

Supplementary information for: Osmolyte Induced Changes in Peptide Conformational Ensemble Correlate with Slower Amyloid Aggregation: a Coarse-Grained Simulation Study

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S1. Simulations

Table S1 – All coarse grained simulations conducted in this study and corresponding composition

	peptide ^a	Water ^a	Sorbitol ^a	Time (μs)	Runs
Water	1	2252	0	2	5
1.5 m sorbitol	1	2311	250	2	5
1.1 m sorbitol	1	2461	200	2	5
0.8 m sorbitol	1	2611	150	2	5
0.65 m sorbitol	1	2686	125	2	5
0.4 m sorbitol	1	2812	83	2	5
Water large	216	486432	0	1	4
Sorbitol large	216	499176	54000	1	3
0.1 m sorbitol	0	36000	256	1	1
0.2 m sorbitol	0	28800	416	1	1
0.3 m sorbitol	0	28800	624	1	1
0.4 m sorbitol	0	28800	832	1	1
0.5 m sorbitol	0	17776	640	1	1
0.6 m sorbitol	0	28800	1248	1	1
0.7 m sorbitol	0	28800	1448	1	1
0.8 m sorbitol	0	28800	1656	1	1
0.9 m sorbitol	0	28800	1864	1	1
1.0 m sorbitol	0	14400	1040	1	1
1.1 m sorbitol	0	28800	2280	1	1
1.2 m sorbitol	0	28800	2488	1	1
1.3 m sorbitol	0	28800	2696	1	1
1.4 m sorbitol	0	28800	2904	1	1
1.5 m sorbitol	0	13328	1440	1	1

^a number of particles in simulations

Simulation details

All simulations were performed with the GROMACS package.¹⁻⁴ The Replica Exchange Molecular Dynamics (REMD) simulation⁵ of the peptide in pure water was conducted using the AMBER ff99sb-ildn-NMR force field⁶⁻⁹ and the TIP4Pew water model.¹⁰ All bond lengths to hydrogen atoms were kept constant with the LINCS algorithm.¹¹ The simulation consisted of 44 replicas spanning the temperature range 277–465K, where temperatures were assigned so as to achieve an acceptance rate of 0.2.¹² The dynamics were carried with periodic boundary conditions using a time-step of 2 fs, and replica exchange attempts every 1 ps. Temperatures were held constant using the Nosé-Hoover thermostat with a 1 ps coupling constant.^{13,14} Electrostatic calculations were performed using the particle-mesh Ewald (PME) method with 1 Å grid spacing.^{15,16} Van der Waals interactions were truncated smoothly with a switching distance of 10 Å and a cutoff distance of 12 Å, and by also accounting for long-range dispersion corrections. The REMD simulation was performed in the NVT ensemble using the volume calibrated by an isobaric-isothermic ensemble simulations at ambient temperature, T = 298 K, and pressure of P = 1 atm. Pressure in this NPT production was kept constant using the Parrinello-Rahman barostat with a 1 ps coupling constant and compressibility of $4.5 \cdot 10^{-5}$ bar⁻¹.^{17,18} Of the 500 ns collected in the REMD production (per replica), only the last 300 ns were used for analysis.

For the molecular dynamics simulations of aqueous sorbitol, we used the TIP4Pew water model and the GLYCAM06 force-field.¹⁹ The partial charges of sorbitol (Figure S1) were reassigned following the standard procedure²⁰⁻²² by using the Gaussian 03 package²³, R.E.D tools,^{24,25} and Resp program.²⁶ The derived partial charges are listed in Table S1. The sorbitol simulations were run in the isobaric-isothermic ensemble, as mentioned above. All other simulation details are as detailed above. For the charge calculations we used snapshots of the latter half of a 200 ns simulation.

Coarse-grained simulations were run using MARTINI forcefield v2.1^{27,28}, with a timestep of 0.04 ps for binary solutions and of 0.01 ps for peptide simulation. The relatively short timestep for peptide simulations is used to allow stable dihedral angles calculations²⁹. Simulation settings were set according to recommended MARTINI parameters. Non-bonded parameters were used with a cutoff of 1.2 nm and a shifted function from 0.9 to 1.2 nm. Temperature was controlled using the V-rescale algorithm, with a time constant of 1 ps. Pressure was controlled using a Parinello-Rachman barostat with a time constant of 12 ps, compressibility of $3 \cdot 10^{-4}$ bar⁻¹, and a reference pressure of 1 bar for all simulation boxes.

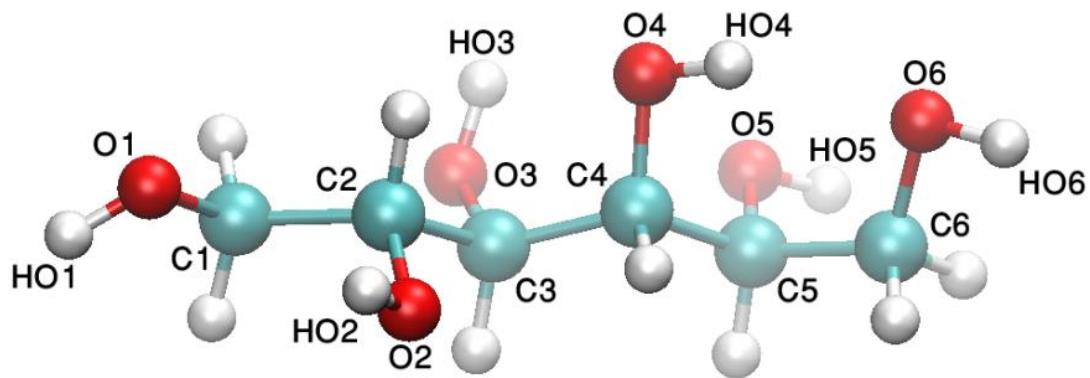


Fig. S1. A sorbitol molecule in the CPK representation.

Table S2. The derived partial charges of sorbitol in the GLYCAM force field. Atom names are as in Figure S1.

Atoms	Partial charge
C1, C6	0.273
O1, O6	-0.696
HO1,HO6	0.423
C2,C3,C4,C5	0.288
O2,O3,O4,O5	-0.714
HO2,HO3,HO4,HO5	0.426
All aliphatic hydrogens	0.0

S2. REMD simulations analysis

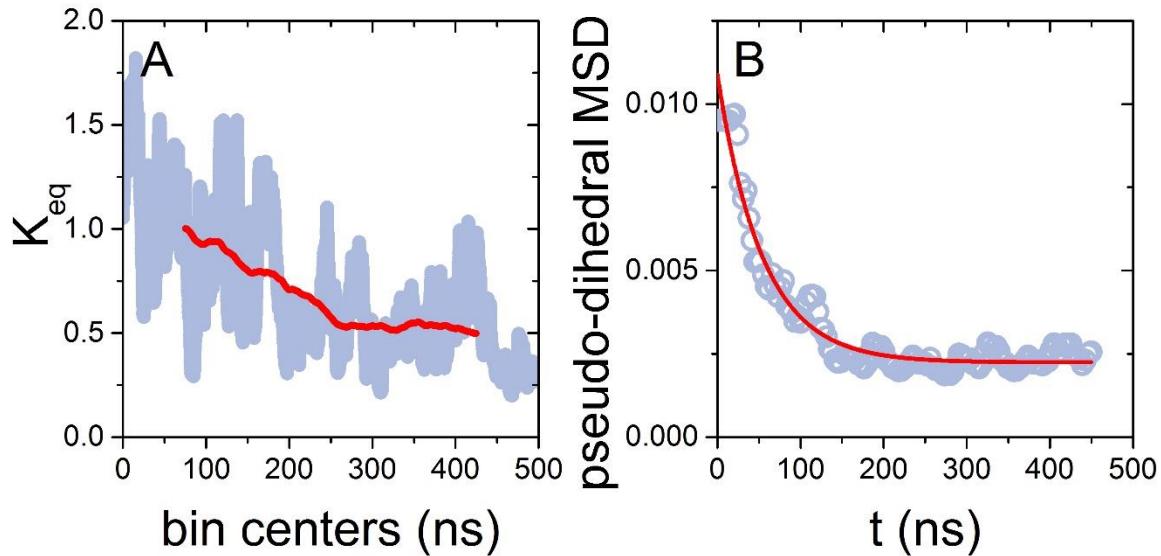


Fig. S2. Convergence of all-atom REMD simulations. (A) Convergence of folding equilibrium constant K_{eq} from the replica at $T = 298K$. K_{eq} was calculated using the RMSD with respect to a representative folded conformation at each frame, and the cutoff for the folded ensemble was set to $\text{RMSD} \leq 2.0 \text{ \AA}$. The plot shows the running average of K_{eq} , where each point represents the center of an averaging window of 150 ns, spanning 75 ns to the left and 75 ns to the right of each point (shown in red). This running average is overlaid on the averaged data of non-overlapping, 5 ns windows (grey). The average taken from the 175-325 ns window (centered at 250 ns) already shows the same K_{eq} as 425 ns (within statistical error), demonstrating the equilibrium constant is well converged. (B) Sum of mean-square-differences of all pseudo-dihedral angle distributions between a 50 ns running time window and the final selected dihedral distributions shown in Fig. S3. The red line is an exponential fit to the data in grey. Final pseudo-dihedral angles are taken from the average of the last 300 ns of this REMD replica where both peptide equilibrium (panel A) and dihedral angles (panel B) are well converged.

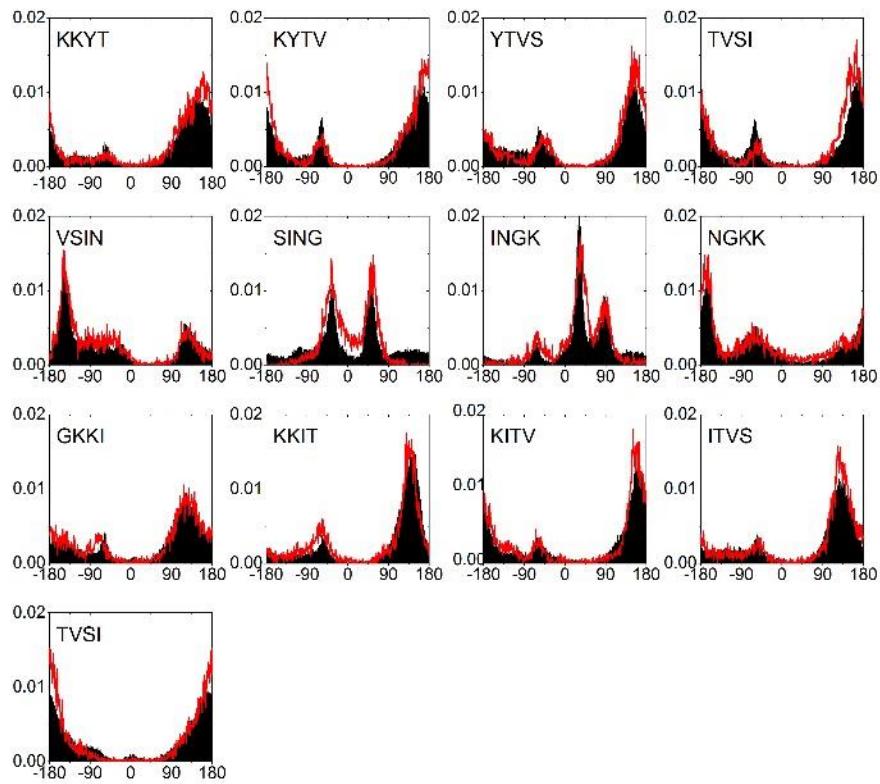


Fig. S3. Resulting pseudo-dihedral angle probability distribution in CG peptide calibrated simulations (red lines) overlaid on distributions from all-atom REMD (black bars) shows good agreement between both models.

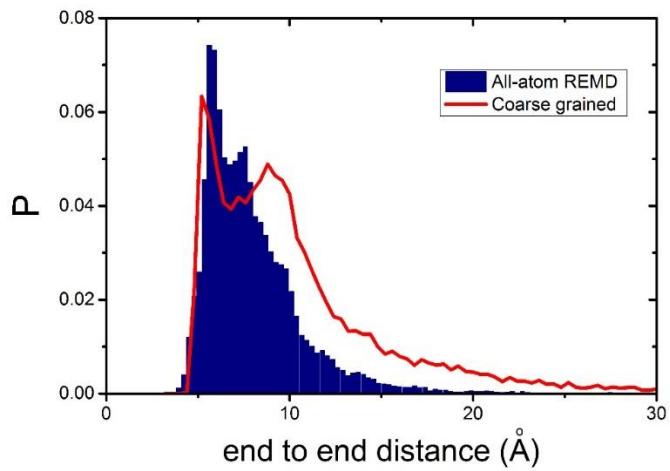


Fig. S4. Comparison of end-to-end distance between CG and all-atom model of met16.

S3. Osmotic pressure of sorbitol model

In order to evaluate how well the sorbitol model reproduces experimentally known solution properties, we calculated the osmotic pressure of the sorbitol model in binary solutions with MARTINI waters using two different methods. The first method uses the sorbitol-sorbitol radial distribution function, $g_{ss}(r)$ to calculate the second virial coefficient B , through the relation³⁰

$$B = -2\pi N_{av} \int_{r=0}^{\infty} (g_{ss}(r) - 1) r^2 dr,$$

with r the distance from the center sorbitol's center of mass, and N_{av} Avogadro's constant. Once determined, B can be used to calculate the osmotic pressure Π at low sorbitol concentrations using sorbitol molality m_s , and the mass to volume density of the solution, d :

$$\Pi \equiv m_s + Bdm_s^2.$$

A second method to obtain the osmotic pressure is through the use of the Kirkwood-Buff theory of solutions, as we describe in Ref. 31. Briefly, the derivative of the osmotic pressure with respect to water density is related to the Kirkwood-Buff integrals \mathcal{G}_{sw}^∞ and \mathcal{G}_{ww}^∞ (the former integral, for example, is shown for 1 mol/kg Fig. 2D, and discussed in the main text) by:

$$\left(\frac{\partial \Pi}{\partial \rho_w} \right) = \frac{-m_w}{\rho_w (1 + \rho_w \mathcal{G}_{ww}^\infty - \rho_w \mathcal{G}_{sw}^\infty)}.$$

Here, ρ_w is the water-bead number density, and m_w is the pure water-bead molality. Integrating over a second order polynomial fit to the resulting values for the derivative with respect to ρ_w , and setting the integration constant so that $\Pi = 0$ in pure water, yields the simulated osmotic pressure. While this method gives the exact osmotic pressure, it is numerically noisier than the first estimate as two KBIs are involved rather than one.

Fig S5 shows the results from both methods, and compares them with experimental (red line) and ideal (van 't Hoff, black line) osmotic pressures. Both methods used to calculate the pressure from simulations carry errors with them. These are mainly associated with radial distributions integrals, which are notoriously difficult to converge. Despite this, a qualitatively good fit, showing a slight yet positive deviation from ideal osmotic behavior is apparent, in agreement with experiments. The deviation from experimental results increases, however, as sorbitol concentrations increase. At the concentration used in our peptide aggregation study (1 mol/kg) the osmotic pressure shows a close agreement with experiment. Important to the coarse-grained aspect of our simulations, all calculations performed here do not utilize any special or arbitrary fitting factors to account for the 4-to-1 mapping native to MARTINI. We conclude that despite

the known drawbacks of the coarse grained representation, our model manages to reproduce experimental solution thermodynamic parameters to good accuracy.

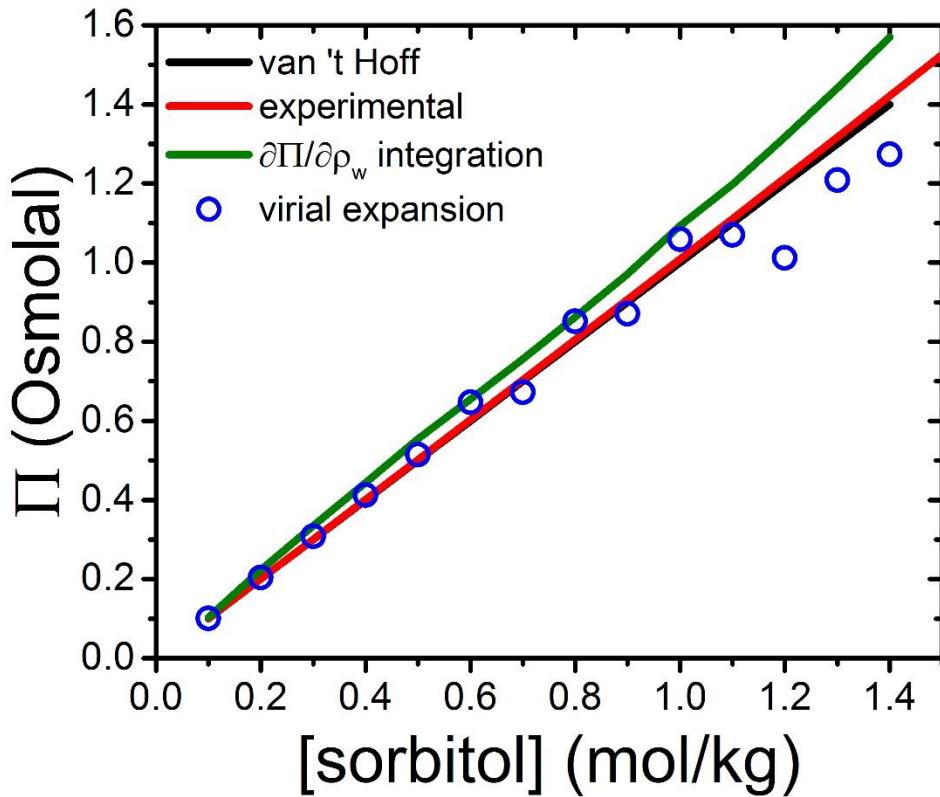


Fig. S5. Comparison between simulated and experimental osmotic pressures. Two methods of calculation for the osmotic pressure, discussed above, are shown as blue symbols and green line, and compared with experimental data shown as the red line. Small positive deviations from ideal osmotic pressure (in black) are observed for both simulated and experimental data at the relevant concentrations used in this study.

S4. Preferential interaction of sorbitol with the peptide

An additional method to obtain the effect of cosolute on protein folding free-energy is through the direct measurement of the number of sorbitol excluding waters, where

$$\Delta\Gamma = \left[N_W \left(1 - \frac{N_S / N_W}{n_S / n_W} \right) \right]_{folded} - \left[N_W \left(1 - \frac{N_S / N_W}{n_S / n_W} \right) \right]_{unfolded}$$

Here, N relates to the number of molecules in the vicinal area of the peptide, while n is the number of molecules in the bulk area, far from the peptide, and the subscripts S and W represent cosolute and water models respectively. A cutoff between the vicinal and bulk regions is determined by the region of $g(r)$ convergence (Fig. 2D). The number for $\Delta\Gamma$ is obtained by averaging the limiting value of the curve shown in Fig. S6 at the region where no more changes (apart from the undulations known to occur in coarse-grained liquids^{32,33}, typically 15-20 Å from the peptide surface). The difference $\Delta\Gamma$ can then be related to the m -value (as described in the main text) through the relation

$$m = \frac{RT}{13.9} \Delta\Gamma,$$

where 13.9 is the number (in moles) of water beads in 1kg of pure MARTINI water, R is the universal gas constant, and T is the temperature.

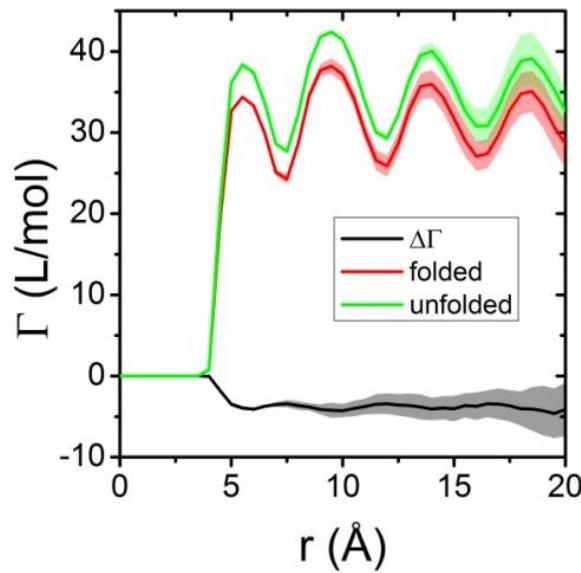


Fig S6. Change in preferential interaction of water with peptide shows difference between the folded and unfolded ensemble in the presence of 1 mol/kg sorbitol. The value of $\Delta\Gamma$ is proportional to the m -value discussed in the text. The overall free energy change upon folding in these conditions is $m = -0.9 \pm 0.2$ kJ/mol.

S5. SASA changes upon folding of CG peptide

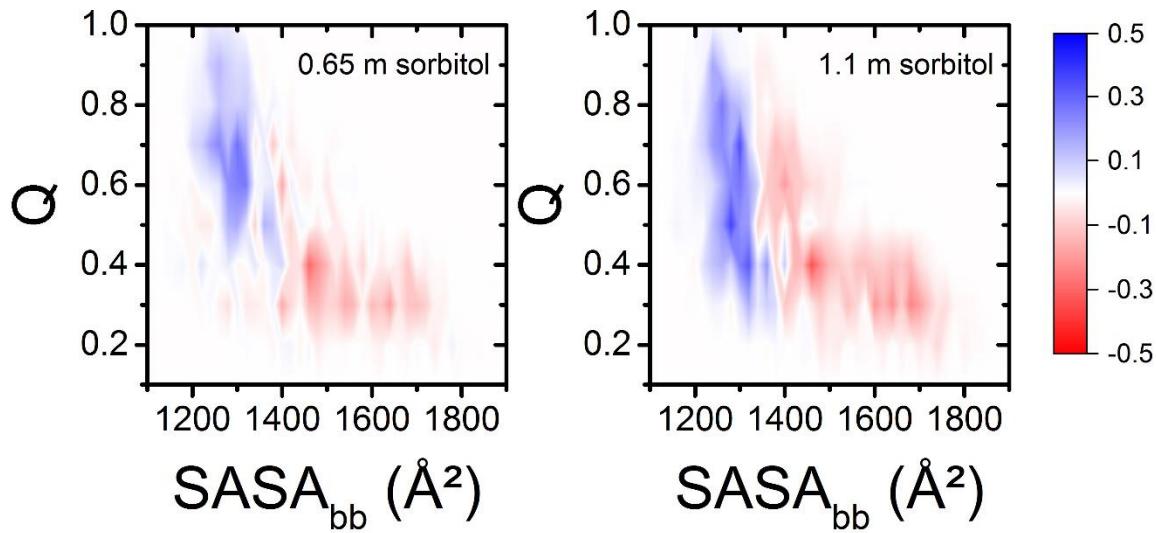


Fig S7. Changes in Q vs. backbone SASA distributions due to the presence of sorbitol. Values are calculated by subtracting the probability distributions in the presence of sorbitol from those in the absence of sorbitol. Red regions show a decrease in probability, while blue regions show an increase, and color intensity indicates the magnitude of distribution change (in percent). The figures clearly show that the presence of sorbitol differentiates high vs. low SASA conformations, but does not have a similarly clear cutoff for Q.

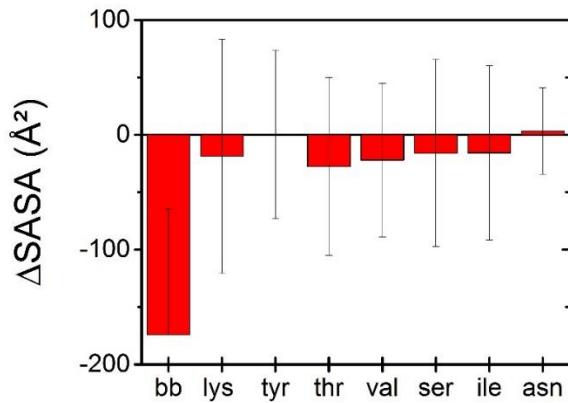


Fig S8. Changes in SASA upon folding pf CG peptide in water for backbone and sidechains of different amino acids. For each ensemble defined by the SASA cutoff specified in the text, an average SASA is obtained using the VMD measure algorithm. The probe size is set to 2.35, the size of the MARTINI water bead. The difference upon folding is shown. Error bars are the propagated standard deviations of the ensemble averages.

S6. Kinetic model for amyloid growth

The master equation used here to numerically fit the experimental result is based on a master equation originally developed by Knowles et al.³⁴ to model amyloid aggregation kinetics. We previously modified this scheme to include monomer detachment from formed fibrils. This addition was necessary to explain our experimental observables for met16 aggregation in the presence of various osmolytes and polymers, including sorbitol.³⁵ Here we add yet another term, the attachment of formed fibrils to each other. This is necessary because our simulated trajectories show that a major part of elongation results from attachment of formed fibrils to each other. The final form of the kinetic scheme is formulated as a discrete master equation for clusters of size i

$$\frac{dP_i}{dt} = k_{on} [P_1(P_{i-1} - P_i)] + k_{off} [P_{i+1} - P_i] + k_b \left[\sum_{j=i+2}^N P_j + P_{2i} - (i-3)P_i \right] + \\ + k_a \left[\sum_{k+j=i} P_j P_k - P_i \left(\sum_{j=2}^N P_j \right) - P_i^2 \right]$$

Here P_i are probabilities of monomers to exist in a cluster of size i , k_{on} and k_{off} are rates of monomer attachment and detachment to formed fibrils, and k_b and k_a are rates of fibril breakage and attachment to other fibrils, respectively. An additional term of the form $k_n P_1^2$ describes nucleation from two monomers, where the nucleus is of size $i=2$, and exists for monomers and dimers only. The rate of fibril-to-fibril attachment, k_a was necessary to fit our simulations results but was not necessary to fit the aggregation data of our experimental work. This is because the ThT fluorescence assay used as a probe in our experiments cannot discern events of fibril attachment.

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Appendix 1: Forcefield parameters for met16 peptide including REMD-calibrated dihedrals in GROMACS format

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; MARTINI 2.1 Coarse Grained topology file for "met16"
; Sequence:
; KKYTVSINGKKITVSI
; Secondary Structure:
; CCCCCCCCCCC

[ moleculetype ]
; Name          Exclusions
Protein        1

[ atoms ]
  1   P5      1   LYS    BB     1  0.0000 ; C
  2   C3      1   LYS    SC1    2  0.0000 ; C
  3   Qd      1   LYS    SC2    3  1.0000 ; C
  4   P5      2   LYS    BB     4  0.0000 ; C
  5   C3      2   LYS    SC1    5  0.0000 ; C
  6   Qd      2   LYS    SC2    6  1.0000 ; C
  7   P5      3   TYR    BB     7  0.0000 ; C
  8   SC4     3   TYR    SC1    8  0.0000 ; C
  9   SC4     3   TYR    SC2    9  0.0000 ; C
 10  SP1     3   TYR    SC3   10  0.0000 ; C
 11  P5      4   THR    BB    11  0.0000 ; C
 12  P1      4   THR    SC1   12  0.0000 ; C
 13  P5      5   VAL    BB    13  0.0000 ; C
 14  AC2     5   VAL    SC1   14  0.0000 ; C
 15  P5      6   SER    BB    15  0.0000 ; C
 16  P1      6   SER    SC1   16  0.0000 ; C
 17  P5      7   ILE    BB    17  0.0000 ; C
 18  AC1     7   ILE    SC1   18  0.0000 ; C
 19  P5      8   ASN    BB    19  0.0000 ; C
 20  P5      8   ASN    SC1   20  0.0000 ; C
 21  P5      9   GLY    BB    21  0.0000 ; C
 22  P5     10   LYS    BB    22  0.0000 ; C
 23  C3     10   LYS    SC1   23  0.0000 ; C
 24  Qd     10   LYS    SC2   24  1.0000 ; C
 25  P5     11   LYS    BB    25  0.0000 ; C
 26  C3     11   LYS    SC1   26  0.0000 ; C
 27  Qd     11   LYS    SC2   27  1.0000 ; C
 28  P5     12   ILE    BB    28  0.0000 ; C
 29  AC1     12   ILE    SC1   29  0.0000 ; C
 30  P5     13   THR    BB    30  0.0000 ; C
 31  P1     13   THR    SC1   31  0.0000 ; C
 32  P5     14   VAL    BB    32  0.0000 ; C
 33  AC2     14   VAL    SC1   33  0.0000 ; C
 34  P5     15   SER    BB    34  0.0000 ; C
 35  P1     15   SER    SC1   35  0.0000 ; C
 36  Qa     16   ILE    BB    36 -1.0000 ; C
 37  AC1     16   ILE    SC1   37  0.0000 ; C

[ bonds ]
; Backbone bonds
  1     4      1  0.35000  400 ; LYS(C)-LYS(C)
  4     7      1  0.35000  400 ; LYS(C)-TYR(C)
  7    11      1  0.35000  400 ; TYR(C)-THR(C)
 11   13      1  0.35000  400 ; THR(C)-VAL(C)
 13   15      1  0.35000  400 ; VAL(C)-SER(C)

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15    17      1   0.35000   400 ; SER(C)-ILE(C)
17    19      1   0.35000   400 ; ILE(C)-ASN(C)
19    21      1   0.35000   400 ; ASN(C)-GLY(C)
21    22      1   0.35000   400 ; GLY(C)-LYS(C)
22    25      1   0.35000   400 ; LYS(C)-LYS(C)
25    28      1   0.35000   400 ; LYS(C)-ILE(C)
28    30      1   0.35000   400 ; ILE(C)-THR(C)
30    32      1   0.35000   400 ; THR(C)-VAL(C)
32    34      1   0.35000   400 ; VAL(C)-SER(C)
34    36      1   0.35000   400 ; SER(C)-ILE(C)

; Sidechain bonds
1     2      1   0.33000   5000 ; LYS
2     3      1   0.28000   5000 ; LYS
4     5      1   0.33000   5000 ; LYS
5     6      1   0.28000   5000 ; LYS
7     8      1   0.32000   5000 ; TYR
15   16      1   0.25000   7500 ; SER
19   20      1   0.32000   5000 ; ASN
22   23      1   0.33000   5000 ; LYS
23   24      1   0.28000   5000 ; LYS
25   26      1   0.33000   5000 ; LYS
26   27      1   0.28000   5000 ; LYS
34   35      1   0.25000   7500 ; SER

[ constraints ]
8     9      1   0.27000 ; TYR
8     10     1   0.27000 ; TYR
9     10     1   0.27000 ; TYR
11   12      1   0.26000 ; THR
13   14      1   0.26500 ; VAL
17   18      1   0.31000 ; ILE
28   29      1   0.31000 ; ILE
30   31      1   0.26000 ; THR
32   33      1   0.26500 ; VAL
36   37      1   0.31000 ; ILE

[ angles ]
; Backbone angles
1     4      7      2   127    25 ; LYS(C)-LYS(C)-TYR(C)
4     7      11     2   127    25 ; LYS(C)-TYR(C)-THR(C)
7     11     13     2   127    25 ; TYR(C)-THR(C)-VAL(C)
11   13     15     2   127    25 ; THR(C)-VAL(C)-SER(C)
13   15     17     2   127    25 ; VAL(C)-SER(C)-ILE(C)
15   17     19     2   127    25 ; SER(C)-ILE(C)-ASN(C)
17   19     21     2   127    25 ; ILE(C)-ASN(C)-GLY(C)
19   21     22     2   127    25 ; ASN(C)-GLY(C)-LYS(C)
21   22     25     2   127    25 ; GLY(C)-LYS(C)-LYS(C)
22   25     28     2   127    25 ; LYS(C)-LYS(C)-ILE(C)
25   28     30     2   127    25 ; LYS(C)-ILE(C)-THR(C)
28   30     32     2   127    25 ; ILE(C)-THR(C)-VAL(C)
30   32     34     2   127    25 ; THR(C)-VAL(C)-SER(C)
32   34     36     2   127    25 ; VAL(C)-SER(C)-ILE(C)

; Backbone-sidechain angles
2     1      4      2   100    25 ; LYS(C)-LYS(C) SBB
1     4      5      2   100    25 ; LYS(C)-LYS(C) SBB
4     7      8      2   100    25 ; LYS(C)-TYR(C) SBB
7     11     12     2   100    25 ; TYR(C)-THR(C) SBB
11   13     14     2   100    25 ; THR(C)-VAL(C) SBB
13   15     16     2   100    25 ; VAL(C)-SER(C) SBB
15   17     18     2   100    25 ; SER(C)-ILE(C) SBB
17   19     20     2   100    25 ; ILE(C)-ASN(C) SBB
21   22     23     2   100    25 ; GLY(C)-LYS(C) SBB
22   25     26     2   100    25 ; LYS(C)-LYS(C) SBB

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25    28    29      2    100    25 ; LYS(C)-ILE(C) SBB
28    30    31      2    100    25 ; ILE(C)-THR(C) SBB
30    32    33      2    100    25 ; THR(C)-VAL(C) SBB
32    34    35      2    100    25 ; VAL(C)-SER(C) SBB
34    36    37      2    100    25 ; SER(C)-ILE(C) SBB
; Sidechain angles
    1    2    3      2    180    25 ; LYS
    4    5    6      2    180    25 ; LYS
    7    8    9      2    150    50 ; TYR
    7    8   10      2    150    50 ; TYR
   22   23   24      2    180    25 ; LYS
   25   26   27      2    180    25 ; LYS

[ dihedrals ]
; Backbone dihedrals
[dihedrals]
; KKYT
1     4     7     11     1     0     3.70061  1
1     4     7     11     1     0     0.88488  2
1     4     7     11     1     0     0.913     3
1     4     7     11     1     0     0.16143  4
1     4     7     11     1     0     0.21993  5
1     4     7     11     1     90    -1.37761  1
1     4     7     11     1     90    2.05615   2
1     4     7     11     1     90    0.01273  3
1     4     7     11     1     90    -0.07164  4
1     4     7     11     1     90    -0.42108  5
; 1     4     7     11     1     90    6.44013   0
; KYTV
4     7     11    13     1     0     3.76407  1
4     7     11    13     1     0     0.92105  2
4     7     11    13     1     0     2.06649  3
4     7     11    13     1     0     0.63389  4
4     7     11    13     1     0     -0.11833 5
4     7     11    13     1     0     -0.46887 6
4     7     11    13     1     90    0.13201   1
4     7     11    13     1     90    2.5362    2
4     7     11    13     1     90    0.48809   3
4     7     11    13     1     90    -0.33137  4
4     7     11    13     1     90    -0.32474  5
4     7     11    13     1     90    0.03125   6
; 4     7     11    13     1     90    3.21116   0
; YTVS
7     11    13    15     1     0     3.91116  1
7     11    13    15     1     0     0.03712  2
7     11    13    15     1     0     0.09689  3
7     11    13    15     1     0     0.41527  4
7     11    13    15     1     0     -0.05057 5
7     11    13    15     1     90    0.99996  1
7     11    13    15     1     90    3.33799  2
7     11    13    15     1     90    0.685     3
7     11    13    15     1     90    0.61899  4
7     11    13    15     1     90    -0.32148 5
; 7     11    13    15     1     90    2.97958   0
; TVSI
11    13    15    17     1     0     3.44444  1
11    13    15    17     1     0     -0.3654   2
11    13    15    17     1     0     0.96655  3
11    13    15    17     1     0     0.30801  4
11    13    15    17     1     0     -0.10031 5
11    13    15    17     1     0     -0.12022 6
11    13    15    17     1     0     -0.02915 7
11    13    15    17     1     90    1.54725   1

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11	13	15	17	1	90	2.4196	2
11	13	15	17	1	90	0.0918	3
11	13	15	17	1	90	-0.4578	4
11	13	15	17	1	90	-0.85695	5
11	13	15	17	1	90	-0.43002	6
11	13	15	17	1	90	-0.11371	7
; 11	13	15	17	1	90	6.65684	0
; VSIN							
13	15	17	19	1	0	1.75741	1
13	15	17	19	1	0	-0.15709	2
13	15	17	19	1	0	-0.99829	3
13	15	17	19	1	0	0.52766	4
13	15	17	19	1	0	-0.49089	5
13	15	17	19	1	0	0.14996	6
13	15	17	19	1	0	-0.23281	7
13	15	17	19	1	90	2.16539	1
13	15	17	19	1	90	0.87817	2
13	15	17	19	1	90	1.17224	3
13	15	17	19	1	90	-0.50514	4
13	15	17	19	1	90	0.56317	5
13	15	17	19	1	90	0.22127	6
13	15	17	19	1	90	-0.27671	7
; 13	15	17	19	1	90	7.03003	0
; SING							
15	17	19	21	1	0	-0.46157	1
15	17	19	21	1	0	0.70728	2
15	17	19	21	1	0	0.941	3
15	17	19	21	1	0	0.65507	4
15	17	19	21	1	0	0.10295	5
15	17	19	21	1	0	-0.16128	6
15	17	19	21	1	0	-0.11663	7
15	17	19	21	1	90	-0.33144	1
15	17	19	21	1	90	0.15752	2
15	17	19	21	1	90	0.44237	3
15	17	19	21	1	90	0.82642	4
15	17	19	21	1	90	0.40483	5
15	17	19	21	1	90	-0.03855	6
15	17	19	21	1	90	-0.43519	7
; 15	17	19	21	1	90	9.171	0
; INGK							
17	19	21	22	1	0	-0.86717	1
17	19	21	22	1	0	0.81001	2
17	19	21	22	1	0	0.6579	3
17	19	21	22	1	0	-0.01495	4
17	19	21	22	1	0	0.10205	5
17	19	21	22	1	0	0.15164	6
17	19	21	22	1	0	-0.01947	7
17	19	21	22	1	0	-0.08432	8
17	19	21	22	1	0	0.02006	9
17	19	21	22	1	90	-2.41202	1
17	19	21	22	1	90	-0.68191	2
17	19	21	22	1	90	-0.39993	3
17	19	21	22	1	90	-0.80828	4
17	19	21	22	1	90	-0.76515	5
17	19	21	22	1	90	0.26492	6
17	19	21	22	1	90	0.63569	7
17	19	21	22	1	90	0.31539	8
17	19	21	22	1	90	-0.02	9
; 17	19	21	22	1	90	15.32339	0
; NGKK							
19	21	22	25	1	0	4.16935	1
19	21	22	25	1	0	-1.32069	2
19	21	22	25	1	0	0.99667	3

19	21	22	25	1	0	-0.25206	4
19	21	22	25	1	0	0.04114	5
19	21	22	25	1	0	-0.10523	6
19	21	22	25	1	0	0.01626	7
19	21	22	25	1	90	0.52452	1
19	21	22	25	1	90	1.18747	2
19	21	22	25	1	90	0.36998	3
19	21	22	25	1	90	-0.56404	4
19	21	22	25	1	90	0.41719	5
19	21	22	25	1	90	-0.34757	6
19	21	22	25	1	90	0.24711	7
; 19	21	22	25	1	90	7.15402	0
; GKKI							
21	22	25	28	1	0	3.35857	1
21	22	25	28	1	0	1.28718	2
21	22	25	28	1	0	0.36333	3
21	22	25	28	1	0	0.06896	4
21	22	25	28	1	0	-0.59184	5
21	22	25	28	1	0	-0.31134	6
21	22	25	28	1	0	0.00339	7
21	22	25	28	1	90	-0.57581	1
21	22	25	28	1	90	1.56454	2
21	22	25	28	1	90	0.39271	3
21	22	25	28	1	90	-0.73144	4
21	22	25	28	1	90	-0.17465	5
21	22	25	28	1	90	0.0753	6
21	22	25	28	1	90	0.09612	7
; 21	22	25	28	1	90	7.03307	0
; KK1T							
22	25	28	30	1	0	4.3381	1
22	25	28	30	1	0	1.08812	2
22	25	28	30	1	0	-0.11046	3
22	25	28	30	1	0	0.27796	4
22	25	28	30	1	0	-0.50607	5
22	25	28	30	1	0	-0.0532	6
22	25	28	30	1	90	-0.97285	1
22	25	28	30	1	90	3.06055	2
22	25	28	30	1	90	-0.53428	3
22	25	28	30	1	90	-0.04908	4
22	25	28	30	1	90	-0.38271	5
22	25	28	30	1	90	-0.40955	6
; 22	25	28	30	1	90	7.23719	0
; KITV							
25	28	30	32	1	0	3.26266	1
25	28	30	32	1	0	-0.35859	2
25	28	30	32	1	0	0.68107	3
25	28	30	32	1	0	0.10521	4
25	28	30	32	1	0	-0.01496	5
25	28	30	32	1	0	-0.15923	6
25	28	30	32	1	0	-0.17012	7
25	28	30	32	1	90	0.63646	1
25	28	30	32	1	90	1.97928	2
25	28	30	32	1	90	-0.14443	3
25	28	30	32	1	90	-0.06725	4
25	28	30	32	1	90	-0.30435	5
25	28	30	32	1	90	-0.06585	6
25	28	30	32	1	90	0.18504	7
; 25	28	30	32	1	90	6.94951	0
; ITVS							
28	30	32	34	1	0	3.12156	1
28	30	32	34	1	0	1.65553	2
28	30	32	34	1	0	0.00546	3
28	30	32	34	1	0	0.45185	4

```

25      28      30      32      1      0      -0.04926 5
25      28      30      32      1      0      -0.23398 6
25      28      30      32      1      0      -0.2555  7
28      30      32      34      1      90     -0.17809 1
28      30      32      34      1      90     2.6502   2
28      30      32      34      1      90     0.25742 3
28      30      32      34      1      90     0.00349 4
25      28      30      32      1      90     -0.26109 5
25      28      30      32      1      90     -0.12253 6
25      28      30      32      1      90     0.12481 7
; 28      30      32      34      1      90     4.95073 0
; TVSI
30      32      34      36      1      0      4.37967 1
30      32      34      36      1      0      -0.29652 2
30      32      34      36      1      0      -0.13548 3
30      32      34      36      1      0      -0.73646 4
25      28      30      32      1      0      -0.36224 5
30      32      34      36      1      90     -0.05251 1
30      32      34      36      1      90     0.69862 2
30      32      34      36      1      90     -0.09365 3
30      32      34      36      1      90     -0.23855 4
25      28      30      32      1      90     -0.18586 5
; 30      32      34      36      1      90     9.44401 0

; Sidechain improper dihedrals
    7      9      10      8      2      0      50 ; TYR

#ifndef POSRES
#include "posre.itp"
#endif

```

Appendix 2: Bonded parameters for sorbitol CG model

```
[ moleculetype ]
; molname nrexcl
SORB 1

[ atoms ]
;id type resnr residu atom cgnr charge mass
1 P4c 1 SORB B1 1 0.000 60.0528
2 P4c 1 SORB B2 2 0.000 60.0528
3 P4c 1 SORB B3 3 0.000 60.0528

[ constraints ]
; i j funct length forcec.
1 2 1 0.27 35000
2 3 1 0.27 35000

[ angles]
; i j k funct angle force.c.
1 2 3 2 142.000 170.0
```

Appendix 3: forcefield parameters for P4c bead type.

P4c 72.0 0.000 A 0.0 0.0

P4c	P4c	1	0.19402E-00	0.20914E-02 ; almost attractive
P5	P4c	1	0.17246E-00	0.18590E-02 ; semi attractive
SP5	P4c	1	0.24145E-00	0.26027E-02 ; supra attractive
P4c	P4	1	0.21558E-00	0.23238E-02 ; attractive
P4c	BP4	1	0.76824E-00	0.26348E-01 ; supra attractive, s=0.57
P4c	SP4	1	0.21558E-00	0.23238E-02 ; attractive
P4c	P3	1	0.21558E-00	0.23238E-02 ; attractive
P4c	SP3	1	0.21558E-00	0.23238E-02 ; attractive
P4c	P2	1	0.19402E-00	0.20914E-02 ; almost attractive
P4c	SP2	1	0.19402E-00	0.20914E-02 ; almost attractive
P4c	P1	1	0.19402E-00	0.20914E-02 ; almost attractive
P4c	SP1	1	0.19402E-00	0.20914E-02 ; almost attractive
P4c	Nda	1	0.17246E-00	0.18590E-02 ; semi attractive
P4c	SNda	1	0.17246E-00	0.18590E-02 ; semi attractive
P4c	Nd	1	0.17246E-00	0.18590E-02 ; semi attractive
P4c	Snd	1	0.17246E-00	0.18590E-02 ; semi attractive
P4c	Na	1	0.17246E-00	0.18590E-02 ; semi attractive
P4c	SNa	1	0.17246E-00	0.18590E-02 ; semi attractive
P4c	NO	1	0.15091E-00	0.16267E-02 ; intermediate
P4c	SN0	1	0.17246E-00	0.18590E-02 ; semi attractive
P4c	C5	1	0.13366E-00	0.14408E-02 ; almost intermediate
P4c	SC5	1	0.13366E-00	0.14408E-02 ; almost intermediate
P4c	C4	1	0.11642E-00	0.12549E-02 ; semi repulsive
P4c	SC4	1	0.11642E-00	0.12549E-02 ; semi repulsive
P4c	C3	1	0.11642E-00	0.12549E-02 ; semi repulsive
P4c	SC3	1	0.11642E-00	0.12549E-02 ; semi repulsive
P4c	C2	1	0.99167E-01	0.10690E-02 ; almost repulsive
P4c	AC2	1	0.99167E-01	0.10690E-02 ; almost repulsive
P4c	SC2	1	0.99167E-01	0.10690E-02 ; almost repulsive
P4c	C1	1	0.86233E-01	0.92953E-03 ; repulsive
P4c	AC1	1	0.86233E-01	0.92953E-03 ; repulsive
P4c	SC1	1	0.86233E-01	0.92953E-03 ; repulsive
P4c	Qda	1	0.24145E-00	0.26027E-02 ; supra attractive
P4c	SQda	1	0.24145E-00	0.26027E-02 ; supra attractive
P4c	Qd	1	0.24145E-00	0.26027E-02 ; supra attractive
P4c	SQd	1	0.24145E-00	0.26027E-02 ; supra attractive
P4c	Qa	1	0.24145E-00	0.26027E-02 ; supra attractive
P4c	SQa	1	0.24145E-00	0.26027E-02 ; supra attractive
P4c	Q0	1	0.24145E-00	0.26027E-02 ; supra attractive
P4c	SQ0	1	0.24145E-00	0.26027E-02 ; supra attractive