK293T cells were plated on 1well chamber slides ( $3 \times 10^{5}$ cells/chamber) (BioExpress). The following day, cells were transfected with both HaloLRH1 isoforms and empty vector. After 24hrs, cells were labeled with HaloTag TMRDirect Ligand (1:200) following Promega's protocol Part\# TM260. Cells were then fixed in 4\% paraformaldehyde per manufacture's protocol and processed for immunocytochemistry. Images were analyzed by confocal microscopy and captured on an Olympus Fluoview 1000 with SIM scanner.

Supplemental Figure 3: Promoter:Reporter assay and QPCR in HEK293T cells confirm all plasmids are functional. $A$ and $B$ ) Transient transfections were performed and luciferase levels were measured. Data was normalized as fold change over vector. Statistically significant differences are depicted by asterisks ( $\mathrm{p}<0.05$ ) as analyzed by one-way ANOVA. C and D) HEK293T cells were transfected with AviLRH1 for 48hrs. RNA was isolated, cDNA was prepared from $2 \mu$ g total RNA, and gene expression was analyzed by QPCR using GAPDH as the housekeeping gene. Data was normalized as fold change over vector. Statistically significant differences are depicted by asterisks ( $p<0.05$ ) as analyzed by one-way ANOVA.

Supplemental Figure 4: Detection of endogenous LRH1 isoform 1-specific peptide in the nuclear fraction by mass spectrometry-based proteomics analysis. LRH1 pull down samples were prepared as described in Figure 3 and the nuclear fraction was analyzed by LC MS/MS. The delta ppm between the theoretical and observed monoisotopic mass was determined to be <1ppm.

Supplemental Figure 5: Detection of endogenous LRH1 isoform 2-specific peptide in the insoluble fraction by mass spectrometry-based proteomics analysis. LRH1 pull down samples were prepared as described in Figure 3 and the insoluble fraction was analyzed by LC MS/MS. The delta ppm between the theoretical and observed monoisotopic mass was determined to be <1ppm.

Supplemental Figure 6: LC MS/MS analysis of endogenous LRH1 immunoprecipitation. Summary of the $b$ and $y$ ions from the MS2 data of the LRH1 v1-specific peptide detected in the nuclear fraction (A) and the LRH1 v2-specific peptide identified in the insoluble fraction (B).

Supplemental Figure 7: LRH1 response element in the promoter region of SERBP1. The Genomatix genome analyzer v2.60912 (www.genomatix.de) was used for identification and extraction of the promoter sequence of SERBP1. Genomatix utilizes a large library of matrix descriptions for transcription factor binding sites. This software was used to search for LRH1 response elements in the promoter sequence, which resulted in identification of two transcription factor binding sites (V\$SF1.01,V\$FTF.01).

Supplemental Table 2: List of identified LRH1-specific proteins from overexpression studies.

| Gene | Forward | Reverse |
| :---: | :---: | :---: |
| hGAPDH | 5'-GAAATCCCATCACCATCTTCCAGG-3' | 5'-GAGCCCCAGCCTTCTCCATG-3' |
| hLRH1 | 5'-CTGATACTGGAACTTTTGAA-3' | $5^{\prime}$ 'CTTCATTTGGTCATCAACCTT-3' |
| hSHP | 5'-AAAGGGACCATCCTCTTCAAC-3' | 5'-CTGGTCGGAATGGACTTGAG-3' |
| hSERBP1 | 5'-GTGACTGAGGAAACACCTGAA-3' | 5'-AGCCTTCCACTCATCCAAAG-3' |
| hilf3 | 5'-CCAGAGGACGACAGTAAAGAAG-3' | 5'-GTTGTGGGCTTCTCCTTACA-3' |

Supplemental Figure 1: Overexpression of both LRH1 isoforms using HaloTag


Supplemental Figure 2: HaloLRH1 localizes to the nucleus


Supplemental Figure 3: Promoter:Reporter assay and QPCR in HEK293T cells confirm all plasmids are functional
A

B

C
LRH1

D
SHP


Supplemental Figure 4: Detection of endogenous LRH1 isoform 1-specific peptide in the nuclear fraction by mass spectrometry-based proteomics analysis.


Supplemental Figure 5: Detection of endogenous LRH1 isoform 2-specific peptide in the insoluble fraction by mass spectrometry-based proteomics analysis.


Supplemental Figure 6: LC MS/MS analysis of endogenous LRH1 immunoprecipitation

A
Nuclear Fraction
B
Insoluble Fraction

LRH1 Variant 1 VETEALGLAR $\mathrm{MH}^{+2}$ (mono) 529.7957

Identified y ions Full MS2 529.80

| dentified y ions Full MS2 529.80 |  |  |  |  |
| :---: | :--- | :--- | :--- | :---: |
| $\mathbf{b}$ |  |  |  | $\mathbf{y}$ |
| - | 1 | V | 10 | - |
| 229.1 | 2 | E | 9 | 959.5 |
| 330.1 | 3 | T | 8 | 830.4 |
| 459.2 | 4 | E | 7 | 729.4 |
| 530.2 | 5 | A | 6 | 600.3 |
| 643.3 | 6 | L | 5 | 529.3 |
| 700.3 | 7 | G | 4 | 416.2 |
| 813.4 | 8 | L | 3 | 359.2 |
| 884.4 | 9 | A | 2 | 246.1 |
| - | 10 | R | 1 | 175.1 |

Identified b and y ions Full MS2 613.85

| b |  |  |  | y |
| :---: | :--- | :--- | :--- | :---: |
| - | 1 | H | 11 | - |
| - | 195.1 | 2 | G | 10 |
| 308.2 | 3 | L | 9 | 1089.6 |
| 409.2 | 4 | T | 8 | 9192.6 |
| 506.3 | 5 | P | 7 | 818.5 |
| 619.4 | 6 | I | 6 | 721.4 |
| 718.4 | 7 | V | 5 | 608.3 |
| 805.5 | 8 | S | 4 | 509.3 |
| 933.5 | 9 | Q | 3 | 422.2 |
| 1080.6 | 10 | F | 2 | 294.2 |
| - | 11 | K | 1 | 147.1 |

Supplemental Figure 7: LRH1 response elements in the promoter of SERBP1
>Promoter A| sym=SERBP1|taxid=9606|spec=Homo
sapiens|chr=1|ctg=NC_000001|str=()|start=67895883|end=67896623|len=741|comm=Promoter Region AGGCTGACGGAGGGCCACTCCAGCAACTGAGCAGCTCGGACTTGGCACAGGCAGGGTCCAGCCTACCCGCGGCCGGCGCG GCGCCTCAGCAACACCCCCTCCCCCGGAAGTTTGTGCGTCTCAGATGTAAGGTCTGGGAGTCACCTTGACCCCCCTGGAG CGGTGGGGAAAGTTAGCCCTGAGCCCGAGGGGGCGTGAGTGAAACCAAAGAGCACTTTCGCGAGTCAGTTACGCCGTCAC GGT1TाTGGTTCAAGGTAGCTGAAGACCTAGAGAGTTAAGGGTGGCCAGAGCAGGTGGCAGGACTCCGCCCACCACC ACGCTTCCCTTCCTTCAGCCCGCACGCTTCACTCCCCTCATGGGGTCTCGCCTCTCAGGCGCGCGCGAAGAGTTGTCGCG CGGTGCGTCCTGGGAATTGTAGTCCCGACGCGGAGAGTCGCCTCAGGAGAAATGACTCTGGCCTACATACCCCACAGTGC CTTGCGGCGCAGGCCGCTCCCGGATGTGTGCCTGGCGCCGGAAGAGAAGACGGCCCCCCTCTCTCGGCCCGGCCATCTTG TGGGAAGAGCTGAAGCAGGCGCTCTTGGCTCGGCGCGGCCCGCTGCAATCCGTGGAGGAACGCGCCGCCGAGCCACCATC ATGCCTGGGCACTTACAGGAAGGCTTCGGCTGCGTGGTCACCAACCGATTCGACCAGTTATTTGACGACGAATCGGACCC CTTCGAGGTGCTGAAGGCAGC

V\$SF1F binding site in Promoter A
V\$FTF. 01 Position:250-264 strand(+)
1 gtttCAAGgtagctg
V\$SF1.01 Position:138-152 strand(-)
gggtCAAGgtgactc



Supplemental Table 2: List of identified LRH1-specific proteins from overexpression studies

| Number | Identified Proteins | Accession Number | Molecular Weight |
| :---: | :---: | :---: | :---: |
| 1 | Ubiquitin-60S ribosomal protein $\mathrm{L40}$ OS=Homo sapiens GN=UBA52 PE=1 SV=2 | RL40_HUMAN ( +3 ) | 15 kDa |
| 2 | Chromobox prote in homolog $30 \mathrm{~S}=$ Homo sapiens $\mathrm{GN}=\mathrm{CBX} 3 \mathrm{PE}=1 \mathrm{SV}=4$ | CBX3 HUMAN | 21 kDa |
| 3 | FACT complex subunit SSRP1 OS=Homo sapiens GN=SSRP1 PE=1 SV=1 | SSRP1_HUMAN | 81 kDa |
| 4 | Regul ator of chromosome condensation OS=Homo sapiens GN=RCC1 PE=1 SV=1 | RCC1 HUMAN | 45 kDa |
| 5 | Plasminogen activator inhibitor 1 RNA-binding protein OS=Homo sapiens GN=SERBP1 PE=1 SV=2 | PAIRB_HUMAN | 45 kDa |
| 6 | Heat shock 70 kDa protein $1 \mathrm{~A} / 1 \mathrm{~B}$ OS=Homo sapiens GN=HSPA1A PE=1 SV $=5$ | HSP71 HUMAN | 70 kDa |
| 7 | Interleukin enhancer-binding factor 20S=Homo sapiens GN=\|LF2 PE=1 SV=2 | ILF2_HUMAN | 43 kDa |
| 8 | GTP-binding nucle ar protein Ran OS=Homo sapiens GN=RAN PE=1 $1 \mathrm{VV}=3$ | RAN_HUMAN | 24 kDa |
| 9 | Coiled-coil domain-containing protein 86 OS=Homo sapiens GN=CCDC $86 \mathrm{PE}=1 \mathrm{SV}=1$ | CCD86 HUMAN | 40 kDa |
| 10 | Heterogeneous nuclear ribonucleo protein A3 OS=Homo sapiens GN=HNRNPA3 PE=1 SV=2 | ROA3_HUMAN | 40 kDa |
| 11 | Probable ATP-dependent RNA helicase DDX17 OS=Homo sapiens GN=DDX17 PE=1 SV=2 | DDX17 HUMAN | 80 kDa |
| 12 | Probable ATP-de pendent RNA helicase DDX5 OS=Homo sapiens GN=DDX5 PE=1 SV=1 | DDX5 HUMAN | 69 kDa |
| 13 | Heat shock 70 kD a prote in $60 \mathrm{~S}=$ Homo sapiens GN=HSPA6 PE= $1 \mathrm{SV}=2$ | HSP76_HUMAN ( +1 ) | 71 kDa |
| 14 | Putative high mobility group protein B1-like $10 \mathrm{~S}=$ Homo sapiens GN=HMGB1P1 PE=5 SV=1 | HGB1A HUMAN ( +1 ) | 24 kDa |
| 15 | Heat shock cognate 71 kDa protein OS=Homo sapiens GN=HSPA8 PE=1 SV=1 | HSP7C_HUMAN | 71 kDa |
| 16 | ATP-dependent RNA helicase $A$ OS $=$ Homo sapiens GN=DHX9 PE= $1 \mathrm{SV}=4$ | DHX9 HUMAN | 141 kDa |
| 17 | Cluster of Core histone macro-H2A. 1 OS=Homo sapiens GN=H2AFY PE=1 SV=4 (H2AY_HUMAN) | H2AY HUMAN | 40 kDa |
| 18 | Interleukin enhancer-binding factor 30 S=Homo sapiens $\mathrm{GN}=1 \mathrm{LF} 3 \mathrm{PE}=1 \mathrm{SV}=3$ | ILF3_HUMAN | 95 kDa |
| 19 | ATP synthase subunit alpha, mitochondrial OS=Homo sapiens GN=ATP5A1 PE=1 SV=1 | ATPA_HUMAN | 60 kDa |
| 20 | High mobility group protein HMG-1/HMG-Y OS=Homo sapiens GN=HMGA1 PE=1 SV=3 | HMGA1_HUMAN | 12 kDa |
| 21 | Poly [ADP-ribose] polymerase 10S=Homo sapiens GN=P ARP1 PE=1 SV=4 | PARP1_HUMAN | 113 kDa |
| 22 | ATP ase inhibitor, mitochondrial OS=Homo sapiens GN=ATPIF1 PE=1 SV=1 | ATIF1_HUMAN | 12 kDa |
| 23 | Drebrin OS=Homo sapiens GN=DBN1 PE=1 SV=4 | DREB HUMAN | 71 kDa |
| 24 | Insulin-like growth factor 2 mRNA -binding protein $1 \mathrm{OS}=$ Homo sapiens GN=IGF2BP1 PE=1 SV=2 | IF2B1 HUMAN | 63 kDa |
| 25 | Cluster of Zinc finger protein 280C OS=Homo sapiens GN=ZNF280C PE=1 SV=1 (Z280C_HUMAN) | Z280C_HUMAN | 83 kDa |
| 26 | Heme oxyge nase 10S=Homo sapiens GN=HMOX1 PE=1 SV=1 | HMOX1_HUMAN | 33 kDa |
| 27 | Dnad homolog subfamily C member 9 OS=Homo sapiens GN=DNAJC9 PE=1 SV=1 | DNJC9 HUMAN | 30 kDa |
| 28 | DNA mismatch repair prote in Msh6 OS=Homo sapiens GN=MSH6 PE=1 SV=2 | MSH6_HUMAN | 153 kDa |
| 29 | Histone acetyltransferase KAT7 OS=Homo sapiens GN=KAT7 PE=1 SV=1 | KAT7_HUMAN | 71 kDa |
| 30 | Pyruvate dehydroge nase E1 component subunit beta, mitochondrial OS=Homo sapiens GN=PDHB PE=1 SV=3 | ODPB_HUMAN | 39 kDa |
| 31 | Sarcoplasmic/endoplasmic reticulum calcium ATPase 20S=Homo sapiens GN=ATP2A2 PE=1 SV=1 | AT2A2 HUMAN | 115 kDa |
| 32 | RNA-binding protein Musashi homolog 2 OS=Homo sapiens GN=MSI2 PE=1 SV=1 | MSI2H_HUMAN | 35 kDa |
| 33 | Telomere-associated protein RIF1 OS=Homo sapiens GN=RIF1 PE=1 SV=2 | RIF1_HUMAN | 274 kDa |
| 34 | Nuclear pore complex protein Nup85 OS=Homo sapiens GN=NUP85 PE=1 SV=1 | NUP85 HUMAN | 75 kDa |
| 35 | Alpha-globin transcription factor CP2 OS=Homo sapiens GN=TFCP2 PE=1 SV=2 | TFCP2_HUMAN | 57 kDa |
| 36 | Cell division control protein 42 homol og OS=Homo sapiens GN=CDC42 PE=1 SV=2 | CDC42_HUMAN | 21 kDa |
| 37 | Sister chromatid cohesion protein PDS5 homolog A OS=Homo sapiens GN=PDS5A PE=1 SV=1 | PDS5A HUMAN | 151 kDa |
| 38 | Transcription termination factor 2 OS=Homo sapiens GN=TTF2 PE=1 SV=2 | TF2 HUMAN | 130 kDa |
| 39 | Glycerol-3-phosphate dehydrogenase, mitochondrial OS=Homo sapiens GN=GPD2 PE=1 SV=3 | GPDM_HUMAN | 81 kDa |
| 40 | Importin subunit al pha-2 $\mathrm{OS}=\mathrm{Homo}$ sapiens GN=KPNA 2 PE=1 SV=1 | IMA2_HUMAN | 58 kDa |
| 41 | Catenin beta-10S=Homo sapiens GN=CTNNB1 PE=1 SV=1 | CTNB1 HUMAN | 85 kDa |
| 42 | CUGBP Elav-like family member 1 OS=Homo sapiens GN=CELF1 PE=1 SV=2 | CELF1 HUMAN | 52 kDa |
| 43 | Multiple myeloma tumor-associate d prote in 2 OS=Homo sapiens GN=MMTAG2 PE=1 SV=1 | MMTA2_HUMAN | 29 kDa |
| 44 | 40 r ribosomal protein $\mathrm{S} 16 \mathrm{OS}=$ Homo sapiens GN=RPS16 PE=1 SV=2 | RS16 HUMAN | 16 kDa |
| 45 | Histone acetyltran sferase KAT6A OS=Homo sapiens GN=KAT6A PE=1 SV=2 | KAT6A_HUMAN | 225 kDa |
| 46 | Bromodomain adjacent to zinc finger domain protein 2A OS=Homo sapiens GN=BAZ2A PE=1 SV=4 | BAZ2A_HUMAN | 211 kDa |
| 47 | MOSC domain-containing protein 1, mitochondrial OS=Homo sapiens GN=MARC1 PE=1 SV=1 | MOSC1_HUMAN | 38 kDa |
| 48 | 60 S ribosomal protein L36a OS=Homo sapiens GN=RPL36A PE=1 SV=2 | RL36A_HUMAN (+1) | 12 kDa |

