

Understanding the Selectivity of a Multi-Channel Fluorescent Probe for Peroxynitrite Over Hypochlorite

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Preparation of stock solutions of various ROS/RNS.

Hypochlorite (ClO^-) stock solution (50 mM in H_2O) was prepared by diluting commercial NaClO solution in deionized water (pH = 10). The solution concentration was determined by UV-Vis spectroscopy ($\epsilon_{293\text{ nm}} = 350\text{ cm}^{-1}\text{M}^{-1}$).

Peroxynitrite (OONO^-) stock solution (37 mM in H_2O at pH > 10) was prepared following procedure by Pryor et al. (Anal. Biochem. 1996, 236, 242-249). The solution concentration was determined by UV-Vis spectroscopy ($\epsilon_{302\text{ nm}} = 1,670\text{ cm}^{-1}\text{M}^{-1}$).

Nitric Oxide (NO) stock solution (1.9 mM in H_2O) was generated by bubbling a stream of nitric oxide gas into deoxygenated deionized H_2O for 15 min. The nitric oxide gas was produced by dropping dilute H_2SO_4 (2 M) solution onto NaNO_2 solid and passed through 5 wt% NaOH solution to remove NO_2 .

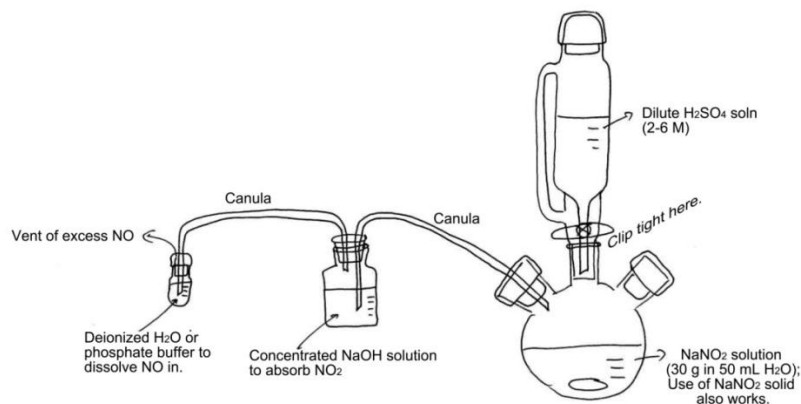


Figure S1. Setup for preparation of a nitric oxide stock solution. *Note: 1) the entire system should be thoroughly deoxygenated with inert gas, i.e. N_2 or Ar ; 2) H_2SO_4 solution should be dropped into a vigorously stirring NaNO_2 solution (or onto NaNO_2 solid) very slowly at the very beginning to avoid sudden build-up of internal gas pressure; 3) NaOH solution to wash off NO_2 can be replaced by NaOH pellets; 4) the system should be set up in a fume hood with good ventilation.*

Superoxide ($\text{O}_2^{\cdot-}$) stock solution (50 mM in DMF) was prepared by dissolving solid KO_2 in anhydrous DMF. Presence of H_2O in DMF leads to disproportionation of $\text{O}_2^{\cdot-}$.

Hydroxyl radical ($\text{HO}^\cdot$) was generated *in situ* by addition of an aliquot of $\text{Fe}(\text{ClO})_4$ solution (200 mM in H_2O) into a solution containing excess H_2O_2 .

Singlet oxygen ($^1\text{O}_2$) was generated *in situ* by addition of an aliquot of hypochlorite stock into a solution containing excess H_2O_2 .

Hydrogen peroxide (H_2O_2) stock solution (50 mM in H_2O) was prepared by dilution of commercial 30% H_2O_2 solution in deionized H_2O .

Spectral studies

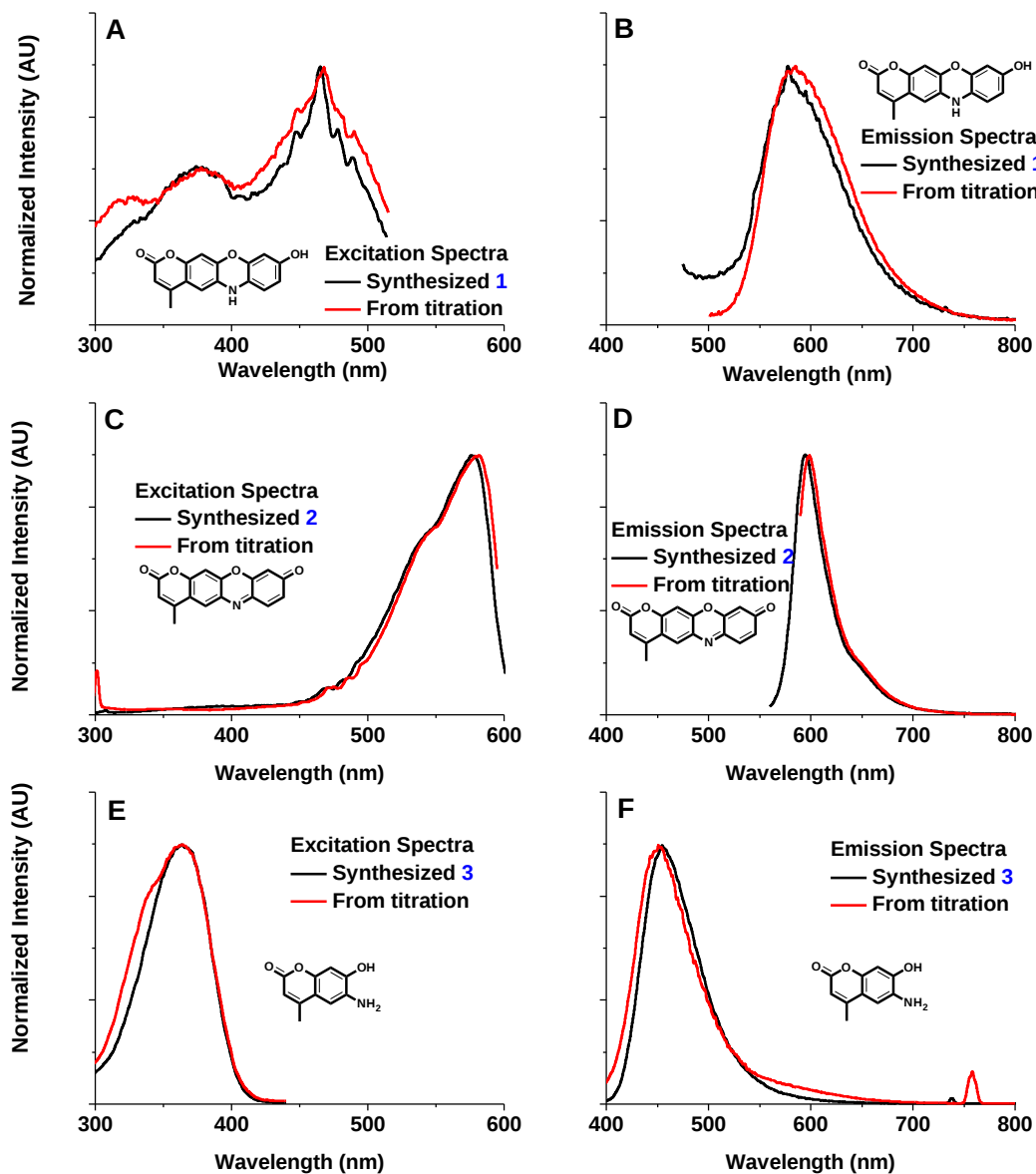


Figure S2. Comparison of fluorescence spectra of independently synthesized **1**, **2** and **3** with those extrapolated from fluorescence titration of probe **PN600** by hypochlorite or peroxynitrite, as a verification of their formation during titration.

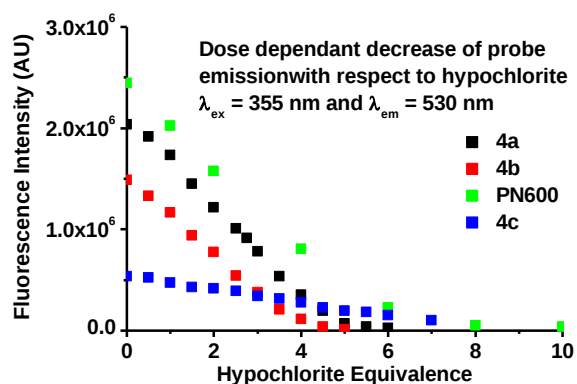


Figure S3. Dose dependant decrease of the probe emission upon addition of hypochlorite. *Note: it took similar amount of hypochlorite (ca. 5-7 equiv.) to consume probe 4a, 4b and PN600. In comparison, consumption of 4c require significantly higher dose of hypochlorite. An explanation to this observation is that the oxidation product of probe 4a, 4b and PN600 are not reactive toward hypochlorite and the oxidation production of 4c is also reactive toward hypochlorite and therefore competes with 4c.*

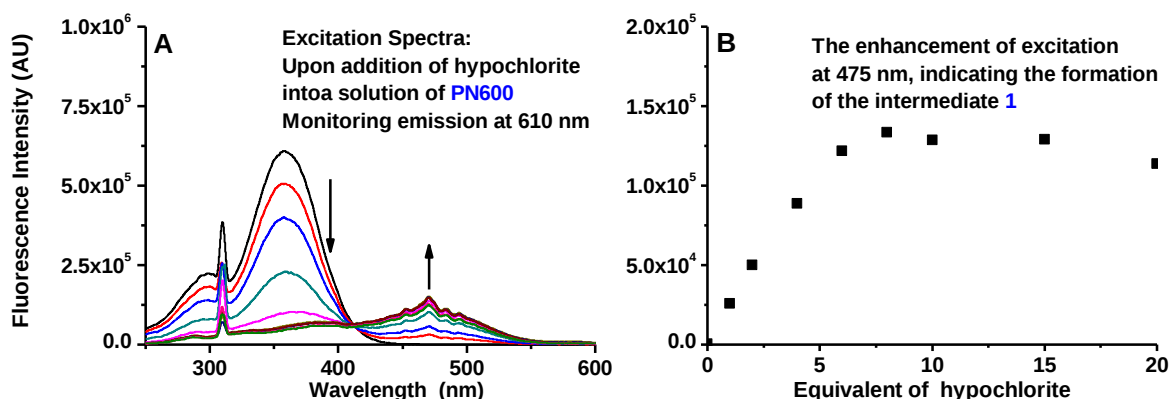


Figure S4. Excitation spectra monitoring emission at 610 nm, upon addition of an aliquot of hypochlorite into a solution of **PN600**, showing the decrease of excitation of **PN600** with a maximum at 350 nm and increase of excitation of **1** with a maximum at 467 nm as the dose of hypochlorite increases.

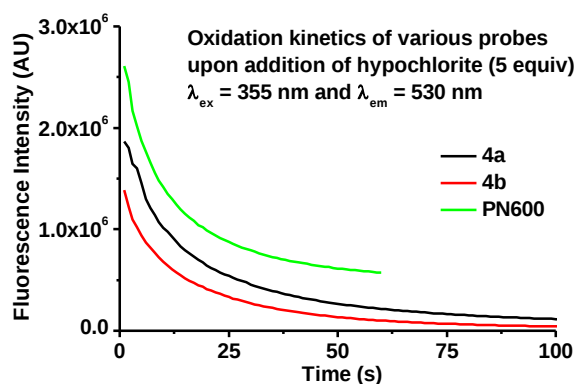


Figure S5. Oxidation kinetics of probe **4a**, **4b** and **PN600**, upon addition of an aliquot of hypochlorite (5 equiv.).

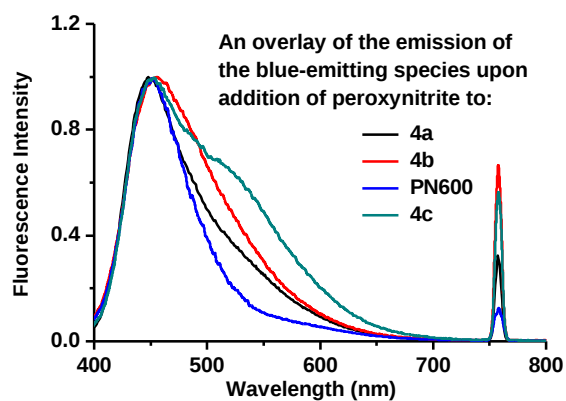


Figure S6. Overlay of the blue emissions (presumably from **3**) generated upon addition of an aliquot of peroxynitrite stock into a solution of probe **4a**, **4b**, **PN600** and **4c**, respectively. *Note: the presence of other species, which are also excitable by light of 355 nm and emit at longer wavelength than **3**, are responsible for the spectral difference in the range of 475 nm to 700 nm.*

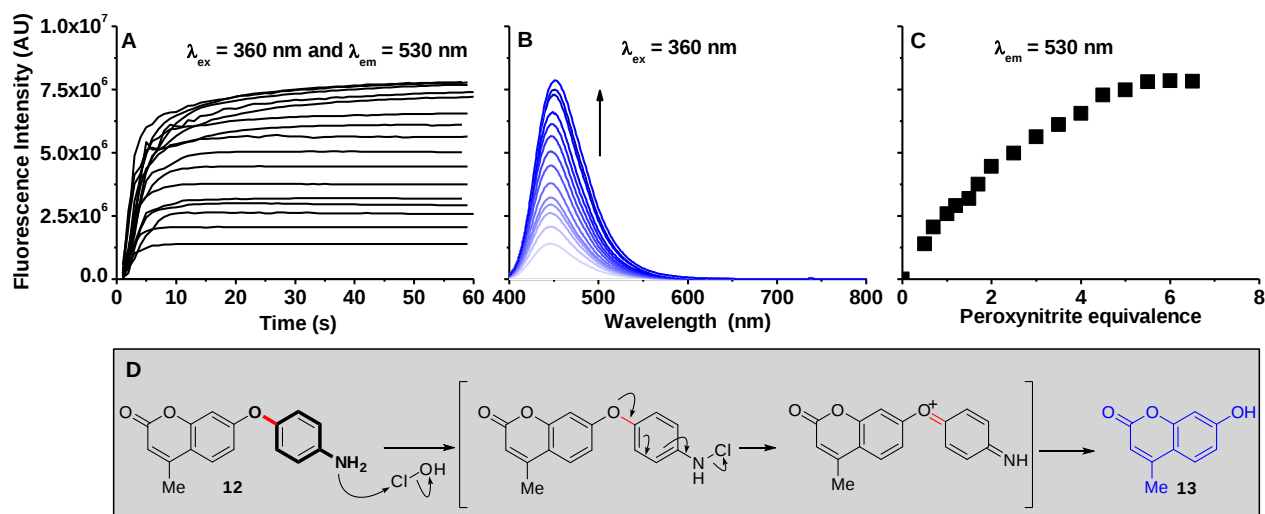


Figure S7. Oxidation of probe **12** by hypochlorite. A) Kinetics of the formation of **13**; B) emission spectra of a solution of probe **12**, upon addition of various equivalence of hypochlorite; C) dose dependent emission enhancement at 450 nm; D) a mechanistic explanation to the formation of **13** upon hypochlorite mediated oxidation of probe **12**.

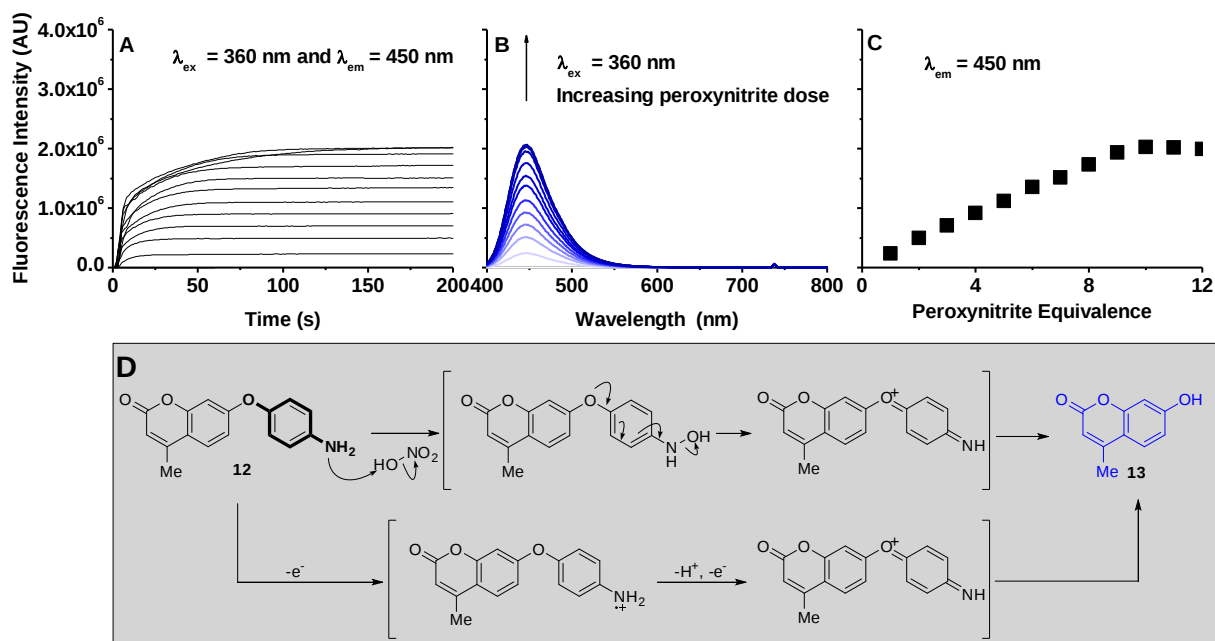


Figure S8. Oxidation of probe **12** by peroxynitrite. A) Kinetics of the formation of **13**; B) emission spectra of a solution of probe **12**, upon addition of various equivalence of peroxynitrite; C) dose dependent emission enhancement at 450 nm; D) a mechanistic explanation to the formation of **13** upon peroxynitrite mediated oxidation of probe **12**.

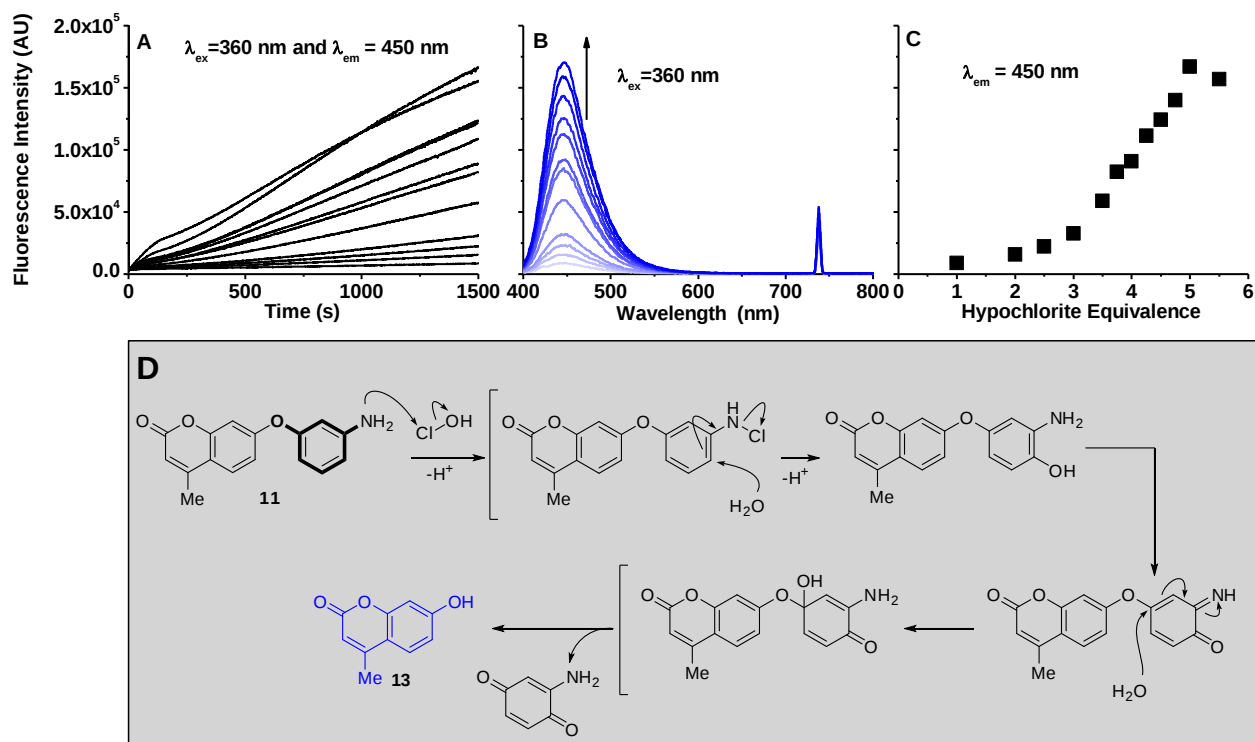


Figure S9. Oxidation of probe **11** by hypochlorite. A) Kinetics of the formation of **13**; B) emission spectra of a solution of probe **11**, upon addition of various equivalence of hypochlorite; C) dose dependent emission enhancement at 450 nm; D) a mechanistic explanation to the formation of **13** upon hypochlorite mediated oxidation of probe **11**. Note: probe **11** can be oxidized, but to a limited extent only and kinetics is much slower compared to peroxynitrite mediated oxidation of probe **11**.

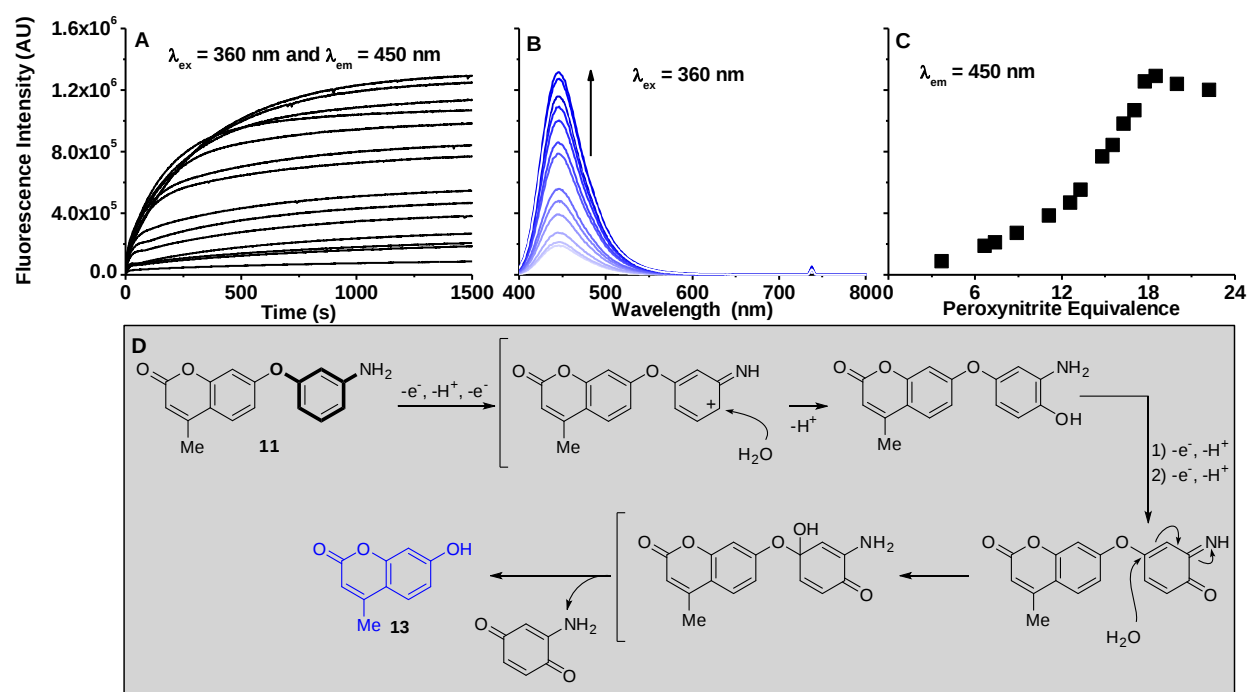


Figure S10. Oxidation of probe **11** by peroxynitrite. A) Kinetics of the formation of **13**; B) emission spectra of a solution of probe **11**, upon addition of various equivalence of peroxynitrite; C) dose dependent emission enhancement at 450 nm; D) a mechanistic explanation to the formation of **13** upon peroxynitrite mediated oxidation of probe **11**.

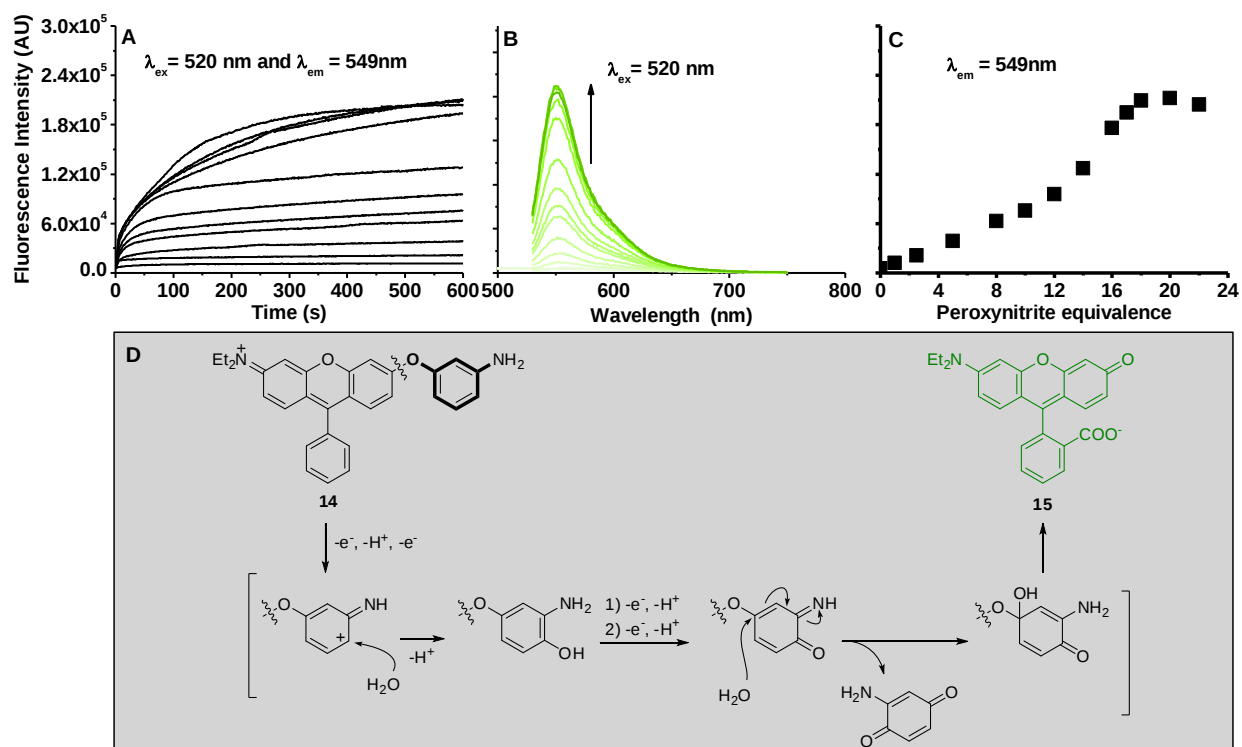


Figure S11. Oxidation of probe **14** by peroxynitrite. A) Kinetics of the formation of **15**; B) emission spectra of a solution of probe **14**, upon addition of various equivalence of peroxynitrite; C) dose dependent emission enhancement at 549 nm; D) a mechanistic explanation to the formation of **15** upon peroxynitrite mediated oxidation of probe **14**.

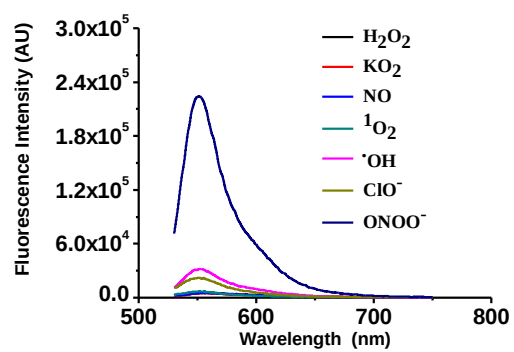


Figure S12. Selectivity study of probe **14** toward various reaction oxygen or nitrogen species.

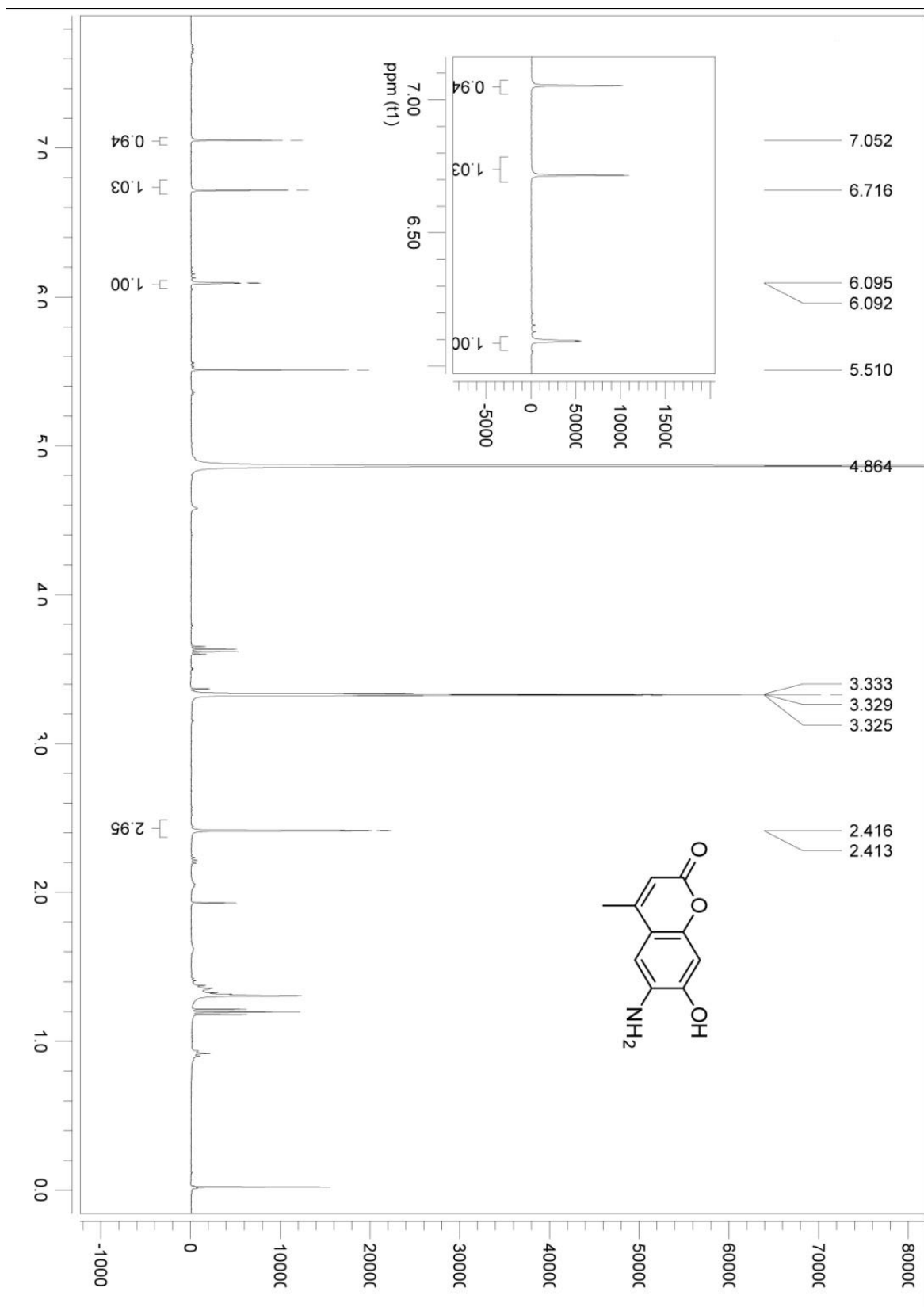


Figure S13. The ^1H -NMR spectrum of **3** in CD_3OD .

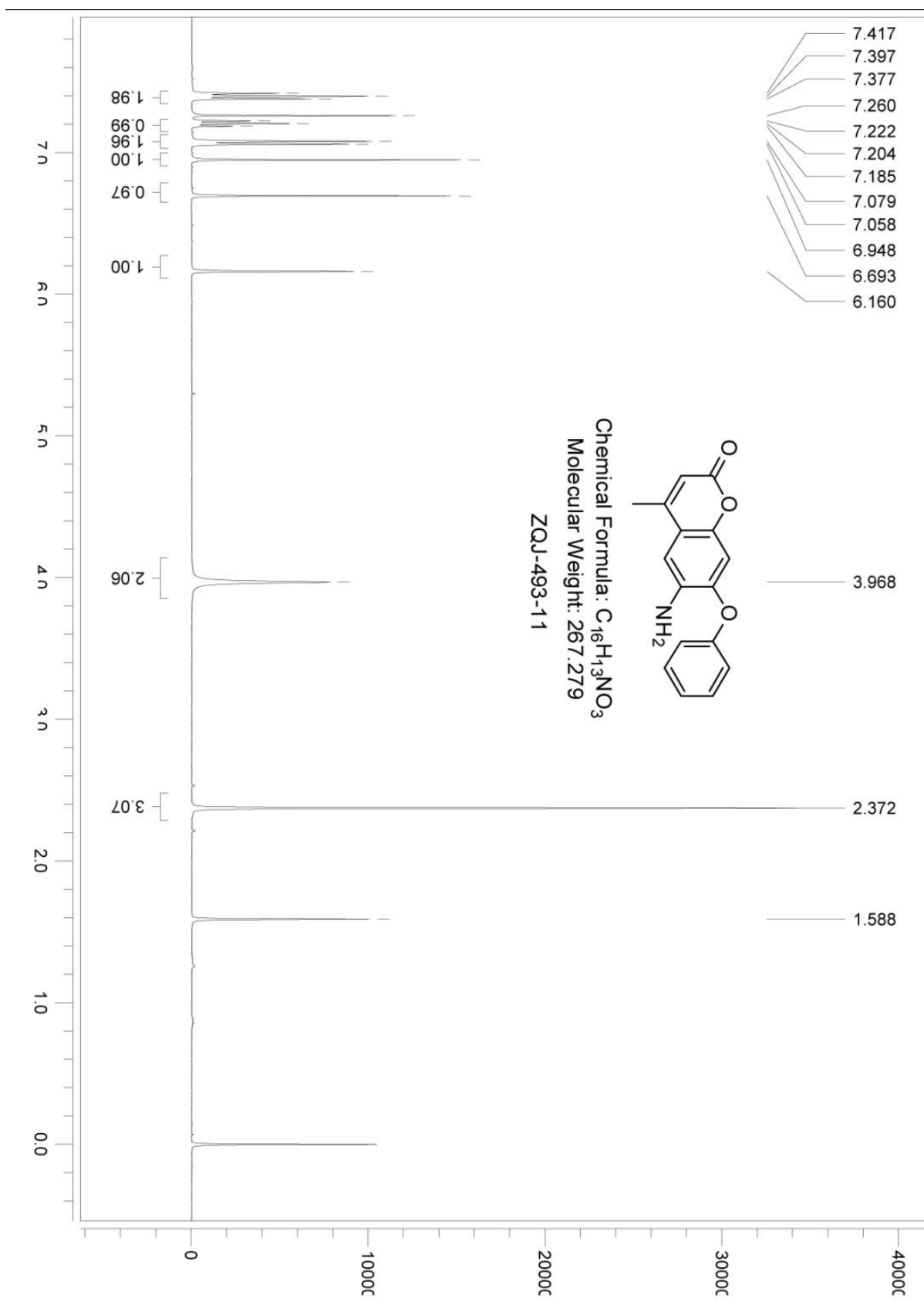


Figure S14. The ¹H-NMR spectrum of **4a** in CDCl₃.

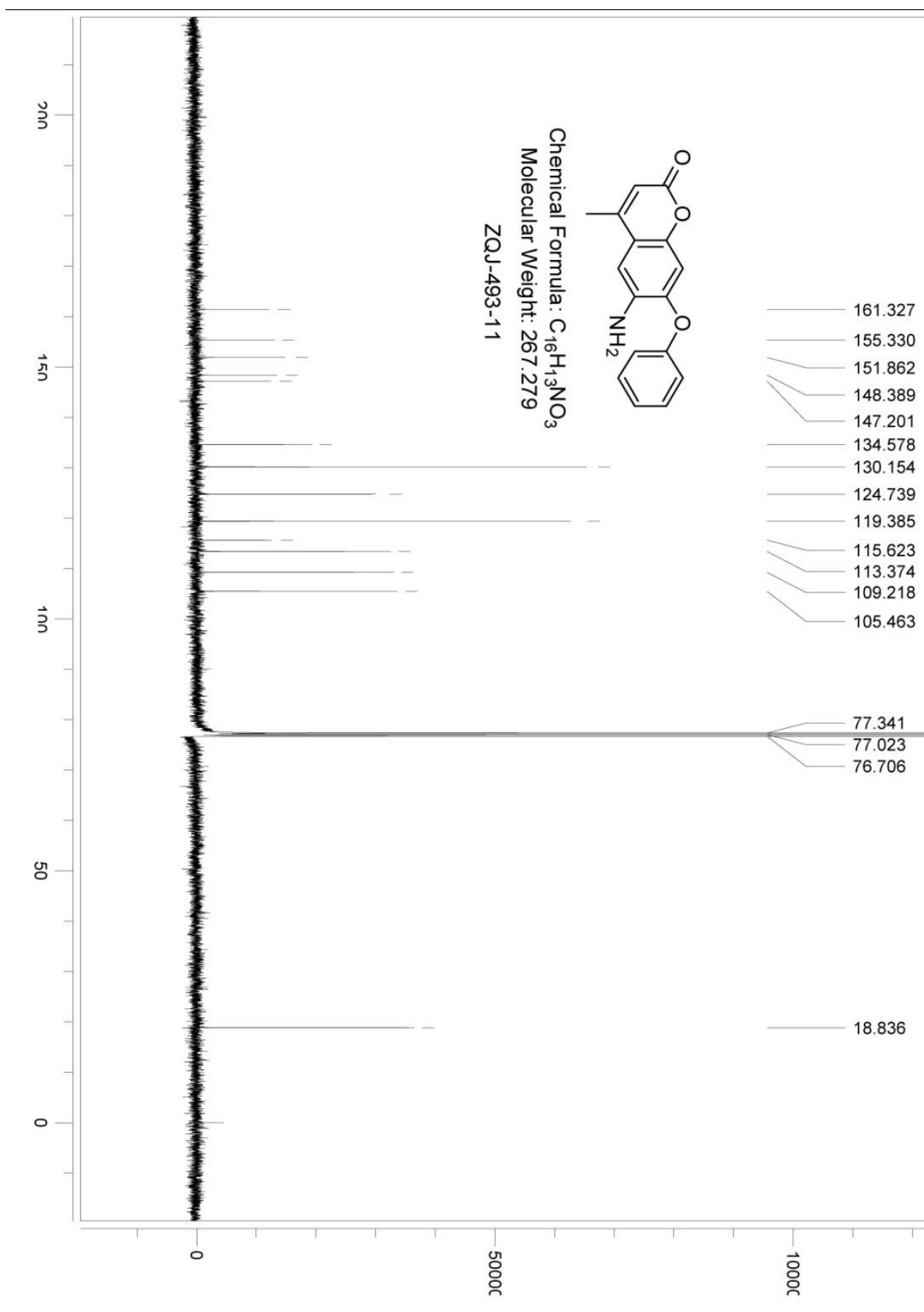


Figure S15. The ^{13}C -NMR spectrum of **4a** in $CDCl_3$.

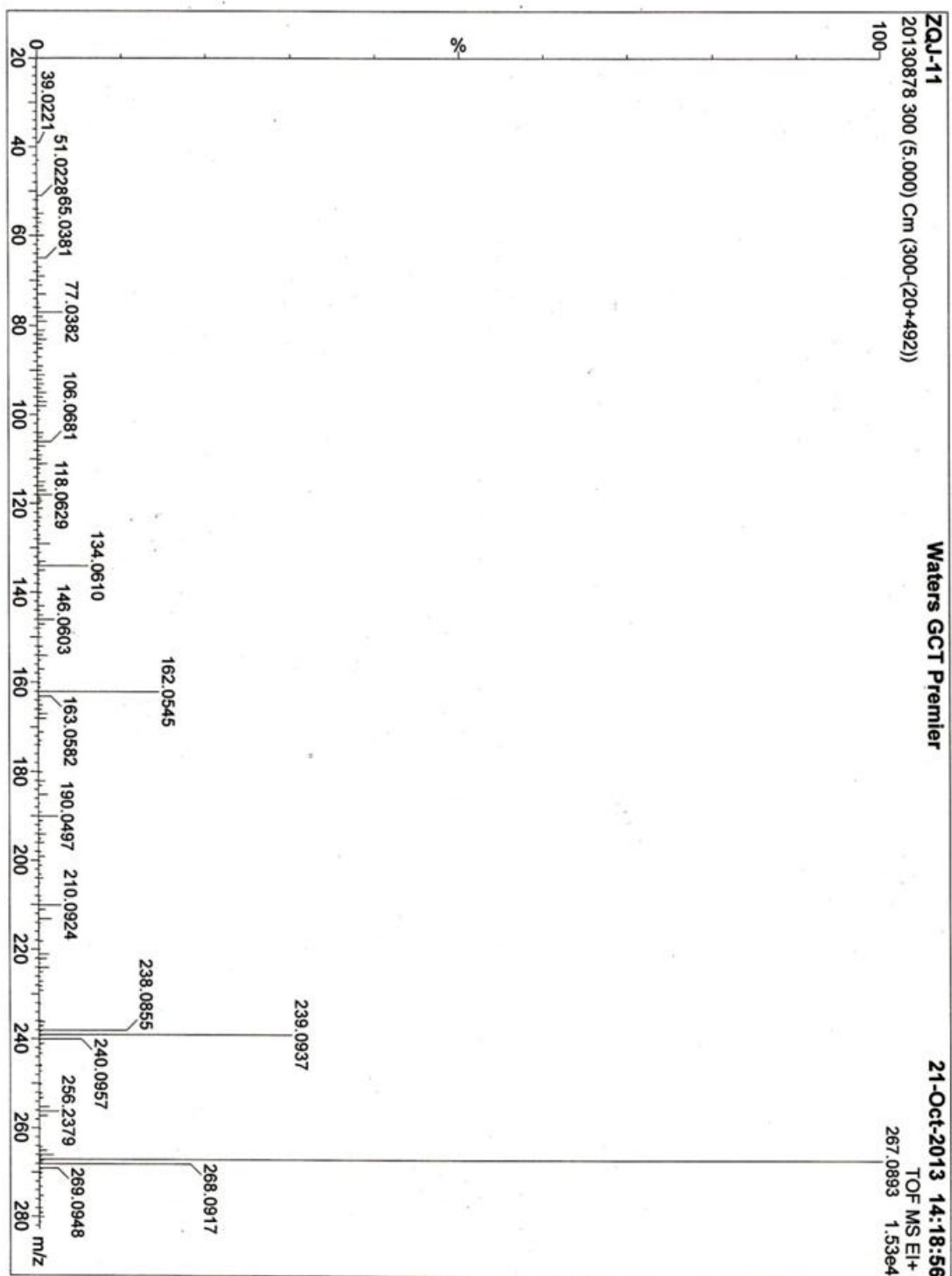


Figure S16. The HRMS spectrum of **4a**.

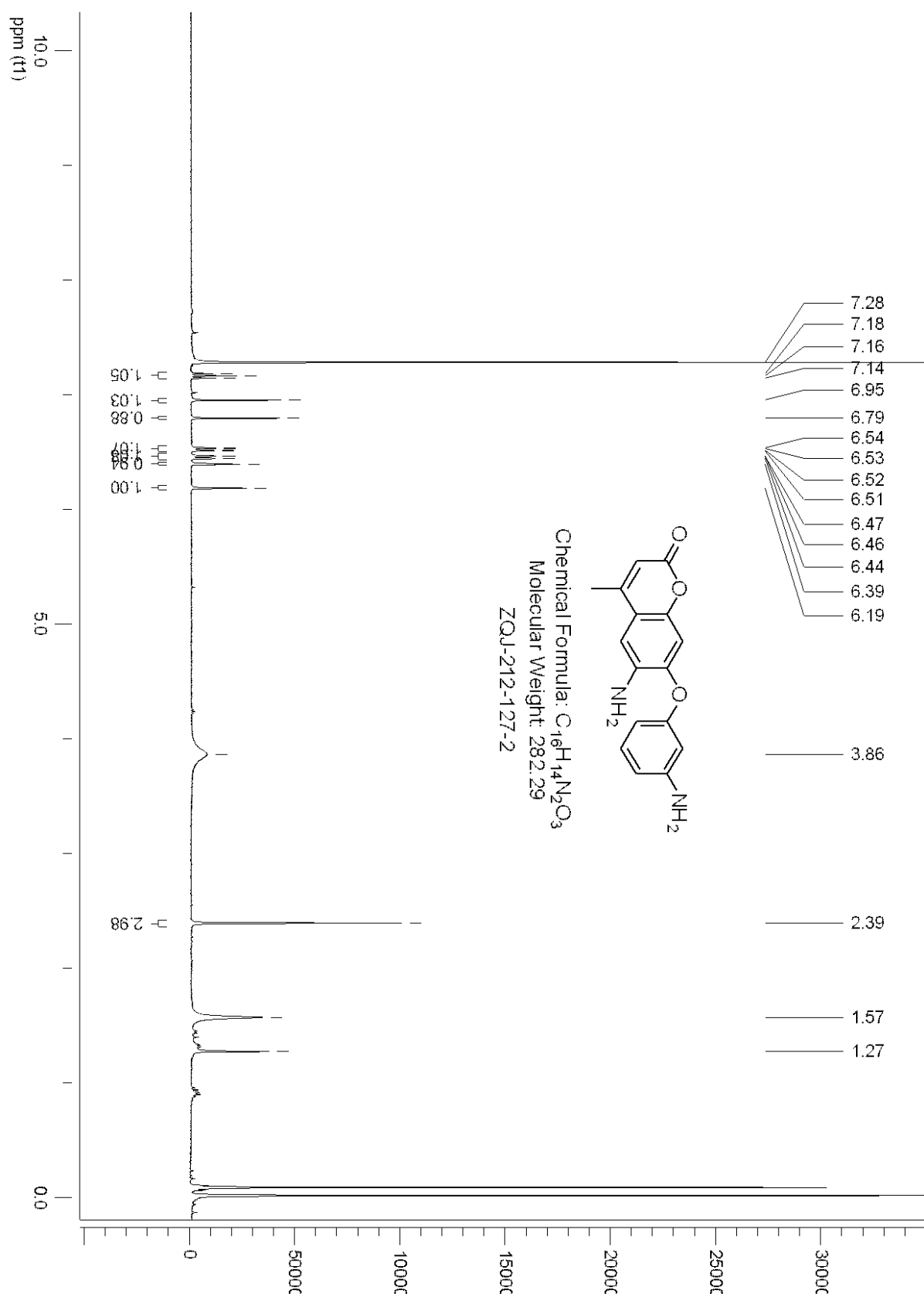


Figure S17. The ¹H-NMR spectrum of **4c** in CDCl₃.

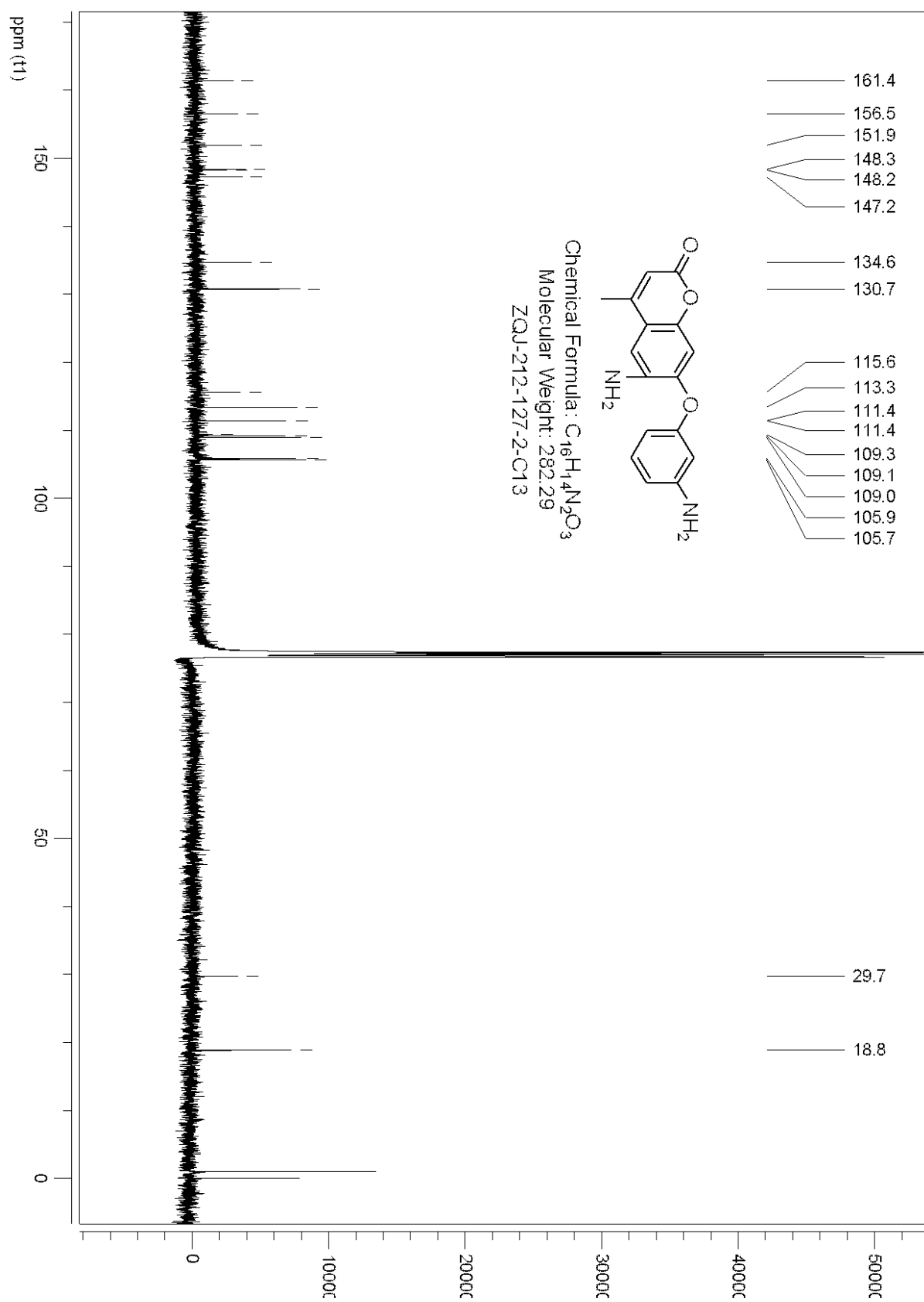


Figure S18. The ^{13}C -NMR spectrum of **4c** in $CDCl_3$.

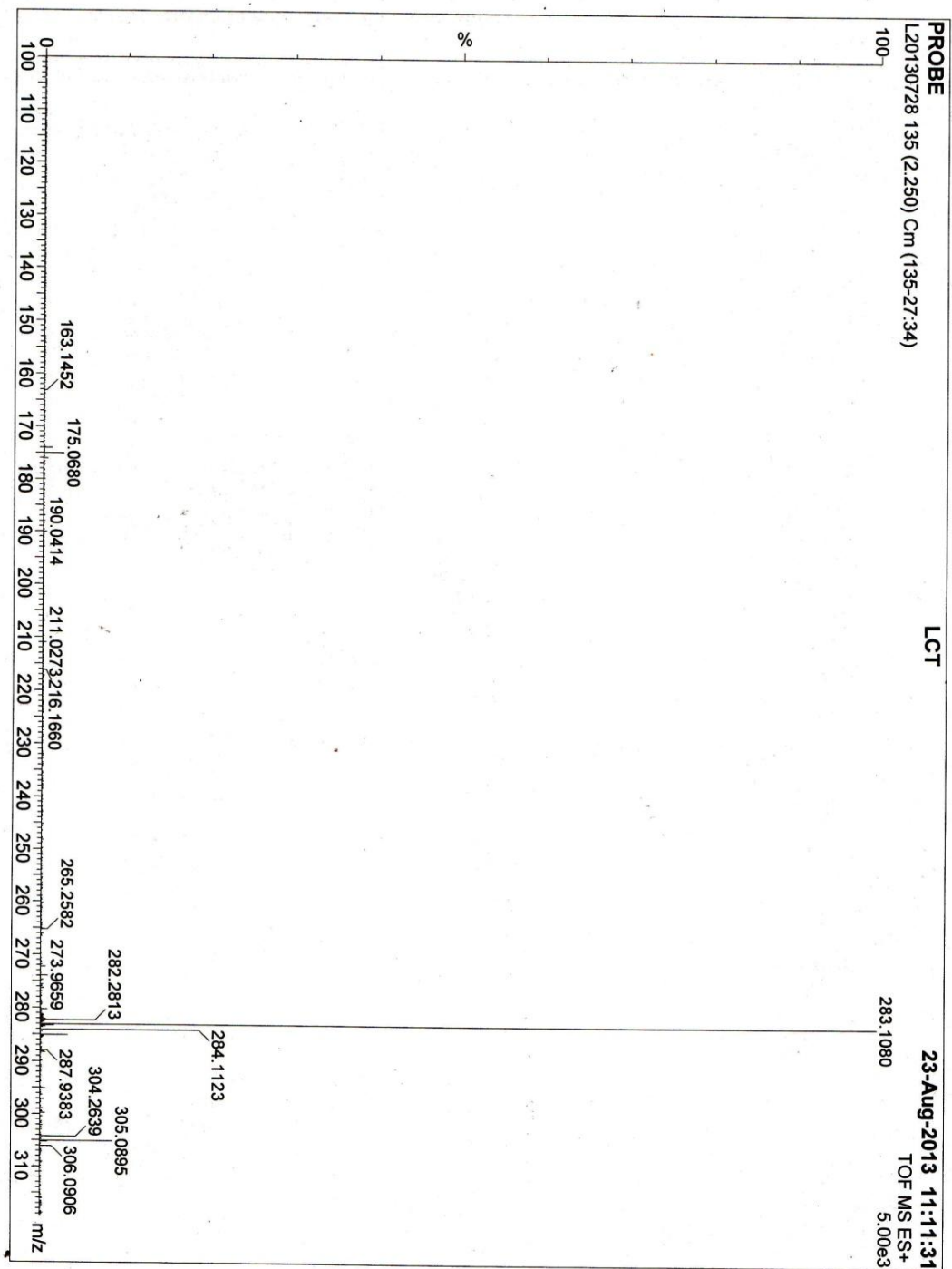


Figure S19. The HRMS spectrum of **4c**.

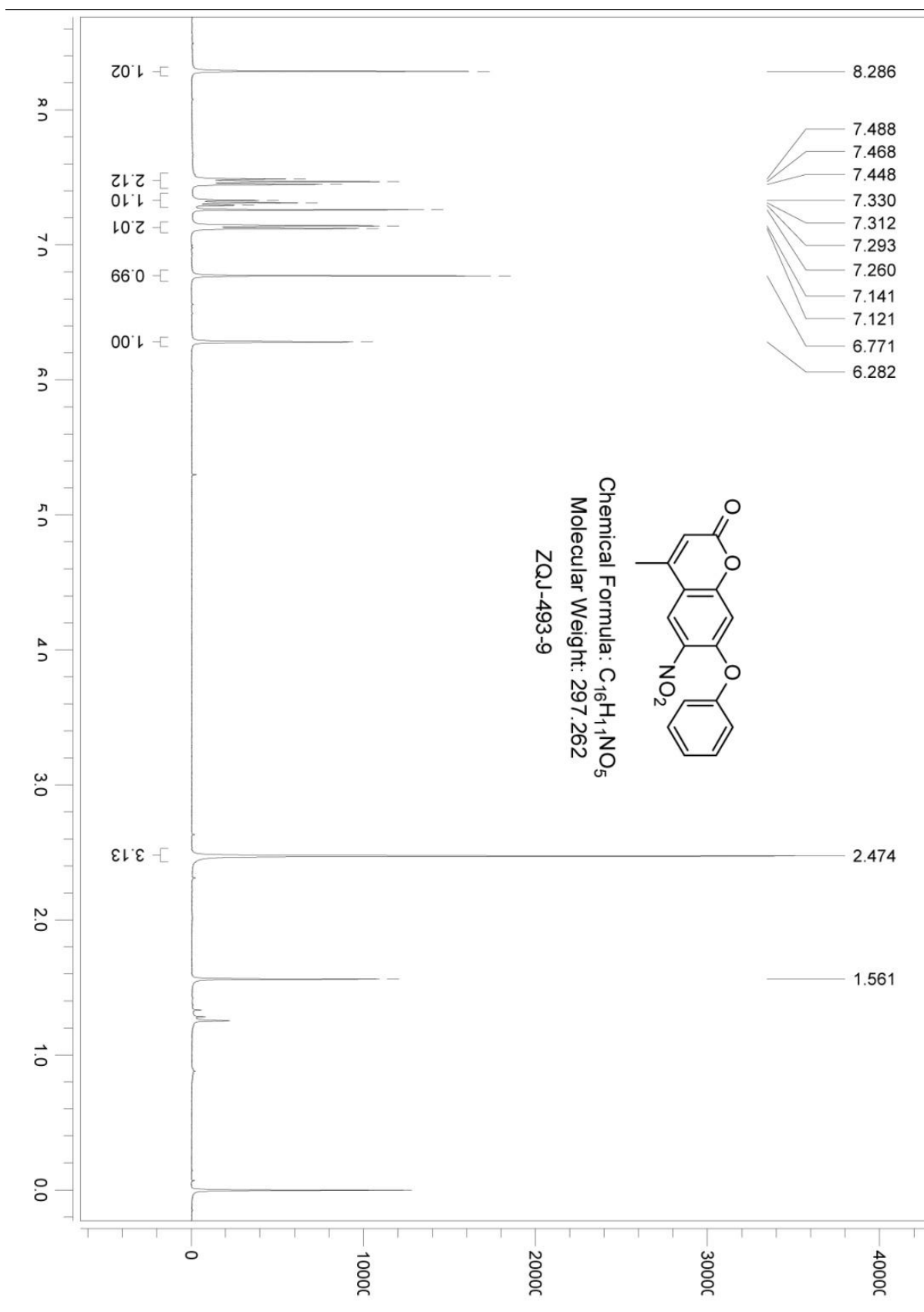


Figure S20. The ^1H -NMR spectrum of **7a** in CDCl_3 .

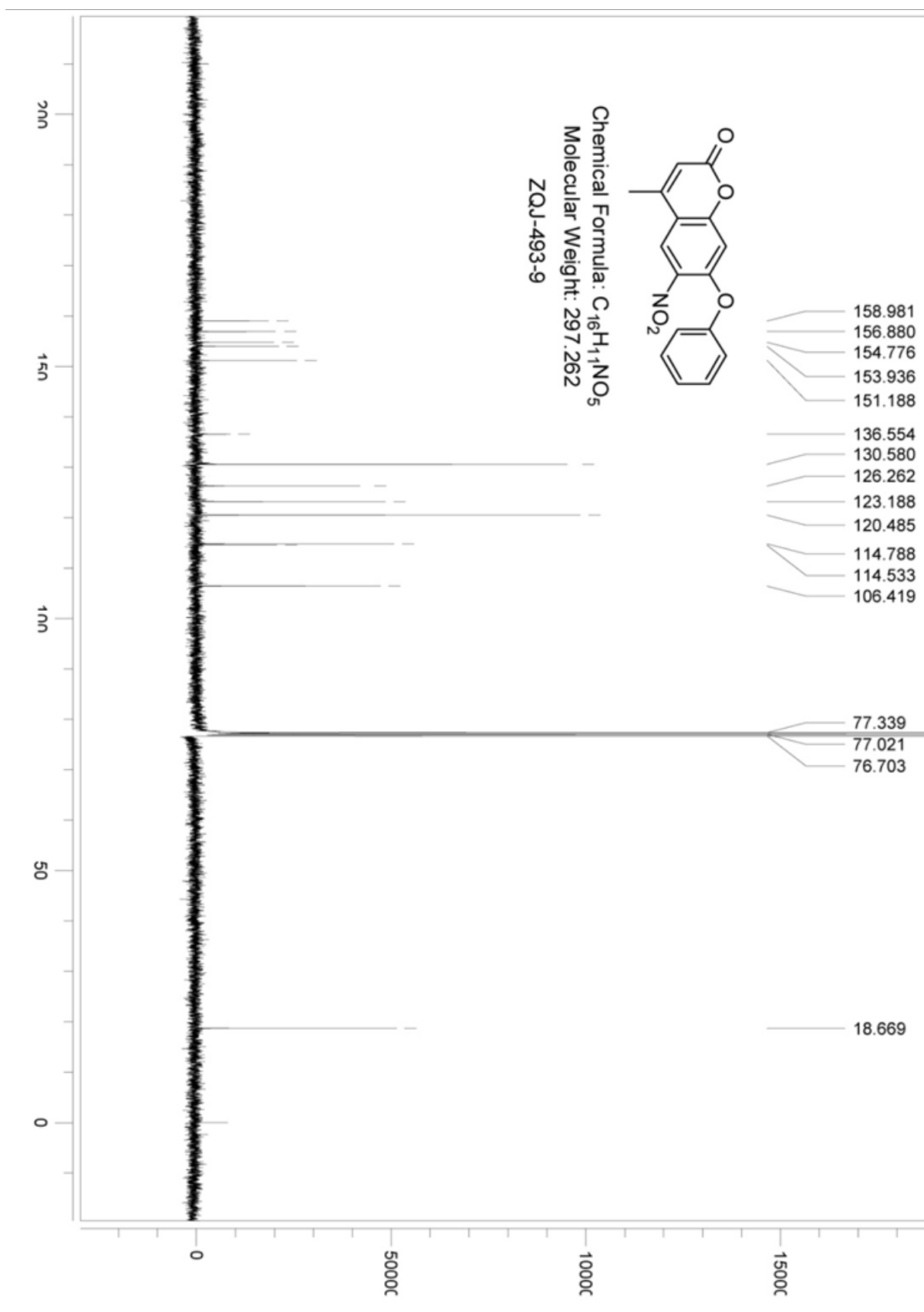


Figure S21. The ^{13}C -NMR spectrum of **7a** in CDCl₃.

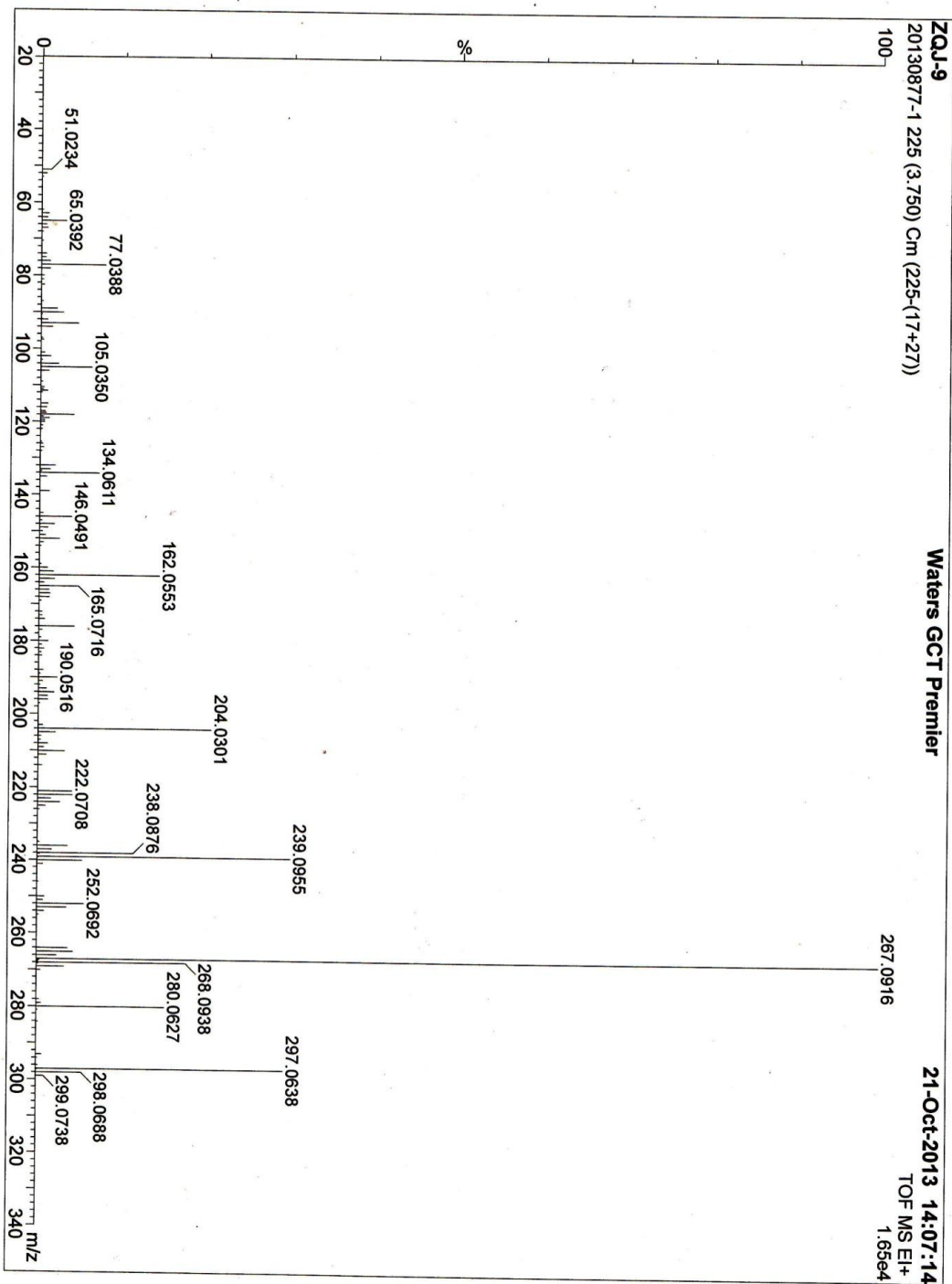


Figure S22. The HRMS spectrum of **7a**.

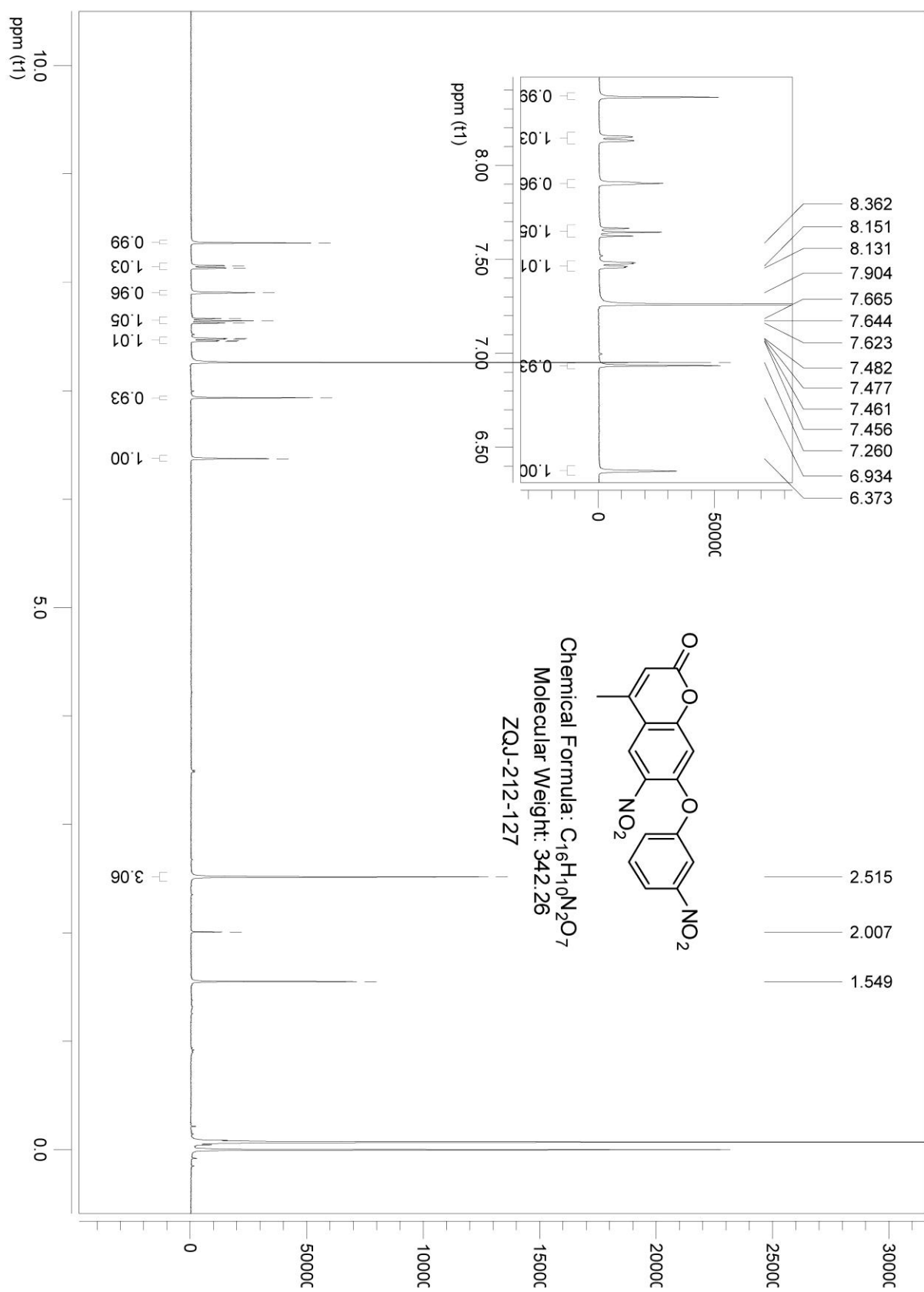


Figure S23. The ^1H -NMR spectrum of **7c** in CDCl_3 .

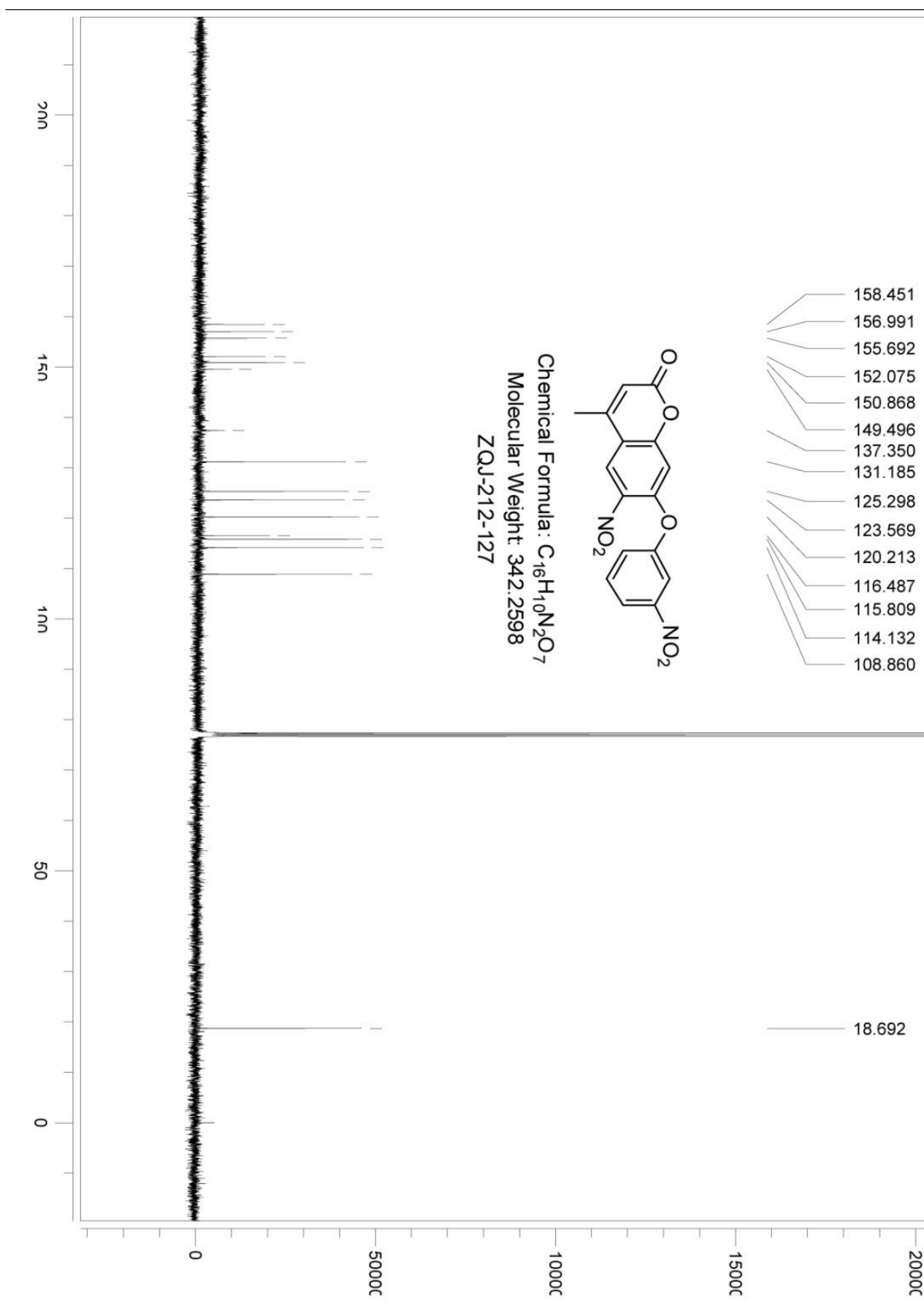


Figure S24. The ^{13}C -NMR spectrum of **7c** in $CDCl_3$.

Elemental Composition Report

Page 1

Single Mass Analysis

Tolerance = 30.0 mDa / DBE: min = -1.5, max = 100.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 2

Monoisotopic Mass, Even Electron Ions

19 formula(e) evaluated with 1 results within limits (up to 1 closest results for each mass)

Elements Used:

C: 0-16 H: 0-11 N: 0-2 O: 0-7

WB-ZHANG

ECUST Institute of Fine Chem

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06-Mar-2014
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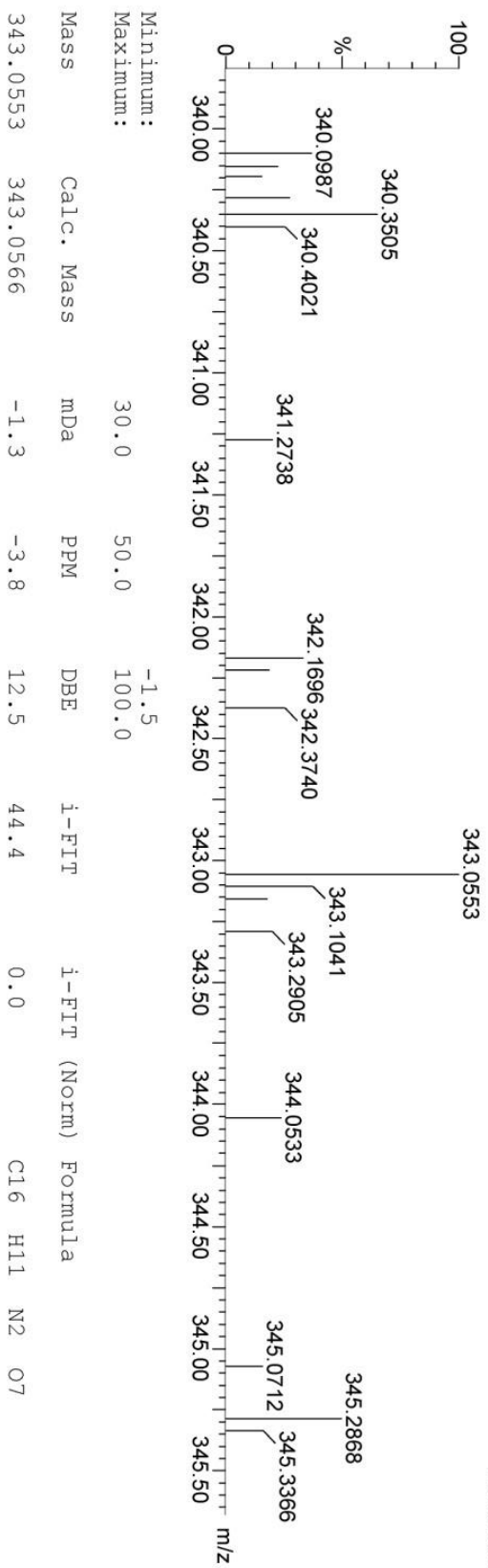


Figure S25. The HRMS spectrum of 7c.

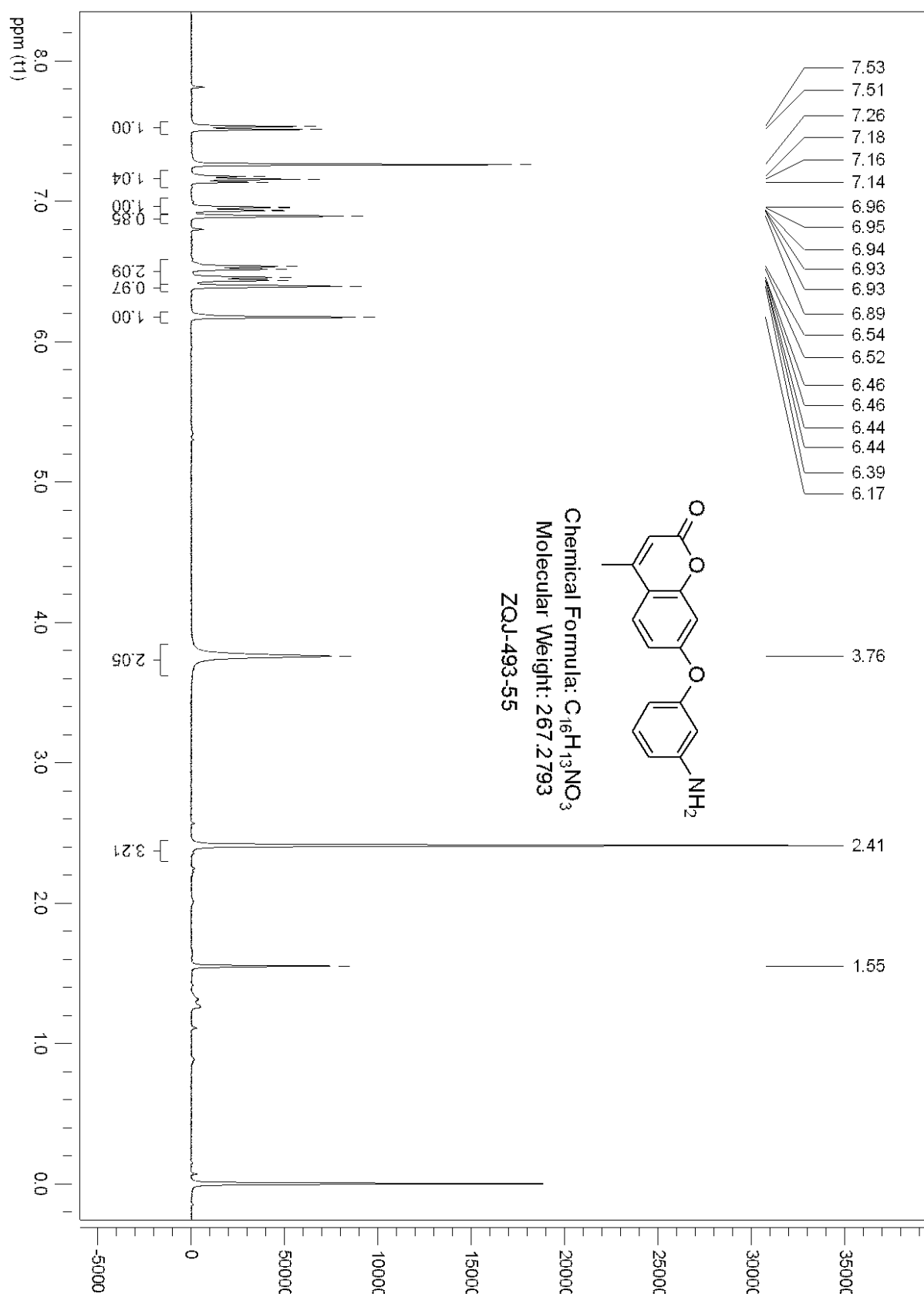


Figure S26. The ^1H -NMR spectrum of **11** in CDCl_3 .

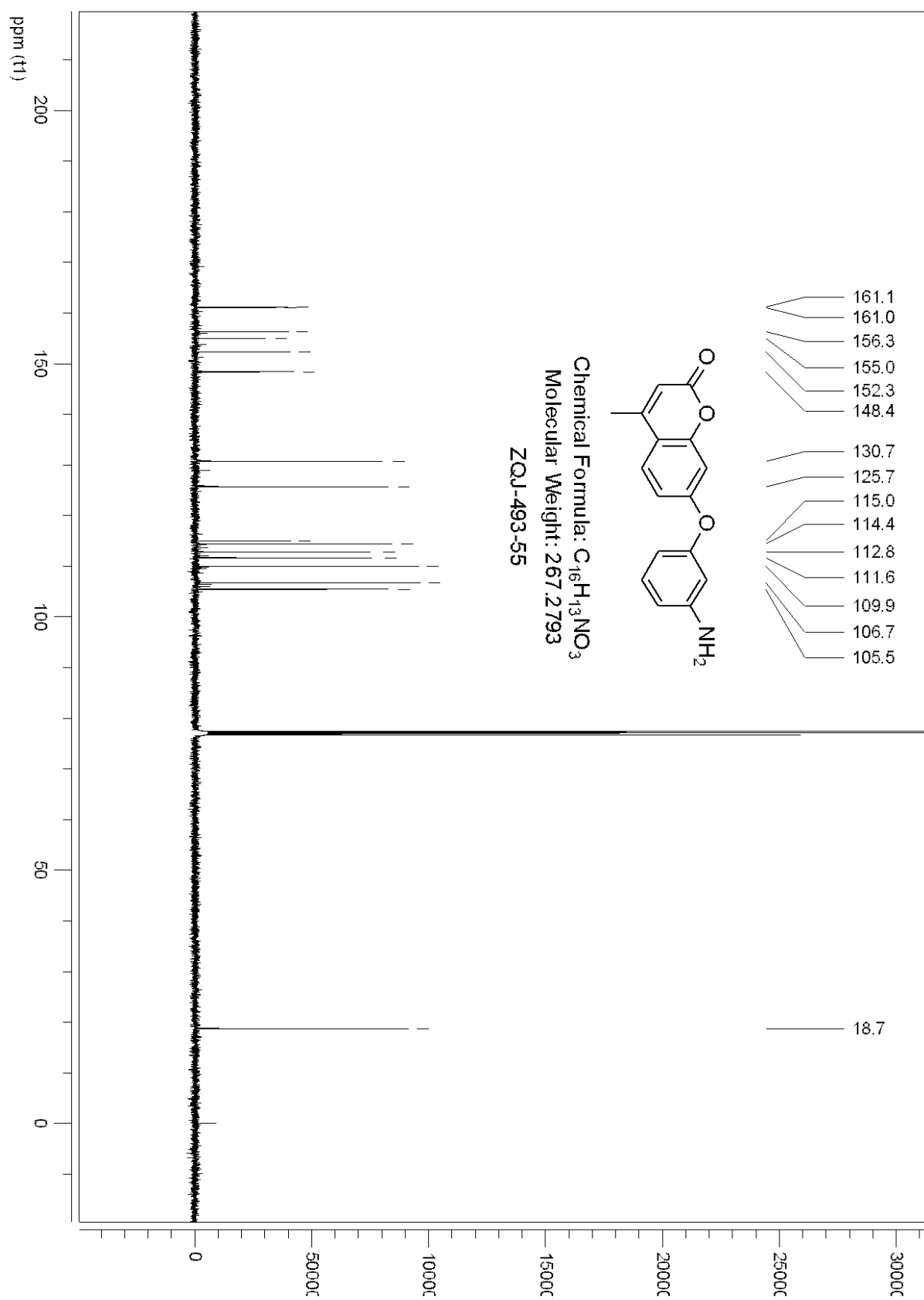


Figure S27. The ^{13}C -NMR spectrum of **11** in $CDCl_3$.

Elemental Composition Report

Page 1

Single Mass Analysis

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Element prediction: Off

Number of isotope peaks used for i-FIT = 2

Monoisotopic Mass, Even Electron Ions

41 formula(e) evaluated with 5 results within limits (up to 1 closest results for each mass)

Elements Used:

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WB-ZHANG

ECUST institute of Fine Chem

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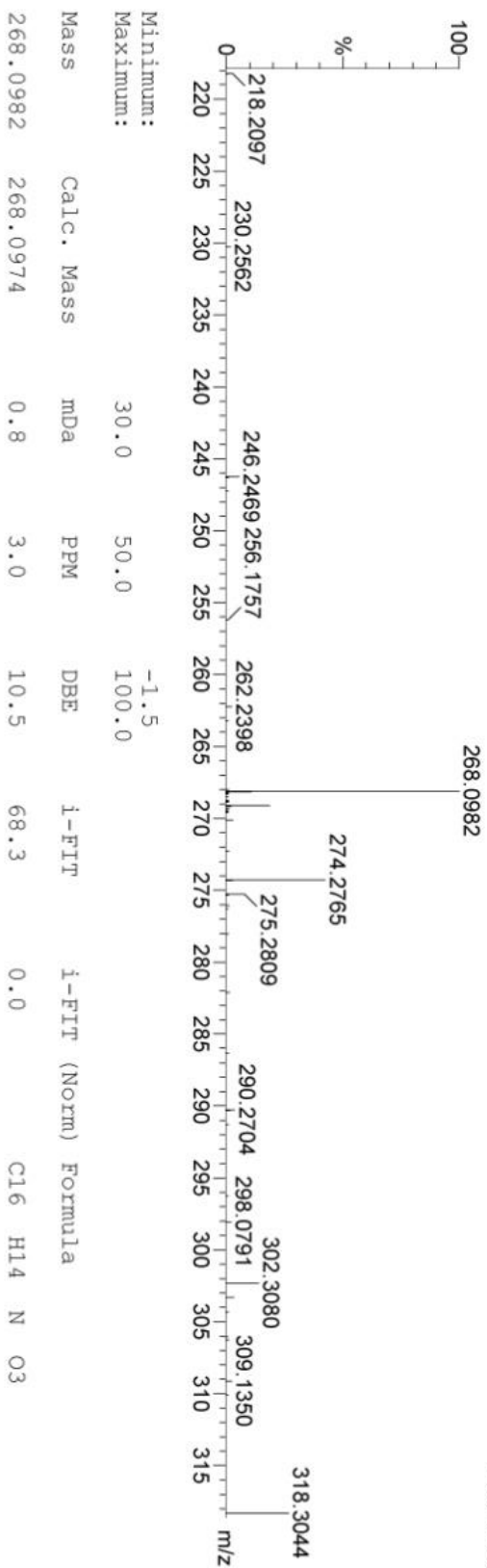


Figure S28. The HRMS spectrum of 11.

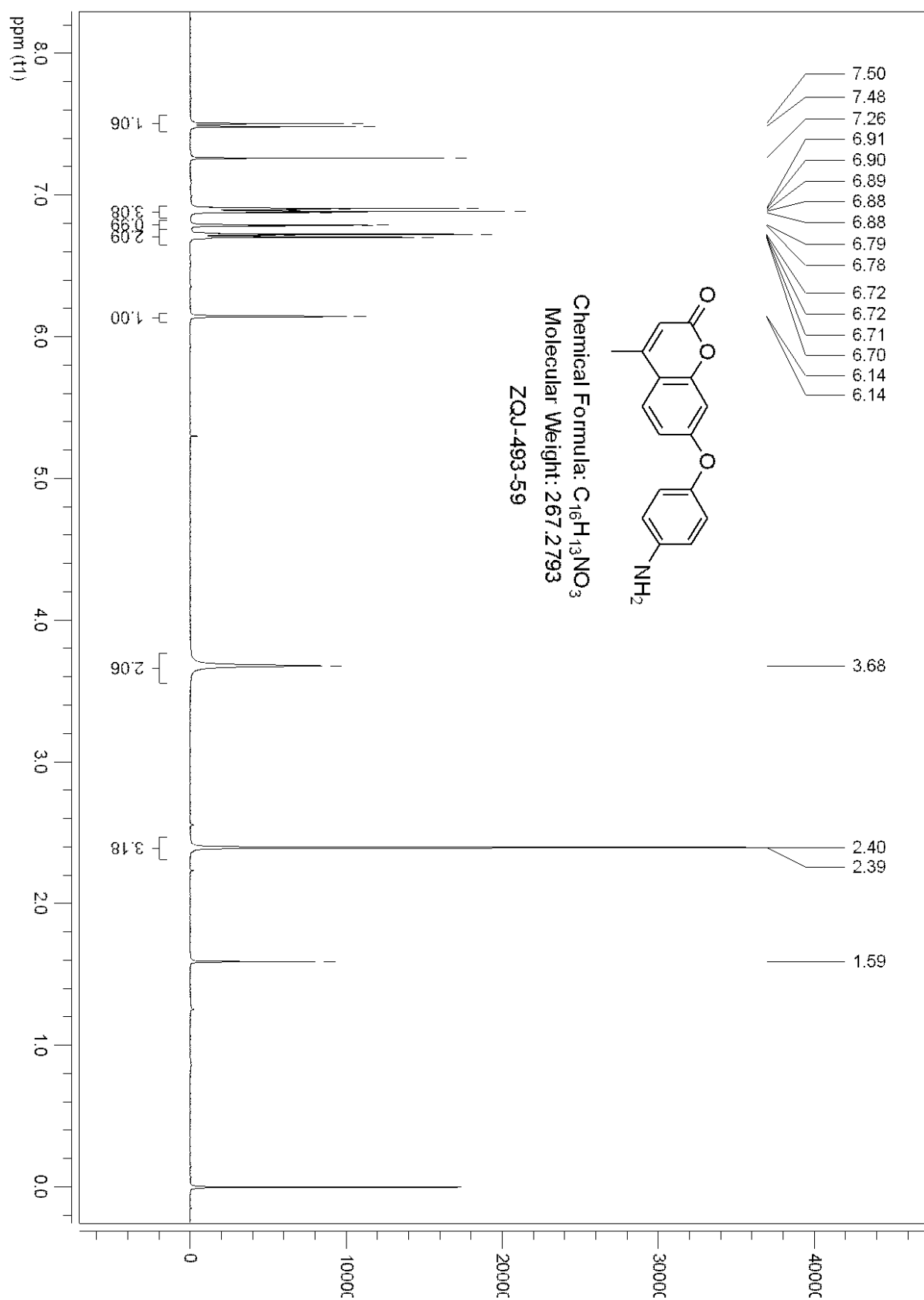


Figure S29. The ^1H -NMR spectrum of **12** in CDCl_3 .

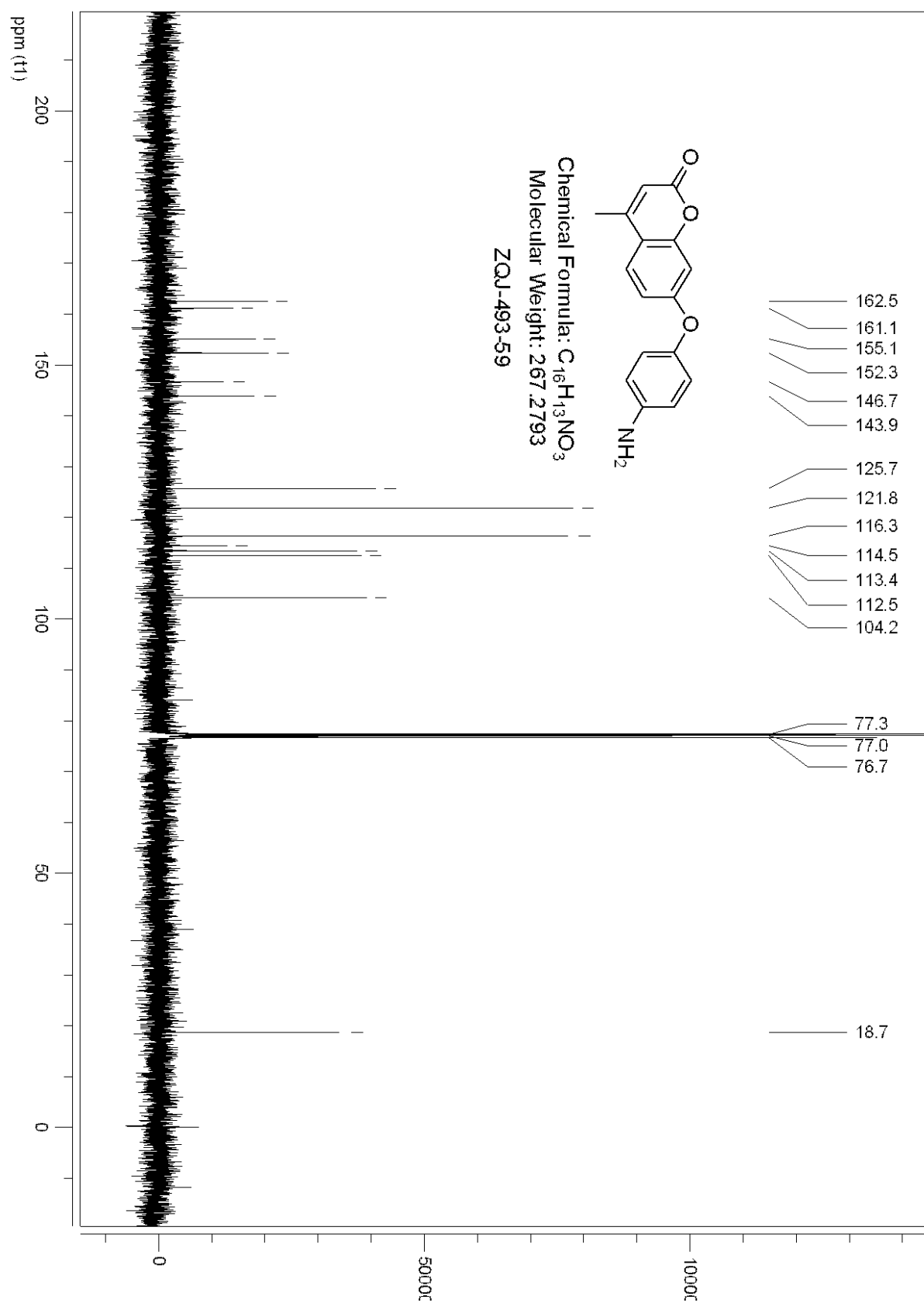


Figure S30. The ^{13}C -NMR spectrum of **12** in $CDCl_3$.

Elemental Composition Report

Page 1

Single Mass Analysis

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Element prediction: Off

Number of isotope peaks used for i-FIT = 2

Monoisotopic Mass, Even Electron Ions
9 formula(e) evaluated with 1 results within limits (up to 1 closest results for each mass)

Elements Used:

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WB-ZHANG

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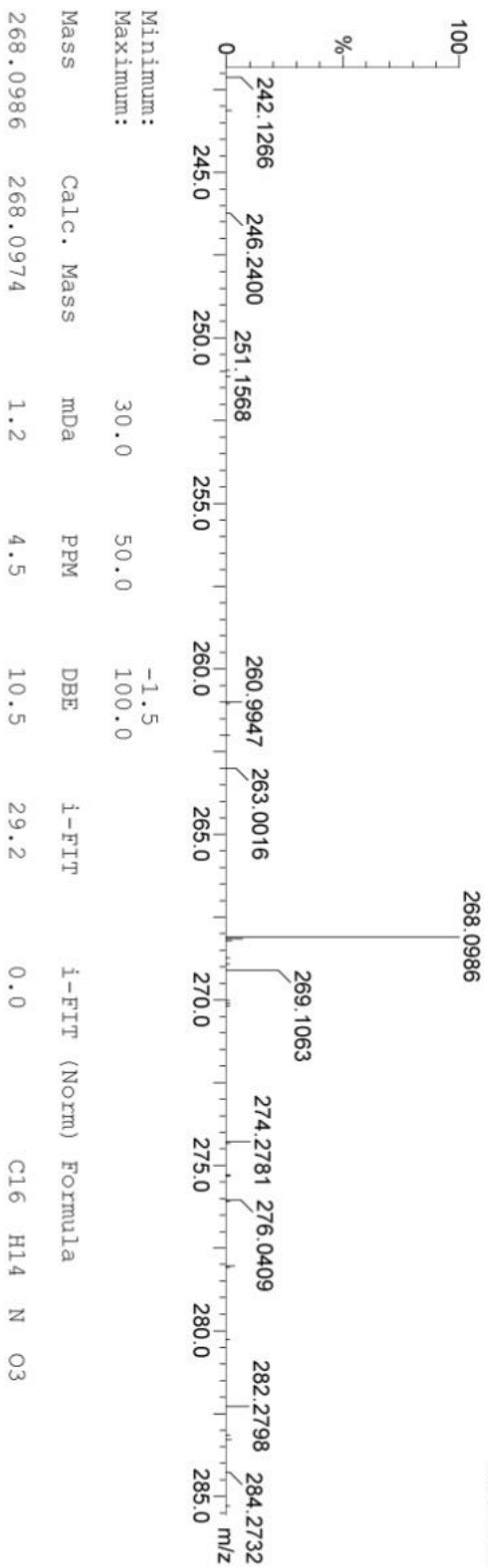


Figure S31. The HRMS spectrum of 12.

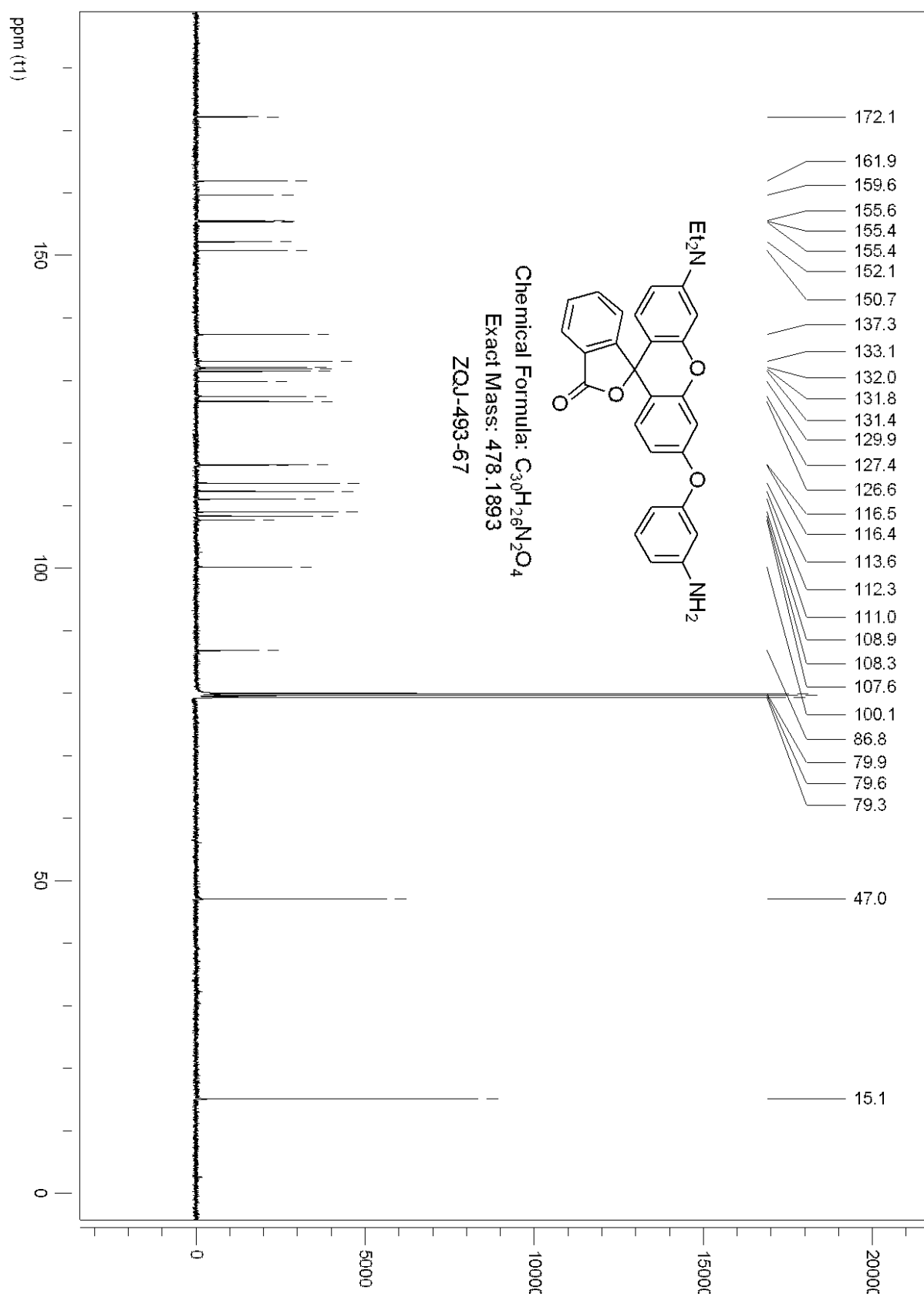


Figure S33. The ¹³C-NMR spectrum of **14** in CDCl₃.

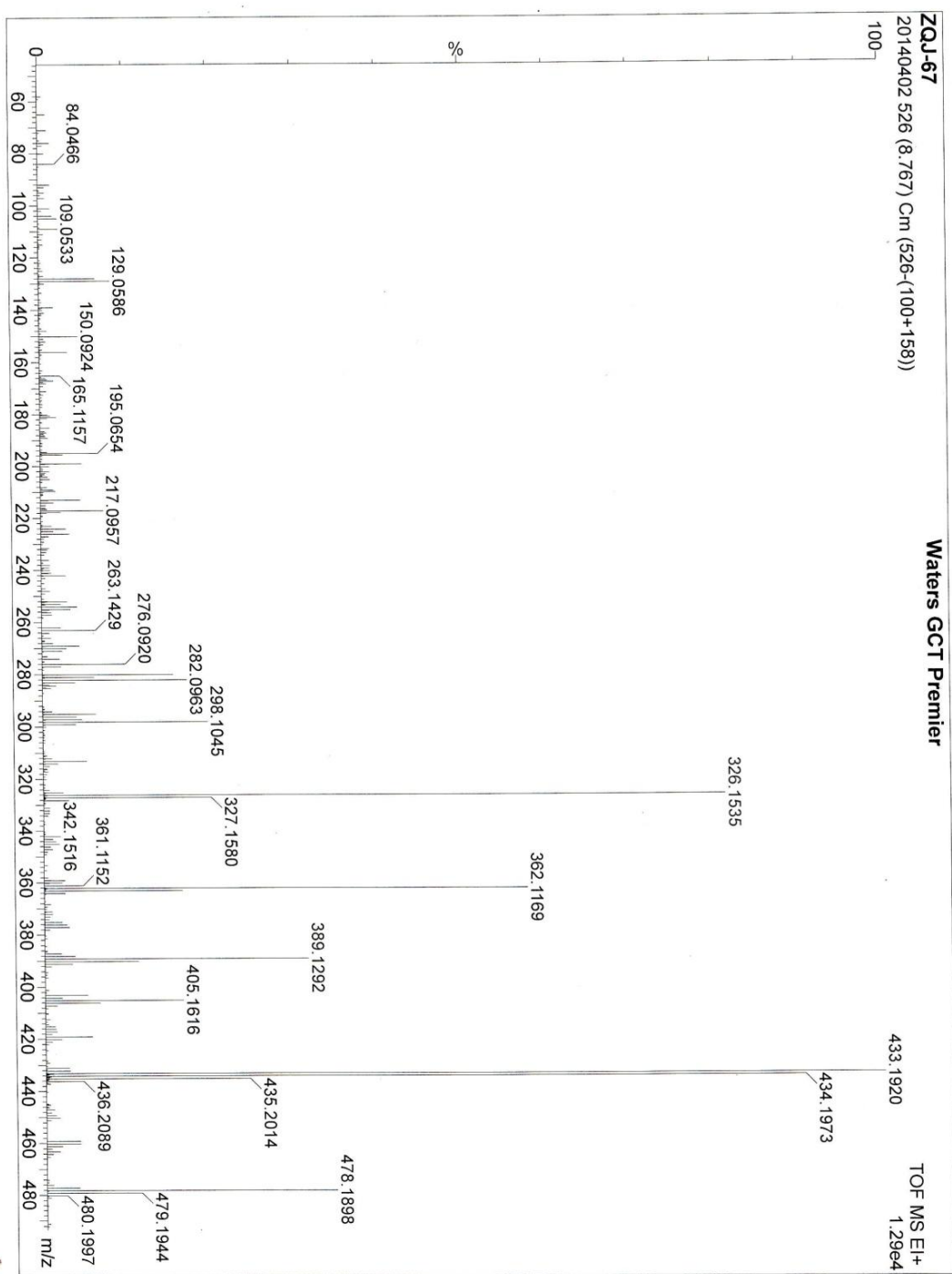


Figure S34. The HRMS spectrum of **14**.

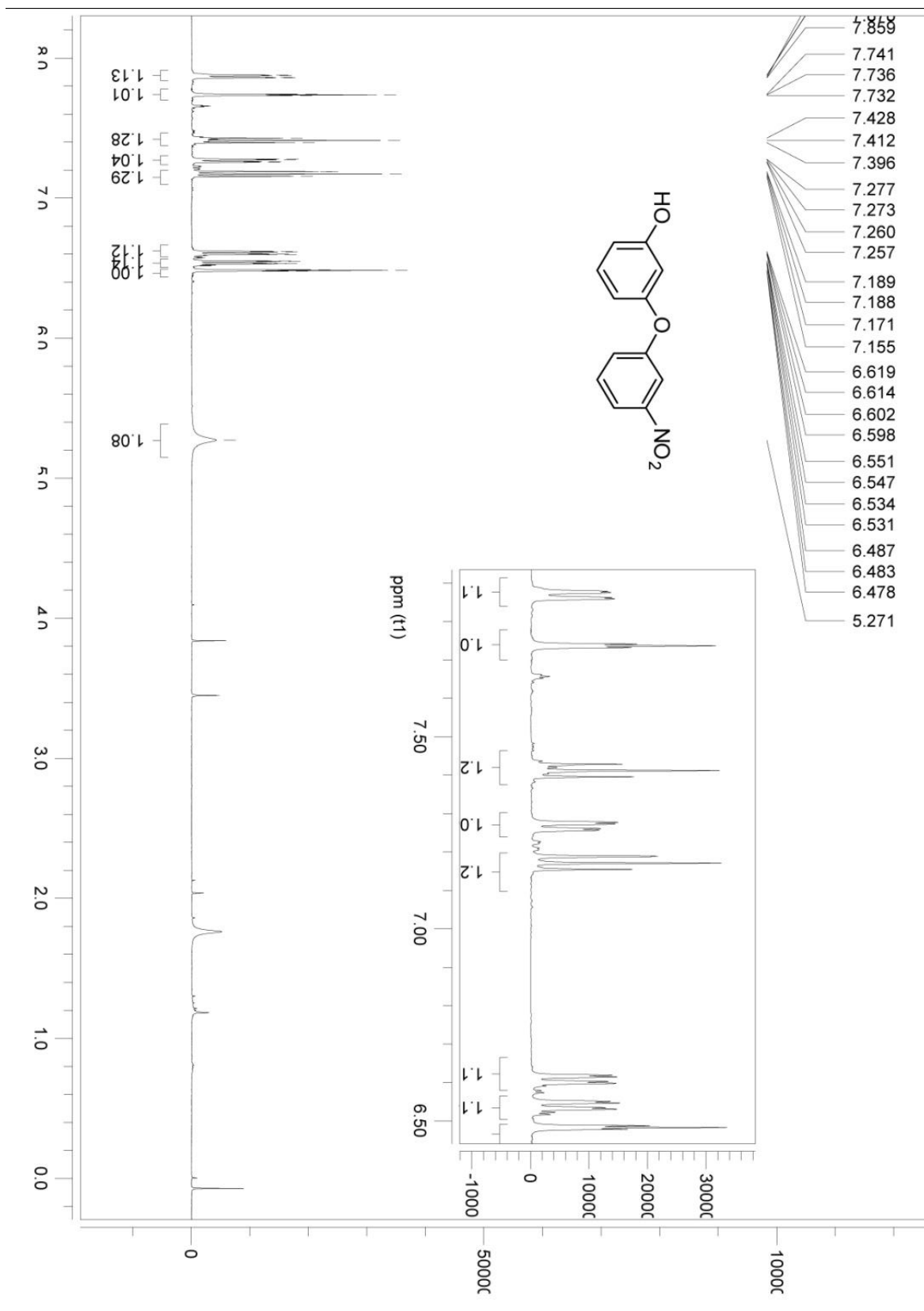


Figure S35. The ¹H-NMR spectrum of **19** in CDCl₃.

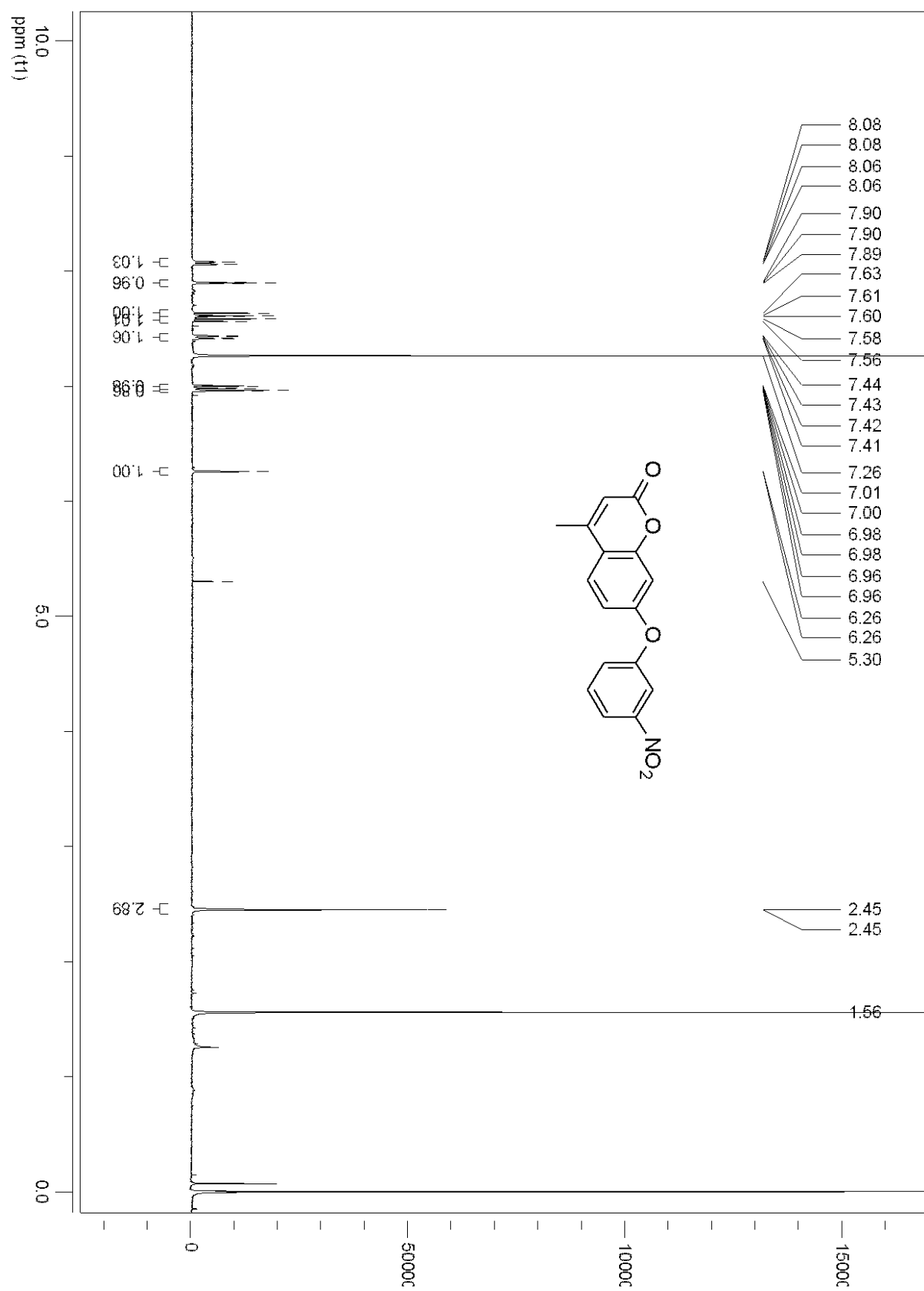


Figure S36. The ¹H-NMR spectrum of **21** in CDCl₃.

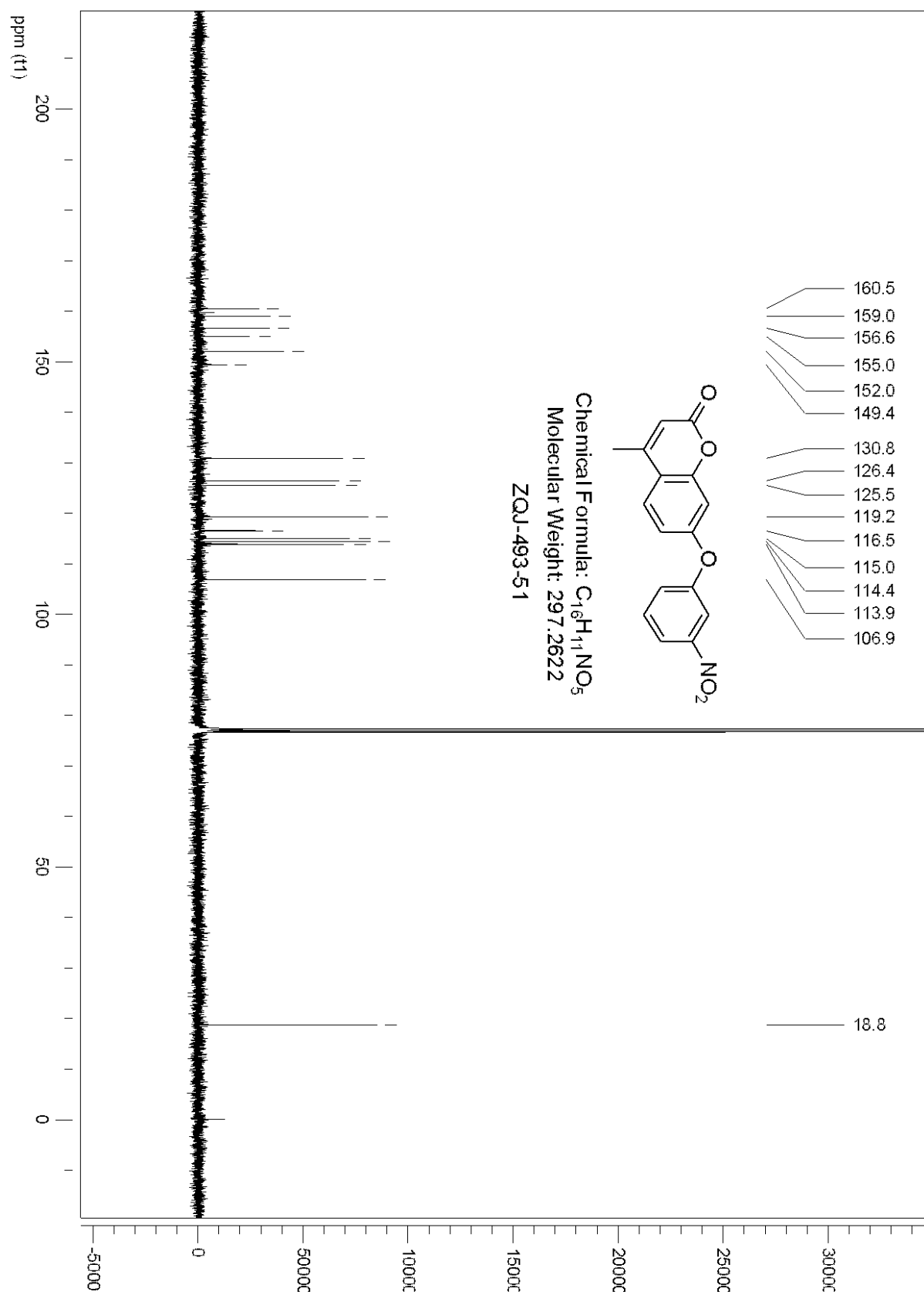


Figure S37. The ^{13}C -NMR spectrum of **21** in CDCl₃.

Elemental Composition Report

Page 1

Single Mass Analysis

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Element prediction: Off

Number of isotope peaks used for i-FIT = 2

Monoisotopic Mass, Even Electron Ions

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Elements Used:

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WB-ZHANG

ECUST Institute of Fine Chem

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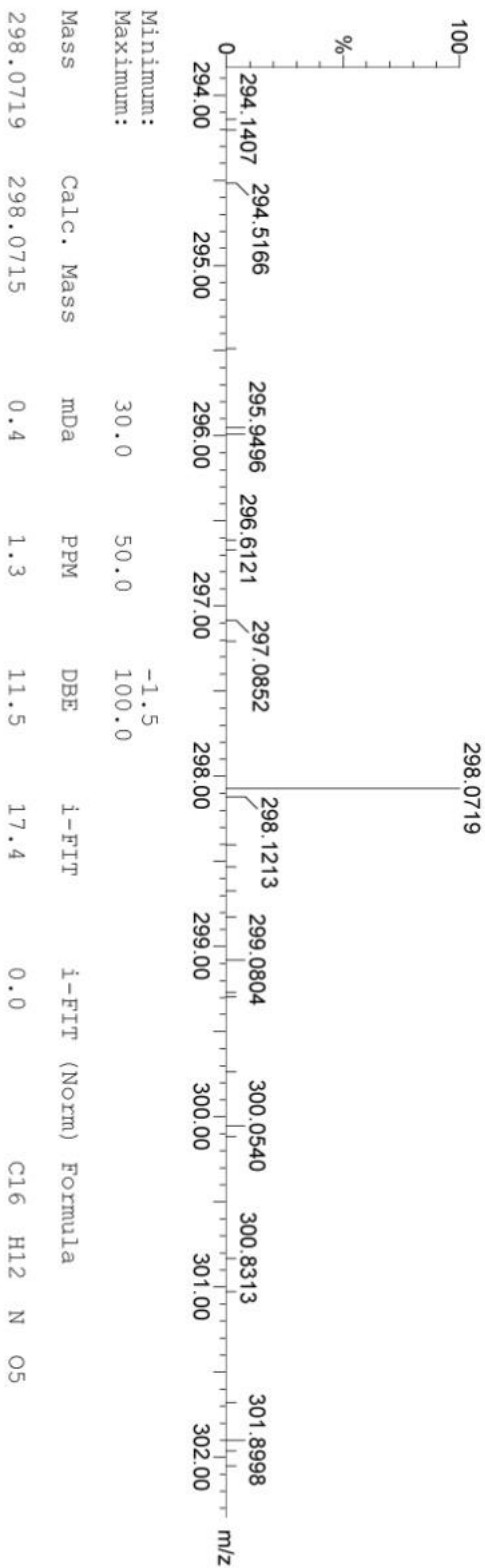


Figure S38. The HRMS spectrum of 21.

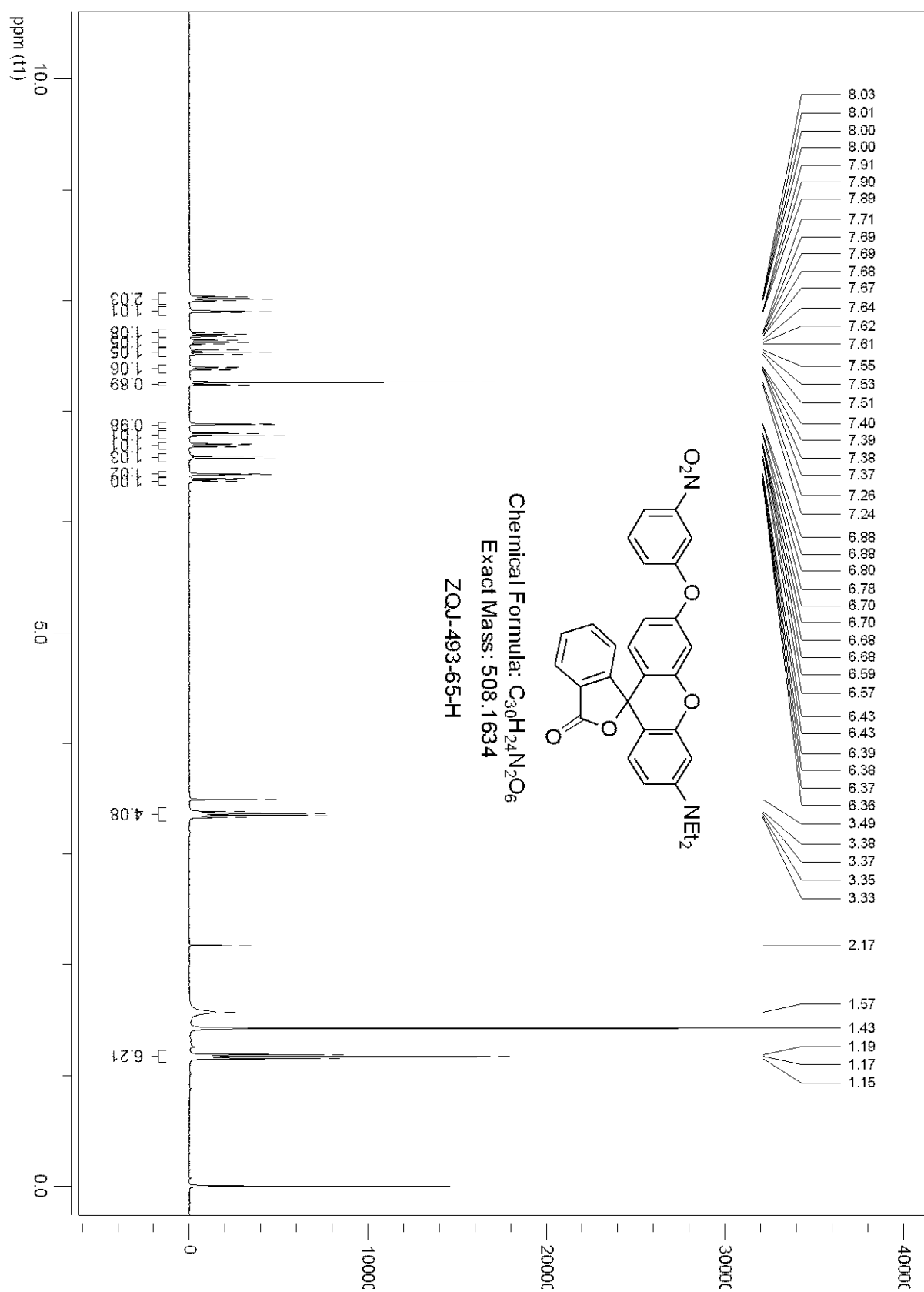


Figure S39. The ^1H -NMR spectrum of **23** in CDCl_3 .

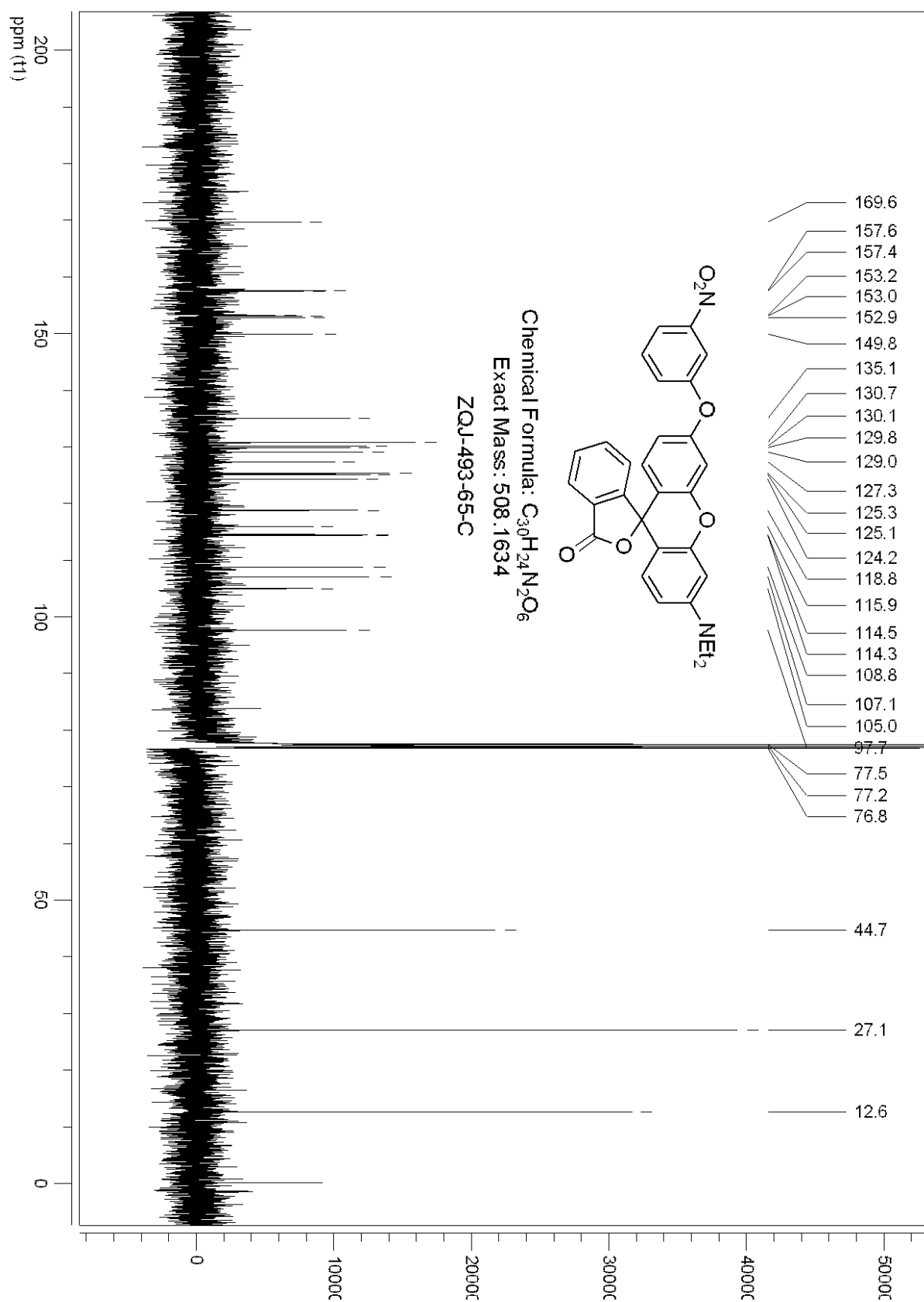


Figure S40. The ¹³C-NMR spectrum of **23** in CDCl₃.

Elemental Composition Report

Page 1

Single Mass Analysis

Tolerance = 30.0 mDa / DBE: min = -1.5, max = 100.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 2

Monoisotopic Mass, Even Electron Ions

13 formula(e) evaluated with 1 results within limits (up to 1 closest results for each mass)

Elements Used:

C: 0-30 H: 0-25 N: 0-2 O: 0-6

WB-ZHANG

ECUST Institute of Fine Chem

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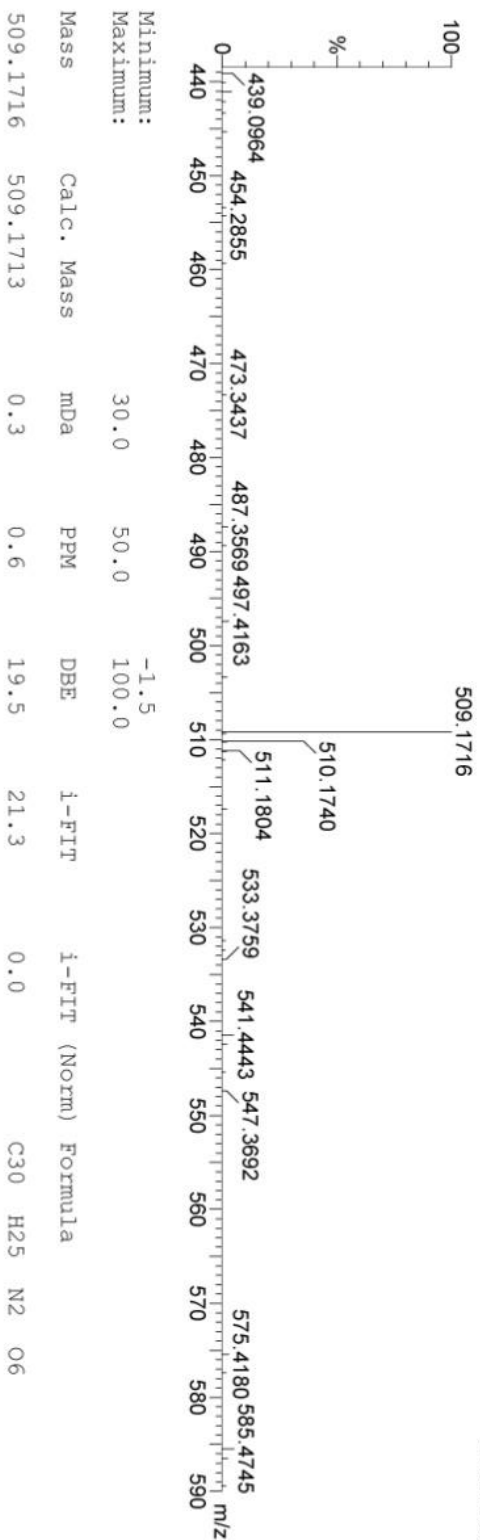


Figure S41. The HRMS spectrum of **23**.