Mesoporous Carbon Nanospheres Featured Fluorescent Aptasensor for Multiple Diagnosis of Cancer *in vitro* and *in vivo*

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Table S1.

The performances of the OMCN/ P_0 -Cy3 aptasensor for detecting MUC1 or MCF-7 cells in different mediums.

Medium	Recovery (%)			Stability ^a (%)	Repeatability ^b (%)	
	Low (1.06 µM)	Middle (5.03 µM)	High (10.6 μM)	(12 h)	FS	MR
Tris-HCl	92.73	95.63	105.18	93.4	4.45	1.65
Serum	90.22	93.81	97.58	95.74	2.44	1.47
Cell	108.85	92.15	96.75	92.72	5.05	2.68

^a The stability of OMCN/P₀-Cy3 aptasensor after storing for 12 h. ^b The RSD of five times measurements (n = 5). FS: the results of fluorescence spectrophotometer. MR: the results of microplate reader.



Figure S1. (A) The IR spectra of MCN and OMCN. (B) The Raman spectra of MCN and OMCN.



Figure S2. (A) Fluorescence quenching and recovering of the OMCN/P0-Cy3 aptasensor incubated without (black line) and with (red line) MUC1 as a function of time. (B) Effect of OMCN concentrations on the fluorescence intensity of the OMCN/P0-Cy3 aptasensor in the absence (black line) and presence (red line) of MUC1. The inset was the ratio of F/F0 as a function of OMCN concentrations. F and F0 are the fluorescence intensities with and without MUC1, respectively.



Figure S3. (A, B) Fluorescent spectra of OMCN/P₀-Cy3 in the presence of MUC1 with different concentrations in Tris-HCl solution and in 4% serum-containing buffer solution, respectively. The insets were their calibration curves for MUC1 detections. (C) Fluorescent spectra of OMCN/P₀-Cy3 in the presence of MCF-7 cells with different concentrations in Hank's buffer solution. The inset was the calibration curve for MCF-7 detection.



Figure S4. (A, B) The calibration curves for MUC1 detections in Tris-HCl solution and 4% serum-containing buffer solution, respectively, using the Infinite M1000 Pro microplate reader. (C) The calibration curve for MCF-7 detection in Hank's buffer solution using the Infinite M1000 Pro microplate reader.



Figure S5. (A-C) Fluorescent spectra of OMCN/P₀-Cy3 upon detecting MUC1 (3.18 μ M) in Tris-HCl solution (A), 4% serum-containing buffer solution (B), and 2 × 10⁵ cells/mL MCF-7 cells in Hank's buffer solution (C) for 5 times, respectively.



Figure S6. (A-C) Fluorescent spectra of the 0-12 h stored OMCN/P₀-Cy3 aptersensor for detecting MUC1 (10 μ M) in Tris-HCl solution (A), 4% serum-containing buffer solution (B), and MCF-7 cells (2 × 10⁶ cells/mL) in Hank's buffer solution (C), respectively.



Figure S7. The F/F_0 ratios of the OMCN/P₀-Cy3 aptasensor in the presence of lysozyme (20 μ M), MUC1 (10 μ M), HSA (20 μ M), and cytochrome C (20 μ M), respectively. F and F₀ are the fluorescence intensities with and without the corresponding proteins, respectively.



Figure S8. The viability of MCF-7 and MCF-10A cells after treating with different concentrations of OMCN for 24 h, respectively.



Figure S9. Different magnified confocal fluorescence microscopy images and corresponding 2.5 dimensional sectional views of MCF-10A cells incubated with OMCN/P₀-Cy3 for different time periods. Scale bar: 50 μ m (200 × images), 10 μ m (900 × images).



Figure S10. The inverted fluorescence microscopy images of liver and kidney sections treated without or with OMCN/P₀-Cy3. Bar = $200 \mu m$.



Figure S11. In vivo fluorescent images of normal nude mice treated with P_0 -Cy3 via intravenous injection. Images were taken at 0 min (A), 5 min (B), 15 min (C), 30 min (D) and 60 min (E), respectively.