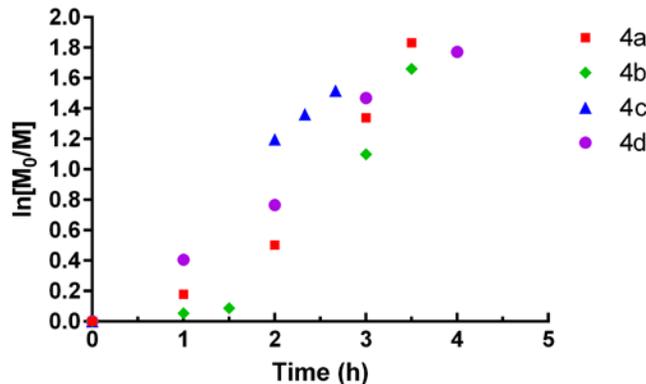


Supplementary information

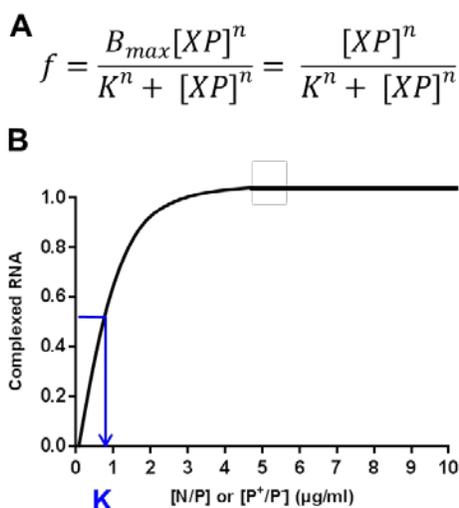
Phosphonium Polymethacrylates for Short Interfering RNA Delivery: Effect of Polymer and RNA Structural Parameters on Polyplex Assembly and Gene Knockdown

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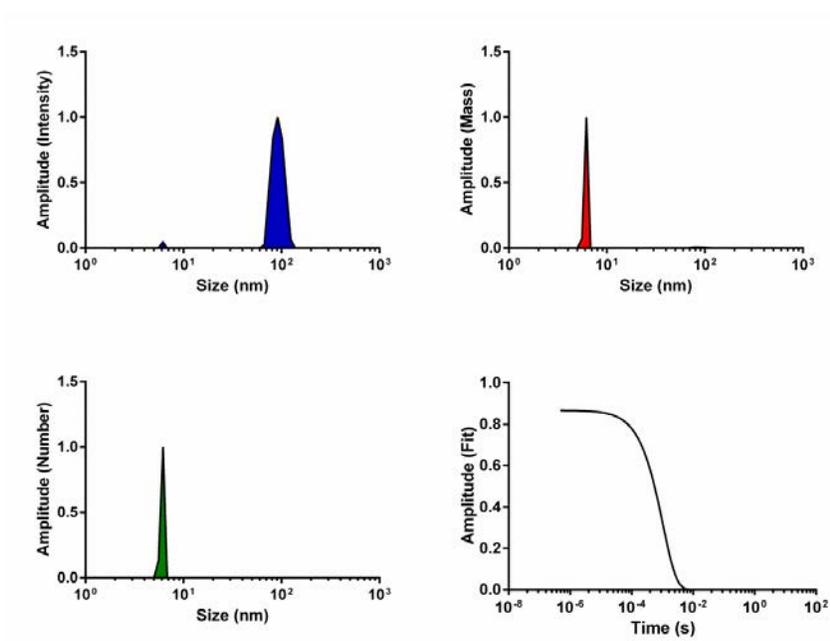
S1: Kinetic plots for the RAFT polymerization of methacrylate monomers yielding polymers **4a** (■), **4b** (◆), **4c** (▲) and **4d** (●). Reactions were performed with CTP, V-501 and monomers (**3a-d**) using a molar ratio of $[M]_0:[CTA]_0:[I]_0=100:1:0.5$ in $D_2O:EtOH$ (3:1 vol/vol).



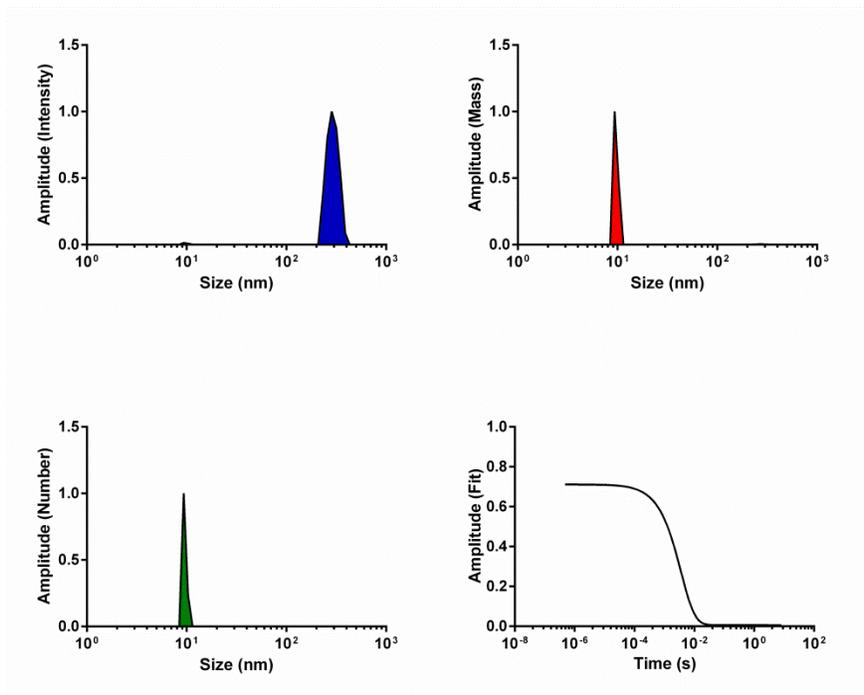
S2: A) Modified Hill's equation. B_{max} = Maximum specific binding ($B_{max}=1$), XP = polymer concentration represented as N^+/P ratio (ammonium polymer) or P^+P^- (phosphonium polymer) ratio, K = Binding constant to achieve a half-maximum binding at 30 minutes, n = Hill coefficient; B) Illustration of Hill's equation model and binding constant K .

S3: Dynamic light scattering and zeta potential measurements for RNA polyplexes at N^+/P^- or P^+/P^- ratio 20 (as analyzed by mass).

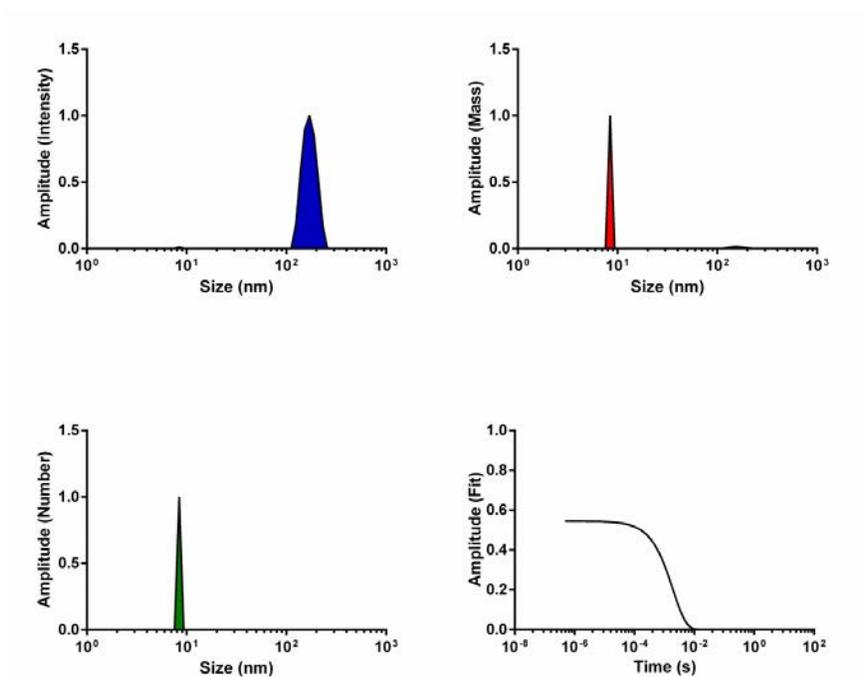
Polyplexes	R_h (nm)	Subpopulation, R_h (nm)	Subpopulation (%)	Zeta- potential (mV)
siRNA	1.02 ± 0.07	–	–	-21.9 ± 5.3
4a	8.8 ± 2.2	95.8 ± 53.5	2.9 ± 1.9	30.1 ± 6.9
4b	13.7 ± 5.6	223.2 ± 138.6	1.4 ± 1.5	24 ± 2.8
4c	14.8 ± 6.9	174.5 ± 85.3	6.3 ± 6.1	30.5 ± 3.7
4d	4 ± 1	–	–	31.3 ± 3.4



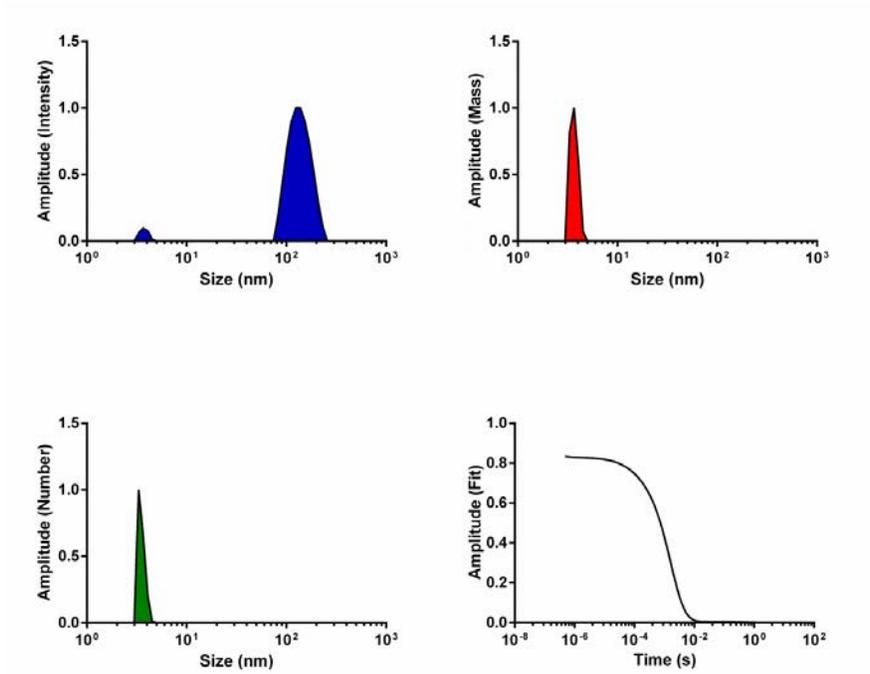
S4: Dynamic light scattering of RNA polyplexes formed with polymer **4a** at N^+/P^- ratio 20. Hydrodynamic radius (R_h , nm) was measured by intensity (A), mass (B) or number (C). (D) Illustration of correlation function. Experiment were performed in triplicate, with three independent measurements (n=3).



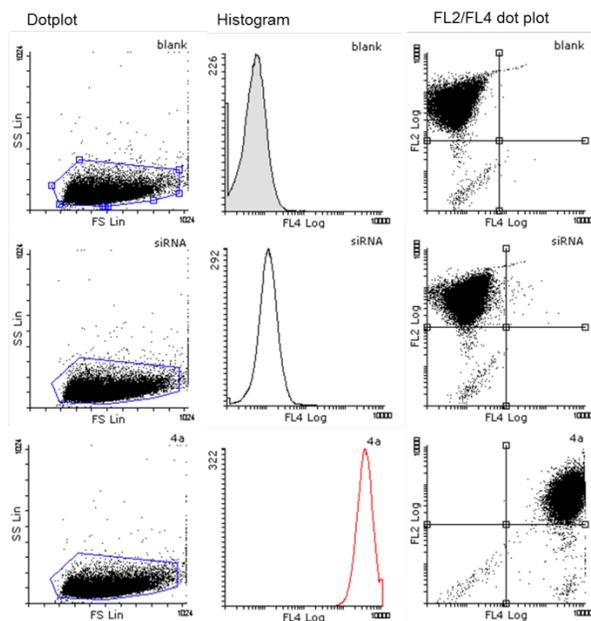
S5: Dynamic light scattering of RNA polyplexes formed with polymer **4b** at N^+/P^- ratio 20. Hydrodynamic radius (R_h , nm) was measured by intensity (A), mass (B) or number (C). (D) Illustration of correlation function. Experiment were performed in triplicate, with three independent measurements ($n=3$).

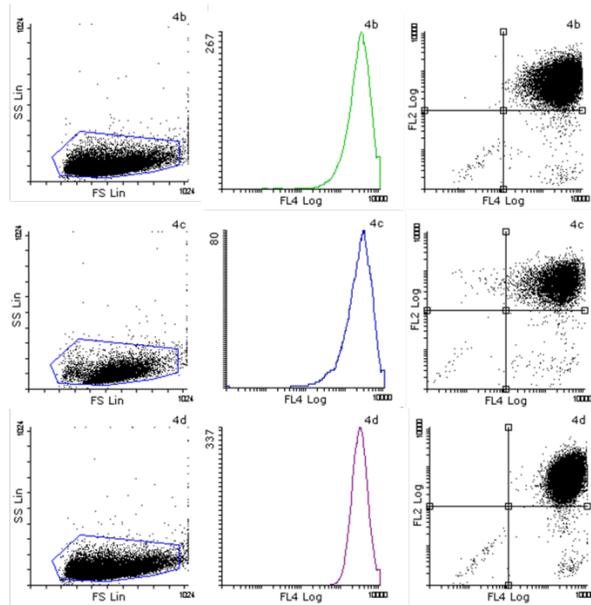


S6: Dynamic light scattering of RNA polyplexes formed with polymer **4c** at N^+/P^- ratio 20. Hydrodynamic radius (R_h , nm) was measured by intensity (A), mass (B) or number (C). (D) Illustration of correlation function. Experiment were performed in triplicate, with three independent measurements ($n=3$).

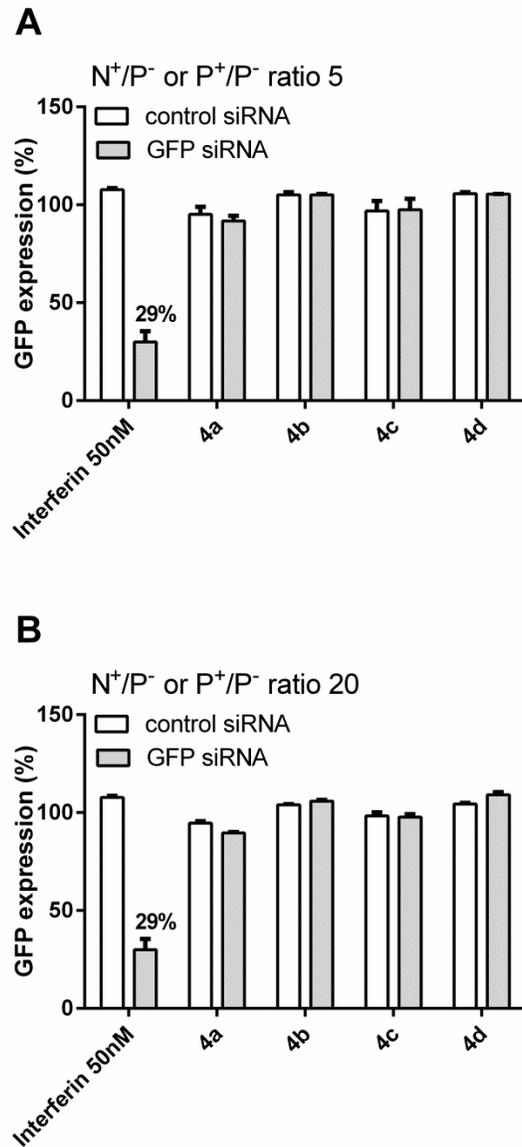


S7: Dynamic light scattering of RNA polyplexes formed with polymer **4d** at N^+/P^- ratio 20. Hydrodynamic radius (R_h , nm) was measured by intensity (A), mass (B) or number (C). (D) Illustration of correlation function. Experiment were performed in triplicate, with three independent measurements ($n=3$).

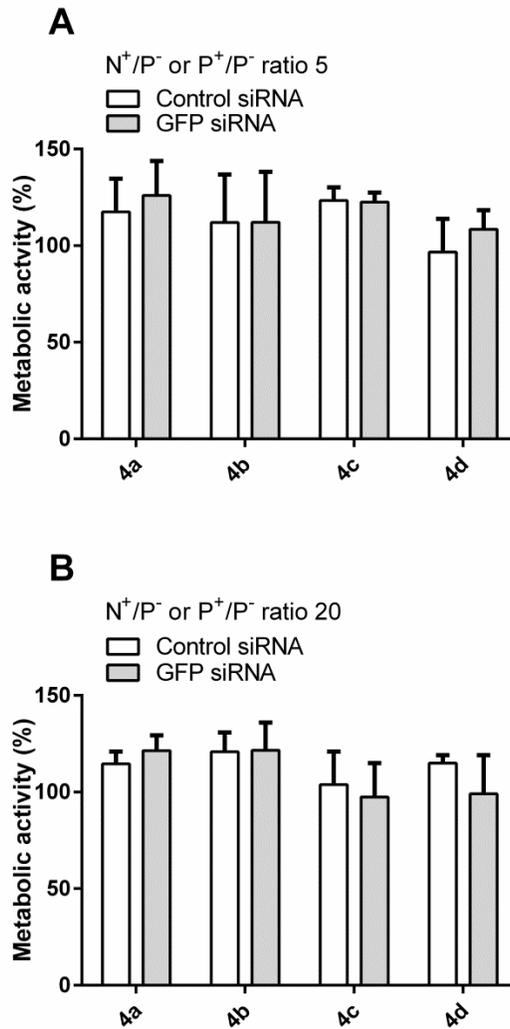




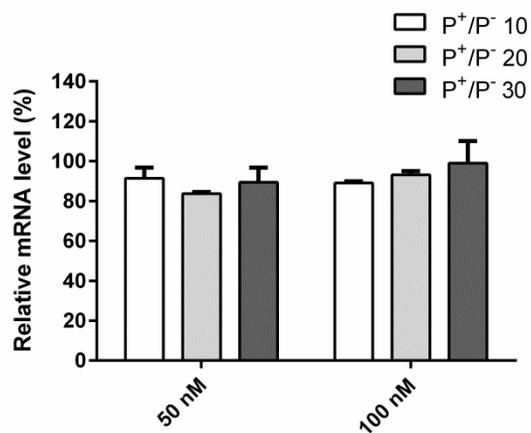
S8: 3T3 cellular uptake using flow cytometry. The main population is plotted in a side scatter/forward scatter dot plot and gated (R-1) to exclude cell debris. The gate population is plotted on a histogram (FL-4 = AF-647) and FL-2/FL-4 dot plot showing an increase of FL-4 fluorescence in samples containing siRNA-polyplexes.



S9: No GFP knockdown is achieved in 3T3 cells when employing polymers (**4a-d**) at different N⁺/P⁻ or P⁺/P⁻ ratio using 187 nM siRNA after 48 hs incubation as analyzed by flow cytometry. A) Polymers at N⁺/P⁻ or P⁺/P⁻ ratio = 5. B) Polymers at N⁺/P⁻ or P⁺/P⁻ ratio = 20. In contrast, commercially available Interferin successfully transfects siRNA at concentration of 50 nM. Data is represented as GFP expression (%), mean ± SEM (n=2).



S10: 3T3 cellular viability after polyplex exposure at different N^+/P^- or P^+/P^- ratio using 187 nM siRNA. No effect on cellular viability was observed when polyplexes were exposed to the cells and are incubated for 48 hs. (A) N^+/P^- or P^+/P^- ratio = 5 (B) N^+/P^- or P^+/P^- ratio = 20. Data is represented as metabolic activity (%), mean \pm SEM (n=2).



S11: Knockdown studies with siRNA targeting Survivin. HeLa cells were transfected with polymer **4d** using a P⁺/P⁻ ratio of 10, 20, 30, with 50 or 100 nM siRNA. Survivin and GAPDH mRNA levels were measured by qRT-PCR 24 hs after transfection. Survivin mRNA levels were normalized to GAPDH mRNA levels. Data is represented as relative mRNA level (%), mean ± SEM (n=2).