

# Stereospecific Synthesis of 23-Hydroxyundecylprodiginines and Analogues, and Conversion to Antimalarial Premarineosins via a Rieske Oxygenase Catalyzed Bicyclization

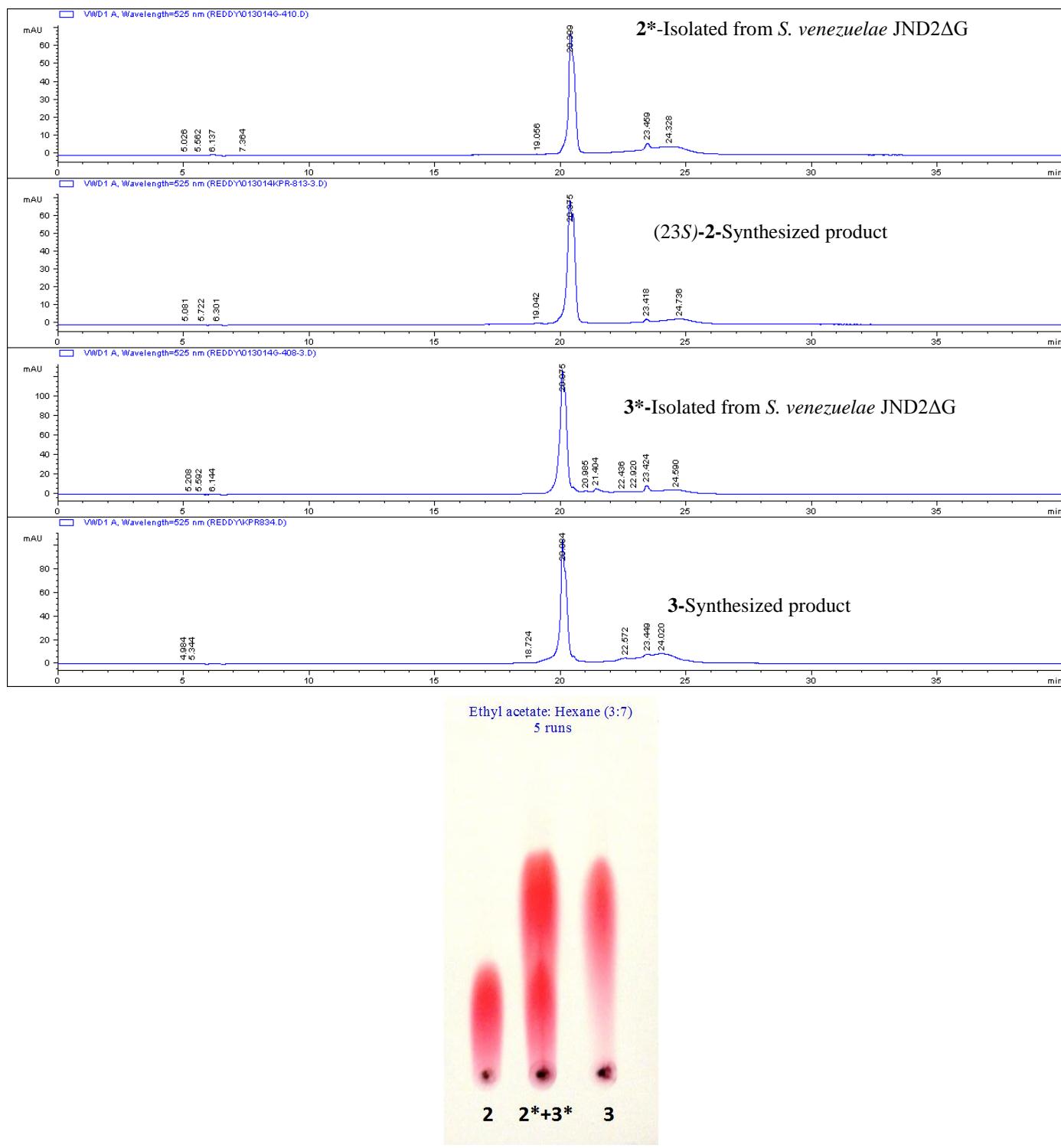
Papireddy Kancharla, Wanli Lu, Shaimaa M. Salem, Jane Xu Kelly,

and Kevin A. Reynolds\*

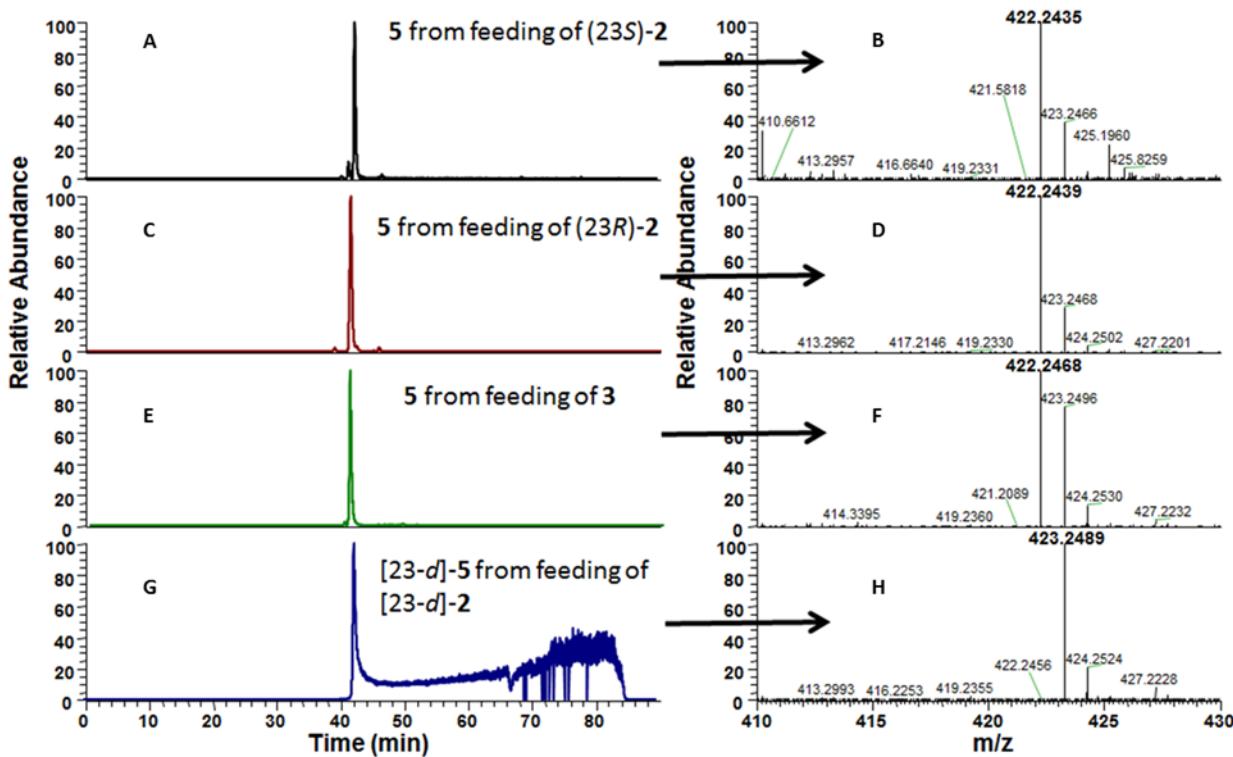
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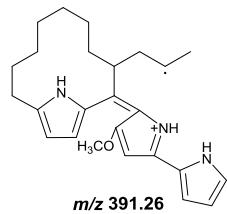
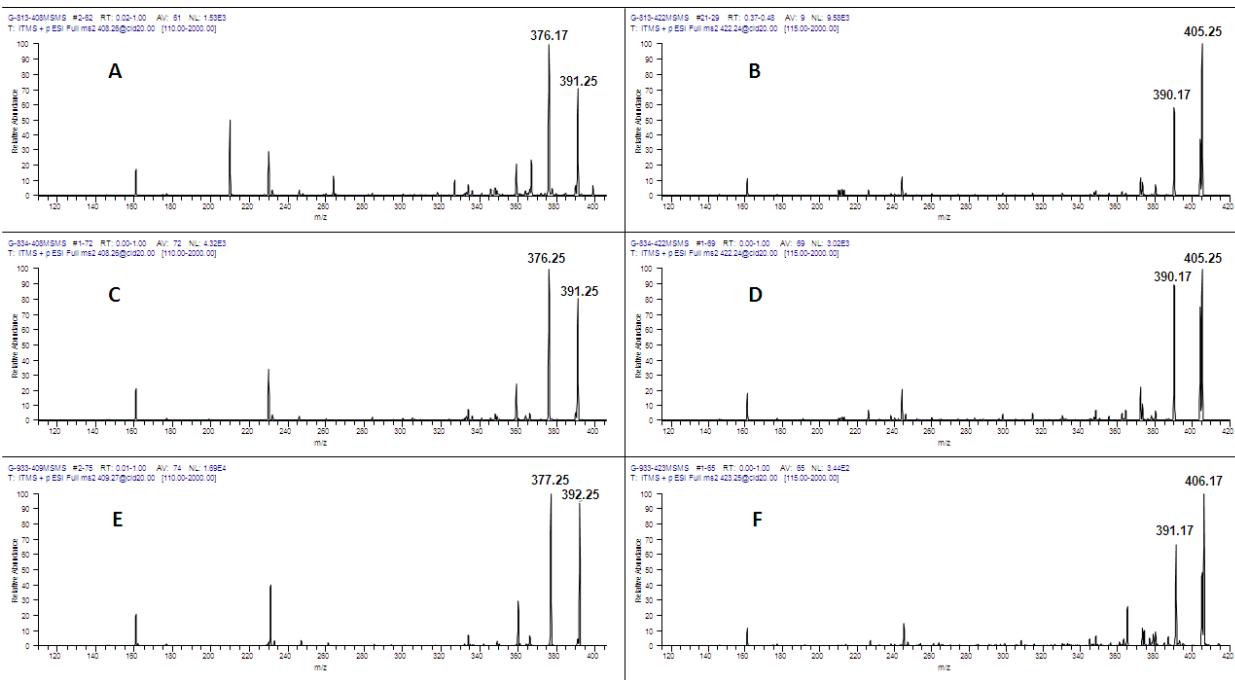


**Figure S1.** HPLC and TLC comparisons between synthetic [(23S)-**2**, and **3**], and natural prodiginines (**2\***, and **3\***)



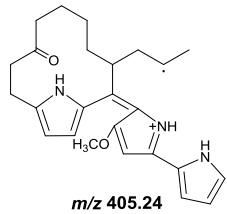
**Figure S2. LC-MS (EIC) profiles of 5 and [23-d]-5 from feeding of (23S)-2, (23R)-2, 3 and [23-d]-2 to *S. venezuelae* MarG.**

(A) EIC for  $m/z$  422.24-422.25, corresponding to  $[M + H]^+$  for **5**, from LC-MS analysis of extracts of *S. venezuelae* MarG fed with (23S)-2. (B) MS spectrometry of **5** from extracts of *S. venezuelae* MarG fed with (23S)-2. (C) EIC for  $m/z$  422.24-422.25, corresponding to  $[M + H]^+$  for **5**, from LC-MS analysis of extracts of *S. venezuelae* MarG fed with (23R)-2. (D) MS spectrometry of **5** from extracts of *S. venezuelae* MarG fed with (23R)-2. (E) EIC for  $m/z$  422.24-422.25, corresponding to  $[M + H]^+$  for **5**, from LC-MS analysis of extracts of *S. venezuelae* MarG fed with 3. (F) MS spectrometry of **5** from extracts of *S. venezuelae* MarG fed with 3. (G) EIC for  $m/z$  423.24-423.25, corresponding to  $[M + H]^+$  for [23-d]-**5**, from LC-MS analysis of extracts of *S. venezuelae* MarG fed with [23-d]-2. (H) MS spectrometry of [23-d]-**5** from extracts of *S. venezuelae* MarG fed with [23-d]-2.

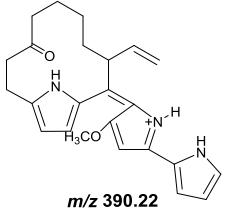


*m/z* 376.24

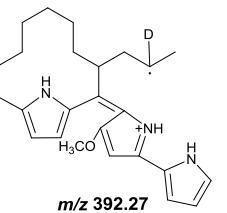
The chemical structure shows a tryptamine core with a methoxy group (*H*<sub>3</sub>*CO*) at position 5 and an allyl group at position 2. The indole nitrogen is protonated, indicated by a plus sign and a hydrogen atom (*N*<sup>+</sup>*H*). The structure is shown in its keto form.



*m/z* 405.24

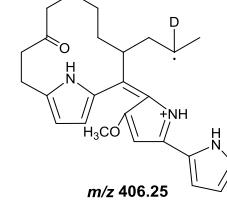


*m/z* 390.22

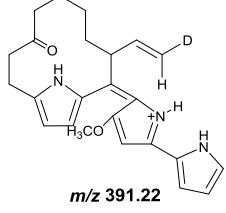


*m/z* 377.24

The chemical structure shows a tryptamine core with a 3-methoxyphenyl group attached to the indole nitrogen atom. The indole ring is substituted with a 3-methoxyphenyl group at the 5-position. The tryptamine side chain is shown as a propyl group.

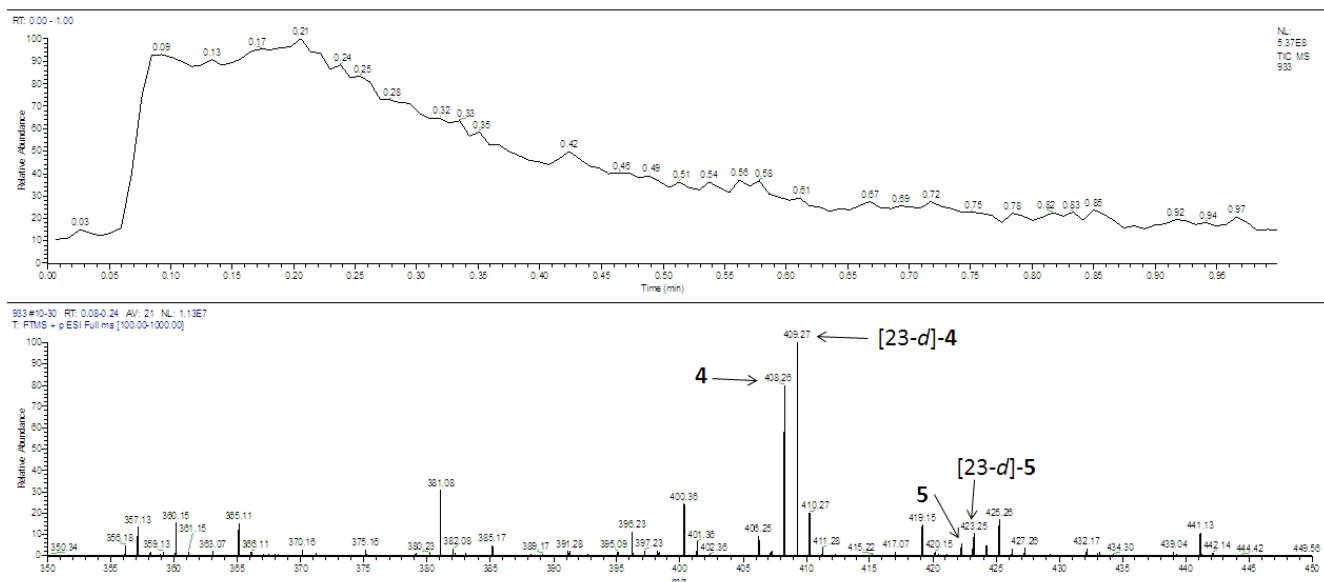


6.25

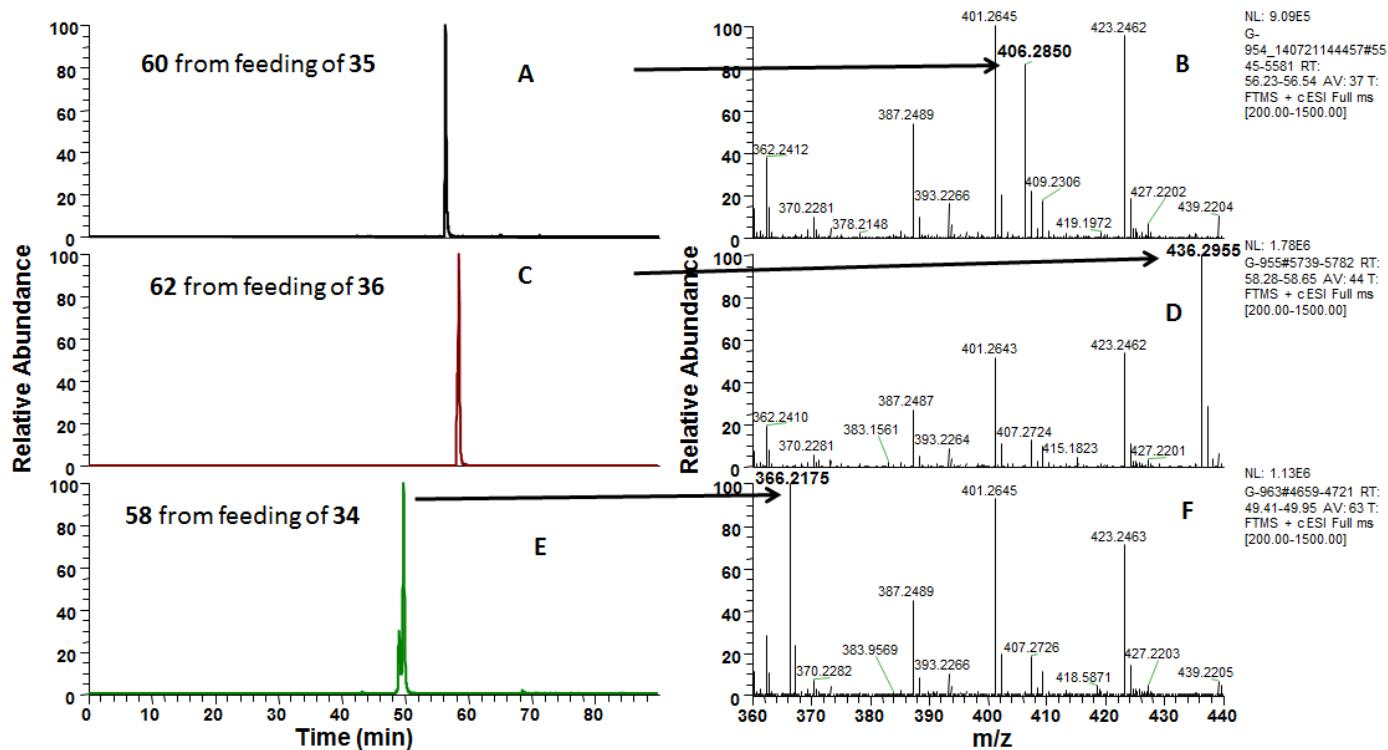


*m/z* 391.22

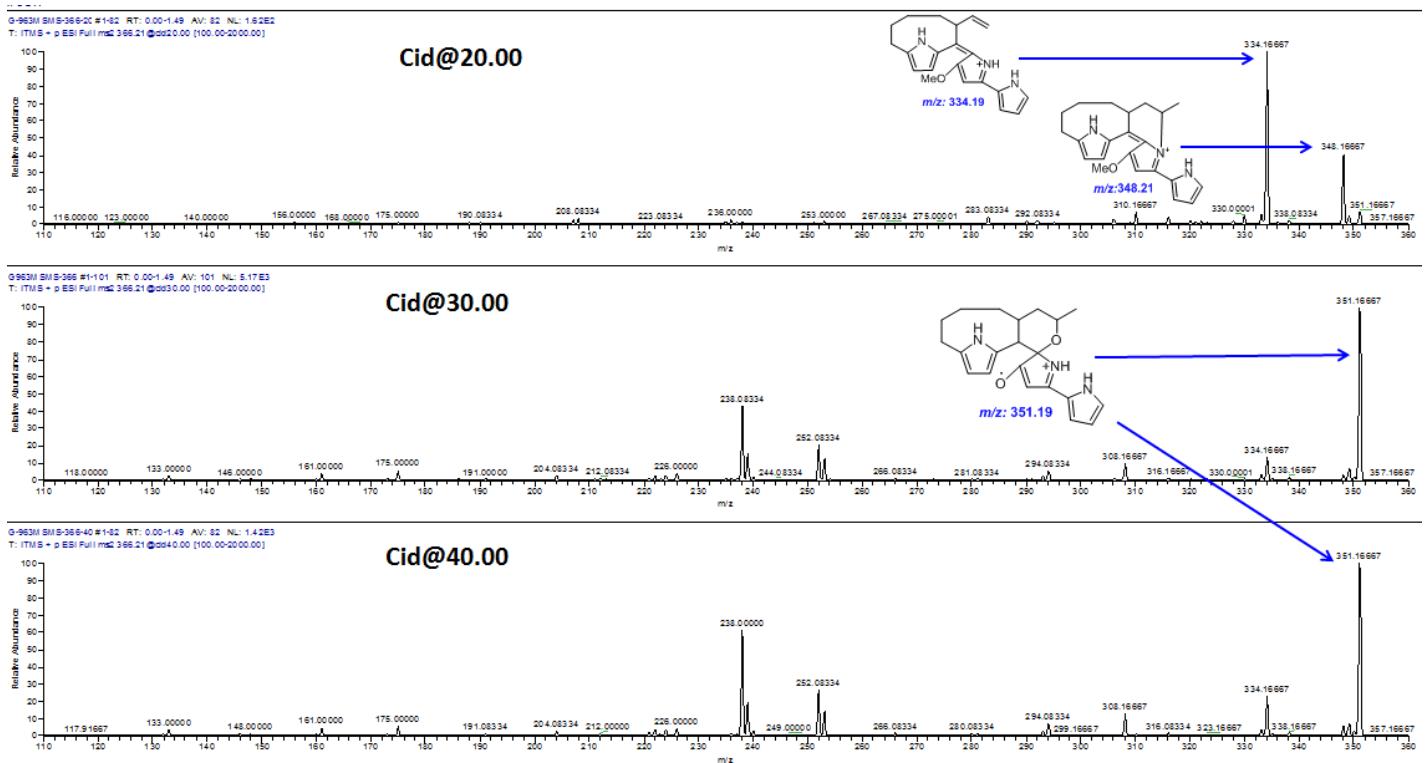
**Figure S3. ESI-CID-MS<sup>2</sup> profile of 4, [23-d]-4, 5, and [23-d]-5 from feeding experiments with *S. venezuelae* MarG:** A. The MS/MS profile of **4** from *S. venezuelae* marG fed with (23S)-2; B. The MS/MS profile of **5** from *S. venezuelae* marG fed with (23S)-2; C. The MS/MS profile of **4** from *S. venezuelae* marG fed with **3**; D. The MS/MS profile of **5** from *S. venezuelae* marG fed with **3**; E. The MS/MS profile of [23-d]-**4** from *S. venezuelae* marG fed with [23-d]-2; F. The MS/MS profile of [23-d]-**5** from *S. venezuelae* marG fed with [23-d]-2.



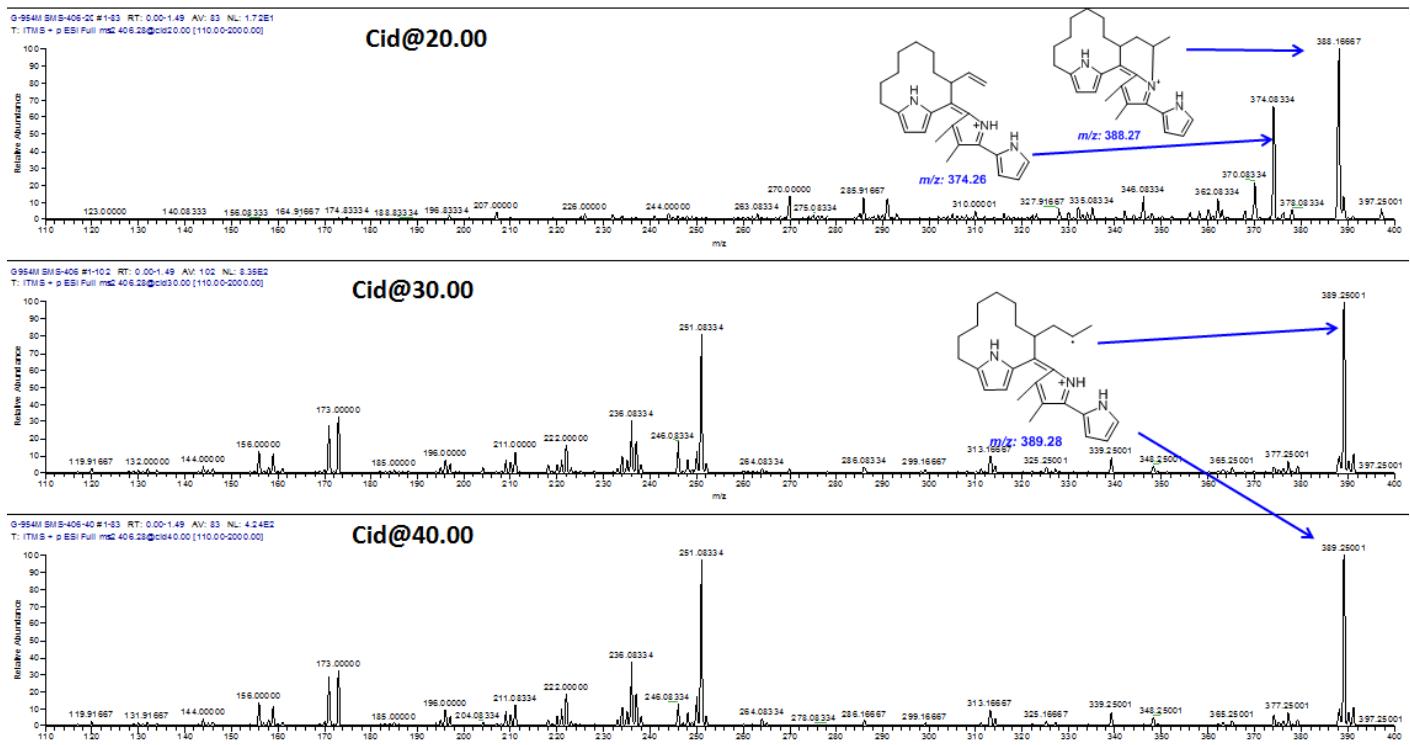
**Figure S4.** MS profile of [23-d]-4, and 4 (~1.2:1 ratio), and [23-d]-5, and 5 from feeding of [23-d]-2 to *S. venezuelae* MarG



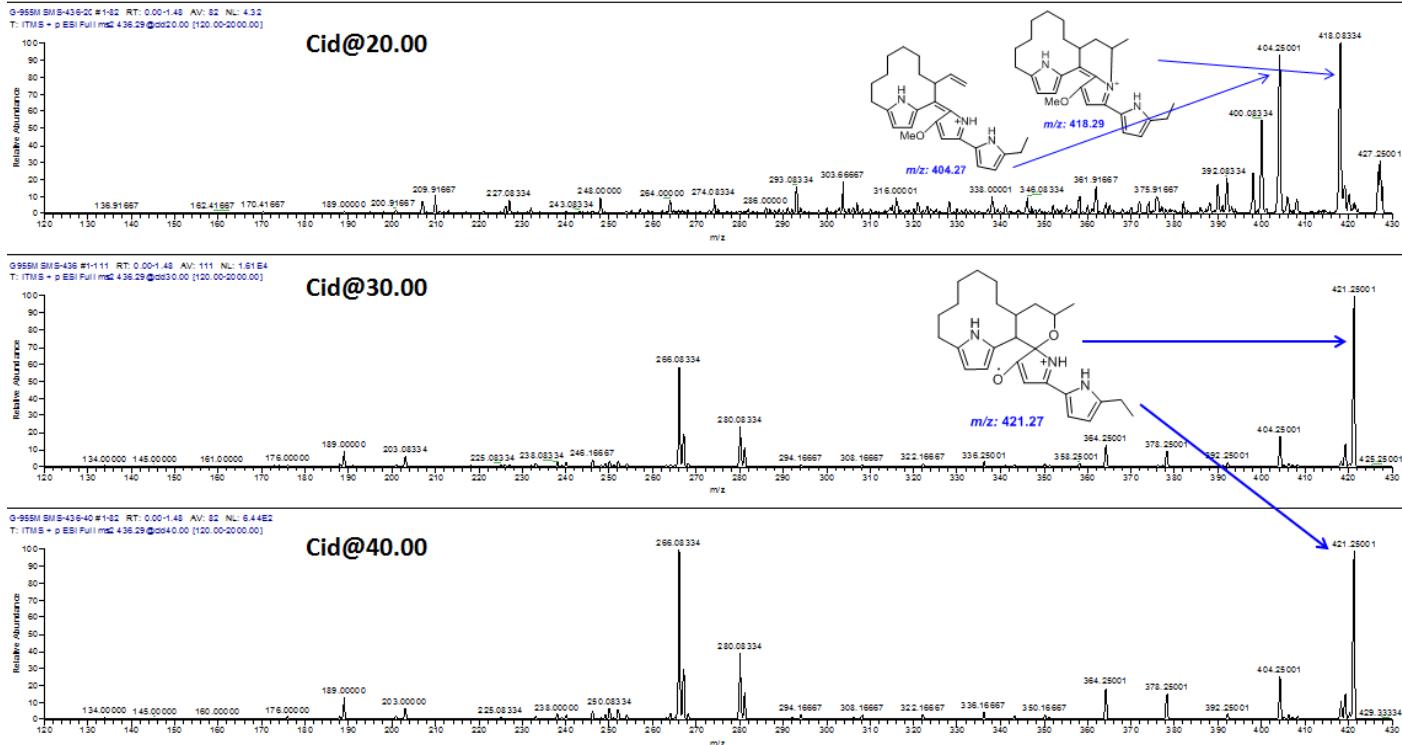
**Figure S5.** LC-MS (EIC) profiles of **58**, **60**, and **62** from feeding of **34–36** to *S. venezuelae* MarG. (A) EIC for  $m/z$  406.28-406.29, corresponding to  $[M + H]^+$  for **60**, from LC-MS analysis of extracts of *S. venezuelae* MarG fed with **35**. (B) MS spectrometry of **60** from extracts of *S. venezuelae* MarG fed with **35**. (C) EIC for  $m/z$  436.29-436.30, corresponding to  $[M + H]^+$  for **62**, from LC-MS analysis of extracts of *S. venezuelae* MarG fed with **36**. (D) MS spectrometry of **62** from extracts of *S. venezuelae* MarG fed with **36**. (E) EIC for  $m/z$  366.21-366.22, corresponding to  $[M + H]^+$  for **58**, from LC-MS analysis of extracts of *S. venezuelae* MarG fed with **34**. (F) MS spectrometry of **58** from extracts of *S. venezuelae* MarG fed with **34**.



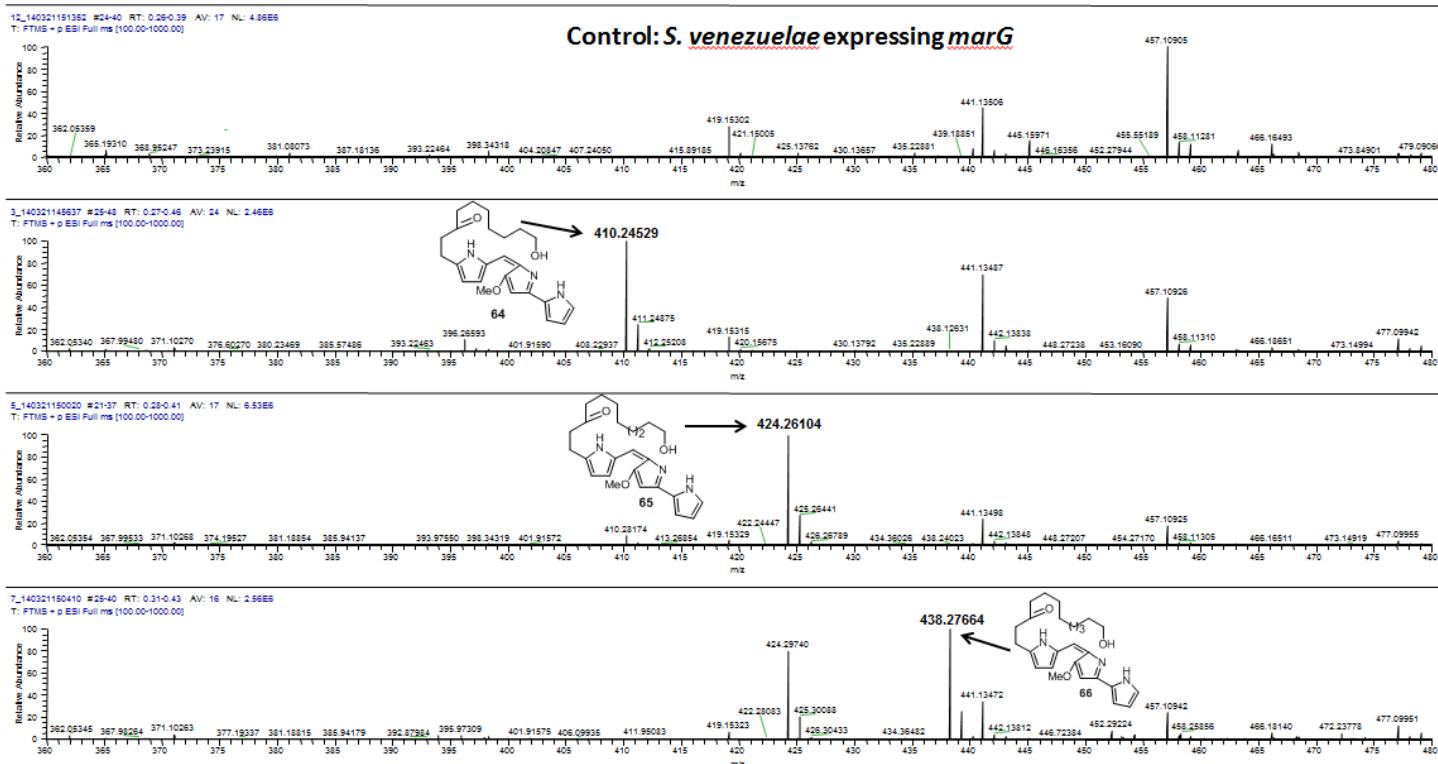
**Figure S6.** ESI-CID@20, 30, 40-MS<sup>2</sup> profiles of **58** from *S. venezuelae* marG fed with **34**.



**Figure S7.** ESI-CID@20, 30, 40-MS<sup>2</sup> profiles of **60** from *S. venezuelae* marG fed with **35**.



**Figure S8.** ESI-CID@20, 30, 40- $\text{MS}^2$  profiles of **62** from *S. venezuelae* marG fed with **36**.



**Figure S9.** MS profile of **64–66** from feeding of **39–41** to *S. venezuelae* MarG

**Table S1.** Strains, plasmids and cosmid used in this study

Strains	Relevant genotype/comments	Source/ Reference
<i>Escherichia coli</i> One Shot TOP10	<i>F mcrA Δ(mrr-hsdRMS-mcrBC) φ80lacZΔM15 ΔlacX74 recA1araD139 Δ(araleu) 7697 galU galK rpsL (StrR) endA1 nupG</i>	Invitrogen
<i>E. coli</i> ET12567(pUZ8002)	<i>dam dcm hsdS</i> , pUZ8002	1
<i>Streptomyces venezuelae</i> ATCC 15439	Wild-type pikromycin, methymycin, and neomethymycin producer	2
<i>S. venezuelae</i> MarG	<i>S. venezuelae</i> host the <i>marG</i> gene under the control of <i>ermE*</i> promoter	This study
<i>Plasmids and cosmid</i>		
pCR8-GW-TOPO	High copy number PCR cloning vector containing T7 and SP6 RNA polymerase promoters flanking a multiple cloning region within the alpha-peptide coding region of the enzyme beta-galactosidase; <i>bla</i> .	Invitrogen
pET15b	T7 promoter, RBS, N terminal 6x His Tag, <i>bla</i> , pBR322 origin	Novagene
pSE34	pWHM3 with <i>ermE*</i> promoter, <i>bla</i> , <i>tsr</i>	Pfizer Inc.
pIJ778	pBluescript KS (+) containing the spectinomycin/streptomycin resistance gene <i>aadA</i> and the <i>oriT</i> of plasmid RP4, flanked by FRT sites	3
pMarG-topo	1.4 kbp DNA fragment containing <i>marG</i> gene in pCR8-GW-TOPO	This study
pMarG-15b	pET15b with 1.4 kbp NdeI-BamHI <i>marG</i> ORF	This study
pMarG-34	pSE34 with 1.4 kbp XbaI-HindIII <i>marG</i> ORF along with RBS	This study
pMarG-34S	<i>bla</i> in pMarG-34 replaced with <i>aadA-oriT cassette</i>	This study
8A7	A SuperCos1 cosmid carrying the mar gene cluster together with 6 kbp of primary metabolic genes	4

**Table S2.** Primers used in this study

Primer	Sequence
<i>marG</i> -15bF	5'- <u>CATATG</u> CTCGCGGGAAACTCGAAAG-3'
<i>marG</i> -15bR	5'- <u>GGATCCT</u> CAGGACC CGCGCCGC-3'
<i>Amp-SpF</i>	5'- TGATAAATGCTTCAATAATATTGAAAAAGGAAGAGTATGATTCCGGGGATCCGTGACC-3'
<i>Amp-SpR</i>	5'- AATCTAAAGTATATGAGTAAACTGGTCTGACAGTTATGTAGGCTGGAGCTGCTTC-3'

**Table S3.** HPLC gradient used for synthetic [(23*S*)-**2** and **3**], and natural prodiginines (**2\*** and **3\***)

Time (min)	Flow rate (ml/min)	% A <sup>a</sup>	% B <sup>b</sup>
0	0.3	100	0
10	0.3	25	75
15	0.3	0	100
20	0.3	15	85
25	0.3	25	75
40	0.3	100	0

<sup>a</sup>Buffer A: water/acetonitrile/methanol/TFA (50:10:40:0.1)

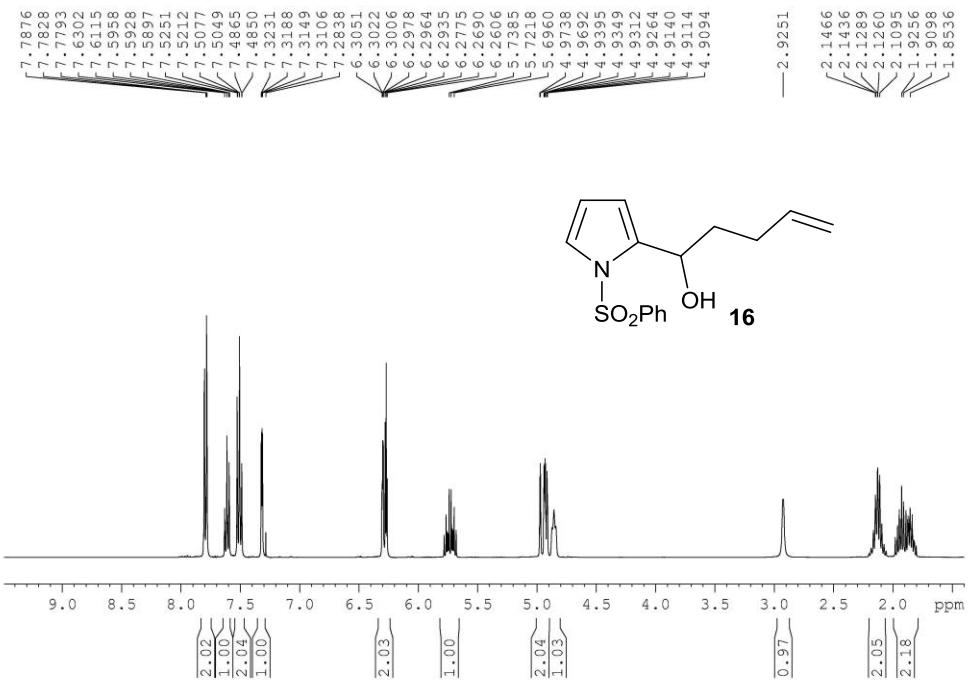
<sup>b</sup>Buffer B: 100% methanol/0.1% TFA

**Table S4.** LC-MS gradient used for analysis of *S. venezuelae* MarG fed with synthesized hydroxyundecylprodiginines

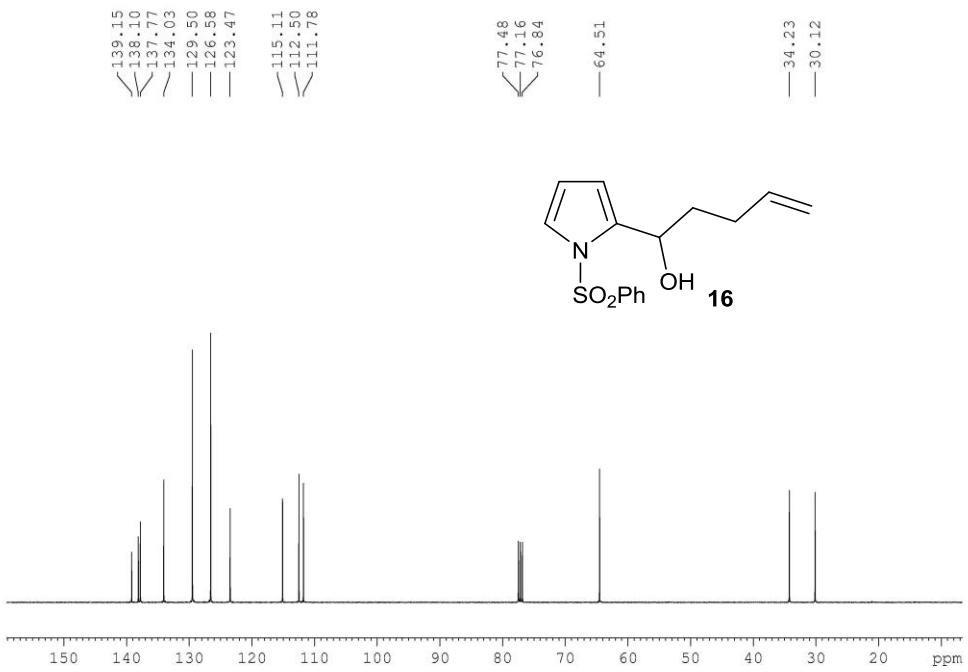
Time (min)	Flow rate (mL/min)	% Water <sup>a</sup>	% Methanol <sup>a</sup>
0	0.25	90	10
5	0.25	90	10
70	0.25	0	100
80	0.25	0	100
81	0.25	90	10
90	0.25	90	10

<sup>a</sup> Contained 0.05% HCOOH

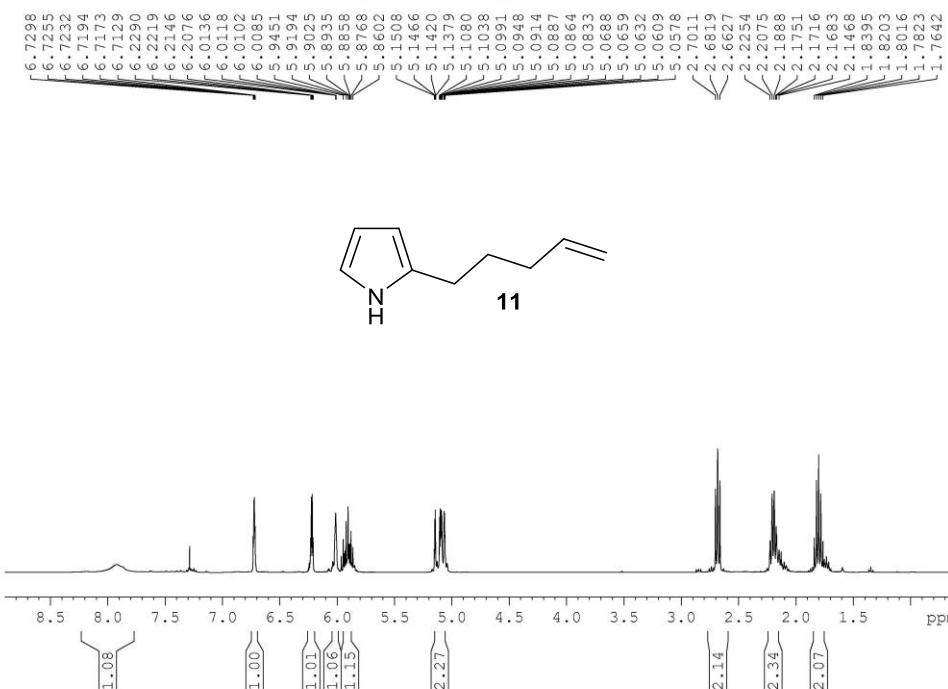
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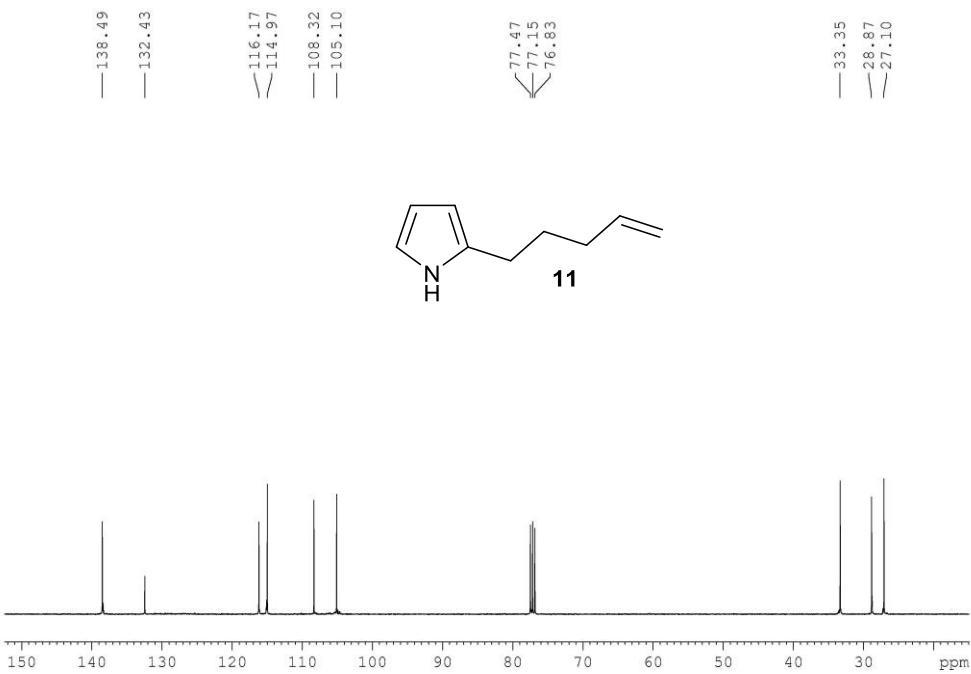
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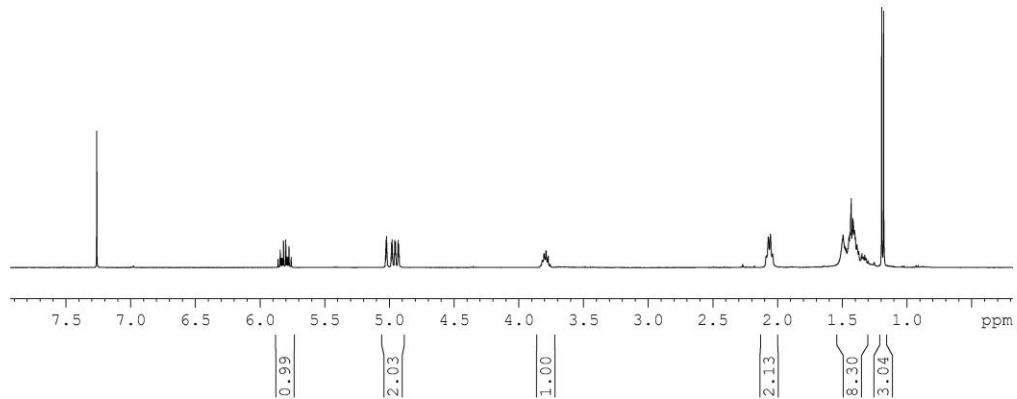
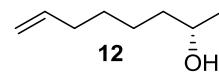
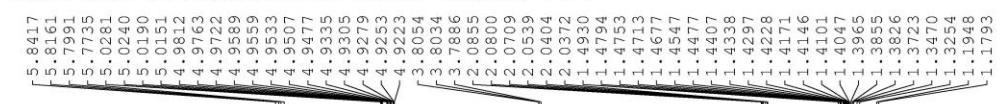
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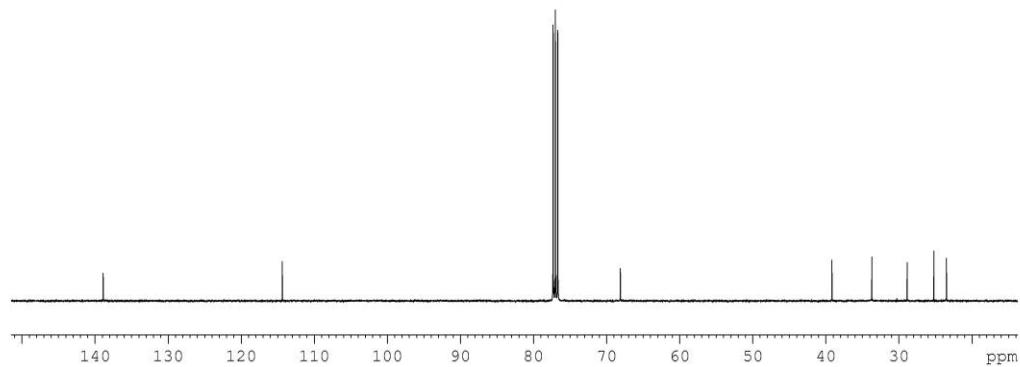
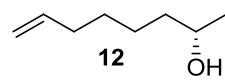
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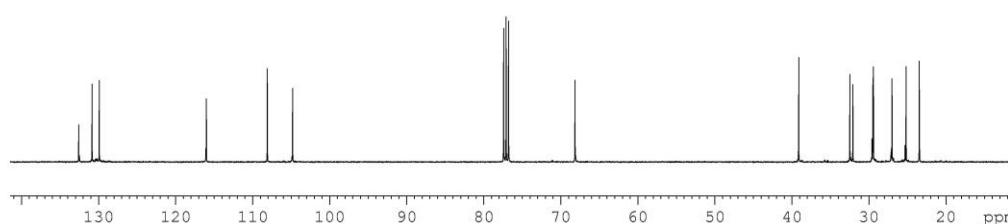
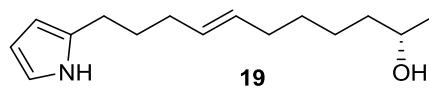
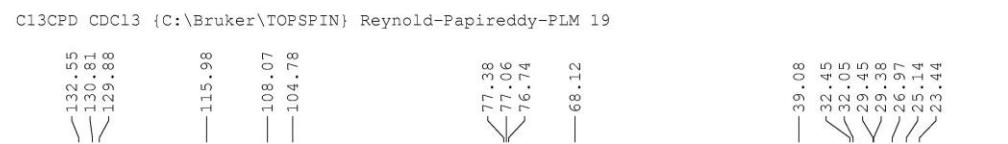
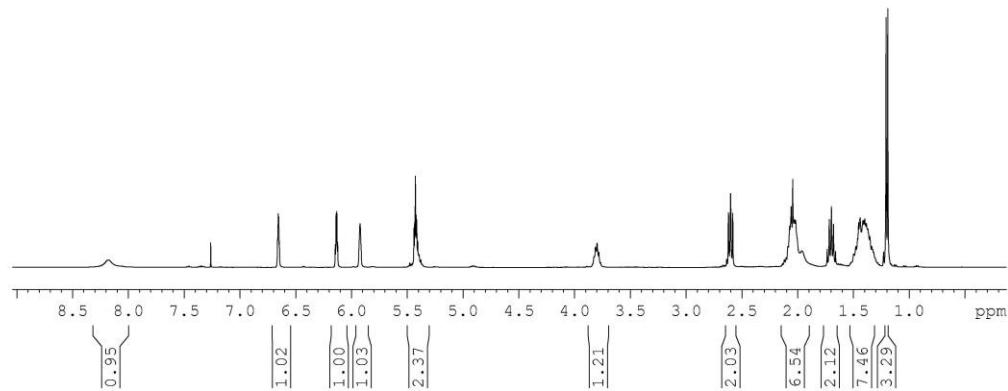
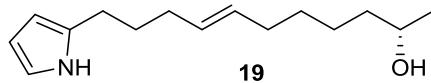
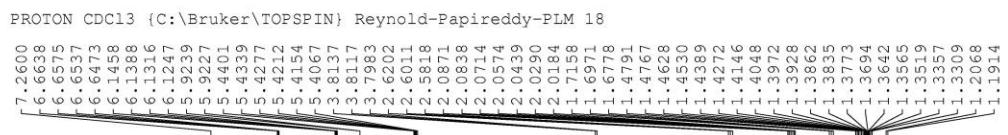


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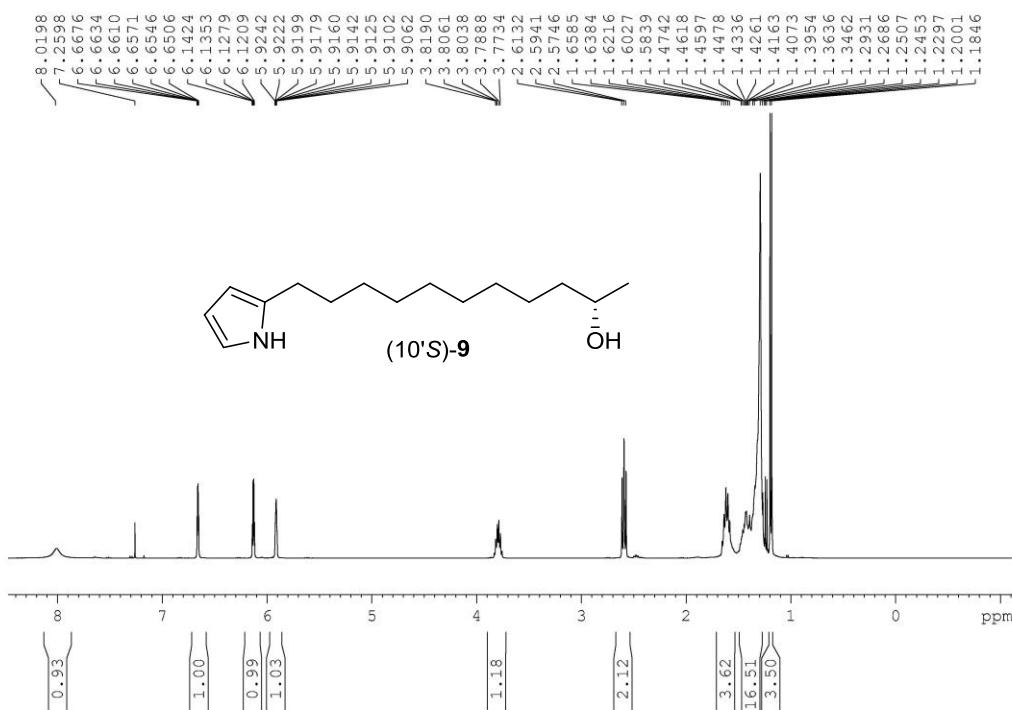


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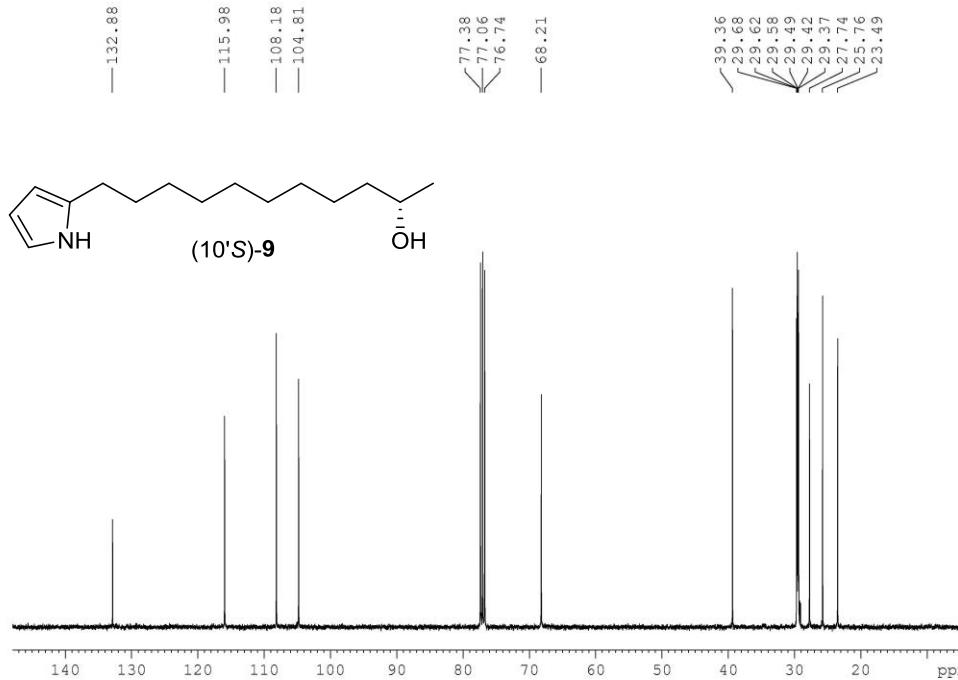




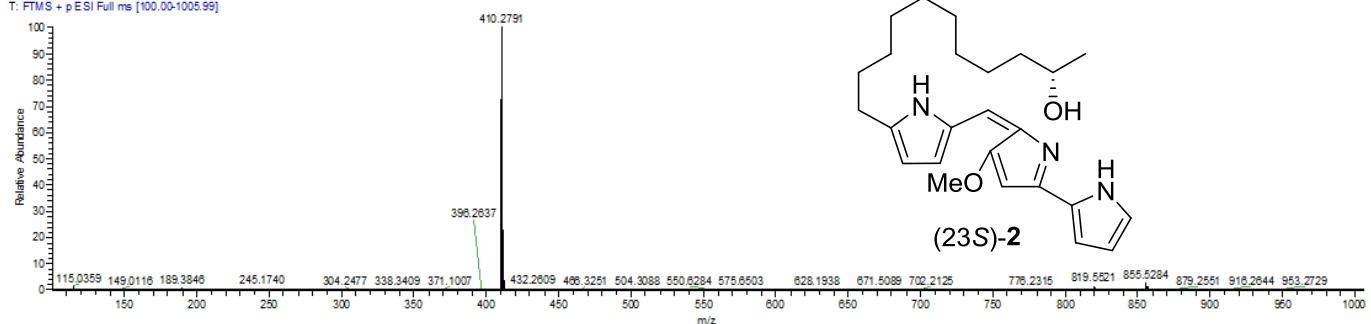
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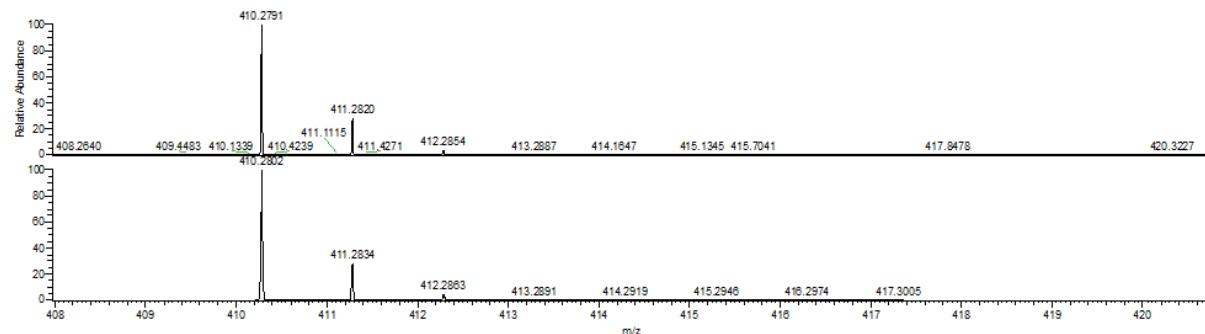


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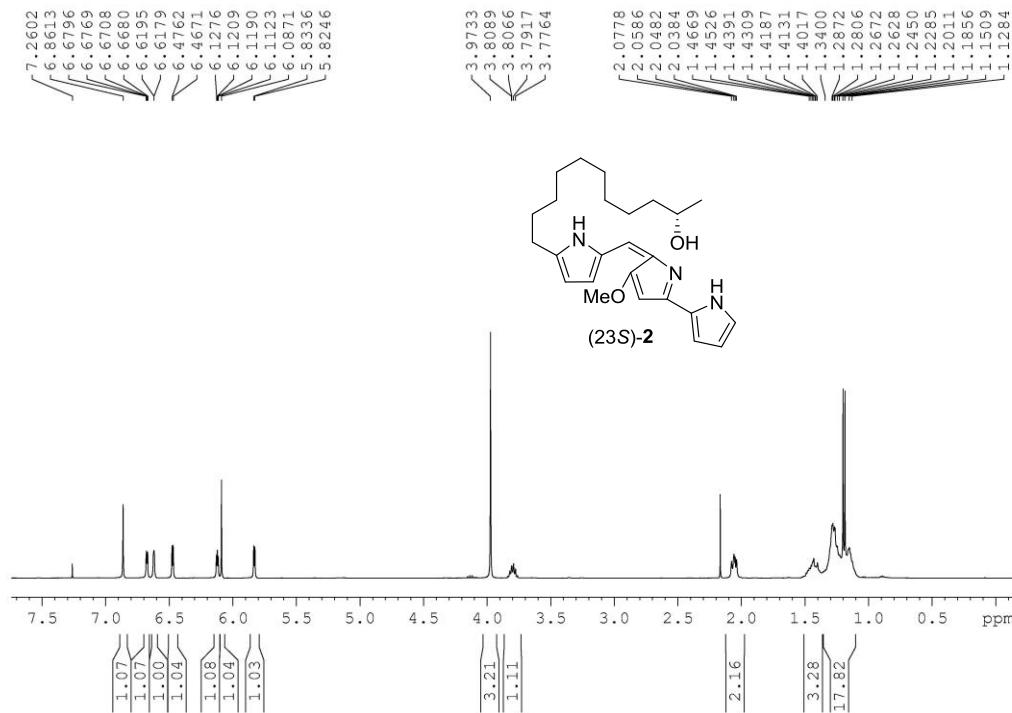


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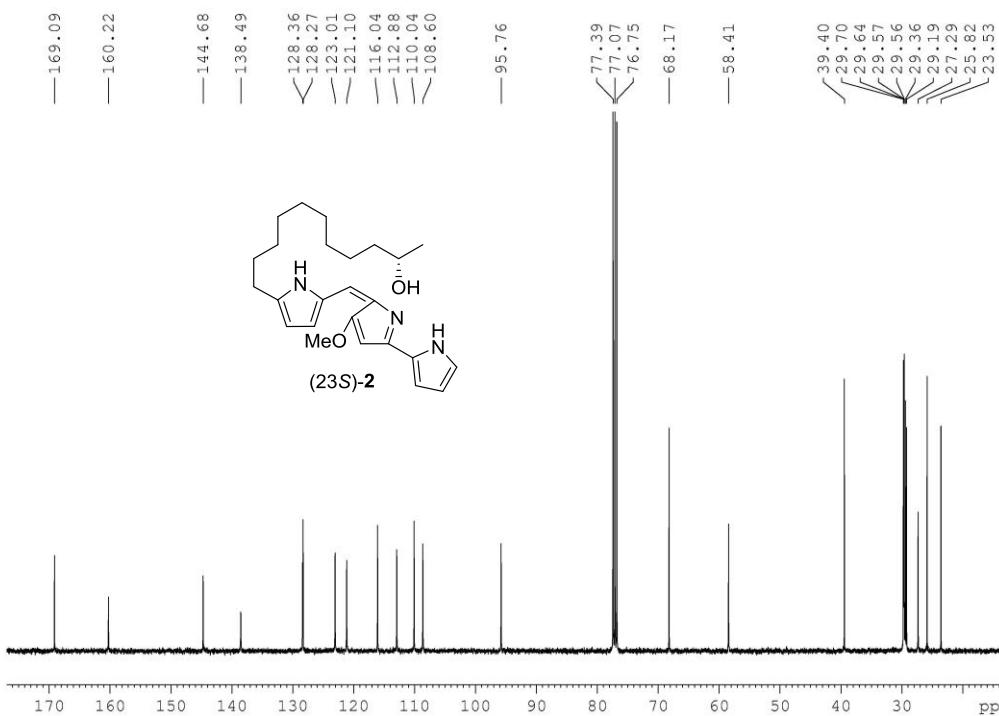
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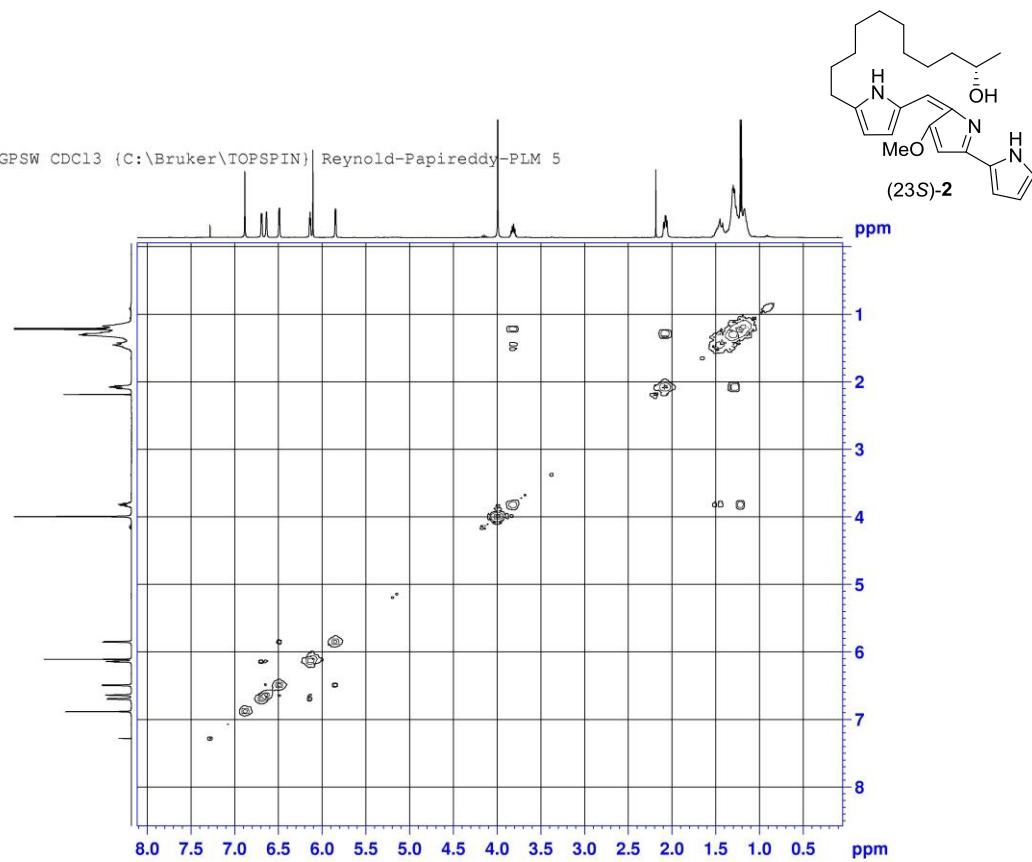
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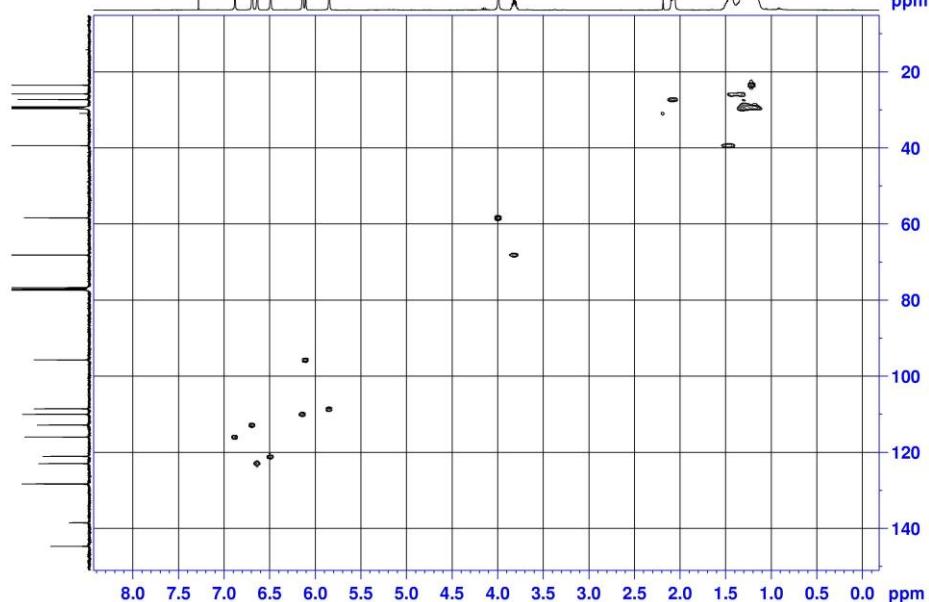
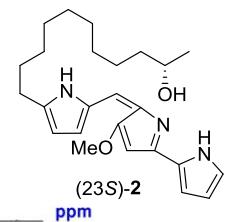
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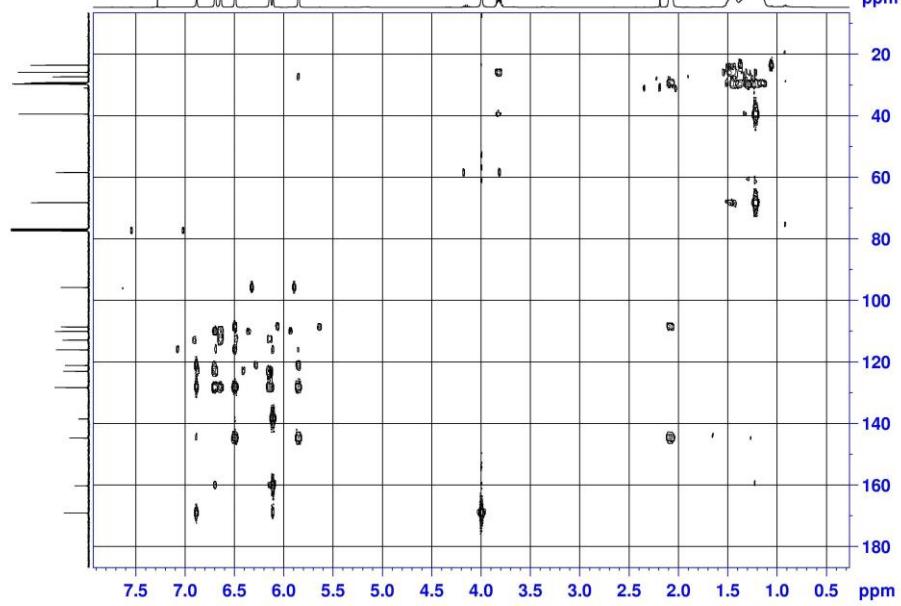
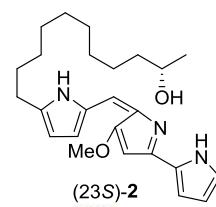
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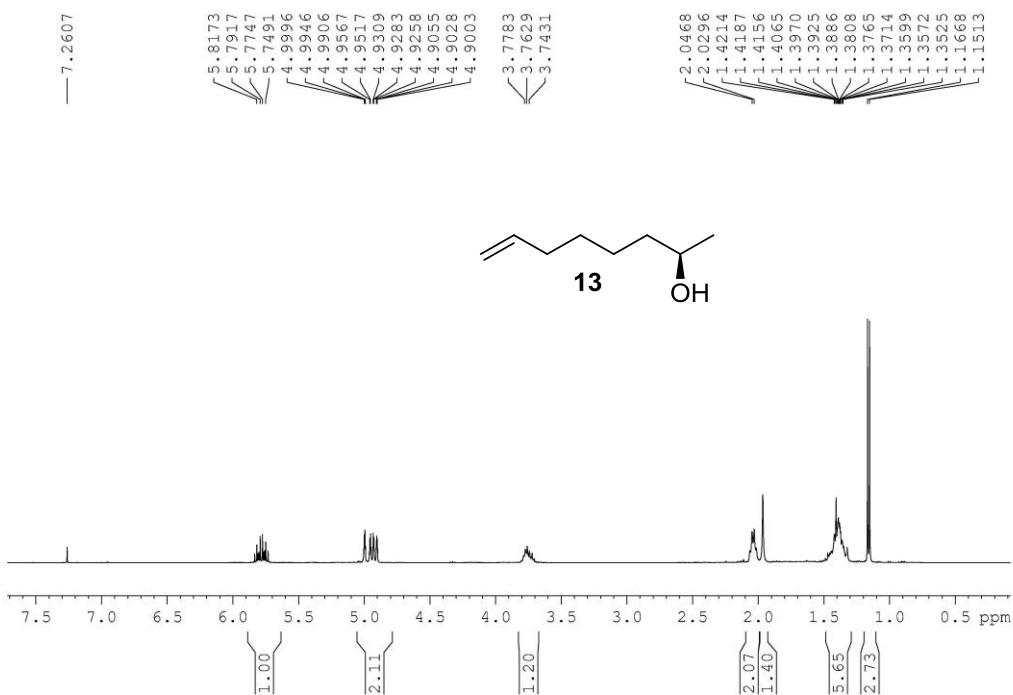
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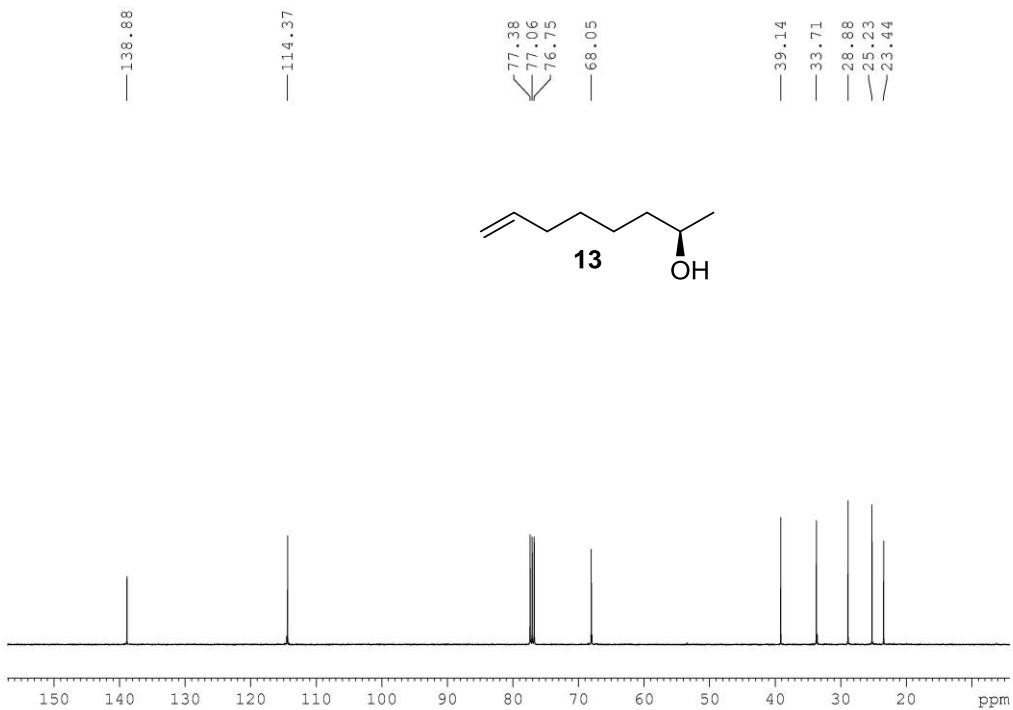
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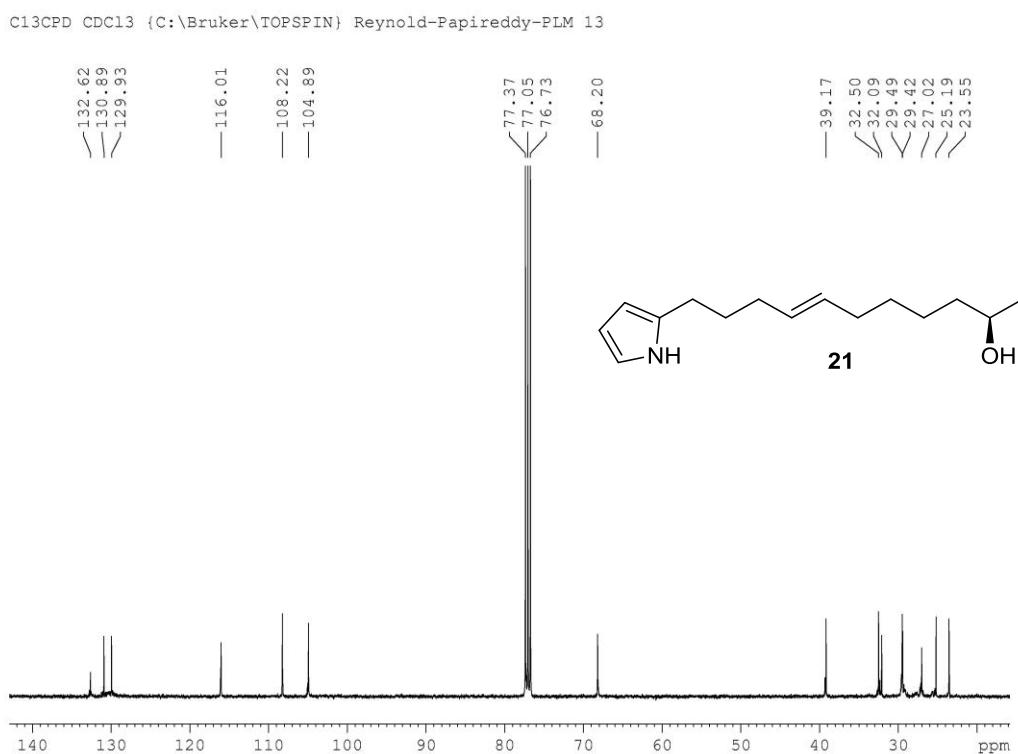
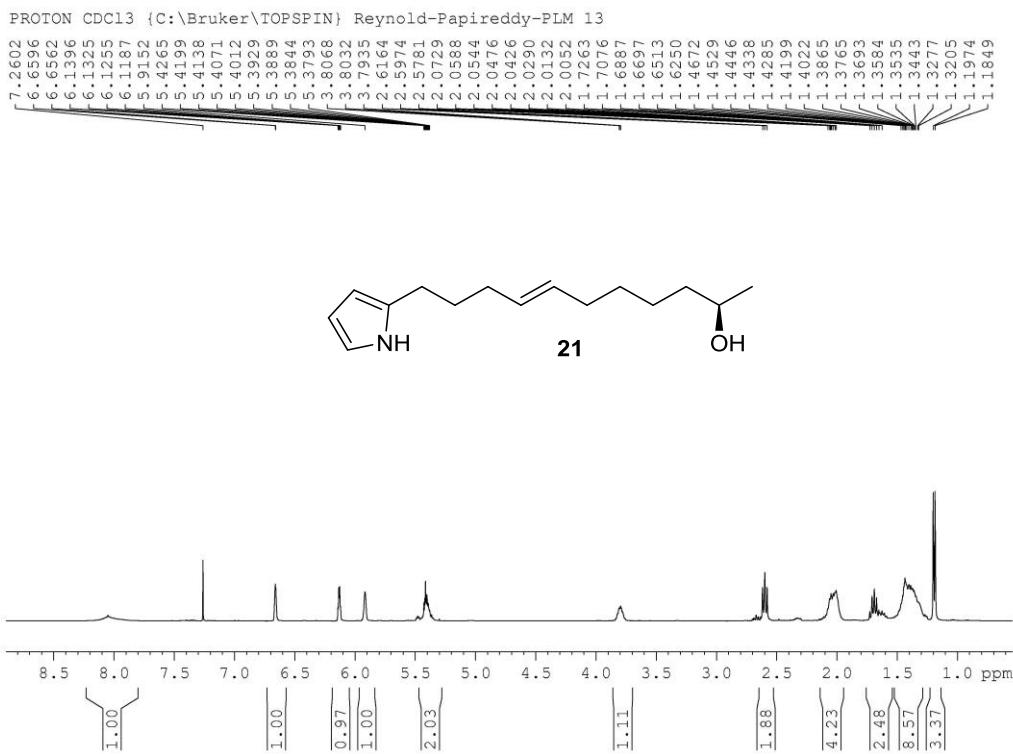


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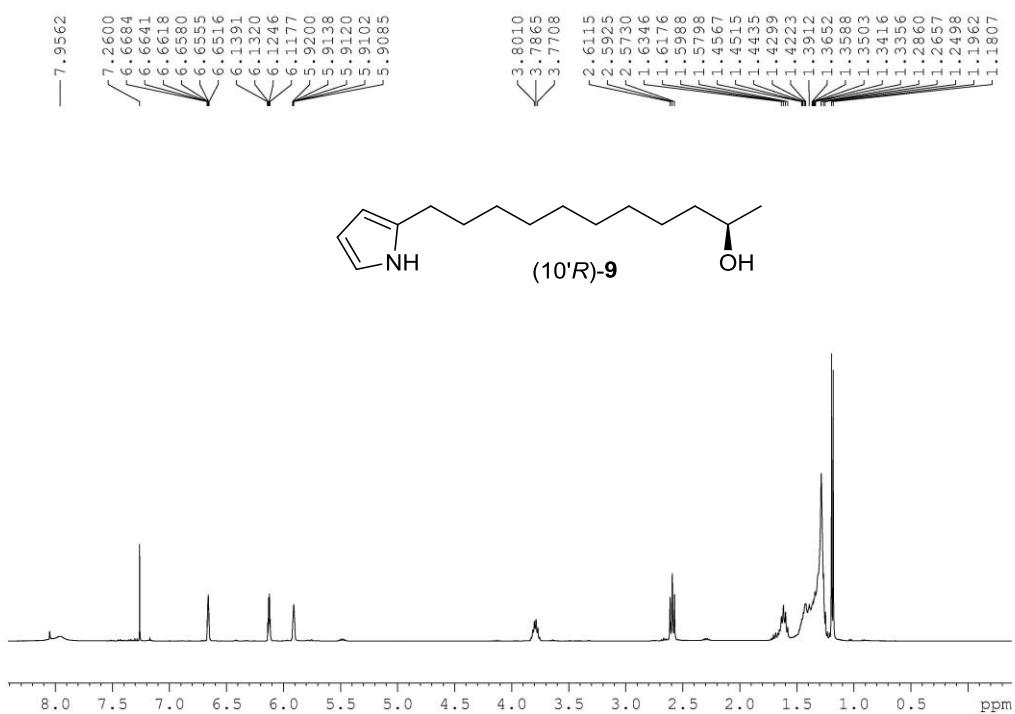


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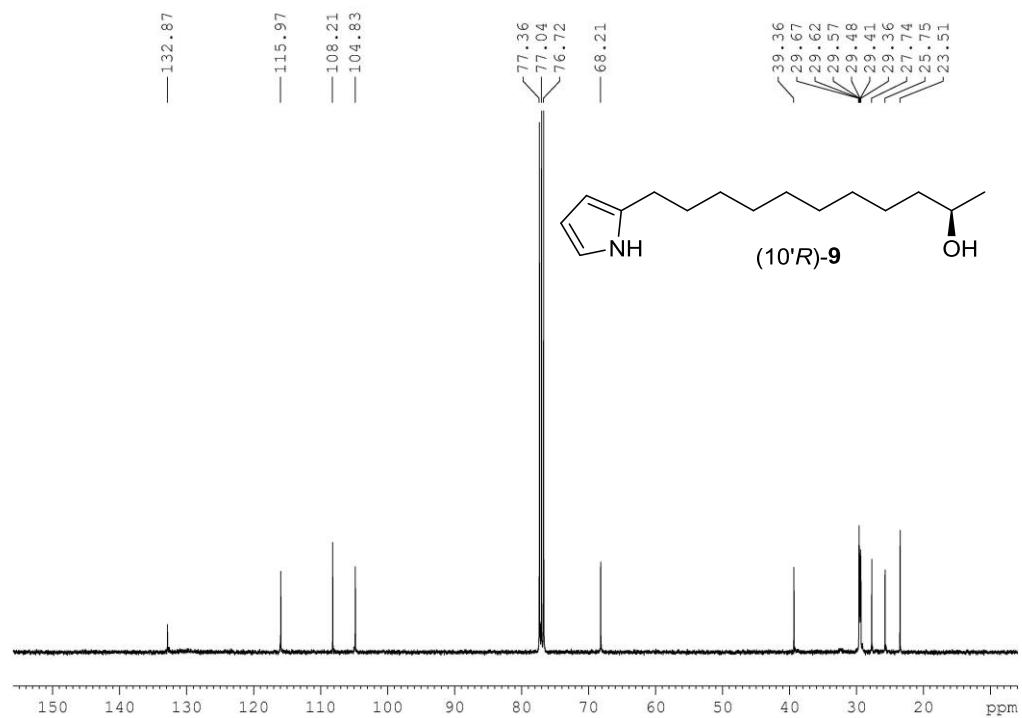




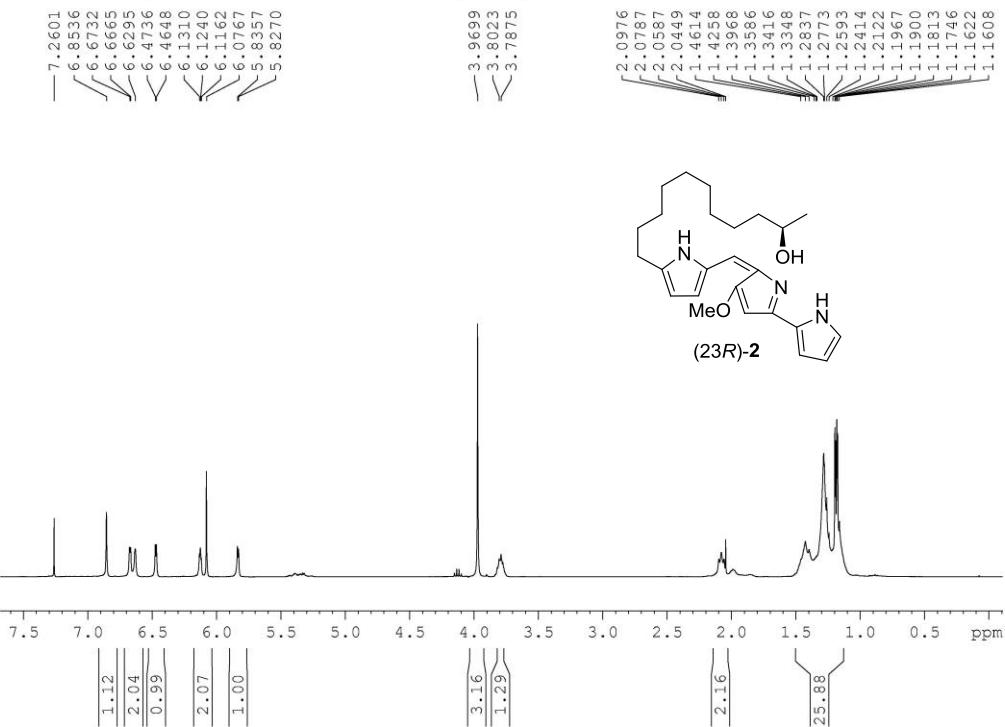
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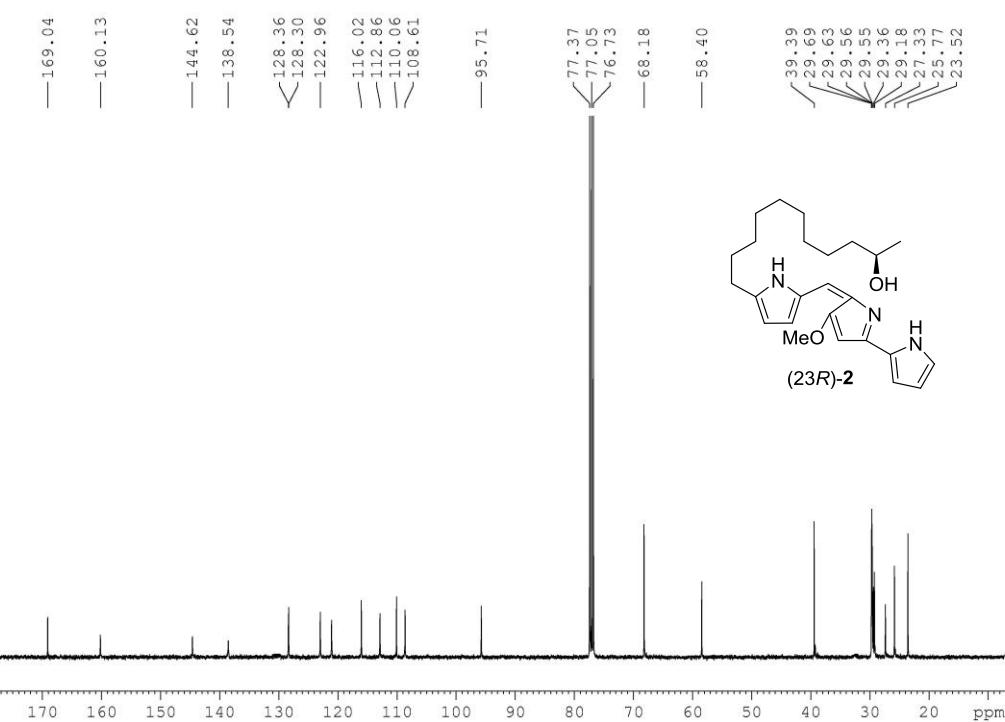
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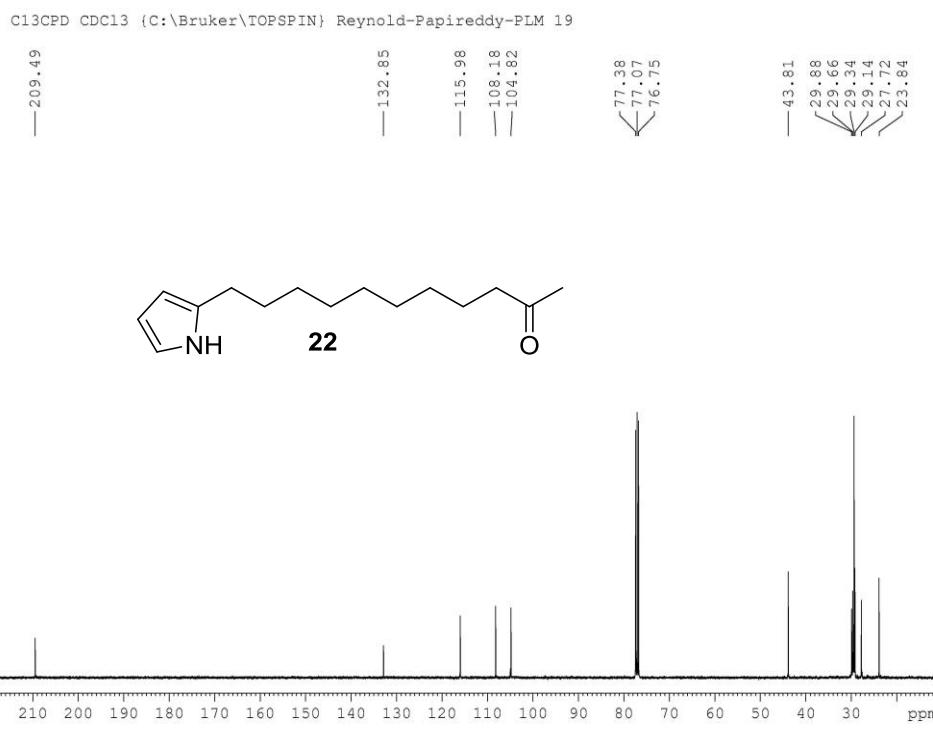
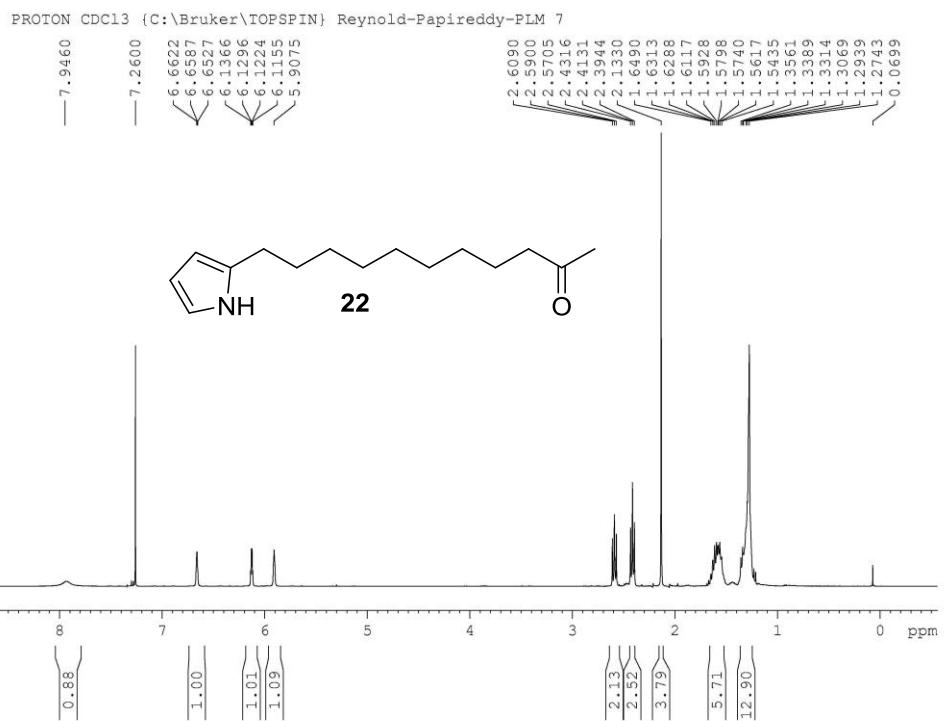


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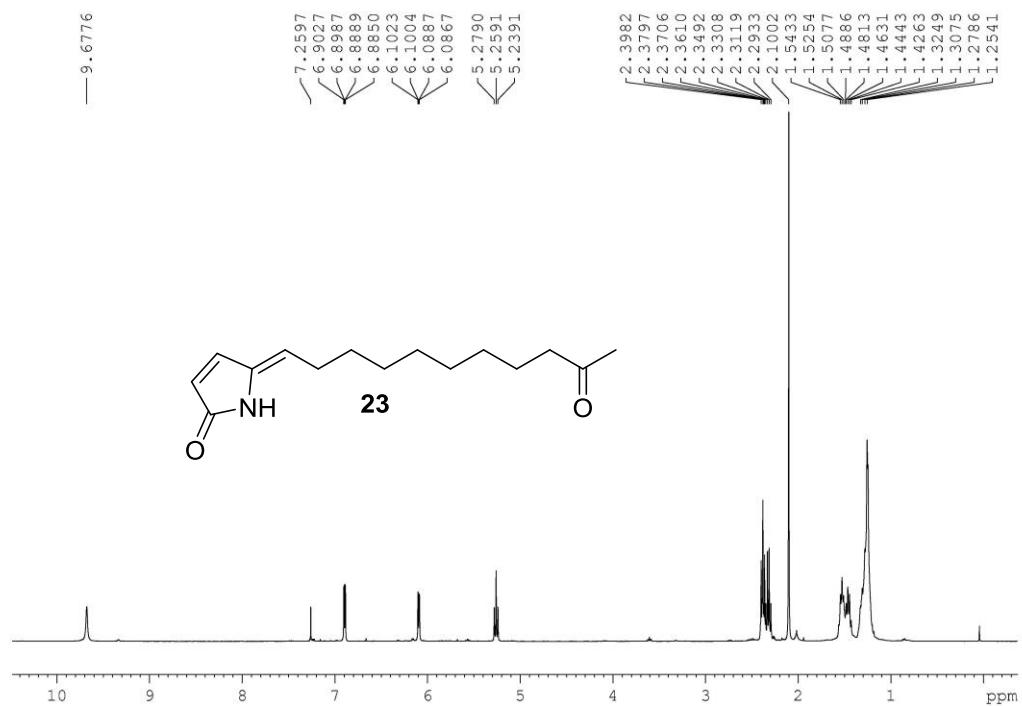


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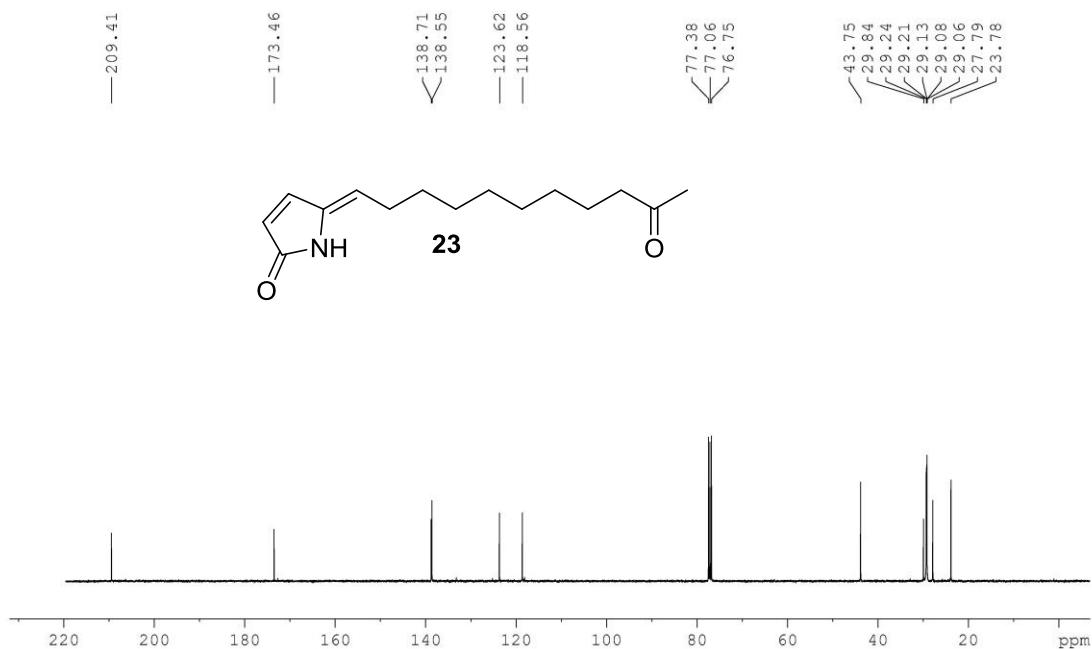


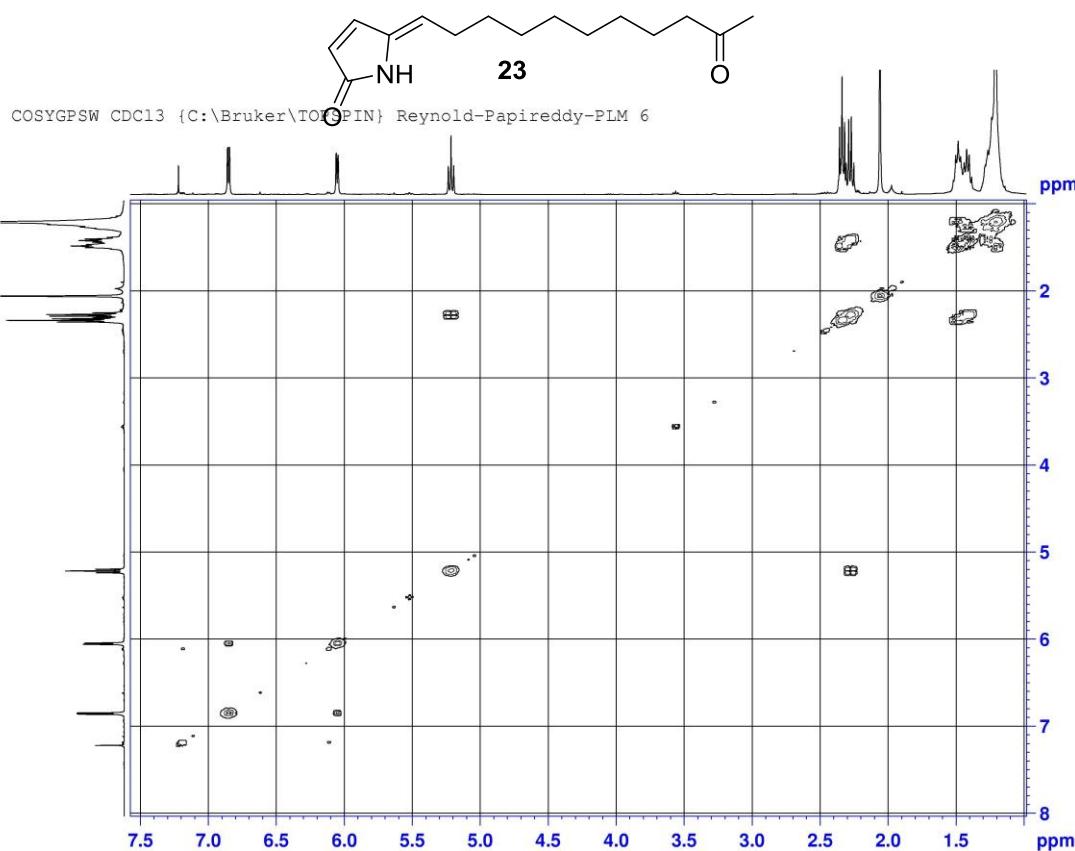
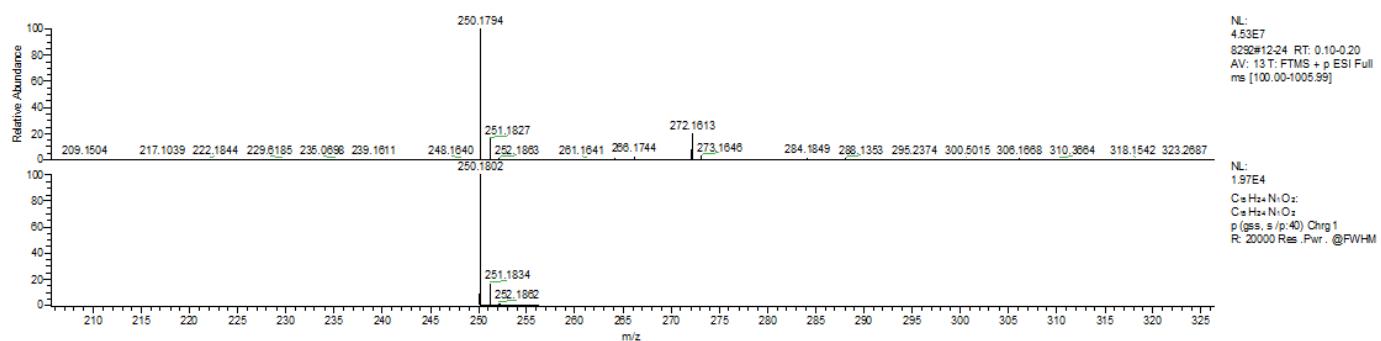
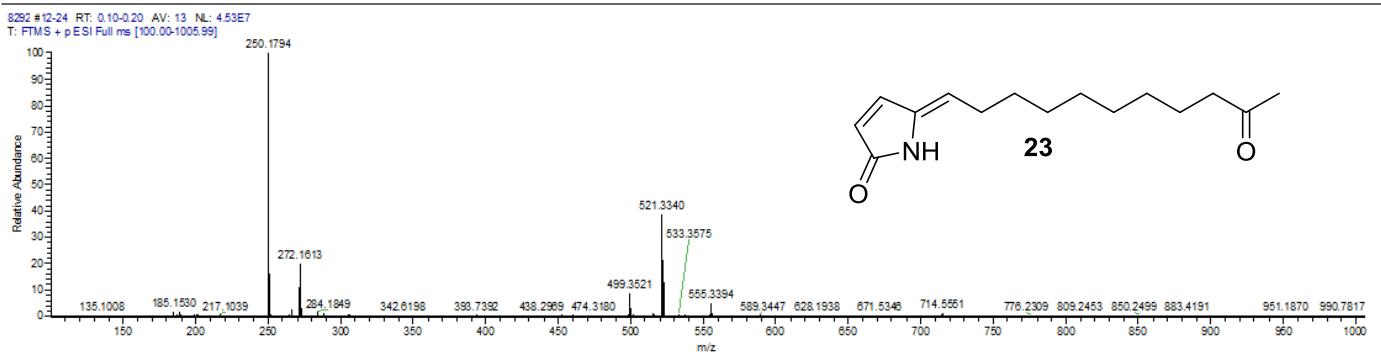


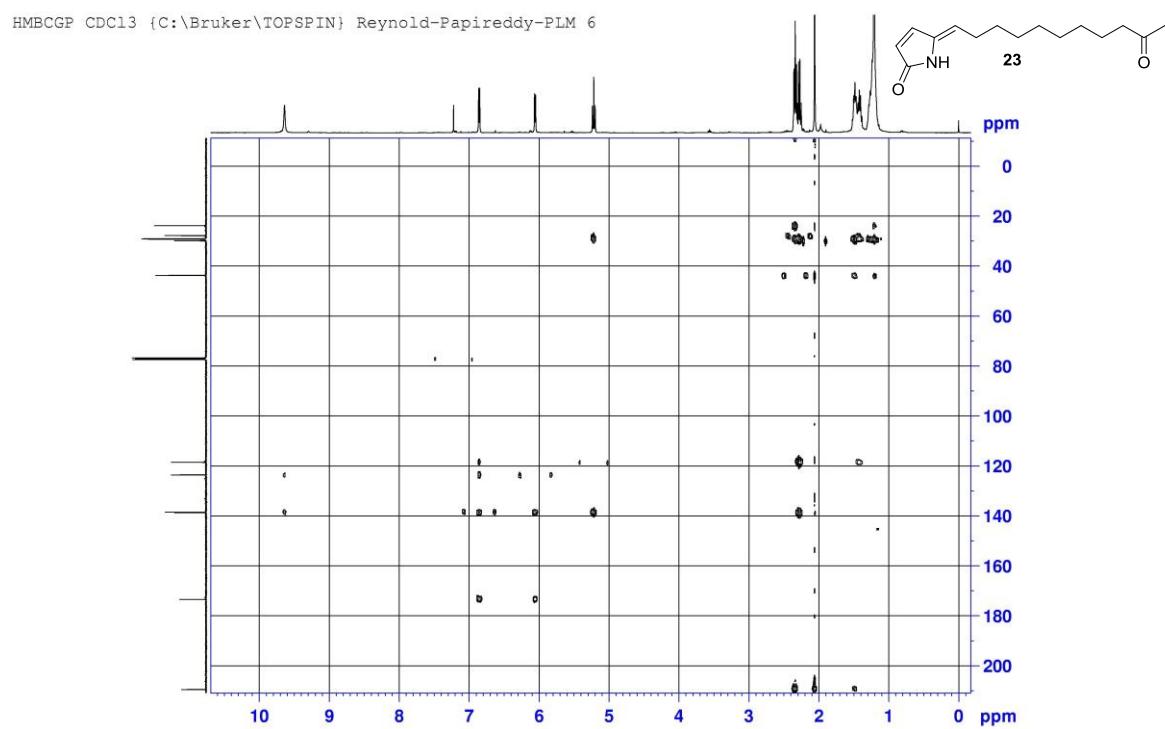
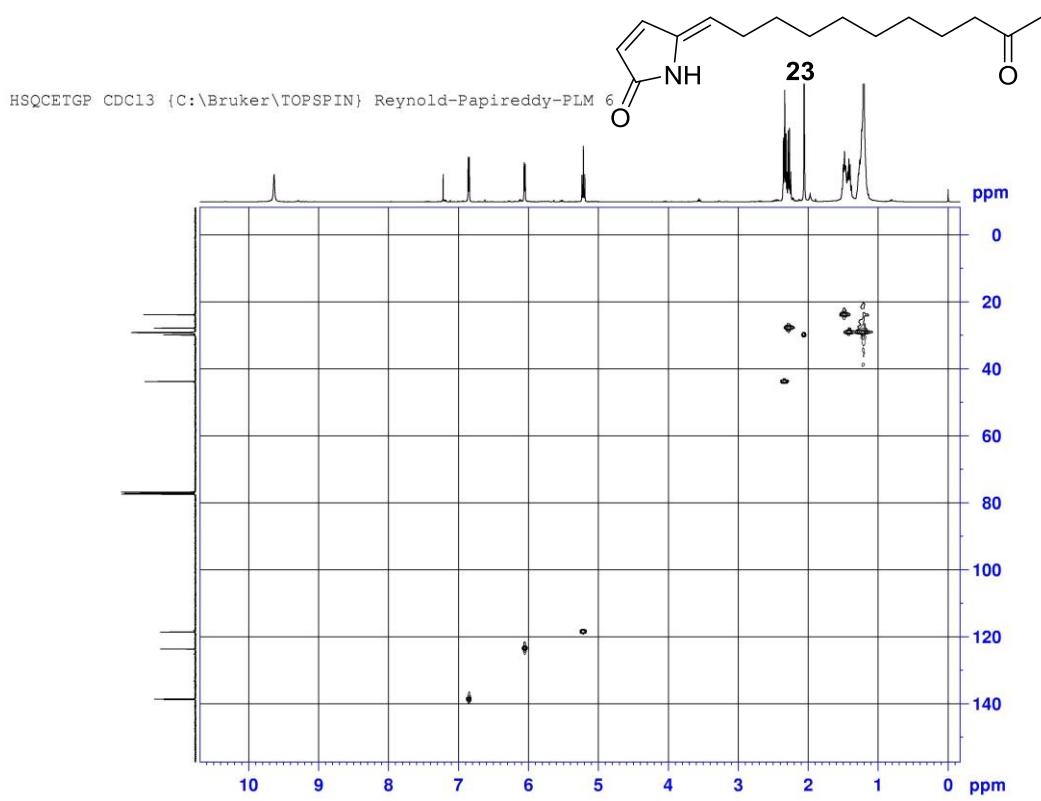
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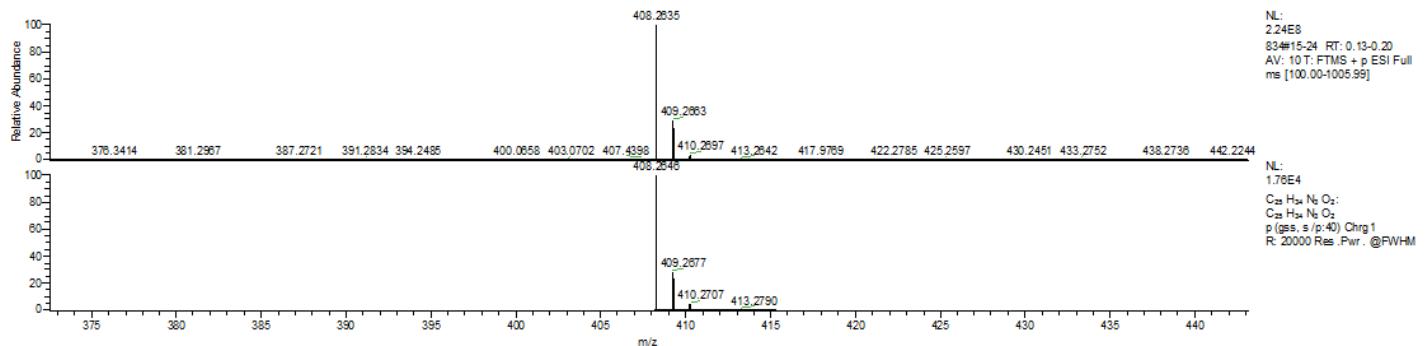
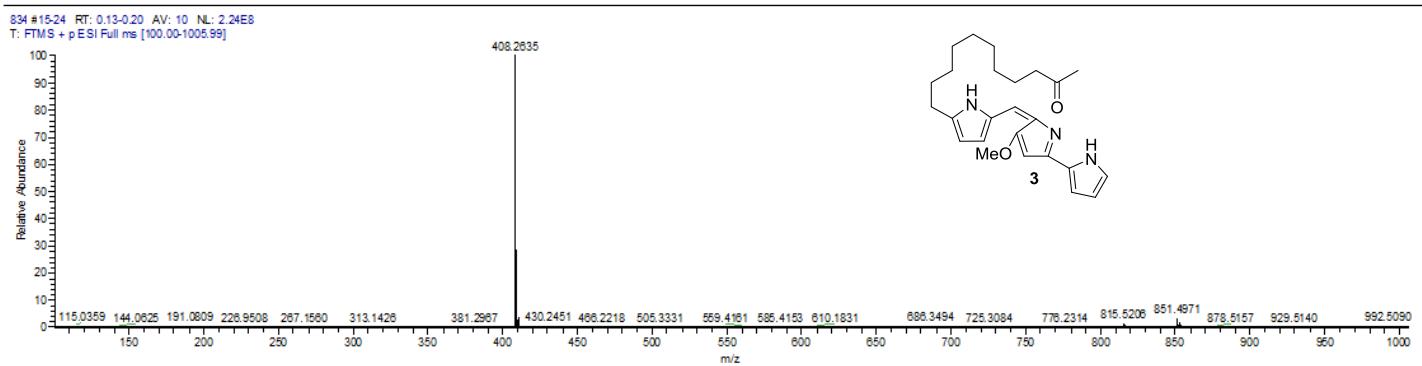
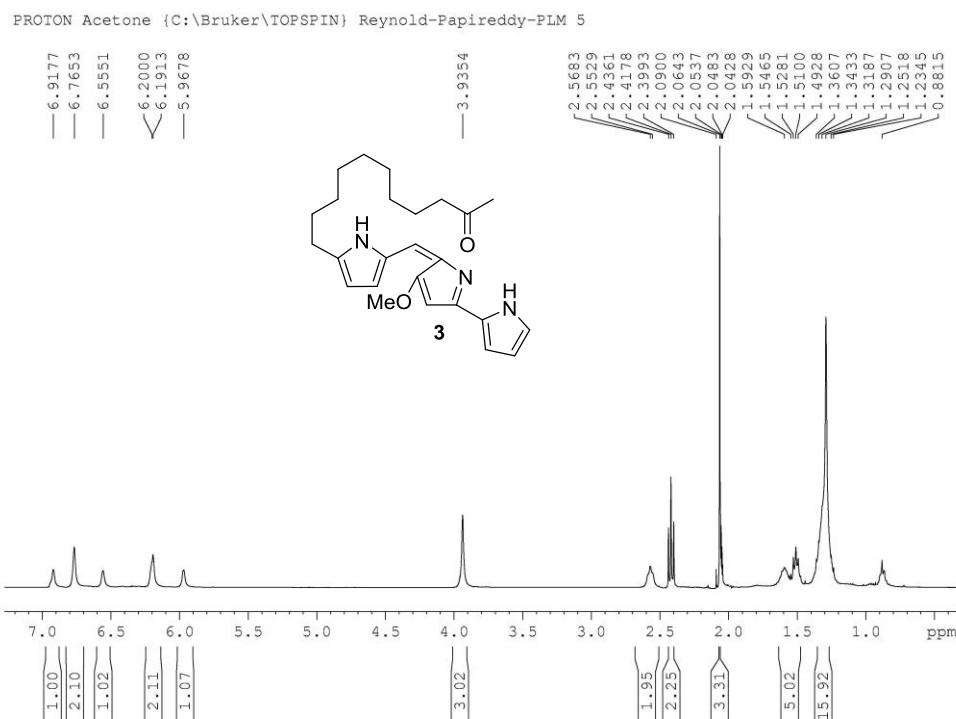


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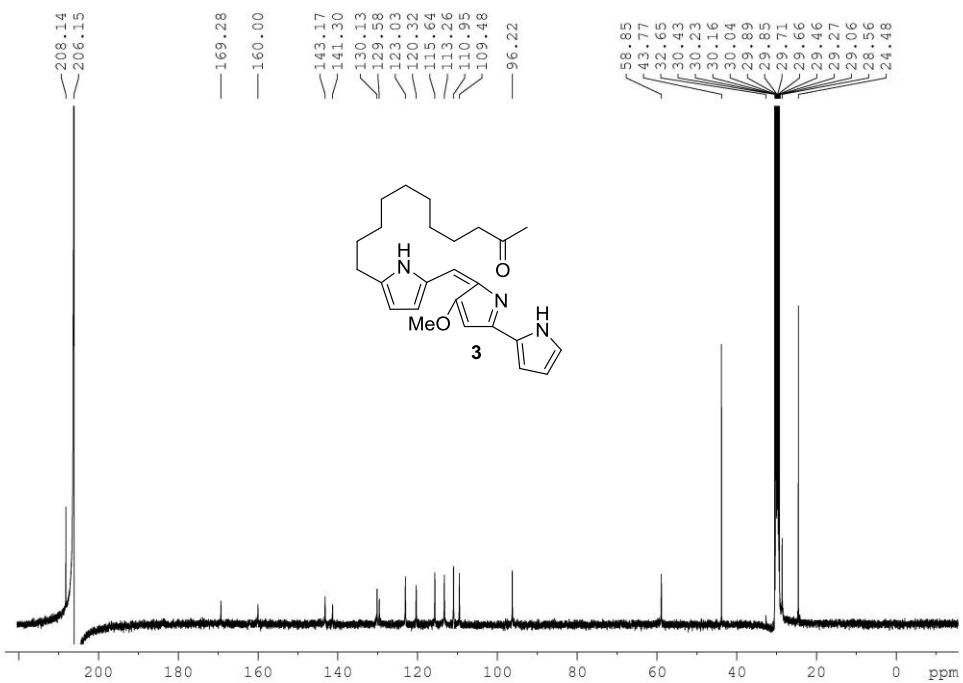




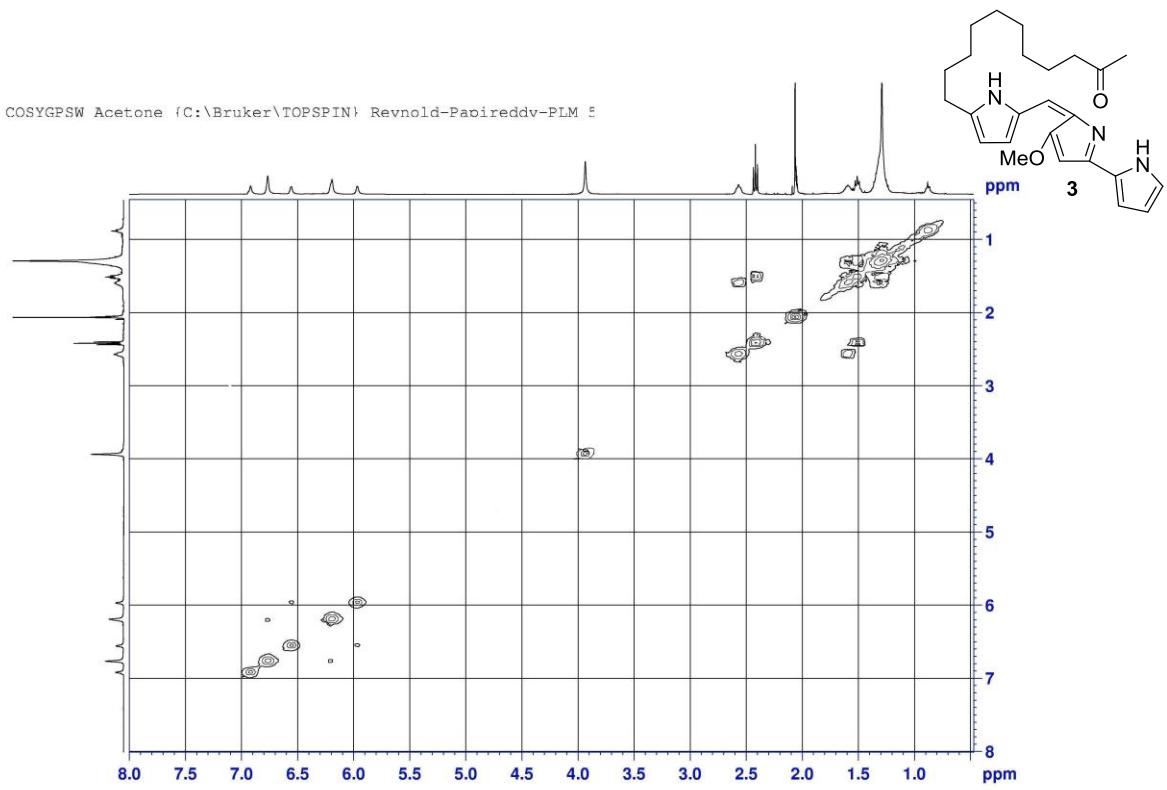


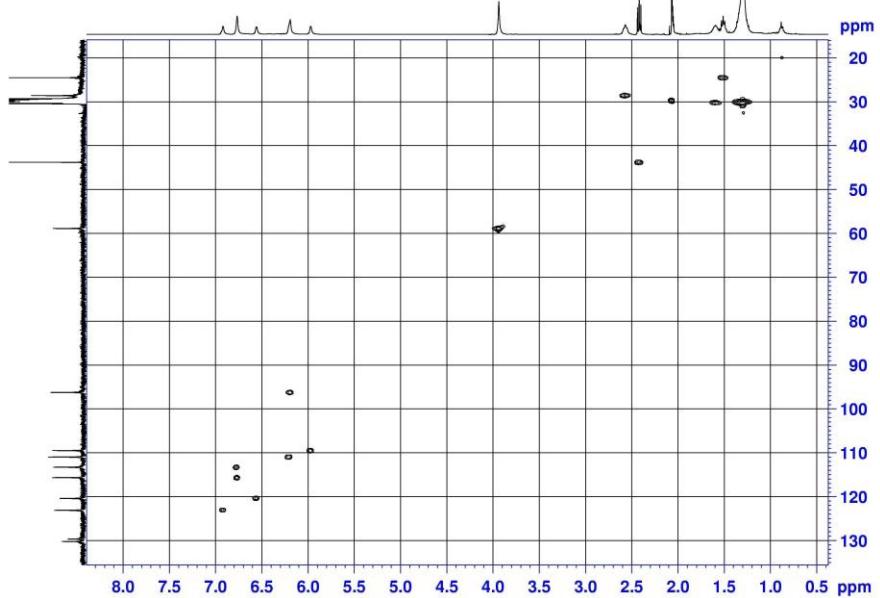
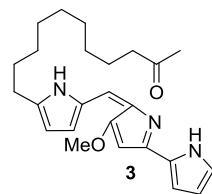


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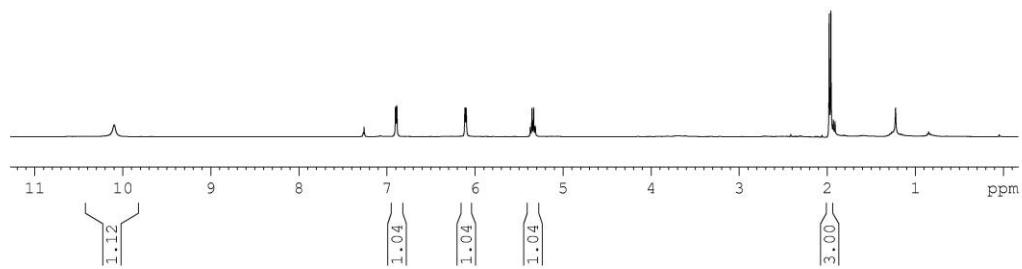
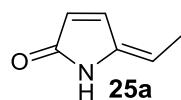
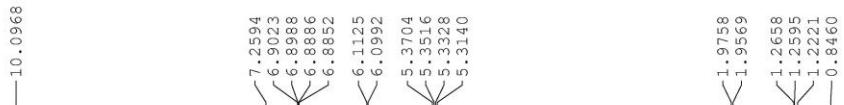


COSYGPSW Acetone {C:\Bruker\TOPSPIN} Revnold-Papireddv-PLM 5

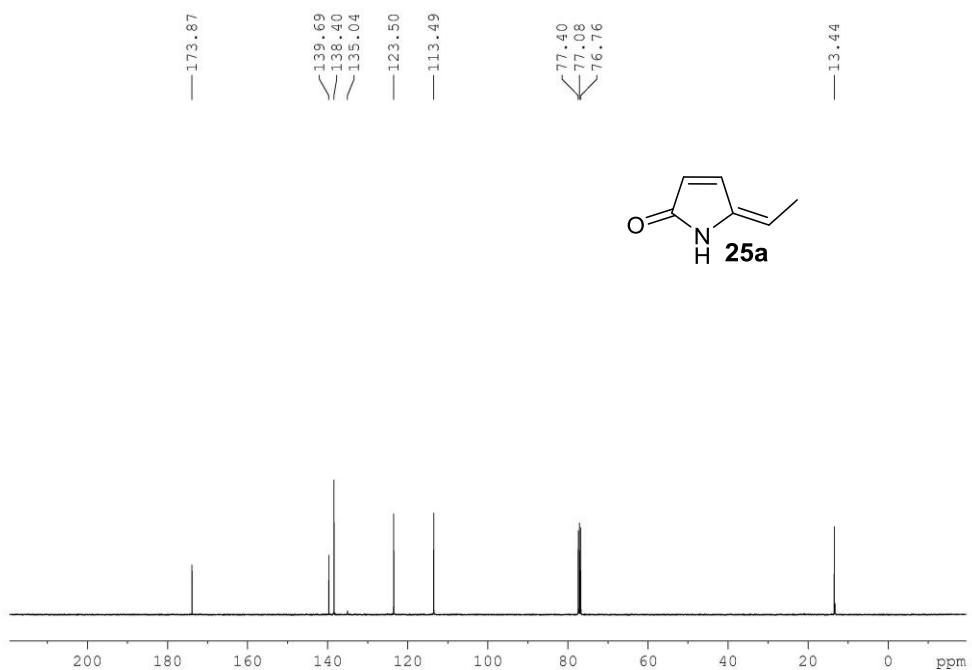




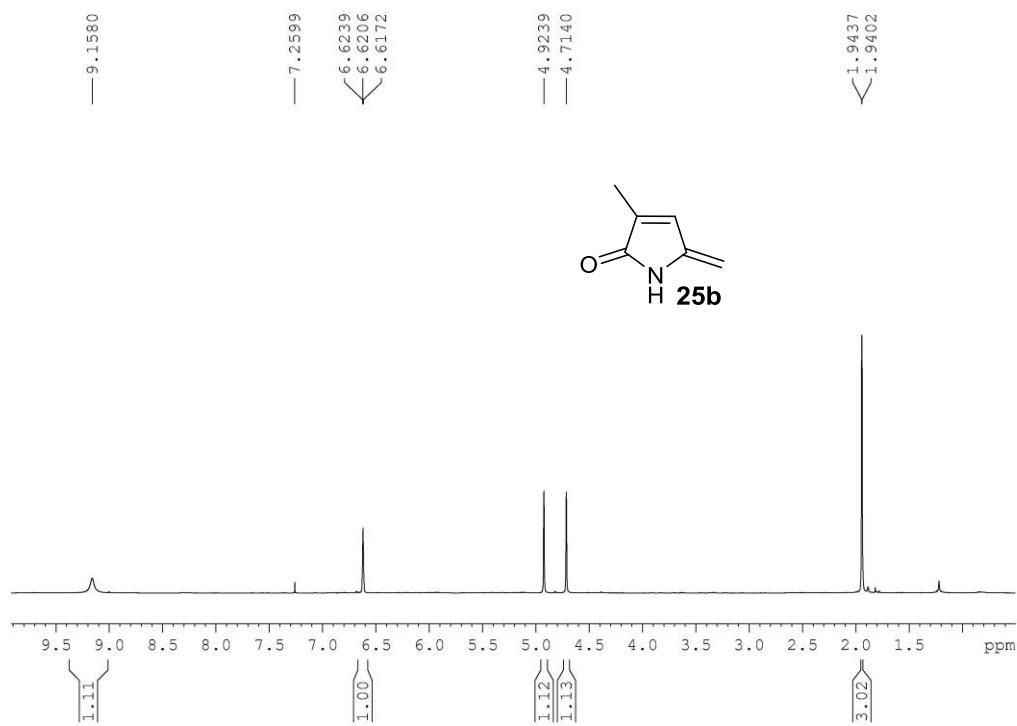
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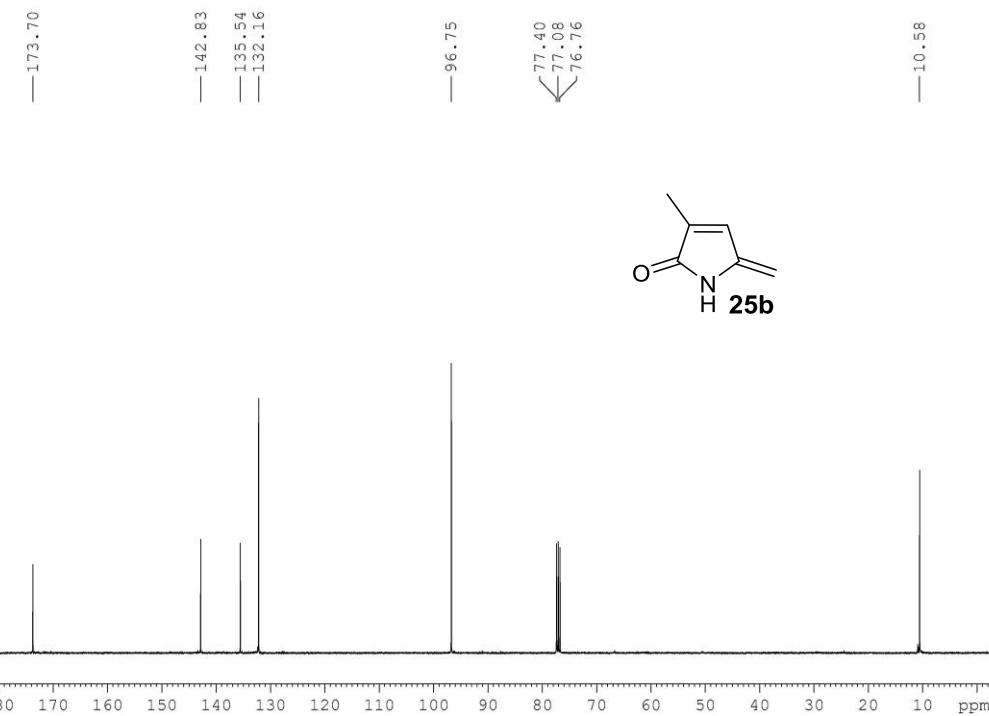
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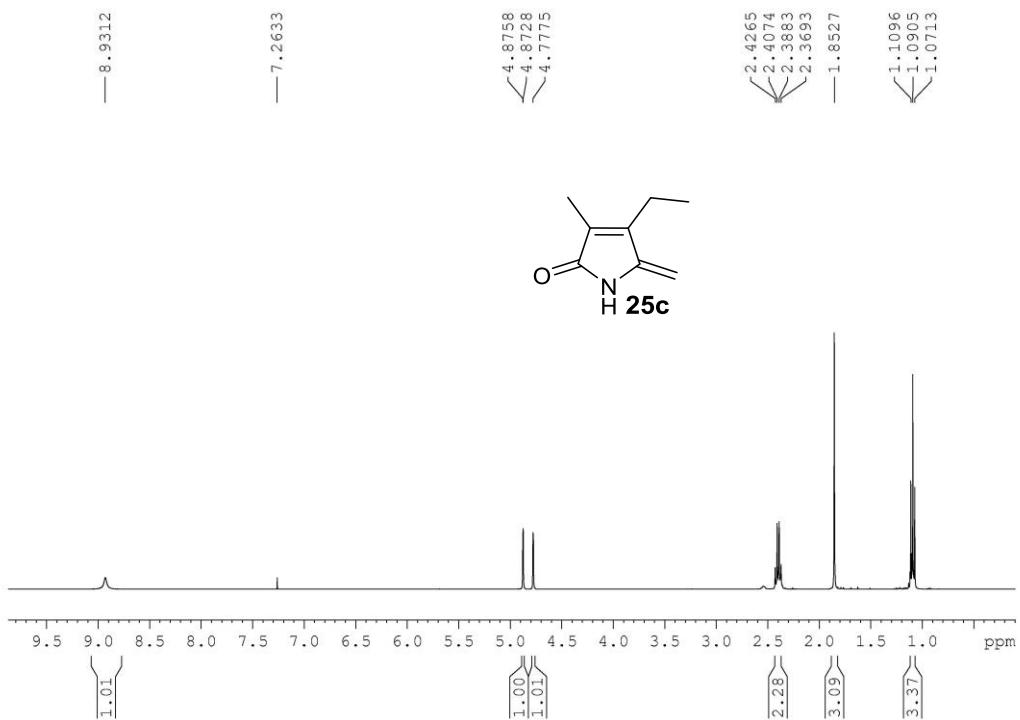
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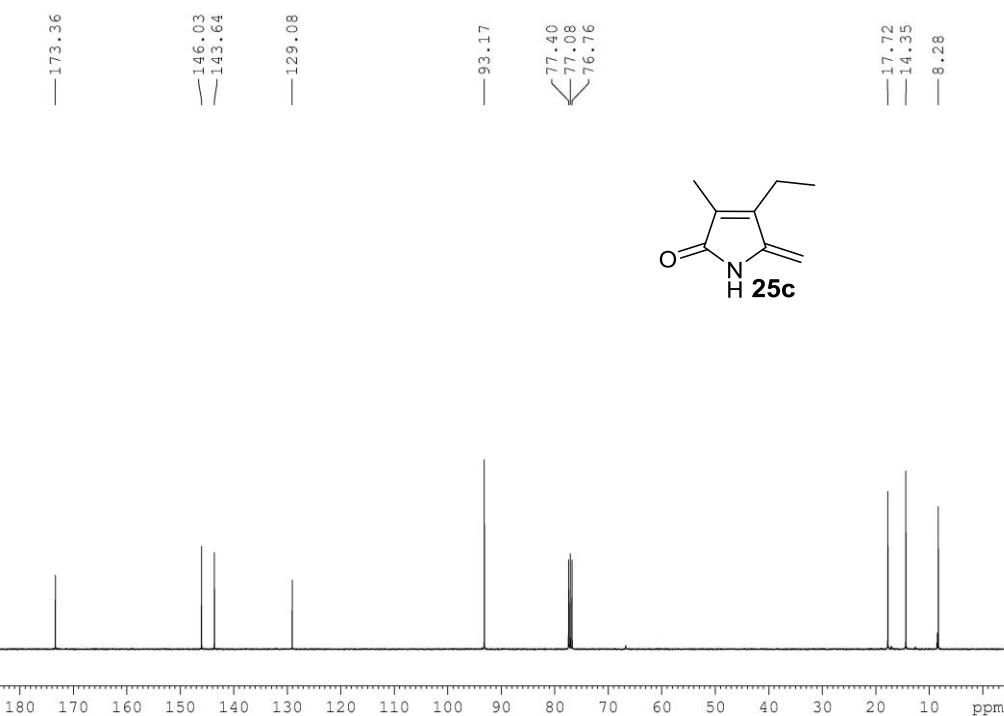
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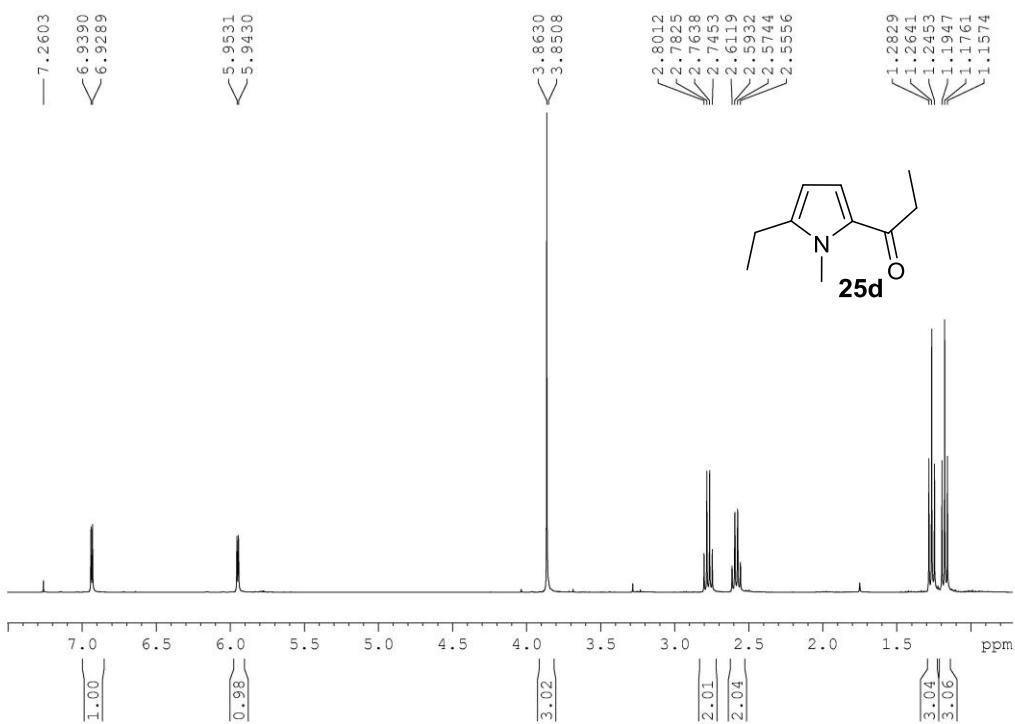
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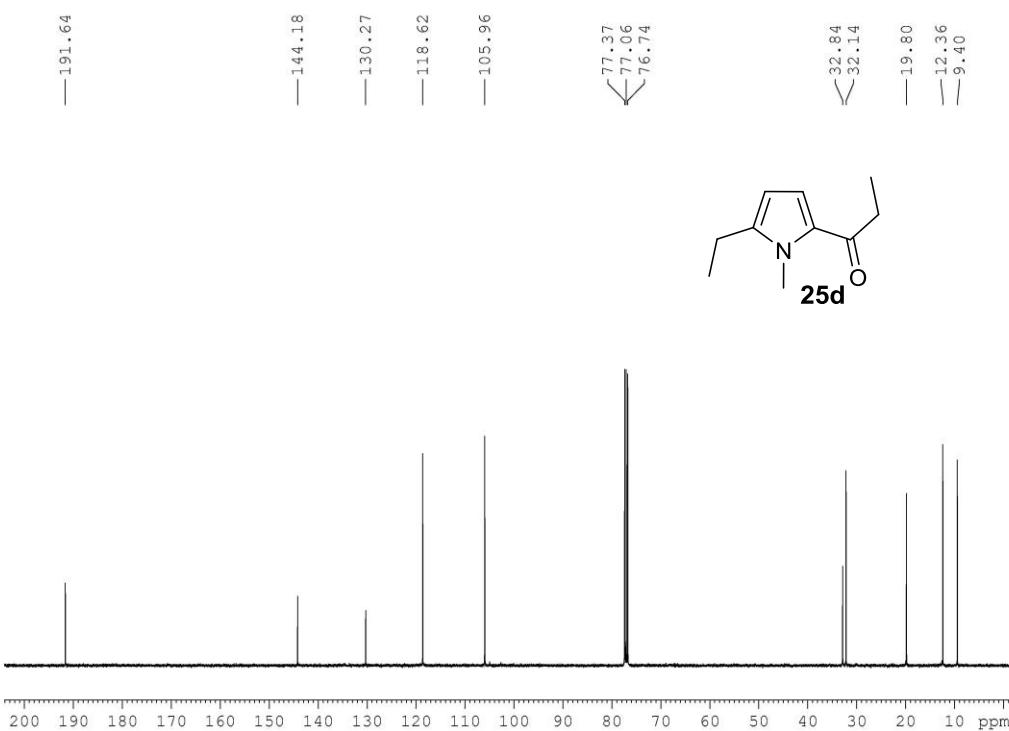
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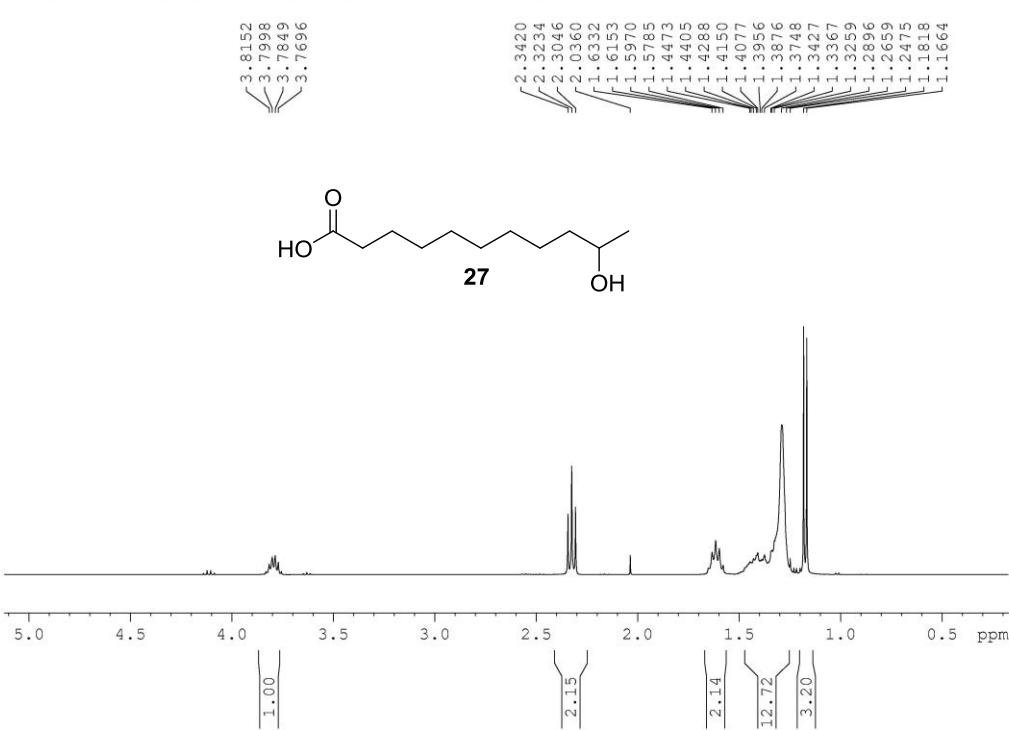
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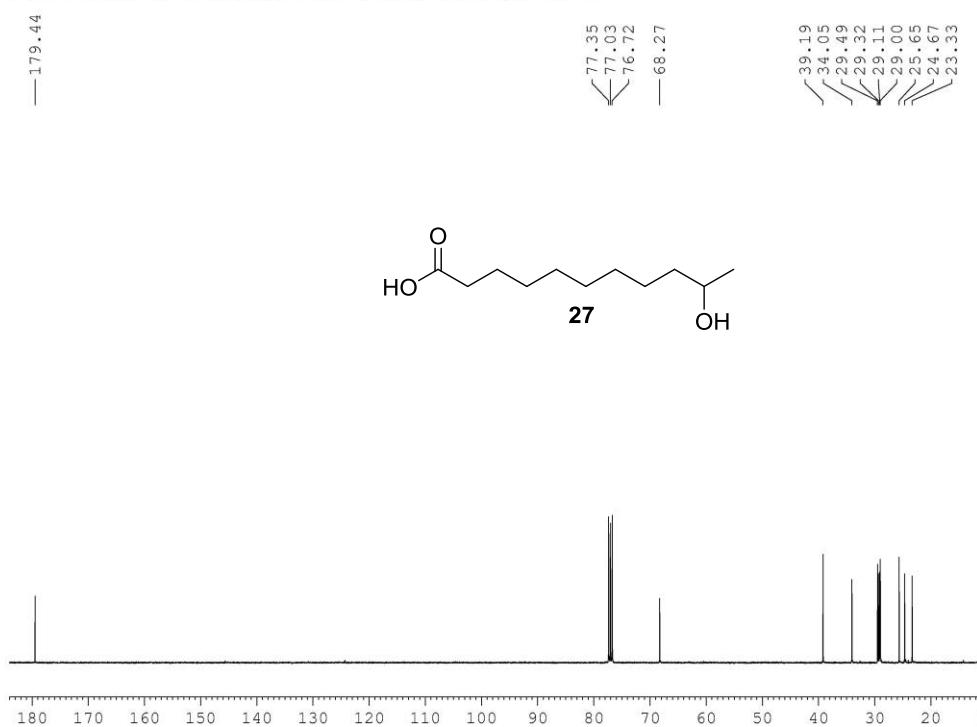
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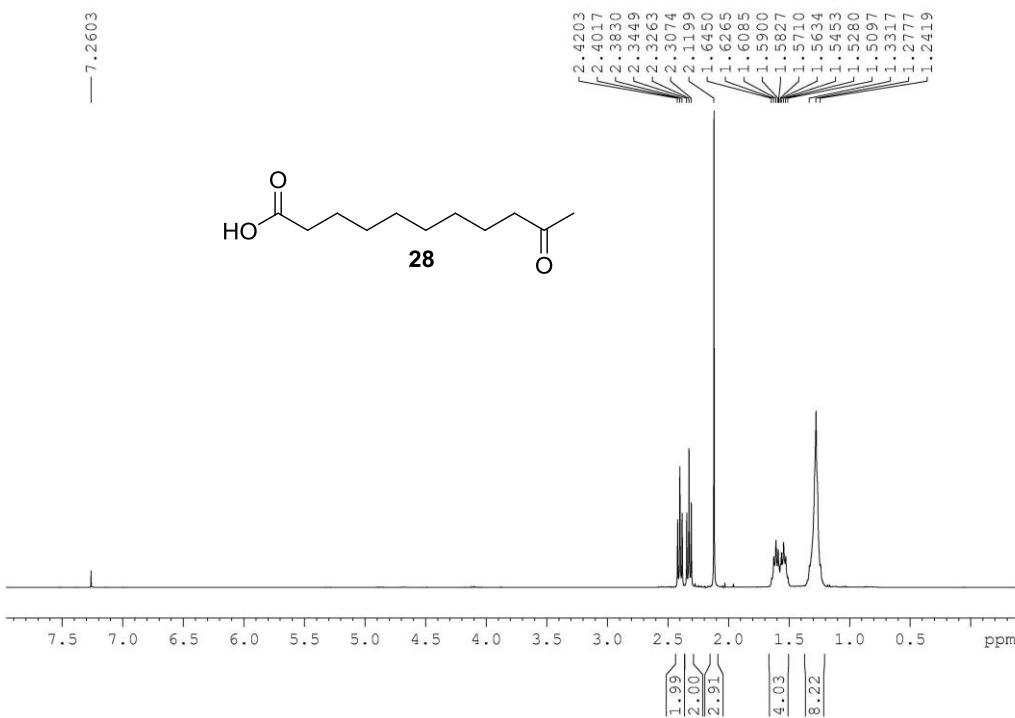
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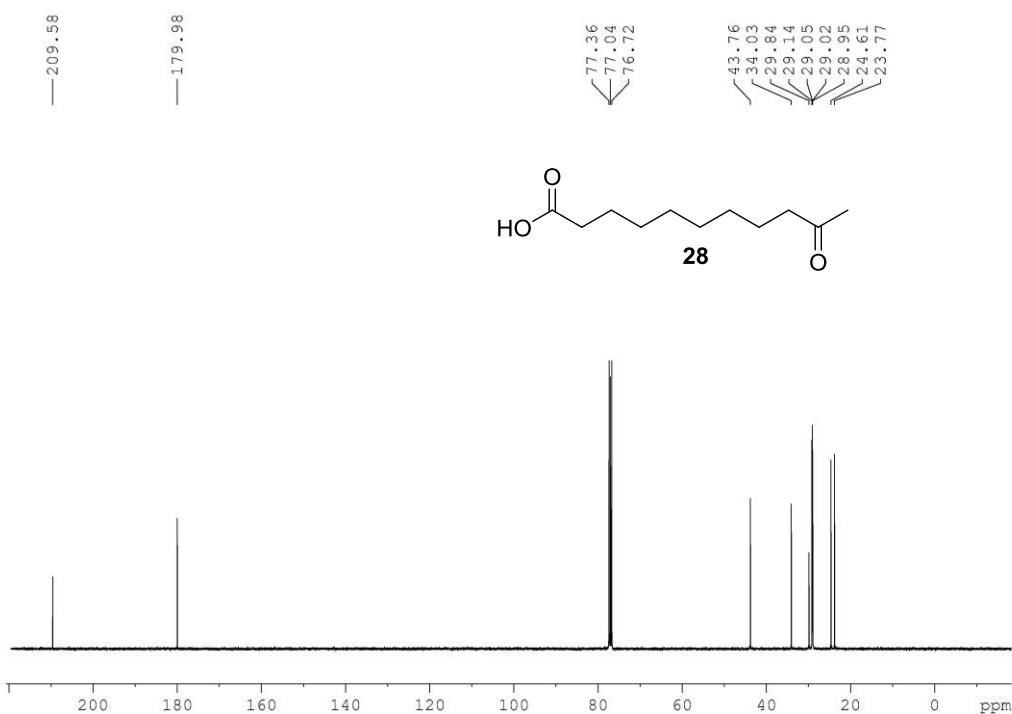
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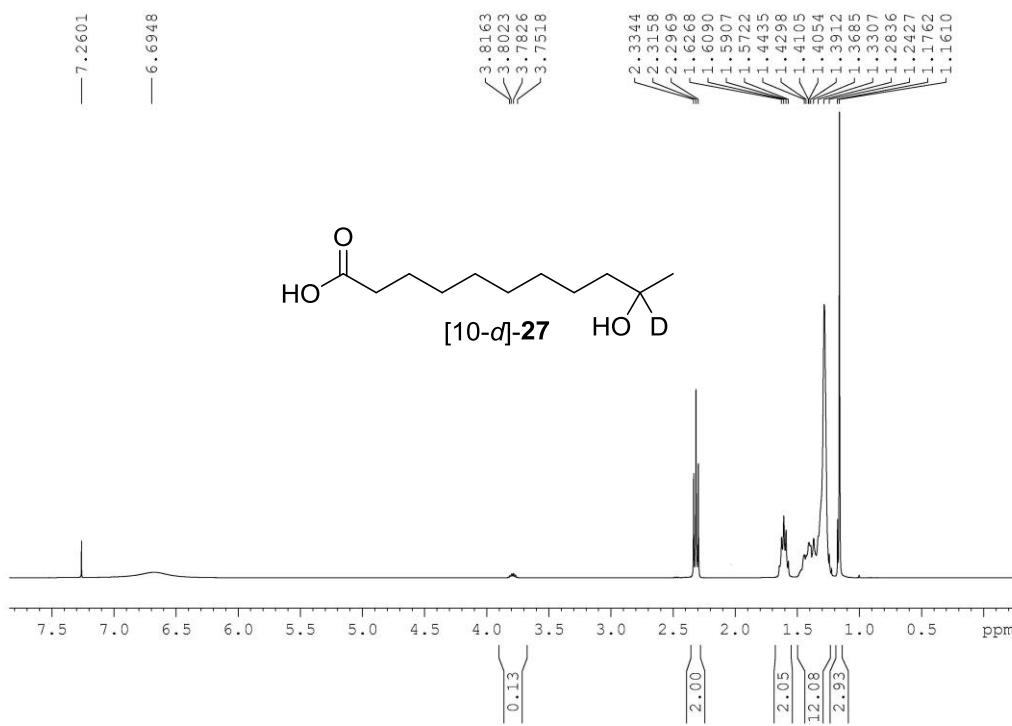
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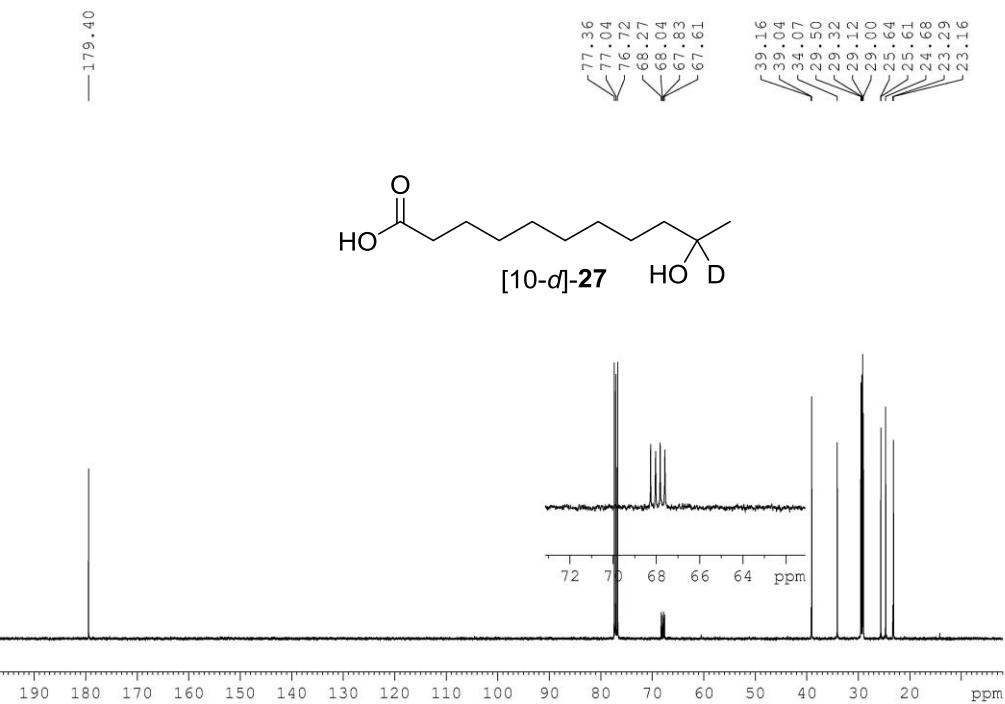
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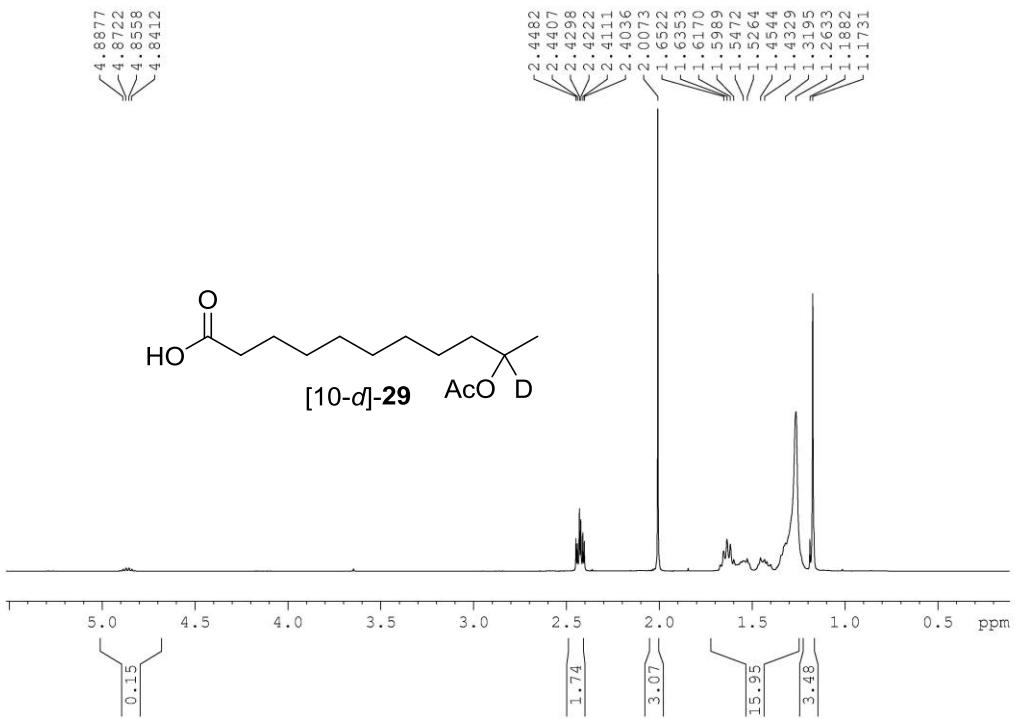
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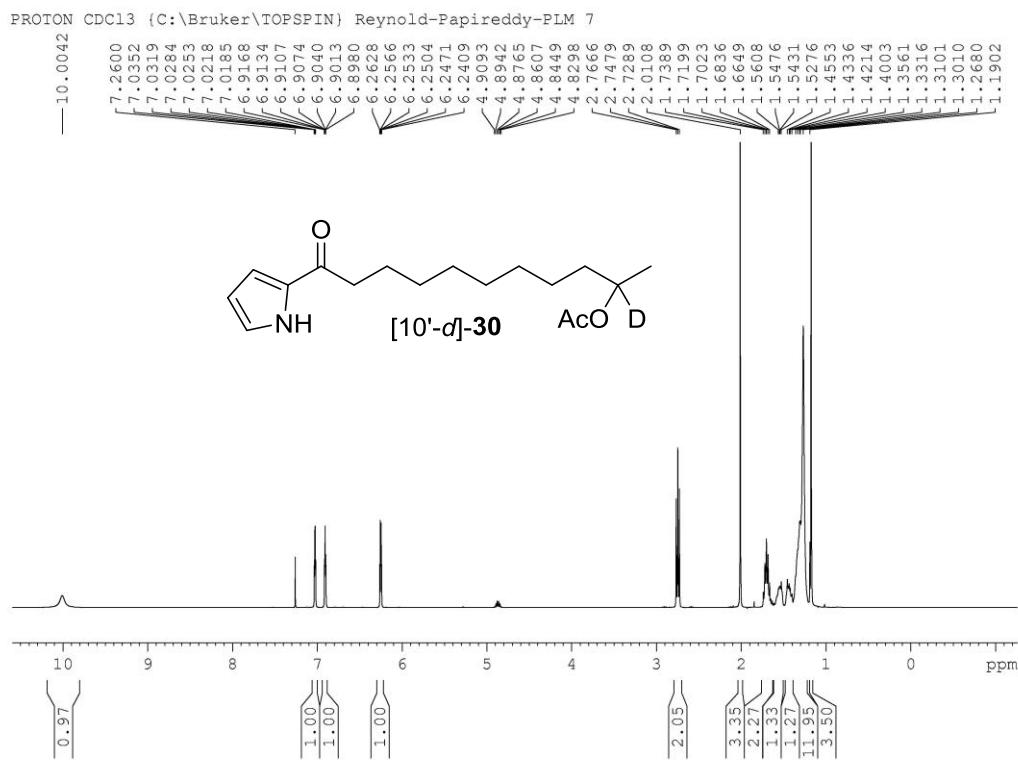


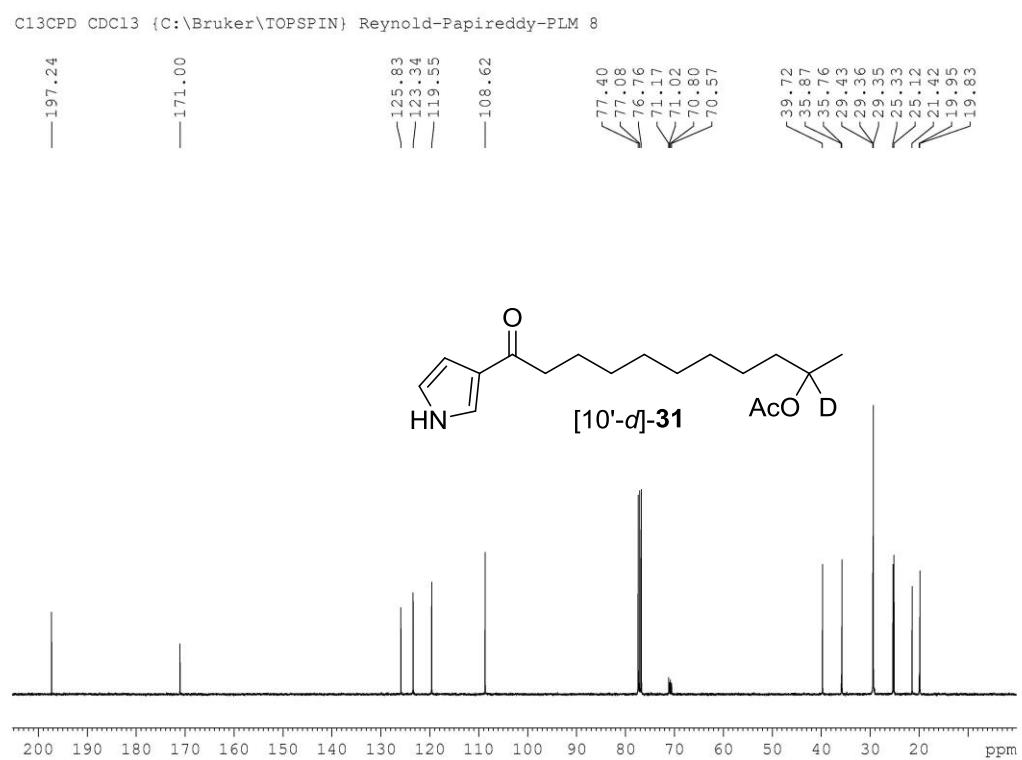
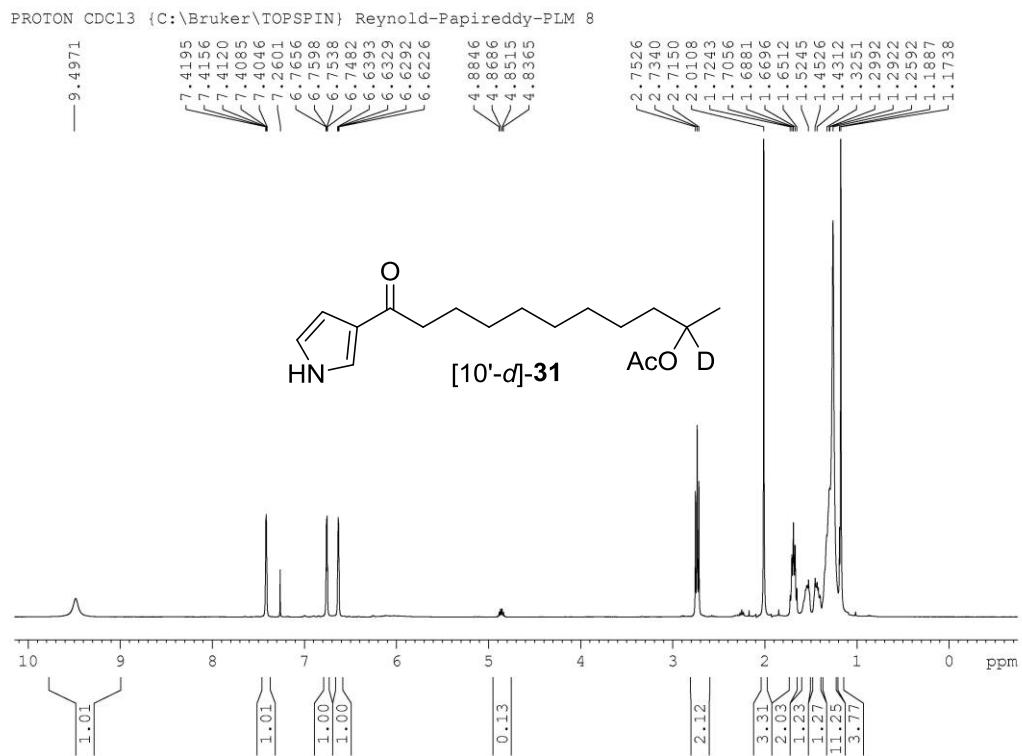
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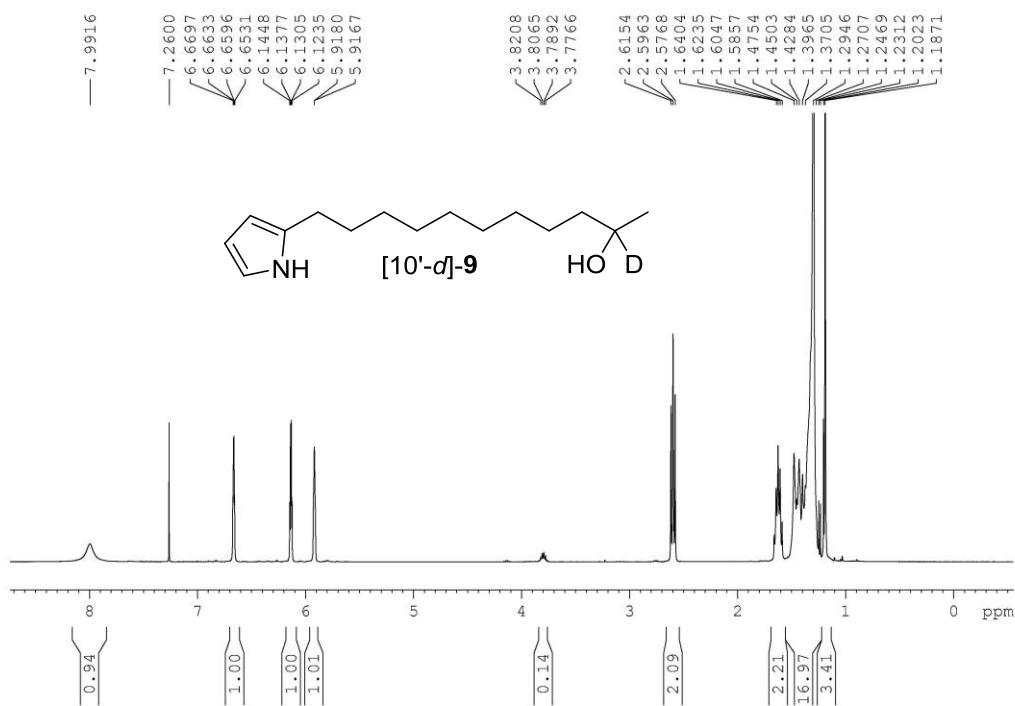
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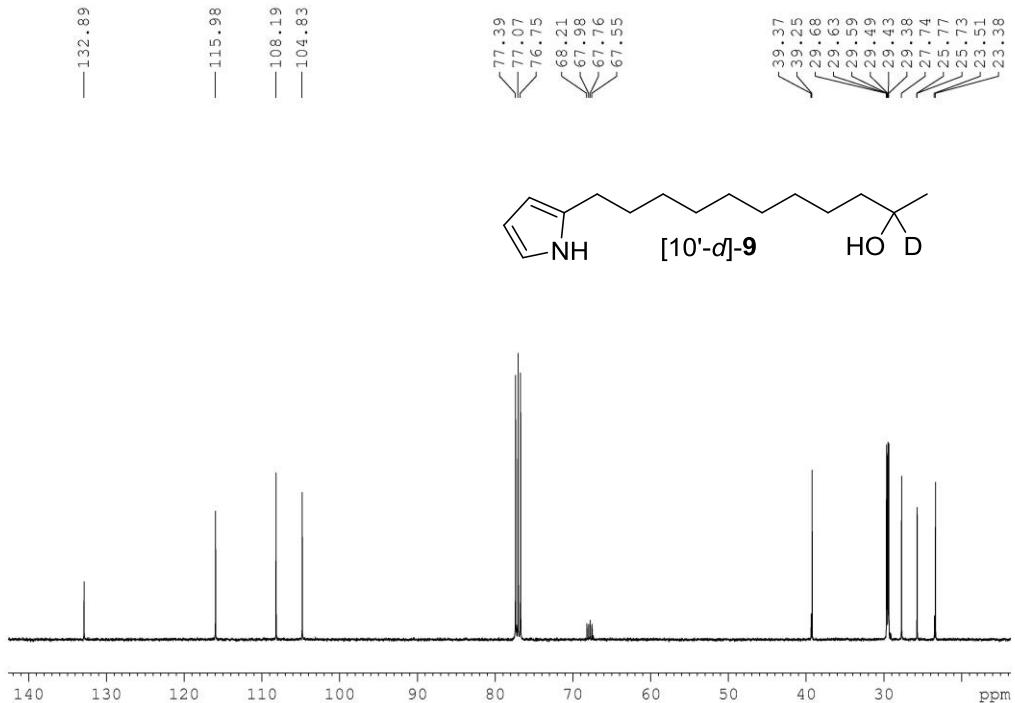




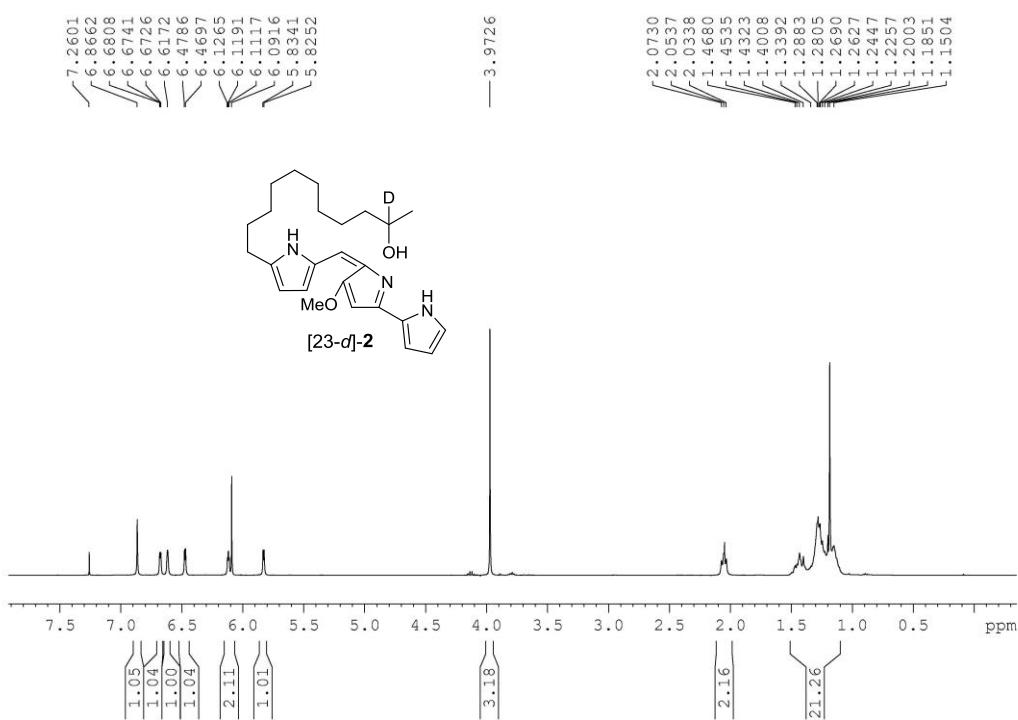
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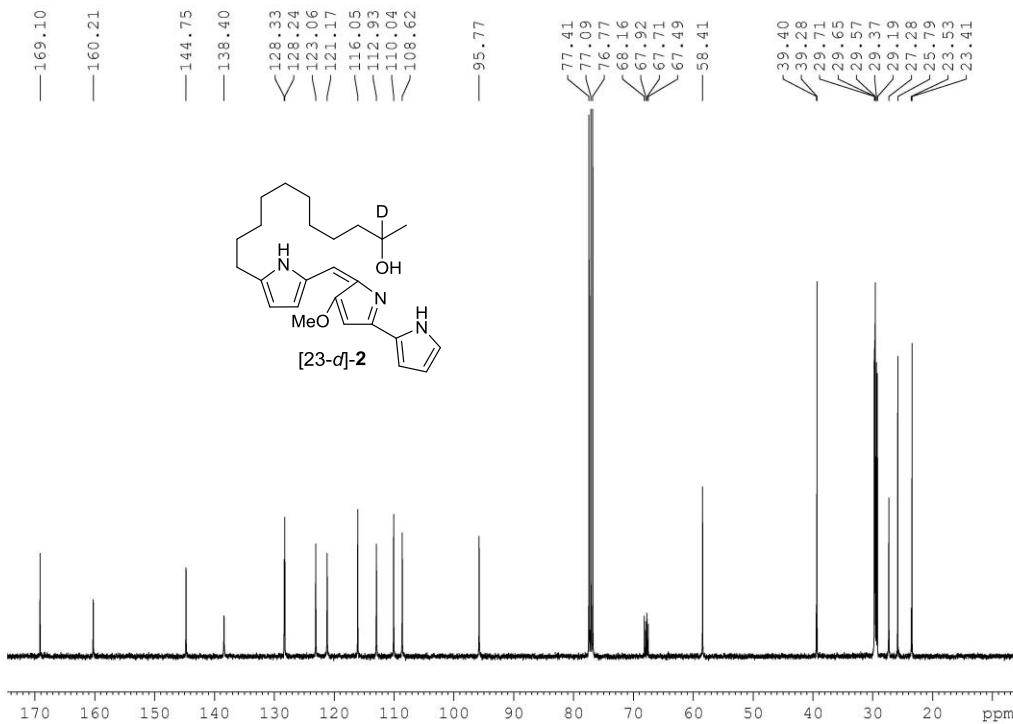
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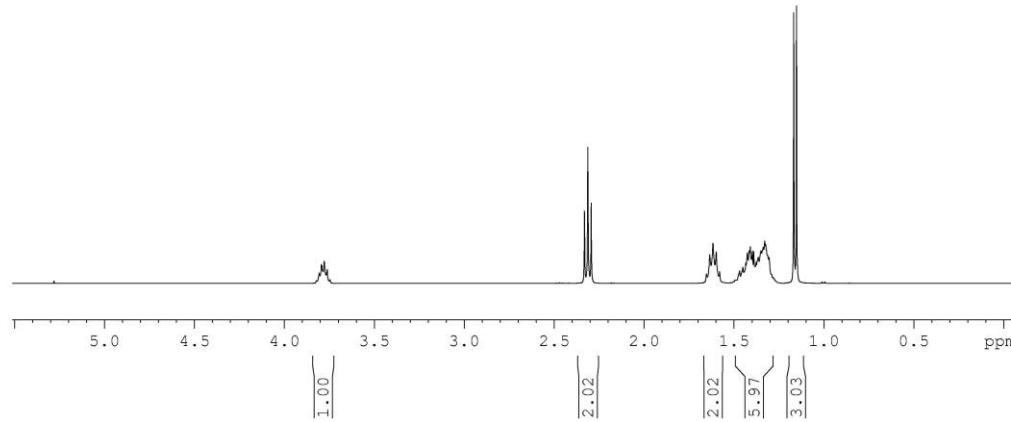
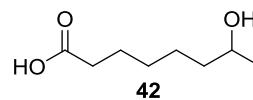
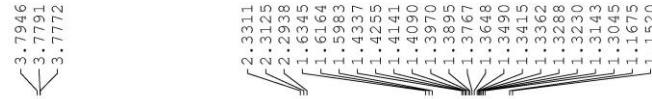
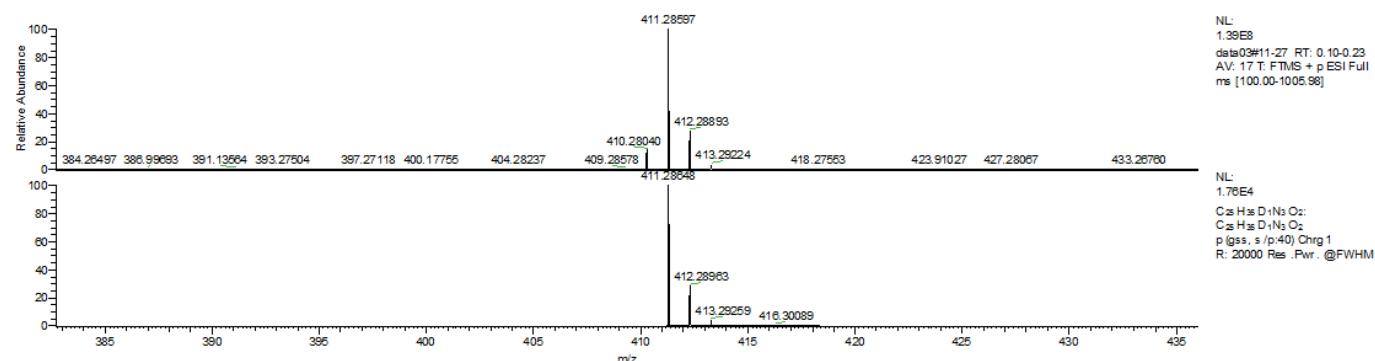
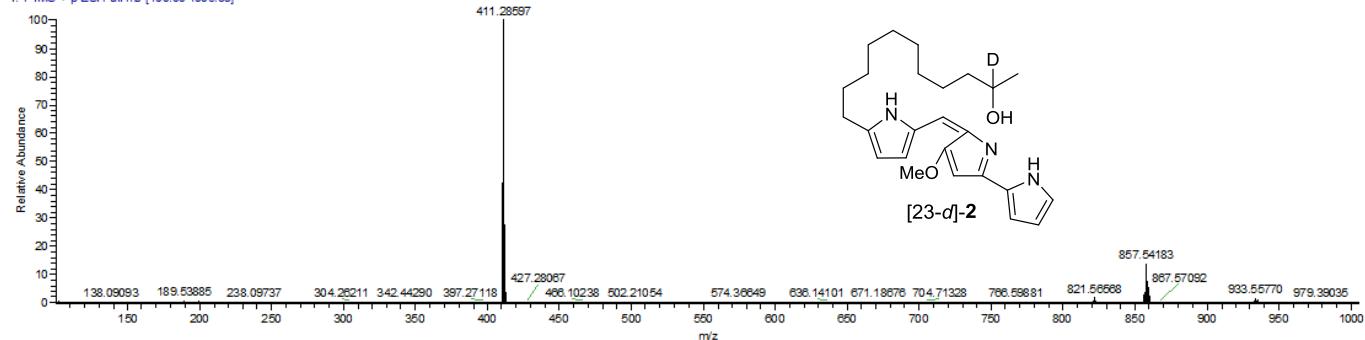
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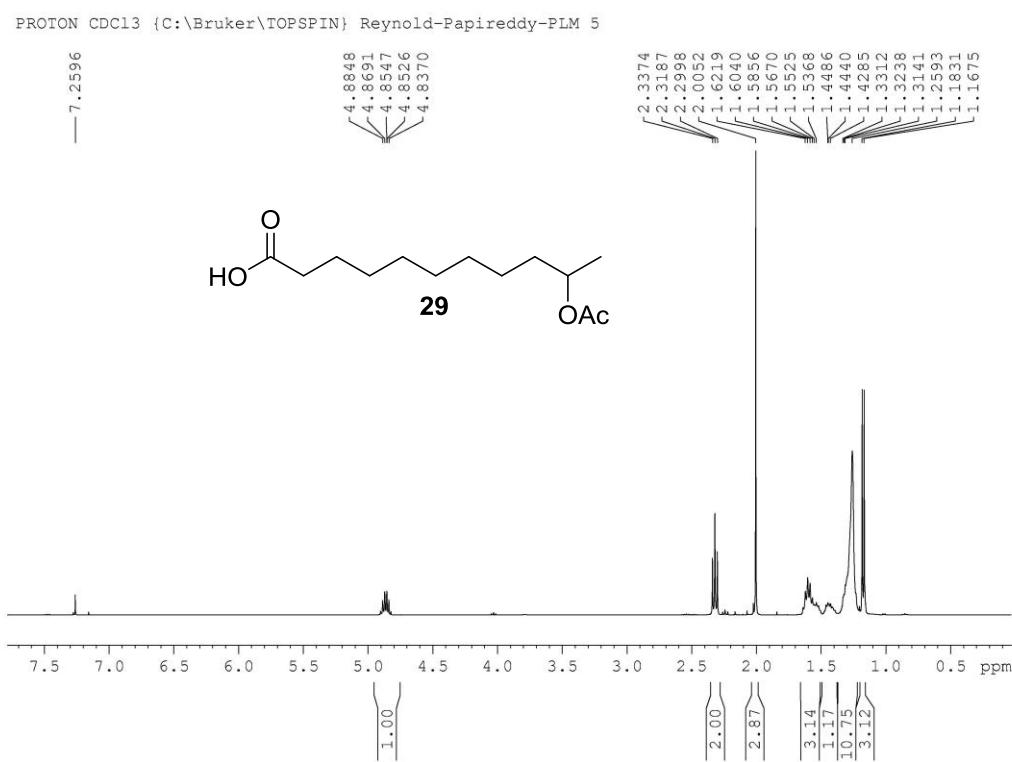
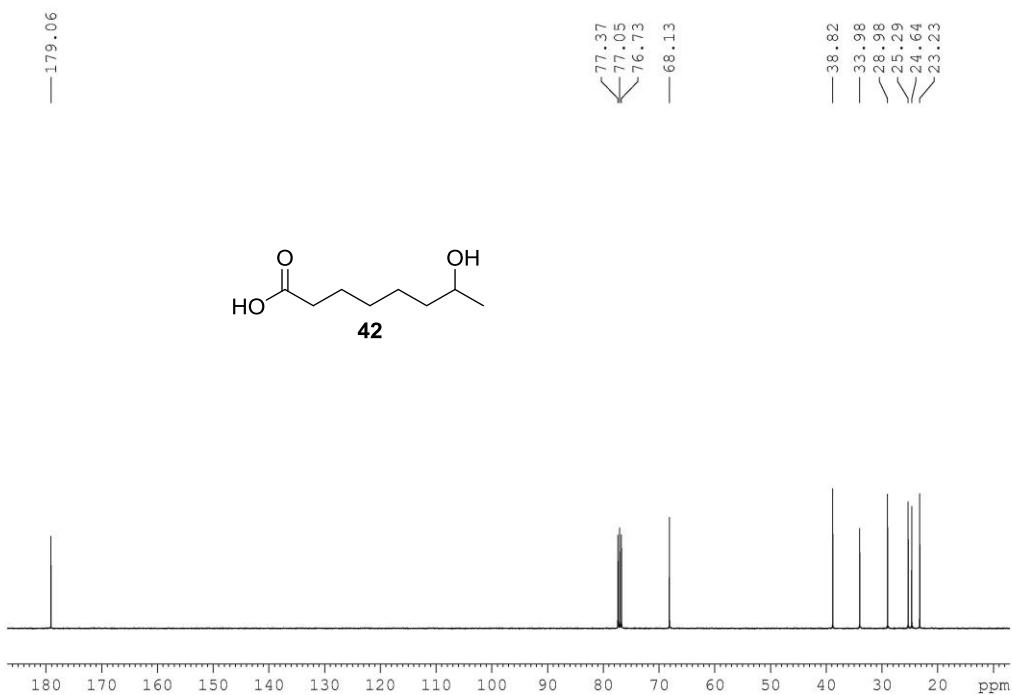


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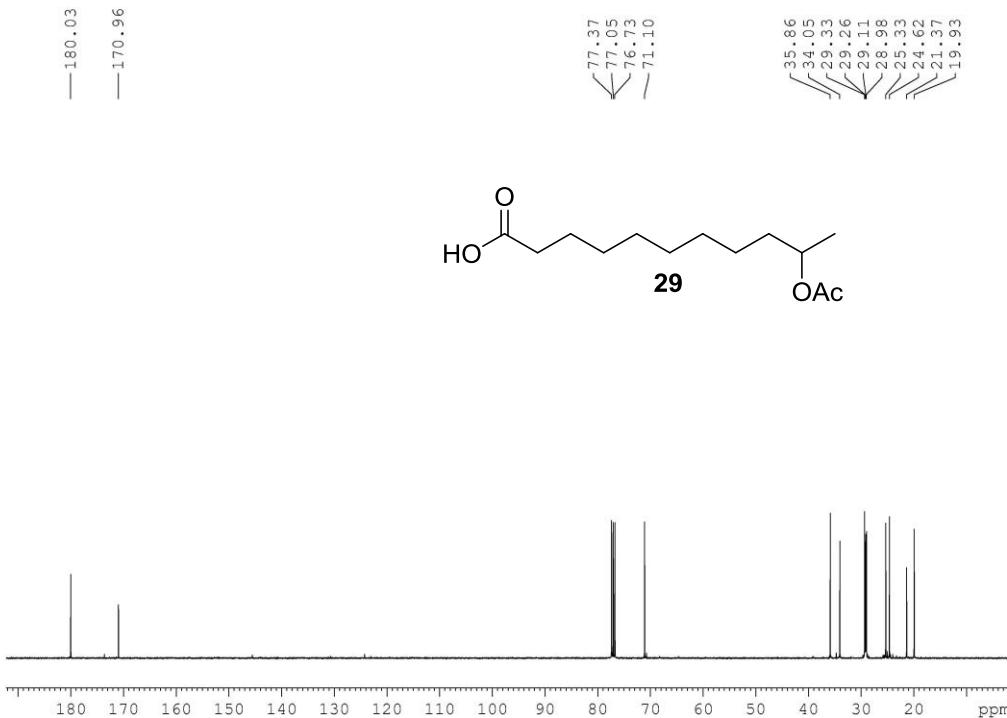


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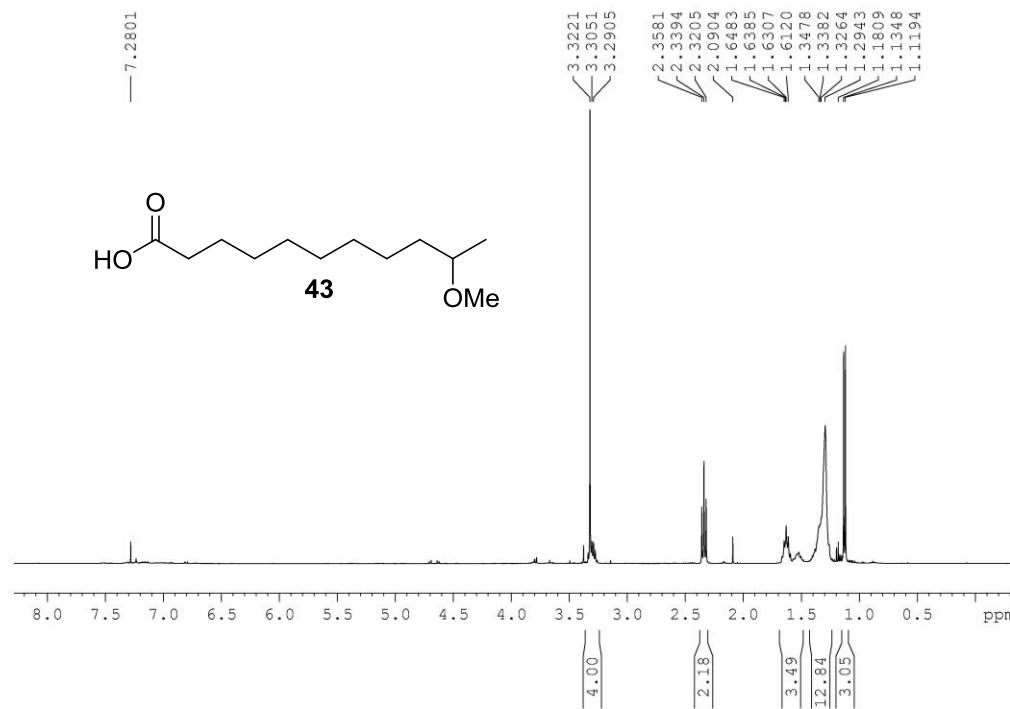




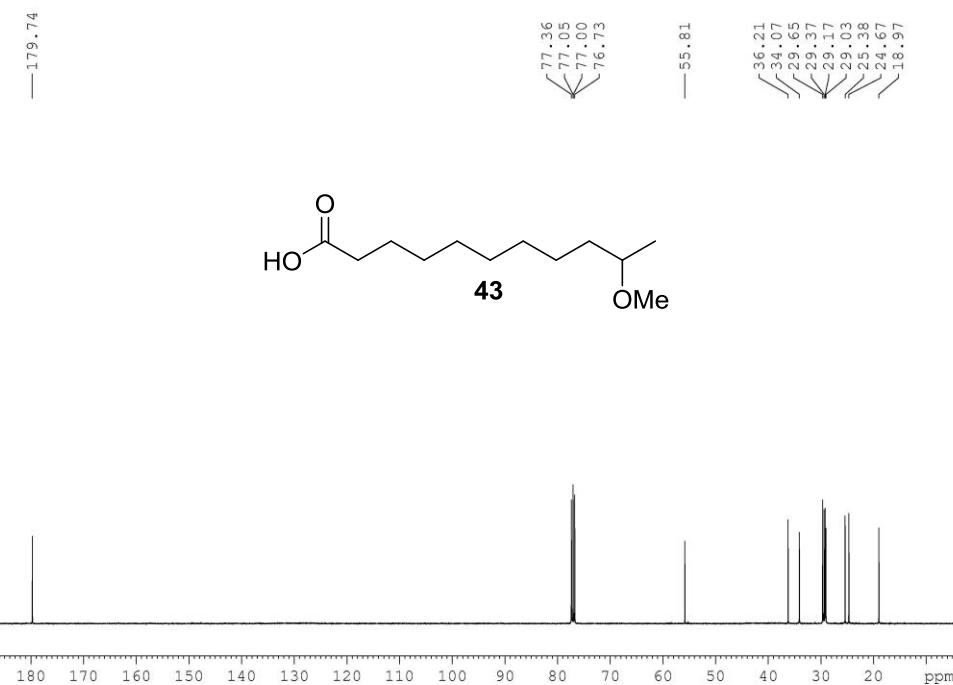
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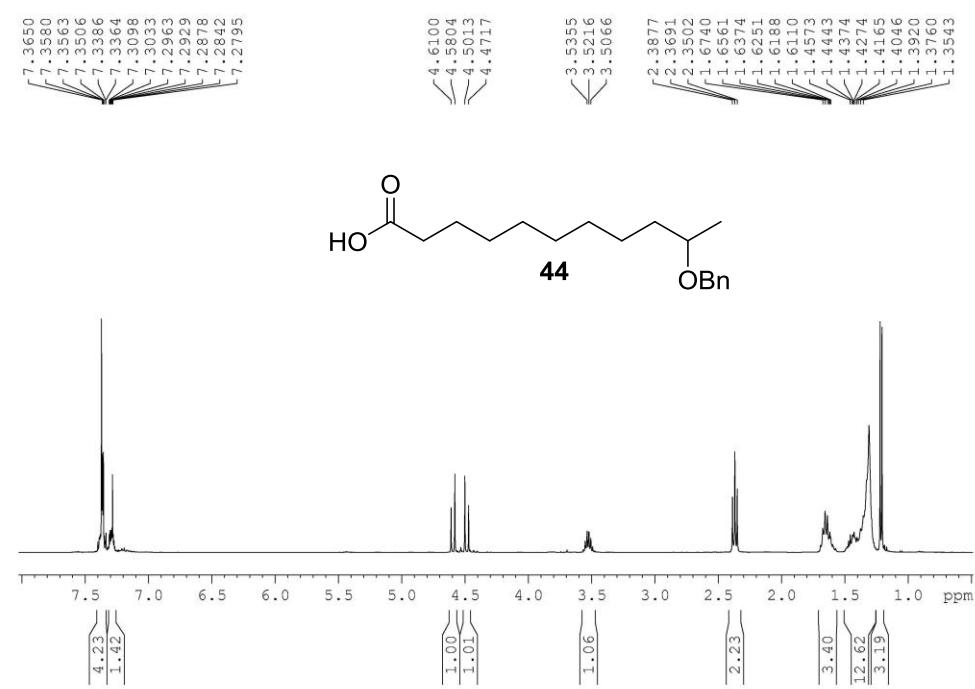
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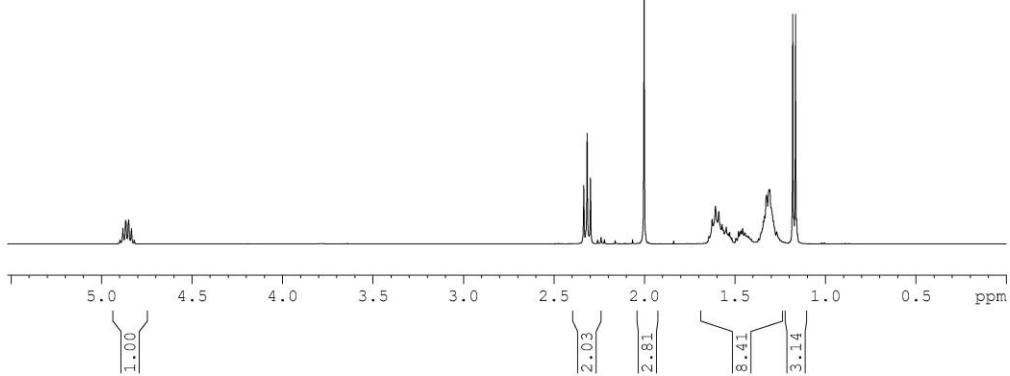
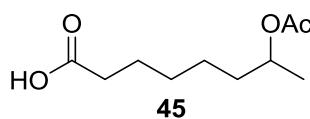
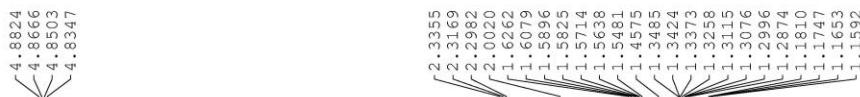
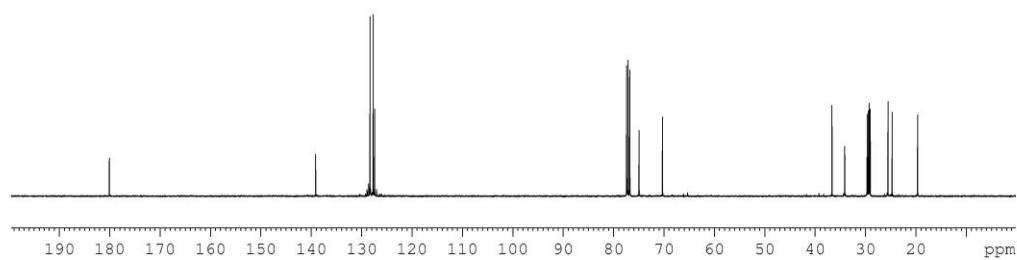
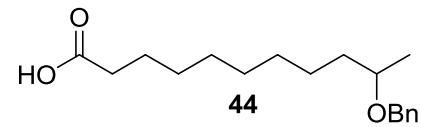


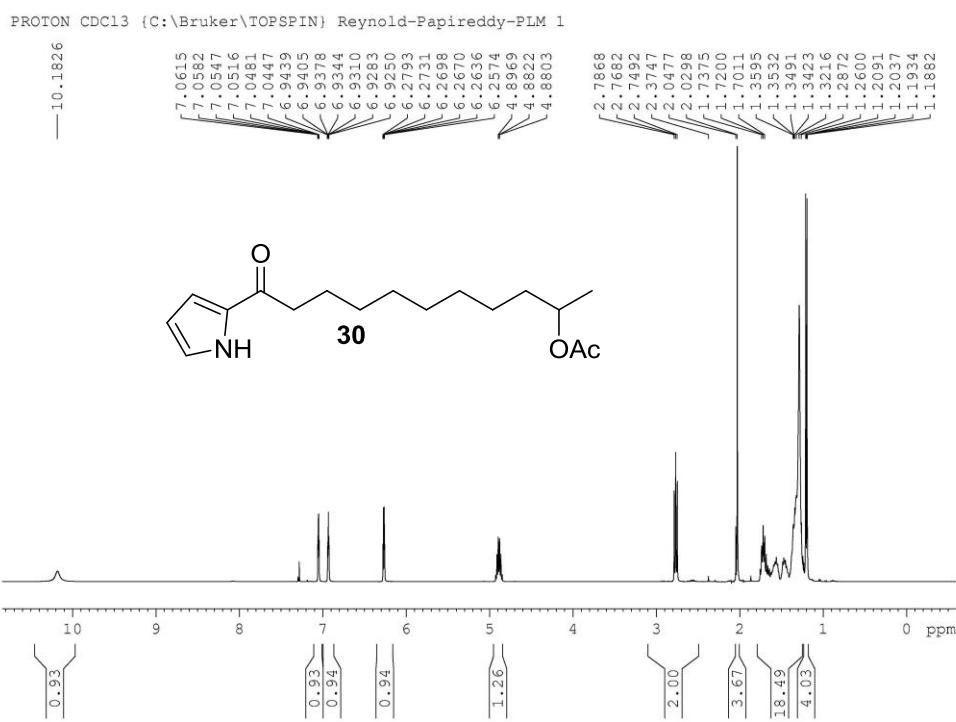
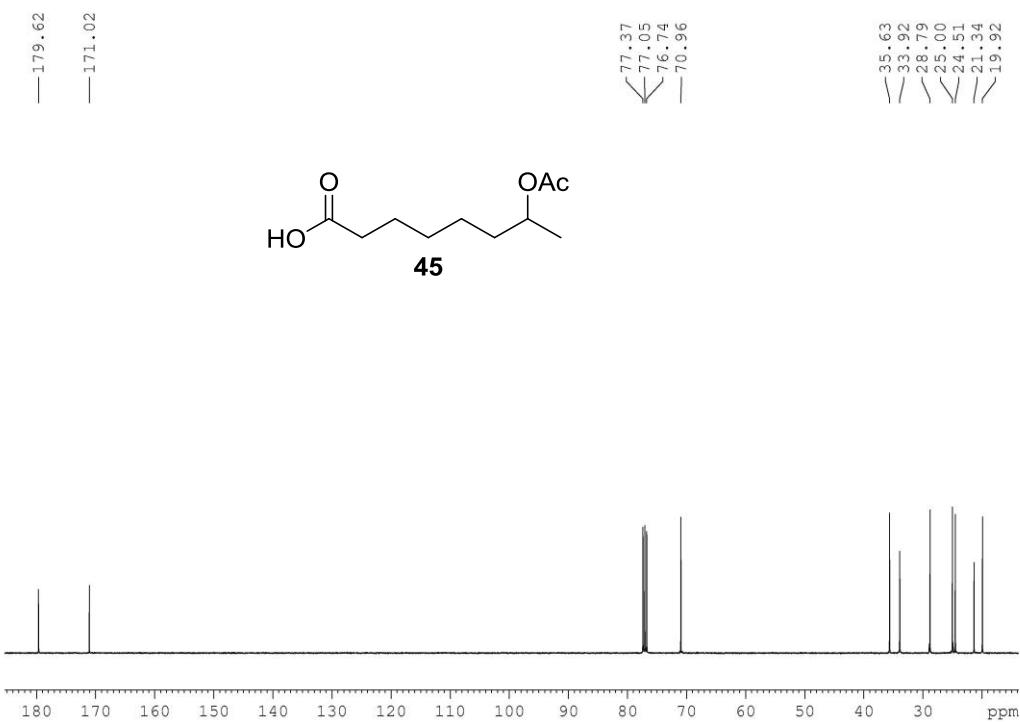
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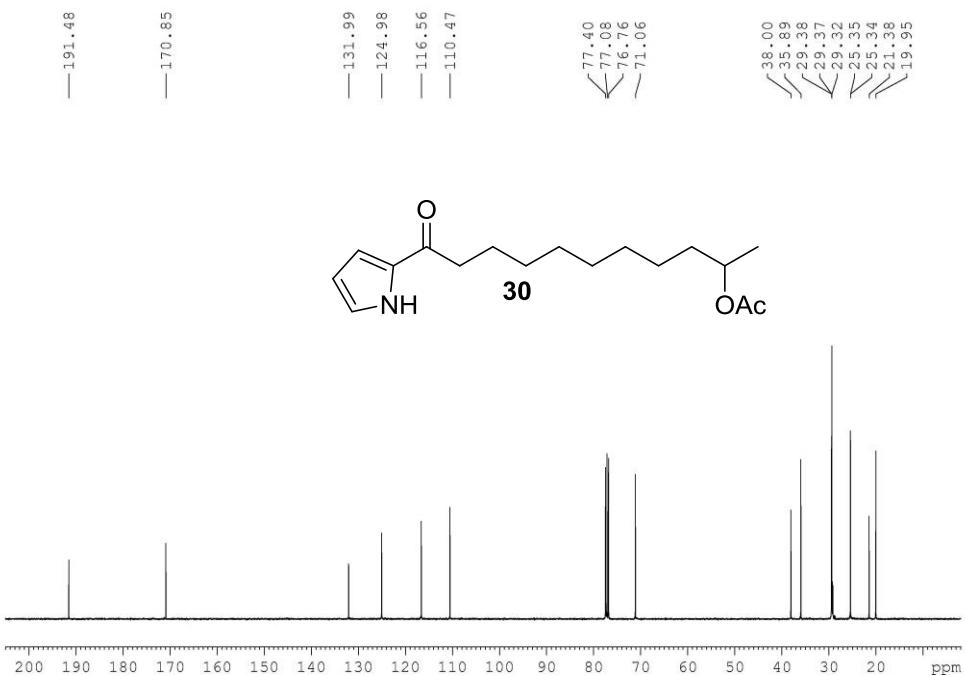
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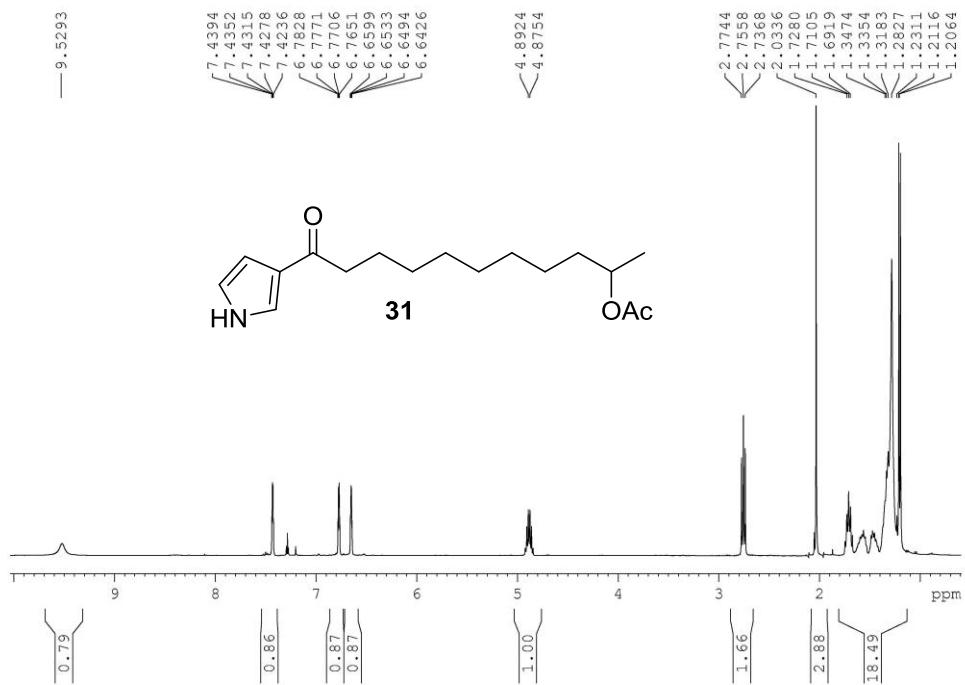




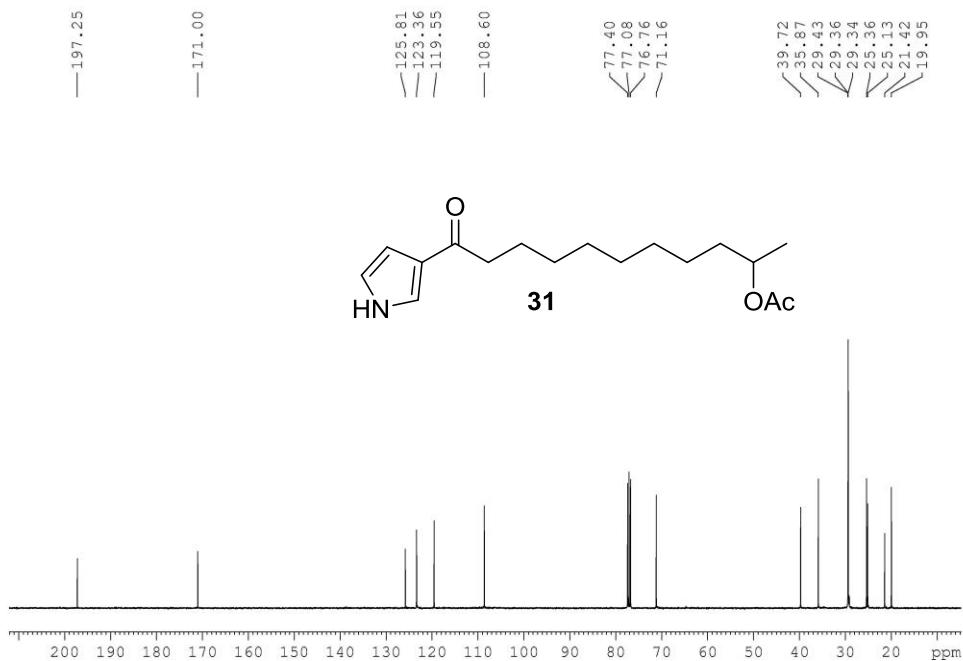
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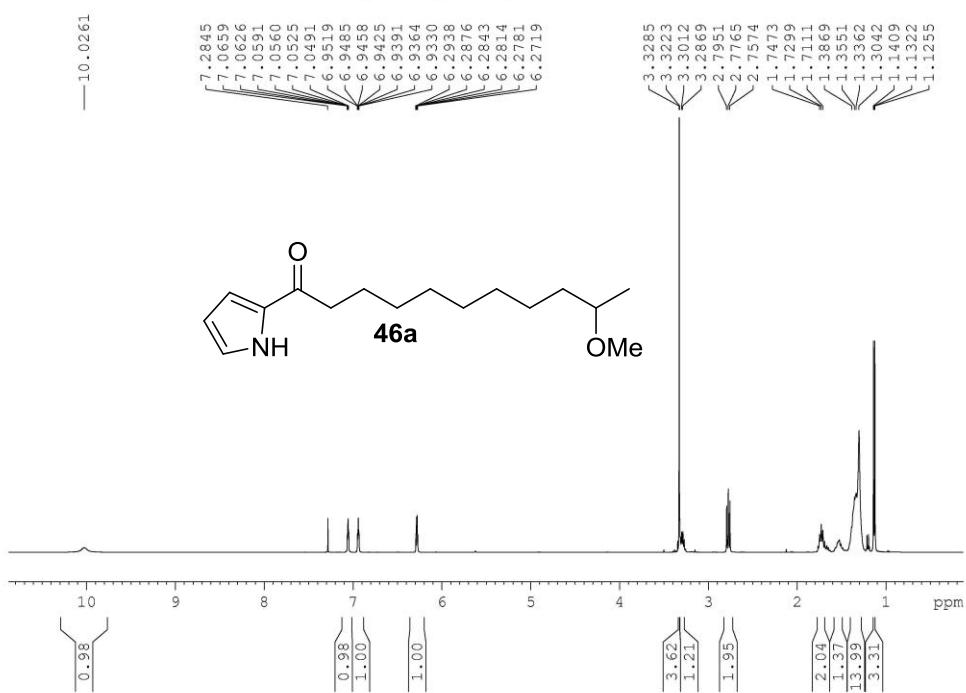
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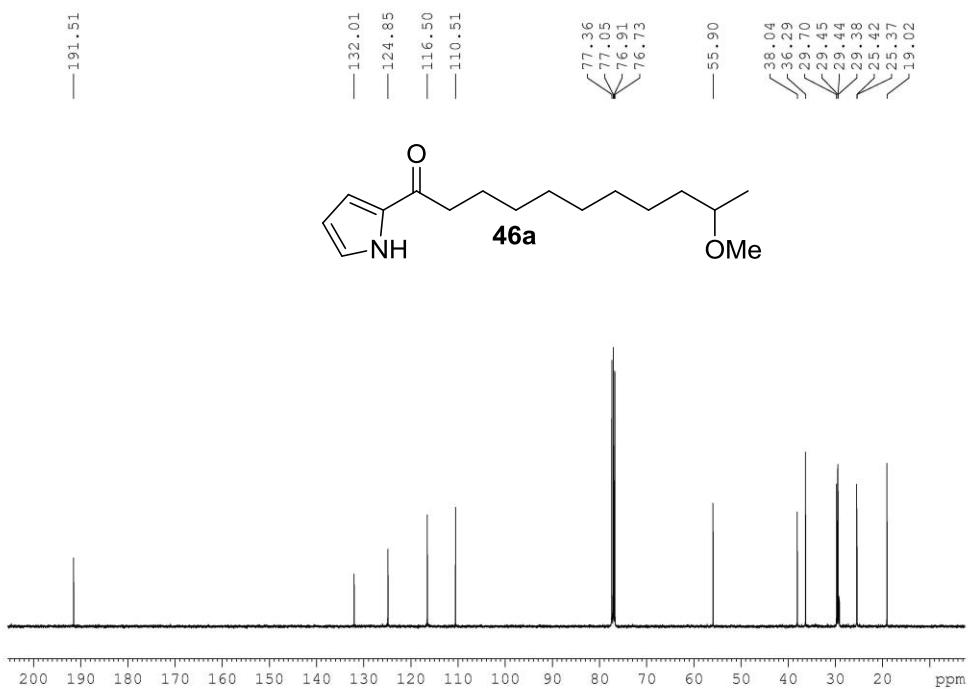
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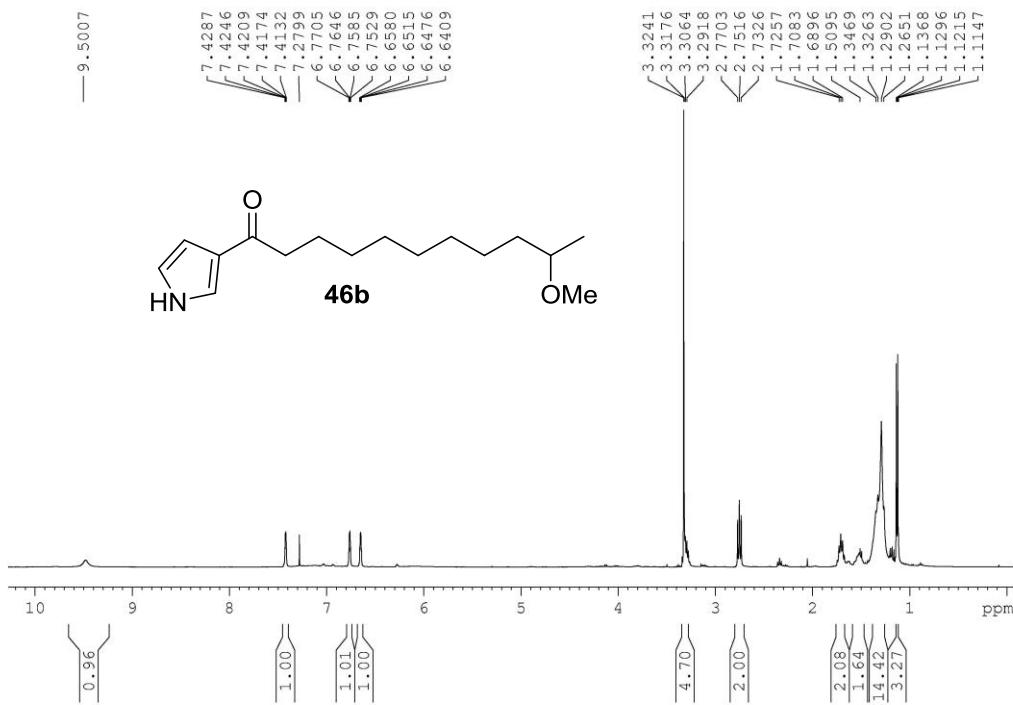
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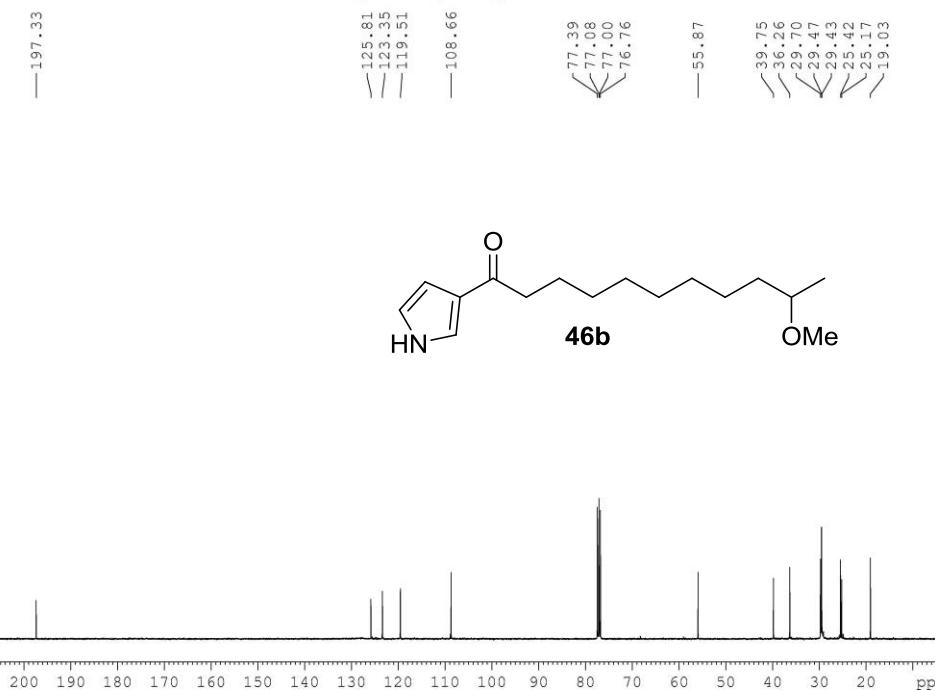
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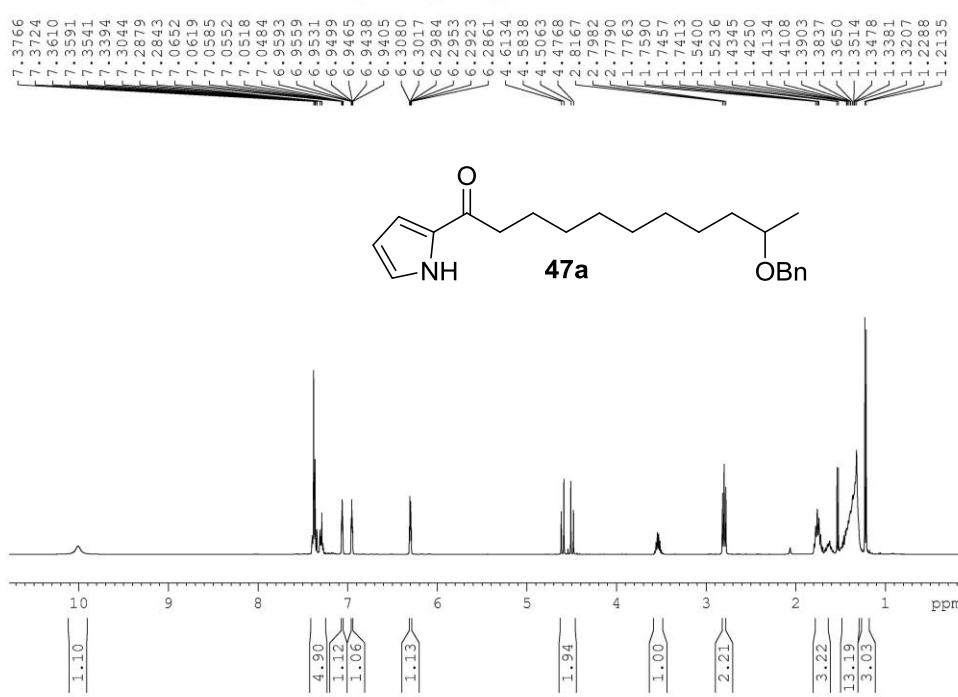
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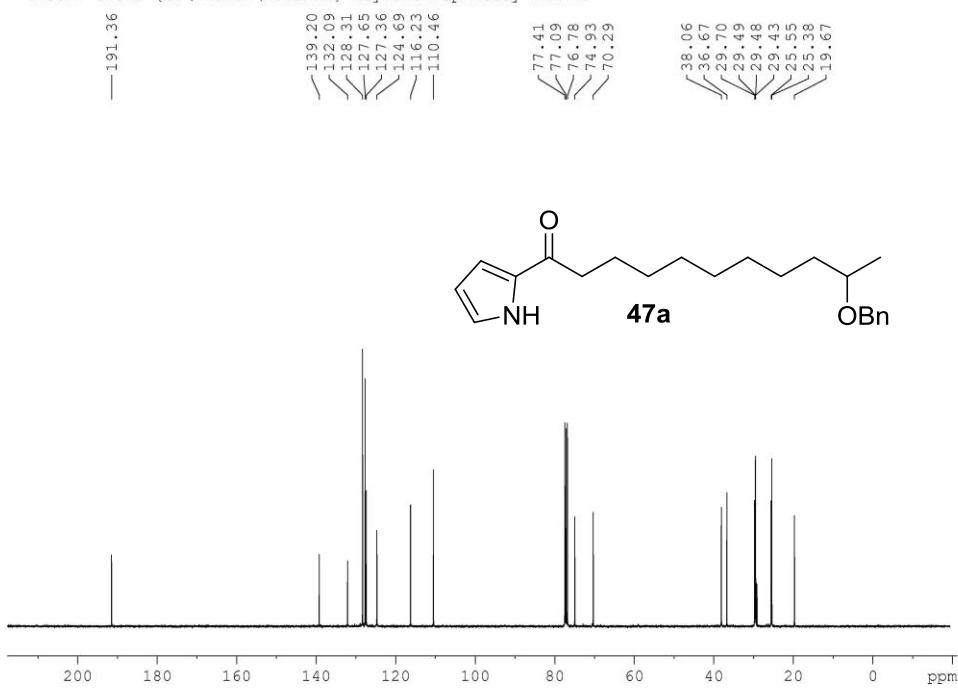
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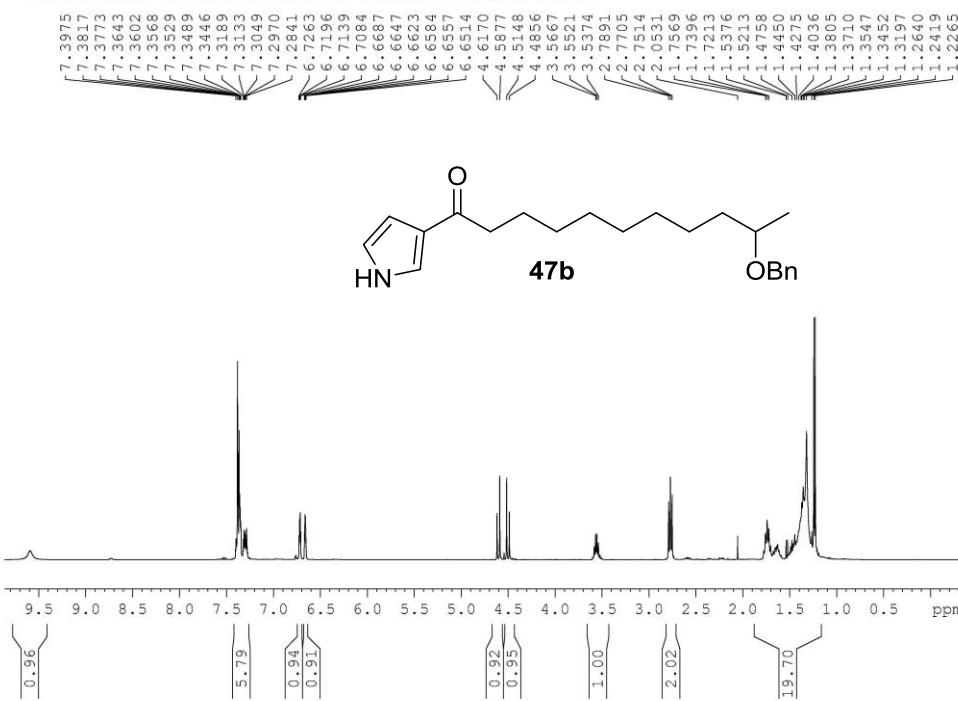
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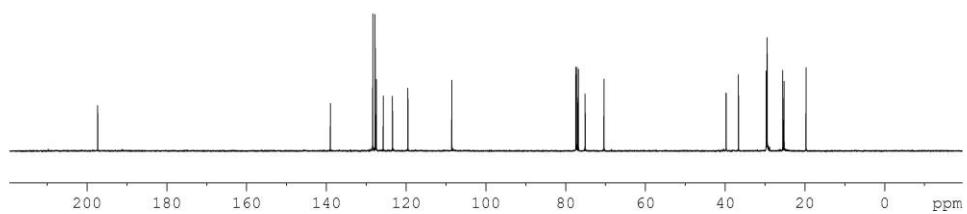
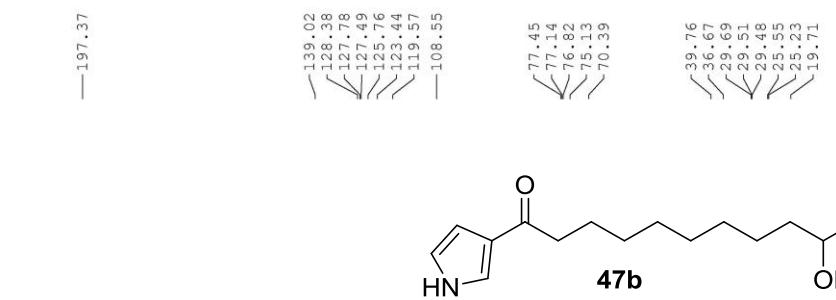
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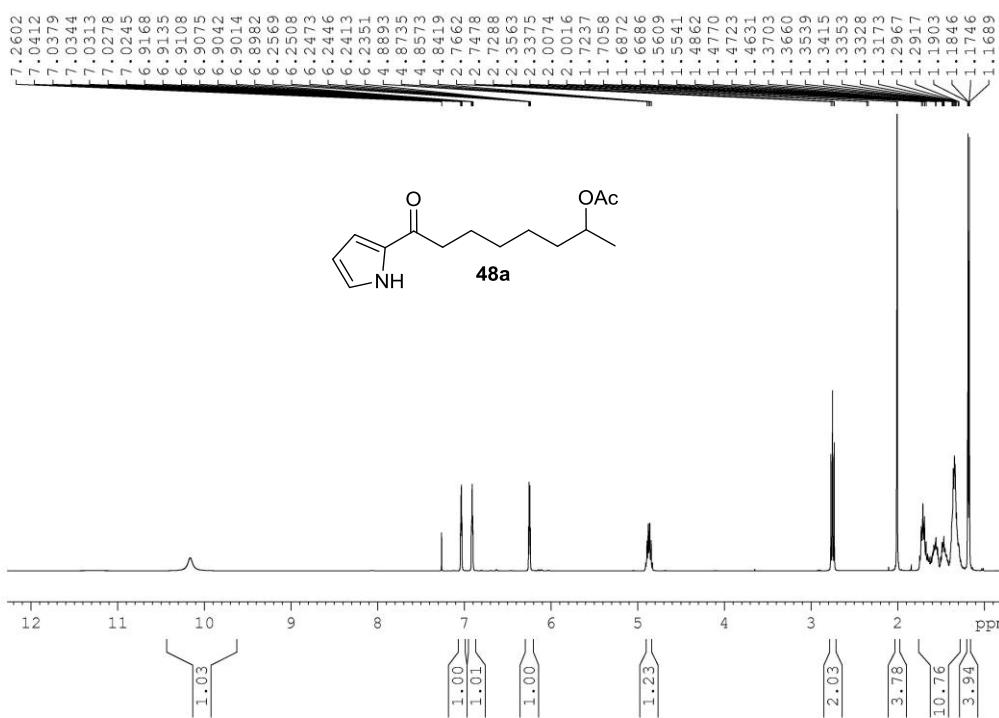
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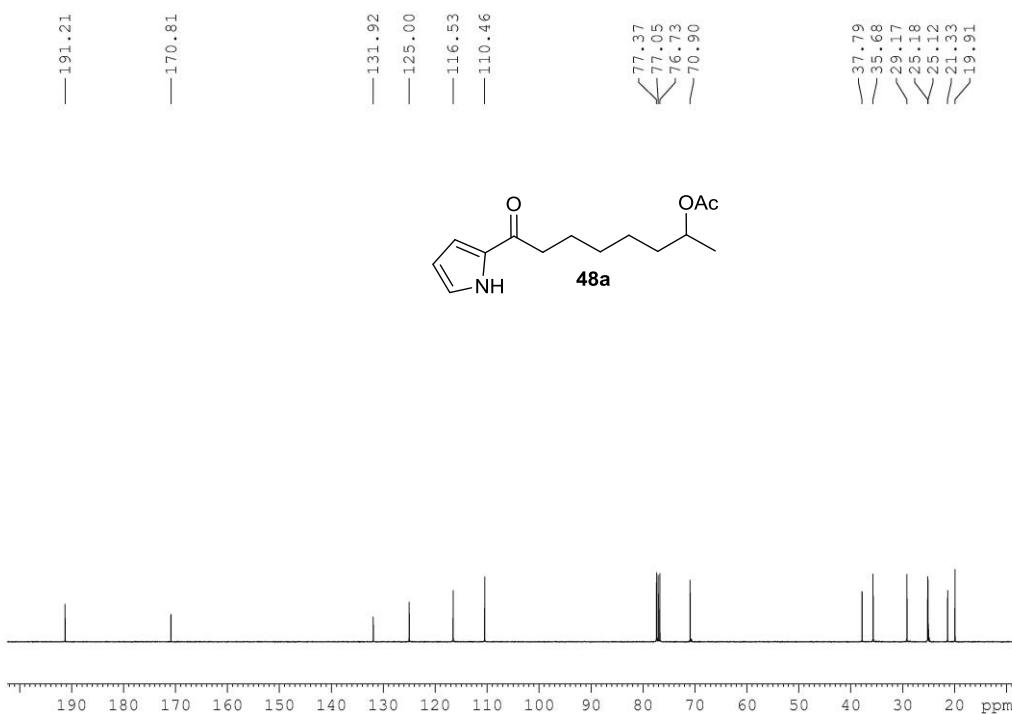
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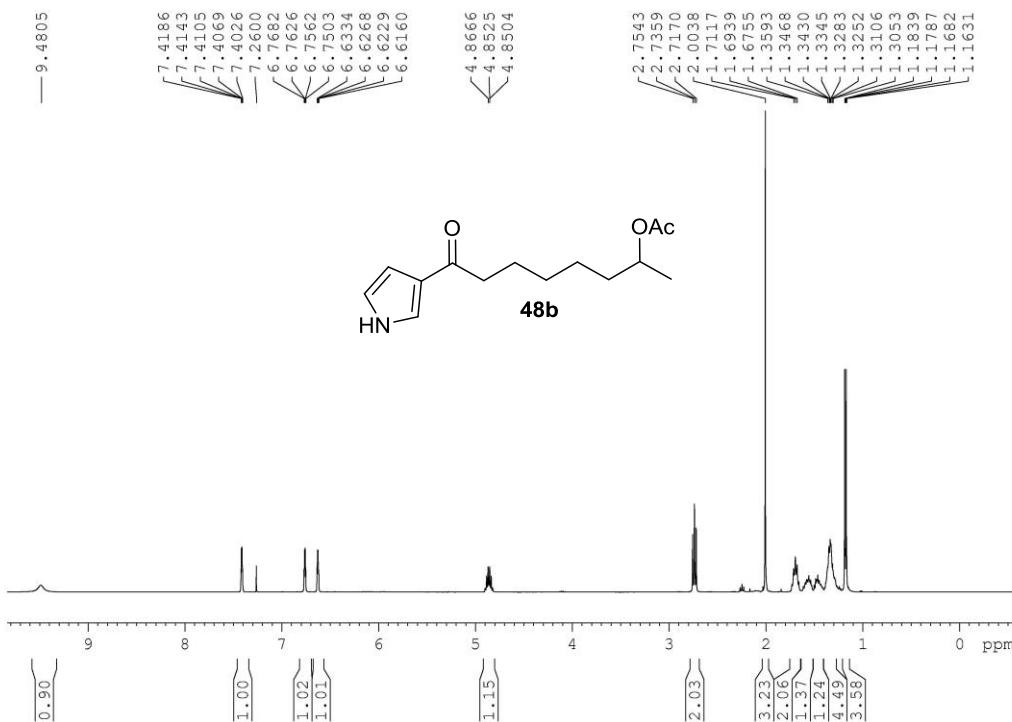
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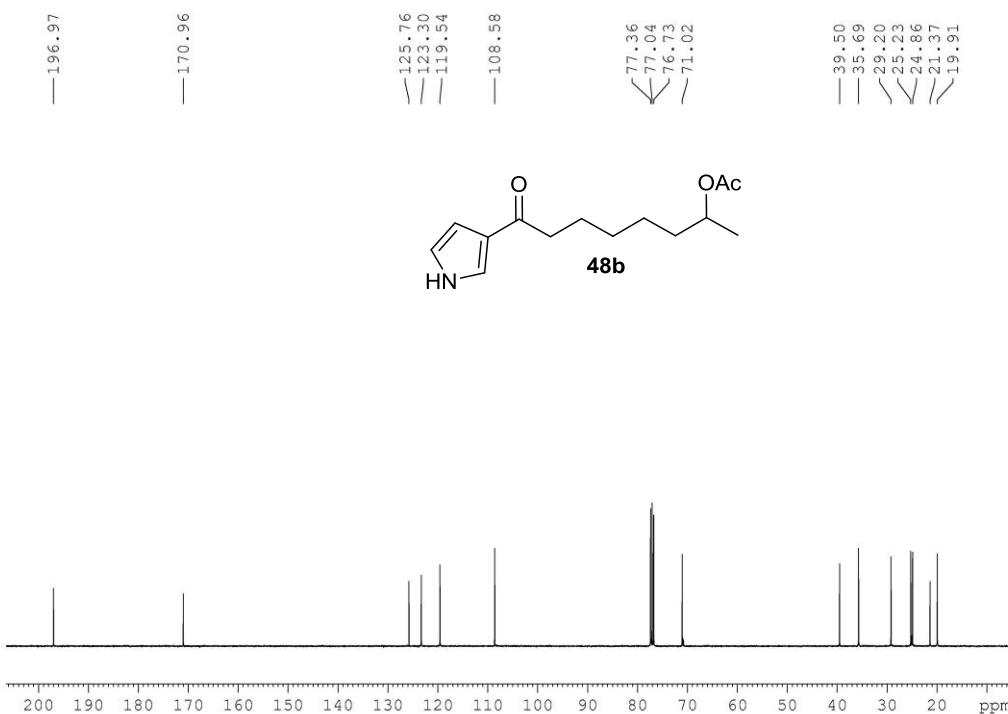
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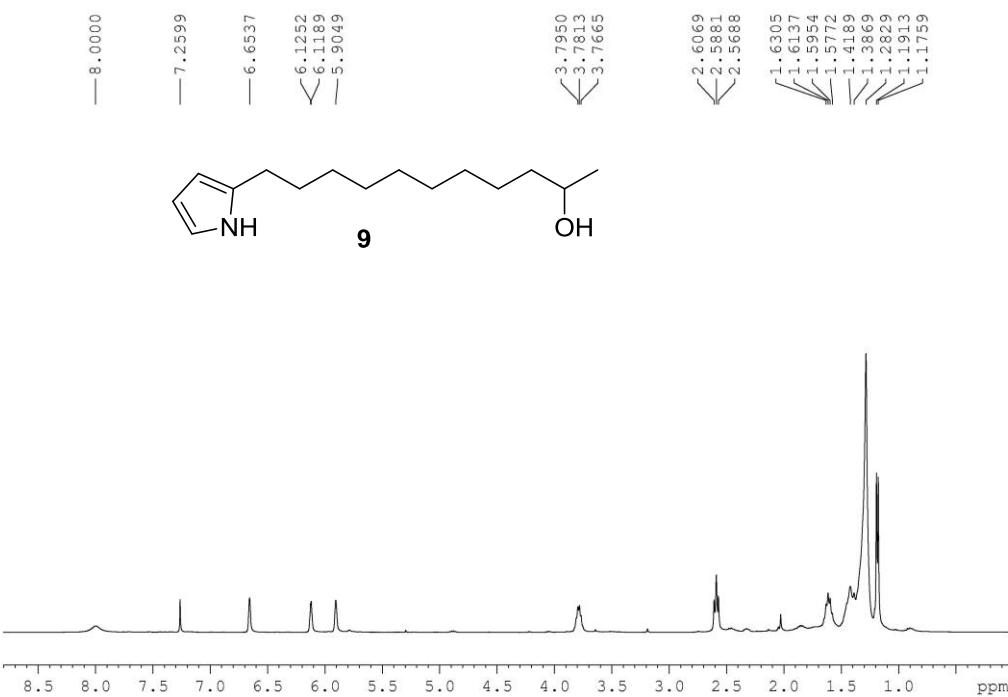
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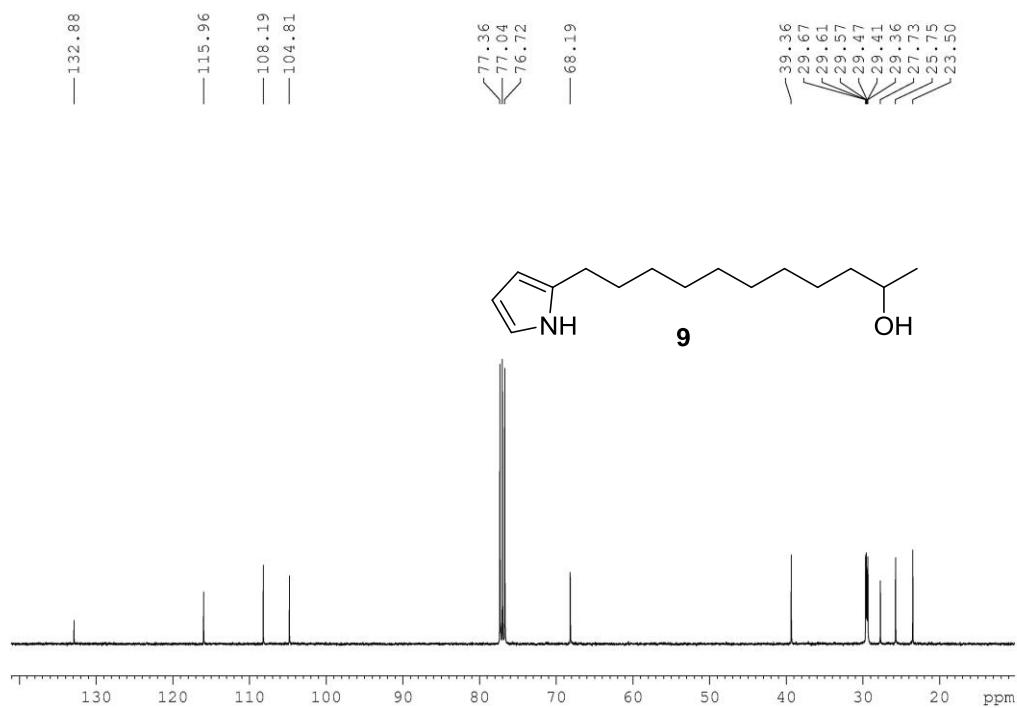
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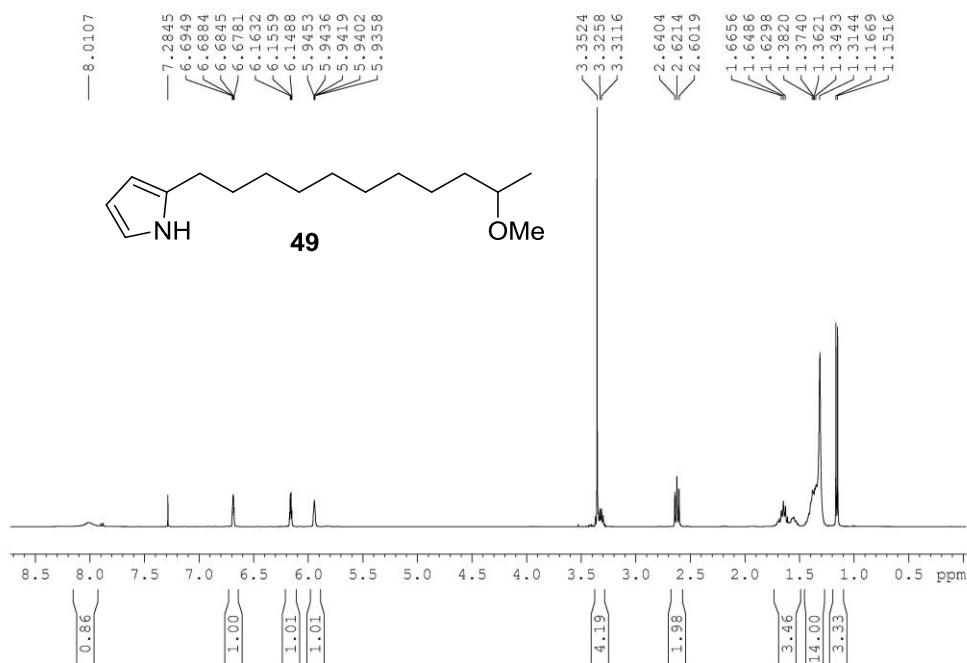
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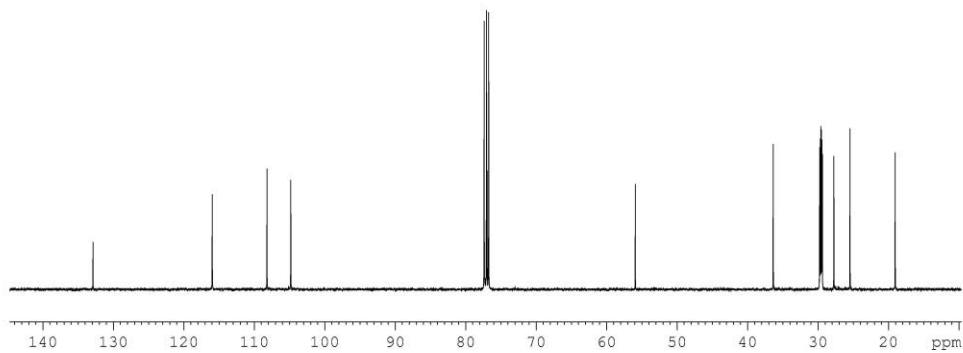
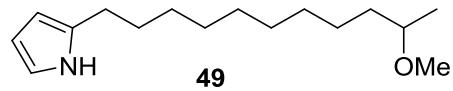
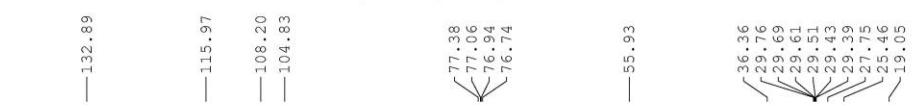
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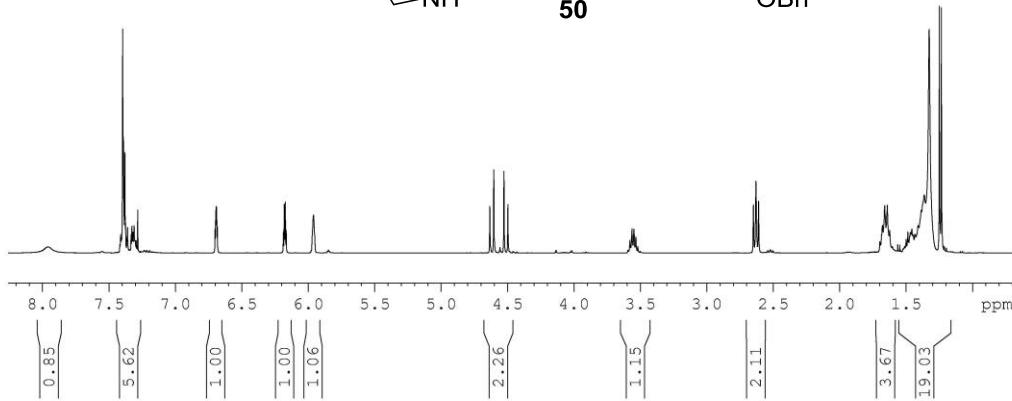
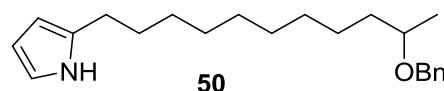
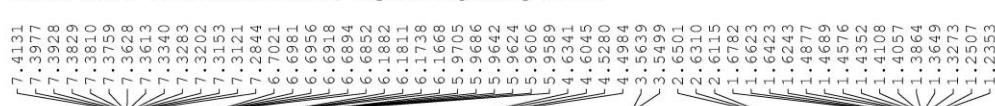
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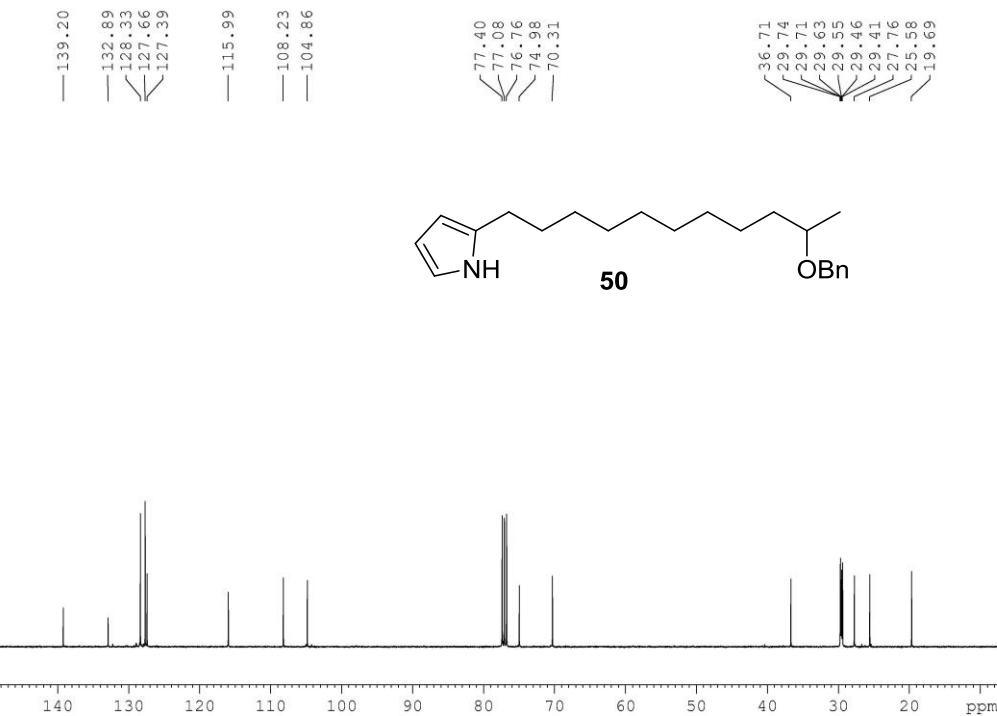
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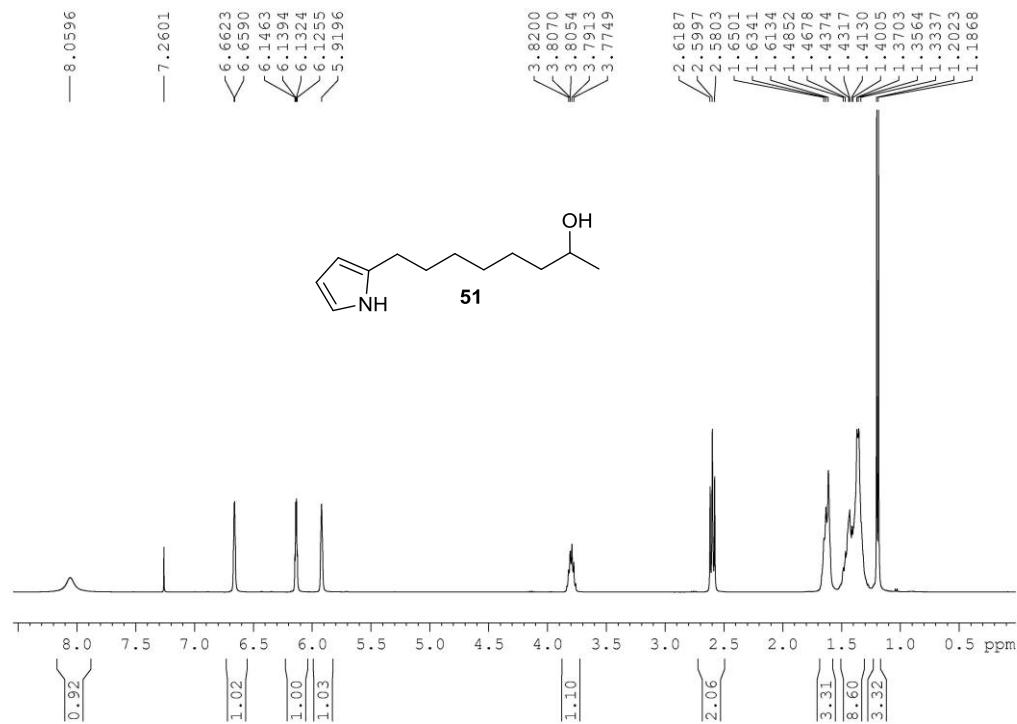
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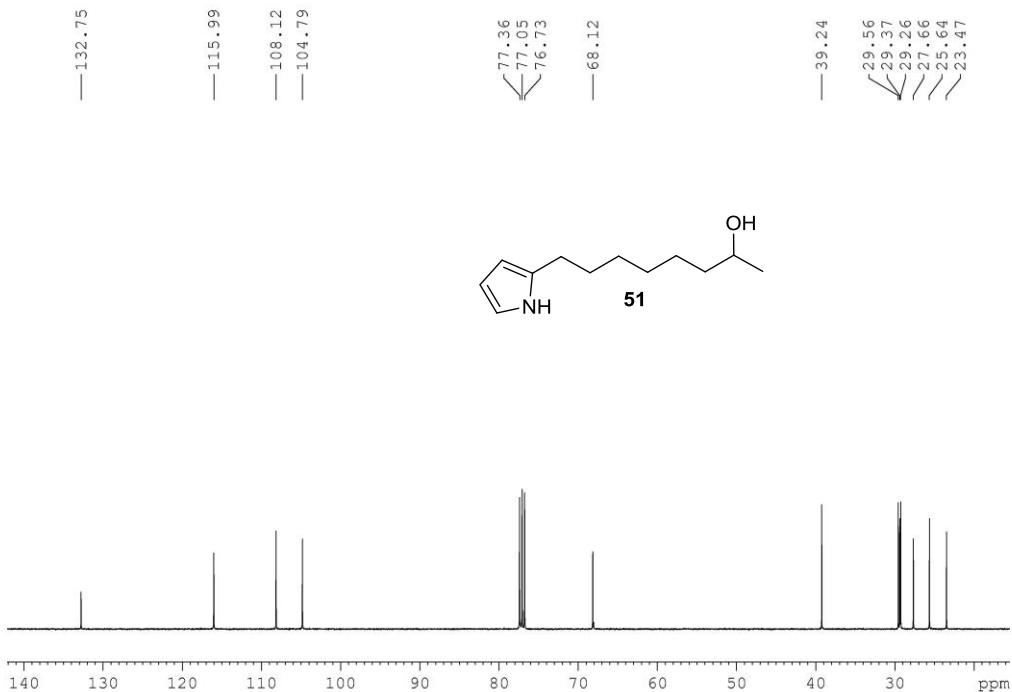
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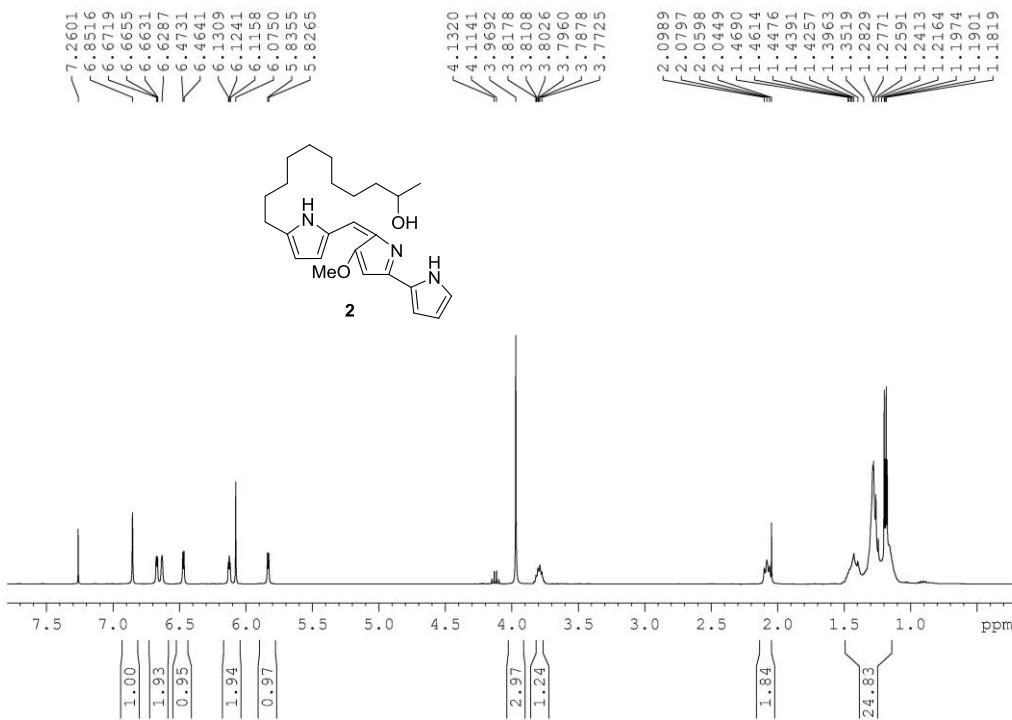
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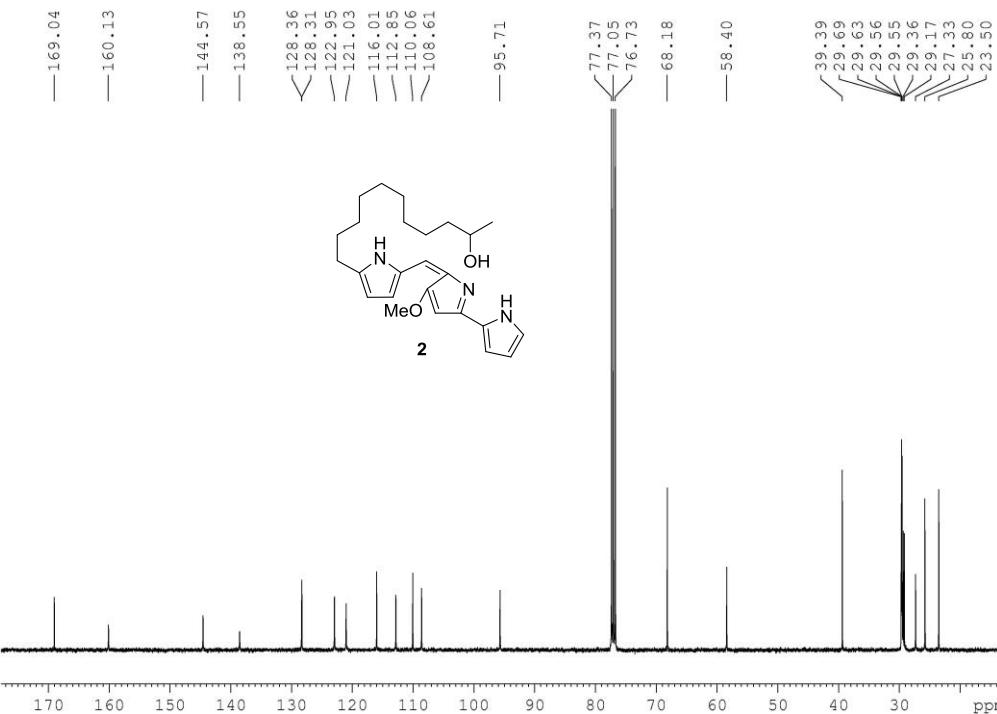
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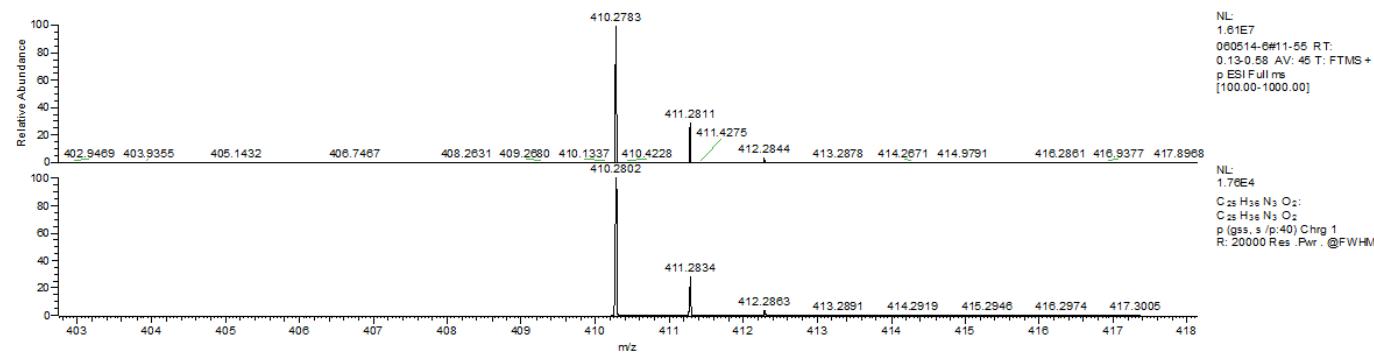
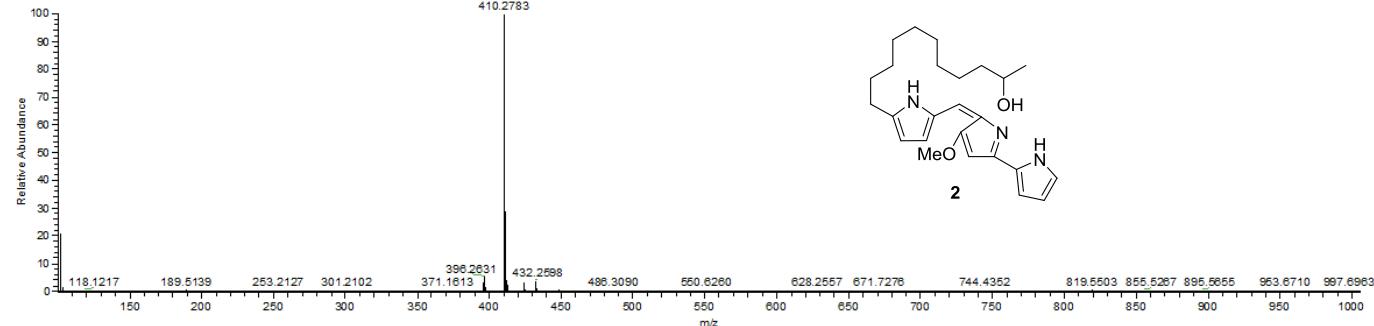
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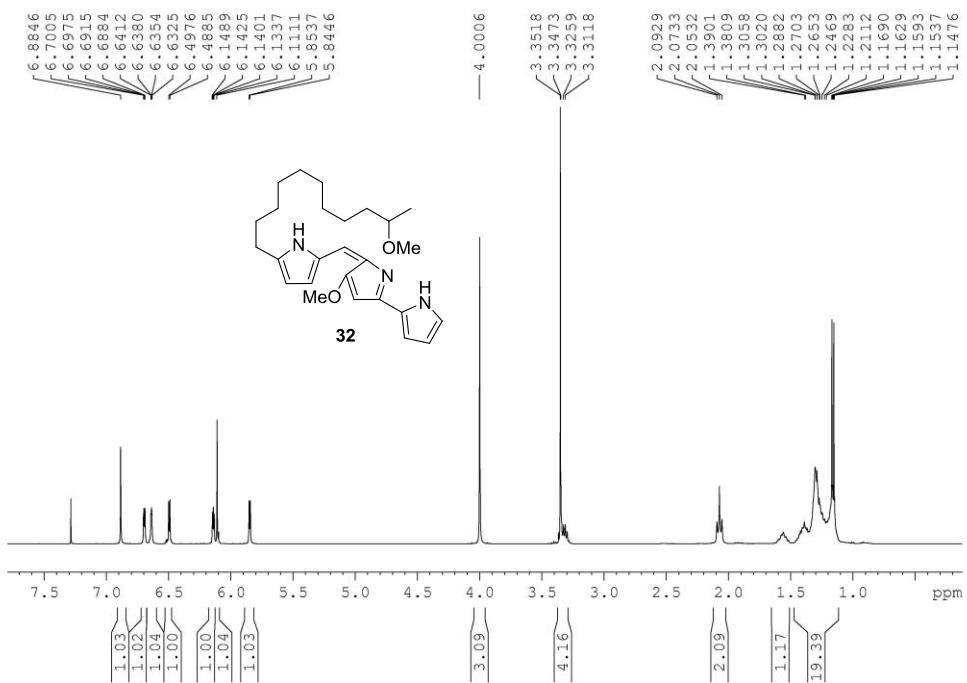
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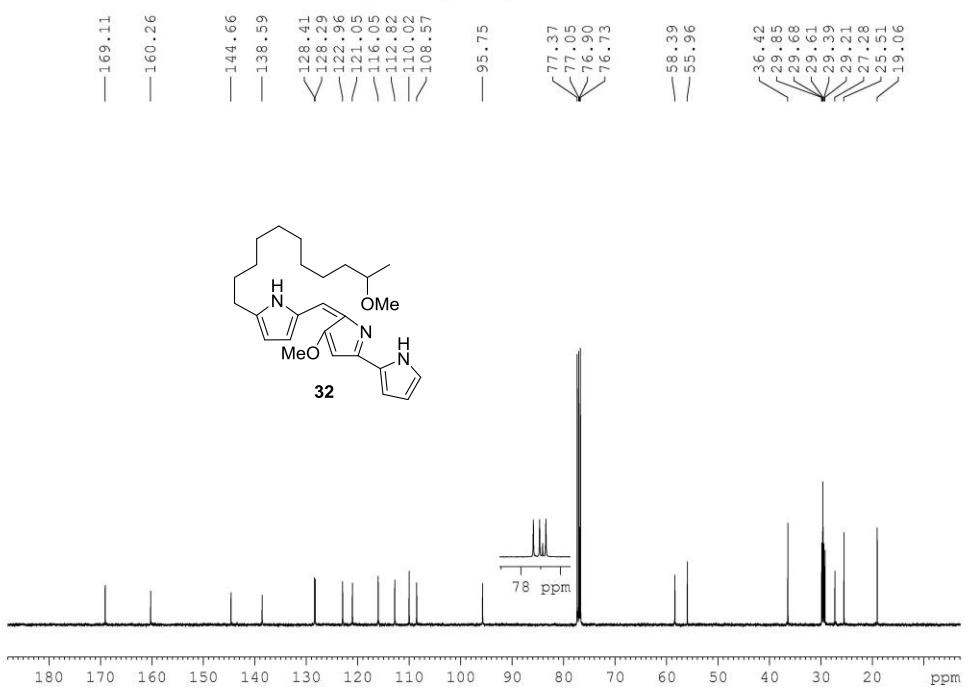
060514-6 #11-55 RT: 0.13-0.58 AV: 45 NL: 1.61E7  
T: FTMS + p ESI Full ms [100.00-1000.00]

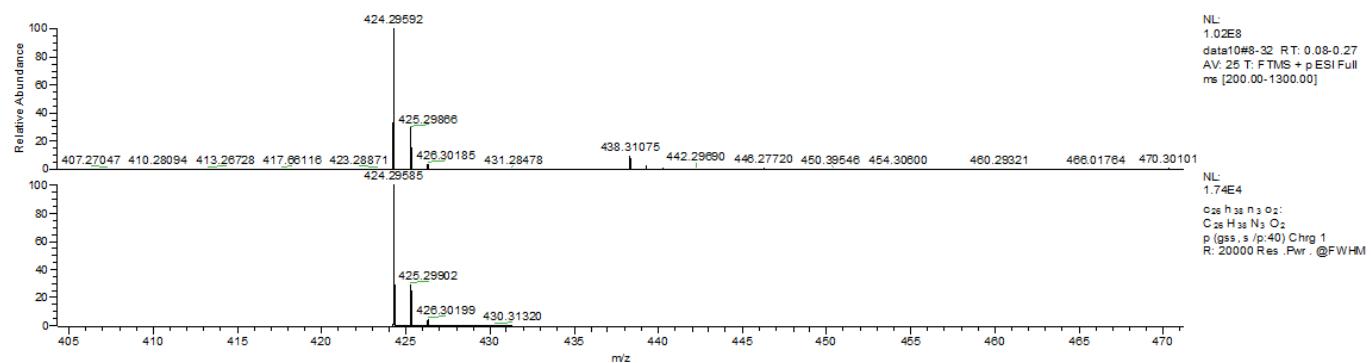
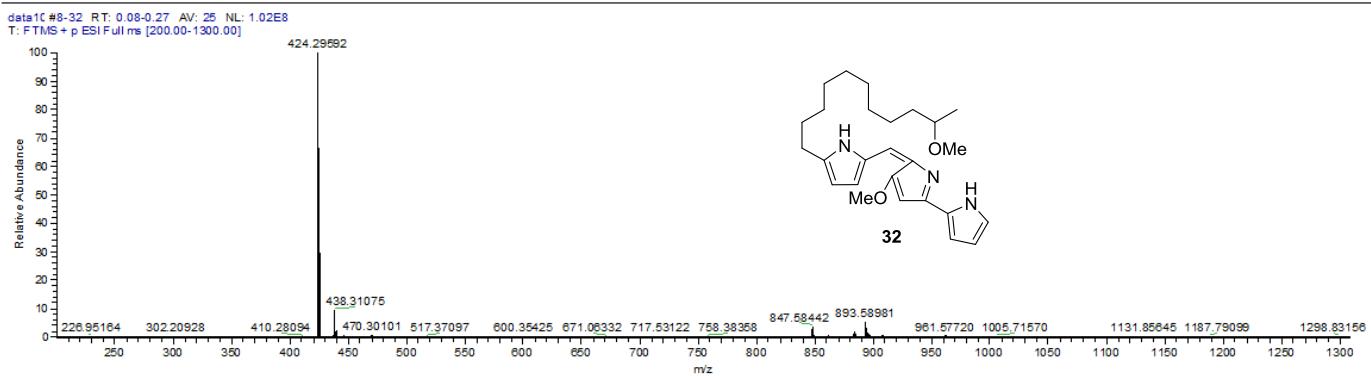


PROTON CDC13 {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 5

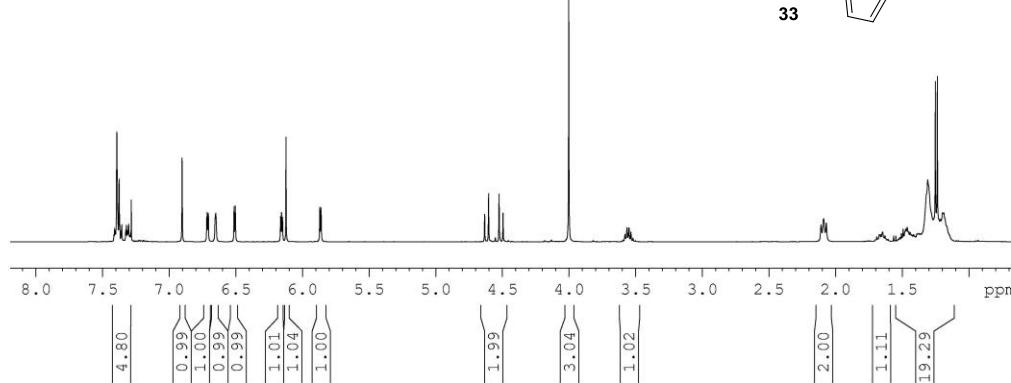
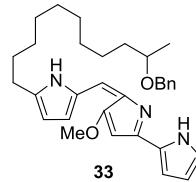
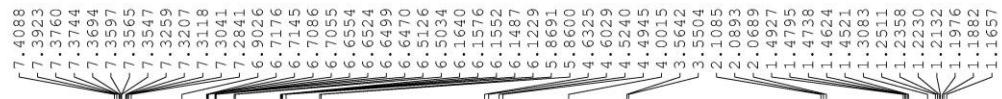


C13CPD CDC13 {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 5

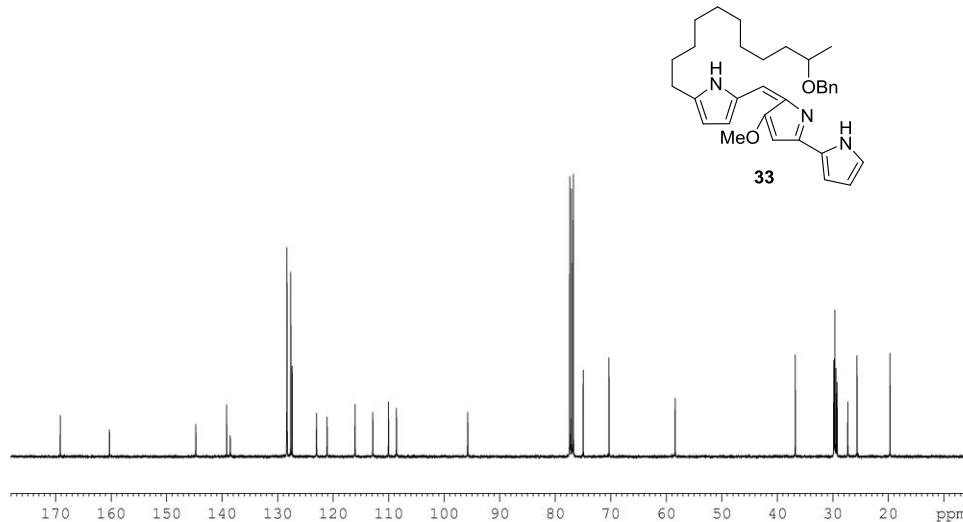
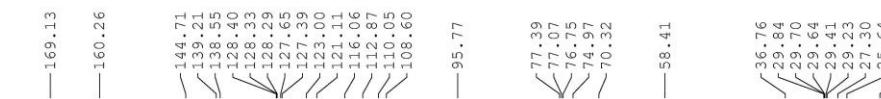




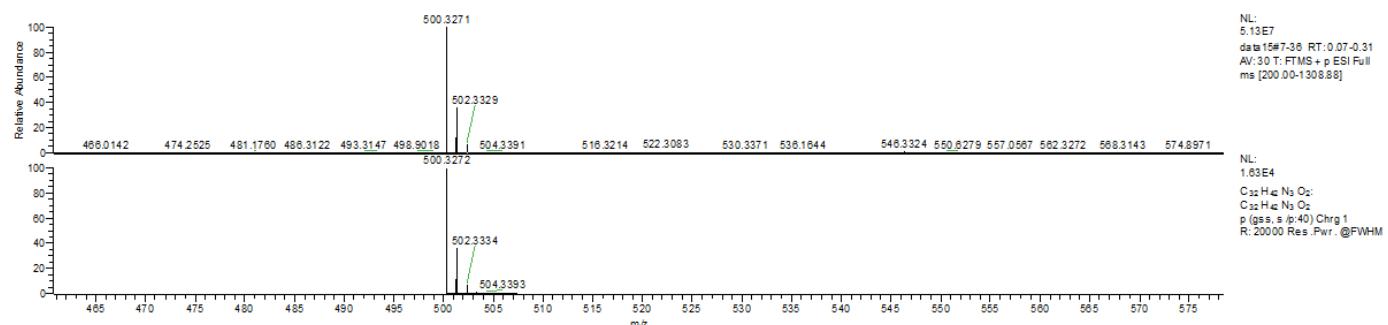
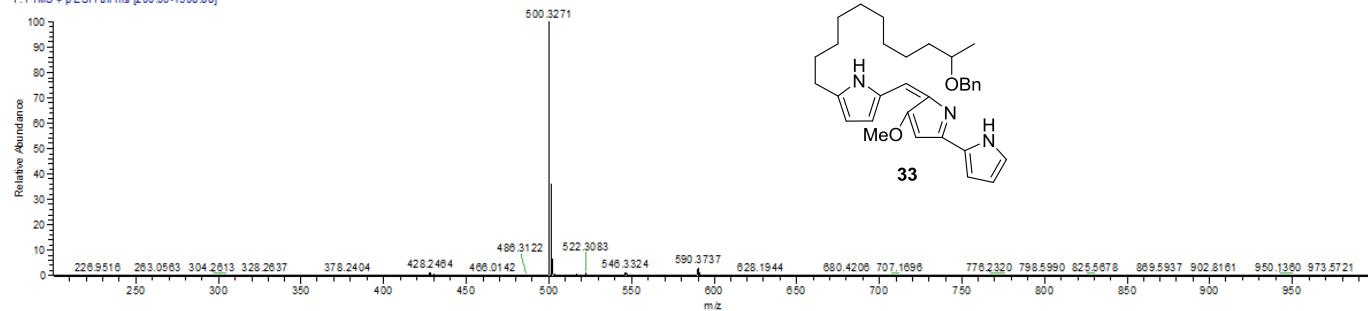
PROTON CDC13 {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 14



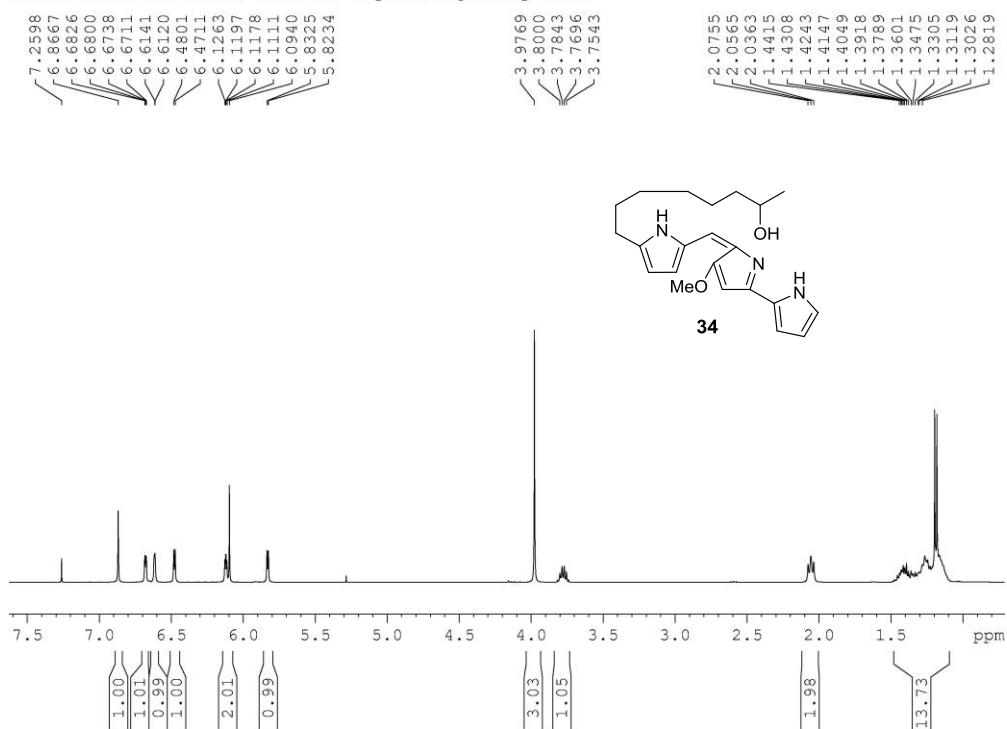
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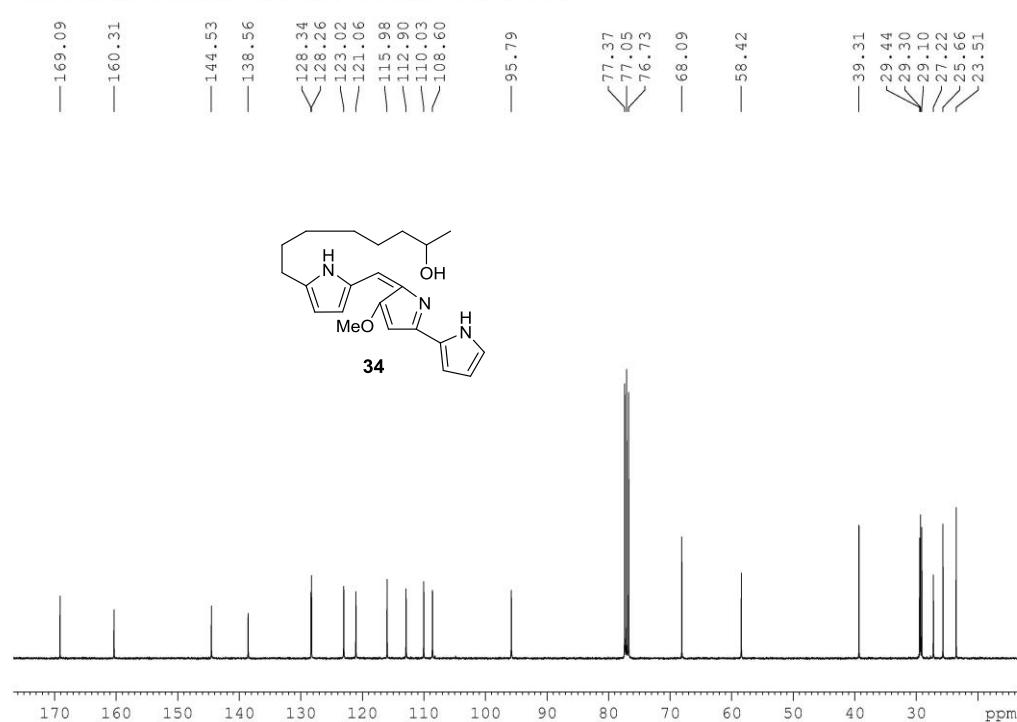
data1#7-38 RT: 0.07-0.31 AV: 30 NL: 5.13E7  
T: FTMS + p ESI Full ms [200.00-1308.88]



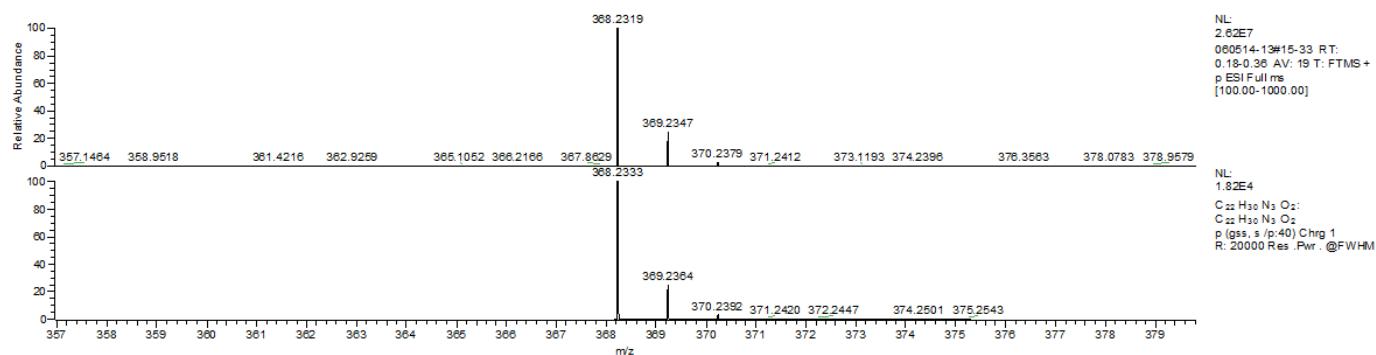
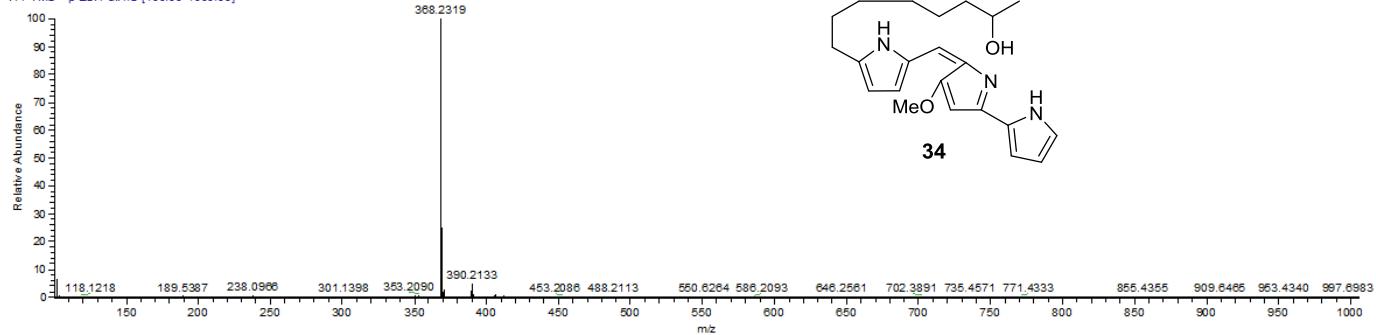
PROTON CDCl<sub>3</sub> {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 2



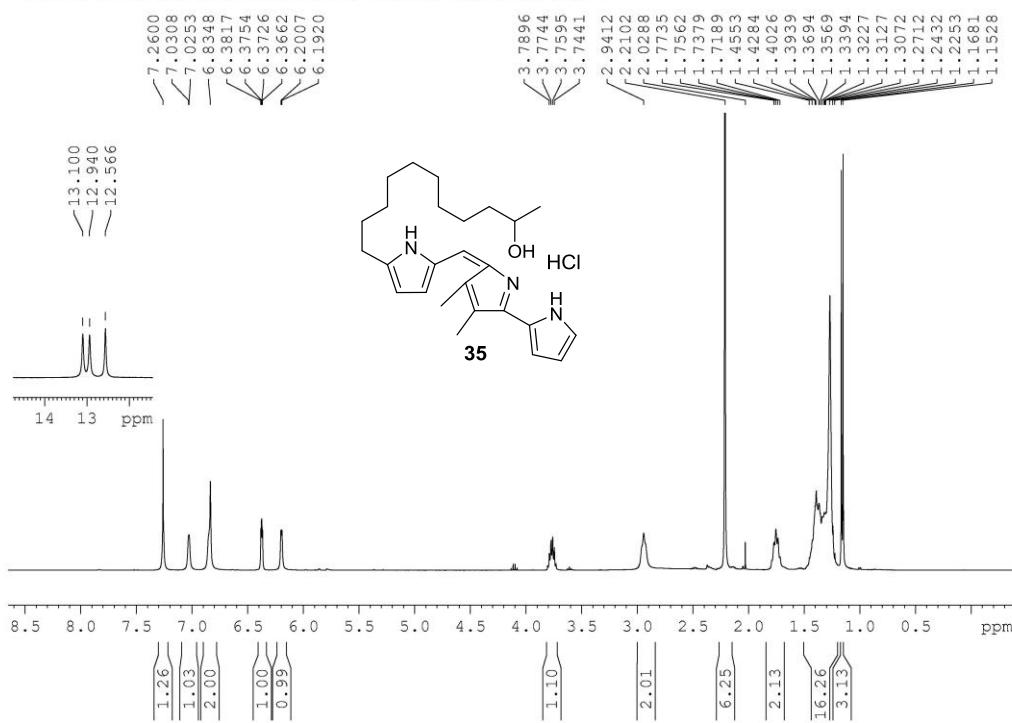
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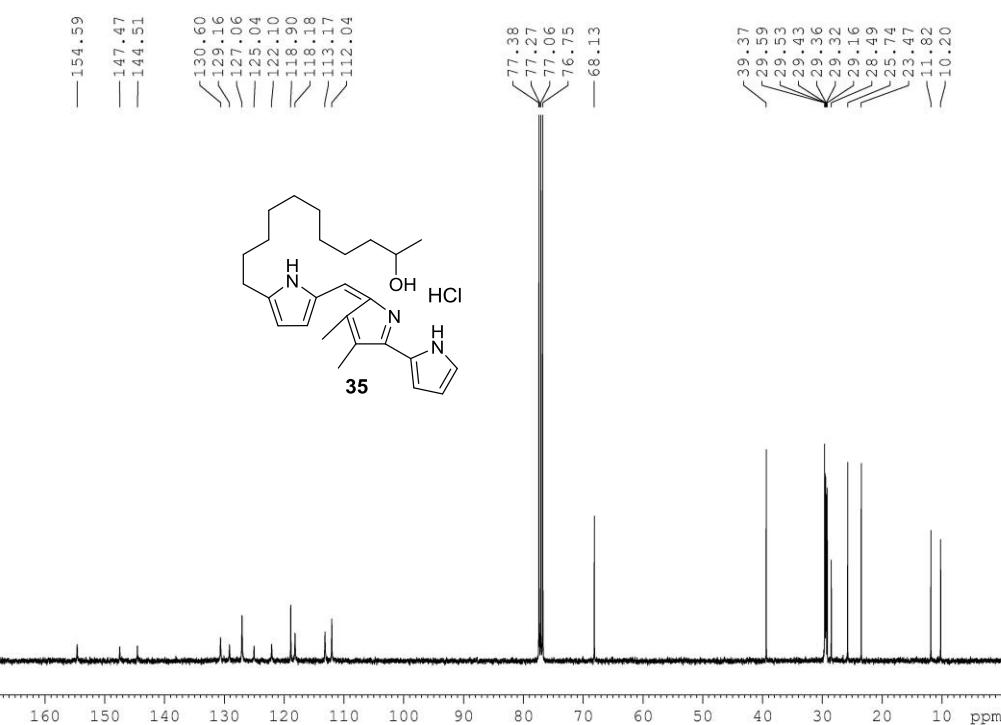
060514-13 #15-33 RT: 0.18-0.36 AV: 19 NL: 2.62E7  
T: FTMS + p ESI Full ms [100.00-1000.00]



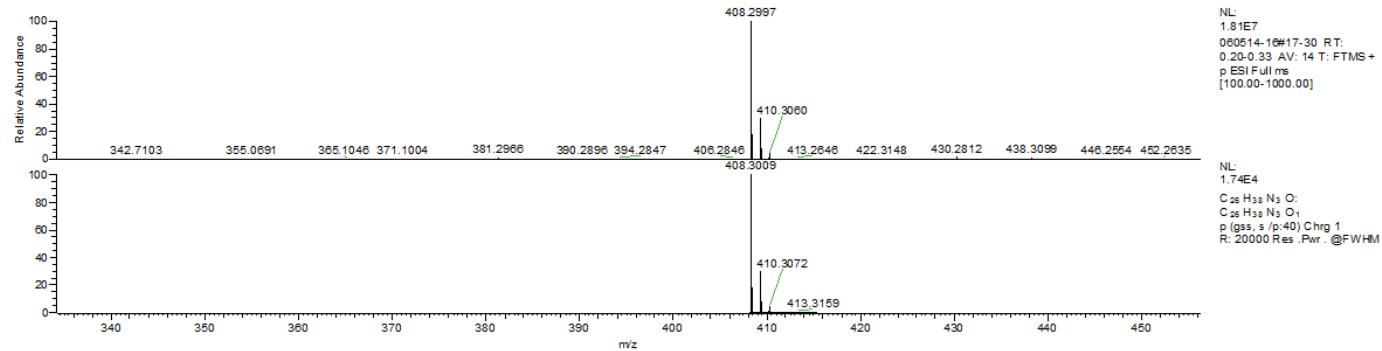
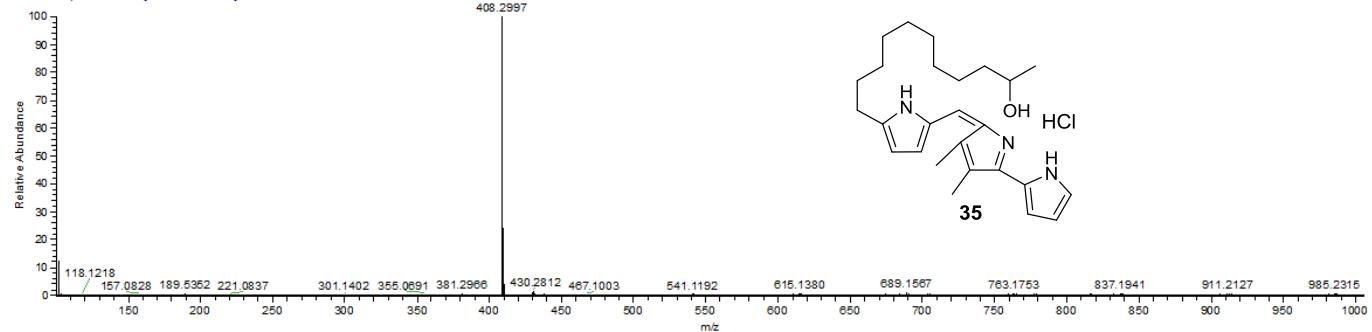
PROTON CDCl<sub>3</sub> {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 1



C13CPD CDCl<sub>3</sub> {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 1



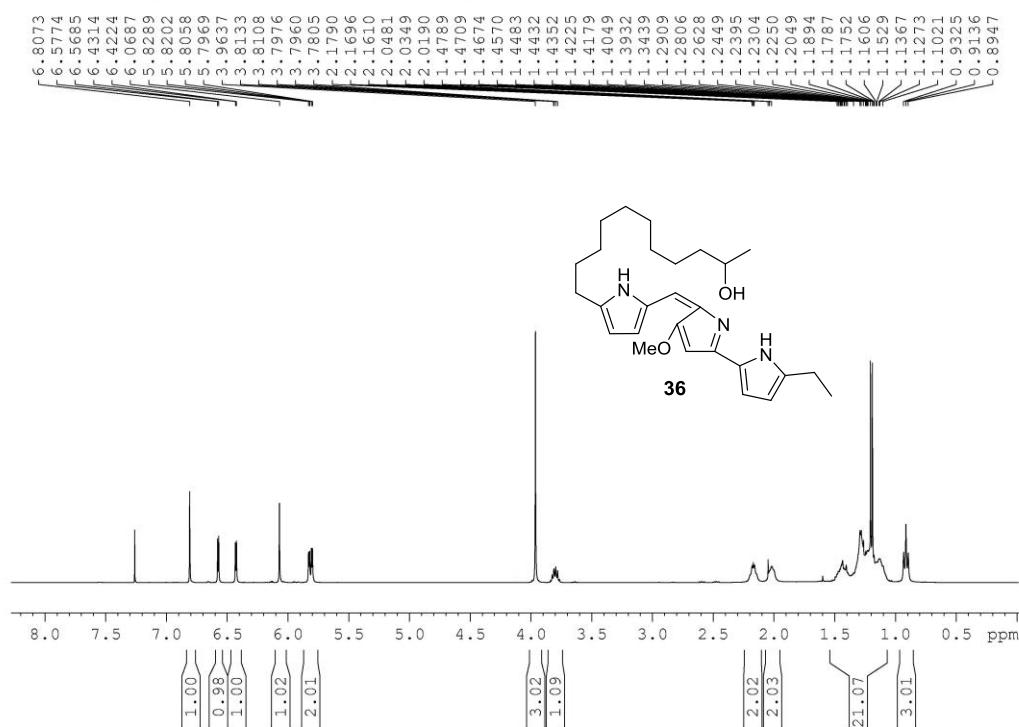
060514-16 #17-30 RT: 0.20-0.33 AV: 14 NL: 1.81E7  
T: FTMS + p ESI Full ms [100.00-1000.00]



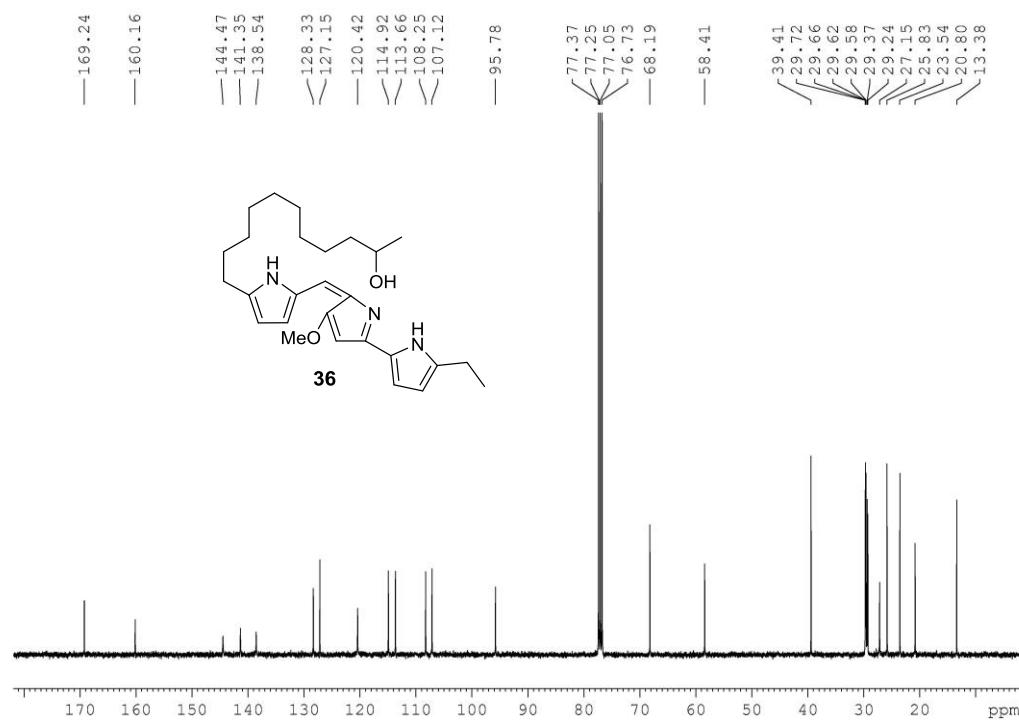
NL:  
1.81E7  
060514-16#17-30 RT:  
0.20-0.33 AV: 14 T: FTMS +  
p ESI Full ms  
[100.00-1000.00]

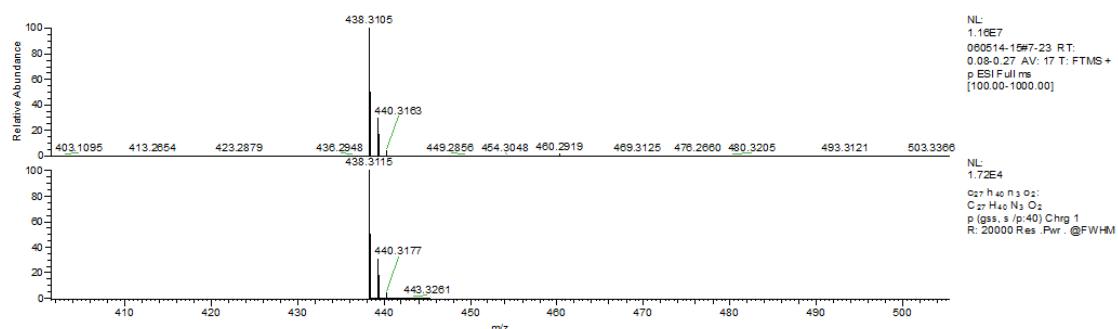
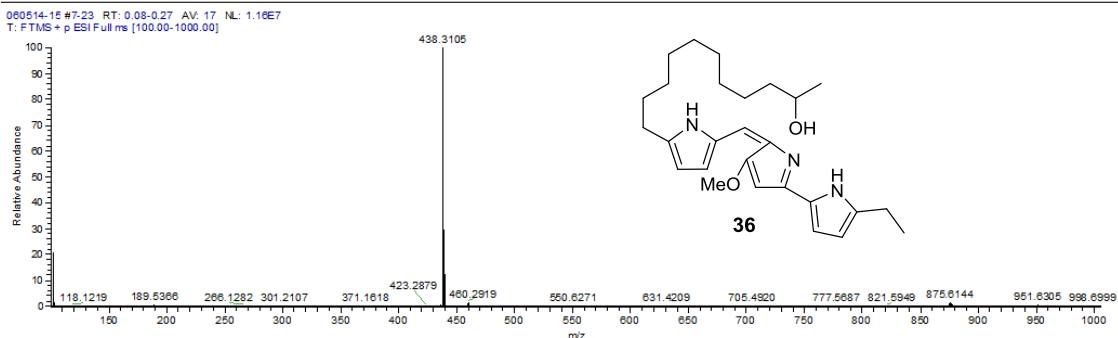
NL:  
1.74E4  
C<sub>26</sub>H<sub>33</sub>N<sub>3</sub>O:  
C<sub>26</sub>H<sub>33</sub>N<sub>3</sub>O<sub>1</sub>  
p (gss, s/p/40) Chrg 1  
R: 20000 Res\_Pwr @FWHM

PROTON CDC13 {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 12

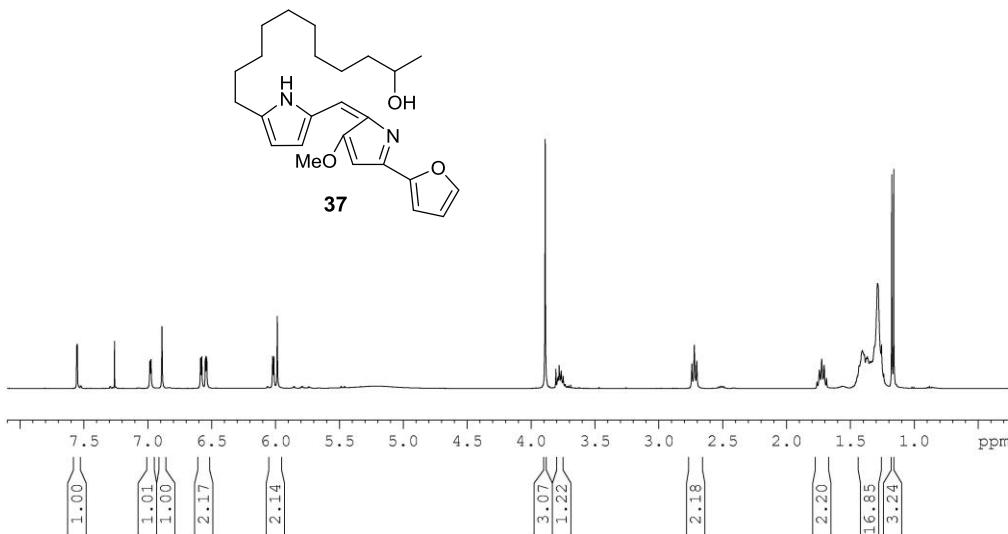


C13CPD CDC13 {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 19

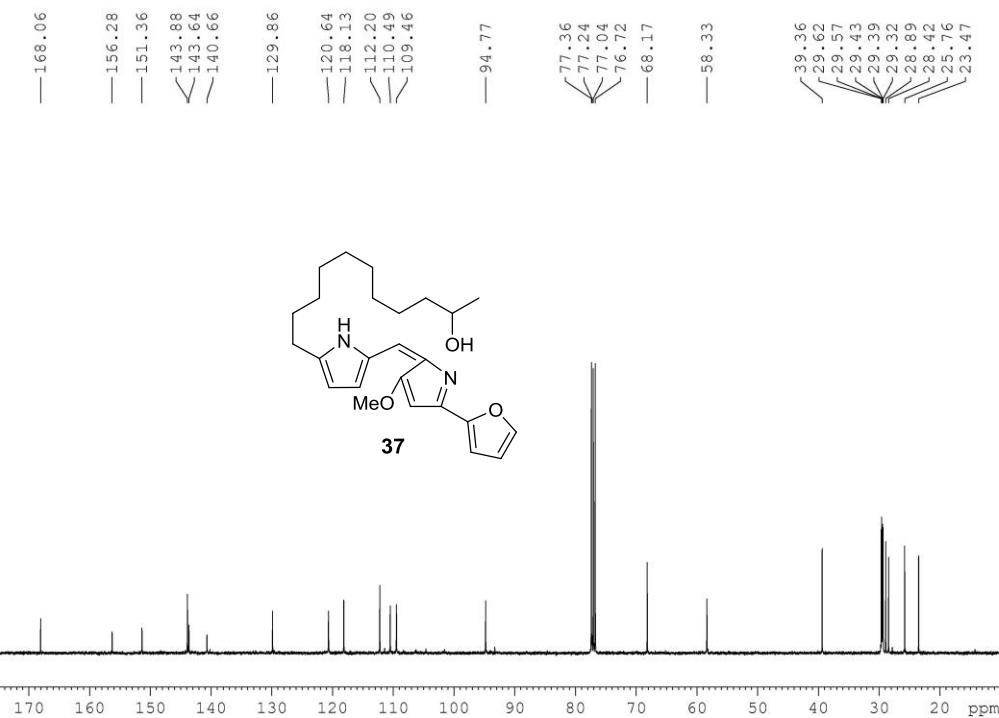




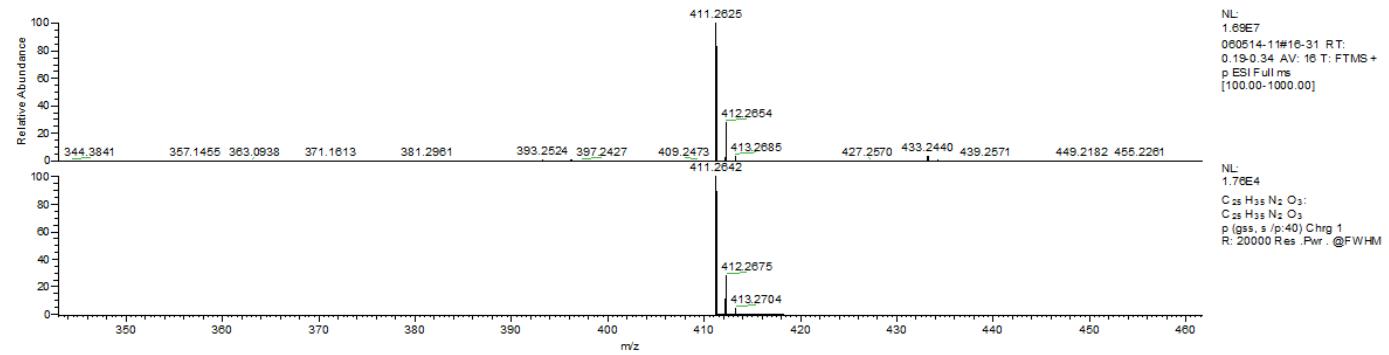
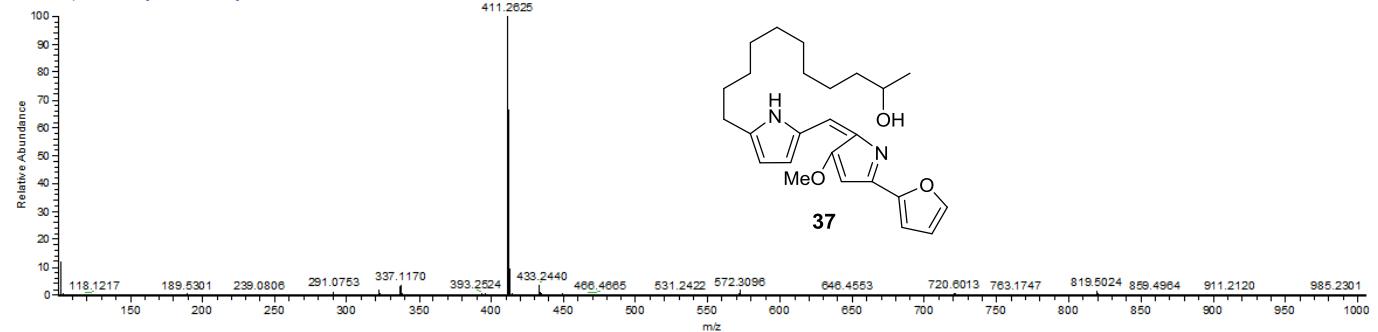
PROTON CDCl3 {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 3



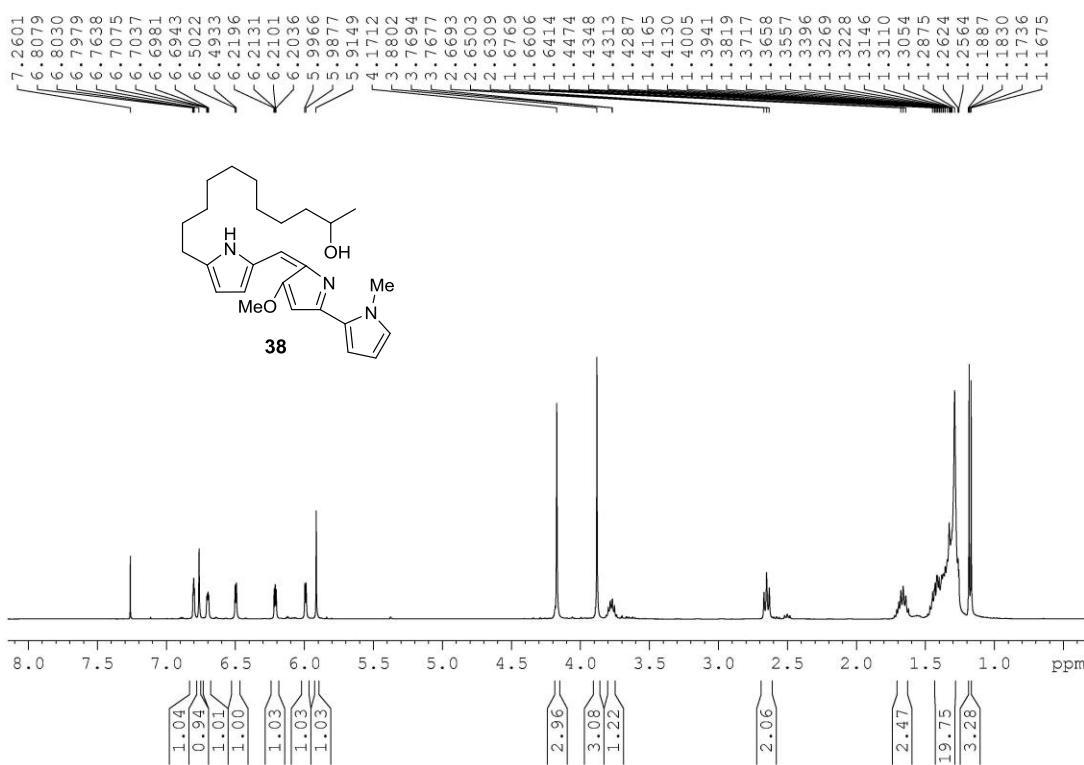
C13CPD CDCl<sub>3</sub> {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 8



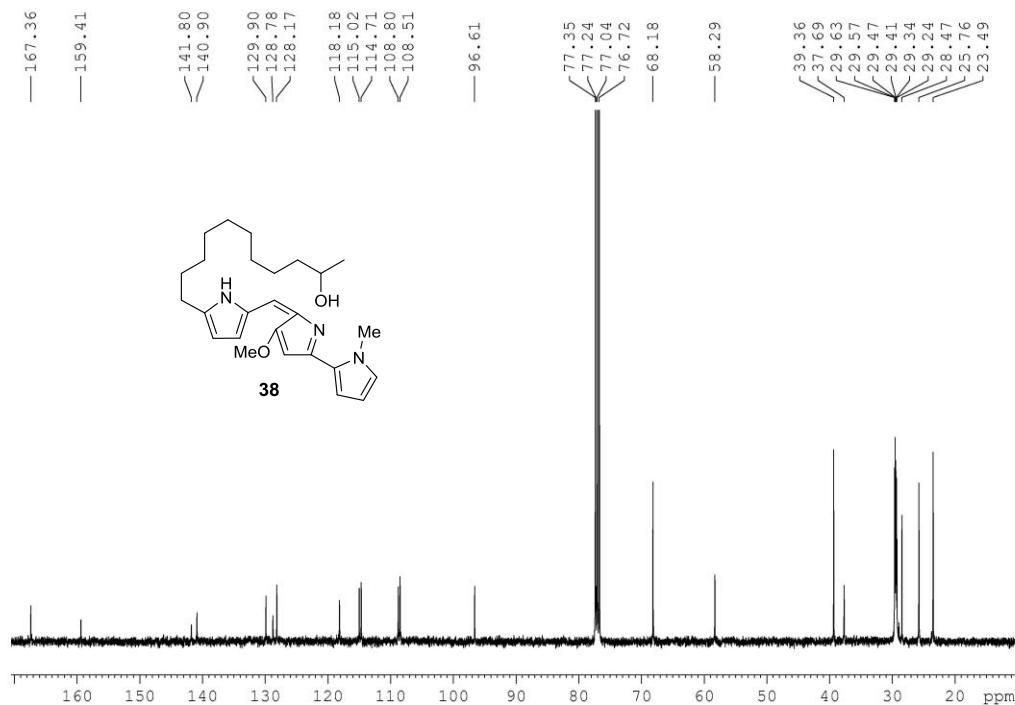
060514-11 #16-31 RT: 0.19-0.34 AV: 16 NL: 1.69E7  
T: FTMS + p ESI Full ms [100.00-1000.00]



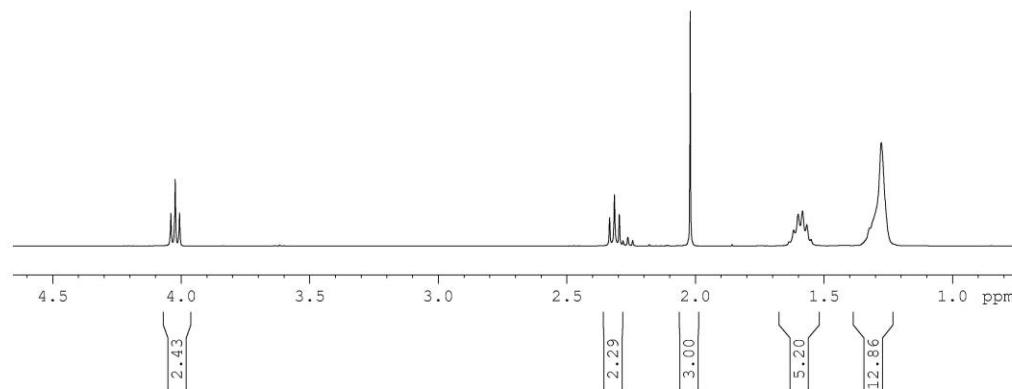
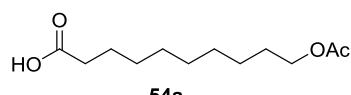
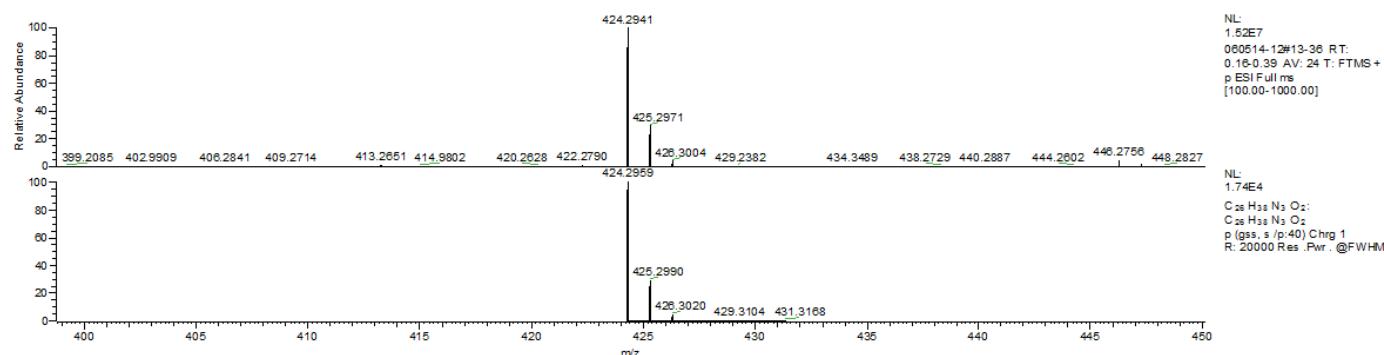
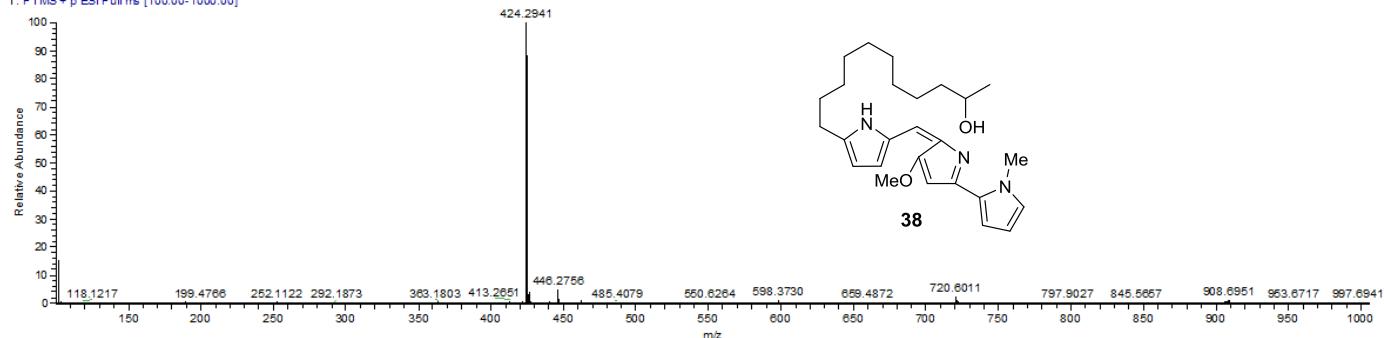
PROTON CDCl<sub>3</sub> {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 7

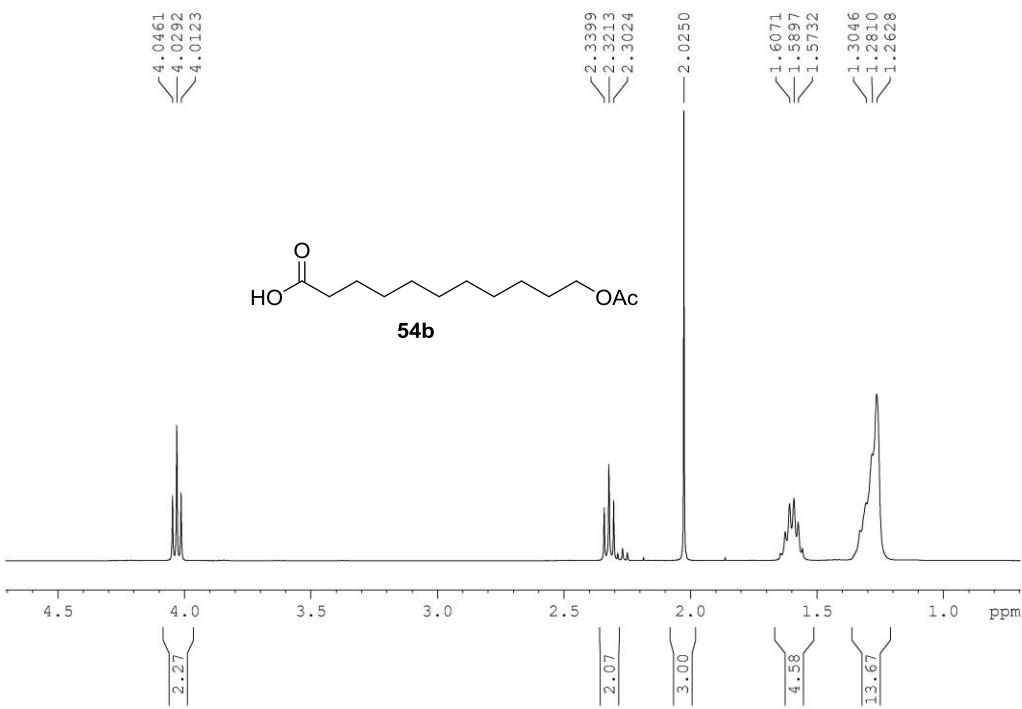


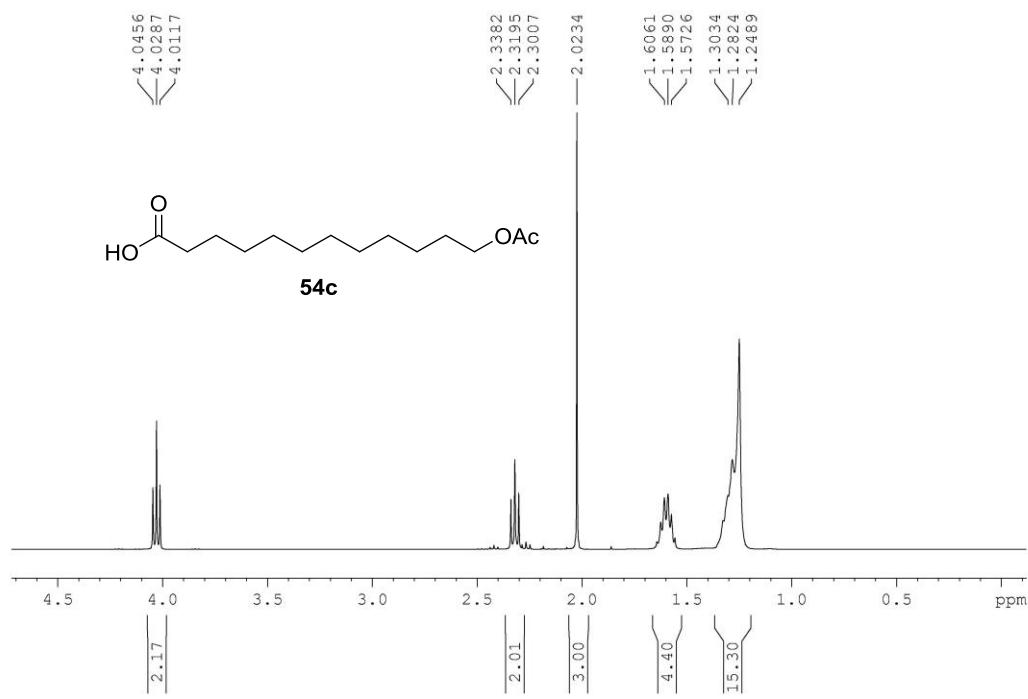
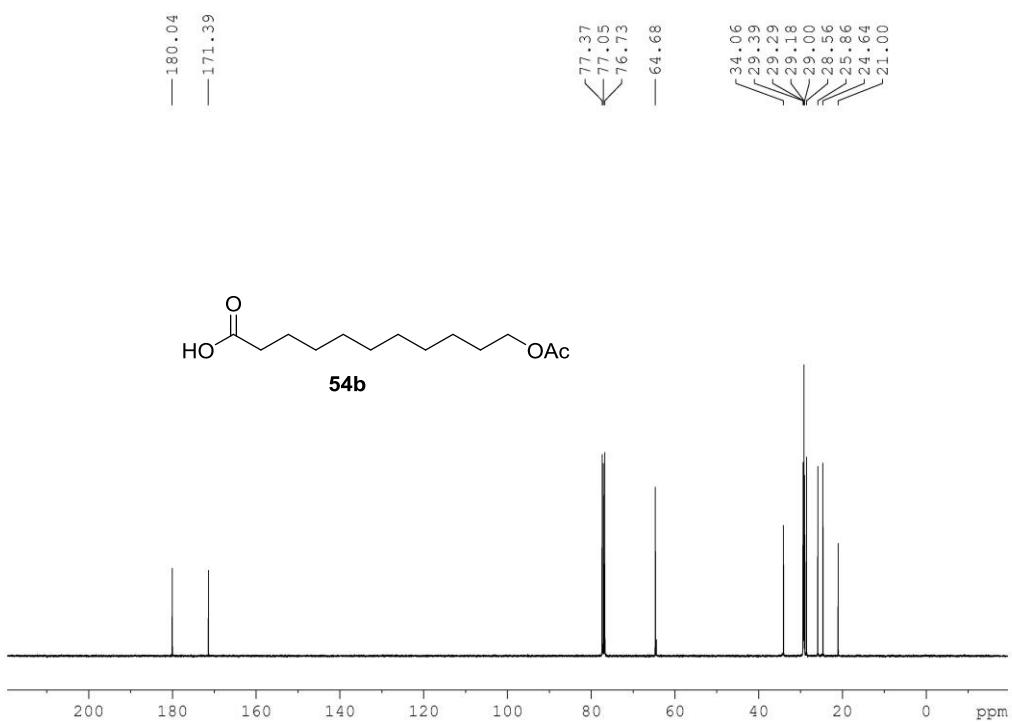
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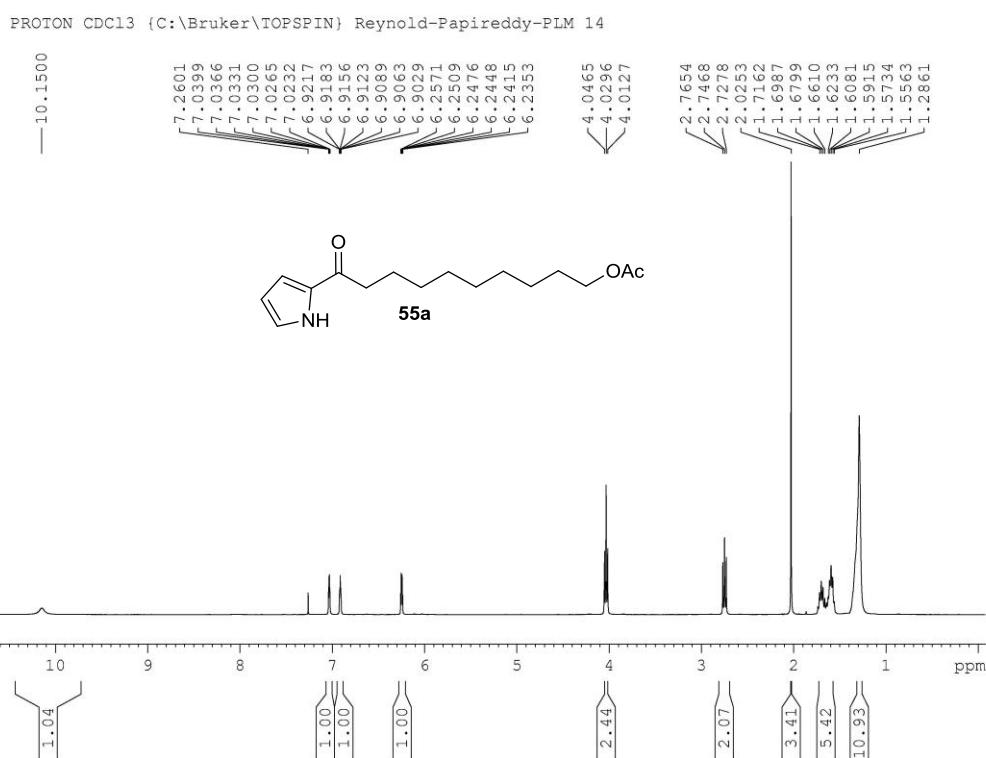
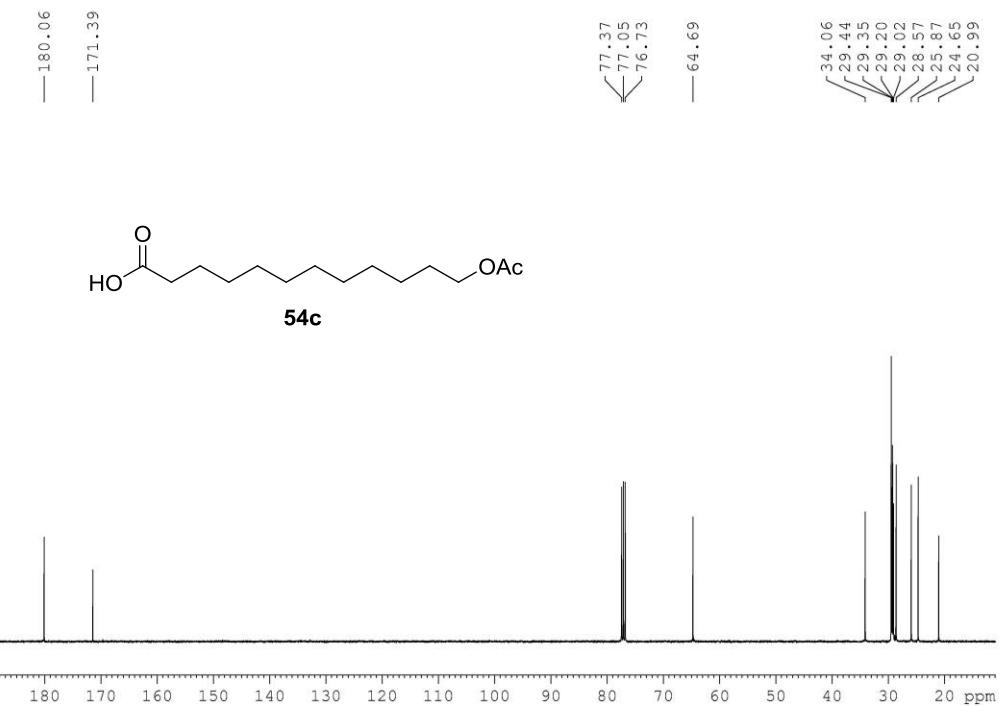


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T: FTMS + p ESI Full ms [100.00-1000.00]

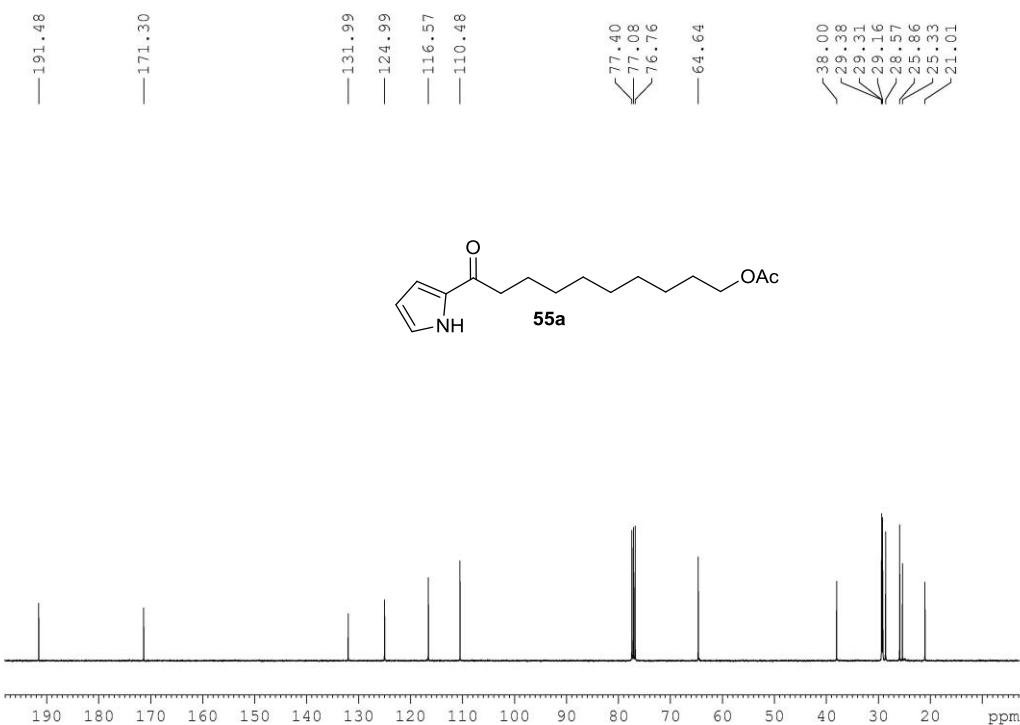




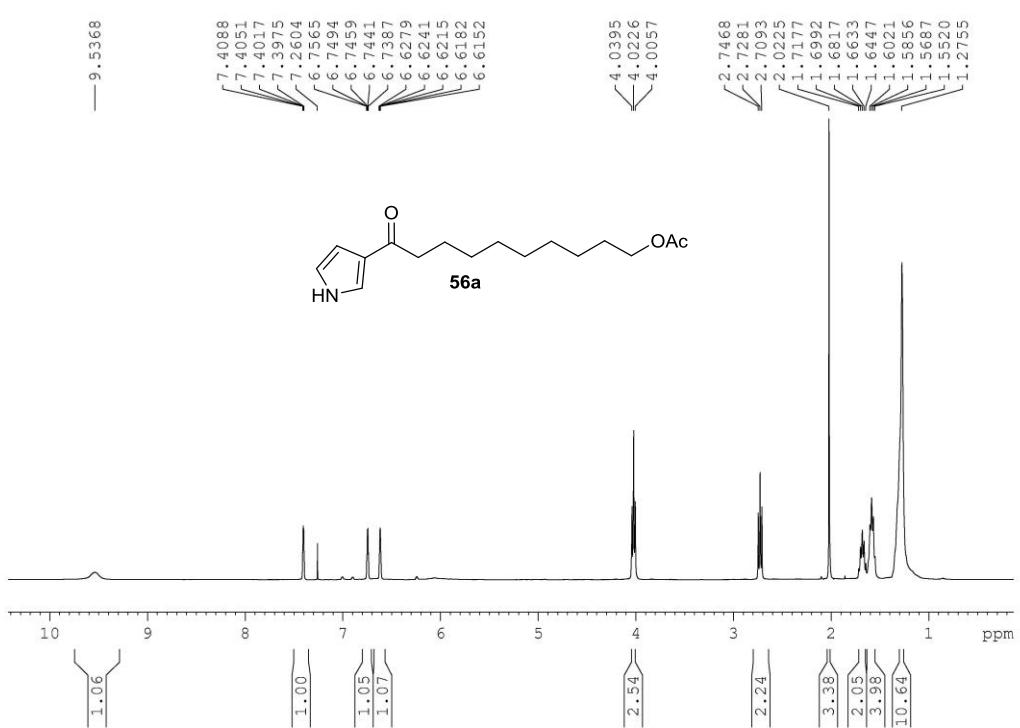




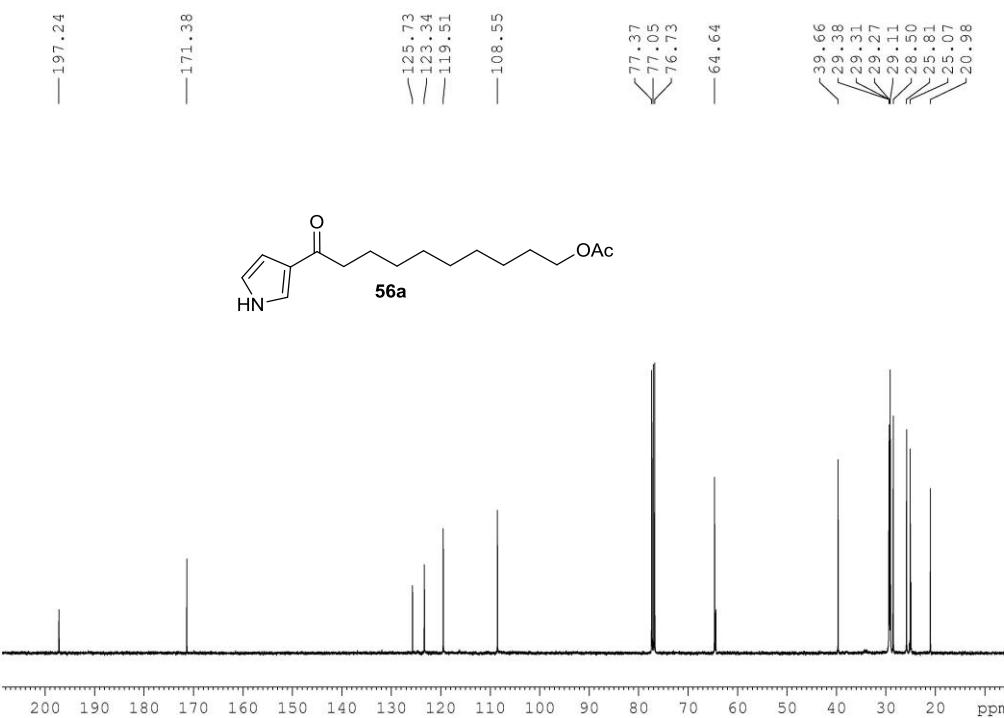
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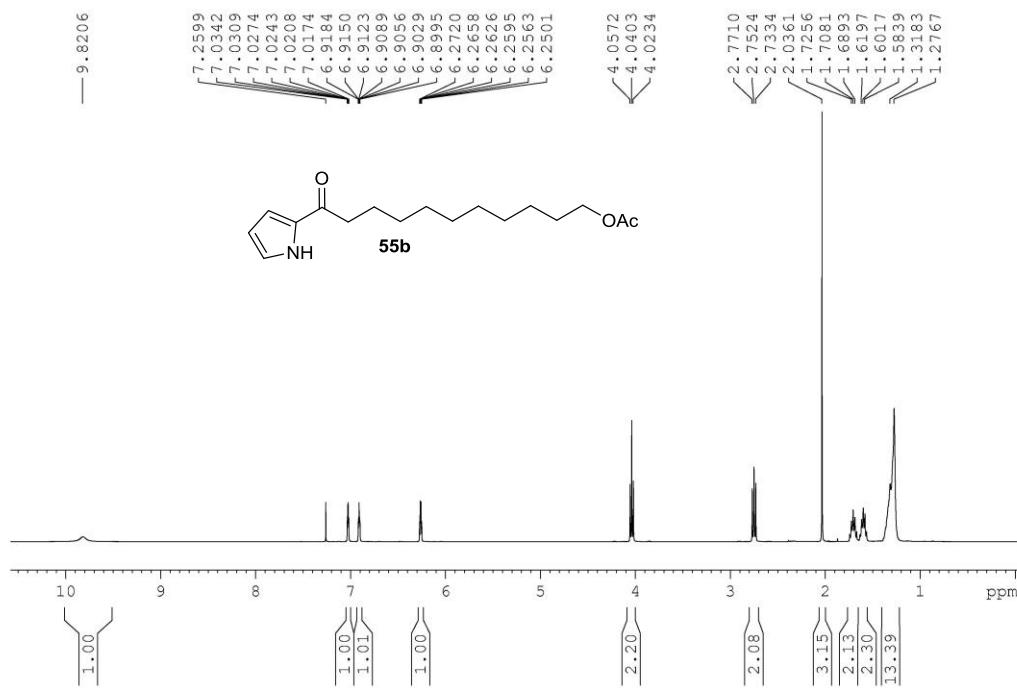
PROTON CDC13 {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 15



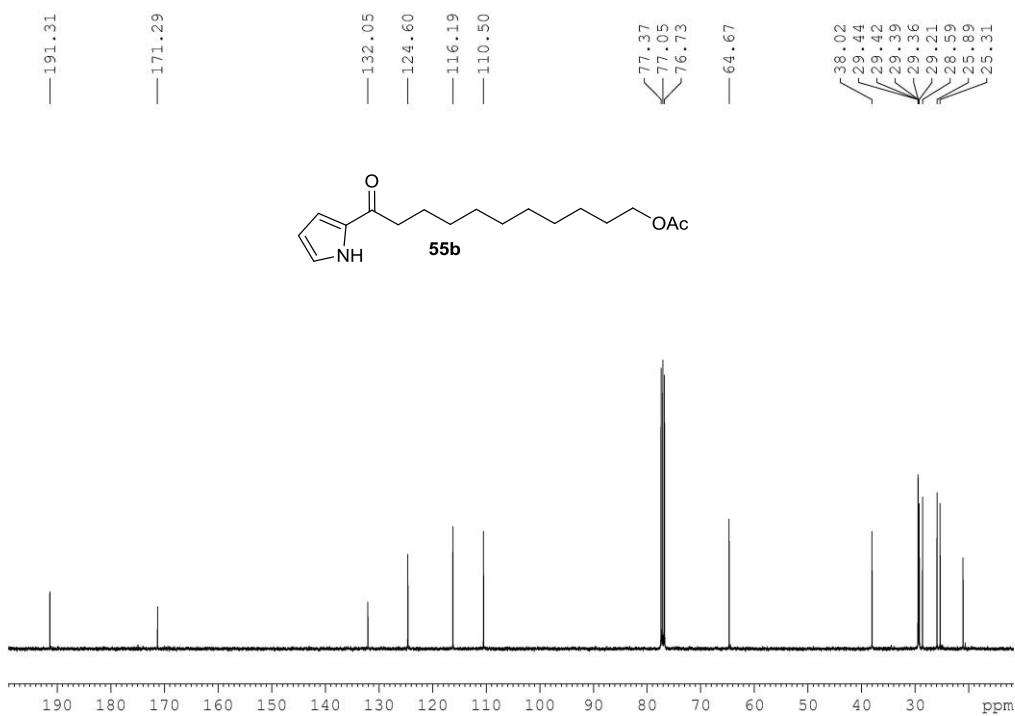
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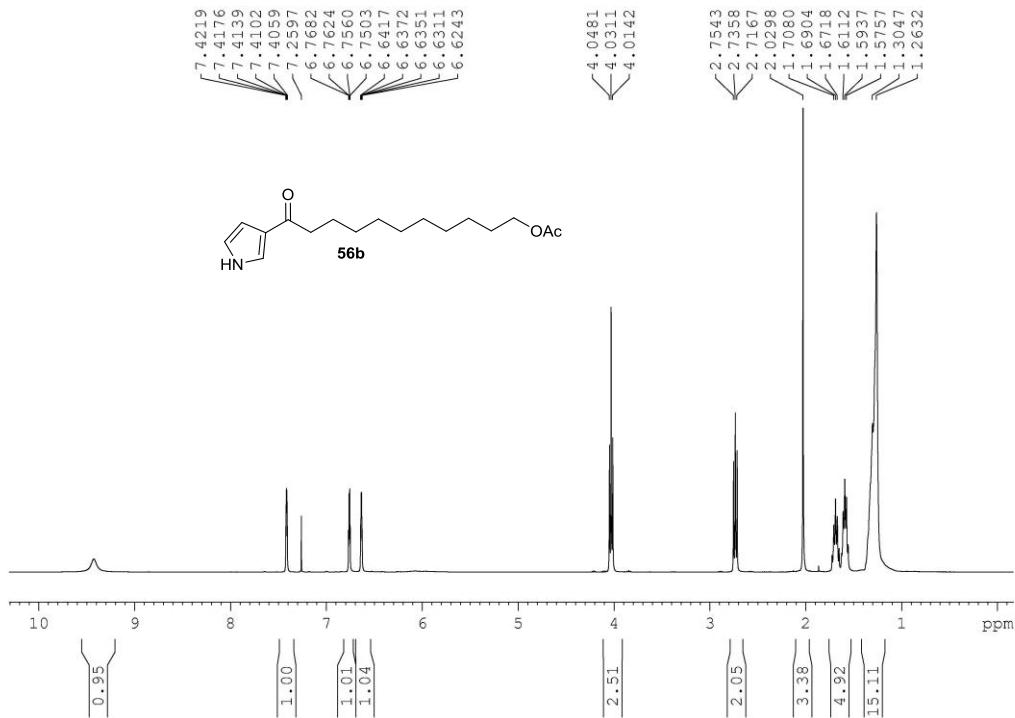
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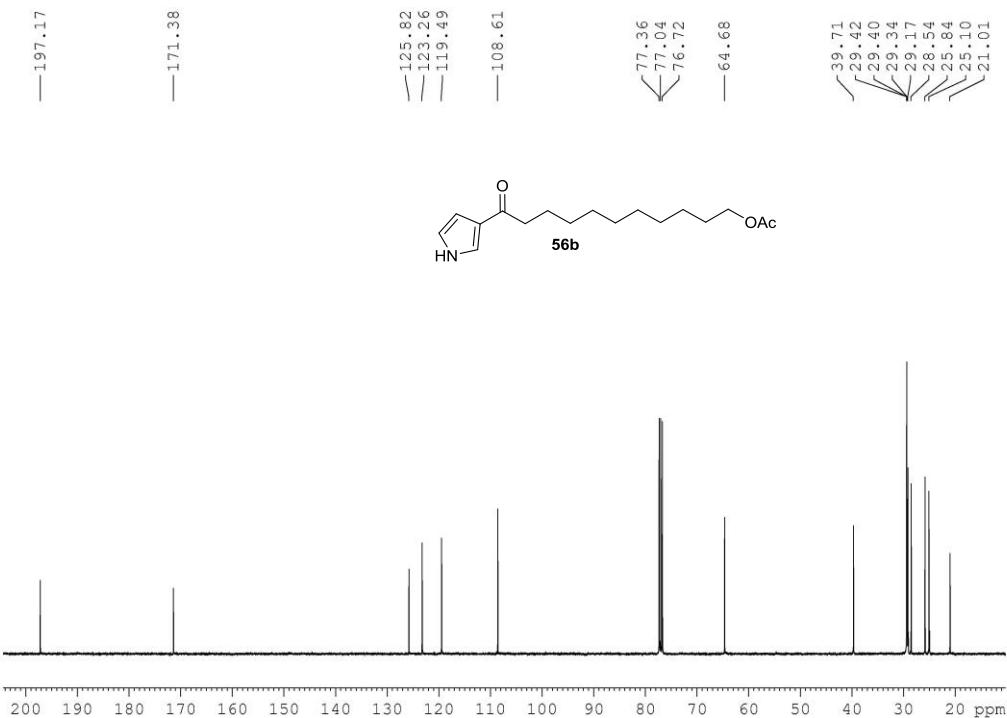
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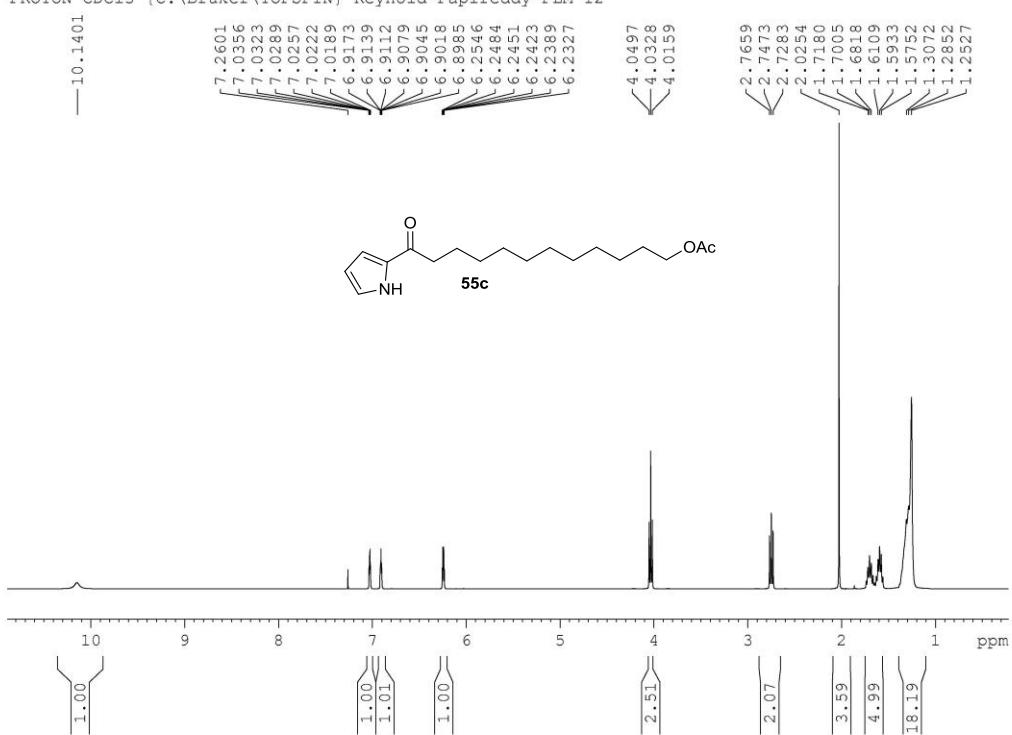
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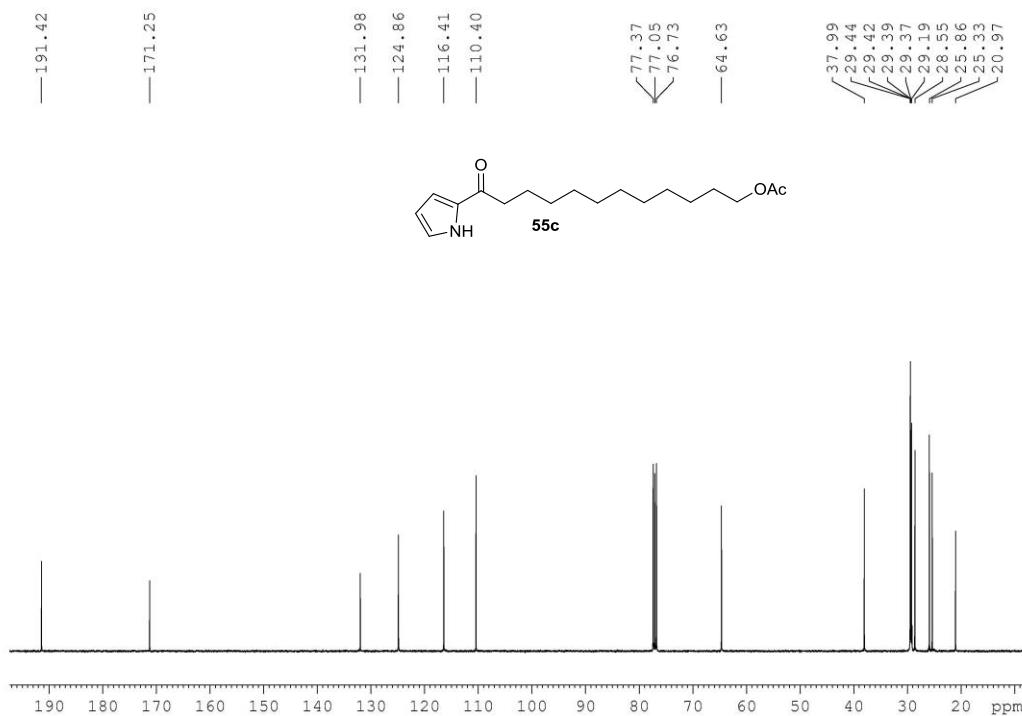
C13CPD CDC13 {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 11



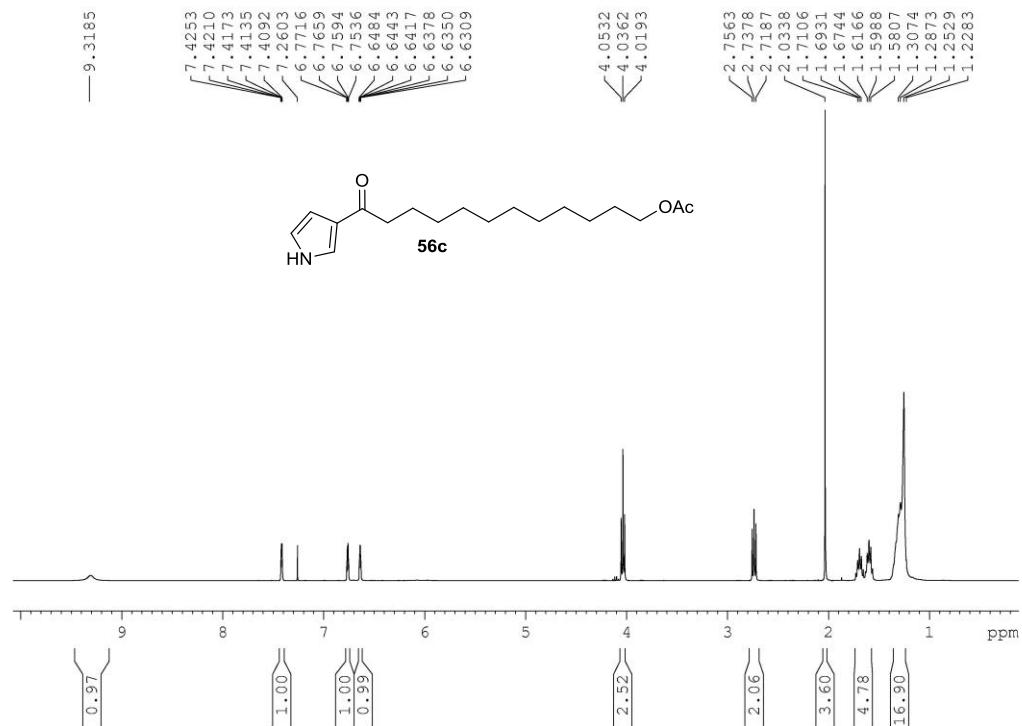
PROTON CDC13 {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 12



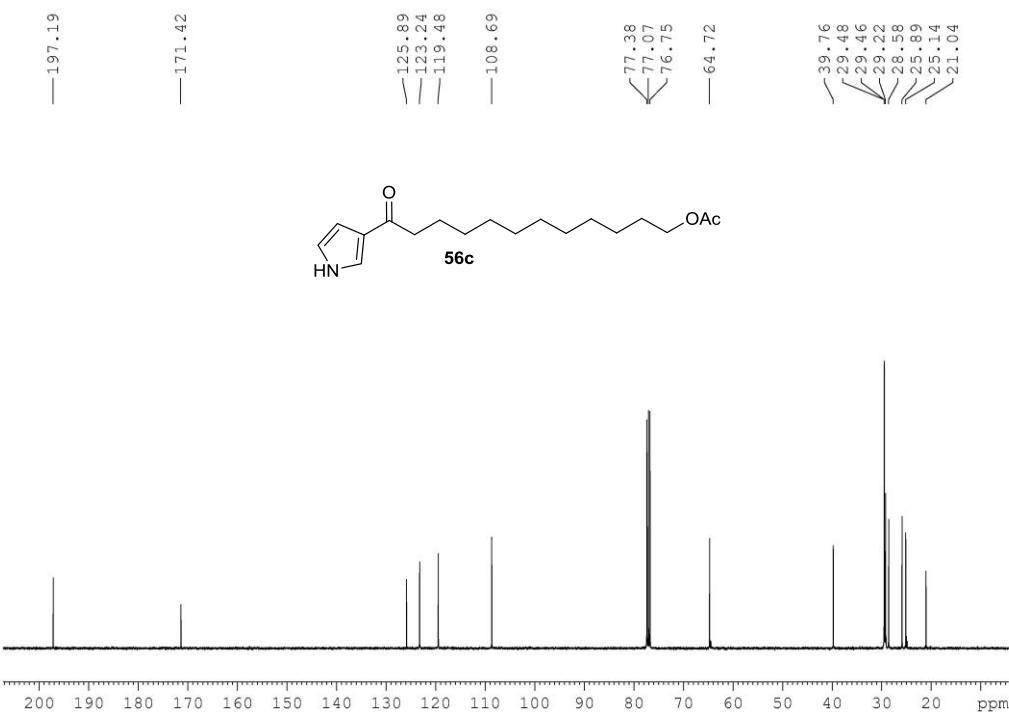
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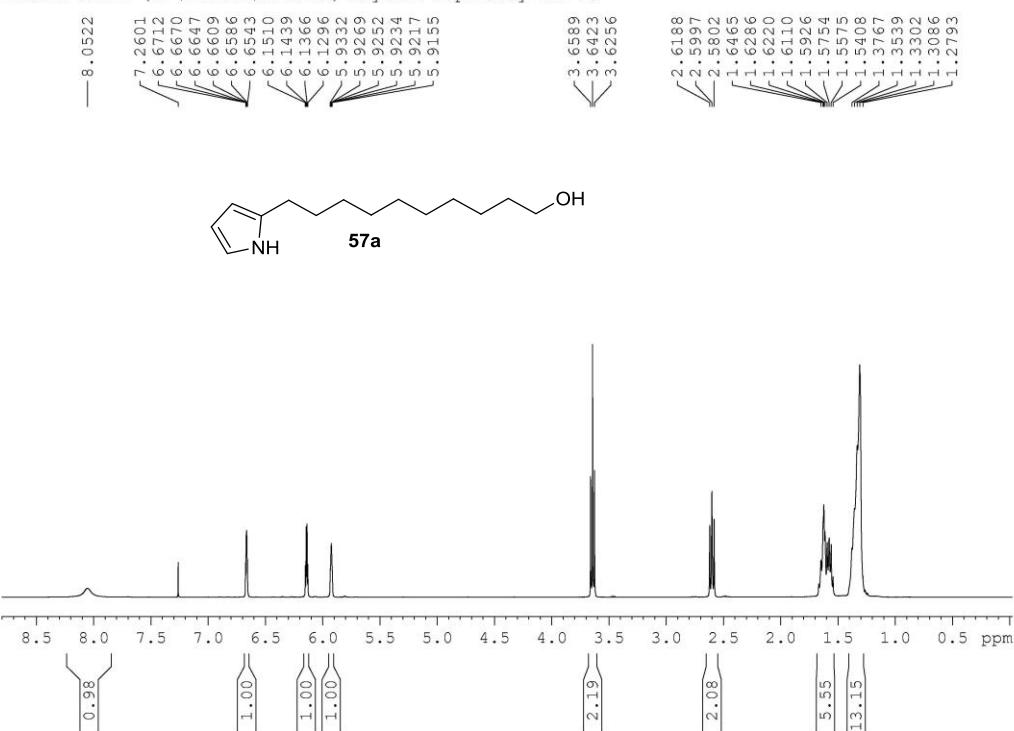
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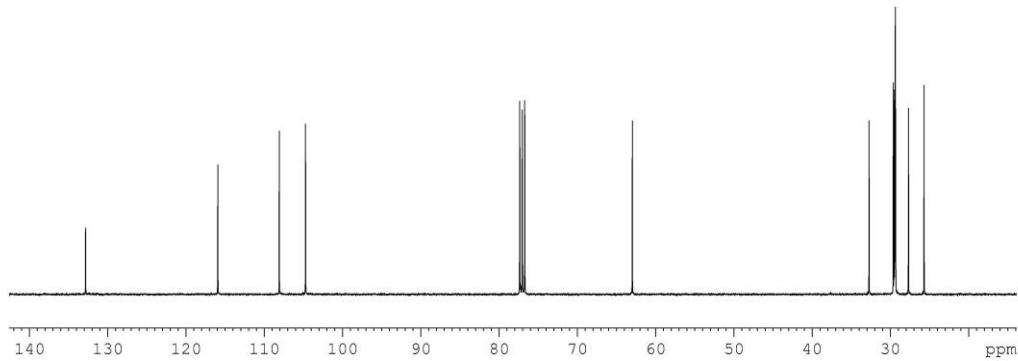
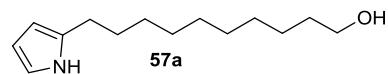
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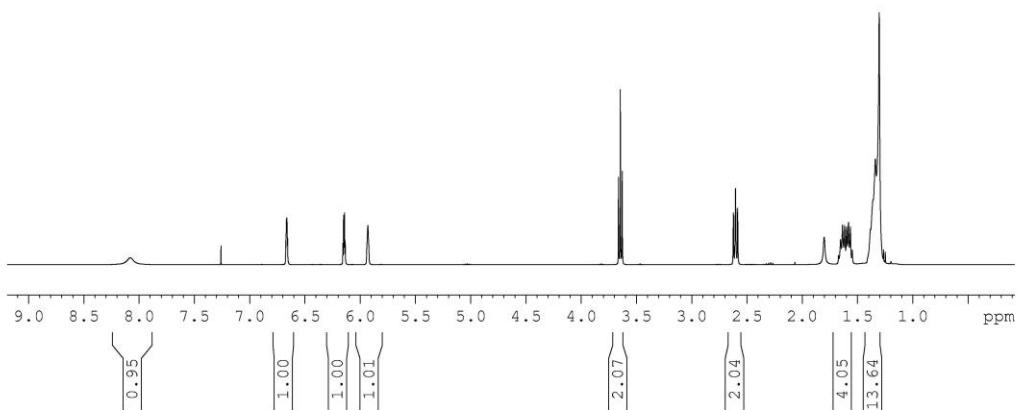
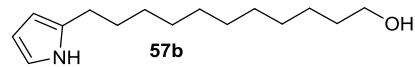
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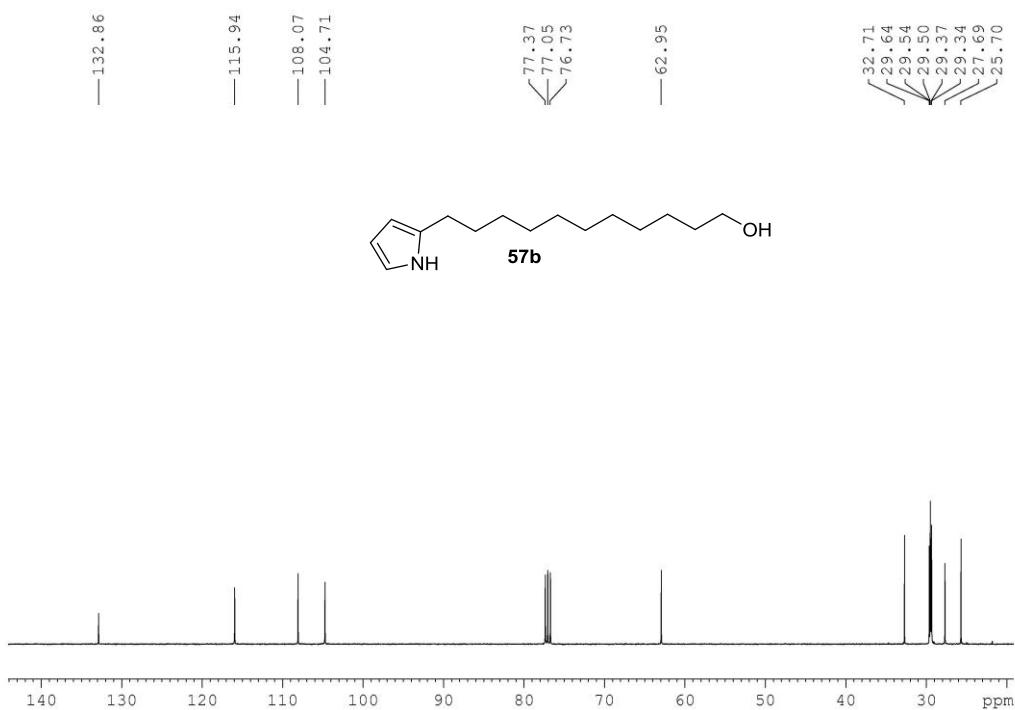
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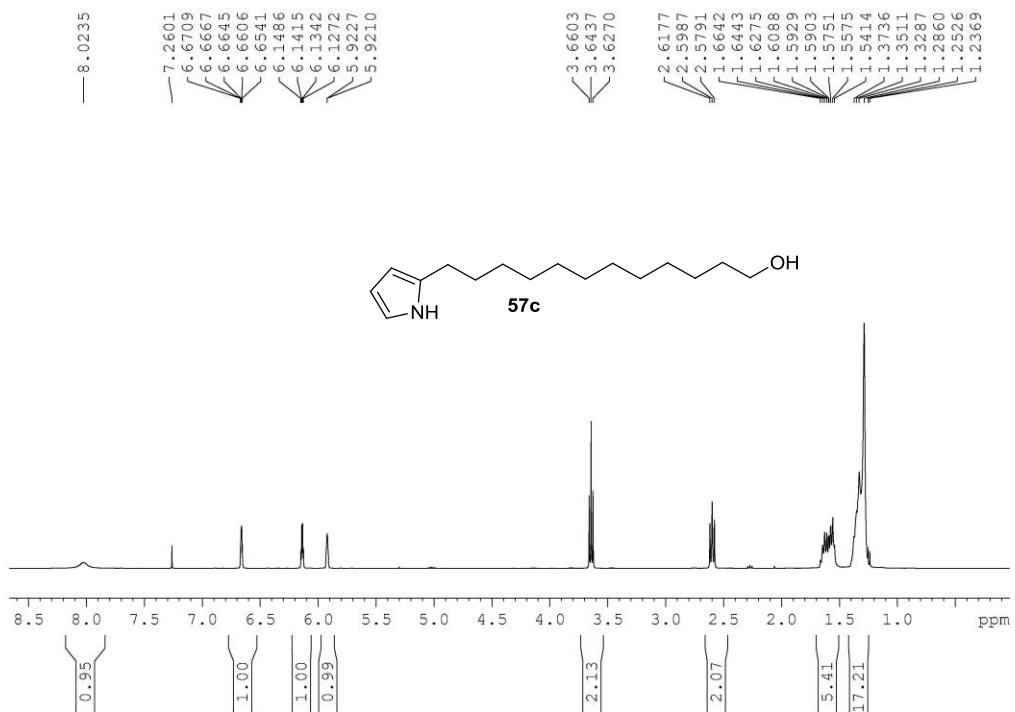
PROTON CDC13 {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 9



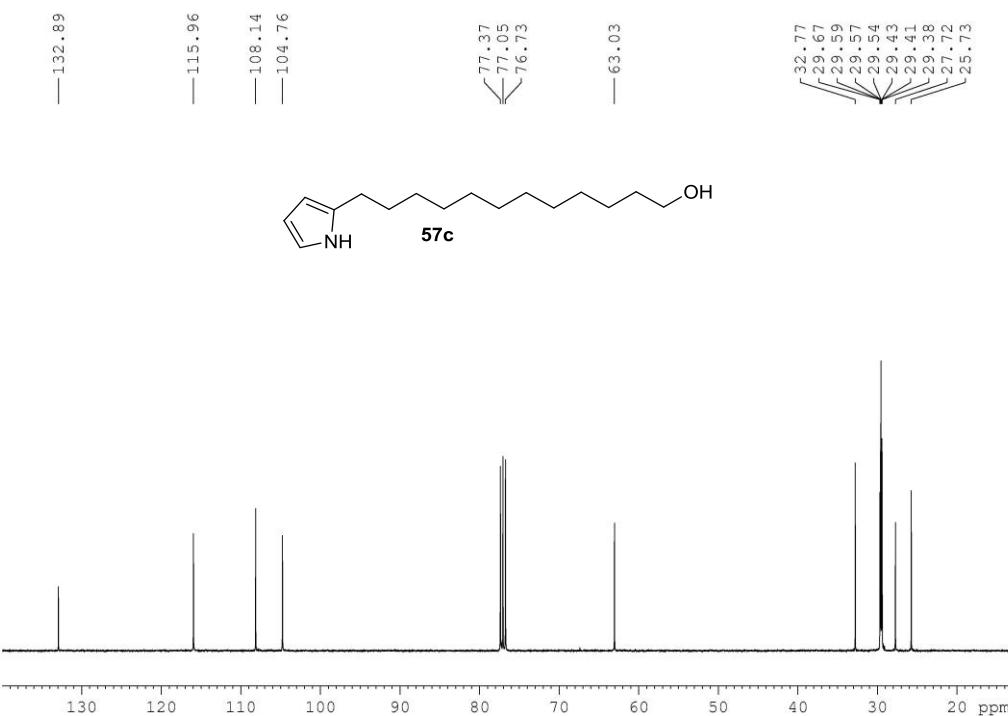
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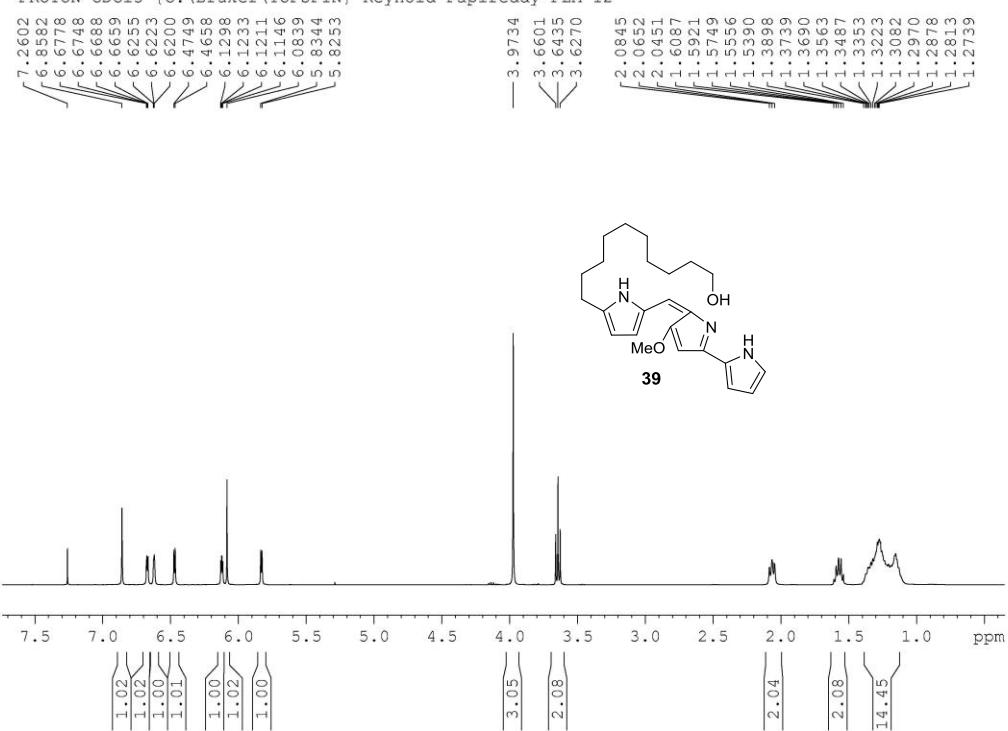
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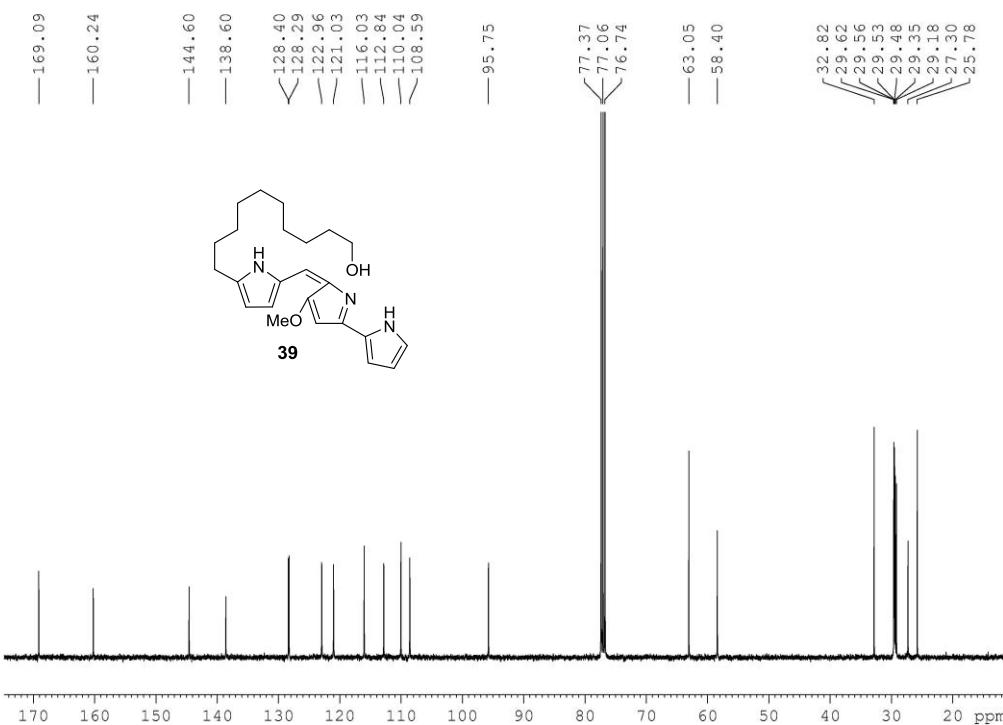
C13CPD CDC13 {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 5



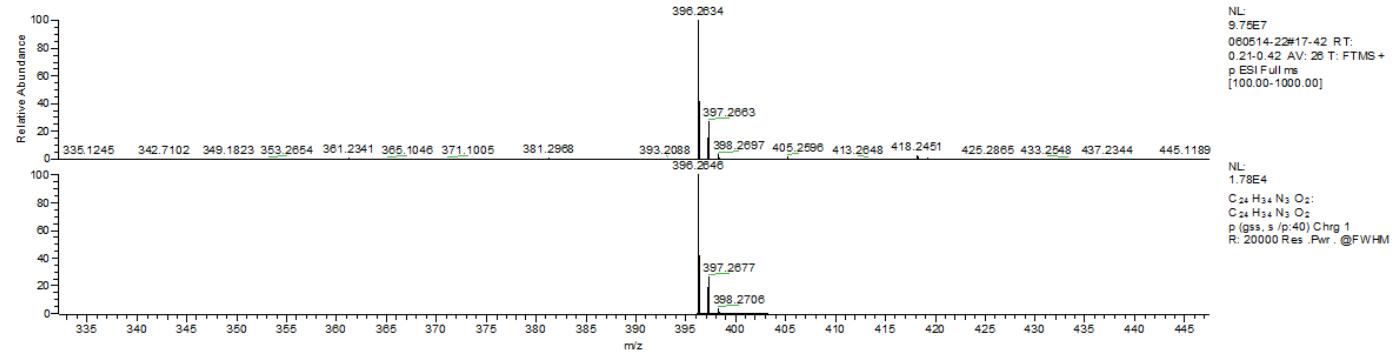
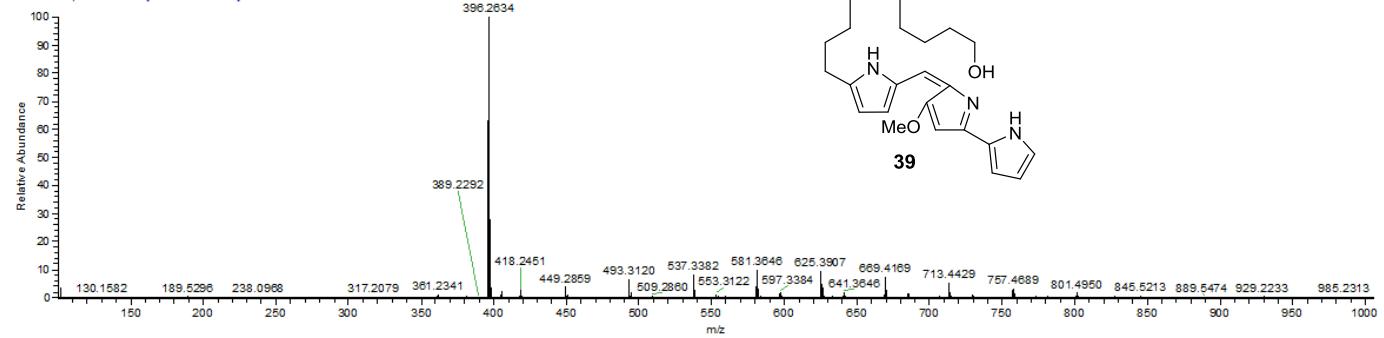
PROTON CDC13 {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 12



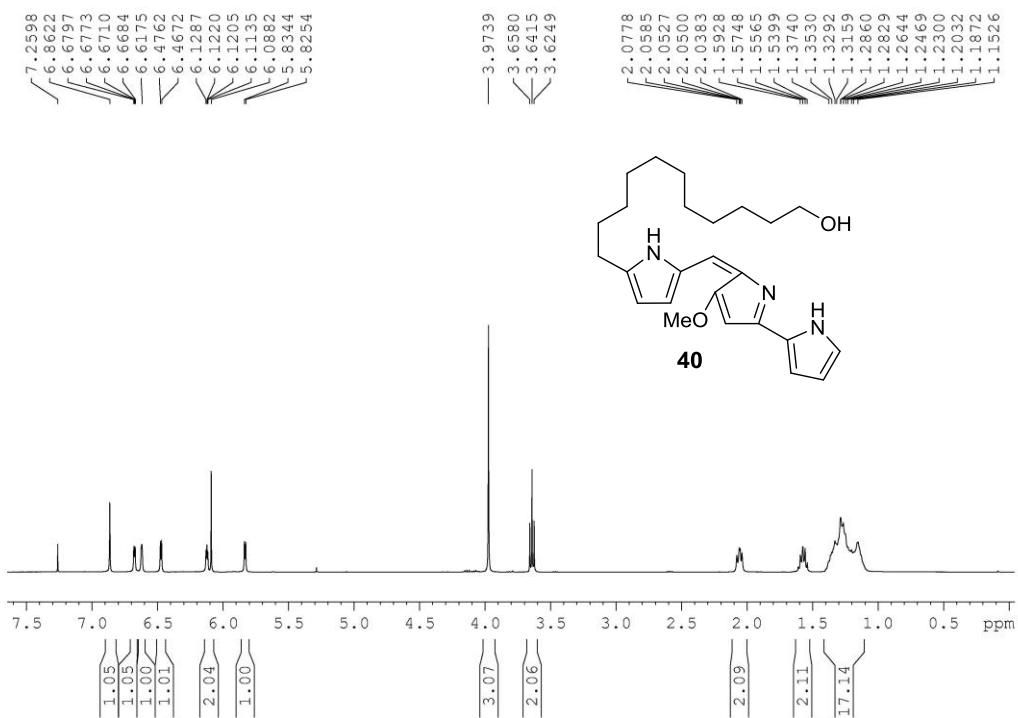
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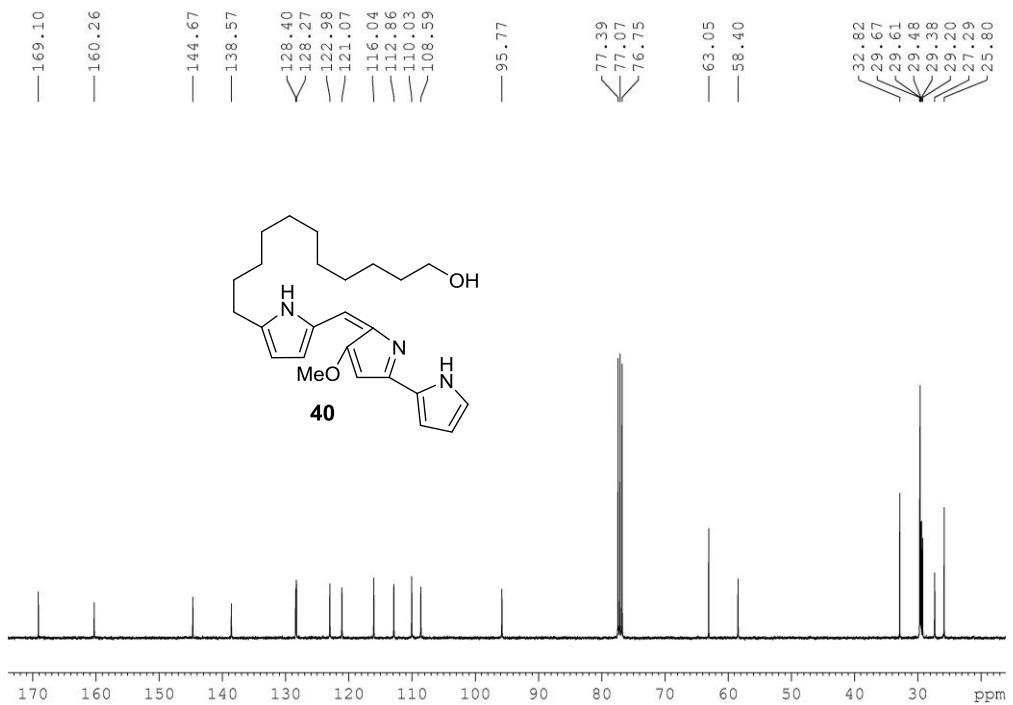
060514-22 #17-42 RT: 0.21-0.42 AV: 26 NL: 9.75E7  
T: FTMS + p ESI Full ms [100.00-1000.00]



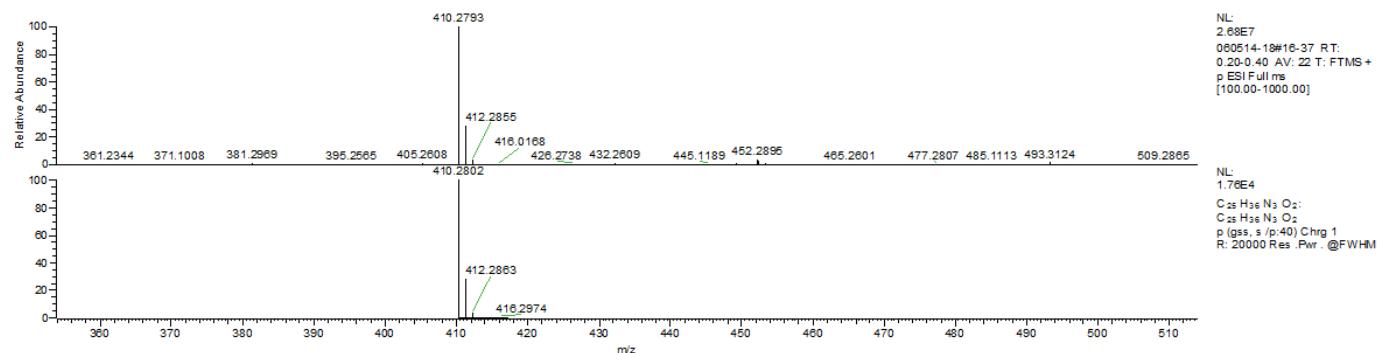
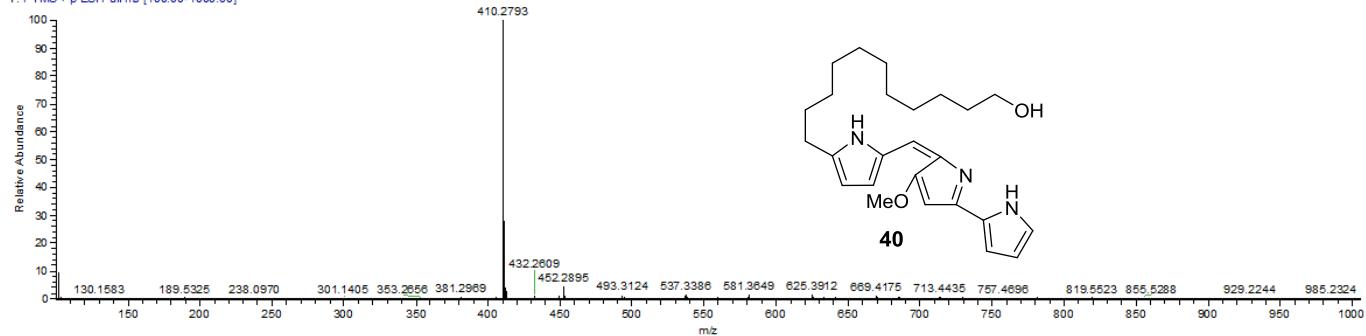
PROTON CDC13 {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 10



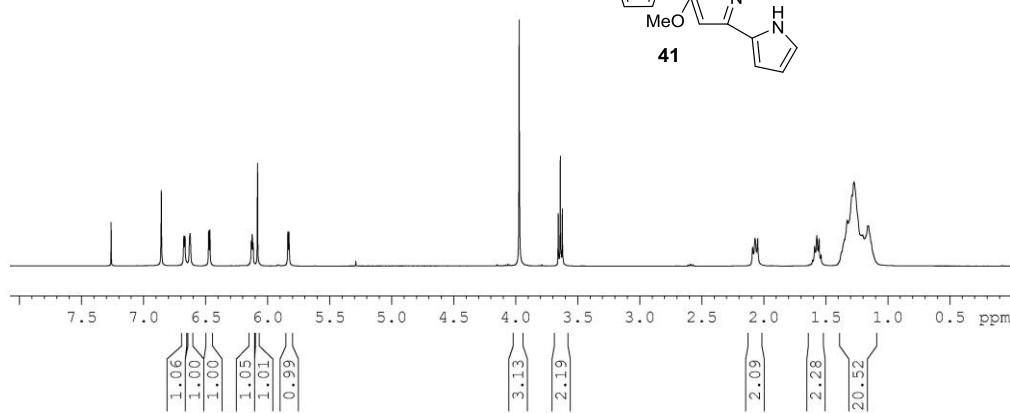
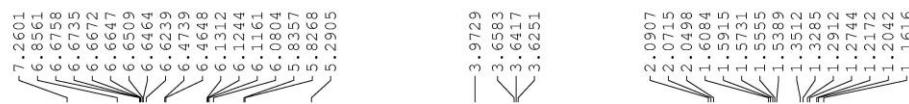
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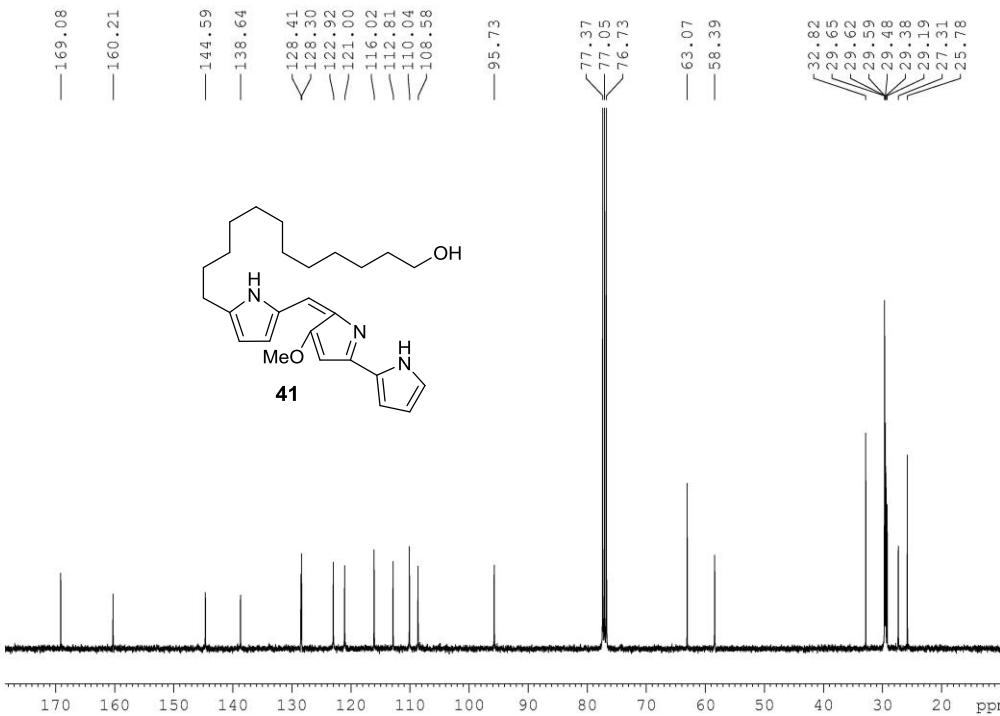
060514-18 #16-37 RT: 0.20-0.40 AV: 22 NL: 2.68E7  
T: FTMS + p ESI Full ms [100.00-1000.00]



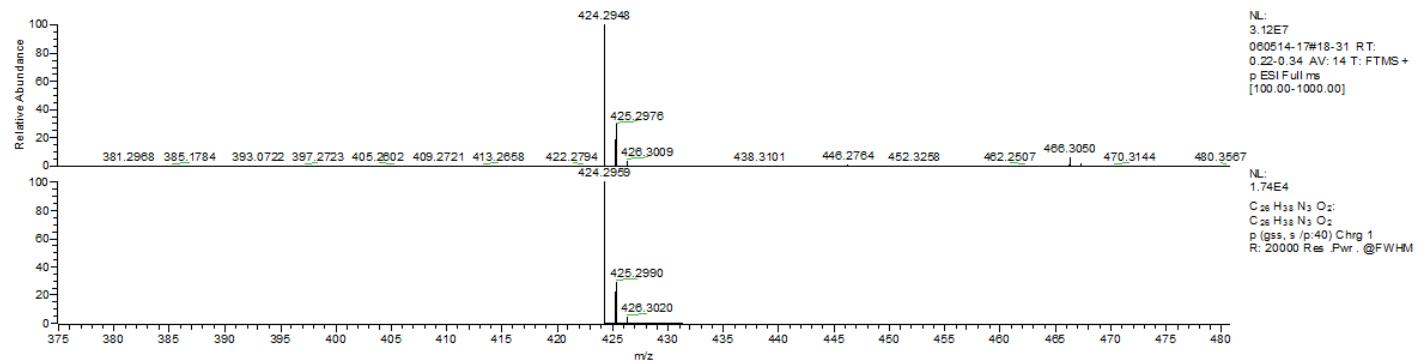
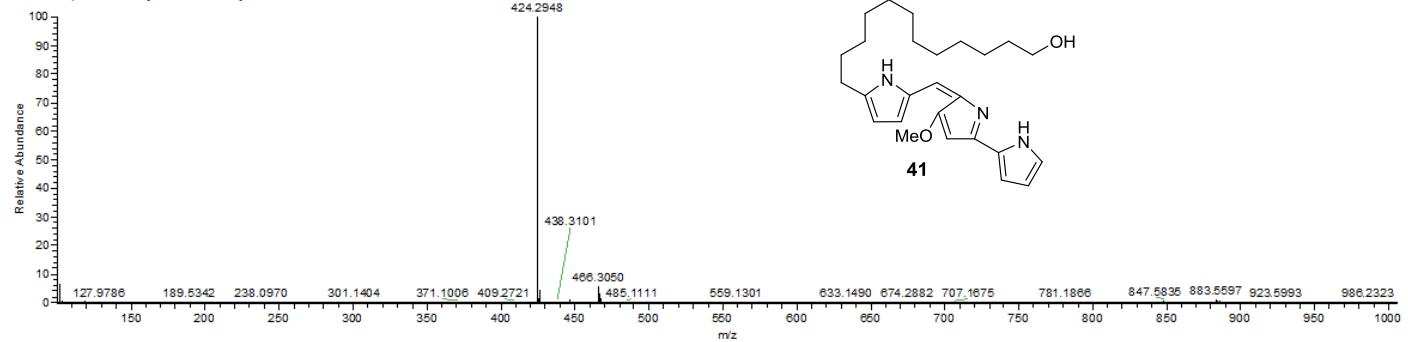
PROTON CDC13 {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 4



C13CPD CDCl<sub>3</sub> {C:\Bruker\TOPSPIN} Reynold-Papireddy-PLM 4



060514-17 #18-31 RT: 0.22-0.34 AV: 14 NL: 3.12E7  
T: FTMS + p ESI Full ms [100.00-1000.00]



## References

1. Paget, M. S.; Chamberlin, L.; Atri, A.; Foster, S. J.; Buttner, M. J. *J. Bacteriol.* **1999**, *181*, 204–211.
2. Xue, Y.; Zhao, L.; Liu, H. W.; Sherman, D. H. *Proc Natl Acad Sci U S A* **1998**, *95*, 12111–12116.
3. Gust, B.; Challis, G. L.; Fowler, K.; Kieser, T.; Chater, K. F. *Proc Natl Acad Sci U S A* **2003**, *100*, 1541–1546.
4. Salem, S. M.; Kancharla, P.; Florova, G.; Gupta, S.; Lu, W.; Reynolds, K. A. *J. Am. Chem. Soc.* **2014**, *136*, 4565–4574.