

# Supporting information

## **Minimization of synthetic polymer ligands for specific recognition and neutralization of a toxic peptide**

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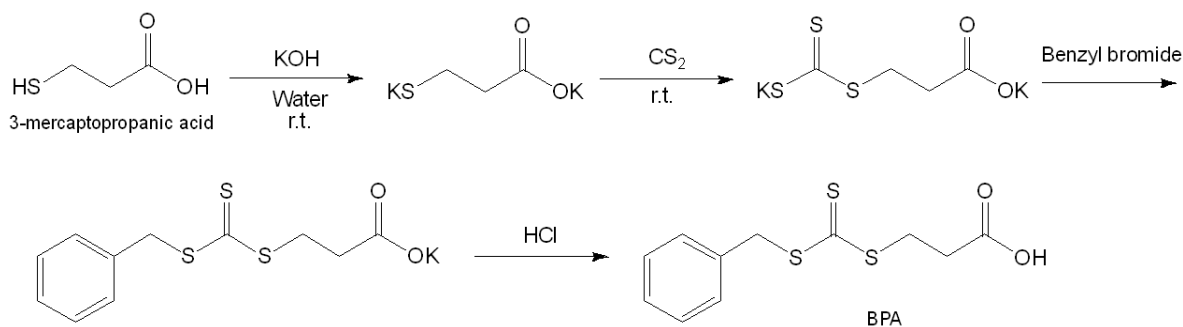
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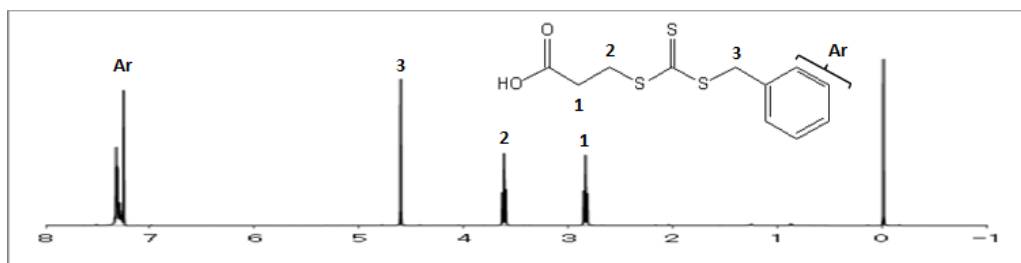
## S1. Preparation of minimized synthetic polymer ligands

### 1. Preparation of BPA



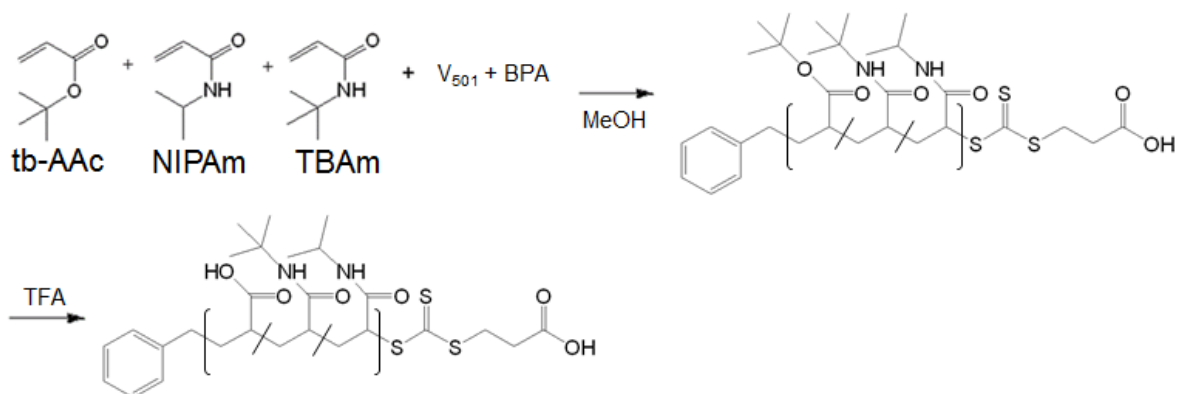
**Scheme S1-1.**

BPA was prepared according to Davis and his coworkers<sup>1</sup>. 3-Mercaptopropionic acid (10 mL, 115 mmol) was added to potassium hydroxide (12.9 g in 125 mL H<sub>2</sub>O, 230 mmol). Subsequently, 15.3 mL CS<sub>2</sub> was added dropwise. The resulting orange-colored solution was stirred for 5 h at room temperature, after which 13.65 mL benzyl bromide (55.4 mmol) was added. Benzyl bromide turned the color of the solution to opaque yellow. The mixture was heated to 80 °C for 12 h and cooled. Chloroform (100 mL) was added next, and 100 mL HCl (1 M in H<sub>2</sub>O) was slowly added until the organic layer turned yellow. The aqueous layer was extracted with chloroform (3 x 100 mL), while the organic layer was washed with distilled water (3 x 100 mL). Finally, the organic layer was dried over MgSO<sub>4</sub>. BPA was recrystallized in hexane. Yield: 65 %.



**Figure S1-1.** <sup>1</sup>H NMR of BPA in CDCl<sub>3</sub>

## 2. Preparation of 300-mer, 30-mer, 15-mer and 9-mer polymer ligands



**Scheme S1-2.**

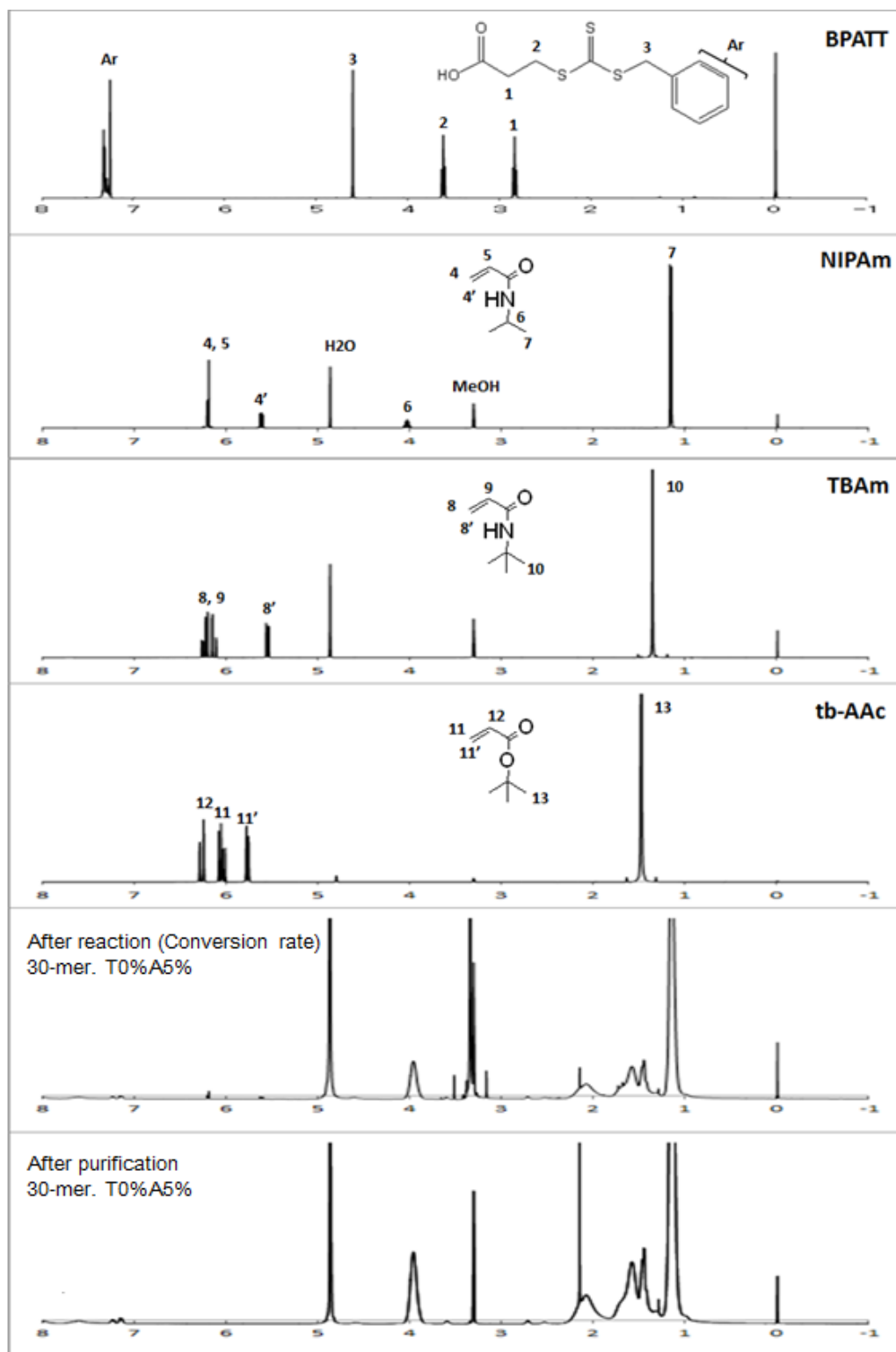
Reversible addition-fragmentation chain-transfer (RAFT) was used to obtain 300-mer, 30-mer, 15-mer and 9-mer polymer ligands with narrow molecular weight distribution. BPA was used as chain transfer agent. *Tert*-butyl acrylate (tb-AAc), *N*-isopropyl acrylamide (NIPAm), *N*-*tert*-butylacrylamide (TBAm), BPA (20.4 or 40.8 mg) and  $V_{501}$  (6.3 mg) were dissolved in methanol (~1mL) according to the feed ratio listed in Table S1-1. Solutions were degassed in three cycles of freeze-evacuate-thaw, sealed in an ampoule, and heated at 70 °C for 3 h in an oil bath. Products were precipitated with brine and distilled water, and conversion rates were calculated from  $^1\text{H}$  NMR in MeOD. After the composition and molecular weight were determined by  $^1\text{H}$  NMR and GPC, polymer ligands were deprotected with TFA.

**Table S1-1.** Feed ratio and characteristics of resulting polymers

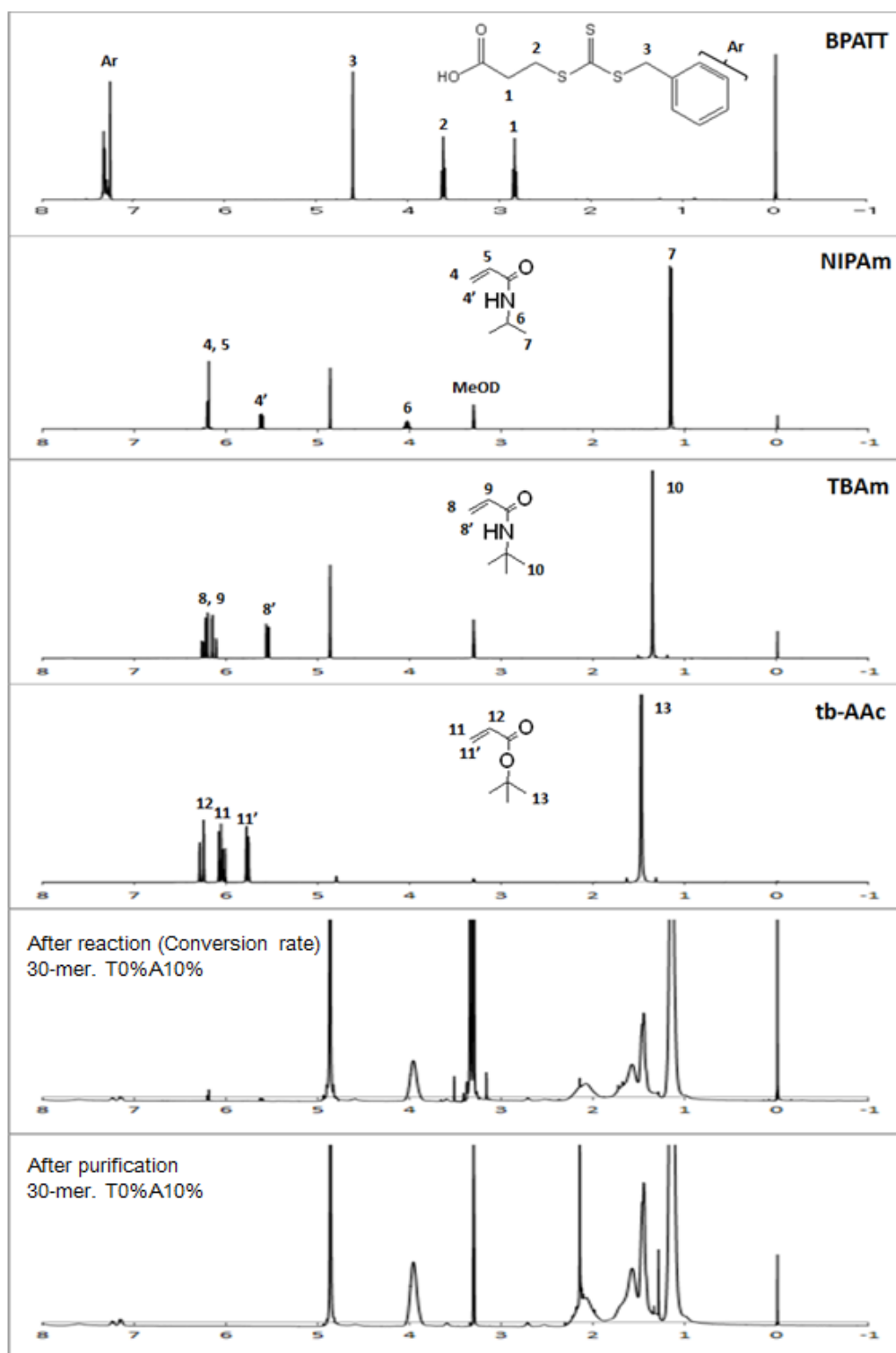
	TBAm (x)	<i>tb</i> -AAc (y)	NIPAm (100-x-y)	Conversion rate	GPC		
					M <sub>n</sub>	M <sub>w</sub>	PDI
30-mer (T0%A5%)	0	5 mol% 14.1 mg, 0.11 mmol	95 mol% 244.2 mg, 2.15 mmol	97	3400	4600	1.4
30-mer (T0%A10%)	0	10 mol% 28.2 mg, 0.22 mmol	90 mol% 231.3 mg, 2.04 mmol	98	2800	3700	1.3
30-mer (T0%A20%)	0	20 mol% 56.4 mg, 0.44 mmol	80 mol% 205.6 mg, 1.82 mmol	98	2600	3400	1.3
30-mer (T10%A0%)	10 mol% 28.6 mg, 0.23 mmol	0	90 mol% 231.3 mg, 2.04 mmol	97	2800	3800	1.4
30-mer (T10%A10%)	10 mol% 28.6 mg, 0.23 mmol	10 mol% 28.2 mg, 0.22 mmol	80 mol% 205.6 mg, 1.82 mmol	94	2700	3600	1.3
30-mer (T20%A0%)	20 mol% 57.2 mg, 0.45 mmol	0	80 mol% 205.6 mg, 1.82 mmol	93	3100	4100	1.4
30-mer (T20%A5%)	20 mol% 57.2 mg, 0.45 mmol	5 mol% 14.1 mg, 0.11 mmol	75 mol% 192.8 mg, 1.7 mmol	98	2500	3300	1.3
30-mer (T20%A10%)	20 mol% 57.2 mg, 0.45 mmol	10 mol% 28.2 mg, 0.22 mmol	70 mol% 179.9 mg, 1.59 mmol	96	2800	3600	1.3
30-mer (T20%A20%)	20 mol% 57.2 mg, 0.45 mmol	20 mol% 56.4 mg, 0.44 mmol	60 mol% 154.2 mg, 1.36 mmol	96	2600	3400	1.3
30-mer (T40%A0%)	40 mol% 114.4 mg, 0.9 mmol	0	60 mol% 154.2 mg, 1.36 mmol	99	2700	3600	1.3
30-mer (T40%A5%)	40 mol% 114.4 mg, 0.9 mmol	5 mol% 14.1 mg, 0.11 mmol	55 mol% 141.35 mg, 1.25 mmol	98	2500	3400	1.4
30-mer (T40%A10%)	40 mol% 114.4 mg, 0.9 mmol	10 mol% 28.2 mg, 0.22 mmol	50 mol% 129.0 mg, 1.14 mmol	95	2600	3500	1.3
30-mer (T40%A20%)	40 mol% 114.4 mg, 0.9 mmol	20 mol% 56.4 mg, 0.44 mmol	40 mol% 102.8 mg, 0.91 mmol	97	2500	3200	1.3
15-mer (T20%A0%)	20 mol% 57.2 mg, 0.45 mmol	0	80 mol% 205.6 mg, 1.82 mmol	93	1500	1700	1.2
15-mer (T20%A5%)	20 mol% 57.2 mg, 0.45 mmol	5 mol% 14.1 mg, 0.11 mmol	75 mol% 192.8 mg, 1.7 mmol	96	1400	1600	1.1
15-mer (T20%A10%)	20 mol% 57.2 mg, 0.45 mmol	10 mol% 28.2 mg, 0.22 mmol	70 mol% 179.9 mg, 1.59 mmol	98	1400	1600	1.1
15-mer (T20%A20%)	20 mol% 57.2 mg,	20 mol% 56.4 mg,	60 mol% 154.2 mg,	99	1600	2000	1.3

	0.45 mmol	0.44 mmol	1.36 mmol				
15-mer	40 mol%	0	60 mol%	96	1500	1700	1.1
(T40%A0%)	114.4 mg, 0.9 mmol		154.2 mg, 1.36 mmol				
15-mer	40 mol%	5 mol%	55 mol%	95	1400	1500	1.1
(T40%A5%)	114.4 mg, 0.9 mmol	14.1 mg, 0.11 mmol	141.35 mg, 1.25 mmol				
15-mer	40 mol%	10 mol%	50 mol%	98	1600	2100	1.3
(T40%A10%)	114.4 mg, 0.9 mmol	28.2 mg, 0.22 mmol	129.0 mg, 1.14 mmol				
15-mer	40 mol%	20 mol%	40 mol%	98	1500	1800	1.2
(T40%A20%)	114.4 mg, 0.9 mmol	56.4 mg, 0.44 mmol	102.8 mg, 0.91 mmol				

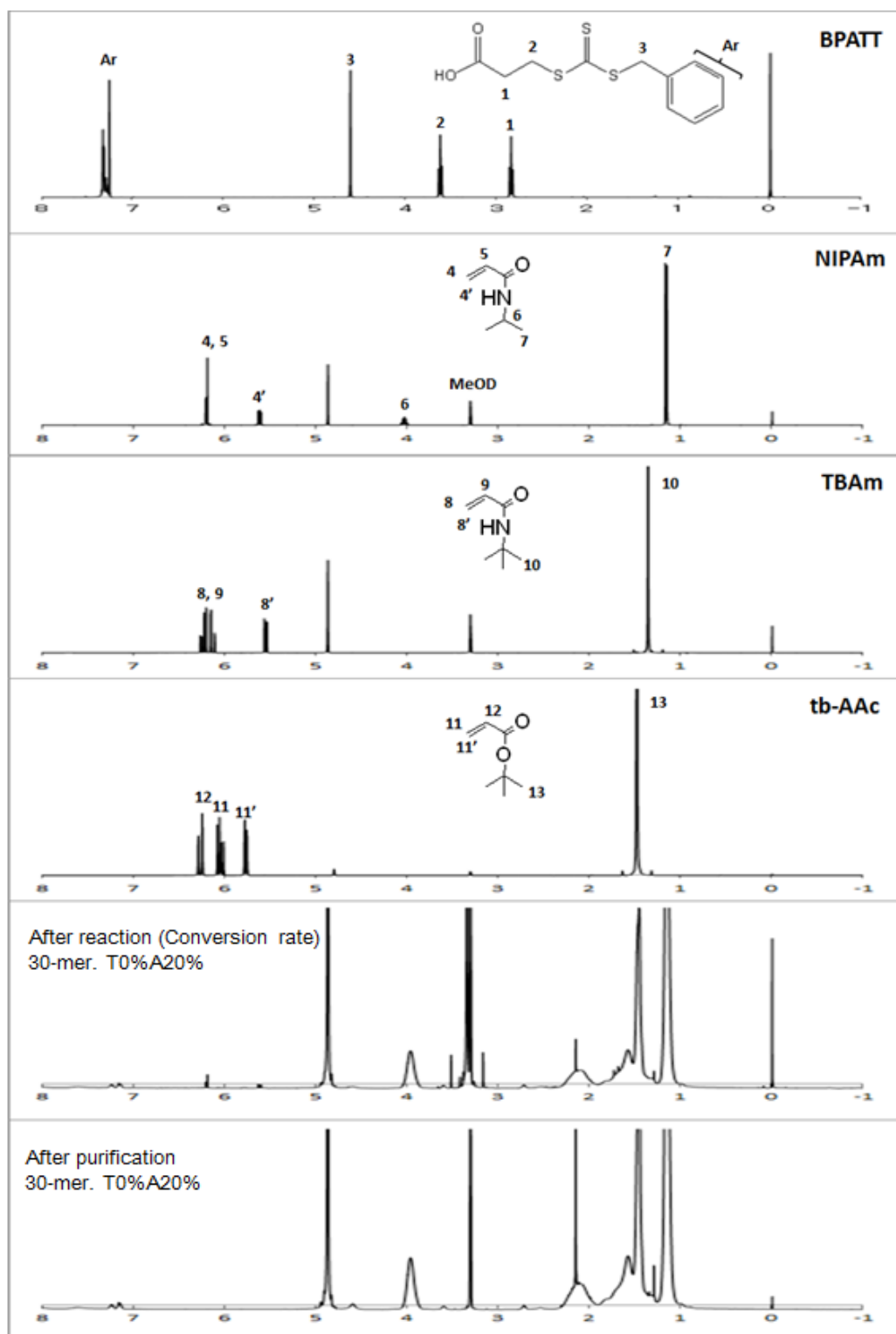
### 3. $^1\text{H}$ NMR of polymer ligands in MeOD



**Figure S1-2.**  $^1\text{H}$  NMR of 30-mer PL, containing 0 % TBAm and 5 % *tb*-AAc

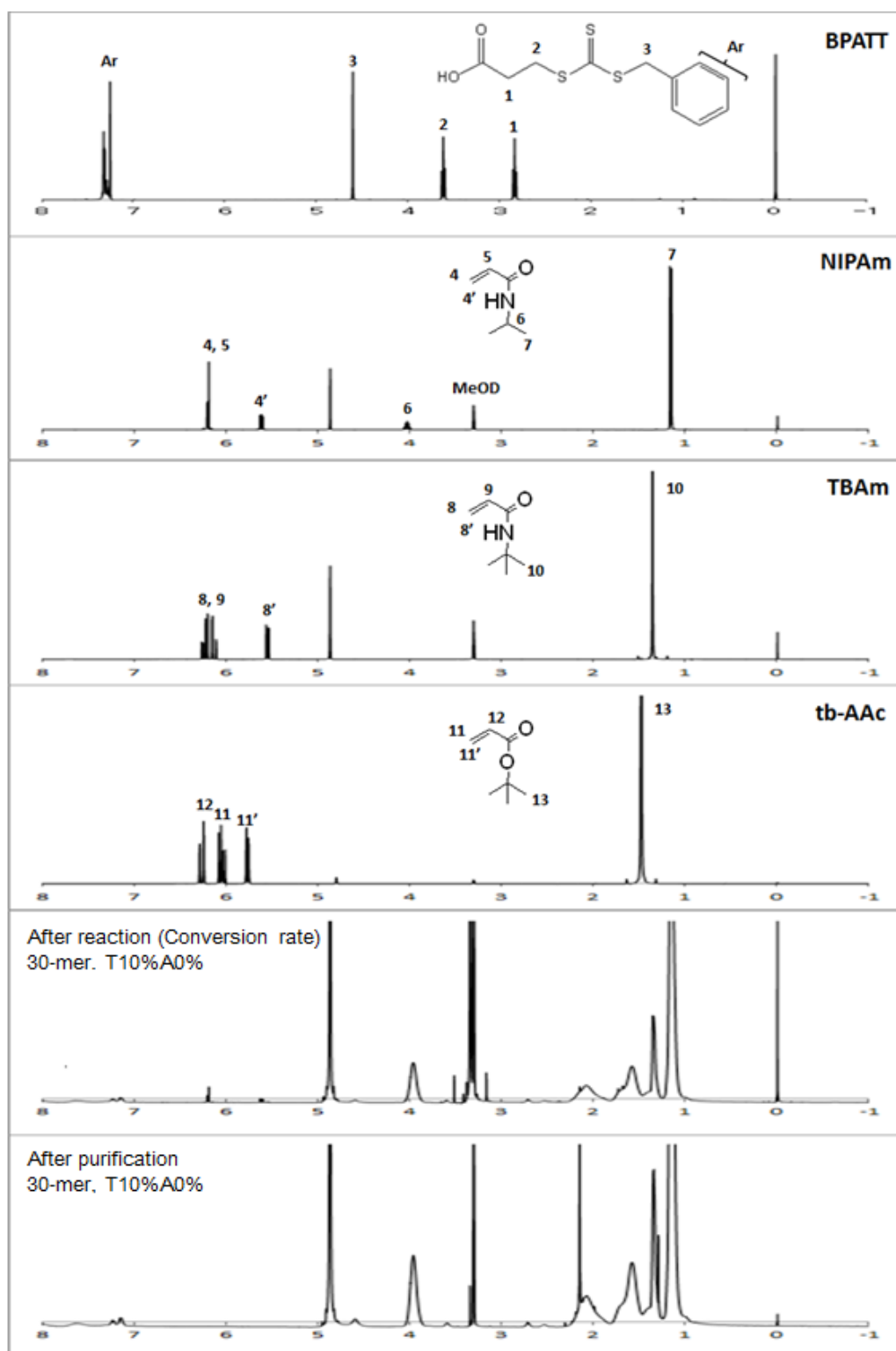


**Figure S1-3.**  $^1\text{H}$  NMR of 30-mer PL, containing 0 % TBAm and 10 % *tb*-AAc

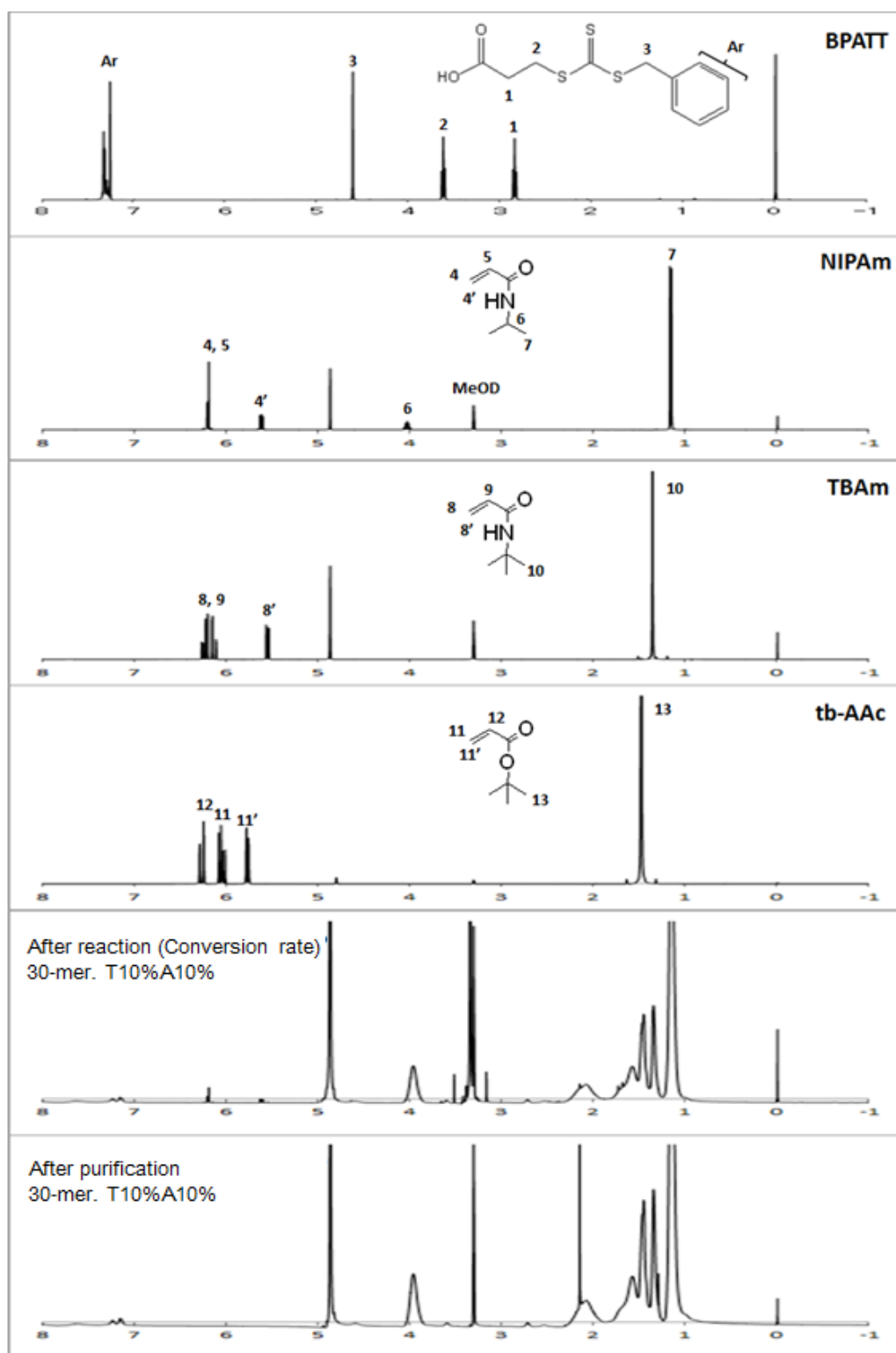


**Figure S1-4.**  $^1\text{H}$  NMR of 30-mer PL, containing 0 % TBAm and 20 % *tb*-AAc

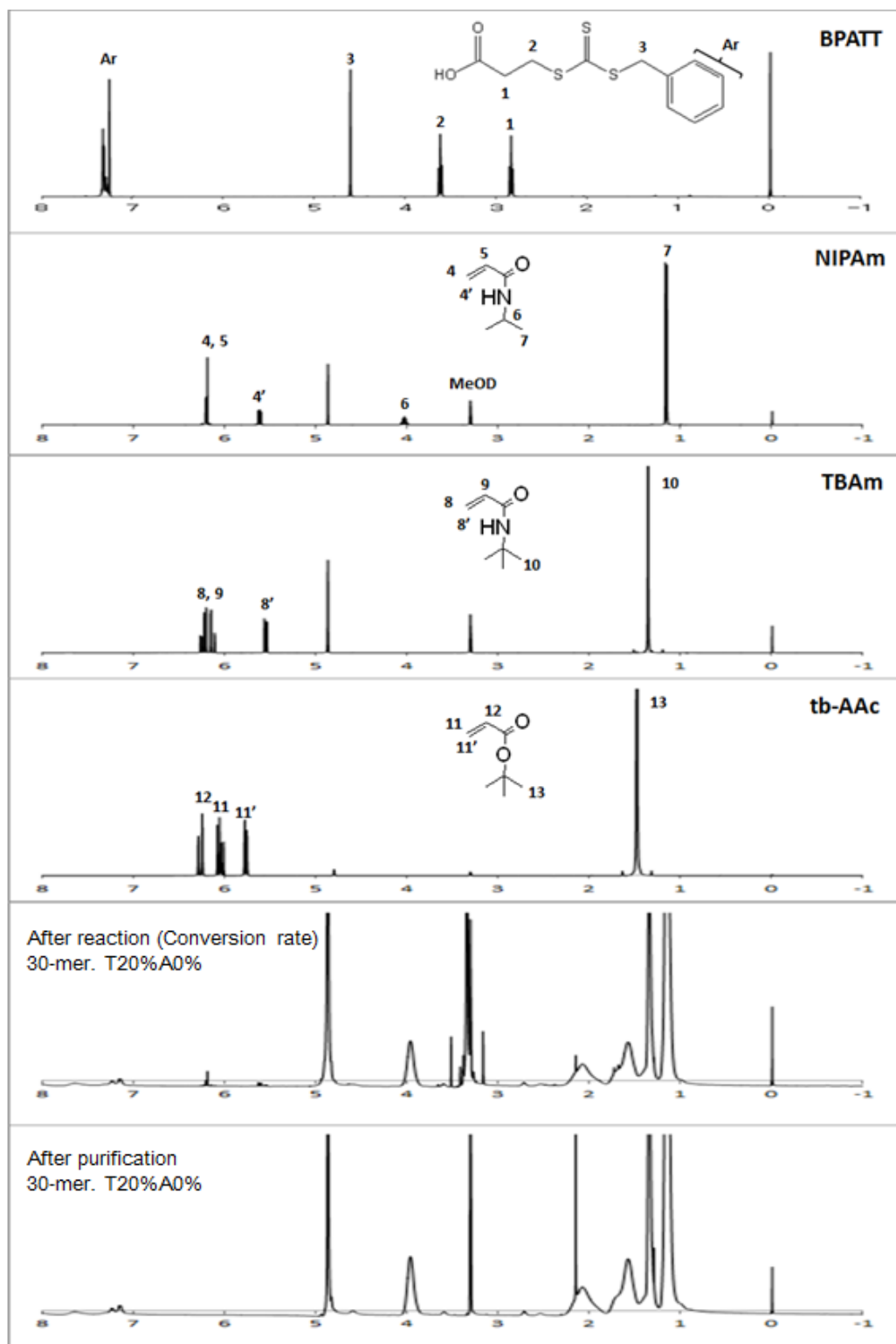




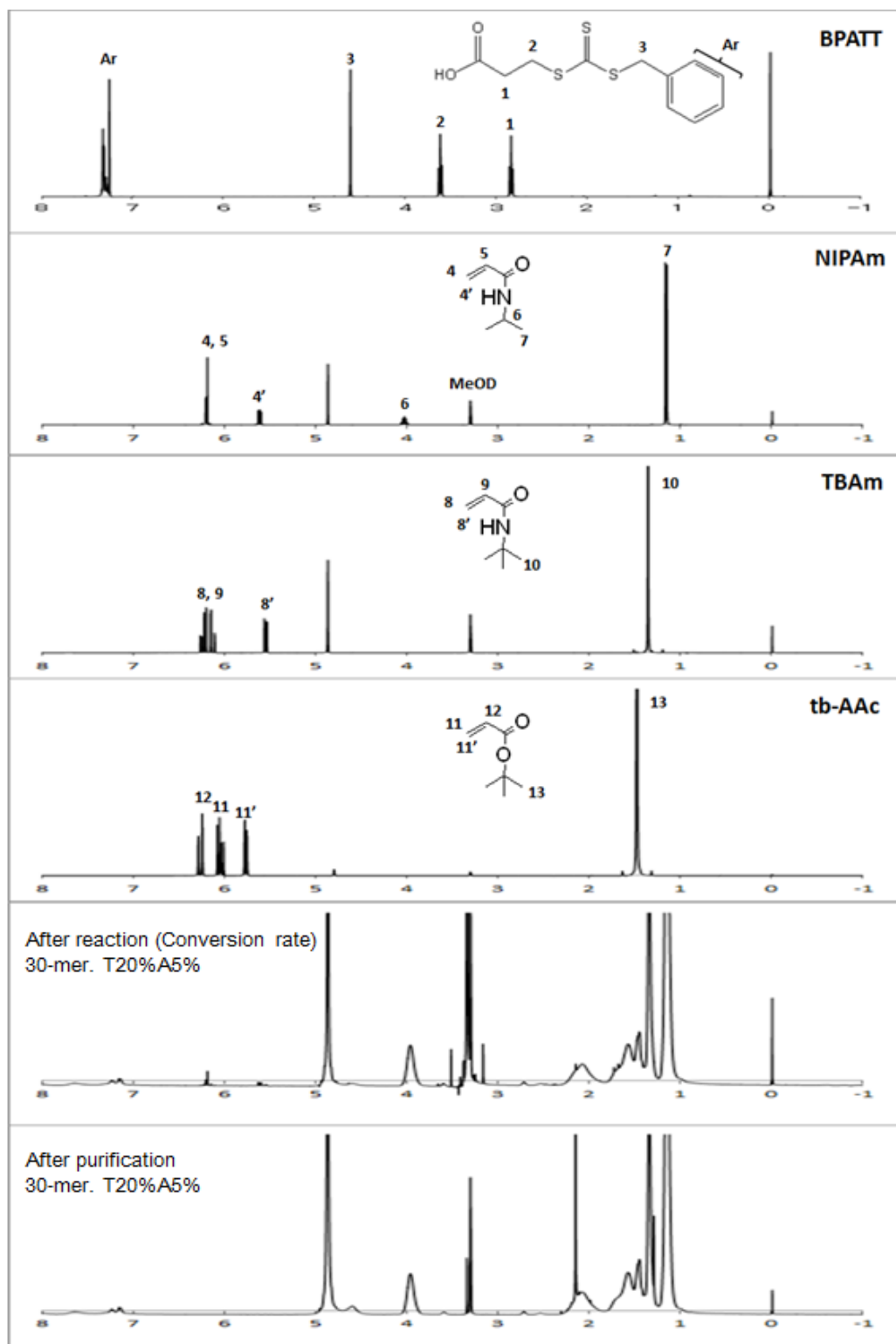
**Figure S1-5.**  $^1\text{H}$  NMR of 30-mer PL, containing 10 % TBAm and 0 % *tb*-AAc



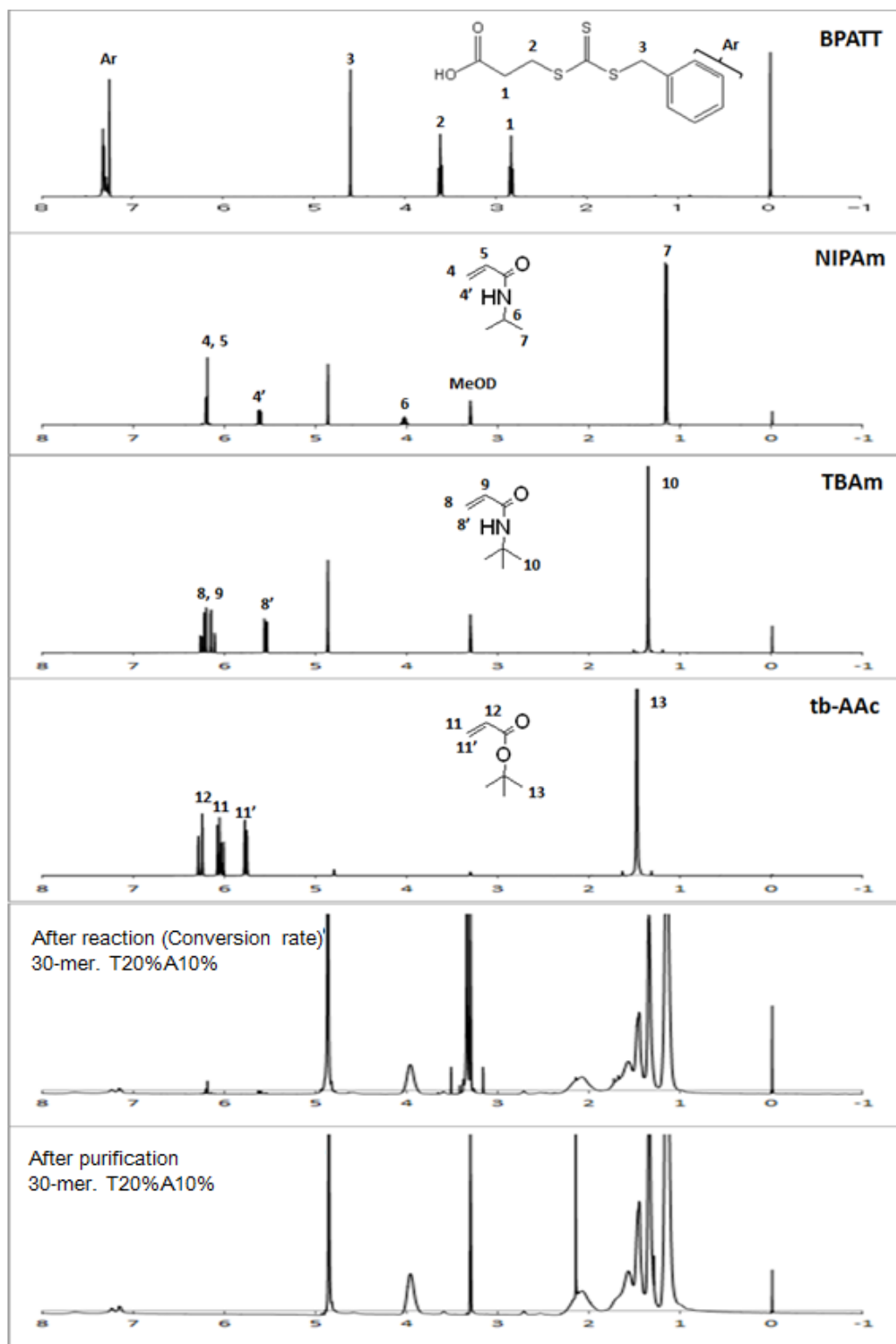
**Figure S1-6.**  $^1\text{H}$  NMR of 30-mer PL, containing 10 % TBAm and 10 % *tb*-AAc



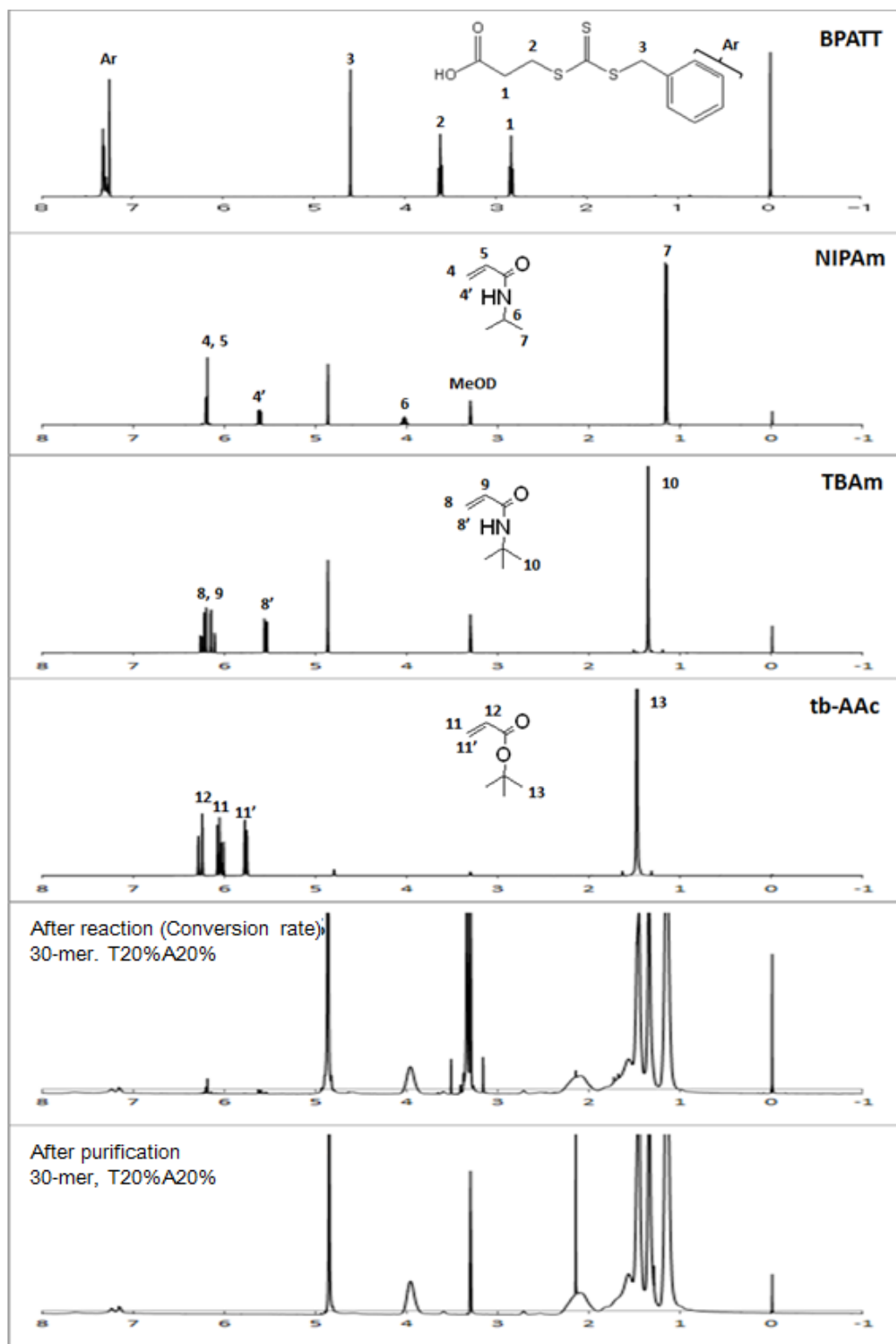
**Figure S1-7.**  $^1\text{H}$  NMR of 30-mer PL, containing 20 % TBAm and 0 % *tb*-AAc



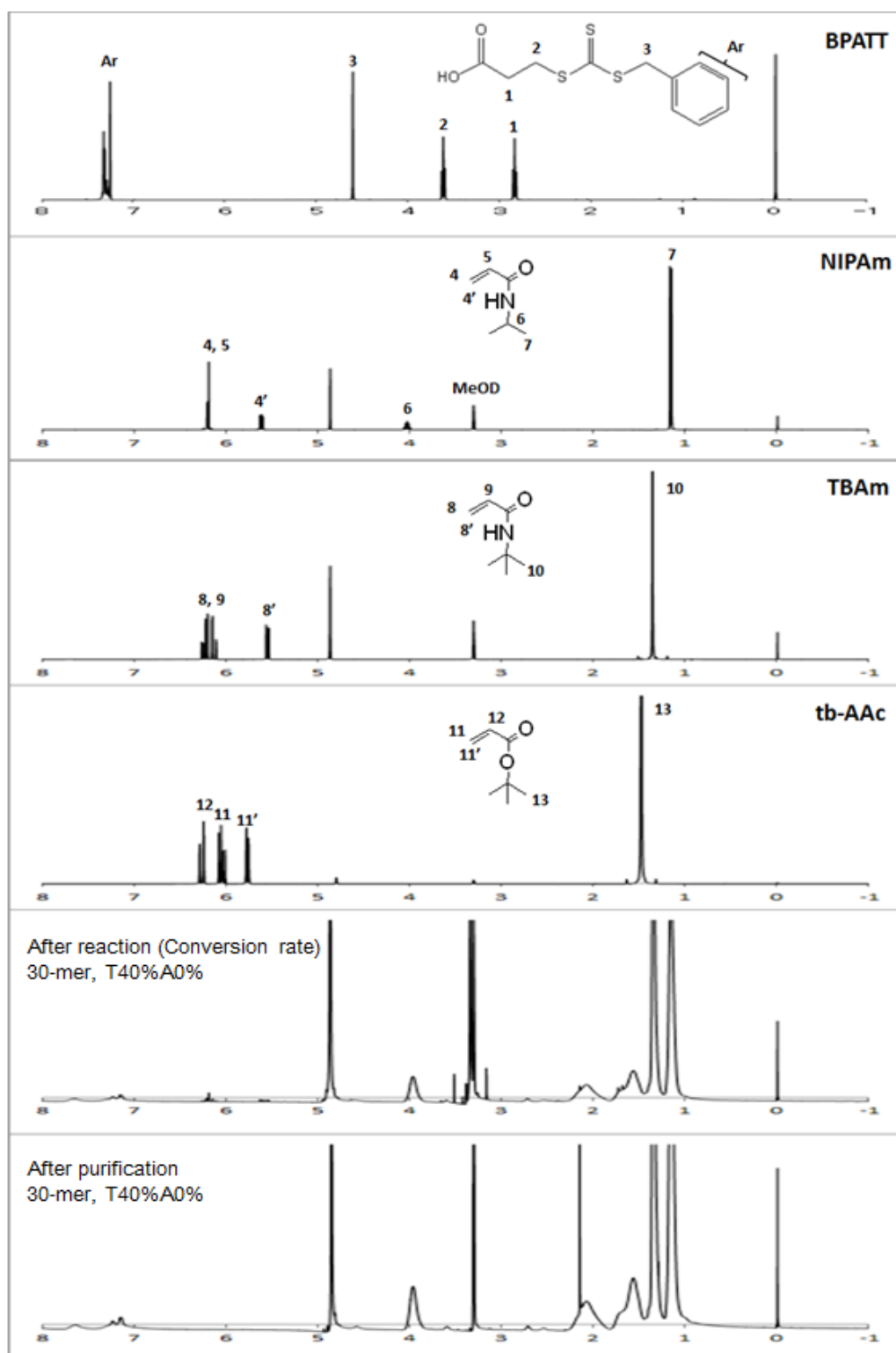
**Figure S1-8.**  $^1\text{H}$  NMR of 30-mer PL, containing 20 % TBAm and 5 % *tb*-AAc



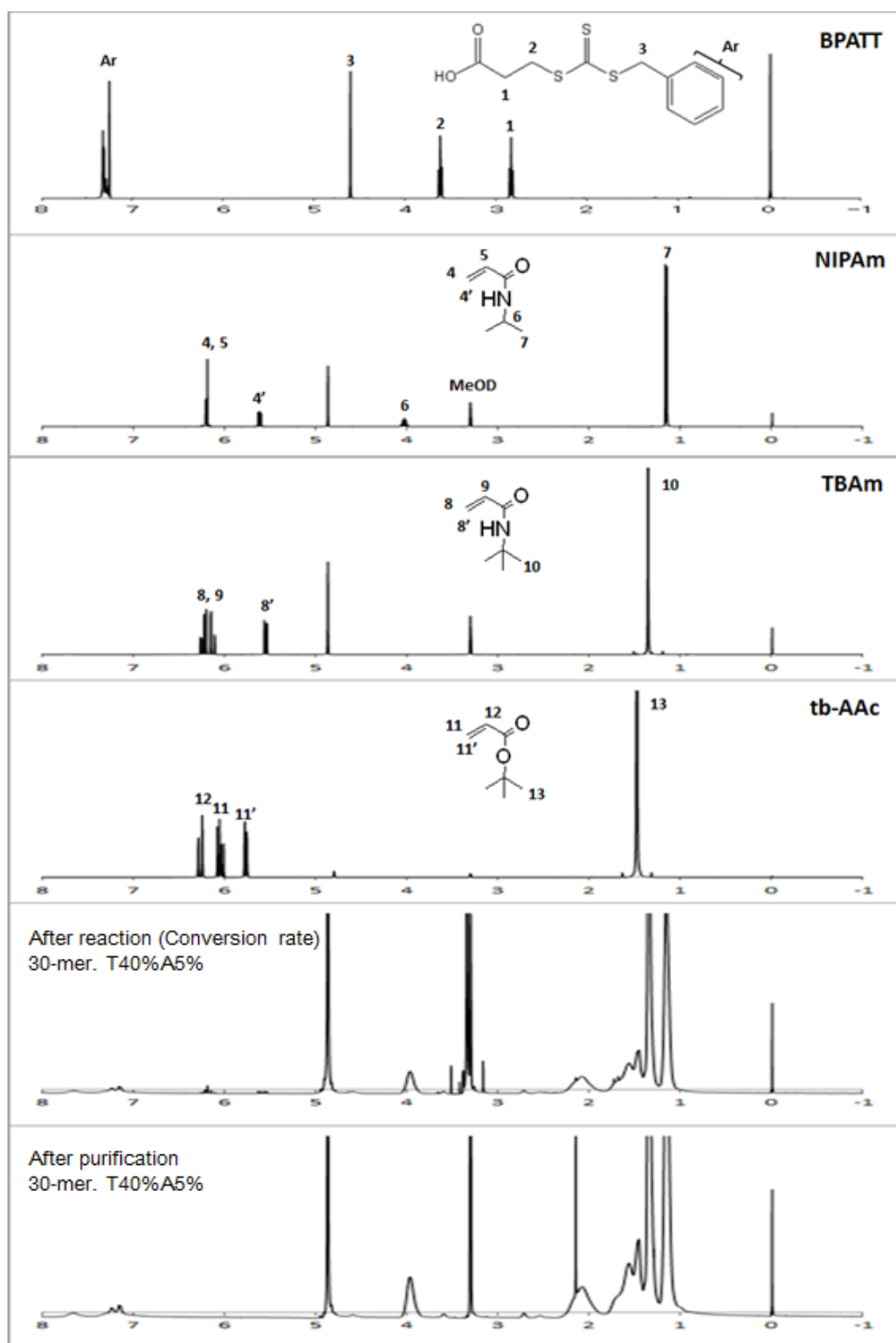
**Figure S1-9.**  $^1\text{H}$  NMR of 30-mer PL, containing 20 % TBAm and 10 % *tb*-AAc



**Figure S1-10.**  $^1\text{H}$  NMR of 30-mer PL, containing 20 % TBAm and 20 % *tb*-AAc

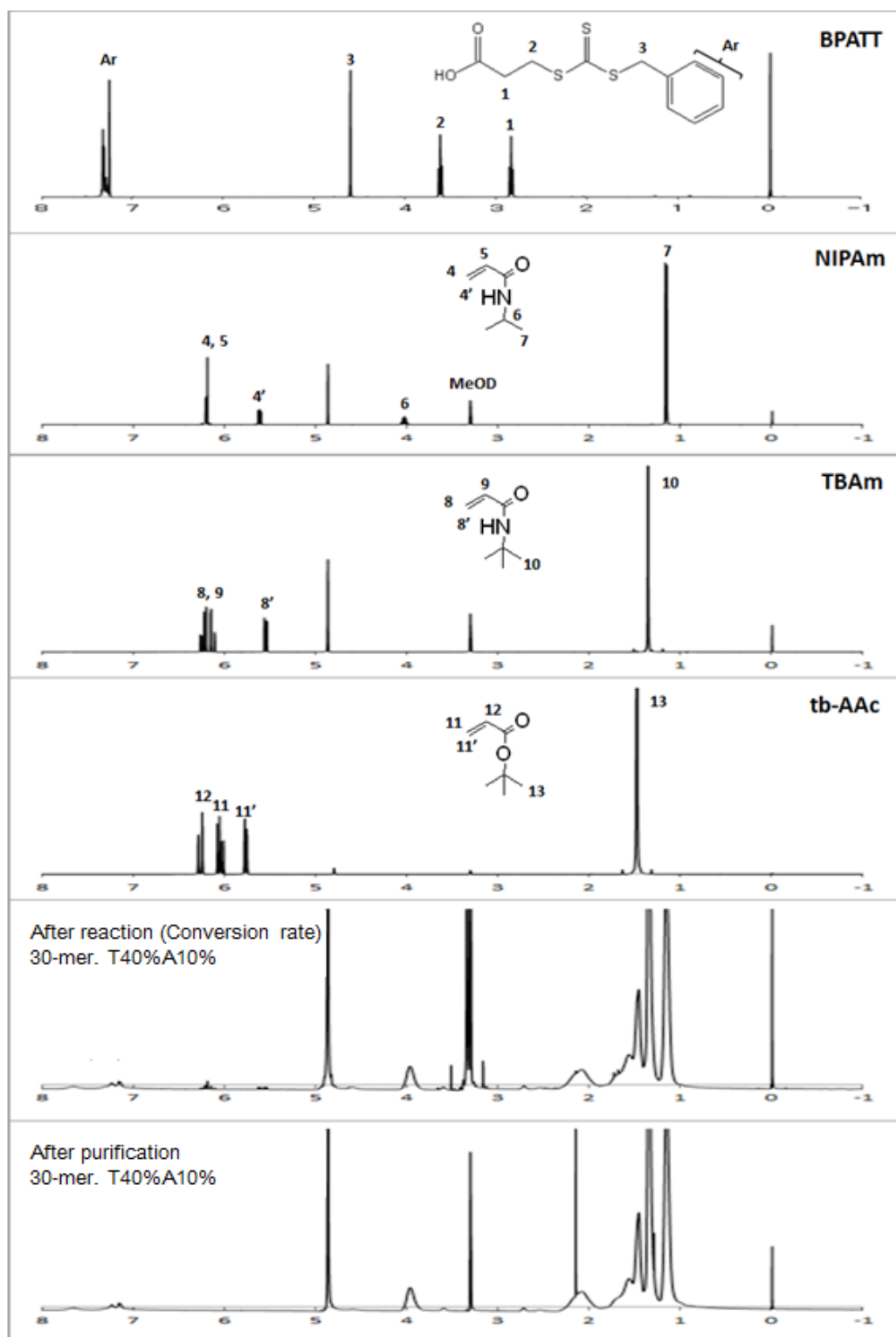


**Figure S1-11.**  $^1\text{H}$  NMR of 30-mer PL, containing 40 % TBAm and 0 % *tb*-AAc

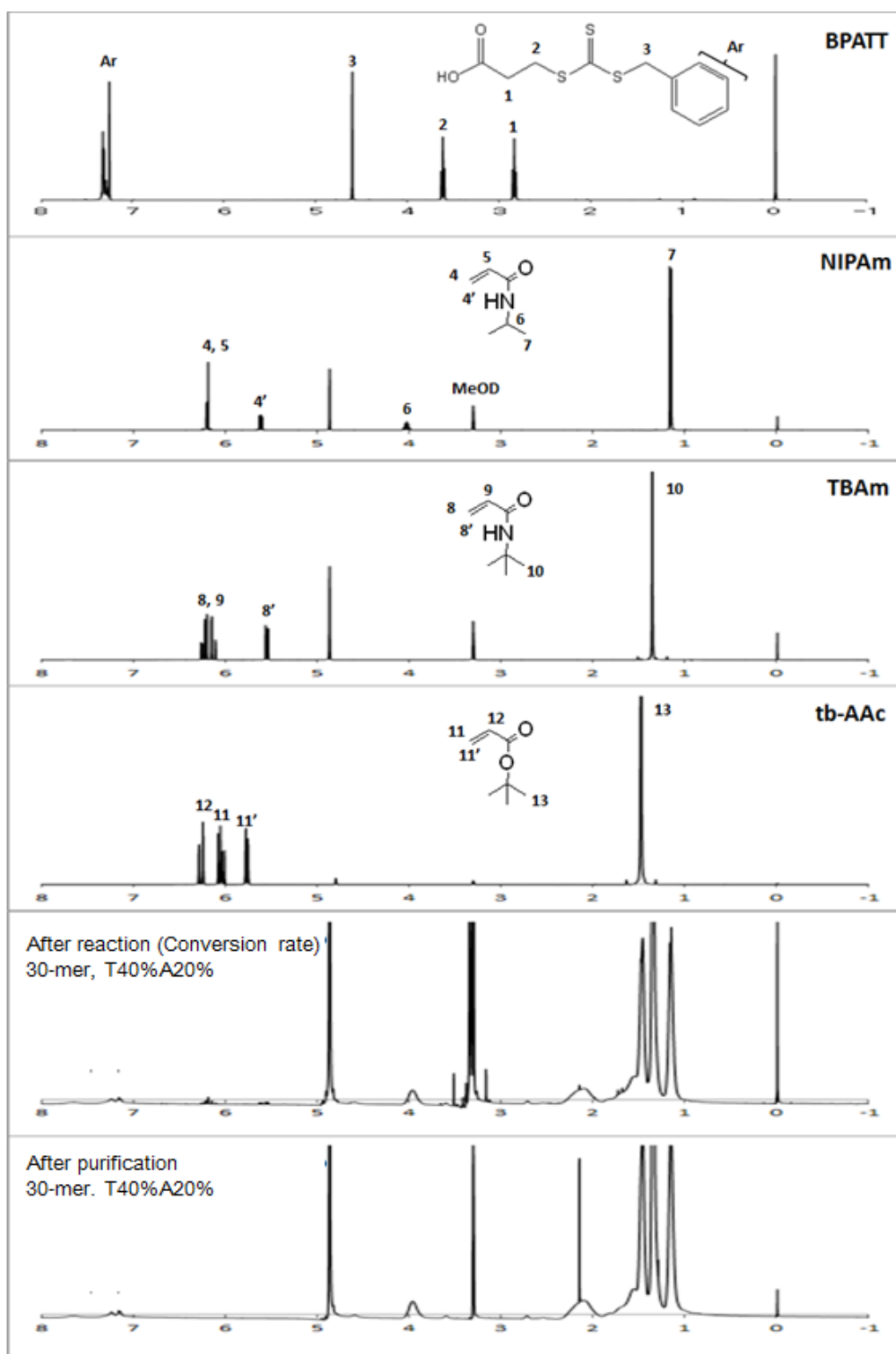


**Figure S1-12.**  $^1\text{H}$  NMR of 30-mer PL, containing 40 % TBAm and 5 % *tb*-AAc

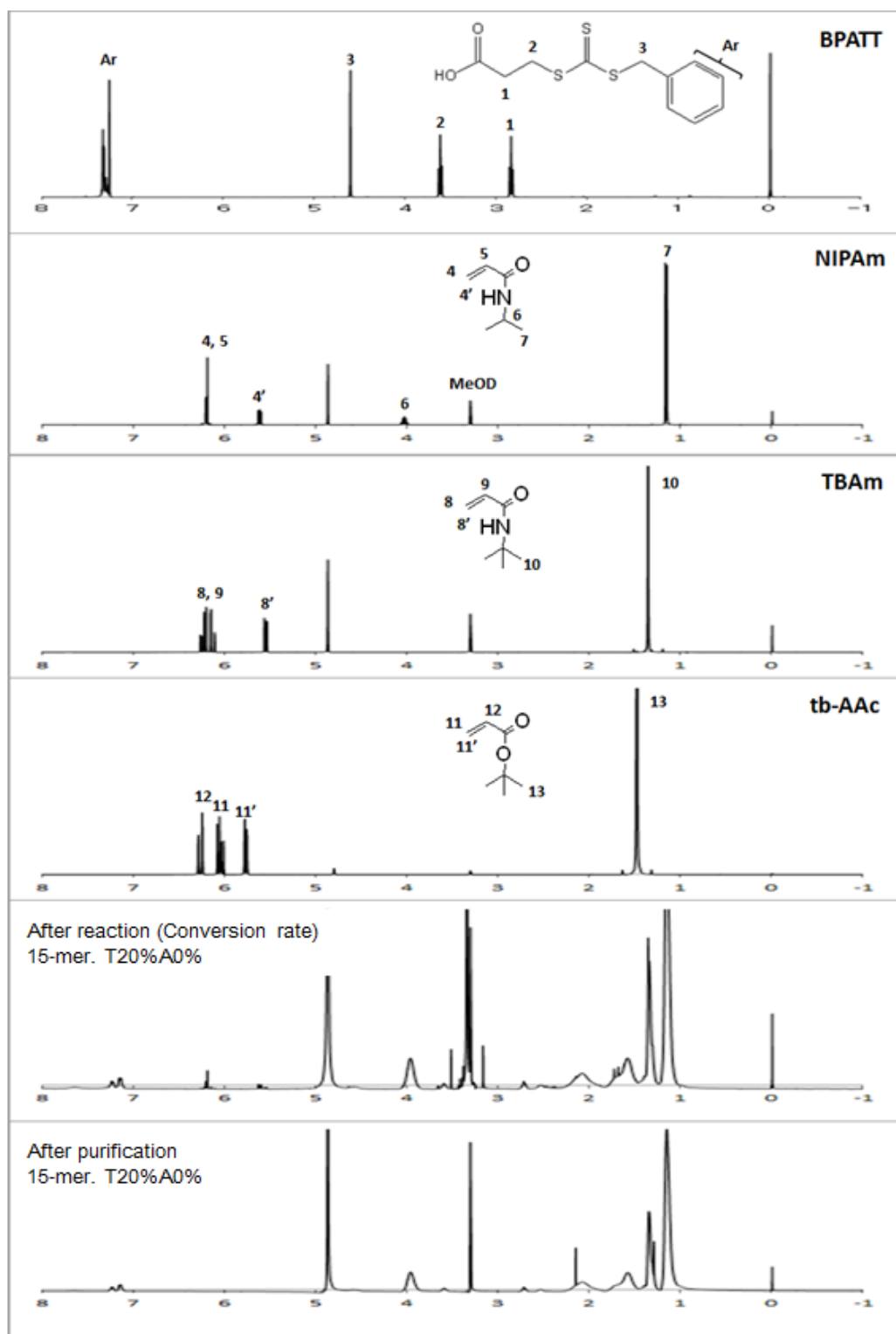




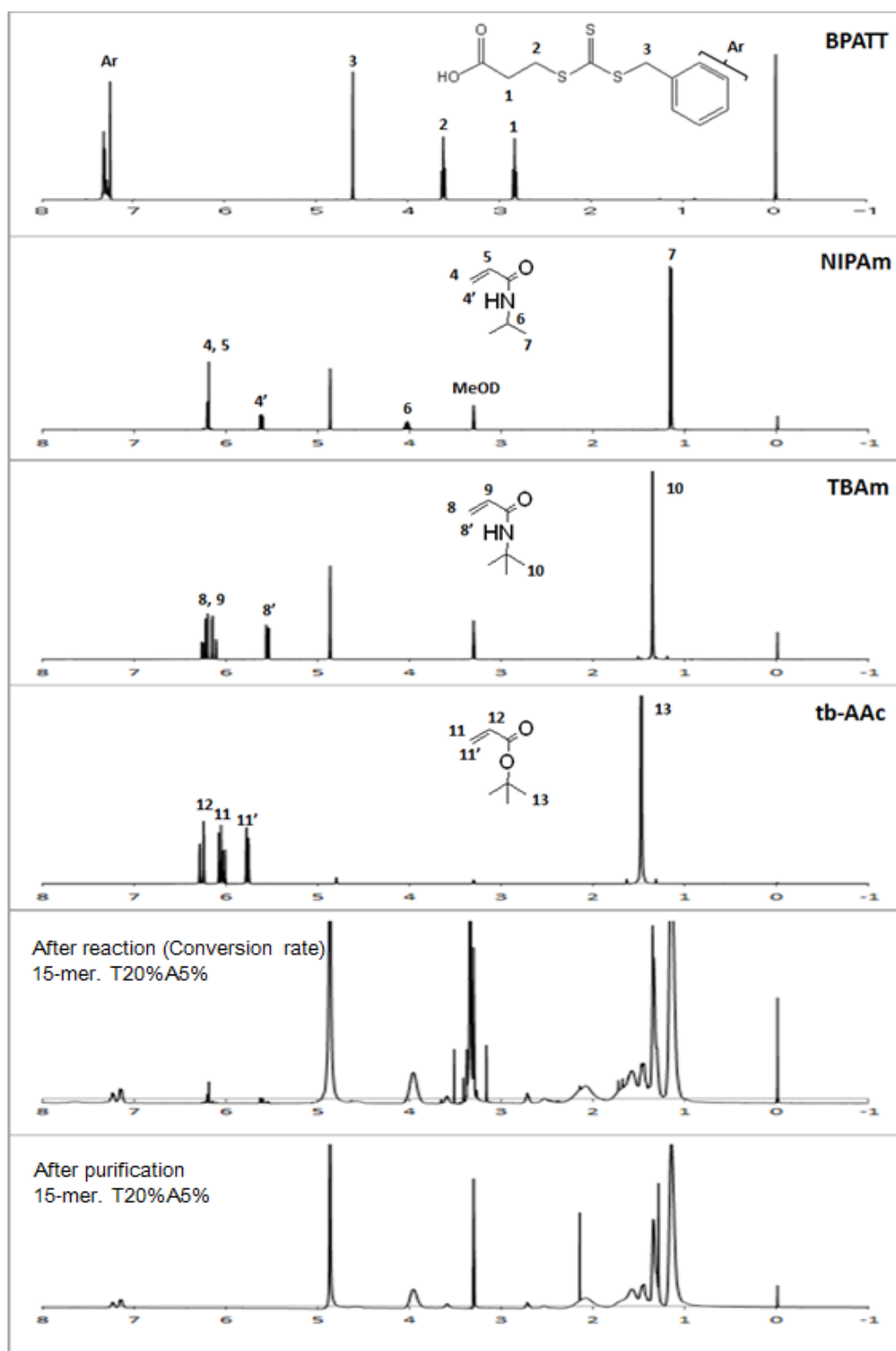
**Figure S1-13.**  $^1\text{H}$  NMR of 30-mer PL, containing 40 % TBAm and 10 % *tb*-AAc



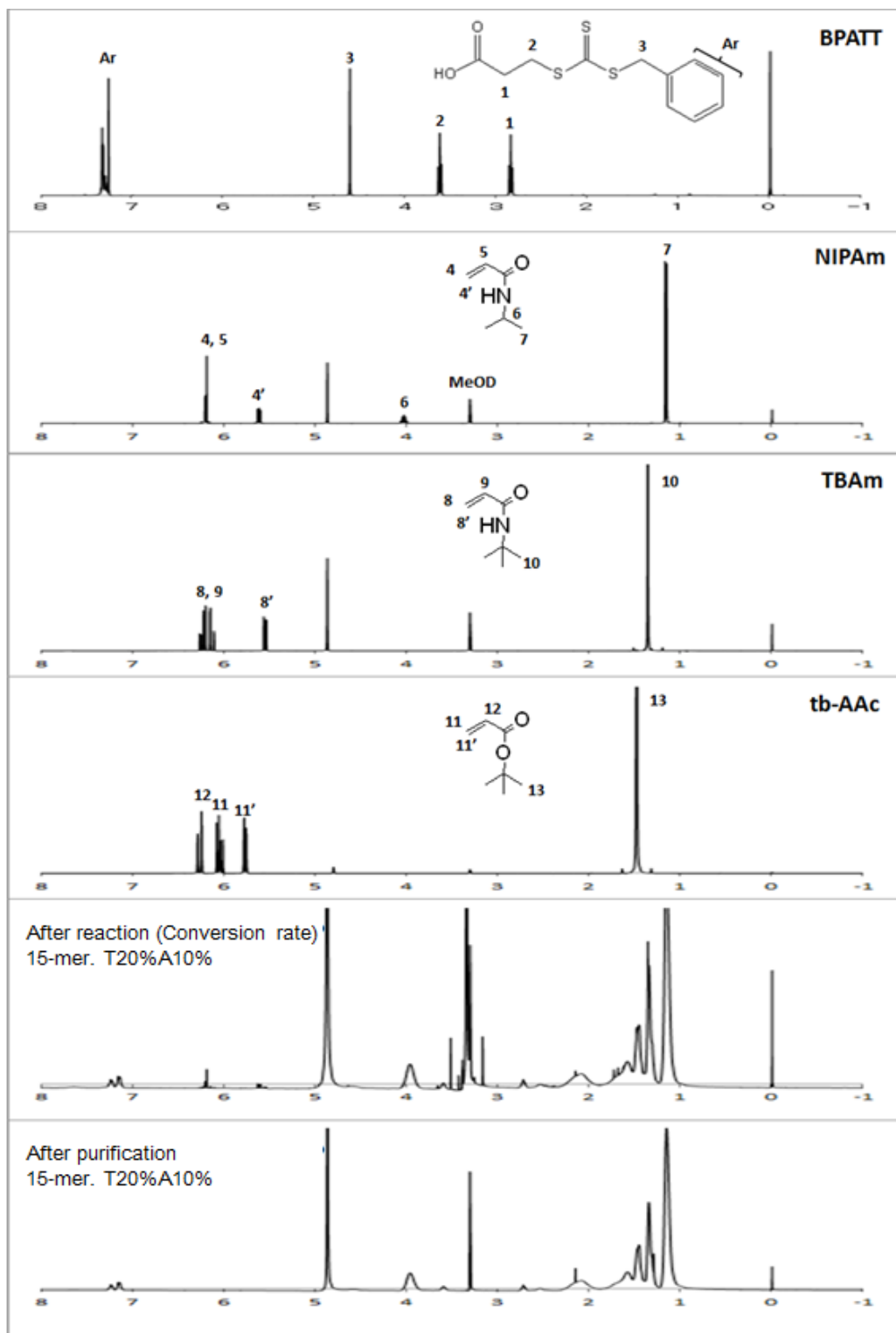
**Figure S1-14.**  $^1\text{H}$  NMR of 30-mer PL, containing 40 % TBAm and 20 % *tb*-AAc



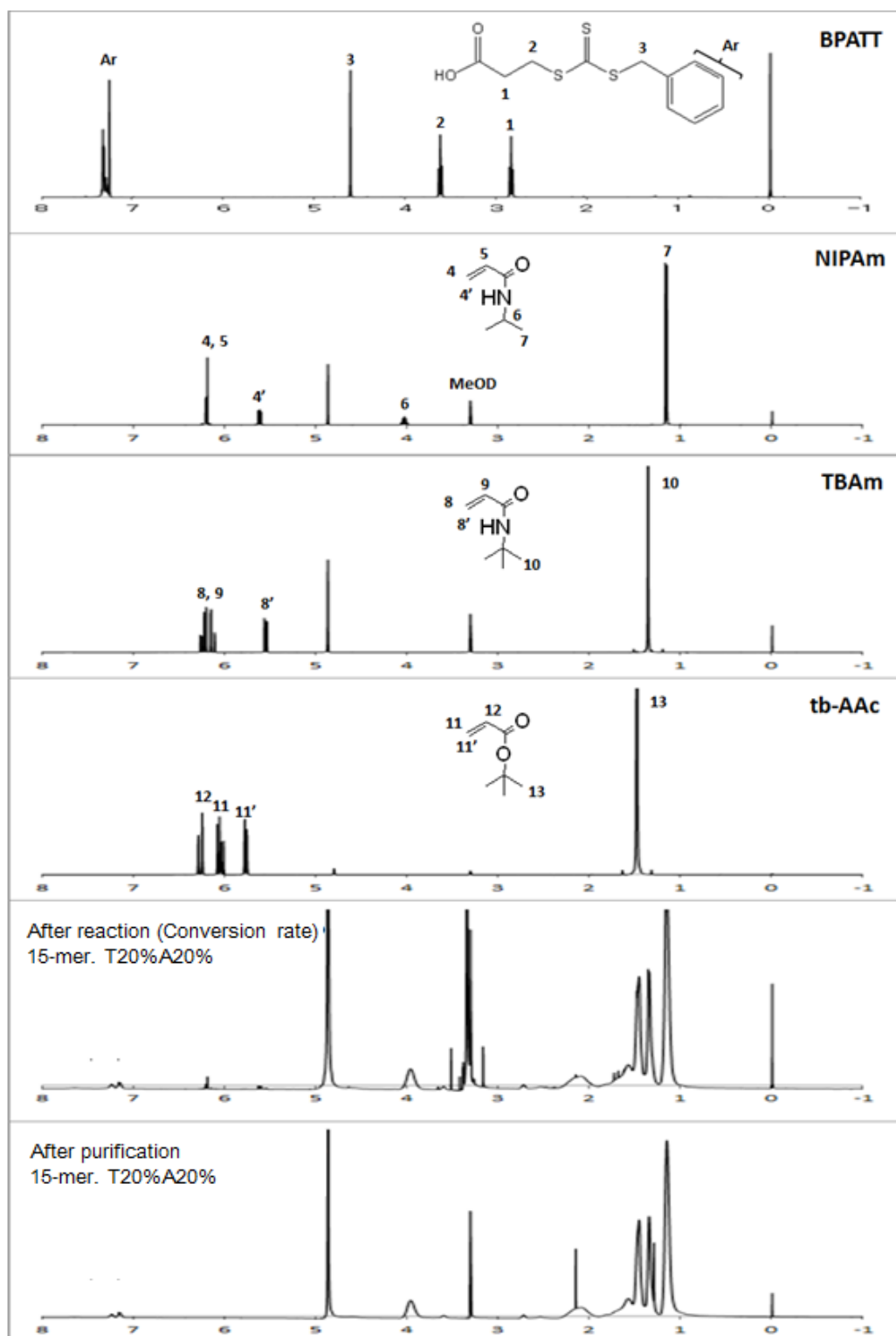
**Figure S1-15.**  $^1\text{H}$  NMR of 15-mer PL, containing 20 % TBAm and 0 % *tb*-AAc



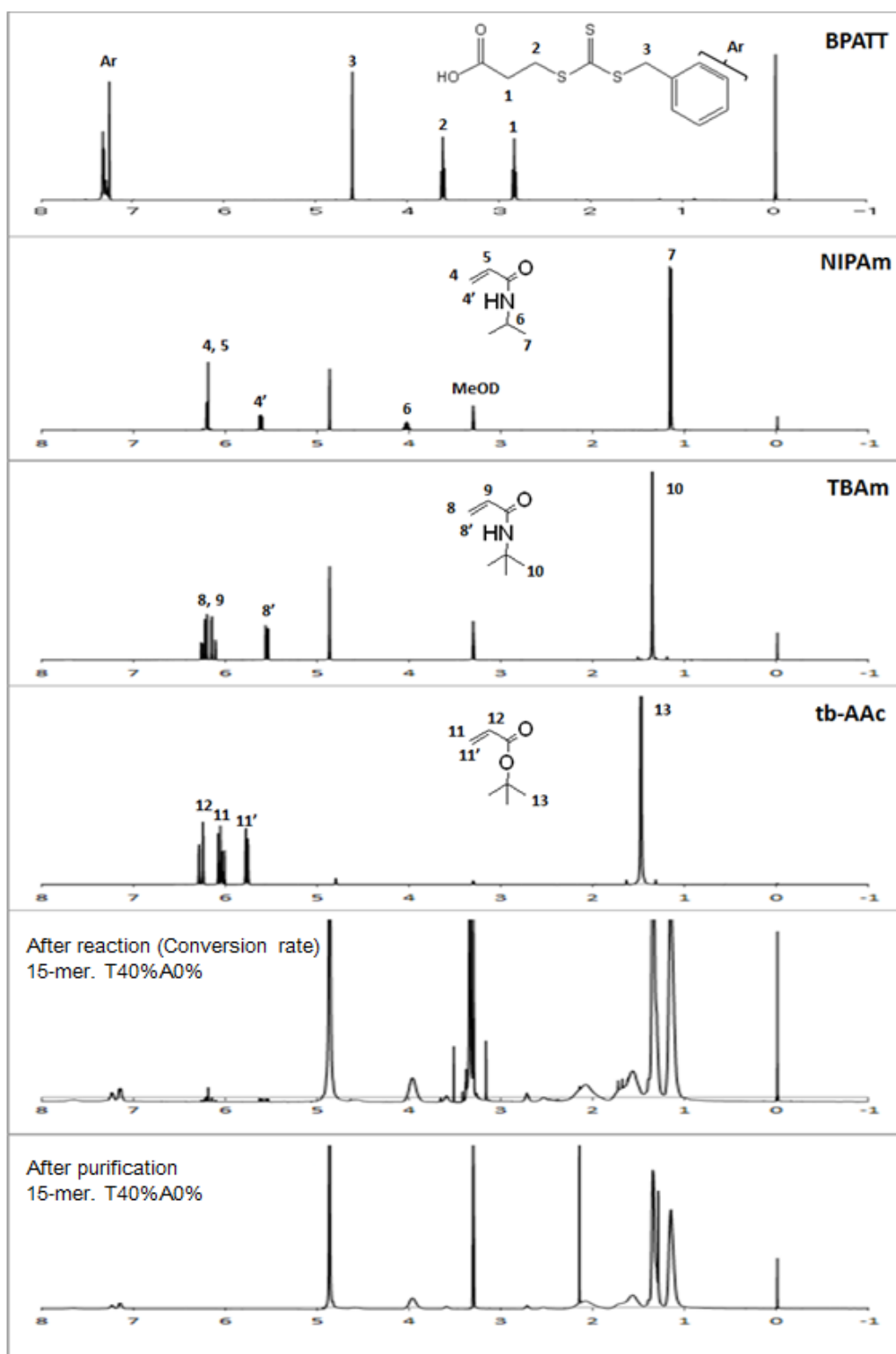
**Figure S1-16.**  $^1\text{H}$  NMR of 15-mer PL, containing 20 % TBAm and 5 % *tb*-AAc



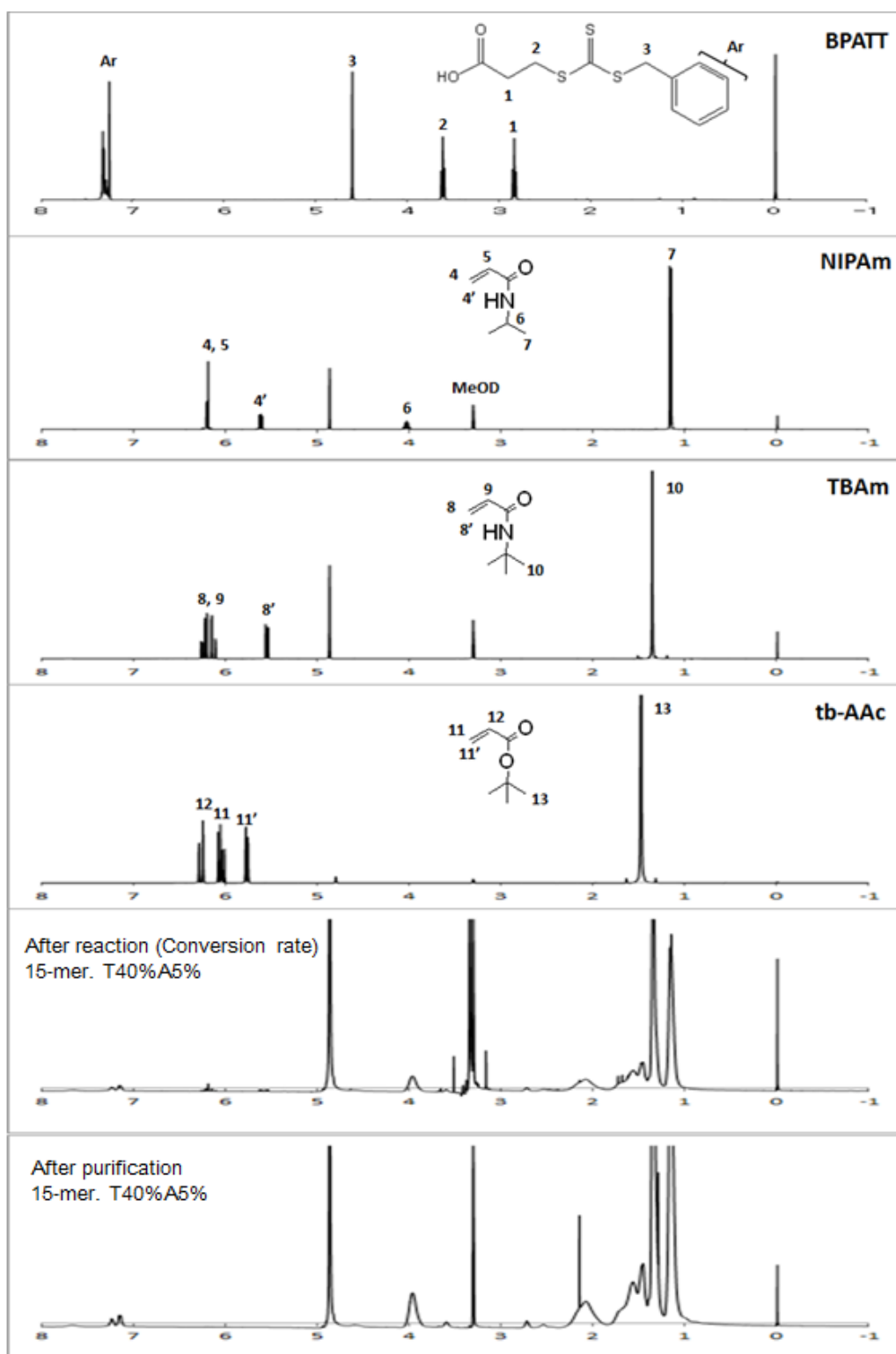
**Figure S1-17.**  $^1\text{H}$  NMR of 15-mer PL, containing 20 % TBAm and 10 % *tb*-AAc



**Figure S1-18.**  $^1\text{H}$  NMR of 15-mer PL, containing 20 % TBAm and 20 % *tb*-AAc

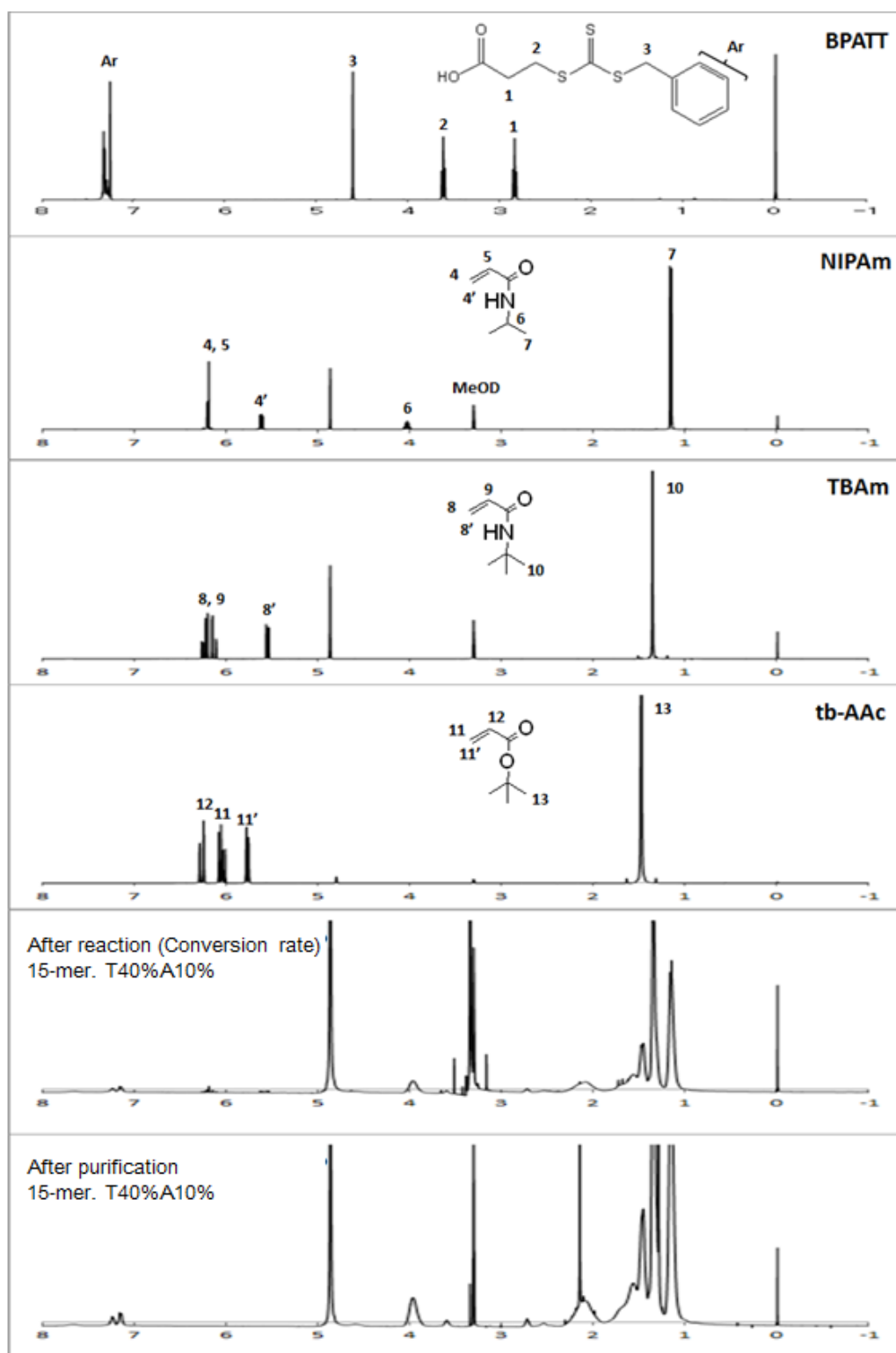


**Figure S1-19.**  $^1\text{H}$  NMR of 15-mer PL, containing 40 % TBAm and 0 % *tb*-AAc

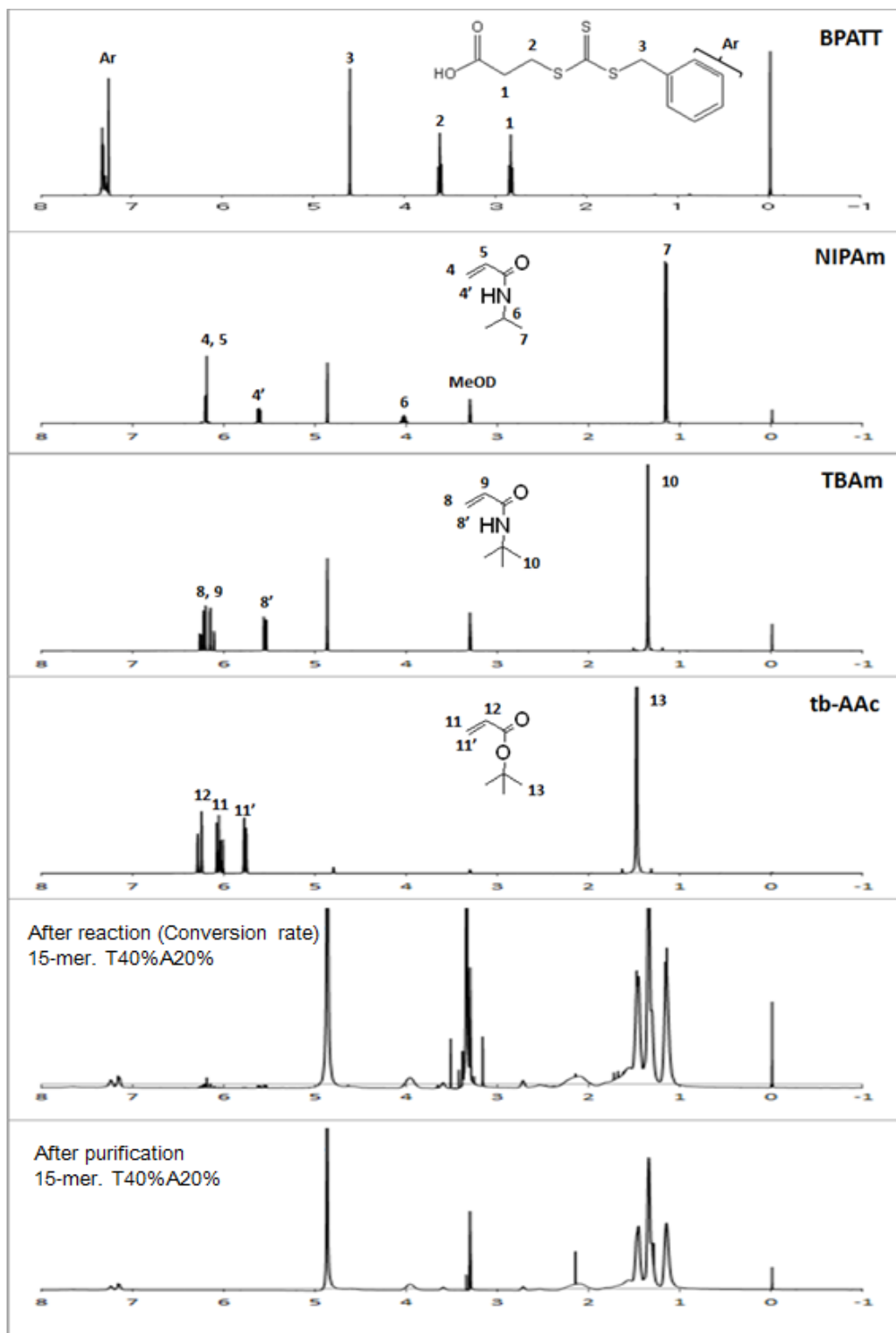


**Figure S1-20**  $^1\text{H}$  NMR of 15-mer PL, containing 40 % TBAm and 5 % *tb*-AAc





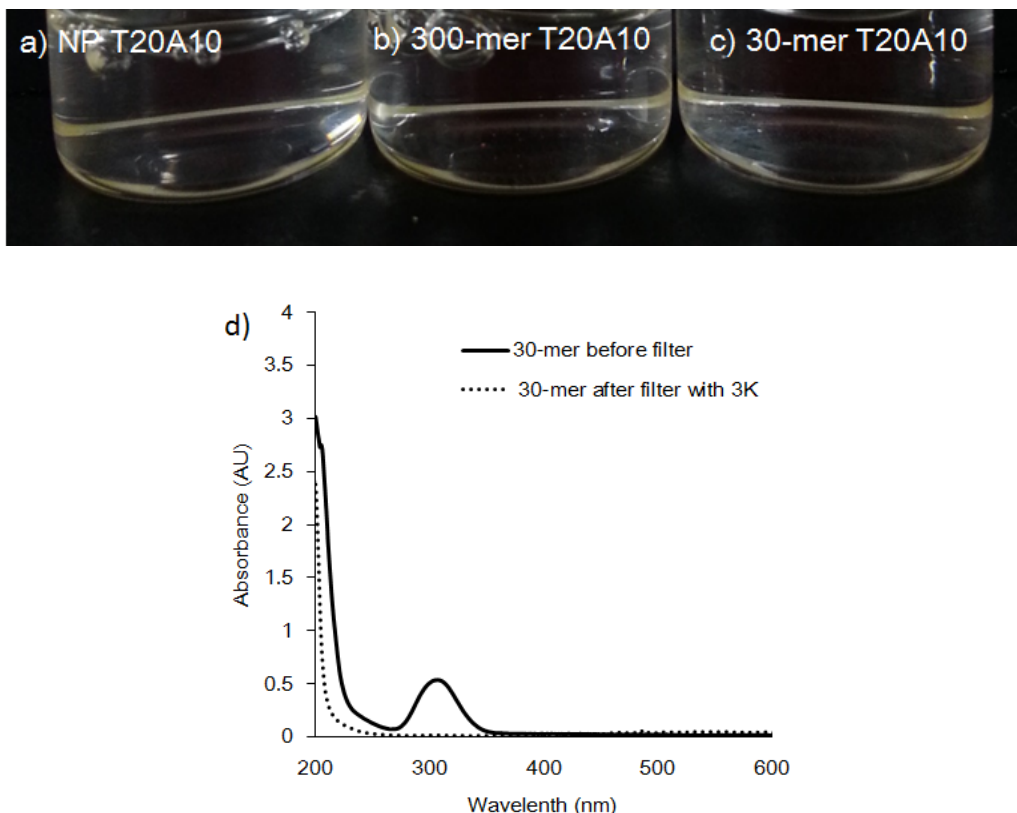
**Figure S1-21.**  $^1\text{H}$  NMR of 15-mer PL, containing 40 % TBAm and 10 % *tb*-AAc



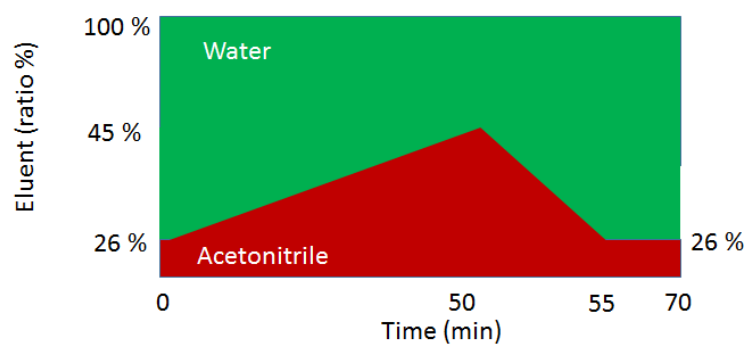
**Figure S1-22.**  $^1\text{H}$  NMR of 15-mer PL, containing 40 % TBAm and 20 % *tb*-AAc

## S2. HPLC analysis of peptides unbound by synthetic PL

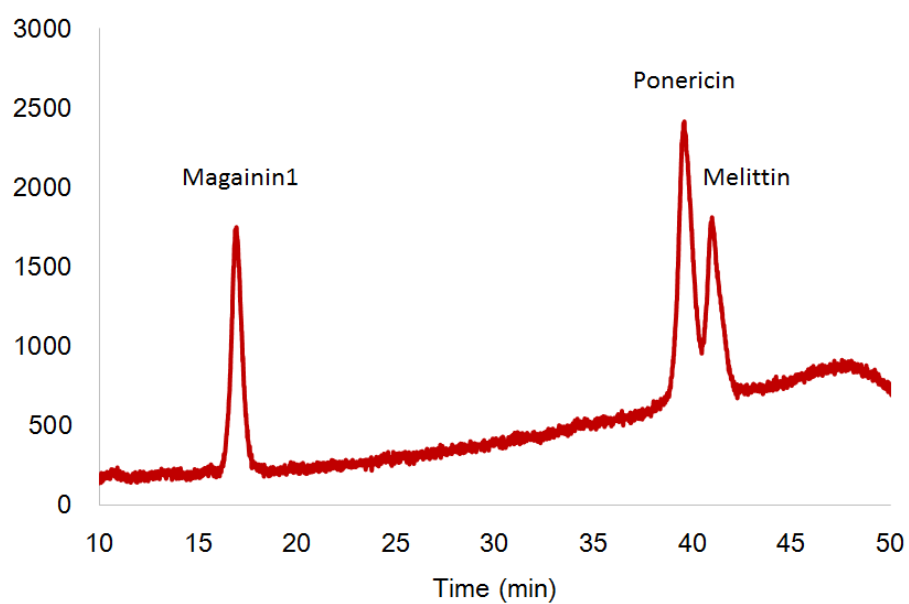
All peptides and polymers were dissolved in 35 mM PBS (0.15 M NaCl, pH 7.3) and stored at 4°C. It was confirmed that all PLs were soluble in the buffer and the solutions were transparent even after incubation at 37 °C for 0.5 hr (Figure S2-0 a-c). 1.9 mg/mL of NP, 300-, and 30-mer PLs were added into the peptide mixed solutions (melittin, ponericin and magainin1, 0.1 mM each) in 35 mM PBS (0.15 M NaCl, pH 7.3) and incubated at 37°C for 0.5 hr. Then, the incubated solutions were filtered with centrifugal filter system for 0.5 hr (Milipore Co., Amicon Ultra-0.5, NMWL 10K, 8000 G at 37°C). We confirmed that PLs were separated from the solution by this filtration process (Figure S2-0 d). After centrifugation, 7  $\mu$ L of the filtrate were injected on a CAPCELL PAK C18 column for separation by high performance liquid chromatography (HPLC). The mobile phase consisted of 0.1 % TFA dissolved in acetonitrile/water. The linear elution gradient is illustrated in **Figure S2-1**. Flow rate of the mobile phase was 1 mL/min. The column was kept at 37 °C, and elution was monitored at 220 nm.



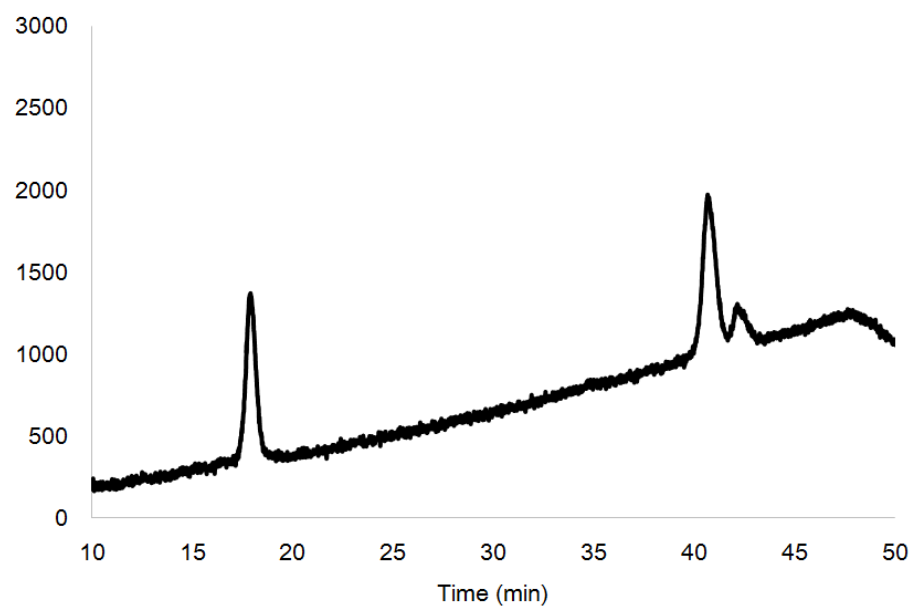
**Figure S2-0.** a-c Photographs of polymer solutions (0.1 mM) after incubation at 37°C for 0.5 hr. d UV-vis spectra of buffer solution of 30-mer PLs before and after filtration.



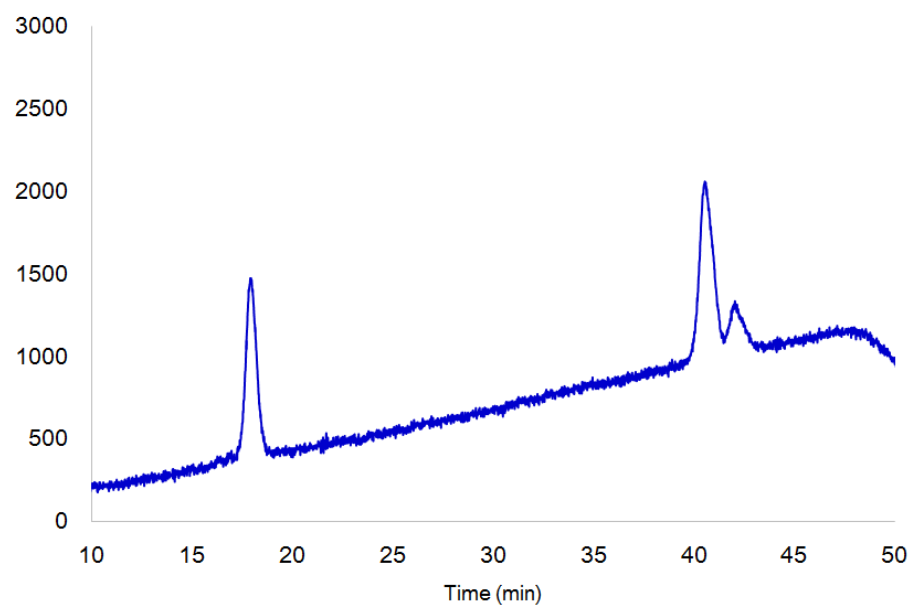
**Figure S2-1.** HPLC scheme



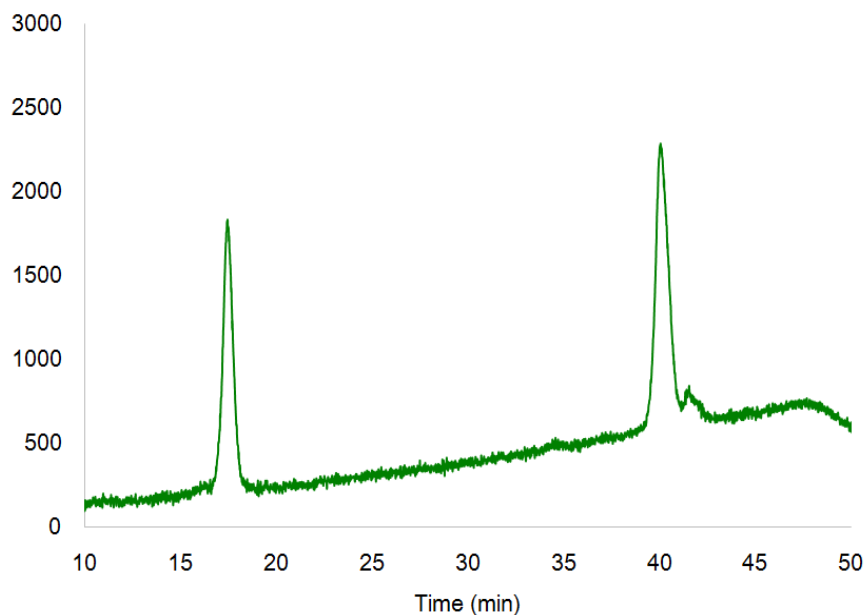
**Figure S2-2.** A mixture of free lytic peptides, consisting of magainin 1, ponericin, and melittin (concentration of peptides: 0.1 mM each).



**Figure S2-3.** Peptides unbound by NP (T20%A10%)  
(concentration of peptides: 0.1 mM each, concentration of NP: 1.9 mg/mL)



**Figure S2-4.** Peptides not bound by 300-mer PL (T20%A10%)  
(concentration of peptides: 0.1 mM each, concentration of 300-mer PL: 1.9 mg/mL)

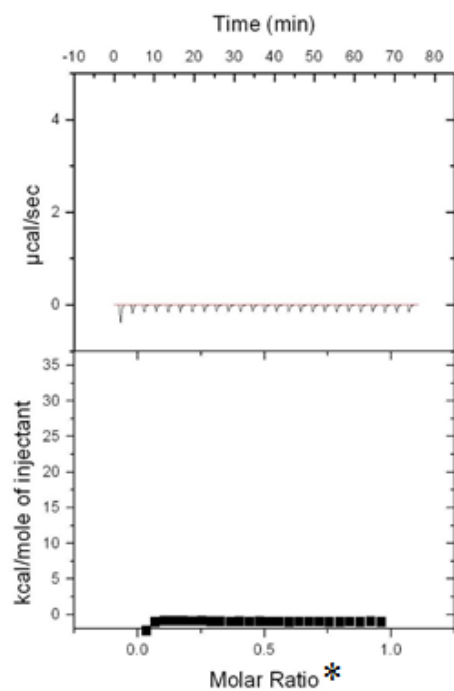


**Figure S2-5.** Peptides not bound by 30-mer PL (T20%A10%)  
(concentration of peptides: 0.1 mM each, concentration of 30-mer PL: 1.9 mg/mL)

### S3. ITC titration of magainin 1 into synthetic PL

#### [General ITC titration procedure]

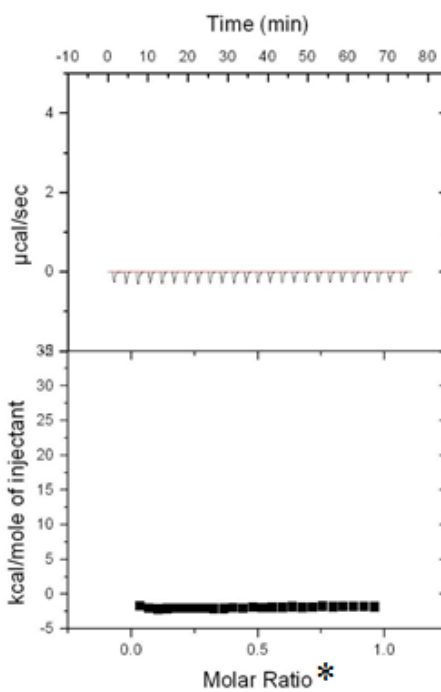
All peptides and polymers were dissolved in 35 mM PBS (0.15 M NaCl, pH 7.3) and stored at 4°C. After degassing the polymer ligands solution (0.38 mg/mL), it was loaded into a ITC reaction cell and the reference cell was loaded with degassed MilliQ water. The instrument was equilibrated at 37°C until the baseline was flat and stable. Peptide solution (0.5 mM) was degassed and loaded in syringe. Each titration was programmed for 25 injections with 10  $\mu$ L. The spacing between injections was 150 s. The reference power was 10  $\mu$ cal/s. Titrations were conducted at 37°C with a stirring speed of 310 rpm. The raw data was analyzed by ORIGIN software.



**Figure S3-1.** ITC titration of magainin 1 into NP (T20%A10%)

Cell	0.38 mg/mL NP with 20 %TBAm and 10 % AAc in 35 mM PBS (pH 7.3, 0.15M NaCl)
syringe	0.5 mM magainin 1 in 35 mM PBS (pH 7.3, 0.15M NaCl)

\* Apparent molar ratio

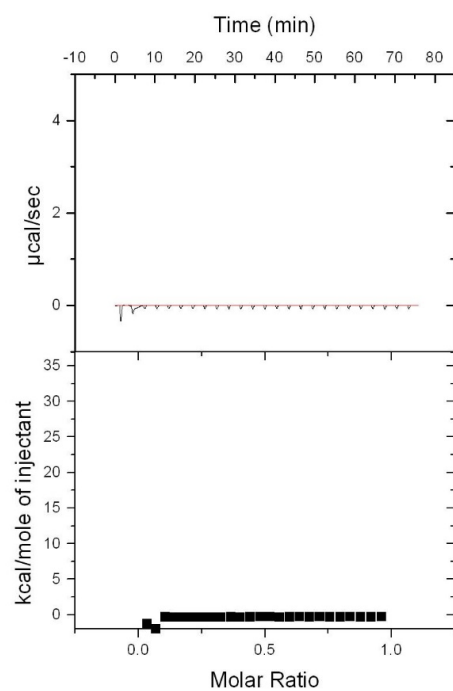


**Figure S3-2.** ITC titration of magainin 1 into 300-mer PL (T20%A10%)

Cell	0.38 mg/mL 300-mer PL with 20 % TBAm and 10 % AAc in 35 mM PBS (pH 7.3, 0.15M NaCl)
syringe	0.5 mM magainin 1 in 35 mM PBS (pH 7.3, 0.15M NaCl)

\* Apparent molar ratio

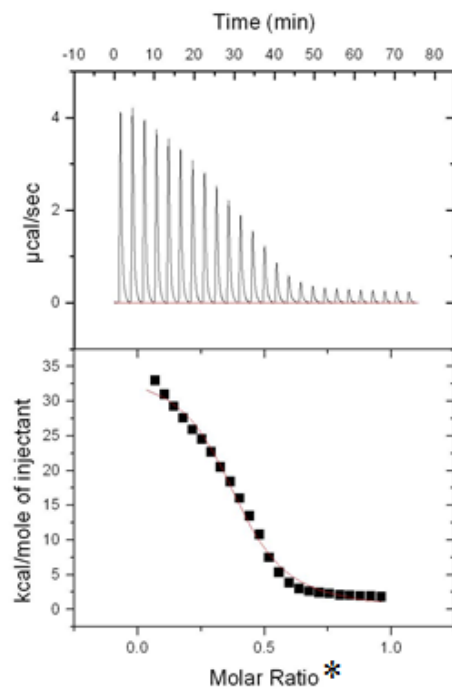




**Figure S3-3.** ITC titration of magainin 1 into 30-mer PL (T20%A10%)

Cell	0.38 mg/mL 30-mer PL with 20 % TBAm and 10 % AAc in 35 mM PBS (pH 7.3, 0.15M NaCl)
syringe	0.5 mM magainin 1 in 35 mM PBS (pH 7.3, 0.15M NaCl)

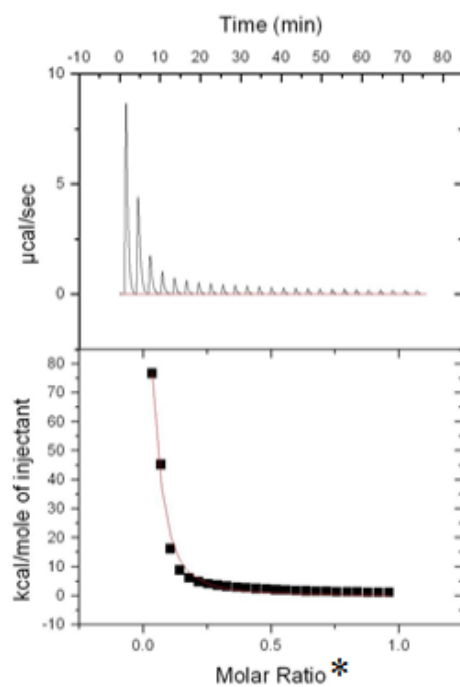
## S4. ITC titration of melittin or ponericin into synthetic PL



**Figure S4-1.** ITC titration of melittin into NP (T20%A10%)

Cell	0.38 mg/mL NP that contains 20 % TBAm and 10 % AAc in 35 mM PBS (pH 7.3, 0.15M NaCl)
syringe	0.5 mM melittin in 35 mM PBS (pH 7.3, 0.15M NaCl)

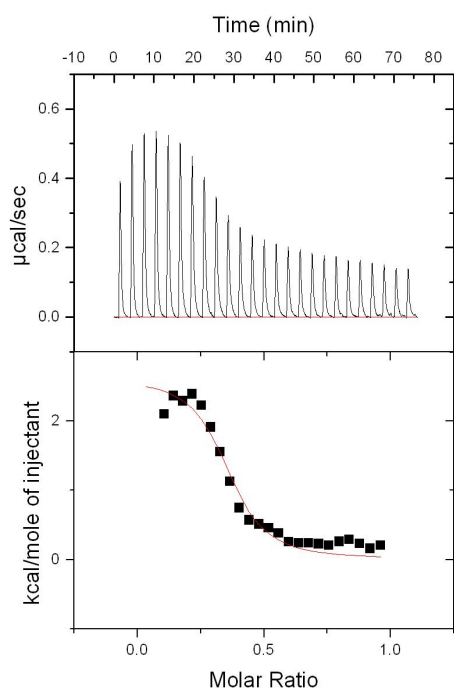
\* Apparent molar ratio



**Figure S4-2.** ITC titration of melittin into 300-mer PL (T20%A10%)

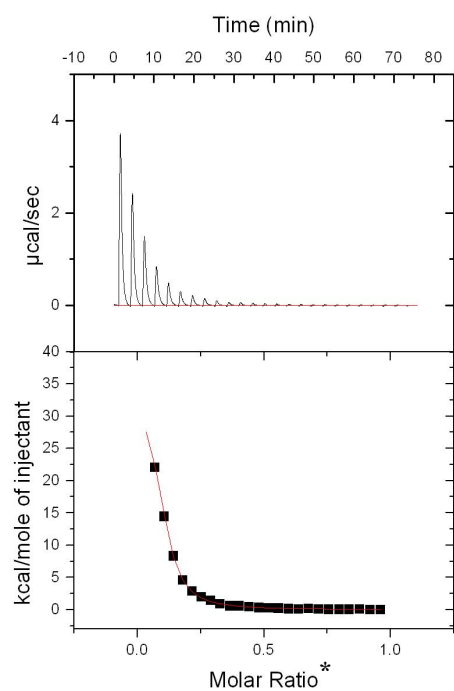
Cell	0.38 mg/mL 300-mer with 20 % TBA <sub>m</sub> and 10 % AAc in 35 mM PBS (pH 7.3, 0.15M NaCl)
syringe	0.5 mM melittin in 35 mM PBS (pH 7.3, 0.15M NaCl)

\* Apparent molar ratio



**Figure S4-3.** ITC titration of melittin into 30-mer PL (T20%A10%)

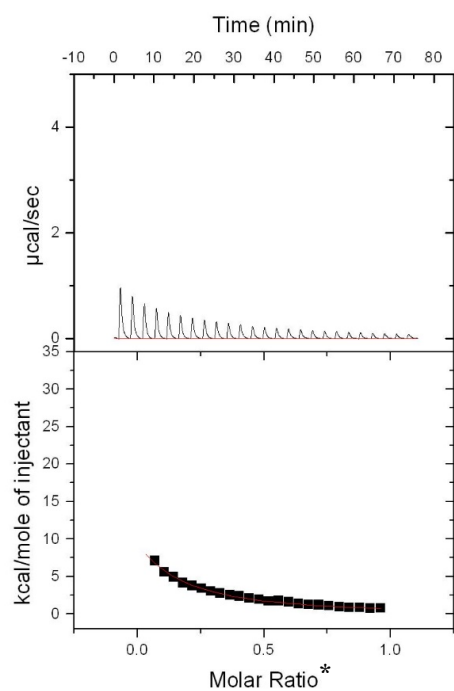
Cell	0.38 mg/mL 30-mer PL with 20 % TBAm and 10 % AAc in 35 mM PBS (pH 7.3, 0.15M NaCl)
syringe	0.5 mM melittin in 35 mM PBS (pH 7.3, 0.15M NaCl)



**Figure S4-4.** ITC titration of ponicin into NP (T20%A10%)

Cell	0.38 mg/mL NP functionalized with 20 % TBAm and 10 % AAc in 35 mM PBS (pH 7.3, 0.15M NaCl)
syringe	0.5 mM ponicin in 35 mM PBS (pH 7.3, 0.15M NaCl)

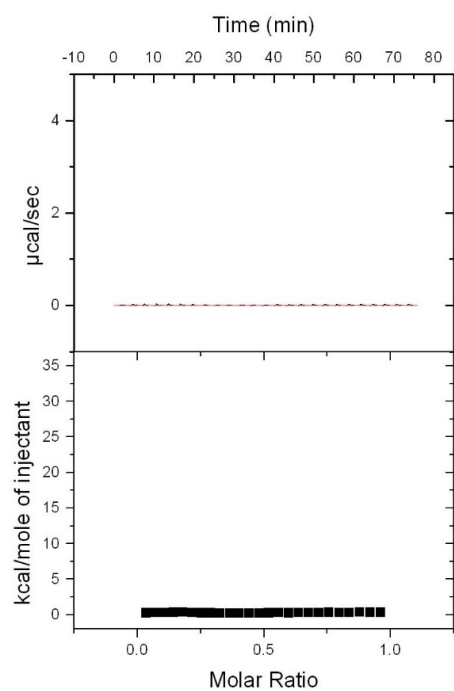
\* Apparent molar ratio



**Figure S4-5.** ITC titration of ponericin into 300-mer PL (T20%A10%)

Cell	0.38 mg/mL 300-mer PL with 20 % TBAm and 10 % AAc in 35 mM PBS (pH 7.3, 0.15M NaCl)
syringe	0.5 mM ponericin in 35 mM PBS (pH 7.3, 0.15M NaCl)

\* Apparent molar ratio



**Figure S4-6.** ITC titration of ponericin into 30-mer PL (T20%A10%)

Cell	0.38 mg/mL 30-mer PL with 20 % TBAm and 10 % AAc in 35 mM PBS (pH 7.3, 0.15M NaCl)
syringe	0.5 mM ponericin in 35 mM PBS (pH 7.3, 0.15M NaCl)

## S5. Hemolysis neutralization assay

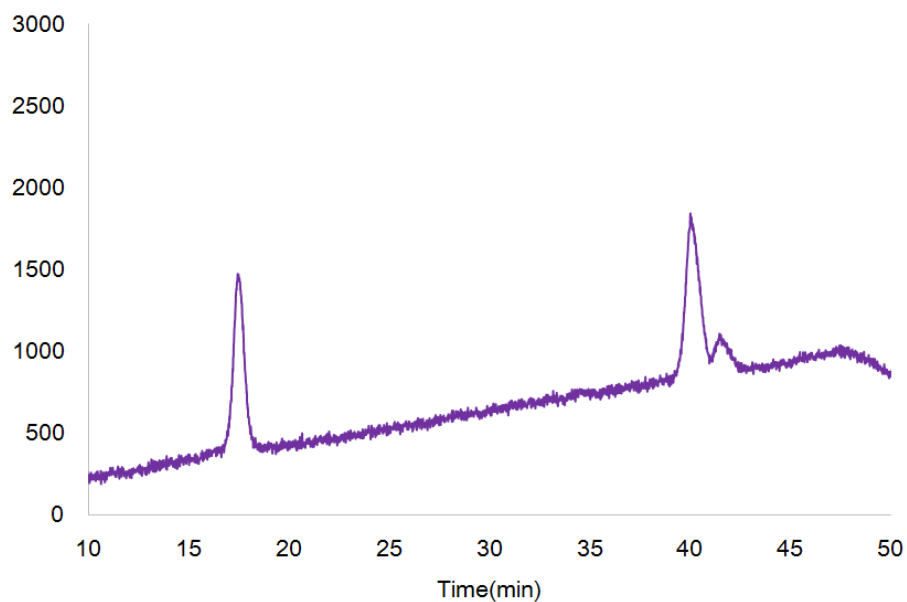
Red blood cells (RBC) were washed with PBS, collected by centrifugation for 10 min at 800× *g*, and then resuspended in PBS. The RBC was then incubated with PLs for 30 min at 37 °C in PBS. A mixture of melittin and PL was then added to 100 mL cells to a final volume of 200 mL. The resulting suspension was incubated at 37 °C for 30 min. Samples were then centrifuged at 800× *g* for 10 min. Released hemoglobin was measured by measuring the absorbance of the supernatant at 415 nm ( $Abs_{\text{polymer ligand}}$ ). Control values for 0% neutralization and 100% neutralization were obtained, respectively, from cells incubated with 1.8 mM melittin only ( $Abs_{0\%}$ ) and RBC only ( $Abs_{100\%}$ ). Percent neutralization was calculated according to S6.

## S6. Formula for % neutralization (E1)

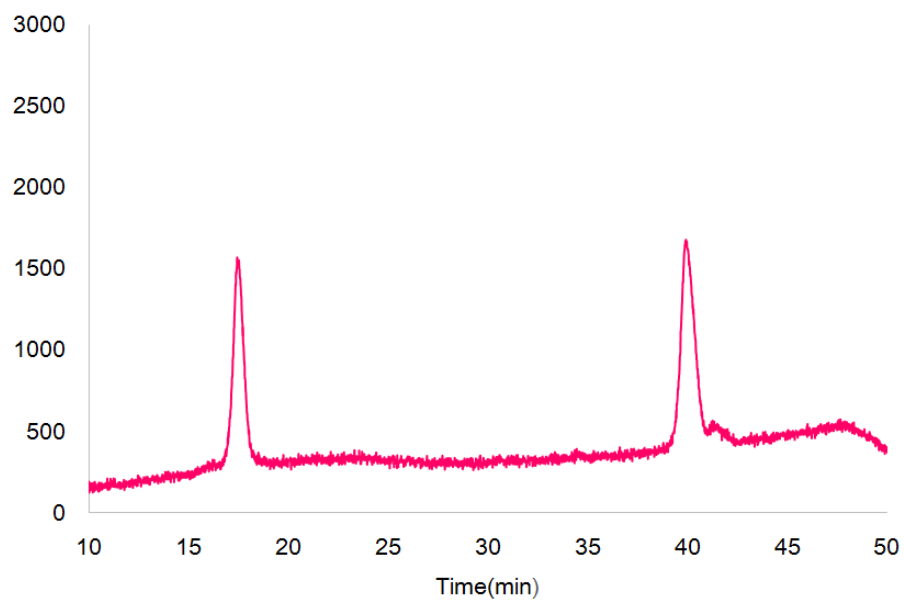
$$\text{Neutralization \%} = \frac{Abs_{100\%} - Abs_{\text{Polymer ligand}}}{Abs_{100\%} - Abs_{0\%}} \times 100$$



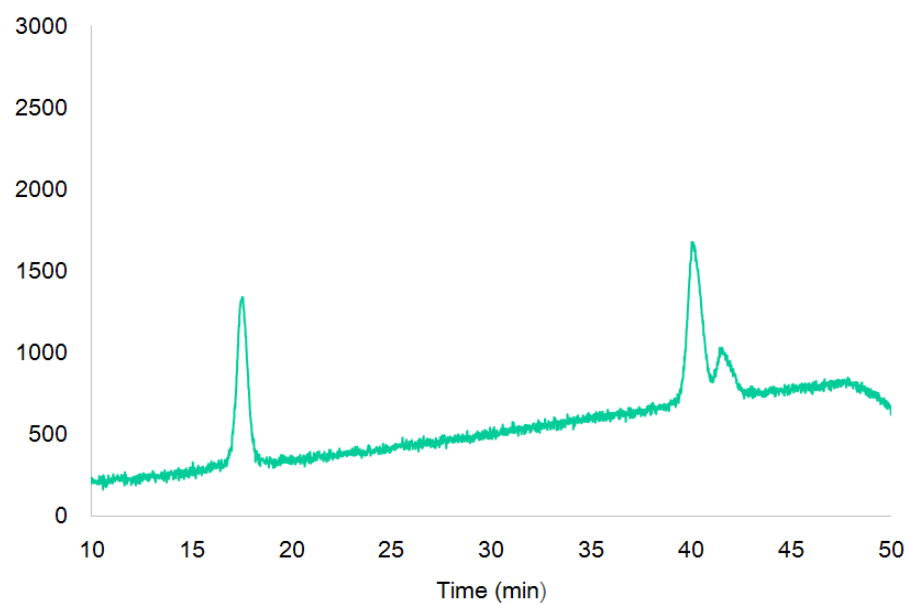
## S7. HPLC analysis of lytic peptides unbound by low-MW PL



**Figure S7-1.** Peptides unbound by 30-mer PL (T20%A20%)  
(concentration of peptides: 0.1 mM each, concentration of 30-mer PL: 1.9 mg/mL)



**Figure S7-2.** Peptides unbound by 30-mer PL (T40%A10%)  
(concentration of peptides: 0.1 mM each, concentration of 30-mer PL: 1.9 mg/mL)



**Figure S7-3.** Peptides unbound by 15-mer PL (T40%A20%)  
(concentration of peptides: 0.1 mM each, concentration of 15-mer PL: 1.9 mg/mL)

#### Reference

1. Stenzel, M.H.; Davis, T. P. *J. Polym. Sci. A.*, **2002**, *40*, 4498-4512