

Supporting information

siRNA-Loaded Polyion Complex Micelle Decorated with Charge- Conversional Polymer Tuned to Undergo Stepwise Response to Intra- Tumoral and Intra-Endosomal pHs for Exerting Enhanced RNAi Efficacy

Montira Tangsangasakri¹, Hiroyasu Takemoto², Mitsuru Naito³, Yoshinori Maeda³, Daiki Sueyoshi¹, Hyun Jin Kim³, Yutaka Miura³, Jooyeon Ahn⁴, Ryota Azuma⁴, Nobuhiro Nishiyama^{2,5}, Kanjiro Miyata^{3,5*}, and Kazunori Kataoka^{1,3,4,5,*}

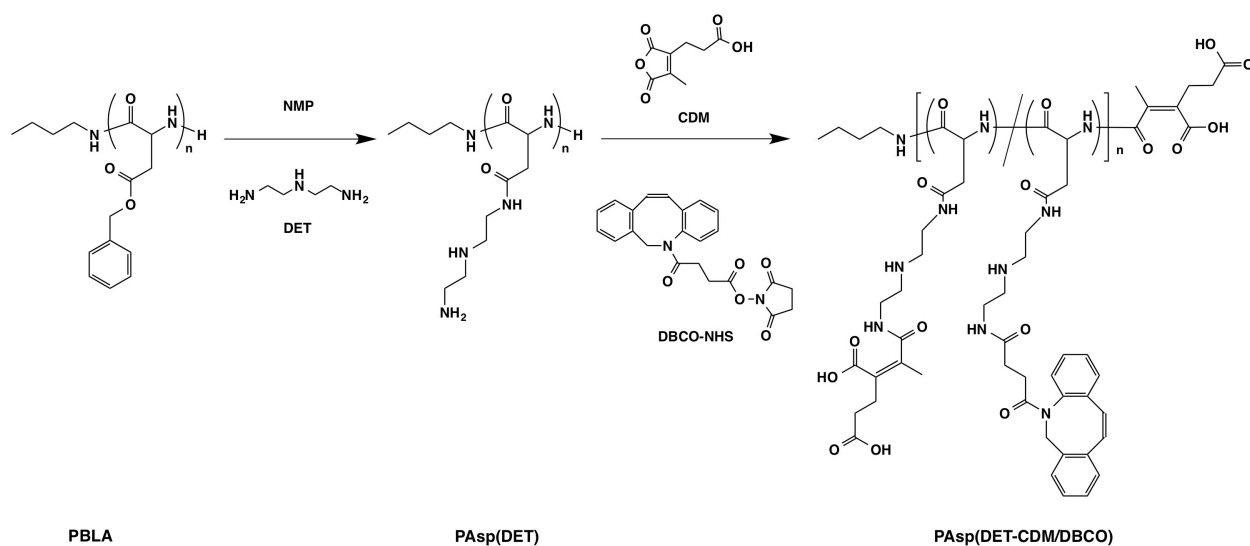
¹Department of Bioengineering, Graduate School of Engineering, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-8656, Japan

²Polymer Chemistry Division, Chemical Resources Laboratory, Tokyo Institute of Technology, R1-11, 4259 Nagatsuta, Midori-ku, Yokohama 226-8503, Japan

³Center for Disease Biology and Integrative Medicine, Graduate School of Medicine, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-0033, Japan

⁴Department of Materials Engineering, Graduate School of Engineering, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-8656, Japan

⁵Innovation Center of NanoMedicine, Institute of Industry Promotion-Kawasaki, 3-25-14 Tonomachi, Kawasaki-ku, Kawasaki 210-0821, Japan.



Scheme S1. Synthesis procedure of PAsp(DET-CDM/DBCO). This polyaspartamide derivative has the mixed sequence of α and β isomers and its ω -end may be modified with either CDM or DBCO moieties. The α isomers of PAsp(DET) and PAsp(DET-CDM/DBCO) and the CDM-modified ω -end are depicted as a representative structure for simplicity.

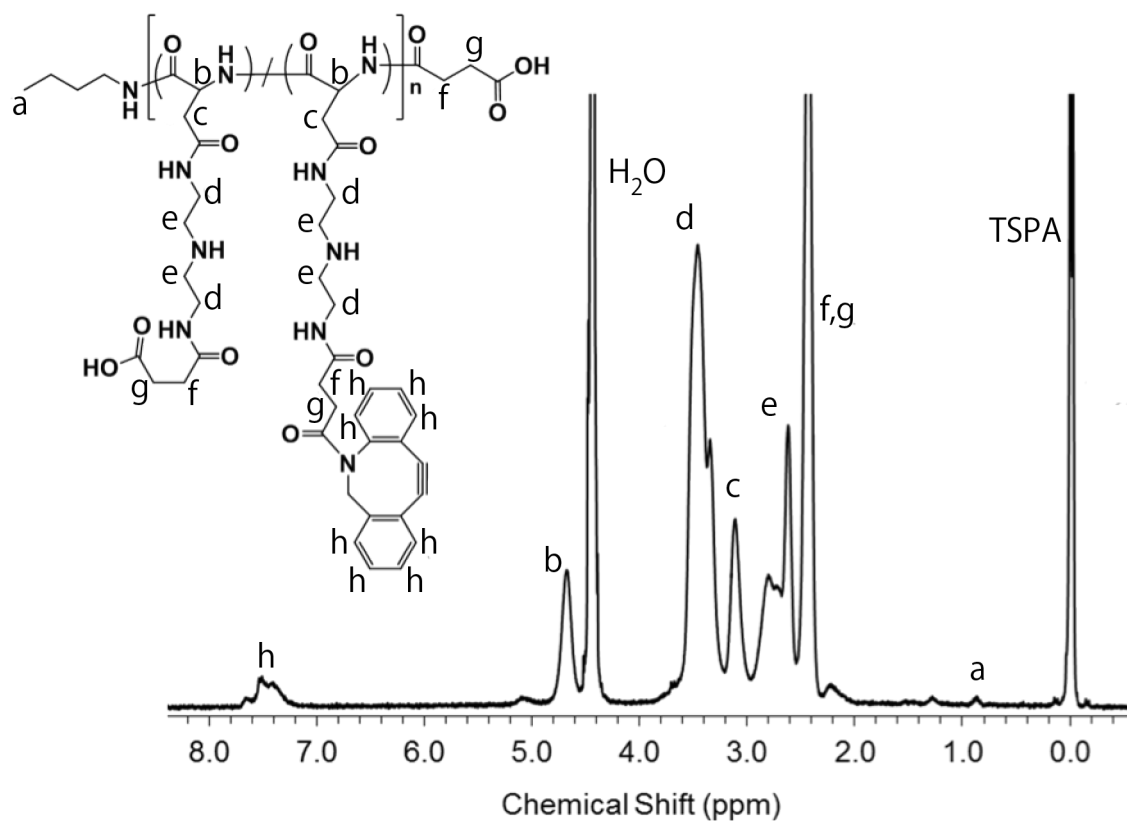


Figure S1. ^1H NMR spectrum of PAsp(DET-SUC/DBCO) in D_2O at 70°C (polymer concentration = 5 mg/mL). This polyaspartamide derivative has the mixed sequence of α and β isomers and its ω -end may be modified with either SUC or DBCO moieties. Only α isomer of PAsp(DET-SUC/DBCO) and SUC-modified ω -end are depicted as a representative structure for simplicity.

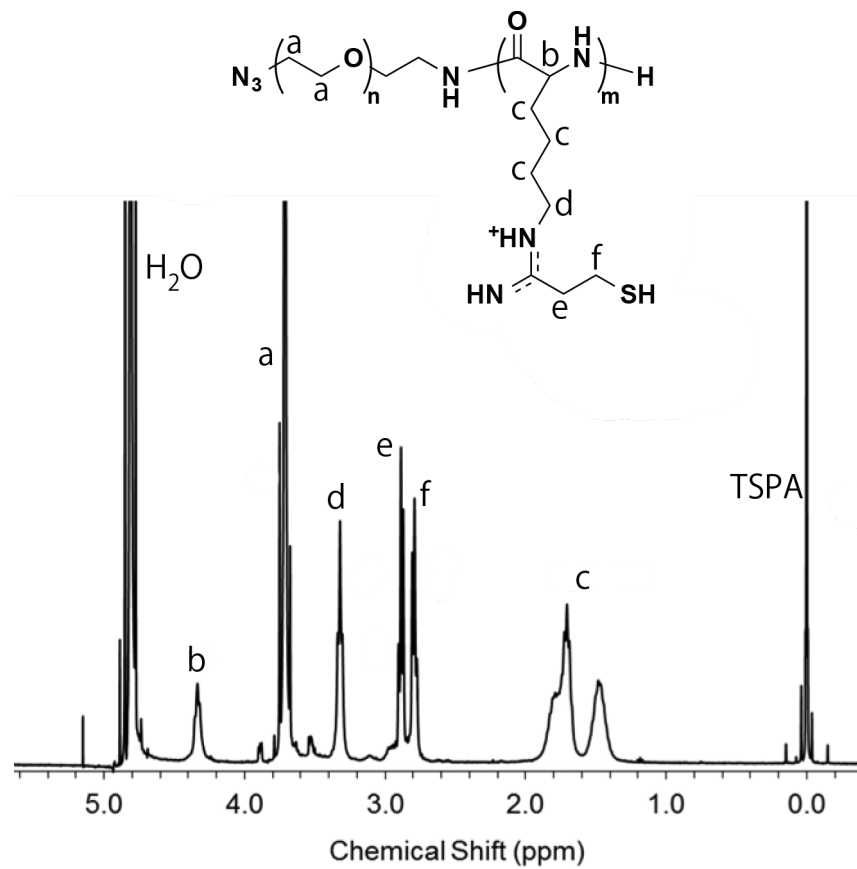


Figure S2. ^1H NMR spectrum of $\text{N}_3\text{-PEG-}b\text{-PLys(MPA)}$ in D_2O at $25\text{ }^\circ\text{C}$ (polymer concentration = 5 mg/mL).

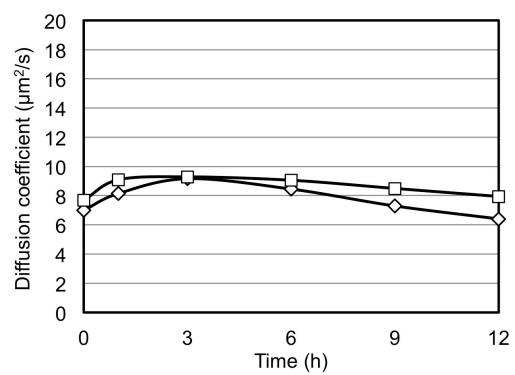


Figure S3. Stability of nonmodified micelles (open diamond) and CDM-micelles (open square) (100 nM Cy3/Chol-siRNA) in 90% FBS-containing HEPES buffer (pH 7.4), determined by FCS.

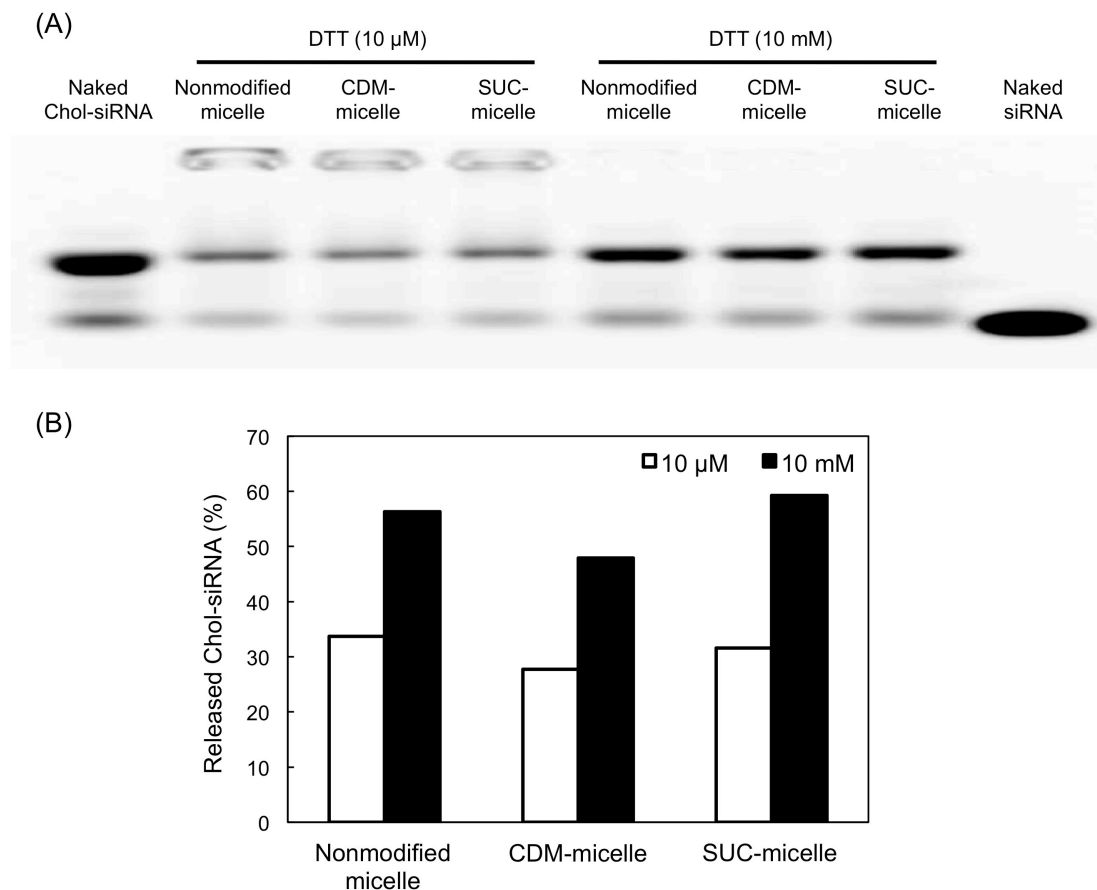


Figure S4. siRNA release profiles of nonmodified micelle, CDM-micelle, and SUC-micelle under two varying reductive conditions. Each micelle sample was prepared with Cy3/Chol-siRNA and mixed with dextran sulfate at a residual molar ratio of sulfate group in dextran sulfate to phosphate group in siRNA = 0.5 in the presence of 10 μ M or 10 mM concentration of dithiothreitol (DTT). After 1 h incubation at 37 $^{\circ}$ C, samples were electrophoresed in 1% agarose gel (100 V, 20 min). The agarose gel was imaged (A) and quantitatively analyzed based on densitometry of detected Cy3 signals (B) using Molecular Imager[®] PharosFX[™] Systems (Bio-Rad Laboratories).

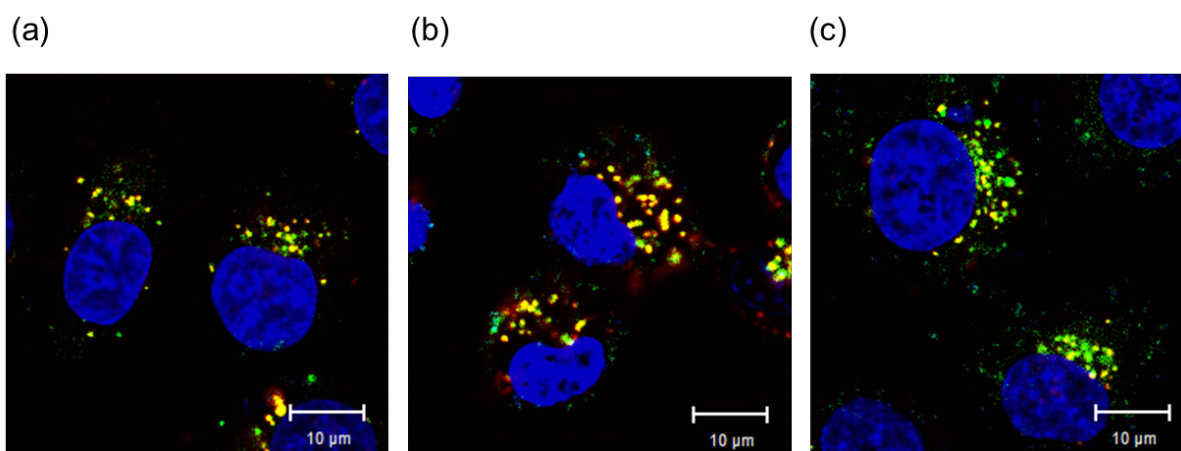


Figure S5. CLSM images of A549 cells after 12 h incubation with (a) nonmodified micelles, (b) CDM-micelles, and (c) SUC-micelles at pH 6.7 (siRNA concentration = 200 nM). Red: Cy3/Chol-siRNA, green: late endosome/lysosome stained with LysoTracker Green, blue: nuclei stained with Hoechst 33342, and yellow: colocalization between red and green pixels.

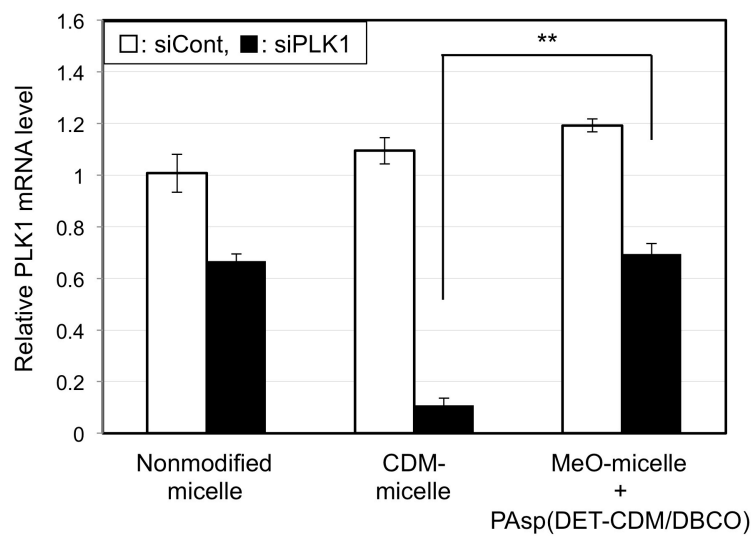


Figure S6. Gene silencing efficiencies of nonmodified micelles prepared from N₃-PEG-*b*-PLys(MPA) without PAsp(DET-CDM/DBCO), CDM-micelles prepared from N₃-PEG-*b*-PLys(MPA) with PAsp(DET-CDM/DBCO), and MeO-micelles prepared from MeO-PEG-*b*-PLys(MPA) with PAsp(DET-CDM/DBCO) in A549 cells cultured at pH 6.7 (siRNA concentration = 200 nM). Data represent the means \pm SEM (n = 4). **: $p < 0.01$.

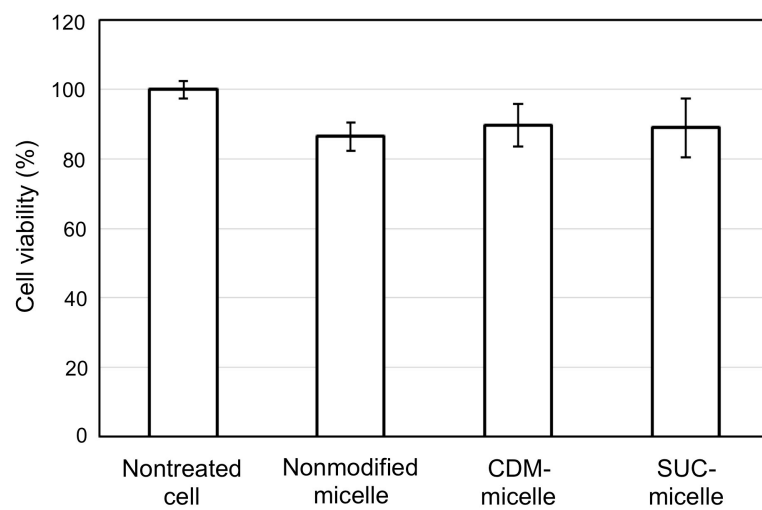


Figure S7. Viability of A549 cells after 48 h incubation with nonmodified micelles, CDM-micelles, and SUC-micelles at pH 7.4 (siRNA concentration = 200 nM). Data represent the means \pm SEM (n = 4).