

ω-Alkynyl Lipid Surrogates for Polyunsaturated Fatty Acids: Free Radical and Enzymatic Oxidations

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Author Notes

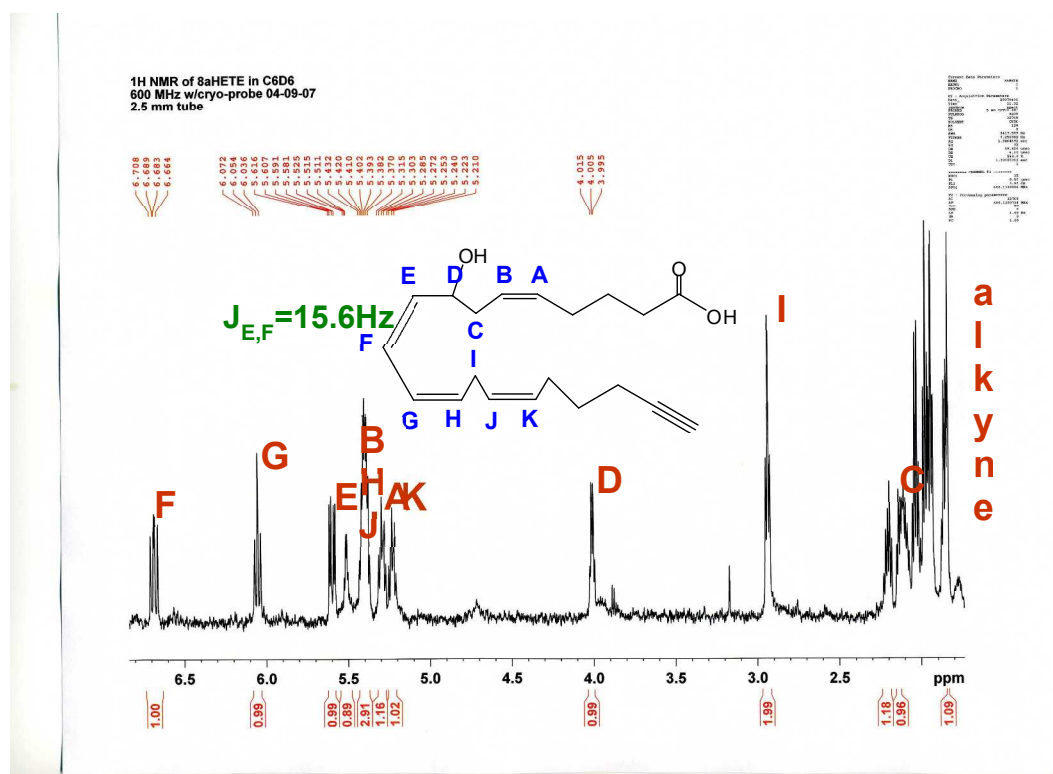
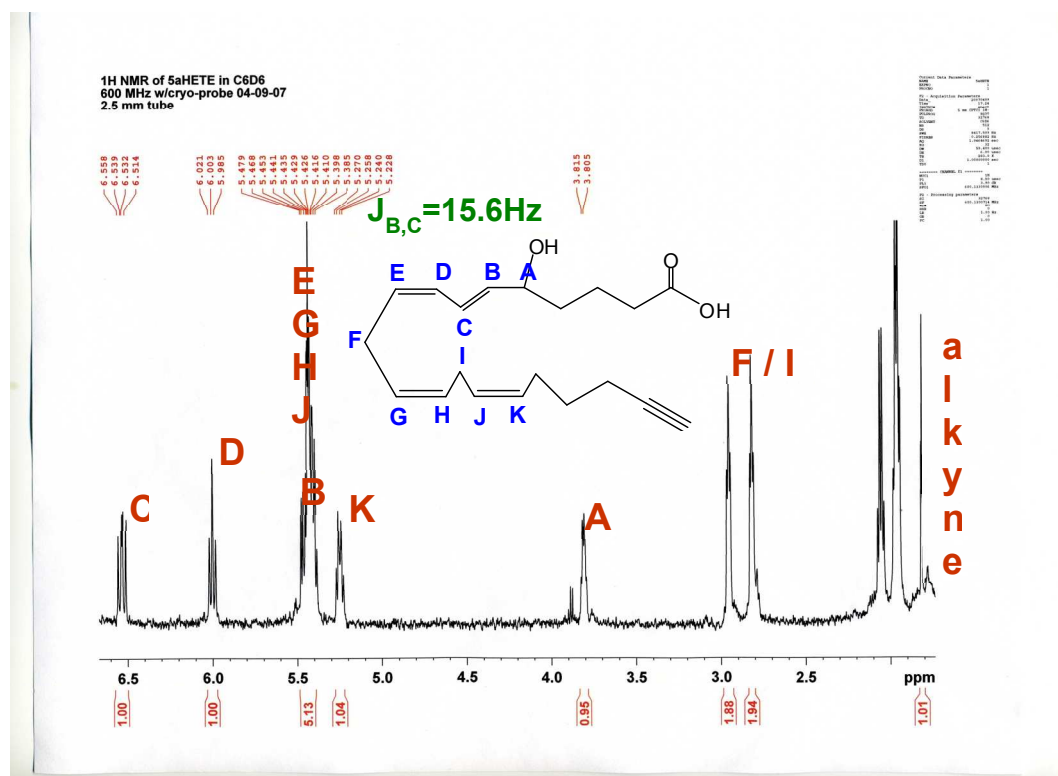
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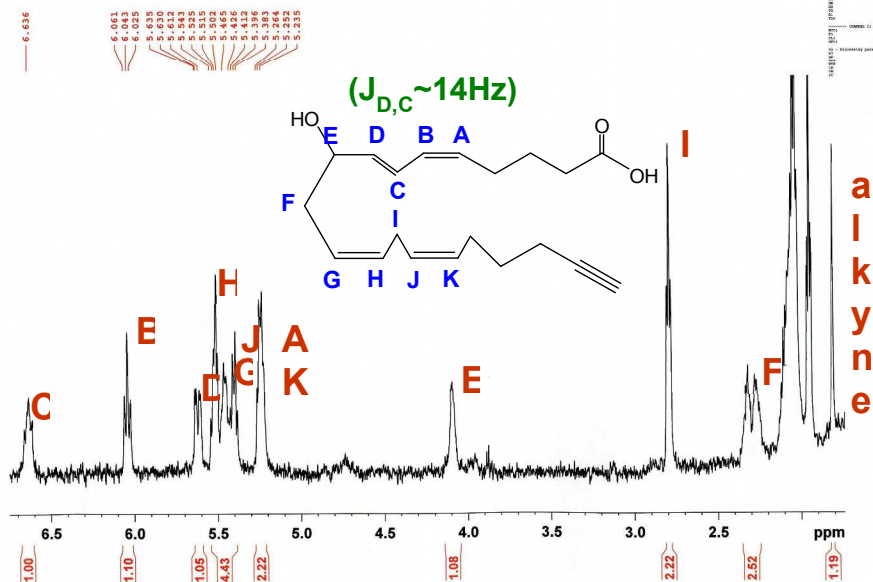
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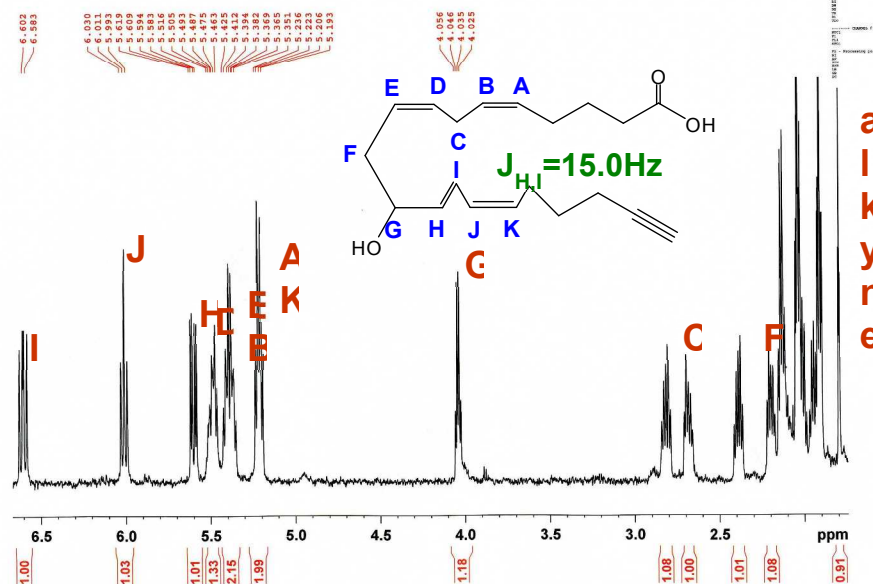
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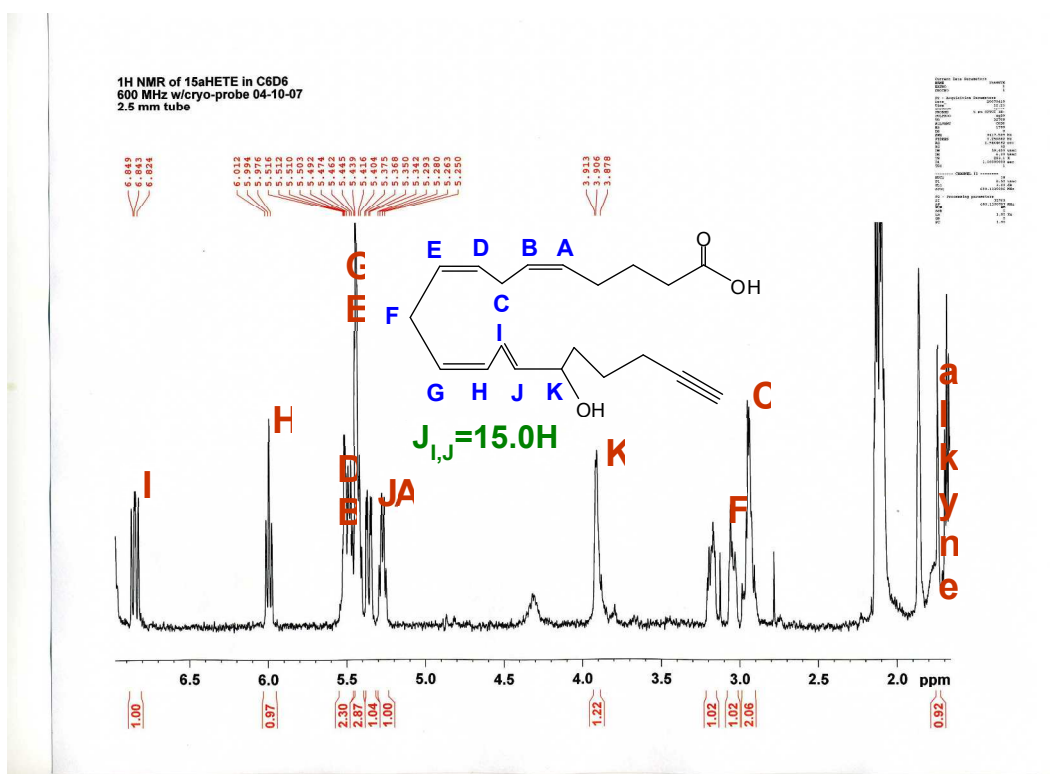
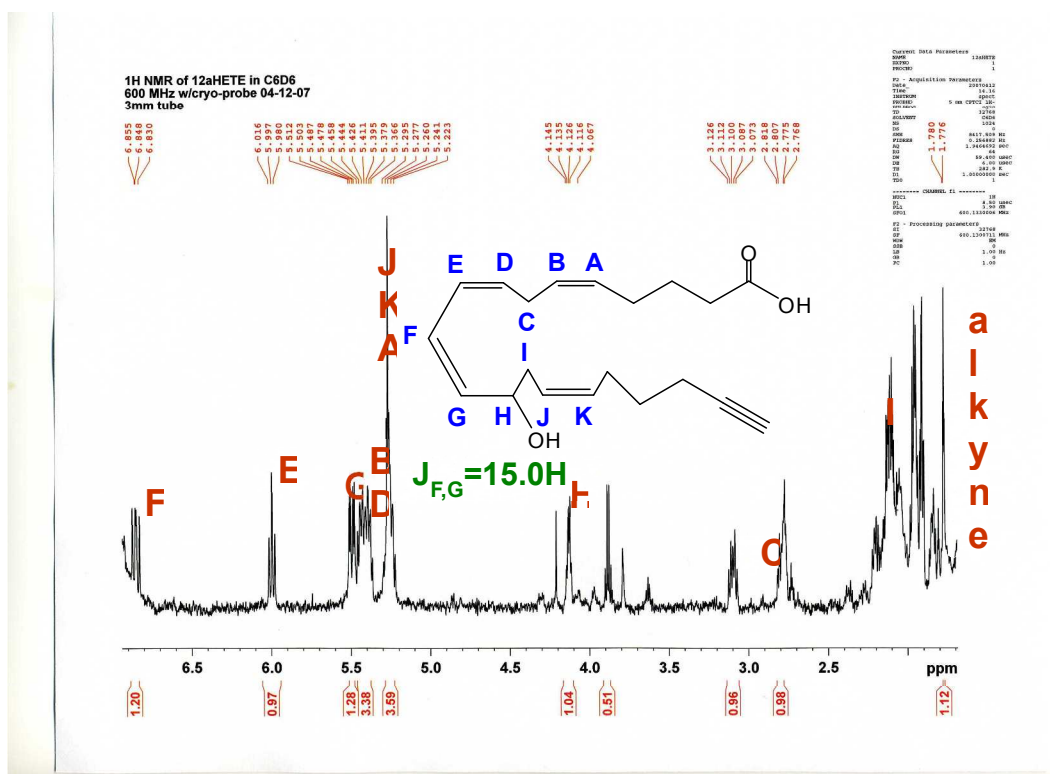


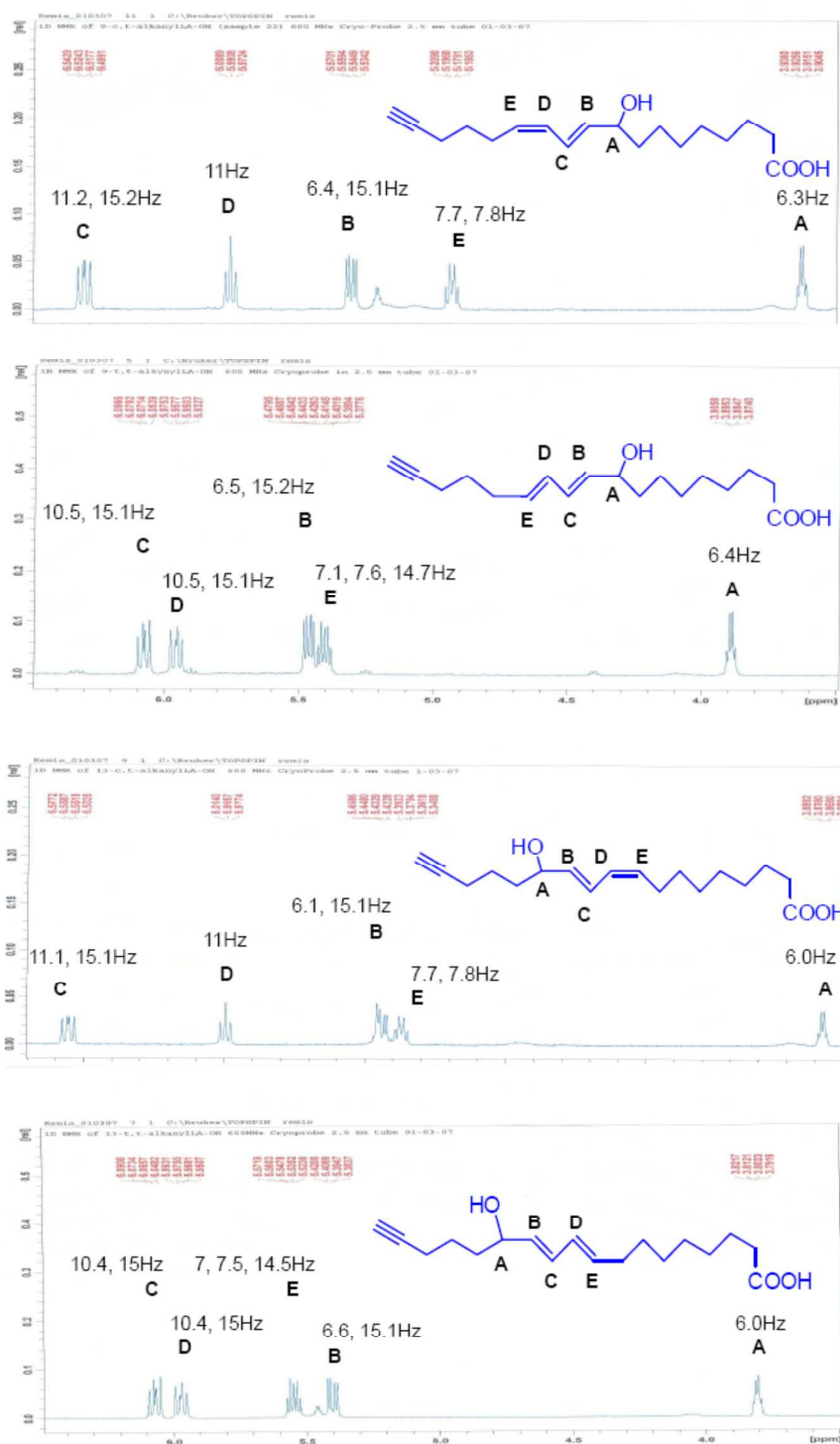
¹H NMR of 9aHETE in C6D6
600 MHz w/cryo-probe 04-09-07
2.5 mm tube



¹H NMR of 11aHETE in C6D6
600 MHz w/cryo-probe 04-09-07
2.5 mm tube



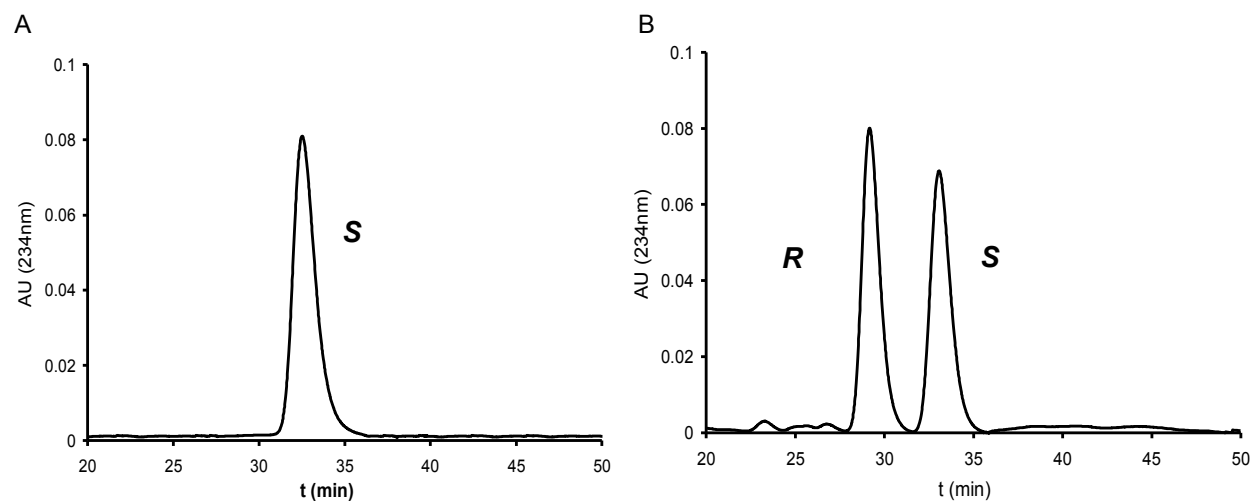




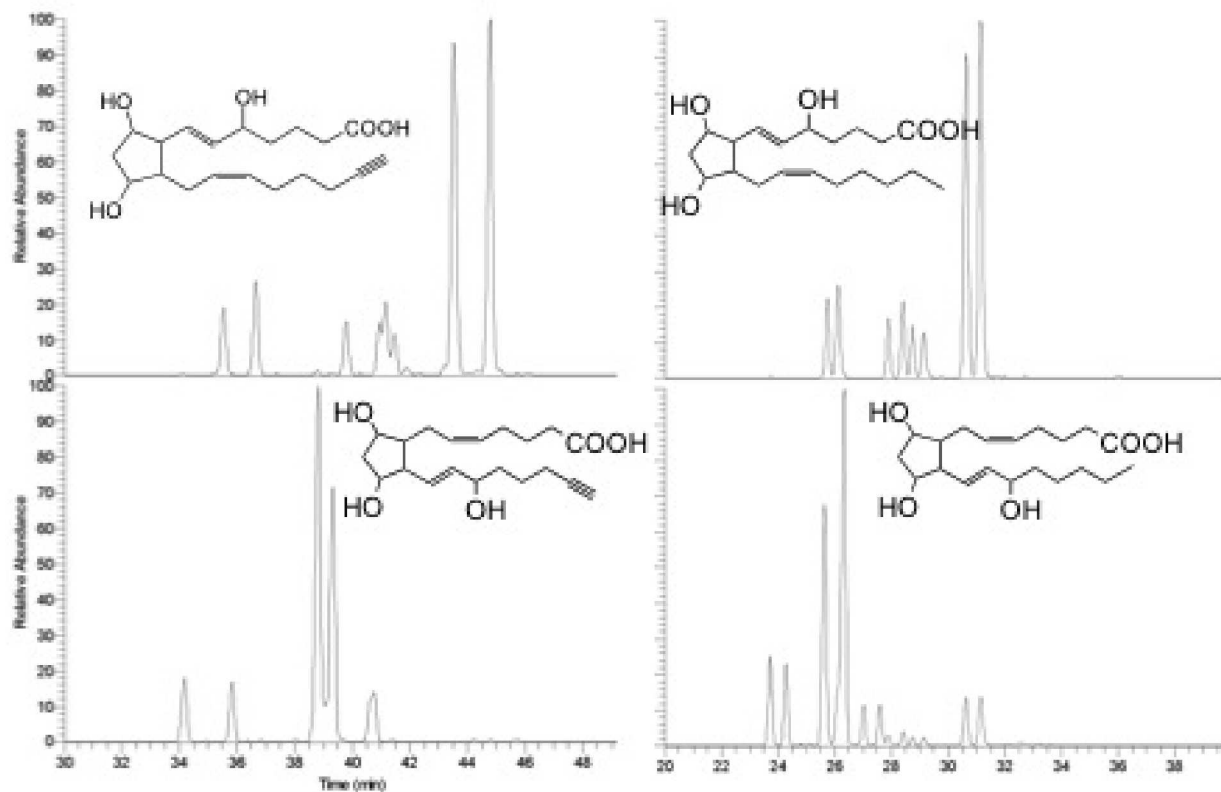
Supplemental Figure 1. ^1H NMR spectra of alkynyl HETE and HODE products, and coupling constants assigning the double bonds as either cis or trans.

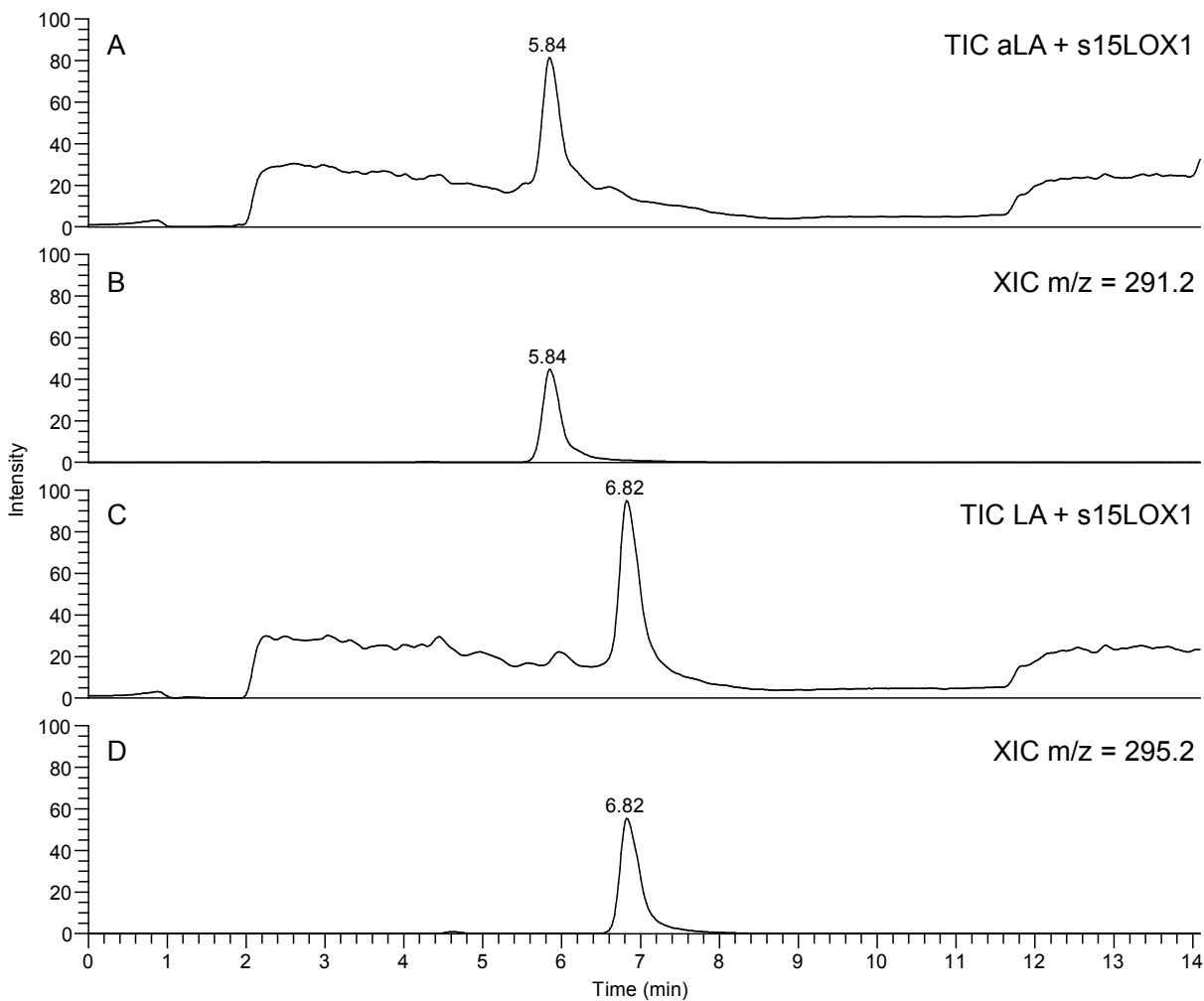
	Parent ion (m/z)	Products of in source dehydration/decarboxylation (m/z)	Fragment ions (m/z)
5- <i>a</i> HETE	314.8	253.2, 297.2	115.2, 199.2
8- <i>a</i> HETE	315.3	253.4, 297.1	155.0, 159.1
9- <i>a</i> HETE	315.0	253.4, 296.9	123.1, 147.0, 166.9, 175.0
11- <i>a</i> HETE	315.1	297.3	166.9
12- <i>a</i> HETE	315.1	253.1, 297.2	179.2, 208.2
15- <i>a</i> HETE	315.2	253.2, 297.3	175.0, 218.9

Supplemental Table 1. MS² fragmentation patterns for *a*HETE (MS signal optimized with 15-HETE). Red and blue numbers represent non-regiospecific fragments.

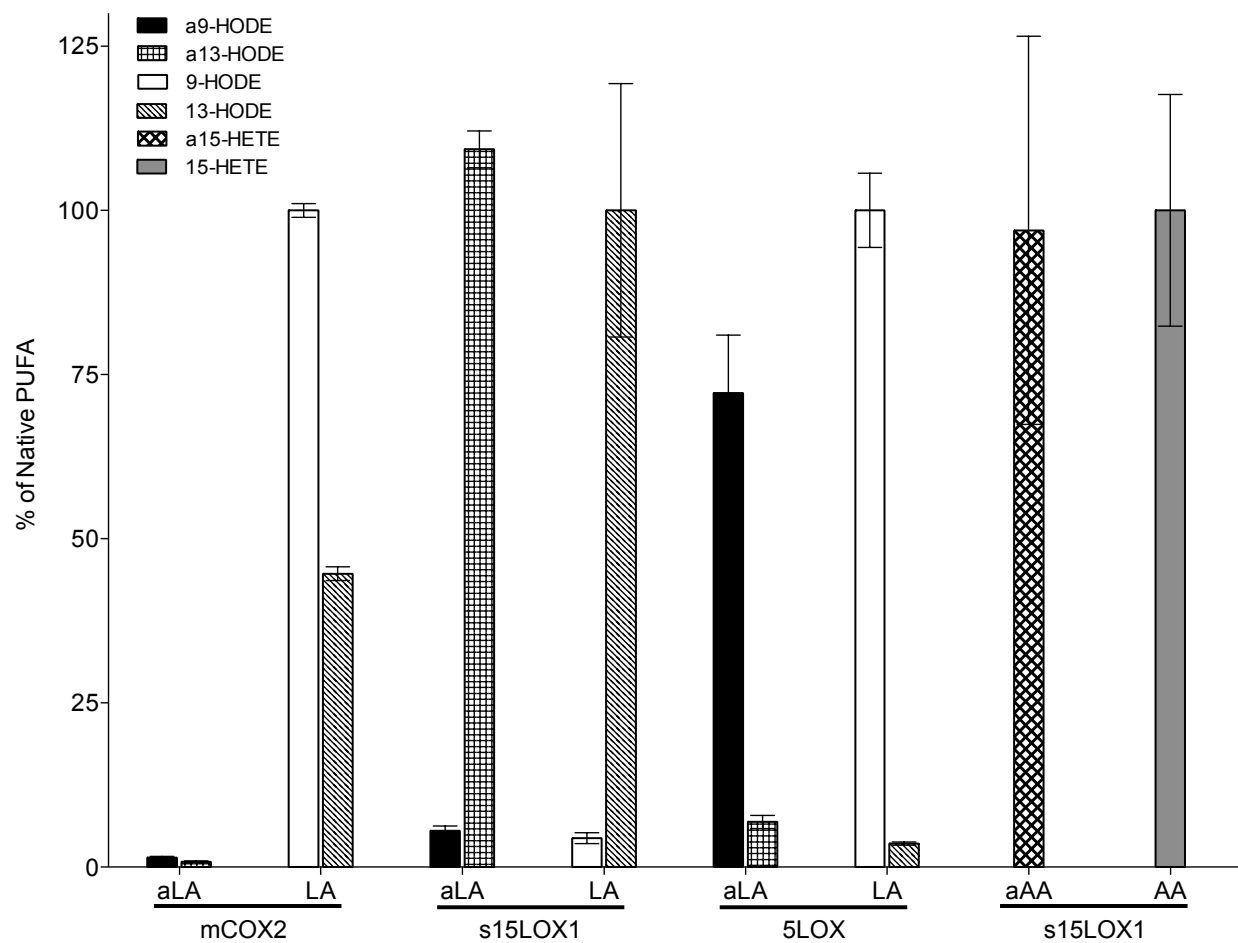


Supplemental Figure 2. Chiral analysis of alkynyl 13-c,t HODE methyl ester produced enzymatically by the reaction with s15LOX1 giving an optically pure product (A), and nonenzymatically giving mixture of optical enantiomers (B).

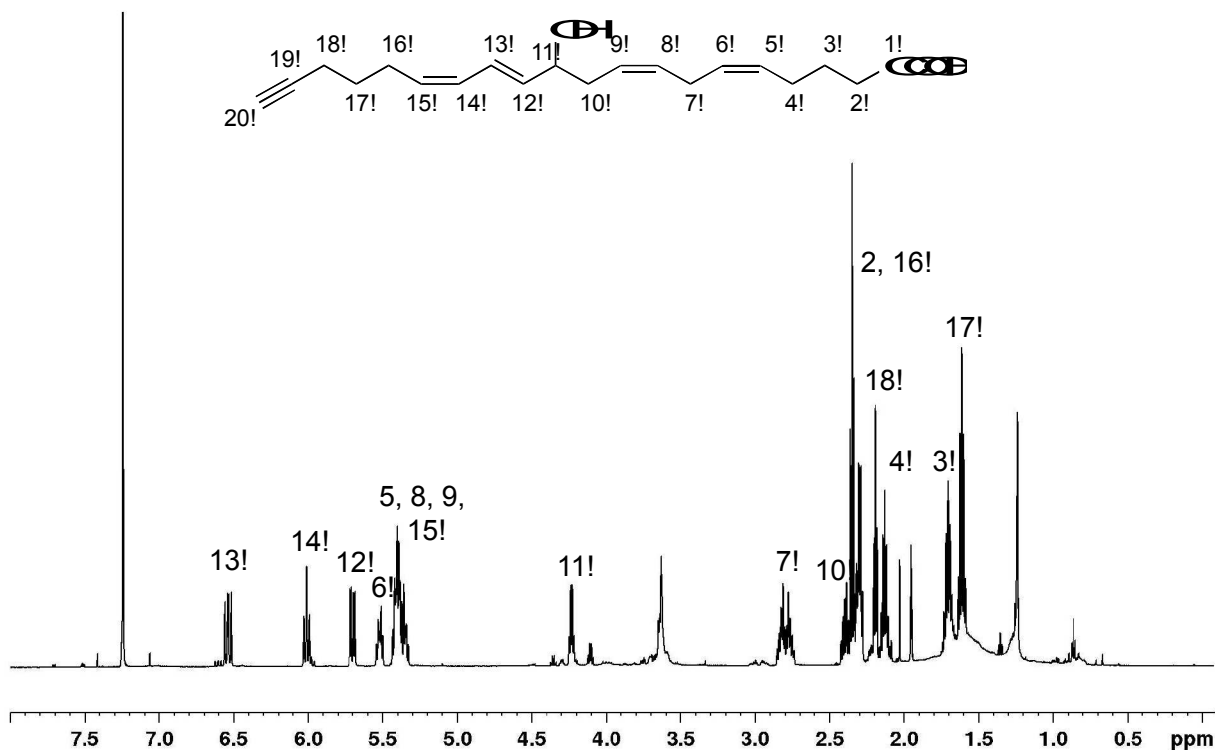




Supplemental Figure 4. A comparison of the metabolite profiles of *a*LA (A) and LA (C) with s15LOX1. The metabolism of *a*LA by s15LOX1 shows a single product showing the addition of oxygen. The metabolism of LA by s15LOX1 also shows a single product showing the addition of oxygen.



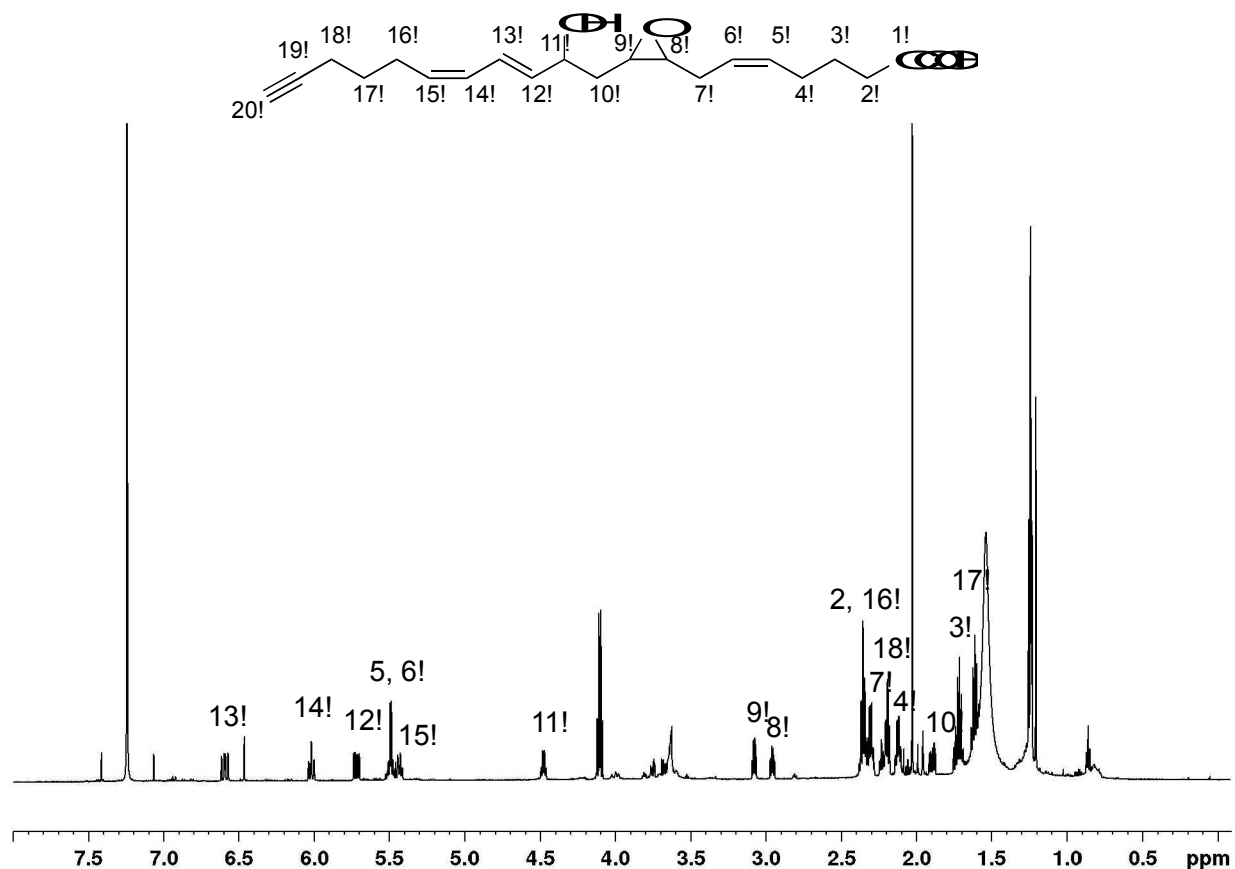
Supplemental Figure 5. Quantification of single oxygenation metabolites for the reaction between the PUFA pairs *a*LA/LA and *a*AA/AA with murine COX2, soybean 15LOX1, and potato 5LOX. Fatty acid pairs are normalized to the most abundant native (LA or AA) metabolite for each enzyme.



Supplemental Figure 6. ^1H NMR spectrum of *a11*-HETE in CDCl_3

Chemical shift	Multiplicity	Proton(s)	Coupling Constant
ppm		carbon no.	Hz
6.54	<i>dd</i>	H13	$J_{12,13} = 15.1$
6.01	<i>t</i>	H14	$J_{14,15} = 11.0$
5.70	<i>dd</i>	H12	$J_{12,13} = 15.2$
5.51	<i>m</i>	H6	
5.39	<i>m</i>	H5, 8, 9, 15	
4.23	<i>q</i>	H11	
2.78	<i>m</i>	H7	
2.34	<i>m</i>	H2a, 10, 16	
2.19	<i>m</i>	H18	
2.12	<i>quint</i>	H4	
1.70	<i>m</i>	H3	
1.61	<i>quint</i>	H17	

Supplemental Table 2. ^1H -NMR chemical shifts and coupling constants of the collected LCMS peak with $m/z = 315.2$, which was identified to be *a11*-HETE.



Supplemental Figure 7. ^1H NMR spectrum of *a*11-8,9-HEET in CDCl_3

Chemical shift	Multiplicity	Proton(s)	Coupling Constant
ppm		carbon no.	Hz
6.59	<i>dd</i>	H13	$J_{12,13} = 15.1$
6.02	<i>t</i>	H14	$J_{14,15} = 11.0$
5.72	<i>dd</i>	H12	$J_{12,13} = 15.2$
5.51	<i>m</i>	H5,6	
5.45	<i>dt</i>	H15	$J_{14,15} = 10.6$
4.48	<i>q</i>	H11	
3.08	<i>quint</i>	H9	$J_{8,9} = 4.2$
2.95	<i>dt</i>	H8	$J_{8,9} = 4.3$
2.24	<i>m</i>	H2,4,7,16,18	
1.90	<i>m</i>	H10a	
1.72	<i>m</i>	H10b	
1.61	<i>quint</i>	H17	

Supplemental Table 3. ^1H -NMR chemical shifts and coupling constants of the collected LCMS peak with $m/z = 331.2$, which was identified to be *a*11-8,9-HEET.