Protein Arginine Methyltransferase 8: Tetrameric Structure and Protein Substrate Specificity

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Supplementary Tables and Figures

Table S1: The statistics table for the tPRMT8 X-ray crystal structure

Name	tPRMT8
PDB code	4X41
Data collection	
Resolution (Å)	25-3.49 (3.62-3.49) [*]
Space group	P222 ₁
Unit-cell	
a / b / c (Å)	68.2/78.2/203.9
$\alpha / \beta / \gamma$ (°)	90/90/90
No. of reflections Measured	13561 (1357)
Completeness (%)	94.8 (96.7)
R _{sym} (%) ^a	27.2 (72.4)
Mean I/ σ (I)	3.9 (2.0)
Multiplicity	2.9 (2.8)
Refinement	
R _{work} (%)	23.1
R _{free} (%)	28.2
Geometry deviations	
Bond lengths (Å)	0.005
Bond angles (°)	1.02
No. of atoms / Mean B-values ($Å^2$)	4998/55.3
Ramachandran plot (%)	
Most favored	93.0
Allowed	7.0
Disallowed	0.0

Values in parentheses are for the highest resolution shell.

^a $R_{sym} = \sum_{hkl} \sum_{i} |I_i(hkl) - \langle I(hkl) \rangle | / \sum_{hkl} \sum_{i} I_i(hkl).$

Data Collection Parameters	
Instrument	SSRL BL4-2
Defining slits size (H mm x V mm)	0.3 x 0.3
Detector distance (m)	2.5
Wavelength (Å); energy (keV)	1.127; 11
Beam current (Å)	500
Q range (Å ⁻¹)	0.007-0.46
Exposure time per frame (s)	1
Size of quarz capillary in diameter (mm)	1.5
Frame per FPLC experiment	800
Amount loaded (µl)	100
Sample concentration (mg/ml)	5.0
Temperature (K)	293
SEC column	Superdex 200PC 3.2/3
FPLC flow rate (ml/min)	0.05
Structural Parameters	
Image frame used for analysis	670-699
Q region (Å ⁻¹)	0.013-0.178
I(0) from Guinier	476.70
Rg (Å) from Guinier	44.31
I(0) from $P(r)$	477.90
Rg (Å) from $P(r)$	44.68
Dmax (Å) from $P(r)$	115.22
Rg (Å) from crystal structure	43.63
Dmax (Å) from crystal structure	119
Porod Volume estimate from $P(r)$	308,674
Excluded volume from DAMMIF (Å ³)	431,578.95
Dry volume calculated from sequence $(Å^3)$	210,564
Calculated tetrameric molecular weight	166.23
Software employed	
Primary data reduction	SasTool
Data processing	PRIMUS
Ab initio analysis	DAMMIF
Validation and averaging	DAMAVER
Computation of model intensities	FoXS

 Table S2: Data collection and scattering-derived parameters for tPRMT8

Figure S1: Sequence alignment of PRMT1, Hmt1, PRMT8 and tPRMT8. The sequence of the two PRMT8 constructs, PRMT8 and tPRMT8 are aligned with the full length PRMT1 and the yeast PRMT1 homolog, Hmt1. The conserved residues are marked with star and the highly similar and less similar residues are indicated by double and single dots respectively (generated by ClustalW2, EMBL-EBI).

PRMT8	MGMKHSSRCLLLRRKMAENAAESTEVNSPPSQPPQPVVPAKPVQCVHHVSTQPSCPGRGK	60
tPRMT8 PRMT1		0
Hmt1		5
PRMT8	MSKLLNPEEMTSRDYYFDSYAHFGIHEEMLKDEVRTLTYRNSMYHNKHVFKDKVVLDVGS	
tPRMT8	MSKLLNPEEMTSRDYYFDSYAHFGIHEEMLKDEVRTLTYRNSMYHNKHVFKDKVVLDVGS	
PRMT1	SSEKPNAEDMISKDYYFDSYAHFGIHEEMLKDEVRILIYRNSMFHNRHLFKDKVVLDVGS	
Hmt1	DYYFDSYDHYGIHEEMLQDTVRTLSYRNAIIQNKDLFKDKIVLDVGC	47
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PRMT8	GTGILSMFAAKAGAKKVFGIECSSISDYSEKIIKANHLDNIITIFKGKVEEVELPVEKVD	180
tPRMT8	GTGILSMFAAKAGAKKVFGIECSSISDYSQKIIKANHLDNIITIFKGKVEEVELPVEKVD	120
PRMT1	GTGILCMFAAKAGARKVIGIECSSISDYAVKIVKANKLDHVVTIIKGKVEEVELPVEKVD	129
Hmt1	GTGILSMFAAKHGAKHVIGVDMSSIIEMAKELVELNGFSDKITLLRGKLEDVHLPFPKVD	107
PRMT8	IIISEWMGYCLFYESMLNTVIFARDKWLKPGGLMFPDRAALYVVAIEDRQYKDFKIHWWE	240
tPRMT8	IIISEWMGYCLFYESMLNTVIFARDKWLKPGGLMFPDRAALYVVAIEDRQYKDFKIHWWE	180
PRMT1	IIISEWMGYCLFYESMLNTVLYARDKWLAPDGLIFPDRATLYVTAIEDRQYKDYKIHWWE	189
Hmt1	IIISEWMGYFLLYESMMDTVLYARDHYLVEGGLIFPDKCSIHLAGLEDSQYKDEKLNYWQ	167
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PRMT8	NVYGFDMTCIRDVAMKEPLVDIVDPKQVVTNACLIKEVDIYTVKTEELSFTSAFCLQIQR	300
tPRMT8	NVYGFDMTCIRDVAMKEPLVDIVDPKQVVTNACLIKEVDIYTVKTEELSFTSAFCLQIQR	240
PRMT1	NVYGFDMSCIKDVAIKEPLVDVVDPKQLVTNACLIKEVDIYTVKVEDLTFTSPFCLQVKR	249
Hmt1	DVYGFDYSPFVPLVLHEPIVDTVERNNVNTTSDKLIEFDLNTVKISDLAFKSNFKLTAKR	227
PRMT8	NDYVHALVTYFNIEFTKCHKKMGFSTAPDAPYTHWKQTVFYLEDYLTVRRGEEIYGTI	
tPRMT8	NDYVHALVTYFNIEFTKCHKKMGFSTAPDAPYTHWKQTVFYLEDYLTVRRGEEIYGTI	
PRMT1	NDYVHALVAYFNIEFTRCHKRTGFSTSPESPYTHWKQTVFYMEDYLTVKTGEEIFGTI	
Hmt1	QDMINGIVTWFDIVFPAPKGKRPVEFSTGPHAPYTHWKQTIFYFPDDLDAETGDTIEGEL	287
PRMT8	SMKPNAKNVRDLDFTVDLDFKGQLCETSVSNDYKMR 394	
tPRMT8	SMKPNAKNVRDLDFTVDLDFKGQLCETSVSNDYKMR 334	
PRMT1	GMRPNAKNNRDLDFTIDLDFKGQLCELSCSTDYRMR 343	
Hmt1	VCSPNEKNNRDLNIKISYKFESNGIDGNSRSRKNEGSYLMH 328	
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Figure S2: Homotetramerization of tPRMT8: The size exclusion chromatography profile of tPRMT8 at various buffers (left) and salt (right) conditions. The SEC profiles of tPRMT8 were almost identical with varying salt concentration and pH range.

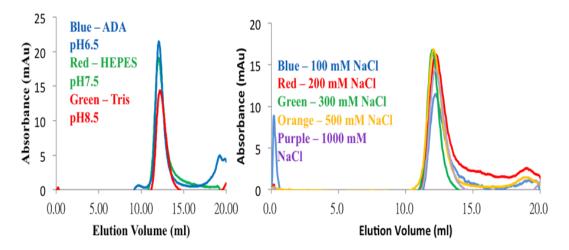
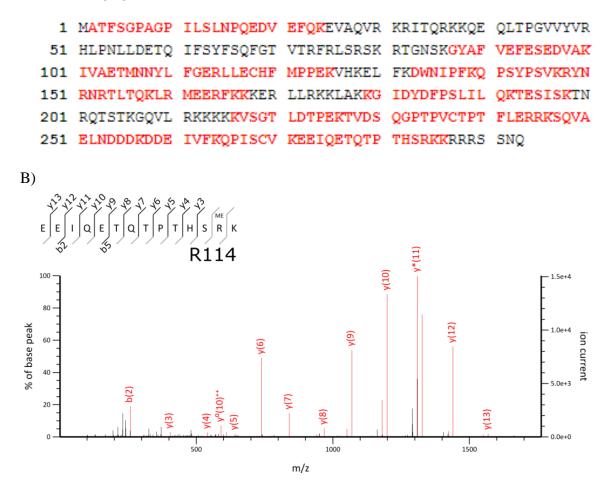


Figure S3: The *in vitro* **methylation of NIFK.** A) The methylated NIFK by tPRMT8 was digested by Lys-C protease followed by LS-MS/MS which shown 64% coverage. B). In LC-MS/MS spectra, Arg114 and Arg284 were mono-methylated. The Arg244 and Arg245 are close neighbors which leads to difficulties in identifying the methylated arginine and the degree of methylation.

A)

Matched peptides shown in **bold red**.



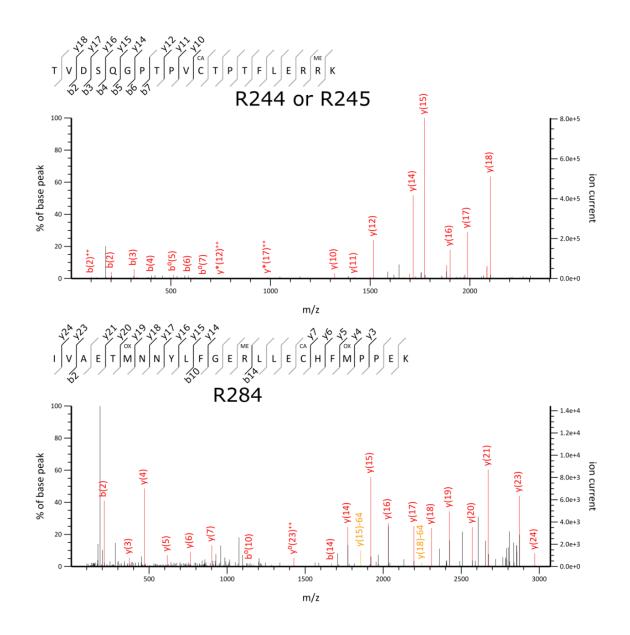


Figure S4: tPRMT8C is structurally similar to tPRMT8

A) The AUC overlay of tPRMT8 (black) and tPRMT8C (in red) indicates a similar profile that represents a major species consistent with a tetrameric form of ~160 kDa. B) The CD spectra (left) of tPRMT8 (red triangle) overlay with tPRMT8C (blue square) suggests the secondary structures are very similar. The comparison of Tm values (right) for tPRMT8 (red triangle) with tPRMT8C (blue square) suggests tPRMT8c is less stable as the Tm value decreased by ~10°C.

