

Synthesis and Chiral Resolution of Twisted Carbon Nanobelts

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⁺These authors contributed equally to this work

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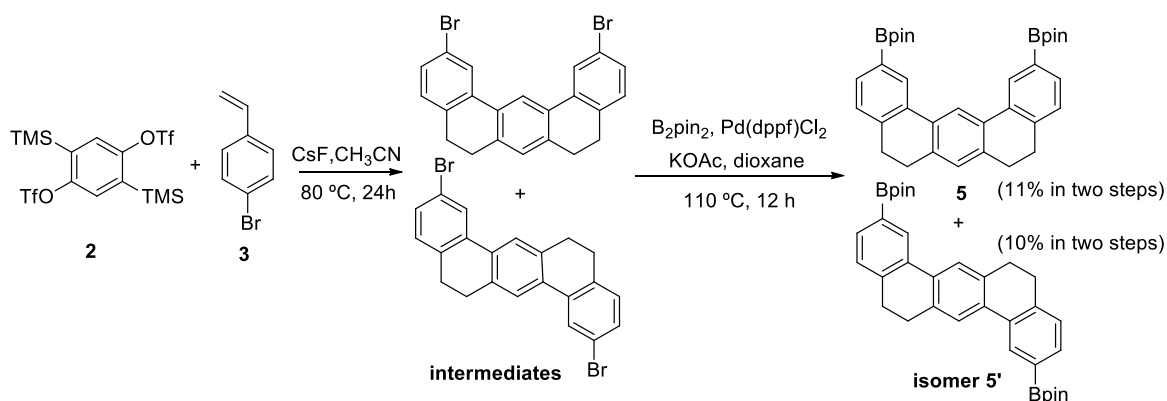
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1. Experimental Section

1.1 General information

The starting material **3** was obtained from commercial suppliers and used without further purification. Compounds **2**¹ and **4**² were synthesized according to previous literatures. ¹H NMR and ¹³C NMR spectra were recorded in deuterated solvents on a Bruker AVANCE 400 NMR Spectrometer and a Bruker AVIII 500WB NMR Spectrometer. ¹H NMR chemical shifts are reported in ppm using the residual protonated solvent as an internal standard and tetramethylsilane (TMS) as reference. High resolution mass spectroscopic measurements were performed on IonSpec 4.7 Tesla Fourier Transform Mass Spectrometer (MALDI-TOF). UV-vis-NIR absorption spectra were recorded on a Shimadzu UV-3600 spectrophotometer. The absolute quantum yields were recorded on a JASCO model FP-6600 spectrofluorometer, equipped with integrating sphere. Time-resolved fluorescence spectroscopic measurements were conducted on a Hamamatsu model compact fluorescence lifetime spectrometer C11367 (Quantaurs-Tau). Cyclic voltammetry (CV) measurements were performed with a Zahner IM6e electrochemical workstation using glassy carbon disc as the working electrode, Pt wire as the counter electrode, Ag/AgCl electrode as the reference electrode. The solution of 0.1 M tetrabutylammonium hexafluorophosphate (TBAPF₆) dissolved in chlorobenzene was employed as the supporting electrolyte. Ferrocene was used as an external reference. The analytical HPLC was performed at 40 °C in a column oven (JASCO CO2060PLUS) under detection of UV-vis absorption (JASCO MD2018PLUS) and circular dichromism (CD) intensities (JASCO CD2095PLUS) with a flow rate of 1.0 mL/min. The preparative HPLC was performed at ambient temperature under detection of UV-vis (JASCO UV2075PLUS) with a flow rate of 10 mL/min. CD spectra were obtained on JASCO J-1500.

1.2 Synthetic procedures and characterization data for all new compounds

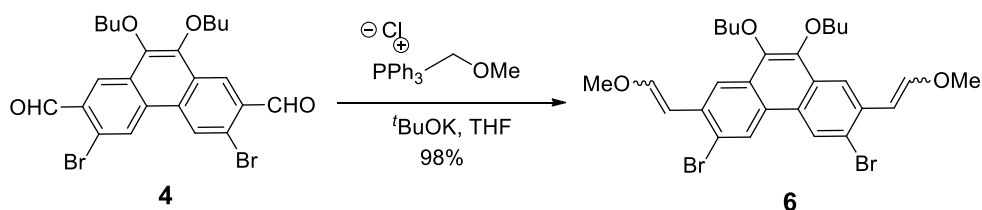


Compound **5**:

Compound **2** (6.0 g, 11.4 mmol), compound **3** (10.04 g, 57 mmol) and CsF (10.8 g, 71.16 mmol) were dissolved in dry acetonitrile (130 mL) under N₂ atmosphere. The reaction mixture was heated for

24 h at 110 °C. After cooling to room temperature, solvent was removed under reduced pressure and the residue was extracted by DCM and water. The combined organic layers were dried over MgSO₄. After removal of the solvent under vacuum, the residue was purified by silica gel chromatography (Hexane) to afford the intermediate isomers as a white solid (1.8 g). The intermediates were conducted to next step without further purification due to the low solubility.

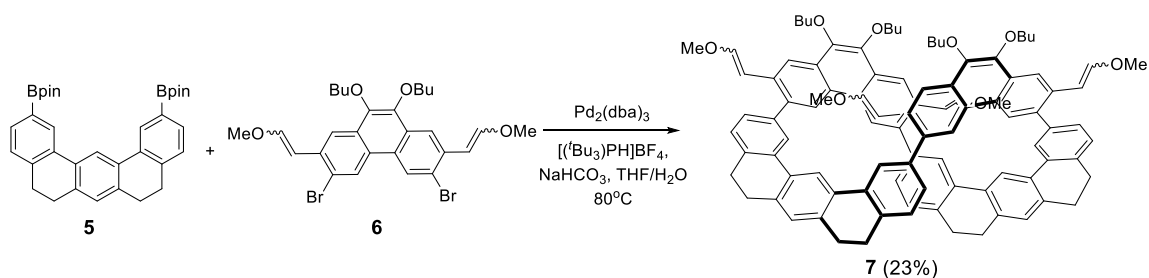
The intermediates with bis(pinacolato)diboron (4.15 g, 16.36 mmol), Pd(dppf)Cl₂ (117.1 mg, 0.16 mmol) and KOAc (803.1 mg, 8.18 mmol) were dissolved in dry dioxane (40 mL) in a sealed tube with stirrer bar. The reaction mixture was heated for 12 h at 110 °C under N₂ atmosphere. After cooling to room temperature, solvent was removed under reduced pressure and the residue was quickly purified via silica gel column chromatography (DCM). The crude product was and then further purified by preparative GPC using CHCl₃ at a rate of 14 mL/min to afford compound **5** (670 mg, 11% in two steps) and isomer **5'** (609 mg, 10% in two steps) as white solid. Compound **5**: ¹H NMR (400 MHz, 298 K, CD₂Cl₂): δ ppm 8.28 (s, 2H), 8.25 (s, 1H), 7.61 (dd, ³J = 5.9 Hz, ⁴J = 0.9 Hz, 2H), 7.25 (d, *J* = 5.9 Hz, 2H), 7.11 (s, 1H), 2.90-2.83 (m, 8H), 1.36 (s, 24H). ¹³C NMR (100 MHz, CDCl₃): δ ppm 140.7, 136.8, 133.9, 133.5, 133.1, 129.9, 127.8, 127.6, 119.2, 83.8, 29.4, 28.6, 24.8, 24.7. Compound **5'**: ¹H NMR (400 MHz, 298 K, CD₂Cl₂): δ ppm 8.16 (s, 2H), 7.17 (s, 2H), 7.61 (dd, ³J = 7.4 Hz, ⁴J = 1.1 Hz, 2H), 7.25 (d, *J* = 7.4 Hz, 2H), 2.91 (s, 8H), 1.34 (s, 24H). ¹³C NMR (100 MHz, CDCl₃): δ ppm 140.9, 136.0, 133.7, 133.4, 129.7, 127.7, 123.4, 83.8, 29.5, 28.7, 24.7. Compound **5**: HRMS analysis (MALDI, 100%, *m/z*) calcd for C₃₄H₄₀B₂O₄: 534.3119, found 534.3109 (error: -0.3 ppm).



Compound 6:

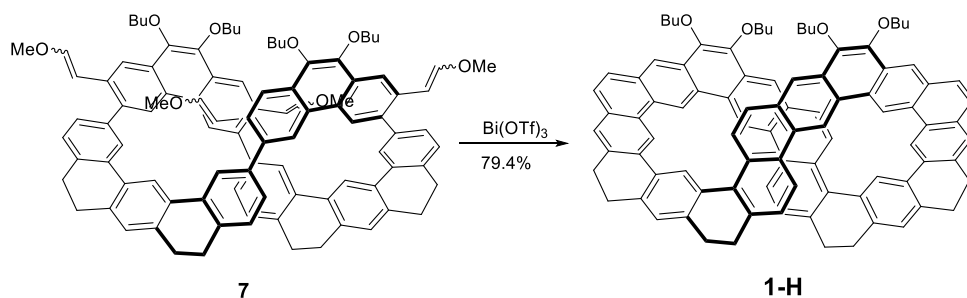
To a solution of methoxymethyltriphenylphosphonium chloride (767 mg, 2.24 mmol) in dry THF (20 mL) was added *t*-BuOK (251 mg, 2.24 mmol) at 0 °C under N₂ atmosphere. After stirring for 30 mins, a solution of compound **4** (400 mg, 0.75 mmol) in THF (20 mL) was added and the mixture was stirred at room temperature for 12 h. The reaction was quenched with water and extracted with DCM. The combined organic layers were washed with water and dried over MgSO₄. After removal of the solvent under vacuum, the residue was purified by silica gel chromatography (Hexane:DCM = 3:1) to afford compound **6** with two isomers (434 mg, 98%) as a white solid. The *cis*-/*trans*- vinyl isomers could not be separated by silica gel chromatography but did not affect the next reaction. Compound **6**: ¹H NMR (400 MHz, 298 K, CD₂Cl₂): δ ppm 8.68 (br, 0.66H), 8.62 and 8.61 (s, 2H), 8.13 and 8.12 (s, 1.25H), 7.15 (d, *J* = 12.7 Hz, 1.25H), 6.40 (d, *J* = 7.2 Hz, 0.75H), 6.23 (d, *J* = 12.8 Hz, 1.25H), 5.75 (d, *J* = 7.2 Hz, 0.75H),

4.20-4.14 (m, 4H), 3.87 (s, 2.2H), 3.77 (s, 3.8H). 1.91-1.82 (m, 4H), 1.68-1.54 (m, 4H), 1.04-0.99 (m, 4H), ^{13}C NMR (100 MHz, CD_2Cl_2): δ ppm 151.1, 150.2, 143.3, 134.0, 135.0, 133.4, 129.2, 129.1, 128.8, 126.7, 126.6, 126.5, 126.4, 126.2, 126.1, 123.4, 121.6, 121.5, 121.3, 118.3, 104.4, 104.4, 103.3, 103.2, 73.3, 73.3, 61.1, 56.8, 32.6, 32.5, 32.5, 19.5, 13.8, 13.7. Compound **6**: HRMS analysis (MALDI, 100%, m/z) calcd for $\text{C}_{28}\text{H}_{32}\text{Br}_2\text{O}_4$: 590.0662, found 590.0661 (error: -0.1 ppm).



Compound 6:

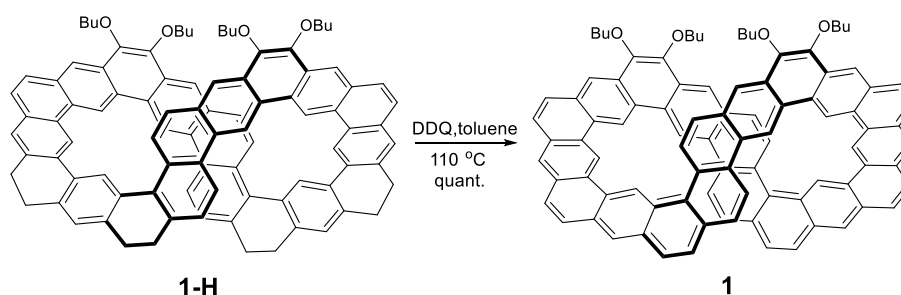
A mixture of **5** (300 mg, 0.56 mmol), **6** (332.6 mg, 0.56 mmol), NaHCO_3 (3.77 g, 44.9 mmol), THF (400 mL) and H_2O (20 mL) was carefully degassed before $\text{Pd}_2(\text{dba})_3$ (51.3 mg, 0.056 mmol) and $[(t\text{-Bu})_3\text{PH}]\text{BF}_4$ (65.2 mg, 0.225 mmol) were added. The mixture was stirred and heated at 80 °C for 3 days under N_2 atmosphere. The organic solvent was removed under reduced pressure, then H_2O and DCM were added. The organic layer was separated, dried over anhydrous sodium sulfate. The solvent was removed under vacuum and the residue was first purified by a short column chromatography (silica gel, DCM/Hexane = 1:1) to remove catalysts, and then further purified by preparative GPC using CHCl_3 at a rate of 14 mL/min to afford compound **7** (90 mg, 23%) as white solid. The NMR spectrum of **7** is complicated due to existence of multiple *cis*-/*trans*- vinyl isomers. Compound **7**: HRMS analysis (MALDI, 100%, m/z) calcd for $\text{C}_{100}\text{H}_{96}\text{O}_8$: 1424.7100, found 1424.7100 (error: 0.1 ppm).



Compound 1-H:

A mixture of **7** (90 mg, 0.063 mmol) and $\text{Bi}(\text{OTf})_3$ (8.3 mg, 0.013 mmol) in 1,2-dichloroethane (DCE, 40 mL) was heated at 60 °C for 2 hours under N_2 atmosphere. The organic solvent was removed under reduced pressure, then H_2O and DCM were added. The organic layer was separated, dried over anhydrous sodium sulfate. The solvent was removed and the residue was purified by silica gel chromatography (Hexane:DCM = 3:1) to afford pure compound **1-H** (65 mg, 79%) as a yellow solid. ^1H NMR (500 MHz,

298 K, CD₂Cl₂): δ ppm 9.67 (s, 2H), 9.61 (s, 2H), 8.89 (s, 2H), 8.55 (s, 2H), 8.45 (d, J = 8.2 Hz, 2H), 8.27 (s, 2H), 8.13 (s, 2H), 8.09 (d, J = 9.2 Hz, 2H), 7.88 (d, J = 8.9 Hz, 2H), 7.86 (s, 2H), 7.78 (d, J = 9.0 Hz, 2H), 7.50 (d, J = 9.2 Hz, 2H), 7.46 (d, J = 8.2 Hz, 2H), 7.42 (s, 2H), 4.32-4.27 (m, 2H), 4.23-4.17 (m, 4H), 4.09-4.04 (m, 2H), 3.25-2.90 (m, 16H), 1.94-1.88 (m, 4H), 1.81-1.75 (m, 4H), 1.65-1.58 (m, 4H), 1.51-1.44 (m, 4H), 1.01 (t, J = 7.4 Hz, 6H), 0.88 (t, J = 7.4 Hz, 6H). ¹³C NMR (125 MHz, 298 K, CD₂Cl₂): δ ppm 142.6, 139.6, 137.6, 137.4, 137.1, 134.1, 133.5, 133.3, 131.6, 130.9, 130.0, 129.0, 128.2, 127.6, 126.9, 126.7, 126.7, 126.5, 126.4, 125.8, 125.4, 123.1, 120.4, 119.7, 119.1, 118.8, 118.6, 73.1, 73.1, 32.6, 32.4, 29.7, 29.1, 28.9, 28.6, 19.5, 19.3, 13.8, 13.6. Compound **1-H**: HRMS analysis (MALDI, 100%, m/z) calcd for C₉₆H₈₀O₄: 1296.6051, found 1296.6047 (error: 0.3 ppm).



Compound 1:

A mixture of **1-H** (91 mg, 0.07 mmol) and DDQ (191 mg, 0.84 mmol) in dry toluene was heated at 110 °C for 24 hours under N₂ atmosphere. The organic solvent was removed under reduced pressure, then H₂O and DCM were added. The organic layer was separated, dried over anhydrous sodium sulfate. The solvent was removed and the crude was washed by methanol and acetonitrile to afford pure compound **1** as yellow solid (90 mg, quantitative). ¹H NMR (500 MHz, 373 K, toluene-*d*₈): δ ppm 10.11 (s, 2H), 10.09 (s, 2H), 9.83 (s, 2H), 9.26 (s, 2H), 8.71 (s, 2H), 8.37 (d, J = 9.1 Hz, 2H), 8.17 (s, 2H), 8.16 (s, 2H), 7.92 (d, J = 8.3 Hz, 2H), 7.87 (s, 2H), 7.84 (d, J = 9.0 Hz, 2H), 7.78 (d, J = 9.1 Hz, 2H), 7.75 (d, J = 9.2 Hz, 2H), 7.84 (d, J = 9.0 Hz, 2H), 7.68 (d, J = 9.0 Hz, 8H), 7.46 (d, J = 8.2 Hz, 2H), 7.28 (d, J = 8.4 Hz, 2H), 7.25 (d, J = 9.1 Hz, 2H), 6.91 (d, J = 8.3 Hz, 2H), 4.44-4.40 (m, 2H), 4.38-4.33 (m, 2H), 4.30-4.26 (m, 2H), 4.20-4.16 (m, 2H), 2.03-1.97 (m, 2H), 1.87-1.81 (m, 2H), 1.72-1.65 (m, 2H), 1.57-1.50 (m, 2H), 1.07 (t, J = 7.4 Hz, 6H), 0.93 (t, J = 7.4 Hz, 6H). ¹³C NMR (125 MHz, THF-*d*₈): δ ppm 143.7, 143.6, 132.1, 131.9, 131.6, 131.3, 131.4, 130.3, 130.1, 130.0, 129.6, 129.2, 129.2, 129.0, 129.0, 128.8, 128.4, 127.9, 127.5, 127.5, 127.4, 127.2, 127.1, 126.9, 126.4, 126.0, 126.0, 125.6, 124.8, 121.7, 121.2, 121.0, 120.5, 120.0, 73.6, 33.4, 33.2, 30.4, 25.4, 25.3, 25.1, 25.0, 24.8, 20.2, 20.1, 14.2, 14.1. Compound **1**: HRMS analysis (MALDI, 100%, m/z) calcd for C₉₆H₇₂O₄: 1288.5425, found 1288.5415 (error: -0.8 ppm).

2. Additional spectra for characterizations of 1-H and 1

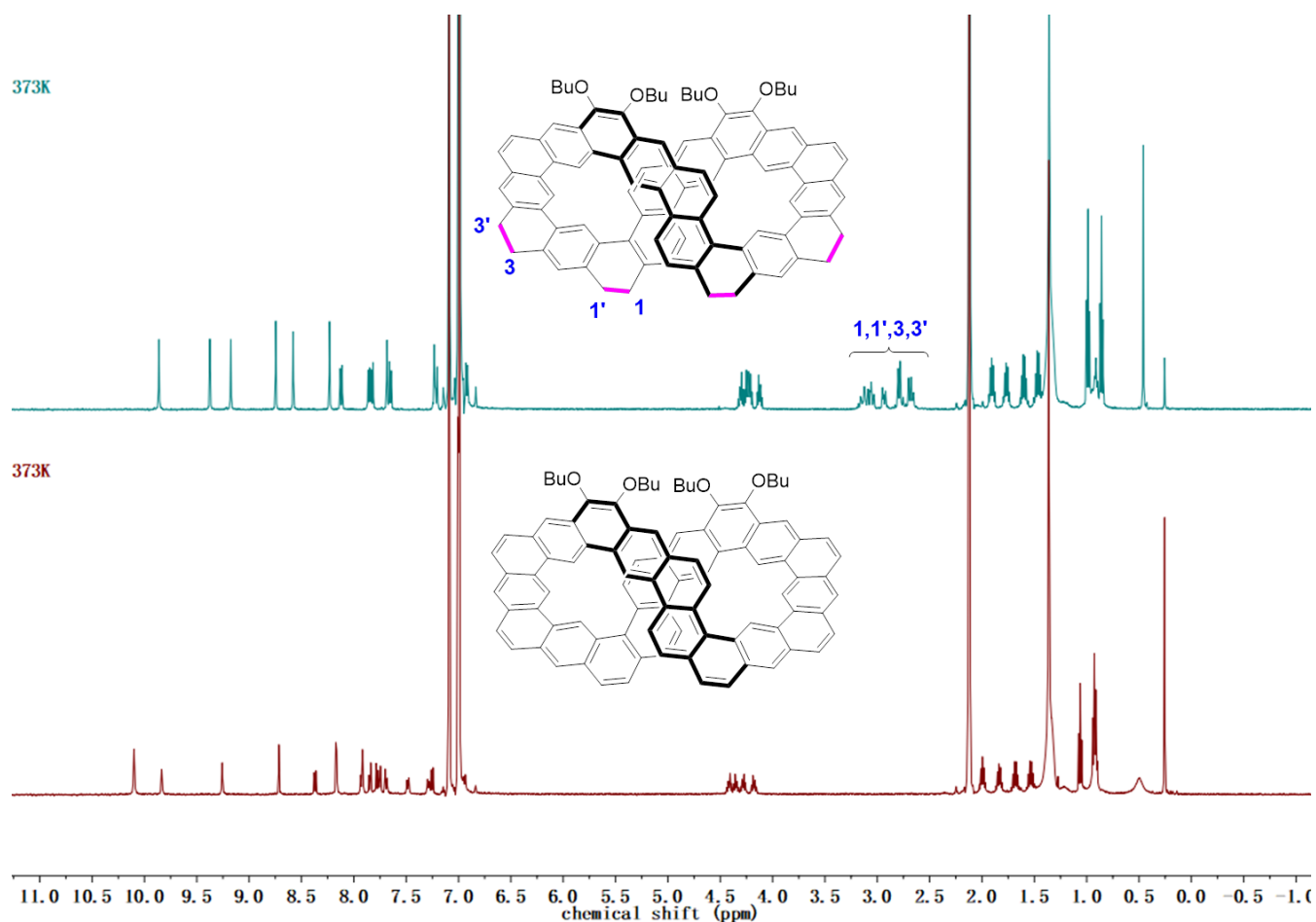


Figure S1. The full range ¹H NMR spectra of **1-H** (top) and **1** (bottom) in toluene-*d*₈ (500 MHz) at 373 K. The multiple split peaks of 1, 1', 3, 3' on the saturated *sp*³ carbons at 2.1~3.2 ppm in **1-H** disappear after dehydrogenation to afford **1**.

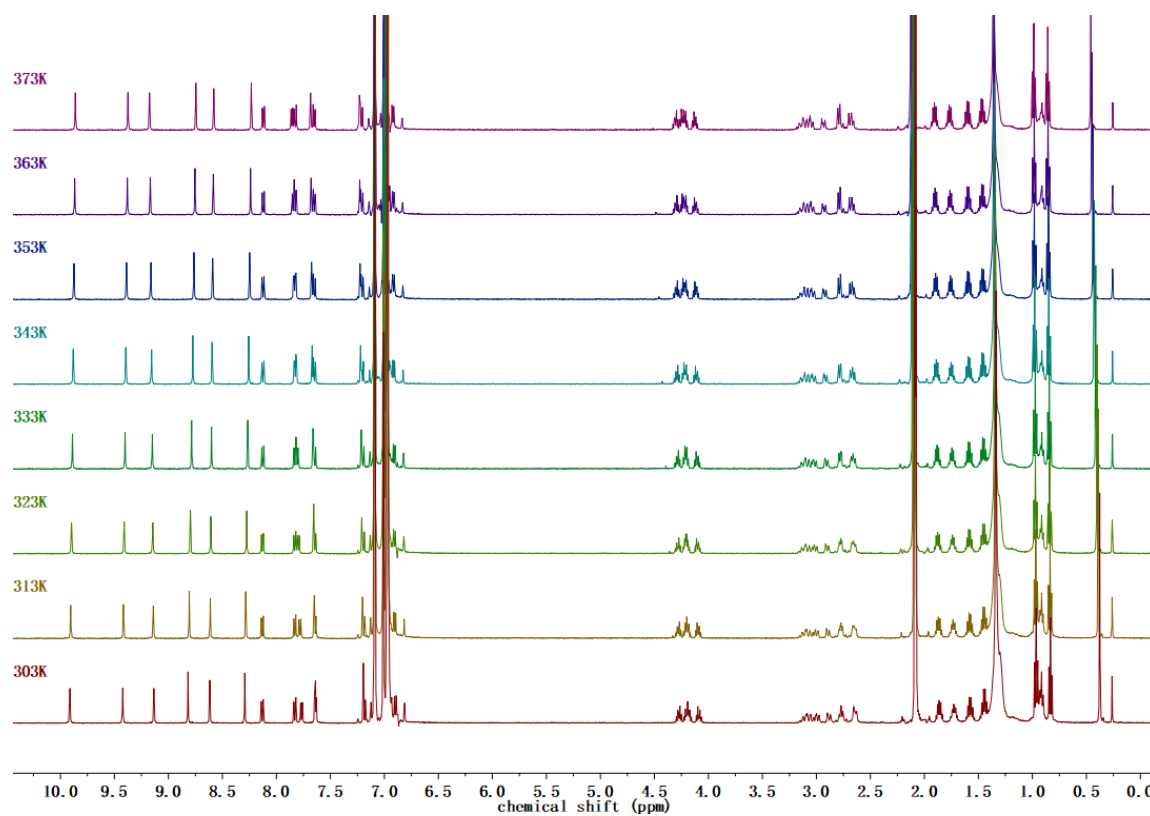


Figure S2. VT ^1H NMR spectra of **1-H** in toluene- d_8 (500 MHz).

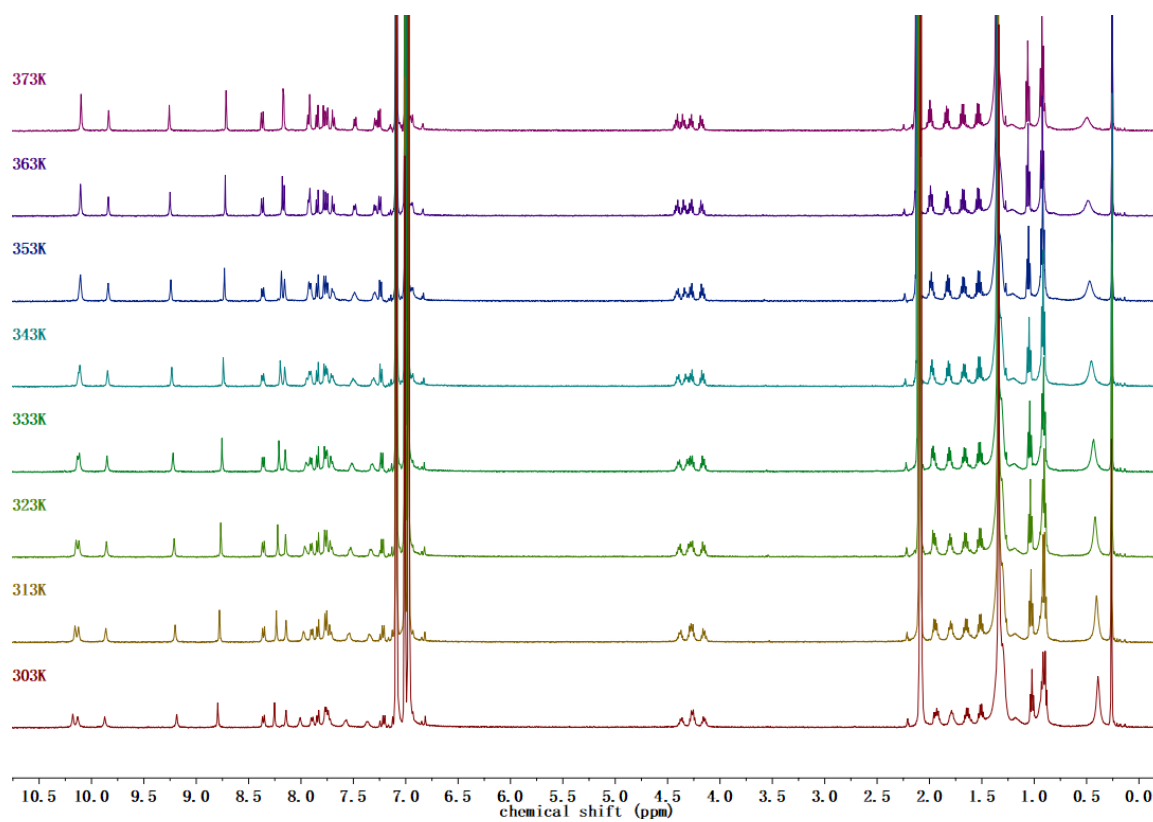


Figure S3. VT ^1H NMR spectra of **1** in toluene- d_8 (500 MHz).

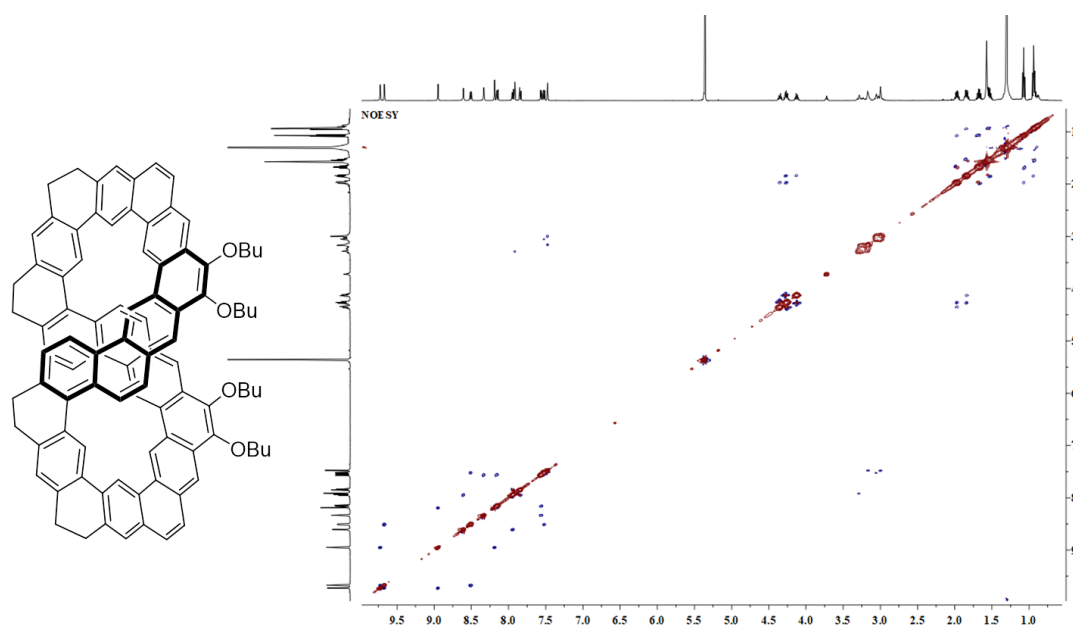


Figure S4. 2D NOESY NMR spectrum of **1-H** in CD_2Cl_2 (500 MHz) at 298 K showing correlations.

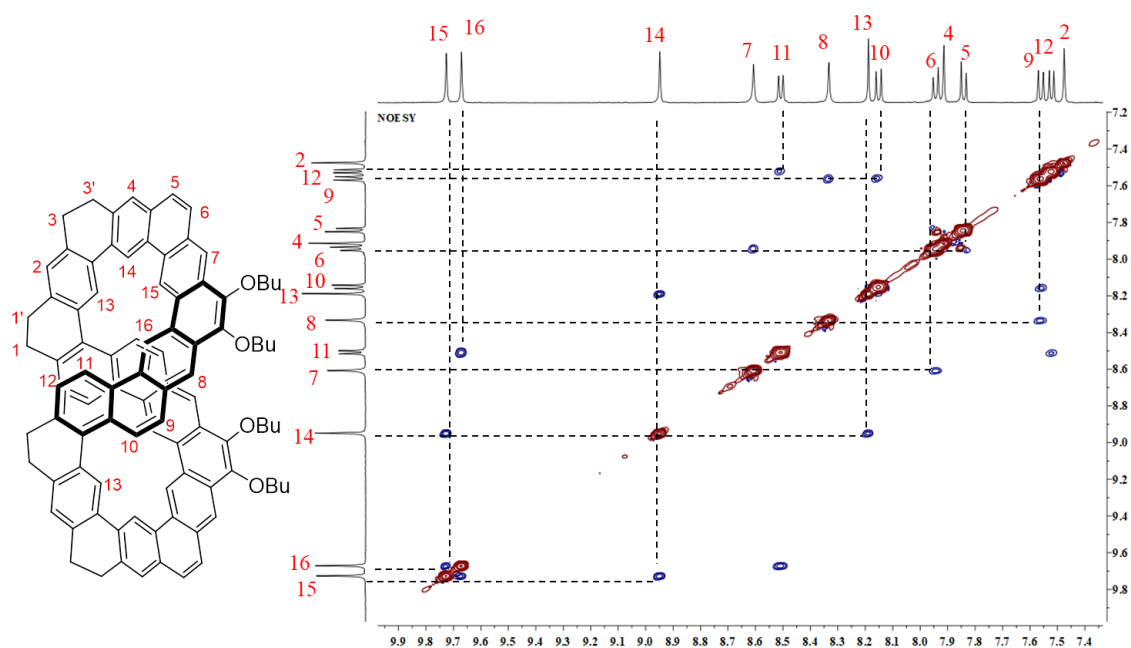


Figure S5. Partial 2D NOESY NMR spectrum of **1-H** in CD_2Cl_2 (500 MHz) at 298 K showing aromatic/aromatic correlations.

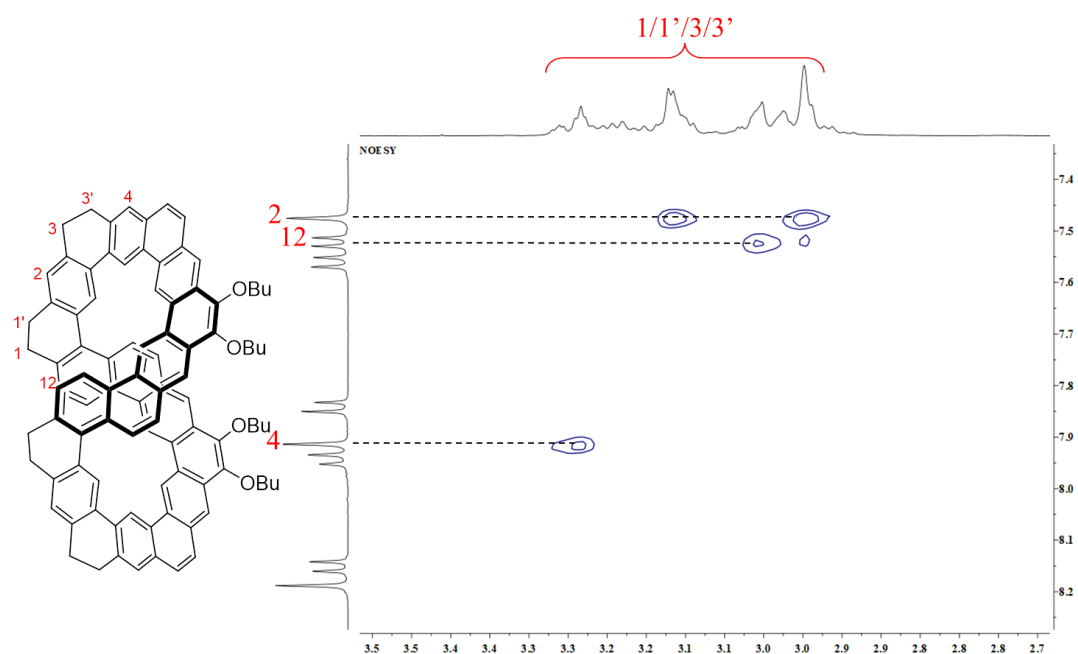


Figure S6. Partial 2D NOESY NMR spectrum of **1-H** in CD₂Cl₂ (500 MHz) at 298 K showing aromatic/aliphatic correlations.

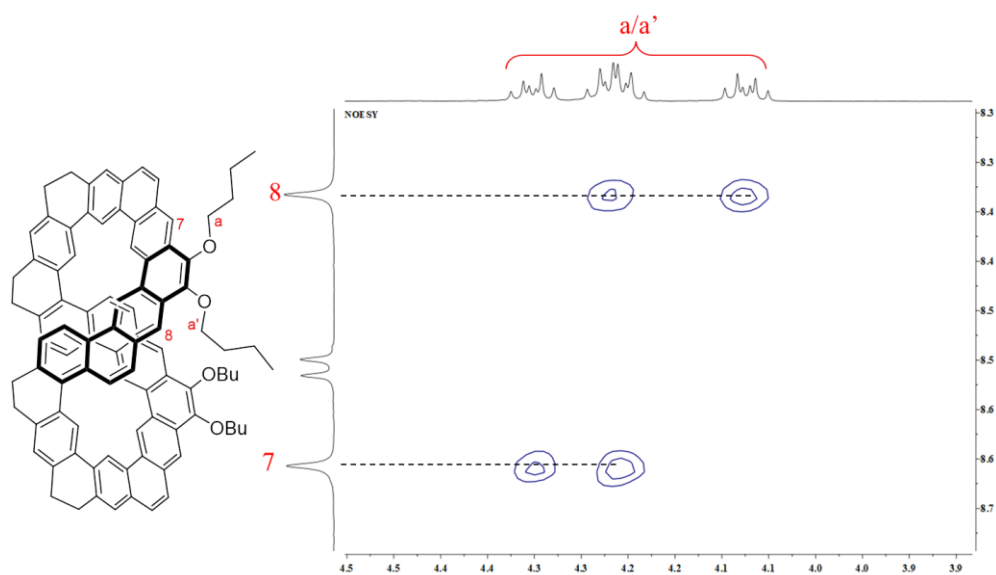


Figure S7. Partial 2D NOESY NMR spectrum of **1-H** in CD₂Cl₂ (500 MHz) at 298 K showing aromatic/aliphatic correlations.

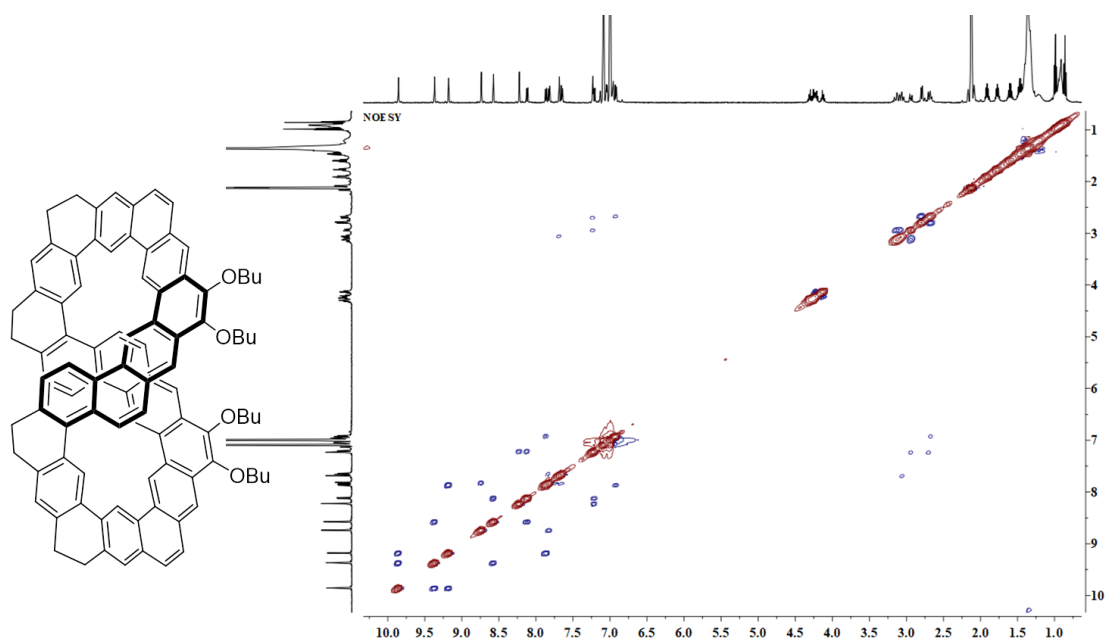


Figure S8. 2D NOESY NMR spectrum of **1-H** in toluene- d_8 (500 MHz) at 373 K showing correlations.

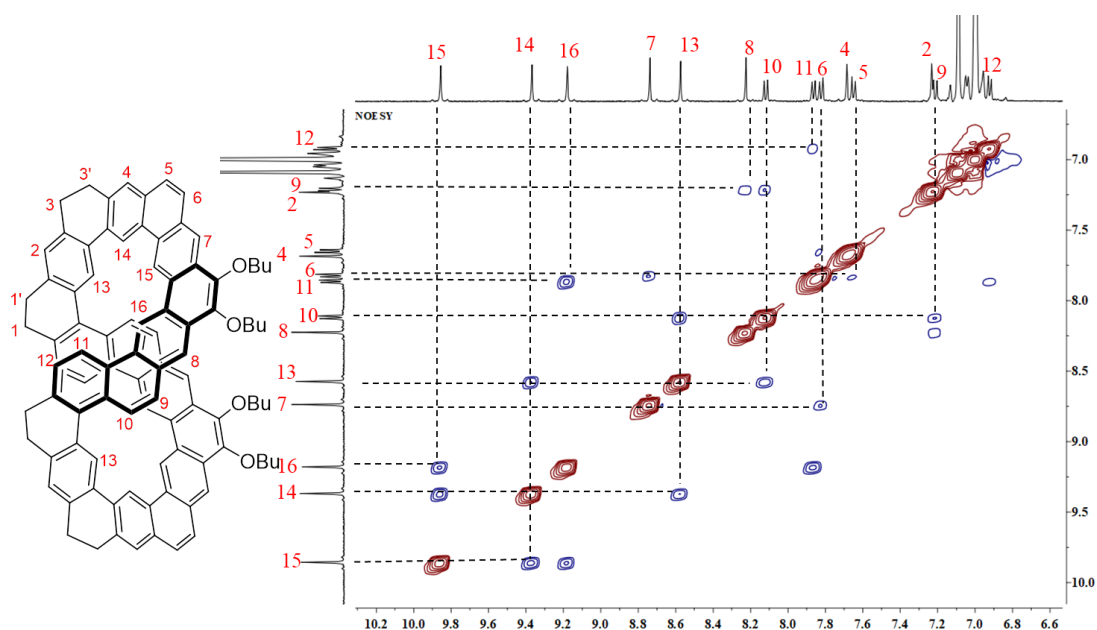


Figure S9. Partial 2D NOESY NMR spectrum of **1-H** in toluene- d_8 (500 MHz) at 373 K showing aromatic/aromatic correlations.

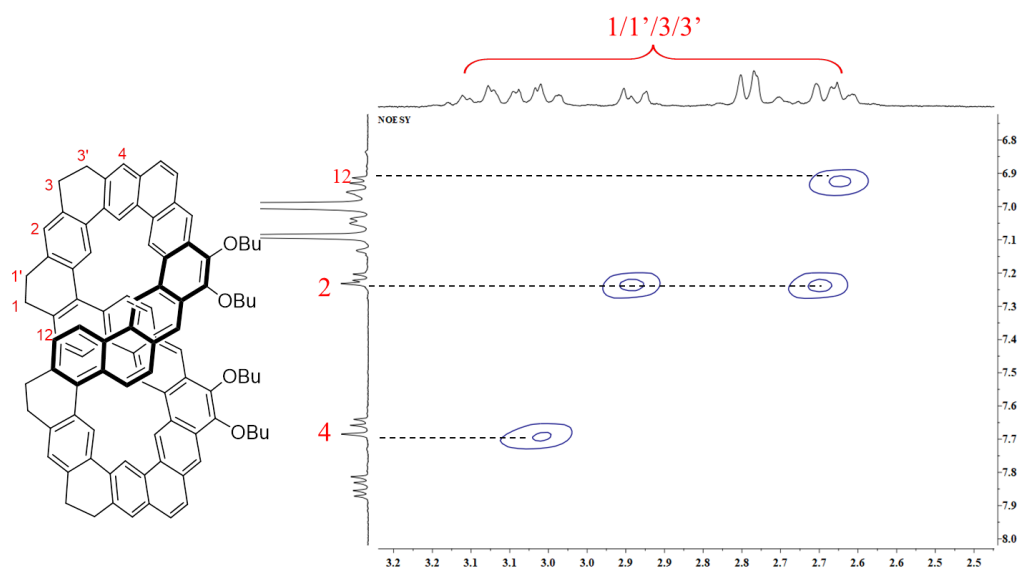


Figure S10. Partial 2D NOESY NMR spectrum of **1-H** in toluene- d_8 (500 MHz) at 373 K showing aromatic/aliphatic correlations.

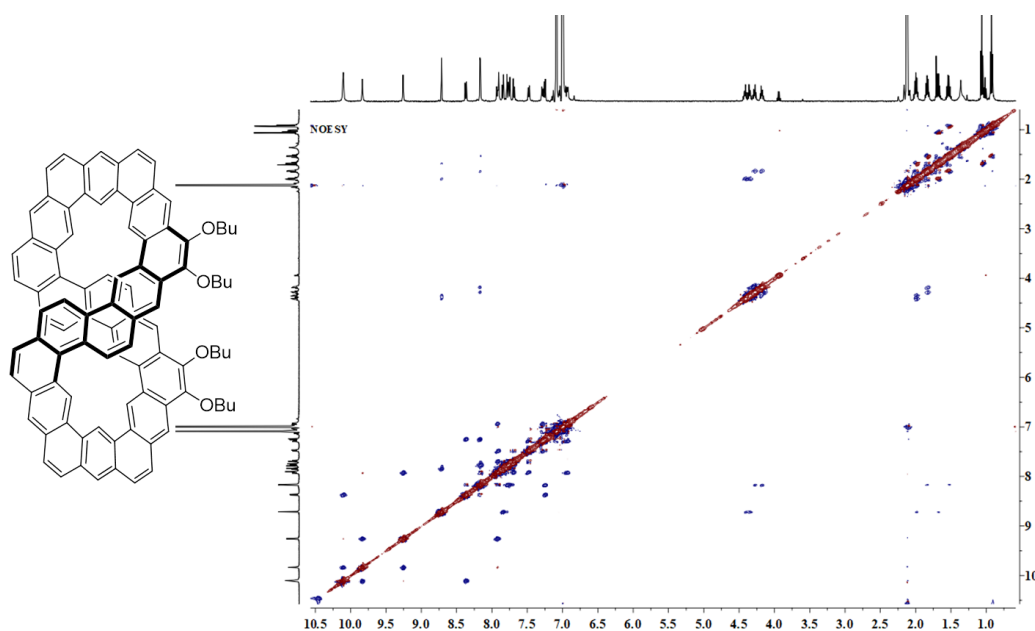


Figure S11. 2D NOESY NMR spectrum of **1** in toluene- d_8 (500 MHz) at 373 K showing correlations.

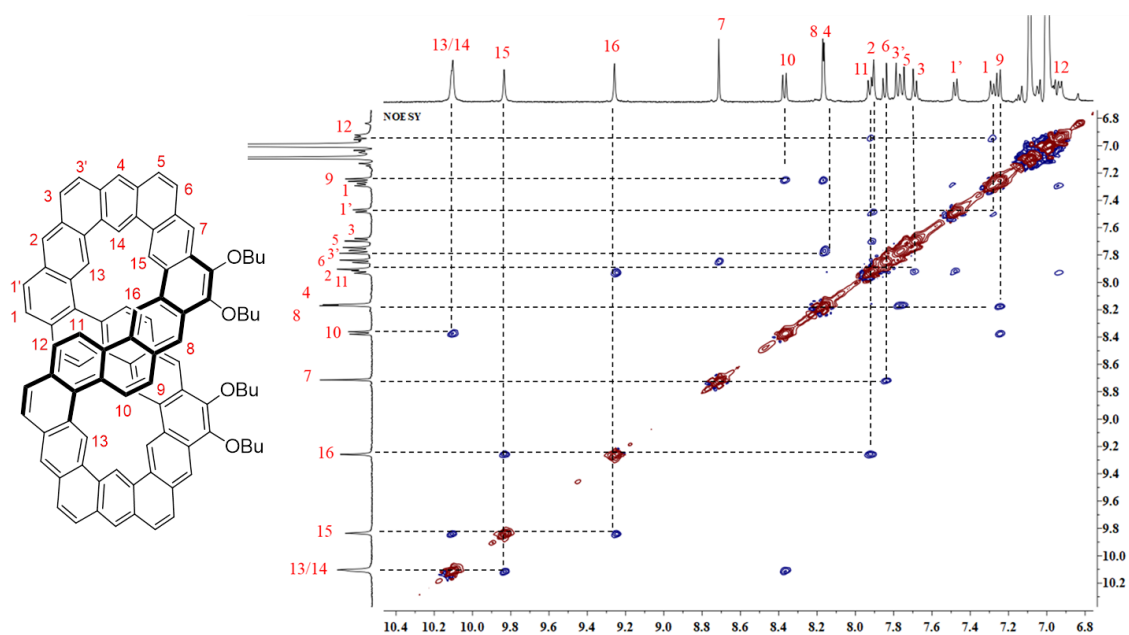


Figure S12. Partial 2D NOESY NMR spectrum of **1** in toluene- d_8 (500 MHz) at 373 K showing aromatic/aromatic correlations.

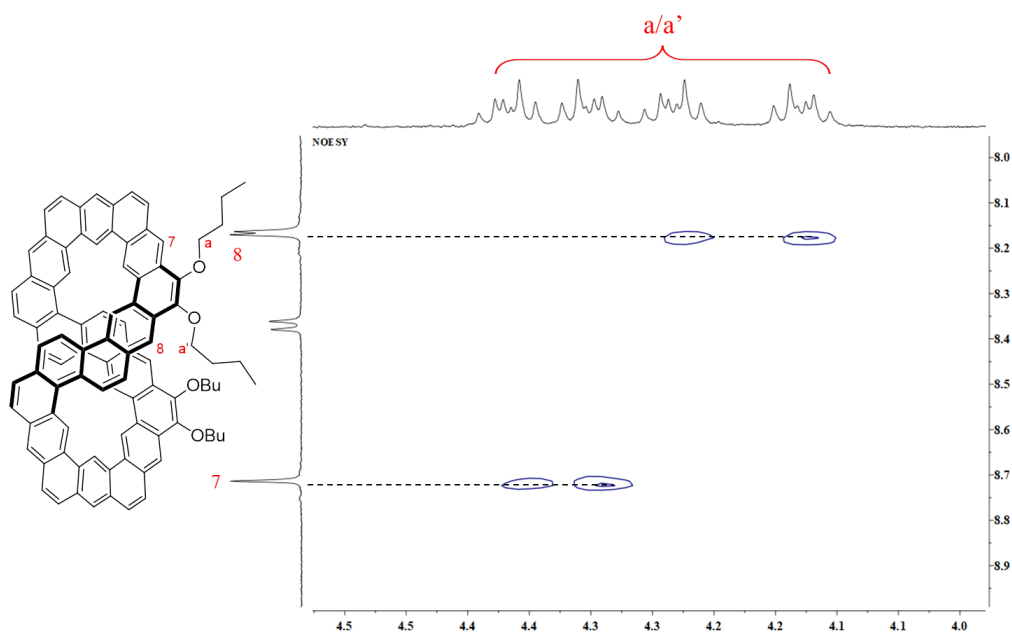


Figure S13. Partial 2D NOESY NMR spectrum of **1** in toluene- d_8 (500 MHz) at 373 K showing aromatic/aliphatic correlations.

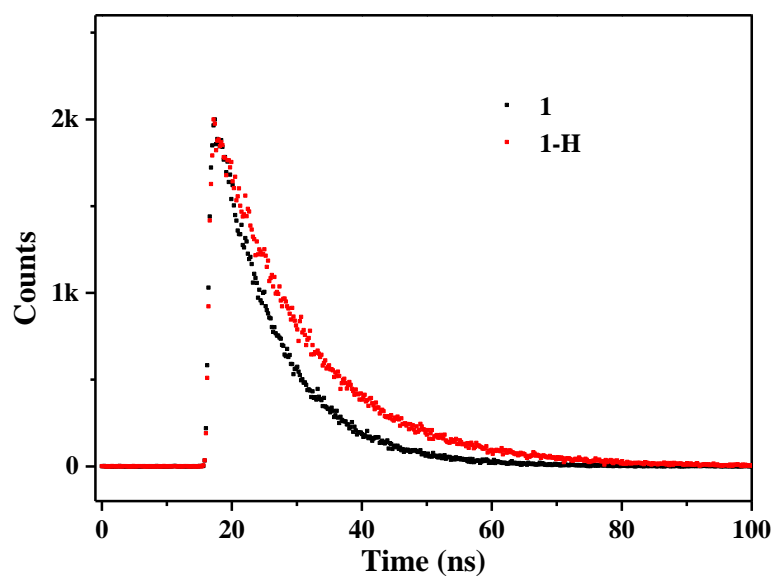


Figure S14. Time-resolved fluorescence decay profiles of **1-H** ($\lambda_{\text{exc}} = 340$ nm, $\lambda_{\text{probe}} = 460$ nm) and **1** ($\lambda_{\text{exc}} = 365$ nm, $\lambda_{\text{probe}} = 473$ nm).

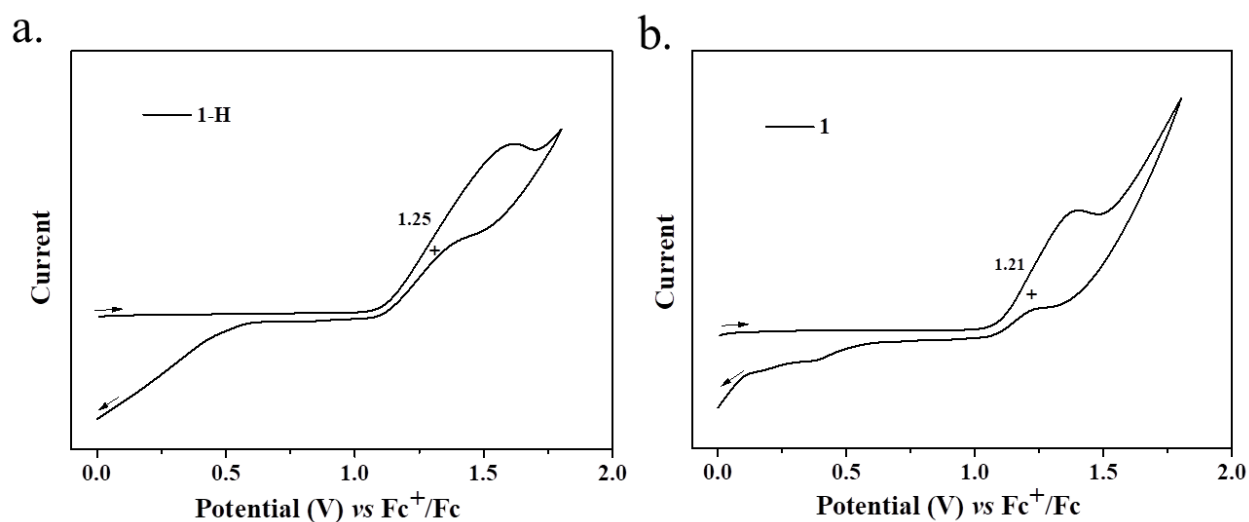
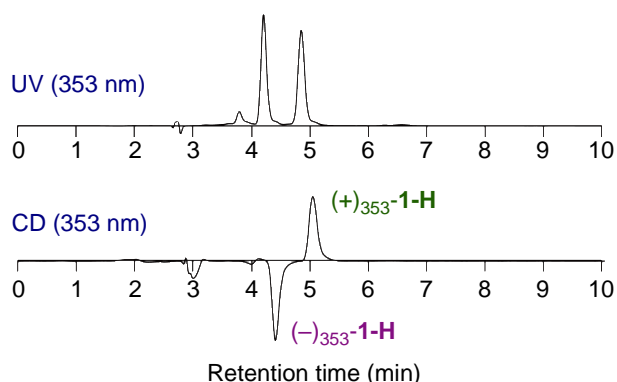


Figure S15. Cyclic voltammogram of (a) **1-H** and (b) **1** measured in chlorobenzene with TBAPF₆ as supporting electrolyte. The downward curves at the end of potential flow may arise from the deposition and coupling of the oxidized species on the surface of work electrode after the first scan.

3. Chiral resolution and chiroptic properties for 1-H and 1

3.1 Chiral HPLC analysis and optical resolution of 1-H and 1

a. Analytical HPLC of 1-H



b. Analytical HPLC of 1

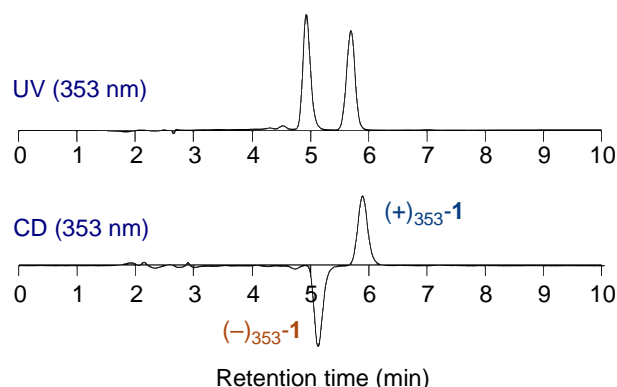
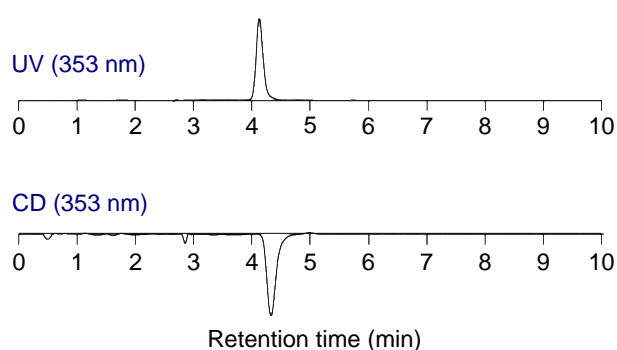
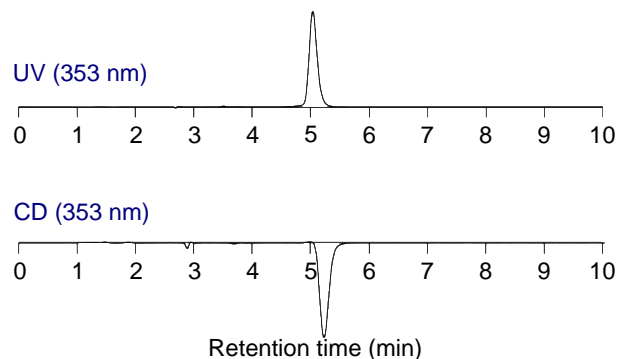


Figure S16. Chiral HPLC analyses of racemic specimens. [Conditions] Column: COSMOSIL Cholester 4.6 ϕ \times 250 mm, Column oven: 40 $^{\circ}$ C, UV detector: 353 nm, CD detector: 353 nm, eluent: CH₂Cl₂/MeOH (7:3 v/v), flow rate: 1.0 mL min⁻¹. (a) Chromatograms of (*rac*)-1-H. (b) Chromatograms of (*rac*)-1.

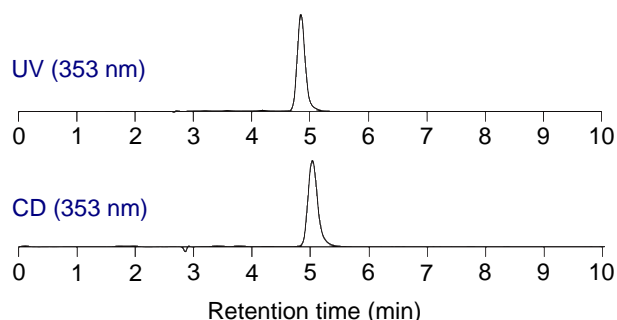
a. Analytical HPLC of (-)₃₅₃-1-H



c. Analytical HPLC of (-)₃₅₃-1



b. Analytical HPLC of (+)₃₅₃-1-H



d. Analytical HPLC of (+)₃₅₃-1

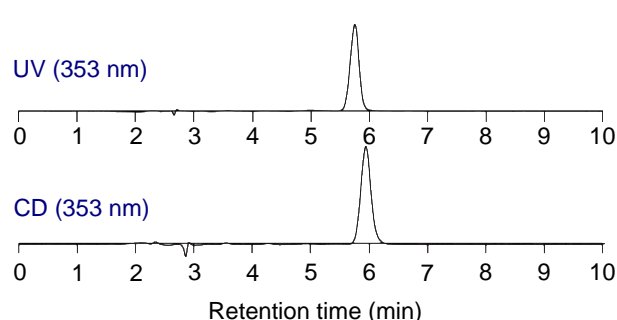
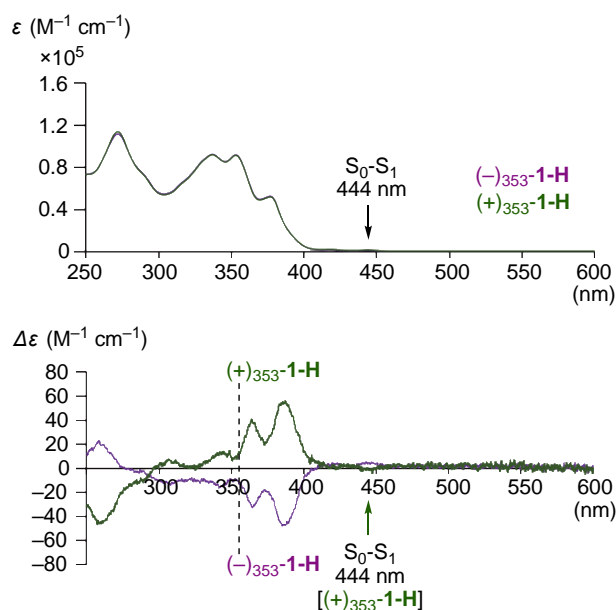


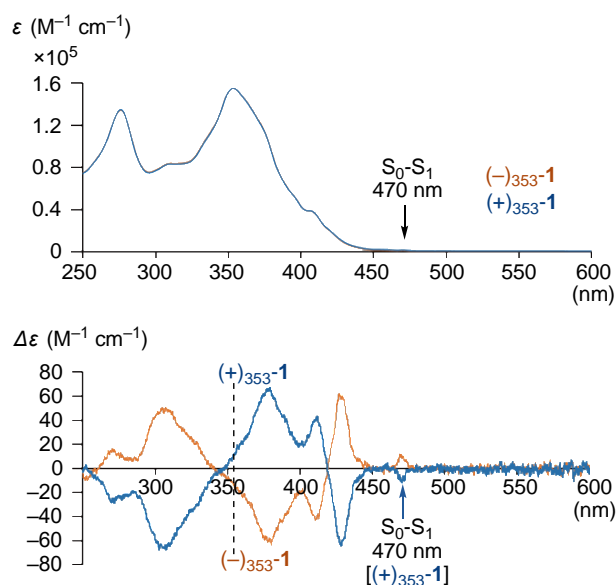
Figure S17. Chiral HPLC analyses of separated enantiomers. [Conditions] Column: COSMOSIL Cholester 4.6 ϕ \times 250 mm, Column oven: 40 $^{\circ}$ C, UV detector: 353 nm, CD detector: 353 nm, eluent: CH₂Cl₂/MeOH (7:3 v/v), flow rate: 1.0 mL min⁻¹. (a) Chromatograms of (-)₃₅₃-1-H. (b) Chromatograms of (+)₃₅₃-1-H. (c) Chromatograms of (-)₃₅₃-1. (d) Chromatograms of (+)₃₅₃-1.

3.2 UV-vis and CD spectra of 1-H and 1

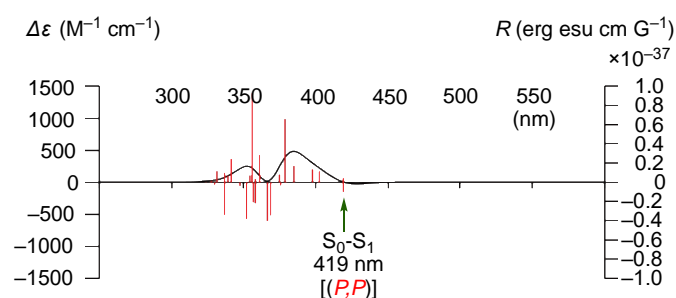
a. UV-vis and CD spectra of 1-H



b. UV-vis and CD spectra of 1



c. Simulated CD spectrum of (P,P)-1-H



d. Simulated CD spectrum of (P,P)-1

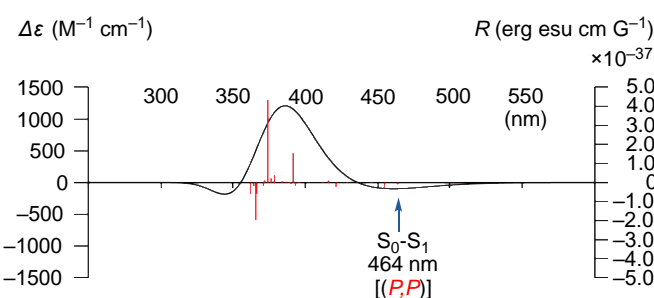
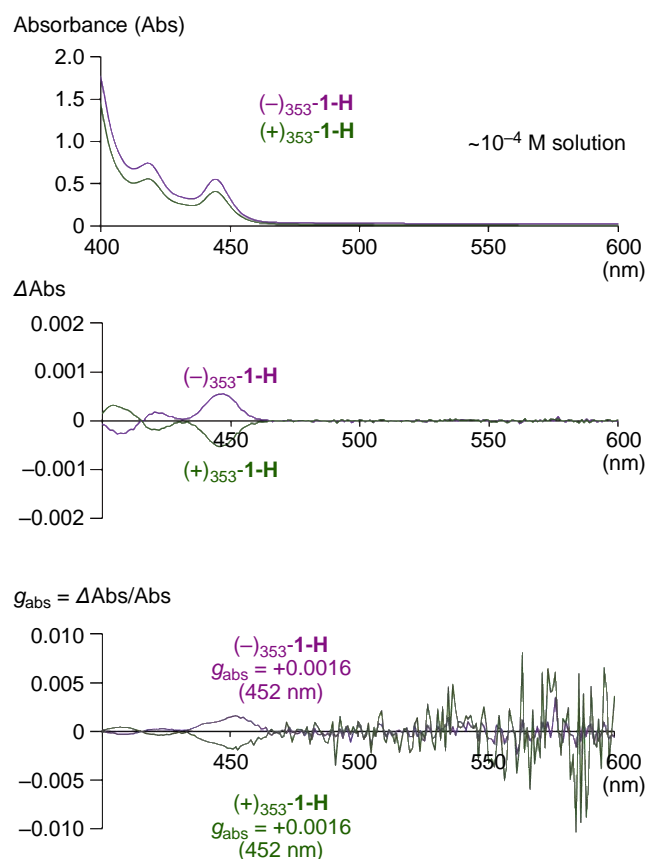


Figure S18. UV-vis absorption and CD spectra in DCM at 298 K and simulation of CD spectra. (a) UV-vis and CD spectra of $(-)\text{353-1-H}$ ($1.06 \times 10^{-5} \text{ M}$) and $(+)\text{353-1-H}$ ($7.73 \times 10^{-6} \text{ M}$). (b) UV-vis and CD spectra of $(-)\text{353-1}$ ($8.99 \times 10^{-6} \text{ M}$) and $(+)\text{353-1}$ ($8.70 \times 10^{-6} \text{ M}$). (c) Simulated CD spectra of (P,P)-1-H. The sign of S_0-S_1 transition was minus, which led us to assign $(+)\text{353-1-H}$ as (P,P)-isomer. (d) Simulated CD spectra of (P,P)-1. The sign of S_0-S_1 transition was minus, which led us to assign $(+)\text{353-1-H}$ as (P,P)-isomer.

3.3 Measurement of g_{abs} values of 1-H and 1

a. UV-vis and CD spectra of 1-H



b. UV-vis and CD spectra of 1

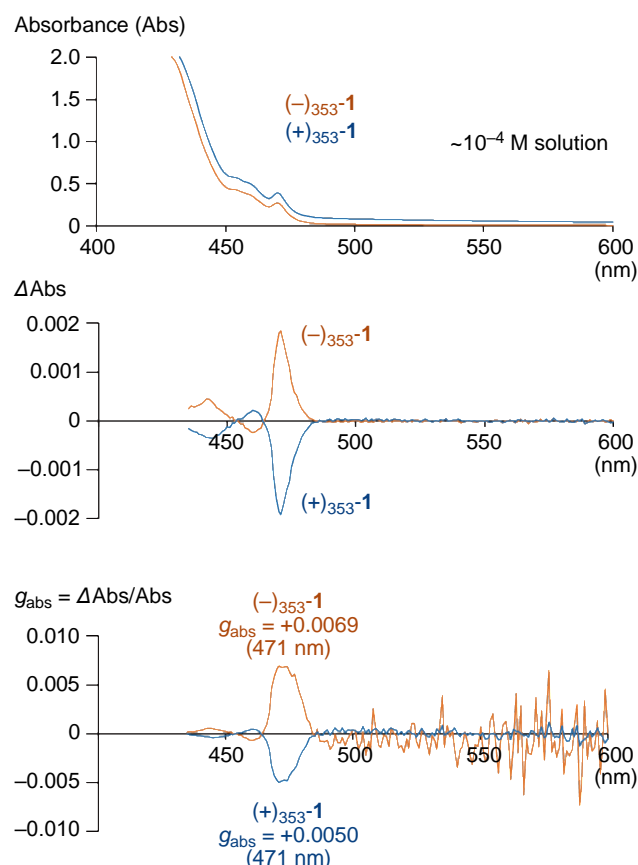


Figure S19. UV-vis absorption and CD spectra in DCM at 298 K. (a) UV-vis and CD spectra of $(-)\text{353-1-H}$ ($\sim 10^{-4}$ M) and $(+)\text{353-1-H}$ ($\sim 10^{-4}$ M). (b) UV-vis and CD spectra of $(-)\text{353-1}$ ($\sim 10^{-4}$ M) and $(+)\text{353-1}$ ($\sim 10^{-4}$ M).

4. DFT calculations for 1-H and 1

Density functional theory (DFT) calculations were performed with the Gaussian09 program suite³ with Becke's three-parameter hybrid exchange functionals and the Lee-Yang-Parr correlation functional (B3LYP) employing the 6-31G(d,p) basis set for all atoms.⁴ Time-dependent DFT (TD-DFT) calculations were performed at the (U)B3LYP/6-31G(d,p) level of theory under vacuum. NICS values were calculated (B3LYP/6-31G(d,p)) using the standard GIAO procedure (NMR pop=NCSall).⁵ ACID plot (B3LYP/6-31G(d,p)) was calculated by using the method developed by Herges based on the optimized ground-state geometries.⁶ The iso-chemical shielding surface (ICSS)⁷ calculations were carried out and the VMD programme⁸ was used to plot ICSS cube files.

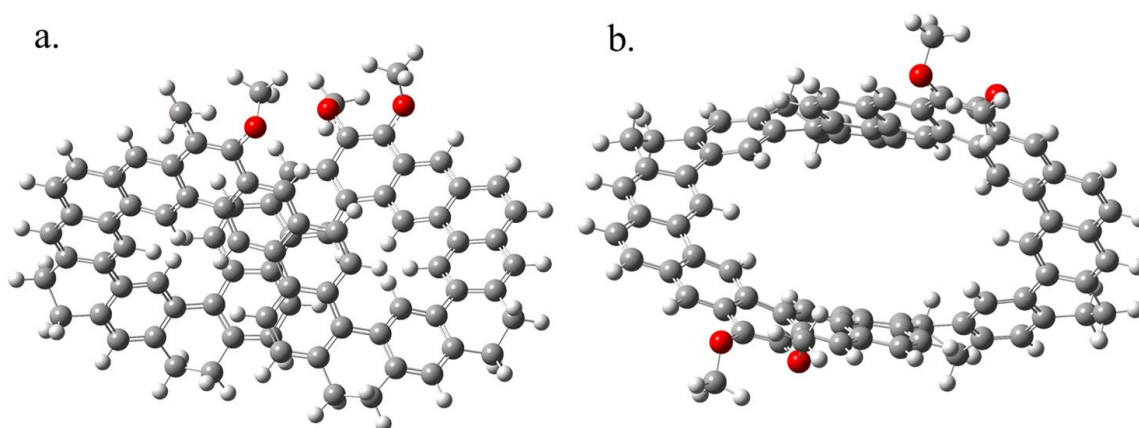


Figure S20. The optimized geometry of **1-H** (left for top-view and right for side-view) at B3LYP/6-31G(d,p) level of theory showing a twisted backbone. The *n*-butoxy substituents are replaced by methoxy groups during the calculations.

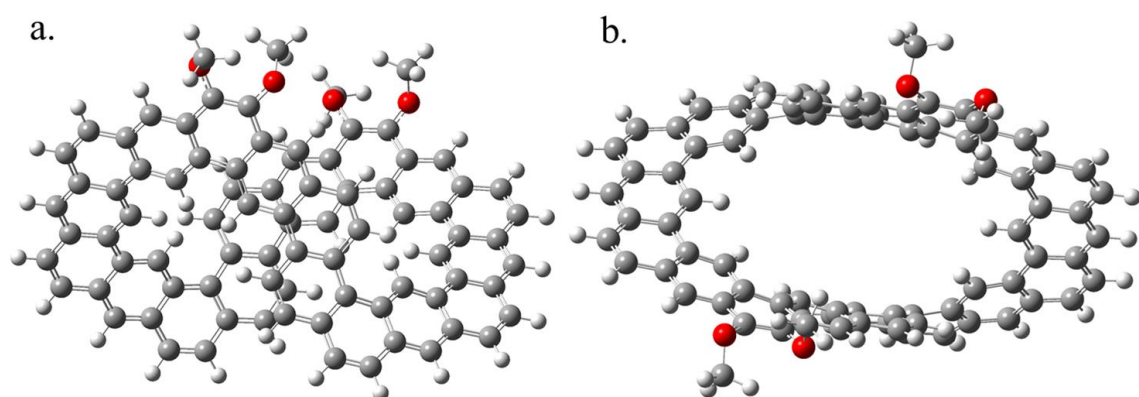
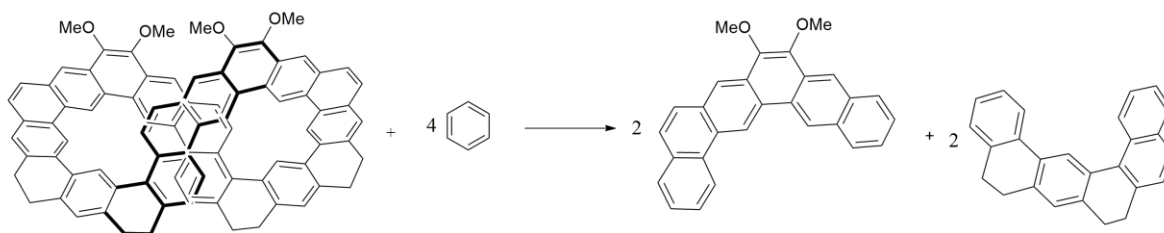


Figure S21. The optimized geometry of **1** (left for top-view and right for side-view) at B3LYP/6-31G(d,p) level of theory showing a twisted backbone. The *n*-butoxy substituents are replaced by methoxy groups during the calculations.

a. Method for 1-H



b. Method for 1

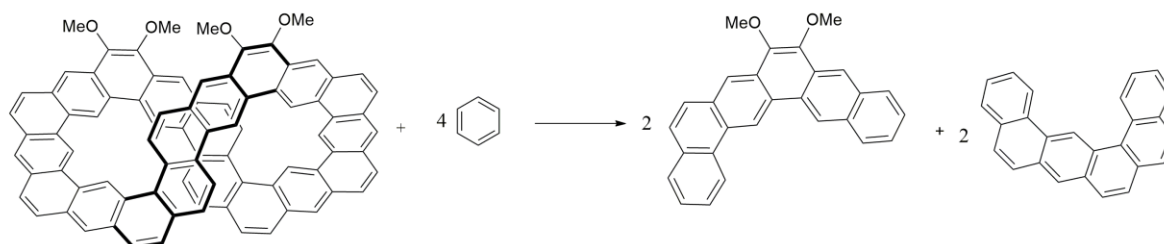


Figure S22. The hypothetical homodesmotic reactions applied in the calculation of strain energy for **1-H** and **1**. The *n*-butoxy substituents are replaced by methoxy groups during the calculations.

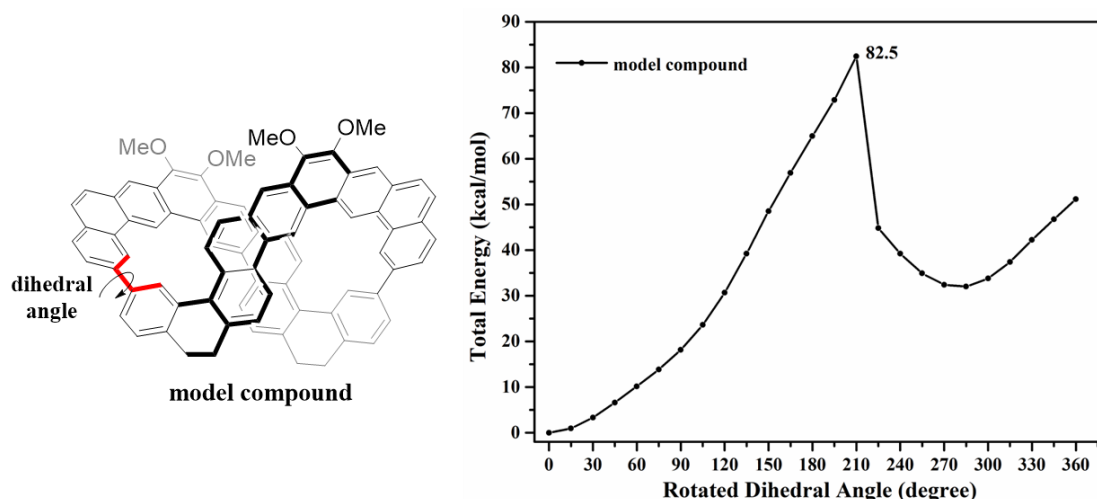


Figure S23. Calculated total energies of an analogue of **1** with a partially fused structure (named as **model compound**) by rotating the dihedral angle (highlight with red). The relatively high energy (up to 82.5 kcal/mol when rotated 210°) indicate a high racemization barrier.

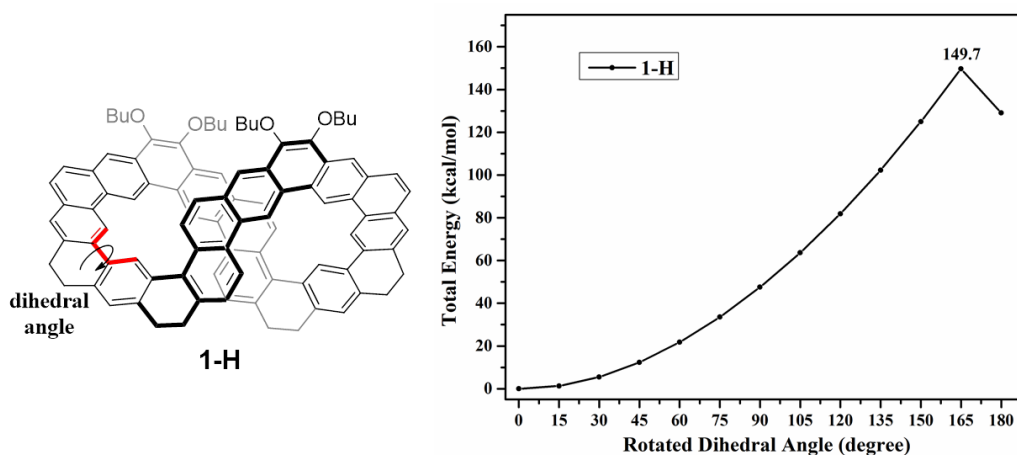


Figure S24. Calculated total energies of **1-H** by rotating the dihedral angle (highlight with red). The very high energy (up to 149.7 kcal/mol when rotated 165°) indicates a very rigid configuration with persistent chirality.

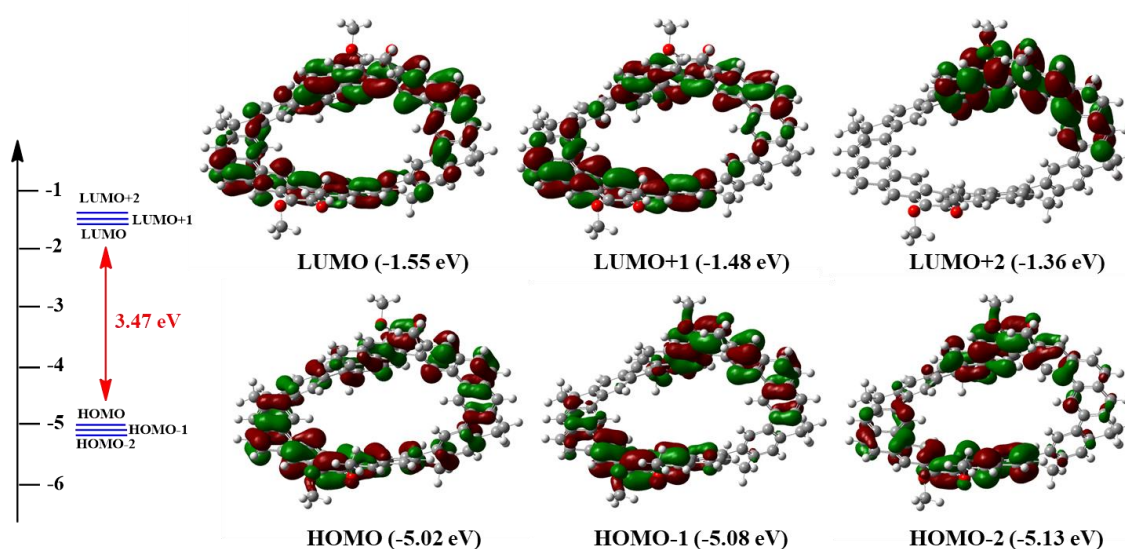


Figure S25. Frontier molecular orbital profiles and energy diagram of **1-H** obtained by B3LYP/6-31G(d,p) level calculation.

Table S1. Selected TD-DFT (B3LYP/6-31G(d,p)) calculated energies, oscillator strength and compositions of major electronic transitions of **1-H**.

Wavelength (nm)	Osc.Strength (f)	Major contributions
420.48495	0.006	H-1->LUMO (31%), HOMO->L+1 (38%)
419.53167	0.0312	H-2->LUMO (16%), H-1->L+1 (26%), HOMO->LUMO (29%)
403.90993	0.0039	H-3->LUMO (37%), H-2->L+1 (26%), HOMO->L+1 (24%)
400.73756	0.1512	H-3->L+1 (10%), H-2->LUMO (20%), HOMO->LUMO (46%)
386.2075	0.0351	H-1->LUMO (39%), HOMO->L+1 (16%)
381.54852	0.0649	H-3->LUMO (11%), H-1->L+1 (16%), H-1->L+3 (15%), HOMO->LUMO (10%), HOMO->L+3 (21%)
380.15635	0.0826	H-3->LUMO (13%), H-1->L+1 (23%), H-1->L+2 (10%), HOMO->L+2 (14%)
376.96623	0.5091	H-2->LUMO (52%), H-1->L+1 (10%)
374.23542	0.1987	H-3->LUMO (24%), H-2->L+1 (41%)
372.70544	0.1765	H-4->L+1 (11%), H-3->L+1 (26%), H-1->L+1 (11%), HOMO->L+2 (16%)
370.14627	0.2217	H-4->LUMO (17%), H-2->L+1 (10%), HOMO->L+2 (13%), HOMO->L+3 (18%)
366.28613	1.0538	H-4->L+1 (22%), H-3->L+1 (39%)
364.38075	0.4371	H-4->LUMO (23%), H-1->L+2 (40%), HOMO->L+2 (13%)
362.97264	0.2613	H-4->LUMO (27%), H-4->L+1 (17%), H-1->L+3 (33%), HOMO->L+3 (11%)
361.2593	0.0581	H-4->LUMO (16%), H-2->L+2 (27%), H-1->L+2 (14%)
359.38489	0.0758	H-4->L+1 (12%), H-3->L+3 (13%), H-2->L+3 (29%), H-1->L+3 (10%)
357.16934	0.0282	H-3->L+2 (55%), H-2->L+2 (21%)
356.30713	0.0141	H-4->L+2 (16%), H-3->L+3 (24%)
354.74733	0.0138	H-4->L+3 (10%), H-3->L+3 (32%), H-2->L+3 (21%)
354.26079	0.0513	H-4->L+1 (17%), H-4->L+2 (20%), H-4->L+3 (13%), H-3->L+3 (11%), H-1->L+4 (16%)
346.64409	0.0266	HOMO->L+4 (80%)
341.75196	0.17	H-5->LUMO (11%), H-4->L+2 (11%), H-4->L+3 (11%), H-3->L+4 (11%), H-1->L+4 (40%)
339.02325	0.0018	H-5->LUMO (36%), HOMO->L+5 (43%)
337.68437	0.0631	H-4->L+2 (19%), H-4->L+3 (17%), H-2->L+4 (39%)
336.43817	0.2532	H-3->L+4 (50%), HOMO->L+5 (11%)
331.46422	0.0209	H-5->LUMO (38%), H-1->L+4 (22%), HOMO->L+5 (18%)
329.06256	0.0029	H-5->L+1 (51%), HOMO->L+7 (13%)
328.18283	0.0577	H-5->L+1 (27%), H-2->L+4 (31%), H-1->L+5 (19%)
326.42882	0.0013	H-3->L+4 (18%), H-2->L+5 (11%), HOMO->L+6 (17%)
322.69903	0.0264	H-2->L+5 (20%), H-1->L+5 (29%)
321.9198	0.0174	H-2->L+5 (35%), H-1->L+5 (21%)
320.55482	0.0274	H-3->L+5 (79%)
319.97572	0.0639	H-5->L+2 (11%), H-5->L+3 (12%), H-4->L+5 (16%), HOMO->L+7 (13%)
319.62927	0.0659	H-6->LUMO (11%), H-5->L+2 (13%), H-5->L+3 (12%), H-2->L+6 (11%), HOMO->L+6 (15%)
317.97341	0.0125	H-4->L+4 (86%)
316.76297	0.0923	H-6->LUMO (26%), H-5->L+2 (28%)
316.02822	0.0162	H-6->L+1 (26%), H-5->L+3 (34%)
315.6983	0.0192	H-4->L+5 (19%), H-2->L+7 (20%), H-1->L+6 (29%)
310.76848	0.1126	H-6->LUMO (36%), H-2->L+6 (14%)
308.98717	0.0161	H-8->LUMO (13%), H-1->L+7 (13%), HOMO->L+6 (36%)
308.61031	0.0159	H-7->LUMO (13%), H-6->L+1 (35%)
306.80044	0.0011	H-4->L+5 (23%), H-1->L+6 (18%), HOMO->L+7 (34%)
303.79347	0.0145	H-3->L+6 (15%), H-2->L+7 (23%), H-1->L+6 (10%)
303.27331	0.0492	H-3->L+6 (13%), H-2->L+6 (30%), H-1->L+7 (12%)
303.21397	0.1112	H-6->L+2 (36%), H-6->L+3 (21%), H-1->L+7 (18%)
302.60713	0.068	H-6->L+3 (21%), H-2->L+7 (17%)
301.39338	0.028	H-6->L+3 (22%), H-3->L+6 (21%), H-2->L+7 (16%), H-1->L+6 (12%)

300.87409	0.0545	H-8->LUMO (17%), H-7->L+1 (17%), H-3->L+7 (15%)
300.55317	0.0624	H-7->L+1 (10%), H-3->L+7 (45%), H-2->L+6 (10%)
299.89888	0.018	H-8->L+1 (25%), H-7->LUMO (20%)

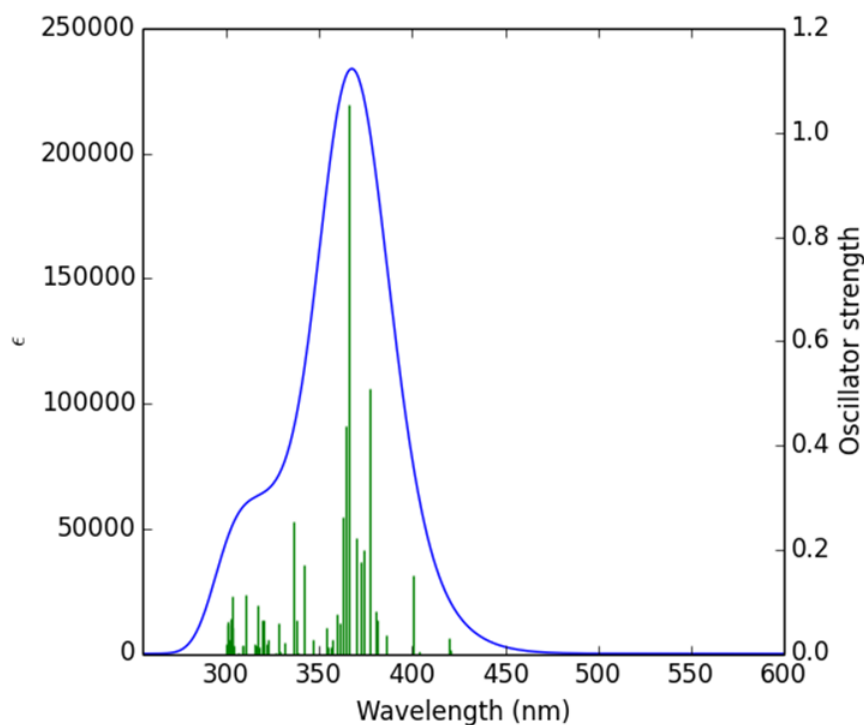


Figure S26. Calculated electronic absorption spectrum (B3LYP/6-31G(d,p)) of **1-H**.

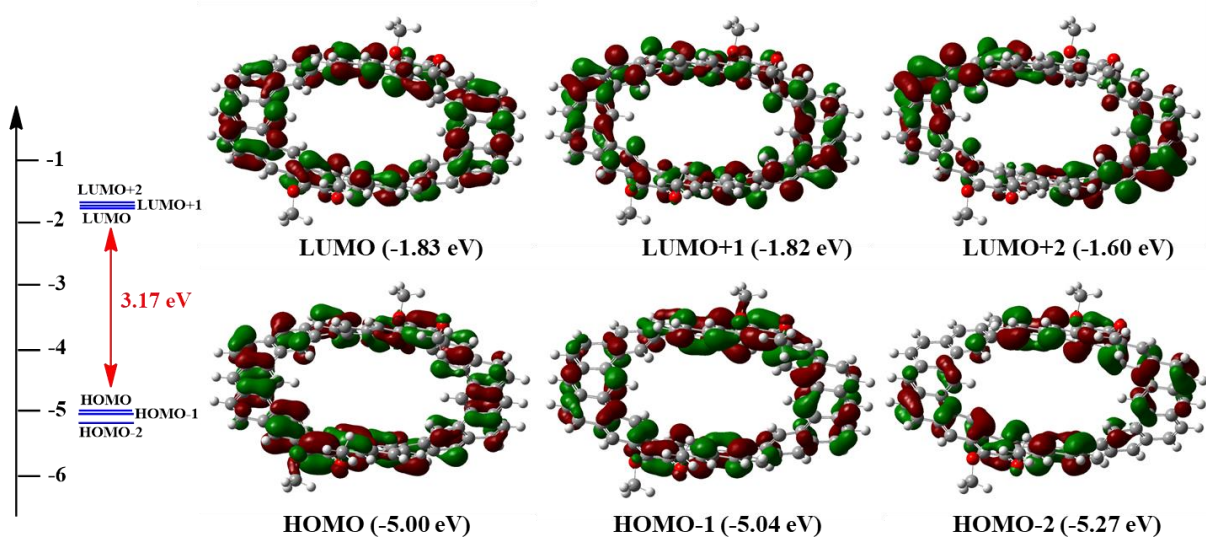


Figure S27. Frontier molecular orbital profiles and energy diagram of **1** obtained by B3LYP/6-31G(d,p) level calculation.

Table S2. Selected TD-DFT (B3LYP/6-31G(d,p)) calculated energies, oscillator strength and compositions of major electronic transitions of **1**.

Wavelength (nm)	Osc.Strength (f)	Major contributions
464.42985	0.0025	HOMO->LUMO (80%)
454.07139	0.0024	H-1->L+1 (79%)
446.53242	0.0518	H-1->LUMO (48%), HOMO->L+1 (44%)

438.75785	0.0177	H-3->LUMO (14%), H-2->L+1 (14%), H-1->LUMO (12%), HOMO->L+1 (16%), HOMO->L+2 (21%)
428.2701	0.0186	H-5->L+1 (15%), H-4->LUMO (21%), H-1->L+3 (18%), HOMO->L+4 (18%)
423.24091	0.2973	H-1->LUMO (36%), HOMO->L+1 (35%), HOMO->L+2 (12%)
416.41766	0.0236	H-3->L+1 (14%), H-2->LUMO (34%), H-1->L+2 (21%), HOMO->L+3 (15%)
412.97779	0.0024	H-5->LUMO (11%), H-2->LUMO (12%), H-1->L+4 (10%), HOMO->L+3 (34%)
402.03701	9E-4	H-2->LUMO (36%), H-1->L+2 (43%)
397.43619	0.0539	H-4->LUMO (31%), H-3->LUMO (10%), HOMO->L+2 (38%)
395.78686	0.618	H-3->LUMO (25%), H-1->L+3 (38%)
395.14355	0.1904	H-4->L+1 (31%), H-1->L+3 (10%)
393.83817	0.563	H-2->L+1 (61%)
391.24075	0.1005	H-3->L+1 (34%), HOMO->L+3 (14%)
389.76483	0.0357	H-4->LUMO (19%), H-3->LUMO (19%), HOMO->L+4 (47%)
388.82364	0.0131	H-4->L+1 (24%), HOMO->L+5 (49%)
386.62902	0.0061	H-2->L+3 (12%), H-1->L+4 (33%)
383.01008	0.0247	H-5->LUMO (14%), H-4->L+2 (10%), H-2->L+3 (17%), H-1->L+4 (23%)
381.72473	0.539	H-5->L+1 (22%), H-1->L+5 (55%)
380.56476	0.0291	H-5->LUMO (23%), H-3->L+1 (17%), H-1->L+4 (17%)
379.26094	0.6979	H-5->L+1 (40%), H-2->L+2 (15%), H-1->L+5 (10%)
378.55457	0.109	H-5->LUMO (27%), H-3->L+2 (20%), HOMO->L+5 (11%)
375.2094	1.2762	H-2->L+2 (52%), H-1->L+5 (13%)
371.4437	0.0461	H-5->L+3 (13%), H-4->L+2 (12%), H-4->L+4 (20%), H-3->L+4 (13%)
370.61097	0.0731	H-4->L+2 (10%), H-4->L+3 (51%), H-2->L+4 (11%)
368.92372	0.0457	H-3->L+3 (60%)
367.54571	0.0811	H-4->L+2 (21%), H-3->L+2 (30%), H-2->L+3 (25%)
365.36864	0.0312	H-5->L+2 (26%), H-2->L+4 (63%)
364.0277	0.0133	H-5->L+3 (21%), H-4->L+4 (11%), H-2->L+5 (43%)
362.87702	0.0333	H-5->L+2 (31%), H-3->L+5 (23%)
361.82861	0.0854	H-5->L+5 (16%), H-4->L+4 (15%), H-3->L+4 (22%), H-2->L+3 (11%), H-2->L+5 (12%)
352.56837	0.029	H-5->L+2 (13%), H-3->L+5 (56%)
349.44812	0.0079	H-4->L+5 (79%)
346.89626	0.0676	H-6->LUMO (19%), H-5->L+3 (11%), H-4->L+2 (10%), H-4->L+4 (16%), H-3->L+4 (22%)
345.44647	0.0285	H-5->L+4 (61%)
343.83703	0.1037	H-6->LUMO (17%), H-4->L+4 (11%), H-3->L+4 (16%), HOMO->L+6 (31%)
343.17084	0.0361	H-7->LUMO (14%), H-1->L+6 (27%), HOMO->L+7 (25%)
342.22361	0.0315	H-7->L+1 (10%), H-6->LUMO (11%), H-5->L+3 (14%), H-2->L+5 (12%), H-1->L+7 (16%), HOMO->L+6 (16%)
341.65778	0.0499	H-7->LUMO (27%), H-6->L+1 (26%), H-1->L+6 (17%), HOMO->L+7 (24%)
339.1067	0.0081	H-6->L+1 (46%)
338.47719	0.1287	H-6->LUMO (16%), H-5->L+5 (49%), HOMO->L+6 (12%)
336.16451	0.0072	H-7->LUMO (15%), H-7->L+1 (14%), H-1->L+6 (18%), H-1->L+7 (23%)
336.05517	0.005	H-7->LUMO (18%), H-1->L+6 (12%), H-1->L+7 (29%)
333.14755	0.1153	H-7->L+1 (46%), HOMO->L+6 (16%)
332.31712	0.0126	H-7->L+3 (11%), H-6->L+2 (24%), H-2->L+7 (16%)
329.57865	0.0108	H-6->L+4 (18%), H-5->L+7 (12%), H-4->L+6 (13%), H-3->L+6 (10%)
325.80264	0.0066	H-8->L+1 (15%), H-6->L+3 (34%), H-2->L+6 (13%)
322.41371	0.0344	H-8->L+1 (22%), HOMO->L+8 (37%)
321.38575	0.0044	H-7->L+3 (26%), H-6->L+2 (31%), H-3->L+6 (12%)
321.22754	7E-4	H-8->LUMO (39%), H-1->L+8 (22%)

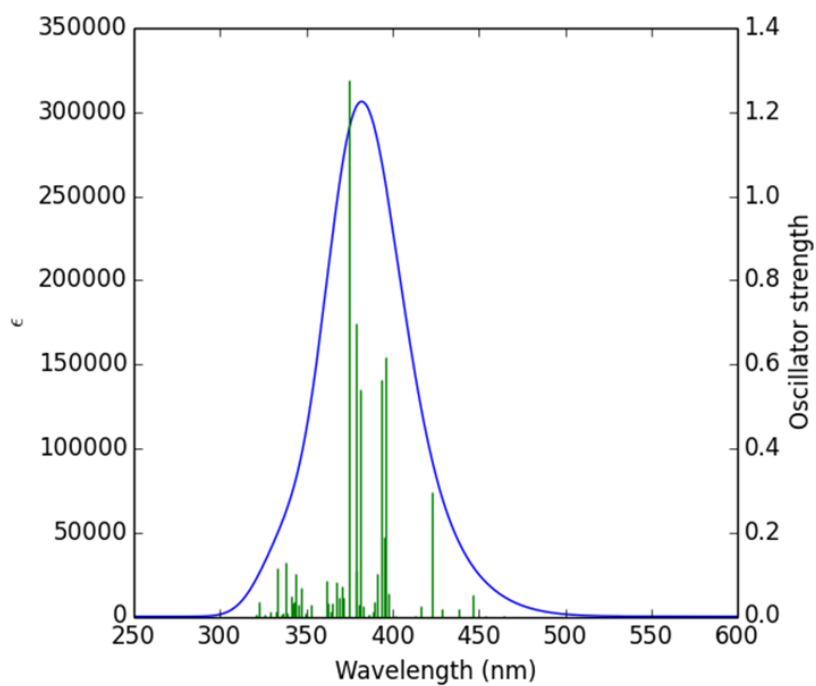


Figure S28. Calculated electronic absorption spectrum (B3LYP/6-31G(d,p)) of **1**.

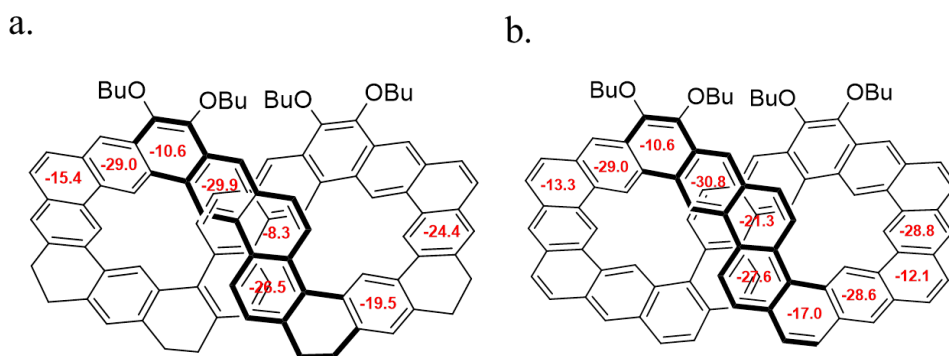


Figure S29. Calculated NICS(1)_{zz} values (ppm) of (a) **1-H** and (b) **1**.

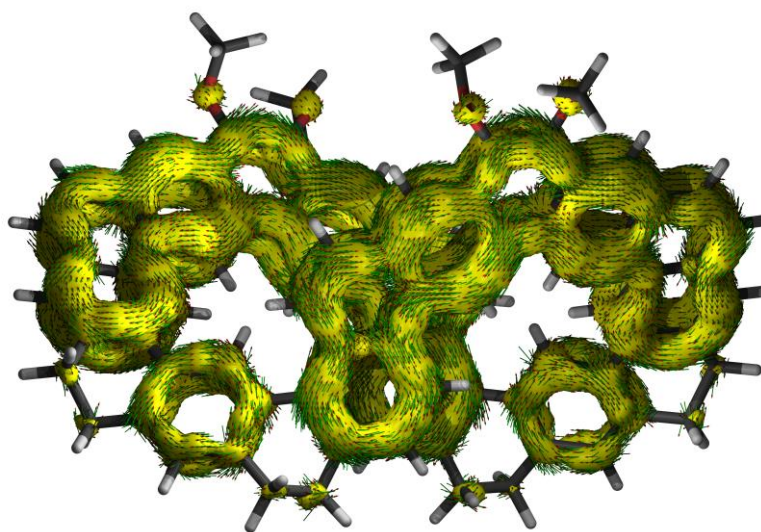


Figure S30. Calculated ACID plots of **1-H** with isovalue of 0.02. The magnetic field is pointing in through the paper.

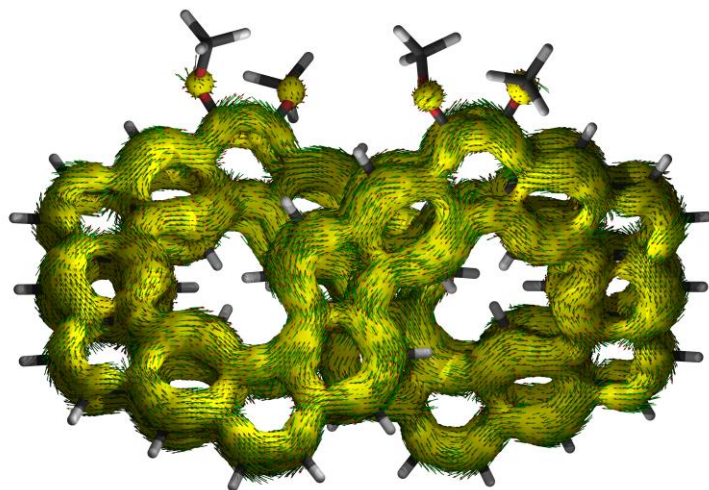


Figure S31. Calculated ACID plots of **1** with isovalue of 0.02. The magnetic field is pointing in through the paper.

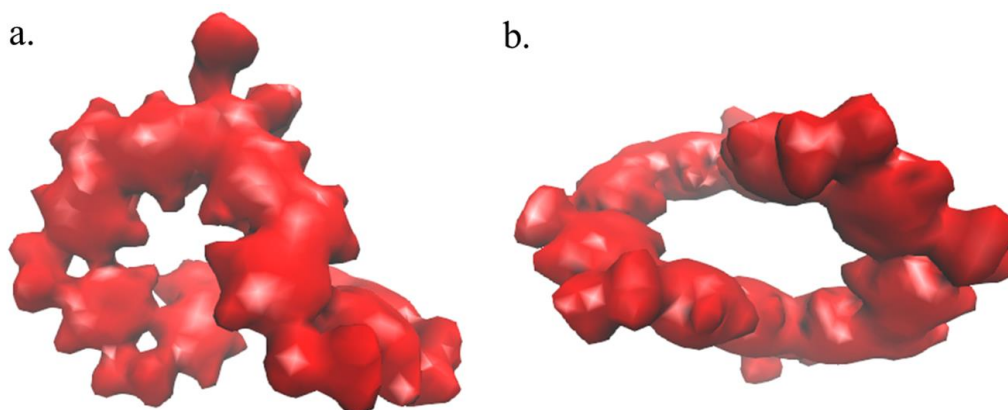


Figure S32. Calculated 3D ICSS maps of **1-H** with isovalue of 6 (left for side-view and right for top-view).

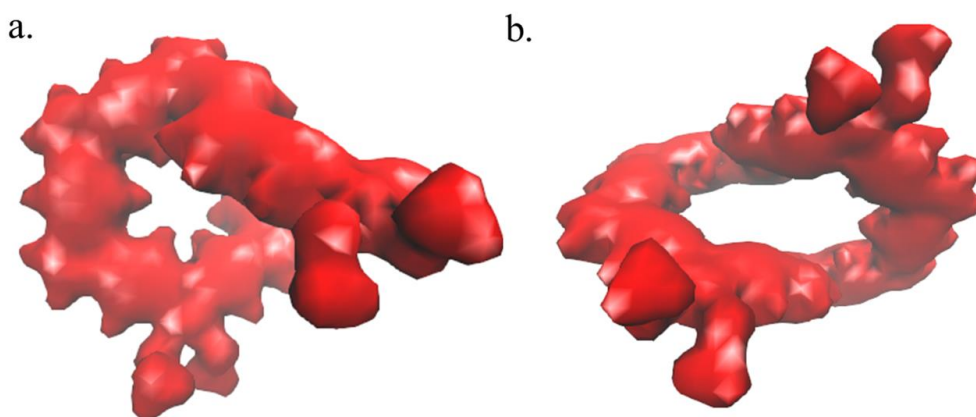


Figure S33. Calculated 3D ICSS maps of **1** with isovalue of 6 (left for side-view and right for top-view).

5. X-ray crystallographic data for 1-H

Crystallographic data have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication no. CCDC 2099885 for **1-H**. The X-ray intensity data were measured at low temperature (T=100K), using a four circles goniometer Kappa geometry, Bruker AXS D8 Venture, equipped with a Photon 100 CMOS active pixel sensor detector. Frames were integrated with the Bruker SAINT8 software package. Data were corrected for absorption effects using the multi-scan method (SADABS).⁹ Molecule was solved with the software SHELXT,¹⁰ using a Dual Space method. Refinement of the structure was performed by least squares procedures on weighted F² values using the SHELXL-version 2014/6¹¹ included in the WinGx system programs for Windows.¹² The crystallographic data were summarized in Table S3.

Table S3. Crystallographic data of **1-H**.

	1-H
Chemical formula	C ₉₈ H ₈₄ Cl ₄ O ₄
Formula weight	1476.45
Crystal system	monoclinic
Space group	-P 2yn
Calculated density	1.345 Mg/m ³
<i>a</i> (Å)	15.1951(5)
<i>b</i> (Å)	22.9211(8)
<i>c</i> (Å)	21.1399(7)
α (°)	90
β (°)	100.292(2)
γ (°)	90
Unit cell volume (Å ³)	7244.3(4)
Temperature (K)	100(2)
No. of formula units per unit cell	4
No. of reflections measured	12746
No. of independent reflections	10416
<i>R</i> _{int}	0.1049
Final R1 [<i>I</i> >2sigma(<i>I</i>)]	0.0919

6. NMR spectra and HR-mass spectra for all new compounds

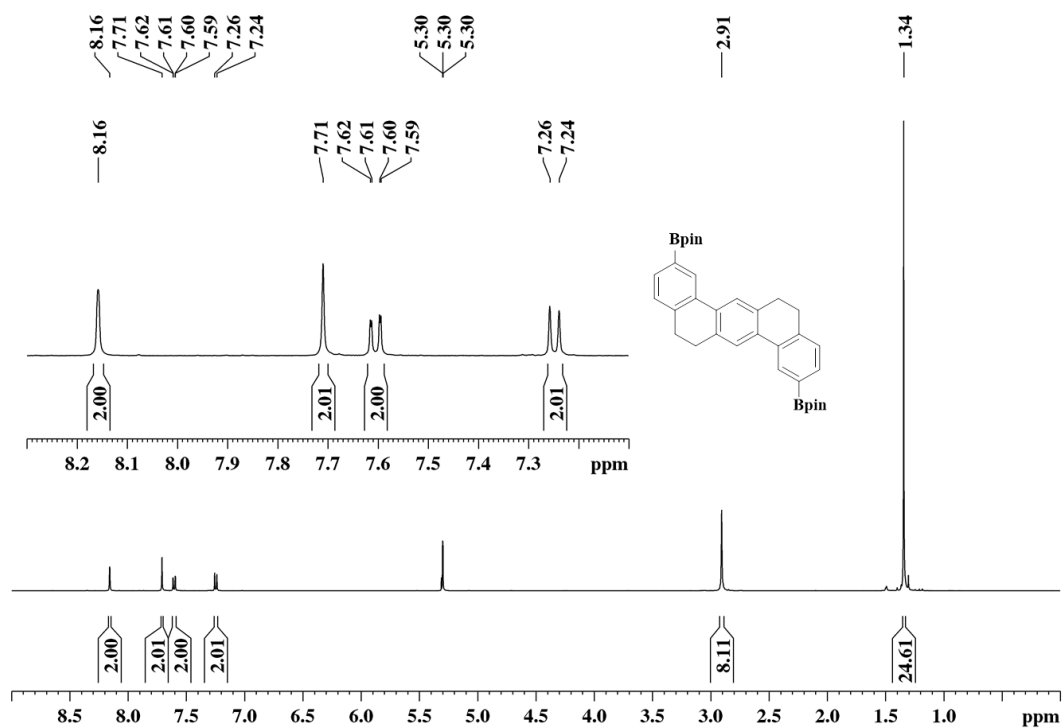


Figure S34. ¹H NMR spectrum of **5'** in CD₂Cl₂ (400 MHz, 298 K)

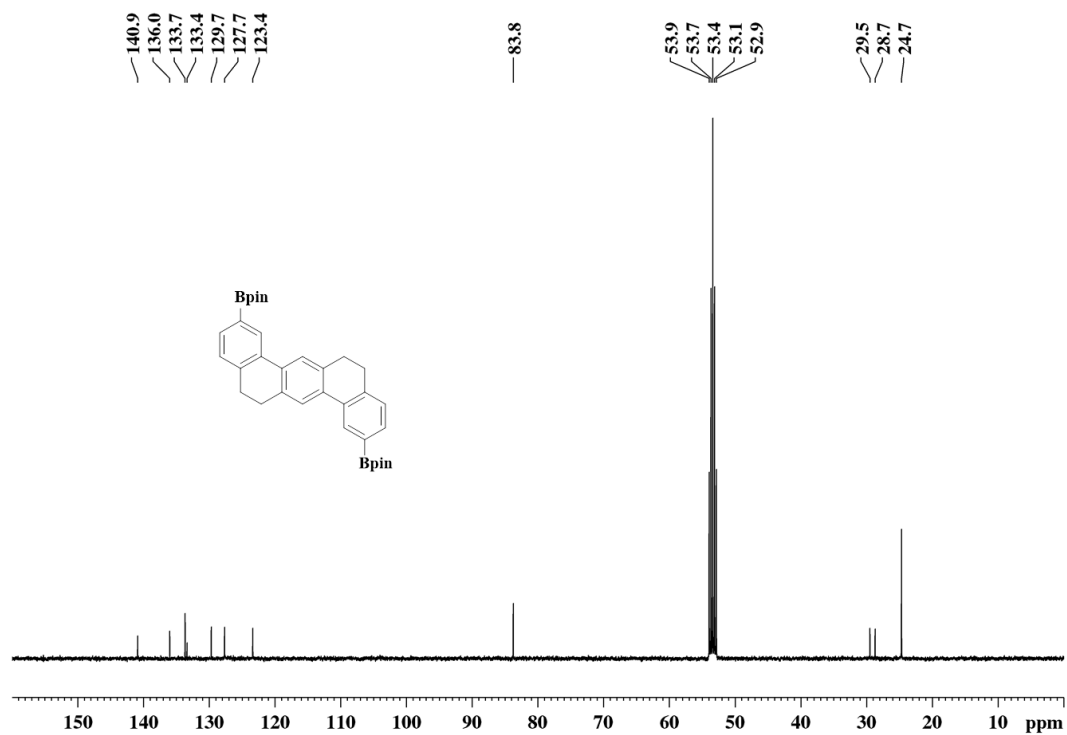


Figure S35. ¹³C NMR spectrum of **5'** in CD₂Cl₂ (100 MHz, 298 K)

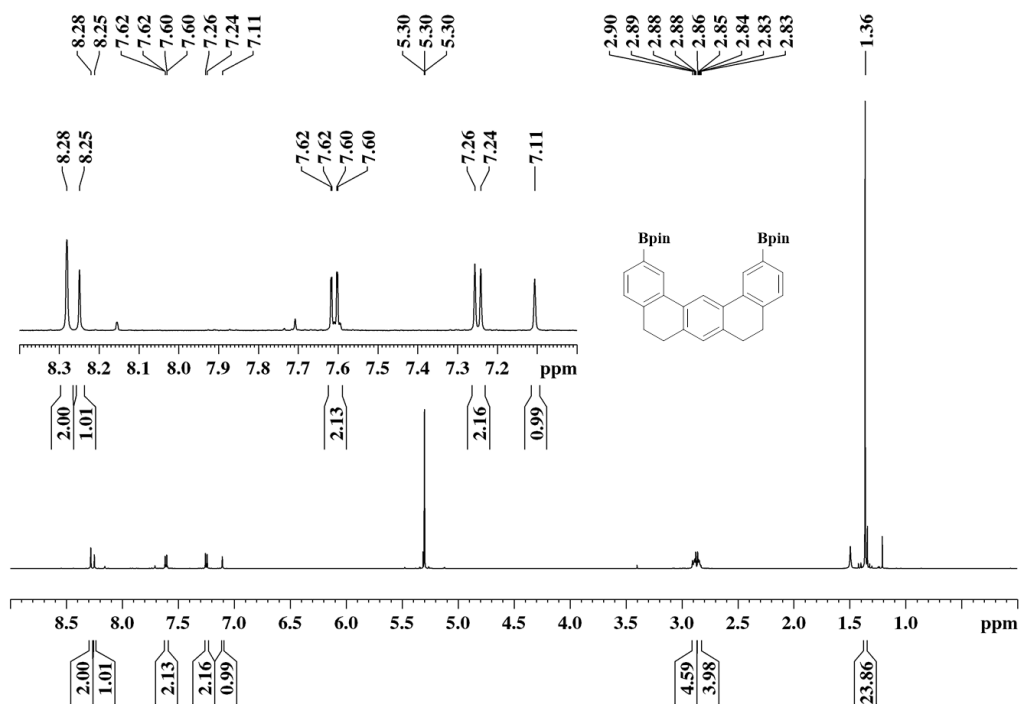


Figure S36. ¹H NMR spectrum of **5** in CD₂Cl₂ (400 MHz, 298 K)

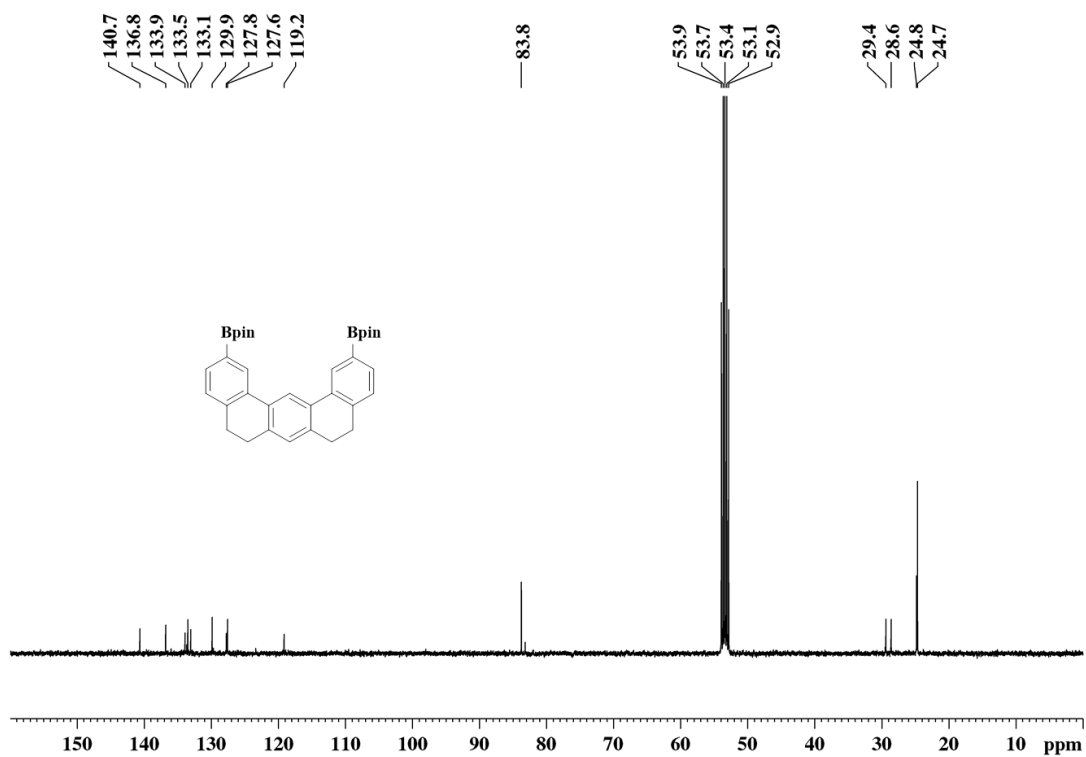


Figure S37. ¹³C NMR spectrum of **5** in CD₂Cl₂ (100 MHz, 298 K)

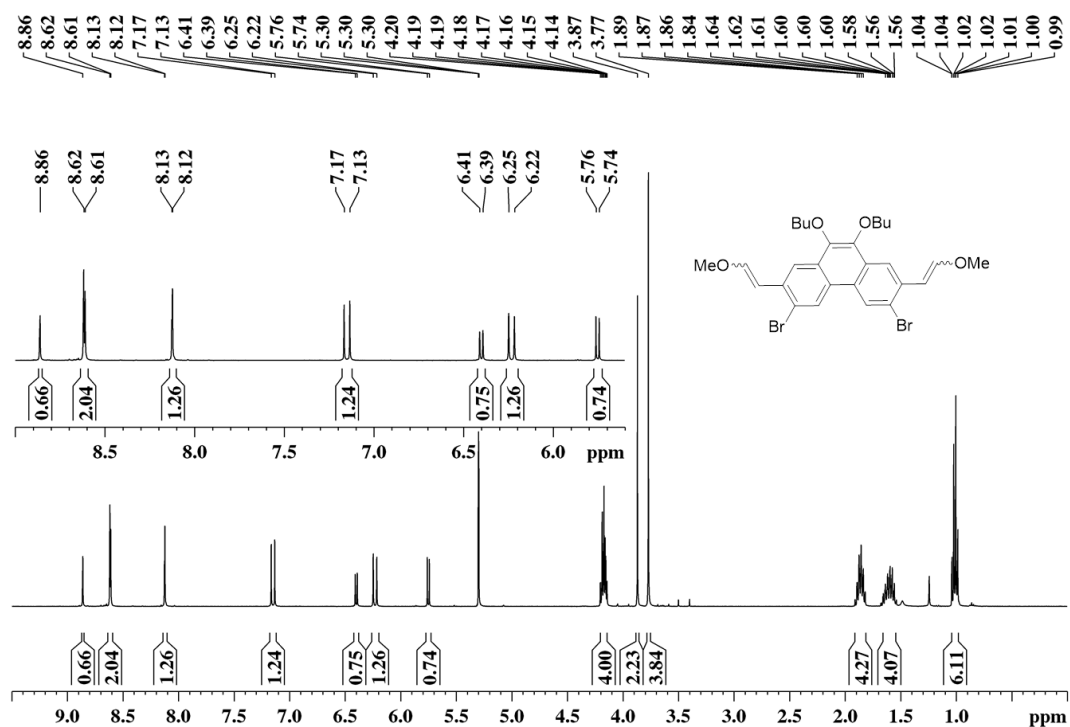


Figure S38. ¹H NMR spectrum of **6** in CD₂Cl₂ (400 MHz, 298 K)

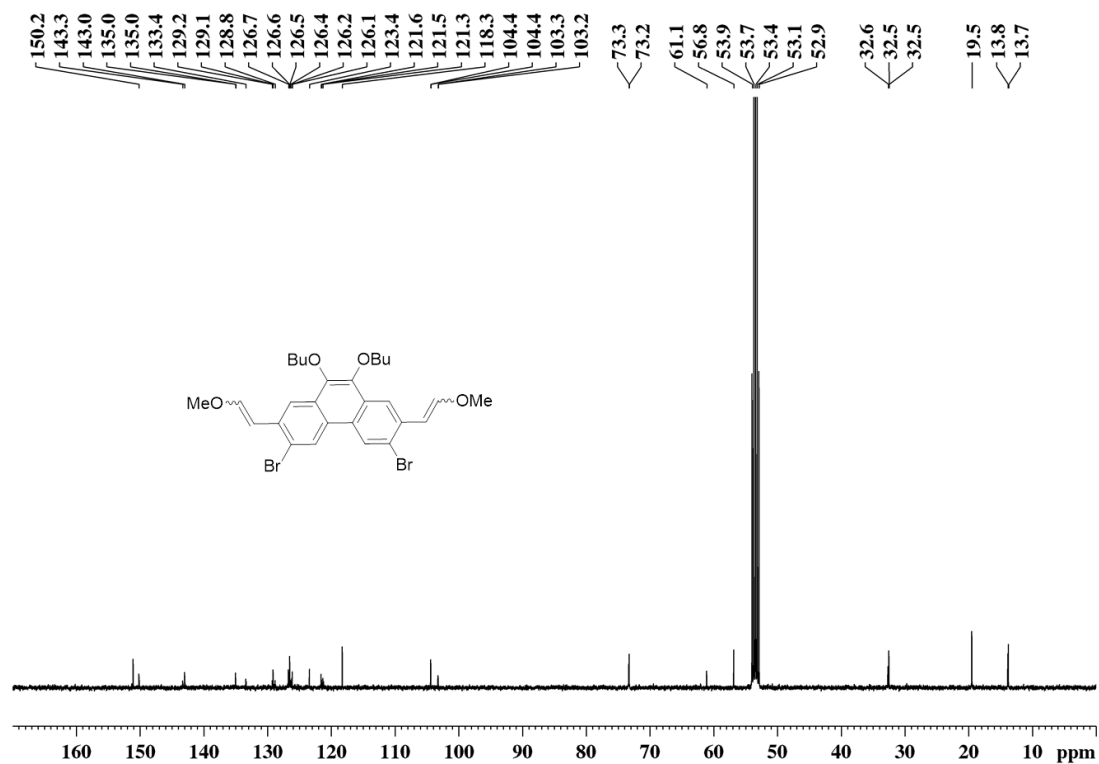


Figure S39. ¹³C NMR spectrum of **6** in CD₂Cl₂ (100 MHz, 298 K)

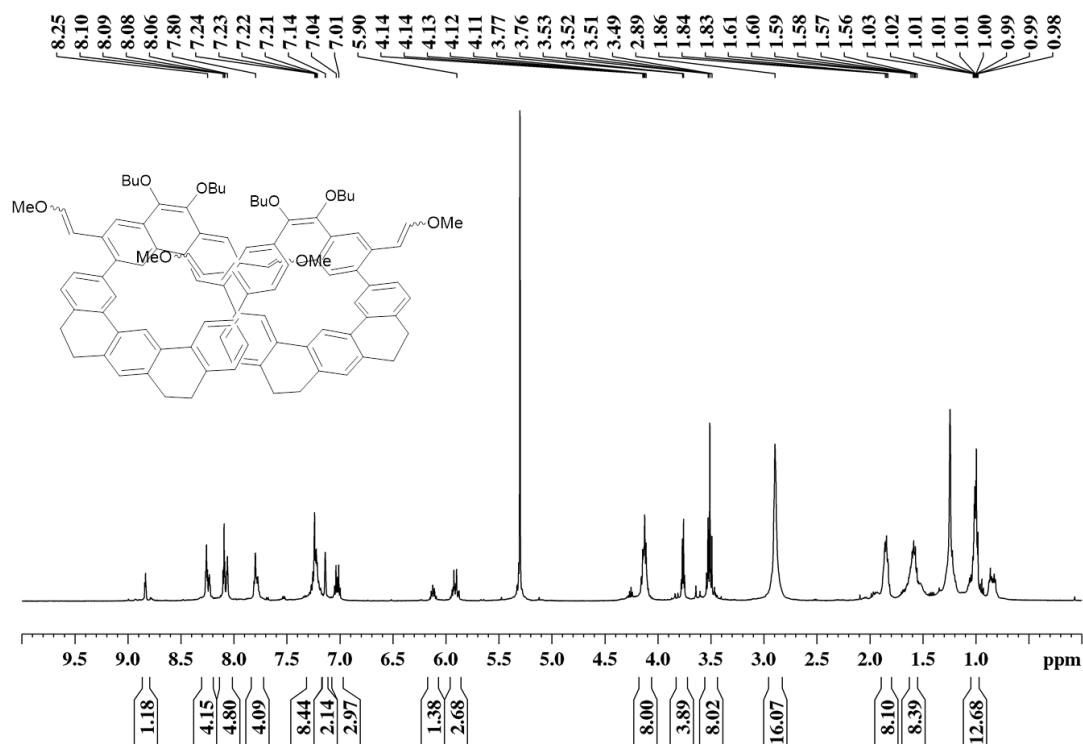
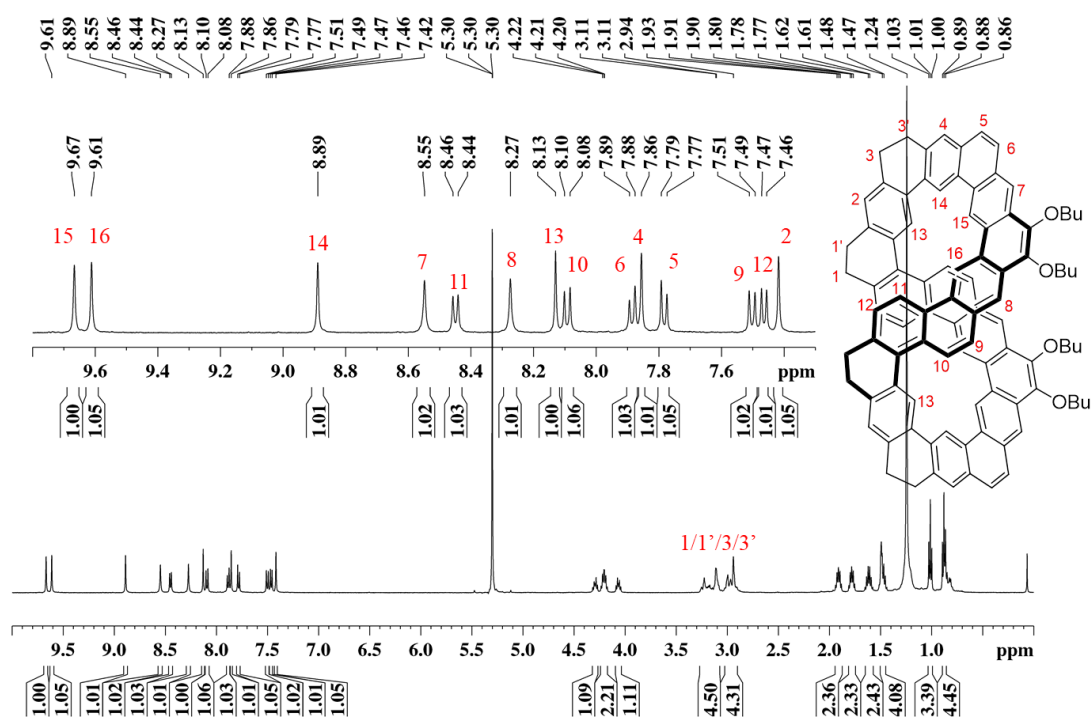


Figure S40. ¹H NMR spectrum of **7** in CD₂Cl₂ (400 MHz, 298 K)



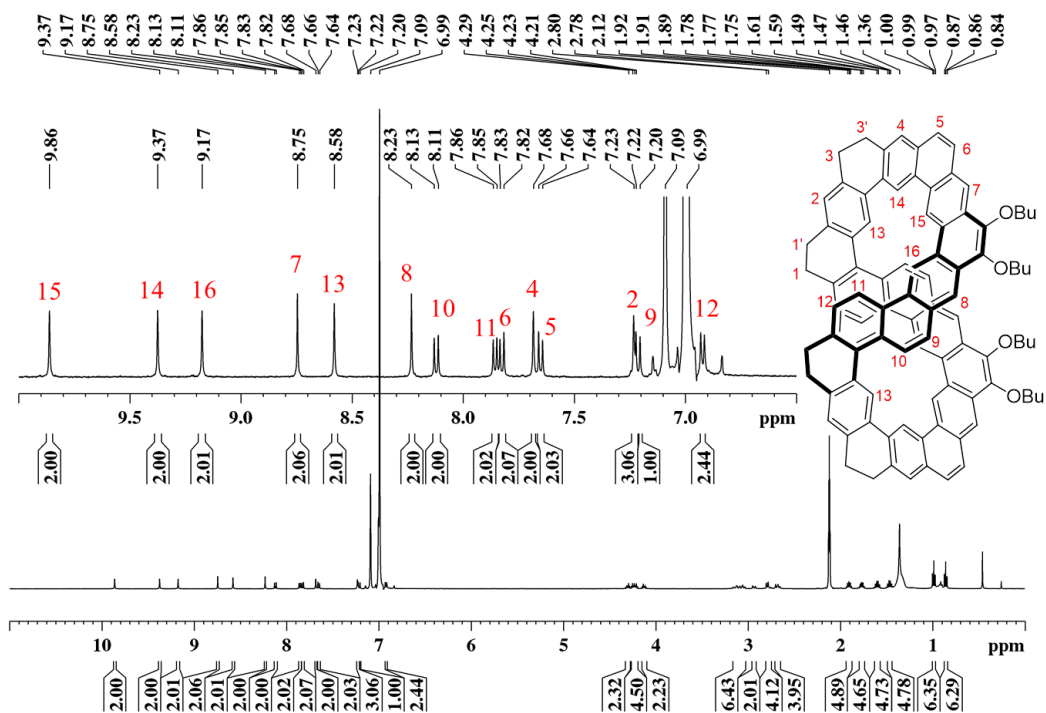


Figure S42. ¹H NMR spectrum of **1-H** in Toluene-*d*₈ (500 MHz, 373 K)

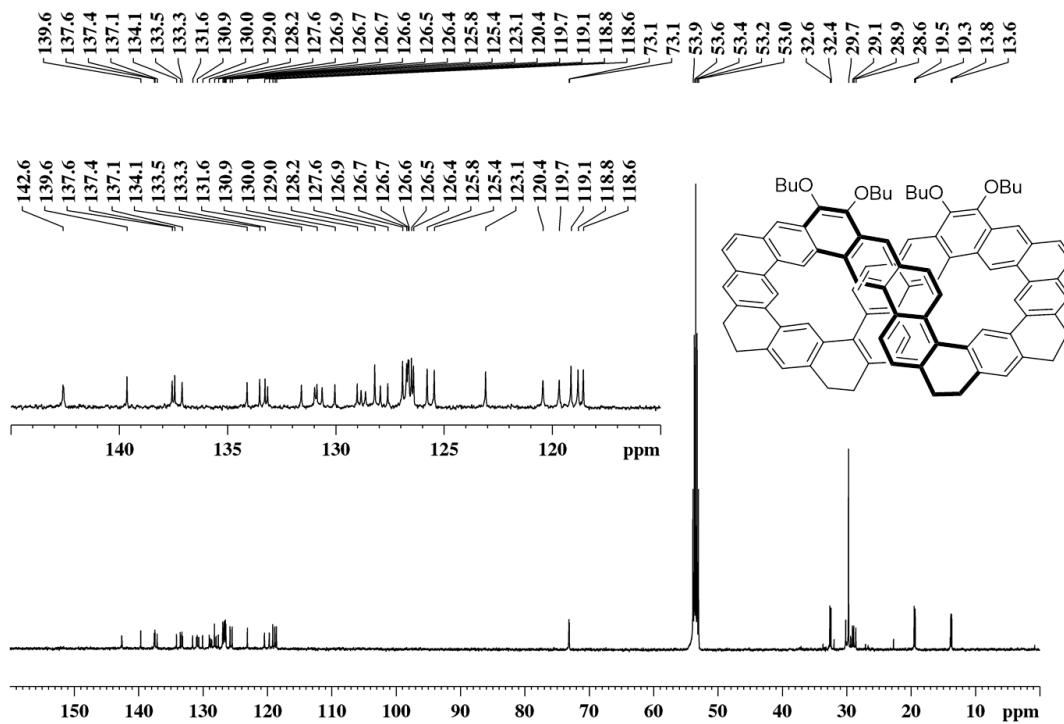


Figure S43. ¹³C NMR spectrum of **1-H** in CDCl₂ (125 MHz, 298 K)

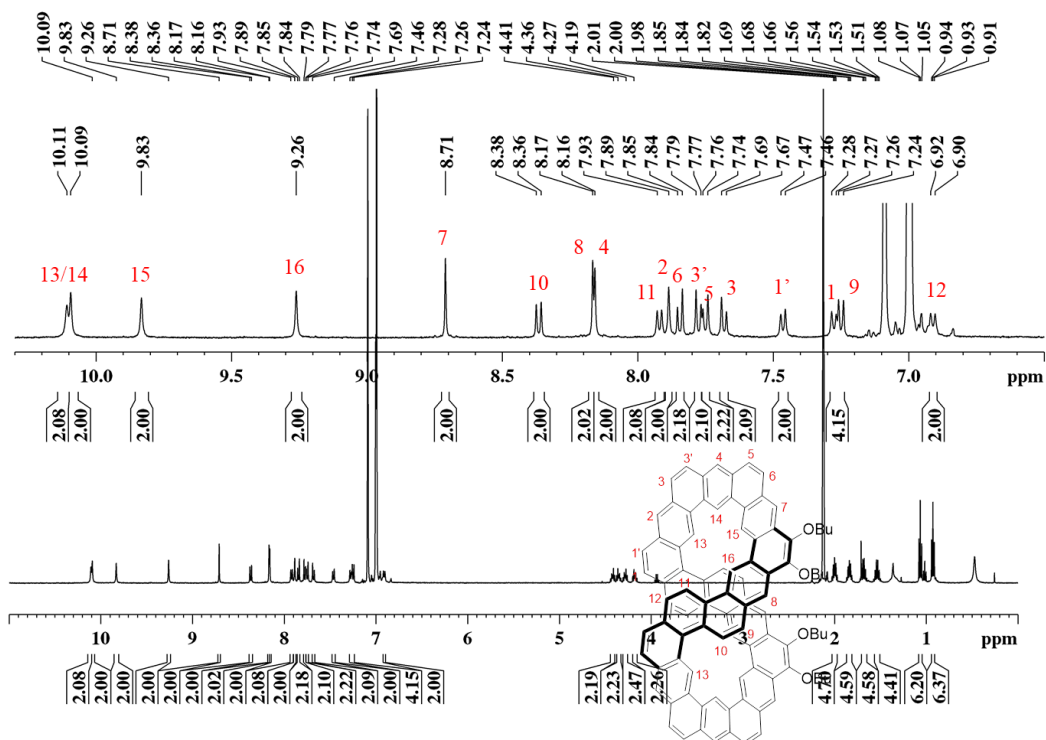


Figure S44. ¹H NMR spectrum of **1** in Toluene-*d*₈ (500 MHz, 373 K)

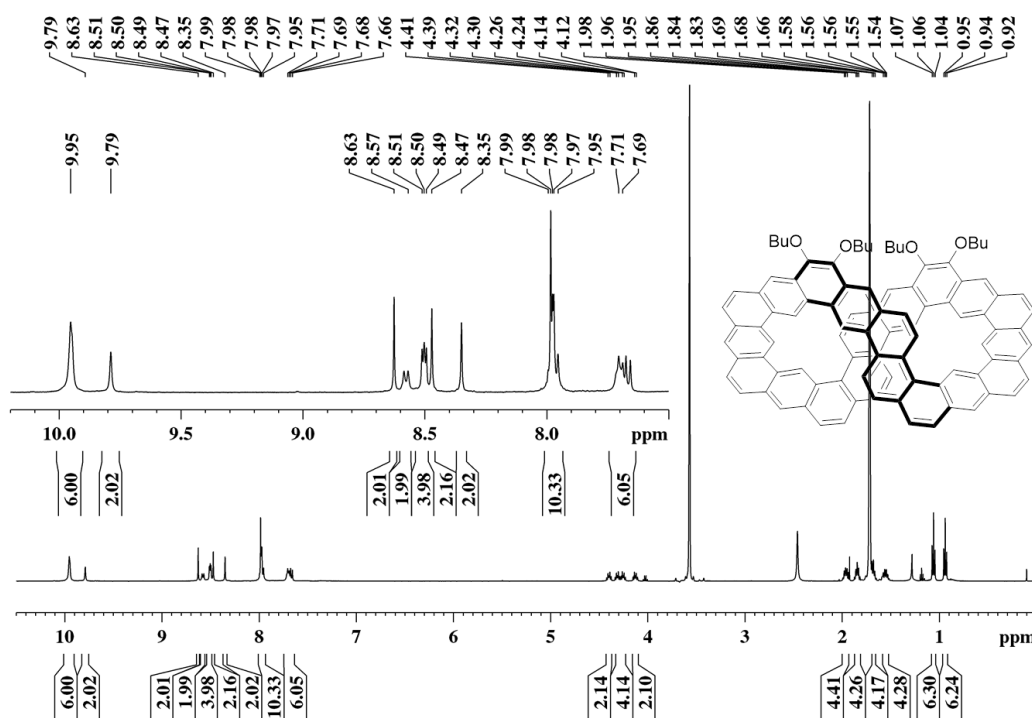


Figure S45. ¹H NMR spectrum of **1** in THF-*d*₈ (500 MHz, 298 K)

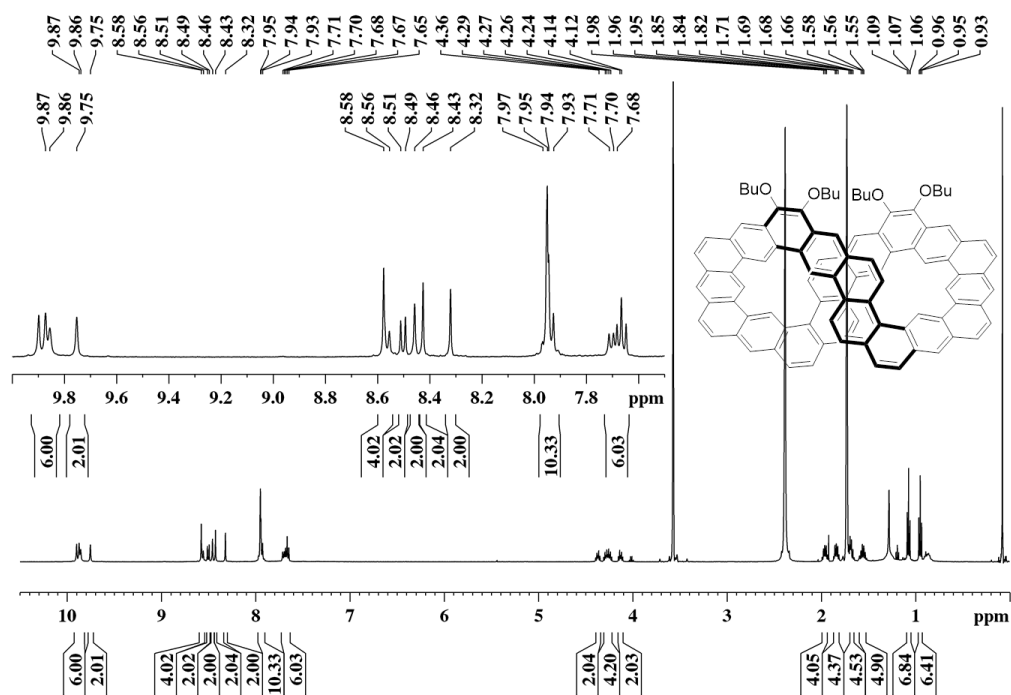


Figure S46. ¹H NMR spectrum of **1** in THF-*d*₈:CS₂ = 2:1 (500 MHz, 298 K)

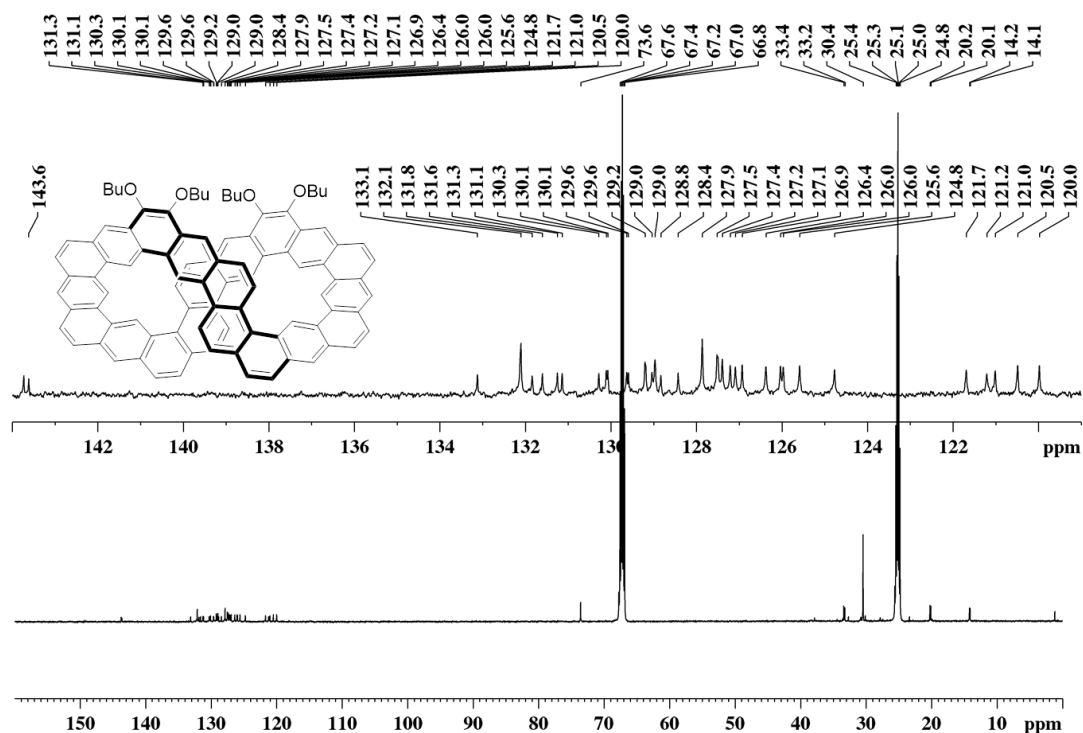


Figure S47. ¹³C NMR spectrum of **1** in Toluene-*d*₈ (500 MHz, 298 K)

MALDI,2,20210622

Analysis Info

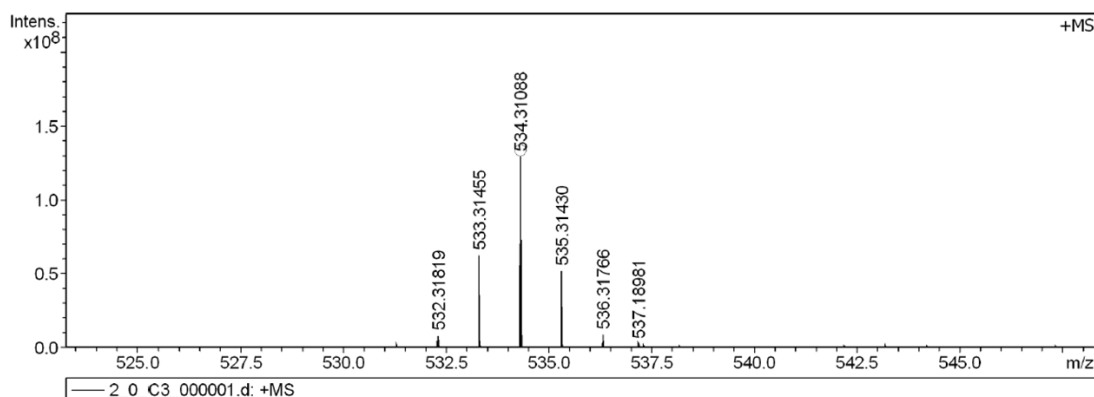
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Method MALDI_P_100-3000
Sample Name MURU-N-ESI
Comment

Acquisition Date 6/24/2021 6:15:22 PM

Operator
Instrument solariX

Acquisition Parameter

Acquisition Mode	Single MS	Acquired Scans	2	Calibration Date	Thu Jun 24 06:06:37
Polarity	Positive	No. of Cell Fills	1	Data Acquisition Size	2027152
Broadband Low Mass	202.1 m/z	No. of Laser Shots	10	Data Processing Size	4194304
Broadband High Mass	1000.0 m/z	Laser Power	55.4 lp	Apodization	Sine-Bell Multiplication
Source Accumulation	0.001 sec	Laser Shot Frequency	0.020 sec		
Ion Accumulation Time	0.100 sec				



Meas. m/z	#	Ion Formula	Score	m/z	err [ppm]	Mean err [ppm]	mSigma	rdB	e ⁻ Conf	N-Rule
534.310885	1	C34H40B2O4	100.00	534.311873	-0.3	1.1	30.7	16.0	odd	ok

Figure S48. HR mass spectrum (MALDI-TOF) of 5

MALDI,3,20210622

Analysis Info

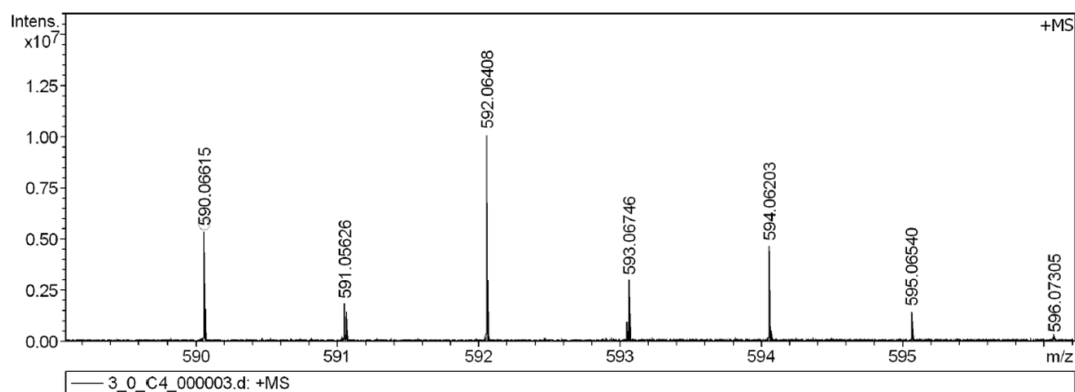
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Method MALDI_P_100-3000
Sample Name MURU-N-ESI
Comment

Acquisition Date 6/24/2021 6:18:17 PM

Operator
Instrument solariX

Acquisition Parameter

Acquisition Mode	Single MS	Acquired Scans	2	Calibration Date	Thu Jun 24 06:06:37
Polarity	Positive	No. of Cell Fills	1	Data Acquisition Size	2027152
Broadband Low Mass	202.1 m/z	No. of Laser Shots	10	Data Processing Size	4194304
Broadband High Mass	1000.0 m/z	Laser Power	48.6 lp	Apodization	Sine-Bell Multiplication
Source Accumulation	0.001 sec	Laser Shot Frequency	0.020 sec		
Ion Accumulation Time	0.100 sec				



Meas. m/z	#	Ion Formula	Score	m/z	err [ppm]	Mean err [ppm]	mSigma	rdB	e ⁻ Conf	N-Rule
590.066147	1	C28H32Br2O4	100.00	590.066185	-0.1	0.5	28.1	12.0	odd	ok

Figure S49. HR mass spectrum (MALDI-TOF) of 6

MALDI,4,20210622

Analysis Info

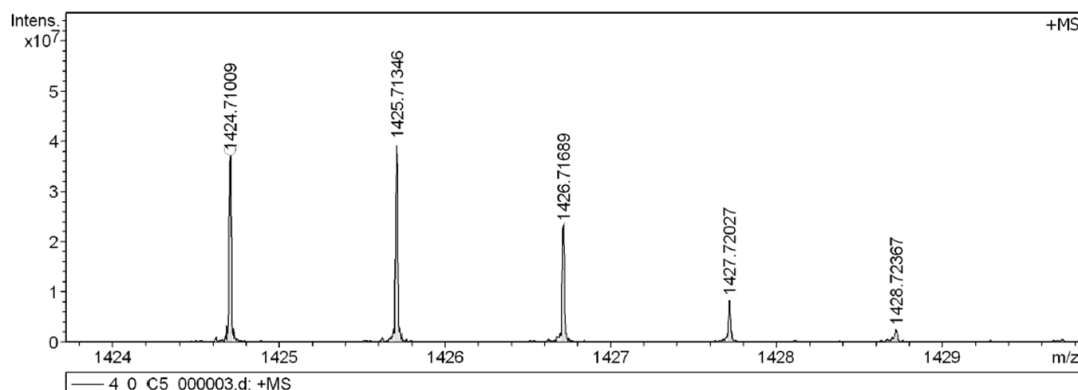
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Method MALDI_P_100-3000
Sample Name MURU-N-ESI
Comment

Acquisition Date 6/24/2021 6:20:56 PM

Operator
Instrument solariX

Acquisition Parameter

Acquisition Mode	Single MS	Acquired Scans	3	Calibration Date	Thu Jun 24 06:06:37
Polarity	Positive	No. of Cell Fills	1	Data Acquisition Size	2027152
Broadband Low Mass	202.1 m/z	No. of Laser Shots	20	Data Processing Size	4194304
Broadband High Mass	2600.0 m/z	Laser Power	52.6 lp	Apodization	Sine-Bell Multiplication
Source Accumulation	0.001 sec	Laser Shot Frequency	0.020 sec		
Ion Accumulation Time	0.100 sec				



Meas. m/z	#	Ion Formula	Score	m/z	err [ppm]	Mean err [ppm]	mSigma	rdb	e ⁻	Conf	N-Rule
1424.710088	1	C100H96O8	100.00	1424.709971	0.1	-0.1	14.5	53.0	odd		ok

Figure S50. HR mass spectrum (MALDI-TOF) of 7

MALDI,5,20210622

Analysis Info

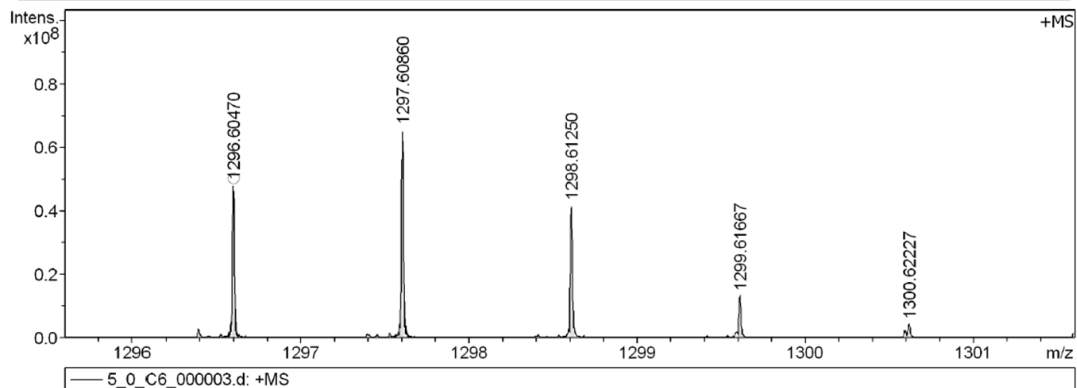
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Method MALDI_P_100-3000
Sample Name MURU-N-ESI
Comment

Acquisition Date 6/24/2021 6:23:53 PM

Operator
Instrument solariX

Acquisition Parameter

Acquisition Mode	Single MS	Acquired Scans	2	Calibration Date	Thu Jun 24 06:06:37
Polarity	Positive	No. of Cell Fills	1	Data Acquisition Size	2027152
Broadband Low Mass	202.1 m/z	No. of Laser Shots	30	Data Processing Size	4194304
Broadband High Mass	2400.0 m/z	Laser Power	52.6 lp	Apodization	Sine-Bell Multiplication
Source Accumulation	0.001 sec	Laser Shot Frequency	0.020 sec		
Ion Accumulation Time	0.100 sec				



Meas. m/z	#	Ion Formula	Score	m/z	err [ppm]	Mean err [ppm]	mSigma	rdb	e ⁻	Conf	N-Rule
1296.604698	1	C96H80O4	100.00	1296.605112	0.3	-0.2	100.5	57.0	odd		ok

Figure S51. HR mass spectrum (MALDI-TOF) of 1-H

MALDI,6,20210622

Analysis Info

Analysis Name D:\Data\MALDI\2021\0624\6_0_C7_000004.d
Method MALDI_P_100-3000
Sample Name MURU-N-ESI
Comment

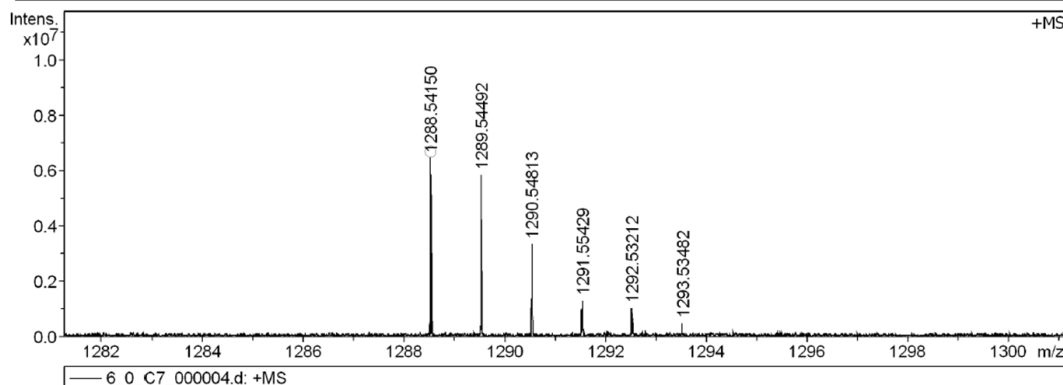
Acquisition Date 6/24/2021 6:28:09 PM

Operator

Instrument solariX

Acquisition Parameter

Acquisition Mode	Single MS	Acquired Scans	2	Calibration Date	Thu Jun 24 06:26:45
Polarity	Positive	No. of Cell Fills	1	Data Acquisition Size	2097152
Broadband Low Mass	202.1 m/z	No. of Laser Shots	10	Data Processing Size	4194304
Broadband High Mass	2400.0 m/z	Laser Power	44.4 lp	Apodization	Sine-Bell Multiplication
Source Accumulation	0.001 sec	Laser Shot Frequency	0.020 sec		
Ion Accumulation Time	0.100 sec				



Meas. m/z	#	Ion Formula	Score	m/z	err [ppm]	Mean err [ppm]	mSigma	rdb	e ⁻ Conf	N-Rule
1288.541498	1	C96H72O4	100.00	1288.542512	-0.8	0.6	43.6	61.0	odd	ok

Figure S52. HR mass spectrum (MALDI-TOF) of **1**

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