

Supporting Information

Mechanism of the App^{ABLUF} Photocycle Probed by Site-Specific Incorporation of Fluorotyrosine Residues: The Effect of the Y21 pKa on the Forward and Reverse Ground-State Reactions

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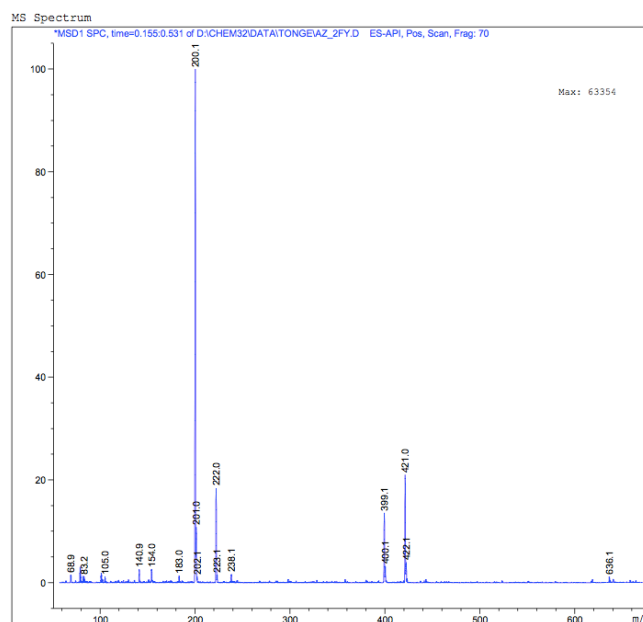
Table S1. pH dependence of the light to dark state recovery in wild-type and FY21 AppA_{BLUF}(Y56F).

	Recovery Rate /s ⁻¹				
	pH6/pD6	pH7/pD7	pH8/pD8	pH9/pD9	pH10/pD10
AppA_{BLUF}	ND	7.3 x 10 ⁻⁴ /ND	6.5 x 10 ⁻⁴ /1.3 x 10 ⁻⁴	7.5 x 10 ⁻⁴ /ND	ND
AppA_{BLUF}(Y56F)	ND	6.0 x 10 ⁻⁴ /ND	6.5 x 10 ⁻⁴ /1.3 x 10 ⁻⁴	6.4 x 10 ⁻⁴ /ND	ND
2-FY21					
AppA_{BLUF}(Y56F)	ND	1.3 x 10 ⁻² /ND	1.1 x 10 ⁻² /1.8 x 10 ⁻³	1.2 x 10 ⁻² /ND	ND
3-FY21					
AppA_{BLUF}(Y56F)	ND	3.6 x 10 ⁻² /4.9 x 10 ⁻³	3.4 x 10 ⁻² /3.9 x 10 ⁻³	3.7 x 10 ⁻² /ND	ND
2,3-F₂Y21					
AppA_{BLUF}(Y56F)	0.26/ND	0.22/0.39	0.22/0.04	0.24/0.045	0.3/0.06
3,5-F₂Y21					
AppA_{BLUF}(Y56F)	0.5/0.15	0.56/0.13	0.63/0.11	0.69/0.13	0.75/0.15
2,3,5-F₃Y21					
AppA_{BLUF}(Y56F)	2.7/1.48	2.9/1.21	2.6/1.15	2.7/1.16	ND/1.18

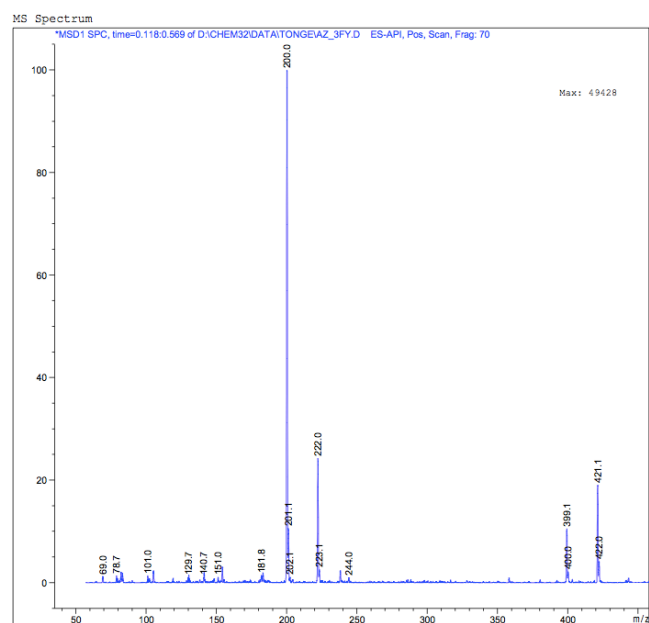
¹ND: The recovery rate was not determined at pH6/pD6 or pH10/pD10 for all samples. In some cases this was due to the impact of pH on protein stability.

Figure S1. Mass spectra of 2-FY, 3-FY, 2,3-F₂Y, 3,5-F₂Y, and 2,3,5-F₃Y.

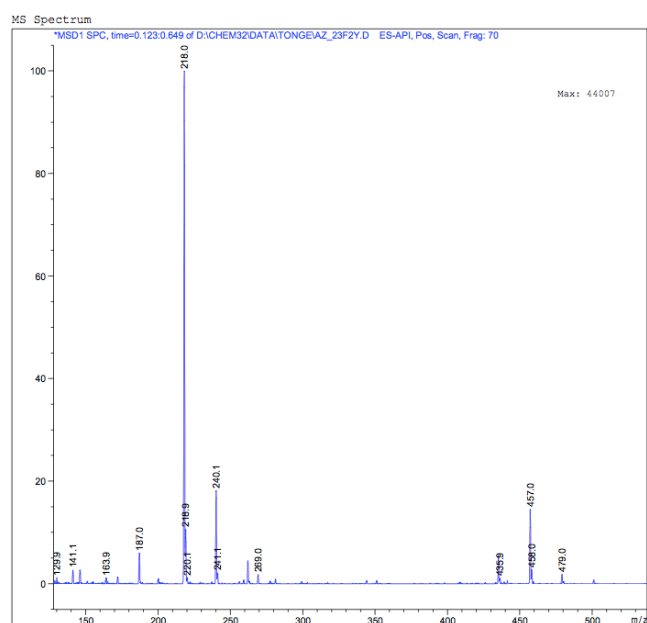
A) 2-FY. Mass (M+H) calculated for C₉H₁₁FNO₃ 200.07, found 200.1.



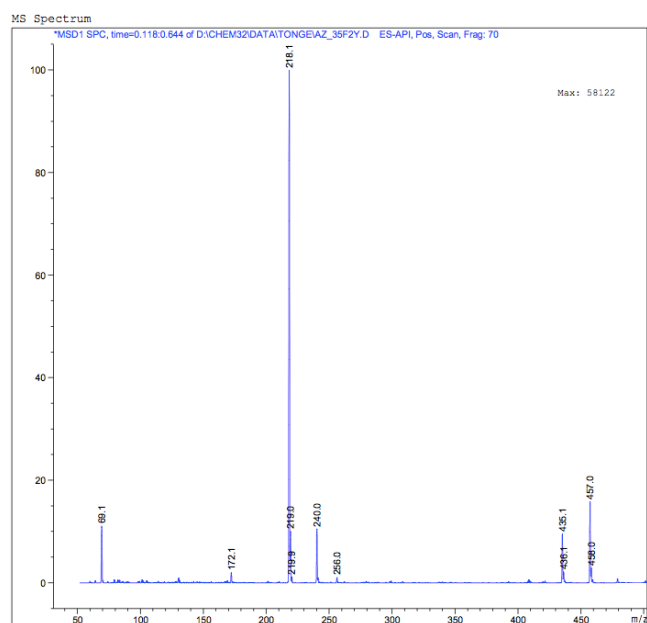
B) 3-FY Mass (M+H) calculated for C₉H₁₁FNO₃ 200.07, found 200.0.



C) 2,3-F₂Y Mass (M+H) calculated for C₉H₁₀F₂NO₃ 218.1, found 218.0.



D) 3,5-F₂Y Mass (M+H) calculated for C₉H₁₀F₂NO₃ 218.1, found 218.1.



E) 2,3,5-F₃Y Mass (M+H) calculated for C₉H₉F₃NO₃ 236.05, found 236.0.

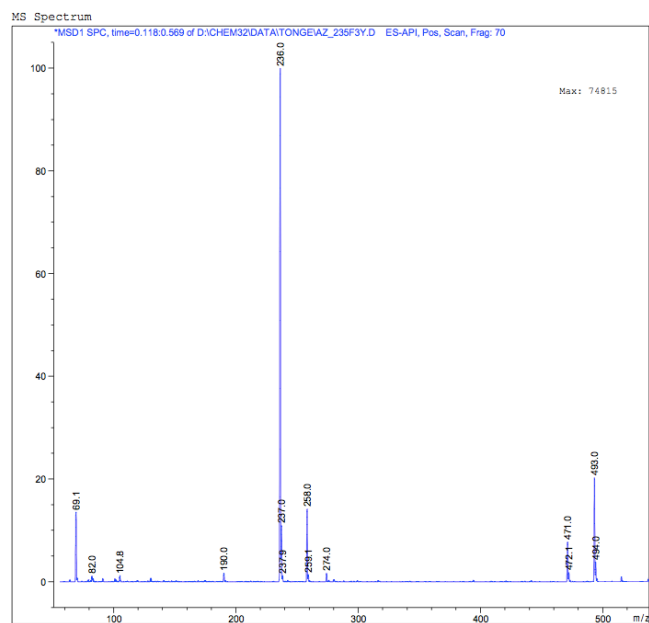
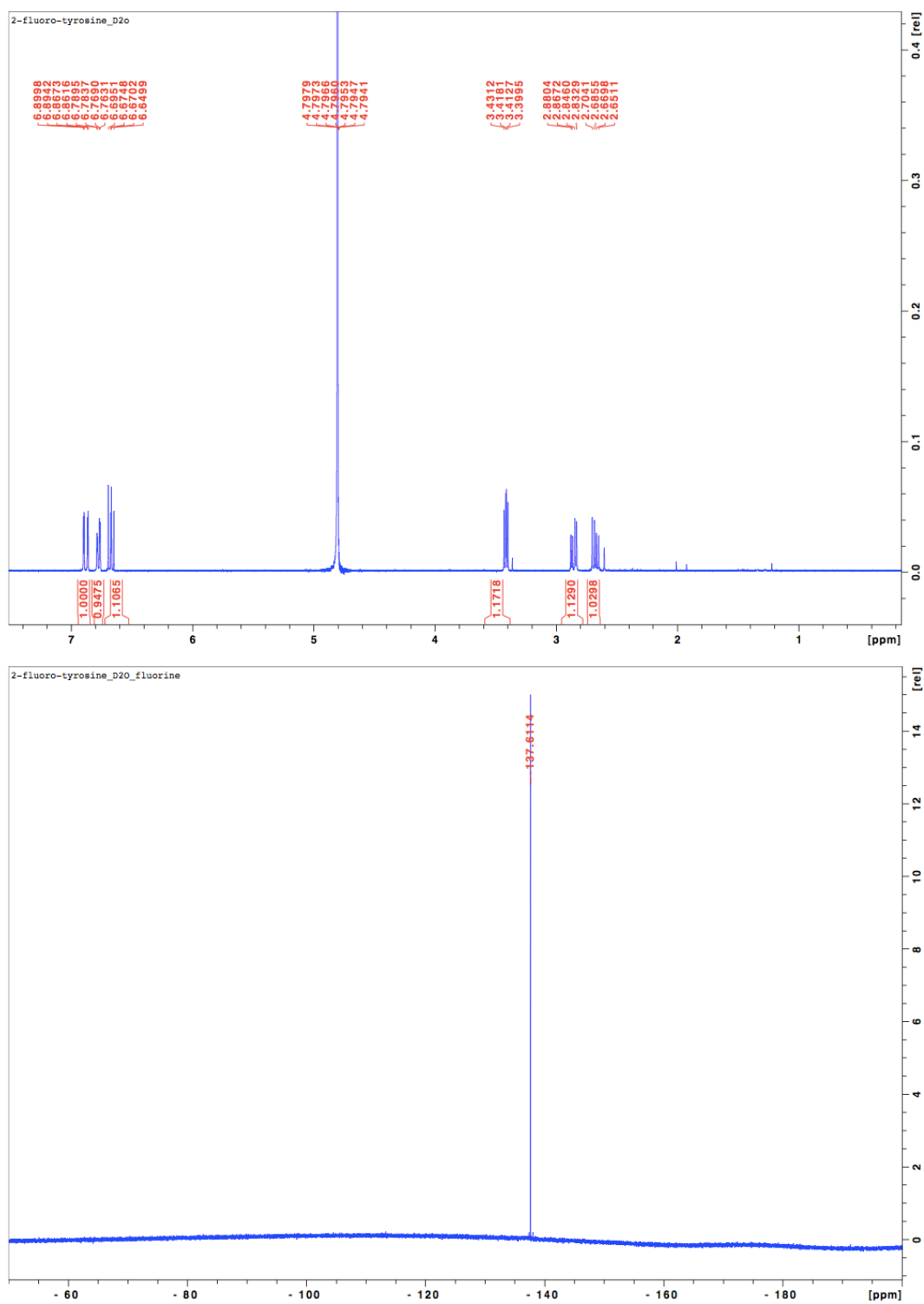
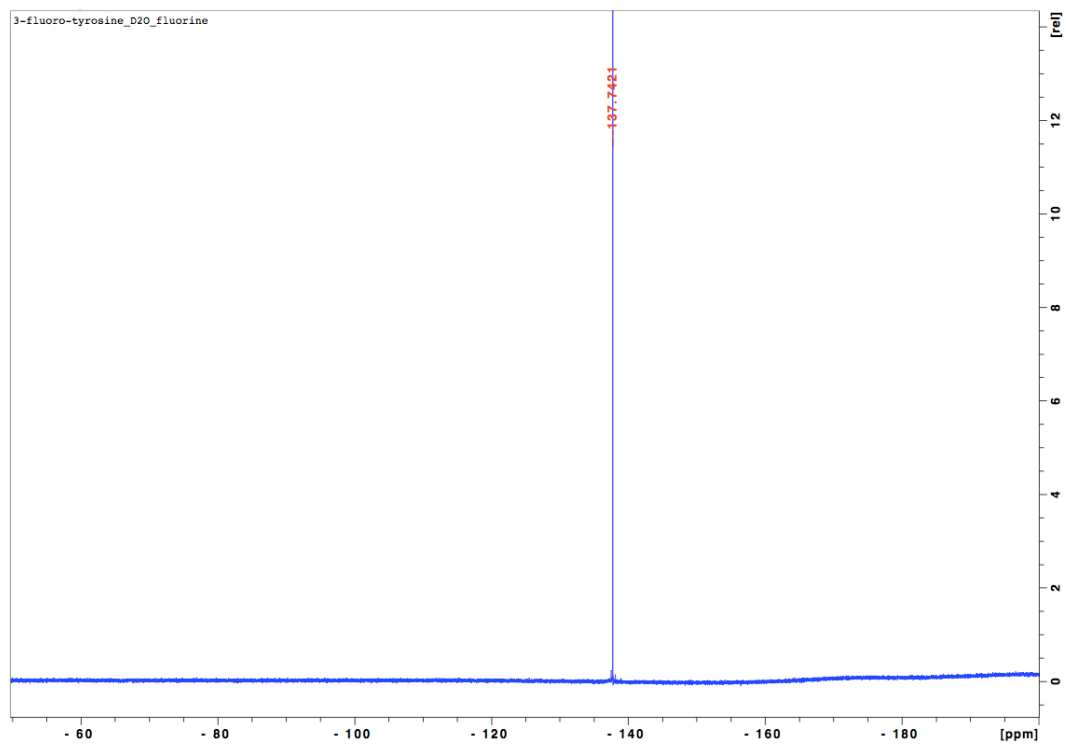
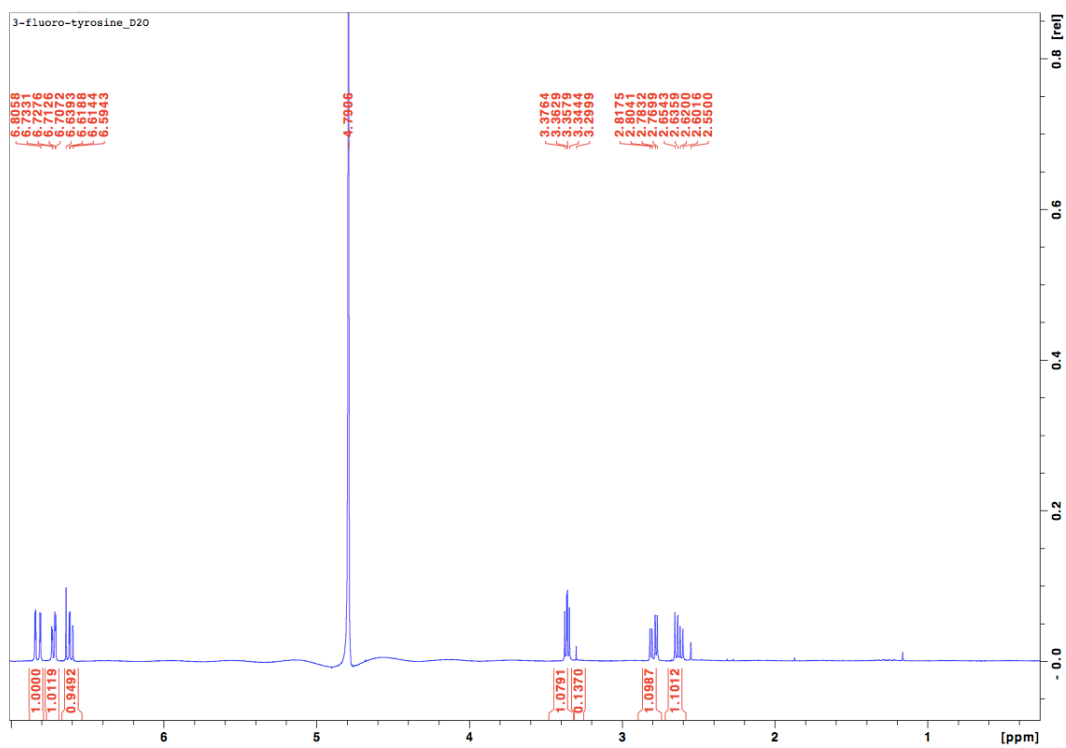


Figure S2. ^1H NMR and ^{19}F NMR of the Fluorotyrosines in D_2O .

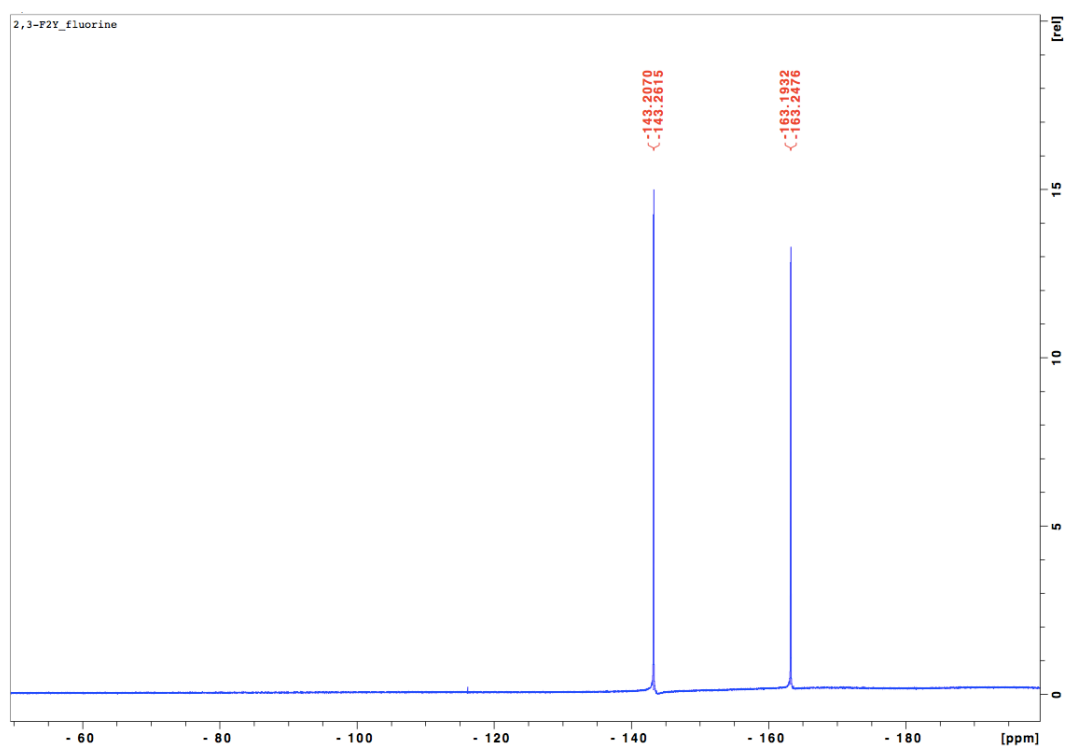
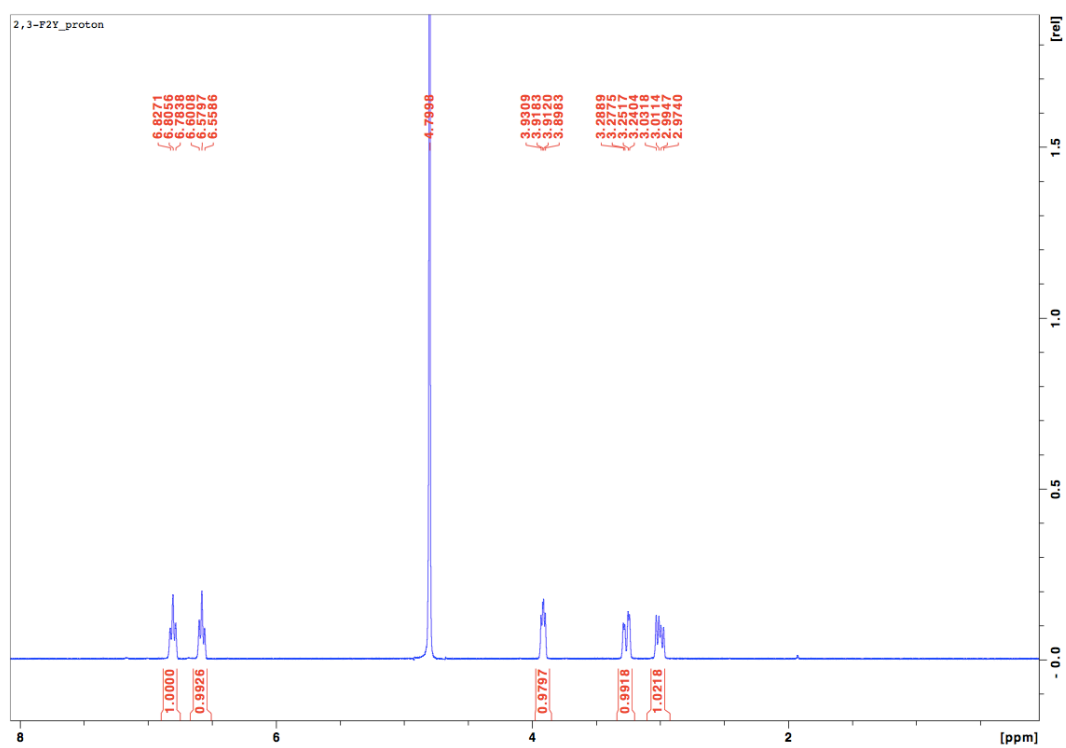
A) 2-FY



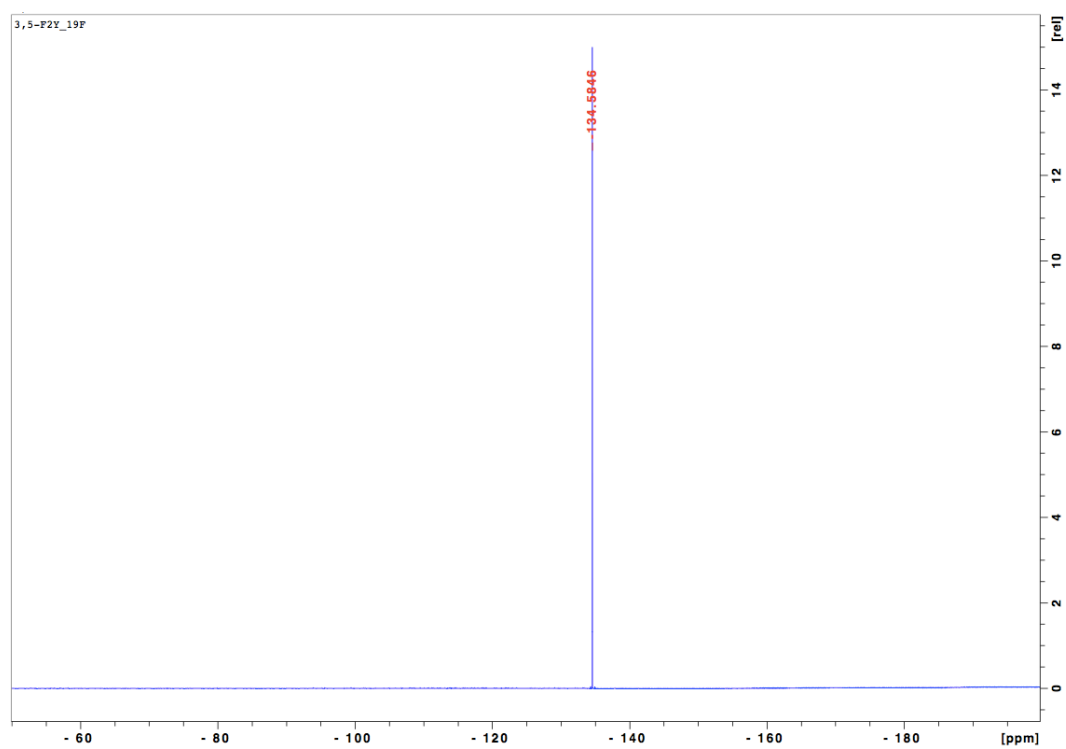
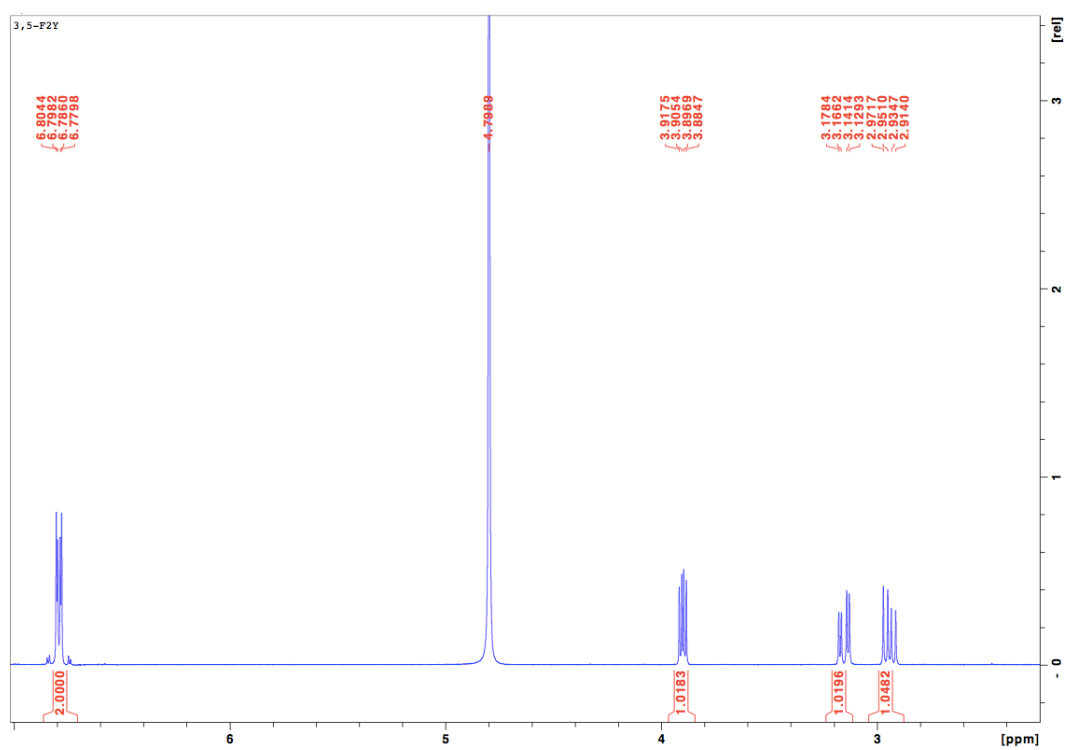
B) 3-FY



C) 2,3-F₂Y



D) 3,5-F₂Y



E) 2,3,5-F₃Y

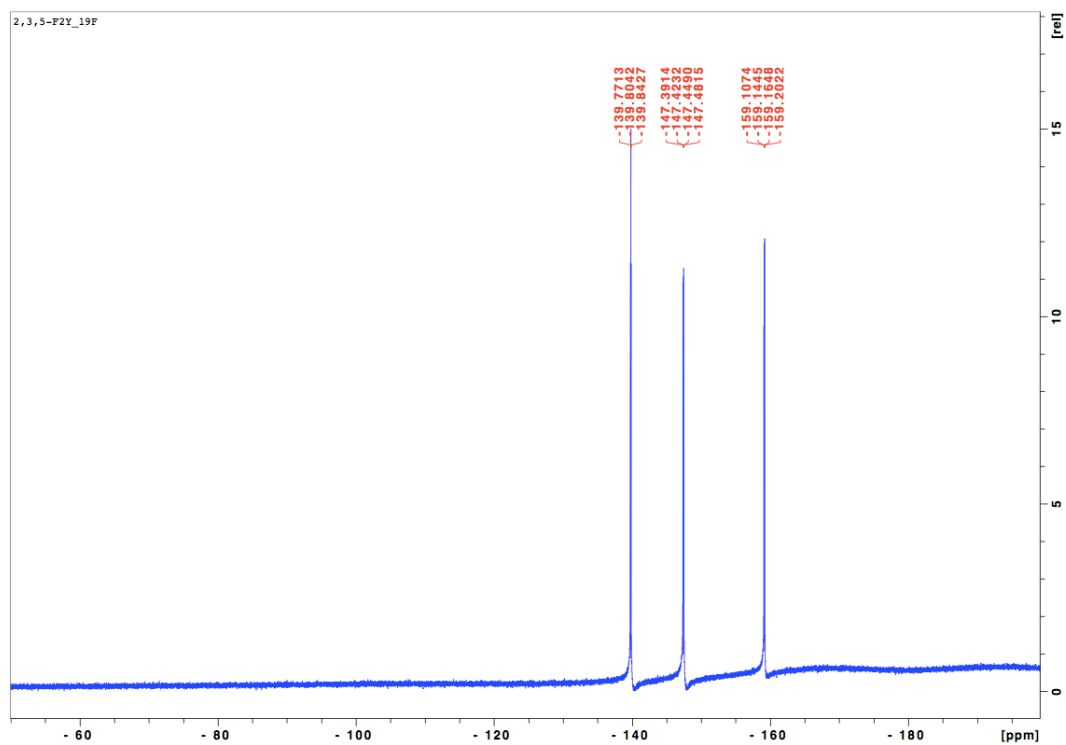
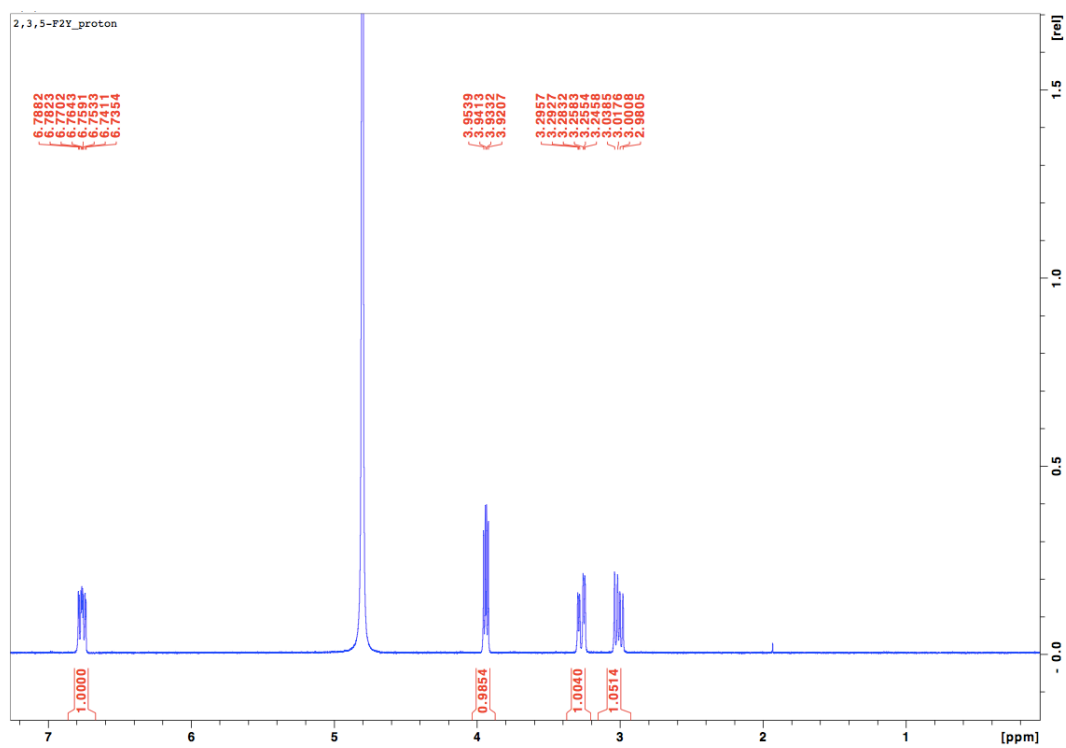


Figure S3. UV-Vis spectrum of AppA_{BLUF(Y56F)} in the dark state showing the protein/flavin peak at 270 nm and flavin peak at 446 nm.

Chromophore content was determined by taking the ratio of the absorbance at 270 nm and 446 nm. In the representative spectrum below the ratio is 4.2 (1.94/0.46).

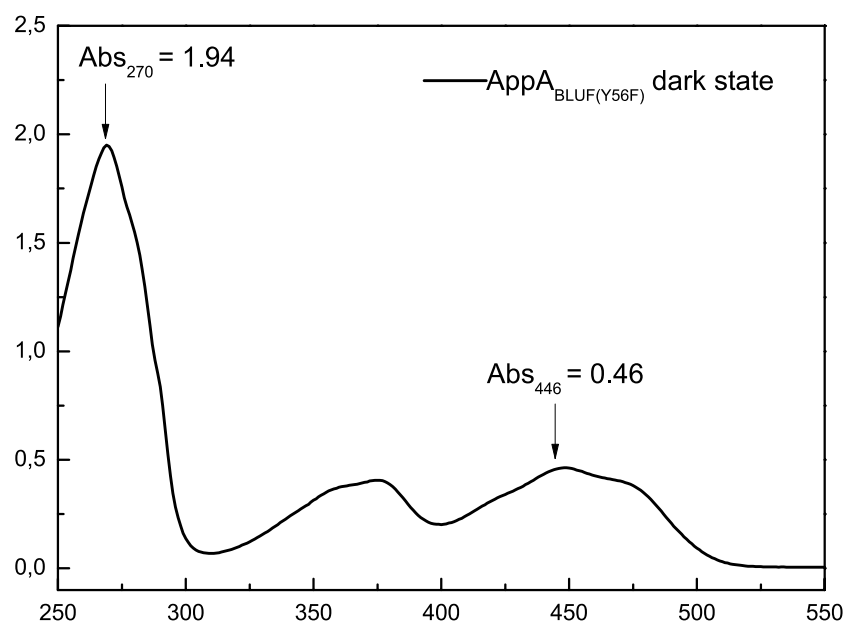


Figure S4. MALDI mass spectra showing 2-FY incorporation in AppA_{BLUF}(Y56F).

The sample was digested with trypsin and treated with iodoacetamide to alkylate the cysteine residues. The calculated mass $[M+H]^+$ of the peptide containing 2-FY (GSHMLEADVMTGSDLVSCCYR) is 2507. The calculated mass of the native peptide containing the natural tyrosine is 2489. The area under the peak was used to calculate the percentage of a non-labeled protein in the sample, and indicated >96% incorporation of 2-FY into AppA_{BLUF}(Y56F).

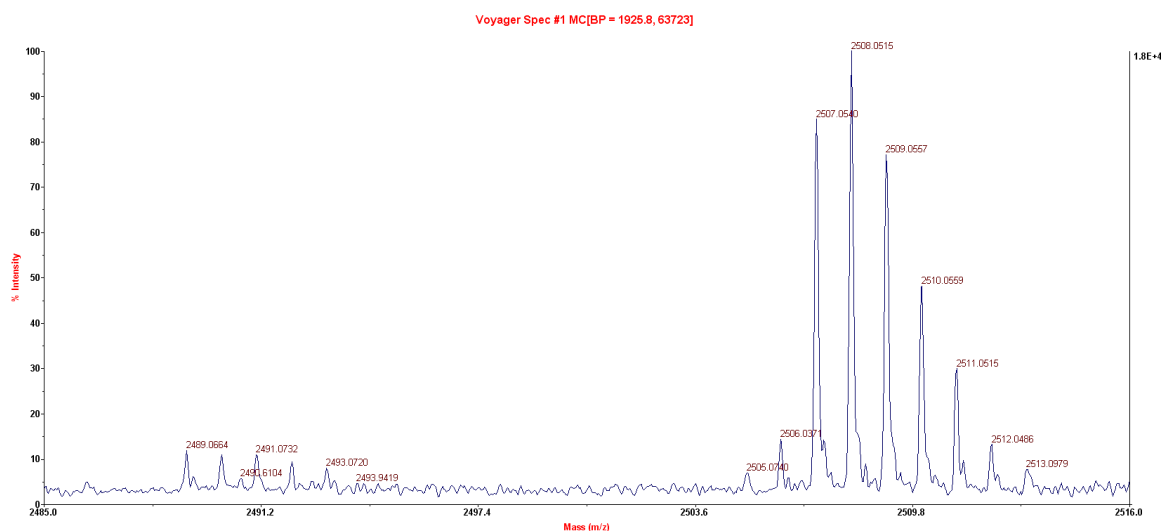


Figure S5. MALDI mass spectra showing incorporation of the 3FY, 2,3-F₂Y, 3,5-F₂Y and 2,3,5-F₃Y fluorotyrosines into AppA_{BLUF}(Y56F).

All samples were digested with trypsin and treated with iodoacetamide to alkylate the cysteine residues. The native peptide MQHDLEADVTMTGSDLVSCCYR has a calculated mass $[M+H]^+$ of 2588. However, the native peptide could only be detected in AppA_{BLUF}(Y56F) 3-FY21 at ~1% the intensity of the peptide arising from the labeled sample. Thus in all samples the incorporation of the fluorotyrosine is $\geq 99\%$.

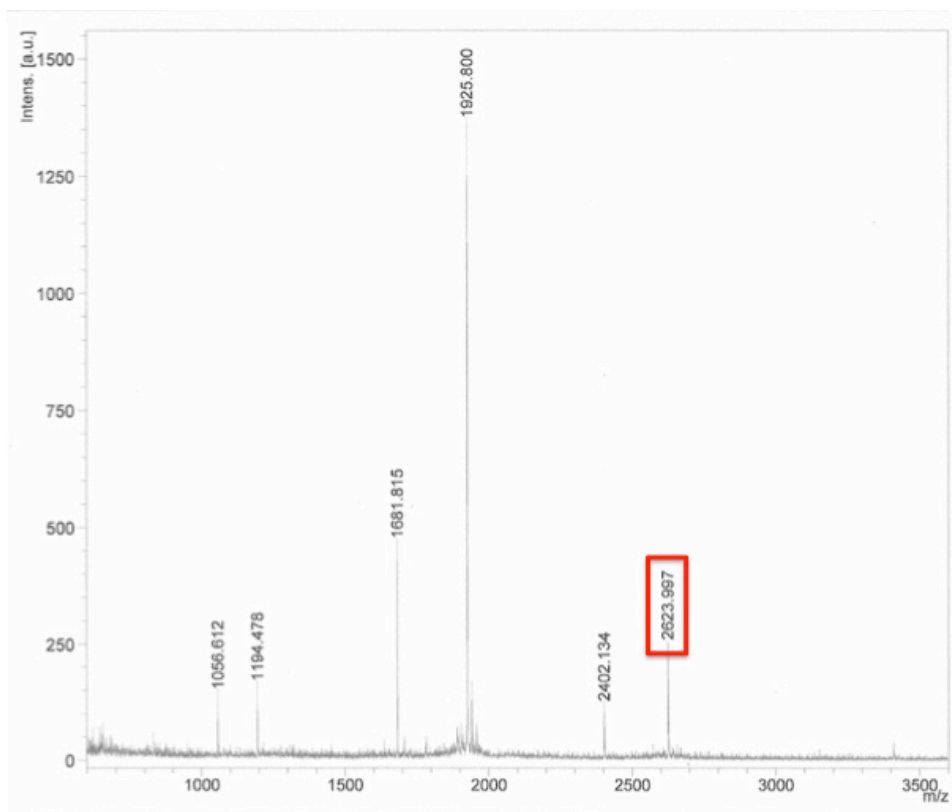
A) AppA_{BLUF}(Y56F) 3-FY21:

The calculated mass $[M+H]^+$ of the 3-FY peptide is 2606.



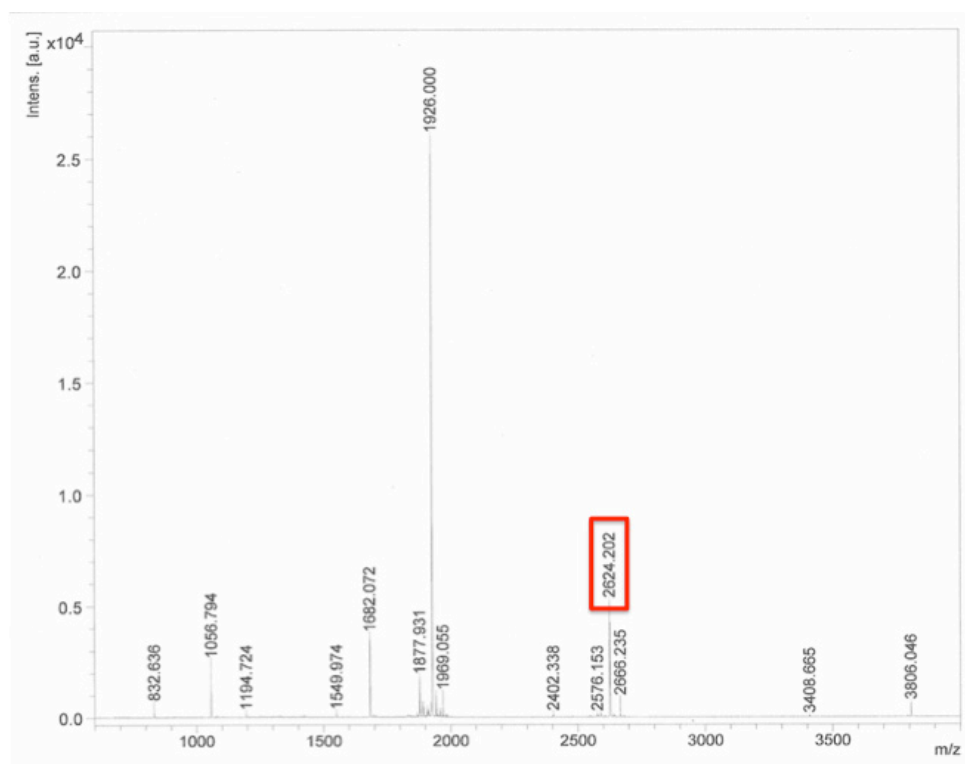
B) AppA_{BLUF}(Y56F) 2,3-F₂Y21

The calculated mass $[M+H]^+$ of the 2,3-F₂Y peptide is 2624.



C) AppABLUF(Y56F) 3,5-F₂Y21

The calculated mass $[M+H]^+$ of the 3,5-F₂Y peptide is 2624.



D) AppAB_{BLUF}(Y56F) 2,3,5-F₃Y21

The calculated mass $[M+H]^+$ of the 2,3,5-F₃Y peptide is 2642.

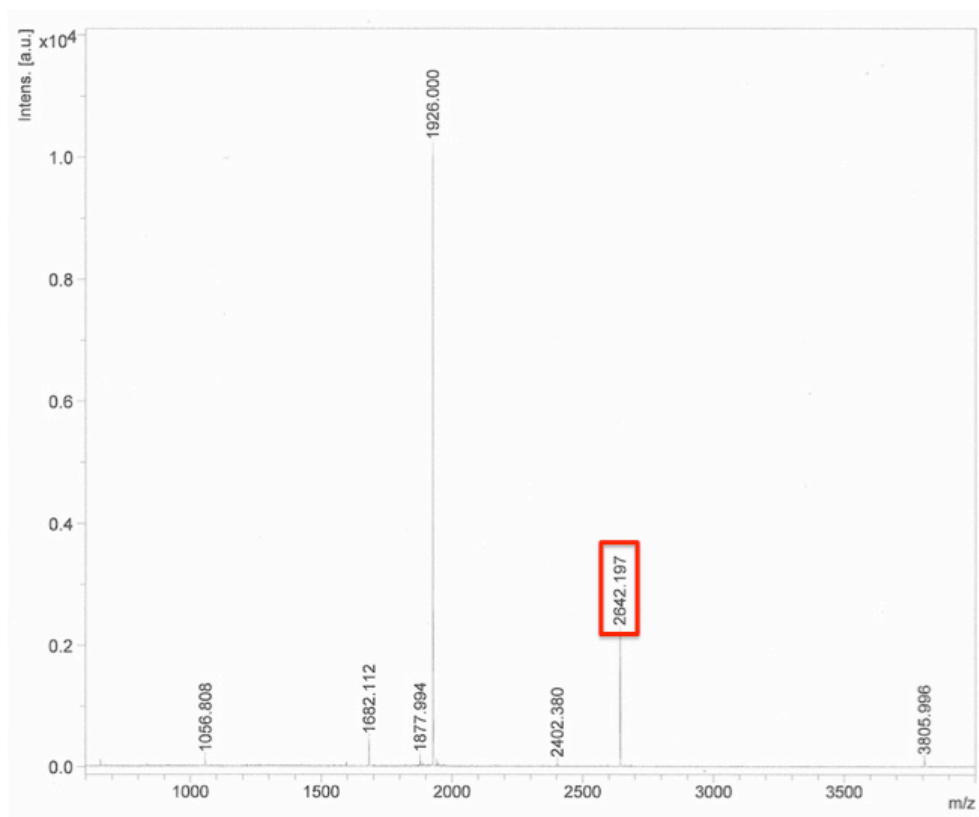


Figure S6. UV-Vis spectra recorded using the Ocean Optics USB2000+ spectrometer.

A: Spectra of AppA_{BLUF} taken over a 2 hr period during continuous illumination with the white light source used to record the data. B: Absorbance at 450 nm recorded during continuous illumination with the white light source. A small increase in the amplitude is observed at 450 nm over the time course of irradiation. This is related to an increase in the background. Importantly these data show that the white light source does not result in formation of the light state or of the sample.

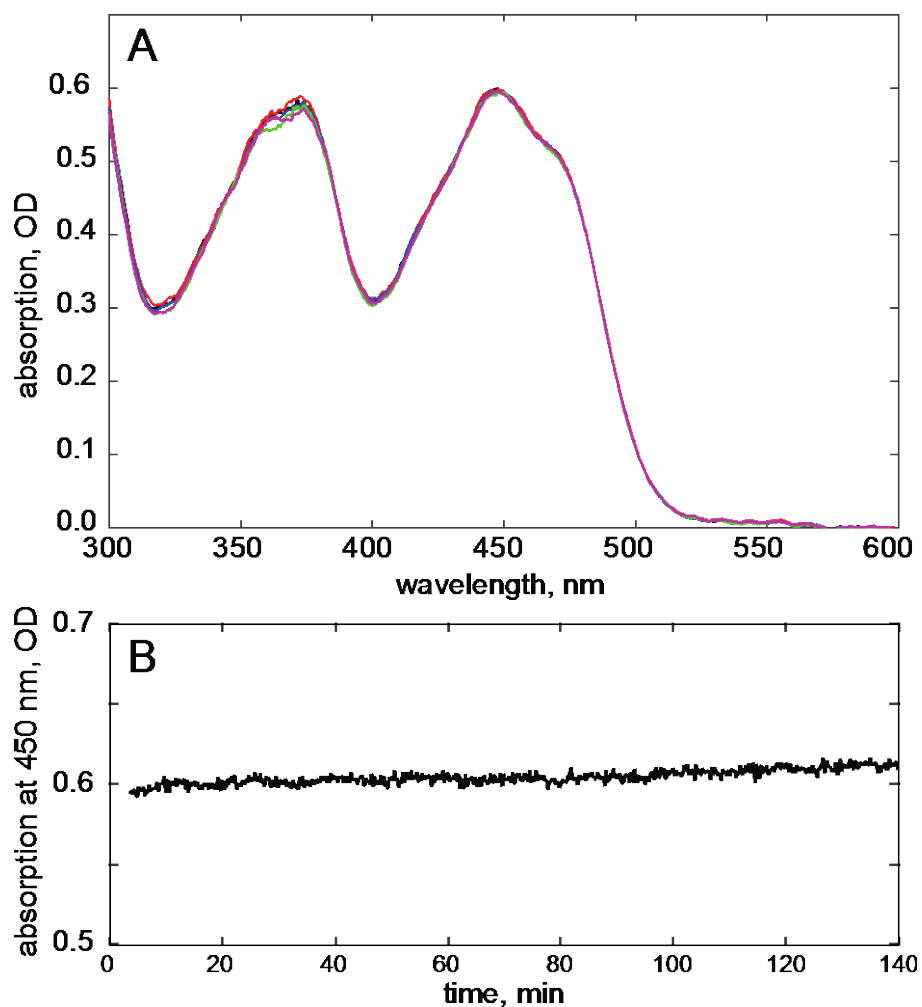


Figure S7. Comparison of the photoactivation portion of the photocycle of App^{ABLUF} and App^{ABLUF(Y56F)}. In the EADS plot solid lines show wild-type App^{ABLUF} and dashed lines show App^{ABLUF(Y56F)}. The spectral features and kinetic parameters confirmed that the Y56F mutation did not introduce changes to the structure nor the photoactivation of App^{ABLUF}.

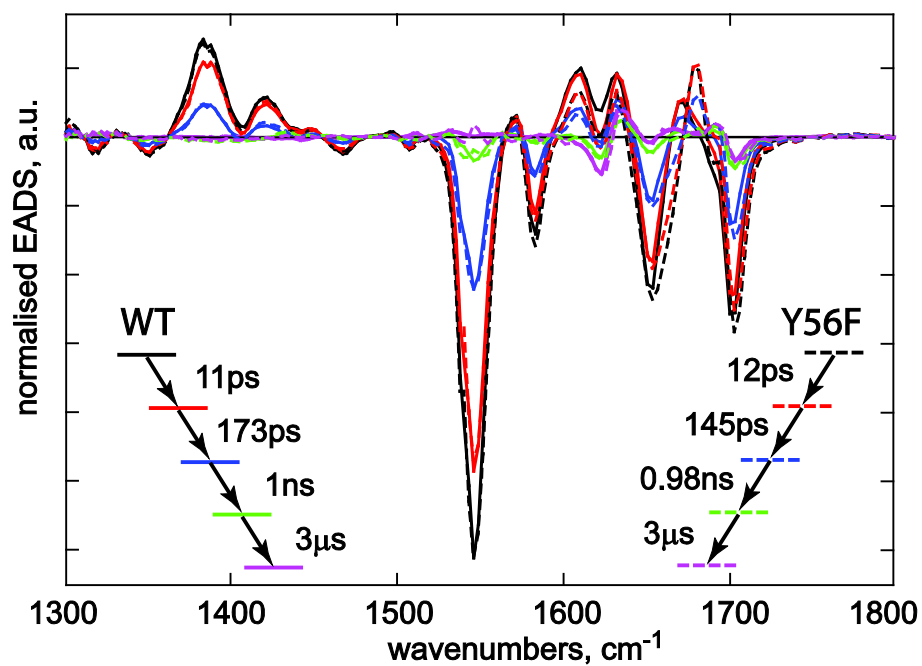


Figure S8. TRMPS dataset of AppA_{BLUF} between 50 μs and 200 μs . These spectra support the observation that no further spectral evolution is observed on that time scale.

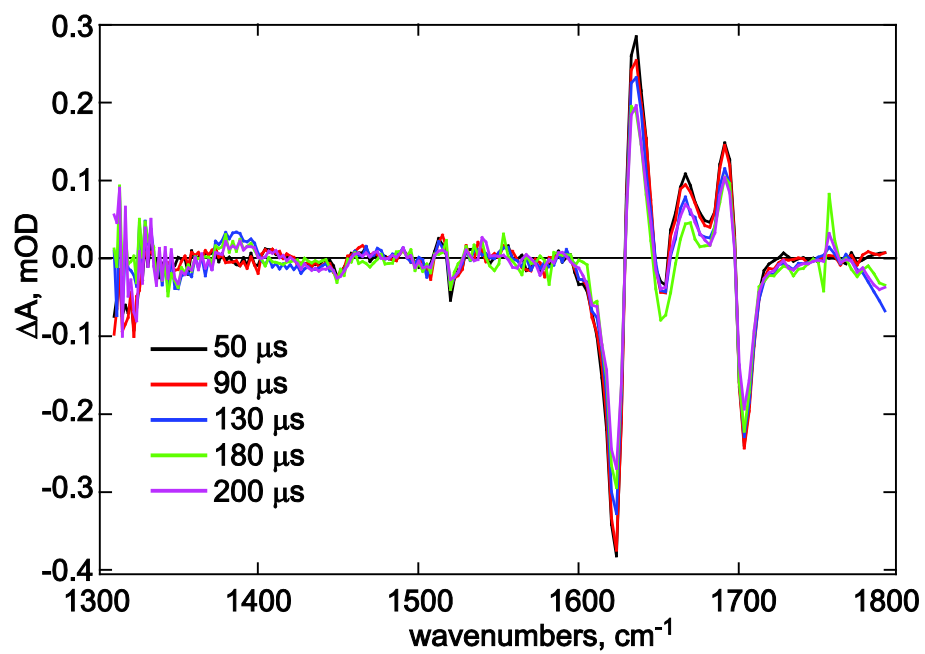


Figure S9. Complete TRMPS dataset of AppA_{BLUF}, AppA_{BLUF}(Y56F), and the FY21 substituted AppA_{BLUF}(Y56F). Left column shows the EADS spectra of all samples with estimated time constants and the right column shows the evolution of the experimental IR spectra as the protein is forming the light signaling state.

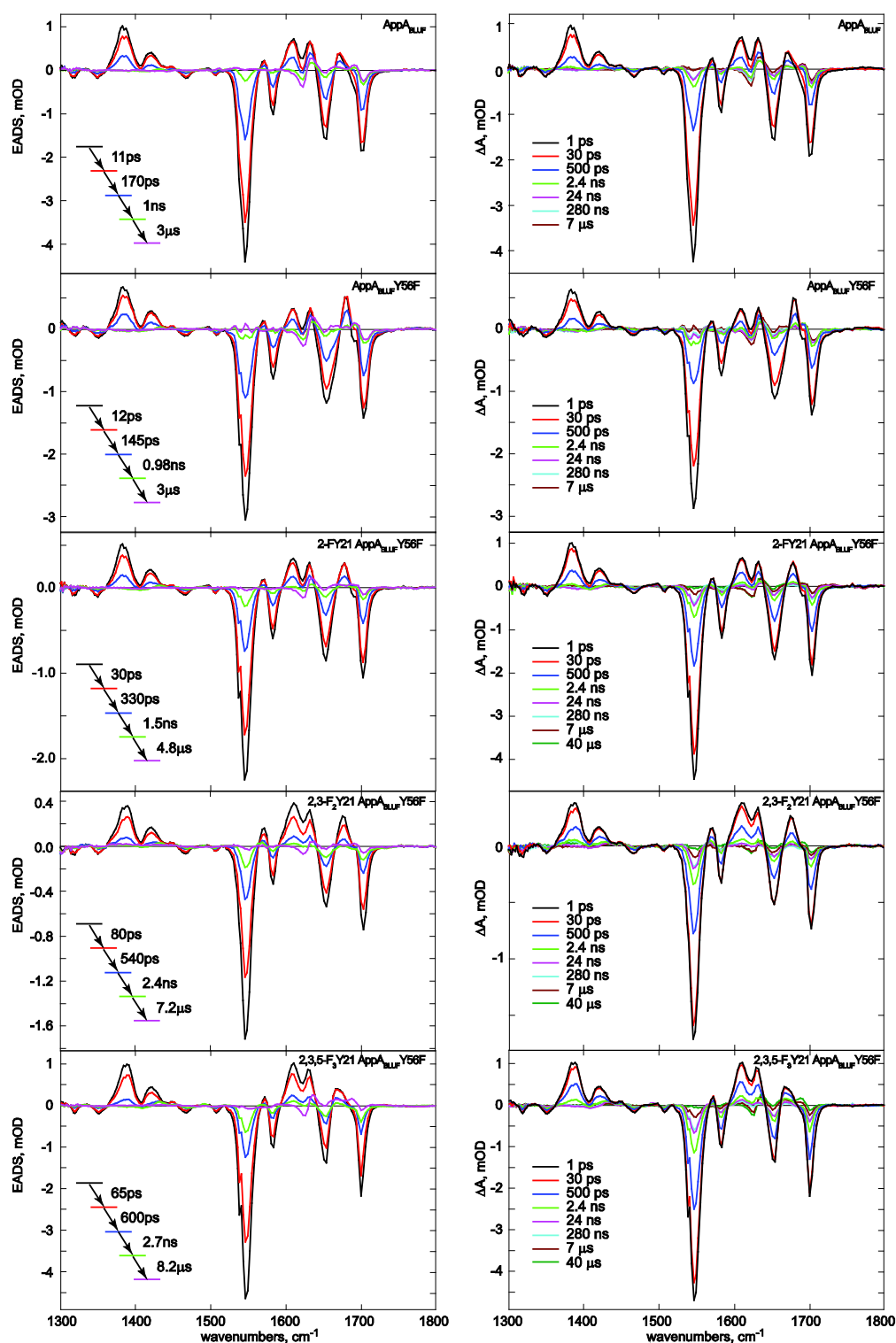


Figure S10. Rapid-scan FTIR difference spectra of AppA_{BLUF(Y56F)} 2,3-F₂Y21. The recovery of the dark state was determined by analysis of the light minus dark state spectra. The estimated rate constant was determined from global analysis of the 1590 – 1710 cm⁻¹ spectral region.

